

Molecular phylogeny of European Runcinida (Gastropoda, Heterobranchia): the discover of an unexpected pool of complex species, with special reference to the case of *Runcina coronata*

ANA KARLA ARAUJO^{1,*}, MARTA POLA^{2,3}, MANUEL ANTONIO E. MALAQUIAS⁴,
MANUEL BALLESTEROS⁵, FABIO VITALE^{6,7} and JUAN LUCAS CERVERA^{1,8}

¹Departamento de Biología, Facultad de Ciencias del Mar y Ambientales, Campus de Excelencia Internacional del Mar (CEIMAR), Universidad de Cádiz, Puerto Real (Cádiz), Spain

²Departamento de Biología, Facultad de Ciencias, Universidad Autónoma de Madrid; Campus de Excelencia Internacional UAM; CSIC, Madrid, Spain

³Centro de Investigación en Biodiversidad y Cambio Global (CIBC-UAM). Campus de Excelencia Internacional UAM; CSIC, Madrid, Spain

⁴Section of Taxonomy and Evolution, Department of Natural History, University Museum of Bergen, University of Bergen, Bergen, Norway

⁵Departamento de Biología Evolutiva, Ecología I Ciències Ambientals, Facultat de Biologia, Universitat de Barcelona, Barcelona, Spain

⁶Museo di Storia Naturale del Salento, Calimera, Lecce, Italy

⁷Associazione Salento Sommerso, Lecce, Italy

⁸Instituto Universitario de Investigación Marina (INMAR), Campus de Excelencia Internacional del Mar (CEIMAR), Universidad de Cádiz, Puerto Real (Cádiz), Spain

Received 31 March 2020; revised 11 April 2021; accepted for publication 22 May 2021

Runcinida is a small heterobranch order of sea slugs with 61 known species distributed worldwide across temperate and tropical latitudes with two established families (Ilbiidae and Runcinidae). Little is known about the phylogenetic relationships within Runcinida. Here, we present the first molecular phylogeny of the order with an emphasis on European species and we discuss the taxonomic status of the type species *Runcina coronata*. Molecular phylogenetics based on the mitochondrial genes cytochrome *c* oxidase subunit I and 16S rRNA and nuclear gene histone *H3* is used to investigate relationships between species. Detailed morpho-anatomical worked was additionally employed to study *Runcina coronata*. Our results suggest the monophyly of Runcinida and showed that *Runcina coronata* is a complex of four species, namely: *R. coronata* proper, *R. aurata* and two new species here formally described (*R. caletensis* sp. nov. and *R. tingensis* sp. nov.).

ADDITIONAL KEYWORDS: biodiversity – cryptic species – Ilbiidae – Runcinidae – sea slugs – species delimitation.

INTRODUCTION

The order Runcinida Burn, 1963 includes two recognized families, namely Ilbiidae Burn, 1963 and Runcinidae H.Adams & A.Adams, 1854 (Bouchet *et al.*, 2017), with 61 valid species distributed in the Atlantic and

Mediterranean Sea (46 species) and the Indian and Pacific Ocean (15 species) (MolluscaBase, 2021). Members of this taxon are mostly characterized by the presence of an undivided dorsal shield (notum), an undivided foot, small external gills around, or to the right side of, the anus (sometimes absent), lack of parapodial lobes and four gizzard plates (Burn, 1963; Miller & Rudman, 1968; Thompson, 1976; Burn & Thompson, 1998).

The family Ilbiidae is composed of three genera: *Fofinha* Moro & Ortea, 2015, *Ilbia* Burn, 1963 and

*Corresponding author. E-mail: anakarla.araujo@uca.es

[Version of record, published online 17 July 2021; <http://zoobank.org/> urn:lsid:zoobank.org:pub:344343E1-010A-4E00-AF6F-59A3234C9361]

Pseudoilbia Miller & Rudman, 1968. Members of this family are characterized by the absence of shell, gills (*Ilbia* and *Pseudoilbia*) and/or gizzard plates (*Pseudoilbia* and *Fofinha*) and presence of a pedal furrow (*Ilbia* and *Pseudoilbia*).

The family Runcinidae includes eight genera: *Edmundsina* Ortea, 2013, *Ildica* Bergh, 1889, *Karukerina* Ortea, 2013, *Lapinura* Marcus & Marcus, 1970, *Metaruncina* Baba, 1967, *Runcina* Forbes in Forbes & Hanley, 1853, *Runcinella* Odhner, 1924 and *Runcinida* Burn, 1963. Members of this family are characterized by having a radula with a bilobed rachidian tooth, presence of gills (with up to four pinnulae) and four gizzard plates. The shell can be present or absent.

The systematics of Runcinida is confusing due to the lack of synapomorphic traits uniting all members in each family. Earlier referred by Odhner (1939) as Peltacea and subsequently named Runcinacea by Burn, 1963, the group was considered a suborder – the most ‘derived’ – within Cephalaspidea only sharing the presence of an external seminal groove (Odhner, 1939; Burn, 1963; Schmekel, 1985). Colosi (1915) first challenged the position of runcinids within Cephalaspidea and later Odhner (Odhner, 1968) suggested that Runcinacea should be ranked as an order. Recently, Bouchet et al. (2017: 330, paragraph e) in their revised *Nomenclator and typification of gastropod and monoplacophoran families* suggested the ending -ida for orders and proposed the name Runcinida to replace Runcinacea.

Over the years, many authors have continued to consider runcinids as part of Cephalaspidea (Marcus & Marcus, 1970; Kress, 1977; Schmekel, 1985; Thompson & Brodie, 1988; Vaught, 1989; Gosliner, 1990, 1991; Cervera et al., 1991; Mikkelsen, 1993, 1996; Millard, 1997; Schmekel & Cappellato, 2001, 2002), while several phylogenetic studies on Heterobranchia based on morphological (Dayrat & Tillier, 2002; Wägele & Klussmann-Kolb, 2005) and molecular data (Dayrat et al., 2001; Grande et al., 2004a, b; Vonnemann et al., 2005) questioned the inclusion of Runcinida in the Cephalaspidea. Malaquias et al. (2009), based on molecular phylogenetics, produced the first sound evidence for the inclusion of runcinids in their own order as early proposed by Odhner (in Franc, 1968). This was later supported by the molecular phylogenetic studies by Jörger et al. (2010), Wägele et al. (2014) and Oskars et al. (2015).

Most known runcinids species are concentrated in European waters (Atlantic Ocean, including the Mediterranean Sea) and belong to the genera *Pseudoilbia*, *Runcina* and *Runcinella*. Among these genera, *Runcina* is the most species-rich (31 species). However, the definition of its type species, *Runcina coronata* (Quatrefages, 1844), is problematic and

remains a matter of debate. The species *R. coronata* is reported from England southwards to the Mediterranean coast of France (Quatrefages, 1844; Alder & Hancock, 1846; Vayssière, 1883; Schmekel & Cappellato, 2002; Cervera et al., 2004; Ballesteros et al., 2016). It is defined by having an almost elliptical body with two small lobes on the front of the head and a rounded posterior end of the body. The notum is brown in colour with lighter edges and with characteristic small, dull, white spots forming a semicircle on the head zone behind the eyes and at the posterior end of the notum. Three small and slightly pinnate gills are situated on the right side of the anus (Quatrefages, 1844; Alder & Hancock, 1846; Forbes & Hanley, 1851).

Runcina coronata (as *Pelta coronata*) was described from Brehat, Bretagne (Atlantic coastline of France). Two years later, Alder & Hancock (1846) found specimens at Torbay (southern England) similar to those described by Quatrefages and also to *Limapontia nigra* Johnston, 1835, but differing from the former by the presence of gills. Alder & Hancock (1846) believed that the lack of gills in *R. coronata* was an erroneous observation by Quatrefages but, nevertheless, they did not designate any specific name for these specimens. Based on additional specimens also collected at Torbay, Forbes (1853) described the species *Runcina hancocki*, which mainly differed from *R. coronata* by the presence of external gills. Vayssière (1883), in his ‘Monographie du *Pelta*’, considered *Runcina* a synonym of *Pelta*, but in 1951, the genus name *Pelta* Quatrefages, 1844 was suppressed and only the name *Runcina* Forbes (in Forbes & Hanley, 1853) was considered valid (see Lemche, 1967: opinion n. 811).

Runcina coronata was first reported from the Mediterranean Sea (Marseille, France) by Vayssière (1883). Pruvot-Fol (1954) suggested that the Mediterranean species *R. calaritana* Colosi, 1915 was conspecific with *R. coronata* and Burn (1963), after comparing the original description of *R. coronata* and the specimens studied by Vayssière, pointed out differences in the shape of the body, coloration and number of crests in the gizzard plates, suggesting these specimens to belong to *R. calaritana*. The species *Runcina aurata* García et al., 1986 (type locality: Club La Hacienda, Bay of Algeciras, Strait of Gibraltar) was synonymized with *R. coronata* by Cervera et al. (1991) after a detailed comparison of specimens of *Runcina aurata* with the original description of *R. coronata*.

This work provides the first approach to the study of the diversity and systematics of the European species of runcinids based on a molecular phylogenetic framework. The taxonomic status of the type species *Runcina coronata* (Quatrefages, 1844) is investigated combining molecular phylogenetics and morpho-anatomical characters.

MATERIAL AND METHODS

TAXON SAMPLING

Samples were obtained from fieldwork conducted by the authors and colleagues, and through the study of museum collections. Voucher specimens are held in the collections of the Zoologische Staatssammlung München, ZSM (Munich, Germany), Museum Victoria, MV (Melbourne, Australia), Museu de Zoologia da Universidade de São Paulo, MZUSP (São Paulo, Brazil), University Museum of Bergen, ZMBN (Bergen, Norway) and Museo Nacional de Ciencias Naturales (Madrid, Spain). Seventy-seven specimens of Runcinida were included for molecular analyses, representing five genera: *Ilbia* (one specimen), *Lapinura* (two specimens), *Metaruncina* (one specimen), *Runcina* (71 specimens) and *Runcinida* (one specimen). The remaining two specimens were referred as ‘runcinid’ sp.. All the species used in this study are listed in Table 1 and were identified by comparison with primary literature. The aplysiid *Aplysia dactylomela* Rang, 1828 and the acteonoid *Micromelo undatus* (Bruguière, 1792) were used as outgroup taxa. In total, 189 novel sequences were generated during the present work and 91 were obtained from GenBank (Table 1).

For the study of the *Runcina coronata* species-complex, we included specimens from Swanage in southern England (c. 210 km from the type locality of *R. coronata*, which is Brehat in France), Cádiz in Andalusia, south-western Spain and Tangier in north-western Morocco.

DNA EXTRACTION, AMPLIFICATION AND SEQUENCING

Tissue samples were taken from the foot and DNA was extracted using the DNeasy Blood and Tissue Kit (Qiagen, Valencia, CA). Partial sequences of the mitochondrial cytochrome *c* oxidase subunit I (*COI*) and 16S rRNA (16S) and nuclear histone H3 (*H3*) genes were amplified by polymerase chain reaction (PCR) using the universal primers: LCO1490 and HCO2198 (Folmer *et al.*, 1994 for *COI*); 16S ar-L and 16br-H (Palumbi *et al.*, 1991 for 16S); and H3aF and H3aR (Colgan *et al.*, 1998 for *H3*). Polymerase chain reactions were conducted in a 25 µL reaction volume containing 1 µL of both forward and reverse primers (10 µmol/L), 2.5 µL of dNTP (2 mmol/L), a gene-dependent amount of magnesium chloride (25 mmol/L), 0.25 µL of Qiagen DNA polymerase (5 units/µL), 5 µL of ‘Q-solution’ (5×), 2.5 µL of Qiagen buffer (10×) (Qiagen Taq PCR Core Kit) and 2 µL of genomic DNA. Amplification of *COI* was performed with an initial denaturation for 5 min at 94 °C, followed by 35–36 cycles of 1 min at 94 °C, 30 s at 45 °C (annealing temperature) and 1 min at 72 °C, with a final extension of 10 min at 72 °C. The

16S amplification began with an initial denaturation for 5 min at 94 °C, followed by 35–36 cycles of 1 min at 94 °C, 30 s at 42 and 49 °C (annealing temperatures) and 1 min at 72 °C, with a final extension of 10 min at 72 °C. Amplification of *H3* was performed with an initial denaturation for 5 min at 94 °C, followed by 35 cycles of 1 min at 94 °C, 30 s at 52 °C (annealing temperature) and 1 min at 72 °C, with a final extension of 10 min at 72 °C. Successful PCR products were sent to Macrogen, Inc for purification and sequencing on a 3730XL DNA sequencer (Applied Biosystems).

PHYLOGENETIC ANALYSES

Sequences were edited in GENEIOUS v.10.2.3 (Kearse *et al.*, 2012) and aligned using MAFFT (Katoh *et al.*, 2009) implemented in GENEIOUS v.10.2.3 with default settings. Sequences from the protein-coding genes *COI* and *H3* were translated into amino acids to check for stop-codons. Hypervariable regions of the 16S alignment where homology could not be confidently established were removed using GBLOCKS under relaxed settings (Talavera & Castresana, 2007). Nevertheless, analyses including and excluding these regions provided similar results. Therefore, final analyses were performed including all nucleotides. Sequences of the *COI*, 16S and *H3* genes were trimmed to 658, 489 and 328 nucleotides, respectively. Single gene and concatenated (*H3* + *COI* + 16S) analyses were performed. Saturation for the first-, second- and third-codon positions of the *COI* and *H3* genes was calculated in MEGA X (Kumar *et al.*, 2018).

The best-fit evolutionary model for each gene was determined in jModelTest v.2.1.6 (Guindon & Gascuel, 2003; Durriba *et al.*, 2012), under the Akaike information criterion (AIC) (Akaike, 1974). The GTR+G+I model was selected for the *COI* and 16S genes, and K80+G for the *H3* gene. Bayesian inference (BI) analyses were performed in MrBayes v.3.2.1 (Ronquist & Huelsenbeck, 2003) with random starting trees and two parallel runs of 10⁷ generations. The models implemented were those estimated with jModelTest v.2.1.6. The combined dataset was partitioned among genes and the ‘unlink’ command was used to allow all parameters to vary independently within each partition. Convergence was checked in TRACER v.1.7.1 (Rambaut *et al.*, 2018) with a burn-in of 25%. Nodes with a posterior probability (PP) ≥ 0.95 (Alfaro *et al.*, 2003) were considered well supported and discussed. Maximum likelihood (ML) analysis was executed using RAxML v.8 (Stamatakis, 2014) and node support was assessed with nonparametric bootstrapping (BS) with 5000 replicates. Nodes with bootstrap values (BS) ≥ 70 (Hillis & Bull, 1993) were considered significant and were discussed. Both BI

Table 1. List of specimens used for phylogenetic analysis. (*) New sequences generated for this study. Museum abbreviations: Museum Victoria collections (NMVF), University Museum of Bergen (ZMBN), Zoologische Staatssammlung München, Germany (ZSM), Museo Nacional de Ciencias Naturales (MNCN), Museu de Zoologia da Universidade de São Paulo (MZUSP) and The Natural History Museum, London, United Kingdom (NHMUK)

Species	Preliminary Ids	Revised Ids	Locality	Voucher no.	16S	COI	H3
<i>Ilbia ilbi</i>	Burn, 1963		Hammers Haven, Australia	NMVF234189	MK323019	-	MK322992
<i>Lapinura divae</i>	(Marcus & Marcus, 1963)		Bermuda (GB)	ZMBN 82997	KJ022825	KF992195	KJ022893
<i>Lapinura divae</i>			Bermuda	ZMBN 82997	MW144769*	-	MW182586*
<i>Metaruncina nhatrangiensis</i>			Vietnam (GB)	-	KP313664	KP313668	-
<i>Metaruncina setoensis</i>	(Baba, 1954)		Okinawa, Japan	ZMBN 88231	MW144770*	MW172338*	-
<i>Pseudoilbia avellana</i>	(Schmekel & Cappellato, 2001)	<i>R. avellana</i>	Palamos, Spain	MNCN 15.05/90415	-	MW172346*	MW182594*
<i>Runcina adriatica</i>	T. Thompson, 1980		Azores Islands, Portugal	MNCN 15.05/91518	MW144774*	MW172339*	MW182587*
<i>Runcina adriatica</i>			Azores Islands, Portugal	ZMBN81756	-	MW172341*	MW182589*
<i>Runcina adriatica</i>			Azores Islands, Portugal	ZMBN81756	MW144773*	MW172344*	MW182591*
<i>Runcina adriatica</i>			Iz Island, Croatia	MNCN 15.05/88099	MK323015	MK322986	MK323036
<i>Runcina adriatica</i>			Ugljan Island, Croatia	MNCN 15.05/88100	MK322996	MK322967	MK323022
<i>Runcina adriatica</i>			Zut Island, Croatia	MNCN 15.05/88101	MK322997	MK322968	-
<i>Runcina adriatica</i>			Otranto, Italy	MNCN 15.05/88102	MK322993	MK322966	-
<i>Runcina adriatica</i>			Palamos, Spain	MNCN 15.05/90416	-	MW172340*	MW182588*
<i>Runcina adriatica</i>			Cadaqués, Spain	MNCN 15.05/88103	MK322994	MK322987	MK323020
<i>Runcina adriatica</i>			Cadaqués, Spain	MNCN 15.05/90417	MW144771*	MW172342*	-
<i>Runcina adriatica</i>			Cadaqués, Spain	MNCN 15.05/90418	MW144772*	MW172343*	MW182590*
<i>Runcina adriatica</i>			Otranto, Italy	MNCN 15.05/90666	MW144775*	MW172345*	MW182592*
<i>Runcina cf. adriatica</i>			Lecce, Italy	Pending number	MW144778*	MW172348*	MW182595*
<i>Runcina cf. adriatica</i>			Lecce, Italy	MNCN 15.05/91515	MW144802*	MW172377*	MW182621*
<i>Runcina cf. adriatica</i>			La Strea, Italy	MNCN 15.05/91516	MW144817*	MW172391*	MW182634*
<i>Runcina cf. adriatica</i>			Ugljan Island, Croatia	MNCN 15.05/91501	MW144810*	MW172384*	MW182627*
<i>Runcina cf. adriatica</i>			Ugljan Island, Croatia	MNCN 15.05/91502	MW144811*	MW172385*	MW182628*
<i>Runcina cf. adriatica</i>			Ugljan Island, Croatia	MNCN 15.05/91503	MW144812*	MW172386*	MW182629*
<i>Runcina africana</i>	Pruvot-Fol, 1953		Tenerife, Canary Islands, Spain	NHMUK 20030791/1	KJ022780	DQ974680	KJ022942
<i>Runcina africana</i>			Port Lligant, Spain	MNCN 15.05/90651	MW144776*	-	MW182593*

Table 1. Continued

Species Preliminary Ids	Revised Ids	Locality	Voucher no.	16S	COI	H3
<i>Runcina cf. bahiensis</i> Cervera et al., 1991	<i>R. hornae</i>	Cadaqués, Spain	MNCN 15.05/88104	-	MK322988	MK323005
<i>Runcina brenkoeae</i> T. Thompson, 1980		Split, Croatia	MNCN 15.05/88086	MK322998	MK322969	MK323023
<i>Runcina brenkoeae</i>		Nin, Croatia	MNCN 15.05/88089	MK323017	MK322972	MK323038
<i>Runcina ferruginea</i>		Nin, Croatia	MNCN 15.05/88088	MK323018	MK322971	MK323039
<i>Runcina ferruginea</i>		Lece, Italy	MNCN 15.05/91509	-	MW172399*	-
<i>Runcina ferruginea</i>		Lece, Italy	MNCN 15.05/91510	-	MW172400*	MW182641*
<i>Runcina ferruginea</i>		Lece, Italy	MNCN 15.05/91511	-	MW172401*	-
<i>Runcina ferruginea</i>		Lece, Italy	MNCN 15.05/91512	-	MW172402*	-
<i>Runcina ferruginea</i>		Lece, Italy	MNCN 15.05/91513	MW144787*	-	MW182644*
<i>Runcina ferruginea</i>		Iz Island, Croatia	MNCN 15.05/90667	MW144788*	MW172361*	MW182607*
<i>Runcina ferruginea</i>		Ugljan Island, Croatia	MNCN 15.05/90668	MW144786*	MW172359*	MW182605*
<i>Runcina ferruginea</i>		Split, Croatia	MNCN 15.05/90669	-	MW172360*	MW182606*
<i>Runcina ferruginea</i>		Banyuls-sur-Mer, France	ZSMML20142074	-	MW172374*	MW182618*
<i>Runcina hansbechi</i> Schmekel & Cappellato, 2001	<i>R. brenkoeae</i>	Banyuls-sur-Mer, France	ZSMML20142071	MW144789*	MW172362*	MW182608*
<i>Runcina cf. hansbechi</i>	<i>R. avellana</i>	Mataró, Spain	MNCN 15.05/88108	MK323028	MK322973	MK32300
<i>Runcina hornae</i> Schmekel & Cappellato, 2002		Palamós, Spain	MNCN 15.05/88110	MK323033	MK322978	-
<i>Runcina cf. hornae</i>		Iz Island, Croatia	MNCN 15.05/91504	MW144802*	-	MW182622*
<i>Runcina lusitanica</i> Araujo et al., 2019		Faro, Portugal	MNCN 15.05/88091	MK323024	MK322979	MK322999
<i>Runcina lusitanica</i>		Faro, Portugal	MNCN 15.05/200065	-	MK322981	MK323002
<i>Runcina lusitanica</i>		Faro, Portugal	MNCN 15.05/88093	-	MK322965	MK323004
<i>Runcina brenkoeae</i>		Roses, Spain	MNCN 15.05/88087	MK323001	MK322970	MK323026
<i>Runcina brenkoeae</i>		Roses, Spain	MNCN 15.05/88090	MK323003	MK322964	-
<i>Runcina brenkoeae</i>		Roses, Spain	MNCN 15.05/90422	-	MW172349*	MW182596*
<i>Runcina brenkoeae</i>		Banyuls-sur-Mer, France	ZSMML20142072	MW144777*	MW172347*	-
<i>Runcina coronata</i> (Quatrefages, 1844)		Swanage, England	MNCN 15.05/88105	MK323031	MK322976	MK323010
<i>Runcina coronata</i>		Swanage, England	MNCN/ADN 118951	-	-	-
<i>Runcina coronata</i>	<i>R. aurata</i>	Cádiz, Spain	MNCN 15.05/90423	-	MW172356*	MW182602*
<i>Runcina coronata</i>	<i>R. aurata</i>	Cádiz, Spain	MNCN 15.05/88106	MN057638	-	MN057641
<i>Runcina coronata</i>	<i>R. aurata</i>	Cádiz, Spain	MNCN 15.05/88107	MN057639	MN057640	MN057642

Table 1. Continued

Species Preliminary Ids	Revised Ids	Locality	Voucher no.	16S	COI	H3
<i>Runcina coronata</i>	<i>R. aurata</i>	Cádiz, Spain	MNCN 15.05/91500	MW144785*	MW172358*	MW182604*
<i>Runcina coronata</i>	<i>R. aurata</i>	Cádiz, Spain	MNCN/ADN 118948	MW144780*	MW172351*	MW182597*
<i>Runcina coronata</i>	<i>R. aurata</i>	Cádiz, Spain	MNCN/ADN 118950	-	MW172353*	MW182599*
<i>Runcina coronata</i>	<i>R. caletensis</i> sp. nov	Cádiz, Spain	MNCN/ADN 118949	MW144781*	MW172352*	MW182598*
<i>Runcina coronata</i>	<i>R. caletensis</i> sp. nov	Cádiz, Spain	MNCN 15.05/200113	MW144783*	MW172355*	MW182601*
<i>Runcina coronata</i>	<i>R. tingensis</i> sp. nov	Tangier, Morocco	MNCN 15.05/91514	MW144782*	MW172354*	MW182600*
<i>Runcina coronata</i>	<i>R. tingensis</i> sp. nov	Tangier, Morocco	MNCN 15.05/200114	MW144784*	MW172357*	MW182603*
<i>Runcina ferruginea</i> Kress, 1977		Newlyn, Cornwall, England	MNCN 15.05/88109	MK323032	MK322977	MK323011
<i>Runcina ferruginea</i>		Roses, Spain	MNCN 15.05/91517	MW144779*	MW172350*	-
<i>Runcina ferruginea</i>		Lecce, Italy	MNCN 15.05/91508	-	MW172397*	MW182639*
<i>Runcina lusitanica</i>		Faro, Portugal	MNCN 15.05/88092	MK323035	MK322980	MK323014
<i>Runcina lusitanica</i>		Faro, Portugal	MNCN 15.05/88094	MK323027	MK322989	-
<i>Runcina lusitanica</i>		Faro, Portugal	MNCN 15.05/90424	MW144790*	MW172363*	MW182609*
<i>Runcina lusitanica</i>		Faro, Portugal	MNCN 15.05/90425	MW144791*	MW172364*	MW182610*
<i>Runcina marcosi</i> Araujo et al., 2019		Roses, Spain	MNCN 15.05/88098	MK323025	MK322982	MK323000
<i>Runcina marcosi</i>		Mataro, Spain	MNCN/ADN 118952			
<i>Runcina marcosi</i>		Roses, Spain	MNCN 15.05/88095	MK323037	MK322983	MK323016
<i>Runcina marcosi</i>		Roses, Spain	MNCN 15.05/88097	-	MK322990	MK323012
<i>Runcina marcosi</i>		Roses, Spain	MNCN 15.05/88096	MK323034	MK322984	MK323013
<i>Runcina marcosi</i>		Cádiz, Spain	MNCN 15.05/200066	-	MK322985	MK323006
<i>Runcina marcosi</i>		Banyuls-sur-Mer, France	ZSM MOL 201442089	MK323021	MK322991	MK322995
<i>Runcina marcosi</i>		Roses, Spain	MNCN 15.05/90426	-	MW172393*	-
<i>Runcina marcosi</i>		Roses, Spain	MNCN 15.05/90427	MW144819*	MW172394*	MW182636*
<i>Runcina ornata</i> (Quatrefages, 1844)		Azores Islands, Portugal	ZMBN87929	MW144796*	MW172368*	-
<i>Runcina ornata</i>		Azores Islands, Portugal	ZMBN87949	MW144795*	MW172367*	-
<i>Runcina</i> sp. 3	<i>R. hornae</i>	Mataro, Spain	MNCN 15.05/90654	MW144818*	MW172392*	MW182635*
<i>Runcina</i> sp. 4	<i>R. hornae</i>	Palamos, Spain	MNCN/ADN 118954	-	MW172395*	MW182637*
<i>Runcina</i> sp. 8	<i>R. hornae</i>	Roses, Spain	MNCN 15.05/90655	MW144801*	MW172376*	MW182620*
<i>Runcina</i> sp. 10	<i>R. hornae</i>	Mataro, Spain	MNCN 15.05/90656	MW144822*	MW172404*	MW182643*
<i>Runcina</i> sp. 11	<i>R. hornae</i>	Mataro, Spain	MNCN 15.05/90657	MW144821*	MW172398*	MW182640*

Table 1. Continued

Species Preliminary Ids	Revised Ids	Locality	Voucher no.	16S	COI	H3
<i>Runcina</i> sp. 13	<i>R. hornae</i>	Mataro, Spain	MNCN 15.05/90658	-	MW172403*	MW182642*
<i>Runcina</i> sp. 15	<i>R. hornae</i>	Cadaqués, Spain	MNCN 15.05/90659	-	MW172369*	MW182614*
<i>Runcina</i> sp. 17	<i>R. adriatica</i>	Cadaqués, Spain	MNCN 15.05/90419	MW144798*	MW172371*	MW182616*
<i>Runcina</i> sp. 18	<i>R. hornae</i>	Cadaqués, Spain	MNCN 15.05/90660	MW144797*	MW172370*	MW182615*
<i>Runcina</i> sp. 19	<i>R. avellana</i>	Roses, Spain	MNCN/ADN 118953	MW144805*	MW172379*	-
<i>Runcina</i> sp. 19	<i>R. avellana</i>	Roses, Spain	MNCN 15.05/90652	MW144806*	MW172380*	MW182624*
<i>Runcina</i> sp. 20	<i>R. hornae</i>	Roses, Spain	MNCN 15.05/90661	MW144813*	MW172387*	MW182630*
<i>Runcina</i> sp. 20	<i>R. hornae</i>	Roses, Spain	MNCN 15.05/90662	MW144814*	MW172388*	MW182631*
<i>Runcina</i> sp. 20	<i>R. hornae</i>	Roses, Spain	MNCN 15.05/90663	MW144815*	MW172389*	MW182632*
<i>Runcina</i> sp. 21	<i>R. hornae</i>	Zut Island, Croatia	MNCN 15.05/91505	MW144816*	MW172390*	MW182633*
<i>Runcina</i> sp. 22		Zut Island, Croatia	MNCN 15.05/91506	MW144807*	MW172381*	-
<i>Runcina</i> sp. 22		Zut Island, Croatia	MNCN 15.05/91507	MW144809*	MW172383*	MW182626*
<i>Runcina</i> sp. 24	<i>R. hornae</i>	Roses, Spain	MNCN 15.05/90664	MW144820*	MW172396*	MW182638*
<i>Runcina</i> sp. 26	<i>R. avellana</i>	Roses, Spain	MNCN 15.05/90665	MW144808*	MW172382*	MW182625*
<i>Runcina</i> sp. 31	<i>R. adriatica</i>	Es Caials (SP)	MNCN 15.05/90420	MW144792*	MW172365*	MW182611*
<i>Runcina</i> sp. 33	<i>R. hornae</i>	Port Lligant (SP)	MNCN 15.05/90665	MW144793*	-	MW182612*
<i>Runcina</i> sp. 34	<i>R. adriatica</i>	Port Lligant (SP)	MNCN 15.05/90421	MW144794*	MW172366*	MW182613*
<i>Runcinida marisae</i> Chernyshev, 1998		Vostok Bay, Russia	-	-	KP313667	-
<i>Runcinida valentinae</i> Chernyshev, 2006		Iturup Island, Russia	-	-	KP313666	-
<i>Runcinida</i> sp. 1		Cap Ferret, France	MNCN 15.05/90670	MW144800*	MW172375*	MW182619*
<i>Runcinid</i> sp. 1		Alagoas, Brazil	MZUSP103407	MW144799*	MW172373*	-
<i>Lapinura</i> sp. 1		Maui, Hawaii	MNCN 15.05/90671	MW144804*	MW172378*	MW182623*
<i>Runcinid</i> sp. 2		Quirimbas, Mozambique	ZMBN105119	-	MW172372*	MW182617*
<i>Aplysia dactylomela</i> Rang. 1828		Cape Verde (GB)	NHMUK 20030795/20030796	KJ022921	KF992168	KJ022798
<i>Micromelo undatus</i> (Bruguère, 1792)		Tenerife, Canary Island (GB)	NHMUK 20030800	KJ022944	DQ974653	KJ022778

and ML trees were visualized in FigTree v.1.4.3 (<http://tree.bio.ed.ac.uk/software/figtree/>). Minimum and maximum pairwise uncorrected *p*-distances of *COI* were calculated in MEGA X (Kumar *et al.*, 2018).

SPECIES DELIMITATION

For species delimitation analyses, the Automatic Barcode Gap Discovery (ABGD) (Puillandre *et al.*, 2012) and Bayesian Poisson Tree Processes (bPTP) (Zhang *et al.*, 2013) methods were used. For the ABGD analyses we used the fast-evolving *COI* gene using the ingroup sequences obtained in this study and those available in GenBank. We used the default setting ($P_{\min} = 0.001$, $P_{\max} = 0.1$, Steps = 10, $X = 1.2$, Nb bins = 20) under the three models of evolution, namely Jukes–Cantor (JC69), Kimura (K80) and Simple Distance. bPTP analyses were run with default parameters using the *COI* trees at the webserver (<https://species.h-its.org/ptp/>) (Zhang *et al.*, 2013).

MORPHOLOGY

The external morphology of the specimens used in our molecular phylogenies were, in most cases, studied based on photographs of live specimens (175 images) and from living animals collected during sampling activities. In the latter case, examination of the shape and coloration was carried out under a stereomicroscope. In order to identify all specimens, we compared these observations with original descriptions of species and specialized literature.

For the internal morphology, the animals were dissected either dorsally or ventrally, and the buccal bulbs and gizzards were extracted and dissolved in a solution of 10% sodium hydroxide to expose and clean them. The radulae and gizzard plates were then immersed in water, dried and mounted for scanning electron microscopy (SEM) with a Nova NanoSEM 450 available at the University of Cadiz (Cadiz, Spain). The reproductive systems were dissected out of the animals and examined and drawn using a stereomicroscope equipped with a camera lucida. All specimens previously identified as *R. coronata* used for morphological comparison are listed in the ‘Examined material’ in the Systematic description.

RESULTS

PHYLOGENETIC ANALYSES

The combined dataset yielded a sequence alignment of 1475 positions. We obtained 189 new sequences, 64 for *H3*, 68 for *COI* and 57 for 16S (Table 1). No saturation was observed in the *COI* and *H3* genes,

not even in the third-codon positions (not shown). The combined tree (*H3* + *COI* + 16S) provided better resolution than *H3*, *COI* or 16S separately (see Supporting Information, Figs S1–S3). The results of ML and BI trees rendered similar topologies, but bootstrap values were lower than posterior probabilities in some clades (Fig. 1).

Both ML and BI supported the monophyly of the order Runcinida (PP = 1; BS = 99), with the species *Ilbia ilbi* Burn, 1963 (family Ilbiidae) sister to Clade A (family Runcinidae). Clade A (PP = 0.96; BS = -) is divided into two main sister subclades, here called Clade B (PP = 0.98, BS = - [no value was recovered]) and Clade C (PP = 1, BS = 78).

Clade B was subdivided into three well-supported clades: the first, Clade D, including only the specimen labelled as runcinid sp. 2 from Mozambique; the second, Clade E (PP = 1; BS = 96), containing *Metaruncina setoensis* (Baba, 1967) and *Metaruncina nhatrangensis* Chernyshev, 2005; and the third, Clade F (PP = 1; BS = 99), with most of *Runcina* specimens and *Pseudoilbia avellana* (Schmekel & Cappellato, 2001). Within Clade F, *Runcina ornata* appears as sister to the remaining *Runcina* specimens. Within the latter, three clades with maximum support correspond to well-established species, such as *R. lusitanica* Araujo *et al.*, 2019, *R. marcosi* Araujo *et al.*, 2019 and the type species, *R. coronata*. However, specimens attributed to *Runcina adriatica* Thompson, 1980, *Runcina ferruginea* Kress, 1977, *Runcina hansbechi* Schmekel & Cappellato, 2001 and *Runcina africana* Pruvot-Fol, 1953 branched off in different clades. In addition, specimens early identified as *Runcina* cf. *bahensis* Cervera *et al.*, 1991 and *Runcina hornae* Schmekel & Cappellato, 2002 clustered together with several unidentified species from Spain (PP = 1; BS = 100).

Clade C (PP = 1; BS = 78) was subdivided into two clades: Clade G, containing specimens identified as *Runcina ferruginea* (PP = 1; BS = 100); and Clade H (PP = 1; BS = 80) with two subgroups, one including *Lapinura divae* (Marcus & Marcus, 1970), *Lapinura* sp. 1, specimens identified as *R. ferruginea* and runcinid sp. 1 (PP = 1; BS = 82), and the other with *Runcinida marisae* Chernyshev, 1998, *Runcinida valentinae* Chernyshev, 2006 and *Runcinida* sp. 1 (PP = 1; BS = 90).

SPECIES DELIMITATION

The ABGD analyses identified 31 groups with all three models of evolution implemented (Fig. 1A). Within these groups and for all models, the specimens identified as *R. coronata* (type species of the genus *Runcina*) split in four distinct groups: RC1, RC2, RC3

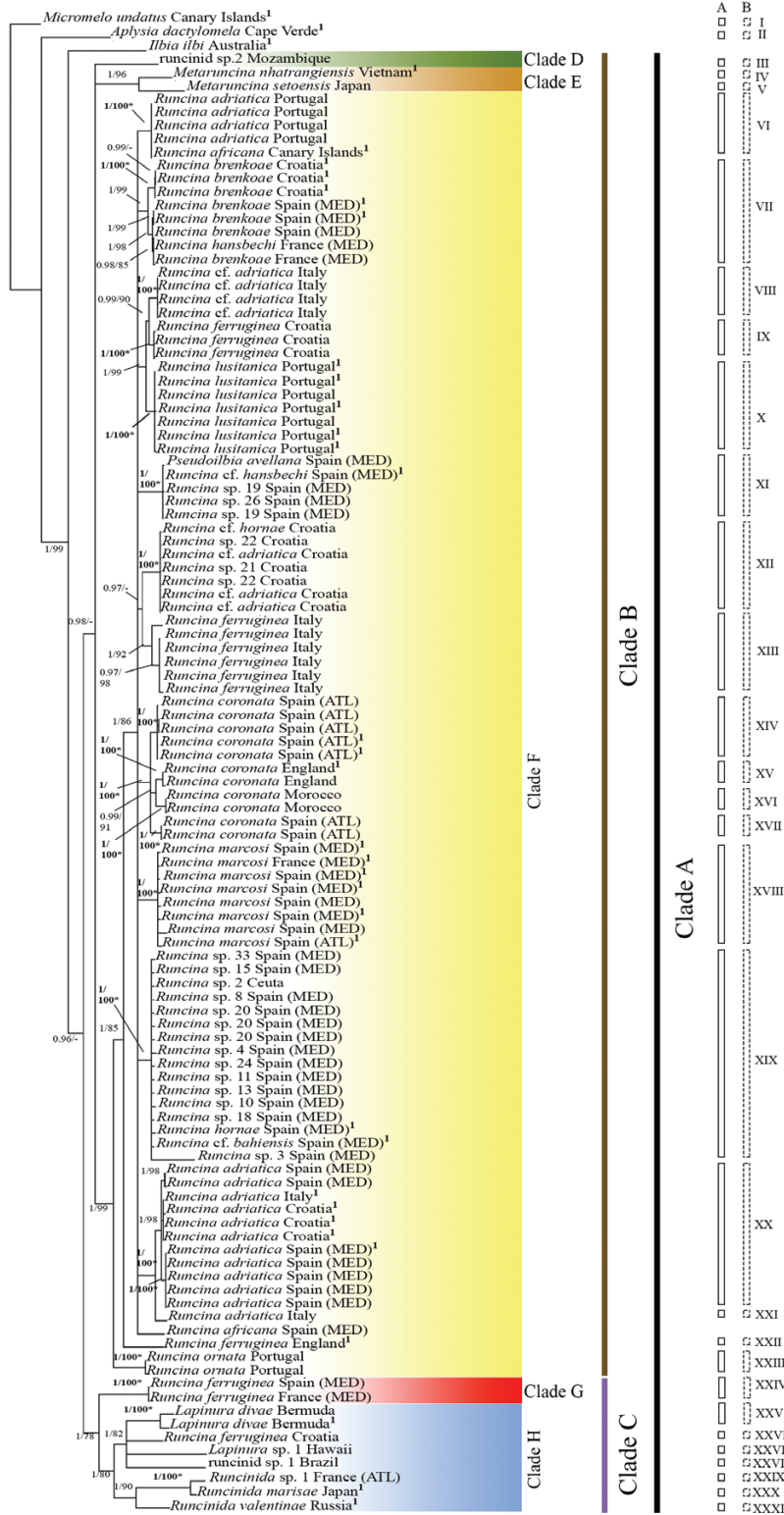


Figure 1. Phylogenetic hypothesis of the order Runcinida based on the combined genes *H3*, *COI*, and 16S inferred by Bayesian analysis. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood analysis. A, ABGD results based on the *COI* dataset. B, bPTP results based on the *COI* dataset. Rectangles in *Ilbia ilbi* are missing since there is no *COI* sequence available. Abbreviations: ATL, Atlantic Ocean; MED, Mediterranean Sea. *, branches with maximum support. 1, refers to sequences from Genbank.

and RC4 (Fig. 2). The bPTP analysis showed the same result (Figs 1B, 2).

The minimum pairwise uncorrected *p*-distances for *COI* among groups of Runcinida recovered by ABGD are presented in Table 2, and ranged from 5.6% to 23.1%. Within the genus *Runcina*, distances ranged from 6% to 17.7% between species (Table 2, groups VI–XXIII; see Fig. 1). Regarding the specimens originally identified as *R. coronata*, the distances between the four groups (Fig. 2) were considerably higher compared with the distance between specimens within each group (Table 3). Groups RC1 (*Runcina aurata*) and RC4 (*Runcina caletensis* sp. nov.) contain specimens from the same locality (Cádiz, Spain) and the minimum distance between them was 7.8%. Comparing both groups with group RC2 (*Runcina coronata*), which contains specimens from Swanage, England, the minimum distances were 6.3% for RC1 and 8.8% for RC4. Finally, the minimum genetic distances when these three groups are compared with group RC3 (*Runcina tingensis* sp. nov.; specimens from Morocco) were 6.4% (RC1), 7.0% (RC2) and 9.9% (RC4) (Table 3).

Our morphological studies (see theme Systematic description below) and molecular results support four different species among the specimens originally identified as *Runcina coronata*. We present a redescription of *R. coronata* and *R. aurata*, and the description of the two new species.

SYSTEMATIC DESCRIPTION

FAMILY RUNCINIDAE H. ADAMS & A. ADAMS, 1854

(FIGS 3A, B, 4A–C, 5A, 6A)

Runcina Forbes in Forbes & Hanley, 1851.

Runcina coronata (Quatrefages, 1844).

Synonymy

Pelta coronata Quatrefages, 1844: 151, pl.3, fig. IV.

Runcina hancocki Forbes (in Forbes & Hanley), 1851: 611, pl. C.C.C, fig. 2.

R. calaritana Colosi, 1915: 1, figs 1–18.

Type locality: Bréhat, Bretagne, France (48°50'59"N; 2°59'47"W).

Examined material: MNCN 15.05/88105, Swanage (50°36'28"N; 1°56'45"W), southern England, coll. Ian F. Smith, Apr 2016, 3 mm living animal, found by brushing the bases of stones with encrusting pink coralline algae (dissected and sequenced). MNCN 15.05/90423, Swanage (50°36'28"N; 1°56'45"W), southern England, coll. Ian F. Smith, Apr 2016, 4 mm living animal, found by brushing the bases of stones

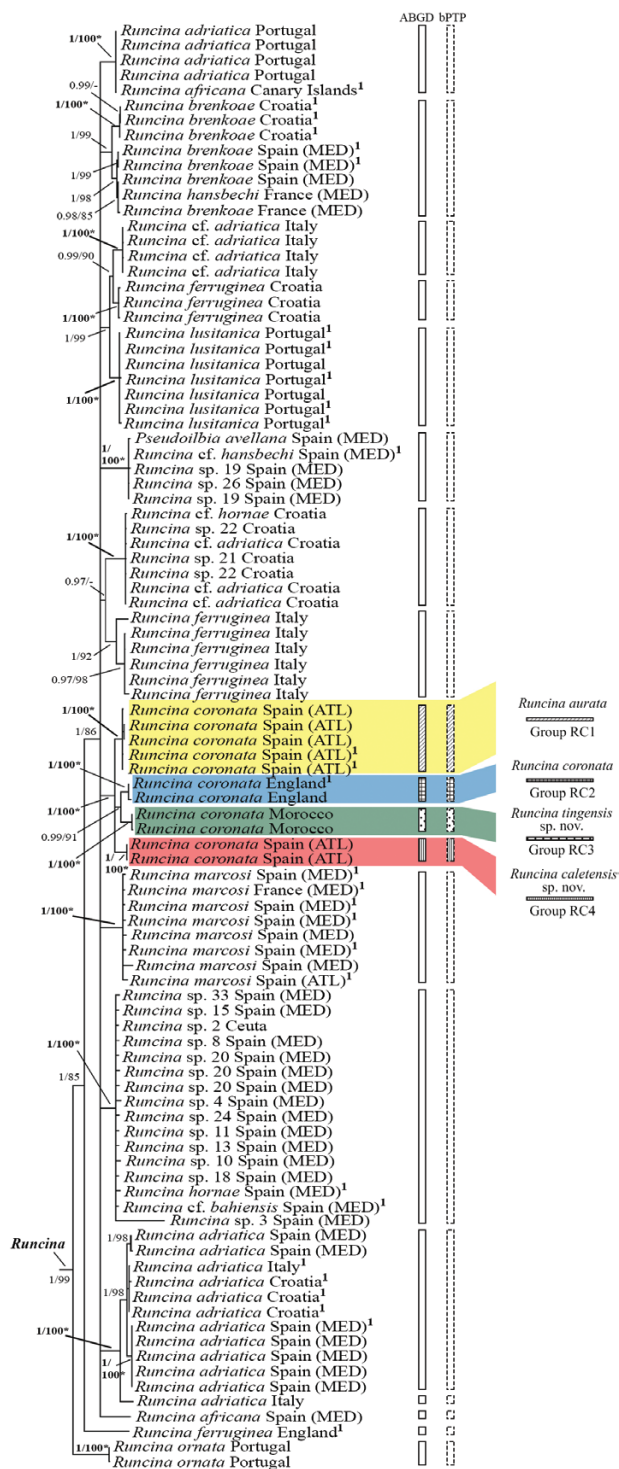


Figure 2. *Runcina coronata* species-complex. Detail of Clade F extracted from the analyses illustrated in Figure 1. Continuous rectangles, ABGD analysis based on the *COI* dataset. Dotted rectangles, bPTP analysis based on the *COI* dataset. *, branches with maximum support. 1, refers to sequences from Genbank.

Table 2. Uncorrected *p*-distances based on the *COI* gene within the order Runcinida. Roman numbers are based on species delimitation groups represented in Figure 1

	III	IV	V	VI	VII	VIII	IX	X	XI	XII	XIII	XIV	XV	XVI
IV	15.5													
V	18	15.8												
VI	12.2-12.7	16.7-18.3	16.6-16.9											
VII	14.9-15.4	17.4-18.3	17.6-18.5	8.2-9										
VIII	14.5-15.2	18.1-19.1	16.8-17.6	9.8-10.9	9.9-12.2									
IX	12.7-13.4	16.7-17.1	15.8-16.1	7.6-8.8	7.7-10.7	5.6-7								
X	15.7-16.9	17.5-18.4	18-19.5	9.1-10.4	9.9-12.4	7.8-9.8	6.1-7.8							
XI	15.5-15.9	17.9-18.7	16.7-17.4	9.8-11	9.8-13	11.1-12.7	9-10.5	11.1-13						
XII	15.9-16.4	17.1-17.5	16.7-17.5	10.6-11.4	10.2-12.2	11.2-12.6	9.4-10.2	10.2-12.5	10.3-11.3					
XIII	14-15	16.2-17.1	17.8-18.5	7.2-9.3	7.2-11.1	9.6-11.7	9-10.5	8.8-11.5	9.6-11.3	8.4-10.4				
XIV	15.4-15.5	18.3-18.7	16.9-17	10.5-10.9	8.9-10.6	11.3-13.1	9.4-10.5	11.8-13.2	11.5-12.2	11.1-12	9.4-12			
XV	15.7	18.9-19.7	16.7-17.4	10.5-11.6	9.3-11.1	11.2-12.3	9.1-10.2	9.9-11	10.9-11.8	10.9-12.4	10.4-12.6	6.3-7.3		
XVI	16	19-19.8	18.4	10.8-12.2	9.2-11.4	12.7-13.4	10.5-11.6	11.8-12.9	11-12.3	12.7-13.7	10.8-12.6	6.4-7.8	7-7.7	
XVII	17	19.7	17.8-17.9	11.7-12.3	10-12.3	13.1-13.7	9.5-10	12.3-13.4	13.4-14.5	12.8-13.7	12.6-13.7	7.8-8	8.8-9.5	9.9-10.2
XVIII	15.6-17.6	17.8-19.4	17.2-19.7	9-11.3	10.2-13.1	10.5-13.6	9.5-11.8	11.6-13.9	11.4-13.8	10.9-14.4	9.4-11.8	11.6-13.5	12.3-14.6	12-13.7
XIX	15.1-15.7	18.2-19	16.7-17.8	10.4-11.2	10.1-12.3	10.2-11.7	8.4-9.9	9.9-12.4	10.7-12.4	11.1-13.2	9.6-12.3	12.1-12.9	12.2-13.7	12.5-13.5
XX	16.9-18.7	17.7-19.2	19.4-20.3	11.5-13	11.6-13.6	13.3-15.3	11.3-13.1	11.3-14.4	12-14.9	11.5-13.7	11.4-13.9	12.8-14	12.8-14.5	12.3-13.8
XXI	17.9	19.4	18.4	12.8-13.6	14.4-14.8	14.4-14.8	13.6-13.9	14.6-15.3	14.1-14.9	12.9-13.3	11.8-13.7	13.9-14.1	14.1	13.4-13.6
XXII	15.4	16.8	16.3	9.6-10.4	10.1-12.1	10.7-11.1	9.6-10.4	12.2-13.9	11.6-12.5	12.4-13	9.8-11.1	12.6-13.3	12.3-13.1	12.7-13
XXIII	17.5-17.6	18.3-18.7	18.6-18.7	13.5-14.3	12.9-14.5	15.2-15.9	13.5-14.1	13.8-14.9	14.6-15.6	14.7-15.6	12.3-13.4	13.5-14.1	15.3-16.1	15.1-15.6
XXIV	18.3-18.5	18.8-19	20.8-21	14.4-15.2	16.7-18.2	18.7-19.3	16.6-17	18.2-19.5	16.6-18.1	15.5-16.3	16.4-17.2	16.1-16.6	17.2-18.1	17-18
XXV	16.9	20.5	19.5	16.7-17.3	16.7-18.8	18.3-18.8	15-15.9	18.1-18.8	15.6-16.7	17-17.5	16.2-17.7	16-16.6	17.2-17.8	16.9-17.8
XXVI	17.5	19.9	19.5	16.1-16.8	15.7-17.1	16.9-17.7	15.3-16.3	17.2-18.6	15.7-16.9	15.7-16.8	15-15.5	17.3-17.6	17.7-18.7	17.9-18
XXVII	16.9	18.3	20.1	16.6-16.9	17.9-18.8	18.5-19	16.7-17.9	17.4-18.6	17.5-18.1	17.2-17.6	16.7-17.9	17.6-17.8	18.5-19.2	18-18.2
XXVIII	21	21.4	20.5	17.8-19.1	20.1-21.1	20.7-21.5	19.6-19.9	20.1-21.1	20.1-21	21.3-21.5	19.6-20.5	19.2-19.3	20.6-21.3	20.8-21
XXIX	20.7	20.7	18.1	16.9-17.3	17.1-18.9	18.4-18.7	16.7-17	18.3-19.5	18.2-19.6	18.2-18.7	17.4-18.8	16.9-17.1	17.3-17.9	17.6-18.2
XXX	20.2	18.3	17.1	17.7-18.9	18.5-19	16.7-17.5	16.1-16.6	17.5-18.9	18-19	18.8-19.8	18-19.1	17.2-17.5	17.1-17.8	18.5-18.7
XXXI	19.3	20.5	19.3	17.4-17.7	17.5-19.8	18.5-19	16.7-17.4	18.5-20	17.9-19.6	18.2-18.6	17.8-19.2	18.2-18.5	18.4-19	19-19.7
XVII	XVIII	XIX	XX	XXI	XXII	XXIII	XXIV	XXV	XXVI	XXVII	XXVIII	XXIX	XXX	XXXI
17	15.6-17.6	15-15.7	16.9-18.7	17.9	15.4	17.5-17.6	18.3-18.5	16.9	17.5	16.9	21	20.7	19.3	19.3
19.7	17.8-19.4	18.2-19	17.7-19.2	19.4	16.8	18.3-18.7	18.8-19	20.5	19.9	18.3	21.4	20.7	18.3	18.3
17.8-17.9	17.2-19.7	16.7-17.8	19.4-20.3	18.4	16.3	18.6-18.7	20.8-21	19.5	19.5	20.1	20.5	18.1	17.1	17.1
11.7-12.3	9-11.3	10.4-11.2	11.5-13	12.8-13.6	9.6-10.4	13.5-14.3	14.4-15.2	16.7-17.3	16.1-16.8	16.6-16.9	17.8-19.1	16.9-17.3	17.7-18.9	17.7-18.9
10-12.3	10.2-13.1	10.1-12.3	11.6-13.6	14.4-14.8	10.1-12.1	12.9-14.5	16.7-18.2	16.7-18.8	15.7-17.1	17.9-18.8	20.1-21.1	17.1-18.9	18.5-19	18.5-19
13.1-13.7	10.5-13.6	10.2-11.7	13.3-15.3	14.4-14.8	10.7-11.1	15.2-15.9	18.7-19.3	18.3-19.3	18.3-18.8	16.9-17.7	20.7-21.5	18.4-18.7	16.1-16.6	16.1-16.6
9.5-10	9.5-11.8	8.4-9.9	11.3-13.1	13.6-13.9	9.6-10.4	13.5-14.1	16.6-17	15-15.9	15.3-16.3	15.3-16.3	16.7-17.9	19.6-19.9	16.7-17	16.7-17.5
12.3-13.4	11.6-13.9	9.9-12.4	14.6-15.3	12.2-13.9	13.8-14.9	13.8-14.9	18.2-19.5	18.1-18.8	18.1-18.8	17.2-18.6	17.4-18.6	20.1-21.1	18.3-19.5	17.5-18.7
13.4-14.5	11.4-13.8	10.7-12.4	14.1-14.9	14.1-14.9	11.6-12.5	12.3-13.4	14.6-15.3	15.6-16.7	15.7-16.9	15.7-16.9	17.5-18.1	20.1-21	18.2-19.6	18-19
12.8-13.7	10.9-14.4	11.1-13.2	12.9-13.3	12.4-13	12.4-13	14.7-15.6	14.6-15.6	18.1-18.8	18.2-18.6	17.2-17.6	17.8-19.2	21.3-21.5	18.2-18.7	18.8-19.8

Table 2. Continued

	XVII	XVIII	XIX	XX	XXI	XXII	XXIII	XXIV	XXV	XXVI	XXVII	XXVIII	XXIX	XXX
XIII	12.6–13.7	9.4–11.8	9.6–12.3	11.4–13.9	11.8–13.7	9.8–11.1	12.3–13.4	16.4–17.2	16.2–17.7	15–15.5	16.7–17.9	19.6–20.5	17.4–18.8	18–18.8
XIV	7.8–8	11.6–13.5	12.1–12.9	12.8–14	13.9–14.1	12.6–13.3	13.5–14.1	16.1–16.6	16–16.6	17.3–17.6	17.6–17.8	19.2–19.3	16.9–17.1	17.2–17.5
XV	8.8–9.5	12.3–14.6	12.2–13.7	12.8–14.5	14.1	12.3–13.1	15.3–16.1	17.2–18.1	17.2–17.8	17.7–18.7	18.5–19.2	20.6–21.3	17.3–17.9	17.1–17.8
XVI	9.9–10.2	12–13.7	12.5–13.5	12.3–13.8	13.4–13.6	12.7–13	15.1–15.6	17–18	16.9–17.8	17.9–18	18–18.2	20.8–21	17.6–18.2	18.5–18.7
XVII														
XVIII	12.7–14.9													
XIX	12.7–13.5	10.8–13.9												
XX	13.6–14.5	11.5–14.8	9.9–11.6											
XXI	16.5	12.4–13.9	13.2–14.1	9.1–9.8										
XXII	12.2–12.4	12.9–14.9	12.4–13.3	13.9–15.5	16.5									
XXIII	15–15.4	13.4–15.7	13.8–15.1	16.3–17.7	16.1–16.4	13.3–13.7								
XXIV	16.6–16.9	17.8–19.2	17.8–18.7	16.3–18.2	17.9–18	18.3–18.5	19.1–19.9							
XXV	17.8	19.4–21.1	17.6–18.3	19.3–20.7	20.3	17.3	18.9–19.1	17.7–17.8						
XXVI	18.6–18.7	17.9–20.2	16.1–17.3	18.3–19	19.6	16.6	16.7–16.2	15.3–15.5	13.1					
XXVII	18.8	18.6–20.9	17.2–17.7	18.6–19.5	18.9	17.9	20.1–20.4	16.3–16.4	13	15.7				
XXVIII	20.5	20.2–22.1	19.8–20.5	22.3–23.1	22	21.1	20.1–20.9	20.7–20.9	18.8	18.1	19.6			
XXIX	16	18–20.2	16.9–17.3	17.2–19.6	20.3	17.4	19.3–19.5	17.3–17.5	16.7	18.1	18.5	19.3		
XXX	17.1	19.6–22	17.5–18	18.9–20.5	21.2	17.9	19.8–19.9	17.4–17.5	16.2	18.6	18.5	19.8	6.5	
XXXI	17.4–17.7	18–20.2	16.7–17.8	18.5–19.8	20.1	18.1	19.9–20.1	16.9–17.1	17.4	19	19.3	19.1	16.6	16.4

Table 3. Uncorrected *p*-distances based on *COI* sequences for specimens previously identified as *Runcina coronata*

	Distance between groups (%)			Distance within groups (%)
	RC1	RC2	RC3	
RC1				0.2–0.5
RC2	6.3–7.3			1.0
RC3	6.4–7.8	7.0–7.7		0.7
RC4	7.8–8.0	8.8–9.5	9.9–10.2	0.0

with encrusting pink coralline algae (dissected and sequenced).

External morphology (Fig. 3A, B): Living specimens 3–4 mm in length. Body elongated and moderately broad. Lateral grooves on both sides between notum and foot. Notum smooth. Posterior part of the notum rounded. Foot slightly wider than the notum. Propodium rounded and wide, metapodium pointed. General ground colour of body dark brown. Front part of the notum dark brown on central zone between the eyes and pale fawn on the sides. White and yellowish spots all over the body, concentrated as semicircle bands behind the eyes and anterior to the notum end, and on the surface of the foot. Eyes small and visible. Three rounded gill laminae present on right of anus; the largest in the middle divided into two arcs. Gills yellowish with slightly brown margins. Anus located in median line of body, beneath the end of notum.

Internal anatomy (Figs 4A–C, 5A, 6A): Radular formula $19 \times 1.1.1$ (MNCN 15.05/88105). Rachidian tooth bilobed with long and smooth lateral wings on each side. Central part of rachidian tooth contains a pair of pads, each possessing 9–11 denticles. Size of denticles variable, with smaller and more developed denticles randomly distributed along pads. Small depression present between pads, with minute denticle present in some rows (Fig. 4A). Lateral teeth denticulate, elongate, hooked shape with 30–34 relatively long denticles (Fig. 4B). Triangular jaws present. Four gizzard plates with ten crests (Fig. 4C). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind the digestive gland divided into two lobes. Female gland opens to exterior through median size common genital duct (Fig. 5A). Male pore opens next to mouth, on the right side. Male copulatory organ elongated and cylindrical. Penial papilla absents. Prostate gland cylindrical, posterior to a narrow atrium. Slender seminal vesicle half size of prostate gland (Fig. 6A).

Distribution: Bréhat, Bretagne, France (Atlantic) (Quatrefages, 1844); Torbay, England. (Alder &

Hancock, 1846, Forbes, 1853); Swanage, England (present study).

Remarks: A discussion of this species is included together with *R. aurata* in the ‘Remarks’ of the latter species.

RUNCINA AURATA GARCÍA ET AL., 1986
(FIGS 3C–F, 4D–F, 5B, 6B)

Type locality: Club La Hacienda, Cádiz, Spain (36°14′18″N; 5°18′36″W)

Examined material: MNCN 15.05/91500, La Caleta (Cádiz) (36°31′59″N; 6°18′31″W), Andalusia, south-western Spain, 8 April 2019, 3.5 mm living animal, depth 0.5–1.0 m (dissected and sequenced). MNCN 15.05/88106, La Caleta (Cádiz) (36°31′59″N; 6°18′31″W), Andalusia, south-western Spain, coll. Josep Romà, 18 April 2015, 2 mm in length preserved, depth 0.5–1.0 m. (dissected and sequenced). MNCN 15.05/88107, La Caleta (Cádiz) (36°31′59″N; 6°18′31″W), Andalusia south-western, Spain, coll. Josep Romà, 18 April 2015, 2 mm in length preserved, depth 0.5–1.0 m (dissected and sequenced). MNCN:ADN 118948, La Caleta (Cádiz) (36°31′59″N; 6°18′31″W), Andalusia south-western, Spain, coll. Josep Romà, 17 May 2015, 1.5 mm in length preserved, depth 0.5–1.0 m (dissected and sequenced). MNCN:ADN 118950, El Chato (Cádiz) (36°28′39″N; 6°15′49″W), Andalusia south-western, Spain, coll. Ana Bartual, 13 April 2015, 1 mm in length preserved, depth 0.5–1.0 m (dissected and sequenced).

External morphology (Fig. 3C–F): Living specimen 3.5 mm length and preserved specimens 1–2 mm length. Body elongated and moderately broad. Lateral grooves on both sides between notum and foot. Anterior part of notum (‘head’) slightly bilobed. Posterior part of notum rounded. Propodium rounded, metapodium pointed. Foot as wide as notum. Foot extended beyond notum on rear part. Ground colour of body translucent pale fawn or yellowish. Digestive system visible as a broad brownish blotch. White spots on central zone of notum, behind eyes forming



Figure 3. Living animals of *Runcina coronata* (A, B) and *Runcina aurata* (C–F). A, MNCN 15.05/88105, 3 mm in length, Swanage (southern England). B, MNCN 15.05/90423, 4 mm in length, Swanage (southern England) (photos Ian F. Smith). C, MNCN/ADN 118948, 1.5mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). D, MNCN/ADN 118950, 1mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). E, MNCN 15.05/88106, 2 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). F, MNCN 15.05/88107, 2mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). Photos A and B, courtesy of Ian F. Smith; photos C–F by Ana Karla Araujo.

triangular patches and anterior to notum end. White spots maybe also absent. Black dots dispersed on notum and more concentrated on head zone. Eyes inconspicuous. Dark band on middle of dorsal surface of foot. Black dots may be present on ventral surface of foot. Four rounded and relatively large gills laminae to the right of anus. Gills yellowish

with slightly brown margins. Anus located in median line of body, beneath the end of notum.

Internal anatomy (Figs 4D–F, 5B, 6B): Radular formulae $12 \times 1.1.1$ (MNCN 15.05/88106) and $13 \times 1.1.1$ (MNCN 15.05/91500). Rachidian tooth bilobed with long and smooth lateral wings on each side. Central

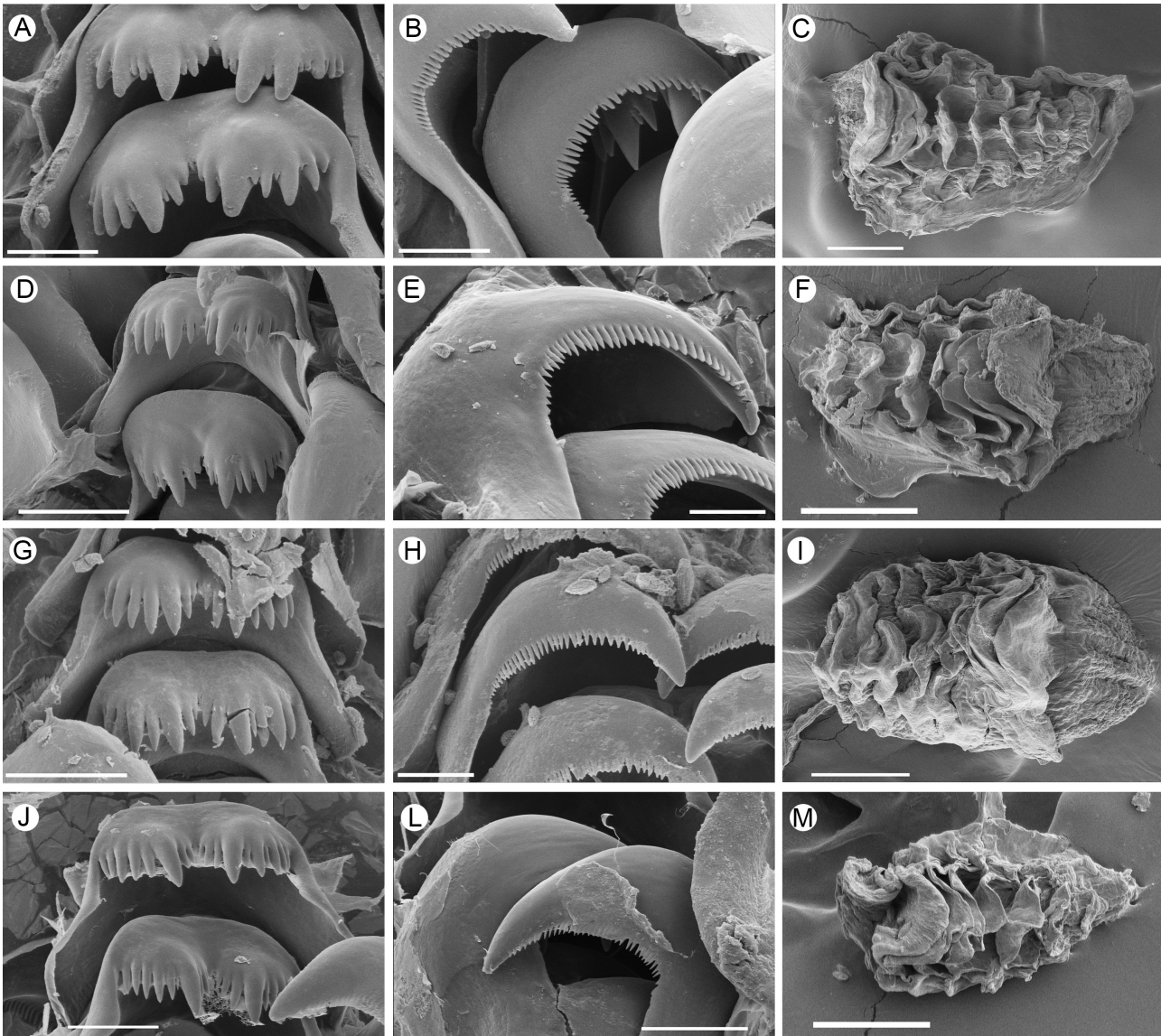


Figure 4. Scanning electron micrographs of radula and gizzard plates. A–C, *Runcina coronata*. A, rachidian teeth (MNCN 15.05/88105). B, lateral teeth (MNCN 15.05/88105). C, gizzard plate (MNCN 15.05/90423). D–F, *Runcina aurata*. D, rachidian teeth (MNCN 15.05/91500). E, lateral teeth (MNCN 15.05/88106). F, gizzard plate (MNCN 15.05/88106). G–I, *Runcina caletensis* (MNCN 15.05/200113) G, rachidian teeth. H, lateral teeth. I, gizzard plate. J–M, *Runcina tingensis*. J, rachidian teeth (MNCN 15.05/200114). L, lateral teeth (MNCN 15.05/200114). M, gizzard plate (MNCN 15.05/91514). Scale bars: A, B, E, J, L = 10 μm ; C, F, I, M = 50 μm ; D, G = 20 μm ; H = 5 μm .

part contains pair of pads, each possessing 10–11 long, slender, pointed denticles. Size of denticles variable. Small denticles between large denticles. Small depression present between pads, with minute denticle present (Fig. 4D). Lateral teeth denticulate, elongate, hooked shape with 35–36 long, pointed and same-size denticles (Fig. 4E). Triangular jaws present. Four gizzard plates with seven to nine crests (Fig. 4F). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind digestive gland,

opening to exterior through small size common genital duct (Fig. 5B). Male pore opens next to mouth, on the right side. Elongated and cylindrical male copulatory organ. Penial papilla not observed. Cylindrical and long prostate gland ends in slender and small seminal vesicle with black pigmentation (Fig. 6B).

Distribution: Cádiz, Strait of Gibraltar, Malaga and Murcia (southern Spain) (Templado, 1984; Garcia *et al.*, 1986) and Azores Islands (Portugal) (Gosliner, 1990).

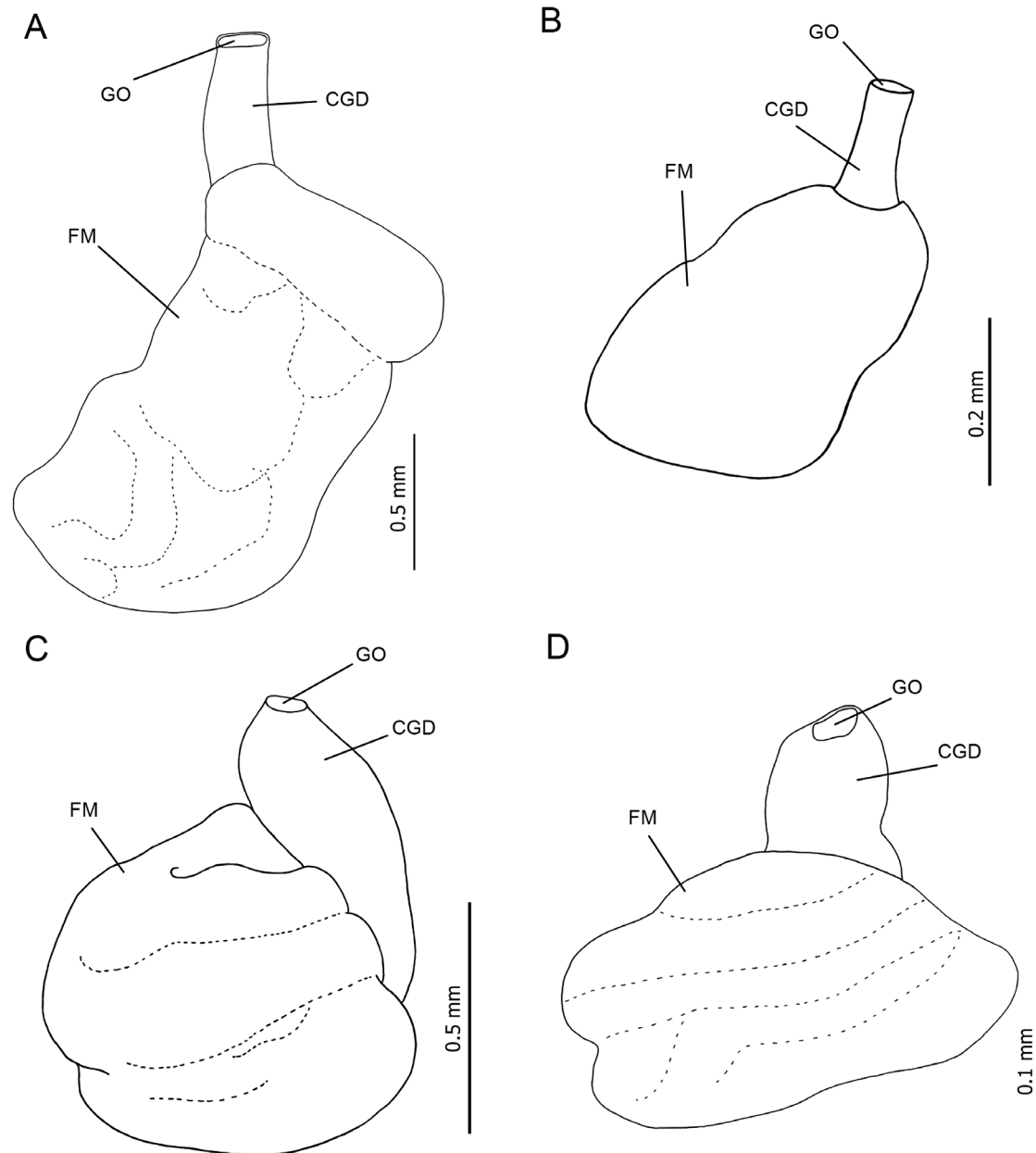


Figure 5. Female reproductive system. A. *R. coronata*, southern England (MNCN 15.05/90423). B. *R. aurata*, La Caleta, Cádiz, south-western Spain, Atlantic Ocean (MNCN 15.05/88106). C. *R. caletensis*, La Caleta, Cádiz, south-western Spain, Atlantic Ocean (MNCN 15.05/200113). D. *R. tingensis*, Tangier, north-western Morocco, Atlantic Ocean (MNCN 15.05/91514). Abbreviations: FM, female mass; CGD, common genital duct; GO, gonopore.

Remarks: *Runcina coronata* has been considered a taxonomically difficult species (see Introduction). Originally described from Brehat (Atlantic coast of France), this species was first reported in the Mediterranean Sea by Vayssière (1883) who identified specimens from Marseille (Mediterranean coast of France) as *R. coronata*. However, Burn (1963), based on morphological differences, especially the shape of the body and the colour pattern, suggested the

specimens identified by Vayssière (1883) could be *R. calaritana*. We cannot confidently attribute those specimens to a specific species, but we agree with Burn (1963) that they probably do not correspond to *R. coronata*. Pruvot-Fol (1954), and Cervera *et al.* (1991) regarded *R. calaritana* (Gulf of Cagliari, Sardinia, Italy) and *R. aurata* (from around the Strait of Gibraltar) conspecific with *R. coronata*. This problematic has ultimately created the perception

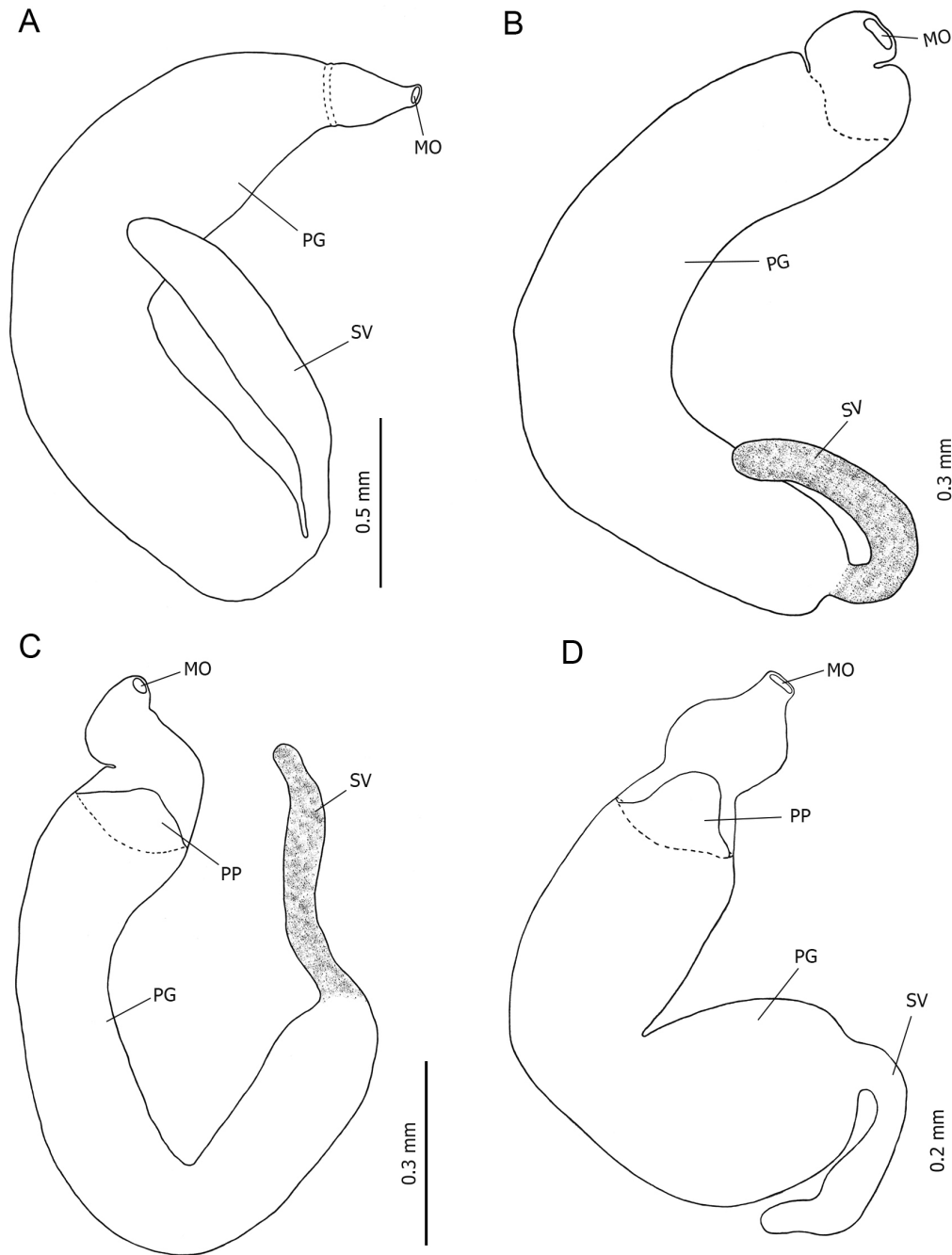


Figure 6. Male reproductive system. A, *Runcina coronata*, southern England (MNCN 15.05/90423). B, *Runcina aurata*, La Caleta, Cádiz, south-western Spain, Atlantic Ocean (MNCN 15.05/88106). C, *Runcina caletensis*, La Caleta, Cádiz, south-western Spain, Atlantic Ocean (MNCN 15.05/200113). D, *Runcina tingensis*, Tangier, north-western Morocco, Atlantic Ocean (MNCN 15.05/91514). Shaded area indicates the presence of sperm. Abbreviations: MO, male opening; PP, penial papilla; PG, prostate gland; SV, seminal vesicle.

that *R. coronata* was present in the Mediterranean Sea (Schmekel & Cappellato, 2002; Cervera *et al.*, 2004; Ballesteros *et al.*, 2016).

In general, the external and internal morphology of our specimens of *R. coronata* from Swanage

(England) are consistent with the original description of the species (Quatrefages, 1844), and with the description provided by Schmekel & Cappellato (2002) based on specimens from Roscoff (Atlantic coast of France) and Plymouth (south of

England). However, compared with the description provided by [García et al. \(1986\)](#); specimens from the Strait of Gibraltar), our animals from England exhibit several differences, mainly in the shape of the body and colour pattern. The anterior and posterior ends of the notum are rounded, while in Spanish specimens it is pointed ([García et al., 1986](#)). The colour pattern of our specimens ([Fig. 3A, B](#)) differs drastically from those from the Strait of Gibraltar, which have a uniformly dark colour pattern, two whitish bands on both sides of the head and one white small band on the posterior right side of the notum ([García et al., 1986](#)). This suggests that likely specimens attributed to *R. coronata* by [García et al. \(1986\)](#) belong to a distinct species.

The original description of *R. coronata* describes briefly the male copulatory organ as ‘a rather short testicular bag in the shape of a “club”, with a seminal vesicle sometimes absent ([Quatrefages, 1844](#)).’ [Kress \(1977\)](#) studied specimens from Plymouth (England) and provided additional anatomical data on the reproductive system. Comparatively, our specimens from Swanage (England) exhibit a similarly long and cylindrical prostate, but a slightly different seminal vesicle and common genital duct. [Kress \(1977\)](#) referred to a seminal vesicle ‘considerably shorter than prostate’ and a common genital duct forming a long loop, whereas in our material the seminal vesicle was approximately half the size of the prostate and the common genital duct was short ([Fig. 6A](#)).

The species *R. aurata* was described by [García et al. \(1986\)](#) from the southern coast of Spain (Cádiz, Strait of Gibraltar and Malaga). [Gosliner \(1990\)](#) reported the species from the Azores and suggested that a specimen illustrated and depicted by [Thompson & Brodie \(1988: fig. 1E\)](#) from Plymouth as *R. coronata* was most likely *R. aurata*. Despite the fact that the description and illustration provided by [Thompson & Brodie \(1988\)](#) are vague and lacking important information, the reference to the presence of a light area surrounding the eyes, suggests their identification as *R. coronata* to be correct.

The features of our specimens collected in Cádiz (Spain) are consistent with the original description of the species *R. aurata* ([García et al., 1986](#)). Externally, they differ from *R. coronata* by having a translucent yellow colour with black spots on the notum and on the ventral surface of the foot ([Fig. 3; Table 4](#)). Also, the number of gills is distinct: three gills in *R. coronata*, while our animals of *R. aurata* from Cádiz have four gills ([Table 4](#)). The original description of *R. aurata* refers, in fact, to three gills only, but the authors did not seem to have thoroughly looked at this character, which is difficult if not examined properly and across several specimens ([García et al., 1986](#)).

Concerning the radula, we observed some subtle differences between *R. aurata* and *R. coronata*, namely in the number of radular rows and shape of the denticles in the pads of the rachidian teeth ([Table 4](#)). However, in runcinids, these features can vary, even within species ([Schmekel & Cappellato, 2001; 2002; Araujo et al. 2019](#)), and are, therefore, difficult to use

Table 4. Summary of diagnostic characters between *Runcina coronata*, *Runcina aurata*, *Runcina caletensis* and *Runcina tingensis*. Data based on [Quatrefages \(1844\)](#), [Vayssière \(1883\)](#), [García et al. \(1986\)](#), [Cervera et al. \(1991\)](#), [Schmekel & Cappellato \(2002\)](#) and present study

	<i>Runcina coronata</i>	<i>Runcina aurata</i>	<i>Runcina caletensis</i>	<i>Runcina tingensis</i>
Colour pattern	Ground colour dark brown. White semi-circles bands behind eyes and anterior to notum end.	Ground colour translucent pale fawn or yellowish. White spots behind eyes and anterior to notum end.	Ground colour translucent pale fawn or yellowish. Black dots on notum end. Two black longitudinal lines on the head.	Ground colour light brown. Triangular white patches behind eyes and anterior to notum end.
Gills	Three rounded	Four rounded and relatively large.	Three rounded	Two rounded
Shell	Absent	Absent	Absent	Absent
Radular formulae	19 × 1.1.1	12–13 × 1.1.1	13 × 1.1.1	12–14 × 1.1.1
Rachidian teeth	9–11 short denticles on each pad	10–11 long, slender, pointed denticles	7–8 denticles stalactite-shaped. Depression between pads absent.	7–10 short triangular denticles.
Lateral teeth	30–34 denticles	35–36 denticles	33–36 denticles	34–37 denticles
Gizzard plate	10 crests	7–9 crests	7–10 crests	8 crests

in species identification. Regarding the gizzard plates, our specimens of *R. coronata* show ten crests in each plate, while in our animals of *R. aurata* it ranges from seven to nine, which is consistent with its original description (García, *et al.*, 1986).

The male reproductive system in our specimens of *R. aurata* resembles the description of this organ by Gosliner (1990) for specimens from the Azores, but we could not observe a penial papilla. The seminal vesicle is shorter than in *R. coronata* and the atrium and male opening are broader (Fig. 6A, B). The female gland mass of *R. aurata* was never studied before and in our specimens consists of one lobe, while in the studied specimens of *R. coronata* it is divided into two lobes (Fig. 5B).

The minimum uncorrected *p*-distance for the *COI* gene between *R. coronata* and *R. aurata* is 6.3% (Table 3) and, in addition to the phylogenetic tree, the species delimitation analyses suggested both species as valid (Fig. 2).

***RUNCINA CALETENSIS* ARAUJO, POLA, MALAQUIAS
& CERVERA, *SP. NOV.***

(Figs 4G–I, 5C, 6C, 7A, B)

Zoobank registration: urn:lsid:zoobank.org:act:EC216698-6700-4607-9E35-515AB69BD17C

Examined material: Holotype: MNCN 15.05/200113, La Caleta (Cádiz) (36°31'59"N; 6°18'31"W), Andalusia, south-western Spain, coll. Josep Romà, 17 May 2015, 3 mm living animal, depth 0.5–1 m (dissected and sequenced). Paratype: MNCN:ADN 118949, La Caleta (Cádiz) (36°31'59"N; 6°18'31"W), Andalusia, south-western Spain, coll. Josep Romà, 16 June 2015, 1.5 mm in length preserved, depth 0.5–1.0 m. (dissected and sequenced).

Etymology: Named after the type locality: the beach of La Caleta in Cádiz, Spain.

External morphology (Fig. 7A, B): Living and preserved specimens 3.0 mm and 1.5 mm in length, respectively. Body elongated and broad. Lateral grooves on both sides between notum and foot. Anterior part of notum ('head') slightly bilobed. Posterior part of notum rounded. Foot as wide as notum. Foot extends posteriorly beyond notum. Ground colour translucent pale fawn or yellowish. Digestive system visible as broad brownish blotch in juvenile specimens. Tiny white, black and yellow spots all over the body. White spots on the lobes of the head and on the anterior ventral surface of the foot. Triangular white patches behind the eyes. White semicircle anterior to the notum end. Very

few white spots in juvenile. Small black dots forming two longitudinal lines on head region. Black spots concentrated on the notum end posteriorly to white semicircle. In juveniles, only a few larger black spots are present along the margin of the notum. Eyes inconspicuous. Dark band on middle of posterior region of foot. Three rounded gill laminae located on right side of anus. Gills yellowish with slightly brown margins. Anus located in the midline of the body beneath the notal edge.

Internal anatomy (Figs 4G–I, 5C, 6C): Radular formula 13 × 1.1.1 (15.05/200113). Rachidian tooth slightly bilobed with long and smooth lateral wings on each side. Central part of rachidian tooth contains a pair of pads, each possessing seven to eight developed denticles. Central depression between pads absent. Denticles long and pointed, decreasing in size towards middle of the tooth (Fig. 4G). Lateral teeth denticulate, elongate, hooked shaped with 33–36 long and pointed denticles (Fig. 4H). Triangular jaws present. Four gizzard plates with seven to ten crests (Fig. 4I). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind digestive gland. Female gland opens to exterior through common genital duct (Fig. 5C). Male pore opens next to mouth, on right side. Elongated and cylindrical male copulatory organ. Short, conical and unarmed penial papilla projects into large atrium. Cylindrical prostate gland strongly curved. Long and slender seminal vesicle with black pigmentation (Fig. 6C).

Distribution: Cádiz, southern Spain (present study).

Remarks: The species *R. caletensis* shares with *R. coronata* the presence of white bands and spots on the notum, and is overall externally similar to *R. aurata*. However, in *R. caletensis* the yellow ground colour is opaque, while in *R. aurata* the colour is translucent (Figs 3E, F, 7B). In addition, *R. caletensis* lacks black spots on the ventral surface of the foot, a feature present in *R. aurata*. Based on available data, *R. caletensis* is the only one among these species with rachidian teeth lacking the depression between the pads and with well-developed denticles of similar length along the masticatory edge of pads (Fig. 4G). The female gland mass of *R. caletensis* has a rounded shape and the common genital duct is larger than in the other two species (Fig. 5A–C). The seminal vesicle in *R. caletensis* is thinner than in *R. coronata* and more elongated than in *R. aurata*. Unlike for *R. coronata* and *R. aurata*, a penial papilla was observed in *R. caletensis*.

The minimum uncorrected *p*-distances for the *COI* gene is 7.8% between *R. caletensis* and *R. coronata*,

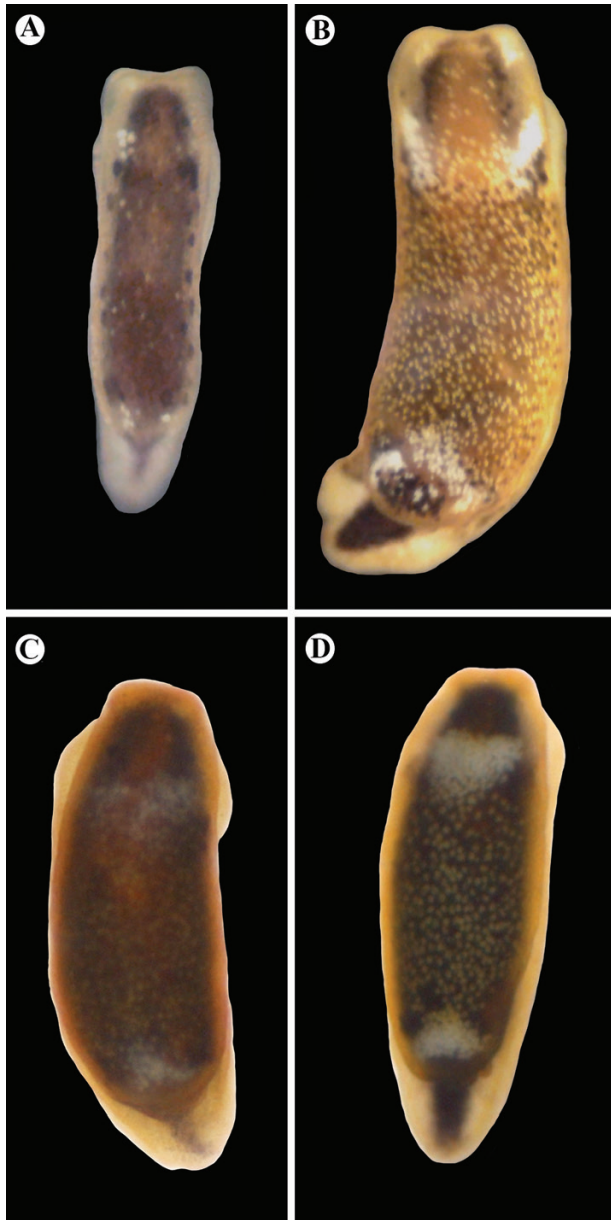


Figure 7. Living animals of *Runcina caletensis* (A, B) and *Runcina tingensis* (C, D). A, MNCN/AND 118949, 1.5 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). B, MNCN 15.05/200113, 3 mm in length, La Caleta, Cádiz (south-western Spain; Atlantic Ocean). C, MNCN 15.05/91514, 1.5 mm in length, Tangier (north-western Morocco; Atlantic Ocean). D, MNCN 15.05/200114, 2 mm in length, Tangier (north-western Morocco; Atlantic Ocean). A, B images by Ana Karla Araujo; C, D images courtesy of Naoufal Tamsouri.

and 8.8% between *R. caletensis* and *R. aurata* (Table 3). Species delimitation analyses recognized *R. caletensis* as a valid species (Fig. 2).

RUNCINA TINGENSIS* ARAUJO, POLA, MALAQUIAS & CERVERA, *SP. NOV.

(FIGS 4J–M, 5D, 6D, 7C, D)

Zoobank registration: urn:lsid:zoobank.org:act:84C29263-8B36-4FA7-A84A-94F4191CE491

Examined material: Holotype: MNCN 15.05/200114, Tangier (35°47'32"N; 5°49'04"W), Morocco, coll. Naoufal Tamsouri, 22 March 2018, 2 mm living animal, depth 0.5–1.0 m (dissected and sequenced). Paratype: MNCN 15.05/91514, Tangier (35°47'32"N; 5°49'04"W), Morocco, coll. Naoufal Tamsouri, 22 March 2018, 1.5 mm fixed animal, depth 0.5–1.0 m (dissected and sequenced).

Etymology: The name *tingensis* refers to Tingi, the Greek name of Tangier (Morocco), the type locality of the species.

External morphology (Fig. 7C, D): Living and preserved specimens 2.0 mm and 1.5 mm in length, respectively. Body elongated. Lateral grooves on both sides between notum and foot present. Anterior part of notum ('head') straight, rounded on posterior end. Foot as wide as notum, with propodium and metapodium rounded; metapodium extends beyond notum. Ground colour of body light brown. Broad continuous light orange line on edge of notum. Digestive system visible as a broad brownish blotch. White spots concentrated behind eyes and on anterior end of notum forming a triangle. Yellowish dots dispersed on middle of notum. Dark small spots can be present behind anterior white spots on head region and rear part of notum. Eyes inconspicuous. Longitudinal mid-dorsal dark band on foot. Two rounded gills laminae on right side of anus. Gills light brown. Anus located on right lateral side beneath edge of notum, approximately in mid-region of body length.

Internal anatomy (Figs 4J–M, 5D, 6D): Radular formulae $14 \times 1.1.1$ (MNCN 15.05/91514) and $12 \times 1.1.1$ (MNCN 15.05/200114). Rachidian tooth bilobed with smooth lateral wings on each side. Central part of rachidian tooth contains pair of pads, each possessing seven to ten denticles. Denticles short, pointed. One inner denticle on each pad conspicuously more developed. Central small depression present between pads; small denticle in-between pads absent (Fig. 4J). Lateral teeth denticulate, elongate and hooked shaped with 34–37 long, thin, pointed denticles (Fig. 4L). Triangular jaws present. Four gizzard plates with eight crests (Fig. 4M). Shell absent. Reproductive system monaulic. Female gland mass placed on right side and behind

digestive gland. Opens to exterior through short and wide common genital duct (Fig. 5D). Male pore opens next to mouth, on right side. Male copulatory organ cylindrical. Short and unarmed penial papilla projects into round atrium. Prostate gland cylindrical and strongly curved with posterior part rounded. Seminal vesicle slender with middle part wider (Fig. 6D).

Distribution: Tangier, Morocco (present study).

Remarks: *Runcina tingensis* resembles *R. coronata* by its dark colour and presence of small, white and yellow spots on the notum. However, *R. tingensis* has a broad, continuous, light orange line along the edge of the notum, while *R. coronata* has lighter regions only on the head and on the back of the notum. Compared with *R. aurata* and *R. caletensis*, the dark colour of *R. tingensis* contrasts with the translucent yellowish colour of these two species. The distribution of white spots is also distinctive among these species. In *R. tingensis* they form a triangular pattern on the notum (Fig. 7C, D), while in the other three species they form a semicircle (Figs 3, 7B). According to our observations, the radula of *R. tingensis* has, compared with *R. aurata*, *R. coronata* and *R. caletensis*, a prominently more developed denticle in each pad of the rachidian teeth (Fig. 4J). The female mass of *R. tingensis* differs from *R. coronata* and *R. aurata* by its elliptical shape and a larger common genital duct (Fig. 5), and its prostate is notably curved and the seminal vesicle enlarged in its central region (Fig. 6). A penial papilla was present as observed for *R. caletensis*.

The minimum uncorrected *p*-distances for the *COI* gene is 7% between *R. tingensis* and *R. coronata*, 6.4% between *R. tingensis* and *R. aurata*, and 9.9% between *R. tingensis* and *R. caletensis* (Table 3).

DISCUSSION

Burn (1963) presented a systematic classification of the order Runcinida (as Runcinacea) based on morpho-anatomical characters: in particular, shell, gills and radula. According to his classification, the rare *Ildica nana* Bergh, 1889 was the most primitive species followed by the genera *Runcina*, *Runcinida* and *Runcinella*, while the genus *Ilbia* was considered the most derived. Our results, suggest a sister-relationship between the genus *Ilbia* and the remaining members of Runcinida with genera *Runcina*, *Runcinida*, *Metaruncina* and *Lapinura* (PP = 1; BS = 99). Unfortunately, samples of *Ildica* and *Runcinella* were not available to us and, therefore, we could not thoroughly test Burn's (1963) hypothesis, but despite

the limitations of our dataset, the results suggest the validity of the families Ilbiidae (with *Ilbia*) and Runcinidae (with *Runcina*, *Runcinida*, *Metaruncina* and *Lapinura*).

RUNCINA DIVAE OR LAPINURA DIVAE?

The genus *Lapinura* (type species: *Ildica divae* Marcus & Marcus, 1963), introduced by Marcus & Marcus (1970) for specimens collected in Curaçao, Bonaire and Florida, is characterized by an external cup-shaped larval shell. This genus was synonymized with *Runcina* by Clark (1984) after examining populations from Bermuda of supposed *Lapinura divae* in which some specimens lacked the external shell. Because of this 'variability', Clark disregarded the external shell and others characteristics that define the genus, emphasizing only the radular formula ($N \times 1.1.1$) and the presence of gizzard plates – common for most runcinids – to assign *Lapinura divae* to the genus *Runcina*.

Representatives of the Caribbean species *Lapinura/Runcina divae* from Bermuda branched off outside Clade F (the one with the type species of the genus *Runcina*) with other runcinids in Clade H. Therefore, we suggest that the genus *Lapinura* should be reinstated as valid and at least applied for the Caribbean species described by Marcus & Marcus (1963). Moreover, we agree with Ortea *et al.* (2017) about a possible existence of more than one species being referred under the name *Lapinura divae*, due to the discrepancies (number of rows of the radula, presence/absence of the shell and number of crests of the gizzard plates) among specimens collected in the Caribbean Sea and Brazil (Marcus & Marcus, 1963; Marcus & Marcus, 1970; Thompson, 1977; Clark, 1984).

WHAT ABOUT EUROPEAN RUNCINIDS?

Three genera have been referred in European waters: *Runcina* (31 species), *Runcinella* (one species) and *Pseudoilbia* (one species). We have studied specimens of *Runcina* and *Pseudoilbia* but, unfortunately, specimens of *Runcinella condio* Moro & Ortea, 2015, described from the Canary Islands, were not available for study.

Runcinida

Here we add a fourth genus to the European fauna, namely *Runcinida*. To date, the genus *Runcinida* was restricted to the western Pacific Ocean and included three species, *R. elioti* (Baba, 1937), *R. valentinae* and *R. marisae*. However, according to our results, a specimen identified as *Runcinida* sp. 1, collected in

Cap Ferret, France (Atlantic coast), clustered together with *R. marisae* and *R. valentinae* (PP = 1, BS = 90). Externally, all species of *Runcinida* can be distinguished by their unique colour pattern, with a dark-brown notum, yellowish or orange edge of notum and foot, and gills arranged in a semicircle above the anus. Our specimen (*Runcinida* sp. 1; Fig. 8) fits this colour pattern and arrangement of the gills, but differs from the other species in the genus by having small, black dots spread on the dorsal and ventral surfaces of the foot and several larger white spots on the notum (Fig. 8). *Runcinida elioti* (Baba, 1937) has fewer white spots too, but lacks small, black dots on the foot (Baba, 1937). *Runcinida marisae* has small, black dots on the ventral surface of the foot but white dots are absent (Chernyshev, 1998). Finally, *Runcinida valentinae* lacks black and white dots, but has a triangular orange patch on the anterior part of the notum (Chernyshev, 2006). The ABDG and bPTP recognized *Runcinida* sp. 1, *R. marisae* and *R. valentinae* as distinct species with *COI* uncorrected *p*-distances ranging between 6.5% to 16.6% (Table 2).

Pseudoilbia avellana or *Runcina avellana*?

The species *Runcina avellana* was originally described by Schmekel & Cappellato (2001) from Banyuls-sur-Mer, French Mediterranean. These authors found an unusual radula ($3 \times 1.0.1$) and no gizzard plates in the single specimen examined, which they pointed out could be a juvenile. Ortea (2013), because of the absence of rachidian teeth and gizzard plates assigned the species to the genus *Pseudoilbia*, proposing the new combination name *Pseudoilbia avellana*.

Our specimens characterized by features consistent with the original description of *R. avellana* (*sensu* Schmekel & Cappellato, 2001; Fig. 9), like the shape



Figure 8. Living animal of *Runcinida* sp. (MNCN 15.05/90670, 3 mm in length, Cap Ferret, north of France, Atlantic Ocean). Image courtesy of Marina Poddubetskaia.

of the body, presence of a round brownish mark on the centre of tail, general colour pattern, absence of gizzard plates and presence of gills, collected at Roses, Spain about 45 km from the type locality, clustered in the phylogenetic analyses among species of the genus *Runcina* (Clade F; Fig. 1), questioning the assignment by Ortea (2013) of this species to the genus *Pseudoilbia*. The latter genus is characterized by animals lacking gills, gizzard plates, shell and with radular formula 2.0.2 (Miller & Rudman, 1968). *Runcina avellana* shares some of these features, but not all, and our phylogenetic results support its inclusion in *Runcina*. Moreover, the lack of rachidian teeth in *R. avellana*, reported by Schmekel & Cappellato (2001), might be an artefact since only one apparently juvenile specimen was studied by these authors. Unfortunately, our effort in preparing the radula of this species was not successful and, thus, we could not study this structure. Additional specimens of *R. avellana* are necessary to permit a detailed study of its anatomy and comparison with other species of *Runcina* and *Pseudoilbia*.

Runcina

The genus *Runcina* has been traditionally defined by the presence of up to four separated gills on the right side of the anus and a triseriate (1.1.1) radula with bilobed rachidian teeth and smooth or denticulated lateral teeth (Burn, 1963; Gosliner, 1991; Schmekel

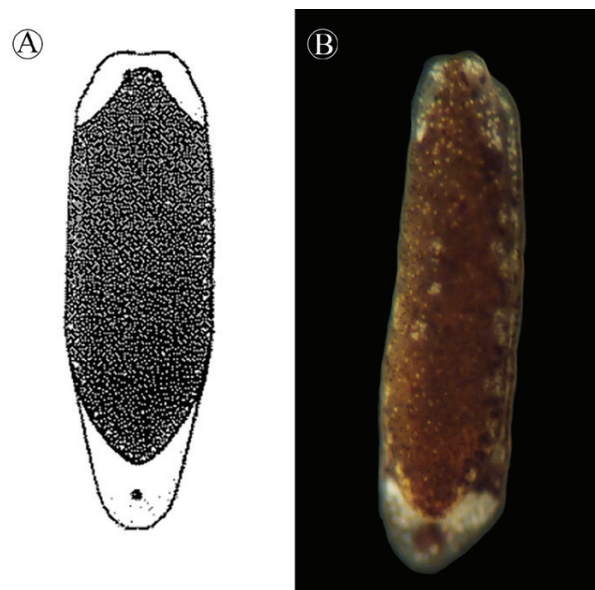


Figure 9. *Runcina avellana*. A, schematic illustration taken from Schmekel & Cappellato, 2001 (original description). B, living animal from Catalonia, north-eastern Spain (Mediterranean Sea) (MNCN 15.05/88108, 1.5 mm in length). Image B by Ana Karla Araujo.

& Cappellato, 2001). Our analysis supports the monophyly of the genus *Runcina* (PP = 1; BS = 99) but with *R. avellana*, which lacks rachidian tooth and, in addition, specimens resembling *R. ferruginea* from the Mediterranean coasts of Spain and France, and from Croatia clustered elsewhere in the tree together with other runcinids from Bermuda, Hawaii, Brazil, Japan, Russia and also from the Atlantic coast of France (PP = 1; BS = 78).

We have included in our phylogenetic analysis about 39% of the nominal species of European *Runcina* (MolluscaBase, 2021) and, in addition, several unidentified specimens from Spain, Italy and Croatia (Fig. 1). Within *Runcina*, we retrieved a clade (PP = 1; BS = 100) containing one specimen that we provisionally have identified as *R. cf. bahiensis*, one specimen provisionally identified as *R. hornae* and several unidentified specimens. Most specimens in this clade were collected in Catalonia (north-eastern Mediterranean Spanish coast) and, despite the remarkable variation in colour pattern (Fig. 10), species delimitation analyses suggest they all belong to the same species (COI uncorrected *p*-distances varied between 0.0–2.0%).

The species *R. bahiensis* was originally described from the Bay of Algeciras (Strait of Gibraltar, Spain) (Cervera *et al.*, 1991) and *R. hornae* from Banyuls-sur-Mer (Mediterranean coast of France) (Schmekel & Cappellato, 2002). Both species have been reported in several localities in Catalonia (Sánchez-Moyano *et al.*, 2000; Ballesteros *et al.*, 2016), and are regarded as differing in colour pattern, number of rows of radular teeth, shape of body and presence of small protuberances (Cervera *et al.*, 1991; Schmekel & Cappellato, 2002). Several of our specimens (Fig. 10B–D) bear a colour pattern and body shape consistent with *R. bahiensis*, but none of them have the small protuberances characteristic of this species. The study of a radula of one specimen from Catalonia (*Runcina* sp. 3) matched the original description of the radula of *R. hornae* (Schmekel & Cappellato, 2002; Fig. 11). Unfortunately, specimens of *R. bahiensis* from the type locality or nearby were not available for this study, but considering the radular similarities and the proximity to the type locality of *R. hornae* (c. 25.5 km), we identify specimens in this clade as *R. hornae*, which is here demonstrated to be a species with remarkable colour variability (Fig. 10).

Runcina coronata species complex

In the last few years, several complexes of species have been detected among heterobranch sea slugs (see, among others: Jörger & Schrödl, 2013; Padula *et al.*, 2014; Carmona *et al.*, 2015; Krug *et al.*, 2016; Korshunova *et al.*, 2017; Austin *et al.*, 2018). In

Runcinida, the first complex of species was unravelled by Araujo *et al.* (2019) for the species *Runcina brenkoae* with the description of two new species, namely *R. marcosi* and *R. lusitanica*.

In the current study, molecular and morphological data have showed that *R. coronata* hides a complex of at least four species, including two new to science and the previously described species *R. aurata*. Due to similarities with the original description by Quatrefages (1844), and the descriptions provided by Forbes (1851) and Schmekel & Cappellato (2002), we regard our specimens from Swanage (England) conspecific with *R. coronata*.

The geographical distribution of *R. coronata* has been reported to extend from England to the French Mediterranean coast (Vayssière, 1883; Cervera *et al.*, 2004). However, our results questioned the presence of *R. coronata* in the Iberian Peninsula, where most likely the records to this species belong to *R. aurata* (see in Results, 'Remarks' section of *R. aurata*). The morphological differences between specimens identified as *R. coronata* in the Mediterranean Sea and those from England and the Atlantic coast of France suggest, as previously stressed by Burn (1963), that animals studied by Vayssière (1883) from the Mediterranean are a distinct species and also that *R. calaritana* (Colosi, 1915) could be a valid name.

Therefore, we here restrict the distribution of *R. coronata* to southern England and the Atlantic coast of France, a limited geographical span supported by the direct development of the species (Schmekel & Cappellato, 2001). The species *R. aurata* and *R. caletensis*, despite subtle differences, are externally difficult to distinguish and coexist in the same geographical area (Cádiz, south of Spain), whereas the species *R. tingensis* is so far only known from the north-western coast of Morocco.

This work has revealed several additional putative cases of hidden diversity among runcinids in Europe (e. g. *R. adriatica* and *R. ferruginea*), and our detailed study of the *R. coronata* species-complex has made it possible to redefine the type species of the genus, to clarify the taxonomic status of *R. aurata* and to describe two new species to science. In addition, we have provided the first modern approach to understanding relationships in the order Runcinida and a provisional framework to discuss the familial and generic classification of the group.

ACKNOWLEDGEMENTS

We are grateful to all colleagues who provided specimens and photographs for this study, including Josep Romà, Naoufal Tamsouri, Ana Bartual, Ian F. Smith, Enric Madrenas, Miquel Pontes, Carles

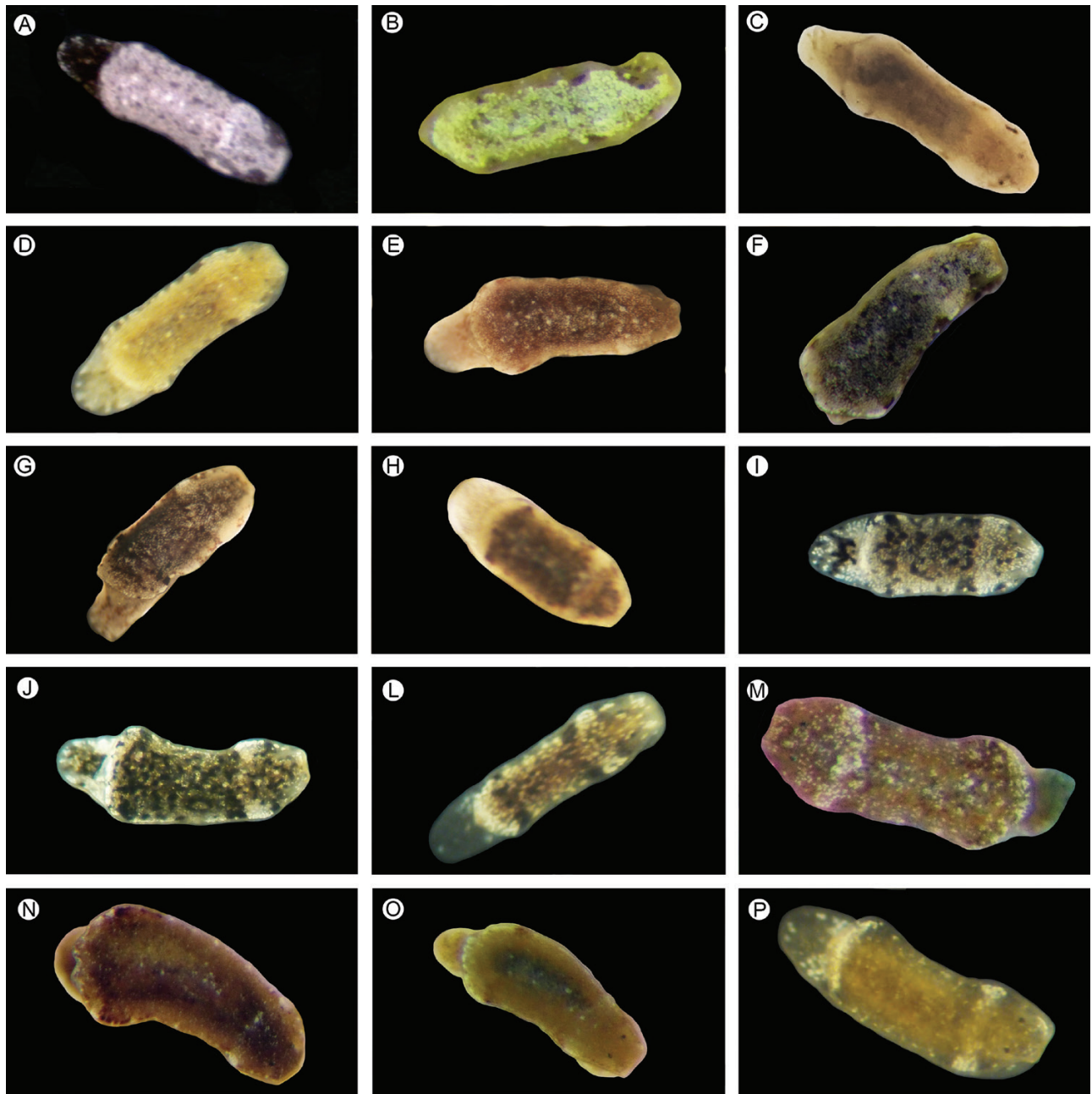


Figure 10. Living animals of *Runcina hornae*. Specimens from Catalonia, north-eastern Spain (Mediterranean Sea). A, MNCN 15.05/88104, 1 mm in length. B, MNCN 15.05/90661, 1 mm in length. C, MNCN/AND 118954, 2 mm in length. D, MNCN 15.05/90656, 1.5 mm in length. E, MNCN 15.05/90660, 3 mm in length. F, MNCN 15.05/90655, 3 mm in length. G, MNCN 15.05/90659, 2.5 mm in length. H, MNCN 15.05/90665, 1.5 mm in length. I, MNCN 15.05/90658, 1.5 mm in length. J, MNCN 15.05/90654, 2 mm in length. L, MNCN 15.05/90657, 1 mm in length. M, MNCN 15.05/90662, 2 mm in length. N, MNCN 15.05/90664, 3 mm in length. O, MNCN 15.05/90663, 1 mm in length. P, MNCN 15.05/88110, 2 mm in length. Images A, E, G, H courtesy of Carlés Galià; images B, F, M, N, O courtesy of Marina Poddubetskaia; images C, D, I, J, L, P by Ana Karla Araujo.

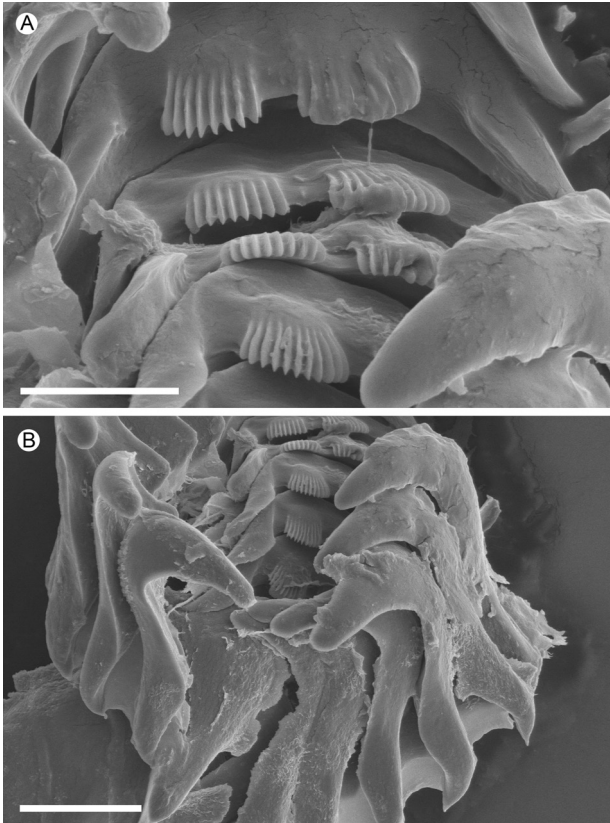


Figure 11. Scanning electron micrographs of radula of *Runcina hornae* (MNCN 15.05/90654, 1 mm in length, Mataro, Spain). A, rachidian teeth. B, lateral teeth. Scale bars: A = 5 μ m; B = 10 μ m.

Galià, Marina Poddubetskaia, Alen Petani, Jakov Prkić, Cory Pittman and Jorge Antonio Domínguez Godino. We thank Bastian Brenzinger, Luis Ricardo Simone, Jon Anders Kongsrud and Melanie Mackenzie for lending specimens and processing loans of the ZSM, MZUSP, ZMBN and Museum Victoria collections, respectively. We also thank Juan González (SC-ICYT, UCA) for assistance with SEM. This study was supported by several research projects, ‘Anfiatlantismo, alopatria y especies crípticas en moluscos opisthobranchios, con especial atención a los de la región Ibero-Balear-Macaronésica’ funded by Ministry of Education and Science (CGL2006-05182/BOS) to J. L. Cervera, ‘Diversidad de moluscos opisthobranchios en Bermuda y biogeografía del Océano Atlántico’ funded by Ministry of Science and Innovation (CTM2008-05228-E/MAR) to J. L. Cervera, ‘Progress in Mediterranean Sea Slugs biodiversity’ ASSEMBLE (7th FP) grant agreement no.22779 to J. L. Cervera, ‘La Caleta (Cádiz) e intermareales rocosos asociados: una ventana permanente al conocimiento de la biodiversidad marina’ funded by Fundación Biodiversidad of the Ministry for

the Ecological Transition and ‘Desentrañando la diversidad críptica en las regiones Lusitánica y Mediterránea: Heterobranquios marinos (Mollusca), Sílidos (Annelida) y Caprélidos (Arthropoda, Pancrustacea) como casos de estudio’ funded by the University of Cadiz (PR2018-039) to J. L. Cervera. A. K. Araujo holds a PhD grant from the Conselho Nacional de Desenvolvimento Científico e Tecnológico – CNPq/Brazil (Processo 205276/2014–8).

REFERENCES

- Akaike H. 1974.** A new look at the statistical model identification. *IEEE Transactions on Automatic Control* **19**: 716–723.
- Alder J, Hancock A. 1846.** Notices of some new and rare British species of naked Mollusca. *Annals and Magazine of Natural History* **18**: 289–294.
- Alfaro ME, Zoller S, Lutzoni F. 2003.** Bayes or bootstraps? A simulation study comparing the performance of Bayesian Markov chain Monte Carlo sampling and bootstrapping in assessing phylogenetic confidence. *Molecular Biology and Evolution* **20**: 255–266.
- Araujo AK, Pola M, Malaquias MAE, Cervera JL. 2019.** To be or not to be? What molecules say about *Runcina brenkoae* Thompson, 1980 (Gastropoda: Heretobranchia: Runcinida). *Scientia Marina* **83**: 223–235.
- Austin J, Gosliner T, Malaquias MAE. 2018.** Systematic revision, diversity patterns, and trophic ecology of the tropical Indo-West Pacific sea slug genus *Phanerophthalmus* A.Adams, 1850 (Cephalaspidea, Haminoeidae). *Invertebrate Systematics* **32**: 1336–1387.
- Baba K. 1937.** Opisthobranchia of Japan (I). *Journal of the Department of Agriculture Kyūshū Imperial University* **5**: 195–236.
- Baba K. 1967.** Supplementary notes on the anatomy of *Metaruncina setoensis* (Baba, 1954), (N.G.) (Opisthobranchia, Cephalaspidea). *Publications of the Seto Marine Biological Laboratory* **15**: 185–197.
- Ballesteros M, Madrenas E, Pontes M. 2016.** Actualización del catálogo de los moluscos opisthobranchios (Gastropoda: Heterobranchia) de las costas catalanas. *Spira* **6**: 1–28.
- Bouchet P, Rocroi JP, Hausdorf B, Kaim A, Kano Y, Nützel A, Parkhaev P, Schrödl M, Strong EE. 2017.** Revised classification, Nomenclator and Typification of Gastropod and Monoplacophoran Families. *Malacologia* **61**: 1–256.
- Burn R. 1963.** Australian Runcinacea (Mollusca, Gastropoda). *Australian Zoologist* **13**: 9–22.
- Burn R, Thompson TE. 1998.** Order Cephalaspidea. In: Beesley PL, Ross GJB, Wells A, eds. *Mollusca: the southern synthesis. Fauna of Australia, Vol. 5*. Melbourne: CSIRO Publishing, Part B viii, pp. 565–1234.
- Carmona L, Pola M, Gosliner TM, Cervera JL. 2015.** *Burnaia* Miller, 2001 (Gastropoda, Heterobranchia, Nudibranchia): a facelinid genus with an Aeolidiidae’s outward appearance. *Helgoland Marine Research* **69**: 285–291.

- Cervera JL, García-Gómez JL, García FJ. 1991.** The genus *Runcina* Forbes & Haley, 1851 (Opisthobranchia: Cephalaspidea) in the Strait of Gibraltar, with the description of a new species from the bay of Algeciras. *Journal of Molluscan Studies* **57**: 199–208.
- Cervera JL, Calado G, Gavaia C, Malaquias MAE, Templado J, Ballesteros M, García-Gómez JC, Megina C. 2004.** An annotated and updated checklist of the opisthobranchs (Mollusca: Gastropoda) from Spain and Portugal (including islands and archipelagos). *Boletín del Instituto Español de Oceanografía* **20**: 5–111.
- Chernyshev AV. 1998.** *Runcinida marisae* sp. nov. (Opisthobranchia, Runcinidae), a new runcinid mollusc from the Japan Sea. *Ruthenica* **8**: 29–32.
- Chernyshev AV. 2006.** New data on molluscs of the Family Runcinidae (Gastropoda: Opisthobranchia) from Russian Far Eastern seas. *Bulletin of the Russian Far East Malacological Society* **10**: 122–125.
- Clark KB. 1984.** New records and synonymies of Bermuda opisthobranchs (Gastropoda). *The Nautilus* **98**: 85–97.
- Colgan D, McLauchlan A, Wilson GDF, Livingston SP, Edgecombe GD. 1998.** Histone H3 and U2 snRNA DNA sequences and arthropod molecular evolution. *Australian Journal of Zoology* **46**: 419–437.
- Colosi G. 1915.** Osservazioni anatomo-istologiche sulla *Runcina calaritana* n. sp. *Memorie della Reale Accademia delle Scienze di Torino* **2**, LXVI: 1–35.
- Darriba D, Taboada GL, Doallo R, Posada D. 2012.** jModelTest2: more models, new heuristics and parallel computing. *Nature Methods* **9**: 696–704.
- Dayrat B, Tillier S. 2002.** Evolutionary relationships of euthyneuran gastropods (Mollusca): a cladistic re-evaluation of morphological characters. *Zoological Journal of the Linnean Society* **135**: 403–470.
- Dayrat B, Tillier A, Lecointre G, Tillie S. 2001.** New clades of Euthyneura gastropods (Mollusca) from 28S rRNA sequences. *Molecular Phylogenetics and Evolution* **19**: 225–235.
- Folmer RO, Black M, Hoeh W, Lutz R, Vrijenhoek R. 1994.** DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* **3**: 294–299.
- Forbes E, Hanley S. 1851.** *A history of British Mollusca and their shells*, 3. London: Van Voorst, 616.
- García JC, López CM, Luque AA, Cervera JL. 1986.** Descripción comparativa de *Runcina aurata* n. sp. y *R. coronata* (Quatrefages, 1844) (Gastropoda: Opisthobranchia). *Cahiers de Biologie Marine* **27**: 457–468.
- Gosliner T. 1990.** Opisthobranch Molluscs from the Azores Islands. I. Runcinidae and Chromodorididae. *Açoreana Supplement* 135–166.
- Gosliner TM. 1991.** Four new species and a new genus of opisthobranch gastropods from the Pacific coast of North America. *The Veliger* **34**: 272–290.
- Grande C, Templado J, Cervera JL, Zardoya R. 2004a.** Phylogenetic relationships among Opisthobranchia (Mollusca: Gastropoda) based on mitochondrial cox 1, trnV, and rrnL genes. *Molecular Phylogenetics and Evolution* **33**: 378–388.
- Grande C, Templado J, Cervera JL, Zardoya R. 2004b.** Molecular phylogeny of Euthyneura (Mollusca: Gastropoda). *Molecular Biology and Evolution* **21**: 303–313.
- Guindon S, Gascuel O. 2003.** A simple, fast, and accurate method to estimate large phylogenies by maximum-likelihood. *Systematic Biology* **52**: 696–704.
- Hillis DM, Bull JJ. 1993.** An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. *Systematic Biology* **42**: 182–192.
- Jörger KM, Schrödl M. 2013.** How to describe a cryptic species? Practical challenges of molecular taxonomy. *Frontiers in Zoology* **10**: 1–27.
- Jörger KM, Stöger I, Kano Y, Fukuda H, Knebelberger T, Schrödl M. 2010.** On the origin of Acochlidia and other enigmatic euthyneuran gastropods, with implications for the systematics of Heterobranchia. *BMC Evolutionary Biology* **10**: 323.
- Katoh K, Asimenos G, Toh H. 2009.** Multiple alignment of DNA sequences with MAFFT. *Methods in Molecular Biology* **537**: 39–64.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012.** Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* **28**: 1647–1649. doi:10.1093/bioinformatics/bts199
- Korshunova T, Martynov A, Bakken T, Picton B. 2017.** External diversity is restrained by internal conservatism: new nudibranch mollusc contributes to the cryptic species problem. *Zoologica Scripta* **46**: 683–692.
- Kress A. 1977.** *Runcina ferruginea* n. sp. (Opisthobranchia: Cephalaspidea), a new runcinid from Great Britain. *Journal of the Marine Biological Association of the United Kingdom* **57**: 201–211.
- Krug PJ, Vendetti JE, Valdés A. 2016.** Molecular and morphological systematics of *Elysia* Risso, 1818 (Heterobranchia: Sacoglossa) from the Caribbean region. *Zootaxa* **4148**: 001–137.
- Kumar S, Stecher G, Knyaz C, Tamura K. 2018.** MEGAX: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution* **35**: 1547–1549.
- Lemche H. 1967.** *Runcina* Forbes, 1851 (Gastropoda): validated under the plenary powers. Opinion 811. *Bulletin of Zoological Nomenclature* **24**: 89–90.
- Malaquias MAE, Mackenzie-Dodds J, Bouchet P, Gosliner T, Reid DG. 2009.** A molecular phylogeny of the Cephalaspidea sensu lato (Gastropoda: Euthyneura): Architectibranchia redefined and Runcinacea reinstated. *Zoologica Scripta* **38**: 23–41.
- Marcus Er, Marcus Ev. 1963.** Opisthobranchs from Lesser Antilles. *Studies on the Fauna of Curaçao and other Caribbean Islands* **19**: 1–76.
- Marcus Er, Marcus Ev. 1970.** Opisthobranchs from Curaçao and Faunistically related regions. *Studies on the Fauna of Curaçao and other Caribbean Islands* **122**: 1–129.

- Mikkelsen PM. 1993.** Monophyly versus the Cephalaspidea (Gastropoda, Opisthobranchia) with an analysis of traditional cephalaspidean characters. *Bolletino Malacologico* **29**: 115–138.
- Mikkelsen PM. 1996.** The evolutionary relationships of Cephalaspidea s.l. (Gastropoda: Opisthobranchia): a phylogenetic analysis. *Malacologia* **37**: 375–442.
- Millard V. 1997.** *Classification of Mollusca. A classification of worldwide Mollusca*. Pretoria: published by the author, 544.
- Miller MC, Rudman WB. 1968.** Two new genera and species of the superfamily Runcinoidea (Mollusca: Gastropoda: Opisthobranchia). *Transactions of the Royal Society of New Zealand (Zoology)* **10**: 183–189.
- Moro L, Ortea J. 2015.** Nuevos taxones de babosas marinas de las Islas Canarias y de Cabo Verde (Mollusca: Heterobranchia). *Vieraea* **43**: 21–86.
- Odhner N. 1939.** Opisthobranchia Mollusca from the western and northern coasts of Norway. *Kongelige Norske Videnskabers Selskabs Skrifter* **1**: 1–93.
- Odhner NH. 1968.** Sous-classe des Opisthobranches. In: Franc A, Grassé PP, eds. *Traité de Zoologie* 5(3). Paris: Masson, 608–893.
- Ortea J. 2013.** Descripción de un nuevo runcináceo (Gastropoda: Opisthobranchia) de las costas de Ghana, África Occidental. *Revista de la Academia Canaria de Ciencias* **XXV**: 9–14.
- Ortea J, Moro L, Espinosa J. 2017.** El género *Lapinura* (Marcus & Marcus, 1963) (Mollusca: Runcinacea) en el Atlántico, con la descripción de nuevas especies de las islas de Cabo Verde y Costa Rica. *Avicennia* **21**: 11–18.
- Oskars TR, Bouchet P, Malaquias MAE. 2015.** A new phylogeny of the Cephalaspidea (Gastropoda: Heterobranchia) based on expanded taxon sampling and gene markers. *Molecular Phylogenetics and Evolution* **89**: 130–150.
- Padula V, Araújo AK, Matthews-Cascon H, Schrödl M. 2014.** Is the Mediterranean nudibranch *Cratena peregrina* (Gmelin, 1791) present on the Brazilian coast? Integrative species delimitation and description of *Cratena minor* n. sp. *Journal of Molluscan Studies* **80**: 575–584.
- Palumbi SR, Martin A, Roman S, Mcmillan W, Stice L, Grabowski G. 1991.** *The simple fools' guide to PCR*. Department of Zoology and Kewalo Laboratory. Available from: palumbi.stanford.edu/SimpleFoolsMaster.pdf.
- Pruvot-Fol A. 1954.** Mollusques Opisthobranches. *Faune de France* **58**: 460.
- Puillandre N, Lambert A, Brouillet S, Achaz G. 2012.** ABGD, Automatic Barcode Gap Discovery for primary species delimitation. *Molecular Ecology* **21**: 1864–1877.
- Quatrefages De J. 1844.** Mémoire sur les Gastéropodes phlébentérés, ordre nouveau de la classe des gastéropodes. *Annales des Sciences Naturelles (Zoologie), Serie 3*, **1**: 129–183.
- Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA. 2018.** Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Systematic Biology* **67**: 901–904.
- Ronquist F, Huelsenbeck JP. 2003.** MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* **19**: 1572–1574.
- Sánchez-Moyano JE, Estacio FJ, García-Adiego EM, García-Gómez JC. 2000.** The molluscan epifauna of the alga *Halopteris scorparia* in southern Spain as a bioindicator of coastal environmental conditions. *Journal of Molluscan Studies* **66**: 431–448.
- Schmekel L. 1985.** *Aspects of evolution within the opisthobranchs*. In: Trueman ER, Clarke MR, eds. *The Mollusca. Evolution 10*. Orlando: New York: Academic Press, 221–267.
- Schmekel L, Cappellato D. 2001.** Contributions to the Runcinidae. I. Six new species of the genus *Runcina* (Opisthobranchia Cephalaspidea) in the Mediterranean. *Vie et Milieu* **51**: 141–160.
- Schmekel L, Cappellato D. 2002.** Contributions to the Runcinidae. II. Three new species and comparative studies on five established species of *Runcina* (Opisthobranchia Cephalaspidea) in the Mediterranean. *Vie et Milieu* **52**: 85–102.
- Stamatakis A. 2014.** RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **30**: 1312–1313.
- Talavera G, Castresana J. 2007.** Improvement of phylogenies after removing divergent and ambiguously aligned blocks from protein sequence alignments. *Systematic Biology* **56**: 564–577.
- Templado J. 1984.** Moluscos de las praderas de *Posidonia oceanica* en las costas del Cabo de Palos (Murcia). *Investigación Pesquera* **48**: 509–526.
- Thompson TE. 1976.** *Biology of Opisthobranch Molluscs*, Vol I. London: The Ray Society, 207.
- Thompson TE. 1977.** Jamaican opisthobranch molluscs I. *Journal of Molluscan Studies* **43**: 93–140.
- Thompson TE, Brodie G. 1988.** Eastern Mediterranean Opisthobranchia: Runcinidae (Runcinacea), with a review of Runcinid classification and a description of a new species from Fiji. *Journal of Molluscan Studies* **54**: 339–346.
- Vaught KC. 1989.** *A classification of the living Mollusca*. Melbourne (FL): American Malacologists, 189.
- Vayssière A. 1883.** Les genres Pelta (*Runcina*) et Tyrodina. *Annales des Sciences Naturelles (Zoologie), Serie 6*, **15**: 1–46.
- Vonnemann V, Schrödl M, Klussmann-Kolb A, Wägele H. 2005.** Reconstruction of the phylogeny of the Opisthobranchia (Mollusca: Gastropoda) by means of 18S and 28S rRNA gene sequences. *Journal of Molluscan Studies* **71**: 113–125.
- Wägele H, Klussmann-Kolb A. 2005.** Opisthobranchia (Mollusca, Gastropoda) — more than just slimy slugs. Shell reduction and its implications on defence and foraging. *Frontiers in Zoology* **2**: 1–18.
- Wägele H, Klussmann-Kolb A, Verbeek E, Schrödl M. 2014.** Flashback and foreshadowing— a review of the taxon Opisthobranchia. *Organisms Diversity & Evolution* **14**: 133–149.
- Zhang J, Kapli P, Pavlidis P, Stamatakis A. 2013.** A general species delimitation method with applications to phylogenetic placements. *Bioinformatics* **29**: 2869–287.

SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.

Figure S1. Phylogenetic hypothesis based on BI of the 16S gene. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood. Unsupported branches not labelled.

Figure S2. Phylogenetic hypothesis based on BI of the *COI* gene. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood. Unsupported branches not labelled.

Figure S3. Phylogenetic hypothesis based on BI of the *H3* gene. Numbers on the left of the slash are posterior probabilities and on the right bootstrap values derived from maximum likelihood. Unsupported branches not labelled.