

Proceraea exoryxae sp. nov. (Annelida, Syllidae, Autolytinae), the first known polychaete miner tunneling into the tunic of an ascidian

Daniel Martin^{1,3}, Arne Nygren² and Edwin Cruz-Rivera³

¹ Centre d'Estudis Avançats de Blanes (CEAB-CSIC), Girona, Spain

² Sjöfartsmuseet Akvariet, Göteborg, Sweden

³ Department of Biological Sciences, University of the Virgin Islands, St. Thomas, Virgin Islands, USA

ABSTRACT

While studying organisms living in association with the solitary tunicate *Phallusia nigra* (Ascidiacea, Ascidiidae) from a shallow fringing reef at Zeytouna Beach (Egyptian Red Sea), one of the collected ascidians showed peculiar perforations on its tunic. Once dissected, the perforations revealed to be the openings of a network of galleries excavated in the inner tunic (atrium) by at least six individuals of a polychaetous annelid. The worms belonged to the Autolytinae (Syllidae), a subfamily that is well known to include specialized predators and/or symbionts, mostly associated with cnidarians. The Red Sea worms are here described as *Proceraea exoryxae* sp. nov., which are anatomically distinguished by the combination of simple chaetae only in anterior chaetigers, and a unique trepan with 33 teeth in one outer ring where one large tooth alternates with one medium-sized tricuspid tooth, and one inner ring with small teeth located just behind the large teeth. Male and female epitokes were found together with atokous individuals within galleries. *Proceraea exoryxae* sp. nov. constitutes the first known miner in the Autolytinae and the second species in this taxon known to live symbiotically with ascidians. The implications of finding this specialized parasite are discussed considering that *Phallusia nigra* has been introduced worldwide, in tropical and sub-tropical ecosystems, where it has the potential of becoming invasive.

Subjects Taxonomy, Zoology

Keywords Parasitism, Red Sea, *Phallusia nigra*, Coral Reef

INTRODUCTION

There are approximately 11,840 polychaete annelids known, spanning a remarkable array of habitats, ecological niches, and trophic modes (*Read & Fauchald, 2016*). Among these, symbiotic species (sensu *Castro, 2015*) span at least 28 different families (*Martin & Britayev, 1998*). These symbiotic interactions, in general, are poorly understood, but cases of inquilinism, commensalism, mutualism and parasitism have been documented. Interestingly, parasitism seems to be among the least common modes of life for polychaetes (<0.5% of known species, spread among 13 families), most of them being found within the Spionidae and most often being shell borers

Submitted 29 March 2017

Accepted 4 May 2017

Published 1 June 2017

Corresponding author

Edwin Cruz-Rivera,
edwin.cruzrivera@uvi.edu

Academic editor

Christopher Glasby

Additional Information and
Declarations can be found on
page 18

DOI 10.7717/peerj.3374

© Copyright
2017 Martin et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

([Martin & Britayev, 1998](#)). Several reports of associations with tunicates (Phylum Chordata) are available ([Okada, 1935](#); [Spooner, Wilson & Trebble, 1957](#); [Fiore & Jutte, 2010](#)), but the polychaetes have not been identified in some of these instances ([Illg, 1958](#); [Monniot, 1990](#)). There are few details known for these associations although consumption of the ascidian host has been reported in one case ([Spooner, Wilson & Trebble, 1957](#)).

Phallusia nigra [Savigny, 1816](#), is a solitary ascidian that has been introduced into tropical and subtropical ecosystems worldwide since it was originally discovered in the Red Sea ([Shenkar, 2012](#); [Vandepas et al., 2015](#); [Zhan et al., 2015](#)). The ascidian hosts a remarkable array of crustacean symbionts, including amphipods and at least eight confirmed copepod species ([Kim et al., 2016](#)). During studies on the ecology of *Phallusia nigra* and its associated fauna in the Egyptian coast of the Red Sea, one of the collected specimens showed various perforations on its tunic. Upon dissection, we discovered a network of excavated galleries resembling the habit of some leaf-mining herbivores in terrestrial and marine habitats ([Brearley & Walker, 1995](#); [Connor & Taverner, 1997](#); [Sinclair & Hughes, 2010](#); [Mejaes, Poore & Thiel, 2015](#)). The galleries were inhabited by several specimens of a small polychaete species belonging to the subfamily Autolytinae (Annelida, Syllidae). Although some bivalves and crustaceans have been reported to live within ascidian tunics ([Lambert, 2005](#); [McClintock et al., 2009](#); [Morton & Dinesen, 2011](#); [Cañete & Rocha, 2013](#)), no previous reports of annelids exhibiting a similar habit are known ([Lambert, 2005](#); [Monniot, 1990](#)).

The Autolytinae are small free-living polychaetes, ranging from 1 to 60 mm long and from 0.1 to 1.2 mm wide. They are distributed worldwide and inhabit shallow waters, mostly restricted to the continental shelf. They often live in a more or less intimate association with sedentary invertebrates on which they supposedly feed, such as cnidarians (usually hydroids), but also bryozoans, sponges and tunicates ([Okada, 1928](#); [Hamond, 1969](#); [Fauchald & Jumars, 1979](#); [Genzano & San Martín, 2002](#); [Nygren, 2004](#); [Nygren & Pleijel, 2007](#); [Martin et al., 2015](#)). Autolytines are commonly found living inside thin, semi-hyaline tubes, either made in association with the host or secreted by the worms and attached directly to the colonial animals with which they associate ([Gidholm, 1967](#); [Fischer, Mewes & Franke, 1992](#); [Genzano & San Martín, 2002](#)).

Autolytinae constitute a phylogenetically well-delineated group of polychaetes in the family Syllidae ([Aguado & San Martín, 2009](#)), characterized by a sinuous pharynx, absence of ventral cirri, presence of simple bayonet-type dorsal chaetae, and reproduction with dimorphic sexes ([Franke, 1999](#); [Nygren & Sundberg, 2003](#); [Nygren, 2004](#)). Since the comprehensive revision by [Nygren \(2004\)](#), numerous new species have been described ([Çinar & Gambi, 2005](#); [Nygren & Pleijel, 2007](#); [Lucas, San Martín & Sikorski, 2010](#); [Nygren et al., 2010](#); [Álvarez-Campos, San Martín & Piotrowski, 2014](#); [Çinar, 2015](#); [Dietrich et al., 2015](#); [Martin et al., 2015](#); [Aguirre, San Martín & Álvarez-Campos, 2016](#)). Currently, the subfamily comprises 180 nominal species, of which 112 are considered valid and distributed among 13 recognized genera ([Nygren & Pleijel, 2007](#); [Nygren et al., 2010](#); [Rivolta, San Martín & Sikorski, 2016](#)). Among them, *Proceraea* Ehlers, 1,864 contains 28 species ([Nygren, 2004](#); [Nygren et al., 2010](#); [Martin et al., 2015](#)).

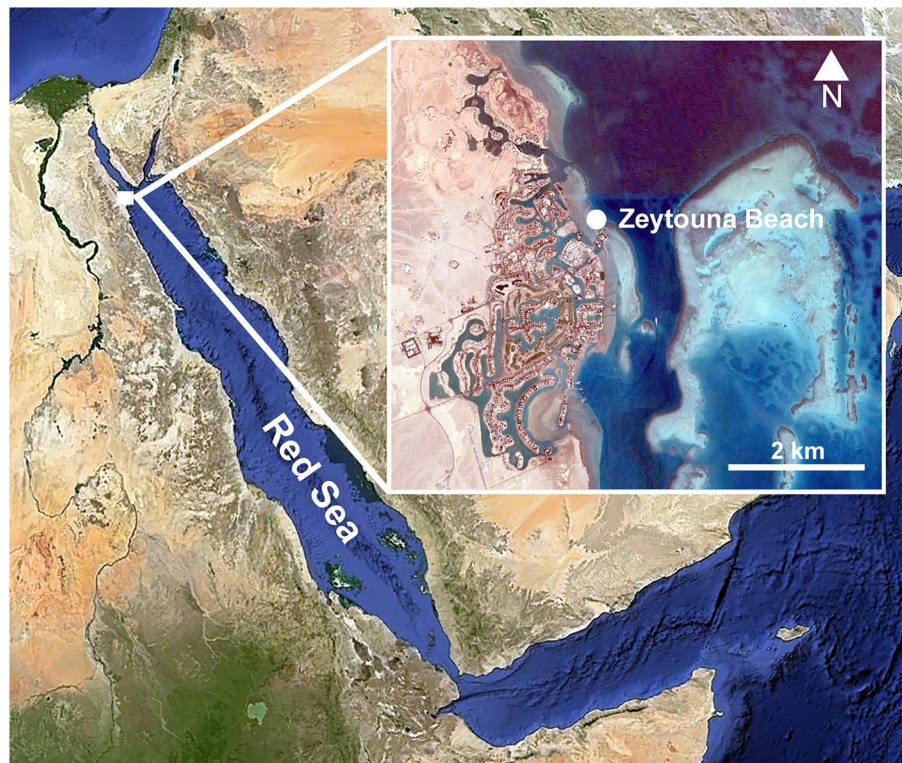


Figure 1 Location of the sampling site. Zeytouna Beach, Egyptian coast of the Red Sea. Maps are from Google Earth Pro, © 2016 DigitalGlobe, © 2016 CNS/Astrium.

It is in *Proceraea* that we place the new species herein described, which occurs inside galleries excavated in the tunic of *Phallusia nigra* and is, thus, the first known miner autolytine. This finding led us to discuss the current knowledge on symbioses involving autolytines, as well as the possible ecological implications of the symbiotic relationship between the polychaete and its host ascidian.

MATERIALS AND METHODS

Individuals of *Phallusia nigra* were collected by SCUBA from the shallow fringing reef at Zeytouna Beach, on the Egyptian Red Sea ($27^{\circ}24'09.2''\text{N}$ $33^{\circ}41'08.5''\text{E}$; Fig. 1) under the auspices of the John D. Gerhart Field Station in El Gouna (American University in Cairo), with permission from the management of Zeytouna Beach. All ascidians were collected on October 7, 2010 at 3–7 m depth and brought to the El Gouna Field Station. In the laboratory, the specimens of *Phallusia nigra* ($N = 50$) were dissected with an incision around the entire periphery of the tunic, and the visceral mass and the pharyngeal sac were removed (Fig. 2A). All of them were inspected for associated animals. Ascidian masses and any abnormalities or damage on the hosts were recorded. Dissected hosts and symbionts from the atrial cavity were photographed with a digital camera equipped with a macro lens.

The entire tunic of the infested ascidian specimen was placed in formaldehyde for a few seconds. Then, the galleries were cut with an angular-tipped scalpel through the atrial

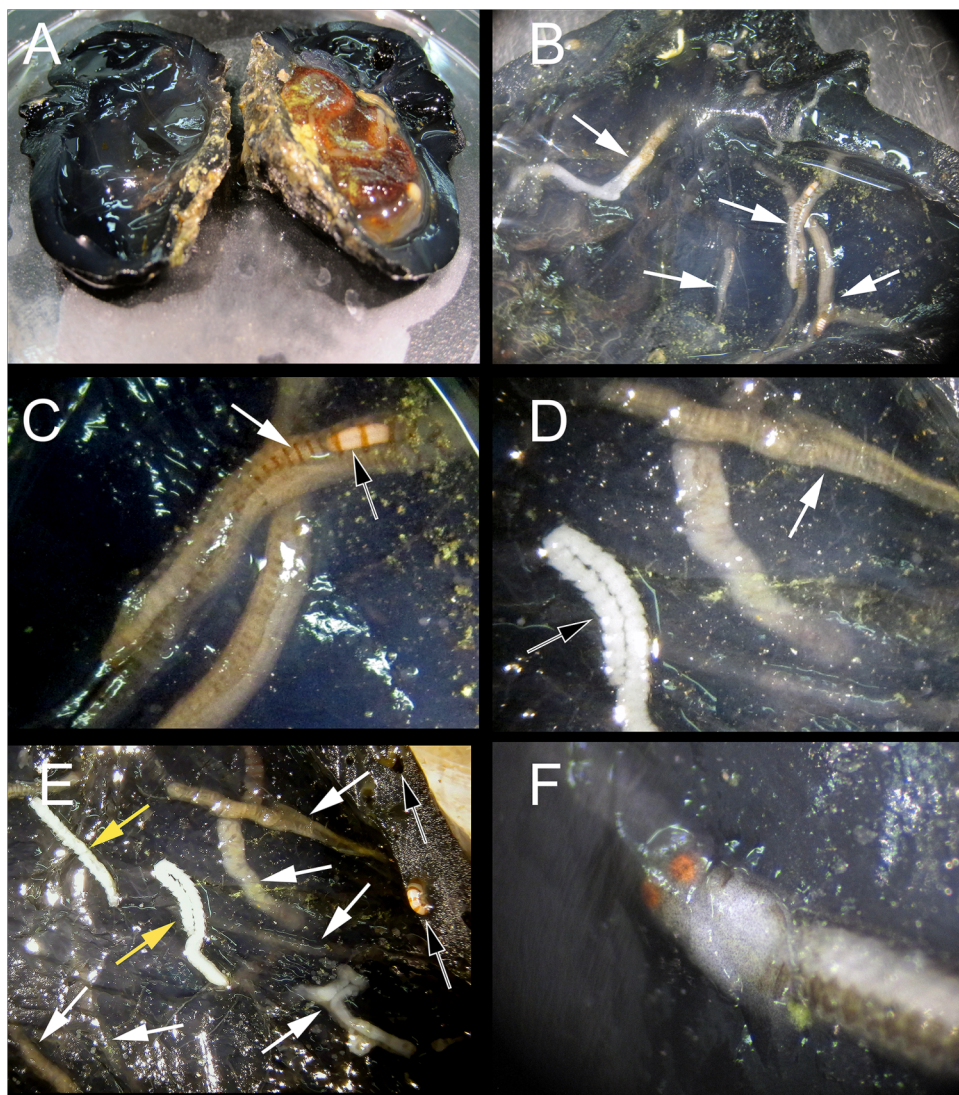


Figure 2 Host dissection and location of mining polychaetes. (A) An uninfected *Phallusia nigra* dissected to show normal atrial surface (left) and internal organs. (B) Inner atrial surface showing the presence of several atokous forms inside their galleries (white arrows). (C) Detail of the color of the anterior region of an atokous form; white arrow: position of pharynx; black arrow: position of proventricle. (D) Detail of color of the mid-body of an atokous form (white arrow) and the posterior region of a male epitokous form (black arrow). (E) Inner atrial surface showing the presence of atokous (white arrows) and male epitokous (yellow arrows) forms inside their galleries, as well as part of the banded body of an atokous form protruding from an external tunic opening and other empty tunic openings (black arrows). (F) Close-up view of the head of a male epitoke in its gallery (specimen not preserved).

surface of the tunic to extract mining autolytines with the help of fine forceps. However, their body was very delicate and all of them broke during extraction. In fact, some stolons were completely destroyed in the process and it was not possible to save them for taxonomic studies. All obtained specimens were fixed and preserved in a 4% formalin-seawater solution and transferred to 70% ethanol prior to observations.

Light microscope photos were taken with a Canon EOS 5D Mark II connected to either a Zeiss KF2 triocular microscope via a LM-Scope TUST42C coupler, or a Canon EF

65 mm macro lens with one to five times magnification. For scanning electron microscope (SEM) observations, the 70% ethanol preserved materials were prepared using standard SEM procedures (*Martin et al., 2003*). Prior to run the SEM procedures to observe the trepan, this structure was carefully dissected and as much as possible cleaned from the external muscular tissue layer. Images were taken in a Hitachi TM3000 TABLETOP microscope at the SEM service of the CEAB–CSIC.

The electronic version of this article in portable document format (PDF) will represent a published work according to the International Commission on Zoological Nomenclature (ICZN), and hence the new names contained in the electronic version are effectively published under that Code from the electronic edition alone. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (life science identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix <http://zoobank.org/>. The LSID for this publication is: urn:lsid:zoobank.org:pub:685CB1C2-CB5B-4A87-9CD7-C04BFFDE03B4. The online version of this work is archived and available from the following digital repositories: PeerJ, PubMed Central and CLOCKSS. Specimen vouchers were deposited at the Museo Nacional de Ciencias Naturales of Madrid, Spain (MNCN).

RESULTS

Taxonomic account

Phylum ANNELIDA Lamarck, 1809

Subclass ERRANTIA Audouin & Milne-Edwards, 1832

Order PHYLLODOCIDA Dales, 1962

Suborder NEREIDIFORMIA

Family SYLLIDAE Grube, 1850

Subfamily AUTOLYTINAE Langerhans, 1879

Tribe PROCERINI *Nygren, 2004*

Genus *Proceraea* Ehlers, 1864

Proceraea exoryxae sp. nov.

LSID. urn:lsid:zoobank.org:act:34373CE6-A0D4-488D-B4A5-12CF4E103504

(Figs. 2–7)

Type material: Holotype. MNCN 16.01/17717: atokous anterior fragment, Zeytouna Beach, Egyptian Red Sea, 27°24'09.2"N 33°41'08.5"E, October 7 2010, 3–7 m depth, E. Cruz-Rivera coll.; fixed in 4% formalin seawater, preserved in 70% ethanol. Paratypes. MNCN 16.01/17718: atokous anterior fragment, pharynx dissected; MNCN 16.01/17719: atokous specimen, anterior fragment (up to chaetiger 10) prepared for SEM, mid-body segments and dissected proventricle preserved in 70% ethanol; MNCN 16.01/17720: atokous anterior fragment, pharynx dissected; MNCN 16.01/17721: male stolon, anterior fragment; MNCN 16.01/17722: female

stolon, anterior fragment; MNCN 16.01/17723: atokous mid-body fragments. MNCN 16.01/17724: atokous posterior fragments. Collection details for all other types deposited are the same as for holotype.

Diagnosis: *Proceraea* with simple chaetae in anterior chaetigers, and a trepan with 33 teeth with one outer ring where one large tooth alternates with one medium-sized tricuspid tooth, and one inner ring with small teeth located just behind the large teeth.

Description: All observations are from preserved specimens if not otherwise stated. Length 3–10.5 mm for 10–68 chaetigers in four anterior fragments, 3–14.5 mm for 19–90 chaetigers in nine median fragments, and 6.5–22 mm for 50–125 chaetigers in three posterior fragments. Width of anterior fragments, excluding parapodial lobes, c. 0.4 mm. Live individuals dorsally with light brown transverse stripes, one per segment, not known whether these are inter- or intrasegmental, or if there is any other additional coloration (Figs. 2B–2E); proventricle white (Figs. 2B and 2C). Formalin preserved specimens without any sign of coloration.

Body shape, excluding parapodial lobes, cylindrical in transection, ventrally flattened. Body long and slender, with slowly tapering end. Nuchal organs ciliated. Prostomium rounded rectangular (Figs. 3A and 3C). Four eyes with lenses, anterior pair larger, confluent in dorsal view, eye spots absent (Fig. 3C). Palps in dorsal view projecting c. half of prostomial length, fused (Figs. 3A and 3B).

Nuchal organs extending to median part of chaetiger 1 (Fig. 3A(A1)). Prostomium with three antennae, median antenna inserted medially on prostomium, lateral antennae on anterior margin. Median antenna reaching chaetiger 8–10, lateral antennae about half as long as median antenna. Tentacular cirri two pairs. Dorsal tentacular cirri about two third as long as median antenna, ventral tentacular cirri about half as long as dorsal tentacular cirri. First dorsal cirri about as long as median antenna, second dorsal cirri as long as ventral tentacular cirri. From chaetiger 3 to chaetiger 20–25, cirri alternate indistinctly in length, shorter cirri slightly shorter and longer cirri equal or slightly longer than body width excluding parapodial lobes (Figs. 3A and 3B), dorsal cirri in more posterior chaetigers more or less equal in length, c. half of body width excluding parapodial lobes; anal cirri as long as half body width, excluding parapodial lobes at level of proventricle.

Cirrophores on tentacular cirri, first and second dorsal cirri (Fig. 3A), otherwise absent. Antennae, tentacular cirri, dorsal cirri, and anal cirri cylindrical. Parapodial lobes rounded. Aciculae 2–3 in anterior chaetigers, 1–2 in median and posterior chaetigers, straight, with a round, swollen distal end (Fig. 4G).

Chaetal fascicle with 9–12 chaetae in anterior chaetigers (Fig. 4A), 4–10 in median and posterior chaetigers. Chaetiger 1–5 with simple chaetae only (Figs. 4B and 5A–5D), chaetiger 6 with simple chaetae only ($n = 3$), or with single compound chaeta in addition to the simple chaetae ($n = 1$). From chaetiger 7 to between chaetiger 10–13 with an increasing proportion of compound chaetae (Fig. 4A). Except for the single, thick, distally denticulated bayonet chaeta (Fig. 5E), starting at the earliest in chaetiger 9,

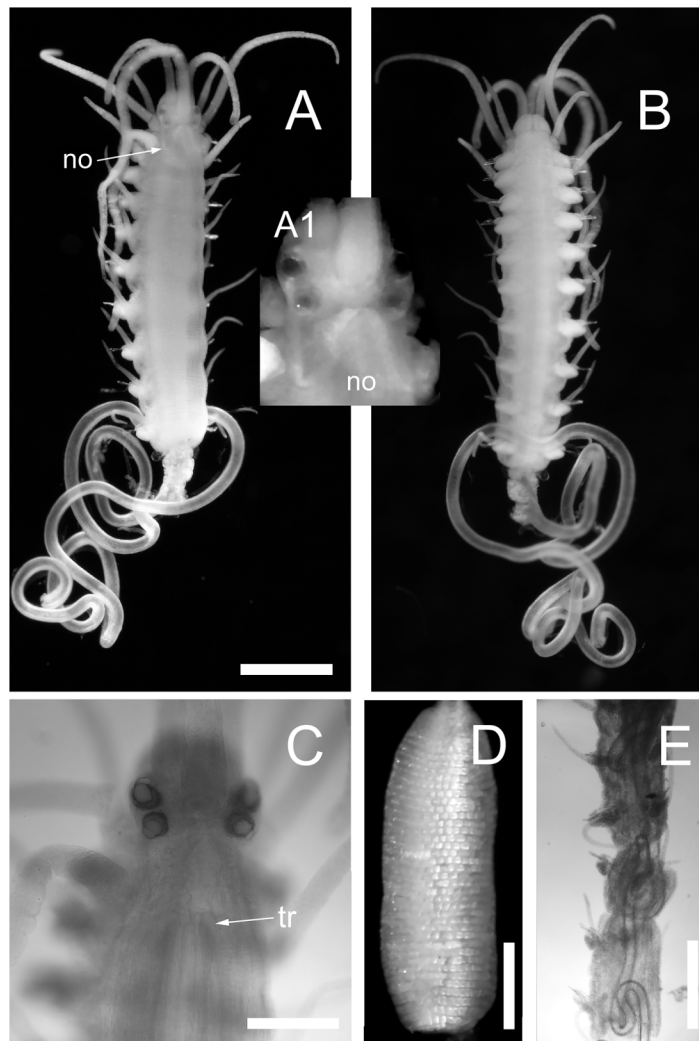


Figure 3 *Proceraea exoryxae* sp. nov. (A) Anterior fragment, dorsal view [MNCN 16.01/17719], and detail of the head from the same specimen (A1). (B) Anterior fragment, ventral view [MNCN 16.01/17719]. Body is broken after chaetiger 10, exposing the pharynx (A and B). (C) Anterior end, dorsal view [MNCN 16.01/17719]. (D) Proventricule [MNCN 16.01/17719]. (E) Pharynx sinuation in chaetigers 9–14, dorsal view [MNCN 16.01/17720]. no, nuchal organs; tr, position of trepan. Scale bars A, B, E = 0.5 mm, C, D = 0.2 mm.

more posterior chaetigers with compound chaetae only. Simple chaetae unidentate with rows of spines subdistally (Figs. 4B, 5A–5D and 5F). In anterior 4–5 chaetigers most simple chaetae with a proportionally long region distal to the swollen neck (Figs. 4B and 5B–5D), one or two of the inferior-most chaetae with a shorter region distal to the swollen neck (Figs. 4B, 5A, 5C and 5D), similar in appearance to the shafts of the compound chaetae found in later chaetigers. Starting from chaetigers 6–7 all simple chaetae (except for the bayonet chaeta) nearly identical to the shafts of the compound chaetae (Figs. 4B and 5E). Blades of compound chaetae serrated, with two large distal teeth, distal-most slightly smaller, becoming smaller to almost disappear in mid-body and posterior chaetigers, shafts with a swollen neck with rows of spines (Figs. 4B and 5E).

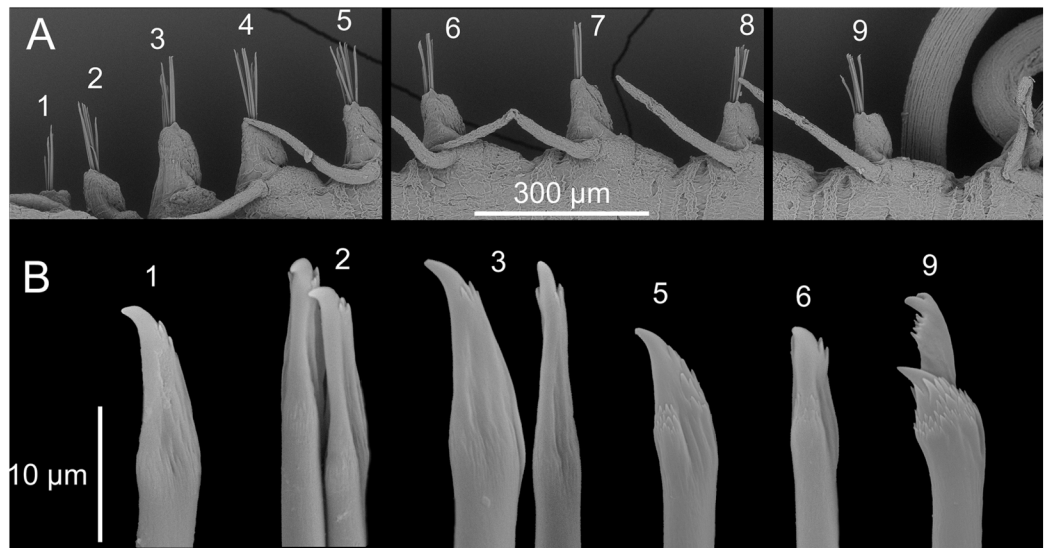


Figure 4 *Proceraea exoryxae* sp. nov. SEM micrographs of chaetae structure [MNCN 16.01/17719]. (A) Chaetigers 1–9. (B) Chaetae: 1–3, simple chaetae with long region distal to the swollen neck from chaetigers 1–3; 5–6, simple chaetae with short region distal to the swollen neck from chaetigers 5 and 6; 9, compound chaetae from chaetiger 9.

Pharynx with several sinuations (Figs. 2C and 3E), mostly anterior to the proventricle, exact sinuation difficult to assess. Trepan at level of chaetiger 1–2 (Fig. 3C), with 33 teeth with one outer ring where one large tooth alternates with one medium-sized tricuspid tooth, and one inner ring with small teeth located just behind the large teeth (Figs. 6A–6C). Basal ring present, infradental spines absent. Proventricle as long as three segments in chaetiger 20–22 (uncertain observation, as the single specimen with proventricle still inside body looks distorted in this region), with 40–45 rows of square-shaped muscle cells ($n = 2$) (Figs. 2C and 3D).

Distribution: Known only from the type locality, Zeytouna Beach (Egyptian coast of the Red Sea).

Etymology: The specific epithet “*exoryxae*” derives from the term εξόρυξη, which means miner in Greek.

Ecology: *Proceraea exoryxae* sp. nov. was extremely rare. It was only found in one *Phallusia nigra* specimen, despite multiple successive collections of this ascidian in the same and other reefs during following years (Kim *et al.*, 2016). The excavated galleries in which the new autolytine resided were visible only through the atrium wall (the internal surface of the tunic; Figs. 2B–2F), whereas the outside surface of the host tunic showed no signs of deformation, aside from the entrance openings of the galleries (Fig. 2E). The inner walls of the galleries were covered by a thin hyaline layer, apparently secreted by the worms. The wet mass of the individuals of *Phallusia nigra* collected in this reef ranged from 7.32 to 13.25 g and the specimen containing *Proceraea exoryxae* sp. nov. was 11.10 g. Two individuals of the amphipod *Leucothoe furina* (Savigny, 1816), a common

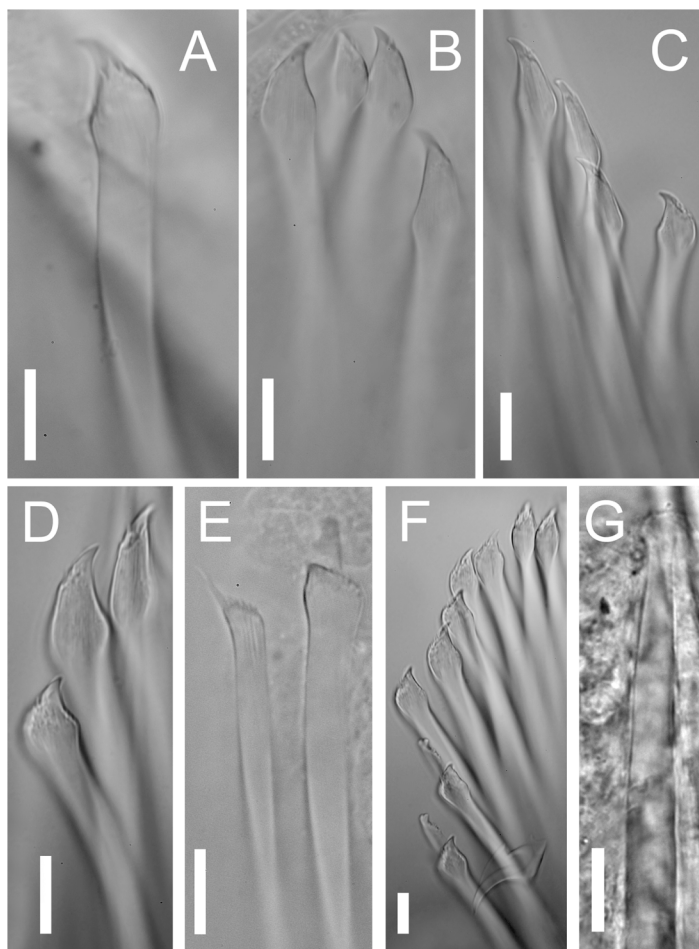


Figure 5 *Proceraea exoryxae* sp. nov. morphology of chaetae and aciculae. (A) Inferior-most simple chaeta, chaetiger 1 [MNCN 16.01/17717]. (B) Simple chaetae, chaetiger 3 [MNCN 16.01/17717]. (C) Simple chaetae, chaetiger 4 [MNCN 16.01/17719]. (D) Simple chaetae, chaetiger 5 [MNCN 16.01/17719]. (E) Simple and compound chaetae, chaetiger 10 [MNCN 16.01/17719]. (F) Bayonet chaeta and compound chaeta, chaetiger 9 [MNCN 16.01/17717]. (G) Mid-body acicula [MNCN 16.01/17723]. Scale bars A–G = 0.1 mm.

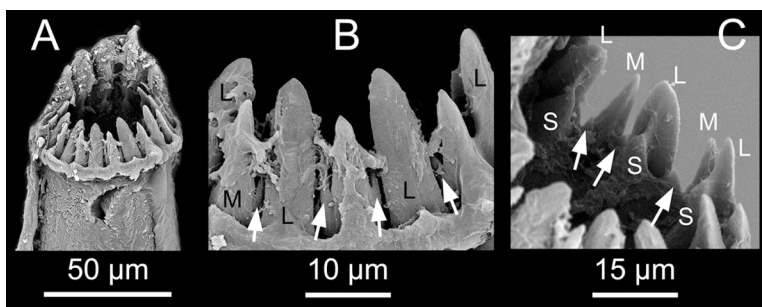


Figure 6 *Proceraea exoryxae* sp. nov. SEM micrographs of trepan structure. (A) Whole view of the trepan (teeth on the back partly covered by tissue) [MNCN 16.01/17720]. (B) Large and medium, tricuspid teeth in external view. (C) Large, medium, tricuspid and small teeth in internal view. L, Large teeth; M, Medium, tricuspid teeth; S, small teeth; white arrows pointing on the lateral cusps.

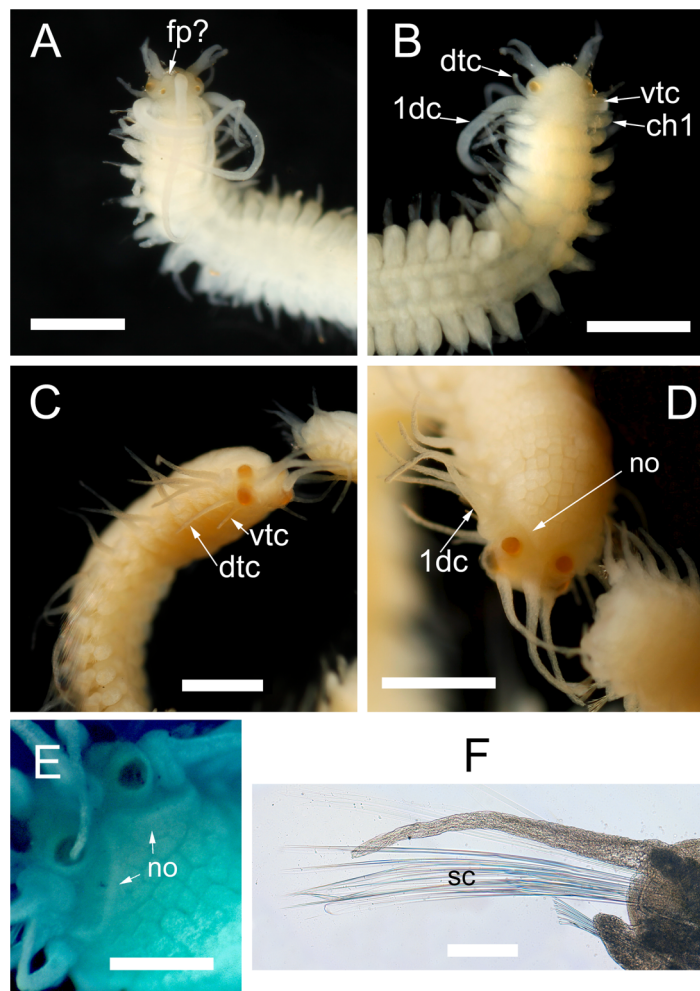


Figure 7 *Proceraea exoryxae* sp. nov. morphology of epitokes. Anterior end of male stolon [MNCN 16.01/17721]: (A) dorsal view; (B) ventral view. Anterior end of female stolon [MNCN 16.01/17722]: (C) anteroventral view; (D) dorsal view; (E) detail of prostomium showing the nuchal organs (stained with methyl blue). (F) Mid-body parapodia of female stolon showing the swimming chaetae [MNCN 16.01/17722]. dte, dorsal tentacular cirri; vtc, ventral tentacular cirri; 1dc, first dorsal cirri; ch1, chaetiger 1; fp, frontal process; no, nuchal organs; sc, swimming chaetae. Scale bars A–D = 0.5 mm, E, F = 100 μ m.

associate of *Phallusia nigra* in the Egyptian Red Sea (White, 2011) were also found in the same host.

Reproduction: Probably with schizogamy, as several male and female stolons were found in the same galleries as the atokous forms (Figs. 2D–2F). Stolons were obtained detached from the corresponding stocks. However, they have bayonet and compound chaetae identical to those in the atokous forms, thus strengthening the hypothesis that they belong to *Proceraea exoryxae* sp. nov. Male and female stolons are described below.

Morphology of the epitokous male. Length 2.7 mm for 6+17 chaetigers in region a and b (Nygren, 2004), incomplete; width in region a 0.4 mm excluding parapodial lobes, in region b 0.7 mm including parapodial lobes. Exact color in vivo unknown, but either the ventral or the dorsal side of region b dark brownish, region a whitish, with diffuse darker

pigmentation (Fig. 2F). Preserved specimen whitish, without color markings, chaetiger 2–6 with paired yellowish sperm glands seen through the body wall (Fig. 7B). Prostomium rounded rectangular, wider than long, anterior margin convex. Four eyes with lenses, situated ventro-lateral and dorsal on prostomium, ventro-lateral pair larger (Figs. 7A and 7B). Palps absent. Nuchal organs not seen. Median antenna inserted medially on prostomium, reaching c. chaetiger 10; lateral bifid antennae, inserted on anterior margin, equal in length to prostomial width; basal part 1/3 of total length, outer ventral rami slightly longer and thinner than inner dorsal rami. Frontal processes possibly absent, or developing (seen as small protuberances on prostomium lateral to the median antenna) (Fig. 7A). Tentacular cirri 2 pairs (Fig. 7B), dorsal tentacular cirri, as long as 1/2 prostomial width, ventral tentacular cirri, 1/3 as long as dorsal pair. First dorsal cirri, equal in length to median antenna. Achaetous knobs absent. Cirri in region a reciprocally equal, equal in length to 1/2 body width excluding parapodial lobes, cirri in region b reciprocally equal, slightly shorter than cirri in region a. Short median ceratophore, and short cirrophores on first dorsal cirri, cirrophores otherwise absent. Median antenna, tentacular cirri, first dorsal cirri, and cirri in region a and b cylindrical. Parapodia in region a uniramous, neuropodial lobes rounded, parapodia in region b with developing notopodial lobes. Single neuropodial acicula in all chaetigers; 2 anterodorsal and 5 posteroventral notopodial aciculae in region b. Neuropodial fascicle with 7–8 compound chaetae and a single bayonet chaeta of the same types described for the atoke. Swimming chaetae absent, indicating a non-fully developed male stolon.

Morphology of the epitokous female: Length 5 mm for 6+27+9 chaetigers in region a, b and c (Nygren, 2004), incomplete; width in region a and c 0.6 mm excluding parapodial lobes, in region b 1 mm including parapodial lobes. Color of living individuals unknown. Preserved specimen yellowish, with body filled by eggs (Fig. 7D); color markings absent. Prostomium rounded rectangular, wider than long, anterior margin straight. Four eyes with lenses, situated ventro-lateral and dorsal on prostomium, ventro-lateral pair larger (Figs. 7C and 7D). Palps absent. Nuchal organs reaching beginning of chaetiger 1 (Figs. 7D and 7E). Median antenna inserted medially on prostomium, reaching c. chaetiger 5; lateral antennae inserted on anterior margin, about two third in length of median antenna. Tentacular cirri 2 pairs (Fig. 7C), dorsal tentacular cirri, as long as prostomial width, ventral tentacular cirri about 1/2 as long as dorsal pair. First dorsal cirri (Fig. 7D), equal in length to lateral antennae. Achaetous knobs absent. Cirri in region a reciprocally equal, slightly shorter than first dorsal cirri, equal in length to body width excluding parapodial lobes, cirri in region b reciprocally equal, slightly longer than cirri in region a, cirri in region c reciprocally equal, slightly shorter than cirri in region a. Ceratophores on median and lateral antennae, cirrophores present on all dorsal cirri, but tentacular cirri without cirrophores. Antennae, tentacular cirri, and dorsal cirri cylindrical. Parapodia in region a uniramous, neuropodial lobes rounded, parapodia in region b with additional notopodial lobes. Single neuropodial acicula in all chaetigers; 2–3 anterodorsal and 6–7 posteroventral notopodial aciculae in region b. Neuropodial fascicle with 7–8 compound chaetae and a single bayonet chaeta of the same types

described for the atokous form. Notopodial chaetal fascicle with 20–25 long and thin swimming chaetae (Fig. 7F).

DISCUSSION

Taxonomic remarks

The combination of morphological features in *Proceraea exoryxae* sp. nov. makes it difficult to place it in any specific genus within the Autolytinae. The thick type of bayonet chaeta, distally denticulated, and the presence of cirrophores only on anterior-most cirri indicate, however, that the new species is a member of the tribe Procerini.

Accordingly, these morphological characters are not found in either of the two other main groups (Autolytini and *Epigamia*).

We place the new species in the genus *Proceraea* even though simple chaetae (apart from bayonet chaetae) are not found in any described member. We base our decision on the fact that the trepan teeth in *Proceraea exoryxae* sp. nov. are arranged in more than one ring, which is only found in *Proceraea* among Procerini. The observation of the trepan rings is clear under SEM, despite part of the dissected pharynx being still covered by tissue. The presence of simple chaetae in a restricted number of anterior chaetigers is a feature shared only with some members of *Procerastea* and *Imajimaea* among the Autolytinae, which differ in having trepans with a single ring, instead of separate rings as in *Proceraea exoryxae* sp. nov. Moreover, simple chaetae in *Proceraea exoryxae* sp. nov. differ from those in these two genera in that there seems to be two types. The first one (Figs. 4B(1–3)) has a peculiar morphology with an enlarged, hooked tip surrounded by a distal half crown of small denticles. In the second type, the hooked tip progressively reduces its length (e.g., Fig. 4(5–6)) to finally acquire a shape almost indistinguishable from the blades of compound chaetae (Fig. 4(9)). Only the first type of chaeta is present in the first chaetigers, and its number is progressively reduced to disappear around chaetiger 9–10. Conversely the second type progressively increased in number to be finally replaced by compound chaetae around chaetiger 10. At this level, it is almost impossible to distinguish between a simple chaeta and a compound one that has lost the blade. However, in mid-body and posterior segments, the presence of compound chaetae without blades is extremely rare. This, together with the fact that there is an antero-posterior gradation in tip length in the second type of simple chaetae is evidence that they are actually simple chaetae instead of compound ones without blade.

Further, all members of *Procerastea* have thick, distally dilated, bayonet chaetae and have dorsal cirri only on the first chaetiger, while *Proceraea exoryxae* sp. nov. has thick bayonet chaetae not distally dilated and dorsal cirri on all segments. *Imajimaea*, on the other hand, shares the presence of dorsal cirri on all its segments, except for *Imajimaea draculai* that lacks dorsal cirri on chaetigers 2–5. However, all species of *Imajimaea* have thin, subdistally denticulated, bayonet chaetae, and not thick bayonet chaetae, distally denticulated as in *Proceraea exoryxae* sp. nov.

Assuming that the assignment of the two stolons to this new species is correct, this may also shed some light on its taxonomic relationships. The type of stolon with six chaetigers in region a, two pairs of tentacular cirri, and no achaetous knobs is found in

Virchowia clavata Langerhans, 1879, *Virchowia pectinans* (Hartmann-Schröder, 1983), *Autolytus emertoni* Verrill, 1881, *Procerastea halleziana* Malaquin, 1893, *Procerastea nematodes* Langerhans, 1884 and *Proceraea picta* Ehlers, 1864 (Nygren, 2004).

The information on the stolons of *Virchowia clavata*, *Procerastea* spp. and *Proceraea picta* is based on the literature only, but even in the species where the stolons are thoroughly described and illustrated, the achaetous knobs are not detailed. All other species assigned to *Proceraea* for which stolons are known, i.e., *Procerastea cornuta* (Agassiz, 1862), *Poecilotheria fasciata* Bosc, 1802, *P. hanssoni* Nygren, 2004, *Pseudozonaria nigropunctata* Nygren & Gidholm, 2001, *P. okadai* (Imajima, 1966), and *P. prismatica* (Müller, 1776), are equipped with achaetous knobs ventral to the first dorsal cirri. It is important to note that molecular phylogenetic studies have found the genus *Proceraea* to be paraphyletic without the inclusion of e.g., *Virchowia* and *Procerastea* (Nygren et al., 2010). *Proceraea picta* and close relatives are found as a sister group to a clade where the *Proceraea* having stolons with achaetous knobs constitute the first subclade, and *Virchowia*, *Procerastea* and other genera, whose known stolons lack achaetous knobs, constitute the second subclade. This indicates that having stolons with achaetous knobs is the derived state, while the lack of achaetous knobs is plesiomorphic. A revision of *Proceraea* is clearly needed, but beyond the scope of this paper. Until then, we consider a generic assignment to *Proceraea* for this new species to be the best option.

Autolytinid symbionts

Despite the hundreds of symbiotic polychaete species known, including >80 species considered parasitic, parasitism is relatively rare in this taxon when compared to other lifestyles (Martin & Britayev, 1998; Britayev & Lyskin, 2002; Britayev et al., 2014). Most recorded symbiotic associations between polychaetes and invertebrates involve sponge, cnidarian, mollusc, or echinoderm hosts, but a few mention ascidians. Some previous reports of polychaetes living among ascidians came from dredged or scraped-quadrat samples, which are usually inadequate to determine symbiont–host associations because they result in specimen mixtures, while soft-bodied animals, like tunicates or annelids, are often greatly damaged. In this context, the term “associated with” most often refers to spatially co-occurring specimens. Nonetheless, high densities of polychaetes, including syllids, are known to occur among the fauna associated with particular benthic tunicates (Allen, 1915; Fielding, Weerts & Forbes, 1994; Cerdá & Castilla, 2001; Fiore & Jutte, 2010; Sepúlveda et al., 2015). Polychaetes were dominant on intertidal (but not subtidal) beds of *Pyura stolonifera* (Heller, 1878), the second most abundant group in beds of the congeneric *Pyura praeputialis* (Heller, 1878) (Fielding, Weerts & Forbes, 1994; Cerdá & Castilla, 2001), and constituted 28% of the fauna associated with didemnid ascidians (Fiore & Jutte, 2010). These reports, however, largely refer to animals living in the sediments accumulated in the crevices among ascidian aggregates and, thus, there is no reason to suspect true symbiotic interactions. Similarly, intraspecific variation in growth form of *Pyura chilensis* Molina, 1782 has been documented in response to the presence of chaetopterid polychaete tubes in the assemblage (Sepúlveda et al., 2015), but this was interpreted as a density-dependent phenomenon not related to symbiosis.

Table 1 List of known autolytinid syllids reported as symbionts.

Symbiont	Host	References
<i>Proceraea</i> sp.	Cn <i>Abietinaria turgida</i> (Clarke, 1877)	Britayev & San Martín (2001)
	Cn <i>Orthopyxis integra</i> (Macgillivray, 1842)	T. A. Britayev, 2015, personal communication
<i>Imajimaea draculai</i> (San Martín & López, 2002)	Cn <i>Funiculina quadrangularis</i> (Pallas, 1766)	Nygren & Pleijel (2010)
<i>Myrianida piningera</i> (Montagu, 1808)	Tu <i>Phallusia mammillata</i> (Cuvier, 1815)	Okada (1935), Spooner, Wilson & Trebble (1957)
	Tu <i>Asciadiella aspersa</i> (O. F. Müller, 1776)	Okada (1935), Spooner, Wilson & Trebble (1957)
<i>Procerastea cornuta</i> Agassiz, 1862	Cn Unidentified hydroid	Pettibone (1963)
	Cn Unidentified Coral	Gardiner (1976)
<i>Procerastea halleziana</i> Malaquin, 1893	Cn <i>Ectopleura crocea</i> (Agassiz, 1862)	Genzano & San Martín (2002)
	Cn <i>Coryne eximia</i> Allman, 1859	Allen (1915, 1923), Spooner, Wilson & Trebble (1957), Alós (1989)
	Cn <i>Tubularia indivisa</i> Linnaeus, 1758	Caullery (1925), Spooner, Wilson & Trebble (1957)
<i>Proceraea penetrans</i> (Wright and Woodwick, 1977)	Cn <i>Stylaster californicus</i> (Verrill, 1869)	Wright & Woodwick (1977)
<i>Proceraea madeirensis</i> Nygren, 2004	Cn <i>Eudendrium carneum</i> Clarke, 1882	E. Cruz-Rivera, 1991, personal observations
<i>Pachyprocerastea hydrozoicola</i> (Hartmann-Schröder, 1992)	Cn <i>Pseudosolanderia</i> sp.	Hartmann-Schröder (1992)
<i>Procerastea parasimpliseta</i> Hartmann-Schröder, 1992	Cn <i>Pseudosolanderia</i> sp.	Hartmann-Schröder (1992)

Note:

Cn, Cnidarians; Tu, tunicates.

Other studies have documented serendipitous observations obtained while searching for other ascidian associates. For example, in his monograph on ascidian-associated copepods, Illg (1958) reported unidentified polychaetes from the atria and branchial sacs of dredged ascidians. Similarly, Monniot, (1990) reported ten unidentified Syllidae from the branchial sac of *Microcosmus anchylodeirus* Traustedt, 1883. In summary, information on the nature of polychaete-ascidian relationships remains very scarce. Most reports of polychaetes (and syllids in particular) do not refer to individuals “living in association with” tunicates (which would imply some degree of specialization and thus suggest any type of symbiotic interaction). In fact, there is only one previous report specifically referring to a symbiosis, in which another autolytine, *Myrianida pinnigera*, was found living in association with *Asciadiella aspersa* and *Phallusia mammilata* (Table 1). Two decades later, Spooner, Wilson & Trebble (1957) stated that this species feeds on the body fluids of these and other ascidians from the British southern coast. While intriguing, this interaction has never been quantitatively evaluated and the evidence for this specialized trophic mode is still unclear. But if so, *Proceraea exoryxae* sp. nov. is the second known polychaete, and the second autolytine too, living in symbiosis with ascidians.

In addition to these two species of tunicate associates, eight more autolytines have been previously reported as living in symbiosis with other invertebrates, all them cnidarians (Table 1). Most of them are considered parasites, although only some are sufficiently studied to be clearly defined as such (Martin & Britayev, 1998). Among the best

documented, *Proceraea penetrans* (Wright & Woodwick, 1977) induces galls on its hydrocoral hosts, while *Proceraea* sp. modifies the theca of polyps in its hydroid hosts in order to live inside, probably feeding on the polyps themselves (Britayev, San Martín & Sheiko, 1998; Britayev & San Martín, 2001).

***Proceraea exoryxae* sp. nov. as a miner**

The association of *Proceraea exoryxae* sp. nov. with *Phallusia nigra* appears to be extremely rare, as there was only one infested host among all those we examined. The presence of a polychaete inside the tunic of *Phallusia nigra* has not been reported in previous studies at the same and other reefs (Kim et al., 2016). As mentioned above, parasitism is an atypical phenomenon among polychaetes, but also parasitic species are, with a few exceptions, extremely rare. In fact, many symbiotic polychaetes are only known from a single specimen (or very few) found only once (Martin & Britayev, 1998). The reasons for this rarity are often unknown. We may speculate that the lack of dedicated studies may be the actual reason in many cases, although that seems unlikely for *Proceraea exoryxae* sp. nov., which was discovered during multi-year monitoring of the associated fauna of the host ascidian (Kim et al., 2016). We could also hypothesize that the parasite is a recent introduction from an unknown origin, but this also seems unlikely because the host is a Red Sea endemic ascidian (Vandepas et al., 2015) and specialist parasites would be expected to occur in areas where hosts have the longest evolutionary history. More reasonably, either the polychaete occurs only infrequently and is thus difficult to find, or its peculiar and hidden habitat may have caused it to be overlooked in previous studies. We can also not discard the possibility that the parasitic mode of life may be just a phase in the life cycle of the worm, possibly connected to reproduction, as inferred from the presence of epitokous forms among atokes. This would add a temporal component to the presence of *Proceraea exoryxae* sp. nov. inside *Phallusia nigra*, that would increase the difficulty in finding it.

Despite (and, maybe, due to) its rarity, *Proceraea exoryxae* sp. nov. is the first polychaete formally defined as miner and, certainly, the first of Autolytinae. We use the term mining as it is often used to describe insects that tunnel through the tissues of their plant hosts (Connor & Taverner, 1997; Sinclair & Hughes, 2010; Mejaes, Poore & Thiel, 2015), but also marine isopods tunneling seagrass leaves (Brearley & Walker, 1995). This is also the mechanism we suggest for the formation of the galleries in the *Phallusia nigra* tunic where *Proceraea exoryxae* sp. nov. was found. The rarity of the polychaete precluded a thorough assessment of the host-symbiont interaction although, as in the case of *M. pinnigera*, the new species possibly feeds on the tissues of the host ascidian. Nonetheless, it represents the first clear example of mechanical damage by a polychaete on an ascidian, and as such, we classify the interaction as a parasitic symbiosis (Castro, 2015). The defensive characteristics attributed to the *Phallusia nigra* tunic, which include the accumulation of vanadium and sulfuric acid, and their derived metabolites (Stoecker, 1980; Hirose, Yamashiro & Mori, 2001; Pisut & Pawlik, 2002; Odate & Pawlik, 2007), did not prevent infestation by *Proceraea exoryxae* sp. nov., while they have been suggested to prevent infestation by the bivalve *Musculus subpictus* (Cantraine, 1835) in

an population introduced in Panama (*Cañete & Rocha, 2013*). Because both the host and polychaete symbiont were likely at their native habitat, and because symbionts are often unaffected by host defensive metabolites, the new partnership here reported may imply a noticeable degree of specialization. The presence of epitokous forms certainly confirms that at least the first phases of the reproductive cycle of the species (i.e., stolon formation) occurred inside the galleries, which may be considered as an additional evidence of specialization. However, whether *Proceraea exoryxae* sp. nov. is an exclusive parasite of *Phallusia nigra* or infests other ascidians awaits further studies.

Although rare for polychaetes, many invertebrates are known to live in symbiotic associations with ascidians, including amphipods, shrimps, copepods, pinnotherid crabs, nemerteans and cnidarians (*Illg, 1958; Stock, 1967; Boxshall, 2005; Lambert, 2005; Monniot, 1990; Thiel, 2000; Baeza & Díaz-Valdés, 2011; White, 2011; Kim et al., 2016*). Most of these animals live in the branchial sac of the host and are often considered commensals, with the exception of some copepod taxa, which are largely classified as ectoparasites on this respiratory organ (*Illg, 1958; Stock, 1967; Boxshall, 2005; Kim et al., 2016*). In contrast, but perhaps not surprisingly, few animals have evolved to inhabit the ascidian tunic, which is often structurally tough, and may contain spicules, inorganic acids, concentrated vanadium, and a variety of defensive secondary chemicals (*Stoecker, 1980; Pisut & Pawlik, 2002; Joullié et al., 2003; Odate & Pawlik, 2007; Kopllovitz et al., 2009*). Some mytilid mussels in the genera *Mytilimeria* and *Musculus* (= *Modiolarca*) are symbiotic bivalves that live completely embedded in the tunic of their ascidian host (*Say, 1822; White, 1949; Lambert, 2005; Morton & Dinesen, 2011; Cañete & Rocha, 2013*). Similarly, two species of amphipods in the genus *Polycheria* live by filter feeding from individual shallow pockets they excavate on the tunic of their host ascidians (*Skogsberg & Vansell, 1928; McClintock et al., 2009*). Recently, the parasitic copepod *Janstockia phallusiella* *Boxshall & Marchenkov, 2005* has been reported as living attached to the atrial wall of *Phallusia nigra* (*Kim et al., 2016*). None of these animals, however, produce a network of tunnels similar to that observed in the specimen of *Phallusia nigra* infested by *Proceraea exoryxae* sp. nov.

Among polychaetes several species are known to inhabit excavated galleries. Probably the best known are polydorid spionids, which include numerous species from different genera that burrow into calcareous substrates, including algae, crustacean carapaces, and mollusc shells. Some of them are simple borers, but others are well-known commensals and parasites, sometimes being even considered as pests when they infest species of commercial interest (*Martin & Britayev, 1998*). Although less diverse, similar habits are also present among cirratulids and sabellids, the latter being also able to infest fresh water invertebrates (*Martin & Britayev, 1998*). Polychaetes are also known to excavate galleries in seagrasses (*Guidetti, 2000; Gambi, van Tussenbroek & Brearley, 2003*), cnidarians (*Martin et al., 2002; Cairns & Bayer, 2008; Cairns, 2006, 2009, 2011, 2012; Mueller et al., 2013; Britayev et al., 2014; Molodtsova, Britayev & Martin, 2016*) and sponges (see *Lattig & Martin, 2011* and references herein). Seagrass associated polychaetes are mainly detritivores that bore into the dead sheath tissues (*Gambi, van Tussenbroek & Brearley, 2003*), but their galleries are also present in living meristems and leaves that have

been reported as “mined” tissues ([Guidetti, 2000](#)). Cnidarian associates (e.g., polynoids, eunicids, syllids) may inhabit depressions in the host skeleton that are usually covered by overgrowing host tissues and/or skeleton to form tunnels or galleries, presumably as a reaction to the symbionts’ presence ([Britayev et al., 2014](#)). A particular case is that of *Haplosyllis anthogorgicola* [Utinomi, 1956](#), which excavates a network of galleries inside the soft tissues of its host gorgonian. Host tissue overgrowths are limited to small tube-like protuberances at the gallery exits, from where the worms supposedly feed by stealing food from the nearby host polyps ([Martin et al., 2002](#)). Polychaete sponge borers are mainly syllids (e.g., *Haplosyllis*, *Haplosyllides*), which may either inhabit the aquiferous channels of the sponge or excavate their own galleries inside the host tissues ([Martin & Britayev, 1998](#); [Martin, Aguado & Britayev, 2009](#); [Lattig & Martin, 2011](#)).

When observing the tunic of the Red Sea specimen of *Phallusia nigra* we did not detect traces of external overgrowths associated to the gallery openings and, when dissecting the excavated galleries, we did not find any induced malformations or defined cavities (like cysts, galls or blisters). Conversely, there was a thin, translucent layer covering the galleries. Likely, this was an inner lining secreted by the worms to cover the tunnel walls, possibly made in a similar fashion as the hyaline tubes that some autolytines build to remain attached to their host cnidarians ([Molodtsova, Britayev & Martin, 2016](#)). At present, the mechanics of excavating tunnels by *Proceraea exoryxae* sp. nov. are unknown, but the typical syllid feeding structures (i.e., trepan, evaginable pharynx and sucking proventricle) seem to be a perfect combination enabling *Proceraea exoryxae* sp. nov. for this particular task.

In addition to possible affectations to host fitness, the parasitic mode of life attributed to *Proceraea exoryxae* sp. nov. may also be relevant for coastal management. Being native from the Red Sea, *Phallusia nigra* has been introduced worldwide in tropical and sub-tropical ecosystems ([Shenkar, 2012](#); [Vandepas et al., 2015](#)) where, as many other tunicates ([Zhan et al., 2015](#)), it has the potential of becoming invasive. Accordingly, three interesting questions remain open for further studies: (1) whether the parasitic *Proceraea exoryxae* sp. nov. may be (or has already been) introduced together with the ascidian, (2) whether it may contribute to control the spreading of *Phallusia nigra* in non-native regions, and (3) whether it may switch its host to infest, and thus cause damage, to native ascidians in the regions where the Red Sea host/parasite partnership has been introduced. In combination with molecular tools to trace the origin of an introduced species, the existence of a specialized parasite known only from the native host population may also help assess whether the host species has been introduced directly from this native population or indirectly from an already introduced population ([MacKenzie, 1993, 2002](#); [Catalano et al., 2014](#)). Nevertheless, the actual relevance of the association may be obscured by its rarity and, thus, will certainly rely on a future confirmation of its actual prevalence, as well as on the assessment of spatial and temporal extension of the infestation.

ACKNOWLEDGEMENTS

EC-R acknowledges the Department of Biological Sciences, University of the Virgin Islands for time release during the preparation of this article. Hussa Al Ajeer, Ali Fahmi,

Maricel Flores-Díaz, and Tamer Hafez, helped with specimen collection and ascidian dissections. We thank the staff of the J. D. Gerhart Field Station for support during our fieldwork and Dr X. Turon from the CEAB–CSIC for his advice on tunicate associates. María García, from the SEM service at the CEAB–CSIC, kindly helped with the SEM observations. This paper is contribution #175 from the Center for Marine and Environmental Sciences, University of the Virgin Islands (EC-R), and a contribution of DM to the research project MarSymBiomics and the Consolidated Research Group on Marine Benthic Ecology.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

EC-R was supported by a Faculty Improvement Grant from the American University in Cairo. DM was supported by research project MarSymBiomics (grant number CTM2013-43287-P) funded by the Ministerio de Educación, Cultura y Deporte, and by the Consolidated Research Group on Marine Benthic Ecology (grant number 2014SGR120) of the Agència de Gestió d'Ajuts Universitaris i de Recerca of the Generalitat de Catalunya. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

American University in Cairo.

MarSymBiomics: CTM2013-43287-P.

Ministerio de Educación, Cultura y Deporte.

Consolidated Research Group on Marine Benthic Ecology, Agència de Gestió d'Ajuts Universitaris i de Recerca of the Generalitat de Catalunya: 2014SGR120.

Competing Interests

The authors declare that they have no competing interests.

Author Contributions

- Daniel Martin conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables and reviewed drafts of the paper.
- Arne Nygren conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables and reviewed drafts of the paper.
- Edwin Cruz-Rivera conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper and collected the specimens.

Data Availability

The following information was supplied regarding data availability:

Specimen vouchers were deposited at the Museo Nacional de Ciencias Naturales of Madrid, Spain (MNCN). Microscope photos are included in the manuscript.

New Species Registration

The following information was supplied regarding the registration of a newly described species:

Species name: urn:lsid:zoobank.org:act:34373CE6-A0D4-488D-B4A5-12CF4E103504

Publication LSID: urn:lsid:zoobank.org:pub:685CB1C2-CB5B-4A87-9CD7-C04BFFDE03B4

REFERENCES

- Agassiz A. 1862.** On alternate generation of annelids and the embryology of *Autolytus cornutus*. *Journal of the Boston Society for Natural History* 7:384–409.
- Aguado MT, San Martín G. 2009.** Phylogeny of Syllidae (Polychaeta) based on morphological data. *Zoologica Scripta* 38(4):379–402 DOI 10.1111/j.1463-6409.2008.00380.x.
- Aguirre L, San Martín G, Álvarez-Campos P. 2016.** Autolytinae from Peru: description of *Myrianida paredesi* sp. nov. and new records of *Myrianida pentadentata* (Imajima, 1966), and *Proceraea micropedata* (Hartmann-Schröder, 1962). *Journal of the Marine Biological Association of the United Kingdom* 96(8):1633–1649 DOI 10.1017/S0025315415002015.
- Allen EJ. 1915.** Polychaeta of Plymouth and the South Devon coast, including a list of the Archannelida. *Journal of the Marine Biological Association of the United Kingdom* 10(4):592–646 DOI 10.1017/S002531540000919X.
- Allen EJ. 1923.** Regeneration and reproduction of the syllid *Procerastea*. *Philosophical Transactions of the Royal Society of London* 211(382–390):131–177 DOI 10.1098/rstb.1923.0003.
- Allman GJ. 1859.** V. Notes on the hydroid zoophytes. *Annals and Magazine of Natural History* 4:137–144.
- Alós C. 1989.** Adiciones a la fauna de Anélidos Poliquetos de la península Ibérica: familia Syllidae. *Cahiers de Biologie Marine* 30:329–337.
- Álvarez-Campos P, San Martín G, Piotrowski C. 2014.** A new species of Myrianida Milne Edwards, 1845 (Autolytinae: Syllidae: Annelida), from Luzón, Philippine Islands. *Special Publication of the California Academy of Sciences, the Coral Triangle–The Hearst Biodiversity Expedition to the Philippines* 2011:145–154.
- Baeza JA, Díaz-Valdés M. 2011.** The symbiotic shrimp *Ascidonia flavomaculata* lives solitarily in the tunicate *Ascidia mentula*: implications for its mating system. *Invertebrate Biology* 130(4):351–361 DOI 10.1111/j.1744-7410.2011.00244.x.
- Bosc LAC. 1802.** *Histoire naturelles des vers, contenant leur description et leur moeurs avec figures dessinees d'apres nature*. Paris: Deterville Libraire.
- Boxshall G. 2005.** Crustacean parasites: Copepoda (copepods). In: Rohde K, ed. *Marine Parasitology*. Wallingford: CABI Publishing, 123–138.
- Boxshall GA, Marchenkov A. 2005.** A new genus of notodelphyid copepod (Crustacea, Copepoda, Cyclopoida) from a compound ascidian host collected in the Suez Canal. *Zoosystema* 27:483–497.
- Brearley A, Walker D. 1995.** Isopod miners in the leaves of two Western Australian *Posidonia* species. *Aquatic Botany* 52(3):163–181 DOI 10.1016/0304-3770(95)00493-9.

- Britayev TA, Gil J, Altuna A, Calvo M, Martin D. 2014.** New symbiotic associations involving polynoids (Polychaeta, Polynoidae) from Atlantic waters, with redescrptions of *Parahololepidella greeffi* (Augener, 1918) and *Gorgoniapolynoe caeciliae* (Fauvel, 1913). *Memoirs of Museum Victoria* **71**:27–43 DOI [10.24199/j.mmv.2014.71.04](https://doi.org/10.24199/j.mmv.2014.71.04).
- Britayev TA, Lyskin SA. 2002.** Feeding of the symbiotic polychaete *Gastrolepidia clavigera* (Polynoidae) and its interactions with its hosts. *Doklady Biological Sciences* **385**:352–356 DOI [10.1023/A:1019964918471](https://doi.org/10.1023/A:1019964918471).
- Britayev TA, San Martín G. 2001.** Description and life-history traits of a new species of *Proceraea* with larvae infecting *Abietinaria turgida* (Polychaeta, Syllidae and Hydrozoa, Sertulariidae). *Ophelia* **54**(2):105–113 DOI [10.1080/00785236.2001.10409458](https://doi.org/10.1080/00785236.2001.10409458).
- Britayev TA, San Martín G, Sheiko O. 1998.** Symbiotic Polychaetes *Proceraea* sp. (Polychaeta, Syllidae) and modification of hydrothecae in hydroid *Abietinaria turgida* (Hydrozoa, Sertulariidae). *Zoologicheskii Zhurnal* **77**:527–532.
- Cairns SD. 2006.** Studies on western Atlantic Octocorallia (Coelenterata: Anthozoa). Part 6: the genera *Primnoella* Gray, 1858; *Thouarella* Gray, 1870; *Dasystemella* Versluys, 1906. *Proceedings of the Biological Society of Washington* **119**(2):161–194 DOI [10.2988/0006-324X\(2006\)119\[161:SOWAOC\]2.0.CO;2](https://doi.org/10.2988/0006-324X(2006)119[161:SOWAOC]2.0.CO;2).
- Cairns SD. 2009.** Review of Octocorallia (Cnidaria: Anthozoa) from Hawaii and adjacent seamounts. Part 2: genera *Paracalyptrophora* Kinoshita, 1908; *Candidella* Bayer, 1954; and *Calyptrophora* Gray, 1866. *Pacific Science* **63**(3):413–448 DOI [10.2984/049.063.0309](https://doi.org/10.2984/049.063.0309).
- Cairns SD. 2011.** A revision of the Primnoidae (Octocorallia: Acyonacea) from the Aleutian Islands and Bering Sea. *Smithsonian Contributions to Zoology* **634**:1–55 DOI [10.5479/si.00810282.634](https://doi.org/10.5479/si.00810282.634).
- Cairns SD. 2012.** New Zealand Primnoidae (Anthozoa: Alcyonacea). Part 1: genera *Narella*, *Narelloides*, *Metanarella*, *Calyptrophora*, and *Helicoprismoa*. *NIWA Biodiversity Memoirs* **126**:1–72.
- Cairns SD, Bayer FM. 2008.** A review of the Octocorallia (Cnidaria: Anthozoa) from Hawaii and adjacent seamounts: the genus *Narella* Gray, 1870. *Pacific Science* **62**(1):83–115 DOI [10.2984/1534-6188\(2008\)62\[83:AROTOC\]2.0.CO;2](https://doi.org/10.2984/1534-6188(2008)62[83:AROTOC]2.0.CO;2).
- Cañete JI, Rocha RM. 2013.** *Modiolarca lateralis* (Pteryomorpha: Mytilidae): bivalve associated to six species of ascidians from Bocas del Toro, Panama. *Latin American Journal of Aquatic Research* **41**(5):1030–1035 DOI [10.3856/vol41-issue5-fulltext-25](https://doi.org/10.3856/vol41-issue5-fulltext-25).
- Cantraine F. 1835.** Diagnoses ou descriptions succinctes de quelques espèces nouvelles de mollusques, qui feront partie de l'ouvrage Malacologie méditerranéenne et littorale, et comparaisons des coquilles qu'on trouve dans les collines subapennines avec celles qui vivnet encore dans nos mers. *Bulletin de l'Académie Royale des Sciences et Belles-lettres de Bruxelles* **2**:380–411.
- Castro P. 2015.** Symbiotic Brachyura. In: Castro P, Davie P, Guinot D, Schram FR, von Vaupel Klein JC, eds. *Treatise on Zoology—Anatomy, Taxonomy, Biology the Crustacea, Volume 9 Part C (2 vols)*. Leiden: Brill, 543–581.
- Catalano SR, Whittington ID, Donnellan SC, Gillanders BM. 2014.** Parasites as biological tags to assess host population structure: guidelines, recent genetic advances and comments on a holistic approach. *International Journal for Parasitology: Parasites and Wildlife* **3**(2):220–226 DOI [10.1016/j.ijppaw.2013.11.001](https://doi.org/10.1016/j.ijppaw.2013.11.001).
- Caullery M. 1925.** Schizogenese et schizogamie de *Procerastea halleziana* Malaquin. Parasitism de ce Syllidien sur les Tubulaires. *Bulletin de la Société Zoologique de France* **50**:204–208.

- Cerdá M, Castilla JC. 2001.** Diversidad y biomasa de macro-invertebrados en matrices intermareales del tunicado *Pyura praeputialis* (Heller, 1878) en la Bahía de Antofagasta, Chile. *Revista Chilena de Historia Natural* **74**(4):841–853 DOI [10.4067/S0716-078X2001000400011](https://doi.org/10.4067/S0716-078X2001000400011).
- Çinar ME. 2015.** A new species of *Myrianida* (Polychaeta: Syllidae: Autolytinae) from Rhodes (Greece, eastern Mediterranean). *Journal of the Marine Biological Association of the United Kingdom* **95**(6):1101–1104 DOI [10.1017/S0025315415000107](https://doi.org/10.1017/S0025315415000107).
- Çinar ME, Gambi MC. 2005.** Cognetti's syllid collection (Polychaeta: Syllidae) deposited at the Museum of the Stazione Zoologica "Anton Dohrn" (Naples, Italy), with descriptions of two new species of *Autolytus*. *Journal of Natural History* **39**(10):725–762 DOI [10.1080/00222930400001327](https://doi.org/10.1080/00222930400001327).
- Clarke SF. 1877.** Report on the hydroids collected on the coast of Alaska and the Aleutian Islands by W.H. Dall, U.S. Coast Survey, and party, from 1871 to 1874 inclusive. *Proceedings of the Academy of Natural Sciences of Philadelphia* **1876**:209–238 plates 207–216.
- Clarke SF. 1882.** New and interesting hydroids from the Chesapeake Bay. *Memoirs of the Boston Society of Natural History* **3**:135142.
- Connor EF, Taverner MP. 1997.** The evolution and adaptive significance of the leaf-mining habit. *Oikos* **79**(1):6–25 DOI [10.2307/3546085](https://doi.org/10.2307/3546085).
- Cuvier G. 1815.** Mémoire sur les Ascidies et sur leur anatomie. *Annales du Muséum d'Histoire Naturelle de Paris* **2**:10–39.
- Dietrich A, Hager T, Bönsch R, Winkelmann C, Schmidt A, Nygren A. 2015.** A new species of *Myrianida* (Autolytinae, Syllidae, Annelida) from the North Sea, with short notes on the distribution and habitat of Northeast Atlantic autolytines. *Marine Biology Research* **11**(8):804–813 DOI [10.1080/17451000.2015.1016971](https://doi.org/10.1080/17451000.2015.1016971).
- Fauchald K, Jumars PA. 1979.** The diet of worms: a study of polychaete feeding guilds. *Oceanography and Marine Biology: An Annual Review* **17**:193–284.
- Fielding P, Weerts K, Forbes A. 1994.** Macroinvertebrate communities associated with intertidal and subtidal beds of *Pyura stolonifera* (Heller) (Tunicata: Ascidiacea) on the Natal coast. *South African Journal of Zoology* **29**(1):46–53 DOI [10.1080/02541858.1994.11448326](https://doi.org/10.1080/02541858.1994.11448326).
- Fiore CL, Jutte PC. 2010.** Characterization of macrofaunal assemblages associated with sponges and tunicates collected off the southeastern United States. *Invertebrate Biology* **129**(2):105–120 DOI [10.1111/j.1744-7410.2010.00184.x](https://doi.org/10.1111/j.1744-7410.2010.00184.x).
- Fischer A, Mewes K, Franke HD. 1992.** Stolonization and mating behavior of *Autolytus prolifer* (Polychaeta). Göttingen: Institute of the Scientific Film.
- Franke HD. 1999.** Reproduction of the Syllidae (Annelida: Polychaeta). *Hydrobiologia* **402**:39–55 DOI [10.1023/A:1003732307286](https://doi.org/10.1023/A:1003732307286).
- Gambi MC, van Tussenbroek BI, Brearley A. 2003.** Mesofaunal borers in seagrasses: world-wide occurrence and a new record of boring polychaetes in the Mexican Caribbean. *Aquatic Botany* **76**(1):65–77 DOI [10.1016/s0304-3770\(03\)00031-7](https://doi.org/10.1016/s0304-3770(03)00031-7).
- Gardiner SL. 1976.** Errant Polychaete annelids from North Carolina. *Journal of the Elisha Mitchell Scientific Society* **91**:77–220.
- Genzano GN, San Martín G. 2002.** Association between the polychaete *Procerastea halleziana* (Polychaeta: Syllidae: Autolytinae) and the hydroid *Tubularia crocea* (Cnidaria: Hydrozoa) from the Mar del Plata intertidal zone, Argentina. *Cahiers de Biologie Marine* **43**:165–170.
- Gidholm L. 1967.** A revision of Autolytinae (Syllidae, Polychaeta) with special reference to Scandinavian species, and with notes on external and internal morphology, reproduction and ecology. *Arkiv för Zoologi* **19**:157–213.

- Guidetti P. 2000.** Invertebrate borers in the Mediterranean seagrass *Posidonia oceanica*: biological impact and ecological implications. *Journal of the Marine Biological Association of the United Kingdom* **80**(40):725–730 DOI [10.1017/s0025315400002551](https://doi.org/10.1017/s0025315400002551).
- Hamond R. 1969.** On the preferred foods of some autolytoids (Polychaeta, Syllidae). *Cahiers de Biologie Marine* **10**:439–445.
- Hartmann-Schröder G. 1992.** Drei neue Polychaeten-arten der familien Polynoidae und Syllidae von Neu-Kaledonien, assoziiert mit einer verkalteten Hydrozoe. *Helgoländer Meeresuntersuchungen* **46**(1):93–101 DOI [10.1007/BF02366214](https://doi.org/10.1007/BF02366214).
- Heller C. 1878.** Beiträge zur nähern Kenntniss der Tunicaten. *Sitzungsberichte der Akademie der Wissenschaften in Wien* **77**:83–109.
- Hirose E, Yamashiro H, Mori Y. 2001.** Properties of tunic acid in the ascidian *Phallusia nigra* (Asciidiidae, Phlebobranchia). *Zoological Science* **18**(3):309–314 DOI [10.2108/zsj.18.309](https://doi.org/10.2108/zsj.18.309).
- Illg PL. 1958.** North American copepods of the family Notodelphyidae. *Proceedings of the United States National Museum* **107**(3390):463–659 DOI [10.5479/si.00963801.108-3390.463](https://doi.org/10.5479/si.00963801.108-3390.463).
- Joullié MM, Leonard MS, Portonovo P, Liang B, Ding X, La Clair JJ. 2003.** Chemical defense in ascidians of the Didemnidae family. *Bioconjugate Chemistry* **14**:30–37 DOI [10.1021/bc025576n](https://doi.org/10.1021/bc025576n).
- Kim I-H, Cruz-Rivera E, Sherif M-E-D, El-Sahhar S. 2016.** Cyclopid copepods (Ascidicolidae, Notodelphyidae) associated with *Phallusia nigra* Savigny, 1816 (Asciacea) in the Red Sea: a new ascidicolid and first descriptions of the males from two notodelphyids. *Journal of Crustacean Biology* **36**(4):553–566 DOI [10.1163/1937240X-00002439](https://doi.org/10.1163/1937240X-00002439).
- Koplovitz G, McClintock JB, Amsler CD, Baker BJ. 2009.** Palatability and chemical anti-predatory defenses in common ascidians from the Antarctic Peninsula. *Aquatic Biology* **7**:81–92 DOI [10.3354/ab00188](https://doi.org/10.3354/ab00188).
- Lambert G. 2005.** Ecology and natural history of the protochordates. *Canadian Journal of Zoology* **83**(1):34–50 DOI [10.1139/z04-156](https://doi.org/10.1139/z04-156).
- Lattig P, Martin D. 2011.** Sponge-associated *Haplosyllis* (Polychaeta: Syllidae: Syllinae) from the Caribbean Sea, with the description of four new species. *Scientia Marina* **75**(4):733–758 DOI [10.3989/scimar.2011.75n4733](https://doi.org/10.3989/scimar.2011.75n4733).
- Linnaeus C. 1758.** *Systema Naturae per Regna Tria Naturae, Secundum Classes, Ordines, Genera, Species, cum characteribus, differentiis, synonymis, locis. Tomus I. Editio Decima, Reformata.* Stockholm: Laurentii Salvii.
- Lucas Y, San Martín G, Sikorski A. 2010.** A new genus and species of Syllidae (Annelida: Polychaeta) from off the coast of Norway with unusual morphological characters and an uncertain systematic position. *Proceedings of the Biological Society of Washington* **123**(4):251–257 DOI [10.2988/09-11.1](https://doi.org/10.2988/09-11.1).
- Macgillivray J. 1842.** L.—catalogue of the marine zoophytes of the neighbourhood of Aberdeen. *Annals and Magazine of Natural History* **9**(60):462–469 DOI [10.1080/03745484209445365](https://doi.org/10.1080/03745484209445365).
- MacKenzie K. 1993.** Parasites as biological indicators. *Bulletin if the Scandinavian Society for Parasitology* **3**:1–10.
- MacKenzie K. 2002.** Parasites as biological tags in population studies of marine organisms: an update. *Parasitology* **124**(7):153–163 DOI [10.1017/S0031182002001518](https://doi.org/10.1017/S0031182002001518).
- Malaquin A. 1893.** Recherches sur les syllidiens: morphologie, anatomie, reproduction, développement. *Mémoires de la Société des Sciences, de l'Agriculture et des Arts de Lille, 4e série* **18**:1–477.
- Martin D, Aguado MT, Britayev TA. 2009.** Review of the symbiotic genus *Haplosyllides*, with description of a new species. *Zoological Science* **26**:646–655 DOI [10.2108/zsj.26.646](https://doi.org/10.2108/zsj.26.646).

- Martin D, Britayev TA. 1998.** Symbiotic polychaetes: review of known species. *Oceanography and Marine Biology: An Annual Review* **36**:217–340.
- Martin D, Britayev TA, San Martín G, Gil J. 2003.** Inter-population variability and character description in the sponge associated *Haplosyllis spongicola* complex (Polychaeta: Syllidae). *Hydrobiologia* **496**(1–3):145–162 DOI [10.1007/978-94-017-0655-1_14](https://doi.org/10.1007/978-94-017-0655-1_14).
- Martin D, Gil J, Abgarian C, Evans E, Turner EM Jr, Nygren A. 2015.** *Proceraea janetae* sp. nov. (Annelida, Syllidae, Autolytinae), a scleractinian coral feeder from Grand Cayman Island. *Journal of the Marine Biological Association of the United Kingdom* **95**(4):703–712 DOI [10.1017/S0025315414001428](https://doi.org/10.1017/S0025315414001428).
- Martin D, Núñez J, Riera R, Gil J. 2002.** On the associations between *Haplosyllis* (Polychaeta, Syllidae) and gorgonians (Cnidaria, Octocorallaria), with a description of a new species. *Biological Journal of the Linnean Society* **77**(4):455–477 DOI [10.1046/j.1095-8312.2002.00117.x](https://doi.org/10.1046/j.1095-8312.2002.00117.x).
- McClintock JB, Amsler MO, Koplovitz G, Amsler CD, Baker BJ. 2009.** Observations on an association between the dexaminid amphipod *Polycheria antarctica* F. *Acanthopoda* and its ascidian host *Distaplia cylindrica*. *Journal of Crustacean Biology* **29**(4):605–608 DOI [10.1651/09-3146.1](https://doi.org/10.1651/09-3146.1).
- Mejaes BA, Poore AG, Thiel M. 2015.** Crustaceans inhabiting domiciles excavated from macrophytes and stone. *Life Styles and Feeding Biology of the Crustacea* **2**:118–144.
- Molina GI. 1782.** *Versuch einer Naturgeschichte von Chili. 4. Viertes Buch. Würmer, Insekten, Amphibien, Fische, Vögel und vierfüssige Thiere von Chili*. Leipzig: Friedrich Gotthold Jacobäer.
- Molodtsova T, Britayev TA, Martin D. 2016.** Chapter 25. Cnidarians and their polychaete symbionts. In: Goffredo S, Dubinsky Z, eds. *The Cnidaria, Past, Present and Future the World of Medusa and her Sisters*. Switzerland: Springer International Publishing, 387–413.
- Monniot C. 1990.** Diseases of Urochordata. In: Kinne O, ed. *Diseases of Marine Animals, Vol. III*. Hamburg: Biologische Anstalt Helgoland, 596–636.
- Montagu G. 1808.** II. Description of several marine animals found on the south coast of Devonshire. *Transactions of the Linnean Society of London* **9**(1):81–114 DOI [10.1111/j.1096-3642.1818.tb00327.x](https://doi.org/10.1111/j.1096-3642.1818.tb00327.x).
- Morton B, Dinesen GE. 2011.** The biology and functional morphology of *Modiolarca subpicta* (Bivalvia: Mytilidae: Musculinae), epizoically symbiotic with *Ascidia aspersa* (Urochordata: Ascidiacea), from the Kattegat, northern Jutland, Denmark. *Journal of the Marine Biological Association of the United Kingdom* **91**(8):1637–1649 DOI [10.1017/S0025315410001980](https://doi.org/10.1017/S0025315410001980).
- Mueller CE, Lundälv T, Middelburg JJ, van Oevelen D. 2013.** The symbiosis between *Lophelia pertusa* and *Eunice norvegica* stimulates coral calcification and worm assimilation. *PLoS ONE* **8**(3):e58660 DOI [10.1371/journal.pone.0058660](https://doi.org/10.1371/journal.pone.0058660).
- Müller OF. 1776.** *Zoologicae Danicae Prodrömus, seu Animalium Daniae et Norvegiae indigenarum characteres, nomina et synonyma imprimis popularium*. Havniae [Copenhagen]: Hallagerii.
- Nygren A. 2004.** Revision of Autolytinae (Syllidae: Polychaeta). *Zootaxa* **680**:1–314 DOI [10.11646/zootaxa.680.1.1](https://doi.org/10.11646/zootaxa.680.1.1).
- Nygren A, Pleijel F. 2007.** A new species of *Myrianida* (Syllidae, Polychaeta) from Belize. *Zootaxa* **1595**:17–29 DOI [10.5281/zenodo.178674](https://doi.org/10.5281/zenodo.178674).
- Nygren A, Pleijel F. 2010.** Redescription of *Imajimaea draculai*—a rare syllid polychaete associated with the sea pen *Funiculina quadrangularis*. *Journal of the Marine Biological Association of the United Kingdom* **90**(7):1441–1448 DOI [10.1017/S0025315409991536](https://doi.org/10.1017/S0025315409991536).

- Nygren A, Sundberg P. 2003. Phylogeny and evolution of reproductive modes in Autolytinae (Syllidae, Annelida). *Molecular Phylogenetics and Evolution* **29**(2):235–249 DOI [10.1016/S1055-7903\(03\)00095-2](https://doi.org/10.1016/S1055-7903(03)00095-2).
- Nygren A, Sundkvist T, Mikac B, Pleijel F. 2010. Two new and two poorly known autolytines (Polychaeta: Syllidae) from Madeira and the Mediterranean sea. *Zootaxa* **2640**:35–52 DOI [10.5281/zenodo.198574](https://doi.org/10.5281/zenodo.198574).
- Odate S, Pawlik JR. 2007. The role of vanadium in the chemical defense of the solitary tunicate, *Phallusia nigra*. *Journal of Chemical Ecology* **33**(3):643–654 DOI [10.1007/s10886-007-9251-z](https://doi.org/10.1007/s10886-007-9251-z).
- Okada YK. 1928. Feeding organs and feeding habits of *Autolytus edwardsi* St. Joseph. *Quarterly Journal of Microscopical Science* **72**:219–245.
- Okada YK. 1935. Stolization in *Myrianida*. *Journal of the Marine Biological Association of the United Kingdom* **20**(1):93–98 DOI [10.1017/S0025315400010079](https://doi.org/10.1017/S0025315400010079).
- Pallas PS. 1766. *Miscellanea Zoologica. Quibus novae imprimis atque obscurae animalium species describuntur et observationibus iconibusque illustrantur*. Hagae Comitum: apud Petrum van Cleef.
- Pettibone MH. 1963. Marine polychaete worms of the New England region. Part 1. Families Aphroditidae through Trochochaetidae. *Bulletin of the United States National Museum* **227**:1–356 DOI [10.5479/si.03629236.227.1](https://doi.org/10.5479/si.03629236.227.1).
- Pisut DP, Pawlik JR. 2002. Anti-predatory chemical defenses of ascidians: secondary metabolites or inorganic acids? *Journal of Experimental Marine Biology and Ecology* **270**(2):203–214 DOI [10.1016/S0022-0981\(02\)00023-0](https://doi.org/10.1016/S0022-0981(02)00023-0).
- Read G, Fauchald K. 2016. World Polychaeta database. Available at <http://www.marinespecies.org/polychaeta/aphia.php?p=taxdetails&id=129262> (accessed 9 December 2016).
- Rivolta A, San Martín G, Sikorski A. 2016. Additions to the description, reproduction and systematic position of the enigmatic species *Acritagasyllis longichaetosus* Lucas, San Martín & Sikorski, 2010 (Annelida: Phyllodocida: Syllidae). *Journal of the Marine Biological Association of the United Kingdom* **96**(8):1709–1716 DOI [10.1017/S0025315415002118](https://doi.org/10.1017/S0025315415002118).
- San Martín G, López E. 2002. New species of *Autolytus* Grube, 1850, *Paraprocerastea* San Martín and Alós, 1989, and *Sphaerosyllis* Claparède, 1863 (Syllidae, Polychaeta) from the Iberian Peninsula. *Sarsia* **87**:135–143 DOI [10.1080/003648202320205210](https://doi.org/10.1080/003648202320205210).
- Savigny J. 1816. *Mémoires sur les animaux sans vertèbres*. Paris: G. Dufour.
- Say T. 1822. An account of some of the marine shells of the United States. *Journal of the Academy of Natural Sciences of Philadelphia* **2**:221–248.
- Sepúlveda RD, Rozbaczylo N, Ibáñez CM, Flores M, Cancino JM. 2015. Ascidian-associated polychaetes: ecological implications of aggregation size and tube-building chaetopterids on assemblage structure in the Southeastern Pacific Ocean. *Marine Biodiversity* **45**(4):733–741 DOI [10.1007/s12526-014-0283-7](https://doi.org/10.1007/s12526-014-0283-7).
- Shenkar N. 2012. Ascidian (Chordata, Ascidiacea) diversity in the Red Sea. *Marine Biodiversity* **42**(4):459–469 DOI [10.1007/s12526-012-0124-5](https://doi.org/10.1007/s12526-012-0124-5).
- Sinclair RJ, Hughes L. 2010. Leaf miners: the hidden herbivores. *Austral Ecology* **35**(3):300–313 DOI [10.1111/j.1442-9993.2009.02039.x](https://doi.org/10.1111/j.1442-9993.2009.02039.x).
- Skogsberg T, Vansell GH. 1928. Structure and behavior of the amphipod *Polycheria osborni*. *Proceedings of the California Academy of Sciences* **17**:267–295.
- Spooner GM, Wilson DP, Trebble N. 1957. Phylum Annelida. *Plymouth Marine Fauna*. Plymouth: Marine Biological Association UK, 109–149.

- Stock JH. 1967.** Report on the Notodelphyidae (Copepoda, Cyclopoida) of the Israel South Red Sea Expedition. *Israel South Red Sea Expedition, 1962, Reports* **27**:3–126.
- Stoecker D. 1980.** Chemical defenses of ascidians against predators. *Ecology* **61**(6):1327–1334 DOI [10.2307/1939041](https://doi.org/10.2307/1939041).
- Thiel M. 2000.** Population and reproductive biology of two sibling amphipod species from ascidians and sponges. *Marine Biology* **137**(4):661–674 DOI [10.1007/s002270000372](https://doi.org/10.1007/s002270000372).
- Utinomi H. 1956.** On the so-called ‘Umi-Utiwa’ a peculiar flabellate gorgonacean, with notes on a syllidean polychaete commensal. *Publications of the Seto Marine Biological Laboratory* **5**(2):243–250 DOI [10.5134/174554](https://doi.org/10.5134/174554).
- Vandepas LE, Oliveira LM, Lee SS, Hirose E, Rocha RM, Swalla BJ. 2015.** Biogeography of *Phallusia nigra*: is it really black and white? *Biological Bulletin* **228**(1):52–64 DOI [10.1086/BBLv228n1p52](https://doi.org/10.1086/BBLv228n1p52).
- Verrill AE. 1869.** Synopsis of the polyps and corals of the North Pacific Exploring Expedition, under Commodore C. Ringgold and Capt. John Rodgers, USN, from 1853 to 1856. Collected by Dr. Wm. Stimpson, Naturalist to the Expedition. With descriptions of some additional species from the west coast of North America. Pt. 3: Madreporaria. *Proceedings of the Essex Institute*, 17–50.
- White KM. 1949.** *Musculus lebourae*, new species. *Journal of Molluscan Studies* **28**:46–49 DOI [10.1093/oxfordjournals.mollus.a064557](https://doi.org/10.1093/oxfordjournals.mollus.a064557).
- White KN. 2011.** A taxonomic review of the Leucothoidae (Crustacea: Amphipoda). *Zootaxa* **3078**:1–113.
- Wright JD, Woodwick KH. 1977.** A new species of *Autolytus* (Polychaeta: Syllidae) commensal on a Californian hydrocoral. *Bulletin of the Southern California Academy of Sciences* **76**:42–48.
- Zhan A, Briski E, Bock DG, Ghabooli S, MacIsaac HJ. 2015.** Ascidians as models for studying invasion success. *Marine Biology* **162**(12):2449–2470 DOI [10.1007/s00227-015-2734-5](https://doi.org/10.1007/s00227-015-2734-5).