



Ecology and Behaviour of the Ladybird Beetles

(*Coccinellidae*)

Edited by I. Hodek, H.F. van Emden
and A. Honek

ECOLOGY AND
BEHAVIOUR OF THE
LADYBIRD BEETLES
(COCCINELLIDAE)

ECOLOGY AND BEHAVIOUR OF THE LADYBIRD BEETLES (COCCINELLIDAE)

Edited by I. Hodek, H.F. van Emden and A. Honěk

 WILEY-BLACKWELL

A John Wiley & Sons, Ltd., Publication

This edition first published 2012 © 2012 by Blackwell Publishing Ltd

Blackwell Publishing was acquired by John Wiley & Sons in February 2007. Blackwell's publishing program has been merged with Wiley's global Scientific, Technical and Medical business to form Wiley-Blackwell.

Registered office: John Wiley & Sons, Ltd, The Atrium, Southern Gate, Chichester, West Sussex, PO19 8SQ, UK

Editorial offices: 9600 Garsington Road, Oxford, OX4 2DQ, UK
The Atrium, Southern Gate, Chichester, West Sussex, PO19 8SQ, UK
111 River Street, Hoboken, NJ 07030-5774, USA

For details of our global editorial offices, for customer services and for information about how to apply for permission to reuse the copyright material in this book please see our website at www.wiley.com/wiley-blackwell.

The right of the author to be identified as the author of this work has been asserted in accordance with the UK Copyright, Designs and Patents Act 1988.

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise, except as permitted by the UK Copyright, Designs and Patents Act 1988, without the prior permission of the publisher.

Designations used by companies to distinguish their products are often claimed as trademarks. All brand names and product names used in this book are trade names, service marks, trademarks or registered trademarks of their respective owners. The publisher is not associated with any product or vendor mentioned in this book. This publication is designed to provide accurate and authoritative information in regard to the subject matter covered. It is sold on the understanding that the publisher is not engaged in rendering professional services. If professional advice or other expert assistance is required, the services of a competent professional should be sought.

Library of Congress Cataloguing-in-Publication Data

Ecology and behaviour of the ladybird beetles (Coccinellidae) / edited by I. Hodek, H.F. van Emden, and A. Honěk.

p. cm.

Includes index.

ISBN 978-1-4051-8422-9 (cloth)

1. Ladybugs. I. Hodek, Ivo. II. Van Emden, Helmut Fritz. III. Honěk, A. (Alois)

QL596.C65E26 2012

595.76'9—dc23

2011045545

A catalogue record for this book is available from the British Library.

Wiley also publishes its books in a variety of electronic formats. Some content that appears in print may not be available in electronic books.

Set in 9/11 pt PhotinaMT by Toppan Best-set Premedia Limited

To all those scientists
who have now passed on
and who laid the foundation
of our present knowledge of Coccinellidae,
particularly to Michael Majerus
who intended to be one of authors
of this volume.

CONTENTS

Detailed contents, ix

Contributors, xvii

Preface, xviii

Introduction, xix

Taxonomic glossary, xx

1. PHYLOGENY AND CLASSIFICATION, 1

Oldřich Nedvěd and Ivo Kovář

2. GENETIC STUDIES, 13

John J. Sloggett and Alois Honěk

3. LIFE HISTORY AND DEVELOPMENT, 54

Oldřich Nedvěd and Alois Honěk

4. DISTRIBUTION AND HABITATS, 110

Alois Honěk

5. FOOD RELATIONSHIPS, 141

Ivo Hodek and Edward W. Evans

6. DIAPAUSE/DORMANCY, 275

Ivo Hodek

7. INTRAGUILD INTERACTIONS, 343

Éric Lucas

8. NATURAL ENEMIES OF LADYBIRD BEETLES, 375

Piotr Ceryngier, Helen E. Roy and Remy L. Poland

9. COCCINELLIDS AND SEMIOCHEMICALS, 444

Jan Pettersson

10. QUANTIFYING THE IMPACT OF COCCINELLIDS ON THEIR PREY, 465

J. P. Michaud and James D. Harwood

11. COCCINELLIDS IN BIOLOGICAL CONTROL, 488

J. P. Michaud

12. RECENT PROGRESS AND POSSIBLE FUTURE TRENDS IN THE STUDY OF COCCINELLIDAE, 520

Helmut F. van Emden and Ivo Hodek

Appendix: List of Genera in Tribes and Subfamilies, 526

Oldřich Nedvěd and Ivo Kovář

Subject index, 532

Colour plate pages fall between pp. 250 and pp. 251

DETAILED CONTENTS

Contributors, xvii	1.4.9 Coccidulinae, 8
Preface, xviii	1.4.10 Scymninae, 9
Introduction, xix	1.5 Future Perspectives, 10
Taxonomic glossary, xx	References, 10

1. PHYLOGENY AND CLASSIFICATION, 1

Oldřich Nedvěd and Ivo Kovář

1.1 Position of the Family, 2	1.1.1 The Cerylonid complex, 2
	1.1.2 Sister families, 2
	1.1.3 Feeding habits, 2
	1.1.4 Monophyly of Coccinellidae, 2
1.2 Characteristics of the Family, 3	
1.3 Changes in the Classification of Subfamilies, 3	1.3.1 Morphologically based classifications, 3
	1.3.2 Split of Sticholotidinae, 3
	1.3.3 Monophyly of other subfamilies, 4
	1.3.3.1 Contribution of immature stages, 4
	1.3.4 Molecular analyses, 4
	1.3.4.1 Alternative molecular methods, 5
	1.3.5 Rejection of the monophyly of subfamilies, 5
1.4 Characteristics of the Subfamilies and Tribes, 5	
	1.4.1 Proposed classification, 5
	1.4.2 Microweiseinae, 5
	1.4.3 Sticholotidinae, 5
	1.4.4 Coccinellinae, 6
	1.4.5 Epilachninae, 7
	1.4.6 Exoplectrinae, 7
	1.4.7 Chilocorinae, 7
	1.4.8 Ortaliinae, 8

2. GENETIC STUDIES, 13

John J. Sloggett and Alois Honěk

2.1 Introduction, 14	
2.2 Genome Size, 14	
2.3 Chromosomes and Cytology, 14	2.3.1 Chromosome numbers and banding, 14
	2.3.2 Sex determination, 14
	2.3.3 Supernumerary (B) chromosomes, 15
	2.3.4 Cytogenetic changes, intraspecific cytogenetic variation and speciation, 16
2.4 Colour Pattern Variation, 18	
	2.4.1 The nature of colour patterns, 19
	2.4.2 Genetic determination of colour patterns, 20
	2.4.3 Geographic variation, 23
	2.4.4 Temporal variation, 25
	2.4.5 Significance and evolution, 25
2.5 The Inheritance of Other Traits, 27	
	2.5.1 Morphological characters: wing polymorphism, 27
	2.5.2 Life history characters: heritability, selection experiments and genetic trade-offs, 28
2.6 Molecular Genetic Studies, 29	
	2.6.1 Sequence evolution, 29
	2.6.1.1 Mitochondrial DNA and the inference of evolutionary history, 29
	2.6.1.2 The ITS1 region, 31

2.6.2	Molecular studies of coccinellid biology, 33	3.5.6.3	Female choice and melanism, 83
2.6.2.1	Species, population and strain identification, 33	3.5.6.4	Hybridization, 85
2.6.2.2	Phylogenetics, 37	3.5.7	Oviposition, 85
2.6.2.3	Population genetic and phylogeographic studies, 38	3.5.7.1	Oviposition substrate, 85
2.6.2.4	Reproductive success, paternity and sperm competition, 42	3.5.7.2	Oviposition rhythmicity, 86
2.7	Conclusions, 43	3.5.7.3	Oviposition rate, 86
	Acknowledgements and Dedication, 44	3.5.7.4	Oviposition period, 86
	References, 44	3.5.8	Fecundity, 88
		3.5.9	Longevity, 90
		3.5.9.1	Voltinism, 90
		3.5.9.2	Effect of temperature, photoperiod and humidity, 90
		3.5.9.3	Effect of food, 91
		3.5.9.4	Effect of sexual activity, 91
3.1	Introduction, 55	3.6	Temperature and Development, 91
3.2	Egg, 55	3.6.1	Thermal constants, 92
3.2.1	Egg morphology, 55	3.6.2	Relationship between LDT and SET, 92
3.2.2	Egg size, 56	3.6.3	Thermal window and development rate isomorphy, 95
3.2.3	Cluster size, 56	3.6.4	Other events affected by temperature, 95
3.2.4	Hatching rate, 68	3.6.4.1	Tolerance to extreme temperatures, 96
3.2.5	Trophic eggs, 69		
3.3	Larva, 71	Acknowledgements, 97	
3.3.1	Larval morphology, 71	References, 97	
3.3.2	Instars, 71		
3.3.3	Development, 72		
3.3.4	Body size, 73		
3.4	Pupa, 73	4. DISTRIBUTION AND HABITATS, 110	
3.4.1	Prepupal stage, 73		
3.4.2	Pupal morphology, 74		
3.4.3	Timing of pupation, 75		
3.4.4	Places of pupation, 75		
3.4.5	Pupal defence, 76		
3.4.6	Colouration and thermal melanism, 76		
3.5	Adult, 76		
3.5.1	Teneral development, 76		
3.5.2	Wings and flight, 77		
3.5.3	Pre-oviposition period, 77		
3.5.4	Size, 78		
3.5.5	Ovarioles, 78		
3.5.6	Mating, 82		
3.5.6.1	Frequency and duration of mating, 82		
3.5.6.2	Sperm competition, 83		

4.3.3	Dominance, diversity and niche differentiation, 127	5.2.12	Tritrophic studies, 196
4.4	Coccinellid Communities of Particular Habitats, 128	5.2.13	Food of phytophagous Coccinellidae, 198
4.5	Conclusion, 132	5.2.14	Food of mycophagous Coccinellidae, 200
Acknowledgements, 133		5.3	Quantitative Aspects of Food Relations, 201
References, 133		5.3.1	Effect of physical factors on consumption, 202
		5.3.1.1	Daily consumption rate, 202
		5.3.1.2	Total food consumption, 202
		5.3.2	Effect of prey density on consumption: functional response, 204
		5.3.3	Effects of consumption on growth and reproduction, 207
		5.3.3.1	Larval development, 207
		5.3.3.2	Adult performance, 208
		5.3.4	Conversion and utilization of consumed food, 209
		5.3.5	Aggregative numerical response, 211
		5.3.5.1	Temporal and spatial patterns, 211
		5.3.5.2	Modeling of aggregative responses, 211
		5.3.5.3	Factors other than focal prey density, 211
5.1	Introduction, 142	5.4	Food-Related Behaviour, 213
5.2	Food Specificity, 142	5.4.1	Foraging behaviour, 213
5.2.1	Food range, 142	5.4.1.1	Indirect factors in foraging, 215
	5.2.1.1 Methods for detection of food range, 144	5.4.1.2	The role of senses in foraging, 218
5.2.2	Nutritional suitability of food, 145	5.4.1.3	Finding an oviposition site, 226
5.2.3	Prey size-density hypothesis, 147	5.4.1.4	Foraging of first instars, 233
5.2.4	Euryphagous and stenophagous species / Generalist and specialist species, 150	5.4.1.5	Movement among habitats/patches in the landscape, 234
	5.2.4.1 Generalists, 152	5.4.1.6	Ladybird foraging and ants, 236
	5.2.4.2 Specialists, 154	5.4.2	Prey capture, 238
5.2.5	Mixed and combined diet, 155	5.4.3	Food intake, 241
	5.2.5.1 Complementation across stages, 156	5.5	Conclusions, 242
	5.2.5.2 Prey switching, 157	Acknowledgements, 242	
	5.2.5.3 Prey specialization through selection, 157	References, 243	
5.2.6	Lower quality prey (mostly aphids), 157		
	5.2.6.1 Toxic prey, 157		
	5.2.6.2 Rejected prey, 165		
	5.2.6.3 'Problematic' prey, 167		
5.2.7	Prey other than aphids/coccids, 169		
	5.2.7.1 Developmental stages of Holometabola, 169		
	5.2.7.2 Non-aphid hemipterans, 174		
5.2.8	Cannibalism, 175		
	5.2.8.1 Sibling egg cannibalism, 175		
	5.2.8.2 Non-sibling egg cannibalism, 177		
5.2.9	Non-insect food (pollen, nectar, spores of fungi), 180		
5.2.10	Substitute diets and food supplements (sprays), 185		
5.2.11	Essential foods, 187		

6. DIAPAUSE/DORMANCY, 275

Ivo Hodek

6.1	Introduction: Mechanisms and Definitions, 276	6.3	6.2.16 Ecophysiological regulation of diapause in coccinellids, 300 Behaviour Patterns Related to Diapause, 301
6.1.1	Hibernation and aestivation, 276	6.3.1	6.3.1 Phases of dormancy behaviour, 301
6.1.2	Termination/completion of diapause, 277	6.3.1.1	Pre-diapause, 301
6.1.3	Phases of dormancy, 277	6.3.1.2	Migration, 302
6.1.4	Endocrinological aspects of adult diapause, 277	6.3.1.3	Flight and methods for its study, 303
6.2	Ecophysiological Regulation of Diapause in Coccinellids, 278	6.3.1.4	Aggregations, 305
6.2.1	<i>Coccinella septempunctata</i> , 278	6.3.1.5	Emergence from dormancy sites, 309
6.2.1.1	Central Europe, 278	6.3.2	Behaviour of individual species, 309
6.2.1.2	Western Europe: France, 282	6.3.2.1	<i>Ceratomegilla</i> (= <i>Semiadalia</i>) <i>undecimnotata</i> , 309
6.2.1.3	Northern Europe, 283	6.3.2.2	Hypsotactic species <i>Harmonia axyridis</i> , <i>Har. conformis</i> and <i>Aiolocaria hexaspilota</i> (= <i>mirabilis</i>), 310
6.2.1.4	Mediterranean region, 284	6.3.2.3	<i>Coccinella septempunctata</i> and other species dormant in the litter, 311
6.2.1.5	Nearctic region, 285	6.3.2.4	<i>Coleomegilla maculata</i> , 313
6.2.1.6	Potential multivoltines in univoltine populations, 286	6.3.2.5	<i>Adalia bipunctata</i> , 313
6.2.2	<i>Coccinella septempunctata</i> <i>brucki</i> , 287	6.3.2.6	<i>Myrrha octodecimguttata</i> and other forest species which hibernate in bark crevices, 314
6.2.2.1	Central Japan (central Honshu), 287	6.3.2.7	<i>Hippodamia convergens</i> , 314
6.2.2.2	Sapporo, Hokkaido (Japan), 288	6.3.2.8	<i>Hippodamia quinquesignata</i> , 316
6.2.2.3	Northern Honshu (Japan), 289	6.3.2.9	Hibernation of mycophagous and phytophagous species, 316
6.2.3	<i>Coccinella novemnotata</i> , 289	6.4	Anatomical and Physiological Changes Related to Dormancy, 316
6.2.4	<i>Adalia bipunctata</i> , 290	6.4.1	Anatomical state, 316
6.2.5	<i>Propylea quatuordecimpunctata</i> and <i>P. dissecta</i> , 291	6.4.1.1	Fat body and digestive tract, 316
6.2.6	<i>Hippodamia tredecimpunctata</i> , 292	6.4.1.2	Ovary, spermatheca, 317
6.2.7	<i>Hippodamia convergens</i> , 292	6.4.1.3	Male gonads, 321
6.2.8	<i>Ceratomegilla</i> (= <i>Semiadalia</i>) <i>undecimnotata</i> , 292	6.4.1.4	Flight muscles, 323
6.2.9	<i>Harmonia axyridis</i> , 295	6.4.2	Metabolic changes related to diapause, 323
6.2.10	<i>Coccinella leonina</i> (= <i>repanda</i>), 297	6.4.2.1	Lipids, 323
6.2.11	<i>Apolinus</i> (= <i>Scymnoides</i>) <i>lividigaster</i> and <i>Illeis</i> (= <i>Leptothoea</i>) <i>galbula</i> , 297	6.4.2.2	Glycogen, 325
6.2.12	<i>Harmonia sedecimnotata</i> , 297		
6.2.13	<i>Chilocorus</i> spp., 298		
6.2.14	<i>Stethorus punctum picipes</i> and <i>S. japonicus</i> , 299		
6.2.15	<i>Scymnus</i> (<i>Neopullus</i>) <i>sinuannulatus</i> , 300		

6.4.2.3	Water, 326	7.9.1	Defensive mechanisms of coccinellids against intraguild predation, 355
6.4.2.4	Metabolic rate, 326	7.9.1.1	Defence of all stages, 355
6.4.3	<i>Corpora allata</i> and regulation of vitellogenesis, 327	7.9.1.2	Defence of eggs, 356
6.4.4	Cold-hardiness, 328	7.9.1.3	Defence of larvae, 356
6.5	Conclusions and Lacunae in Knowledge, 332	7.9.1.4	Defence of moulting individuals and pupae, 357
	Acknowledgements, 333	7.9.1.5	Defence of adults, 357
	References, 333	7.9.2	Intraguild predation on coccinellids, 358
		7.9.2.1	Intraguild predation by intraguild predators, 358
7. INTRAGUILD INTERACTIONS, 343		7.9.2.2	Intraguild predation by intraguild parasitoids, 358
Éric Lucas		7.9.2.3	Intraguild predation by intraguild pathogens, 359
7.1	Scope, 344	7.10	Coccinellids Interacting with Intraguild Ants, 359
7.2	Coccinellids as Guild Members, 344	7.10.1	Non-myrmecophilous coccinellid species, 359
7.3	Coccinellids as Neutralists, 344	7.10.2	Myrmecophilous coccinellid species, 360
7.3.1	Temporal guild partition, 344	7.11	Applied Aspects of Intraguild Interactions, 360
7.3.2	Spatial guild partition, 344	7.11.1	Conservation, 360
7.3.3	Thermal guild partition, 344	7.11.2	Biological control, 361
7.3.4	Body size guild partition, 345	7.11.2.1	Intraguild predation and biocontrol, 361
7.4	Coccinellids as Interacting Organisms, 345	7.11.2.2	Facilitation and biocontrol, 361
7.5	Coccinellids as Intraguild Commensalists and Mutualists, 345	7.11.2.3	Ants and biocontrol, 362
7.6	Coccinellids as Competitors, 347	7.11.2.4	Intraguild interactions and biocontrol approaches, 362
7.6.1	Exploitative competition, 347	7.12	Interguild Effects, 362
7.6.2	Apparent competition, 349	7.13	Conclusion, 363
7.7	Coccinellids as Vectors of Male-Killing Bacteria, 349	References, 364	
7.8	Coccinellids as Intraguild Predators, 349		
7.8.1	General rules of intraguild predation involving coccinellid predators, 349	8. NATURAL ENEMIES OF LADYBIRD BEETLES, 375	
7.8.2	Occurrence of intraguild predation by coccinellids, 350		
7.8.3	Intraguild predation on intraguild coccinellids (by coccinellids), 351		
7.8.4	Intraguild predation on intraguild non-coccinellid predators, 352		
7.8.4.1	Intraguild neuropterans, 352		
7.8.4.2	Intraguild dipterans, 352		
7.8.4.3	Intraguild hemipterans, 353		
7.8.4.4	Other intraguild predators, 353		
7.8.5	Intraguild predation on intraguild parasitoids, 353		
7.8.6	Intraguild predation on intraguild pathogens, 354		
7.8.7	Coccinellids as top predators, 354		
7.9	Coccinellids as Intraguild Prey, 355		
		8.1	Introduction, 376
		8.2	Predation and Related Phenomena, 376
		8.2.1	Anti-predator defences, 376
		8.2.1.1	Aposematic colouration and other visual signals, 376
		8.2.1.2	Reflex bleeding, 376

	8.2.1.3	Morphological anti-predator adaptations, 377	8.4	Parasites and Pathogens, 411
	8.2.2	Vertebrate predators, 377	8.4.1	Acarina, 411
	8.2.3	Invertebrate predators, 377	8.4.1.1	Phoretic mites, 411
	8.2.4	Hemiptera-tending ants, 379	8.4.1.2	<i>Coccipolipus</i> Husband (Prostigmata: Podapolipidae), 411
	8.2.5	Social aphids with a soldier caste, 382	8.4.2	Nematodes, 414
8.3	Parasitoids, 383		8.4.2.1	Allantonematidae (Tylenchida), 414
	8.3.1	General characteristics of parasitoids of ladybirds, 390	8.4.2.2	Mermithidae (Mermithida), 415
	8.3.1.1	Host specificity of parasitoids recorded from Coccinellidae, 390	8.4.3	Fungal pathogens, 415
	8.3.1.2	Parasitoids of different subfamilies of Coccinellidae, 390	8.4.3.1	Hypocreales (Ascomycota), 415
	8.3.1.3	Parasitism in different developmental stages of Coccinellidae, 390	8.4.3.2	<i>Hesperomyces</i> spp. (Ascomycota: Laboulbeniales, Laboulbeniaceae), 416
	8.3.2	Review of the more important taxa, 391	8.4.3.3	Nosematidae (Microsporidia), 418
	8.3.2.1	<i>Dinocampus</i> Foerster (Hymenoptera: Braconidae, Euphorinae), 391	8.4.4	Protozoan pathogens, 419
	8.3.2.2	<i>Uga</i> Girault (Hymenoptera: Chalcididae), 399	8.4.4.1	Septate eugregarines (Apicomplexa: Eugregarinida: Septatorina), 419
	8.3.2.3	<i>Cowperia</i> Girault (Hymenoptera: Encyrtidae), 399	8.4.5	Bacteria, 421
	8.3.2.4	<i>Homalotylus</i> Mayr (Hymenoptera: Encyrtidae), 400	8.4.5.1	General pathogenic bacteria, 421
	8.3.2.5	<i>Nothoserphus</i> Brues (Hymenoptera: Proctotrupidae), 404	8.4.5.2	Male-killing bacteria, 421
	8.3.2.6	<i>Metastenus</i> Walker (Hymenoptera: Pteromalidae), 405	8.5	Impact of Natural Enemies on Ladybird Populations, 425
	8.3.2.7	<i>Phalacrotophora</i> Enderlein (Diptera: Phoridae), 406	8.5.1	Impact on phytophagous Coccinellidae, 426
	8.3.2.8	<i>Oomyzus scaposus</i> (Thomson) (Hymenoptera: Eulophidae, Tetrastichinae), 408	8.5.2	Impact on predatory Coccinellidae, 426
	8.3.2.9	<i>Pediobius foveolatus</i> (Crawford) (Hymenoptera: Eulophidae, Entedoninae), 409	8.5.3	Concluding note, 428
				References, 428
				9. COCCINELLIDS AND SEMIOCHEMICALS, 444
				<i>Jan Pettersson</i>
			9.1	Introduction, 445
			9.2	Aposematism and Reflex Bleeding Chemistry, 445
			9.2.1	Reflex bleeding, 445
			9.2.2	Reflex bleeding substances, 445
			9.2.2.1	Experiments, 445
			9.2.2.2	Identification, 445
			9.2.2.3	Sources, 445

9.2.2.4	Production cost, 446	9.7	Hibernation and Aggregation, 457	
9.2.2.5	Age and stage modifications, 447	9.8	Habitat Preferences: Responses to Plants and Plant Volatiles, 457	
9.2.2.6	Species specific chemistry, 448	9.8.1	Habitat selection, 457	
9.2.3	Relation to enemies and competitors, 449	9.8.2	Plant stand traits, 457	
9.2.3.1	Parasitoids, 449	9.8.3	Avoided plants, 458	
9.2.3.2	Ants, 449	9.8.4	Plant–plant interactions/attractive plants, 458	
9.2.3.3	Spiders, 449	9.9	Conclusions and Future Challenges, 459	
9.3	Semiochemicals Related to Food, 449		References, 460	
9.3.1	Plant volatiles, 449	10. QUANTIFYING THE IMPACT OF COCCINELLIDS ON THEIR PREY, 465		
9.3.1.1	Herbivore-induced plant volatiles, 449	<i>J. P. Michaud and James D. Harwood</i>		
9.3.1.2	Adult receptors for food semiochemicals, 450	10.1	Introduction, 466	
9.3.1.3	Responses of larvae, 450	10.2	Assays of Consumption, 467	
9.3.1.4	Prey sex pheromones: predator kairomones, 451	10.3	Indirect Impacts, 468	
9.3.2	Prey alarm pheromones, 451	10.4	Traditional Approaches to the Study of Predation, 468	
9.3.2.1	Aphid alarm pheromone, 451	10.4.1	Selective exclusion, 469	
9.3.2.2	Mechanism for modified responses, 451	10.4.2	Field cages, 469	
9.3.2.3	EBF release: aphid individual risks, 451	10.4.3	Cage inclusion, 470	
9.3.3	Toxic substances in prey, 451	10.4.4	Manual removal, 471	
9.3.4	Feeding stimulants for phytophagous coccinellids, 452	10.5	Statistical Approaches, 471	
9.3.5	Learning, 452	10.6	Resolving Coccinellid Impact within Complex Communities, 472	
9.4	Mating and Sex Pheromones, 453	10.6.1	Multi-species combinations, 472	
9.4.1	Chemoreceptors on the antennae, 453	10.6.2	Demographic-based estimates, 473	
9.4.2	Hydrocarbons on elytra, 454	10.7	Post-Mortem Analysis of Predation, 474	
9.4.3	Ultrastructure of the integumentary glands, 454	10.7.1	Antibody-based analysis of predation, 474	
9.5	Oviposition, 454	10.7.2	Protein marking for predation analysis, 477	
9.5.1	Oviposition deterrence: oviposition deterrence pheromones, 454	10.7.3	Detection of prey-specific DNA, 478	
9.5.2	Species-specificity of oviposition deterrence pheromone substances, 454	10.8	Conclusions, 481	
9.5.3	Active substances in oviposition deterrence pheromone tracks, 455		References, 482	
9.5.4	Relation to other aphid enemies, 455	11. COCCINELLIDS IN BIOLOGICAL CONTROL, 488		
9.5.5	Aphid abundance, 456	<i>J. P. Michaud</i>		
9.6	Egg and Pupa Protection, 456	11.1	Introduction, 489	
9.6.1	Protection of eggs, 456	11.2	The Roles of Coccinellidae in Biological Control, 489	
9.6.2	Protection of pupae, 456	11.2.1	Prey specificity, 489	
		11.2.2	Generalist coccinellids, 490	
		11.2.3	Intraguild predation, 490	

- 11.3 Scale Insects Versus Aphids as Targets of Exotic Introductions, 491
 11.3.1 Coccidophagous coccinellids, 491
 11.3.2 Aphidophagous coccinellids, 492
 11.3.3 Invasive coccinellids, 493
 11.3.4 Competitive displacement, 494
- 11.4 Augmentation of Coccinellids, 495
 11.4.1 The mealybug destroyer, 496
 11.4.2 Redistribution of coccinellids, 496
 11.4.3 Selection of source material for augmentation, 497
 11.4.4 Voltinism and diapause, 497
 11.4.5 Dietary requirements, 497
 11.4.6 Life stages for release, 498
- 11.5 Conservation, 498
 11.5.1 Alternative or supplementary food, 499
 11.5.2 Hibernation refuges, 500
 11.5.3 Habitat management, 500
 11.5.3.1 Strip-harvesting, 501
 11.5.3.2 Floral diversity (non-crop plants), 501
 11.5.3.3 Intercropping, 501
 11.5.3.4 Reduced tillage, 502
- 11.6 Ancillary Factors Influencing Biological Control by Coccinellids, 502
 11.6.1 Ant-attendance of aphid colonies, 502
 11.6.2 Timing of arrival in annual crops, 503
- 11.6.3 Interaction of biological control by coccinellids with plant structure and chemistry, 504
 11.6.4 Selective use of pesticides, 506
- 11.7 Conclusions, 509
References, 509
- 12. RECENT PROGRESS AND POSSIBLE FUTURE TRENDS IN THE STUDY OF COCCINELLIDAE, 520**
Helmut E. van Emden and Ivo Hodek
Acknowledgements, 525
References, 525
- APPENDIX: LIST OF GENERA IN TRIBES AND SUBFAMILIES, 526**
Ondřich Nedvěd and Ivo Kovář
Coccinellidae Latreille, 1807, 527
Chilocorinae Mulsant, 1846 [2:25], 527
Coccidulinae Mulsant, 1846 [6:46], 527
Coccinellinae Latreille, 1807 [5:94], 528
Epilachninae Mulsant, 1846 [4:24], 529
Exoplectrinae Crotch, 1874 [2:24], 529
Microweiseinae Leng, 1920 [4:23], 529
Ortaliinae Mulsant, 1850 [2:14], 530
Scymninae Mulsant, 1846 [11:51], 530
Sticholotidinae Weise, 1901 [6:58], 531
Subject index, 532

Colour plate pages fall between pp. 250 and pp. 251

CONTRIBUTORS

PIOTR CERYNGIER *Centre for Ecological Research,
Polish Academy of Sciences, Dziekanow Lesny, 05-092
Lomianki, POLAND*

HELMUT F. VAN EMDEN *School of Biological Sciences,
University of Reading, Whiteknights, Reading RG6
6AS UK*

EDWARD W. EVANS *Department of Biology, Utah State University, Logan, UT 84322 USA*

JAMES D. HARWOOD *Department of Entomology,
University of Kentucky, Lexington, KY 40546-0091
USA*

IVO HOĐEK *Institute of Entomology, Academy of Sciences, CZ 37005 České Budějovice, CZECH REPUBLIC*

ALOIS HONĚK *Department of Entomology, Crop Research Institute, CZ 16106 Prague 6, CZECH REPUBLIC*

IVO KOVÁŘ *Emer. Scientist of the National Museum, Prague; Current address: Zichovec, CZECH REPUBLIC*

ERIC LUCAS *Département des Sciences Biologiques, Université du Québec à Montréal, C.P. 8888 Succ. Centre-ville, Montréal, Québec H3C 3P8 CANADA*

J.P. MICHAUD *Department of Entomology, Kansas State University, 1232 240th Ave., Hays, KS 67601 USA*

OLDŘICH NEDVĚD *Faculty of Science, University of South Bohemia and Institute of Entomology, Academy of Sciences, CZ 37005 České Budějovice, CZECH REPUBLIC*

JAN PETTERSSON *Department of Ecology, Swedish University of Agricultural Sciences, Box 7044, SE-750 07 Uppsala, SWEDEN*

REMY L. POLAND *Department of Genetics, University of Cambridge, Cambridge CB2 3EH UK, recent address: Clifton College, 32 College Road, Clifton, Bristol, BS8 3JH, UK*

HELEN E. ROY *NERC Centre for Ecology and Hydrology, Crowmarsh Gifford, Oxfordshire OX10 8BB UK*

JOHN J. SLOGGETT *Maastricht Science Programme, Maastricht University, P.O. Box 616, 6200 MD Maastricht, THE NETHERLANDS*

PREFACE

For more than a decade, no volume on the general aspects of ladybirds has been published, although the predaceous major part of this coleopteran family represents an important component of the natural enemies of Sternorrhyncha (aphids, coccids, aleyrodids and psyllids). These sucking insects are among the most dangerous pests of crops, as under suitable conditions their populations increase exponentially, especially when parthenogenesis and viviparity occur.

Although classical biological control with coccinellids has recently been mostly abandoned, the increasing concern about chemical pest control has increased the need for modern types of biological control, mostly involving conservation and augmentation, within the framework of integrated pest management. For the success of these sophisticated methods of control, precise knowledge of behaviour and ecological relations is indispensable.

Such knowledge, particularly in the areas of ethology and molecular genetics, has accumulated in the last decade and reviewing these areas is an important novel contribution of this book. We hope that, at this stage, it will help to improve conservation and augmentation control, but also that it will stimulate further research.

The book was very unfortunately interrupted by the premature death of Professor Michael Majerus, Department of Genetics, Cambridge University, who was our proposed author for Chapters 2 and 8. We were extremely lucky to find very able successors, John Sloggett (Chapter 2), and Helen Roy and Remy Poland

(for parasites and pathogens in Chapter 8), who fulfilled their task with great success. Many thanks!

We should like to thank the other authors of chapters for contributing their expertise so readily and for reacting so positively and constructively to our suggestions for revision. We are also grateful for the expert help we have received with taxonomic nomenclature: Dr O. Nedvěd of the University of South Bohemia, Czech Republic, and from the UK Drs V.F. Eastop, C.H. Lyal and D.J. Williams (Natural History Museum), Drs R.T.V. Fox and S.L.Jury (University of Reading) and Professor D.L.J. Quicke (Imperial College). Dr Lyal deserves additional thanks for his advice in relation to Chapter 1.

We should also like to thank Dr Ward Cooper of Wiley-Blackwell for enthusiastically agreeing to publish our book and Mr Kelvin Matthews for his help with the publication process. We should also like to thank the other Wiley-Blackwell staff who have worked so helpfully and efficiently on the production of this book.

Ivo Hodek
České Budějovice, Czech Republic

Helmut F. van Emden
Reading, UK

Alois Honěk
Prague, Czech Republic

March 2012

INTRODUCTION

The reader may find the following information helpful in order to use the book more easily.

To keep the flow of the text unbroken and to save repetition and space, taxonomic affiliations of organisms and species authorities are given only in the Taxonomic Glossary (following section, while the Subject Index is at the end of book, as usual). Readers should also note that the Latin names as given in this glossary are used in the text and tables, and therefore may not be the same as the older Latin names given in the original papers cited. However, because the species names

have usually not changed, and the older names are also listed in the glossary, any confusion will be avoided.

The very common generic names are always abbreviated as follows throughout the text of all chapters: *A.* for *Adalia*; *C.*, *Coccinella*; *Cer.*, *Ceratomegilla*; *Chil.*, *Chilocorus*; *Col.*, *Coleomegilla*; *Har.*, *Harmonia*; *Hip.*, *Hippodamia*; *P.*, *Propylea*.

Some phenomena are discussed in more than one chapter in the book, but from different angles; the reader's attention is directed to this by cross-references to the section number.

TAXONOMIC GLOSSARY

With common names where appropriate

Coleoptera: Family Coccinellidae

Synonyms [in square brackets] as well as currently valid names are listed in alphabetical order.

Adalia bipunctata (L.) – two spot ladybird [*Adalia fasciatopunctata* (Faldermann)]

Adalia conglomerata (L.)

Adalia decempunctata (L.) – ten spot ladybird

Adalia deficiens Mulsant

[*Adalia fasciatopunctata* (Faldermann)] = *Adalia bipunctata* (L.)

[*Adalia flavomaculata* (De Geer)] = *Lioadalia flavomaculata* (De Geer)

Adalia tetraspilota (Hope)

[*Adonia*] = *Hippodamia*

[*Adonia arctica* (Schneider)] = *Hippodamia arctica* (Schneider)

[*Adonia variegata* (Goeze)] = *Hippodamia variegata* (Goeze)

Afidenta misera (Weise) [*Afidenta mimetica* Dieke]

Aphidentula bisquadripunctata (Gyllenhal) [*Epilachna bisquadripunctata* (Gyllenhal)]

Afissula rana Kapur

Afissula sanscrita (Crotch)

Aiolocaria hexaspilota (Hope) [*Aiolocaria mirabilis* (Motschulsky)]

[*Aiolocaria mirabilis* (Motschulsky)] = *Aiolocaria hexaspilota* (Hope)

Alloneda dodecaspiota (Hope)

Anatis halonis Lewis

Anatis labiculata (Say) – fifteen spotted lady beetle

Anatis mali (Say) – eyespotted lady beetle

Anatis ocellata (L.) – eyed ladybird

Anatis quindecimpunctata (DeGeer)

Anegleis cardoni (Weise)

[*Anisolemmia dilatata* (F.)] = *Megalocaria dilatata* (F.)

Anisolemmia tetrasticta Fairmaire

Anisosticta bitriangularis (Say)

Anisosticta novemdecimpunctata (L.) – water ladybird

Anisosticta sibirica Bielawski

Aphidecta oblitterata (L.) – larch ladybird

Apolinus lividigaster (Mulsant) [*Scymnoides lividigaster* (Mulsant)]

Axinoscymnus cardilobus Ren & Pang

[*Azya trinitatis* (Marshall)] = *Pseudoazyta trinitatis* (Marshall)

Azya orbignera Mulsant

Brachiacantha quadripunctata Melsheimer

Brachiacantha ursina (F.)

Brumoides septentrionis (Weise)

Brumoides suturalis (F.) – three-striped lady beetle [*Brumus suturalis* (Mani)]

[*Brumus quadripustulatus* (L.)] = *Exochomus quadripustulatus* (L.)

[*Brumus suturalis* (Mani)] = *Brumoides suturalis* (F.)

Bulaea lichatschovi (Hummel)

Callicaria superba (Mulsant)

Calvia albida Bielawski

Calvia decemguttata (L.)

Calvia duodecimmaculata Gebler

Calvia muiri Timberlake [*Eocaria muiri* Timberlake]

Calvia quatuordecimguttata (L.) – cream-spot ladybird

Calvia quindecimguttata (F.)

Calvia shiva Kapur

Calvia shiva pasupati Kapur

Calvia shiva pinaki Kapur

Calvia shiva trilocana Kapur

Ceratomegilla barovskii kiritschenkoi (Semenov-Tian-Shanski) [*Spiladelpha barovskii kiritschenkoi* Semenov-Tian-Shanski]

- Ceratomegilla notata* (Laicharting)
Ceratomegilla undecimnotata (Schneider) [*Semiadalia undecimnotata* (Schneider)]
Cheiromenes lunata (F.)
Cheiromenes propinqua vicina (Mulsant) [*Cheiromenes vicina* (Mulsant), *Cydonia vicina nilotica* Mulsant]
[*Cheiromenes sexmaculata* (F.)] = *Menochilus sexmaculatus* (F.)
Cheiromenes sulphurea (Olivier)
[*Cheiromenes vicina*] = *Cheiromenes propinqua vicina* (Mulsant)
[*Chilocorus baylei* (Blackburn)] = *Chilocorus malasiae* Crotch
Chilocorus bijugus Mulsant
Chilocorus bipustulatus (L.) – heather ladybird
Chilocorus braeti Weise
Chilocorus cacti (L.)
Chilocorus circumdatus (Gyllenhal)
Chilocorus discoideus Crotch
Chilocorus distigma Klug
Chilocorus geminus Zaslavsky
Chilocorus hauseri Weise
Chilocorus hexacyclus Smith
Chilocorus infernalis Mulsant
Chilocorus inornatus Weise
Chilocorus kuwanae Silvestri
Chilocorus malasiae Crotch [*Chilocorus baylei* (Blackburn)]
Chilocorus nigripes Mader
Chilocorus nigritus (F.)
Chilocorus orbus Casey
Chilocorus quadrimaculatus (Weise)
Chilocorus renipustulatus (Scriba) – kidney-spot ladybird
Chilocorus rubidus Hope
Chilocorus similis (Rossi)
Chilocorus stigma (Say) – twice-stabbed lady beetle
Chilocorus tricyclus Smith
Chnootriba similis (Thunberg)
Cleobora mellyi (Mulsant) – Tasmanian ladybird
Clitostethus arcuatus (Rossi)
Clitostethus oculatus (Blatchley) [*Nephaspis oculatus* (Blatchley)]
Coccidophilus citricola Brèthes
Coccidula rufa (Herbst)
Coccidula scutellata (Herbst)
[*Coccinella algerica* Kovář] = *Coccinella septempunctata algerica* Kovář
Coccinella californica Mannerheim
Coccinella explanata Miyatake
Coccinella hieroglyphica L. – hieroglyphic ladybird
Coccinella leonina transversalis F. [*Coccinella repanda* Thunberg, *Coccinella transversalis* F.]
Coccinella luteopicta (Mulsant)
Coccinella magnifica Redtenbacher – scarce seven spot ladybird [*Coccinella divaricata* Olivier)]
Coccinella monticola Mulsant
Coccinella nigrovittata Kapur [*Tytthaspis trilineata* (Weise)]
Coccinella novemnotata Herbst – nine-spotted lady beetle
Coccinella quinquepunctata L. – five spot ladybird
Coccinella reitteri Weise
[*Coccinella repanda* Thunberg] = *Coccinella leonina* F.
Coccinella septempunctata L. – seven spot ladybird
Coccinella septempunctata algerica Kovář [*Coccinella algerica* Kovář]
Coccinella septempunctata brucki Mulsant
[*Coccinella sinuatomarginata* Faldermann] = *Coccinula sinuatomarginata* (Faldermann)
[*Coccinella transversalis* F.] = *Coccinella leonina transversalis* F.
Coccinella transversoguttata Faldermann – transverse lady beetle
Coccinella transversoguttata richardsoni Brown
Coccinella trifasciata L. – three-banded lady beetle
Coccinella undecimpunctata L. – eleven spot ladybird
[*Coccinella undecimpunctata aegyptiaca* Reiche] = *Coccinella undecimpunctata menetriesi* Mulsant
Coccinella undecimpunctata menetriesi Mulsant
[*Coccinella undecimpunctata aegyptiaca* Reiche]
Coccinula crotchi (Lewis)
Coccinula quatuordecimpustulata (L.)
Coccinula redimita (Weise)
Coccinula sinensis (Weise)
Coccinula sinuatomarginata (Faldermann) [*Coccinella sinuatomarginata* Faldermann]
Coelophora biplagiata Swartz [*Lemnia biplagiata* (Swartz)]
Coelophora bissellata Mulsant
Coelophora duvauclii (Mulsant)
Coelophora inaequalis (F.) – common Australian lady beetle
Coelophora mulsanti (Montrouzier)
Coelophora quadrivittata Fauvel
Coelophora saucia Mulsant
Coleomegilla maculata (DeGeer) – spotted lady beetle
Coleomegilla maculata fuscilabris (Mulsant)

- Coleomegilla maculata lengi* Timberlake – twelve-spotted ladybeetle
Coleomegilla quadrifasciata (Schoenherr)
Cryptognatha simillima Sicard
Cryptognatha nodiceps Marshall
Cryptognatha signata Korschefsky
Cryptogonus ariasi (Mulsant)
Cryptogonus kapuri Ghorpade
Cryptogonus orbiculus (Gyllenhal)
Cryptogonus postmedialis Kapur
Cryptogonus quadriguttatus (Weise)
Cryptolaemus montrouzieri Mulsant – mealybug destroyer
Curinus coeruleus Mulsant – metallic blue lady beetle
Cyclonedda ancoralis (Germar)
Cyclenedda limbifer Casey
Cyclenedda munda (Say) – polished lady beetle
Cyclenedda polita Casey
Cyclenedda sanguinea (L.) – blood-red lady beetle
[*Cydonia vicina nilotica* Mulsant] = *Cheilomenes propinqua vicina* (Mulsant)
Declivitata spp.
Delphastus catalinae (Horn)
Delphastus pusillus (LeConte)
Diomus austrinus Gordon
Diomus hennesseyi Fürsch
Diomus pumilio Weise
Diomus seminulus (Mulsant)
Diomus thoracicus F.
[*Eocaria muiri* Timberlake] = *Calvia muiri* (Timberlake)
Epilachna admirabilis Crotch
[*Epilachna bisquadripunctata* (Gyllenhal)] = *Aphidentula bisquadripunctata* (Gyllenhal)
[*Epilachna boisduvali* (Mulsant)] = *Henosepilachna boisduvali* Mulsant
Epilachna borealis (F.) – squash beetle
Epilachna canina (F.)
[*Epilachna 'chrysomelina'* (F.)]; used variably either for
Henosepilachna argus (Geoffroy) or *Henosepilachna elaterii* (Rossi) or *Henosepilachna vigintioctopunctata* (F.)
[*Epilachna cucurbitae*] = *Henosepilachna sumbana* Bielawski
Epilachna defecta Mulsant
Epilachna dregei Mulsant – potato ladybird
Epilachna dumerili Mulsant
Epilachna eckloni Mulsant
[*Epilachna enneasticta* Mulsant] = *Henosepilachna enneasticta* (Mulsant)
Epilachna eusema (Weise)
- Epilachna karisimbica* Weise
Epilachna marginella (F.)
Epilachna marginicollis (Hope)
Epilachna mexicana (Guérin-Méneville)
Epilachna mystica Mulsant
Epilachna nigrolimbata Thomson
[*Epilachna niponica* Lewis] = *Henosepilachna niponica* (Lewis)
Epilachna paenulata (Germar)
[*Epilachna philippinensis* Dieke] = *Henosepilachna vigintisexpunctata* (Boisduval)
[*Epilachna pusillanima* Mulsant] = *Henosepilachna pusillanima* (Mulsant)
[*Epilachna pustulosa* Kôno] = *Henosepilachna pustulosa* (Kôno)
Epilachna quadricollis (Dieke)
[*Epilachna septima* Dieke] = *Henosepilachna septima* (Dieke)
[*Epilachna sparsa orientalis* Dieke] = *Henosepilachna vigintioctopunctata* (F.)
Epilachna undecimvariolata (Boisduval)
Epilachna varivestis Mulsant – mexican bean beetle
[*Epilachna vigintioctomaculata* Motschulsky] = *Henosepilachna vigintioctomaculata* (Motschulsky)
[*Epilachna vigintioctomaculata* Motschulsky] = *Henosepilachna vigintioctomaculata* (Motschulsky)
Epilachna vigintisexpunctata (Boisduval)
[*Epilachna yasutomii* (Katakura)] = *Henosepilachna yasutomii* Katakura
Epiverta chelonia (Mader)
Eriopis connexa (Germar)
Exochomus childreni Mulsant
[*Exochomus concavus* Fürsch] = *Parexochomus troberti concavus* (Fürsch)
Exochomus flavipes (Thunberg)
Exochomus flavidiventris Mader
Exochomus fulvimanus Weise
[*Exochomus lituratus* (Gorham)] = *Priscibrumus lituratus* (Gorham)
[*Exochomus melanocephalus* (Zoubkoff)] = *Parexochomus melanocephalus* (Zoubkoff)
[*Exochomus nigromaculatus* (Goeze)] = *Parexochomus nigromaculatus* (Goeze)
Exochomus quadripustulatus (L.) – pine ladybird
[*Brumus quadripustulatus* (L.)]
[*Exochomus troberti* Mulsant] = *Parexochomus troberti* (Mulsant)
Halmus chalybeus (Boisduval) – steelblue lady beetle
[*Orcus chalybeus* (Boisduval)]
[*Halyzia hauseri* (Mader)] = *Macroilleis hauseri* (Mader)

- Halyzia sanscrita* Mulsant
Halyzia sedecimguttata (L.) – orange ladybird
Halyzia straminea (Hope)
Halyzia tschitscherini Semenov
Harmonia antipoda (Mulsant in White) – antipodean ladybird
Harmonia axyridis (Pallas) – harlequin ladybird or Asian multi-colored ladybeetle
[*Harmonia breiti* Mader] = *Harmonia expalliata* Sicard
Harmonia conformis (Boisduval)
Harmonia dimidiata (F.) [*Leis dimidiata* (F.)]
Harmonia eucharis (Mulsant)
Harmonia expalliata Sicard [*Harmonia breiti* Mader]
Harmonia quadripunctata (Pontoppidan) – cream-streaked ladybird
Harmonia octomaculata (F.)
Harmonia sedecimnotata (F.)
Harmonia yedoensis (Takizawa)
Henosepilachna argus (Geoffroy) – bryony ladybird
Henosepilachna bifasciata (L.)
Henosepilachna boisduvali (Mulsant) [*Epilachna boisduvali* Mulsant]
Henosepilachna dodecastigma (Wiedemann)
Henosepilachna elaterii (Rossi) [*Epilachna chrysomelina* (F.)]
Henosepilachna enneasticta (Mulsant) [*Epilachna enneasticta* Mulsant]
Henosepilachna guttatópustulata (F.)
Henosepilachna indica (Mulsant)
Henosepilachna niponica (Lewis) [*Epilachna niponica* Lewis]
Henosepilachna ocellata (Redtenbacher)
Henosepilachna processa Li
Henosepilachna pusillanima (Mulsant) [*Epilachna pusillanima* Mulsant]
Henosepilachna pustulosa (Kôno) [*Epilachna pustulosa* Kôno]
Henosepilachna septima (Dieke) [*Epilachna septima* Dieke]
Henosepilachna sumbana Bielawski [*Epilachna cucurbitae* Richards, *Henosepilachna cucurbitae* Richards]
Henosepilachna vigintioctomaculata (Motschulsky)
[*Epilachna vigintioctomaculata* Motschulsky]
Henosepilachna vigintioctopunctata (F.) [*Epilachna vigintioctopunctata* (F.), *Epilachna sparsa orientalis* Dieke]
Henosepilachna vigintisexpunctata (Boisduval)
[*Epilachna philippinensis* Dieke]
Henosepilachna yasutomii Katakura [*Epilachna yasutomii* (Katakura)]
- Hippodamia arctica* (Schneider) [*Adonia arctica* (Schneider)]
Hippodamia caseyi Johnson
Hippodamia convergens Guerin – convergent ladybeetle
Hippodamia glacialis (F.) – glacial lady beetle
Hippodamia parenthesis (Say) – parenthesis lady beetle
Hippodamia quinquesignata (Kirby)
Hippodamia quinquesignata punctulata Le Conte
[*Hippodamia quinquesignata ambigua* LeConte]
Hippodamia septemmaculata (DeGeer)
Hippodamia sinuata Mulsant
Hippodamia tredecimpunctata (L.) – thirteen spot ladybird
Hippodamia variegata (Goeze) – Adonis ladybird, variegated lady beetle [*Adonia variegata* (Goeze)]
Hyperaspis aestimabilis Mader
Hyperaspis bigeminata (Randall)
Hyperaspis binotata (Say)
Hyperaspis campestris (Herbst)
[*Hyperaspis congressis* Watson] = *Hyperaspis conviva* Casey
Hyperaspis conviva Casey [*Hyperaspis congressis* Watson]
Hyperaspis desertorum Weise
Hyperaspis lateralis Mulsant
Hyperaspis notata Mulsant
Hyperaspis pantherina Fürsch
Hyperaspis raynevali Mulsant
Hyperaspis reppensis (Herbst)
Hyperaspis senegalensis Mulsant
Hyperaspis senegalensis hottentotta Mulsant
Hyperaspis sphaeridoides Mulsant
Hyperaspis undulata (Say)
Illeis bielawskii Ghorpade
Illeis cincta (F.)
Illeis galbula (Mulsant) [*Leptothea galbula* (Mulsant)]
Illeis koebelei Timberlake
Jauravia quadrinotata Kapur
[*Leis dimidiata* (F.)] = *Harmonia dimidiata* (F.)
[*Leis*] = *Harmonia*
[*Lemnia biplagiata* (Swartz)] = *Coelophora biplagiata* Swartz
[*Leptothea galbula* (Mulsant)] = *Illeis galbula* (Mulsant)
Lindorus lophanthae (Blaisdell) [*Rhyzobius lophantae* (Blaisdell), *Rhyzobius lorophantae* (Blaisdell)]
Liodalda flavomaculata (De Geer) [*Adalia flavomaculata* (De Geer)]
Macroilleis hauseri (Mader) [*Halyzia hauseri* (Mader)]
Macronaemia hauseri (Weise)

Megalocaria dilatata (F.) [*Anisolemnia dilatata* (F.)]
 [*Menochilus quadriplagiatus* (Swartz)] = *Menochilus sexmaculatus* (F.)
Menochilus sexmaculatus (F.) [*Cheilomenes sexmaculata* (F.), *Menochilus quadriplagiatus* (Swartz)]
Micraspis allardi (Mulsant)
Micraspis discolor (F.)
Microweisea sp.
Mulsantina hudsonica (Casey)
Mulsantina picta (Randall)
Myrrha octodecimguttata (L.) – eighteen spot ladybird
Myzia oblongoguttata (L.) – striped ladybird
Myzia subvittata (Mulsant)
Neda marginalis Mulsant
Neocalvia anastomozans Crotch
 [*Nephaspis oculatus* (Blatchley)] = *Clitostethus oculatus* (Blatchley)
Nephush bilucernarius (Mulsant)
Nephush bisignatus (Boheman)
Nephush flavifrons (Melsheimer) [*Scymnus flavifrons* Melsheimer, North America, not *Scymnus flavifrons* Blackburn, Australia]
Nephush guttulatus (LeConte) [*Scymnus guttulatus* LeConte]
Nephush includens (Kirsch)
Nephush kiesenwetteri (Mulsant) [*Scymnus kiesenwetteri* Mulsant]
Nephush ornatus (LeConte) [*Scymnus ornatus* LeConte]
Nephush quadrimaculatus (Herbst) [*Scymnus quadrimaculatus* (Herbst)]
Nephush redtenbacheri (Mulsant)
Nephush soudanensis (Sicard) [*Scymnus soudanensis* Sicard]
Oenopia billieti (Mulsant)
Oenopia conglobata (L.) [*Synharmonia conglobata* (L.)]
Oenopia kirbyi Mulsant
Oenopia lyncea (Olivier)
Oenopia sexareata (Mulsant)
 [*Olla abdominalis* (Say)] = *Olla v-nigrum* (Mulsant)
Olla v-nigrum (Mulsant) – ash-gray lady beetle [*Olla abdominalis* (Say)]
Orcus australasiae (Boisduval)
 [*Orcus chalybeus* (Boisduval)] = *Halmus chalybeus* (Boisduval)
Palaeoneda auriculata (Mulsant) [*Paleoneda miniata* (Hope)]
 [*Paleoneda miniata* (Hope)] = *Palaeoneda auriculata* (Mulsant)
Pania luteopustulata Mulsant
Paranaemia vittigera (Mannerheim)

Parastethorus nigripes (Kapur) [*Stethorus loxtoni* Britton & Lee]
Parexochomus melanocephalus (Zoubkoff) [*Exochomus melanocephalus* (Zoubkoff)]
Parexochomus nigromaculatus (Goeze) [*Exochomus nigromaculatus* (Goeze)]
Parexochomus troberti (Mulsant) [*Exochomus troberti* (Mulsant)]
Parexochomus troberti concavus (Fuersch) [*Exochomus concavus* Fuersch]
Pentilia insidiosa Mulsant
Pharoscymnus anchorago (Fairmaire)
Pharoscymnus numidicus (Pic)
Pharoscymnus ovoideus Sicard
Phymatosternus lewisii (Crotch)
Platynaspis luteorubra (Goeze)
Priscibrumus lituratus (Gorham) [*Exochomus lituratus* (Gorham)]
Priscibrumus uropygialis (Mulsant)
Propylea dissecta (Mulsant)
Propylea japonica (Thunberg)
Propylea quatuordecimpunctata (L.) – fourteen spot ladybird [*Propylaea quatuordecimpunctata* (L.)]
Pseudoazya trinitatis (Marshall) [*Azya trinitatis* (Marshall)]
 [*Pseudoscydmus tsugae*] Sasaji & McClure] = *Sasajiscymnus tsugae* (Sasaji & McClure)
 [*Pseudoscydmus kurohime* (Mityake)] = *Sasajiscymnus kurohime* (Miyatake)
Psyllobora confluens (F.)
Psyllobora vigintiduopunctata (L.) – twenty two spot ladybird [*Thea vigintiduopunctata* (L.)]
Psyllobora vigintimaculata (Say) – twenty-spotted lady beetle
 [*Pullus auritus* (Thunberg)] = *Scymnus auritus* Thunberg
 [*Pullus mediterraneus* (F.)] = *Scymnus marinus* (Mulsant)
 [*Pullus subvillosus* (Goeze)] = *Scymnus subvillosus* (Goeze)
Rhyzobius litura (F.)
 [*Rhyzobius lophanthae* (Blaisdell)] = *Lindorus lophanthae* (Blaisdell)
 [*Rhyzobius lorophanthae* (Blaisdell)] = *Lindorus lophanthae* (Blaisdell)
Rhyzobius ventralis (Erichson) – black lady beetle
Rodatus major (Blackburn)
Rodolia cardinalis (Mulsant) – vedalia beetle
Rodolia fumida Mulsant
Rodolia guerini (Crotch)

- Rodolia iceryae* Janson
Rodolia occidentalis Weise
Sasajiscymnus kurohime (Miyatake) [*Pseudoscymnus kurohime* (Miyatake)]
 [*Sasajiscymnus ningshanensis* (Sasaji & McClure)] =
 Scymnus ningshanensis Yu & Yao
Sasajiscymnus tsugae (Sasaji & McClure)
 [*Pseudoscymnus tsugae* Sasaji & McClure]
 [*Scymnodes lividigaster* (Mulsant)] = *Apolinus lividigaster* (Mulsant)
Scymnus abietis (Paykull)
 [*Scymnus aeneipennis* Sicard] = *Zagloba aeneipennis* (Sicard)
Scymnus apetzi Mulsant
Scymnus ater Kugelann
Scymnus auritus Thunberg [*Pullus auritus* (Thunberg)]
Scymnus coccivora Ayyar
Scymnus creperus Mulsant
Scymnus dorcatomoides Weise
Scymnus flavifrons Blackburn (Australia)
 [*Scymnus flavifrons* Melsheimer] (North America) = *Nephus flavifrons* (Melsheimer)
Scymnus frontalis (F.)
 [*Scymnus guttulatus* LeConte] = *Nephus guttulatus* (LeConte)
Scymnus haemorrhoidalis Herbst
Scymnus hilaris Motschulsky
Scymnus hoffmanni Weise
Scymnus impexus Mulsant
Scymnus interruptus (Goeze)
 [*Scymnus kiesenwetteri* Mulsant] = *Nephus kiesenwetteri* (Mulsant)
Scymnus lacustris LeConte
Scymnus levallanti Mulsant
Scymnus loewii Mulsant – dusky lady beetle
Scymnus louisianae Chapin
Scymnus marginicollis Mannerheim
Scymnus marinus (Mulsant) [*Scymnus mediterraneus* Iablokoff-Khnzorian, *Pullus mediterraneus* (Iablokoff-Khnzorian)]
 [*Scymnus mediterraneus*] = *Scymnus marinus* (Mulsant)
Scymnus moreletti Mulsant
Scymnus nigrinus Kugelann
Scymnus ningshanensis Yu & Yao [*Sasajiscymnus ningshanensis* (Sasaji & McClure)]
 [*Scymnus ornatus* LeConte] = *Nephus ornatus* (LeConte)
Scymnus otohime Kamiya
Scymnus posticalis Sicard
Scymnus postpictus Casey
- Scymnus pyrocheilus* Mulsant
Scymnus quadrillum Motschulsky
 [*Scymnus (Nephus) quadrimaculatus* (Herbst)] = *Nephus quadrimaculatus* (Herbst)
Scymnus rubromaculatus (Goeze)
Scymnus sinuanodus Yu & Yao
Scymnus smithianus Silvestri
Scymnus soudanensis Sicard
Scymnus subvillosus (Goeze) [*Pullus subvillosus* (Goeze)]
Scymnus suturalis Thunberg
Scymnus syriacus (Marsuel)
Scymnus tardus Mulsant
 [*Semiadalia undecimnotata* (Schneider)] = *Ceratomegilla undecimnotata* (Schneider)
Serangium parcesetosum Sicard
 [*Sidis*] = *Nephus*
[*Spiladelpha barovskii* kiritschenkoi Semenov-Tian-Shanski] = *Ceratomegilla barovskii* kiritschenkoi (Semenov-Tian-Shanski)
Sospita vigintiguttata (L.)
Stethorus bifidus Kapur
Stethorus gilvifrons (Mulsant)
Stethorus japonicus Kamiya
[*Stethorus loxtoni* Britton & Lee] = *Parastethorus nigripes* (Kapur)
Stethorus madecassus Chazeau
[*Stethorus picipes* Casey] = *Stethorus punctum picipes* Casey
[*Stethorus punctillum* Weise] = *Stethorus pusillus* (Herbst)
Stethorus punctum (LeConte) – spider-mite destroyer
Stethorus punctum picipes Casey [*Stethorus picipes* Casey]
Stethorus pusillus (Herbst) [*Stethorus punctillum* (Weise)]
Stethorus tridens Gordon
Stethorus vegans (Blackburn)
Subcoccinella vigintiquatuorpunctata (L.) – twenty four spot ladybird
[*Synharmonia conglobata* (L.)] = *Oenopia conglobata* (L.)
Synona obscura Poorani, Ślipiński & Booth
Synonycha grandis (Thunberg)
Thalassa saginata Mulsant
[*Thea vigintiguopunctata* (L.)] = *Psyllobora vigintiguopunctata* (L.)
Tytthaspis sedecimpunctata (L.) – sixteen spot ladybird
[*Tytthaspis trilineata* (Weise)] = *Coccinella nigrovittata* Kapur
[*Verania*] = *Micraspis*

Vibidia duodecimguttata (Poda)
Zagloba aeneipennis (Sicard) [*Scymnus aeneipennis* Sicard]

OTHER ORGANISMS

In alphabetical order within each taxon, and with Family given in italics within brackets after the authority

OTHER INSECTS

Dermoptera

Anechura harmandi (Burr) (*Forficulidae*) – hump earwig
Forficula auricularia L. (*Forficulidae*) – European earwig

Hemiptera: Heteroptera

Calocoris norvegicus (Gmelin) (*Miridae*) – strawberry bug
Campylomma verbasci (Meyer) (*Miridae*) – mullein bug
Catenaaultiella rugosa (Schoutenden) (*Plataspidae*)
Eurygaster integriceps Puton (*Pentatomidae*) – sunn pest or corn bug
Geocoris punctipes (Say) (*Lygaeidae*) – big-eyed bug
Hyaliodes vitripennis (Say) (*Miridae*)
Lygus hesperus (Knight) (*Miridae*) – western plant bug
Lygus lineolaris (Palidot de Beauvois) (*Miridae*) – tarnished plant bug
Lygus (*Miridae*)
Nabis (Reduvius) americiferus Carayon (*Nabidae*) – common damsel bug
Nysius huttoni White (*Lygaeidae*) – wheat bug
Orius insidiosus (Say) (*Anthocoridae*) – insidious flower bug
Podisus maculiventris (Say) (*Pentatomidae*) – spined soldier bug
Pyrrhocoris apterus (L.) (*Pyrrhocoridae*) – firebug
Sidnia kinbergi (Stål) (*Miridae*)

Hemiptera: Auchenorrhyncha

Homalodisca vitripennis (Germar) (*Cicadellidae*) – glassy-winged sharpshooter

Nilaparvata lugens (Stål) (*Delphacidae*) – brown planthopper
Philaenus spumarius (L.) (*Cercopidae*) – common froghopper or meadow spittlebug

Hemiptera: Sternorrhyncha: Aphidoidea

(Synonyms are in square brackets)

Acyrthosiphon caraganae (Cholodkovsky) (*Aphididae*)
Acyrthosiphon ignotum Mordvilko (*Aphididae*)
Acyrthosiphon kondoi Shinji (*Aphididae*) – blue alfalfa aphid
[*Acyrthosiphon nipponicum* (Essig & Kuwana)] = *Neoaulacorthum nipponicum* (Essig & Kuwana) (*Aphididae*)
Acyrthosiphon pisum (Harris) (*Aphididae*) – pea aphid
Adelges cooleyi (Gillette) (*Adelgidae*) – Cooley spruce gall adelgid
Adelges laricis Vallot (*Adelgidae*) – larch adelgid
Adelges nordmannianae (Eckstein) (*Adelgidae*)
[*Adelges nusslini* (Boerner)] = *Adelges nordmannianae* (Eckstein) (*Adelgidae*)
Adelges piceae (Ratzeburg) (*Adelgidae*) – balsam woolly adelgid
Adelges tsugae Annand (*Adelgidae*) – hemlock woolly adelgid
Aphis carduella Walsh (*Aphididae*)
[*Aphis cirsiiacanthoidis* Boerner] = *Aphis fabae cirsiiacanthoidis* Scopoli (*Aphididae*)
Aphis craccivora Koch (*Aphididae*) – cowpea aphid or groundnut aphid
Aphis cytisorum Hartig (*Aphididae*)
Aphis fabae Scopoli (*Aphididae*) – black bean aphid
Aphis fabae cirsiiacanthoidis Scopoli (*Aphididae*)
Aphis farinosa J.F.Gmelin (*Aphididae*)
Aphis glycines Matsumura (*Aphididae*) – soybean aphid
Aphis gossypii Glover (*Aphididae*) – cotton aphid or melon aphid
Aphis hederae Kaltenbach (*Aphididae*)
[*Aphis helianthi* Monell] = *Aphis carduella* Walsh (*Aphididae*)
Aphis jacobaeae Schrank (*Aphididae*)
[*Aphis laburni* Kaltenbach] = *Aphis cytisorum cytisorum* Hartig (*Aphididae*)
Aphis nerii Boyer de Fonscolombe (*Aphididae*) – oleander aphid
Aphis pomi De Geer (*Aphididae*) – green apple aphid
Aphis punicae Passerini (*Aphididae*)

- Aphis sambuci* L. (*Aphididae*) – elder aphid
Aphis spiraecola Patch (*Aphididae*) – spiraea aphid or green citrus aphid
Aphis spiraephaga F.P. Müller (*Aphididae*)
Aphis spiraephila Patch (*Aphididae*)
Aphis urticata J.F. Gmelin (*Aphididae*)
[*Aulacorthum magnoliae* (Essig & Kuwana)] = *Neoaulacorthum magnoliae* (Essig & Kuwana) (*Aphididae*)
Aulacorthum solani (Kaltenbach) (*Aphididae*) – glasshouse potato aphid or foxglove aphid
Betulaphis brevipilosa Boerner (*Aphididae*)
Betulaphis quadriflora (Kaltenbach) (*Aphididae*)
Brachycaudus helichrysi (Kaltenbach) (*Aphididae*) – leaf-curling plum aphid
Brachycaudus persicae (Passerini) (*Aphididae*) – black peach aphid
Brachycaudus prunicola (Kaltenbach) (*Aphididae*)
Brachycaudus tragopogonis (Kaltenbach) (*Aphididae*)
Brevicoryne brassicae (L.) (*Aphididae*) – cabbage aphid
Callipterinella calliptera (Hartig) (*Aphididae*)
Capitophorus elaeagni (Del Guercio) (*Aphididae*)
Cavariella konoi Takahashi (*Aphididae*)
Ceratovacuna lanigera Zehntner (*Aphididae*) – sugar cane woolly aphid
Cervaphis quercus Takahashi (*Aphididae*)
Chaetosiphon fragaefolii (Cockerell) (*Aphididae*) – strawberry aphid
Chaitophorus capreae (Mosley) (*Aphididae*)
Chaitophorus leucomelas Koch (*Aphididae*)
[*Chaitophorus versicolor* Koch] = *Chaitophorus leucomelas* Koch (*Aphididae*)
Chromaphis juglandicola (Kaltenbach) (*Aphididae*) – walnut aphid
Cinara palaestinensis Hille Ris Lambers (*Aphididae*)
Delphinioibium junackianum (Karsch) (*Aphididae*)
Diuraphis noxia (Kurdjumov) (*Aphididae*) – Russian wheat aphid
Drepanosiphum platanoidis (Schrank) (*Aphididae*) – sycamore aphid
Dysaphis crataegi (Kaltenbach) (*Aphididae*) – hawthorn–parsnip aphid
Dysaphis devecta (Walker) (*Aphididae*) – rosy leaf-curling aphid
Dysaphis plantaginea (Passerini) (*Aphididae*) – rosy apple aphid
Elatobium abietinum (Walker) (*Aphididae*) – spruce aphid
Eriosoma lanigerum (Hausmann) (*Aphididae*) – woolly apple aphid
Eucallipterus tiliae (L.) (*Aphididae*) – lime aphid
Euceraphis betulae (Koch) (*Aphididae*)
Euceraphis punctipennis (Zetterstedt) (*Aphididae*)
Hyalopterus pruni (Geoffroy) (*Aphididae*) – mealy plum aphid
Hyperomyzus carduellinus (Theobald) (*Aphididae*)
Hyperomyzus lactucae (L.) (*Aphididae*) – blackcurrant-sowthistle aphid
Laingia psammae Theobald (*Aphididae*)
Liosomaphis berberidis (Kaltenbach) (*Aphididae*)
Lipaphis pseudobrassicae (Davis) (*Aphididae*) – turnip aphid or mustard aphid
[*Longiunguis donacis* (Passerini)] = *Melanaphis donacis* (Passerini) (*Aphididae*)
Macrosiphoniella artemisiae (Boyer de Fonscolombe) (*Aphididae*)
Macrosiphoniella sanborni (Gillette) (*Aphididae*) – chrysanthemum aphid
Macrosiphum albifrons Essig (*Aphididae*) – lupin aphid
Macrosiphum euphorbiae (Thomas) (*Aphididae*) – potato aphid
[*Macrosiphum ibarae* (Matsumura)] = *Sitobion ibarae* (Matsumura) (*Aphididae*)
Macrosiphum rosae (L.) (*Aphididae*) – rose aphid
Megoura viciae Buckton (*Aphididae*) – vetch aphid
Melanaphis donacis (Passerini) (*Aphididae*)
Metopolophium dirhodum (Walker) (*Aphididae*) – rose-grain aphid
Metopolophium festucae (Theobald) (*Aphididae*) – fescue aphid
Microlophium carnosum (Buckton) (*Aphididae*)
Microsiphoniella artemisiae (Gillette) (*Aphididae*)
Mindarus abietinus Koch (*Aphididae*)
Myzocallis boernerii Stroyan (*Aphididae*)
Myzocallis carpini (Koch) (*Aphididae*)
Myzocallis castanicola Baker (*Aphididae*)
Myzocallis coryli (Goetze) (*Aphididae*) – hazel aphid or filbert aphid
Myzus cerasi (F.) (*Aphididae*) – cherry blackfly
Myzus persicae (Sulzer) (*Aphididae*) – peach-potato aphid
Myzus persicae nicotianae Blackman (*Aphididae*)
Neoaulacorthum magnoliae (Essig & Kuwana) (*Aphididae*)
Neoaulacorthum nipponicum (Essig & Kuwana) (*Aphididae*)
Neomyzus circumflexus (Buckton) (*Aphididae*)
Neophyllaphis podocarpi Takahashi (*Aphididae*)
Periphyllus californiensis (Shinji) (*Aphididae*)
Periphyllus lyropictus (Kessler) (*Aphididae*)

- Periphyllus testudinaceus* (Fernie) (*Aphididae*)
Phorodon humuli (Schrank) (*Aphididae*) – damson–hop aphid
Phyllaphis fagi (L.) (*Aphididae*)
Phylloxera glabra (von Heyden) (*Phylloxeridae*)
Pineus pini (Macquart) (*Adelgidae*)
Pseudoregma alexanderi (Takahashi) (*Aphididae*)
Pseudoregma bambucicola (Takahashi) (*Aphididae*)
Pterocallis alni (deGeer) (*Aphididae*)
Rhopalosiphum maidis (Fitch) (*Aphididae*) – corn leaf aphid
Rhopalosiphum padi (L.) (*Aphididae*) – bird cherry–oat aphid
Schizaphis graminum (Rondani) (*Aphididae*) – greenbug
Schizolachnus pineti (F.) (*Aphididae*)
Schizolachnus piniradiatae (Davidson) (*Aphididae*)
Sitobion akebiae (Shinji) (*Aphididae*)
Sitobion avenae (F.) (*Aphididae*) – grain aphid
Sitobion ibarae (Matsumura) (*Aphididae*)
Symydobius oblongus (von Heyden) (*Aphididae*)
Thelaxes dryophila (Schrank) (*Aphididae*)
[*Theroaphis maculata* (Buckton)] = *Theroaphis trifolii* (Monell) (*Aphididae*) – spotted alfalfa aphid
Theroaphis trifolii (Monell) (*Aphididae*) – spotted alfalfa aphid
Toxoptera aurantii (Boyer de Fonscolombe) (*Aphididae*) – black citrus aphid or tea aphid
Toxoptera citricidus (Kirkaldy) (*Aphididae*) – brown citrus aphid
[*Toxoptera graminum* (Rondani)] = *Schizaphis graminum* (Rondani) (*Aphididae*) – greenbug
Tuberculatus annulatus (Hartig) (*Aphididae*)
Tuberolachnus salignus (J.F. Gmelin) (*Aphididae*) – willow aphid
Uroleucon aeneum (Hille Ris Lambers) (*Aphididae*)
Uroleucon ambrosiae (Thomas) (*Aphididae*)
Uroleucon cichorii (Koch) (*Aphididae*)
Uroleucon cirsii (L.) (*Aphididae*)
Uroleucon compositae (Theobald) (*Aphididae*)
Uroleucon formosanum (Takahashi) (*Aphididae*)
Uroleucon jaceae (L.) (*Aphididae*)
Uroleucon nigrotuberculatum (Olive) (*Aphididae*)
Vesiculaphis caricis (Fullaway) (*Aphididae*)
- Agonoscena pistaciae* Burckhardt et Lauterer (*Psyllidae*)
Aleurodicus cocois (Curtis) (*Aleyrodidae*)
Aleurodicus dispersus Russell (*Aleyrodidae*) – spiralling whitefly
Aleurotuba jelinekii (Frauenfeld) (*Aleyrodidae*)
Aleyrodes proletella (L.) (*Aleyrodidae*)
Aonidiella aurantii (Maskell) (*Diaspididae*) – California red scale
Aonidiella orientalis (Newstead) (*Diaspididae*)
Aonidimytilus albus (Cockerell) (*Diaspididae*) – cassava scale
Aspidiotus destructor Signoret (*Diaspididae*) – coconut scale
Aspidiotus nerii Bouché (*Diaspididae*)
Asterolecanium sp. (*Asterolecaniidae*)
Aulacaspis tegalensis (Zehntner) (*Diaspididae*) – sugar cane scale
Aulacaspis tubercularis Newstead (*Diaspididae*)
Bemisia tabaci (Gennadius) (*Aleyrodidae*)
[*Bemisia argentifolii* Bellows] = *Bemisia tabaci* Gennadius (*Aleyrodidae*)
Chionaspis alnus Kuwana (*Diaspididae*)
Chionaspis salicis (L.) (*Diaspididae*)
Chrysomphalus aonidum (L.) (*Diaspididae*) – Florida red scale
Chrysomphalus bifasciculatus Ferris (*Diaspididae*)
Coccus hesperidum L. (*Coccidae*) – soft brown scale
Coccus viridis (Green) (*Coccidae*) – green coffee scale or soft green scale
Dactylopius opuntiae (Cockerell) (*Dactylopiidae*)
Diaphorina citri Kuwayama (*Psyllidae*) – Asian citrus psyllid
Diaspidiotus perniciosus (Comstock) (*Diaspididae*)
Dysmicoccus (*Pseudococcidae*)
Eriococcus coriaceus Maskell (*Eriococcidae*)
Eulecanium caraganae Borchsenius (*Coccidae*)
Ferrisia virgata (Cockerell) (*Pseudococcidae*) – striped mealybug
Hemberlesia lataniae (Signoret) (*Diaspididae*)
Heteropsylla cubana Crawford (*Psyllidae*)
Icerya purchasi Maskell (*Monophlebidae*) – cottony cushion scale
Lepidosaphes beckii (Newman) (*Diaspididae*) – citrus mussel scale
Lepidosaphes cornutus Ramakrishna Ayyar (*Diaspididae*)
Lepidosaphes ulmi (L.) (*Diaspididae*) – mussel scale or oystershell scale
Maconellicoccus hirsutus (Green) (*Pseudococcidae*) – pink hibiscus mealybug