



Molecular phylogeny and classification of the chemosymbiotic bivalve family Lucinidae (Mollusca: Bivalvia)

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A new molecular phylogeny of the chemosymbiotic bivalve family Lucinidae is presented. Using sequences from the nuclear 18S and 28S rRNA genes and the mitochondrial gene cytochrome *b*, 105 specimens were analysed representing 87 separate species classified into 47 genera. Samples were collected from a wide range of habitats including mangroves, seagrass beds, shallow sands, offshore muds, and hydrocarbon seeps at depths ranging from the intertidal to over 2000 m. A chronogram, derived from the combined molecular tree, was calibrated using ten lucinid fossils. The trees show five well-supported clades and two single branches of *Fimbria fimbriata* (Linnaeus, 1758) and *Monitilora ramsayi* (Smith, 1885). A new classification of Lucinidae is proposed with seven subfamilial divisions: three new subfamilies – Pegophyseminae, Leucosphaerinae, and Monitilorinae – are introduced and Codakiinae, usually treated as a synonym of Lucininae, is revived to include the *Lucinoma*, *Codakia*, and *Ctena* subclades. Membership of the Lucininae and Myrteinae is considerably revised compared with Chavan's commonly employed 'Treatise' classification. Previously considered as a separate family, Fimbriinae is now regarded as a subfamily within Lucinidae. The status of Milthinae is presently equivocal pending further analysis and Divari-cellinae is recognized as polyphyletic, and is therefore abandoned, with species and genera now grouped in various places within the Lucininae. Deeper water Lucinidae mainly belong to Leucosphaerinae, Codakiinae (*Lucinoma* clade), and Myrteinae, with *Lucinoma* species being most frequently associated with hydrocarbon seeps. Species occurring in seagrass habitats derive largely from Pegophyseminae, Codakiinae, and Lucininae, and species from mangrove habitats derive from the Pegophyseminae and Lucininae.

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INTRODUCTION

Bivalves of the family Lucinidae are remarkable for their obligate symbiosis with sulphide-oxidising Proteobacteria, housed in the ctenidia, from which they gain much of their nutrition (e.g. Distel & Felbeck, 1987; Distel, 1998; Gros, Liberge & Felbeck, 2003; Ball *et al.*, 2009). Morphological and palaeoecological evidence from fossils suggests that the chemosymbiosis is ancient (Liljedahl, 1991a, b; Taylor & Glover, 2000), with the implication that the course of lucinid

evolution and their habitat occupation patterns have been strongly constrained by the association. Although chemosymbiosis has been recorded in six bivalve families, Lucinidae are by far the most diverse (Taylor & Glover, 2010). They are particularly varied and abundant in the tropics, with a depth range from the intertidal zone to around 2500 m, and occupying a wide range of habitats, including mangrove muds, intertidal sands, seagrass beds, subtidal sites of organic enrichment, oxygen minimum zones, cold seeps, mud volcanoes, and hydrothermal vents.

There are estimated to be 330 documented living species of Lucinidae classified into 88 genera (J.

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Taylor & E. Glover, pers. database), but we are aware of many undescribed species, especially from tropical offshore habitats. Burgeoning interest in chemosymbiosis and deep-sea exploration, particularly around hydrocarbon seeps and vents, has stimulated much taxonomic research. Continuing systematic studies of Lucinidae from both shallow and deep-water habitats have demonstrated that diversity has been grossly underestimated, with a plethora of new species and genera described within the last 10 years. For example, Cosel & Bouchet (2008) described 32 new species and nine new genera from bathyal depths of the Indo-West Pacific, Glover & Taylor (2007) reported 18 new species and nine new genera from around New Caledonia, Cosel (2006) introduced eight new species and six new genera from waters off West Africa, and Taylor & Glover (2009a) described four new species and a new genus from hydrocarbon seeps in the western Atlantic.

In addition to the diversity of living lucinids the family has a long fossil history dating back at least to the Silurian (Ludlovian, *c.* 420 Ma; Liljedahl, 1991a, b). There are surprisingly few records from the late Palaeozoic, but diversity increased significantly throughout the Mesozoic, followed by a major radiation in the early Cenozoic, and with many living genera making first appearances in the Neogene.

Our aim in this study is to provide a new molecular phylogeny for the Lucinidae. As well as reconstructing the evolutionary history of the modern Lucinidae, a robust molecular phylogeny from a wide range of taxa will form the basis of a new classification, providing a framework for the study of chemosymbiosis in the family, including the distribution of bacterial symbionts and the recognition of possible co-evolutionary pathways, and additionally, tracking morphological characters associated with symbiosis. Patterns of habitat occupation and the association of distinct clades of lucinids with, for example, mangroves, seagrass beds, cold water, deep water, hydrocarbon seeps, and hydrothermal vents can also be examined. Finally, a strong molecular phylogeny should allow us to evaluate the phylogenetic signal of shell characters, such as the presence of divaricate ribbing, ligamental structure, patterns of hinge teeth, shape of the anterior adductor muscle scar, and form of lunule, which might aid the study and reinterpretation of the rich fossil record of Lucinidae.

Prior to 2004, the superfamily Lucinoidea was generally considered to comprise the families Lucinidae, Fimbriidae, Thyasiridae, Ungulinidae, Cyrenoididae, and Mactromyidae (Chavan, 1969). Molecular analyses have revised this concept, demonstrating that three families, Ungulinidae, Thyasiridae, and Cyrenoididae, are not closely related to Lucinidae, and should be removed from the Lucinoidea and clas-

sified elsewhere within euheterodont bivalves (Williams, Taylor & Glover, 2004; Taylor & Glover, 2006; Taylor *et al.*, 2007; Taylor, Glover & Williams, 2009). As for the Fimbriidae, *Fimbria fimbriata* (Linnaeus, 1758) nested amongst a range of Lucinidae species, with no support for separate familial status (Williams *et al.*, 2004), although *Fimbria*-like bivalves have a long fossil history into the Mesozoic. Thus, for living taxa, Lucinoidea now comprises only Lucinidae, with the positions of the extinct Mactromyidae and Paracyclidae remaining equivocal. The monophyly of the Lucinidae was established by previous molecular analyses using species from a wide variety of heterodont families as out-groups, and using independent lucinid data sets (Williams *et al.*, 2004; Taylor, Glover & Williams, 2005; Taylor, Williams & Glover, 2007).

The first comprehensive study of Recent and fossil Lucinidae to develop ideas of relationship and phylogeny was that of Chavan (1937–1938), which, as well as including an extensive discussion of the affinities of genera, included a geological range chart of recent and fossil genera that outlined his ideas. Later, Chavan (1951) reviewed the lucinids possessing divaricate shell ribbing, and these were subsequently classified by Glibert & Van de Poel (1967) into a separate new subfamily: Divaricellinae. Thirty years later Chavan's ideas had changed significantly (Chavan, 1969), and he divided the Lucinidae into four subfamilies, Lucininae, Myrteinae, Milthinae, and Divaricellinae, but with Fimbriidae as a separate family. Bretsky (1976), working with mostly North American taxa, combined results from a phenetic analysis (Bretsky, 1970) with data from fossils to produce a series of phylogenetic trees for different lucinid lineages. The classification based on these studies used no suprageneric categories, but divided lucinids into seven broad genera with numerous subgenera. There are major conflicts between the shell-based phylogenies and classifications of Chavan and Bretsky, probably resulting from the homoplasy of shell characters (for a more detailed review, see Taylor & Glover, 2006). Despite these problems, further compounded by poor illustrations, Chavan's (1969) classification of lucinid genera remains the most widely used. However, an initial molecular analysis using 31 species from 21 genera of Lucinidae (Williams *et al.*, 2004) revealed major incongruence with the morphology-based phylogenies and classifications, suggesting that a major revision of ideas concerning lucinid relationships was needed. Notable results from this preliminary molecular analysis were the identification of a distinct clade of *Anodontia* species [excepting *Anodontia alba* (Link, 1807)], two major clades of shallow-water lucinids, and the basal position of *Myrtea* and *NotoMyrtea*. Since then, further molecular analyses of lucinids have investi-

gated the status and relationships of some individual species (Glover, Taylor & Rowden, 2004) and genera (Glover, Taylor & Williams, 2008). Where appropriately preserved material was available, taxonomic decisions based on shell morphology have been corroborated by molecular results (Taylor & Glover, 2005; Glover *et al.*, 2008). Additionally, it has been recognized that some living species have been 'shoehorned' into inappropriate genera, for instance the abundant western Atlantic species '*Lucina*' *costata* (d'Orbigny, 1846), has been variously classified as *Codakia*, *Ctena*, or *Parvilucina*. Molecular data should clarify the positions of such taxa.

This new molecular analysis using the nuclear 18S and 28S rRNA genes, and the mitochondrial gene cytochrome *b*, is based on 105 samples, including 87 recognized species from 47 genera (53% of living genera 47/88), and encompassing most of the major groups of living Lucinidae from around the world collected from a wide range of habitats at depths from 0 to 2050 m. The analysis includes longer sequences for 18S and 28S genes, and new cytochrome *b* data for many of the taxa previously analysed by Williams *et al.* (2004). Despite the sampling effort, important taxa such as *Miltha* and *Eomiltha*, which represent living but rare survivors of once diverse groups of lucinids from the Cenozoic, are unfortunately missing from the analysis.

MATERIAL AND METHODS

Species analysed, with collection localities, genes sequenced, and GenBank numbers, are listed in Table 1. Taxa previously referred to as '*Lucina*' new genus and species and '*Lucina*' *dalli* Lyngé, 1909 in our previous molecular analysis (Williams *et al.*, 2004) are now classified as *Bathyaustriella thionipta* Glover *et al.*, 2004 and *Indoaustriella scarlatoi* (Zorina, 1974), respectively (Glover *et al.*, 2004, 2008).

The DNA extraction and amplification protocols described by Williams & Ozawa (2006) and Taylor, Glover & Williams (2008) were used to amplify portions of three genes: the nuclear 18S and 28S rRNA genes, and the mitochondrial gene cytochrome *b* (*cytb*). Sequence reactions were performed directly on purified PCR products using a BigDye Terminator v1.1 Cycle Sequencing Kit (Applied Biosystems, Carlsbad, CA, USA), run on an Applied Biosystems 3730 DNA Analyser automated capillary sequencer. Sequencing and PCR primers are listed in Table 2.

Complete double-stranded forward and reverse sequences for each gene fragment were assembled and edited using SEQUENCHER v4.8 (Gene Codes Corporation, Ann Arbor, MI, USA). The alignment of *cytb* sequences was unambiguous and no stop codons

were observed. Sequences of ribosomal genes were aligned using ClustalX v2.0.9 (Thompson, Higgins & Gibson, 1994; Thompson *et al.*, 1997) (delay divergent sequence = 95%, gap-opening penalty = 15, gap-extension penalty = 7, and with iterations at the end of the alignment), with minor adjustments made by eye in MACCLADE v4.08 OSX (Maddison & Maddison, 2003). Poorly aligned sites in the ribosomal gene alignments were identified using GBLOCKS v0.91b (Castresana, 2000) and removed from the analyses. The parameters for GBLOCKS were set as: minimum number of sequences for a conserved position, 70% of total number of sequences; minimum number of sequences for a flanking position, 90% of the total number of sequences; maximum number of contiguous non-conserved positions, 3; minimum length of a block, 10; and all gap positions allowed. After the removal of ambiguous blocks of data, selected using GBLOCKS, a total of 980 bp of 18S rRNA (84% of 1156 bp in the original alignment) and 1351 bp of 28S rRNA (75% of 1796 bp in the original alignment) remained to be used in the phylogenetic analyses. Alignments are available on request from the authors.

Phylogenies (Figs 1–4) were constructed using Bayesian methods (MRBAYES v3.1.2; Huelsenbeck & Ronquist, 2001). Models used in the Bayesian analyses were determined by MRMODELTEST v2.1 (J. A. A. Nylander, 2004. Program distributed by the author. Evolutionary Biology Centre, Uppsala University). The *cytb* data set was further tested to see whether variation across codon positions would result in an improved likelihood. The best model for all data sets was determined to be GTR + G + I using the hierarchical likelihood-ratio tests. In the Bayesian analysis the GTR + G + I model was used for each gene partition, four chains were used, and the starting tree was random. As suggested by the model, base frequencies were estimated, rate variation was gamma-distributed among sites and approximated with four categories (α shape estimated) and an allowance was made for invariant sites. In the combined analyses variation was partitioned among genes, and gene-specific model parameters were used (with all parameters free to vary independently within each partition). In addition, each gene was allowed to evolve at a different rate. The analysis for each data set was run for 3 500 000 generations (7 500 000 for combined gene analyses), with a sample frequency of 100. Each analysis was run twice. The first 15 000 trees from each run were discarded so that the final consensus tree was based on the combination of accepted trees from each run (a total of 40 000 trees; 80 000 for the combined gene tree). Convergence between the two runs was tested by examining the potential scale reduction factors (PSRFs) produced by the 'sump' command in MRBAYES. Support for nodes

Table 1. Species included in the analysis classified in new subfamily classification (Table 2), collection localities, museum registration numbers for vouchers and GenBank accession numbers for the genes sequenced for each taxon

Species	Locality	Locality code	Museum number	18S rRNA	28S rRNA	Cytb
Pegophyseminae						
<i>Pegophysema philippiana</i> (Reeve, 1850)	King Bay, Dampier, Western Australia		BMNH 20070286	AJ581846	AM779673	
<i>Cavatidens bullula</i> (Reeve, 1850)	Roebuck Bay, Broome, Western Australia	BR	BMNH 20100264	FR686688	FR686769	FR686599
<i>Cavatidens bullula</i> (Reeve, 1850)	Kungkaben Bay, Thailand	KK	BMNH 20070284	AM774495	AM779668	FR686598
<i>Cavatidens omissa</i> (Iredale, 1930)	Moreton Bay, Queensland, Australia	MB	BMNH 20100278	AJ581847	AJ581881	FR686601
<i>Cavatidens omissa</i> (Iredale, 1930)	Gunnamatta Bay, New South Wales, Australia	SYD	BMNH 20100285	AJ581843	AJ581878	FR686626
<i>Cryptophysema vesicula</i> (Gould, 1850)	Dili, East Timor	TM	BMNH 20100288	FR686768	FR686768	FR686603
<i>Cryptophysema vesicula</i> (Gould, 1850)	Onna, Okinawa, Japan	OK	BMNH 20100289	FR686767	FR686767	FR686604
<i>Euanodontia ovum</i> (Reeve, 1850)	Lizard Island, Queensland, Australia	LI	BMNH 20100290	AJ581844	AJ581879	FR686600
<i>Euanodontia ovum</i> (Reeve, 1850)	Rodrigues, Indian Ocean	ROD	BMNH 20100295	AJ581845	FR687321	FR686602
<i>Loripinus fragilis</i> (Philippi, 1836)	Embiez Island, France		BMNH 20100270	AJ581842	AJ581877	
Leucosphaerinae						
<i>Leucosphaera</i> cf. <i>diaphana</i> Glover & Taylor, 2007	Ubajan, Bohol Island, Philippines. PANGLAO 2004, st. S27, 9°41.5'N, 123°51.0'E. 12 m		MNHN IM-2009-10359	FR686699	FR686781	FR686683
<i>Anodontia alba</i> Link, 1804	Lower Matecumbe Key, Florida, USA		BMNH 20070286	AJ581840	AM779672	FR686597
<i>Pseudolucinisca lacteola</i> (Tate, 1897)	Esperance, Western Australia		BMNH 20100248	AJ581867	AJ581900	FR686624
<i>Dulcina</i> sp.	Bohol Sea, Philippines. PANGLAO 2005, St. CP 2335, 9°34.3'N, 123°37.8'E. 729–733 m		MNHN IM-2009-10371	FR686695	FR686777	FR686615
<i>Dulcina karubari</i> Cosel & Bouchet (2008)	Maribojoc Bay, Bohol Sea, Philippines. PANGLAO 2005, st CP 2331, 9°39.2'N, 123°47.5'E. 255–268 m		MNHN IM-2009-10372	FR686697	FR686776	FR686608
Undescribed genus/species (UGS-1)	Maribojoc Bay, Bohol Sea, Philippines. PANGLAO 2005, st CP 2397, 9°34.9'N, 123°41.7'E. 642–669 m		MNHN IM-2009-10373	FR686696	FR686778	FR686611
Undescribed genus/species (UGS-2)	Manga, Bohol Island, Philippines. PANGLAO 2004, st S20, 9°41.8'N, 123°51.1'E. 10 m		MNHN IM-2009-10360	FR686698	FR686780	FR686681
Undescribed genus/species (UGS-3)	Chesterfield Bank, New Caledonia. EBISCO 2005, st CP 2614, 19°39'S, 158°47'E. 608–642 m		MNHN IM-2009-10376	FR686701	FR686779	FR686606

Myrteinae									
<i>Myrtea spinifera</i> (Montagu, 1803)	Shetland Islands, UK 60.1956°N, 1.1253°W. 48 m.	BMNH 20100296	FR686690	FR686774	FR686668				
<i>Notomyrtea botanica</i> (Hedley, 1918)	Victor Harbour, South Australia	BMNH 20100285	AJ581862	AJ581896	FR686666				
<i>Myrtea flabelliformis</i> (Prashad, 1932)	Bohol Sea, Philippines. PANGLAO 2005, st CP 2388, 9°26.9'N, 123°34.5'E. 762–786 m	MNHN IM-2009-10375	FR686694	FR686775	FR686614				
<i>Myrtea</i> sp.	off Pamilacan Island Philippines. PANGLAO 2005, st CP 2348, 9°29.6'N, 123°52.5'E, 196–216 m	MNHN IM-2009-10369	FR686693	FR686770	FR686613				
<i>Myrtea</i> sp.	Safaga, Egypt, Red Sea	BMNH 20100300	FR686689	FR686773	FR686612				
<i>Gloverina rectangularis</i> Cosel & Bouchet (2008)	Bohol Sea, Philippines. PANGLAO 2005, st CP 2389, 9°27.9'N, 123°38.4'E, 784–786 m	MNHN IM-2009-11084	FR686692	FR686771	FR686612				
<i>Gloverina</i> cf. <i>rectangularis</i> Cosel & Bouchet (2008)	Bohol Sea, Philippines. PANGLAO 2005, st CP 2388, 9°26.9'N, 123°34.5'E, 762–786 m	MNHN IM-2009-10368	FR686691	FR686772	FR686607				
Codakiinae									
<i>Codakia orbicularis</i> (Linnaeus, 1758)	Little Duck Key, Florida, USA	BMNH 20100281	AM774500	AM779674	FR686625				
<i>Codakia paytenorum</i> (Iredale, 1937)	Miyako Island, Okinawa, Japan	BMNH 20100283	FR686709	FR686816	FR686630				
<i>Codakia paytenorum</i> (Iredale, 1937)	Lord Howe Island, Australia	BMNH 20100279	FR686708	FR686815	FR686629				
<i>Codakia rugifera</i> (Reeve, 1850)	Ulladulla, New South Wales, Australia	BMNH 20100291	AJ581851	AJ581885	FR686632				
<i>Codakia tigrina</i> (Linnaeus, 1758)	Lizard Island, Queensland, Australia	BMNH 20100286	AJ581852	AJ581886	FR686641				
<i>Codakia interrupta</i> (Lamarck, 1816)	Alona Beach, Pangelao Island, Philippines. PANGLAO 2004, st M1, 9°32.9'N, 123°46.6'E. 0–1 m	MNHN IM-200910357	FR686702	FR686817	FR686664				
<i>Ctena bella</i> (Conrad, 1837)	Moreton Bay, Queensland, Australia	BMNH 20100274	FR686711	FR686823	FR686639				
<i>Ctena bella</i> (Conrad, 1837)	Yonahisamatsu Ryuku Islands, Japan	BMNH 20100297	FR686704	FR686824	FR686618				
<i>Ctena decussata</i> (da Costa, 1829)	Embiez Islands, France	BMNH 20100271	FR686706	FR686826	FR686621				
<i>Ctena decussata</i> (da Costa, 1829)	Mali Ston Bay, Croatia	BMNH 20100261	FR686705	FR686825	FR686620				
<i>Ctena delicatula</i> (Pilsbry, 1904)	Hong Kong	BMNH 20100261	FR686710	FR686825	FR686620				
<i>Ctena delicatula</i> (Pilsbry, 1904)	Kungkrabaen Bay, Thailand	BMNH 20070283	AM774494	AM779667	FR686638				
<i>Ctena eburnea</i> (Gmelin, 1791)	Mao Island, Cape Verde Islands	BMNH 20100287	FR686707	FR686827	FR686622				
<i>Ctena mexicana</i> (Dall, 1901)	Bahias de Punta, Soliman, Mexico	BMNH 20070285	AM774496	AM779669	FR686640				
<i>Ctena orbiculata</i> (Montagu, 1808)	Long Key, Florida, USA	BMNH 20100254	AJ581853	AJ581887	FR686627				
<i>Ctena orbiculata</i> (Montagu, 1808)	Bermuda	BMNH 20100249	FR686714	FR686822	FR686635				
<i>Ctena imbricatula</i> (C. B. Adams 1845)	Bocas, Panama	BMNH 20100263	FR686715	FR686829	FR686636				

Table 1. Continued

Species	Locality	Locality code	Museum number	18S rRNA	28S rRNA	Cytb
<i>Ctena chiquita</i> (Dall, 1901)	Cabo Blanco, Costa Rica	COS	BMNH 20100256	FR686716	FR686830	FR686637
<i>Ctena</i> sp.	Pamilacan Island, Philippines. PANGLAO 2004, st S12, 9°29.4'N, 123°56.0'E, 6–8 m	PAN	MNH IM-2009-10365	FR686703	FR686821	FR686679
<i>Epicodakia tatei</i> (Angas, 1879)	Victor Harbor, South Australia		BMNH 20100275	FR686712	FR686828	FR686645
<i>Lucinoma annulata</i> (Reeve, 1850)	Southern California, USA		BMNH 20100251	FR686717		
<i>Lucinoma aequizonata</i> (Stearns, 1890)	500m, Santa Barbara Basin, California, USA		BMNH 20070307	FR686719	FR686820	FR686649
<i>Lucinoma borealis</i> (Linnaeus, 1767)	Salcombe, Devon, UK		BMNH 20070289	AM774501	AM779675	FR686662
<i>Lucinoma myriamae</i> Cosel, 2006	cold seep, 1700 m, off Nigeria	NIG	BMNH 20100293	FR686718	FR686818	FR686656
<i>Lucinoma myriamae</i> Cosel, 2006	off Angola, 6°34.15'S 7°8.48'E, 2050 m	ANG	BMNH 20100292	FR686713	FR686819	FR686661
Fimbrinae						
<i>Fimbrina fimbriata</i> (Linnaeus, 1758)	Lizard Island, Queensland, Australia		BMNH 20070293	AM774505	AM779679	FR686646
Monitilorinae						
<i>Monitilora ramsayi</i> (Smith, 1885)	Moreton Bay, Queensland, Australia		BMNH 20070292	AM774504	AM779678	FR686667
Lucininae						
<i>Austriella corrugata</i> (Deshayes, 1843)	King Bay, Dampier, Western Australia	DMP	BMNH 20070306	AJ581848	AJ581882	AM774139
<i>Austriella corrugata</i> (Deshayes, 1843)	Sungai Buloh, Singapore	SIN	BMNH 20070290	AM774128	AM779676	AM774140
<i>Bathyaustriella thionipta</i> Glover <i>et al.</i> , 2004	480 m, Kermadec Ridge, New Zealand		BMNH 20070337	AJ581858	AJ581892	AM774147
<i>Bretskya scapula</i> Glover & Taylor, 2007	Cortes, Bohol Island, Philippines. PANGLAO 2004, st T19, 9°42.2'N, 123°50.8'E, 10–26 m		IM-2009-10825	FR686722		FR686616
<i>Cardiolumina australopilula</i> Taylor & Glover, 1997	Rat Island, Houtman Abrolhos Islands, Western Australia		BMNH 20100262	FR686723	FR686785	FR686617
<i>Cardiolumina pisiformis</i> (Thiele, 1930)	Withnell Bay, Dampier, Western Australia		BMNH 20100269	AJ581849	AJ581883	FR686631
<i>Cardiolumina quadrata</i> (Prashad, 1932)	Off Balicasag Island, Bohol Sea, Philippines. PANGLAO 2005, st CP 2340, 9°29.4'N, 123°44.4'E. 271–318 m		MNH IM-2009-10374	FR686721	FR686787	FR686610
<i>Cardiolumina semperiana</i> (Issel, 1869)	Safage, Egypt		BMNH 20100301	FR686728	FR686786	FR686633
<i>Cardiolumina siquijorensis</i> Taylor & Glover, 1997	Taloto, Bohol Island, Philippines. PANGLAO 2004, st. M26, 9°40.9'N, 123°51.1'E. 0–2 m		MNH IM-2009-10366	FR686740	FR686789	FR686665
<i>Cardiolumina</i> undescribed species	Manga, Bohol Island, Philippines. PANGLAO 2004, st. S20, 9°41.8'N, 123°51.1'E. 10 m		MNH IM-2009-10361	FR686748	FR686788	FR686682

<i>Cavilinga blanda</i> (Dall & Simpson, 1901)	Lower Matecumbe Key, Florida, USA	BMNH 20100246	FR686724	FR686802	FR686623
<i>Chavania</i> sp.	Moreton Bay, Queensland, Australia	MB	FR686726	FR686790	FR686634
<i>Chavania striata</i> (Tokunaga, 1906)	Lizard Island, Queensland, Australia		AJ581850	AJ581884	FR686660
<i>Discolucina virginea</i> (Deshayes, 1832)	Tagbilaran, PANGLAO channel, Philippines. PANGLAO 2004, st. D13, 9°38.0'N, 123°51.4'E. 2–3 m		AM774497	AM779671	
<i>Divalunga bardwelli</i> (Iredale, 1936)	Denham, Shark Bay, Western Australia		FR686729	FR686793	FR686642
<i>Divalunga quadrilucata</i> (d'Orbigny, 1842)	Guadeloupe, French West Indies	tissue sample	AJ581854	AJ581888	FR686644
<i>Divaricella irpex</i> (Smith, 1885)	Roebuck Bay, Broome, Western Australia	BR	FR686730	FR686784	FR686643
<i>Divaricella irpex</i> (Smith, 1885)	Lizard Island, Queensland, Australia	LI	AJ581855	AJ581889	
<i>Funafutia levukana</i> (Smith, 1885)	Pamilacan Island, Philippines. PANGLAO 2004, st. B30, 9°29.4'N, 123°56.1'E. 25 m		FR686700	FR686782	FR686605
<i>IndoaustrIELla dalli</i> (Lynge, 1909)	Kungkraben Bay, Thailand		AM774130	AM774135	AM774143
<i>IndoaustrIELla lamprelli</i> Glover et al., 2008	Brisbane, Queensland, Australia		AM774131	AM774136	AM774144
<i>IndoaustrIELla plicifera</i> (Adams, 1855)	Tajong Laboh, Malaysia		AM774132	AM774137	AM774145
<i>IndoaustrIELla scarlatoi</i> (Zorina, 1974)	Tai Tam, Hong Kong		AJ581859	AJ581893	AM774146
<i>Lepidolucina venusta</i> (Philippi, 1847)	Port Dickson, Malaysia		FR686739	FR686806	FR686663
<i>Loripes clausus</i> (Philippi, 1848)	Ibo Island, Mozambique		FR686737	FR686792	FR686658
<i>Loripes lucinalis</i> (Lamarck, 1818)	Embiez Islands, France	FR	FR686735	FR686794	FR686655
<i>Loripes lucinalis</i> (Lamarck, 1818)	Weymouth, UK	UK	AJ581857	AJ581891	FR686654
<i>Lucina adansonii</i> d'Orbigny, 1839	Boavista Island, Cape Verde Islands		FR686731	FR686803	FR686648
<i>Lucina pensylvanica</i> (Linnaeus, 1758)	Lower Matecumbe Key, Florida	FK	AM774127	AM774138	AM774148
<i>Lucina pensylvanica</i> (Linnaeus, 1758)	Guadeloupe, French West Indies	GDL	AJ581856	AJ581890	FR686653
<i>Lucina pensylvanica</i> (Linnaeus, 1758)	Los Roquas, Venezuela	VEN	FR686738	FR686805	FR686659
<i>Lucina pensylvanica</i> (Linnaeus, 1758)	Bocas, Panama	BOC	FR686744	FR686804	FR686672
' <i>Lucina</i> ' <i>desiderata</i> Smith, 1885	Cortes, Bohol Island, Philippines. PANGLAO 2004, st T19, 9°42.2'N, 123°50.8'E. 10–26 m		FR686750	FR686783	
<i>Lucinella divaricata</i> (Linnaeus 1758)	Embiez Islands, France		FR686733	FR686801	FR686651
<i>Luciniscia centrifuga</i> (Dall, 1901)	Islas Perlas, Panama, Eastern Pacific		FR686732	FR686810	FR686650
<i>Luciniscia fenestrata</i> (Hinds, 1845)	Islas Perlas, Panama, Eastern Pacific		FR686734	FR686811	FR686652
<i>Luciniscia nassula</i> (Conrad, 1846)	Little Duck Key, Florida, USA		FR686736	FR686812	FR686657
<i>Parvilucina crenella</i> (Dall, 1901)	Ramrod Key, Florida, USA		FR686741	FR686799	FR686669

Table 1. Continued

Species	Locality	Locality code	Museum number	18S rRNA	28S rRNA	Cytb
<i>Parvilucina pectinella</i> (Adams, 1852)	Bocas, Panama		BMNH 20100257	FR686742	FR686798	FR686670
' <i>Parvilucina</i> ' <i>costata</i> (d'Orbigny, 1842)	Bocas, Panama	BOC	BMNH 20100252	FR686727	FR686809	FR686628
' <i>Parvilucina</i> ' <i>costata</i> (d'Orbigny, 1842)	Ile Cochon, Guadeloupe, French West Indies	GDL	BMNH 20100267	FR686725	FR686808	FR686619
<i>Phacoides pectinatus</i> (Gmelin, 1792)	Fort Pierce, Florida, USA		BMNH 20070291	AM774503	AM779677	FR686674
<i>Pillucina australis</i> Glover & Taylor, 2001	Rat Island, Houtman Abrolhos Islands, Western Australia		BMNH 20100294	FR686743	FR686800	FR686671
<i>Pillucina pisdium</i> (Dunker, 1860)	Gunnamatta Bay, New South Wales Australia		BMNH 20100284	AJ581865	AJ581898	
<i>Pillucina vietnamica</i> Zorina, 1978	Abu Dhabi, Arabian Gulf		BMNH 20080847		FR686796	FR686675
<i>Pillucina vietnamica</i> Zorina, 1978	Kungkraben Bay, Thailand		BMNH 20070282	AM774493	AM779666	FR686673
<i>Pillucina</i> sp.	Manga, Bohol Island, Philippines, PANGLAO 2004, st S19, 9°42.1'N, 123°51.4'E. 3–4 m	PAN	MNHN IM-2009-10362	FR686747	FR686795	FR686680
<i>Rasta lamyi</i> (Abrard, 1942)	Aqaba, Jordan					
<i>Radiolucina amianta</i> (Dall, 1901)	Ramrod Key, Florida, USA		BMNH 20070294	AJ581868	AM779680	FR686678
<i>Radiolucina cancellaris</i> (Philippi, 1846)	Islas Perlas, Panama, Eastern Pacific		BMNH 20100247	FR686745	FR686813	FR686676
<i>Stewartia floridana</i> (Conrad, 1833)	Cedar Key, West Florida, USA		BMNH 20100268	FR686746	FR686814	FR686677
<i>Troendleina</i> cf. <i>musculator</i> Cosel & Bouchet, 2008	Bohol/Sulu Seas sill, Philippines. PANGLAO 2005, st CP 2362, 8°56.5'N, 123°32.7'E. 679–740 m		BMNH 20100260	FR686749	FR686797	FR686684
<i>Wallucina assimilis</i> (Angas, 1867)	Jervis Bay, New South Wales, Australia		MNHN IM-2009-10367	FR686720	FR686807	FR686609
Out-groups			BMNH 20100260	AJ581869	FR686791	FR686687
<i>Eucrasatella cumingi</i> (Adams, 1852)	Moreton Bay, Queensland, Australia		BMNH20070212	AM774479	AM779653	
<i>Parathyasira equalis</i> (Verrill & Bush, 1898)	Gullmarsfjorden, Sweden		BMNH20070296	AM392453	AM392437	FR686685
<i>Thyasira polygona</i> (Jeffreys, 1864)	Northern North Saa		BMNH20070298	AM774484	AM392433	FR686686

Locality codes for Figures 1–5.

Table 2. Forward (F) and reverse (R) PCR primers (also used in sequencing), and forward (FS) and reverse (RS) internal sequencing primers

Gene/Primer	Sequence 5' → 3'	Annealing temp	[MgCl ₂]	Primer reference
18S rRNA		54 °C	3.0 mM	
18S-5' (F)	CTGGTTGATYCTGCCAGT			Winnepenninckx, Reid & Bacheljau, 1998
18S1100R (RS)	CTTCGAACCTCTGACTTTTCG			Williams, Reid & Littlewood, 2003
18S600F (FS)	GGTGCCAGCAGCCGCGGT			Williams & Ozawa, 2006
18S600R (RS)	ACCGCGGCTGCTGGCACC			Modified from Littlewood, Curini-Galletti & Herniou, 2000
28S rRNA		52 °C	2.5 mM	
LSU5 (F)	TAGGTCGACCCGCTGAAYTTAAGCA			Littlewood <i>et al.</i> , 2000
LSU1600R (R)	AGCGCCATCCATTTTCAGG			Williams <i>et al.</i> , 2003
900F (FS)	CCGTCTTGAAACACGGACCAAG			Lockyer <i>et al.</i> , 2003
ECD2S (RS)	CTTGGTCCGTGTTTCAAGACGG			Modified from Littlewood <i>et al.</i> , 2000
LSU330F (FS)	CAAGTACCGTGAGGGAAAGTTG			Littlewood <i>et al.</i> , 2000
cytochrome <i>b</i>		45 °C	3.0 mM	
cytB-F (F)	GRGGKGCTACKGTAATTACTAA			Taylor <i>et al.</i> , 2008
cytB-R (R)	AAATAYCAYTCNGGCTGRATATG			Taylor <i>et al.</i> , 2008
cytbR_new (R)	AAATAYCACTCGGGCTGGATATG			This study

Annealing temperatures and concentration of magnesium chloride [MgCl₂] used in polymerase chain reactions.

was determined using posterior probabilities (PP, calculated by MRBAYES).

We also analysed the combined data set ($N = 80$ taxa) using BEAST v1.5.1 (Drummond & Rambaut, 2007) to produce a phylogenetic hypothesis based on three genes (concatenated sequences from 18S, 28S and *cytb* genes) using Bayesian inference with an uncorrelated relaxed, lognormal clock. This method allows for the co-estimation of both phylogeny and divergence times, and is thought to result in better trees than Bayesian analysis alone (Drummond *et al.*, 2006). It also calculates the 95% highest posterior density (HPD) interval for node heights (ages). The 95% HPD is the shortest interval that contains 95% of the sampled values. Only one exemplar of each species was included, although in some cases it was necessary to concatenate sequences from different individuals of the same species. The Yule tree prior was used, which assumes a constant speciation rate among lineages, with a lognormal prior for the birth rate. Sequence variation was partitioned among genes, and gene-specific model parameters were used with each gene allowed to evolve at a different rate. Models were chosen after several preliminary analyses (GTR + G + I for 18S and 28S, and SRD06 for *cytb*). Based on MRBAYES analyses, we set the in-group to be monophyletic to ensure the correct placement of the root.

The starting trees for the BEAST analyses were neighbour-joining trees with branches transformed using nonparametric rate smoothing (NPRS) and node heights scaled so that all calibration points fell within the limits defined by the priors (NPRS transformed and scaled using TREEEDIT v1.0a10; A. Rambaut & M. Charleston, <http://evolve.zoo.ox.ac.uk>). The analysis ran for 125 000 000 generations with a sample frequency of 10 000. All effective sample size (ESS) values were greater than 100 (some were orders of magnitude greater). The final tree (Fig. 5) is a maximum clade credibility tree with node heights based on median values of 12 400 trees (after a burn-in of 100 generations), and support for nodes was determined using posterior probability (PP; calculated by BEAST).

MOLECULAR CLOCK CALIBRATION

The chronogram was calibrated using ten fossil records. Details for each calibration are listed below and positions are indicated on Figure 5. Calibrations with fossils using lognormal priors:

1. Date 1: '*Loripinus' conili* (de Raincourt, 1877), early Eocene, late Ypresian (55.8–48.6 Ma), Herouval, Paris Basin, France (Cossmann & Pissarro, 1904–06: fig. 82iv–3). To date crown Pegophyseminae (median, 48.6 Ma; SD 0.089 Myr;

- zero offset, 7.78 Myr; 95% interval, 65.6–48.6 Ma).
2. Date 2: '*Lucina*' *blanckenhorni* Chavan, 1947, late Cretaceous, Campanian (83.5–70.6 Ma), Mount of Olives, Jerusalem, Israel (Fig. 6D). To date Myrteinae stem group (median, 55.8 Ma; SD 0.125 Myr; zero offset, 12.13 Myr; 95% interval, 83.4–55.8 Ma).
 3. Date 3: *Lucinoma* sp., undescribed species (BMNH L64868–70), Palaeocene, Selandian–Thanetian, 62–59 Ma; Aquia Formation, Aquia Creek, Potomac River, Virginia, USA (Fig. 6H, I). To date first appearance of *Lucinoma* (median, 59 Ma; SD 0.105 Myr; zero offset, 10.98 Myr; 95% interval, 83.5–59 Ma).
 4. Date 4: '*Phacoides*' *dejaeri* (Vincent, 1930), Palaeocene (Danian 65–61 Ma), Calcaire de Mons, Belgium (Fig. 6K). Considered as possible *Codakia* stem group (median, 61 Ma; SD 0.0915 Myr; zero offset, 10.02 Myr; 95% interval, 83–61 Ma).
 5. Date 5: *Codakia leonina* (Basterot, 1825), early Miocene, Aquitanian (23–20.4 Ma) Saucats, Aquitaine Basin, France (Fig. 6L). To date crown group *Codakia* (median, 23 Ma; SD 0.06 Myr; zero offset, 2.55 Myr; 95% interval, 28.4–23 Ma).
 6. Date 6: *Epicodakia* sp., middle to late Eocene (Bartonian–Priabondian 40.4–33.9 Ma), Kalbarri, Western Australia (Fig. 6N) (Darragh & Kendrick, 2008). To date crown group *Ctena*/*Epicodakia* (median, 33.9 Ma; SD 0.11 Myr; zero offset, 6.57 Myr; 95% interval, 48.6–33.9 Ma).
 7. Date 7: *Lucinisca plesiolopha* Dall, 1900, middle Miocene (16–11.6 Ma), Alum Bluff Group, Florida (Gardner, 1926: pl. 19, figs 1, 2). To date crown group *Lucinisca* (median, 11.6 Ma; SD 0.345 Myr; zero offset, 5.701 Myr; 95% interval, 28.7–11.6 Ma).
 8. Date 8: *Loripes dujardini* (Deshayes, 1850), early Miocene, Aquitanian (23–20 Ma), Saucats, Aquitaine Basin, France (Fig. 6P). To date the stem of the *Loripes* clade (median, 20 Ma; SD 0.175 Myr; zero offset, 5.81 Myr; 95% interval, 34–20 Ma).
 9. Date 9: *Parvilucina sphaeriola* (Dall, 1903), middle Miocene (16–11.6 Ma), Alum Bluff Group, Florida, USA (Gardner, 1926: pl. 20, fig. 6). Date for the stem for North American *Parvilucina* (median, 11.6 Ma; SD 0.436 Myr; zero offset, 6.664 Myr; 95% interval, 33.9–11.6 Ma).
 10. Date 10: *Cardiolucina agassizi* (Michelotti, 1839), late Miocene (Tortonian 11.6–7.2 Ma), Southern Europe (Taylor & Glover, 1997: fig. 1). To date crown group *Cardiolucina* (median, 7.5 Ma; SD 0.275 Myr; zero offset, 3.125 Myr; 95% interval, 16–7.5 Ma).

RESULTS

Phylogenetic trees constructed from the separate gene data sets are shown in Figures 1–3. Not all taxa are represented in all three trees: for example, we failed to obtain 18S sequences for *Cryptophysema vesicula* (Gould, 1850) or *cytb* for *Pegophysema philippiana* (Reeve, 1850).

The tree derived from 18S rRNA data (Fig. 1) shows a clade of *Myrtea*, *Notomyrtea*, and *Gloverina* species. A clade containing species usually classified in the *Anodontia* group form a highly supported long branch, but within the clade species are also separated from each other on long branches. Notably, *Anodontia alba* does not group in this clade, but falls within a sister clade of morphologically heterogeneous taxa, including *Pseudolucinisca lacteola* (Tate, 1897), *Leucosphaera*, *Dulcina* species, and three separate undescribed taxa (UGS-1, UGS-2, and UGS-3).

Fimbriata fimbriata and *Monitilora ramsayi* (Smith, 1885) form separate branches sister to the rest of the lucinid species that are resolved into two large clades. One of these clades comprises species of *Ctena*, *Codakia*, and *Lucinoma*, and the second consists of a wide range of mainly shallow water lucinids. Sequences for species within this latter clade were highly conserved and there was little resolution of subclades. The position of *Phacoides pectinatus* (Gmelin, 1792) is notable on a long branch within this large clade.

The 28S rRNA tree (Fig. 2) showed a similar topology, but with higher resolution of the major clades and their internal structure. Ten '*Anodontia*' exemplars, representing five subgenera, are separated as a highly supported clade, with long internal branches, especially that of *Cryptophysema vesicula*. Again, *Anodontia alba* lies outside this clade. *Monitilora* and *Fimbria* form isolated branches. As in the 18S tree there are two major clades of lucinids, but subclade and generic divisions are much better resolved, as are the *Codakia*, *Ctena*, and *Lucinoma* subclades, for example. Again, *Phacoides pectinatus* occupies a long branch within the larger clade.

The tree derived from *cytb* data (Fig. 3) had essentially the same topography as the 18S and 28S trees, but provides finer detail at shallower nodes and clearer resolution of genera. The position of the long branch of *Euanodontia ovum* (Reeve, 1850), separate from the *Cavatidens*/*Cryptophysema* clade, is anomalous, but there is poor support for this topography. Notable within the largest lucinid clade is the well-supported subclade consisting of the genera *Loripes*, *Lucinella*, *Wallucina*, *Pillucina*, and *Chavania*, the subclade comprising species of *Austriella* and *Indoaustricola*, and another with six species of *Cardiolucina*.



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Figure 1. Molecular phylogeny of the Lucinidae based on the single gene analysis of 18S rRNA produced by Bayesian analysis using MRBAYES. Support values are Bayesian posterior probabilities (%). Details of the taxa are given in Table 1.



Figure 2. Molecular phylogeny of the Lucinidae based on the single gene analysis of 28S rDNA produced by Bayesian analysis using MRBAYES. Support values are Bayesian posterior probabilities (%). Details of taxa are given in Table 1.

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Figure 3. Molecular phylogeny of the Lucinidae based on the single gene analysis of cytochrome *b* produced by Bayesian analysis using MRBAYES. Support values are Bayesian posterior probabilities (%). Details of taxa are given in Table 1.

In the tree derived from the combined data set of the three genes (Fig. 4), the topology was similar to the individual gene trees, but support values were in most cases higher. The species analysed group into five highly supported clades (PP > 98%): the *Euanodontia/Cavatidens* group; the *Anodontia/Dulcina/Pseudolucinisca* group; the *Myrtea/Gloverina* group; the *Ctena/Lucinoma/Codakia* group; and the very large clade of other lucinids, with *Funafutia levukana* (Smith, 1885) as the sister group. Additional to these are *Monitilora ramsayi* and *Fimbria fimbriata*, which form separate unstable branches in the basal part of the tree. Within the two major clades generic divisions are well supported. Further discussion of the positions of genera within the major clades follows below.

With a smaller taxon set used in the BEAST chronogram analysis (Fig. 5), the five highly supported clades are very similar in topography to those in the combined tree.

DISCUSSION

Preliminary molecular results (Williams *et al.*, 2004; Taylor & Glover, 2006) showed major incongruence with previous classifications of Lucinidae based on shell characters (Chavan, 1969; Bretsky, 1976). Although traditional shell characters have long been and continue to be universally used for the recognition of genera and species, they have proven to be a poor guide to suprageneric relationships. Using the molecular results from the larger data set outlined in Figures 3 and 4, we propose a new hypothesis of relationships and classification of the Lucinidae (Table 3). Although we have sampled only 53% of living genera, the major groups identified are robust and well supported in our analysis. We classify the major clades recognized as subfamilies of Lucinidae. Family group names are already available for five of these clades, and we introduce three new names. Additionally, included in the classification are living genera for which no molecular data is yet available (Table 3). The placement of these is provisional but testable by future molecular analysis.

PEGOPHYSEMINAE – NEW SUBFAMILY

Except for *Anodontia alba* (type species of *Anodontia*), all other species analysed that have been previously classified as *Anodontia* are separate on a highly supported long branch. Within this clade, the species previously classified into various subgenera of *Anodontia*, namely *Loripinus*, *Pegophysema*, *Euanodontia*, *Cryptophysema*, and *Cavatidens* also lie on long internal branches (most taxa included in 28S tree; Fig. 2). Because of their phylogenetic distinctive-

ness we propose a new subfamily Pegophyseminae, and in consideration of the differences from *Anodontia alba* we elevate the subgenera listed above, together with *Afropysema* and *Bythosphaera* (Taylor & Glover, 2005), to full generic status. The type species of *Pegophysema* is *Pegophysema schrammi* (Crosse, 1876), and an 18S sequence from this species (donated by D. Distel, specimen from Bermuda) grouped with other pegophysemine species in the analysis of Williams *et al.* (2004). A closely similar species from the Indo-West Pacific, *Pegophysema philippiana*, is included in the 28S and 18S trees herein.

Pegophyseminae are united by some unusual morphological characters, including thin, globular, smooth shells, a narrow edentulous hinge with an internal and laterally extended ligament in most species, and, anatomically, the presence of a mantle septum, digitate mantle gills, and extensive posterior mantle fusion (Taylor & Glover, 2005).

Although the chronogram (Fig. 5) suggests a late Cretaceous origin, there are no convincing records of Pegophyseminae bivalves in pre-Cenozoic rocks. Records for the Cenozoic are frequent, and contemporaneously in the Eocene of the Paris Basin and southern USA (Cossmann & Pissarro, 1904–06; Gardner, 1951), there were lucinids with characters of living Pegophyseminae such as *Anodontia augustana* Gardner, 1951, '*Lucina*' *conili* de Raincourt, 1877, and '*Lucina*' *parnensis* Deshayes, 1857. Pre-Cenozoic species with a superficial resemblance to pegophyseminae include the hydrocarbon seep-associated '*Cryptolucina*' *kuhnpassetensis* Kelly, 2000 from the Cretaceous of Greenland, which is inflated and edentulous (Kelly *et al.*, 2000). From the Palaeozoic, Boyd & Newell (1979) compared the Permian *Gigantocyclus zidensis* Termier & Termier, 1977, from the Permian of Tunisia with *Anodontia*, and the Devonian *Paracyclas* species, placed by Johnston (1993) into the family Paracyclidae, are similar in general shape to living taxa. There is insufficient evidence to include any of these genera in the Pegophyseminae.

LEUCOSPHERINAE NEW SUBFAMILY

This group of species forms a distinct, well-supported clade in the molecular analysis, but there are no clear and consistent morphological characters that unite the group. We propose a new subfamily name Leucospherinae for this clade, based on the genus *Leucosphera*. Unfortunately, an obvious subfamily name based on *Anodontia* could not be used because of the prior use for the freshwater mussel *Anodonta*.

Lucinids in this clade have no radial shell sculpture, cardinal teeth are usually small or absent, lateral teeth are lacking, whereas the lunule is prominent and highly asymmetrical in *Pseudolucinisca*,

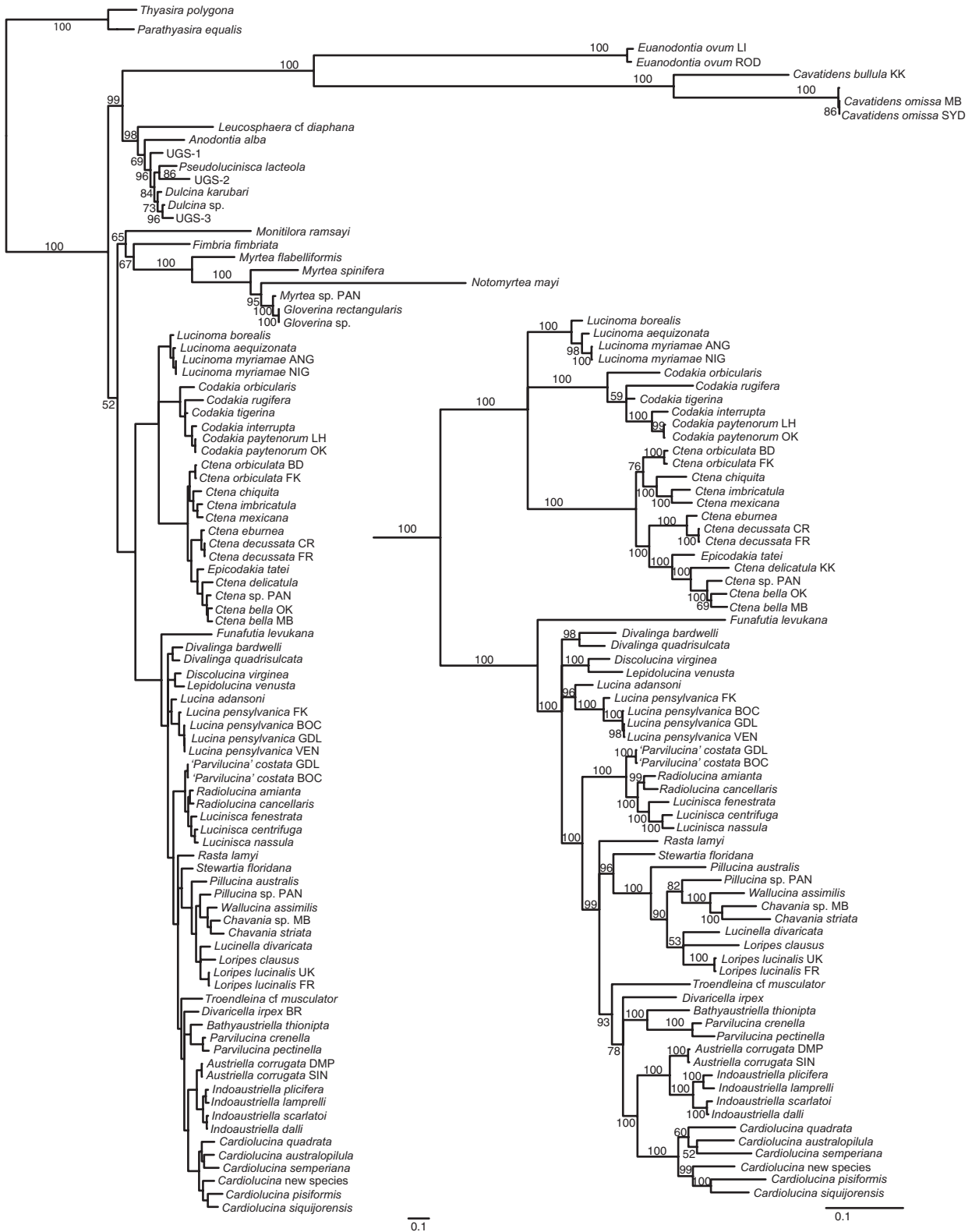


Figure 4. Molecular phylogeny of Lucinidae produced by Bayesian analysis of the combined data set from concatenated sequences of the nuclear 18S rRNA and 28S rRNA, and mitochondrial cytochrome *b* genes, using Thyasiridae species as out-groups. *Phacoides pectinatus* is excluded from this tree because of its unstable position within the large clade. The inset to the right shows details of the larger clades. Support values are Bayesian posterior probabilities (%). Details of taxa in Table 1.

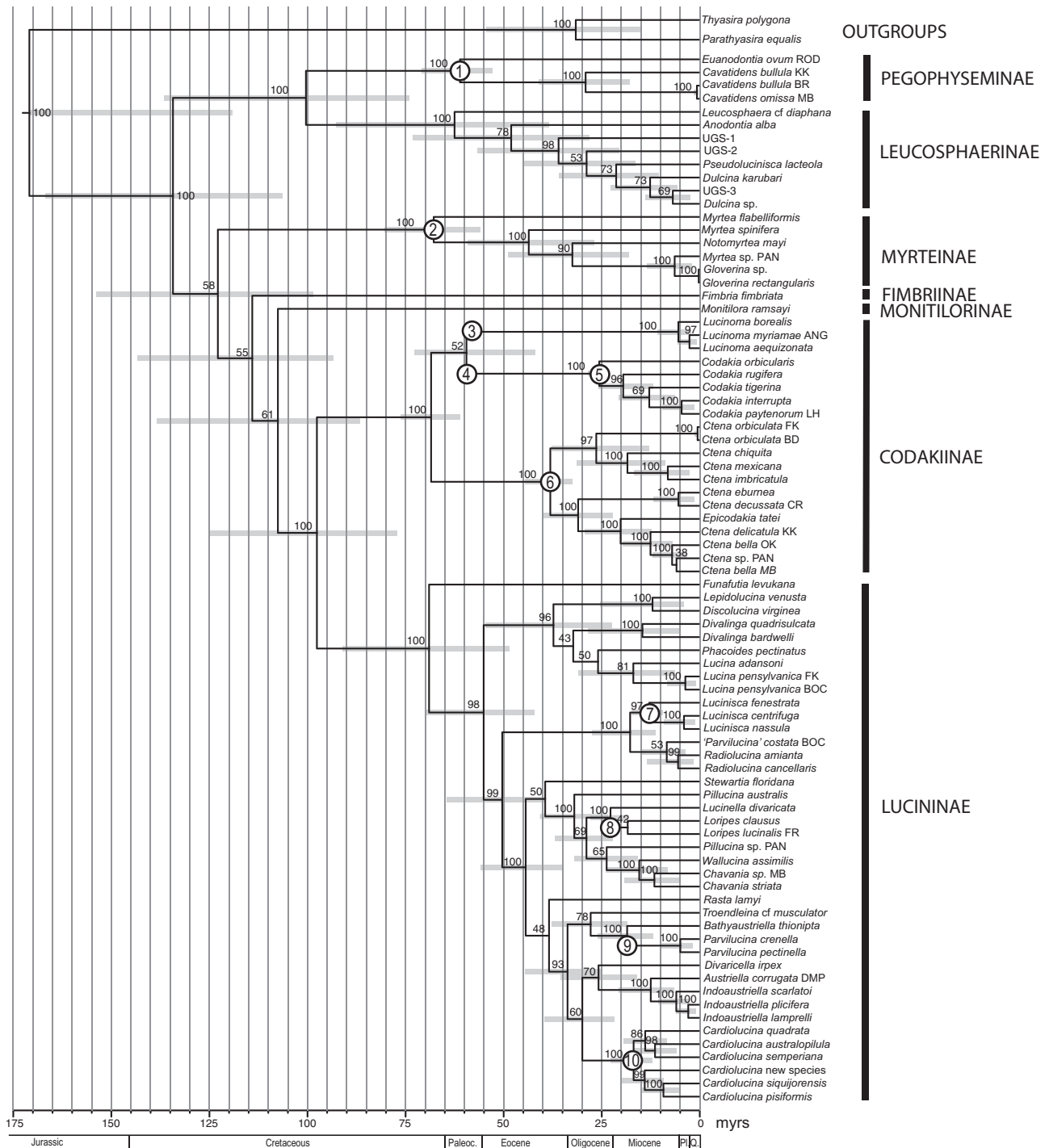


Figure 5. Chronogram generated by BEAST for species of Lucinidae, and calibrated using ten fossil records. The position of each calibration point is indicated by the numbers in circles (see the Material and methods and Discussion sections for details about the fossils). Error bars for node ages are the 95% highest posterior density intervals; clades with less than 50% support have no error bars; the root node has node bar trimmed by 50% for clarity. New subfamily divisions are shown on the right-hand side, with details presented in Table 3.

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Table 3. Proposed classification of Lucinidae based on the molecular analyses and including those genera with living representatives only

Lucinidae Fleming, 1828

PEGOPHYSEMINAE new subfamily Taylor & Glover herein

Pegophysema Stewart, 1930 (*Lucina schrammi* Crosse, 1876)

Cavatidens Iredale, 1930 (*Cavatidens omissa* Iredale, 1830)

Cryptophysema Taylor & Glover, 2005 (*Lucina vesicula* Gould, 1850)

Euanodontia Taylor & Glover, 2005 (*Lucina ovum* Reeve, 1850)

Loripinus Monterosato, 1883 (*Lucina fragilis* Philippi, 1836)

Unconfirmed:

Afrophysema Taylor & Glover, 2005 (*Cryptodon eutornus* Tomlin, 1921)

Bythosphaera Taylor & Glover, 2005 (*Cryptodon watsoni* Smith, 1885)

Meganodontia Bouchet & Cosel 2004 (*Meganodontia acetabulum* Bouchet & Cosel, 2004)

LEUCOSPHERINAE new subfamily Taylor & Glover herein

Leucosphaera Taylor & Glover, 2005 (*Lucina (Loripinus) salamanensis* Thiele & Jaekel, 1931)

Anodontia Link, 1807 (*Anodonta alba* Link, 1807)

Dulcina Cosel & Bouchet, 2008 (*Dulcina guidoi* Cosel & Bouchet, 2008)

Pseudolucinisca Chavan, 1959 (*Lucina lacteola* Tate, 1897)

Undescribed genera – UGS-1, UGS-2, UGS-3

Unconfirmed:

Afrolucina Cosel, 2006 (*Afrolucina discontinua* Cosel, 2006)

Alucinoma Habe, 1958 (*Alucinoma soyae* Habe, 1958)

Callucina Dall, 1901 (*Lucina radians* Conrad, 1841)

Epidulcina, Cosel & Bouchet, 2008 (*Epilucina delphinae* Cosel & Bouchet, 2008)

Minilucina, Cosel & Bouchet, 2008 (*Minilucina coriolis* Cosel & Bouchet, 2008)

Neophysema Taylor & Glover, 2005 (*Neophysema aphanes* Taylor & Glover, 2005)

Semelilucina Cosel & Bouchet, 2008 (*Semelilucina semeliformis* Cosel & Bouchet, 2008)

Tinalucina Cosel, 2006 (*Tinalucina aequatorialis* Cosel, 2006)

MYRTEINAE Chavan, 1969

Myrtea Turton, 1822 (*Venus spinifera* Montagu, 1803)

Gloverina Cosel & Bouchet, 2008 (*Gloverina vestifex* Cosel & Bouchet, 2008)

Notomyrtea Iredale, 1924 (*Myrtea botanica* Hedley, 1918)

Unconfirmed:

Elliptiolucina Cosel & Bouchet, 2008 (*Elliptiolucina magnifica* Cosel & Bouchet, 2008)

Eulopia Dall, 1901 (*Lucina sagrinata* Dall, 1886)

Gonimyrtia Marwick, 1929 (*Loripes concinna* Hutton, 1885)

Graecina Cosel, 2006 (*Graecina karinae* Cosel, 2006)

Jorgenia Taylor & Glover, 2009a (*Jorgenia louisiana* Taylor & Glover, 2009a)

Myrtina, Glover & Taylor, 2007 (*Myrtina porcata* Glover & Taylor, 2007)

Poumea Glover & Taylor, 2007 (*Poumea coselia* Glover & Taylor, 2007)

Rostrilucina, Cosel & Bouchet, 2008 (*Rostrilucina garuda* Cosel & Bouchet, 2008)

Solelucina Glover & Taylor, 2007 (*Solelucina koumacia* Glover & Taylor, 2007)

Taylorina, Cosel & Bouchet, 2008 (*Taylorina alata* Cosel & Bouchet, 2008)

MONITILORINAE new subfamily Taylor & Glover herein

Monitilora Iredale, 1930 (*Lucina ramsayi* Smith, 1885)

Unconfirmed:

Prophetilora Iredale, 1930 (*Prophetilora arizela* Iredale, 1930)

FIMBRIINAE Nicol, 1950

Fimbria Mühlfeld, 1811 (*Fimbria magna* Mühlfeld, 1811)

CODAKIINAE Korobokov, 1954

Codakia Scopoli 1777 (*Venus orbicularis* Linnaeus, 1758)

Ctena Mörch, 1861 (*Codakia mexicana* Dall, 1901)

Epicodakia Iredale, 1930 (*Epicodakia conssettiana* Iredale, 1930)

Epilucina Dall, 1901 (*Lucina californica* Conrad, 1887)

Lucinoma Dall, 1901 (*Lucina filosa* Stimpson, 1851)

Unconfirmed:

Epilucina Dall, 1901 (*Lucina californica* Conrad, 1887)

Table 3. *Continued*

LUCININAE Fleming, 1828

- Austriella* Tenison-Woods, 1881 (*Austriella sordida* Tenison-Woods, 1881)
Bathyaustriella Glover, Glover *et al.* 2004 (*Bathyaustriella thionipta* Glover *et al.*, 2004)
Bretskyia Glover & Taylor, 2007 (*Bretskyia scapula* Glover & Taylor, 2007)
Cardiolucina Sacco, 1901 (*Cardium agassizi* Michelotti, 1839)
Cavilinga Chavan, 1937 (*Lucina trisulcata* Conrad, 1841)
Chavania Glover & Taylor, 2001 (*Lucina erythraea* Issel, 1869)
Discolucina Glover & Taylor, 2007 (*Lucina virginiae* Deshayes, 1832)
Divalinga Chavan, 1951 (*Lucina quadrisulcata* d'Orbigny, 1846)
Divaricella Martens, 1880 (*Lucina (Divaricella) angulifera* Martens, 1880)
Funafutia Glover & Taylor, 2001 (*Lucina levukana* Smith, 1885)
Indoaustriella Glover *et al.*, 2008 (*Lucina plicifera* Adams, 1855)
Lepidolucina Glover & Taylor, 2007 (*Lucina venusta* Philippi, 1847)
Loripes Poli, 1791 (*Loripes lacteus* Poli, 1791)
Lucina Bruguière, 1797 (*Venus pensylvanica* Linnaeus, 1758)
Lucinella Monterosato, 1883 (*Lucina commutata* Philippi, 1836)
Lucinisca Dall, 1901 (*Lucina nassula* Conrad, 1846)
Parvilucina Dall, 1901 (*Lucina tenuisculpta* Carpenter, 1864)
Phacoides Gray, 1847 (*Tellina pectinata* Gmelin, 1791)
Pillucina Pilsbry, 1921 (*Pillucina spaldingi* Pilsbry, 1921)
Radiolucina Britton, 1972 (*Phacoides amiantis* Dall, 1901)
Rasta Taylor & Glover, 2000 (*Rastafaria thiophila* Taylor & Glover, 1997)
Stewartia Olsson & Harbison, 1953 (*Lucina anodonta* Say, 1824)
Troendleina Cosel & Bouchet, 2008 (*T. marquesana* Cosel & Bouchet, 2008)
Wallucina Iredale, 1930 (*Lucina jacksoniensis* Smith, 1885)
 Unconfirmed:
Barbierella Chavan, 1938 (*Lucina barbieri* Deshayes, 1857)
Bourdotia Dall, 1901 (*Lucina bourdoti* Cossmann, 1882)
Cavilucina Fischer, 1887 (*Lucina sulcata* Lamarck, 1806)
Divalucina Iredale, 1936 (*Cyclas cumingi* A. Adams, 1864)
Falsolucinoma Cosel, 2006 (*Lucina leloeuffi* Cosel, 1989)
Ferrocina Glover & Taylor 2007 (*Ferrocina multiradiata* Glover & Taylor, 2007)
Gibbolucina Cossmann, 1904 (*Venus callosa* Lamarck, 1806)
Here Gabb, 1866 (*Lucina richtofeni* Gabb, 1866)
Joellina Cosel, 2006 (*Joellina dosiniformis* Cosel, 2006)
Keletistes Oliver, 1986 [*Loripes (Keletistes) rhizoecus* Oliver, 1986]
Lamellolucina Taylor & Glover, 2006 (*Lamellolucina pilbara* Taylor & Glover, 2002)
Lamylucina Cosel, 2006 [*Phacoides (Lucinoma) gainei* Lamy, 1920]
Liralucina Glover & Taylor, 2007 (*Phacoides sperabilis* Hedley, 1909)
Nevenulora Iredale, 1930 (*Lucinida hilaira* Hedley, 1917)
Parvidontia Glover & Taylor, 2007 (*Parvidontia laevis* Glover & Taylor, 2007)
Pleurolucina Dall, 1901 (*Lucina leucocyma* Dall, 1886)
Plicolucina Glover *et al.*, 2004 (*Plicolucina flabellata* Glover *et al.*, 2004)
Pompholigina Dall, 1901 (*Lucina gibba* Gray, 1825)
 ?Milthinae Chavan, 1969
Miltha H. & A. Adams, 1857 (*Lucina childreni* Gray, 1824)
Eomiltha Cossmann, 1912 (*Lucina contorta* DeFrance, 1823)

Genera listed as 'Unconfirmed' are without molecular confirmation of position. Type species of genera in parentheses.

Leucosphaera, and *Dulcina*. The clade includes *Anodontia alba* from the western Atlantic, *Pseudolucinisca lacteola* from southern Australia (Glover & Taylor, 2008), and the small, semitransparent, thin-shelled *Leucosphaera*, a common but little recorded

lucinid in offshore fine sediments of the Indo-West Pacific, and previously confounded with juvenile 'Anodontia' (see Taylor & Glover, 2005; Glover & Taylor, 2007). Additionally, the group includes some deeper water forms including species of *Dulcina*,

together with three undescribed species and genera (UGS-1, UGS-2, and UGS-3).

It was initially surprising that *Anodontia alba* (Figs 6B, 7F) did not group with the superficially similar taxa, usually called 'Anodontia' (Williams *et al.*, 2004), now included here in the Pegophyseminae (see above). *Anodontia alba* and the eastern Pacific sister species *Anodontia edentuloides* (Verrill, 1870), however, differ from pegophysemines in both shell characters and internal anatomy, for example lacking the mantle septum and digitate mantle gills (Taylor & Glover, 2005). The Eocene (Ypresian) species *Eophysema cossmanni* Chavan, 1941, from the Paris Basin, classified by Strougo (1975) into a new genus *Rawya* (type species *Lucina pharaonis* Bellardi, 1854), has many characters of *Anodontia alba*, but, in addition, has small cardinal teeth (Fig. 6A). We regard this species as a possible early representative of the *Anodontia alba* lineage. Cardinal teeth are absent in recent species, but they possess a smooth triangular area beneath the umbo ('aire dentaire' of Chavan, 1937–1938), corresponding to the position of teeth in dentate taxa. From this evidence we suggest that the *Anodontia alba* clade was separate from the Pegophyseminae at least by the mid-Eocene. The chronogram (Fig. 5) suggests a much older divergence of the clades in the late Cretaceous, at 100 ± 25 Ma.

Other genera in this clade have a poor or unrecognized fossil record, including the laterally compressed, edentulous deep-water *Dulcina* and *Alucinoma* (Cosel & Bouchet (2008). Two Miocene species, *Lucina strigosa* Michelotti, 1861 and *Lucina michelotti* Mayer, 1858 were assigned to *Pseudolucinisca* by Chavan (1959), but these species possess lateral teeth, unlike any living forms (Glover & Taylor, 2008).

MYRTEINAE CHAVAN, 1969

Myrteinae comprise a diverse, poorly studied group of lucinids, usually associated with deeper water, offshore habitats. Chavan (1969) included 11 genera within his subfamily Myrteinae: five of these (*Myrtea*, *Gonimyrrtea*, *Monitilora*, *Lucinoma*, and *Prophetilora*) with living species and six others including only fossil species. Our molecular results indicate that *Monitilora* and *Lucinoma* group in separate subfamilies, Monitilorinae and Codakiinae, respectively, and are discussed in detail below. The position of *Gonimyrrtea* and *Prophetilora* remains uncertain, pending further molecular analysis. We analysed six species classified in the genera *Myrtea*, *Notomyrrtea*, and *Gloverina*, the latter a recently described genus (Cosel & Bouchet, 2008) included here in the Myrteinae for the first time. Our results (Figs 1–5)

show that Myrteinae form a highly supported clade sister to all other lucinids, except the Pegophyseminae and Leucosphaerinae.

In our new concept, Myrteinae share some common morphological features, such as thin, elongate shells and regularly spaced commarginal lamellae that are often elevated above the dorsal shell margin. Some species have radial folds between the lamellae, but never strong radial ribbing. Additionally, they have short, anterior adductor muscle scars, narrow hinges with small to obscure cardinal teeth, and in some species a clearly asymmetric lunule.

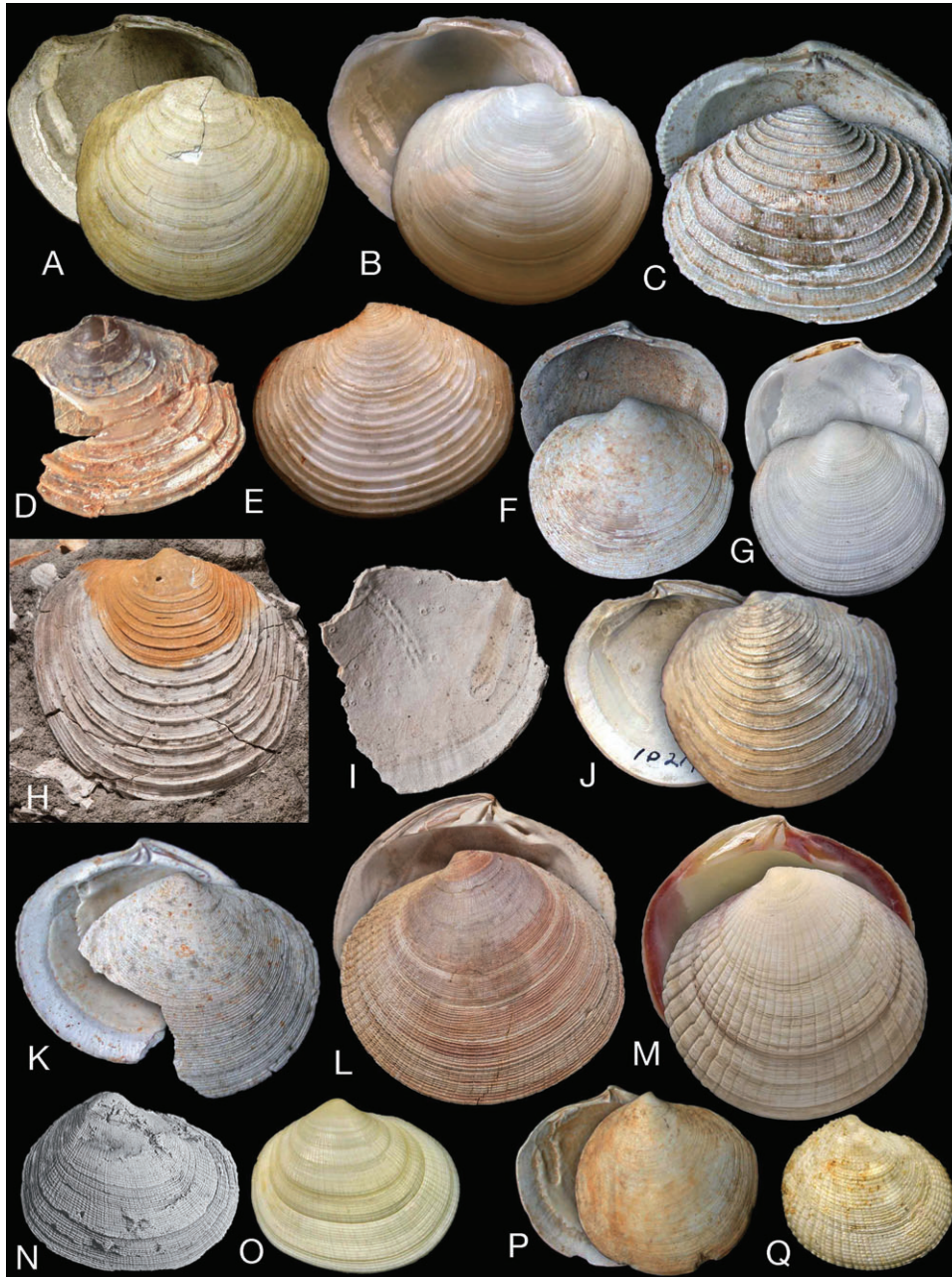
Other newly described genera from the deep sea, including *Elliptolucina*, *Taylorina*, *Rostrilucina*, *Graecina* (Cosel, 2006; Cosel & Bouchet, 2008), and *Jorgenia* (Taylor & Glover, 2009a), may also belong in Myrteinae, but no suitable material for molecular analysis was available. Similarly, some shallow water genera, such as *Myrtina* and *Solelucina* (Glover & Taylor, 2007), have shell characters consistent with inclusion in Myrteinae.

Myrteinae are poorly documented in the fossil record, but some *Myrtea*-like bivalves are present throughout the Cenozoic, and *Lucina blanckenhorni* Chavan, 1947 from the late Cretaceous (Campanian 83.5–70.6 Ma) may be part of a 'Myrtea' stem group. Somewhat earlier, *Paramyrtea sabulosa* Kendrick & Vartak, 2007, from the mid-Cretaceous (Cenomanian) of India, has shell characters compatible with inclusion in the Myrteinae. The chronogram (Fig. 5) indicates a late Cretaceous radiation for the subfamily, with a possible separation from other lucinids earlier in the Cretaceous.

FIMBRIINAE NICOL, 1950

Although represented today by only two living species of *Fimbria*, the group has a long fossil history dating back to the early Jurassic. Chavan (1969) separated living and fossil genera as a distinct family Fimbriidae, but our molecular results for *Fimbria fimbriata* indicate that it falls within the Lucinidae. Living *Fimbria* have distinctive thick, ovate, anteriorly extended shells, with a sculpture of narrow radial ribs that are more prominent at the anterior, and posteriorly these are crossed by closely spaced, rounded commarginal lamellae. The hinge is broad, with large cardinal teeth, and with anterior and posterior lateral teeth in both valves, whereas the anterior adductor scar is very short and scarcely detached from the pallial line.

In the chronogram (Fig. 5) *Fimbria fimbriata* aligns as a sister taxon to *Monitilora ramsayi*, Codakiinae, and Lucininae, with a late Jurassic–early Cretaceous divergence. There is a remarkable continuity of shell morphology of *Fimbria*-like



species throughout the Cenozoic and Mesozoic, with bivalves possessing some characters of living *Fimbria* (but not necessarily congeneric) recorded back to the early Jurassic (Pliensbachian–Toarcian) (Dubar, 1948; Monari, 2003). Less certain are the even earlier fossils, for example the late Triassic *Schafhaeutlia* with huge cardinal teeth (Hautmann, 2001) that have been placed in Fimbriidae (e.g. Chavan, 1969), although we can see no convincing evidence for inclusion. Equally, we have doubts concerning the lucinid status of other Mesozoic genera

Sphaera, *Sphaeriola*, and *Haastina* that have been classified in the Fimbriidae.

MONITILORINAE NEW SUBFAMILY

Living *Monitilora* are known from eastern Australia, the type species *Monitilora ramsayi* (Smith, 1885), and two unnamed species from South-East Asia. The single species included in the molecular analysis, *Monitilora ramsayi* (Fig. 6G), forms a separate branch with an unstable position, but never aligns

Figure 6. Some fossil lucinids used to calibrate the chronogram or mentioned in the Discussion section, and comparable Recent species. Not to scale. Actual shell lengths (L). Institutional abbreviations: ANSP, Academy of Natural Sciences, Philadelphia; BMNH, the Natural History Museum, London; MNHN, Museum National d'Histoire Naturelle, Paris; RBINS, Royal Belgian Institute of Natural Sciences, Brussels; WAM, Western Australian Museum. A. *Eophysema cossmanni* Chavan, 1941. Eocene (Ypresian), St-Gobain, Aisne, France (MNHN B 53241). Also named *Rawya cossmanni* by Strougo (1975). L = 29.6 mm. B. *Anodontia alba* (Link, 1807). Recent, western Atlantic. Syntype of *Lucina anatelloides* Reeve, 1850 (BMNH 1963553/1). L = 38.6 mm. C. *Fimbria montensis* (Cossmann, 1908). Palaeocene (Danian), Calcaire de Mons, Belgium (RBINS IG5496), exterior and interior of right valve. L = 32.4 mm. D. '*Lucina*' *blanckenhorni* (Chavan, 1947). Cretaceous (Campanian), Mount of Olives, Israel (RBINS IG 21.735). L = 13.2 mm. E. *Myrtea spinifera* (Montagu, 1803). Recent, Milford Haven, UK (BMNH). L = 25.7 mm. F. *Monitilora duponti* (Cossmann, 1908). Palaeocene (Danian), Calcaire de Mons, Belgium. Exterior and interior of left valve (RBINS IG 6544). L = 19.2 mm. G. *Monitilora ramsayi* (Smith, 1885). Botany Bay, New South Wales, Australia (BMNH 1963207). L = 14.5 mm. H. *Lucinoma* sp. undescribed species. Palaeocene (Selandian–Thanetian), 62–59 Ma, Aquia Formation, Aquia Creek, Potomac River, Virginia, USA (BMNH L64868–70). Exterior of left valve. L = 29.5 mm. I. *Lucinoma* sp., details as in (H). Interior of left valve with detail of elongate anterior adductor scar and impression of pallial blood vessel. Muscle scar length = 15.2 mm. J. *Lucinoma filosa* (Stimpson, 1851). Recent, off Martha's Vineyard, MA, USA (ANSP 102172). Exterior of left and interior of right valves. L = 37.0 mm. K. '*Phacoides*' *dejaeri* (Vincent, 1930). Palaeocene (Danian), Calcaire de Mons, Belgium (RBINS IG5611). Exterior and interior of right valve. Possible member of *Codakia* stem group. L = 22.8 mm. L. *Codakia leonina* (Basterot, 1825). Miocene (Aquitainian), Saucats, France (BMNH) L = 45.4 mm. M. *Codakia punctata* (Linnaeus, 1758). Recent, Tuoho, New Caledonia (MNHN). L = 64.0 mm. N. *Epicodakia* sp. Eocene (Bartonian–Priabondian), Kalbarri, Western Australia. Exterior right valve (WAM 05.182). From Darragh & Kendrick, 2008: fig. 1.20). L = 10.4 mm. O. *Epicodakia consettiana* Iredale, 1930. Recent, Houtman Abrolhos Islands, Western Australia (BMNH). L = 18.0 mm. P. *Loripes dujardini* (Deshayes, 1850). Middle Miocene, Manthelan, Loire Basin, France. Exterior and interior of right valve (RBINS IG 10591). L = 18 mm. Q. *Ctena squamosa* (Lamarck, 1806). Oligocene (Rupelian), Etampes, France (BMNH). L = 6.2 mm.

within any other of the major clades. In the BEAST chronogram (Fig. 5) it forms a sister branch to the combined Codakiinae and Lucininae. Chavan (1969) placed *Monitilora* within the Myrteinae, but our analysis shows that *Monitilora* is distinct from Myrteinae, and in view of its phylogenetic position on a separate branch we propose a new subfamily Monitilorinae for this distinctive and long-lived clade. In shell characters *Monitilora* species are subcircular, moderately inflated, with regularly spaced, low, commarginal lamellae that have fine radial folds in the interspaces. The lunule is large and asymmetrical, the cardinal teeth are small, and there is an anterior lateral tooth in the right valve only.

Apart from *Monitilora ramsayi*, the species from southern Australia that have been classified as *Monitilora* (e.g. Lamprell & Whitehead, 1992) should be placed in other genera. '*Monitilora*' *adelaidiana* Cotton & Godfrey, 1938 is similar to *Callucina*, whereas '*Monitilora*' *paupera* (Tate, 1892) probably belongs with the *Myrtea* group. Another little-studied genus, *Prophetilora* (type species *Prophetilora arizela* Iredale, 1930 = *Lucina simplex* Reeve, 1850), has been classified as a subgenus of *Monitilora* (Lamprell & Whitehead, 1992), but no specimens were available for molecular analysis, and its position is uncertain.

Shells with characters very similar to the living *Monitilora ramsayi* can be recognized up to the early Palaeocene (Danian), for example, *Monitilora duponti*

(Cossmann, 1908) (Fig. 6F) from the Calcaire de Mons, Belgium, was previously classified in the genus by Chavan (1937–1938). Our analysis suggests that Monitilorinae split from other lucinids at least by the mid-Cretaceous (Fig. 5). Some *Monitilora*-like bivalves were present in the Mesozoic, but have been placed in the genus *Mesomiltha* Chavan, 1952 (type species *Lucina pulchra* Zittel & Goubert, 1861, Jurassic, Oxfordian): an example being *Mesomiltha* cf. *fallax* (Forbes, 1846), from the Cretaceous (Cenomanian) of India (Kendrick & Vartak, 2007: fig. 131–M), but a more detailed study of these species is needed.

CODAKIINAE KOROBKOV, 1954

This family group name was proposed by Korobkov (1954) to include *Codakia* and *Ctena*, but not *Lucinoma*. Molecular results (Figs 1–5) demonstrate that this is a highly supported, large clade that includes species of *Lucinoma*, *Codakia*, *Ctena*, and *Epicodakia*. Prior morphological analyses (Bretsky, 1970, 1976) did not recognize this grouping, first reported from molecular results by Williams *et al.* (2004). The clade encompasses three distinct, highly supported subclades: one comprising species of *Lucinoma*, another comprising species of *Codakia*, and a third consisting of *Ctena* and *Epicodakia*. The two latter groups include some of the most abundant, tropical, shallow water lucinids, particularly from seagrass habitats.

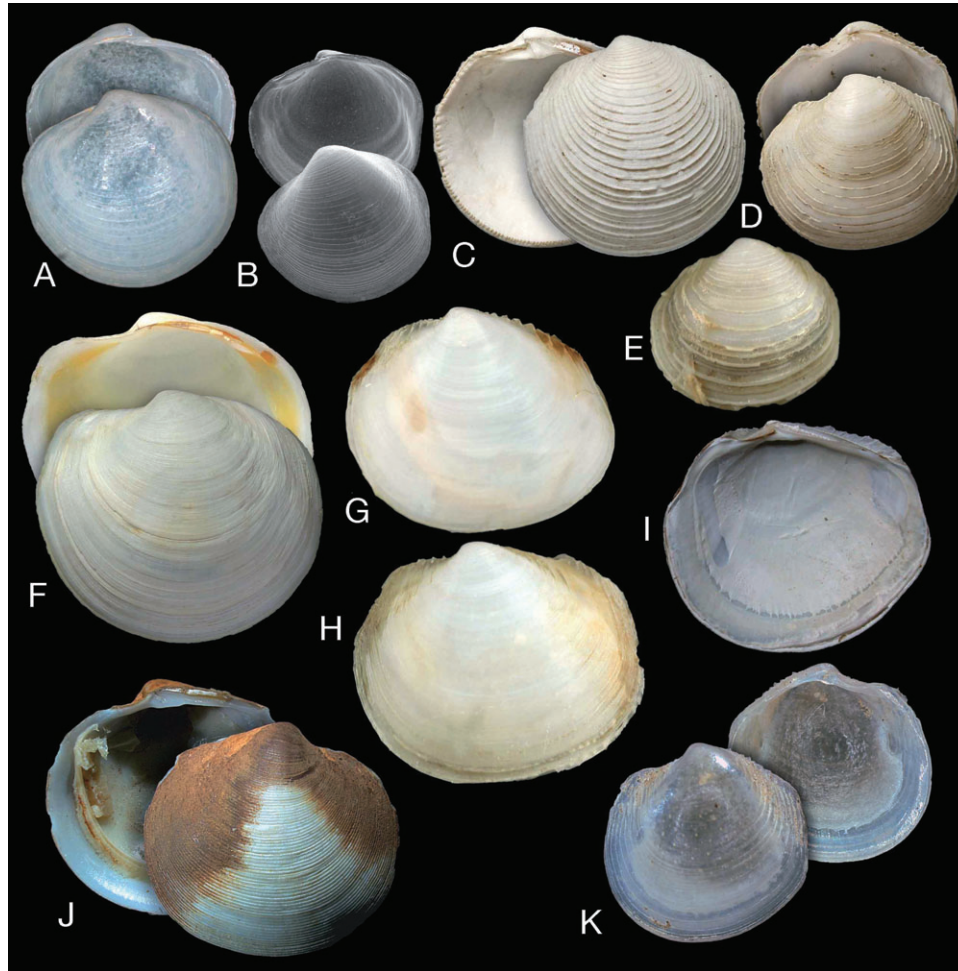


Figure 7. Representatives of the new subfamily Leucosphaerinae including some of the sequenced specimens. Not to scale. L = actual shell lengths. A. *Leucosphaera* cf. *diaphana* Glover & Taylor 2007. Shell from same station as sequenced specimen. Ubajan, Bohol Island, Philippines. PANGLAO 2004, st. S27, 9°41.5'N, 123°51.0'E, 12 m (MNHN). L = 4.9 mm. B. *Leucosphaera diaphana* Glover & Taylor 2007, holotype, New Caledonia (MNHN). L = 6.0 mm. C. *Pseudolucinisca lacteola* (Tate, 1897) (syntype of *Lucina concentrica* Reeve, 1850, St Vincent's Gulf, South Australia (BMNH 1870.10.26.33). L = 21.5 mm. D. Undescribed genus and species (UGS-1). Shells from same station as sequenced specimen figured in E (MNHN). L = 10.3 mm. E. Undescribed genus and species (UGS-1). Maribojoc Bay, Bohol Sea, PANGLAO 2005, st CP 2397, 9°34.9'N, 123°41.7'E. 642–669 m (MNHN IM-2009-10373). L = 7.0 mm. F. *Anodontia alba* Link 1807 'West Indies' (BMNH 1963476-2). L = 53mm. G. *Dulcinea* sp. Bohol Sea, Philippines. PANGLAO 2005, st. CP 2335, 9°34.3'N, 123°37.8'E. 729–733 m (MNHN IM-2009-10371). L = 17 mm. H. *Dulcinea karubari* Cosel & Bouchet, 2008. Maribojoc Bay, Bohol Sea, Philippines. PANGLAO 2005, st. CP 2331, 9°39.2'N, 123°47.5'E. 255–268 m (MNHN IM-2009-10372). L = 16 mm. I. *Dulcinea karubari* Cosel & Bouchet, 2008. Holotype, Tanimbar Islands, Indonesia. KARUBAR, st. CP 63, 8°00'S, 132°58'E. 215 m (MNHN 20729). Right valve to show internal details of shell. L = 27.9 mm. J. Undescribed genus and species (UGS-3). Chesterfield Bank, New Caledonia (MNHN IM-2009-10376). L = 23.5 mm. K. Undescribed genus and species (UGS-2), shell from same station as sequenced specimen. Manga, Bohol Island, Philippines. PANGLAO 2004, st. S20, 9°41.8'N, 123°51.1'E. 10 m specimen. L = 4.0 mm.

From shell characters, *Codakia* and *Ctena* have long been thought to be related, and indeed Bretsky (1976) classified *Ctena* as a subgenus of *Codakia*, but there have been no previous suggestions of a relationship between *Codakia* and *Lucinoma*. Chavan (1969) for example classified *Codakia* and *Ctena* in the subfam-

ily Lucininae, but placed *Lucinoma* in Myrteinae, and other authors have treated *Lucinoma* as a subgenus of *Phacoides* (Dall, 1901; Britton, 1970). Alternatively, *Lucinoma* was classified by Bretsky (1976) as a subgenus of *Miltha*, deriving it from the mid-Eocene genus *Plastomiltha*. Molecular results show no close

relationship between *Lucinoma* species and the *Myrtea* group. Similarly, we found no support for any relationship between *Phacoides* and *Lucinoma*, but, lacking *Miltha* in the analyses, we could not test any relationship between it and *Lucinoma*.

The four species of *Lucinoma* analysed form a well-supported group. Although some species range into the intertidal zone, most *Lucinoma* species live in deeper offshore habitats down to 2050 m. They have shells with regularly spaced, commarginal lamellae, prominent cardinal teeth, and long, narrow, anterior adductor muscle scars. Although fossil *Lucinoma* are frequent in Miocene–Recent deposits, the earliest claimed (Chavan, 1937–1938) is *Lucinoma hannibali* (Clark, 1925) from the Oligocene of Washington, USA (Blakeley Formation, 33.9–23.05 Myr). However, we recognize an earlier undescribed species, *Lucinoma* sp., from the Palaeocene (Selandian–Thanetian, 62–59 Ma), Aquia Formation, Aquia Creek, Potomac River, Virginia, USA (BMNH L64868–70), which has an external sculpture of regularly spaced commarginal lamellae and a long, narrow, anterior adductor muscle scar, features very similar to living *Lucinoma* species (Fig. 6J).

The *Codakia* clade is identified as sister to *Lucinoma* species in the molecular tree, but with low support (Fig. 4). *Codakia* species today are amongst the most well known of Lucinidae, including *Codakia orbicularis* (Linnaeus, 1758) from the western Atlantic (Frenkiel & Mouëza, 1995; Gros *et al.*, 2003). The earliest known *Codakia* that closely resembles living species is *Codakia leonina* (Basterot, 1825) from the late Miocene of southern Europe (Burdigalian, 20.4–15.9 Ma) (Fig. 6L). Earlier fossils have been claimed as part of the *Codakia* group, but all lack radial sculpture. These may form part of a *Codakia* stem group, and include ‘*Phacoides*’ *concentricus* (Lamarck, 1806) from the Eocene (Lutetian 48.6–40.4 Ma) of the Paris Basin, and the earlier ‘*Phacoides*’ *dejaeri* (Vincent, 1930) from the Palaeocene (Danian 65–61 Ma), Calcaire de Mons, Belgium (Glibert & Van de Poel, 1973) (Fig. 6K). Both of these species resemble living *Codakia* (Fig. 6M) in having a subcircular shape, an anterior lateral tooth close to the cardinals, a triangular resilifer, and a short, pointed, anterior adductor muscle scar. Chavan (1937–1938) claimed the Palaeocene ‘*Lucina*’ *gravesi* (Deshayes, 1857) as an early *Codakia*, but the preservation is poor. The late Jurassic–early Cretaceous species *Lucina crassa* Sowerby, 1827 was placed in *Codakia* by Kelly (1992), mainly on the basis of the fine radial grooves and some similarity of hinge teeth, but it has strong resemblance to species of the Jurassic genus *Jagonoma*.

Previous discussions concerning the history of the *Codakia* group centre on the position of *Epilucina*

(Chavan, 1937–1938; Bretsky, 1976). The name *Epilucina* has been widely used in the literature for fossils as old as the late Cretaceous (Speden, 1970), through the Palaeocene (Glibert & Van de Poel, 1967, 1973), Eocene (Pacaud & Le Renard, 1995), and Neogene (Kurihara, 2007), but there is considerable confusion concerning the relationships of the living type species *Epilucina californica* (Conrad, 1837) from the north-eastern Pacific (Kurihara, 2007). For example, Chavan (1969) places it as a subgenus of *Codakia*, whereas Bretsky (1976) argued for a relationship with *Myrtea*. The shell characters are rather equivocal, and it is clear that without molecular evidence the wider relationships of this species and genus are difficult to determine.

The *Ctena* species analysed form a distinct group separate from *Codakia* and *Lucinoma*. There has been some confusion over the concepts of *Ctena* and *Epicodakia*, names often used interchangeably, because Chavan (1969: fig. E3.9a,b) illustrated *Ctena divergens* (Philippi, 1850) as the example of *Epicodakia* rather than the type species, *Epicodakia consettiana* (Iredale, 1930). The two genera are probably closely related, but we lack molecular data for *Epicodakia consettiana*. The species we analysed, *Epicodakia tatei* (Angas, 1879), has shell characters that are more similar to *Ctena*. The earliest fossil *Ctena* with characters similar to living species is probably *Ctena squamosa* (Lamarck, 1806) from the Oligocene (Rupelian, 33.9–28.4 Ma) of Etampes, France (Fig. 6Q). The earlier *Lucina mayeri* Deshayes, 1857 (Eocene, Bartonian) was claimed by Chavan (1937–1938) as a *Ctena*, but this species has been classified as *Parvilucina* by Pacaud & Le Renard (1995). Convincing fossil *Epicodakia* species (Fig. 6N) with shell characters similar to living southern Australian forms (Fig. 6O) are recorded from the middle–late Eocene (Bartonian–Priabondian, 40.4–33.9 Ma) deposits of Western Australia (Darragh & Kendrick, 1980, 2008). *Jagonoma circumcisa* (Zittel & Goubert, 1861) from the late Jurassic (Oxfordian) of France has been claimed as an ancestor of the *Ctena* clade (Chavan, 1952), but lacks any radial sculpture. Further study of Mesozoic lucinids is needed to establish the possible, if any, relationships with Cenozoic and living taxa.

Fossil evidence of *Lucinoma* and *Codakia* clades places both back to the Palaeocene, suggesting an earlier divergence. Similarly, the origin of the *Ctena*/*Epicodakia* group is likely to be Palaeocene or earlier. The BEAST chronogram (Fig. 5) suggests a late Cretaceous–Palaeocene divergence of the three clades.

LUCININAE FLEMING, 1828

This is a large subfamily encompassing many shallow water lucinids, including *Lucina*, *Loripes*, *Austriella*,

Phacoides, *Divalinga*, the speciose genera *Parvilucina*, *Cardiolucina*, and *Pillucina*, with a few taxa from deeper water. They are extremely disparate in shell morphology, including, for example, heavily sculptured forms with both radial and commarginal elements, and others with almost smooth shells. Most have well-developed hinge teeth, both cardinals and laterals, but there are also edentulous species. The subfamily also includes several species with divaricate sculpture that have sometimes been classified in a separate subfamily Divaricellinae. A number of well-supported subgroups can be identified, and some of these are highlighted below.

Funafutia levukana, a small species from fore-reef habitats in the Indo-West Pacific (Glover & Taylor, 2007), has an unstable position. It lies outside and sister to all other Lucininae in the combined gene tree (Fig. 4), although in the 18S tree (Fig. 1) it lies within Lucininae, and in the *cytb* tree (Fig. 3) it groups as a sister to *Fimbria*. We would like to have confirmed this result with a sequence from a second specimen, but no additional material was available. The species was previously thought to be related to the *Loripes* subclade, as it has a short internal ligament (Glover & Taylor, 2001), but does not align within this group in the present analysis, and with hindsight is not very similar in other shell characters.

The subclade containing *Lucina* species also includes *Divalinga*, *Lepidolucina*, and *Discolucina*, as well as *Phacoides* (Figs 4, 5). Two species of *Lucina* were analysed: *Lucina adansoni* d'Orbigny, 1839 from the Cape Verde Islands and *Lucina pennsylvanica* (Linnaeus, 1758) from four locations in the western Atlantic, with the sample from Florida Keys differing from the other three sites. Morphological evidence from periostracal characters has suggested a complex of species in the *Lucina pennsylvanica* group of the western Atlantic (Gibson-Smith & Gibson-Smith, 1982; Taylor *et al.*, 2004). The earliest fossil that is usually attributed to *Lucina* is *Lucina carinifera* Conrad, 1833 from the middle Eocene (Bartonian) of eastern USA. Prominent later forms include the abundant *Lucina columbella* Lamarck, 1818 from the Miocene of southern Europe. The chronogram (Fig. 5) indicates a divergence of *Lucina* from the rest of the clade during the Oligocene rather than the Eocene. *Lucina oligocaenica* Cossmann, 1922 from the Oligocene (Rupelian) of Gironde, France is similar in shell characters to living species.

The position of *Phacoides pectinatus*, a well-studied, haemoglobin-rich species from mangrove fringes of the western Atlantic (Frenkiel, Gros & Mouéza, 1996), is unstable on a long branch in the 18S and 28S trees, but always nests within the Lucininae, and in the chronogram within the *Lucina*

subclade (Fig. 5). Despite the widespread use of the name in the past for various lucinids now consigned to other genera and subfamilies, there appears to be only one living species of *Phacoides*, and its relationships with other living genera are uncertain, although Bretsky (1976) classified it as a subgenus of *Lucina*. Chavan (1937–1938) claimed a continuous record of *Phacoides* from the early Eocene, but included in his generic concept were species that are now recognized as belonging to other genera, such as *Lamellolucina* (Taylor & Glover, 2002), so that all records need re-evaluation.

From the western Atlantic and eastern Pacific, three species of *Lucinisca*, two of *Radiolucina*, and '*Parvilucina*' *costata* form another well-supported subclade. *Radiolucina amiantus* (Dall, 1901) has been associated with *Cardiolucina* species (formerly *Bellucina*) from the Indo-Pacific by shell characters (Britton, 1972; Bretsky, 1976), but they are distant in our molecular tree. '*Parvilucina*' *costata* (d'Orbigny, 1842), a common western Atlantic species, has been variously placed in *Codakia*, *Ctena*, and *Parvilucina* (see Bretsky, 1976), but there is no molecular support for any of these possibilities. A new generic name is needed to accommodate this species and some similar fossils.

A large, highly supported (PP = 100%) subclade includes species of *Loripes*, *Pillucina*, *Chavania*, *Wallucina*, and *Lucinella* (hereafter *Loripes* subclade). Their geographical range covers the eastern Atlantic, Mediterranean, Indo-West Pacific, and southern Australia, but not the western Atlantic and eastern Pacific [with the exception of the rare *Loripes brasiliensis* (d'Orbigny, 1842) from Brazil]. All species within this clade possess a short, obliquely inset, internal ligament that differs structurally from the internal ligament in the Pegophyseminae (Taylor & Glover, 2005: fig. 28H), and the deeply inset resilifer of *Codakia* species. Shell sculpture is variable: *Loripes* and *Wallucina* have relatively smooth shells with fine commarginal lamellae; *Lucinella* has divaricate ribbing; and *Pillucina* species have both radial and commarginal elements. We have not been able to repeat our previous result (Williams *et al.*, 2004), where a specimen of *Pillucina vietnamica* Zorina, 1978 from Port Douglas, Australia aligned in the Codakiinae clade. In the present analysis other samples of *P. vietnamica* from Thailand and the Arabian Gulf aligned within the *Loripes* clade. Using evidence largely from ligament morphology, Chavan (1937–1938) thought the lucinids of the *Loripes* subclade were related to *Ctena* and *Codakia*, whereas Bretsky (1976) suggested a relationship between *Loripes* and *Anodontia*. From the molecular analysis there is no support for either of these suggestions, with the *Loripes* clade nesting within the Lucininae,

and distant from both *Codakia* and 'Anodontia' clades.

The earliest *Loripes* with characters similar to the living species is *Loripes dujardini* (Deshayes, 1850) (Fig. 6P) from the early Miocene (Aquitainian 23–20 Ma), and the chronogram suggests a divergence of the whole *Loripes* subclade in the late Eocene (Fig. 5). Species of the extinct genus *Microloripes* (type species *Loripes dentata* DeFrance, 1823) that range from the Palaeocene to the Pliocene also possessed an obliquely inset internal ligament similar to that of *Loripes*, and may represent a stem group of the clade. Although some lucinid species from the late Jurassic have been placed in *Loripes* and a subgenus *Discoloripes* (Wellnhofer, 1964), these lack the oblique internal ligament that characterizes the *Loripes* group, and should be excluded.

The Indo-West Pacific, mangrove-associated *Austriella* and *Indoaustriella* species form a robust clade: *Austriella corrugata* (Deshayes, 1843) lacks hinge teeth, whereas the four *Indoaustriella* species analysed possess lateral teeth (Glover *et al.*, 2008). There is no fossil record for any of these mangrove species, but a Miocene origin is suggested by the chronogram (Fig. 5). In our previous analyses (Glover *et al.*, 2004; Williams *et al.*, 2004), the hydrothermal vent species *Bathyaustriella thionipta* aligned with *Austriella corrugata*, with which it shares some shell and anatomical similarities. With the larger taxon set of the present analysis it groups closely with *Parvilucina* species (Figs 4, 5), but there is little obvious morphological resemblance.

The genus *Cardiolucina* was represented in our analysis by six species that formed a robust clade, sister to the *Indoaustriella/Austriella* group (Figs 4, 5). *Cardiolucina* species are abundant in the Indo-West Pacific ocean, ranging from the intertidal [*Cardiolucina pisiformis* (Thiele, 1930) and *Cardiolucina semperiana* (Issel, 1869)] to depths of more than 800 m [*Cardiolucina quadrata* (Prashad, 1932)], with two species known from the southern Atlantic (Taylor & Glover, 1997). *Cardiolucina* (as *Bellucina*) has previously been related to *Parvilucina* (e.g. Bretsky, 1976), but Chavan (1969) classified it as a subgenus of *Lucina* (as *Linga*). Neither possibility is supported by our molecular analysis. The type species, *Cardiolucina agassizi* (Michelotti, 1839) occurs in the middle and late Miocene of southern Europe, whereas the chronogram (Fig. 5) suggests an Oligocene origin for the clade.

Polyphyly of Divaricellinae

Lucinids possessing divaricate sculpture species were formerly united in the subfamily Divaricellinae Korobkov, 1954 (see also Glibert & Van de Poel, 1967; Chavan, 1969), with living species classified

into six genera: *Bourdotia*, *Divalinga*, *Divalucina*, *Divaricella*, *Lucinella*, and *Pompholigina*. From morphological evidence and preliminary molecular analysis, doubts have been expressed concerning the monophyly of the group (Dekker & Goud, 1994; Williams *et al.*, 2004). In our present analysis, *Divalinga bardwelli* (Iredale, 1936) and *Divalinga quadrisulcata* (d'Orbigny, 1842) align together, but are widely separated within the Lucininae from *Divaricella irpex* (Smith, 1885) and *Lucinella divaricata* (Linnaeus 1758). The latter species has a hinge and an oblique internal ligament similar to taxa in the *Loripes* group, where it nests in the molecular analysis. For *Divalinga* and *Divaricella*, detailed morphology of the divaricate ribs reveals constructional differences, as demonstrated by Checa (2002), in addition to dissimilarity in hinge and ligament structure. Our evidence suggests a separate evolution of divaricate shell sculpture in the three groups, and we corroborate the remarks of Dekker & Goud (1994) that Divaricellinae is not a valid division of the Lucinidae. Species from other genera with divaricate sculpture, *Divalucina*, *Bourdotia*, and *Pompholigina*, have yet to be analysed. The fossil genus *Paralucina* (type species *Lucina undulata* Deshayes, 1857) from the Oligocene of France has a divaricate sculpture, but lacks an internal ligament, and can be excluded from a relationship with *Lucinella*.

Uncertain status of Milthinae

Chavan (1969) introduced the subfamily Milthinae, including eight genera with living species and 14 fossil genera, grouping together taxa with commarginal sculpture and long anterior adductor muscle scars. From our molecular analysis most of the included living genera, such as *Anodontia*, *Pegophysema*, and *Austriella*, align in other clades, leaving only *Miltha* and *Eomiltha*, which were not included in our analysis, being represented today by three rare living species: *Miltha childreni* (Gray, 1825) from the western Atlantic, *Miltha xantusi* (Dall, 1905) from the eastern Pacific, and *Eomiltha voorhoevi* (Deshayes, 1857) from the Mozambique Channel. The position of both of these genera is uncertain and, despite their names, may not be related. Both *Miltha* and *Eomiltha* have fossil records extending to the Palaeocene, with fossil species often abundant throughout the Cenozoic, with a *Miltha* species common in shallow water deposits of south-western Australia as recently as the early Pleistocene (Ludbrook, 1978). The fossil genera included by Chavan in Milthinae, such as *Pseudomiltha*, are frequent in the Eocene, and possess very long anterior adductor muscle scars. Other fossils such as *Bulacanities* (Kase & Aguilar, 2007) and *Superlucina* (Taylor & Glover, 2009b) also had long

adductor scars, but again their relationship to *Miltha* or any other lucinids is uncertain. In conclusion, the status of Milthinae as conceived by Chavan (1969) is polyphyletic, but the positions of *Miltha* and *Eomiltha* are yet to be resolved.

Incongruence of molecular results with classifications based in shell and morphological characters

Molecular analyses have demonstrated that traditional shell characters are a less satisfactory guide to suprageneric relationships, with major incongruencies apparent between the molecular trees and phylogenetic hypotheses derived from shell characters (Chavan, 1937–1938; Bretsky, 1970, 1976). These include the position of the ‘*Anodontia*’ group, relationships of *Pseudolucinisca*, the relationships of *Lucinoma* with the *Codakia/Ctena* group, the polyphyly of the shell-based Myrteinae, and non-monophyly of genera with divaricate sculpture. However, no rigorous cladistic analysis of shell characters has yet been attempted, but determining homology would be difficult, with very similar features occurring in several major clades. Hinge teeth characters have been extensively used in discussions of fossil lucinids, but their numbers and morphology seem labile, and edentulous forms appear in several different clades (Glover *et al.*, 2008). An example of the difficulties in using shell characters is in the form of the ligament. In most lucinids the ligament is external and set in a shallow groove, but in a few species part of the ligament is internal and set in an oblique groove that extends posteriorly and laterally into the left and right valves. Such a ligament, exemplified by *Loripes lucinalis* (Allen, 1960; Glover & Taylor, 2001), also occurs in species of *Wallucina*, *Pillucina*, *Chavania*, and *Lucinella*, which form a monophyletic subclade within the Lucininae. A similar internal ligament also occurs in *Funafutia levukana*, a small species from coral reef slope habitats placed within the *Pillucina/Wallucina* group by Glover & Taylor (2001). However, the molecular analysis placed this species as a sister taxon to all other Lucininae and distant from the *Loripes/Pillucina* subclade, suggesting an independent derivation of this form of internal ligament. Some species in the Pegophyseminae clade, such as *Euanodontia ovum* (Taylor & Glover, 2005: fig. 28) also possess laterally extended internal ligaments, but these differ in morphology from the *Loripes* type and are unlikely to be homologous. In summary, a revised and more objective study of shell characters is needed so that the rich fossil record of Lucinidae might be better incorporated into the phylogenetic analysis.

Some anatomical characters, such as the extent and type of posterior mantle fusion (Taylor & Glover,

2006), also have potential for phylogenetic analysis, but are poorly known for the majority of taxa. Different morphologies of the mantle respiratory organs, the ‘mantle gills’, were reported by Taylor & Glover (2000, 2006). These are sporadically developed amongst the Lucinidae in Pegophyseminae, *Codakia* species, and, within Lucininae, in *Phacoides pectinatus* and *Lucina* species. These organs differ structurally in the different clades suggesting separate derivations. Ctenidial structure has been widely investigated by electron microscopy in several major groups of Lucinidae, but differences are minor with no clear phylogenetic signal (e.g. Frenkiel & Mouëza, 1995; Gros, Frenkiel & Mouëza, 1998; Ball *et al.*, 2009).

Diversification history of Lucinidae

There is a good fossil record of Lucinidae throughout the Mesozoic and Cenozoic, and as demonstrated in the discussion of our recognized subfamilies it is possible to trace morphological lineages of living taxa through the Cenozoic. However, the relationships of some extinct taxa from the early Cenozoic are problematic, as is also the case for most Mesozoic lucinids.

There is only a patchy fossil record of Lucinidae through the Palaeozoic. *Illionia prisca* (Hisinger, 1837), a species that was abundant in the late Silurian of Gotland, Sweden (Liljedahl, 1991a, b), possesses convincing lucinid characters (Taylor & Glover, 2006). *Paracyclas* and *Phenacocyclas* species are known from the Devonian (LaRocque, 1950; Bailey, 1983; Johnston, 1993). *Paracyclas* species are common in some facies (Bailey, 1983), and some of these have ventrally detached, anterior adductor muscle scars characteristic of Lucinidae (LaRocque, 1950: pl. 12, fig. 12; Taylor & Glover, 2006: fig. 14B), but different from the elongate non-detached scar illustrated by Johnston (1993; figs 81A, 82D), who placed them in a separate family Paracyclidae. An earlier *Paracyclas* species from the Ordovician is claimed by Zong-Jie & Cope (2004), but the preservation is poor. *Palaeolucina* is widely reported from the Carboniferous of China (Chen, 1976), but few details of morphology are available. Later, *Gigantocyclus* (Boyd & Newell, 1979) from the Permian of Tunisia can be confidently placed in the Lucinidae.

For the Mesozoic there is a dearth of confirmed lucinids from the Triassic: Hautmann & Nützel (2005) suggested the small early Triassic species, *Sinbadilla pygmaea* Hautmann & Nützel, 2005, as a possible lucinid, but without strong evidence. ‘*Lucina*’ *deshayesii* Klipstein, 1845 (holotype, BMNH Palaeontology Department 35194) from the St Cassian Formation (late Triassic, Carnian) may be a true lucinid, but the preservation is poor. *Schafklaeutlia* is usually

classified in the Fimbriinae (Chavan, 1969; Hautmann, 2001), but we regard this placement as uncertain. By the late Triassic (Rhaetian), lucinids resembling modern forms are more frequent (Ivimey-Cook *et al.*, 1999). Throughout the Jurassic and Cretaceous lucinids are more diverse, but are generally under-researched. Preservation is often poor, with many nominal species based on internal moulds, with hinge and muscle scar details often unknown. Many species are still named '*Lucina*' or are 'shoe-horned' into the few available generic names, and most need critical study. A few studies of well-preserved faunas indicate that diversity and abundance of Mesozoic lucinids is greater than was commonly thought, with several new genera introduced over last few years (Chavan, 1952; Fischer, 1969; Kelly, 1992; Kendrick & Vartak, 2007; Amano *et al.*, 2008; Kiel, Campbell & Gaillard, 2010). Furthermore, only a few small species (<1cm in length) have been described, although they undoubtedly existed as evidenced in Cenozoic and Recent faunas. At present it is difficult to assess the relationships of most Mesozoic lucinids to later Cenozoic and modern clades.

By contrast to earlier periods, Cenozoic faunas are much better documented. Nonetheless, there is a significant increase in the diversity and abundance of lucinids from the Palaeocene onwards. For example, from the Palaeocene (Danian) of Belgium, 24 species of lucinids were recorded, comprising a remarkable 19% of the total bivalve species found (Glibert & Van de Poel, 1973). Ninety-six species in 28 genera are recorded from the Eocene of the Paris Basin (Cossmann & Pissaro, 1904–06; Pacaud & Le Renard, 1995), and similarly diverse faunas occur in the Miocene (Sacco, 1901; Cossmann & Peyrot, 1909–1912; Gardner, 1926). The chronogram (Fig. 5) demonstrates that many of the clades of living lucinids originated in the Eocene, with radiations in the Miocene. Notable appearances in the late Cenozoic were the ecologically important *Codakia* and *Ctena* species in shallow tropical habitats, and also the dominance of *Lucinoma* species at hydrocarbon seeps, especially in the northern Pacific (Majima, Nobuhara & Kitazaki, 2005). Some lucinids that were abundant during the early Cenozoic have either become extinct, such as *Pseudomiltha* species, or, are now extremely rare, as for example *Eomiltha* and *Miltha*.

In pre-Miocene times the external sculpture of lucinids was usually in the form of low commarginal lamellae, but then species with radial and cancellate sculpture became much more frequent in the Miocene and in younger faunas. Genera with prominent radial ribbing include shallow water tropical species in the Codakiinae and Lucininae clades such as *Codakia*, *Ctena*, *Cardiolucina*, *Lucinisca*, *Pleurolucina*, *Parvilucina*, and *Radiolucina*.

Patterns of habitat occupation

Most lucinids occur in tropical, subtropical, or warm temperate waters, the few species inhabiting cool temperate latitudes are mainly species of *Lucinoma*, for example *Lucinoma filosa* (Stimpson, 1851), from the eastern seaboard of North America to New Brunswick, Canada (Britton, 1970), *Lucinoma annulata* (Reeve, 1850) from the eastern Pacific to 61°N in Alaska (Coan, Valentich Scott & Bernard, 2000), *Lucinoma borealis* from northern Europe to the Lofoten Islands (68°N), Norway (BMNH collections), and *Lucinoma lamellata* (Smith, 1881) at Cape Horn (55°45'S) (Holmes, Oliver & Sellanes, 2005). Additional higher latitude species include *Myrtea spinifera* (Montagu, 1803), which ranges from the Mediterranean to 64°48'N in Norway, and *Loripes lucinalis* from north western Africa to southern Britain, whereas *Parvilucina tenuisculpta* (Carpenter, 1864) occurs from the north-eastern Pacific to southern Alaska (61°N) (Coan *et al.*, 2000). The tiny '*Epicodakia*' *falklandica* (Dell, 1964) is recorded from the Southern Ocean (Zelaya, 2005), but the generic placement of this species is uncertain.

Lucinids have often been reported from various marine habitats with an elevated input of organic material, such as seagrass beds or hydrocarbon seeps. In this section we address the question of whether there is any phylogenetic pattern to the habitat preferences of lucinids: are some individual clades associated with particular habitats?

Seagrass beds: There is a well-known strong association of lucinids with seagrass beds both in tropical and temperate seas (reviewed by Barnes & Hickman, 1999). Seagrass beds entrap organic detritus, and have higher rates of sulphate reduction and sulphide retention than surrounding unvegetated sediment (Holmer, Duarte & Marba, 2003; Holmer *et al.*, 2009). Root and rhizome masses create stable habitats and release oxygen through the roots. Notable amongst the lucinids that inhabit seagrass beds are several *Codakia* species [*Codakia orbicularis*, *Codakia punctata* (Linnaeus, 1758), *Codakia tigerina* (Linnaeus, 1758), and *Codakia paytenorum* (Iredale, 1937)] that are abundant in *Thalassia*-dominated habitats of the western Atlantic and Indo-West Pacific (Moore *et al.*, 1968; Taylor & Lewis, 1970; Jackson, 1972). As well as *Codakia orbicularis*, seagrass beds of the western Atlantic also support populations of *Ctena* species, *Lucina pennsylvanica*, *Anodontia alba*, and *Lucinisca nassula* (Conrad, 1846) (J. D. Taylor & E. A. Glover, pers. observ.), and *Stewartia floridana* (Fisher & Hand, 1984). In the Indo-West Pacific, *Euanodontia ovum*, *Cavatidens omissa* Iredale, 1930, and other pegophysemes, *Pillucina* species (Nakaoka, Mukai & Chuhabundit, 2002; Meyer *et al.*, 2008), *Chavania striata*

(Tokunaga, 1906), and *Divaricella irpex* (E. A. Glover & J. D. Taylor, unpubl. data) are widely associated with seagrass habitats. From temperate latitudes, examples of the association include *Loripes lacteus* (Linnaeus, 1758), which is abundant in Mediterranean *Posidonia* beds (Johnson, Fernandez & Pergent, 2002), *Wallucina assimilis* (Angas, 1868) from southern Australia (Barnes & Hickman, 1999), *Lucinoma borealis* from southern England (Dando, Southward & Southward, 1986; E. A. Glover & J. D. Taylor, pers. observ.), and *Pillucina pisidium* (Dunker, 1860) from Japan (Uede & Takahashi, 2008). From this brief survey it is clear that although *Codakia* species are the largest and most conspicuous species, lucinids associated with seagrass derive from four major clades: the Pegophyseminae, Leucosphaerinae, Codakiinae, and Lucininae. Species of Myrteinae usually live at water depths below those colonized by seagrasses, but we have collected *Notomyrtea mayi* from an intertidal seagrass habitat in South Australia.

Seagrass-dominated shallow water communities became widespread during the Eocene (Brasier, 1975), and the association with lucinids dates from this time. Various Eocene deposits of the Paris Basin have been interpreted as representing seagrass-dominated communities (Murray & Wright, 1974), with lucinids found frequently amongst the molluscs (Pacaud & Le Renard, 1995). From the Eocene (late Lutetian) Selsey Formation of southern England lucinids occur in association with plant remains of seagrasses (Collinson, 1996; Tracey *et al.*, 1996). Additionally, the giant *Superlucina megameris* (Dall, 1901) from the Eocene, White Limestone of Jamaica, has been interpreted as an inhabitant of seagrass communities (Taylor & Glover, 2009b). Such associations have continued through the Cenozoic, as for example in the early Oligocene of the Etampes area of the Paris Basin (Gitton, Lozouet & Maestrati, 1986), and in the Roe Calcarenite of south-west Australia (late Pliocene–early Pleistocene), which has been interpreted as a seagrass-dominated deposit (James & Bone, 2007), where *Miltha hamptonensis* Ludbrook, 1969 was common, along with other lucinids (Ludbrook, 1978).

Mangrove association: Mangrove communities are another shallow water marine ecosystem with a high organic input. In the present day, only a few lucinids are closely associated with mangrove environments. Notable amongst these are *Austriella corrugata* and *Indoaustriella* species from the central Indo-West Pacific (Glover *et al.*, 2008), *Pegophysema philippiana* from the Indo-West Pacific (Lebata & Primavera, 2001; Taylor & Glover, 2005), *Phacoides pectinatus* from the western Atlantic, which inhabits black muds amongst mangroves (Frenkiel *et al.*, 1996), and *Kelestistes rhizoecus* (Oliver, 1986) from West Africa, which

lives in mangrove root peat in the Niger Delta (Zabbey, Hart & Wollf, 2010). In South-East Asia, lucinids can also be abundant in communities peripheral to mangrove stands (Meyer *et al.*, 2008). Although organic-rich, the suboxic sediments are a less favourable habitat than seagrass beds for burrowing bivalves, because the low pH causes extensive corrosion of shells and the substrata are often very soft. In summary, mangrove-associated lucinids derive from two main clades, Pegophyseminae and Lucininae, and within the latter the well-defined *Austriella/Indoaustriella* subclade and *Phacoides pectinatus*. Mangrove systems diversified during the Palaeocene, and by the mid to late Eocene were widespread in the tropics and subtropics (Plaziat *et al.*, 2001). Although some mangrove-associated gastropods are recorded from Cenozoic deposits (Reid *et al.*, 2008), lucinids have not been documented.

Deeper water habitats: Most lucinid species have been described from shallow water habitats, generally from water depths of less than 100 m. Until recently, relatively few lucinids were known from deeper water, but increased sampling of bathyal habitats in the tropics and hydrocarbon seeps is revealing an unexpected diversity, with lucinids now known to depths of 2570 m (Cosel, 2006; Cosel & Bouchet, 2008; Taylor & Glover, 2009a). The relationships of some of these newly described taxa are uncertain, but they most likely can be classified in Leucosphaerinae or the Myrteinae clades. Deep-water lucinids that have been analysed molecularly derive from: Myrteinae (*Myrtea* and *Gloverina*); Leucosphaerinae, with genera such as *Dulcina* and several undescribed species; the *Lucinoma* clade of the Codakiinae; and a few species from the Lucininae such as *Troendleina musculator* Cosel & Bouchet, 2008, *Bathyaustriella thionipta*, and *Cardiolucina quadrata*. The deepest recorded lucinid is *Elliptiolucina labeyriei* Cosel & Bouchet, 2008 from 2570 m in the Sulu Sea. Nevertheless, *Lucinoma* species are the lucinids most widely reported from deeper water habitats, and several have been found in association with hydrocarbon seeps and mud volcanoes. These include *Lucinoma atlantis* Mclean, 1936 (Cordes *et al.*, 2007), *Lucinoma anemiophila* Holmes, Oliver & Sellanes, 2005, *Lucinoma yoshidai* Habe, 1958 (Okutani & Hashimoto, 1997), *Lucinoma kazani* Salas & Woodside, 2002, and *Lucinoma myriamae* (Cosel, 2006) from off West Africa, included in our analysis from 2050 m. Additionally, two species, *Lucinoma aequizonata* (Stearns, 1890) and *Lucinoma gagei* Oliver & Holmes, 2006 are known from sediments of oxygen minimum zones (Cary *et al.*, 1989; Oliver & Holmes, 2006). Other lucinids recorded as associated with hydrocarbon seeps include *Graecina karinae* Cosel, 2006, *Graecina*

colombiensis Taylor & Glover, 2009a, *Meganodontia acetabulum* Bouchet & Cosel 2004, *Mesolinga soliditesta* Okutani & Hashimoto 1997, *Myrtea amorpha* (Olu-Le Roy *et al.*, 2004), and *Jorgenia* species from the Louisiana Slope (Taylor & Glover, 2009a). The only lucinid recorded as yet from a hydrothermal vent is *Bathyaustriella thionipta* from the Kermadec Ridge off New Zealand (Glover *et al.*, 2004). As well as hydrocarbon seeps, Cosel & Bouchet (2008) suggest that around the Philippines and Indonesia the many semi-enclosed basins with poor water circulation and accumulations of decomposing continental plant debris create favourable habitats for chemosymbiotic animals.

The fossil record indicates that lucinids have been present at putative hydrocarbon seeps since at least the late Jurassic (Gaillard *et al.*, 1992; Kiel *et al.*, 2010), through the Cretaceous with *Ezolucina*, *Nipponothracia*, and *Nymphalucina* species (Speden, 1970; Kauffman *et al.*, 1996; Kelly *et al.*, 2000; Amano *et al.*, 2008; Kiel, Amano & Jenkins, 2008), and in the Cenozoic with, for example, species of *Cryptolucina*, *Nipponothracia*, *Megaxinus*, and *Lucinoma* (Taviani, 1994; Saul, Squires & Godert, 1996; Majima *et al.*, 2003; Campbell, 2006; Kase, Kurihara & Hagino, 2007).

Symbionts and co-evolution

The new lucinid phylogeny will provide a framework to examine the distribution of the bacterial symbiont phylotypes. No comprehensive molecular analysis of symbionts and their hosts has yet been attempted, although details of some lucinid symbionts are available, mainly from western Atlantic host species (Distel, Felbeck & Cavanaugh, 1994; Durand & Gros, 1996; Dubilier, Bergin & Lott, 2008). All symbiont phylotypes recorded from lucinid ctenidia to date belong to the gamma group of proteobacteria. Experimental evidence shows that symbionts are acquired by environmental transmission from the sediment (Gros *et al.*, 1998), suggesting that a close co-evolutionary history between host and symbiont is unlikely. Indeed, several lucinids from seagrass beds around Guadeloupe shared the same symbiont (Gros *et al.*, 2003). Although few symbiont sequences from lucinids have yet been published, preliminary results (Cavanaugh *et al.*, 2006; Dubilier *et al.*, 2008; Ball *et al.*, 2009) show some differentiation between hosts, so that the symbionts of *Phacoides pectinatus*, *Euanodontia ovum*, and *Pegophysema schrammi* (as *Anodontia philippiana* in the cited papers) are distinct from the symbionts associated with other lucinids (e.g. *Lucinisca*, *Codakia*, *Stewartia*, and *Lucinoma*) that group more closely together. The addition of further taxa from different host clades and from a wider range of geographical locations and habitats should result in an

improved understanding of host–symbiont relationships and the diversity of symbionts.

Phylogeography

In the present analysis, *Ctena* species provide the most informative data set for phylogeographic analysis. The species form two distinct groups, one consisting of both western Atlantic [*Ctena orbiculata* (Montagu, 1808) and *Ctena imbricatula* (Adams, 1845); note the latter species is usually synonymized with *Ctena orbiculata*, but molecular data shows that they are distinct species] and eastern Pacific species [*Ctena chiquita* (Dall, 1901) and *Ctena mexicana* (Dall, 1901)], and the other comprising West African [*Ctena eburnea* (Gmelin, 1791)], Mediterranean [*Ctena decussata* (Costa, 1829)], and Indo-West Pacific species [*Ctena bella* (Conrad, 1834), *Ctena delicatula* (Pilsbry, 1904), *Epicodakia tatei*, and *Ctena* sp.]. Albeit with only limited sampling, this phylogeny suggests an Atlantic origin with later radiations in the Indo-West Pacific, consistent with the pattern observed in many other taxa (e.g. Williams & Reid, 2004). The chronogram (Fig. 5) estimates that the western Atlantic/eastern Pacific clade diverged from the eastern Atlantic/Mediterranean and Indo-West Pacific clades in the late Eocene. In the western Atlantic/eastern Pacific clade, *Ctena orbiculata* is sister to the eastern Pacific species and *Ctena imbricatula*, with a divergence in the middle Oligocene. In the other clade, the Atlantic and Mediterranean species (*Ctena eburnea* and *Ctena decussata*) are sister to the Indo-West Pacific species, with a divergence in the early Oligocene, earlier than estimates of the closure of the Tethys Seaway, approximately 19–18 Ma (Vrielynck, Odin & Dercourt, 1997; Rögl, 1998; Harzhauser *et al.*, 2002, 2007), although 95% HPD ranges suggest a more recent separation is also possible. The Indo-West Pacific clade diversified in the early Miocene, consistent with age estimates of many other tropical, shallow water Indo-West Pacific radiations (e.g. Williams & Duda, 2008).

Codakia is the only other genus with sufficient interspecific sampling to make any biogeographic comment. In this clade the western Atlantic *Codakia orbicularis* occupies a position sister to the other species from the Indo-West Pacific that may have separated in the late Oligocene, slightly later than that of *Ctena* and *Epicodakia* (although the 95% HPD ranges overlap). Divergence times between eastern Pacific/Atlantic and Indo-West Pacific clades have been noted to differ in several studies (e.g. Frey & Vermeij, 2008). Some of this variation may reflect uncertainty in molecular clock analyses or real differences, resulting from multiple vicariant events or the fact that the constriction of the Tethys Seaway was not an instantaneous event, but extended over several

millions of years (Bellwood, van Herwerden & Konow, 2004). The latter seems particularly pertinent, given a similar pattern of protracted speciation events between marine species either side of the Isthmus of Panama (Lessios, 2008). Other possible explanations for disparate estimates include the extinction of true sister lineages (Paulay, 1997) or the role of alternate vicariant events (Reid, Dyal & Williams, 2010).

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REFERENCES

- Allen JA. 1960.** The ligament of the Lucinacea. *Quarterly Journal of Microscopical Science* **101**: 25–37.
- Amano K, Jenkins RG, Kurihara Y, Kiel S. 2008.** A new genus for *Vesicoma inflata* Kanie & Nishida, a lucinid shell convergent with that of vesicomids, from Cretaceous strata of Hokkaido, Japan. *Veliger* **50**: 255–262.
- Bailey JB. 1983.** Middle Devonian Bivalvia from the Solsville Member (Marcellus Formation), central New York State. *Bulletin of the American Museum of Natural History* **174**: 193–326.
- Ball AD, Purdy KJ, Glover EA, Taylor JD. 2009.** Ctenidial structure and three bacterial symbiont morphotypes in *Anodontia (Euanodontia) ovum* (Reeve, 1850) from the Great Barrier Reef, Australia (Bivalvia: Lucinidae). *Journal of Molluscan Studies* **75**: 175–185.
- Barnes PAG, Hickman CS. 1999.** Lucinid bivalves and marine angiosperms: a search for causal relationships. In: Walker DI, Wells FE, eds. *The Seagrass Flora and Fauna of Rottnest Island, Western Australia*. Perth: Western Australian Museum, 215–238.
- Bellwood DR, van Herwerden L, Konow N. 2004.** Evolution and biogeography of marine angelfishes (Pisces: Pomacanthidae). *Molecular Phylogenetics and Evolution* **33**: 140–155.
- Bouchet P, Cosel R. 2004.** The world's largest lucinid is an undescribed species from Taiwan (Mollusca: Bivalvia). *Zoological Studies* **43**: 704–711.
- Boyd DW, Newell ND. 1979.** Permian pelecypods from Tunisia. *American Museum Novitates* **2686**: 1–22.
- Brasier MD. 1975.** An outline history of seagrass communities. *Palaeontology* **18**: 681–702.
- Bretsky SS. 1970.** Phenetic and phylogenetic classifications of the Lucinidae (Mollusca, Bivalvia). *Bulletin of the Geological Institute of the University of Upsala new series* **2**: 5–23.
- Bretsky SS. 1976.** Evolution and classification of the Lucinidae (Mollusca; Bivalvia). *Palaeontographica Americana* **8**: 219–337.
- Britton JC. 1970.** The Lucinidae (Mollusca: Bivalvia) of the Western Atlantic Ocean. PhD dissertation, George Washington University. University Microfilms 71-12, 288.
- Britton JC. 1972.** Two new species and a new subgenus of Lucinidae (Mollusca: Bivalvia), with notes on certain aspects of lucinid phylogeny. *Smithsonian Contributions to Zoology* **19**: 1–19.
- Campbell KA. 2006.** Hydrocarbon seep and hydrothermal vent paleoenvironments and paleontology: past developments and future research directions. *Palaeogeography, Palaeoclimatology, Palaeoecology* **232**: 362–407.
- Cary SC, Fry B, Felbeck H, Vetter RD. 1989.** Habitat characterization and nutritional strategies of the endosymbiont-bearing bivalve *Lucinoma aequizonata*. *Marine Ecology Progress Series* **55**: 31–45.
- Castresana J. 2000.** Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution* **17**: 540–552.
- Cavanaugh CM, McKiness ZP, Newton ILG, Stewart FJ. 2006.** Marine chemosynthetic symbioses. In: Dworkin M, Falkow SI, Rosenberg E, Schleifer KH, Stackebrandt E, eds. *The prokaryotes 1*. New York: Springer, 475–507.
- Chavan A. 1937–1938.** Essai critique de classification des lucines. *Journal de Conchyliologie* **81**: 237–281; **82**: 215–241.
- Chavan A. 1947.** La faune campanienne du Mont des Oliviers d'après les matériaux Vignal-Massé. *Journal de Conchyliologie* **88**: 125–197.

- Chavan A. 1951.** Essai critique de classification des *Divaricella*. *Institut Royal des Sciences Naturelle de Belgique* **27**: 1–27.
- Chavan A. 1952.** Les pélicypodes des Sables Astartiens de Cordebugle (Calvados). *Schweizerische Palaeontologische Abhandlungen* **69**: 1–132.
- Chavan A. 1959.** Quelques intéressantes subdivisions. *Cahiers Géologiques* **53**: 515–516.
- Chavan A. 1969.** Superfamily Lucinacea Fleming, 1828. In: Moore RC, ed. *Treatise on invertebrate paleontology, part N, mollusca 6, Bivalvia*, Vol. 2. Boulder, CO: Geological Society of America and University of Kansas, N491–N518.
- Checa A. 2002.** Fabricational morphology of oblique ribs in bivalves. *Journal of Morphology* **254**: 195–209.
- Chen CC. 1976.** *Lamellibranch fossils of China*. Nanjing Institute of Geology and Palaeontology. Beijing: Science Press Beijing.
- Coan EV, Valentich Scott P, Bernard FR. 2000.** *Bivalve seashells of Western North America. Marine bivalve mollusks from Arctic Alaska to Baja California*. Santa Barbara: Santa Barbara Museum of Natural History.
- Collinson ME. 1996.** Plant microfossils from the Bracklesham Group (Early and Middle Eocene), Bracklesham Bay, West Sussex, England: review and significance in the context of coeval British Tertiary floras. *Tertiary Research* **16**: 175–202.
- Cordes EE, Carney SL, Hourdez S, Carney RS, Brooks JM, Fisher CR. 2007.** Cold seeps of the deep Gulf of Mexico: community structure and biogeographic comparisons to Atlantic equatorial belt seep communities. *Deep-Sea Research I* **54**: 637–653.
- Cosel R. 2006.** Taxonomy of West African bivalves VIII. Remarks on Lucinidae, with descriptions of five new genera and nine new species. *Zoosystema* **28**: 805–851.
- Cosel R, Bouchet P. 2008.** Tropical deep-water lucinids (Mollusca: Bivalvia) from the Indo-Pacific: essentially unknown, but diverse and occasionally gigantic. In: Héros V, Cowie R, Bouchet P, eds. *Tropical deep sea benthos, volume 25. Mémoires du Muséum, national d'Histoire naturelle, Paris* **196**: 115–213.
- Cossmann M, Peyrot A. 1909–1912.** Conchologie néogène de l'Aquitaine. *Actes de la Société Linnéenne de Bordeaux* **63**: 1–718.
- Cossmann M, Pissarro G. 1904–06.** *Iconographie complète des coquilles fossiles de l'Éocène des environs de Paris. Volume 1 Pélécypodes*. Paris: M. Pissarro.
- Dall WH. 1901.** Synopsis of the Lucinacea and of the American species. *Proceedings of the United States National Museum* **23**: 779–833.
- Dando PR, Southward AJ, Southward EC. 1986.** Chemoautotrophic symbionts in the gills of the bivalve mollusc *Lucinoma borealis* and the sediment chemistry of its habitat. *Proceedings of the Royal Society of London B* **227**: 227–247.
- Darragh TA, Kendrick GW. 1980.** Eocene bivalves from the Pallinup Siltstone near Walpole, Western Australia. *Journal of the Royal Society of Western Australia* **63**: 5–20.
- Darragh TA, Kendrick GW. 2008.** Silicified Eocene molluscs from the Lower Murchison district, Southern Carnarvon Basin, Western Australia. *Records of the Western Australian Museum* **24**: 217–246.
- Dekker H, Goud J. 1994.** Review of the living Indo-West-Pacific species of *Divaricella* sens auct. With descriptions of two new species and a summary of species from other regions. *Vita Marina* **42**: 115–136.
- Distel DL. 1998.** Evolution of chemoautotrophic endosymbioses in bivalves. *Bioscience* **48**: 277–286.
- Distel DL, Felbeck H. 1987.** Endosymbiosis in the lucinid clams *Lucinoma aequizonata*, *Lucinoma annulata* and *Lucina floridana*: a reexamination of the functional morphology of the gills as bacteria-bearing organs. *Marine Biology* **96**: 79–86.
- Distel DL, Felbeck H, Cavanaugh CM. 1994.** Evidence for phylogenetic congruence among sulfur-oxidising chemoautotrophic bacterial symbionts and their bivalve hosts. *Journal of Molecular Evolution* **38**: 533–542.
- Drummond AJ, Ho SYW, Phillips MJ, Rambaut A. 2006.** Relaxed phylogenetics and dating with confidence. *PLoS Biology* **4**: e88.
- Drummond AJ, Rambaut A. 2007.** BEAST: bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology* **7**: 214.
- Dubar G. 1948.** La faune Domérienne du Djebel Bou-Dahar près de Béni-Tajjite étude suivie de celle de quelques Mollusques d'autres gisements marocains. *Notes et Mémoires Service Mines Carte Géologique du Maroc* **68**: 1–250.
- Dubilier N, Bergin C, Lott C. 2008.** Symbiotic diversity in marine animals: the art of harnessing chemosynthesis. *Nature Reviews in Microbiology* **6**: 725–740.
- Durand P, Gros O. 1996.** Bacterial host specificity of Lucinacea endosymbionts: interspecific variation in 16SrRNA sequences. *FEMS Microbiology Letters* **140**: 193–198.
- Fischer J-C. 1969.** Géologie, paléontologie et paléocéologie du Bathonien au sud-ouest du Massif Ardennais. *Mémoires du Muséum National d'Histoire Naturelle sér C* **20**: 1–319.
- Fisher MR, Hand SC. 1984.** Chemoautotrophic symbionts in the bivalve *Lucina floridana* from seagrass beds. *Biological Bulletin* **167**: 445–459.
- Fleming J. 1828.** *A history of British animals, exhibiting the descriptive characters and systematical arrangement of the genera and species of quadrupeds, birds, reptiles, fishes, Mollusca and Radiata of the United Kingdom; including the indigenous, extirpated and extinct kinds; together with periodical and occasional visitants*. Edinburgh: Bell & Bradfute.
- Frenkiel L, Gros O, Mouëza M. 1996.** Gill structure in *Lucina pectinata* (Bivalvia: Lucinidae) with reference to hemoglobin in bivalves with symbiotic sulphur-oxidising bacteria. *Marine Biology* **125**: 511–524.
- Frenkiel L, Mouëza M. 1995.** Gill ultrastructure and symbiotic bacteria in *Codakia orbicularis* (Bivalvia, Lucinidae). *Zoomorphology* **115**: 51–61.
- Frey MA, Vermeij GJ. 2008.** Molecular phylogenies and historical biogeography of a circumtropical group of gastro-

- pods (genus: *Nerita*): implications for regional diversity patterns on the marine tropics. *Molecular Phylogenetics and Evolution* **48**: 1067–1086.
- Gaillard C, Rios M, Rolin Y, Roux M. 1992.** Fossil chemosynthetic communities related to vents and seeps in sedimentary basins: the pseudobioherms of southeastern France compared to other world examples. *Palaios* **7**: 451–465.
- Gardner JA. 1926.** The molluscan fauna of the Alum Bluff Group of Florida. Part III. Lucinacea, Leptonacea, Cardiacia. *United States Geological Survey Professional Paper 142-C*: 101–149.
- Gardner JA. 1951.** Two new guide fossils from the Tallahatta formation of the south-eastern states. *Journal of the Washington Academy of Sciences* **41**: 8–12.
- Gibson-Smith J, Gibson-Smith W. 1982.** *Lucina* s.s. (Mollusca: Bivalvia) in the western Atlantic: a reappraisal. *Veliger* **25**: 139–148.
- Gitton JL, Lozouet P, Maestrati P. 1986.** Biostratigraphie et paléocéologie des gisements types du Stampien de la région d'Etampes (Essonne). *Géologie de La France* **1**: 1–101.
- Glibert M, Van de Poel L. 1967.** Les Bivalvia fossiles du Cénozoïque étranger V. Oligodontina. I. Lucinacea, Cyamiacea, Leptonacea, Dreissenacea, Tellinacea. *Mémoires de l'Institut Royal des Sciences Naturelles de Belgique* **83**: 3–152.
- Glibert M, Van de Poel L. 1973.** Les Bivalvia Danien et du Montien de la Belgique. *Mémoires de l'Institut Royale des Sciences Naturelles de Belgique* **175**: 1–89.
- Glover EA, Taylor JD. 2001.** Systematic revision of Australian and Indo-Pacific Lucinidae (Mollusca: Bivalvia): *Pillucina*, *Wallucina* and descriptions of two new genera and four new species. *Records of the Australian Museum* **53**: 263–292.
- Glover EA, Taylor JD. 2007.** Diversity of chemosymbiotic bivalves on coral reefs: Lucinidae of New Caledonia and Lifou (Mollusca, Bivalvia). *Zoosystema* **29**: 109–181.
- Glover EA, Taylor JD. 2008.** *Callucina* and *Pseudolucinisca* (Mollusca: Bivalvia: Lucinidae) from Australia: revision of genera and description of three new species. *Records of the Western Australian Museum* **24**: 443–457.
- Glover EA, Taylor JD, Rowden AA. 2004.** *Bathyaustriella thionipta*, a new lucinid bivalve from a hydrothermal vent on the Kermadec Ridge, New Zealand and its relationship to shallow-water taxa (Bivalvia: Lucinidae). *Journal of Molluscan Studies* **70**: 283–295.
- Glover EA, Taylor JD, Williams ST. 2008.** Mangrove-associated lucinid bivalves of the central Indo-West Pacific: review of the 'Austriella' group with a new genus and species (Mollusca: Bivalvia: Lucinidae). *Raffles Museum Bulletin of Zoology* **18** (Suppl.): 25–40.
- Gros O, Frenkiel L, Mouéza M. 1998.** Gill filament differentiation and experimental colonization by symbiotic bacteria in aposymbiotic juveniles of *Codakia orbicularis* (Bivalvia: Lucinidae). *Invertebrate Reproduction and Development* **34**: 219–231.
- Gros O, Liberge M, Felbeck H. 2003.** Interspecific infection of aposymbiotic juveniles of *Codakia orbicularis* by various tropical lucinid gill-endosymbionts. *Marine Biology* **142**: 57–66.
- Harzhauser M, Kroh A, Mandic O, Piller WE, Göhlich U, Reuter M, Berning B. 2007.** Biogeographic responses to geodynamics: a key study all around the Oligo-Miocene Tethyan Seaway. *Zoologischer Anzeiger* **246**: 241–256.
- Harzhauser M, Piller WE, Steininger FF. 2002.** Circum-Mediterranean Oligo-Miocene biogeographic evolution – the gastropods' point of view. *Palaeogeography. Palaeoclimatology. Palaeoecology* **183**: 103–133.
- Hautmann M. 2001.** Die Muschelfauna der Nayband-Formation (Obertrias, Nor-Rhät) des östlichen Zentraliran. *Beringeria* **29**: 1–181.
- Hautmann M, Nützel A. 2005.** First record of a heterodont bivalve (Mollusca) from the Early Triassic: palaeontological significance and implications for the 'Lazarus problem'. *Palaeontology* **48**: 1131–1138.
- Holmer M, Duarte CM, Marba N. 2003.** Sulfur cycling and seagrass (*Posidonia oceanica*) status in carbonate sediments. *Biogeochemistry* **66**: 223–239.
- Holmer M, Pedersen O, Krause-Jensen D, Olesen B, Petersen MH, Schopmeyer S, Koch M, LomsTein BA, Jensen HS. 2009.** Sulfide intrusion in the tropical seagrasses *Thalassia testudinum* and *Syringodium filiforme*. *Estuarine, Coastal and Shelf Science* **85**: 319–326.
- Holmes AM, Oliver PG, Sellanes J. 2005.** A new species of *Lucinoma* (Bivalvia: Lucinoidea) from a methane gas seep off the southwest coast of Chile. *Journal of Conchology* **38**: 673–682.
- Huelsenbeck JP, Ronquist F. 2001.** MRBAYES: bayesian inference of phylogenetic trees. *Bioinformatics* **17**: 754–755.
- Ivimey-Cook HC, Hodges P, Swift A, Radley JD. 1999.** Bivalves. In: Swift A, Martill DM, eds. *Fossils of the Rhaetian Penarth group. Palaeontological Association field guide to fossils*, Vol. 9. London: Palaeontological Association, 83–128.
- Jackson JBC. 1972.** The ecology of molluscs of *Thalassia* communities, Jamaica, West Indies. II. Molluscan population variability along an environmental stress gradient. *Marine Biology* **14**: 304–337.
- James NP, Bone Y. 2007.** A Late Pliocene-Early Pleistocene, inner-shelf, subtropical, seagrass-dominated carbonate: roe Calcarene, Great Australian Bight, Western Australia. *Palaios* **22**: 343–359.
- Johnson MA, Fernandez C, Pergent G. 2002.** The ecological importance of an invertebrate chemoautotrophic symbiosis to phanerogam seagrass beds. *Bulletin of Marine Science* **71**: 1343–1351.
- Johnston PA. 1993.** Lower Devonian Pelecypoda from south-eastern Australia. *Memoirs of the Association of Australasian Palaeontologists* **14**: 1–134.
- Kase T, Aguilar YM. 2007.** *Bulacanites obtusiplicatus* gen. and sp. nov., a large lucinid bivalve (Mollusca) from the Pliocene of Central Luzon, Philippines. *Memoirs of the National Science Museum, Tokyo* **44**: 175–183.
- Kase T, Kurihara Y, Hagino K. 2007.** Middle Miocene chemosymbiotic thraacid *Nipponothracia gigantea*

- (Shikama, 1968) from central Japan is a large lucinid bivalve (Lucinoidea: Mollusca). *Veliger* **49**: 294–302.
- Kauffman EG, Arthur MA, Howe B, Scholle PA. 1996.** Widespread venting of methane-rich fluids in Late Cretaceous (Campanian) submarine springs (Teepee Buttes), Western Interior seaway. *Geology* **24**: 799–802.
- Kelly SRA. 1992.** Bivalvia of the Spilsby Sandstone and Sandringham Sands (Late Jurassic-Early Cretaceous) of Eastern England. Part 2. *Monograph of the Palaeontological Society London* **146**: 95–125.
- Kelly SRA, Blanc E, Price SP, Whitham AG. 2000.** Early Cretaceous giant bivalves from seep-related limestone mounds, Wollaston Forland, Northeast Greenland. In: Harper EM, Taylor JD, Crame JA, eds. *The evolutionary biology of the Bivalvia*, Geological Society of London Special Publication 177, 227–246.
- Kendrick GW, Vartak AV. 2007.** Middle Cretaceous (Cenomanian) bivalves from the Karai Formaton, Uttattur Group, of the Cauvery Basin, south India. *Records of the Western Australian Museum, Supplement* **72**: 1–101.
- Kiel S, Amano K, Jenkins RG. 2008.** Bivalves from Cretaceous cold-seep deposits on Hokkaido, Japan. *Acta Palaeontologica Polonica* **53**: 525–537.
- Kiel S, Campbell KA, Gaillard C. 2010.** New and little known mollusks from ancient chemosynthetic environments. *Zootaxa* **2390**: 26–48.
- Korobkov GA. 1954.** *Handbook and methodical guide to Tertiary Mollusca. Lamellibranchiata*. Leningrad: State and Scientific Technical Publishing Agency of Oil and Mineral Fuel Literature.
- Kurihara Y. 2007.** Occurrence of *Epilucina californica* (Conrad) (Bivalvia: Lucinidae) from the Neogene of Japan, with notes on the biogeographic history of *Epilucina*. *Paleontological Research* **11**: 29–39.
- Lamprell K, Whitehead T. 1992.** *Bivalves of Australia*. Bathurst, Australia: Crawford House Press.
- LaRocque A. 1950.** Pre-Devonian Devonian pelecypods of Michigan. *Contributions from the Museum of Paleontology University of Michigan* **7**: 271–366.
- Lebata MJHL, Primavera JH. 2001.** Gill structure, anatomy and habitat of *Anodontia edentula*; evidence of endosymbiosis. *Journal of Shellfish Research* **20**: 1273–1278.
- Lessios HA. 2008.** The great American schism: divergence of marine organisms after the rise of the Central American Isthmus. *Annual Review of Ecology, Evolution and Systematics* **39**: 63–91.
- Liljedahl L. 1991a.** Contrasting feeding strategies in bivalves from the Silurian of Gotland. *Palaeontology* **34**: 219–235.
- Liljedahl L. 1991b.** The Silurian *Iliona prisca*, oldest known deep-burrowing suspension-feeding bivalve. *Journal of Paleontology* **66**: 206–210.
- Littlewood DTJ, Curini-Galletti M, Herniou EA. 2000.** The interrelationships of *Proseriata* (Platyhelminthes: Seriata) tested with molecules and morphology. *Molecular Phylogenetics and Evolution* **16**: 449–466.
- Lockyer AE, Olsen PD, Østergaard P, Rollinson D, Johnston DA, Attwood SW, Southgate VR, Horak P, Snyder SD, Agatsuma T H Le T, McManus DP, Carmichael AC, Naem S, Littlewood DTJ. 2003.** The phylogeny of the Schistosomatidae based on three genes with emphasis on the interrelationships of *Schistosoma* (Weinland, 1858). *Parasitology* **126**: 203–224.
- Ludbrook NH. 1978.** Quaternary molluscs of the western part of the Eucla Basin. *Geological Survey of Western Australia Bulletin* **125**: 1–286.
- Maddison DR, Maddison WP. 2003.** *MacClade. Version 4.06 OSX*. Sunderland, MA: Sinauer Associates.
- Majima R, Ikeda K, Wada H, Kato K. 2003.** An outer-shelf cold-seep assemblage in forearc basin fill, Pliocene Takanabe Formation, Kyushu Island, Japan. *Paleontological Research* **7**: 297–311.
- Majima R, Nobuhara T, Kitazaki T. 2005.** Review of fossil chemosynthetic assemblages in Japan. *Palaeogeography, Palaeoclimatology, Palaeoecology* **227**: 86–123.
- Meyer E, Nilkerd B, Glover EA, Taylor JD. 2008.** Ecological Importance of chemoautotrophic lucinid bivalves in a peri – mangrove community in Eastern Thailand. *Raffles Museum Bulletin of Zoology, Supplement* **18**: 41–55.
- Monari S. 2003.** A new genus and species of fimbriid bivalve from the Kimmeridgian of the western Pontides, Turkey, and the phylogeny of the Jurassic Fimbriidae. *Palaeontology* **46**: 857–884.
- Moore HB, Davies LT, Fraser TH, Gore RH, Lopez NR. 1968.** Some biomass figures from a tidal flat in Biscayne Bay, Florida. *Bulletin of Marine Science* **18**: 2261–2279.
- Murray JW, Wright CA. 1974.** Palaeogene Foraminiferida and palaeoecology, Hampshire and Paris Basins and the English Channel. *Special Papers in Palaeontology* **14**: 1–129.
- Nakaoka M, Mukai H, Chunhabundit S. 2002.** Impacts of dugong foraging on benthic animal communities in a Thailand seagrass bed. *Ecological Research* **17**: 625–638.
- Nicol D. 1950.** Recent species of the lucinoid pelecypod *Fimbria*. *Journal of the Washington Academy of Sciences* **40**: 82–87.
- Okutani T, Hashimoto J. 1997.** A new species of lucinid bivalve (Heterodonta: Lucinidae) from Kanesu-no-Se bank near the mouth of Suruga Bay, with a review of the Recent species of the chemosynthetic genus *Lucinoma* from Japan. *Venus* **56**: 271–280.
- Oliver PG, Holmes AM. 2006.** A new species of *Lucinoma* (Bivalvia: Lucinoidea) from the oxygen minimum zone of the Oman margin, Arabian Sea. *Journal of Conchology* **39**: 63–77.
- Olu-Le Roy K, Sibuet M, Fiala-Médioni A, Gofas S, Salas C, Mariotti A, Foucher J-P, Woodside J. 2004.** Cold seep communities in the deep eastern Mediterranean Sea: composition, symbiosis and spatial distribution on mud volcanoes. *Deep Sea Research I* **51**: 1915–1936.
- Pacaud J-M, Le Renard J. 1995.** Révision des mollusques Paléogènes du Bassin de Paris IV – Liste systématique actualisée. *Cossmanniana* **3**: 151–187.
- Paulay G. 1997.** Diversity and distribution of reef organisms. In: Birkeland C, ed. *Life and death of coral reefs*. New York: Chapman and Hall, 298–353.

- Plaziat J-C, Cavagnetto C, Koeniguer J-C, Baltzer F. 2001.** History and biogeography of the mangrove ecosystem, based on a critical reassessment of the paleontological record. *Wetland Ecology and Management* **9**: 161–179.
- Reid DG, Dyal P, Lozouet P, Glaubrecht M, Williams ST. 2008.** Mudwhelks and mangroves: the evolutionary history of an ecological association (Gastropoda: Potamididae). *Molecular Phylogenetics and Evolution* **47**: 680–699.
- Reid DG, Dyal P, Williams ST. 2010.** Global diversification of mangrove fauna: a molecular phylogeny of Littoraria (Gastropoda: Littorinidae). *Molecular Phylogenetics and Evolution* **55**: 185–201.
- Rögl F. 1998.** Palaeogeographic considerations for Mediterranean and Paratethys seaways (Oligocene to Miocene). *Annalen des Naturhistorischen Museums in Wien* **99A**: 279–310.
- Sacco F. 1901.** *I molluschi de terreni terziarii del Piemonte e della Liguria. Part 29.* Torino: C. Clausen.
- Salas C, Woodside J. 2002.** *Lucinoma kazani* n.sp. (Mollusca: Bivalvia: evidence of a living benthic community associated with a cold seep in the eastern Mediterranean Sea. *Deep-Sea Research I* **49**: 991–1005.
- Saul LR, Squires RL, Godert JE. 1996.** A new genus of cryptic lucinid bivalve from Eocene cold seeps and turbidite influenced mudstone, Western Washington. *Journal of Paleontology* **70**: 788–794.
- Speden IG. 1970.** The type Fox Hills Formation, Cretaceous (Maestrichtian), South Dakota. Part 2. Systematics of the Bivalvia. *Bulletin Peabody Museum of Natural History* **33**: 1–222.
- Strougo A. 1975.** *Lucina pharaonis* Bellardi, 1854. *Bulletin Société Géologique de France* ser. 7 **17**: 746–751.
- Taviani M. 1994.** The ‘calcarei a *Lucina*’ macrofauna reconsidered: deep-sea faunal oases from Miocene-age cold vents in the Romagna Apennine, Italy. *Geo-Marine Letters* **14**: 185–191.
- Taylor JD, Glover EA. 1997.** The lucinid bivalve genus *Cardiolucina* (Mollusca, Bivalvia, Lucinidae): systematics, anatomy and relationships. *Bulletin of the Natural History Museum, London (Zoology)* **63**: 93–122.
- Taylor JD, Glover EA. 2000.** Functional anatomy, chemosymbiosis and evolution of the Lucinidae. In: Harper EM, Taylor JD, Crame JA, eds. *The Evolutionary Biology of the Bivalvia*, Geological Society of London Special Publication 177, 207–225.
- Taylor JD, Glover EA. 2002.** *Lamellolucina*: a new genus of lucinid bivalve with four new species from the Indo-West Pacific. *Journal of Conchology* **37**: 317–336.
- Taylor JD, Glover EA. 2005.** Cryptic diversity of chemosymbiotic bivalves: a systematic revision of worldwide *Anodontia* (Mollusca: Bivalvia: Lucinidae). *Systematics and Biodiversity* **3**: 281–338.
- Taylor JD, Glover EA. 2006.** Lucinidae (Bivalvia) – the most diverse group of chemosymbiotic molluscs. *Zoological Journal of the Linnean Society* **148**: 421–438.
- Taylor JD, Glover EA. 2009a.** New lucinid bivalves from hydrocarbon seeps of the Western Atlantic (Mollusca: Bivalvia: Lucinidae). *Steenstrupia* **30**: 127–140.
- Taylor JD, Glover EA. 2009b.** A giant lucinid bivalve from the Eocene of Jamaica – systematics, life habits and chemosymbiosis (Mollusca: Bivalvia: Lucinidae). *Palaeontology* **52**: 95–109.
- Taylor JD, Glover EA. 2010.** Chemosymbiotic bivalves. In: Kiel S, ed. *The vent and seep biota – from microbes to ecosystems. Topics in Geobiology, Vol. 33.* Heidelberg: Springer, 107–135.
- Taylor JD, Glover EA, Peharda M, Bigatti G, Ball A. 2004.** Extraordinary flexible shell sculpture; the structure and formation of calcified periostracal lamellae in *Lucina pennsylvanica* (Bivalvia: Lucinidae). *Malacologia* **46**: 277–294.
- Taylor JD, Glover EA, Williams ST. 2005.** Another bloody bivalve: anatomy and relationships of *Eucrassatella donacina* from south Western Australia (Mollusca: Bivalvia: Crassatellidae). In: Wells FE, Walker DI, Kendrick GA, eds. *The marine fauna and flora of Esperance, Western Australia.* Perth: Western Australian Museum, 261–288.
- Taylor JD, Glover EA, Williams ST. 2008.** Ancient shallow water chemosymbiotic bivalves: systematics of Solemyidae of eastern and southern Australia. *Memoirs of the Queensland Museum-Nature* **54**: 75–104.
- Taylor JD, Glover EA, Williams ST. 2009.** Phylogenetic position of the bivalve family Cyrenoididae – removal from (and further dismantling of) the superfamily Lucinoidea. *Nautilus* **123**: 1–5.
- Taylor JD, Lewis MS. 1970.** The flora, fauna and sediments of the marine grass beds of Mahé, Seychelles. *Journal of Natural History* **4**: 199–220.
- Taylor JD, Williams ST, Glover EA. 2007.** Evolutionary relationships of the bivalve family Thyasiridae (Mollusca: Bivalvia), monophyly and superfamily status. *Journal of the Marine Biological Association of the United Kingdom* **87**: 565–574.
- Taylor JD, Williams ST, Glover EA, Dyal P. 2007.** A molecular phylogeny of heterodont bivalves (Mollusca: Bivalvia: Heterodonta): new analyses of 18S rRNA and 28S rRNA genes. *Zoologica Scripta* **36**: 587–606.
- Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG. 1997.** The CLUSTAL_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research* **25**: 4876–4882.
- Thompson JD, Higgins DG, Gibson TJ. 1994.** CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties, and weight matrix choice. *Nucleic Acids Research* **22**: 4673–4680.
- Tracey S, Todd JA, Le Renard J, King C, Goodchild M. 1996.** Distribution of Mollusca in units S1 to S9 of the Selsey Formation (Middle Lutetian), Selsey Peninsula, West Sussex. *Tertiary Research* **16**: 97–139.
- Uede T, Takahashi Y. 2008.** Benthos communities of the *Zostera japonica* meadows in the intertidal zone of Takinai and Uchinoura, Tanabe Bay, Wakayama Prefecture, Japan. *Japanese Journal of Benthology* **63**: 42–55.

- Vrielynck B, Odin GS, Dercourt J. 1997.** Miocene paleogeography of the Tethys Ocean; potential global correlations in the Mediterranean. In: Montanari A, Odin G, Coccioni R, eds. *Miocene stratigraphy: an integrated approach*. Amsterdam: Elsevier Science, 157–165.
- Wellnhofer P. 1964.** Zur Pelecypodenfauna der Neuburger Bankkalke (Mittel-Tithon). *Abhandlungen der Bayerischen Akademie der Wissenschaften. Mathematisch-Naturwissenschaftliche Klasse. Neue Folge* **119**: 1–143.
- Williams ST, Duda TF. 2008.** Did tectonic activity stimulate Oligo-Miocene speciation in the Indo-West Pacific? *Evolution* **62**: 1618–1634.
- Williams ST, Ozawa T. 2006.** Molecular phylogeny suggests polyphyly of both the turban shells (family Turbinidae) and the superfamily Trochoidea (Mollusca: Vetigastropoda). *Molecular Phylogenetics and Evolution* **39**: 33–51.
- Williams ST, Reid DG. 2004.** Speciation and diversity on tropical rocky shores: a global phylogeny of snails of the genus *Echinolittorina*. *Evolution* **58**: 2227–2251.
- Williams ST, Reid DG, Littlewood DTJ. 2003.** A molecular phylogeny of the Littorininae (Gastropoda: Littorinidae): unequal evolutionary rates, morphological parallelism and biogeography of the Southern Ocean. *Molecular Phylogenetics and Evolution* **28**: 60–86.
- Williams ST, Taylor JD, Glover EA. 2004.** Molecular phylogeny of the Lucinoidea (Bivalvia): non-monophyly and separate acquisition of bacterial chemosymbiosis. *Journal of Molluscan Studies* **70**: 187–202.
- Winnepenninckx BMH, Reid DG, Backeljau T. 1998.** Performance of 18S rRNA in littorinid phylogeny (Gastropoda: Caenogastropoda). *Journal of Molecular Evolution* **47**: 586–596.
- Zabbey N, Hart AI, Wolff WJ. 2010.** Population structure, biomass and production of the West African lucinid *Kelettistes rhizoecus* (Bivalvia, Mollusca) in Sivivilagbara swamp at Bodo Creek, Niger Delta, Nigeria. *Hydrobiologia* **654**: 193–203.
- Zelaya DG. 2005.** The bivalves from the Scotia Arc Islands: species richness and faunistic affinities. *Scientia Marina* **69**(Suppl. 2): 113–122.
- Zong-Jie F, Cope JCW. 2004.** Early Ordovician bivalves from Dali, West Yunnan, China. *Palaeontology* **47**: 1121–1158.