# Acanthochondria hoi, a new species of parasitic copepod (Poecilostomatoida: Chondracanthidae) on the California halibut, Paralichthys californicus, from Santa Monica Bay, California, with an amended key to the genus Acanthochondria

Julianne E. Kalman

Department of Organismic Biology, Ecology and Evolution, University of California, Los Angeles (UCLA), 621 Charles E. Young Drive South, Box 951606, Los Angeles, California 90095-1606

Abstract.—A new species of parasitic copepod, Acanthochondria hoi, is described from specimens collected within the gill cavity of the California halibut, *Paralichthys californicus* (Ayers), from Santa Monica Bay, California. Acanthochondria hoi can be distinguished from its congeners by the combination of a Type B-V antennule and Type A leg 2, in addition to leg 1 ornamentation. A revision of the key of Acanthochondria prepared by previous authors is provided and includes three new species.

Santa Monica Bay is located in the Southern California Bight and is an open coastal embayment bounded by Point Dume to the north and Palos Verdes Point to the south. The Hyperion Treatment Plant (Bureau of Sanitation, Department of Public Works, City of Los Angeles) provides secondary treatment and disposal of treated wastewater through a 5-mile effluent outfall located in Santa Monica Bay. The Environmental Monitoring Division conducts quarterly otter trawls to monitor the effects of the effluent on the fishes and macroinvertebrates living in the vicinity of the outfall (Dojiri & Brantley 1991). During the July/ August and November 1998, and February and May 1999 trawls, several specimens of California halibut, Paralichthys californicus (Ayers), were collected with parasitic copepods within the gill cavity. These parasites represent a new species of Acanthochondria, which is described below.

*Materials and methods.*—The fishes were collected in Santa Monica Bay, California. Quarterly otter trawls were made aboard the R/V *La Mer* in association with the Environmental Monitoring Division,

Bureau of Sanitation, Department of Public Works, City of Los Angeles. Immediately after the catch was brought on board, the fishes were placed in plastic bags and kept on ice in a cooler for a later examination in the laboratory. The copepods were removed and preserved in 70% isopropyl alcohol, then cleared in 85% lactic acid. They were measured with an ocular micrometer and selected specimens were dissected. Illustration were drawn with the aid of a camera lucida. Holotype and paratypes were deposited in the National Museum of Natural History, Smithsonian Institution, Washington, D.C. (1001623-1001628). Additional specimens are in the collection of the author.

## Systematic Account

Order Poecilostomatoida Thorell, 1859 Family Chondracanthidae Milne-Edwards, 1840 Genus Acanthochondria Oakley, 1927 Acanthochondria hoi, new species Figs. 1–3

*Material examined.*—A total of seven nonovigerous and 22 ovigerous females



Fig. 1. Acanthochondria hoi n. sp., female. A, habitus, dorsal; B, habitus, lateral; C, genito-abdomen, lateral; D, caudal ramus; E, antennule; F, antenna; G, mandible. Scale: 1.0 mm in A, B; 0.1 mm in C, F; 0.05 mm in D, G; 0.2 mm in E.

(each with an attached male) was collected from within the gill cavity (inner side of operculum and floor of oral cavity at the base of gill arches) of the California halibut, *Paralichthys californicus* (Ayers).

Female.—The trunk (Fig. 1A, B) is long and slender. The cephalosome is slightly longer than wide. Neck region consisting of first and second pedigers. Trunk with a single mid-lateral indentation and bearing a pair of posterior processes, which are moderately long and slender. The genital segment (Fig. 1C) is longer than wide; and the abdomen (Fig. 1C) is shorter than the genital segment and bears two dorsal setules. The caudal ramus (Fig. 1D) has three setae, a knob, and a large spinulated terminal process. The antennule (Fig. 1E) is of Type B-V (Ho & Kim 1995), consisting of a large unarmed basal portion and a small cylindrical distal portion with an armature formula of 2-2-8. The antenna (Fig. 1F) is 2segmented; the basal segment is large and unarmed; the terminal claw possesses a small mid-lateral seta and a minute basal setule. The mandible (Fig. 1G) is 2-segmented; convex margin armed with 31-35 teeth, concave margin has 21-24 teeth.

The maxillule (Fig. 2A) is a lobe bearing two processes produced at the distal margin of the appendage. The maxilla (Fig. 2B) is 2-segmented, with the basal segment unarmed. The terminal segment carries a small seta, a large seta, and a large process armed with 15 teeth along its posterior margin. The maxilliped (Fig. 2C) is 3-segmented; the first segment is unarmed; the second segment bears a protruded portion on which one patch of spinules is located and a row of 12 teeth on the distal margin; the terminal segment is clawlike, bearing a proximal patch of small spinules and a subterminal accessory process. Leg 1 (Fig. 2D) is biramous with the exopod bearing an outer seta. The anterior surface is covered with irregular patches of spinules. Leg 2 (Fig. 2E) is biramous with long rami. The exopod carries an outer seta. The rami bear spinules at the distal tips of the anterior surfaces in addition to several setules.

*Measurements.*—Total length (tip of cephalosome to tip of posterior process) 6.84 mm; trunk width 0.44 mm; cephalosome 0.39 mm  $\times$  0.35 mm; genital segment 0.47 mm  $\times$  0.44 mm; abdomen 0.19 mm  $\times$  0.16 mm; posterior process 1.32 mm.

Male.—The body (Fig. 3A), 0.67 mm  $\times$ 0.34 mm, is ventrally flexed. The cephalosome and the first pedigerous segment comprise more than half the total length. The antennule (Fig. 3B) is slender and bears an armature formula of 1-1-2-2-8. The antenna (Fig. 3C) is 2-segmented. The basal segment possesses a rounded knob near the articulation with the terminal claw. The terminal segment bears two setae on the basal portion. The mandible (Fig. 3D) is 2-segmented; terminal segment armed with 20 teeth along convex margin, 9 teeth along concave margin. The maxilla (Fig. 3E) is 2segmented and exhibits the usual sexual dimorphism for this genus by possessing a naked terminal process. Leg 1 (Fig. 3F) is larger than leg 2 (Fig. 3G). Both legs are similarly armed, with the protopod bearing a long outer seta, the exopod with two small elements, and the endopod a smaller unarmed lobe. However, the two elements on the exopod of leg 2 are unequal in size.

*Etymology.*—This species is named after Dr. Ju-Shey Ho, an expert in parasitic copepod research and my mentor and former advisor.

*Remarks.*—*Acanthochondria hoi,* new species, was previously reported by Dojiri (1977) as *Acanthochondria* sp. C. However, a literature search revealed that a description of this species was never published. Ho (1975) tentatively identified a badly damaged specimen of *Acanthochondria* from the California halibut as *A. soleae* (?). In addition, Haaker (1975) and Allen (1990) reported *A. soleae* to occur on the California halibut, directly and indirectly citing Ho (1975), respectively. Ho's (1975) specimen is probably identifiable with *A. hoi.* Kabata (1979) comments that the literature contains



Fig. 2. *Acanthochondria hoi* n. sp., female. A, maxillule; B, maxilla; C, maxilliped; D, leg 1; E, leg 2. Scale: 0.05 mm in A, B, C; 0.3 mm in D, E.

a number of erroneous host listings of *A. soleae*. *A. soleae* is parasitic mainly on the sole, *Solea solea*, and its distribution is restricted to the Atlantic Ocean (Kabata 1979). *A. hoi* differs from *A. soleae* in the relative lengths of the endopod and exopod, leg 1 ornamentation, and structure of the maxilliped.

Acanthochondria hoi is distinguished from its congeners by the combination of a Type B-V antennule and Type A leg 2 (Ho & Kim 1995), in addition to leg 1 ornamentation. Legs 1 and 2 have relatively long rami with the endopod noticeably longer than the exopod. A check into the key of *Acanthochondria* prepared by Ho and Kim (1995) revealed that this specimen collected from the California halibut is new to science. This specimen keyed out to step 33a, which is equivalent to *A. exilipes* (Ho 1971). Table 1 lists differences between *A. hoi* and *A. exilipes*.

### Key to the Species of Acanthochondria

The following revised key includes all accepted species of *Acanthochondria* (Ho & Kim 1995). Three new species were add-



Fig. 3. Acanthochondria hoi n. sp., male. A, habitus, lateral; B, antennule; C, antenna; D, mandible; E, maxilla; F, leg 1; G, leg 2. Scale: 0.1 mm in A; 0.02 mm in B, C, D, E, F, G.

ed, *A. kajika* (Ho & Kim 1996), *A. zebriae* (Ho et al. 2000), and *A. hoi* (Kalman, this report), increasing the number of species to 46. In addition, all typographical errors have been corrected (most notable, from Ho and Kim (1995): step 16b should lead to step 29, not step 28 as previously noted; *A. cyclopsetta, A. exilipes, A. galerita,* and *A.* 

*physidis* should all be cited as Ho 1971, not Ho 1970 as previously noted).

Poly and Mah (2001) deeply criticize some of the characters used in the key by Ho and Kim (1995). However, this revised key is still valid and useful until further morphological characters can be discovered for some nominal species where the host

Table 1.—Differences between Acanthochondria exilipes and A. hoi.

	A. exilipes	A. hoi
Antennule type	B-III	B-V
Teeth on mandible	37–41 on convex margin	31-35 on convex margin
	32–34 on concave margin	21-24 on concave margin
Maxillule	2 patches of spinules	naked
Leg 1 ornamentation	naked	patches of spinules
2nd segment of	2 patches of spinules	1 patch of spinules
maxilliped		and 12 teeth on
		outer margin

family is used as a "character". Thus, the publication containing the best information to aid in species identification is provided in parentheses after each species name.

The males of *Acanthochondria* do not show species differences; therefore, the characters used in this key refer strictly to adult ovigerous females (Ho 1970). For types of antennule and leg 2 found in this key, refer to Ho and Kim (1995).

Neck region consisting of first pediger
only 2
Neck region consisting of first and sec-
ond pedigers 4
Neck region consisting of second pe-
diger only; first pediger incorporated
into head region triglae
(Herrera-Cubilla & Raibaut 1990:82-87)
Second pediger indistinguishably fused
to trunk
Second pediger distinctly separated
from trunk and bearing a pair of large
rounded swellings limandae
(Kabata 1979:127–128)
Antenna of B-VII type laemonemae
(Capart 1959:102-103)
Antenna of B-III type lepidionis
(Ho 1972a:147–149)
Antenna of B-I type zebriae
(Ho et al. 2000:711–713)
Neck very long, at least 8 times longer
than wide 5
Neck moderately long, at most about 3
times as long as wide; leg 2 with ex-
tremely long protopod (Type E) 6
Neck short; at most slightly longer than
wide; protopod of leg 2 not greatly
elongated 8
Posterolateral processes short and
blunt; terminal process of maxilla bear-
ing a short row of fine denticles
diastema (Ho & Dojiri 1988:273–279)
Posterolateral processes long and slen-
der; terminal process of maxilla bear-
ing a long row of large teeth uranoscopi
(Ho & Kim 1995:48–51)
Endopod of leg 2 much reduced, rep-
resented by a little knob tchangi
•
(Shiino 1959:361)

as exopod .....

7

7a	Cephalosome about as long as wide; parasite of Platycephalidae <i>platycephali</i>
	(Но 1973:127–130)
b	Cephalosome distinctly longer than
	wide; parasite of fishes other than Pla-
	tycephalidae inimici
80	(Dojiri & Ho 1988:47–53)
oa	longer than wide) and without lateral
	indentations
b	Trunk appearance otherwise 11
9a	Posterolateral processes shorter than
	head; caudal ramus shorter than abdo-
	men 10
b	Posterolateral processes longer than
	head; caudal ramus distinctly longer
	(Pillai 1985:125_127)
10a	Cephalosome with small rounded knob
	at each anterior corner; hook-like an-
	tenna curved in distal region fraseri
	(Ho 1972b:523–527)
b	Cephalosome with large anterolateral
	swellings; hook-like antenna curved in
	(Vii & Wu 1032:66 68)
11a	Trunk trapezoidal: postoral region
	elongated dilatata
	(Shiino 1955:107–110)
b	Trunk shaped otherwise; leg 1 close to
	oral region 12
12a	Cephalosome bearing a pair of lateral
	horn-like projections; trunk without
	(Shiino 1955:103–107)
b	Cephalosome without such projections:
	trunk mostly with lateral indentations
	13
13a	Leg 2 long and slender (Type D) 14
b	Leg 2 shaped otherwise 15
14a	Posterolateral processes long, as long
	(Kabata 1970:128, 120)
b	Posterolateral processes short distinct-
U	ly shorter than 4th pediger cyclopsetta
	(Ho 1971:3)
15a	Legs 1 and 2 indistinctly bilobated
	(Type B) physidis
k	(Ho 1971:11–15)
D 160	Legs I and 2 distinctly bilobated 16
	A DEDUDE WITTOTT TOTALED DASAL DAD

nu	7 micimule	without	mateu	Dasar part	
	(Type A)				17

# VOLUME 116, NUMBER 3

b	Antennule with inflated basal part
	(Type B) 30
17a	Leg 2 slightly larger than leg 1 18
b	Leg 2 distinctly larger than leg 1 24
18a	Both legs 1 and 2 covered with spi-
	nules 19
b	Both legs 1 and 2 without spinules or
	bearing at most only patches of spi-
	nules
19a	First pediger with lateral protuberance
	sixteni (Dojiri & Ho 1988:53–56)
b	First pediger without such protuber-
	ance dojirii (Kabata 1984:1708–1910)
20a	Cephalosome distinctly longer (at least
	1.38 times) than wide 21
b	Cephalosome about as long as wide 22
21a	Distal part of leg rami covered with
214	spinules vancouverensis
	(Kabata 1984:1710)
h	Distal part of leg rami not covered with
U	spinules alaudicans
	(Shiino 1055.03 06)
$\gamma\gamma_{n}$	(Sinnio 1955.95–90) Trunk about as long as wide
22a h	Trunk about as long as while 25
U	mulk distinctly foliger than wide
222	Demoitie en Silleginidee
258	Parastuc on Sinaginidae shawi
1.	(10.1935:7-9)
b	Parasitic on Gobiidae $\dots$ yui
24	(Shiino 1964:30–33)
24a	lerminal process of maxilla armed
	with a long row of teeth (about 15) 25
b	Terminal process of maxilla armed
	with a short row of teeth (at most 11)
25a	Cephalosome round in dorsal view
	brevicorpa (Yamaguti 1939:535)
b	Cephalosome pear-shaped in dorsal
	view longifrons
	(Shiino 1955:86-89)
26a	Both legs bearing large, prominent
	patches of spinules margolisi
	(Kabata 1984:1705)
b	Both legs naked or with small patches
	of spinules 27
27a	Trunk distinctly longer than wide and
	with prominent lateral indentations 28
b	Trunk about as long as wide with slight
-	lateral indentations 29
28a	Parasitic on Serranidae constricta
20u	(Shiino 1955.96_100)
b	Parasitic on Pleuronectidae kinnedossi
0	(Kabata 1087-215)
200	Labrum with lateral protrucion lage 1

b	and 2 tipped with spinules on both rami <i>kajika</i> (Ho & Kim 1996:276–279) Labrum without lateral protrusion, legs 1 and 2 naked <i>fissicauda</i>
	(Shiino 1955:90–93)
30a	Cephalosome with two lateral round
	swellings on ventral surface of head;
	antennule with prominent ventral pro-
	(Vabata 1070:126 127)
h	(Kabata 1979:120–127) Cephalosome and antennula without
U	such features
31a	Both rami of leg 2 large coniform
514	(Type C): a pair of large protuberances
	lateral to labrum in oral area galerita
	(Ho 1971:8–11)
b	Leg 2 and oral area without such fea-
	tures
32a	Leg 2 distinctly larger than leg 1 33
b	Leg 2 only slightly larger than leg 1
33a	Trunk wider than long; posterolateral
	processes short and blunt tasmaniae
	(Heegaard 1962:154–155)
b	Trunk about as long as wide; postero-
	lateral processes long priacanthi
	(Ho & Kim 1995:53–56)
с	Trunk distinctly longer than wide; pos-
	terolateral processes either long or
24.	short
34a	Endopod of leg 2 about as long as pro-
h	Endered of log 2 distinctly shorter
U	than protopod of leg 2 distinctly shorter
350	Antennule of B III type: leg 1 noked
<i>JJa</i>	erilines (Ho 1971:3_7)
h	Antennule of B-V type: leg 1 with
0	patches of spinules hoi
	(Kalman this report)
36a	Terminal process of maxilla bearing
	less than 10 teeth epachthes
	(Kabata 1968:339–344)
b	Terminal process of maxilla armed
	with at least 15 teeth oralis
	(Yamaguti 1939:536–537)
37a	Trunk as long as wide or slightly lon-
	ger than wide 38
b	Trunk distinctly longer than wide 40
38a	Cephalosome large, as wide as trunk
	and bearing a pair of anterolateral pro-
	tuberances macrocephala

(Ho & Kim 1995:46–48)

- b Cephalosome distinctly narrower than trunk, without protuberance ..... 39
- 39a Cephalosome slightly longer than wide; endopod of leg 2 distinctly longer than exopod ..... incisa (Shiino 1955:83–86)
- b Cephalosome distinctly longer than wide; endopod of leg 2 about as long as exopod ... *ophidii* (Ho 1977:158–160)
- 40a Cephalosome about as long as wide; both legs covered with spinules ..... ...... rectangularis (Kabata 1984:1705)
- b Cephalosome wider than long; both legs with spinules on rami only .... 41
- 41a Cephalosome with two prominent lateral protrusions; antennule of Type B-II ..... sicyasis (Ho 1977:160–164)
- b Cephalosome with swollen oral region; antennule of Type B-V ..... cornuta (Ho 1970:121–127)

#### Acknowledgements

This paper represents a modified portion of a thesis completed at California State University, Long Beach (CSULB). I thank Dr. Ju-Shey Ho (CSULB) for his guidance; Dr. Masahiro Dojiri (Bureau of Sanitation, Department of Public Works, City of Los Angeles), Dr. Donald G. Buth (University of California, Los Angeles), and Dr. Alan Miller (CSULB) for their valuable suggestions; the Environmental Monitoring Division staff (City of Los Angeles) for their field assistance; Danny Tang for his input; and my parents, Marjatta and Marvin Kalman, for their support.

## Literature Cited

- Allen, M. J. 1990. The biological environment of the California halibut, *Paralichthys californicus*.
  Pp. 7–29 in C. W. Haugen, ed., The California halibut, *Paralichthys californicus*, resource and fisheries. California Department of Fish and Game, Fish Bulletin 174.
- Capart, A. 1959. Copépodes parasites. *In* Results Expedition oceanographique Belge dans les Ezux Cotieres Africines de l'Atlantique sud (1948– 1949).—Institut Royal des Sciences naturelles de Belgique 3(5):57–126.
- Dojiri, M. 1977. Copepod parasites of Orange County sewer outfall. M.A. thesis, California State University, Long Beach, 295 pp.

- —, & J. S. Ho. 1988. Two species of Acanthochondria (Copepoda: Poecilostomatoida) parasitic on fishes of Japan.—Report of the Sado Marine Biological Station Niigata University 18:47–56.
- , & R. A. Brantley. 1991. Lepeophtheirus spatha, a new species of copepod (Siphonostomatoida: Caligidae) parasitic on the California halibut from Santa Monica Bay, California.—Proceedings of the Biological Society of Washington 104:727–735.
- Haaker, P. L. 1975. The biology of the California halibut, *Paralichthys californicus* (Ayres), in Anaheim Bay, California. Pp. 137–151 in E. D. Lane and C. W. Hill, eds., The marine resources of Anaheim Bay. California Department of Fish and Game, Fish Bulletin 165.
- Heegaard, P. E. 1962. Parasitic Copepoda from Australian waters.—Records of Australian Museum 25(9):149–234.
- Herrera-Cubilla, A., & A. Raibaut. 1991. Acanthochondria triglae n. sp., copépode parasite des fosses nasals de poisons Triglidae.—Crustaceana 59(1):82–88.
- Ho, J. S. 1970. Revision of the genera of the Chondracanthidae, a copepod family parasitic on marine fishes.—Beaufortia 17(229):105–218.
- ———. 1971. Parasitic copepods of the family Chondracanthidae from fishes of Eastern North America.—Smithsonian Contributions to Zoology 87:1–39.

- . 1973. Chondracanthid copepods parasitic on platycephalid fishes of Australia, with discussion of known species occurring on flatheads.— Parasitology 67:123–131.
- —. 1975. Parasitic Crustacea. Pages 69–72 in E.
   D. Lane and C. W. Hill, eds., The marine resources of Anaheim Bay. California Department of Fish and Game, Fish Bulletin 165.
- . 1977. Parasitic copepods of the family Chondracanthidae from fishes of the south-eastern Pacific (Crustacea, Copepoda).—Steenstrupia 4(13):157–166.
- —, & M. Dojiri. 1988. Copepods of the family Chondracanthidae parasitic on Australian marine fishes.—Australian Journal of Zoology 36: 273–291.
- —, & I. H. Kim. 1995. Acanthochondria (Copepoda: Chondracanthidae) parasitic on fishes of Sado Island in the Sea of Japan, with a preliminary review of the genus.—Report of the Sado

Marine Biological Station, Niigata University 25:45–67.

- ——, I. H. Kim, & A. B. Kumar. 2000. Chondracanthid copepods parasitic on flatfishes of Kerala, India.—Journal of Natural History 34:709– 735.
- Kabata, Z. 1968. Some Chondracanthidae (Copepoda) from fishes of British Columbia.—Journal Fisheries Research Board of Canada 25(2):321–345.
  —, 1979. Parasitic Copepoda of British Fishes. Ray Society, London, 468 pp.
- , 1984. A contribution to the knowledge of Chondracanthidae (Copepoda: Poecilostomatoida) parasitic on fishes of British Columbia.
   Canadian Journal of Zoology 62:1703–1713.
  - —, 1987. Acanthochondria hippoglossi sp. nov. (Copepoda, Chondracanthidae), a crustacean parasitic of some flatfishes off the Pacific coast of North America.—Canadian Journal of Zoology 65:213–216.
- Pillai, N. K. 1985. The fauna of India—copepod parasites of marine fishes.—Zoological Survey of India 900 pp.
- Poly, W. J., and C. L. Mah. 2001. New host distribution records for parasitic copepods in the North-

east Pacific Ocean with a discussion of taxonomy of the genus *Acanthochondria*.—Bulletin of Marine Science 69(3):1121–1127.

- Shiino, S. M. 1955. Copepods parasitic on Japanese fishes, 9. Family Chondracanthidae, subfamily Chondracanthinae.—Report of Faculty of Fisheries, Prefectural University of Mie 2(1):70– 111.
- ——. 1964. On two new species of the genus Acanthochondria Oakley (Crustacea Copepoda) found in Japan.—Zoologische Mededelingen 34:30–36.
- Yamaguti, S. 1939. Parasitic copepods from fishes of Japan. 6, Lernaeopodida I.—Volumen Jubilare Professor Sadao Yoshida 2:529–578.
- Yü, S. C. 1935. Studies on the parasitic copepods of China belonging to the family Chondracanthidae.—Bulletin of the Fan Memorial Institue of Biology 6(1):1–15.
  - , & H. W. Wu. 1932. Parasitic copepods of the flatfishes from China.—Bulletin of the Fan Memorial Institue of Biology 3(4):55–75.