

RESEARCH ARTICLE

TUBI (TURkish Benthic Index): A new biotic index for assessing impacts of organic pollution on benthic communities

Melih Ertan Çinar^{1*}, Kerem Bakır¹, Bilal Öztürk¹,
Tuncer Katağan¹, Ertan Dağlı¹, Şermin Açık², Alper Doğan¹,
Banu Bitlis Bakır²

¹ Department of Hydrobiology, Faculty of Fisheries, Ege University, 35100, Bornova, İzmir, TURKEY

² Institute of Marine Sciences and Technology, Dokuz Eylül University, Inciraltı, 35340, İzmir, TURKEY

*Corresponding autor: melih.cinar@ege.edu.tr

Abstract

A new biotic index, TUBI (TURkish Benthic Index) is proposed here to assess the impacts of organic enrichments on benthic community structures. This new index has two metrics; the Shannon-Weaver's diversity index (metric 1) and the relative abundance of ecological groups (metric 2). The ecological groups of species, which include five categories, were re-organized under three major categories here, namely, sensitive species (including GI and GII), tolerant species (GIII) and opportunistic species (GIV and GV). The metric 2 considers these groups with different weights and eliminates sensitive species in the calculation. Scores of TUBI vary between 0 and 5, and the benthic quality status increases with increasing TUBI scores. Benthic samples collected from Izmir Bay, and the Aegean and Levantine Seas were analyzed by using different biotic indices such as AMBI, M-AMBI, BENTIX, MEDOCC and TUBI, based on the national database for the ecological groups of benthic species. All biotic indices used, with some exceptions, discriminated poor and bad ecological status in the regions. The correlation analysis performed between the total organic carbon in sediment and total inorganic nitrogen in deep water, and biotic indices indicated that TUBI possessed the highest negative correlation values with these environmental variables, therefore better detecting the variability across a gradient of pollution-mediated impacts on benthic communities.

Keywords: biotic index, benthic index, organic enrichment, Mediterranean, Turkey

Introduction

Deteriorations in benthic environments due to waste-water discharges trigger changes in the compositions and functions of community structures that enable

us to predict and estimate the magnitude of the impacts by using some biological tools such as the indicator species concept (Pockington & Wells 1992; Dean 2008; Marques *et al.* 2009). Indicator or opportunistic species are known to have a r-strategy life history trait (Pianka 1970; Heip 1995) and broader ecological valence (Vrijenhoek 1979), but a weak competitive ability that hinders them to form dense populations in pristine, healthy environments. However, they outburst their populations steadily to a level where they utilize optimally sources emerged in the newly re-established environment (Grassle and Grassle 1974; Tsutsumi *et al.* 1990). Both native and alien species can become opportunistic species. Through loading ballast water in polluted harbors and discharging it in the recipient, polluted harbor, ocean-going commercial ships enable opportunistic species to disperse across the world's oceans, and some of them (e.g. *Polydora cornuta* and *Pseudopolydora paucibranchiata*) have been classified both as invasive alien species and new pollution indicator species in the Mediterranean Sea (Çinar *et al.* 2012; Çinar and Bakir 2014).

The European Water Framework Directive (WFD: 2000/60/EC) commits member states to achieve at least a Good Environmental Status (GES) for all European water bodies by 2020 at the latest. The directive sets out eleven qualitative descriptors. The descriptor 6, namely the sea-floor integrity, has 6 indicators, of which two are closely related to the ecological assessments of water bodies by using the presence of sensitive and tolerant species, and multi-metric indices (Rice *et al.* 2010). The directive encourages the usage of biotic indices to determine the benthic quality status across Europe (Blanchet *et al.* 2008; Van Hoey *et al.* 2010). For this aim, a number of biotic indices have been developed, most of which are based on the relative abundances of ecological groups of species within benthic communities (Borja *et al.* 2000; Simboura and Zenetos 2002). However, abundance-based indices such as the Shannon-Weaver's Diversity Index, which has been used traditionally in pollution monitoring studies in marine environments, were also used for classifying water bodies (Dauvin *et al.* 2007; Albayrak *et al.* 2010). Although there are some doubts in considering it as a biotix index, as its approach to produce a score does not match with the species indicator concept, it has been still used as a community descriptor together with the Pielou's Evenness Index (Çinar *et al.* 2006), as a complementary tool to predict possible effects of human-mediated pressures on marine benthic ecosystems (Rosenberg, 1976; Kocatas *et al.* 1985; Ergen *et al.* 2006; Simboura *et al.* 2014) or as a metric in multi-metric indices such as M-AMBI (Mixuka *et al.* 2007).

The usage of biotic indices to assess the benthic ecological status of water bodies is widespread in the Mediterranean, but choosing the most adequate one changes according to habitats, regions and countries. For instance, Greece prefers using BENTIX, Italy and Slovenia M-AMBI, and Spain AMBI and MEDOCC (Borja *et al.* 2009). These biotic indices, except for M-AMBI, are based on one metric that refers to the relative abundances of ecological groups

in samples, to which species have been assigned according to literature knowledge and expert judgement. However, weights of the ecological groups in the calculations and groupings of ecological groups vary among biotic indices, which somewhat lead to different evaluations of water bodies. For example, BENTIX considers two groups, namely sensitive and tolerant ecological groups, and gives three times higher weight to sensitive species in the calculation (Simboura and Zenetos 2002). In contrast, AMBI and MEDOCC consider five and four (GIV and GV were joined together) ecological groups, respectively, and give higher weights to opportunistic species (Borja *et al.* 2000; Pinedo *et al.* 2012). In addition, these indices are recommended to be estimated by using software or an Excel macro, which have their own databases for ecological groups of benthic species. However, these databases represent some differences in attaining species to ecological groups. Therefore, it has been stressed the importance of using a similar, intercalibrated database for the calculations of biotic indices that would minimize bias derived from using different databases (Çinar *et al.* 2012).

Biotic indices based on one metric might lead to wrong conclusion if stations include only sensitive and the first order of opportunistic species. For example, as AMBI and MEDOCC eliminate sensitive species in their calculations, in this scenario, they would indicate a bad ecological status. Therefore, a balance in the formula is needed to consider also the presence of sensitive species in the area. M-AMBI has been developed for filling this gap, but the calculation of this index is very difficult as it needs scores of a factorial analysis from a special statistical programme. Although there is software at AZTI's web page (<http://www.azti.es>) to calculate M-AMBI, it uses the AMBI's database for ecological groups of species, thus it could be useless if a modified or different database (for different ecoregions) is required. Therefore, we have developed a new index here, which has two metrics and is easy to calculate and easy to use with different databases.

This paper describes this two-metric benthic index (TUBI) that involves both the Shannon-Weaver's Diversity Index and the relative abundances of ecological groups. The scores of TUBI were compared with those of the widely-used uni- and multi-metric indices in the Mediterranean and were validated using the environmental variables.

Materials and Methods

1. Sampling sites

The new index, TUBI, was tested along the coasts of Turkey. Macrobenthic data derived from two projects were used; 1) seasonal samples collected at eight stations in 2009 in Izmir Bay, and 2) samples taken from the Aegean (27 stations) and Levantine (26 stations) Seas within the framework of the national pollution monitoring study in 2011 (Figure 1). The faunistic data of the samples

from Izmir Bay and the features of the stations have already given in Çınar *et al.* (2012). Polychaete data collected during the pollution monitoring study were also published by Çınar and Dagli (2013). The main features of the stations along the Aegean and Levantine Seas are given in Table 1.

The stations located in the outer-most part of Izmir Bay are far from any source of pollution and those located in the inner part of the bay are near pollution sources. Three replicates were collected seasonally at each station of Izmir Bay (except for station 4 in fall), whereas only one sample was taken at stations during the pollution monitoring study.

Benthic samples were taken by a standard Van Veen Grap, sampling an area of 0.1 m², and passed through 0.5 mm mesh, and the retained material on sieve was fixed with 4% formaldehyde in the field. In the laboratory, samples were washed with tap water and then sorted according to taxonomic groups under a stereomicroscope and then preserved with 70% ethanol. Specimens were identified under a stereo- and dissecting microscopes and counted.

Specimens were deposited in the Museum of Faculty of Fisheries, Ege University (ESFM).

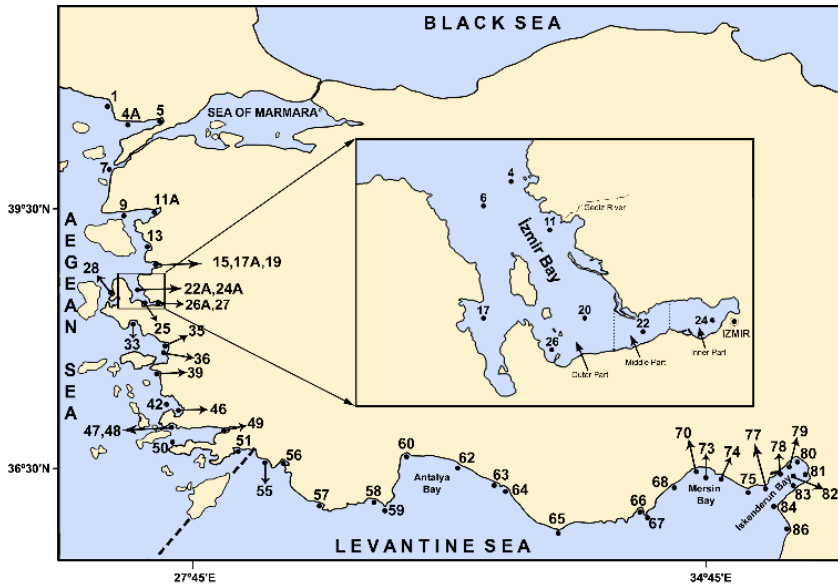


Figure 1. Map of the studied area with the locations of sampling sites

Table 1. The locality name, depth, salinity and temperature recorded at sampling stations in the Aegean and Levantine Seas

| Station Number | Locality | Depth (m) | Salinity (‰) | Temperature (°C) |
|----------------------|--------------------|-----------|--------------|------------------|
| AEGEAN SEA | | | | |
| 1 | Edirne Enez | 20 | 38.41 | 17.91 |
| 4A | Gelibolu Peninsula | 108 | 39.02 | 16.05 |
| 5 | Gelibolu Peninsula | 44 | 39.00 | 16.58 |
| 7 | Bababurnu | 18 | 39.14 | 18.20 |
| 9 | Altınoluk | 34 | 39.13 | 19.37 |
| 11A | Edremit | 38 | 39.21 | 17.80 |
| 13 | Dikili Bay | 23 | 39.17 | 19.19 |
| 15 | Bakırçay | 4 | 34.83 | 18.71 |
| 17A | Çandarlı Bay | 23 | 39.19 | 17.80 |
| 19 | Aliağa | 13 | 39.33 | 18.89 |
| 22A | Gediz | 56 | 39.17 | 17.96 |
| 24A | İzmir Bay | 39 | 39.19 | 18.47 |
| 25 | İzmir Bay | 24 | 39.61 | 23.03 |
| 26A | İzmir Bay | 11 | 39.70 | 24.74 |
| 27 | İzmir Bay | 13 | 39.72 | 26.01 |
| 28 | Çeşme | 65 | 39.23 | 18.17 |
| 33 | Sığacık | 12 | 39.32 | 21.87 |
| 35 | Küçük Menderes | 23 | 39.33 | 22.41 |
| 36 | Kuşadası | 55 | 39.25 | 20.78 |
| 39 | Büyük Menderes | 45 | 39.16 | 20.93 |
| 42 | Güllük | 71 | 39.21 | 18.63 |
| 46 | Güllük | 51 | 39.18 | 20.52 |
| 47 | Bodrum | 37 | 39.06 | 22.65 |
| 48 | Bodrum | 19 | 39.22 | 24.56 |
| 49 | Gökova Bay | 73 | 39.15 | 19.45 |
| 50 | Datça Bay | 80 | 39.05 | 19.34 |
| 51 | Marmaris | 13 | 39.36 | 27.33 |
| LEVANTINE SEA | | | | |
| 55 | Dalaman | 5 | 39.07 | 27.33 |
| 56 | Fethiye Bay | 325 | 38.89 | 14.33 |
| 57 | Kaş | 23 | 39.23 | 27.83 |
| 58 | Finike Bay | 55 | 39.01 | 22.55 |
| 59 | Yardımcı Cape | 108 | 39.08 | 18.45 |
| 60 | Antalya Bay | 45 | 38.99 | 24.98 |
| 62 | Manavgat | 5 | 39.21 | 29.20 |
| 63 | Alanya | 23 | 39.23 | 28.57 |
| 64 | Dildare Cape | 21 | 39.24 | 28.41 |
| 65 | Anamur Cape | 28 | 39.12 | 27.43 |
| 66 | Taşucu Bay | 24 | 39.31 | 29.13 |
| 67 | Göksu | 35 | 39.15 | 28.43 |
| 68 | Erdemli | 35 | 39.26 | 28.37 |
| 70 | Mersin Bay | 18 | 39.25 | 28.60 |
| 73 | Mersin Bay | 5 | 38.90 | 29.94 |
| 74 | Mersin Bay | 12 | 39.12 | 29.46 |
| 75 | Karataş | 22 | 39.26 | 28.45 |
| 77 | İskenderun Bay | 5 | 38.98 | 29.87 |
| 78 | İskenderun Bay | 17 | 39.10 | 29.33 |
| 79 | İskenderun Bay | 23 | 39.05 | 29.60 |
| 80 | İskenderun Bay | 22 | 39.08 | 29.55 |
| 81 | İskenderun Bay | 22 | 39.22 | 29.53 |
| 82 | İskenderun Bay | 60 | 39.07 | 26.60 |
| 83 | İskenderun Bay | 34 | 39.28 | 28.82 |
| 84 | Akıncı Cape | 22 | 39.21 | 28.85 |
| 86 | Samandağ | 4 | 39.19 | 28.90 |

2. Metrics for TUBI

Two metrics representing various features of macrobenthic communities were selected. Metric 1 is the Shannon-Weaver diversity index (H' , \log_2 base) and metric 2 is the relative abundance of ecological groups. Metric 1 generally varies between 0 and 5 in marine benthic habitats, but sometimes receives scores higher than 5, even reaches to 6 in mixed sediments. To stabilize the metric scores, the maximum score of H' is fixed at the score 5 here.

Macro-zoobenthic species within a benthic community can be classified into five ecological groups according to their sensitiveness to disturbances (Glémarec 1986; Borja *et al.* 2000); Group I (GI, sensitive species), Group II (GII, indifferent species), Group III (GIII, tolerant species), Group IV (GIV, second order of opportunistic species) and GV (GV, first order of opportunistic species). In the calculation of the metric 2 of TUBI, three major ecological groups were considered; Group 1 includes sensitive and indifferent species (GI and GII), Group 2 includes tolerant species (GIII), and Group 3 includes opportunistic species (GIV and GV).

3. The index (TUBI) and its boundaries according to the ecological status

The formula of Turkish Biotic Index (TUBI) is as follows:

$$\text{TUBI} = \frac{\text{Metric1} + (5 - \text{Metric2})}{2}$$
$$\text{TUBI} = \frac{H'^{\dagger} + [5 - (\frac{0 \times G1\% + 3 \times G2\% + 5 \times G3\%}{100})]}{2}$$

$\dagger H' > 5 \Rightarrow H' = 5$

This index produces scores from 0 to 5 and indicates the high ecological status when it goes towards the score 5, and denotes the azoic condition when it equals to 0. The class boundaries among the ecological status (from bad to high status) were estimated using the changes of the percentages of ecological groups (G1-G3) across the graded values of TUBI (Figure 2), from the least impacted situation to the most impacted one in seasonal samples taken (92 samples) from Izmir Bay. The point (TUBI=3) where G2 attained maximum values, and the curves of G1 and G3 meet each other (the junction point) is considered as a border between the moderate and good ecological status. The good-high ecological status boundary is formed at the score 4, where the sum of relative abundances of sensitive and indifferent species (placed in G1) at least account for 50% of total faunal populations and the opportunistic species comprise lesser than 20% of total faunal populations. The boundary between poor and moderate status is formed at a point (TUBI= 2) where the benthic community is largely dominated by opportunistic species, with low abundances of G1 and G2. If the percent dominance of opportunistic species are higher than 90% of total faunal populations (TUBI<1), the benthic ecological status can be classified as

bad. The class boundaries and ecological quality ratio (EQR) of TUBI are shown in Table 2.

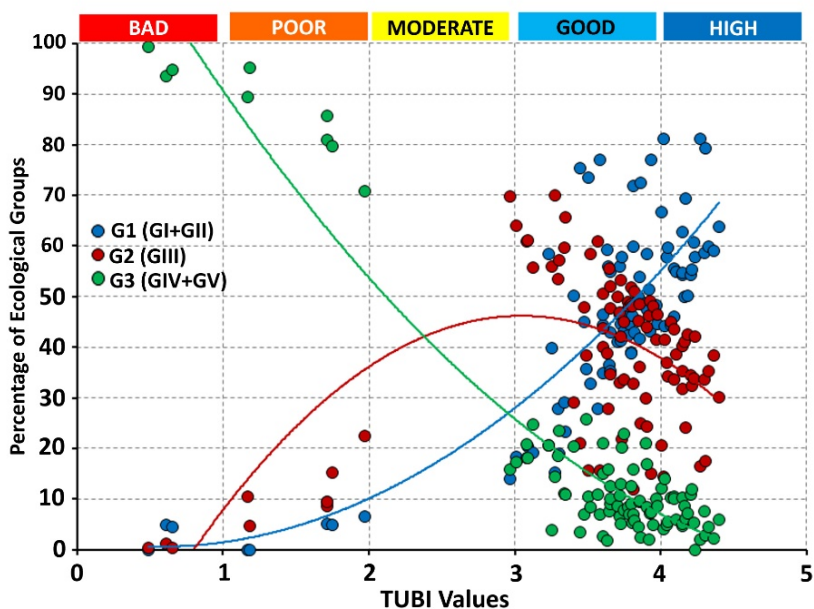


Figure 2. The percentages of ecological groups across the TUBI values

Table 2. The class boundaries of the ecological status and the ecological quality ratios based on TUBI.

| WFD Status | Impact Status | Boundaries | EQR |
|-----------------|---------------------|-----------------------------|--------------------------------|
| HIGH | Non-affected | $4 \leq \text{TUBI} \leq 5$ | $0.80 \leq \text{TUBI} \leq 1$ |
| GOOD | Slightly affected | $3 \leq \text{TUBI} < 4$ | $0.60 \leq \text{TUBI} < 0.80$ |
| MODERATE | Moderately affected | $2 \leq \text{TUBI} < 3$ | $0.40 \leq \text{TUBI} < 0.60$ |
| POOR | Heavily affected | $1 \leq \text{TUBI} < 2$ | $0.20 \leq \text{TUBI} < 0.40$ |
| BAD | Extremely affected | $0 \leq \text{TUBI} < 1$ | $0 \leq \text{TUBI} < 0.20$ |

4. Validation of the TUBI

The TUBI was evaluated by comparing it to the chemical properties in deep water and sediments at replicated seasonal samples collected at eight stations in Izmir Bay. Total organic carbon (TOC) concentrations in sediment, and total inorganic nitrogen (TIN), silica and dissolved oxygen concentrations in deep water were compared with TUBI, and other biotic indices that have been widely used in the Mediterranean Sea, such as the diversity index (H'), AMBI, M-AMBI, BENTIX and MEDOCC. The calculations of these indices were made by using the formula or methods given by Shannon and Weaver (1949) (for H'), Borja *et al.* (2000) (for AMBI), Mixuka *et al.* 2007 (for M-AMBI), Simboura and Zenetos (2002) (for BENTIX), and Pinedo *et al.* (2012) (for MEDOCC). For the calculation of M-AMBI, the reference conditions for the number of

species (S), H' and AMBI in the Aegean and Levantine Seas were taken as 90, 5 and 0, respectively. The class boundaries of these indices were indicated in Table 3. The national database including ecological groups of species, which was prepared by the Turkish experts during a project (Dekos 2014), was used to avoid the incompatibility existing in the BENTIX and AMBI databases. The ecological groups of species found during the present study are indicated in Appendix 1.

Table 3. H', BENTIX, AMBI, M-AMBI and MEDOCC class boundaries and ecological quality ratios (EQR) associated with the different ecological quality status proposed for the European Water Framework Directive (WFD).

| WFD Status | HIGH | GOOD | MODERATE | POOR | BAD |
|------------|----------------------------------|----------------------------------|----------------------------------|----------------------------------|-------------------------------|
| H' | $5 \geq H \geq 4$ | $4 > H \geq 3$ | $3 > H \geq 2$ | $2 > H \geq 1$ | $1 > H \geq 0$ |
| EQR | $1 \geq H \geq 0.80$ | $0.80 > H \geq 0.60$ | $0.60 > H \geq 0.40$ | $0.40 > H \geq 0.20$ | $0.20 > H \geq 0$ |
| AMBI | $0 \leq \text{AMBI} \leq 1.2$ | $1.2 < \text{AMBI} \leq 3.3$ | $3.3 < \text{AMBI} \leq 4.3$ | $4.3 < \text{AMBI} \leq 5.5$ | $5.5 < \text{AMBI} \leq 7$ |
| EQR | $1 \geq \text{AMBI} \geq 0.83$ | $0.83 > \text{AMBI} \geq 0.53$ | $0.53 > \text{AMBI} \geq 0.39$ | $0.39 > \text{AMBI} \geq 0.21$ | $0.21 > \text{AMBI} \geq 0$ |
| BENTIX | $6 \geq \text{BENTIX} \geq 4.5$ | $4.5 > \text{BENTIX} \geq 3.5$ | $3.5 > \text{BENTIX} \geq 2.5$ | $2.5 > \text{BENTIX} \geq 2$ | $2 > \text{BENTIX} \geq 0$ |
| EQR | $1 \geq \text{BENTIX} \geq 0.75$ | $0.75 > \text{BENTIX} \geq 0.58$ | $0.58 > \text{BENTIX} \geq 0.42$ | $0.42 > \text{BENTIX} \geq 0.33$ | $0.33 > \text{BENTIX} \geq 0$ |
| MEDOCC | $0 \leq \text{MEDOCC} \leq 1.6$ | $1.6 < \text{MEDOCC} \leq 3.2$ | $3.2 < \text{MEDOCC} \leq 4.7$ | $4.7 < \text{MEDOCC} \leq 5.5$ | $5.5 < \text{MEDOCC} \leq 6$ |
| EQR | $1 \geq \text{MEDOCC} \geq 0.73$ | $0.73 > \text{MEDOCC} \geq 0.47$ | $0.47 > \text{MEDOCC} \geq 0.20$ | $0.20 > \text{MEDOCC} \geq 0.08$ | $0.08 > \text{MEDOCC} \geq 0$ |
| M-AMBI | $1 \geq \text{M-AMBI} \geq 0.83$ | $0.83 > \text{M-AMBI} \geq 0.62$ | $0.62 > \text{M-AMBI} \geq 0.41$ | $0.41 > \text{M-AMBI} \geq 0.20$ | $0.20 > \text{M-AMBI} \geq 0$ |
| EQR | $1 \geq \text{M-AMBI} \geq 0.83$ | $0.83 > \text{M-AMBI} \geq 0.62$ | $0.62 > \text{M-AMBI} \geq 0.41$ | $0.41 > \text{M-AMBI} \geq 0.20$ | $0.20 > \text{M-AMBI} \geq 0$ |

5. Statistical analyses

Scores of all biotic indices were calculated by using the Microsoft Excel software, except for M-AMBI, which was estimated by using the software at AZTI's web page (<http://www.azti.es>). After estimating the scores of the relative abundances of ecological groups (GI-GV), AMBI, S (number of species) and H', separately, an Excel file was constructed including these scores and then in the software, using the function of "go to M-AMBI", M-AMBI scores were calculated based on the Excel file prepared. The Pearson's moment correlation analysis was used to determine the correlations among biotic indices and environmental variables. The results of biotic indices were mapped using Surfer 11 and data were interpolated via the *kriging* method.

Results

1. Benthic samples from İzmir Bay

Faunistic analysis of seasonal samples taken at eight stations located in different parts of İzmir Bay in 2009 revealed a total of 427 macrobenthic species belonging to 11 taxonomic groups. The most dominant species in the area were *Aricidea claudiae* (8.7% of total number of specimens), *Streblospio gynobranchiata* (8.2%), *Levinsenia demiri* (7.8%), *Sternaspis scutata* (6.2%) and *Lumbrineris geldiaiyi* (5.2%) (Appendix 1).

According to the national database for ecological groups of species, Group 3 (opportunistic species, including GIV and GV) was represented by 29 species in the area. Percentages of three ecological groups (sensitive, tolerant and opportunistic species) of the metric 2 of TUBI varied considerably among

seasonal samples (Figure 3). Stations located in the outer parts of İzmir Bay had the lowest relative abundance of opportunistic species, whereas the station (station 24) in the inner part of the bay had the highest percentages of these species. At stations 4, 6, 11 and 17, the dominances of G1 were higher than 40%, except for fall samples taken from station 17, where G2 (tolerant species) dominated the benthic community. It is interesting to note that the polluted station (station 24) had higher percentages of G1 in summer samples, indicating a dynamic and fluctuated environment in this part of İzmir Bay. Except for some samples, winter samples had higher percentages of G1 at all stations. The highest percentages of tolerant species (G3) were encountered at station 22.

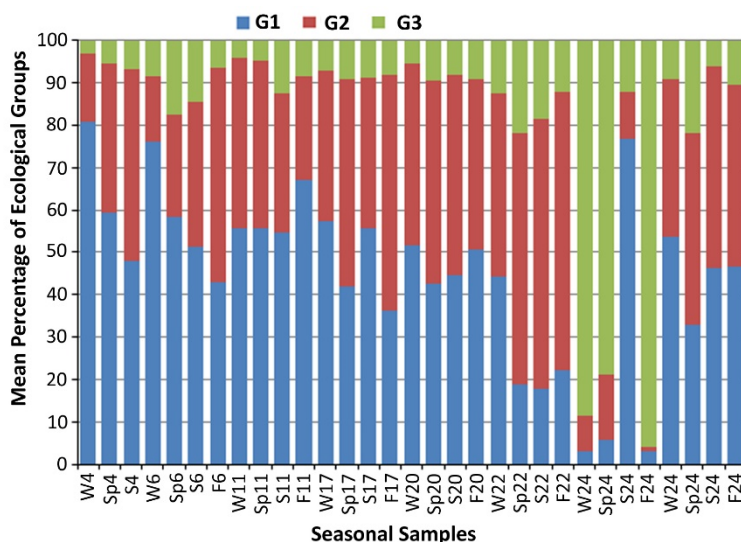


Figure 3. Mean percentages of the ecological groups G1 (including sensitive and indifferent species), G2 (tolerant species) and G3 (opportunistic species) at seasonal samples from İzmir Bay. W: Winter, Sp: Spring, S: Summer, F: Fall.

The mean values of TUBI in seasonal samples and the ecological status of stations are indicated in Figures 4 and 5. The standart error of the mean indicates little variations of TUBI values among replicates. In the area, three samples (station 24, fall) had the lowest TUBI scores, thus classifying the benthic quality status of the water body as bad, six samples (station 24, winter and spring) possessed TUBI scores that indicated poor ecological status. Moderate ecological status was rare in the area, but the majority of samples had TUBI values higher than 4, indicating good or high ecological status. Generally speaking, except for station 24, stations of İzmir Bay had good or high ecological status. TUBI values at station 22, which is situated between the outer and inner parts of the bay, were lower than those estimated at stations situated in the outer parts of the bay.

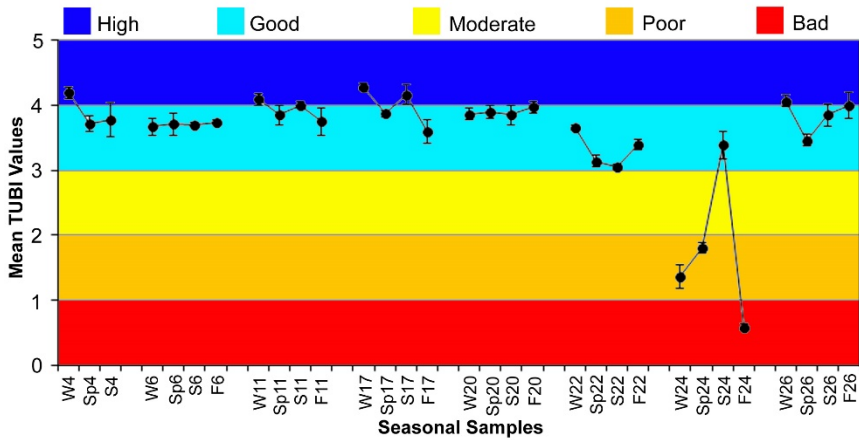


Figure 4. The mean values and \pm SE of the TUBI calculated seasonally at stations of İzmir Bay. W: Winter, Sp: Spring, S: Summer, F: Fall.

In Figure 5, the mean values of biotic indices (H' , AMBI, M-AMBI, BENTIX, MEDOCC and TUBI) and the benthic quality status at seasonal samples from İzmir Bay were compared. According to the scores of AMBI, M-AMBI and MEDOCC, there was no high benthic quality status in İzmir Bay, whereas H' , BENTIX and TUBI detected high ecological status at stations in the outer bay. The inner part of İzmir Bay had generally poor or bad ecological status in spring, fall and winter according to all biotic indices used in this study. However, the ecological status of summer samples were classified as good or moderate (H' and M-AMBI), except for BENTIX that recognized the ecological status of the area as high.

Figure 6 shows the distributions of scores of biotic indices among seasonal samples. Samples were mainly clustered at good ecological status, which were mainly assigned by AMBI, MEDOCC and TUBI. At high ecological status, the majority of scores belonged to H' . M-AMBI scores were mainly concentrated at moderate ecological status, where TUBI classified only one sample (station 22, summer) as moderate. The benthic quality status classified as poor by TUBI was also classified as poor or bad by other biotic indices, except for some stations which had moderate ecological status according to the H' scores.

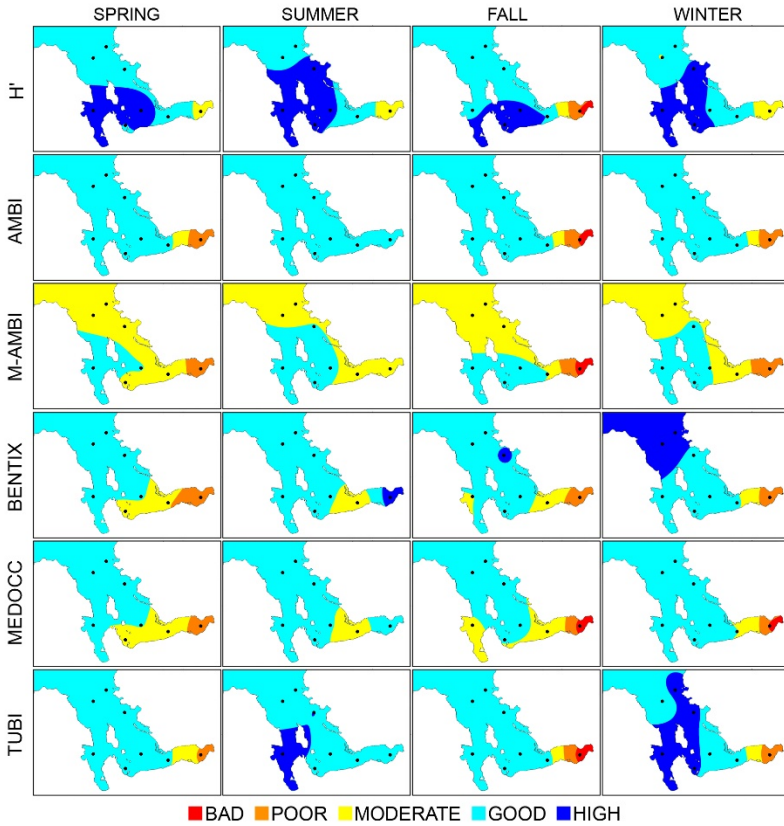


Figure 5. Seasonal benthic quality status of stations in İzmir Bay according to H', AMBI, M-AMBI, BENTIX, MEDOCC and TUBI

The correlation values and regression formulas between the ecological quality ratios (EQR) of TUBI and those of other biotic indices were indicated in Figure 7. The correlations between biotic indices were significant ($p < 0.05$) and positive. The highest correlation values ($r > 0.90$) were estimated between TUBI, and AMBI ($r = 0.93$) and M-AMBI ($r = 0.91$), the lowest values between TUBI and BENTIX ($r = 0.75$) (Figure 7). The EQRs of MEDOCC were well correlated with the lowest scores of EQRs of TUBI, but those of H' were well correlated with the highest scores of EQRs of TUBI.

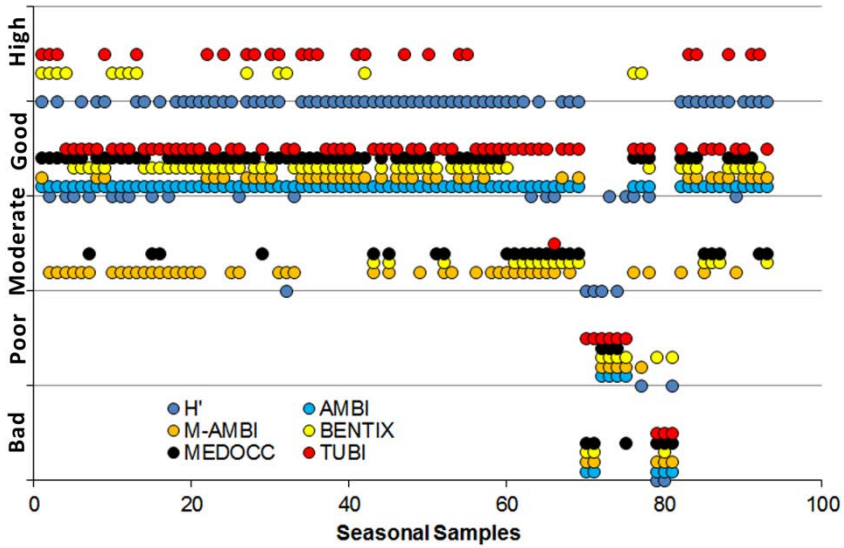


Figure 6. Distributions of samples to different ecological status according to scores of biotic indices

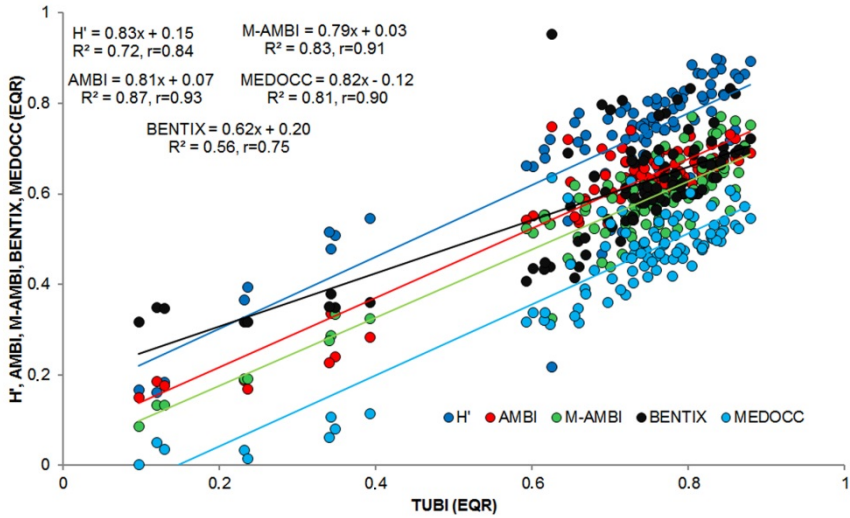


Figure 7. Correlations between ecological quality ratio (EQR) of TUBI and those of other biotic indices (H', AMBI, M-AMBI, BENTIX and MEDOCC).

The efficiency of the biotic indices was tested with the environmental variables in İzmir Bay, especially with the total organic carbon concentrations (TOC) in sediment and total inorganic nitrogen concentrations (TIN) in deep water. The

correlation values between the biotic indices and TOC were negative and significant (Figure 8). The highest correlation value was found between TUBI and TOC ($r = -0.77, p < 0.05$), and the lowest between BENTIX and TOC ($r = -0.48$) (Figure 8). TUBI and H' had the highest correlation value ($r = -0.77$) with TIN, whereas BENTIX and MEDOCC had weak but significant correlations with TIN (Figure 9). The silica concentration in deep water was also negatively and significantly correlated with all biotic indices, but the highest correlation values were found between silica, and TUBI ($r = -0.73$) and H' ($r = -0.70$). The correlation analysis performed between the dissolved oxygen concentrations (DOC) in deep water and biotic indices indicated that there was a positive and significant correlation between DOC and, H' ($r = 0.37, p < 0.05$), M-AMBI ($r = 0.32, p < 0.05$) and TUBI ($r = 0.30, p < 0.05$), and a positive but insignificant correlation between between DOC and, BENTIX ($r = 0.11, p > 0.05$), MEDOCC ($r = 0.12, p > 0.05$) and AMBI ($r = 0.14, p > 0.05$).

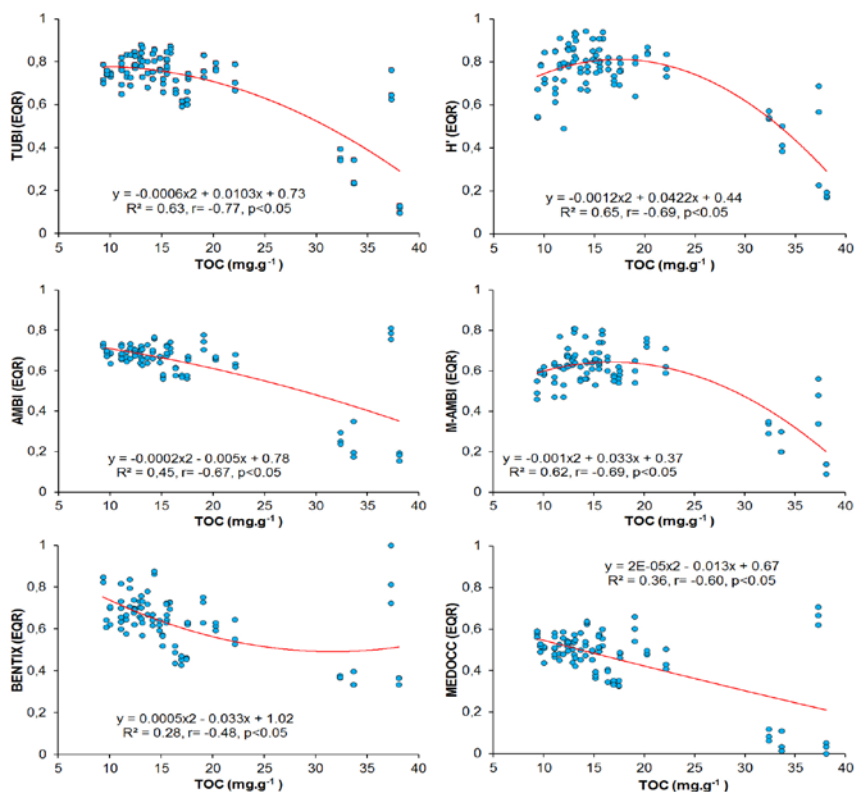


Figure 8. Correlations between total organic carbon (TOC) concentrations in sediment and ecological quality ratios (EQRs) of the biotic indices

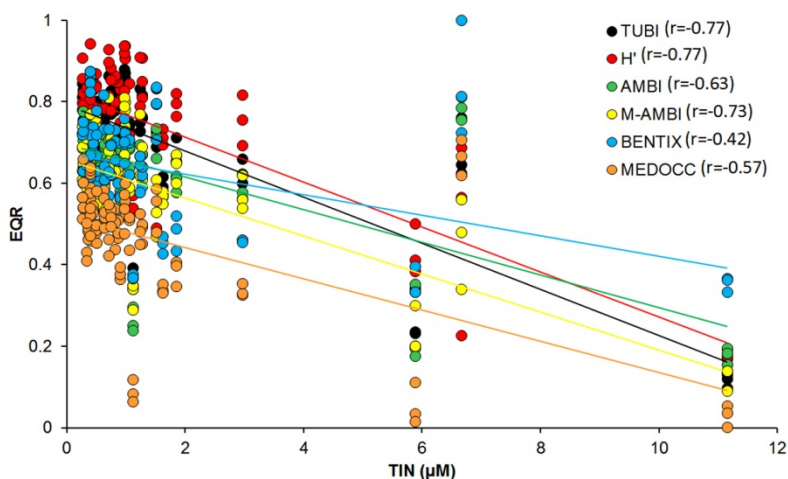


Figure 9. Correlations between total inorganic nitrogen (TIN) concentrations in deep water and the ecological quality ratios (EQRs) of biotic indices

2. Benthic samples from the Aegean and Levantine Seas

Faunistic analysis of benthic samples collected along the Aegean Sea revealed a total of 520 species belonging to 15 systematic groups (Appendix 1). Among the species, *Corbula gibba* (5.2% of total number of specimens), *Bittium reticulatum* (4.5%), *Lumbrineris geldiyi* (4%) and *Streblospio gynobranchiata* (3.3%) were the most dominant species, whereas *Monticellina heterochaeta* (present in 69% of samples), *L. geldiyi* (69%), *C. gibba* (66%), *Turritella communis* (55%) and *Leptochelia savignyi* (52%) were the most frequent species in the area. In the Aegean Sea, a total of 22 species belonged to the ecological group 3, of which *Polydora cornuta*, *S. gynobranchiata* and *Prionospio pulchra* were invasive alien species, dominating the station near the Alsancak Harbour located in the polluted inner part of İzmir Bay.

At stations along the Levantine coast of Turkey, 315 species belonging to 11 systematic groups were encountered, of which *Owenia fusiformis* (14.4% of the total number of specimens), *Sigambra tentaculata* (7%), *Bittium latreilli* (5%), *Monticellina heterochaeta* (4.4%) and *Prionospio depauperata* (4%) were the dominant species (Appendix 1). The most frequent species in the area were *P. depauperata* (present in 46% of samples), *Lumbrineris geldiyi* (42%), *Glycinde bonhourei* (42%), *S. tentaculata* (42%) and *Leptochela pugnax* (38%). In the area, 19 opportunistic species were found, of which *Ophiodromus pallidus*, *Schistomeringos rudolphi*, *Prionospio fallax*, *Pseudopolydora paucibranchiata*, *Heteromastus filiformis* and *Jassa marmorata* belongs to the first-order of opportunistic species.

The benthic quality status of stations in the Aegean and Levantine Seas were variable according to different biotic indices used in the present study (Figure 10). According to TUBI, there was no station representing bad ecological status, but one station from İskenderun Bay was in poor status and four stations from İskenderun and İzmir Bays had moderate status. The other stations had good or high ecological status. H' gave a more optimistic result from the area, determined only good or high ecological status at all stations of the Aegean Sea, whereas it detected 10 stations with moderate ecological status and one station with poor ecological status in the Levantine Sea. In contrast, M-AMBI produced scores indicating moderate ecological status at the majority of stations in the Aegean and Levantine Seas, and detected poor ecological status in the inner parts of İzmir and İskenderun Bays. The ecological status of some stations in İskenderun Bay was only recognized as poor by M-AMBI and TUBI.

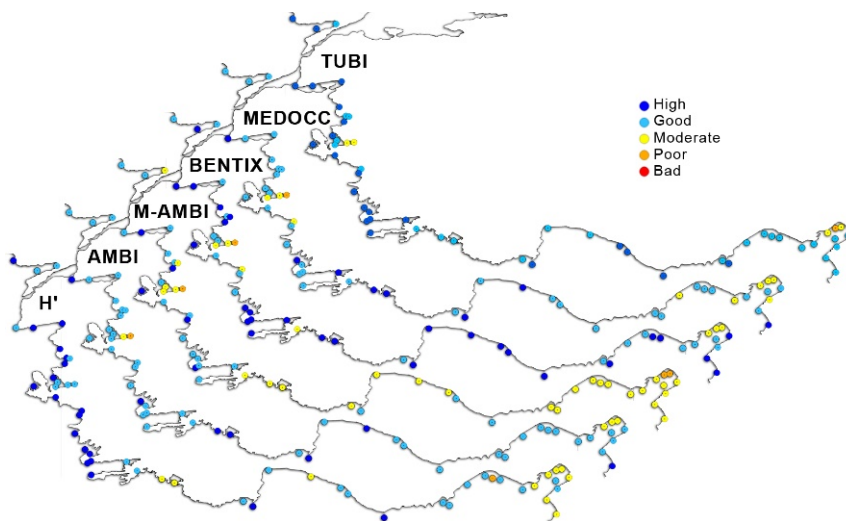


Figure 10. Benthic quality status of stations in the Aegean and Levantine Seas according to H' , AMBI, M-AMBI, BENTIX, MEDOCC and TUBI.

The correlation values among biotic indices and the number of species (S) estimated at stations of the Aegean and Levantine Seas are indicated in Table 4. TUBI and M-AMBI were positively and significantly correlated with S and other biotic indices. However, TUBI was relatively well correlated with M-AMBI ($r=0.84$), AMBI ($r=0.77$) and MEDOCC ($r=0.76$), whereas M-AMBI was well correlated with S ($r=0.90$) and H' ($r=0.92$).

Table 4. Correlation between biotic indices and the number of species (S) estimated at stations from the Aegean and Levantine Seas. Values in bold are statistically significant ($p < 0.05$).

| | H' | AMBI | M-AMBI | BENTIX | MEDOCC | TUBI |
|---------------|-------------|-------------|---------------|---------------|---------------|-------------|
| S | 0.83 | 0.11 | 0.90 | 0.09 | 0.10 | 0.57 |
| H' | | 0.15 | 0.92 | 0.03 | 0.14 | 0.71 |
| AMBI | | | 0.42 | 0.91 | 0.99 | 0.77 |
| M-AMBI | | | | 0.32 | 0.41 | 0.84 |
| BENTIX | | | | | 0.92 | 0.69 |
| MEDOCC | | | | | | 0.76 |

Discussion

The present study proposed a new biotic index, TUBI (TURKISH Benthic Index), to assess the ecological status of water bodies based on the macro-zoobenthic assemblages of the soft bottom environments. The performance of this index has not been tested in hard substrata yet. TUBI have two metrics; the Shannon-Weaver's diversity index and the relative abundance of ecological groups. The second metric has been widely used in many biotic indices, including BENTIX, AMBI and MEDOCC. However, the assessment of ecological groups in formulas varies among them. BENTIX re-categorized the ecological groups under three groups, namely sensitive (Group I and Group II), tolerant (GIII and GIV) and the first order of opportunistic species (GV). However, it takes two major groups [GS (sensitive species, including GI and GII) and GT (tolerant species, including GIII-GV)] into account in the calculation (Simboura and Zenetos 2002). The ecological groups are considered separately in AMBI and have different weights in the formula (Borja *et al.* 2000). MEDOCC is also similar to AMBI, but combines the first and second orders of opportunistic species into one group (GIV) and gives different weights to ecological groups in formula. In TUBI, the number of ecological groups is decreased from five to three (sensitive, tolerant and opportunistic species) like BENTIX, but differs from it in the following ways; 1) TUBI estimates GIII (tolerant species) separately, whereas BENTIX combines its scores with the opportunistic species; 2) TUBI eliminates the percent abundance of sensitive species, gives moderate weight to G2 and high weight to G3 in the calculation, whereas BENTIX gives a high weight to the sensitive and indifferent species (GI and GII) and low weight to GIII-GV. This structure in BENTIX does not allow classifying water bodies as bad ecological status unless azoic zone is developed, whereas other biotic indices including TUBI can recognize water bodies with bad ecological status if opportunistic species heavily dominate the area. Decreasing ecological groups from five to three in TUBI has some advantages, mainly simplifying the calculation. In addition to this, the transition between some ecological groups is not sharp, hindering us to take an accurate decision in the assignments of species to the ecological groups. In the TUBI estimation, the first and second orders of opportunistic species were put into one large category (G3) as it is hard to categorize species as the first order or the second order. Borja *et al.*

(2000) explained that the group IV includes small-sized polychaetes which are sub-surface deposit feeders such as cirratulids and that the group V comprises surface deposit feeders that proliferate in reduced sediments. However, most of cirratulids and spionids that were attained to group IV in the AMBI database are in fact surface deposit-feeders, with palps or tentacles fully extending over sediments to capture detritic material from ambient waters (Fauchald and Jumars 1979). Similarly, the separation of sensitive and indifferent species is not easy, because of insufficient knowledge about the biological and ecological features of many marine species. Therefore, combining these categories into major groups would decrease a possible wrong judgement related to the assignment of species to the ecological groups, thus would increase the reliability of the index to reflect the benthic quality status.

Among indices used in the present study, only M-AMBI is a multi-metric index. It combines the scores of the number of species, and H' and AMBI values of samples, and produces a score along a gradient from the highest and lowest scores by using factor analysis. As it uses more than one metric, it represents some advantages to reflect the ecological status of benthic environments in relation to the magnitude of deteriorations (Bigot *et al.* 2008; Borja *et al.* 2008) and has been proposed in some countries as a main index to assess the ecological quality status of water bodies. However, it is hard, sometimes impossible to calculate this index without using its software developed by AZTI. Therefore, users strictly bound to the AMBI's database for ecological groupings of species and cannot use the regional or national database to calculate M-AMBI scores or change the ecological groups of species when new data are accumulated for their life-history traits. In TUBI, there are two metrics and the estimation is based on the equal contribution of these metrics. However, this new index fixes the maximum score of H' at the score 5 in order to synchronize it with the other metric which varies from 0 to 5, and to restrict its weight in the estimation as it gets high scores in ecotone points, mixed sediments and even in samples which have abundance data homogeneously distributed to the low number of species. In M-AMBI, the maximum and minimum scores of the diversity index should be specified for the region and the high scores of H' change the overall estimation of the index.

The benthic quality status of stations in Izmir Bay was evaluated differently by the biotic indices. For example, M-AMBI and MEDOCC did not detect any high ecological status in the area, whereas H' and TUBI recognized high benthic quality status at stations located in the outer bay especially in summer and winter. Bad ecological status was only determined at station 24 in Izmir Bay, but in summer this station had relatively high index scores and its ecological status was classified as good or moderate. However, according to the BENTIX scores, this station possessed a high ecological status in summer. It was mainly attributed to the fact that BENTIX gave a higher weight to the sensitive species (including GI and GII) rather than to the tolerant species in its formula, so as the

summer samples had lesser number of species (<17 species), but had a high proportion of GII, the ecologic status of the benthic area was classified as high by BENTIX. This shows that if samples have a few number of species but a high percentage of GII, BENTIX leads to a wrong judgement. The other indices including AMBI and MEDOCC eliminate scores of GI in their calculations. Like BENTIX, TUBI creates a score by adding the scores of GI and GII in samples, but does not take it into account in the estimation. If a sample has a few number of species composed of GI and GII, the metric 2 takes 0, but H' detects the paucity in the species numbers and produces a low score, indicating bad or poor ecological status. Therefore, TUBI has two controls (metrics) in defining the ecological status of water bodies. At stations along the Aegean and Levantine coasts of Turkey, a similar pattern was encountered. The ecological status of the majority of stations was classified as moderate by M-AMBI and as high by H'. A consensus was reached among indices, except for H', that samples from the inner parts of İzmir and İskenderun Bays had moderate or poor ecological status. Unlike other indices, BENTIX and TUBI classified the ecological status of many samples as high in the area.

TOC (Total Organic Carbon) in sediments due to organic pollution has been used as an indicator for marine benthic quality (Hyland, *et al.* 2005; Çınar *et al.* 2012a). In İzmir ($r = -0.54$) and Mersin ($r = -0.75$) Bays, a negative but relatively high correlation was found between TOC and H'. TOC becomes toxic to benthic invertebrates at concentrations over 35 mg.g^{-1} due to the deoxygenating effect of organic matter (Hyland *et al.* 2005). However, at low concentrations, like below 10 mg.g^{-1} , the benthic invertebrates cannot be much affected. In the present study, TOC concentrations varied from 9.3 (station 6) to 38 (station 24) mg.g^{-1} at stations of Izmir Bay and bad ecological status was determined at TOC concentrations above 32 mg.g^{-1} . However, there is a discontinuity in the TOC concentrations between 32 and 22 mg.g^{-1} , so the ecotone point where scores of indices drop significantly from good to moderate or moderate to poor ecological status has not been determined, as the biotic indices indicated good or high ecological status at TOC concentration of 22 mg.g^{-1} (station 17, summer). The indices that represented a high correlation with TOC were H', TUBI and M-AMBI. Magni (2003) found that H' was strongly correlated with TOC concentrations following a polynomial function and determined an initial increase of the curve at TOC concentrations around 10 mg.g^{-1} , followed by a marked decrease at TOC concentrations around 35 mg.g^{-1} . The increase in TIN (Total Inorganic Nitrogen) concentrations in deep water decreased the scores of all biotic indices in İzmir Bay and a sharp decrease was observed at TIN concentrations above $3 \text{ }\mu\text{M}$. Among the indices, TUBI was negatively but significantly correlated with TIN, indicating its power to represent the ecological quality status of benthic environments adequately.

In the present study, the biotic indices were estimated by using a similar database of ecological groups of species, and therefore the correlations among

the biotic indices (BENTIX, AMBI and MEDOCC) with one metric were found to be high ($r>0.90$). The scores of AMBI was strongly correlated with those of MEDOCC ($r=0.99$) in the area. In contrast, low correlations ($r<0.40$) were reported between BENTIX and AMBI when different databases were used (Ponti *et al.* 2008; Simonini *et al.* 2009). The difference between the AMBI's and BENTIX's databases is obvious. For example, the polychaete species *Pectinaria koreni* is a sensitive species according to the BENTIX database, whereas it is a second-order opportunistic species according to the AMBI database. Similarly, all *Ancistrosyllis* and *Praxillella* species were classified as sensitive species by the BENTIX database, whereas they were considered as tolerant species by the AMBI database. If these species dominate a benthic habitat, the evaluation of its benthic quality status by these indices differs significantly.

TUBI, with two metrics, reflected well the actual status of benthic ecological status of the eastern Mediterranean sites in accordance with the environmental variables such as TOC and TIN values. This index discriminated stations according to different levels of degradation, therefore better confirming variability across a gradient of human-mediated impacts. Based on this study, we recommend to use TUBI to assess and monitor impacts of organic pollution on benthic communities. However, further studies are required to properly evaluate its strengths and weaknesses on a larger scale.

TUBI (Türk Bentik İndeks): Bentik komuniteler üzerine organik kirliliğin etkilerini belirlemek amacıyla yeni bir biyotik indeks

Özet

Bentik komünite yapıları üzerine organik zenginleşmenin etkilerini belirlemek amacıyla yeni bir biyotik indeks olan TUBI (Türk Bentik İndeks) bu çalışmada önerilmiştir. Bu yeni indeks 2 metriğe sahiptir; Shannon-Weaver çeşitlilik indeksi (metrik 1) ve ekolojik grupların nisbi bollukları (metrik 2). Toplam 5 kategori içeren türlerin ekolojik grupları bu çalışmada, duyarlı türler (GI ve GII'yi içerir), toleranslı türler (GIII) ve fırsatçı türler (GIV ve GV) olmak üzere 3 ana kategori altında toplanmıştır. Metrik 2 bu grupları farklı ağırlıklarda ele almakta ve hesaplamalarında duyarlı türleri gözardı etmektedir. TUBI değerleri 0 ile 5 arasında değişmekte ve bentik kalite durumu, TUBI değerleri arttıkça artmaktadır. İzmir Körfezi, Ege ve Levanten Denizi'nde toplanan bentik örnekler, türlerin ekolojik grupları için hazırlanan ulusal bir veritabanı kullanılarak AMBI, M-AMBI, BENTIX, MEDOCC ve TUBI gibi çeşitli biyotik indeksler kullanılarak analiz edilmiştir. Bazı istisnalar hariç, kullanılan tüm biyotik indeksler bölgelerdeki kötü ve çok kötü ekolojik durumları belirlemiştir. Sedimentteki toplam organik karbon ve dip suyundaki toplam inorganik azot ile biyotik indeksler arasında yapılan korelasyon analizi, TUBI'nin bu çevresel değişkenlerle en yüksek negatif korelasyon değerlerine sahip olduğunu göstermiştir. Bu nedenle bu indeks bentik

kommuniteleri üzerine kirlilik kaynaklı etkilerin gradasyonu boyunca oluşan deęişimleri daha iyi belirlemektedir.

Acknowledgments

This work has been financially supported by TUBITAK Project (SINHA 107G066) and by the Ministry of Environment and Urbanization/General Directorate of Environmental Management (Integrated pollution monitoring project in Aegean and Mediterranean Seas, coordinated by Derinsu Underwater Engineering).

References

Albayrak, S., Balkıs, N., Balkıs, H., Zenetos, A., Kurun, A., Karhan, S.Ü., Çaęlar, S., Balcı, M. (2010) Golden Horn Estuary: description of the ecosystem and an attempt to assess its ecological quality status using various classification metrics. *Med. Mar. Sci.* 11: 295-313.

Bigot, L., Grémare, A., Amouroux, J.-M., Frouin, P., Maire, O., Gaertner, J.C. (2008) Assessment of the ecological quality status of soft-bottoms in Reunion Island (tropical Southwest Indian Ocean) using AZTI marine biotic indices. *Mar. Pollut. Bull.* 56: 704-722.

Blanchet, H., Lavesque, N., Ruellet, T., Dauvin, J.C., Sauriau, P.G., Desroy, N., Desclaux, C., Leconte, M., Bachelet, G., Janson, A.L., Bessineton, C., Duhamel, S., Jourde, J., Mayot, S., Simon, S., Montaudouin, X. (2008) Use of biotic indices in semi-enclosed coastal ecosystems and transitional waters habitats-Implications for the implementations of the European Water Framework Directive. *Ecological Indicators* 8: 360-372.

Borja, A., Bricker, S.B., Dauer, D.M., Demetriades, N.T., Ferreira, J.G., Forbes, A.T., Hutchings, P., Kenchington, R., Marques, J.C., Zhu, C. (2008) Overview of integrative tools and methods in assessing ecological integrity in estuarine and coastal systems worldwide. *Mar. Pollut. Bull.* 56: 1519-1537.

Borja, A., Franco, J., Pérez, V. (2000) A marine biotic index to establish the ecology quality of soft-bottom benthos within European estuarine coastal environments. *Mar. Pollut. Bull.* 40: 1100-1114.

Borja, A., Miles, A., Occhipinti-Ambrogi, A., Berg, T. (2009) Current status of macroinvertebrate methods used for assessing the quality of European marine waters: implementing the Water Framework Directive. *Hydrobiologia* 633: 181-196.

Çınar, M.E., Bakir, K. (2014). Alien Biotic Index (ALEX) - a new index for assessing impacts of alien species on benthic communities. *Mar. Poll. Bull.* 87: 171-179.

Çinar, M.E., Dagli, E. (2013) Polychaetes (Annelida: Polychaeta) from the Aegean and Levantine coasts of Turkey, with descriptions of two new species. *J. Nat. Hist.* 47: 911-947.

Çinar, M.E., Katagan, T., Öztürk, B., Bakir, K., Dagli, E., Açık, S., Dogan, A., Bitlis, B. (2012) Spatio-temporal Distributions of zoobenthos in soft substratum of Izmir Bay (Aegean Sea, eastern Mediterranean), with special emphasis on alien species and ecological quality status. *J. Mar. Biol. Ass. U.K.* 92: 1457-1477.

Çinar, M. E., Katagan, T., Öztürk, B., Egemen, Ö., Ergen, Z., Kocatas, A., Önen, M., Kirkim, F., Bakir, K., Kurt, G., Dagli, E., Kaymakçı, A., Açık, S., Dogan, A., Özcan, T. (2006) Temporal changes of soft bottom zoobenthic communities in and around Alsancak Harbor (Izmir Bay, Aegean Sea), with special attention to the autoecology of exotic species. *Mar. Ecol.* 27: 229-246.

Dauvin, J.-C., Ruellet, T., Desroy, N., Janson, A-L. (2007) The ecological quality status of the Bay of Seine and the Seine estuary: use of biotic indices. *Mar. Pollut. Bull.* 55: 241-257.

Dean, H.K. (2008) The use of polychaetes (Annelida) as indicator species of marine pollution: a review. *Rev. Biol. Trop.* 56: 11-38.

Dekos (2014) Marine and Coastal Water Quality Determination and Classification of Project Cases (DeKoS). CTUE 5118703, Report No. CTUE. 13.155 (final report), February 2014, Gebze-Kocaeli. (in Turkish).

Ergen, Z., Çinar, M.E., Dağlı, E., Kurt, G. (2006) Seasonal dynamics of soft-bottom polychaetes in Izmir Bay (Aegean Sea, eastern Mediterranean). *Sci. Mar.* 70S3: 197-207.

Fauchald, K., Jumars, P. A. (1979) The diet of worms: a study of polychaete feeding guilds. *Oceanogr. Mar. Biol. Ann. Rev.* 17: 193-284.

Glémarec, M. (1986) Ecological impact of an oil-spill: utilisation of biological indicators. IAWPRC-NERC Conference, July 1985. *IAWPRC Journal* 18: 203-211.

Grassle, J.F., Grassle, J.P. (1974) Opportunistic life histories and genetic systems in marine benthic polychaetes. *J. Mar. Res.* 32: 253-284.

Heip, C. (1995) Eutrophication and zoobenthos dynamics. *Ophelia* 41: 113-136.

Hyland, J., Balthis, L., Karakassis, I., Magni, P., Petrov, A., Shine, J., Vestergaard, O., Warwick, R. (2005) Organic carbon content of sediments as an indicator of stress in the marine benthos. *Mar. Ecol. Prog. Ser.* 295: 91-103.

Kocatas, A., Ergen, Z., Katagan, T. (1985) Changes in the benthic communities due to various pollutants in İzmir Bay (Turkey). Meeting on the Effects of Pollution on Marine Environments, Blanes, FAO, 9: 1-19.

Magni, P. (2003) Biological benthic tools as indicators of coastal marine ecosystems health. *Chemistry and Ecology* 19: 363-372.

Marques, J.C., Salas, F., Patricio, J., Teixeira, H., Neto, J.M. (2009) Ecological Indicators for Coastal and Estuarine Environmental Assessment. A User Guide. WIT Press, Boston, 183 pp.

Mixuka, I., Borja, A., Bald, J. (2007) Using historical data, expert judgement and multivariate analysis in assessing reference conditions and benthic ecological status, according to the European water framework directive. *Mar. Pollut. Bull.* 55: 16-29.

Pianka, E.R. (1970) On r- and K- Selection. *Amer. Natur.* 104: 592-597.

Pinedo, S., Jordana, E., Salas, F., Subida, M.D., García Adiego, E., Torres, J. (2012) Testing MEDOCC and BOPA indices in shallow soft-bottom communities in the Spanish Mediterranean coastal waters. *Ecological Indicators* 19: 98-105.

Pocklington, P., Wells, P.G. (1992) Polychaetes key taxa for marine environmental quality monitoring. *Mar. Pollut. Bull.* 24: 593-598.

Ponti, M., Pinna, M., Basset, A., Moncheva, S., Trayanova, A., Georgescu, L.P., Beqiraj, S., Orfanidis, S., Abbiati, M. (2008) Quality assessment of Mediterranean and Black Sea transitional waters: comparing responses of benthic biotic indices. *Aquat. Conserv.: Mar. Freshwat. Ecosyst.* 18: 62-75.

Rice, J., Arvanitidis, C., Borja, A., Frid, C., Hiddink, J., Krause, J., Lorange, P., Ragnarsson, S.A., Sköld, M., Trabucco, B. (2010) Marine Strategy Framework Directive. Task Group 6 Report. Seafloor Integrity. JRC Scientific and Technical Reports, 73 pp.

Rosenberg, R. (1976) Benthic faunal dynamics during succession following pollution abatement in a Swedish estuary. *Oikos* 27: 414-427.

Simboura, N., Zenetos, A. (2002) Benthic indicators to use in ecological quality classification of Mediterranean soft bottoms marine ecosystems, including a new biotic index. *Medit. Mar. Sci.* 3: 77-111.

Simboura, N., Zenetos, A., Pancucci-Papadopoulou, M.A. (2014) Benthic community indicators over a long period of monitoring (2000-2012) of the Saronikos Gulf, Greece, eastern Mediterranean. *Environ. Monit. Assess.* 186: 3809-3821.

Simonini, R., Grandi, V., Massamba-N'Siala, G., Iotti, M., Montanari, G., Perevedelli, D. (2009) Assessing the ecological status of the north-western Adriatic within the European Water Framework Directive: a comparison of Bentix, AMBI and M-AMBI methods. *Mar. Ecol.* 30: 241-254.

Tsutsumi, H., Fukunaga, S., Fujita, N., Sumida, M. (1990) Relationship between growth of *Capitella* sp. and organic enrichment of the sediment. *Mar. Ecol. Prog. Ser.* 63: 157-162.

Van Hoey, G., Borja, A., Birchenough, S., Buhl-Mortensen, L., Degraer, S., Fleischer, D., Kerckhof, F., Magni, P., Muxika, I., Reis, H., Schröder, A., Zettler, M.I. (2010) The use of benthic indicators in Europe: from the Water Framework Directive to the Marine Strategy Framework Directive. *Mar. Pollut. Bull.* 60: 2187-2196.

Vrijenhoek, R.C. (1979) Factors affecting clonal diversity and coexistence. *Amer. Zool.* 19: 787-797.

Received: 14.05.2015

Accepted: 01.06.2015

Appendix 1. Species found in the present study and their total abundances in İzmir Bay (Iz), the Aegean Sea (Ag) and Levantine Sea (Le), and the ecological groups (EG) (GI-GV) they were assigned to. TUBI includes three ecological groups; G1 (GI+GII), G2 (GIII) and G3 (GIV+GV).

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|-----|----|-----|--|-----|-----|----|----|
| <i>Abra alba</i> (W. Wood, 1802) | III | 106 | 10 | - | <i>Mangelia attenuata</i> (Montagu, 1803) | I | 8 | 6 | - |
| <i>Abra nitida</i> (O. F. Müller, 1776) | III | 30 | - | - | <i>Mangelia costata</i> (Pennant, 1777) | I | - | 5 | - |
| <i>Abra prismatica</i> (Montagu, 1808) | III | - | 22 | 8 | <i>Mangelia costulata</i> Risso, 1826 | I | 9 | 4 | 2 |
| <i>Acanthocardia paucicostata</i> (G. B. Sowerby II, 1834) | II | 1 | 2 | - | <i>Mangelia fieldeni</i> (van Aartsen & Fehr-de Wal, 1978) | I | - | 1 | - |
| <i>Acanthocardia tuberculata</i> (Linnaeus, 1758) | I | - | 1 | - | <i>Mangelia stosciana</i> Brusina, 1869 | I | - | 2 | - |
| <i>Acteon tornatilis</i> (Linnaeus, 1758) | I | 1 | 5 | 1 | <i>Mangelia unifasciata</i> (Deshayes, 1835) | II | 10 | 13 | 3 |
| <i>Aglaophamus agilis</i> (Langerhans, 1880) | I | - | 1 | - | <i>Manzonina crassa</i> (Kammacher, 1798) | I | 1 | - | - |
| <i>Alitta succinea</i> (Leuckart, 1847) | V | 5 | 5 | - | <i>Marphysa bellii</i> (Audouin & Milne Edwards, 1833) | II | 31 | 32 | - |
| <i>Alpheus glaber</i> (Olivi, 1792) | II | 4 | 3 | 2 | <i>Marphysa cinari</i> Kurt-Sahin, 2014 | I | - | 1 | - |
| <i>Alvania cancellata</i> (da Costa, 1778) | I | - | 1 | - | <i>Marphysa fallax</i> Marion & Bobretzky, 1875 | I | - | 2 | - |
| <i>Alvania cimex</i> (Linnaeus, 1758) | I | - | 5 | 1 | <i>Marphysa sanguinea</i> (Montagu, 1815) | II | 1 | - | - |
| <i>Alvania colossophilus</i> Oberling, 1970 | I | - | 2 | 4 | <i>Marshallora adversa</i> (Montagu, 1803) | I | 2 | 1 | - |
| <i>Alvania geryonia</i> (Nardo, 1847) | I | 3 | 39 | - | <i>Medicorophium aculeatum</i> (Chevreux, 1908) | III | 10 | - | - |
| <i>Alvania punctura</i> (Montagu, 1803) | I | - | 3 | - | <i>Medicorophium runcicorne</i> (Della Valle, 1893) | III | 6 | 2 | - |
| <i>Alvania testae</i> (Aradas & Maggiore, 1844) | I | - | 3 | - | <i>Mediomastus cirripes</i> Ben-Eliahu, 1976 | IV | 14 | 3 | 91 |
| <i>Ampelisca brevicornis</i> (Costa, 1853) | I | - | 9 | 140 | <i>Megalomma vesiculosum</i> (Montagu, 1815) | I | - | 1 | - |
| <i>Ampelisca diadema</i> (Costa, 1853) | II | 2 | - | - | <i>Megaluropus massiliensis</i> Ledoyer, 1976 | I | - | 1 | 9 |
| <i>Ampelisca jaffaensis</i> Bellan-Santini & Kaim-Malka, 1977 | I | 22 | - | - | <i>Megastomia conoidea</i> (Brocchi, 1814) | III | 85 | 16 | 7 |
| <i>Ampelisca multispinosa</i> Bellan-Santini & Kaim-Malka, 1977 | I | - | 14 | 6 | <i>Melinna palmata</i> Grube, 1870 | III | 62 | 29 | 5 |
| <i>Ampelisca pseudosarsi</i> Bellan-Santini & Kaim-Malka, 1977 | II | - | 10 | 2 | <i>Melita valesi</i> Karaman, 1955 | I | - | - | 15 |
| <i>Ampelisca pseudospinimana</i> Bellan-Santini & Kaim-Malka, 1977 | I | 3 | - | - | <i>Metaphoxus simplex</i> (Bate, 1857) | I | 11 | 10 | 5 |
| <i>Ampelisca ruffoi</i> Bellan-Santini & Kaim-Malka, 1977 | I | - | 9 | 2 | <i>Metaxia metaxa</i> (Delle Chiaje, 1828) | I | - | - | 1 |
| <i>Ampelisca sarsi</i> Chevreux, 1888 | II | 1 | - | - | <i>Microdeutopus versiculatus</i> (Bate, 1856) | II | - | 9 | - |
| <i>Ampelisca tenuicornis</i> Lilljeborg, 1855 | I | 86 | - | - | <i>Microjassa cumbrensis</i> (Stebbing & Robertson, 1891) | I | - | 6 | - |
| <i>Ampelisca truncata</i> Bellan-Santini & Kaim-Malka, 1977 | I | - | 35 | 14 | <i>Micronephthys stammeri</i> (Augener, 1932) | III | 107 | 10 | 2 |
| <i>Ampelisca typica</i> (Bate, 1856) | I | 73 | - | - | <i>Microspio mecznikowianus</i> (Claparède, 1869) | III | 1 | 1 | - |
| <i>Ampharete acutifrons</i> (Grube, 1860) | II | 3 | - | - | <i>Mimachlamys varia</i> (Linnaeus, 1758) | I | - | 2 | - |
| <i>Ampharete octocirrata</i> (Sars, 1835) | II | - | 3 | - | <i>Mitrella gervillii</i> (Payraudeau, 1826) | I | - | 1 | - |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|---|-----|-----|----|----|---|-----|-----|-----|-----|
| <i>Amphicteis gunneri</i> (M. Sars, 1835) | II | 1 | 9 | - | <i>Mitromorpha olivoidea</i> (Cantraine, 1835) | I | - | - | 1 |
| <i>Amphictene auricoma</i> (O.F. Müller, 1776) | I | 2 | - | - | <i>Modiolula phaseolina</i> (Philippi, 1844) | I | - | 2 | 1 |
| <i>Amphictene auricoma</i> (O.F. Müller, 1776) | I | - | 1 | - | <i>Moerella distorta</i> (Poli, 1791) | III | 27 | 2 | - |
| <i>Amphiglena mediterranea</i> (Leydig, 1851) | I | - | - | 3 | <i>Monocorophium acherusicum</i> (Costa, 1853) | III | 6 | - | - |
| <i>Amphiodia obtecta</i> Mortensen, 1940 | III | - | - | 1 | <i>Monodaes couchii</i> (Couch, 1851) | I | - | 1 | - |
| <i>Amphipholis squamata</i> (Delle Chiaje, 1828) | I | 9 | 12 | 1 | <i>Monticellina dorsobranchialis</i> (Kirkegaard, 1959) | III | - | 6 | 159 |
| <i>Amphitrite cirrata</i> (Müller, 1771 in 1776) | I | - | 3 | - | <i>Monticellina heterochaeta</i> Laubier, 1961 | IV | 458 | 143 | 35 |
| <i>Amphiura chiajei</i> Forbes, 1843 | III | 38 | 30 | 6 | <i>Monticellina tessellata</i> (Hartman, 1960) | III | - | - | 2 |
| <i>Amphiura filiformis</i> (O.F. Müller, 1776) | III | 16 | 11 | - | <i>Musculus costulatus</i> (Risso, 1826) | I | - | 5 | - |
| <i>Amphioe ramondi</i> Audouin, 1826 | III | 4 | 1 | - | <i>Musculus subpictus</i> (Cantraine, 1835) | I | - | 4 | - |
| <i>Anapagurus bicorniger</i> A. Milne-Edwards & Bouvier, 1892 | II | 1 | - | - | <i>Myrianida brachycephala</i> (Marenzeller, 1874) | I | 1 | - | - |
| <i>Anapagurus petiti</i> Dechancé & Forest, 1962 | I | 1 | - | - | <i>Myrianida langerhansi</i> (Gidholm, 1967) | I | - | 6 | - |
| <i>Ancistrosyllis groenlandica</i> McIntosh, 1879 | II | 10 | 4 | - | <i>Myrtea spinifera</i> (Montagu, 1803) | II | 23 | 11 | - |
| <i>Ancistrosyllis hamata</i> McIntosh, 1879 | II | 43 | - | - | <i>Mysia undata</i> (Pennant, 1777) | I | 1 | - | - |
| <i>Anobothrus gracilis</i> (Malmgren, 1866) | II | 145 | 20 | 1 | <i>Mysta picta</i> (Quatrefages, 1866) | II | 2 | 3 | 2 |
| <i>Anomia ephippium</i> Linnaeus, 1758 | III | - | 4 | - | <i>Nannastacus longirostris</i> G.O. Sars, 1879 | II | - | 9 | - |
| <i>Antalis dentalis</i> (Linnaeus, 1758) | I | 56 | 20 | 3 | <i>Nannastacus unguiculatus</i> (Bate, 1859) | II | - | 3 | - |
| <i>Antalis inaequicostata</i> (Dautzenberg, 1891) | I | 30 | 3 | - | <i>Nassarius corniculum</i> (Olivieri, 1792) | II | 2 | - | - |
| <i>Aonides oxycephala</i> (Sars, 1862) | II | 8 | 8 | - | <i>Nassarius cuvierii</i> (Payraudeau, 1826) | II | - | - | 1 |
| <i>Aphelochaeta filiformis</i> (Kefervein, 1862) | III | 13 | 12 | - | <i>Nassarius incrassatus</i> (Strøm, 1768) | II | 35 | - | - |
| <i>Apherusa alacris</i> Krapp-Schickel, 1969 | I | - | - | 4 | <i>Nassarius lima</i> (Dillwyn, 1817) | I | - | 1 | 4 |
| <i>Apherusa chierieghinii</i> Giordani-Soika, 1949 | I | - | 6 | - | <i>Nassarius nitidus</i> (Jeffreys, 1867) | II | - | - | 9 |
| <i>Apionsoma misakianum</i> (Ikeda, 1904) | II | - | 1 | 4 | <i>Nassarius pygmaeus</i> (Lamarck, 1822) | IV | 130 | 106 | 23 |
| <i>Apistobranchius tullbergi</i> (Théel, 1879) | I | - | 1 | - | <i>Nassarius reticulatus</i> (Linnaeus, 1758) | II | 1 | - | - |
| <i>Apocorophium acutum</i> (Chevreux, 1908) | III | 1 | 2 | - | <i>Neanthes nubila</i> (Savigny, 1822) | III | - | 11 | - |
| <i>Aponuphis bilineata</i> (Baird, 1870) | II | - | 3 | 6 | <i>Nemertopsis bivittata</i> (Delle Chiaje, 1841) | II | 2 | - | - |
| <i>Aponuphis brementi</i> (Fauvel, 1916) | II | 19 | 21 | 11 | <i>Neogyptis mediterranea</i> (Pleijel, 1993) | I | 3 | 1 | - |
| <i>Aporrhais pesselecani</i> (Linnaeus, 1758) | II | 9 | 8 | - | <i>Nephtys caeca</i> (Fabricius, 1780) | I | 1 | - | - |
| <i>Apsudopsis latreillii</i> (Milne Edwards, 1828) | III | 95 | 58 | 62 | <i>Nephtys hombergii</i> Savigny in Lamarck, 1818 | IV | 4 | 31 | 15 |
| <i>Arabella iricolor</i> (Montagu, 1804) | I | - | 4 | - | <i>Nephtys hystrix</i> McIntosh, 1900 | II | 23 | - | - |
| <i>Arbacia lixula</i> (Linnaeus, 1758) | II | - | 1 | - | <i>Nephtys incisa</i> Malmgren, 1865 | II | 61 | 36 | 4 |
| <i>Arichlidon reyssei</i> (Katzmann, Laubier & Ramos, 1974) | I | - | 2 | 8 | <i>Nereis rava</i> Ehlers, 1864 | III | - | 1 | - |
| <i>Aricidea suecica meridionalis</i> Laubier & Ramos, 1974 | II | 1 | 1 | 2 | <i>Neverita josephina</i> Risso, 1826 | I | - | - | 2 |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|------|-----|-----|---|-----|-----|----|-----|
| <i>Aricidea annae</i> Laubier, 1967 | I | 2 | - | - | <i>Ninoe armoricana</i> Glémarec, 1968 | I | 11 | 1 | - |
| <i>Aricidea assimilis</i> Tebble, 1959 | III | 7 | 11 | 3 | <i>Nothria conchylega</i> (Sars, 1835) | II | 2 | 1 | - |
| <i>Aricidea catherinae</i> Laubier, 1967 | I | - | 2 | 7 | <i>Notomastus aberans</i> Day, 1957 | III | 21 | 42 | 15 |
| <i>Aricidea cerrutii</i> Laubier, 1966 | II | 1 | - | 2 | <i>Notomastus latericeus</i> Sars, 1851 | III | 25 | 14 | - |
| <i>Aricidea claudiae</i> Laubier, 1967 | III | 1366 | 97 | 1 | <i>Notomastus lineatus</i> (Claparède, 1869) | III | - | 7 | - |
| <i>Aricidea lopezi</i> Berkeley & Berkeley, 1956 | II | 15 | - | - | <i>Notomastus mossambicus</i> (Thomassin, 1970) | III | - | - | 45 |
| <i>Aricidea pseudoarticulata</i> Hobson, 1972 | III | 85 | 70 | - | <i>Nucula nitidosa</i> Winckworth, 1930 | II | 19 | 20 | - |
| <i>Aricidea simonae</i> Laubier & Ramos, 1974 | I | 3 | - | - | <i>Nucula nucleus</i> (Linnaeus, 1758) | II | 8 | - | - |
| <i>Ascobulla fragilis</i> (Jeffreys, 1856) | I | - | 1 | - | <i>Nuculana pella</i> (Linnaeus, 1767) | II | 1 | 2 | - |
| <i>Aspidosiphon muelleri</i> Diesing, 1851 | I | 5 | - | - | <i>Obesotoma laevigata</i> (Dall, 1871) | I | - | 4 | 3 |
| <i>Aspidosiphon mexicanus</i> (Murina, 1967) | I | 9 | - | 3 | <i>Ocinebrina aciculata</i> (Lamarck, 1822) | I | 1 | - | - |
| <i>Astarte sulcata</i> (da Costa, 1778) | I | - | 1 | - | <i>Octobranchus lingulatus</i> (Grube, 1863) | II | - | 1 | - |
| <i>Asterina gibbosa</i> (Pennant, 1777) | I | - | 2 | 1 | <i>Odontosyllis fulgurans</i> (Audouin & Milne Edwards, 1833) | II | - | 1 | - |
| <i>Astropecten bispinosus</i> (Otto, 1823) | I | 1 | - | - | <i>Odontosyllis gibba</i> Claparède, 1863 | II | - | 3 | - |
| <i>Athanas nitescens</i> (Leach, 1813 [in Leach, 1813-1814]) | I | 1 | 1 | - | <i>Odostomella doliolum</i> (Philippi, 1844) | I | - | 2 | - |
| <i>Arys jeffreysi</i> (Weinkauff, 1866) | I | - | - | 2 | <i>Odostomia unidentata</i> (Montagu, 1803) | I | - | 1 | - |
| <i>Aurospio banyulensis</i> (Laubier, 1966) | II | - | 16 | - | <i>Ostergrenia digitata</i> (Montagu, 1815) | III | 145 | 9 | - |
| <i>Aurospio dibranchiata</i> Maciolek, 1981 | I | - | 6 | - | <i>Ogyrides mjoeborgi</i> (Balss, 1921) | I | - | - | 3 |
| <i>Axiobella constricta</i> (Claparède, 1869) | I | - | - | 2 | <i>Onchnesoma steenstrupii</i> (Danielsen, 1876) | II | 134 | 54 | 109 |
| <i>Barleeia unifasciata</i> (Montagu, 1803) | I | - | - | 2 | <i>Ondina vitrea</i> (Brusina, 1866) | I | 3 | 5 | - |
| <i>Batharca pectunculoides</i> (Scacchi, 1835) | I | - | - | 1 | <i>Ondina warreni</i> (Thompson, 1845) | I | - | 1 | - |
| <i>Bathyporeia megalops</i> Chevreux, 1911 | I | - | 11 | 1 | <i>Onuphis eremita</i> Audouin & Milne Edwards, 1833 | II | - | 3 | 11 |
| <i>Bela brachystoma</i> (Philippi, 1844) | II | 96 | 23 | 8 | <i>Ophelina acuminata</i> Örsted, 1843 | II | 3 | - | - |
| <i>Bela nebula</i> (Montagu, 1803) | I | 1 | - | - | <i>Ophelina cylindricaudata</i> (Hansen, 1879) | II | 24 | 5 | - |
| <i>Bitium latreillii</i> (Payraudeau, 1826) | II | 4 | 134 | 168 | <i>Ophelina modesta</i> Støp-Bowitz, 1958 | II | 34 | - | - |
| <i>Bitium reticulatum</i> (da Costa, 1778) | II | 56 | 248 | 135 | <i>Ophiopsila aranea</i> Forbes, 1843 | I | - | 1 | - |
| <i>Bitium submamillatum</i> (de Rayneval & Ponzi, 1854) | I | - | 10 | - | <i>Ophiothrix fragilis</i> (Abildgaard, in O.F. Müller, 1789) | I | - | 6 | - |
| <i>Bodotria scorpioides</i> (Montagu, 1804) | II | - | - | 2 | <i>Ophiura albida</i> Forbes, 1839 | IV | 4 | - | 6 |
| <i>Brachystomia eulimoides</i> (Hanley, 1844) | II | 1 | - | - | <i>Ophiura ophiura</i> (Linnaeus, 1758) | II | 1 | 2 | - |
| <i>Brachystomia scalaris</i> (MacGillivray, 1843) | I | - | 1 | - | <i>Ophryotrocha labronica</i> Bacci & La Greca, 1961 | IV | 1 | - | - |
| <i>Branchiostoma lanceolatum</i> (Pallas, 1774) | I | - | 3 | 14 | <i>Ophryotrocha puerilis</i> Claparède & Metschnikow, 1869 | IV | 1 | - | - |
| <i>Brevicirrosyllis weismanni</i> (Langerhans, 1879) | I | - | - | 2 | <i>Opisthosyllis brunnea</i> Langerhans, 1879 | II | - | - | 1 |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|----|----|----|--|-----|----|-----|-----|
| <i>Brissopsis lyrifera</i> (Forbes, 1841) | II | 6 | 1 | 2 | <i>Orchomene humilis</i> (Costa, 1853) | II | 8 | - | - |
| <i>Bulla striata</i> Bruguière, 1792 | I | - | - | 1 | <i>Ova canaliferus</i> (Lamarck, 1816) | I | - | 1 | - |
| <i>Callianassa subterranea</i> (Montagu, 1808) | III | - | 4 | 2 | <i>Owenia fusiformis</i> Delle Chiaje, 1844 | II | 1 | 3 | 523 |
| <i>Calliostoma laugierii</i> (Payraudeau, 1826) | I | - | - | 1 | <i>Oxydromus flexuosus</i> (Delle Chiaje, 1827) | III | 4 | - | - |
| <i>Calocaris macandreae</i> Bell, 1853 | II | - | 1 | - | <i>Oxydromus pallidus</i> Claparède, 1864 | V | 5 | 1 | 12 |
| <i>Calyptraea chinensis</i> (Linnaeus, 1758) | II | 3 | 9 | - | <i>Pagurus cuanensis</i> Bell, 1846 | II | - | 3 | - |
| <i>Campylaspis glabra</i> Sars, 1878 | II | - | 2 | - | <i>Papillicardium papillosum</i> (Poli, 1791) | II | 1 | - | - |
| <i>Campylaspis legendrei</i> Fage, 1951 | II | - | 1 | - | <i>Paradiatylchone filicaudata</i> (Southern, 1914) | II | 34 | 16 | 1 |
| <i>Capitella minima</i> Langerhans, 1881 | IV | - | - | 1 | <i>Paradoneis ilvana</i> Castelli, 1985 | III | - | - | 1 |
| <i>Capitella telata</i> Blake, Grassle, Eckelbarger, 2009 | V | 66 | 2 | - | <i>Paradoneis lyra</i> (Southern, 1914) | III | 33 | 14 | 2 |
| <i>Caprella acanthifera</i> Leach, 1814 | II | - | 7 | - | <i>Paraehlersia ferrugina</i> (Langerhans, 1881) | II | - | 1 | 1 |
| <i>Carangoliopsis spinulosa</i> Ledoyer, 1970 | I | 3 | - | - | <i>Paralacydonia paradoxa</i> Fauvel, 1913 | I | 30 | 132 | 3 |
| <i>Caullella alata</i> (Southern, 1914) | II | - | 1 | 1 | <i>Paranthura costana</i> Bate & Westwood, 1866 | I | - | - | 2 |
| <i>Ceratia proxima</i> (Forbes & Hanley, 1850) | I | 4 | 5 | - | <i>Paraonis tenera</i> Grube, 1872 | I | 1 | - | - |
| <i>Ceratonereis (Composetia) costae</i> (Grube, 1840) | I | - | 1 | - | <i>Paraphoxus oculatus</i> (Sars, 1879) | I | 4 | 9 | - |
| <i>Ceratonereis (Composetia) hircinicola</i> (Eisig, 1870) | I | 41 | 1 | 1 | <i>Parapionosyllis brevicirra</i> Day, 1954 | I | - | 6 | 2 |
| <i>Ceratonereis mirabilis</i> Kinberg, 1865 | I | - | 1 | 5 | <i>Parapionosyllis elegans</i> (Pierantoni, 1903) | I | 2 | - | 1 |
| <i>Cerithidium diplax</i> (Watson, 1886) | II | - | - | 28 | <i>Parapionosyllis minuta</i> (Pierantoni, 1903) | I | 1 | - | 1 |
| <i>Cerithiopsis minima</i> (Brusina, 1865) | I | - | - | 1 | <i>Parapriospio coora</i> Wilson, 1990 | I | 66 | 6 | - |
| <i>Cerithiopsis tubercularis</i> (Montagu, 1803) | I | 1 | - | - | <i>Parexogone caribensis</i> (San Martin, 1991) | I | 6 | 3 | - |
| <i>Cerithium scabridum</i> Philippi, 1848 | II | - | - | 6 | <i>Parexogone hebes</i> (Webster & Benedict, 1884) | I | 2 | - | - |
| <i>Cerithium vulgatum</i> Bruguière, 1792 | I | - | 1 | - | <i>Parhyale eburnea</i> Krapp-Schickel, 1974 | I | - | 1 | - |
| <i>Chaetopterus variopedatus</i> (Renier, 1804) | I | - | 1 | - | <i>Parougia cf. caeca</i> (Webster & Benedict, 1884) | II | 1 | - | - |
| <i>Chaetozone corona</i> Berkeley & Berkeley, 1941 | III | 18 | 1 | - | <i>Parthenina dollfusi</i> (Kobelt, 1903) | I | 1 | 1 | - |
| <i>Chaetozone gibber</i> Woodham & Chambers, 1994 | III | 7 | 7 | - | <i>Parthenina emaciata</i> (Brusina, 1866) | I | 2 | 1 | - |
| <i>Charybdis hellerii</i> (A. Milne-Edwards, 1867) | II | - | - | 2 | <i>Parthenina interstincta</i> (J. Adams, 1797) | I | 3 | 9 | - |
| <i>Chauvetia brunnea</i> (Donovan, 1804) | I | - | 1 | - | <i>Parthenina juliae</i> (de Folin, 1872) | I | 2 | - | - |
| <i>Chauvetia turritellata</i> (Deshayes, 1835) | I | - | 5 | - | <i>Parthenina palazzii</i> (Micali, 1984) | I | 1 | - | - |
| <i>Cheiriphotis mediterranea</i> Myers, 1983 | II | - | - | 23 | <i>Parthenina suturalis</i> (Philippi, 1844) | I | - | 1 | - |
| <i>Chone collaris</i> Langerhans, 1881 | II | 6 | 2 | - | <i>Parthenina terebellum</i> (Philippi, 1844) | I | 3 | 3 | 3 |
| <i>Chone dumeri</i> Malmgren, 1867 | II | - | 2 | 1 | <i>Parvicardium exiguum</i> (Gmelin, 1791) | II | - | 4 | - |
| <i>Chone dumerificta</i> Tovar-Hernández, Licciano, Giangrande, 2007 | I | - | - | 1 | <i>Parvicardium pinnulatum</i> (Conrad, 1831) | I | - | 1 | - |
| <i>Chone longiseta</i> Giangrande, 1992 | I | - | 1 | - | <i>Parvioris ibizenca</i> (Nordsieck, 1968) | I | - | 3 | - |
| <i>Cirolana neglecta</i> Hansen, 1890 | II | 10 | - | - | <i>Parvipalpus linea</i> Mayer, 1890 | I | - | 2 | - |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|---|-----|-----|-----|----|--|-----|-----|----|----|
| <i>Cirrophorus branchiatus</i> Ehlers, 1908 | III | 36 | 2 | - | <i>Perioculodes aequimanus</i> (Korssman, 1880) | II | 2 | - | - |
| <i>Cirrophorus furcatus</i> (Hartman, 1957) | III | 19 | 49 | 2 | <i>Perioculodes longimanus</i> (Bate & Westwood, 1868) | II | 15 | 11 | 5 |
| <i>Clanculus cruciatus</i> (Linnaeus, 1758) | I | - | 4 | - | <i>Phascolion strombus</i> (Montagu, 1804) | I | 2 | 3 | - |
| <i>Clausinella fasciata</i> (da Costa, 1778) | I | 1 | 1 | 1 | <i>Phaxas pellucidus</i> (Pennant, 1777) | II | 10 | 5 | - |
| <i>Clorida albolitura</i> Ah Yong & Naiyanetr, 2000 | II | - | - | 1 | <i>Pherusa plumosa</i> (Müller, 1776) | II | - | 1 | - |
| <i>Conomurex persicus</i> (Swainson, 1821) | I | - | - | 16 | <i>Philine scabra</i> (O. F. Müller, 1784) | II | - | 1 | - |
| <i>Conus ventricosus</i> Gmelin, 1791 | I | - | - | 2 | <i>Philocheras monacanthus</i> (Holthuis, 1961) | I | - | 1 | - |
| <i>Corbula gibba</i> (Olivi, 1792) | IV | 74 | 290 | 14 | <i>Pholoe inornata</i> Johnston, 1839 | II | 1 | - | - |
| <i>Cossura soyeri</i> Laubier, 1964 | III | 779 | 92 | 35 | <i>Phoronis psammophila</i> Cori, 1889 | III | 6 | - | - |
| <i>Crassopleura maravignae</i> (Bivona Ant. in Bivona And., 1838) | I | - | - | 3 | <i>Photis longipes</i> (Della Valle, 1893) | I | - | 1 | - |
| <i>Cumella limicola</i> Sars, 1879 | II | - | 1 | - | <i>Phoxocephalus aquosus</i> Karaman, 1985 | I | 9 | - | - |
| <i>Cyathura carinata</i> (Krøyer, 1847) | III | - | 2 | 10 | <i>Phthisica marina</i> Slabber, 1769 | III | 1 | 16 | - |
| <i>Cylichna cylindracea</i> (Pennant, 1777) | II | 82 | 20 | 1 | <i>Phyllodoce lineata</i> (Claparède, 1870) | II | 2 | 1 | - |
| <i>Cymodoce spinosa</i> (Risso, 1816) | I | - | 1 | - | <i>Phyllodoce maculata</i> (Linnaeus, 1767) | III | 1 | 3 | - |
| <i>Cymodoce truncata</i> Leach, 1814 | I | - | 5 | - | <i>Phyllodoce mucosa</i> Örsted, 1843 | III | 1 | - | - |
| <i>Cymodoce tuberculata</i> Costa in Hope, 1851 | I | - | 3 | - | <i>Phyllodoce rosea</i> (McIntosh, 1877) | III | 1 | - | 1 |
| <i>Deflexilodes acutipes</i> (Ledoyer, 1983) | I | - | 1 | - | <i>Phylo foetida</i> (Claparède, 1869) | I | 1 | - | - |
| <i>Deflexilodes gibbosus</i> (Chevreux, 1888) | I | 1 | 11 | - | <i>Pilargis verrucosa</i> Saint-Joseph, 1899 | III | 98 | 11 | 3 |
| <i>Dexamine spinosa</i> (Montagu, 1813) | II | 1 | 13 | - | <i>Piromis eruca</i> (Claparède, 1869) | III | - | 12 | - |
| <i>Diastylis cornuta</i> (Boeck, 1864) | I | - | 2 | - | <i>Pisidia bluteli</i> (Risso, 1816) | II | 1 | - | - |
| <i>Diastylis neapolitana</i> Sars, 1879 | I | - | 2 | - | <i>Pisidia longicornis</i> (Linnaeus, 1767) | II | 1 | - | - |
| <i>Diastylis rugosa</i> Sars, 1865 | I | - | 1 | - | <i>Pisione guanche</i> San Martín, López & Núñez, 1999 | I | - | - | 7 |
| <i>Diogenes pugilator</i> (Roux, 1829) | II | - | 1 | - | <i>Pista cristata</i> (Müller, 1776) | I | 3 | 12 | - |
| <i>Diopatra neapolitana</i> Delle Chiaje, 1841 | III | - | 1 | - | <i>Pista unibranchia</i> Day, 1963 | II | 5 | 1 | - |
| <i>Diplocirrus glaucus</i> (Malmgren, 1867) | II | 112 | 6 | 1 | <i>Pitar rudis</i> (Poli, 1795) | I | - | 6 | - |
| <i>Diplydora coeca</i> (Örsted, 1843) | III | 18 | 1 | - | <i>Platynereis dumerilii</i> (Audouin & Milne Edwards, 1834) | III | - | 1 | 2 |
| <i>Dischides politus</i> (S. Wood, 1842) | I | - | 2 | - | <i>Podarkeopsis galangau</i> Laubier, 1961 | IV | 55 | 21 | 9 |
| <i>Ditrupa arietina</i> (O. F. Müller, 1776) | I | - | - | 6 | <i>Podocerus variegatus</i> Leach, 1814 | III | - | 1 | 1 |
| <i>Donax semistriatus</i> Poli, 1795 | I | - | - | 17 | <i>Poecilochaetus fauchaldi</i> Pilato & Cantone, 1976 | II | 59 | - | 1 |
| <i>Dosinia lupinus</i> (Linnaeus, 1758) | III | 1 | 6 | - | <i>Poecilochaetus serpens</i> Allen, 1904 | II | 15 | 48 | 2 |
| <i>Drilliola loprestiana</i> (Calcara, 1841) | I | - | - | 1 | <i>Polydora cornuta</i> Bosc, 1802 | V | 133 | 12 | - |
| <i>Drilonereis filum</i> (Claparède, 1868) | I | 12 | 8 | 4 | <i>Polydora hoplura</i> Claparède, 1869 | III | - | 5 | - |
| <i>Dynamene torelliae</i> Holdich, 1968 | II | - | - | 1 | <i>Polygordius appendiculatus</i> Fraipont, 1887 | I | 7 | - | - |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|----|----|----|---|-----|-----|----|-----|
| <i>Ebalia tuberosa</i> (Pennant, 1777) | II | - | 1 | 1 | <i>Polygordius lacteus</i> Schneider, 1868 | I | 29 | 1 | 1 |
| <i>Echinocyamus pusillus</i> (O.F. Müller, 1776) | I | - | 1 | - | <i>Polyophthalmus pictus</i> (Dujardin, 1839) | I | 1 | 2 | 4 |
| <i>Edwardsia claparedii</i> (Panceri, 1869) | III | 49 | 4 | 2 | <i>Pontogenia chrysocoma</i> (Baird, 1865) | II | - | 2 | - |
| <i>Ekleptostylis walkeri</i> (Calman, 1907) | I | 1 | - | - | <i>Praxillella gracilis</i> (M. Sars, 1861) | II | 1 | 9 | 2 |
| <i>Elasmopus pocillimanus</i> (Bate, 1862) | II | - | - | 7 | <i>Praxillella praetermissa</i> (Malmgren, 1865) | II | 21 | 5 | - |
| <i>Electroma vexillum</i> (Reeve, 1857) | II | - | - | 2 | <i>Prionospio anatolica</i> Dagli & Çınar, 2011 | I | - | - | 5 |
| <i>Eocuma sarsii</i> (Kossmann), 1880 | I | - | 1 | 1 | <i>Prionospio auklandica</i> Augener, 1923 | IV | - | - | 12 |
| <i>Epitonium clathrus</i> (Linnaeus, 1758) | I | 1 | 1 | - | <i>Prionospio caspersi</i> Laubier, 1962 | III | - | - | 11 |
| <i>Epitonium muricatum</i> (Risso, 1826) | I | - | 1 | - | <i>Prionospio cirrifera</i> Wirén, 1883 | II | - | 9 | - |
| <i>Epitonium turtonis</i> (Turton, 1819) | I | 2 | - | 1 | <i>Prionospio depauperata</i> Imajima, 1990 | II | 10 | 6 | 157 |
| <i>Erichonius argenteus</i> Krapp-Schickel, 1993 | I | - | - | 5 | <i>Prionospio dubia</i> Day, 1961 | I | 31 | 7 | 1 |
| <i>Erichonius punctatus</i> (Bate, 1857) | II | 3 | 2 | - | <i>Prionospio ehlersi</i> Fauvel, 1928 | I | 4 | 6 | - |
| <i>Eriopisa elongata</i> (Bruzelius, 1859) | I | 5 | - | - | <i>Prionospio ergeni</i> Dagli & Çınar, 2009 | III | - | - | 19 |
| <i>Euchone rosea</i> Langerhans, 1884 | II | 3 | 2 | - | <i>Prionospio fallax</i> Söderström, 1920 | V | 208 | 18 | 12 |
| <i>Euclymene collaris</i> (Claparède, 1869) | I | - | 2 | - | <i>Prionospio maciolekae</i> Dagli & Çınar, 2011 | II | 174 | 24 | - |
| <i>Euclymene lombricoides</i> (Quatrefages, 1866) | II | 8 | 5 | - | <i>Prionospio paucipinnulata</i> Blake & Kudenov, 1978 | II | - | - | 4 |
| <i>Euclymene oerstedii</i> (Claparède, 1863) | II | 2 | 1 | - | <i>Prionospio pulchra</i> Imajima, 1990 | V | 72 | 25 | - |
| <i>Euclymene palermitana</i> (Grube, 1840) | I | - | 2 | - | <i>Prionospio saccifera</i> Mackie & Hartley, 1990 | II | - | - | 123 |
| <i>Eudorella truncatula</i> (Bate, 1856) | I | 6 | 4 | - | <i>Prionospio sexoculata</i> Augener, 1918 | IV | - | - | 15 |
| <i>Eulalia clavigera</i> (Audouin & Milne Edwards, 1833) | I | 2 | - | - | <i>Prionospio steenstrupi</i> Malmgren, 1867 | III | 243 | 76 | 2 |
| <i>Eulalia mustela</i> Pleijel, 1987 | I | - | 1 | - | <i>Proceracea aurantiaca</i> Claparède, 1868 | II | - | 1 | - |
| <i>Eulima glabra</i> (da Costa, 1778) | I | 13 | 8 | - | <i>Processa modica modica</i> Williamson in Williamson & Rochanaburanon, 1979 | I | - | 2 | 1 |
| <i>Eulimella acicula</i> (Philippi, 1836) | I | 11 | 1 | - | <i>Processa noveli noveli</i> Al-Adhub & Williamson, 1975 | II | 23 | - | - |
| <i>Eulimella ventricosa</i> (Forbes, 1844) | I | - | 1 | - | <i>Prosphaerosyllis marmarae</i> Çınar, Dagli & Acik, 2011 | I | - | 1 | - |
| <i>Eumida sanguinea</i> (Örsted, 1843) | II | 2 | 1 | - | <i>Prosphaerosyllis xariffae</i> (Hartmann-Schröder, 1960) | I | 18 | 1 | - |
| <i>Eunereis longissima</i> Johnston, 1840 | II | 1 | - | 1 | <i>Protodorvillea kefersteini</i> (McIntosh, 1869) | III | 7 | 19 | 1 |
| <i>Eunice vittata</i> (Delle Chiaje, 1828) | II | 89 | 24 | 2 | <i>Psamathe fusca</i> Johnston, 1836 | II | - | 2 | - |
| <i>Euparthenia humboldti</i> (Risso, 1826) | I | - | 1 | - | <i>Psammecchinus microtuberculatus</i> (Blainville, 1825) | I | - | 1 | - |
| <i>Eurydice affinis</i> Hansen, 1905 | I | - | 1 | 13 | <i>Pseudofabriciella analis</i> Fitzhugh, Giangrande & Simbora, 1994 | II | - | 1 | - |
| <i>Eurydice pulchra</i> Leach, 1815 | I | - | 2 | - | <i>Pseudofabriciella longipyga</i> Fitzhugh, Giangrande & Simbora, 1994 | I | 3 | 9 | 1 |
| <i>Eurydice spinigera</i> Hansen, 1890 | I | - | 2 | 1 | <i>Pseudoleiocapitella fauveli</i> Harmelin, 1964 | III | 27 | 34 | 2 |
| <i>Eurynome aspera</i> (Pennant, 1777) | I | - | 1 | - | <i>Pseudomystidea limbata</i> (Saint-Joseph, 1888) | II | - | 1 | 1 |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|----|----|----|---|-----|-----|----|----|
| <i>Eurysyllis tuberculata</i> Ehlers, 1864 | I | - | 1 | 1 | <i>Pseudomystides spinachia</i> Petersen & Pleijel in Pleijel, 1993 | I | 13 | 1 | 2 |
| <i>Euspira nitida</i> (Donovan, 1804) | II | 15 | 3 | - | <i>Pseudopolydora antennata</i> (Claparède, 1869) | II | - | - | 1 |
| <i>Eusyllis assimilis</i> Marenzeller, 1875 | I | - | 2 | - | <i>Pseudopolydora</i> <i>paucibranchiata</i> (Okuda, 1937) | V | 129 | - | 2 |
| <i>Eusyllis lamelligera</i> Marion & Bobretzky, 1875 | I | - | 1 | - | <i>Pseudopolydora pulchra</i> (Carazzi, 1893) | IV | 9 | 5 | - |
| <i>Exogone dispar</i> (Webster, 1879) | I | 1 | - | - | <i>Pteria hirundo</i> (Linnaeus, 1758) | I | - | 2 | - |
| <i>Exogone cognettii</i> Castelli, Badalamenti & Lardici, 1987 | I | 3 | 2 | - | <i>Pterocirrus macroceros</i> (Grube, 1860) | I | 1 | - | - |
| <i>Exogone gambiae</i> Lanera, Sordino & San Martín, 1994 | I | - | 3 | 1 | <i>Pusillina inconspicua</i> (Alder, 1844) | II | 1 | 31 | 1 |
| <i>Exogone naidina</i> Örsted, 1845 | II | 7 | 1 | - | <i>Pusillina lineolata</i> (Michaud, 1830) | III | 8 | 8 | - |
| <i>Exogone rostrata</i> Naville, 1933 | II | - | 5 | - | <i>Pusillina marginata</i> (Michaud, 1830) | I | - | 3 | 2 |
| <i>Exogone verugera</i> (Claparède, 1868) | I | 20 | 5 | 6 | <i>Pusillina radiata</i> (Philippi, 1836) | I | - | 18 | 2 |
| <i>Fabricia stellaris</i> (Müller, 1774) | II | 3 | 1 | - | <i>Pyrgiscus rufus</i> (Philippi, 1836) | II | 2 | 2 | - |
| <i>Fauveliopsis adriatica</i> Katzmann & Laubier, 1974 | I | 37 | - | 2 | <i>Pyrgostylus striatulus</i> (Linnaeus, 1758) | I | - | 1 | - |
| <i>Finella pupoides</i> A. Adams, 1860 | II | - | 6 | 17 | <i>Pyrrunculus fourieri</i> (Audouin, 1826) | II | - | 1 | - |
| <i>Flabelligera affinis</i> M. Sars, 1829 | I | - | 1 | - | <i>Pyrrunculus hoernesii</i> (Weinkauff, 1866) | I | - | 1 | - |
| <i>Flexopecten hyalinus</i> (Poli, 1795) | I | - | 2 | - | <i>Raphitoma aequalis</i> (Jeffreys, 1867) | I | - | 7 | - |
| <i>Folinella excavata</i> (Philippi, 1836) | I | - | 1 | - | <i>Raphitoma echinata</i> (Brocchi, 1814) | I | 1 | 1 | - |
| <i>Fulvia fragilis</i> (Forsskål in Niebuhr, 1775) | III | 1 | 14 | - | <i>Raphitoma linearis</i> (Montagu, 1803) | I | 1 | 8 | - |
| <i>Fusinus rostratus</i> (Olivi, 1792) | I | 1 | - | - | <i>Retusa crebrisculpta</i> (Monterosato, 1884) | I | - | 1 | - |
| <i>Fusinus rusticulus</i> (Monterosato, 1880) | I | - | 1 | - | <i>Retusa laevisculpta</i> (Granata- Grillo, 1877) | I | - | 2 | - |
| <i>Fustiaria rubescens</i> (Deshayes, 1825) | I | - | 1 | 1 | <i>Retusa minutissima</i> (Monterosato, 1878) | I | 2 | 2 | - |
| <i>Galathea bolivari</i> Zariquiey Álvarez, 1950 | I | - | 5 | - | <i>Retusa truncatula</i> (Bruguère, 1792) | I | 1 | 2 | - |
| <i>Galathea intermedia</i> Lilljeborg, 1851 | I | - | 2 | - | <i>Retusa umbilicata</i> (Montagu, 1803) | I | 1 | 3 | - |
| <i>Galathowenia oculata</i> (Zachs, 1923) | I | 7 | 5 | 1 | <i>Rhinoclavis kochi</i> (Philippi, 1848) | II | - | - | 1 |
| <i>Gammarella fucicola</i> (Leach, 1814) | III | - | 1 | - | <i>Rhodine loveni</i> Malmgren, 1865 | III | 44 | 16 | 9 |
| <i>Gastrosaccus sanctus</i> (Van Beneden, 1861) | I | 1 | 1 | 7 | <i>Ringicula auriculata</i> (Ménard de la Groye, 1811) | I | 3 | 1 | - |
| <i>Gibbula adansonii</i> (Payraudeau, 1826) | I | - | - | 1 | <i>Ringicula conformis</i> Monterosato, 1877 | I | 24 | 6 | - |
| <i>Gibbula ardens</i> (Salis Marschlin, 1793) | I | 1 | 5 | 2 | <i>Rissoa auriscalpium</i> (Linnaeus, 1758) | I | - | 1 | - |
| <i>Gibbula divaricata</i> (Linnaeus, 1758) | I | 2 | - | - | <i>Rissoa monodonta</i> Philippi, 1836 | I | - | 1 | - |
| <i>Glans trapezia</i> (Linnaeus, 1767) | I | - | 1 | - | <i>Rissoa rodhensis</i> Verduin, 1985 | I | - | - | 2 |
| <i>Glycera alba</i> (O.F. Müller, 1776) | III | 13 | 14 | 7 | <i>Rissoa variabilis</i> (Von Mühlfeldt, 1824) | I | - | 4 | - |
| <i>Glycera fallax</i> Quatrefages, 1850 | III | 47 | 32 | 16 | <i>Rissoa violacea</i> Desmarest, 1814 | I | - | 1 | - |
| <i>Glycera tessellata</i> Grube, 1840 | II | - | - | 2 | <i>Roxania utriculus</i> (Brocchi, 1814) | I | - | 2 | - |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|-----|----|----|--|-----|-----|----|-----|
| <i>Glycera tridactyla</i> Schmarda, 1861 | III | - | 3 | - | <i>Rullierinereis anoculata</i> Cantone, 1983 | I | 2 | - | - |
| <i>Glycera unicornis</i> Lamarck, 1818 | III | 9 | 22 | 1 | <i>Saccella commutata</i> (Philippi, 1844) | II | - | 1 | 10 |
| <i>Glycinde bonhourei</i> Gravier, 1904 | II | 1 | - | 55 | <i>Saccocirrus papilloecercus</i> Bobretzky, 1872 | I | - | - | 1 |
| <i>Glycinde nordmanni</i> (Malmgren, 1866) | II | 2 | - | - | <i>Salvatoria clavata</i> (Claparède, 1863) | II | - | - | 1 |
| <i>Gnathia vorax</i> (Lucas, 1849) | I | 5 | 8 | - | <i>Scalibregma celticum</i> Mackie, 1991 | II | - | 1 | - |
| <i>Goneplax rhomboides</i> (Linnaeus, 1758) | III | 8 | 3 | - | <i>Schistomeringos rudolphi</i> (Delle Chiaje, 1828) | V | 5 | - | 1 |
| <i>Goniada maculata</i> Örsted, 1843 | II | 2 | 14 | 2 | <i>Sclerocheilus minutus</i> Grube, 1863 | I | - | 2 | - |
| <i>Gouldia minima</i> (Montagu, 1803) | I | 1 | 3 | - | <i>Scolecipis tridentata</i> (Southern, 1914) | III | 11 | - | - |
| <i>Gourretia denticulata</i> (Lutze, 1937) | II | 5 | 3 | 2 | <i>Scoletoma emandibulata</i> <i>mabiti</i> (Ramos, 1976) | II | 34 | 6 | 8 |
| <i>Granulina clandestina</i> (Brocchi, 1814) | I | - | 3 | - | <i>Scoletoma impatiens</i> (Claparède, 1868) | II | 5 | 4 | 1 |
| <i>Granulina marginata</i> (Bivona, 1832) | I | 2 | - | - | <i>Scoloplos armiger</i> (Müller, 1776) | III | - | 1 | - |
| <i>Granulina occulta</i> (Monterosato, 1869) | I | - | 11 | - | <i>Scoloplos chevalieri</i> <i>canadiensis</i> Harmelin, 1969 | I | 2 | - | 81 |
| <i>Guernea (Guernea) coalita</i> (Norman, 1868) | I | 9 | 3 | - | <i>Scrobicularia plana</i> (da Costa, 1778) | I | 1 | - | - |
| <i>Gyptis propinqua</i> Marion & Bobretzky, 1875 | I | 1 | - | - | <i>Sigalion mathildae</i> Audouin & Milne Edwards in Cuvier, 1830 | II | - | 2 | 12 |
| <i>Haedropleura septangularis</i> (Montagu, 1803) | I | - | - | 1 | <i>Sigambra tentaculata</i> (Treadwell, 1941) | IV | 566 | 79 | 249 |
| <i>Halocynthia papillosa</i> (Linnaeus, 1767) | II | - | 2 | - | <i>Siphonoecetes</i> (<i>Centraloecetes</i>) <i>dellavallei</i> Stebbing, 1899 | I | - | - | 1 |
| <i>Haminoea hydatis</i> (Linnaeus, 1758) | I | - | 5 | - | <i>Siriella clausii</i> G.O. Sars, 1877 | I | - | 2 | - |
| <i>Haplosyllis spongicola</i> (Grube, 1855) | I | - | 2 | - | <i>Sirpus zariquieyi</i> Gordon, 1953 | I | - | 1 | - |
| <i>Harmothoe antilopes</i> McIntosh, 1876 | II | 5 | 2 | 1 | <i>Sosane sulcata</i> Malmgren, 1866 | II | - | 2 | - |
| <i>Harmothoe goreensis</i> Augener, 1918 | I | 1 | - | - | <i>Spatangus purpureus</i> O.F. Müller, 1776 | I | 2 | 2 | 2 |
| <i>Harmothoe spinifera</i> (Ehlers, 1864) | I | - | 1 | - | <i>Sphaerodoridium claparedei</i> (Greeff, 1866) | II | 1 | 1 | - |
| <i>Harpinia crenulata</i> (Boeck, 1871) | II | 42 | 21 | - | <i>Sphaerodoropsis minuta</i> (Webster & Benedict, 1887) | II | 1 | - | - |
| <i>Harpinia dellavallei</i> Chevreux, 1910 | II | 100 | 5 | 8 | <i>Sphaerosyllis glandulata</i> Perkins, 1981 | I | 4 | 2 | - |
| <i>Harpinia truncata</i> Sars, 1891 | II | 4 | - | - | <i>Sphaerosyllis hystrix</i> Claparède, 1863 | II | 2 | 11 | 5 |
| <i>Hermodice carunculata</i> (Pallas, 1766) | II | - | - | 1 | <i>Sphaerosyllis pirifera</i> Claparède, 1868 | II | - | 1 | 1 |
| <i>Hestospina aurantiaca</i> (M. Sars, 1862) | I | - | 12 | 5 | <i>Sphaerosyllis taylora</i> Perkins, 1981 | I | 42 | 1 | - |
| <i>Heteromastus filiformis</i> (Claparède, 1864) | V | 21 | 16 | 1 | <i>Sphaerosyllis thomasi</i> San Martín, 1984 | I | 16 | 2 | 1 |
| <i>Hexaplex trunculus</i> (Linnaeus, 1758) | I | 1 | - | - | <i>Spio decoratus</i> Bobretzky, 1870 | IV | 15 | 2 | 1 |
| <i>Hiatella arctica</i> (Linnaeus, 1767) | I | - | 1 | - | <i>Spio filicornis</i> (Müller, 1776) | III | - | 1 | 1 |
| <i>Hilbigneris gracilis</i> (Ehlers, 1868) | III | - | 2 | 1 | <i>Spiochaetopterus costarum</i> (Claparède, 1869) | III | 19 | 3 | 7 |
| <i>Hippomedon bidentatus</i> Chevreux, 1903 | I | 1 | - | - | <i>Spiophanes afer</i> Meißner, 2005 | I | 1 | 3 | 25 |
| <i>Hippomedon massiliensis</i> Bellan-Santini, 1965 | I | - | 1 | 2 | <i>Spiophanes bombyx</i> (Claparède, 1870) | II | 1 | - | 3 |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|-----|-----|----|--|-----|------|-----|----|
| <i>Holothuria tubulosa</i> Gmelin, 1791 | I | - | 2 | - | <i>Spiophanes kroyeri</i> Grube, 1860 | II | 4 | 1 | 2 |
| <i>Hyla vitrea</i> (Montagu, 1803) | II | 231 | 129 | 1 | <i>Spiralinella incerta</i> (Milaschewich, 1916) | I | 25 | 6 | - |
| <i>Hyalea camptonyx</i> (Heller, 1866) | I | - | - | 9 | <i>Spirobranchus triqueter</i> (Linnaeus, 1758) | II | - | 1 | - |
| <i>Inachus parvirostris</i> (Risso, 1816) | I | 1 | - | - | <i>Spirobranchus triqueter</i> (Linnaeus, 1758) | III | - | 1 | - |
| <i>Inermonephys inermis</i> (Ehlers, 1887) | II | - | 1 | - | <i>Spisula subtruncata</i> (da Costa, 1778) | III | 3 | 12 | - |
| <i>Iphimedia gibbula</i> Ruffo & Schiecke, 1979 | I | - | 1 | - | <i>Sternaspis scutata</i> Ranzani, 1817 | III | 971 | 108 | - |
| <i>Iphimedia minuta</i> G.O. Sars, 1882 | I | - | 1 | - | <i>Sthenelais boa</i> (Johnston, 1833) | II | - | 3 | 2 |
| <i>Iphinoe doumae</i> Ledoyer, 1965 | III | 29 | 24 | 9 | <i>Streblospio gynobranchiata</i> Rice & Levin, 1998 | V | 1278 | 184 | - |
| <i>Iphinoe tenella</i> Sars, 1878 | II | 41 | - | - | <i>Striarca lactea</i> (Linnaeus, 1758) | I | - | - | 1 |
| <i>Janira maculosa</i> Leach, 1814 | I | - | 1 | - | <i>Subadyte pellucida</i> (Ehlers, 1864) | II | 6 | - | 1 |
| <i>Japonactaeon pusillus</i> (Forbes, 1844) | I | 1 | - | 4 | <i>Sycon raphanus</i> Schmidt, 1862 | I | - | 2 | - |
| <i>Jassa marmorata</i> Holmes, 1905 | V | 2 | 9 | 5 | <i>Syllides edentatus</i> Westheide, 1974 | II | - | 1 | - |
| <i>Jujubinus exasperatus</i> (Pennant, 1777) | I | 1 | 9 | 1 | <i>Syllides fulvus</i> (Marion & Bobretzky, 1875) | II | - | 2 | - |
| <i>Jujubinus montagui</i> (Wood, 1828) | I | - | 1 | - | <i>Syllides japonicus</i> Imajima, 1966 | II | 2 | - | - |
| <i>Jujubinus striatus</i> (Linnaeus, 1758) | I | - | 21 | - | <i>Syllidia armata</i> Quatrefages, 1866 | IV | 3 | 1 | - |
| <i>Kuriella bidentata</i> (Montagu, 1803) | III | 125 | 56 | - | <i>Syllis alternata</i> Moore, 1908 | I | - | 2 | - |
| <i>Labioleannira yhleni</i> (Malmgren, 1867) | I | 1 | - | - | <i>Syllis armillaris</i> (O.F. Müller, 1776) | III | 3 | 2 | - |
| <i>Lacydonia miranda</i> Marion & Bobretzky, 1875 | I | - | 1 | 2 | <i>Syllis beneliahuae</i> (Campoy & Alquézar, 1982) | II | - | 1 | - |
| <i>Laetmonice hystrix</i> (Savigny in Lamarck, 1818) | I | - | 4 | - | <i>Syllis cruzi</i> Núñez & San Martín, 1991 | I | - | 2 | 1 |
| <i>Lagis koreni</i> Malmgren, 1866 | II | 6 | - | - | <i>Syllis ergeni</i> Çınar, 2005 | III | 4 | - | - |
| <i>Lanice conchilega</i> (Pallas, 1766) | IV | 6 | 2 | - | <i>Syllis garciai</i> (Campoy, 1982) | III | 47 | 41 | 5 |
| <i>Laonice bahusiensis</i> Söderström, 1920 | I | 48 | 7 | - | <i>Syllis gerlachi</i> (Hartmann-Schröder, 1960) | II | 2 | 1 | - |
| <i>Laonice cirrata</i> (M. Sars, 1851) | II | 89 | 28 | 1 | <i>Syllis gerundensis</i> (Alós & Campoy, 1981) | I | - | - | 1 |
| <i>Laubieriellus salzi</i> (Laubier, 1970) | I | - | 2 | - | <i>Syllis gracilis</i> Grube, 1840 | II | 1 | 1 | - |
| <i>Leiocapitella glabra</i> Hartman, 1947 | II | - | 1 | - | <i>Syllis hyalina</i> Grube, 1863 | I | 1 | 8 | - |
| <i>Leiochone leiopygos</i> (Grube, 1860) | II | 9 | 4 | 1 | <i>Syllis krohni</i> Ehlers, 1864 | II | 2 | - | - |
| <i>Leonnates aylaoberi</i> (Çınar & Dagli, 2013) | I | - | 4 | 1 | <i>Syllis pontxioi</i> San Martín & López, 2000 | I | 5 | - | - |
| <i>Leonnates persicus</i> Wesenberg-Lund, 1949 | II | 16 | - | 8 | <i>Syllis prolifera</i> Krohn, 1852 | II | 2 | - | 1 |
| <i>Leptocheirus mariae</i> Karaman, 1973 | III | 27 | 2 | 3 | <i>Syllis rosea</i> (Langerhans, 1879) | II | - | - | 2 |
| <i>Leptocheirus pectinatus</i> (Norman, 1869) | III | 10 | 5 | 4 | <i>Synchelidium haplocheles</i> (Grube, 1864) | II | 1 | - | 1 |
| <i>Leptochela pugnax</i> de Man, 1916 | II | - | 1 | 26 | <i>Syrnola fasciata</i> Jickeli, 1882 | II | - | 1 | 1 |
| <i>Leptochelia savignyi</i> (Krøyer, 1842) | III | 11 | 101 | 20 | <i>Tanais dulongii</i> (Audouin, 1826) | III | 1 | - | - |
| <i>Leptopentacta elongata</i> (Düben & Koren, 1846) | II | 1 | - | - | <i>Tellina albicans</i> Gmelin, 1791 | II | 2 | 1 | - |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|-----|------|-----|-----|--|-----|-----|-----|----|
| <i>Leptopentacta tergestina</i> (M. Sars, 1857) | I | - | 2 | - | <i>Tellina pulchella</i> Lamarck, 1818 | III | 29 | 16 | - |
| <i>Leucothoe liljeborgi</i> Boeck, 1861 | I | 34 | 1 | - | <i>Tellina serrata</i> Brocchi, 1814 | I | 1 | - | - |
| <i>Leucothoe occulta</i> Krapp-Schickel, 1975 | I | - | 4 | - | <i>Terebellides stroemii</i> Sars, 1835 | III | 37 | 7 | 1 |
| <i>Leucothoe serraticarpa</i> Della Valle, 1893 | I | - | 2 | 1 | <i>Teretia teres</i> (Reeve, 1844) | I | - | 3 | - |
| <i>Leucothoe spinicarpa</i> (Abildgaard, 1789) | I | - | 4 | - | <i>Thelepus cincinnatus</i> (Fabricius, 1780) | II | - | 2 | - |
| <i>Levinsenia demiri</i> Çinar, Dagli & Acik, 2011 | II | - | 57 | 5 | <i>Thracia phaseolina</i> (Lamarck, 1818) | I | 1 | - | - |
| <i>Levinsenia gracilis</i> (Tauber, 1879) | II | 1214 | - | - | <i>Thyasira flexuosa</i> (Montagu, 1803) | II | 27 | 36 | - |
| <i>Levinsenia marmarensis</i> Çinar, Dagli & Acik, 2011 | II | - | 1 | - | <i>Thysanocardia procera</i> (Möbius, 1875) | I | 3 | - | - |
| <i>Levinsenia materi</i> Çinar & Dagli 2013 | II | - | 3 | 5 | <i>Timoclea ovata</i> (Pennant, 1777) | I | 3 | 1 | 1 |
| <i>Levinsenia tribranchiata</i> (Çinar, Dagli & Açik, 2011) | II | - | 3 | - | <i>Tragula fenestrata</i> (Jeffreys, 1848) | I | - | 3 | - |
| <i>Liljeborgia dellavallei</i> Stebbing, 1906 | I | 4 | - | - | <i>Trichobranchus glacialis</i> Malmgren, 1866 | II | - | 6 | - |
| <i>Lineus cf. ruber</i> (Müller, 1974) | III | 2 | - | - | <i>Trophonopsis muricata</i> (Montagu, 1803) | I | - | - | 3 |
| <i>Linucula harvigiana</i> (Dohrn, 1864) | II | 2 | 1 | - | <i>Trypanosyllis (Trypanosyllis) coeliaca</i> Claparède, 1868 | II | - | 1 | - |
| <i>Liocarcinus maculatus</i> (Risso, 1827) | I | 1 | 2 | - | <i>Tryphosa nana</i> (Krøyer, 1846) | II | 9 | 1 | - |
| <i>Litocorsa stremma</i> Pearson, 1970 | I | 64 | 2 | 150 | <i>Tubulanus linearis</i> (McIntosh, 1874) | III | 155 | 19 | 1 |
| <i>Loripinus fragilis</i> (Philippi, 1836) | III | 5 | 2 | 1 | <i>Tubulanus polymorphus</i> Renier, 1804 | III | 64 | - | - |
| <i>Lucifer typus</i> H. Milne Edwards, 1837 [in H. Milne Edwards, 1834-1840] | III | 1 | - | - | <i>Turbonilla acuta</i> (Donovan, 1804) | I | 1 | - | - |
| <i>Lucinella divaricata</i> (Linnaeus, 1758) | I | 1 | 8 | 1 | <i>Turbonilla delicata</i> Monterosato, 1874 | I | - | 13 | - |
| <i>Lumbrineriopsis paradoxa</i> (Saint-Joseph, 1888) | I | - | 1 | 1 | <i>Turbonilla gradata</i> Bucquoy, Dautzenberg & Dollfus, 1883 | I | 3 | 1 | - |
| <i>Lumbrineris coccinea</i> (Renier, 1804) | I | - | 4 | - | <i>Turbonilla hamata</i> Nordsieck, 1972 | I | 1 | - | - |
| <i>Lumbrineris geldiyai</i> Carrera-Parra, Çinar & Dagli, 2011 | II | 821 | 214 | 93 | <i>Turbonilla jeffreysii</i> (Jeffreys, 1848) | I | 1 | - | - |
| <i>Lumbrineris latreilli</i> Audouin & Milne Edwards, 1834 | II | 12 | 18 | 3 | <i>Turritella communis</i> Risso, 1826 | II | 427 | 109 | 1 |
| <i>Lumbrineris nonatoi</i> Ramos, 1976 | II | 115 | 56 | 5 | <i>Turritella turbona</i> Monterosato, 1877 | I | - | 5 | 1 |
| <i>Lumbrineris zonata</i> Johnson, 1901 | II | 1 | - | - | <i>Upogebia pusilla</i> (Petagna, 1792) | II | 1 | - | - |
| <i>Lysianassa caesarea</i> Ruffo, 1987 | I | - | 1 | - | <i>Upogebia tipica</i> (Nardo, 1869) | II | 4 | - | 1 |
| <i>Lysianassa costae</i> (Milne Edwards, 1830) | I | - | 7 | - | <i>Urothoe elegans</i> (Bate, 1857) | I | - | 6 | - |
| <i>Lysidice ninetta</i> Audouin & H Milne Edwards, 1833 | II | - | 3 | - | <i>Urothoe grimaldii</i> Chevreux, 1895 | I | - | - | 5 |
| <i>Lysidice unicornis</i> (Grube, 1840) | II | 8 | 13 | 3 | <i>Urothoe intermedia</i> Bellan-Santini & Ruffo, 1986 | I | - | 20 | 2 |
| <i>Lysilla loveni</i> Malmgren, 1866 | II | - | 6 | - | <i>Vaunthompsonia cristata</i> Bate, 1858 | I | - | 4 | - |
| <i>Macrochaeta clavicornis</i> (M. Sars, 1835) | II | - | 4 | - | <i>Vermiliopsis infundibulum</i> (Philippi, 1844) | II | - | 1 | - |
| <i>Macrophthalmus graeffei</i> A. Milne-Edwards, 1873 | II | - | - | 10 | <i>Vexillum ebenus</i> (Lamarck, 1811) | I | - | 3 | 1 |
| <i>Maera grossimana</i> (Montagu, 1808) | I | - | 1 | - | <i>Vexillum granum</i> (Forbes, 1844) | I | 1 | - | 1 |

| Species | EG | Iz | Ag | Le | Species | EG | Iz | Ag | Le |
|--|----|-----|----|----|--|-----|----|----|----|
| <i>Maera schmidti</i> Stephensen, 1915 | I | 8 | 1 | 2 | <i>Vexillum tricolor</i> (Gmelin, 1791) | I | - | - | 1 |
| <i>Magelona alleni</i> Wilson, 1958 | II | 55 | 1 | 1 | <i>Vitreolina philippi</i> (de Rayneval & Ponzi, 1854) | I | 1 | - | - |
| <i>Magelona johnstoni</i> Fiege, Licher & Mackie, 2000 | I | - | - | 1 | <i>Volvarina mitrella</i> (Risso, 1826) | I | - | 1 | - |
| <i>Magelona minuta</i> Eliason, 1962 | II | 295 | 42 | 13 | <i>Volvulella acuminata</i> (Bruguière, 1792) | I | - | 1 | 2 |
| <i>Maldane glebifex</i> Grube, 1860 | I | 2 | - | - | <i>Websterinereis glauca</i> (Claparède, 1870) | III | 1 | - | - |
| <i>Malmgreniella lilianae</i> Pettibone, 1993 | II | 34 | 4 | - | <i>Weinkauffia turgidula</i> (Forbes, 1844) | I | - | 2 | 7 |
| <i>Malmgreniella lunulata</i> (Delle Chiaje, 1830) | II | 13 | 1 | - | <i>Westwoodilla rectirostris</i> (Della Valle, 1893) | I | 1 | 8 | 1 |
| <i>Malmgreniella polypapillata</i> Barnich & Fiege, 2001 | II | 11 | - | - | | | | | |