

A Revised List of the Freshwater Mussels (Mollusca: Bivalvia: Unionida) of the United States and Canada

Authors: Williams, James D., Bogan, Arthur E., Butler, Robert S., Cummings, Kevin S., Garner, Jeffrey T., et al.

Source: Freshwater Mollusk Biology and Conservation, 20(2): 33-58

Published By: Freshwater Mollusk Conservation Society

URL: https://doi.org/10.31931/fmbc.v20i2.2017.33-58

BioOne Complete (complete.BioOne.org) is a full-text database of 200 subscribed and open-access titles in the biological, ecological, and environmental sciences published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Complete website, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/terms-of-use.

Usage of BioOne Complete content is strictly limited to personal, educational, and non - commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

REGULAR ARTICLE

A REVISED LIST OF THE FRESHWATER MUSSELS (MOLLUSCA: BIVALVIA: UNIONIDA) OF THE UNITED STATES AND CANADA

James D. Williams^{1*}, Arthur E. Bogan², Robert S. Butler^{3,4}, Kevin S. Cummings⁵, Jeffrey T. Garner⁶, John L. Harris⁷, Nathan A. Johnson⁸, and G. Thomas Watters⁹

ABSTRACT

We present a revised list of freshwater mussels (order Unionida, families Margaritiferidae and Unionidae) of the United States and Canada, incorporating changes in nomenclature and systematic taxonomy since publication of the most recent checklist in 1998. We recognize a total of 298 species in 55 genera in the families Margaritiferidae (one genus, five species) and Unionidae (54 genera, 293 species). We propose one change in the Margaritiferidae: the placement of the formerly monotypic genus *Cumberlandia* in the synonymy of *Margaritifera*. In the Unionidae, we recognize three new genera, elevate four genera from synonymy, and place three previously recognized genera in synonymy. We recognize for the first time two species (one native and one nonindigenous) in the Asian genus *Sinanodonta* as occurring in North America. We recognize four new species and one subspecies and elevate 21 species from synonymy. We elevate 10 subspecies to species status and no longer recognize four subspecies. We change common names for five taxa, correct spelling for eight species, and correct the date of publication of original descriptions for four species.

KEY WORDS: Unionidae, Margaritiferidae, taxonomy, systematics, nomenclature, mussel scientific names, mussel common names

INTRODUCTION

During the past 50 yr, there has been considerable interest in freshwater mussels (order Unionida) in the United States and Canada. Much of this interest was brought about by passage of the U.S. Endangered Species Acts of 1966, 1969, and 1973 and the Canadian Species at Risk Act of 2002. These legislative actions and the environmental movement that accompanied them focused conservation attention on all animals and plants, as well as their habitats. This in turn led

¹ Florida Museum of Natural History, Museum Road and Newell Drive, Gainesville, FL 32611 USA

² North Carolina Museum of Natural Sciences, MSC 1626, Raleigh, NC 27699 USA

³ U.S. Fish and Wildlife Service, 212 Mills Gap Road, Asheville, NC 28803 USA

⁴ Retired.

⁵ Illinois Natural History Survey, 607 East Peabody Drive, Champaign, IL 61820 USA

⁶ Alabama Division of Wildlife and Freshwater Fisheries, 350 County Road 275, Florence, AL 35633 USA

⁷ Department of Biological Sciences, Arkansas State University, State University, AR 71753 USA

⁸ U.S. Geological Survey, Wetland and Aquatic Research Center, 7920 NW 71st Street, Gainesville, FL 32653 USA

⁹ Museum of Biological Diversity, The Ohio State University, 1315 Kinnear Road, Columbus, OH 43212 USA

^{*}Corresponding Author: fishwilliams@gmail.com

to assessment of species conservation status and the development of faunal lists for many states and provinces. The task of developing species lists was difficult for most invertebrates, including mussels, because so little attention had been given to the study of their biology, ecology, and systematics. In 1970, only six U.S. states had recent lists or books covering their mussel fauna. The first modern attempt to provide a comprehensive list of freshwater mussels of North America was published by Burch (1973, 1975).

The first comprehensive list of freshwater mussels of the United States and Canada was compiled in Turgeon et al. (1988) and revised a decade later (Turgeon et al. 1998). Williams et al. (1993) was another important resource during this period; although mainly an assessment of species conservation status, this paper also provided a comprehensive and widely used species list similar to those of Turgeon et al. (1988, 1998). These lists standardized and provided taxonomic stability to mussel common and scientific names to an extent that was previously unavailable. However, systematic taxonomy of mussels was poorly known at that time, and classifications at all taxonomic levels were based largely on concepts from the early 1900s.

Since publication of Turgeon et al. (1988, 1998) and Williams et al. (1993), many studies have refined our understanding of mussel systematic taxonomy. Several major publications have addressed systematic relationships within the class Bivalvia, including the order Unionida (Bieler et al. 2010; Carter et al. 2011; Bolotov et al. 2016; Araujo et al. 2017; Combosch et al. 2017). Major studies specific to the Unionida include Graf and Ó Foighil (2000), Hoeh et al. (2001, 2002, 2009), Roe and Hoeh (2003), Campbell et al. (2005), Walker et al. (2006), Graf and Cummings (2007, 2017), Cummings and Graf (2010), and Campbell and Lydeard (2012a, 2012b). In addition, many studies have examined systematic relationships at lower taxonomic levels (e.g., Serb et al. 2003; Jones et al. 2006; Lane et al. 2016). Together, this body of work depicts a view of mussel taxonomy that differs substantially from that of previous lists of the North American fauna.

We present a revised classification and list of the freshwater mussels of the United States and Canada (Tables 1 and 2). The primary purpose of this revision is to provide in a single resource a comprehensive list and taxonomic classification that reflects recent refinement of mussel systematics.

METHODS

We used as a starting point the list of Turgeon et al. (1998). We revised this list and its taxonomic classification based on a review of peer-reviewed mussel taxonomic and nomenclatural literature produced since 1998, unpublished research by the authors, and discussions with other experts on mussel systematics. We also corrected the spelling of specific epithets and publication dates of original descriptions based on the International Code of Zoological Nomenclature (http://www.

iczn.org/iczn/index.jsp). Species mentioned in the text, but not included in Table 2, have author and date of publication following the name. Author and date of publication for all other species are given in Table 2.

Mussel common names follow Turgeon et al. (1998) with minor exceptions, but they are capitalized as is now the practice for many other animal groups (e.g., birds, reptiles, amphibians, fishes). Capitalization of common names helps avoid confusion by identifying standardized common names. For example, reference to a "fragile papershell" could apply to several thin-shelled species, but the capitalized "Fragile Papershell" is unambiguously recognized as the common name for *Leptodea fragilis*. We note and explain other instances where we changed common names from those of Turgeon et al. (1998) or where recognition of previously unrecognized species necessitated creation of a new common name

We provide a rationale for and discussion of all taxonomic changes in the following accounts for each family and genus and in Table 2. There is a degree of uncertainty and subjectivity in our revised list that is unavoidable given our still imperfect understanding of mussel systematics. We attempted to reconcile divergent views regarding mussel systematics based on our assessment of the strength of evidence for these views. In cases where evidence did not allow reconciliation, we attempted to provide a plausible conclusion based on our professional judgment and experience; these conclusions were based on consensus among the authors to the extent possible.

Subspecies is a taxonomic category applied to populations that are morphologically distinct and geographically separated but that exhibit intergradation in contact zones (Mayr et al. 1953; Gilbert 1961). We evaluated morphological and molecular evidence relating to the status of subspecies recognized by Turgeon et al. (1998) and subsequent workers (Jones and Neves 2010). In most cases, recent evidence did not support recognition of subspecies but supported either subsuming subspecies under the nominal species or elevating subspecies to species status; we discuss this evidence for each case. However, strong evidence with which to evaluate their status was lacking for several, mostly extinct, subspecies (see Epioblasma). The designation of subspecies versus species is arbitrary and inconsistent for many animal groups (Huang and Knowles 2016), and this has historically been the case for mussels (e.g., Ortmann 1918, 1920). For subspecies that lacked strong evidence for synonymization or elevation, we recognize all as species to provide more consistent null hypotheses regarding potential diversity in these groups.

This work has been registered with ZooBank and a copy has been archived at Zenodo.org.

RESULTS

Freshwater bivalve higher classification continues to evolve as more data are generated and new techniques are developed. Fossil and modern bivalve higher classification has

Table 1. Higher classification of the Unionoidea present in the United States and Canada.

Table 1, continued.

CLASS Bivalvia Linnaeus, 1758

INFRACLASS Heteroconchia Hertwig, 1895

COHORT Uniomorphi Gray, 1854 [=Paleoheterodonta]

ORDER Unionida Gray, 1854

SUPERFAMILY Unionoidea Rafinesque, 1820

MARGARITIFERIDAE Henderson, 1929

Margaritifera Schumacher, 1816

UNIONIDAE Rafinesque, 1820

ANODONTINAE Rafinesque, 1820

Anodontini Rafinesque, 1820

Alasmidonta Say, 1818

Anodonta Lamarck, 1799

Anodontoides Simpson in Baker, 1898

Arcidens Simpson, 1900

Lasmigona Rafinesque, 1831

Pegias Simpson, 1900

Pyganodon Crosse and Fischer, 1894

Simpsonaias Frierson, 1914

Strophitus Rafinesque, 1820

Utterbackia Baker, 1927

Utterbackiana Frierson, 1927

Cristariini Lopes-Lima, Bogan, and Froufe, 2017

Sinanodonta Modell, 1945

GONIDEINAE Ortmann, 1916

Gonideini Ortmann, 1916

Gonidea Conrad, 1857

AMBLEMINAE Rafinesque, 1820

Amblemini Rafinesque, 1820

Amblema Rafinesque, 1820

Lampsilini Ihering, 1901

Actinonaias Crosse and Fischer, 1894

Cyprogenia Agassiz, 1852

Cyrtonaias Crosse and Fischer, 1894

Dromus Simpson, 1900

Ellipsaria Rafinesque, 1820

Epioblasma Rafinesque, 1831

Glebula Conrad, 1853

Hamiota Roe and Hartfield, 2005

Lampsilis Rafinesque, 1820

Lemiox Rafinesque, 1831

Leptodea Rafinesque, 1820

Ligumia Swainson, 1840

Medionidus Simpson, 1900

Obliquaria Rafinesque, 1820

Obovaria Rafinesque, 1819

Plectomerus Conrad, 1853

Potamilus Rafinesque, 1818

Ptychobranchus Simpson, 1900

Toxolasma Rafinesque, 1831

Truncilla Rafinesque, 1819

Venustaconcha Frierson, 1927

Villosa Frierson, 1927

Pleurobemini Hannibal, 1912

Elliptio Rafinesque, 1819

Elliptoideus Frierson, 1927

Eurynia Rafinesque, 1820

Fusconaia Simpson, 1900

Hemistena Rafinesque, 1820

Parvaspina Perkins, Gangloff, and Johnson, 2017

Plethobasus Simpson, 1900

Pleurobema Rafinesque, 1819

Pleuronaia Frierson, 1927

Quadrulini Ihering, 1901

Cyclonaias Pilsbry in Ortmann and Walker, 1922

Megalonaias Utterback, 1915

Quadrula Rafinesque, 1820

Theliderma Swainson, 1840

Tritogonia Agassiz, 1852

Uniomerus Conrad, 1853

AMBLEMINAE (incertae sedis)

Disconaias Crosse and Fischer, 1894

Popenaias Frierson, 1927

Reginaia Campbell and Lydeard, 2012

recently been summarized by Carter et al. (2011), with standardized endings for higher taxa within Bivalvia. Recent evidence supports the order Unionida as a monophyletic clade (Combosch et al. 2017). There have been two recent assessments of the taxonomy for Margaritiferidae (Bolotov et al. 2016; Araujo et al. 2017). Higher level relationships within the Unionidae have recently been reviewed by Lopes-Lima et al. (2017). Based on these publications, we provide our assessment of higher classification of the Unionida and its position in the class Bivalvia (Table 1).

There is general agreement on the three subfamily divisions within the Unionidae in North America and seven subfamilies worldwide, but there remains some uncertainty regarding classification at lower levels. We adopted a subfamily-, tribe-, and generic-level classification for the United States and Canada based on recent phylogenetic research (Table 1). We recognize the Anodontinae as a subfamily with two tribes in the United States and Canada. We recognize the subfamily Gonideinae, containing the genus Gonidea. We recognize the subfamily Ambleminae as consisting of four tribes: Amblemini, Lampsilini, Pleurobemini, and Quadrulini. The placement of many genera within tribes in the Ambleminae is well supported and consistent among studies, but the placement of others is less certain and varies among studies (e.g., Plectomerus, Campbell et al. 2005). The Mexican and Central American genera Disconaias and Popenaias and North American Reginaia lack sufficient phylogenetic information to be confidently assigned to a classification, and we placed them in Ambleminae incertae sedis (Table 1).

Our revised list includes many taxonomic changes at the

Table 2. List of Margaritiferidae and Unionidae of the United States and Canada. Currently recognized taxa are bolded. Taxa preceded by an asterisk and not bolded appeared in Turgeon et al. (1998) but are no longer recognized or reassigned to other genera.

Scientific Name	Common Name	Changes in Scientific and Common Names
Security Pulity	Common Ivanic	and Common rames
MARGARITIFERIDAE Henderson, 1929		
*Cumberlandia Ortmann, 1912		Synonym of Margaritifera
*Cumberlandia monodonta (Say, 1829)	Spectaclecase	Reassigned to Margaritifera
Margaritifera Schumacher, 1816		
Margaritifera falcata (Gould, 1850)	Western Pearlshell	
Margaritifera hembeli (Conrad, 1838)	Louisiana Pearlshell	
Margaritifera margaritifera (Linnaeus, 1758)	Eastern Pearlshell	
Margaritifera marrianae Johnson, 1983	Alabama Pearlshell	
Margaritifera monodonta (Say, 1829)	Spectaclecase	Reassigned from Cumberlandia
UNIONIDAE Rafinesque, 1820	•	
Actinonaias Crosse and Fischer, 1894		
Actinonaias ligamentina (Lamarck, 1819)	Mucket	
Actinonaias pectorosa (Conrad, 1834)	Pheasantshell	
Alasmidonta Say, 1818		
Alasmidonta arcula (Lea, 1838)	Altamaha Arcmussel	
Alasmidonta atropurpurea (Rafinesque, 1831)	Cumberland Elktoe	
Alasmidonta heterodon (Lea, 1829)	Dwarf Wedgemussel	Publication date corrected
Alasmidonta marginata Say, 1818	Elktoe	Tubication date corrected
Alasmidonta marginata Say, 1010 Alasmidonta marginata Say, 1010	Coosa Elktoe	
Alasmidonta raveneliana (Lea, 1834)	Appalachian Elktoe	
Alasmidonta robusta Clarke, 1981	Carolina Elktoe	
	Southern Elktoe	
Alasmidonta triangulata (Lea, 1858)		
Alasmidonta undulata (Say, 1817)	Triangle Floater Brook Floater	
Alasmidonta varicosa (Lamarck, 1819)		
Alasmidonta viridis (Rafinesque, 1820)	Slippershell Mussel	
Alasmidonta wrightiana (Walker, 1901)	Ochlockonee Arcmussel	
Amblema Rafinesque, 1820		
Amblema elliottii (Lea, 1856)	Coosa Fiveridge	
Amblema neislerii (Lea, 1858)	Fat Threeridge	
Amblema plicata (Say, 1817)	Threeridge	
Anodonta Lamarck, 1799		
*Anodonta beringiana Middendorff, 1851	Yukon Floater	Reassigned to Sinanodonta
Anodonta californiensis Lea, 1852	California Floater	
*Anodonta couperiana Lea, 1840	Barrel Floater	Reassigned to Utterbackiana
*Anodonta dejecta Lewis, 1875	Woebegone Floater	Synonym of Anodonta californiens
*Anodonta heardi Gordon and Hoeh, 1995	Apalachicola Floater	Reassigned to Utterbackiana
*Anodonta implicata Say, 1829	Alewife Floater	Reassigned to Utterbackiana
Anodonta kennerlyi Lea, 1860	Western Floater	
Anodonta nuttalliana Lea, 1838	Winged Floater	
Anodonta oregonensis Lea, 1838	Oregon Floater	
*Anodonta suborbiculata Say, 1831	Flat Floater	Reassigned to Utterbackiana
Anodontoides Simpson in Baker, 1898		
Anodontoides denigrata (Lea, 1852)	Cumberland Papershell	Elevated from synonymy
Anodontoides ferussacianus (Lea, 1834)	Cylindrical Papershell	J J J
Anodontoides radiatus (Conrad, 1834)	Rayed Creekshell	
Arcidens Simpson, 1900	,	
Arcidens confragosus (Say, 1829)	Rock Pocketbook	
Arcidens wheeleri (Ortmann and Walker, 1912)	Ouachita Rock Pocketbook	Reassigned from Arkansia
*Arkansia Ortmann and Walker, 1912	Suucinu Rock I OCKCLOOOK	Synonym of <i>Arcidens</i>
		Symonym Or michically

O to the M	C N	Changes in Scientific
Scientific Name	Common Name	and Common Names
Cyclonaias Pilsbry in Ortmann and Walker, 1922		
Cyclonaias archeri (Frierson, 1905)	Tallapoosa Orb	Elevated from synonymy
Cyclonaias asperata (Lea, 1861)	Alabama Orb	Reassigned from Quadrula
Cyclonaias aurea (Lea, 1859)	Golden Orb	Reassigned from Quadrula
Cyclonaias houstonensis (Lea, 1859)	Smooth Pimpleback	Reassigned from Quadrula
Cyclonaias infucata (Conrad, 1834)	Sculptured Pigtoe	Reassigned from Quincuncina
Cyclonaias kieneriana (Lea, 1852)	Coosa Orb	Elevated from synonymy
Cyclonaias kleiniana (Lea, 1852)	Florida Mapleleaf	Elevated from synonymy
Cyclonaias mortoni (Conrad, 1835)	Western Pimpleback	Species elevated from subspecies; reassigned from <i>Quadrula</i>
Cyclonaias nodulata (Rafinesque, 1820)	Wartyback	Reassigned from Quadrula
Cyclonaias petrina (Gould, 1855)	Texas Pimpleback	Reassigned from Quadrula
Cyclonaias pustulosa (Lea, 1831)	Pimpleback	Reassigned from Quadrula
Cyclonaias refulgens (Lea, 1868)	Purple Pimpleback	Reassigned from Quadrula
Cyclonaias succissa (Lea, 1852)	Purple Pigtoe	Reassigned from <i>Fusconaia</i>
Cyclonaias tuberculata (Rafinesque, 1820)	Purple Wartyback	
Cyprogenia Agassiz, 1852		
Cyprogenia aberti (Conrad, 1850)	Western Fanshell	
Cyprogenia stegaria (Rafinesque, 1820)	Fanshell	
Cyrtonaias Crosse and Fischer, 1894		
Cyrtonaias tampicoensis (Lea, 1838)	Tampico Pearlymussel	
Disconaias Crosse and Fischer, 1894	Tumpres Tearly musser	
Disconaias fimbriata (Frierson, 1907)	Fringed Mucket	Elevated from synonymy
*Disconaias salinasensis (Simpson, 1908)	Salina Mucket	Synonym of <i>Disconaias fimbriata</i>
Dromus Simpson, 1900		Syllenyin of 2 iscollaria yillio lara
Dromus dromas (Lea, 1834)	Dromedary Pearlymussel	
Ellipsaria Rafinesque, 1820	Diomedaly Tearlymasser	
Ellipsaria lineolata (Rafinesque, 1820)	Butterfly	
Elliptio Rafinesque, 1819	Buttering	
Elliptio ahenea (Lea, 1843)	Southern Lance	
Elliptio angustata (Lea, 1831)	Carolina Lance	
Elliptio arca (Conrad, 1834)	Alabama Spike	
Elliptio arctata (Conrad, 1834)	Delicate Spike	
*Elliptio buckleyi (Lea, 1843)	Florida Shiny Spike	Synonym of Elliptio jayensis
Elliptio chipolaensis (Walker, 1905)	Chipola Slabshell	Syllonym of Empho Juyensis
Elliptio cistellaeformis (Lea, 1863)	Box Spike	
Elliptio complanata (Lightfoot, 1786)	Eastern Elliptio	
Elliptio companaea (Lea, 1831)	Carolina Slabshell	
Elliptio crassidens (Lamarck, 1819)	Elephantear	
Elliptio dariensis (Lea, 1842)	Georgia Elephantear	
*Elliptio dilatata (Rafinesque, 1820)	Spike	Reassigned to Eurynia
Elliptio downiei (Lea, 1858)	Satilla Elephantear	Reassigned to Euryma
*Elliptio errans (Lea, 1856)	Oval Elliptio	Synonym of <i>Elliptio icterina</i> ; publication date corrected
Elliptio fisheriana (Lea, 1838)	Northern Lance	
Elliptio folliculata (Lea, 1838)	Pod Lance	
Elliptio fraterna (Lea, 1852)	Brother Spike	
Elliptio fumata (Lea, 1857)	Gulf Slabshell	Elevated from synonymy
*Elliptio hepatica (Lea, 1859)	Brown Elliptio	Synonym of <i>Elliptio icterina</i>
Elliptio hopetonensis (Lea, 1838)	Altamaha Slabshell	Synonym of Luipuo www.mu
Elliptio icterina (Conrad, 1834)	Variable Spike	

Table 2, continued.

cientific Name	Common Name	Changes in Scientific and Common Names
Elliptio jayensis (Lea, 1838)	Florida Spike	Common name changed from Flat Spike
*Elliptio judithae Clarke, 1986	Plicate Spike	Synonym of Elliptio roanokensis
Elliptio lanceolata (Lea, 1828)	Yellow Lance	
*Elliptio lugubris (Lea, 1834)	Sad Elliptio	Synonym of Elliptio icterina
Elliptio marsupiobesa Fuller, 1972	Cape Fear Spike	
Elliptio mcmichaeli Clench and Turner, 1956	Fluted Elephantear	
Elliptio monroensis (Lea, 1843)	St. Johns Elephantear	
Elliptio nigella (Lea, 1852)	Winged Spike	
Elliptio occulta (Lea, 1843)	Hidden Spike	Elevated from synonymy
Elliptio producta (Conrad, 1836)	Atlantic Spike	
Elliptio pullata (Lea, 1856)	Gulf Spike	Elevated from synonymy
Elliptio purpurella (Lea, 1857)	Inflated Spike	Elevated from synonymy
*Elliptio raveneli (Conrad, 1834)	Carolina Spike	Synonym of <i>Elliptio icterina</i>
Elliptio roanokensis (Lea, 1838)	Roanoke Slabshell	3 J
Elliptio shepardiana (Lea, 1834)	Altamaha Lance	
Elliptio spinosa (Lea, 1836)	Altamaha Spinymussel	
*Elliptio steinstansana Johnson and Clarke, 1983	Tar River Spinymussel	Reassigned to Parvaspina
*Elliptio waccamawensis (Lea, 1863)	Waccamaw Spike	Synonym of <i>Elliptio congaraea</i>
*Elliptio waltoni (Wright, 1888)	Florida Lance	Synonym of Elliptio ahenea
Elliptoideus Frierson, 1927	Tiorida Edilee	Synonym of Empho unched
Elliptoideus sloatianus (Lea, 1840)	Purple Bankclimber	
Epioblasma Rafinesque, 1831	Turple Bankeninoer	
Epioblasma ahlstedti Jones and Neves, 2010	Duck River Dartersnapper	Described as new species
Epioblasma arcaeformis (Lea, 1831)	Sugarspoon	Described as new species
Epioblasma aureola Jones and Neves, 2010	Golden Riffleshell	Species elevated from subspecies
Epioblasma biemarginata (Lea, 1857)	Angled Riffleshell	species elevated from subspecies
Epioblasma brevidens (Lea, 1831)	Cumberlandian Combshell	
-		
Epioblasma capsaeformis (Lea, 1834)	Oyster Mussel	El4-1 f
Epioblasma cincinnatiensis (Lea, 1840)	Ohio Riffleshell	Elevated from synonymy
Epioblasma curtisii (Frierson and Utterback, 1916)	Curtis Pearlymussel	Species elevated from subspecies
Epioblasma flexuosa (Rafinesque, 1820)	Leafshell	
Epioblasma florentina (Lea, 1857)	Yellow Blossom	
*Epioblasma florentina aureola Jones and Neves, 2010	Golden Riffleshell	Described as new subspecies; elevated t species
*Epioblasma florentina curtisii (Frierson and Utterback, 1916)	Curtis Pearlymussel	Subspecies elevated to species
*Epioblasma florentina florentina (Lea, 1857)	Yellow Blossom	Nominotypical subspecies not required
*Epioblasma florentina walkeri (Wilson and Clark, 1914)	Tan Riffleshell	Subspecies elevated to species
Epioblasma gubernaculum (Reeve, 1865)	Green Blossom	Species elevated from subspecies
Epioblasma haysiana (Lea, 1834)	Acornshell	
Epioblasma lenior (Lea, 1842)	Narrow Catspaw	
Epioblasma lewisii (Walker, 1910)	Forkshell	
Epioblasma metastriata (Conrad, 1838)	Upland Combshell	
Epioblasma obliquata (Rafinesque, 1820)	Catspaw	
*Epioblasma obliquata obliquata (Rafinesque, 1820)	Catspaw	Nominotypical subspecies not required
*Epioblasma obliquata perobliqua (Conrad, 1836)	White Catspaw	Subspecies elevated to species
Epioblasma othcaloogensis (Lea, 1857)	Southern Acornshell	•
Epioblasma penita (Conrad, 1834)	Southern Combshell	
Epioblasma perobliqua (Conrad, 1836)	White Catspaw	Species elevated from subspecies
Epioblasma personata (Say, 1829)	Round Combshell	
Epioblasma propinqua (Lea, 1857)	Tennessee Riffleshell	
Epioblasma rangiana (Lea, 1838)	Northern Riffleshell	Species elevated from subspecies

cientific Name	Common Name	Changes in Scientific and Common Names
Clentine Ivanie	Common Ivanie	and Common Names
Epioblasma sampsonii (Lea, 1861)	Wabash Riffleshell	
Epioblasma stewardsonii (Lea, 1852)	Cumberland Leafshell	
Epioblasma torulosa (Rafinesque, 1820)	Tubercled Blossom	
*Epioblasma torulosa gubernaculum (Reeve, 1865)	Green Blossom	Subspecies elevated to species
*Epioblasma torulosa rangiana (Lea, 1838)	Northern Riffleshell	Subspecies elevated to species
*Epioblasma torulosa torulosa (Rafinesque, 1820)	Tubercled Blossom	Nominotypical subspecies not required
Epioblasma triquetra (Rafinesque, 1820)	Snuffbox	
Epioblasma turgidula (Lea, 1858)	Turgid Blossom	
Epioblasma walkeri (Wilson and Clark, 1914)	Tan Riffleshell	Species elevated from subspecies
Eurynia Rafinesque, 1820		Elevated from synonymy
Eurynia dilatata Rafinesque, 1820	Spike	Reassigned from Elliptio
Fusconaia Simpson, 1900		
*Fusconaia askewi (Marsh, 1896)	Texas Pigtoe	Synonym of Fusconaia chunii
*Fusconaia barnesiana (Lea, 1838)	Tennessee Pigtoe	Reassigned to Pleuronaia
Fusconaia burkei (Walker, 1922)	Tapered Pigtoe	Reassigned from Quincuncina
Fusconaia cerina (Conrad, 1838)	Gulf Pigtoe	Common name changed from Southern Pig
Fusconaia chunii (Lea, 1861)	Texas Pigtoe	Elevated from synonymy
Fusconaia cor (Conrad, 1834)	Shiny Pigtoe	
Fusconaia cuneolus (Lea, 1840)	Finerayed Pigtoe	
*Fusconaia ebena (Lea, 1831)	Ebonyshell	Reassigned to Reginaia
Fusconaia escambia Clench and Turner, 1956	Narrow Pigtoe	
Fusconaia flava (Rafinesque, 1820)	Wabash Pigtoe	
*Fusconaia lananensis (Frierson, 1901)	Triangle Pigtoe	Synonym of Fusconaia chunii
Fusconaia masoni (Conrad, 1834)	Atlantic Pigtoe	
Fusconaia mitchelli (Simpson, 1895)	False Spike	Reassigned from Quincuncina
Fusconaia ozarkensis (Call, 1887)	Ozark Pigtoe	
Fusconaia subrotunda (Lea, 1831)	Longsolid	
*Fusconaia succissa (Lea, 1852)	Purple Pigtoe	Reassigned to Cyclonaias
Glebula Conrad, 1853		
Glebula rotundata (Lamarck, 1819)	Round Pearlshell	
Gonidea Conrad, 1857		
Gonidea angulata (Lea, 1838)	Western Ridged Mussel	
Hamiota Roe and Hartfield, 2005	C	Described as new genus
Hamiota altilis (Conrad, 1834)	Finelined Pocketbook	Reassigned from Lampsilis
Hamiota australis (Simpson, 1900)	Southern Sandshell	Reassigned from Lampsilis
Hamiota perovalis (Conrad, 1834)	Orangenacre Mucket	Reassigned from <i>Lampsilis</i>
Hamiota subangulata (Lea, 1840)	Shinyrayed Pocketbook	Reassigned from Lampsilis
Hemistena Rafinesque, 1820		•
Hemistena lata (Rafinesque, 1820)	Cracking Pearlymussel	
Lampsilis Rafinesque, 1820	Ç ,	
Lampsilis abrupta (Say, 1831)	Pink Mucket	
*Lampsilis altilis (Conrad, 1834)	Finelined Pocketbook	Reassigned to Hamiota
*Lampsilis australis Simpson, 1900	Southern Sandshell	Reassigned to <i>Hamiota</i>
Lampsilis binominata Simpson, 1900	Lined Pocketbook	C
Lampsilis bracteata (Gould, 1855)	Texas Fatmucket	
Lampsilis brittsi Simpson, 1900	Northern Brokenray	Species elevated from subspecies
Lampsilis cardium Rafinesque, 1820	Plain Pocketbook	
Lampsilis cariosa (Say,1817)	Yellow Lampmussel	
Lampsilis dolabraeformis (Lea, 1838)	Altamaha Pocketbook	
Lampsilis fasciola Rafinesque, 1820	Wavyrayed Lampmussel	

Table 2, continued.

cientific Name	Common Name	Changes in Scientific and Common Names
Lampsilis floridensis (Lea, 1852)	Florida Sandshell	Elevated from synonymy
*Lampsilis fullerkati Johnson, 1984	Waccamaw Fatmucket	Synonym of Lampsilis radiata
*Lampsilis haddletoni Athearn, 1964	Haddleton Lampmussel	Reassigned to <i>Obovaria</i>
Lampsilis higginsii (Lea, 1857)	Higgins Eye	
Lampsilis hydiana (Lea, 1838)	Louisiana Fatmucket	
Lampsilis ornata (Conrad, 1835)	Southern Pocketbook	
Lampsilis ovata (Say, 1817)	Pocketbook	
*Lampsilis perovalis (Conrad, 1834)	Orangenacre Mucket	Reassigned to Hamiota
Lampsilis powellii (Lea, 1852)	Arkansas Fatmucket	
Lampsilis radiata (Gmelin, 1791)	Eastern Lampmussel	
*Lampsilis radiata conspicua (Lea, 1872)	Carolina Fatmucket	Subspecies no longer recognized
*Lampsilis radiata radiata (Gmelin, 1791)	Eastern Lampmussel	Nominotypical subspecies not require
Lampsilis rafinesqueana Frierson, 1927	Neosho Mucket	1
Lampsilis reeveiana (Lea, 1852)	Arkansas Brokenray	
*Lampsilis reeveiana brevicula (Call, 1887)	Ozark Brokenray	Subspecies no longer recognized
*Lampsilis reeveiana brittsi Simpson, 1900	Northern Brokenray	Subspecies elevated to species
*Lampsilis reeveiana reeviana (Lea, 1852)	Arkansas Brokenray	Nominotypical subspecies not require
Lampsilis satura (Lea, 1852)	Sandbank Pocketbook	
Lampsilis siliquoidea (Barnes, 1823)	Fatmucket	
Lampsilis splendida (Lea, 1838)	Rayed Pink Fatmucket	
Lampsilis straminea (Conrad, 1834)	Rough Fatmucket	
*Lampsilis straminea claibornensis (Lea, 1838)	Southern Fatmucket	Subspecies no longer recognized
*Lampsilis straminea straminea (Conrad, 1834)	Rough Fatmucket	Nominotypical subspecies not require
Lampsilis streckeri Frierson, 1927	Speckled Pocketbook	71 1
*Lampsilis subangulata (Lea, 1840)	Shinyrayed Pocketbook	Reassigned to Hamiota
Lampsilis teres (Rafinesque, 1820)	Yellow Sandshell	
Lampsilis virescens (Lea, 1858)	Alabama Lampmussel	
Lasmigona Rafinesque, 1831	•	
Lasmigona alabamensis Clarke, 1985	Alabama Heelsplitter	Species elevated from subspecies
Lasmigona complanata (Barnes, 1823)	White Heelsplitter	
*Lasmigona complanata alabamensis Clarke, 1985	Alabama Heelsplitter	Subspecies elevated to species
*Lasmigona complanata complanata (Barnes, 1823)	White Heelsplitter	Nominotypical subspecies not require
Lasmigona compressa (Lea, 1829)	Creek Heelsplitter	
Lasmigona costata (Rafinesque, 1820)	Flutedshell	
Lasmigona decorata (Lea, 1852)	Carolina Heelsplitter	
Lasmigona etowaensis (Conrad, 1849)	Etowah Heelsplitter	Elevated from synonymy
Lasmigona holstonia (Lea, 1838)	Tennessee Heelsplitter	J J J
Lasmigona subviridis (Conrad, 1835)	Green Floater	
Lemiox Rafinesque, 1831		
Lemiox rimosus (Rafinesque, 1831)	Birdwing Pearlymussel	
Leptodea Rafinesque, 1820	g ,	
Leptodea fragilis (Rafinesque, 1820)	Fragile Papershell	
Leptodea leptodon (Rafinesque, 1820)	Scaleshell	
Leptodea ochracea (Say, 1817)	Tidewater Mucket	
*Lexingtonia Ortmann, 1914		Synonym of Fusconaia
*Lexingtonia dolabelloides (Lea, 1840)	Slabside Pearlymussel	Reassigned to <i>Pleuronaia</i>
*Lexingtonia subplana (Conrad, 1837)	Virginia Pigtoe	Synonym of Fusconaia masoni
Ligumia Swainson, 1840	<i>C G</i>	, ,
Ligumia nasuta (Say, 1817)	Eastern Pondmussel	
Ligumia recta (Lamarck, 1819)	Black Sandshell	
Ligumia subrostrata (Say, 1831)	Pondmussel	

Crismaticis Niema	Camara Nama	Changes in Scientific
cientific Name	Common Name	and Common Names
Medionidus Simpson, 1900		
Medionidus acutissimus (Lea, 1831)	Alabama Moccasinshell	
Medionidus conradicus (Lea, 1834)	Cumberland Moccasinshell	
*Medionidus mcglameriae van der Schalie, 1939	Tombigbee Moccasinshell	Synonym of Leptodea fragilis
Medionidus parvulus (Lea, 1860)	Coosa Moccasinshell	
Medionidus penicillatus (Lea, 1857)	Gulf Moccasinshell	
Medionidus simpsonianus Walker, 1905	Ochlockonee Moccasinshell	
Medionidus walkeri (Wright, 1897)	Suwannee Moccasinshell	
Megalonaias Utterback, 1915		
Megalonaias nervosa (Rafinesque, 1820)	Washboard	
Obliquaria Rafinesque, 1820		
Obliquaria reflexa Rafinesque, 1820	Threehorn Wartyback	
Obovaria Rafinesque, 1819	·	
Obovaria arkansasensis (Lea, 1862)	Southern Hickorynut	Reassigned from Villosa
Obovaria choctawensis (Athearn, 1964)	Choctaw Bean	Reassigned from Villosa
Obovaria haddletoni (Athearn, 1964)	Haddleton Lampmussel	Reassigned from Lampsilis
*Obovaria jacksoniana (Frierson, 1912)	Southern Hickorynut	Synonym of <i>Obovaria arkansasen</i>
Obovaria olivaria (Rafinesque, 1820)	Hickorynut	, ,
Obovaria retusa (Lamarck, 1819)	Ring Pink	
*Obovaria rotulata (Wright, 1899)	Round Ebonyshell	Reassigned to Reginaia
Obovaria subrotunda (Rafinesque, 1820)	Round Hickorynut	reassigned to reginate
Obovaria unicolor (Lea, 1845)	Alabama Hickorynut	
Parvaspina Perkins, Gangloff, and Johnson, 2017	Thabana Thekerynat	Described as new genus
Parvaspina collina (Conrad, 1836)	James Spinymussel	Reassigned from <i>Pleurobema</i> ;
Turraspina comia (Contaa, 1050)	James Spinymasser	publication date corrected
Parvaspina steinstansana (Johnson and Clarke, 1983)	Tar River Spinymussel	Reassigned from <i>Elliptio</i>
Pegias Simpson, 1900	Tar River Spinymusser	Reassigned from Empiro
Pegias fabula (Lea, 1838)	Littlewing Pearlymussel	
Plectomerus Conrad, 1853	Littlewing Tearlymusser	
Plectomerus dombeyanus (Valenciennes, 1827)	Bankclimber	
Plethobasus Simpson, 1900	Bankenmoer	
Plethobasus cicatricosus (Say, 1829)	White Wartyback	
Plethobasus cooperianus (Lea, 1834)	Orangefoot Pimpleback	
Plethobasus cyphyus (Rafinesque, 1820)	Sheepnose	
Pleurobema Rafinesque, 1819	III ala ant	Caraidanal a manna dubina
*Pleurobema altum (Conrad, 1854)	Highnut	Considered a nomen dubium
Pleurobema athearni Gangloff, Williams, and	Canoe Creek Clubshell	Described as new species
Feminella, 2006	H IB:	
*Pleurobema avellanum Simpson, 1900	Hazel Pigtoe	Synonym of Pleurobema rubellum
Pleurobema beadleianum (Lea, 1861)	Mississippi Pigtoe	
*Pleurobema bournianum (Lea, 1840)	Scioto Pigtoe	Synonym of Pleurobema clava
*Pleurobema chattanoogaense (Lea, 1858)	Painted Clubshell	Synonym of Pleurobema decisum
Pleurobema clava (Lamarck, 1819)	Clubshell	
*Pleurobema collina (Conrad, 1836)	James Spinymussel	Reassigned to Parvaspina
Pleurobema cordatum (Rafinesque, 1820)	Ohio Pigtoe	
Pleurobema curtum (Lea, 1859)	Black Clubshell	
Pleurobema decisum (Lea, 1831)	Southern Clubshell	
Pleurobema fibuloides (Lea, 1859)	Kusha Pigtoe	Elevated from synonymy
*Pleurobema flavidulum (Lea, 1861)	Yellow Pigtoe	Synonym of Pleurobema perovatu
*Pleurobema furvum (Conrad, 1834)	Dark Pigtoe	Synonym of Pleurobema rubellum
Pleurobema georgianum (Lea, 1841)	Southern Pigtoe	

Table 2, continued.

cientific Name	Common Name	Changes in Scientific and Common Names
*Pleurobema gibberum (Lea, 1838)	Cumberland Pigtoe	Reassigned to Pleuronaia
*Pleurobema hagleri (Frierson, 1900)	Brown Pigtoe	Synonym of Pleurobema rubellum
Pleurobema hanleyianum (Lea, 1852)	Georgia Pigtoe	, ,
Pleurobema hartmanianum (Lea, 1860)	Cherokee Pigtoe	Elevated from synonymy
*Pleurobema johannis (Lea, 1859)	Alabama Pigtoe	Synonym of Pleurobema perovatum
Pleurobema marshalli Frierson, 1927	Flat Pigtoe	
*Pleurobema murrayense (Lea, 1868)	Coosa Pigtoe	Synonym of Pleurobema stabile
*Pleurobema nucleopsis (Conrad, 1849)	Longnut	Synonym of Pleurobema georgianum
Pleurobema oviforme (Conrad, 1834)	Tennessee Clubshell	
Pleurobema perovatum (Conrad, 1834)	Ovate Clubshell	
Pleurobema plenum (Lea, 1840)	Rough Pigtoe	
Pleurobema pyriforme (Lea, 1857)	Oval Pigtoe	
Pleurobema riddellii (Lea, 1861)	Louisiana Pigtoe	
Pleurobema rubellum (Conrad, 1834)	Warrior Pigtoe	
Pleurobema rubrum (Rafinesque, 1820)	Pyramid Pigtoe	
Pleurobema sintoxia (Rafinesque, 1820)	Round Pigtoe	
Pleurobema stabile (Lea, 1861)	Coosa Pigtoe	Elevated from synonymy
Pleurobema strodeanum (Wright, 1898)	Fuzzy Pigtoe	Elevated from synonymy
Pleurobema taitianum (Lea, 1834)	Heavy Pigtoe	
*Pleurobema troschelianum (Lea, 1852)	Alabama Clubshell	Synonym of Pleurobema georgianum
Pleurobema verum (Lea, 1861)	True Pigtoe	Synonym of Tieurobema georgianum
Pleuronaia Frierson, 1927	True Figure	Elevated from synonymy
	Tampagga Diatas	Reassigned from <i>Fusconaia</i>
Pleuronaia barnesiana (Lea, 1838) Pleuronaia dolabelloides (Lea, 1840)	Tennessee Pigtoe Slabside Pearlymussel	Reassigned from <i>Lexingtonia</i>
Pleuronaia gibber (Lea, 1838)	Cumberland Pigtoe	Reassigned from <i>Pleurobema</i> ; spelling correction of species name
Popenais Frierson, 1927		•
Popenais popeii (Lea, 1857)	Texas Hornshell	
Potamilus Rafinesque, 1818		
Potamilus alatus (Say, 1817)	Pink Heelsplitter	
Potamilus amphichaenus (Frierson, 1898)	Texas Heelsplitter	
Potamilus capax (Green, 1832)	Fat Pocketbook	
Potamilus inflatus (Lea, 1831)	Inflated Heelsplitter	Common name changed from Alabam Heelsplitter
Potamilus metnecktayi Johnson, 1998	Salina Mucket	Described as new species
Potamilus ohiensis (Rafinesque, 1820)	Pink Papershell	
Potamilus purpuratus (Lamarck, 1819)	Bleufer	
Ptychobranchus Simpson, 1900		
Ptychobranchus fasciolaris (Rafinesque, 1820)	Kidneyshell	
Ptychobranchus foremanianus (Lea, 1842)	Rayed Kidneyshell	Elevated from synonymy
Ptychobranchus greenii (Conrad, 1834)	Triangular Kidneyshell	J J J
Ptychobranchus jonesi (van der Schalie, 1934)	Southern Kidneyshell	
Ptychobranchus occidentalis (Conrad, 1836)	Ouachita Kidneyshell	
*Ptychobranchus subtentum (Say, 1825)	Fluted Kidneyshell	Incorrect spelling of species name
Ptychobranchus subtentus (Say, 1825)	Fluted Kidneyshell	Spelling correction of species name
Pyganodon Crosse and Fischer, 1894	Times Titolies silen	Spenning contention of species manie
Pyganodon cataracta (Say, 1817)	Eastern Floater	
Pyganodon fragilis (Lamarck, 1819)	Newfoundland Floater	
Pyganodon gibbosa (Say, 1824)	Inflated Floater	
Pyganodon grandis (Say, 1824)	Giant Floater	
		Publication date corrected
Pyganodon lacustris (Lea, 1857)	Lake Floater	Publication date corrected

ciontifia Nama	Common Nomo	Changes in Scientific and Common Names
cientific Name	Common Name	and Common Names
Quadrula Rafinesque, 1820		
Quadrula apiculata (Say, 1829)	Southern Mapleleaf	
*Quadrula asperata (Lea, 1861)	Alabama Orb	Reassigned to Cyclonaias
*Quadrula aurea (Lea, 1859)	Golden Orb	Reassigned to Cyclonaias
Quadrula couchiana (Lea, 1860)	Rio Grande Monkeyface	
*Quadrula cylindrica cylindrica (Say, 1817)	Rabbitsfoot	Nominotypical subspecies not required; reassigned to <i>Theliderma</i>
*Quadrula cylindrica strigillata (Wright, 1898)	Rough Rabbitsfoot	Subspecies no longer recognized
Quadrula fragosa (Conrad, 1835)	Winged Mapleleaf	
*Quadrula houstonensis (Lea, 1859)	Smooth Pimpleback	Reassigned to Cyclonaias
*Quadrula intermedia (Conrad, 1836)	Cumberland Monkeyface	Reassigned to Theliderma
*Quadrula kieneriana (Lea, 1852)	Coosa Orb	Reassigned to Cyclonaias
*Quadrula metanevra (Rafinesque, 1820)	Monkeyface	Reassigned to Theliderma
Quadrula nobilis (Conrad, 1854)	Gulf Mapleleaf	Elevated from synonymy
*Quadrula nodulata (Rafinesque, 1820)	Wartyback	Reassigned to Cyclonaias
*Quadrula petrina (Gould, 1855)	Texas Pimpleback	Reassigned to Cyclonaias
*Quadrula pustulosa mortoni (Conrad, 1835)	Western Pimpleback	Subspecies elevated to species; reassigned to <i>Cyclonaias</i>
*Quadrula pustulosa pustulosa (Lea, 1831)	Pimpleback	Nominotypical subspecies not required; reassigned to <i>Cyclonaias</i>
Quadrula quadrula (Rafinesque, 1820)	Mapleleaf	
*Quadrula refulgens (Lea, 1868)	Purple Pimpleback	Reassigned to Cyclonaias
Quadrula rumphiana (Lea, 1852)	Ridged Mapleleaf	,
*Quadrula sparsa (Lea, 1841)	Appalachian Monkeyface	Reassigned to Theliderma
*Quadrula stapes (Lea, 1831)	Stirrupshell	Reassigned to <i>Theliderma</i>
*Quadrula tuberosa (Lea, 1840)	Rough Rockshell	Synonym of Theliderma metanevra
*Quincuncina Ortmann, 1922		Synonym of Fusconaia
*Quincuncina burkei Walker, 1922	Tapered Pigtoe	Reassigned to Fusconaia
*Quincuncina infucata (Conrad, 1834)	Sculptured Pigtoe	Reassigned to Cyclonaias
*Quincuncina mitchelli (Simpson, 1895)	False Spike	Reassigned to Fusconaia
Reginaia Campbell and Lydeard, 2012		Described as new genus
Reginaia apalachicola (Williams and Fradkin, 1999)	Apalachicola Ebonyshell	Described as new species; reassigned from Fusconaia
Reginaia ebenus (Lea, 1831)	Ebonyshell	Reassigned from <i>Fusconaia</i> ; spelling correction of species name
Reginaia rotulata (Wright, 1899)	Round Ebonyshell	Reassigned from <i>Obovaria</i>
Simpsonaias Frierson, 1914	,	
Simpsonaias ambigua (Say, 1825)	Salamander Mussel	
Sinanodonta Modell, 1945		Not previously reported from North Americ
Sinanodonta beringiana (Middendorff, 1851)	Yukon Floater	Reassigned from <i>Anodonta</i>
Sinanodonta woodiana (Lea, 1834)	Chinese Pondmussel	Introduced and established in New Jersey
Strophitus Rafinesque, 1820		
Strophitus connasaugaensis (Lea, 1858)	Alabama Creekmussel	
Strophitus subvexus (Conrad, 1834)	Southern Creekmussel	
Strophitus undulatus (Say, 1817)	Creeper	
Theliderma Swainson, 1840	Стеерег	Elevated from synonymy
Theliderma cylindrica (Say, 1817)	Rabbitsfoot	Reassigned from <i>Quadrula</i>
Theliderma intermedia (Conrad, 1836)	Cumberland Monkeyface	Reassigned from <i>Quadrula</i>
Theliderma metanevra (Rafinesque, 1820)	Monkeyface	Reassigned from <i>Quadrula</i>
Theliderma sparsa (Lea, 1841)	Appalachian Monkeyface	Reassigned from <i>Quadrula</i>
Theliderma stapes (Lea, 1831)	Stirrupshell	Reassigned from <i>Quadrula</i>

Table 2, continued.

Scientific Name	Common Name	Changes in Scientific and Common Names
Scientific Ivanie	Common Name	and Common Names
Toxolasma Rafinesque, 1831		
Toxolasma corvunculus (Lea, 1868)	Southern Purple Lilliput	
Toxolasma cylindrellus (Lea, 1868)	Pale Lilliput	
Toxolasma lividum Rafinesque, 1831	Purple Lilliput	Spelling correction of species name; parentheses unnecessary
*Toxolasma lividus (Rafinesque, 1831)	Purple Lilliput	Incorrect spelling of species name
*Toxolasma mearnsi (Simpson, 1900)	Western Lilliput	Synonym of Toxolasma texasiense
Toxolasma parvum (Barnes, 1823)	Lilliput	Spelling correction of species name
*Toxolasma parvus (Barnes, 1823)	Lilliput	Incorrect spelling of species name
Toxolasma paulum (Lea, 1840)	Iridescent Lilliput	Spelling correction of species name
*Toxolasma paulus (Lea, 1840)	Iridescent Lilliput	Incorrect spelling of species name
Toxolasma pullus (Conrad, 1838)	Savannah Lilliput	
Toxolasma texasiense (Lea, 1857)	Texas Lilliput	Spelling correction of species name
*Toxolasma texasiensis (Lea, 1857)	Texas Lilliput	Incorrect spelling of species name
Tritogonia Agassiz, 1852	1	1 5 1
Tritogonia verrucosa (Rafinesque, 1820)	Pistolgrip	
Truncilla Rafinesque, 1819	8 T	
Truncilla cognata (Lea, 1860)	Mexican Fawnsfoot	
Truncilla donaciformis (Lea, 1828)	Fawnsfoot	
Truncilla macrodon (Lea, 1859)	Texas Fawnsfoot	
Truncilla truncata Rafinesque, 1820	Deertoe	
Uniomerus Conrad, 1853	Decitoe	
Uniomerus carolinianus (Bosc, 1801)	Eastern Pondhorn	Common name changed from Florida Pondhorn
Uniomerus columbensis (Lea, 1857)	Apalachicola Pondhorn	Elevated from synonymy
Uniomerus declivis (Say, 1831)	Tapered Pondhorn	Elevated from synonymy
Uniomerus tetralasmus (Say, 1831)	Pondhorn	
	FOIIGHOTH	
Utterbackia Baker, 1927	D D d-h-11	
Utterbackia imbecillis (Say, 1829)	Paper Pondshell	
Utterbackia peggyae (Johnson, 1965)	Florida Floater	
Utterbackia peninsularis Bogan and Hoeh, 1995	Peninsular Floater	TI 1.6
Utterbackiana Frierson, 1927		Elevated from synonymy
Utterbackiana couperiana (Lea, 1840)	Barrel Floater	Reassigned from Anodonta
Utterbackiana hartfieldorum (Williams, Bogan, and Garner, 2009)	Cypress Floater	Described as new species; reassigned from Anodom
Utterbackiana heardi (Gordon and Hoeh, 1995)	Apalachicola Floater	Reassigned from Anodonta
Utterbackiana implicata (Say, 1829)	Alewife Floater	Reassigned from Anodonta
Utterbackiana suborbiculata (Say, 1831)	Flat Floater	Reassigned from Anodonta
Venustaconcha Frierson, 1927		
Venustaconcha ellipsiformis (Conrad, 1836)	Ellipse	
Venustaconcha pleasii (Marsh, 1891)	Bleedingtooth Mussel	
Venustaconcha trabalis (Conrad, 1834)	Tennessee Bean	Reassigned from <i>Villosa</i> ; common name changed from Cumberland Bean
Venustaconcha troostensis (Lea, 1834)	Cumberland Bean	Elevated from synonymy
Villosa Frierson, 1927		
*Villosa amygdala (Lea, 1843)	Florida Rainbow	Incorrect spelling of species name
Villosa amygdalum (Lea, 1843)	Florida Rainbow	Spelling correction of species name
*Villosa arkansasensis (Lea, 1862)	Ouachita Creekshell	Reassigned to <i>Obovaria</i>
*Villosa choctawensis Athearn, 1964	Choctaw Bean	Reassigned to Obovaria
Villosa constricta (Conrad, 1838)	Notched Rainbow	6
Villosa delumbis (Conrad, 1834)	Eastern Creekshell	
Villosa fabalis (Lea, 1831)	Rayed Bean	

Table 2, continued.

Scientific Name	Common Name	Changes in Scientific and Common Names
Villosa iris (Lea, 1829)	Rainbow	
Villosa lienosa (Conrad, 1834)	Little Spectaclecase	
Villosa nebulosa (Conrad, 1834)	Alabama Rainbow	
Villosa ortmanni (Walker, 1925)	Kentucky Creekshell	
*Villosa perpurpurea (Lea, 1861)	Purple Bean	Synonym of Venustaconcha trabalis
Villosa sima (Lea, 1838)	Caney Fork Rainbow	Elevated from synonymy
Villosa taeniata (Conrad, 1834)	Painted Creekshell	
*Villosa trabalis (Conrad, 1834)	Cumberland Bean	Reassigned to Venustaconcha
Villosa umbrans (Lea, 1857)	Coosa Creekshell	Species elevated from subspecies
*Villosa vanuxemensis umbrans (Lea, 1857)	Coosa Creekshell	Subspecies elevated to species
Villosa vanuxemensis (Lea, 1838)	Mountain Creekshell	
*Villosa vanuxemensis vanuxemensis (Lea, 1838)	Mountain Creekshell	Nominotypical subspecies not required
Villosa vaughaniana (Lea, 1838)	Carolina Creekshell	
Villosa vibex (Conrad, 1834)	Southern Rainbow	
Villosa villosa (Wright, 1898)	Downy Rainbow	

genus, species, and subspecies levels relative to previous lists. We recognize in total 298 freshwater mussel species from the United States and Canada. These comprise the families Margaritiferidae with one genus and five species and Unionidae with 54 genera and 293 species (Table 2). Turgeon et al. (1998) recognized in total 304 taxa: Margaritiferidae with two genera and five species and Unionidae with 49 genera, 286 species, and 13 subspecies. We summarize our changes to Turgeon et al. (1998) as follows. We recognize eight additional genera, including three recently described (Hamiota, Parvaspina, and Reginaia), four elevated from synonymy (Eurynia, Pleuronaia, Theliderma, and Utterbackiana), and one newly reported from North America (Sinanodonta). We place in synonymy four genera, including one in the Margaritiferidae (Cumberlandia) and three in the Unionidae (Arkansia, Lexingtonia, and Quincuncina). We recognize 25 additional species (all Unionidae), including four newly described species and 21 species elevated from synonymy. We place in synonymy 29 species and consider *Pleurobema* altum a nomen dubium, and we reassigned 41 species to other genera. We corrected the specific epithet spelling for eight species, corrected the date of publication for four, and changed the common names of five. Last, we recognized no subspecies, elevating 10 subspecies to species status and subsuming four subspecies into their nominal species (see Methods).

Margaritiferidae Henderson, 1929

Turgeon et al. (1998) recognized two genera in Margaritiferidae, *Cumberlandia* (one species) and *Margaritifera* (four species). On the basis of shell morphology and soft anatomy, Smith (2001) placed *Cumberlandia* in *Margaritanopsis* and *Margaritifera* (in part) in *Pseudunio*, but this classification was not widely accepted. In a molecular phylogenetic analysis, Huff et al. (2004) considered *Cumberlandia* a junior synonym

of *Margaritifera*, and this classification was followed by some subsequent authors (e.g., Graf and Cummings 2007, 2017; Cummings and Graf 2010), but others continued to recognize the genus as valid (e.g., Williams et al. 2008; Watters et al. 2009; Haag 2012). A more comprehensive phylogeny of the Margaritiferidae that included eight of 13 currently recognized species (three from North America) retained the use of *Cumberlandia* (Bolotov et al. 2015). However, based on more recent evidence (Bolotov et al. 2016; Araujo et al. 2017), we consider *Cumberlandia* a junior synonym of *Margaritifera*.

Cumberlandia *Ortmann*, 1912.—Turgeon et al. (1998) recognized one species, *Cumberlandia monodonta*. We place *Cumberlandia* in the synonymy of *Margaritifera* (see Margaritiferidae).

Margaritifera *Schumacher*, 1816.—Turgeon et al. (1998) recognized four species of *Margaritifera*. Placement of *Cumberlandia* in the synonymy of *Margaritifera* brings the number of recognized species to five (see Margaritiferidae).

Unionidae Rafinesque, 1820

Turgeon et al. (1998) recognized 49 genera, 286 species, and 13 subspecies in Unionidae. We recognize 54 genera, 293 species, and no subspecies. We provide support for and discussion of these changes in the following assessments of genera.

Actinonaias *Crosse and Fischer*, 1894.—Turgeon et al. (1998) recognized two species, *Actinonaias ligamentina* and *Actinonaias pectorosa*. Molecular analyses (e.g., Campbell et al. 2005; Zanatta and Murphy 2006) found that the two species of *Actinonaias* together did not represent a monophyletic grouping, but the position of each of these lineages within the Lampsilini was unresolved. The type locality for *Actinonaias* is central Mexico, and 10 recognized species are restricted to this region (Graf and Cummings 2017), but no species

attributable to *Actinonaias* occur between Mexico and the range of *ligamentina* and *pectorosa* in the central United States and southern Canada. No phylogenetic research has examined relationships among Mexican *Actinonaias* and *ligamentina* and *pectorosa*, but it is unlikely they are closely related considering the disjunct distribution and lack of precedent for such a geographical pattern in other freshwater taxa (e.g., Miller et al. 2005). *Actinonaias ligamentina* and *pectorosa* require placement in two different genera, but at this time we retain these two species in the genus *Actinonaias* pending the outcome of further phylogenetic research.

Alasmidonta *Say*, *1818*.—Turgeon et al. (1998) recognized 12 species, and recent evidence supports no changes to this classification.

Amblema *Rafinesque*, 1820.—Turgeon et al. (1998) recognized three species, and recent evidence supports no changes to this classification.

Anodonta Lamarck, 1799.—Turgeon et al. (1998) recognized 10 species. Mock et al. (2004) and Zanatta et al. (2007) found Anodonta to be polyphyletic, with eastern North American species forming a monophyletic clade distinct from the one that includes the type species (Anodonta cygnea, which occurs in Eurasia) and western North American Anodonta. Without discussion, Graf and Cummings (2007) and Cummings and Graf (2010) placed Anodonta couperiana, A. heardi, and A. suborbiculata in Utterbackia, and A. implicata in Pyganodon. Because no supporting evidence was provided, we do not recognize these changes. The next available genus for the eastern North American clade (A. couperiana, A. heardi, A. suborbiculata, and A. implicata) identified as distinct by Mock et al. (2004) is Utterbackiana. Anodonta hartfieldorum Williams, Bogan, and Garner, 2009, was described subsequently and also belongs to Utterbackiana (see *Utterbackiana*).

In a phylogenetic analysis of western North American *Anodonta*, Chong et al. (2008) found *A. beringiana* to be more closely related to the Asian species *Sinanodonta woodiana* than to North American species. Based on this evidence, we reassign *beringiana* to *Sinanodonta* (see *Sinanodonta*).

We retain the remaining four western North American species within *Anodonta* (*A. californiensis*, *A. kennerlyi*, *A. nuttalliana*, and *A. oregonensis*) based on their phylogenetic affinity to Eurasian *Anodonta* (Mock et al. 2004; Zanatta et al. 2007; Chong et al. 2008). *Anodonta dejecta* was recognized by Turgeon et al. (1998), Graf and Cummings (2007), and Cummings and Graf (2010). This species is treated as a synonym of *A. californiensis* by Bequaert and Miller (1973) and the Arizona Game and Fish Department (2017). We do not recognize *A. dejecta*, which is here placed in synonymy of *A. californiensis*.

Anodontoides *Simpson in Baker, 1898.*—Turgeon et al. (1998) recognized two species. One additional species, *Anodontoides denigrata*, was recognized without discussion by Neves et al. (1997) and Cicerello and Schuster (2003). Haag and Cicerello (2016) recognized *A. denigrata* on the basis of molecular data showing that upper Cumberland River

drainage populations were distinct from A. ferussacianus (Bogan and Raley 2013), and we recognize this species for the same reason. Bogan and Raley (2013) referred to A. denigrata as A. argenteus (Lea, 1840), for which the type locality is Stones River, Tennessee. The Stones River is a tributary of the middle Cumberland River and well downstream of the putative distribution of A. denigrata and other species considered endemic to the upper Cumberland River drainage upstream of the hypothesized original location of Cumberland Falls (Haag and Cicerello 2016). Until further research delineates this species' distribution more precisely, we use A. denigrata, for which the type locality is in the upper Cumberland River drainage (Clear Fork, Campbell County, Tennessee; see Ortmann 1918). Ahlstedt et al. (2016) reported a possibly distinct Anodontoides species from the Powell River, Virginia, but further work is needed to determine its validity and taxonomy.

Arcidens Simpson, 1900.—Turgeon et al. (1998) recognized one species, Arcidens confragosus. Clarke (1981) considered Arkansia (see Arkansia) a junior synonym of Arcidens (see also Graf and Cummings 2007), and this classification was supported by morphological and molecular data (Inoue et al. 2014). We recognize two species of Arcidens.

Arkansia *Ortmann and Walker*, 1912.—Arkansia was described as a monotypic genus including A. wheeleri, which was recognized by Turgeon et al. (1998). We place Arkansia in the synonymy of Arcidens (see Arcidens).

Cyclonaias *Pilsbry in Ortmann and Walker, 1922.*—Turgeon et al. (1998) recognized *Cyclonaias*, which has long been considered a monotypic genus for *C. tuberculata*. *Cyclonaias tuberculata* has been aligned with the Quadrulini based on morphological (e.g., Frierson 1927; Modell 1964) and protein polymorphism data (Davis and Fuller 1981). Heard and Guckert (1971) placed *Cyclonaias* in the Pleurobemini based on its ectobranchous brooding (see also Graf and Cummings 2007). However, it appears that ectobranchy arose multiple times (Davis and Fuller 1981; Graf 2002; Roe and Hoeh 2003), meaning that this trait does not necessarily exclude *Cyclonaias* from the Quadrulini, and some female *C. tuberculata* brood glochidia in all four gills (Frierson 1927).

Recent molecular studies consistently supported inclusion of *Cyclonaias* in the Quadrulini, but they further show that it is a member of a monophyletic clade including *Q. pustulosa* and related species (Campbell et al. 2005; Campbell and Lydeard 2012b). Serb et al. (2003) did not support this relationship, but these results were later attributed to an error in sample labeling (Campbell and Lydeard 2012b). However, Serb et al. (2003) as well as Campbell et al. (2005) and Campbell and Lydeard (2012b) support the monophyly of the *Quadrula pustulosa* clade and its distinctiveness from other species of *Quadrula* (see *Quadrula* and *Theliderma*). In addition to *Cyclonaias tuberculata*, the *Quadrula pustulosa* clade identified by these studies includes the following species recognized by Turgeon et al. (1998): *Q. asperata*, *Q. aurea*, *Q. houstonensis*, *Q. nodulata*, *Q. petrina*, *Q. pustulosa*, and *Q. refulgens*, as well

as Fusconaia succissa and Quincuncina infucata (see Fusconaia and Quincuncina).

The name Quadrula is not available for the Q. pustulosa clade because the type species, Q. quadrula, is a member of another distinct, monophyletic clade (see Quadrula). Graf and Cummings (2007) elevated the generic name Amphinaias Crosse and Fischer, 1894, for the Q. pustulosa clade. The type species for Amphinaias (by original designation) is Unio couchianus Lea, 1860, which has a quadrate shell and sulcus (but lacks pustules) similar to the Q. quadrula clade. This morphology is very different from the rounded, pustulose shells of the Q. pustulosa clade. Quadrula couchiana is considered extinct and genetic data are unavailable; however, we do not consider *Amphinaias* an available name for the Q. pustulosa clade because of the strongly divergent morphology of the type species. Campbell and Lydeard (2012b) proposed Rotundaria Rafinesque, 1820, as a name for the Q. pustulosa clade, presuming its availability based on statements in Valenciennes (1827). However, Valenciennes noted that Rafinesque had confused two species, one for which he kept Rafinesque's name *Unio verrucosa* and named the other *Unio* tuberculosa [sic]. As such, Valenciennes's statement cannot be accepted as a subsequent designation of Obliquaria tuberculata Rafinesque, 1820, as the type species of Rotundaria (P. Bouchet, Muséum National d'Histoire Naturelle, Paris, personal communication), and Herrmannsen (1848) later designated Obliquaria subrotunda Rafinesque, 1820, as the type species of Rotundaria. Rafinesque did not select a type species for Rotundaria and because more than one species was included by him in the genus, the type species cannot be fixed by monotypy. Therefore, *Rotundaria* is not available for the Q. pustulosa clade. Frierson (1927) erected the subgenus Bullata for Q. pustulosa but realized this was preoccupied and created the replacement name *Pustulosa* with the same type species. Thus, Cyclonaias becomes the oldest available name for this

Of the 10 species discussed above as members of Cyclonaias, three were not recognized by Turgeon et al. (1998) (C. archeri, C. kieneriana, and C. kleiniana), and one was considered a subspecies (C. mortoni, as Quadrula pustulosa mortoni). Graf and Cummings (2007) elevated Q. archeri from synonymy with Q. asperata, but they provided no justification for this change. The distinctiveness of C. archeri was recognized by Williams et al. (2008) based on its morphology, absence of intergrades, and isolated and restricted distribution. We recognize C. archeri. The distinctiveness of C. kieneriana was recognized by Williams et al. (2008) based on shell morphology; however, it was not supported by molecular data (Serb et al. 2003), but that study included only one specimen of this putative taxon. We recognize C. kieneriana until additional information becomes available (see Williams et al. 2008). Cyclonaias kleiniana was synonymized under Quincuncina infucata by Clench and Turner (1956), but molecular studies supported the distinctiveness of these species and their inclusion in Cyclonaias (Lydeard et al. 2000; Campbell and Lydeard 2012b).

Molecular data supported the distinctiveness of *C. mortoni* from *C. pustulosa* (Serb et al. 2003). In summary, we recognize *Cyclonaias* as including 14 species: *C. tuberculata*, seven species recognized by Turgeon et al. (1998) under *Quadrula*, one subspecies recognized by Turgeon et al. (1998) but now elevated to species status (*C. mortoni*), two species recognized by Turgeon et al. (1998) in different genera (*C. infucata* and *C. succissa*), and three species elevated from synonymy (*C. archeri*, *C. kieneriana*, and *C. kleiniana*).

Cyprogenia Agassiz, 1852.—Turgeon et al. (1998) recognized two species. Subsequent molecular data suggested cryptic species diversity in the genus (Serb and Barnhart 2008; Grobler et al. 2011). The most recent molecular analysis of Cyprogenia identified three independent evolutionary lineages: C. aberti in the Ozark drainages of Arkansas, Missouri, and Kansas; C. stegaria in the Ohio River Basin; and a third lineage in the Ouachita River drainage in Arkansas (Chong et al. 2016). Confusion regarding the type locality of Unio lamarckianus Lea, 1852, requires resolution to determine whether that name is available for the Ouachita River drainage population. We recognize the distinctiveness of this species but defer including it in our list until a specific epithet can be designated.

Cyrtonaias *Crosse and Fischer*, 1894.—Turgeon et al. (1998) recognized one species, *Cyrtonaias tampicoensis*, and recent evidence supports no changes to this classification. Five other species are recognized, all of which occur in Mexico or Central America (Graf and Cummings 2017).

Disconaias *Crosse and Fischer, 1894.*—Turgeon et al. (1998) recognized one species, *Disconaias salinasensis* Simpson in Dall, 1908, which was subsequently placed in the synonymy of *Disconaias fimbriata* by Graf and Cummings (2007). Five other species are recognized, all of which occur in Mexico (Graf and Cummings 2017). We recognize *Disconaias fimbriata* as the only species of the genus occurring in the United States (Rio Grande drainage).

Dromus *Simpson*, 1900.—Turgeon et al. (1998) recognized one species, *Dromus dromas*, and recent evidence supports no changes to this classification.

Ellipsaria *Rafinesque*, 1820.—Turgeon et al. (1998) recognized one species, *Ellipsaria lineolata*, and recent evidence supports no changes to this classification.

Elliptio *Rafinesque*, 1819.—Turgeon et al. (1998) recognized 36 species, making it the largest unionid genus in the United States and Canada, but species concepts within this group remain mostly untested, and their highly variable shell morphology precludes traditional approaches for species diagnosis. Recent molecular studies have largely supported the monophyly of *Elliptio* with two exceptions (Campbell et al. 2005; Campbell and Lydeard 2012b; Perkins et al. 2017). *Elliptio dilatata*, which is morphologically and anatomically similar to many *Elliptio*, is not a member of this group; we recognize reassignment of this species to *Eurynia* (Campbell and Lydeard 2012b). We also recognize reassignment of *Elliptio steinstansana* to *Parvaspina* based on molecular data (Perkins et al. 2017). It is important to note that phylogenetic

affinities remain unknown for most species that we currently recognize under *Elliptio* and some may prove to be members of other genera (e.g., *Eurynia*; Elderkin et al. 2008; Campbell and Lydeard 2012b).

Because of our poor understanding of species diversity within *Elliptio*, we largely retain the classification of Turgeon et al. (1998) with the following exceptions. We stress that this classification is provisional and meant to provide a stable, working hypothesis for diversity within the genus. We elevate from synonymy four species of *Elliptio*: *E. fumata* (from *E*. complanata), E. occulta and E. pullata (from E. icterina), and E. purpurella (from E. arctata and E. strigosa); these changes are based primarily on differences in shell morphology (Brim Box and Williams 2000; Williams et al. 2008, 2011, 2014). We place eight species into synonymy. Four Atlantic Slope species (E. errans, E. hepatica, E. lugubris, and E. raveneli) were recognized by Turgeon et al. (1998) based on Davis and Mulvey (1993). The research by Davis and Mulvey (1993) was confined almost exclusively to the Savannah River drainage and has no context within the greater Atlantic Coast region. The validity of these species has not been evaluated further. We return these species to synonymy following Johnson (1970) as follows: E. errans is synonymized under E. complanata; and E. hepatica, E. lugubris, and E. raveneli are synonymized under E. icterina. We place Elliptio waccamawensis into the synonymy of E. congaraea based on molecular data (McCartney et al. 2016). We place the following species into synonymy based on examination of shell type material by Clarke (1992) and Williams et al. (2011, 2014): E. waltoni (synonymized under E. ahenea), E. judithae (synonymized under E. roanokensis), and E. buckleyi (synonymized under E. jayensis). After these changes, we recognize 30 species of Elliptio, and it remains the largest unionid genus in the United States and Canada.

Turgeon et al. (1998) listed the common names Flat Spike and Florida Shiny Spike for *Elliptio jayensis* and *E. buckleyi*, respectively. We follow the recommendation of Williams et al. (2014) that the common name of *E. jayensis* be changed to Florida Spike because the species is largely endemic to that state and is neither consistently flat nor shiny.

Elliptoideus *Frierson*, 1927.—Turgeon et al. (1998) recognized one species, *Elliptoideus sloatianus*, and recent evidence supports no changes to this classification.

Epioblasma *Rafinesque*, 1831.—Turgeon et al. (1998) recognized 20 species and five subspecies. Our changes to this classification involve recognition of two newly described cryptic species, elevating one species from synonymy, and elevating subspecies to species status. We recognize *Epioblasma ahlstedti* Jones and Neves, 2010, a cryptic species formerly included within *E. capsaeformis*, and we recognize and elevate to species status *Epioblasma aureola* Jones and Neves, 2010, formerly identified as *E. florentina walkeri* but described as *E. florentina aureola* Jones and Neves, 2010.

Epioblasma cincinnatiensis was not recognized by Turgeon et al. (1998), and it has been considered a synonym (e.g., Parmalee and Bogan 1998) or a subspecies (e.g., Morrison

1942) of *Epioblasma torulosa*. Williams et al. (2008) elevated this species from synonymy based on examination of shell type material. Watters et al. (2009) also recognized this taxon but placed it in the synonymy of *Epioblasma phillipsii* (Conrad, 1835). However, *E. phillipsii* is considered a synonym of *Obliquaria reflexa* (see Williams et al. 2008). We follow Williams et al. (2008) in recognizing *E. cincinnatiensis*.

Turgeon et al. (1998) recognized eight subspecies of Epioblasma in three nominal species: florentina (three), obliquata (two), and torulosa (three). A conclusive assessment of the taxonomic status of these taxa may be impossible at this time because half are considered extinct (E. florentina florentina, E. f. curtisii, E. torulosa torulosa, and E. t. gubernaculum). Cummings and Berlocher (1990) found no evidence of intergradation between E. t. torulosa and E. t. rangiana and both taxa co-occurred at many sites; based on this evidence, we elevate these subspecies to species status. Epioblasma aureola and E. walkeri represent morphologically and genetically distinct sister taxa (Jones and Neves 2010, as E. florentina aureola and E. florentina walkeri). These taxa appear to be restricted to two different river systems (Tennessee and Cumberland, respectively); based on the low probability of exchange between these populations and their distinctiveness, we recognize and elevate to full species status E. aureola and E. walkeri. There is little information with which to assess the taxonomic status of E. florentina florentina, E. florentina curtisii, E. obliquata obliquata, E. obliquata perobliqua, and E. torulosa gubernaculum, but all have distinctive shell morphology or occupy distinct geographical regions and we recognize all these taxa as distinct species (see Methods).

We recognize 28 *Epioblasma* species, making it the second largest unionid genus in the United States and Canada.

Eurynia Rafinesque, 1820.—Eurynia was not recognized in Turgeon et al. (1998). Eurynia was elevated from synonymy by Campbell and Lydeard (2012b) to accommodate Elliptio dilatata, which consistently falls outside the Elliptio clade in molecular analyses (see also Perkins et al. 2017). We consider Eurynia monotypic at this time, but more inclusive molecular studies may identify other species that belong to this genus, including some now assigned to Elliptio (Elderkin et al. 2008; Campbell and Lydeard 2012b).

Fusconaia Simpson, 1900.—Turgeon et al. (1998) recognized 13 species. Several studies showed that the genus Fusconaia as portrayed by Turgeon et al. (1998) was polyphyletic (Lydeard et al. 2000; Serb et al. 2003; Campbell et al. 2005; Campbell and Lydeard 2012a, 2012b; Pfeiffer et al. 2016). Based on these results, we reassign three species recognized by Turgeon et al. (1998) to other genera: F. succissa to Cyclonaias, F. barnesiana to Pleuronaia, and F. ebenus to Reginaia. Pleuronaia was resurrected to accommodate F. barnesiana, along with two other species in the clade (Williams et al. 2008; Campbell and Lydeard 2012a, 2012b; see Pleuronaia). Reginaia was described to accommodate F.

ebenus and two other species (Campbell and Lydeard 2012a; see Reginaia).

These studies also showed that several species assigned to other genera belonged in *Fusconaia*. Based on these results, *Quincuncina* is a junior synonym of *Fusconaia*, and we reassign *Q. burkei* and *Q. mitchelli* to *Fusconaia* (Lydeard et al. 2000; Serb et al. 2003; Campbell et al. 2005; Pfeiffer et al. 2016; see *Cyclonaias*, *Quadrula*, and *Quincuncina*). *Lexingtonia* was placed in the synonymy of *Fusconaia* when its type species, *L. subplana*, was determined a junior synonym of *Fusconaia masoni* based on molecular data (Bogan et al. 2003).

Fusconaia chunii was not recognized by Turgeon et al. (1998), but they recognized two other Fusconaia from Texas: F. askewi and F. lananensis. Subsequent molecular data showed that all Fusconaia in Texas drainages from the Sabine River west belonged to a single species (Burlakova et al. 2012). However, Unio chunii Lea, 1861, has priority over Unio askewi Marsh, 1896, and Quadrula lananensis Frierson, 1901, so we place F. askewi and F. lananensis in the synonymy of F. chunii.

We adopt the former common name for *F. askewi*, Texas Pigtoe, for *F. chunii* because it is descriptive of the species' range. Turgeon et al. (1988) listed the common name Gulf Pigtoe for *Fusconaia cerina*, but it was changed to Southern Pigtoe in Turgeon et al. (1998) without comment. However, Turgeon et al. (1998) also used Southern Pigtoe as the common name of *Pleurobema georgianum*. We designate the common name Gulf Pigtoe for *F. cerina*.

In summary, we recognize 11 species of *Fusconaia*, including eight species recognized by Turgeon et al. (1998) under *Fusconaia*, two species recognized by Turgeon et al. (1998) in other genera, and one species elevated from synonymy.

Glebula *Conrad*, 1853.—Turgeon et al. (1998) recognized one species, *Glebula rotundata*, and recent evidence supports no changes to this classification.

Gonidea *Conrad*, 1857.—Turgeon et al. (1998) recognized one species, *Gonidea angulata*, and recent evidence supports no changes to this classification.

Hamiota Roe and Hartfield, 2005.—Hamiota was described subsequent to Turgeon et al. (1998) to accommodate a monophyletic clade of four species that produce superconglutinates (Roe et al. 2001). They were previously recognized under Lampsilis: L. altilis, L. australis, L. perovalis, and L. subangulata (Roe and Hartfield 2005). We recognize all four of these species under Hamiota.

Hemistena *Rafinesque*, 1820.—Turgeon et al. (1998) recognized one species, *Hemistena lata*, and recent evidence supports no changes to this classification.

Lampsilis *Rafinesque*, 1820.—Turgeon et al. (1998) recognized 28 species and four subspecies. Molecular data indicated that *Lampsilis*, as presented by Turgeon et al. (1998), is polyphyletic (Graf and Ó Foighil 2000; Campbell et al. 2005). There are likely unrecognized taxa in the genus *Lampsilis* (e.g., in Arkansas; Harris et al. 2009). The genus

Hamiota was described to accommodate a monophyletic clade of four species, Lampsilis altilis, L. australis, L. perovalis, and L. subangulata (Roe and Hartfield 2005), and we recognize reassignment of these species from Lampsilis to Hamiota. We also recognize reassignment of Lampsilis haddletoni to Obovaria (Williams et al. 2008; see Obovaria). In addition to Hamiota, molecular data suggested the existence of at least two other paraphyletic clades within Lampsilis as recognized by Turgeon et al. (1998). Lampsilis cardium, L. ornata, and L. ovata formed a monophyletic clade sister to Hamiota, and L. siliquoidea and L. teres were members of a clade sister to the latter two groups; however, these groupings were not consistently or strongly supported, and the analyses did not include other species of putative Lampsilis (Campbell et al. 2005). Additional generic-level changes regarding Lampsilis will likely occur in the future, but we retain traditional use of this genus for all species except those reassigned to Hamiota and Obovaria.

Lampsilis floridensis was not recognized by Turgeon et al. (1998), and formerly it was recognized as a subspecies (Clench and Turner 1956) or synonym (Burch 1975) of Lampsilis teres. We recognize L. floridensis as a full species based on shell morphology, unpublished molecular data, and its allopatric distribution (Williams et al. 2008).

Turgeon et al. (1998) recognized nominal *Lampsilis reeveiana* along with two subspecies, *L. r. brevicula* and *L. r. brittsi*. Molecular data showed that *brittsi* populations from the Missouri River drainage formed a well-supported monophyletic clade separate from nominal *reeveiana*, but there was no morphological or genetic distinction between nominal *L. reeveiana* and *L. r. brevicula* (Harris et al. 2004). Based on these data, we follow McMurray et al. (2012) in recognizing *L. brittsi* and *L. reeveiana* as species and placing *L. reeveiana brevicula* into the synonymy of *L. reeveiana*.

Turgeon et al. (1998) recognized nominal *Lampsilis radiata* and one subspecies, *L. r. conspicua*. However, molecular and shell morphology data did not support the distinctiveness of *L. r. conspicua* (Stiven and Alderman 1992), and we place this taxon into the synonymy of *Lampsilis radiata*. Turgeon et al. (1998) also recognized *Lampsilis fullerkati*, but we recognize placement of that species into the synonymy of *L. radiata* based on molecular data (McCartney et al. 2016).

Turgeon et al. (1998) recognized nominal Lampsilis straminea and one subspecies, L. s. claibornensis. Lampsilis straminea straminea is restricted to the Black Belt Prairie region of Alabama and Mississippi and is characterized by a profusion of fine, concentric ridges on the shell, which are absent in L. s. claibornensis. However, concentric ridges are present in some other mussels inhabiting streams in the Black Belt Prairie region and are most likely environmentally induced and not due to genetic differences (Williams et al. 2008). We do not recognize the taxonomic validity of these shell forms and place L. s. claibornensis in the synonymy of Lampsilis straminea. The common name of Lampsilis s. straminea, Rough Fatmucket (Turgeon et al. 1998), is

descriptive of individuals in only a small portion of its range (i.e., the Black Belt Prairie). Therefore, we retain the common name for *L. straminea claibornensis*, Southern Fatmucket, for *L. straminea*.

In summary, we recognize 24 species of *Lampsilis* including one species elevated from synonymy and two species elevated from subspecies. *Lampsilis* is the third largest genus in the family Unionidae following *Elliptio* (30) and *Epioblasma* (28).

Lasmigona Rafinesque, 1831.—Turgeon et al. (1998) recognized six species and one subspecies. Williams et al. (2008) elevated Lasmigona complanata alabamensis to species status based on examination of museum shell material, and molecular data supported the distinctiveness of this taxon (King et al. 1999). Williams et al. (2008) also recognized Mobile Basin populations of Lasmigona holstonia as a distinct species based on unpublished molecular data and the occurrence of these populations in two different river systems. They resurrected from synonymy Lasmigona etowaensis to refer to Mobile Basin populations and retained L. holstonia to refer to Tennessee and Ohio River drainage populations. We recognize all three of these species.

Molecular studies showed that *Lasmigona* is polyphyletic: L. alabamensis, L. complanata, and L. costata formed a monophyletic clade, and L. compressa and L. subviridis represented another monophyletic clade more closely related to Alasmidonta (King et al. 1999). However, this study did not include all species of Lasmigona, and a broader study within the context of the tribe Anodontini is needed to clarify these relationships. Populations of Lasmigona costata in the Ozark Highlands represented a monophyletic clade strongly differentiated from populations east of the Mississippi River, suggesting the presence of at least one cryptic species within this taxon; additional investigation across the range of L. costata is needed to better understand these patterns (Hewitt et al. 2016). An endemic form of Lasmigona in the Barrens region of the upper Caney Fork drainage in Tennessee was recognized by Layzer et al. (1993), but the status of this putative taxon has not been evaluated further.

Lemiox *Rafinesque*, 1831.—Turgeon et al. (1998) recognized one species, *Lemiox rimosus*, and recent evidence supports no changes to this classification.

Leptodea *Rafinesque*, 1820.—Turgeon et al. (1998) recognized three species, and recent evidence supports no changes to this classification. Smith (2000) proposed moving *Leptodea ochracea* into the genus *Ligumia* based on mantle margin pigment and size of glochidia. We do not accept this proposal due to the limited number of taxa (four species in two genera) in that analysis, and we retain *ochracea* in *Leptodea*.

Lexingtonia *Ortmann*, 1914.—Turgeon et al. (1998) recognized two species. However, the type species, *Lexingtonia subplana*, was subsequently relegated to the synonymy of *Fusconaia masoni* based on Johnson (1970) and Bogan et al. (2003). As such, *Lexingtonia* is a junior synonym of *Fusconaia*. The other species recognized by Turgeon et al. (1998), *Lexingtonia dolabelloides*, did not group with

Fusconaia in molecular analyses but formed a monophyletic clade with two other species (Campbell et al. 2005; Campbell and Lydeard 2012a, 2012b). *Pleuronaia* was resurrected by Williams et al. (2008) to accommodate this clade (see *Pleuronaia*).

Ligumia *Swainson*, *1840*.—Turgeon et al. (1998) recognized three species. Subsequent molecular studies indicated the genus is not monophyletic, but further research is needed to fully elucidate these patterns (Campbell et al. 2005; Kuehnl 2009). We retain the classification of Turgeon et al. (1998), but as additional information becomes available taxa assigned to this genus will likely change (see Raley et al. 2007). Gangloff et al. (2013) identified a genetically divergent clade of *Ligumia recta* from the Mobile Basin that may warrant recognition as a distinct taxon.

Medionidus *Simpson*, 1900.—Turgeon et al. (1998) recognized seven species. We no longer recognize *Medionidus mcglameriae*, which was placed in the synonymy of *Leptodea fragilis* based on examination of the type specimen (Williams et al. 2008). Campbell et al. (2005) found some evidence for polyphyly of *Medionidus*, but this evidence was not conclusive and we make no other changes to this genus.

Megalonaias *Utterback*, 1915.—Turgeon et al. (1998) recognized one species, *Megalonaias nervosa*, and recent evidence supports no changes to this classification.

Obliquaria *Rafinesque*, 1820.—Turgeon et al. (1998) recognized one species, *Obliquaria reflexa*, and recent evidence supports no changes to this classification.

Obovaria *Rafinesque*, 1819.—Turgeon et al. (1998) recognized six species. Molecular data showed that *Obovaria* as depicted by Turgeon et al. (1998) is polyphyletic (Campbell et al. 2005). Notably, *Obovaria rotulata* was not a member of this group, and it was later reassigned to *Reginaia* (Campbell and Lydeard 2012b); we recognize this reassignment. In an analysis by Campbell et al. (2005), *O. olivaria* fell outside the clade containing other *Obovaria* and *Epioblasma*, but this conclusion was not consistently supported. We retain *olivaria* within *Obovaria*, but further work on this species is needed to resolve its generic assignment.

Evidence also supports reassignment to Obovaria of species recognized by Turgeon et al. (1998) under other genera. We reassign Villosa arkansasensis and V. choctawensis to Obovaria based on molecular data (Kuehnl 2009; Inoue et al. 2013) and marsupial morphology (Williams et al. 2011, for choctawensis). We also recognize reassignment of Lampsilis haddletoni to Obovaria based on shell morphology of the type lot (Williams et al. 2008, 2011), but this species is considered extinct and there are no available soft parts for anatomical or molecular study. Obovaria jacksoniana was recognized in Turgeon et al. (1998) but is synonymous with Villosa arkansasensis (Inoue et al. 2013). Unio jacksoniana Frierson, 1912, is a junior synonym of *Unio arkansasensis* Lea, 1862, and we place O. jacksoniana in the synonymy of Obovaria arkansasensis. There is also potential for unrecognized taxa within O. arkansasensis in central Gulf Slope drainages (Inoue et al. 2013).

In summary, we recognize seven species of *Obovaria*, including four species recognized by Turgeon et al. (1998) and three species reassigned from other genera, one from *Lampsilis* and two from *Villosa*.

Parvaspina Perkins, Gangloff, and Johnson, 2017.— Parvaspina was described subsequent to Turgeon et al. (1998) to accommodate a monophyletic clade of two species previously recognized as *Elliptio steinstansana* and *Pleurobema collina* (Perkins et al. 2017). We recognize these species as *Parvaspina steinstansana* and *Parvaspina collina*.

Pegias *Simpson*, 1900.—Turgeon et al. (1998) recognized one species, *Pegias fabula*, and recent evidence supports no changes to this classification.

Plectomerus *Conrad*, 1853.—Turgeon et al. (1998) recognized one species, *Plectomerus dombeyanus*, and recent evidence supports no changes to this classification.

Plethobasus *Simpson*, 1900.—Turgeon et al. (1998) recognized three species, and recent evidence supports no changes to this classification.

Pleurobema Rafinesque, 1819.—Turgeon et al. (1998) recognized 32 species, making it one of the largest unionid genera. Molecular data largely support the monophyly of Pleurobema as depicted by Turgeon et al. (1998) with two exceptions (Campbell et al. 2005, 2008; Campbell and Lydeard 2012b). These studies support reassignment of P. collina to Parvaspina and P. gibberum to Pleuronaia (Campbell et al. 2005, 2008; Campbell and Lydeard 2012b; see Parvaspina and Pleuronaia). However, Campbell et al. (2008) and Campbell and Lydeard (2012b) provided evidence that Pleurobema includes two distinct lineages, one including P. sintoxia, P. cordatum, P. plenum, P. riddellii, and P. rubrum and the other including all other species. Further research is needed to elucidate these relationships; we retain traditional use of Pleurobema.

Pleurobema rivals Elliptio in its large number of described species and the intractability of many species concepts, particularly in the Mobile Basin, but these problems are compounded for Pleurobema because many putative taxa are considered extinct. Based on a comprehensive comparison of shell type specimens and other available material, Williams et al. (2008) placed into synonymy nine species of Mobile Basin Pleurobema recognized by Turgeon et al. (1998): P. chattanoogaense (into P. decisum); P. murrayense (into P. stabile); P. nucleopsis and P. troschelianum (into P. georgianum); P. flavidulum and P. johannis (into P. perovatum); and P. avellanum, P. furvum, and P. hagleri (into P. rubellum). Some of these synonyms are further supported by molecular data (e.g., P. chattanoogaense, P. furvum; Campbell et al. 2008), and we recognize all of these changes. We do not recognize Pleurobema altum since it was deemed a nomen dubium because it is not identifiable due to incomplete description, vague type locality, and lack of type material (Williams et al. 2008). One Ohio River drainage species, Pleurobema bournianum, was placed into the synonymy of Pleurobema clava based on shell morphology (Watters et al. 2009), and we recognize this change.

We recognize four additional Mobile Basin species of *Pleurobema* not recognized by Turgeon et al. (1998). Williams et al. (2008) recognized three species based on examination of shell type specimens: *P. fibuloides, P. hartmanianum,* and *P. stabile.* We correct the spelling of *P. stabilis* as used by Williams et al. (2008) to *stabile* based on Lee (2008). *Pleurobema athearni* Gangloff, Williams, and Feminella, 2006, was described subsequent to Turgeon et al. (1998) based on morphological data (Gangloff et al. 2006). In addition, preliminary findings identified an undescribed species in the upper Tennessee River drainage (Schilling 2015).

In summary, we recognize 23 species of *Pleurobema*, including 19 species recognized by Turgeon et al. (1998), three species elevated from synonymy, and one newly described species.

Pleuronaia Frierson, 1927.—Pleuronaia was not included in Turgeon et al. (1998). This was the senior available name for a monophyletic clade of three species—Fusconaia barnesiana, Lexingtonia dolabelloides, and Pleurobema gibberum—identified in a molecular study by Campbell et al. (2005). We recognize resurrection of Pleuronaia to accommodate this group and reassignment of these three species to Pleuronaia as proposed previously (Williams et al. 2008; Campbell and Lydeard 2012a, 2012b). There are likely cryptic taxa of Pleuronaia in the upper Tennessee River drainage (Schilling 2015). We correct the gender agreement of the specific name of Pleuronaia gibberum to gibber (H. Lee, Jacksonville, Florida, personal communication).

Popenais *Frierson*, 1927.—Turgeon et al. (1998) recognized one species, *Popenais popeii*, and recent evidence supports no changes to this classification.

Potamilus *Rafinesque*, 1818.—Turgeon et al. (1998) recognized six species. One additional species, *Potamilus metnecktayi* Johnson, 1998, was described subsequently, and we recognize this species. *Potamilus inflatus* was referred to as the Inflated Heelsplitter by Turgeon et al. (1988) but was changed to Alabama Heelsplitter by Turgeon et al. (1998) without comment. Alabama Heelsplitter is the established common name for *Lasmigona alabamensis*, and we adopt the original common name Inflated Heelsplitter for *P. inflatus*. Roe and Lydeard (1998) found the Amite River population of *P. inflatus* to be genetically divergent, and it may warrant recognition as a distinct taxon.

Ptychobranchus *Simpson*, 1900.—Turgeon et al. (1998) recognized five species. *Ptychobranchus foremanianus* was elevated from the synonymy of *Ptychobranchus greenii* (in part) by Williams et al. (2008) based on shell morphology and periostracum color. A molecular analysis of this genus included insufficient material to resolve the relationship between these two taxa (Roe 2013), but we recognize both species. We correct the gender agreement of *Ptychobranchus subtentum* to *P. subtentus* following Lee (2008).

Pyganodon *Crosse and Fischer*, 1894.—Turgeon et al. (1998) recognized five species. Graf and Cummings (2007) without comment moved *Anodonta implicata* to *Pyganodon*

and omitted *P. fragilis* and *P. lacustris*. However, molecular data demonstrated the validity of *P. fragilis* and *P. lacustris* (Doucet-Beaupré et al. 2012). Based on these results and the lack of justification for movement of *A. implicata* to *Pyganodon*, we retain the classification of Turgeon et al. (1998) for *Pyganodon*.

Quadrula Rafinesque, 1820.—Turgeon et al. (1998) recognized 18 species and two subspecies. Molecular studies generally supported the monophyly of Quadrula as depicted by Turgeon et al. (1998), but they also showed that it is composed of three deeply divergent monophyletic clades plus Tritogonia verrucosa, each of which warranted generic recognition (Serb et al. 2003; Campbell et al. 2005; Campbell and Lydeard 2012b). The type species for Quadrula is Q. quadrula, and the clade containing this species also includes Q. apiculata, Q. fragosa, Q. nobilis, and Q. rumphiana. Quadrula nobilis was elevated from synonymy based on shell morphology and unspecified genetic data (Howells et al. 1996) but not recognized by Turgeon et al. (1998). Relationships among species in the Q. quadrula group were not clearly resolved by Serb et al. (2003), but we recognize all five species. We also recognize within this group Q. couchiana on the basis of its shell morphology, which is similar to that of Q. quadrula (see Cyclonaias).

Based on molecular data, we reassign to *Cyclonaias* 10 taxa recognized by Turgeon et al. (1998) under *Quadrula*, and we reassign 5 species to *Theliderma* (Serb et al. 2003; Campbell et al. 2005; Campbell and Lydeard 2012b; see also Graf and Cummings 2007). We also synonymize two taxa recognized by Turgeon et al. (1998) under *Quadrula* (see *Theliderma*). In summary, we recognize six species of *Quadrula*, including five recognized under this genus by Turgeon et al. (1998) and one elevated from synonymy (*Q. nobilis*).

Quincuncina *Ortmann*, 1922.—Turgeon et al. (1998) recognized three species. Molecular data showed that the type species, *Quincuncina burkei*, belongs in *Fusconaia* (Lydeard et al. 2000; Serb et al. 2003; Campbell et al. 2005). As such, *Quincuncina* is a junior synonym of *Fusconaia*, and we reassign to this genus *Q. burkei* and *Q. mitchelli* (see also Pfeiffer et al. 2016). Based on these findings, we also reassign *Q. infucata* to *Cyclonaias* (see *Cyclonaias*).

Reginaia Campbell and Lydeard, 2012.—Reginaia was described subsequent to Turgeon et al. (1998) to accommodate a monophyletic clade of two species identified in a phylogenetic analysis of Ambleminae (Campbell and Lydeard 2012b). The two Reginaia species were included in Turgeon et al. (1998) as Fusconaia ebena and Obovaria rotulata (Campbell and Lydeard 2012b); we recognize assignment of these species to Reginaia. We follow Watters et al. (2009) in correcting the spelling of the species name ebena to ebenus. A third species, Fusconaia apalachicola Williams and Fradkin, 1999, was described subsequent to Turgeon et al. (1998) from archaeological material; we reassign this species to Reginaia based on its shell characters, which are similar to those of R. ebenus and R. rotulata.

Simpsonaias *Frierson*, 1914.—Turgeon et al. (1998) recognized one species, *Simpsonaias ambigua*, and recent evidence supports no changes to this classification.

Sinanodonta Modell, 1945.—Sinanodonta was not included in Turgeon et al. (1998). This genus was previously considered to be confined to Asia and not part of the North America fauna. Molecular data showed that A. beringiana is more closely related to the Asian species Sinanodonta woodiana than to other western North American Anodonta (Chong et al. 2008; see *Anodonta*). Based on this evidence, we reassign beringiana to Sinanodonta. In 2010 S. woodiana, Chinese Pondmussel, was found in Wickecheoke Creek, a tributary of the Delaware River, New Jersey (Bogan et al. 2011a). Several known glochidial host fishes, native and introduced species, occur in the watershed (Bogan et al. 2011b). The species appears to have become established in that stream despite eradication efforts (J. Bowers-Altman, New Jersey Division of Fish and Wildlife, personal communication). We recognize S. woodiana as established in New Jersey (Table 2). This is the only nonindigenous unionid mussel known to have become established in the United States or Canada.

Strophitus *Rafinesque*, 1820.—Turgeon et al. (1998) recognized three species, and recent evidence supports no changes to this classification. *Strophitus undulatus*, one of the most wide-ranging species in the United States and Canada, likely contains unrecognized cryptic taxa (Watters et al. 2009).

Theliderma Swainson, 1840.—Theliderma was not recognized by Turgeon et al. (1998). This genus was resurrected from synonymy by Graf and Cummings (2007) to accommodate a monophyletic clade of five species recognized by Turgeon et al. (1998) under Quadrula (Q. cylindrica, Q. intermedia, Q. metanevra, Q. sparsa, and Q. stapes; see Serb et al. 2003). Theliderma is the oldest available name for this clade and has T. metanevra as its type species. We recognize placement of all five of these species in Theliderma. No molecular data are available for Theliderma stapes, but its shell morphology is very similar to that of other Theliderma, and we include it in this genus following Graf and Cummings (2007).

Turgeon et al. (1998) recognized Quadrula tuberosa, but we place this taxon in the synonymy of Theliderma metanevra following Parmalee and Bogan (1998, as Q. metanevra). However, the relationship of tuberosa to other species is uncertain, and if it represents a valid species, it is considered extinct (see Haag and Cicerello 2016). Quadrula cylindrica was recognized in Turgeon et al. (1998) as containing two subspecies, Theliderma cylindrica cylindrica and T. cylindrica strigillata. These subspecies traditionally were distinguished from each other based on shell morphology and distribution, with *strigillata* being confined mainly to the upper Tennessee River system in Tennessee and Virginia (Parmalee and Bogan 1998). However, the distributional limits of strigillata have never been clearly defined as it grades into typical T. c. cylindrica in larger streams, suggesting that the shell forms represent ecophenotypic variation (Ortmann 1920), and molecular data provide no support for recognition of *T. c. strigillata* (Serb et al. 2003; Sproules et al. 2006). Based on this evidence, we do not recognize subspecies within *T. cylindrica*. Both *T. c. cylindrica* (threatened) and *T. c. strigillata* (endangered) are federally protected taxa. Synonymizing *strigillata* under *T. cylindrica* will not remove the protection provided by the Endangered Species Act but may impact the status of populations formerly recognized as *strigillata*.

Toxolasma *Rafinesque*, 1831.—Turgeon et al. (1998) recognized eight species. Recent evidence supports no changes at the genus level, but species boundaries within *Toxolasma* remain uncertain. Howells et al. (1996) placed *Toxolasma mearnsi* in the synonymy of *Toxolasma texasiense* based on electrophoretic analysis, a change overlooked by Turgeon et al. (1998); we recognize placement of *T. mearnsi* in the synonymy of *T. texasiense*. Undescribed species of *Toxolasma* have been recognized (e.g., Gulf Lilliput) but have yet to be formerly described (Williams et al. 2008, 2014).

Lee (2006) concluded that *Toxolasma* has a neuter gender, which necessitates correction of spellings from *lividus* to *lividum*, *parvus* to *parvum*, and *paulus* to *paulum*, without change to *corvunculus*, *cylindrellus*, or *pullus*; we recognize these spelling changes. Lee (2006) provided an incorrect spelling of *Toxolasma texasiense* (as *texasense*), but we correct it based on the spelling presented in the original description.

Tritogonia *Agassiz*, *1852*.—Turgeon et al. (1998) recognized one species, *Tritogonia verrucosa*. Molecular data clearly supported inclusion of *T. verrucosa* within the tribe Quadrulini, but its placement within that group was unresolved, and Serb et al. (2003) recommended its placement within *Quadrula* (*sensu lato*) until relationships were better understood (e.g., see Williams et al. 2008; Haag and Cicerello 2016). Regardless of its relationship to other clades within the Quadrulini, *Tritogonia* represents a deeply divergent lineage (Serb et al. 2003; Campbell et al. 2012b), and our recognition of three other genera within this tribe (*Cyclonaias*, *Theliderma*, and *Quadrula* sensu stricto) warrants retention of *Tritogonia* as a monotypic genus (e.g., see Watters et al. 2009; Sietman et al. 2012).

Truncilla *Rafinesque*, 1819.—Turgeon et al. (1998) recognized four species, and recent evidence supports no changes to this classification.

Uniomerus *Conrad, 1853.*—Turgeon et al. (1998) recognized three species. Recent evidence supports no changes at the genus level, but species concepts within *Uniomerus* are uncertain (see Williams et al. 2008). *Uniomerus columbensis* was not recognized by Turgeon et al. (1998) but was elevated from synonymy by Williams et al. (2008) based on unpublished molecular data and shell morphology; we recognize this change. Species boundaries for other taxa (e.g., *Uniomerus declivis*) remain unresolved.

The inappropriate and misleading common name for *Uniomerus carolinianus*, Florida Pondhorn, was changed to Eastern Pondhorn by Williams et al. (2014) because the

species occurs not only in Florida but northward along the Atlantic Coast; we recognize this change.

Utterbackia *Baker*, 1927.—Turgeon et al. (1998) recognized three species and recent evidence supports no changes to this classification.

Utterbackiana Frierson, 1927.—Utterbackiana was not recognized by Turgeon et al. (1998). We resurrect this genus as the senior available name for a monophyletic clade of four eastern North American species included in Turgeon et al. (1998) under Anodonta (A. couperiana, A. heardi, A. implicata, and A. suborbiculata; Mock et al. 2004; Zanatta et al. 2007; see Anodonta). The type species for the genus is Anodonta suborbiculata Say, 1831. In addition to the four taxa mentioned above, a new species was described subsequent to Turgeon et al. (1998), Anodonta hartfieldorum (Williams et al. 2009). We also place this species in Utterbackiana because it appears closely related to U. suborbiculata and was formerly associated with that species.

Venustaconcha Frierson, 1927.—Turgeon et al. (1998) recognized two species. Molecular data showed that Villosa perpurpurea and Villosa trabalis also are members of Venustaconcha (Kuehnl 2009; Lane et al. 2016). Molecular data further showed that Venustaconcha perpurpurea is a junior synonym of Venustaconcha trabalis, and populations of this species in the Tennessee River drainage are genetically and morphologically distinct from those in the Cumberland River drainage (Lane et al. 2016). Based on the type locality of trabalis, Flint River, Alabama, this name is applicable to the Tennessee River drainage species. *Unio troostensis* Lea, 1834, is the oldest available name for the Cumberland drainage species (type locality is Stones River, Tennessee), and we recognize this species as Venustaconcha troostensis (see Haag and Cicerello 2016; Lane et al. 2016). Cumberland Bean was the common name used for V. trabalis by Turgeon et al. (1998), but Lane et al. (2016) proposed Tennessee Bean for Venustaconcha trabalis and Cumberland Bean for Venustaconcha troostensis; we follow this use. Venustaconcha sima was not included in Turgeon et al. (1998) but was elevated from synonymy by Gordon (1995) based on shell coloration and conchological characters, and its distinctiveness is supported by molecular data (Kuehnl 2009). This species was synonymized under Villosa iris by Parmalee and Bogan (1998), and molecular data support its relationship to Villosa (Kuehnl 2009). We recognize sima as a species of Villosa.

Villosa *Frierson*, 1927.—Turgeon et al. (1998) recognized 17 species and one subspecies. Molecular data show that *Villosa*, as depicted by Turgeon et al. (1998), is wildly polyphyletic, with species occurring in as many as seven different clades within the Lampsilini (Kuehnl 2009). These and other data support reassignment of *Villosa trabalis* to *Venustaconcha*, synonymization of *Villosa perpurpurea* under *Venustachoncha trabalis* (see *Venustaconcha*), and reassignment of *Villosa choctawensis* and *V. arkansasensis* to *Obovaria* (see *Obovaria*). Most other species will require reassignment to existing genera (e.g., *V. vaughniana* to *Ligumia*; Raley et al. 2007; Kuehnl 2009) or resurrected or newly described genera, potentially with only *Villosa amygdala*

and *V. villosa* remaining in *Villosa* (Kuehnl 2009). However, these relationships are not fully understood, and currently synonymized or newly described generic names have not been proposed. With the exception of *Villosa trabalis*, *V. perpurpurea*, *V. choctawensis*, and *V. arkansasensis*, we retain all other species recognized by Turgeon et al. (1998) in *Villosa*.

Villosa vanuxemensis umbrans was elevated to species status by Williams et al. (2008) based on shell characters and preliminary molecular data, and subsequent molecular data support this change (Kuehnl 2009); based on this evidence, we recognize *V. umbrans*. There are several undescribed taxa within Villosa (Kuehnl 2009; Harris et al. 2009). We recognize correction of gender agreement for Villosa amygdala, as given by Turgeon et al. (1998), to Villosa amygdalum following Williams et al. (2011, 2014). We recognize fifteen species of Villosa.

DISCUSSION

Changes in mussel taxonomy compared to Turgeon et al. (1998) reflect our better understanding of mussel phylogenetic relationships obtained mainly from molecular genetic data (e.g., Serb et al. 2003; Campbell and Lydeard 2012a, 2012b; Inoue et al. 2013, 2014; Pfeiffer et al. 2016). Molecular genetics continues to be one of the most important tools for understanding unionoid relationships and taxonomy, but other data sets (e.g., life history, host use, soft anatomy, shell morphology, zoogeography) are informative and should not be overlooked when constructing phylogenies and conducting taxonomic studies (e.g., Roe et al. 2001; Jones and Neves 2010; Lane et al. 2016).

We recognize a larger number of genera than Turgeon et al. (1998; 56 vs. 49), but the number of currently recognized species is similar. However, recent studies show that considerable cryptic biodiversity exists in the Unionidae (e.g., *Cyprogenia*, *Lampsilis*, *Villosa*). Most of this biodiversity remains to be discovered, and its future recognition may result in increased numbers of species in the United States and Canada (see Haag 2012). Currently unrecognized species may be narrowly distributed (e.g., one river system) and in need of conservation measures. Development of additional molecular markers, more inclusive taxon sampling, advancements in phylogenetic analyses, and other techniques for species delineation are facilitating taxonomic recognition of species. More thorough understanding of life histories with improved husbandry techniques should also help facilitate species recognition.

Future research will most likely reveal unrecognized taxa. Conversely, additional synonymy may be warranted for some currently recognized species. Much more research is needed to delineate true diversity of the mussels of the United States and Canada.

ACKNOWLEDGMENTS

We thank the following individuals who were always very responsive to our questions regarding names of freshwater mussels: John Alderman, Gerry Dinkins, Mike Gangloff, Dan Graf, Jordan Holcomb, Bob Howells, Sarina Jepsen, Paul Johnson, Stephen McMurray, Terry Myers, Charles Randklev, Kevin Roe, Tim Savidge, Daniel Schilling, Brian Watson, and Jason Wisniewski. We acknowledge Harry G. Lee (Jacksonville, Florida) for providing expert advice on the proper terminations for numerous species names. We also thank Sherry L. Bostick for assistance in preparation and review of several drafts of the manuscript. Although the individuals mentioned here provided assistance and input, we bear full responsibility for any errors. The findings and conclusions in this article are those of the authors and do not necessarily represent the views of their agencies and institutions. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

LITERATURE CITED

- Ahlstedt, S. A., M. T. Fagg, R. S. Butler, J. F. Connell, and J. W. Jones. 2016. Quantitative monitoring of freshwater mussel populations from 1979–2004 in the Clinch and Powell Rivers of Tennessee and Virginia, with miscellaneous notes on the fauna. Freshwater Mollusk Biology and Conservation 19:1–18.
- Araujo, R., S. Schneider, K. J. Roe, D. Erpenbeck, and A. Machrodom. 2017.
 The origin and phylogeny of Margaritiferidae (Bivalvia, Unionoida): A synthesis of molecular and fossil data. Zoologica Stripta 46:289–307. doi: 10.1111/zsc.12217
- Arizona Game and Fish Department. 2017. Heritage data management system. Anodonta californiensis, California Floater. Available at http://www.azgfd.gov/pdfs/w_c/hdms/Invertibrates/Anodcali.fo.pdf (accessed June 15, 2017).
- Bequaert, J. C., and W. B. Miller. 1973. The Mollusks of the Arid Southwest, with an Arizona Check List. The University of Arizona Press, Tucson. 271 pp.
- Bieler, R., J. G. Carter, and E. V. Coan. 2010. Classification of bivalve families. Pages 113–133 in P. Bouchet, J.-P. Rocroi, Rüdiger Bieler, J. G. Carter, and E. V. Coan, editors. Nomenclator of Bivalve Families with a Classification of Bivalve Families. Malacologia 52:1–184.
- Bogan, A. E., J. Bowers-Altman, and M. E. Raley. 2011a. A new threat to conservation of North American freshwater mussels: Chinese Pond Mussel Sinanodonta woodiana in the United States. Tentacle 19:39–40.
- Bogan, A. E., J. Bowers-Altman, and M. E. Raley. 2011b. The first confirmed record of the Chinese Pond Mussel (*Sinanodonta woodiana*) (Bivalvia: Unionidae) in the United States. The Nautilus 125:41–43.
- Bogan, A. E., and M. E. Raley. 2013. Taxonomic status of the Cumberland Papershell, Anodontoides argenteus (Lea, 1840) [formerly Anodontoides denigrata (Lea, 1852)] (Mollusca: Bivalvia: Unionidae). Unpublished report submitted to U.S. Fish and Wildlife Service, Frankfort, Kentucky. 32 pp.
- Bogan, A. E., M. Raley, and J. Levine. 2003. Determination of the systematic position and relationships of the Atlantic Pigtoe, *Fusconaia masoni* (Conrad, 1834) (Mollusca: Bivalvia: Unionidae) with distributions in Virginia, North and South Carolina, and Georgia. Unpublished report submitted to U.S. Fish and Wildlife Service, Asheville, North Carolina. 14 pp.
- Bolotov, I. N., Y. V. Bespalaya, I. V. Vikhrev, O. V. Aksenova, P. E. Aspholm, M. Y. Gofarov, O. K. Klishko, Y. S. Kolosova, A. V. Kondakov, A. A. Lyubas, I. S. Paltser, E. S. Konopleva, S. Tumpeesuwan, N. N. Bolotov, and I. S. Voroshilova. 2015. Taxonomy

- and distribution of the freshwater pearl mussels (Unionoida: Margaritiferidae) in the Far East of Russia. PLoS ONE 10:e0122408. doi: 10.1371/journal.pone.0122408
- Bolotov, I. N., I. V. Vikhrev, Y. V. Bespalaya, M. Y. Gofarov, A. V. Kondakov, E. S. Konopleva, N. N. Bolotov, and A. A. Lyubas. 2016. Multi-locus fossil-calibrated phylogeny, biogeography and a subgeneric revision of the Margaritiferidae (Mollusca: Bivalvia: Unionoida). Molecular Phylogenetics and Evolution 103:104–121.
- Brim Box, J., and J. D. Williams. 2000. Unionid mollusks of the Apalachicola Basin in Alabama, Florida, and Georgia. Alabama Museum of Natural History Bulletin 21:1–143.
- Burch, J. B. 1973. Freshwater unionacean clams (Mollusca: Pelecypoda) of North America. Biota of Freshwater Ecosystems. Identification Manual 11, U.S. Environmental Protection Agency, Washington, D.C. 176 pp.
- Burch, J. B. 1975. Freshwater unionacean clams (Mollusca: Pelecypoda) of North America. Revised edition. Malacological Publications, Hamburg, Michigan. 204 pp.
- Burlakova, L. E., D. Campbell, A. Y. Karatayev, and D. Barclay. 2012. Distribution, genetic analysis and conservation priorities for rare Texas freshwater molluscs in the genera *Fusconaia* and *Pleurobema* (Bivalvia: Unionidae). Aquatic Biosystems 8:1–15.
- Campbell, D. C., P. D. Johnson, J. D. Williams, A. K. Rindsberg, J. M. Serb, K. K. Small, and C. Lydeard. 2008. Identification of 'extinct' freshwater mussel species using DNA barcoding. Molecular Ecology Resources 8:711–724. doi: 10.1111/j.1755-0998.2008.02108.x
- Campbell, D. C., and C. Lydeard. 2012a. Molecular systematics of *Fusconaia* (Bivalvia: Unionidae: Ambleminae). American Malacological Bulletin 30:1–17.
- Campbell, D. C., and C. Lydeard. 2012b. The genera of Pleurobemini (Bivalvia: Unionidae: Ambleminae). American Malacological Bulletin 30:19–38.
- Campbell, D. C., J. M. Serb, J. E. Buhay, K. J. Roe, R. L. Minton, and C. Lydeard. 2005. Phylogeny of North American amblemines (Bivalvia, Unionoida): Prodigious polyphyly proves pervasive across genera. Invertebrate Biology 124:131–164.
- Carter, J. G., C. R. Altaba, L. C. Anderson, R. Araujo, A. S. Biakov, A. E. Bogan, D. C. Campbell, M. Campbell, C. Jin-hua, J. C. W. Cope, G. Delvene, H. H. Dijkstra, F. Zong-jie, R. N. Gardner, V. A. Gavrilova, I. A. Goncharova, P. J. Harries, J. H. Hartman, M. Hautmann, W. R. Hoeh, J. Hylleberg, J. Bao-yu, P. Johnston, L. Kirkendale, K. Kleemann, J. Koppka, J. Kříž, D. Machado, N. Malchus, A. Márquez-Aliaga, J.-P. Masse, C. A. McRoberts, P. U. Middelfart, S. Mitchell, L. A. Nevesskaja, S. Özer, J. Pojeta, Jr., I. V. Polubotko, J. M. Pons, S. Popov, T. Sánchez, A. F. Sartori, R. W. Scott, I. I. Sey, J. H. Signorelli, V. V. Silantiev, P. W. Skelton, T. Steuber, J. B. Waterhouse, G. L. Wingard, and T. Yancey. 2011. A synoptical classification of the Bivalvia (Mollusca). Paleontological Contributions No. 4. Kansas University Paleontological Institute. The University of Kansas, Lawrence. 47 pp.
- Chong, J. P., J. C. Brim Box, J. K. Howard, D. Wolf, T. L. Myers, and K. E. Mock. 2008. Three deeply divided lineages of the freshwater mussel genus *Anodonta* in western North America. Conservation Genetics 9:1303–1309.
- Chong, J. P., J. L. Harris, and K. J. Roe. 2016. Incongruence between mtDNA and nuclear data in the freshwater mussel genus *Cyprogenia* (Bivalvia: Unionidae) and its impact on species delineation. Ecology and Evolution 6:2439–2452. doi: 10.1002/ece3.2071
- Cicerello, R. R., and G. A. Schuster. 2003. A guide to the freshwater mussels of Kentucky. Kentucky State Nature Preserves Commission, Scientific and Technical Series, No. 7. 62 pp.
- Clarke, A. H. 1981. The tribe Alasmidontini (Unionidae: Anodontinae). Part I: Pegias, Alasmidonta, and Arcidens. Smithsonian Contributions to Zoology, No. 326. 101 pp.

- Clarke, A. H. 1992. Brief communications. Malacology Data Net 3:98.
- Clench, W. J., and R. D. Turner. 1956. Freshwater mollusks of Alabama, Georgia, and Florida from the Escambia to the Suwannee River. Bulletin of the Florida State Museum, Biological Sciences 1:97–239, plates 1–9.
- Combosch, D. J., T. M. Collins, E. A. Glover, D. L. Graf, E. M. Harper, J. M. Healy, G. Y. Kawauchi, S. Lemer, E. McIntyre, E. E. Strong, J. D. Taylor, J. D. Zardus, P. M. Mikkelsen, G. Giribet, and R. Bieler. 2017. A family-level Tree of Life for bivalves based on a Sanger-sequencing approach. Molecular Phylogenetics and Evolution 107:191–208.
- Cummings, K. S., and J. M. K. Berlocher. 1990. The naiades or freshwater mussels (Bivalvia: Unionidae) of the Tippecanoe River, Indiana. Malacological Review 23:83–98.
- Cummings, K. S., and D. L. Graf. 2010. Mollusca: Bivalvia. Pages 309–384 in J. H. Thorp and A. P. Covich, editors. Ecology and Classification of North American Freshwater Invertebrates. 3rd ed. Elsevier, Amsterdam, The Netherlands.
- Davis, G. M., and S. L. H. Fuller. 1981. Genetic relationships among Recent Unionacea (Bivalvia) of North America. Malacologia 20:217–253.
- Davis, G. M., and P. Mulvey. 1993. Species status of Mill Creek *Elliptio*. Savannah River Plant National Environment Research Park, SRO–NERP 22:4–58.
- Doucet-Beaupré, H., P. U. Blier, E. G. Chapman, H. Piontkivska, F. Dufresne, B. E. Sietman, R. S. Mulcrone, and W. R. Hoeh. 2012. *Pyganodon* (Bivalvia: Unionoida: Unionidae) phylogenetics: A male- and female-transmitted mitochondrial DNA perspective. Molecular Phylogenetics and Evolution 63:430–444.
- Elderkin, C. L., A. D. Christian, J. L. Metcalfe-Smith, and D. J. Berg. 2008.
 Population genetics and phylogeography of freshwater mussels in North America, *Elliptio dilatata* and *Actinonaias ligamentina* (Bivalvia: Unionidae). Molecular Ecology 17:2149–2163.
- Frierson, L. S. 1927. A Classification and Annotated Check List of the North American Naiades. Baylor University Press, Waco, Texas. 111 pp. Errata et Corrigenda.
- Gangloff, M. M., B. A. Hamstead, E. F. Abernethy, and P. D. Hartfield. 2013.
 Genetic distinctiveness of *Ligumia recta*, the Black Sandshell, in the Mobile River Basin and implications for its conservation. Conservation Genetics 14:913–916. doi: 10.1007/s10592-013-0480-0
- Gangloff, M. M., J. D. Williams, and J. W. Feminella. 2006. A new species of freshwater mussel (Bivalvia: Unionidae), *Pleurobema athearni*, from the Coosa River drainage of Alabama, USA. Zootaxa 1118:43–56.
- Gilbert, C. R. 1961. Hybridization versus intergradation: An inquiry into the relationship of two cyprinid fishes. Copeia 1961:181–192.
- Gordon, M. E. 1995. Venustaconcha sima (Lea), an overlooked freshwater mussel (Bivalvia: Unionoidea) from the Cumberland River basin of central Tennessee. The Nautilus 108:55–60.
- Graf, D. L. 2002. Molecular phylogenetic analysis of two problematic freshwater mussel genera (*Unio* and *Gonidea*) and a re-evaluation of the classification of Nearctic Unionidae (Bivalvia: Palaeoheterodonta: Unionoida). Journal of Molluscan Studies 68:65–71.
- Graf, D. L., and K. S. Cummings. 2007. Review of the systematics and global diversity of freshwater mussel species (Bivalvia: Unionoida). Journal of Molluscan Studies 73:291–314.
- Graf, D. L., and K. S. Cummings. 2017. The freshwater mussels (Unionoida) of the world (and other less consequential bivalves). MUSSELp database. Available at http://mussel-project.uwsp.edu/db/ (accessed March 25, 2017).
- Graf, D. L., and D. Ó Foighill. 2000. The evolution of brooding characters among the freshwater pearly mussels (Bivalvia: Unionoidea) of North America. Journal of Molluscan Studies 66:157–170.
- Grobler, J. P., J. W. Jones, N. A. Johnson, R. J. Neves, and E. M. Hallerman.
 2011. Homogeneity at nuclear microsatellite loci masks mitochondrial

haplotype diversity in the endangered Fanshell Pearlymussel (*Cyprogenia stegaria*). Journal of Heredity 102:196–206.

- Haag, W. R. 2012. North American Freshwater Mussels: Natural History, Ecology, and Conservation. Cambridge University Press, New York. 505 pp.
- Haag, W. R., and R. R. Cicerello. 2016. A distributional atlas of the freshwater mussels of Kentucky. Scientific and Technical Series 8. Kentucky State Nature Preserves Commission, Frankfort. 299 pp.
- Harris, J. L., W. R. Hoeh, A. D. Christian, J. Walker, J. L. Farris, R. L. Johnson, and M. E. Gordon. 2004. Species limits and phylogeography of Lampsilinae (Bivalvia; Unionoida) in Arkansas with emphasis on species of *Lampsilis*. Unpublished final report to Arkansas Game and Fish Commission and U.S. Fish and Wildlife Service. 70 pp, 10 plates.
- Harris, J. L., W. R. Posey, 2nd, C. L. Davidson, J. L. Farris, S. R. Oetker, J. N. Stoeckel, M. G. Crump, S. Barnett, H. C. Martin, J. H. Seagraves, N. J. Wentz, R. Winterringer, C. Osborne, and A. D. Christian. 2009. Unionoida (Mollusca: Margaritiferidae, Unionidae) in Arkansas, third status review. Journal of the Arkansas Academy of Science 63:50–86.
- Heard, W. H., and R. H. Guckert. 1971. A re-evaluation of the Recent Unionacea (Pelecypoda) of North America. Malacologia 10:333–355.
- Herrmannsen, A. N. 1848. Indicis generum Malacozoorum primordia. Nomina subgenerum, familiarum, tribuum, ordinum, classium; adjectis auctoribus, temporibus, locis systematicis atque literariis, etymis, synonymis. Praetermittuntur Cirripedia, Tunicata et Rhizopoda. 2:353–492.
- Hewitt, T. L., J. L. Bergner, D. A. Woolnough, and D. T. Zanatta. 2016. Phylogeography of the freshwater mussel species *Lasmigona costata*: Testing post-glacial colonization hypotheses. Hydrobiologia. doi: 10. 1007/s10750-016-2834-3
- Hoeh, W. R., A. E. Bogan, K. S. Cummings, and S. I. Guttman. 2002. Evolutionary relationships among the higher taxa of freshwater mussels (Bivalvia: Unionoida): Inferences on phylogeny and character evolution from analyses of DNA sequence data. Malacological Review 31–32:123– 141
- Hoeh, W. R., A. E. Bogan, and W. H. Heard. 2001. A phylogenetic perspective on the evolution of morphological and reproductive characteristics in the Unionoida. Pages 257–280 in G. Bauer and K. Wächtler, editors. Ecology and Evolution of the Freshwater Mussels Unionoida. Ecological Studies, Vol. 145. Springer-Verlag, Berlin.
- Hoeh, W. R., A. E. Bogan, W. H. Heard, and E. G. Chapman. 2009. Palaeoheterodont phylogeny, character evolution, diversity and phylogenetic classification: A reflection on methods of analysis. Malacologia 51:307–317.
- Howells, R. G., R. W. Neck, and H. D. Murray. 1996. Freshwater Mussels of Texas. Texas Parks and Wildlife Department, Inland Fisheries Division, Austin. 218 pp.
- Huang, J., and L. L. Knowles. 2016. The species versus subspecies conundrum: Quantitative delimitation from integrating multiple data types within a single Bayesian approach in Hercules Beetles. Systematic Biology 65:685–699.
- Huff, S. W., D. Campbell, D. L. Gustafson, C. Lydeard, C. R. Altaba, and G. Giribet 2004. Investigations into the phylogenetic relationships of freshwater pearl mussels (Bivalvia: Margaritiferidae) based on molecular data: Implications for their taxonomy and biogeography. Journal of Molluscan Studies 70:379–388.
- Inoue, K., D. M. Hayes, J. L. Harris, and A. D. Christian. 2013. Phylogenetic and morphometric analyses reveal ecophenotypic plasticity in freshwater mussels *Obovaria jacksoniana* and *Villosa arkansasensis* (Bivalvia: Unionidae). Ecology and Evolution 3:2670–2683.
- Inoue, K., A. L. McQueen, J. L. Harris, and D. J. Berg. 2014. Molecular phylogenetics and morphological variation reveal recent speciation in freshwater mussels of the genera *Arcidens* and *Arkansia* (Bivalvia: Unionidae). Biological Journal of the Linnean Society 112:535–545.

- Johnson, R. I. 1970. The systematics and zoogeography of the Unionidae (Mollusca: Bivalvia) of the southern Atlantic Slope Region. Bulletin of the Museum of Comparative Zoology 140:263–449.
- Johnson, R. I. 1998. A new mussel, *Potamilis metnecktayi* (Bivalvia: Unionidae), from the Rio Grande system, Mexico and Texas with notes on Mexican *Disconaias*. Occasional Papers on Mollusks 5:427–455, plates 22–27.
- Jones, J. W., and R. J. Neves. 2010. Descriptions of a new species and a new subspecies of freshwater mussels, *Epioblasma ahlstedti* and *Epioblasma* florentina aureola (Bivalvia: Unionidae), in the Tennessee River drainage, USA. The Nautilus 124:77–92.
- Jones, J. W., R. J. Neves, S. A. Ahlstedt, and E. M. Hallerman. 2006. A holistic approach to taxonomic evaluation of two closely related endangered freshwater mussel species, the Oyster Mussel *Epioblasma* capsaeformis and Tan Riffleshell *Epioblasma florentina walkeri* (Bivalvia: Unionidae). Journal of Molluscan Studies 72:267–283. doi: 10.1093/ mollus/eyl004
- King, T. L., M. S. Eackles, B. Gjetvaj, and W. R. Hoeh. 1999. Intraspecific phylogeography of *Lasmigona subviridis* (Bivalvia: Unionidae): Conservation implications of range discontinuity. Molecular Ecology 8:S65–S78.
- Kuehnl, K. F. 2009. Exploring levels of genetic variation in the freshwater mussel genus *Villosa* (Bivalvia: Unionidae) at different spatial and systematic scales: Implications for biogeography, taxonomy, and conservation. Doctoral dissertation, The Ohio State University, Columbus
- Lane, T. W., E. M. Hallerman, and J. W. Jones. 2016. Phylogenetic and taxonomic assessment of the endangered Cumberland Bean, *Villosa* trabalis and Purple Bean, *Villosa perpurpurea* (Bivalvia: Unionidae). Conservation Genetics 17:1109–1124. doi: 10.1007/s10592-016-0847-0
- Layzer, J. B., M. E. Gordon, and R. M. Anderson. 1993. Mussels: The forgotten fauna of regulated rivers. A case study of the Caney Fork River. Regulated Rivers: Research and Management 8:63–71.
- Lee, H. G. 2006. Musings on a local specimen of *Toxolasma paulum* (I. Lea, 1840), the Iridescent Lilliput. Shell-O-Gram 47:3–6.
- Lee, H. G. 2008. Book review: Freshwater Mussels of Alabama and the Mobile Basin in Georgia, Mississippi and Tennessee. The Nautilus 122:261–263.
- Lopes-Lima, M., E. Froufe, V. T. Do, M. Ghamizi, K. E. Mock, U. Kebapci, O. Klishko, S. Kovitvadhi, U. Kovitvadhi, O. S. Paul, J. M. Pfeiffer, 3rd, M. Raley, N. Riccardi, H. Sereflisan, R. Sousa, A. Teirxeira, S. Varandas, X. P. Wu, D. T. Zanatta, A. Zieritz, and A. E. Bogan. 2017. Phylogeny of the most species rich freshwater bivalve family (Bivalvia: Unionida: Unionidae): Defining modern subfamilies and tribes. Molecular Phylogeny and Evolution 106:174–191. Available at http://dx.doi.org/10.1016/j. ympev.2016.08.021
- Lydeard, C., R. L. Minton, and J. D. Williams. 2000. Prodigious polyphyly in imperiled freshwater pearly-mussels (Bivalvia: Unionidae): A phylogenetic test of species and generic designations. Pages 145–158 in E. M. Harper, J. D. Taylor, and J. A. Crane, editors. The Evolutionary Biology of the Bivalvia. Geological Society Special Publication, No. 177.
- Mayr, E., E. G. Linsley, and R. L. Usinger. 1953. Methods and Principles of Systematic Zoology. McGraw-Hill, New York. 336 pp.
- McCartney, M. A., A. E. Bogan, K. M. Sommer, and A. E. Wilbur. 2016. Phylogenetic analysis of Lake Waccamaw freshwater mussel species. American Malacological Bulletin 34:109–120.
- McMurray, S. E., J. S. Faiman, A. Roberts, B. Simmons, and M. C. Barnhart. 2012. A guide to Missouri's freshwater mussels. Missouri Department of Conservation, Jefferson City, 94 pp.
- Miller, R. R., W. L. Minckley, and S. M. Norris. 2005. Freshwater Fishes of México. University of Chicago Press, Chicago, Illinois. 490 pp.
- Mock, K. E., J. C. Brim Box, M. P. Miller, M. E. Downing, and W. R. Hoeh. 2004. Genetic diversity and divergence among freshwater mussel

- (Anodonta) populations in the Bonneville Basin of Utah. Molecular Ecology 13:1085–1098.
- Modell, H. 1964. Das natürliche system der Najaden. 3. Archiv für Molluskenkunde 93:71–126.
- Morrison, J. P. E. 1942. Preliminary report on mollusks found in the shell mounds of the Pickwick Landing Basin in the Tennessee River Valley. Pages 337–392 in W. S. Webb and D. L. DeJarnette, editors. An archaeological survey of Pickwick Basin in the adjacent portions of the states of Alabama, Mississippi and Tennessee. Bureau of American Ethnology, Bulletin 129.
- Neves, R. J., A. E. Bogan, J. D. Williams, S. A. Ahlstedt, and P. W. Hartfield. 1997. Status of aquatic mollusks in the southeastern United States: A downward spiral of diversity. Pages 43–85 in G. A. Benz and D. E. Collins, editors. Aquatic Fauna in Peril: The Southeastern Perspective. Special Publication No. 1, Southeast Aquatic Research Institute. Lenz Design & Communications, Decatur, Georgia.
- Ortmann, A. E. 1918. The nayades (freshwater mussels) of the upper Tennessee drainage. With notes on synonymy and distribution. Proceedings of the American Philosophical Society 57:521–626.
- Ortmann, A. E. 1920. Correlation of shape and station in freshwater mussels (Naiades). Proceedings of the American Philosophical Society 59:269–312
- Parmalee, P. W., and A. E. Bogan. 1998. The Freshwater Mussels of Tennessee. The University of Tennessee Press, Knoxville. 328 pp.
- Perkins, M. A., N. A. Johnson, and M. M. Gangloff. 2017. Molecular systematics of the critically-endangered North American spinymussels (Unionidae: *Elliptio* and *Pleurobema*) and description of *Parvaspina* gen. nov. Conservation Genetics 18:745–757. doi: 10.1007/s10592-017-0924-z
- Pfeiffer, J. M., 3rd, N. A. Johnson, C. R. Randklev, R. G. Howells, and J. D. Williams. 2016. Generic reclassification and species boundaries in the rediscovered freshwater mussel "Quadrula" mitchelli (Simpson in Dall, 1896). Conservation Genetics 17:279–292. doi: 0.1007/s10592-015-0780-7
- Raley, M. E., A. E. Bogan, C. B. Eads, and J. F. Levine. 2007. Molecular evidence for a novel placement of the Carolina Creekshell, *Villosa* vaughaniana (Lea, 1836). Page 41 in Freshwater Mollusk Conservation Society Symposium, Little Rock, Arkansas.
- Roe, K. J. 2013. Molecular phylogenetics and zoogeography of the freshwater mussel genus *Ptychobranchus* (Bivalvia: Unionidae). Bulletin of the American Malacological Society 31:257–265.
- Roe, K. J., and P. D. Hartfield. 2005. *Hamiota*, a new genus of freshwater mussel (Bivalvia: Unionidae) from the Gulf of Mexico drainages of the southeastern United States. The Nautilus 119:1–10.
- Roe, K. J., P. D. Hartfield, and C. Lydeard. 2001. Phylogenetic analysis of the threatened and endangered superconglutinate-producing mussels of the genus *Lampsilis* (Bivalvia: Unionidae). Molecular Ecology 10:2225– 2234.
- Roe, K. J., and W. R. Hoeh. 2003. Systematics of freshwater mussels (Bivalvia: Unionoida). Pages 91–122 in C. Lydeard and D. R. Lindberg, editors. Molecular Systematics and Phylogeography of Mollusks. Smithsonian Books, Washington, D.C.
- Roe, K. J., and C. Lydeard. 1998. Species delineation and the identification of evolutionarily significant units: Lessons from the freshwater mussel genus *Potamilus* (Bivalvia: Unionidae). Journal of Shellfish Research 17:1359– 1363.
- Schilling, D. E. 2015. Assessment of morphological and molecular genetic variation of freshwater mussel species belonging to the genera *Fusconaia*, *Pleurobema*, and *Pleuronaia* in the upper Tennessee River basin. Master's thesis, Virginia Polytechnic Institute and State University, Blacksburg.
- Serb, J. M., and M. C. Barnhart. 2008. Congruence and conflict between molecular and reproductive characters when assessing biological diversity

- in the Western Fanshell *Cyprogenia aberti* (Bivalvia, Unionidae). Annals of the Missouri Botanical Garden 95:248–261.
- Serb, J. M., J. E. Buhay, and C. Lydeard. 2003. Molecular systematics of the North American freshwater bivalve genus *Quadrula* (Unionidae: Ambleminae) based on mitochondrial ND1 sequences. Molecular Phylogenetics and Evolution 28:1–11.
- Sietman, B. E., J. M. Davis, and M. C. Hove. 2012. Mantle display and glochidia release behaviors of five quadruline freshwater mussel species (Bivalvia: Unionidae). American Malacological Bulletin 30:39–46.
- Smith, D. G. 2000. On the taxonomic placement of *Unio ochraceus* Say, 1817 in the genus *Ligumia* (Bivalvia: Unionidae). The Nautilus 114:115–160.
- Smith, D. G. 2001. Systematics and distribution of the recent Margaritiferidae. Pages 33–49 in G. Bauer and K. Wächtler, editors. Ecology and Evolution of Freshwater Mussels Unionoida. Ecological Studies, Vol. 145. Springer-Verlag, Berlin.
- Sproules, J., P. Grobler, N. Johnson, J. W. Jones, R. J. Neves, and E. M. Hallerman. 2006. Genetic analysis of selected populations of the Rabbitsfoot Pearlymussel (*Quadrula cylindrica cylindrica*) (Bivalvia: Unionidae). Unpublished final report submitted to U.S. Fish and Wildlife Service, Frankfort, Kentucky. 16 pp.
- Stiven, A. E., and J. Alderman. 1992. Genetic similarities among certain freshwater mussel populations of the *Lampsilis* genus in North Carolina. Malacologia 34:355–369.
- Turgeon, D. D., A. E. Bogan, E. V. Coan, W. K. Emerson, W. G. Lyons, W. L. Pratt, C. F. E. Roper, A. Scheltema, F. G. Thompson, and J. D. Williams. 1988. Common and Scientific Names of Aquatic Invertebrates from the United States and Canada: Mollusks. American Fisheries Society, Special Publication 16. 277 pp., 12 plates.
- Turgeon, D. D., J. F. Quinn, A. E. Bogan, E. V. Coan, F. G. Hochberg, W. G. Lyons, P. Mikkelsen, R. J. Neves, C. F. E. Roper, G. Rosenberg, B. Roth, A. Scheltema, F. G. Thompson, M. Vecchione, and J. D. Williams. 1998. Common and Scientific Names of Aquatic Invertebrates from the United States and Canada: Mollusks, 2nd ed. American Fisheries Society, Special Publication 26, 526 pp.
- Valenciennes, A. 1827. Coquilles fluviatiles bivalves du Nouveau-Continent, recueillies pendant le voyage de MM. De Humboldt et Bonpland. In A. von Humboldt and A. J. A. Bonpland, editors. Recueil d'observations de zoologie et d'anatomie compare, faites dans l'ocean Atlantique, dans l'intérieur du nouveau continent et dans la mer du sud pendant les années 1799, 1800, 1801, 1802 et 1803; par Al. de Humbodt et A. Bonpland. J. Smith and Gide, Paris, 2:225–237, colored plates 48, 50, 53, 54.
- Walker, J. M., J. P. Curole, D. E. Wade, E. G. Chapman, A. E. Bogan, G. T. Watters, and W. R. Hoeh. 2006. Taxonomic distribution and phylogenetic utility of gender-associated mitochondrial genomes in the Unionoida (Bivalvia). Malacologia 48:265–282.
- Watters, G. T., M. A. Hoggarth, and D. H. Stansbery. 2009. The Freshwater Mussels of Ohio. The Ohio State University Press, Columbus. 421 pp.
- Williams, J. D., A. E. Bogan, and J. T. Garner. 2008. The Freshwater Mussels of Alabama and the Mobile Basin of Georgia, Mississippi, and Tennessee. University of Alabama Press, Tuscaloosa. 908 pp.
- Williams, J. D., A. E. Bogan, and J. T. Garner. 2009. A new species of freshwater mussel, Anodonta hartfieldorum (Bivalvia: Unionidae), from the Gulf Coastal Plain drainages of Alabama, Florida, Louisiana and Mississippi, USA. The Nautilus 123:25–33.
- Williams, J. D., R. S. Butler, G. L. Warren, and N. A. Johnson. 2014. Freshwater Mussels of Florida. University of Alabama Press, Tuscaloosa. 498 pp.
- Williams, J. D., R. S. Butler, and J. M. Wisniewski. 2011. Annotated synonymy of the recent freshwater mussel taxa of the families Margaritiferidae and Unionidae described from Florida and drainages contiguous with Alabama and Georgia. Bulletin of the Florida Museum of Natural History 51:1–84.

- Williams, J. D., and A. Fradkin. 1999. Fusconaia apalachicola, a new species of freshwater mussel (Bivalvia: Unionidae) from pre-Columbian archaeological sites in the Apalachicola basin of Alabama, Florida, and Georgia. Tulane Studies in Zoology 31:51–62.
- Williams, J. D., M. L. Warren, Jr., K. S. Cummings, J. L. Harris, and R. J. Neves. 1993. Conservation status of the freshwater mussels of the United States and Canada. Fisheries 18:6–22.
- Zanatta, D. T., and R. W. Murphy. 2006. Evolution of active host-attraction
- strategies in the freshwater mussel tribe Lampsilini (Bivalvia: Unionidae). Molecular Phylogenetics and Evolution 41:195–208. doi: 10.1016/j. ympev.2006.05.030
- Zanatta, D. T., A. Ngo, and J. Lindell. 2007. Reassessment of the phylogenetic relationships among *Anodonta*, *Pyganodon*, and *Utterbackia* (Bivalvia: Unionoida) using mutation coding of allozyme data. Proceedings of the Academy of Natural Sciences of Philadelphia 156:211–216.