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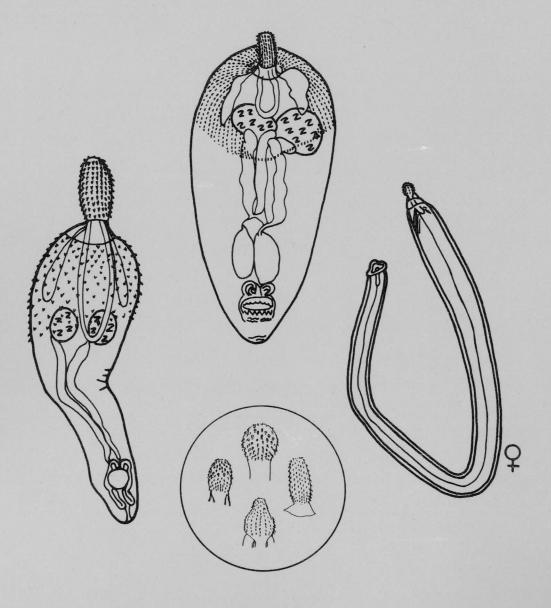
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Key to Acanthocephala Reported in Waterfowl



UNITED STATES DEPARTMENT OF THE INTERIOR Fish and Wildlife Service / Resource Publication 173

Resource Publication

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Key to Acanthocephala Reported in Waterfowl

By Malcolm E. McDonald



UNITED STATES DEPARTMENT OF THE INTERIOR FISH AND WILDLIFE SERVICE

Resource Publication 173 Washington, D.C. • 1988

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Key to Acanthocephala Reported in Waterfowl

by

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Introduction

This is the third part of a continuing series on helminths reported in waterfowl (McDonald 1974, 1981). Coots and moorhens (in Family Rallidae, Order Gruiformes) are included with the Anatidae of Anseriformes. The goal of these studies is complete coverage of waterfowl helminths of the world, although the original incentive—inadequate knowledge of the parasites of North American waterfowl—is less true now. World coverage is desirable because the world distribution of the family, tribes, and even many species of waterfowl often results in world distribution of parasites.

The format of this key follows that of the others in the series: a checklist of species, a general key to families and genera, a guide to identification of species, and keys to species of each genus that has more than one species reported in waterfowl. This group is the smallest of the series, with 52 species, of which 11 may be considered accidental, normally belonging in birds of other orders (two also normally mature only in marine mammals—although recorded in many birds—and one in freshwater rodents). One species is reported only from domestic waterfowl and one only experimentally from domestic ducklings, whereas six other species have been reported from both wild and domestic waterfowl. The frequency of occurrence and status of hosts in the checklist have been based on the literature. All Acanthocephala in waterfowl are found in the small intestine (usually in the posterior portion) and the large intestine.

Probosces usually are ovoid or sometimes elliptical. They may be elongate—narrow or wide, or

ported the eggs among the vegetation, where they are more likely to be ingested.

Intermediate hosts of Acanthocephala have been identified as crustacea of only a few orders: Amphipoda, Isopoda, and Decapoda; they are almost totally adjustic, intermediate in size, and feed on dead plant.

aquatic, intermediate in size, and feed on dead plant and animal remains. Several use fish as transport or paratenic hosts; one is known to use snakes and frogs as transport hosts to birds.

Many species (perhaps a majority) are disting

even spherical. The proboscis may be swollen in a narrow or wide band; expansion may produce a pear-

shaped (pyriform) structure, of which the narrow

end may be pointed or rounded, short or long (see

Acanthocephalan Anatomy). An important feature

of the keys is the number of rows of hooks on the

proboscis and the number of hooks in each row. As

collections have increased in the number of speci-

mens examined, they have also increased in the

amount of variation recorded, so that the range in the number of hooks has increased. The hooks are

always numbered from the anterior end; because the

arrangement of rows generally shows no difference,

it is rarely indicated. Length of a hook is indicated

from the anterior tip to the posterior end; the hook

is external on the proboscis, the root is internal and

Eggs, which are routinely used to identify species

in the keys, are really developing embryos. Three

membranes, referred to as "shells,," are present;

the middle shell in eggs of *Polymorphus* is thickest,

has an elongate shape, and shows extensions at each

pole of the embryo. No taxonomic significance is

presently known for the fibrils on the outer surface

of eggs that were recently recorded in several

species; hypothetically, they might serve to entangle

is given separately.

Many species (perhaps a majority) are distinguished by a yellow or orange color and may readily be seen with the unaided eye as orange bodies in

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the intermediate host; however, at least one common acanthocephalan of waterfowl in Eurasia (Filicollis anatis in Asellus communis) appears as a white body. Several species cause a distinct change in the behavior of the intermediate host, which renders it more vulnerable to predation by the final host (Bethel and Holmes 1973, 1974, 1977; Holmes and Bethel 1972). Life cycles generally require at least 2 months for completion. The intermediate host species are preferred foods of many juvenile and adult waterfowl.

Van Cleave (1918) believed it was significant that only one species of Acanthocephala had ever been found in an individual host, and there were no records of the occurrence of more than one genus in any host species. This is no longer true. Corynosoma and Polymorphus, for example, repeatedly occur in the same bird, and several species of Polymorphus have been recorded for the same host (at least three are recorded in the same individual).

A few members of Acanthocephala cause considerable damage and even death in waterfowl. Two species in particular, *Polymorphus minutus* and *Filicollis anatis*, have repeatedly been cited in Europe and Asia, in domestic waterfowl as well as in wild birds (Petrochenko 1958, 1971b; McDonald 1969b; Macdonald et al. 1978); The pathogenic effects were described by Petrochenko and others (McDonald 1969b). *Polymorphus minutus* is the only species that definitely occurs in North America.

Filicollis species of Eurasia (reported also in cranes in Alaska, perhaps because of the number of cranes that cross the Bering Strait between the two continents) are particularly noteworthy. The proboscis of the female perforates the intestine wall and enlarges to form a pea-sized bulb lying against the outside of the intestine in the body cavity; the body of the worm remains within the intestinal lumen. The bulb ultimately becomes embedded in connective tissue. Excellent accounts are provided in Petrochenko (1958, 1971b) and Petrochenko and Kotelńikov (1962).

Eiders are the only wild birds in which repeated outbreaks of disease and death from Acanthocephala have been reported (Grenquist 1951, 1970; Thom and Garden 1955; Clark et al. 1958; Garden et al. 1964), but dead individual birds of other species have been recorded, and at least one lake and marsh area in the United States routinely shows heavy infections with some losses. (I observed from 1,500 to 10,000 Acanthocephala in dead swans, and noted

pronounced pathological effects on their intestinal walls.) Few of the previously cited papers included detailed studies of the dead birds, and the causes of death were usually assumed. In a study of parasite numbers in eiders in Scotland in which Acanthocephala regularly occurred, Thompson (1985) commented, "In this study there is no evidence to show that P. botulus increased the mortality rate of eiders." Although he collected dead eiders regularly, no determination of the cause of death was attempted. An experimental study of antagonistic reactions between classes of helminths during multiple infections (Petrov and Egizbaeva 1972) showed significant reduction in the size and number of hymenolepidids when present with polymorphids in the small intestine.

As with other groups of helminths of waterfowl, the recent major references to Acanthocephala originated in Russia: Petrochenko (1958, 1971b [English translation]) and Petrochenko and Kotel'nikov (1962). Then a series of studies by Khokhlova (1966a, 1966b, 1971, and 1977) terminated in a partial update of Petrochenko (Khokhlova 1978), actually using the data of Schmidt (cited below). Studies of the biology and life histories of these forms up to that time are listed in McDonald (1969a, 1969b). The taxonomy of portions of the group has been revised by Schmidt (1972, 1973, 1975) and Schmidt and Kuntz (1967); Amin (1982) recently reviewed the entire phylum. Denny (1969) and Podesta and Holmes (1970) discussed the biology of Acanthocephala and other helminths carried by the intermediate hosts, Gammaridae, in Canada. Many papers on the same topic for Europe and Asia are included in the literature (McDonald 1969a). Kontrimavichus and Atrashkevich (1982) described parasitic systems and their position in the study of population biology of helminths. Bullock (1969) gave an excellent account of the techniques of preservation and mounting on microscope slides.

I could detect no distinct differences between Corynosoma sudsuche Belopol'skaya 1959 and Corynosoma mandarinca Oshmarin 1963, both reported from ducks of Primorsk, and I finally eliminated the second as a synonym. The same thing was true for Corynosoma iheringi Machado Filho 1961a, Corynosoma longilemniscatus Machado Filho 1961b, and Corynosoma molfifernandesi Machado Filho 1962a, all reduced to synonyms of Corynosoma enriettii Molfi and Fernandes 1953, of Brazil.

Polymorphus magnus Skrjabin 1913 has been merged with P. minutus Goeze 1782, following the lead of Bezubik 1957a. Bezubik showed that specimens of *P. minutus* described by Petrochenko were actually P. contortus; the true P. minutus then included P. magnus. Worms 2-12 mm long showed only one peak of variation in length for each sex, indicating only one species. Females 4-7 mm long more frequently possessed mature eggs than lacked them (precisely the range that Petrochenko said lacked them)—too large for P. minutus (actually P. contortus) and too young for P. magnus. Only researchers in England (Crompton and Harrison 1965) have followed Bezubik's lead; P. magnus has been recorded repeatedly since 1957, 34 times in wild ducks and 40 times in domestic ducks, mostly in Russia.

The form listed as *Polymorphus trochus* of Khokhlova is distinct from *P. trochus* Van Cleave 1945. *P. trochus* Van Cleave is almost universal in *Fulica americana*, and rarely occurs in ducks in North America, except occasionally when they are severely ill (twice in mallards). The form of the proboscis is strikingly different in the two sexes. *P. trochus* of Khokhlova has been reported repeatedly in wild ducks in Asia, but has not been recorded in the European coot. The form of the proboscis shows little difference between males and females. I have refrained from renaming this form, but a new name is necessary.

Common names of hosts (Table 1) used in this publication follow those of Scott (1977).

Table 1. Names of hosts.

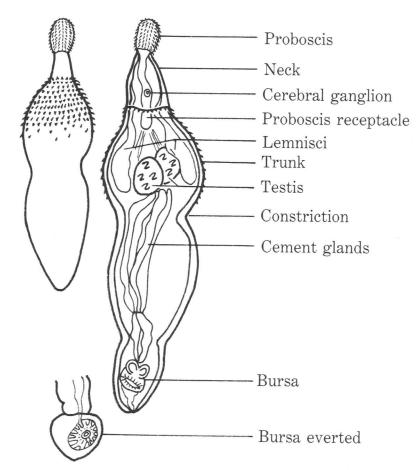
Common name	Scientific name	Common name	Scientific name
American coot	Fulica americana	Long-tailed duck	Clangula hyemalis
	(order Gruiformes)	Loon	Genus Gavia
Bahama pintail	Anas bahamensis		(order Gaviiformes)
Bay duck	Tribe Aythyini	Mallard	Anas platyrhynchos
Brazilian teal	Amazonetta brasiliensis	Mandarin duck	Aix galericulata
Brown dipper	Cinclus pallasii (order Passeriformes)	Common moorhen	Gallinula chloropus (order Gruiformes)
Coot	Genus Fulica	Musk duck	Biziura lobata
	(order Gruiformes)	Northern pintail	Anas acuta
Domestic duck	Anas platyrhynchos (dom.)	Common pochard	Aythya ferina
Domestic muscovy	Cairina moschata (dom.)	Puddle duck	Tribe Anatini
Eider	Tribe Somateriini	Rosybill	Netta peposaca
Northern gannet	Sula bassanus (order Pelecaniformes)	Sea duck	Genera Melanitta, Histrionicus, Clangula
Common goldeneye	Bucephala clangula	Swan	Genus Cygnus
Greater scaup	Aythya marila	Tufted duck	Aythya fuligula
Harlequin duck	Histrionicus histrionicus	Waterfowl	Family Anatidae
Hooded merganser	Mergus cucullatus		

Checklist of Acanthocephala Reported in Waterfowl

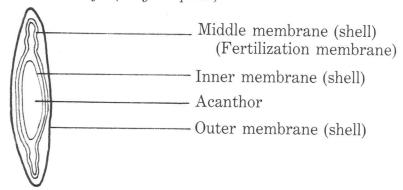
Species	Frequency of records	Status of host (wild = W)
Class Archiacanthocephala		
Family Oligacanthorthynchidae		
Neoncicola avicula (Travassos 1917)	Rare	W
ORDER PALAEACANTHOCEPHALA		
Family Echinorhynchidae		
Acanthocephalus anguillae (Müller 1789) Acanthocephalus lucii (Müller 1776) Acanthocephalus ranae (Schrank 1788)	Accidental Accidental Accidental	W W W
Family Centrorhynchidae		
Centrorhynchus aluconis (Müller 1780) Centrorhynchus teres (Westrumb 1821)	Accidental Accidental	W W
Family Polymorphidae		
Andracantha mergi (Lundström 1942) Andracantha phalacrocoracis (Yamaguti 1939) Arhythmorhynchus frassoni (Molin 1858) Arhythmorhynchus invaginabilis (Linstow 1902) Arhythmorhynchus teres Van Cleave 1929 Corynosoma anatarium Van Cleave 1945 Corynosoma constrictum Van Cleave 1918 Corynosoma enriettii Molfi and Fernandes 1953 Corynosoma peposacae (Porta 1914) Corynosoma semerme (Forssell 1904) Corynosoma strumosum (Rudolphi 1802) Corynosoma sudsuche Belopol'skaya 1959 Corynosoma tunitae Weiss 1914	Infrequent Accidental Accidental Accidental Accidental Rare Characteristic Infrequent Rare Infrequent Infrequent Rare Rare Rare	W W W W W W W, Domestic W W W
Filicollis anatis (Schrank 1788) Filicollis trophimenkoi Atrashkevich 1982 Hexaglandula paucihamatus (Heinze 1936) Polymorphus actuganensis Petrochenko 1949 Polymorphus acutis Van Cleave and Starrett 1940 Polymorphus biziurae Johnston and Edmonds 1948 Polymorphus cincli Belopol'skaya 1959 Polymorphus contortus (Bremser 1821) Polymorphus corynoides Skrjabin 1913 Polymorphus cucullatus Van Cleave and Starrett 1940 Polymorphus diploinflatus Lundström 1942 Polymorphus kostylewi Petrochenko 1949 Polymorphus marilis Van Cleave 1939	Characteristic Infrequent Accidental Frequent Infrequent Rare Experimental Common Infrequent Rare Frequent Infrequent	W, Domestic W W W W W Domestic W W W W Domestic W W W W W W W W Domestic W W

Species	Frequency of records	Status of host (wild = W)
Polymorphus mathevossianae Petrochenko 1949	Frequent	\mathbf{w}
Polymorphus meyeri Lundström 1942	Rare	W
Polymorphus miniatus (Linstow 1896)	Rare	W
Polymorphus minutus (Goeze 1782)	Characteristic	W, Domestic
Polymorphus obtusus Van Cleave 1918	Infrequent	W
Polymorphus paradoxus Connell and Corner 1957	Infrequent	W
Polymorphus phippsi Kostylev 1922	Frequent	W
Polymorphus pupa (Linstow 1905)	Frequent	W
Polymorphus striatus (Goeze 1782)	Frequent	W, Domestic
Polymorphus strumosoides Lundström 1942	Frequent	W
Polymorphus swartzi Schmidt 1965	Rare	W
Polymorphus trochus Van Cleave 1945	Common	W, coot
Polymorphus (trochus of Khokhlova 1966)	Frequent	W
Profilicollis altmani (Perry 1942)	Rare	W
Profilicollis arcticus (Van Cleave 1920)	Infrequent	W
Profilicollis botulus (Van Cleave 1916)	Frequent	W
Profilicollis formosus (Schmidt and Kuntz 1967)	Rare	Domestic
Profilicollis major (Lundström 1942)	Infrequent	W
Family Plagiorhynchidae		
Plagiorhynchus cylindraceus (Schrank 1788)	Accidental	W
Plagiorhynchus gracilis Petrochenko 1958	Infrequent	W

Acanthocephalan Anatomy



Ovum with Embryo (Polymorphus)



Variations of Proboscis



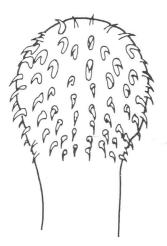




Elongate ovoid



Pyriform



Spherical

Key to Families and Genera

(Acanthocephala Reported in Waterfowl)

1.	Body (trunk) without spines
2.	Trunk with at least a few spines, usually on anterior portion of body(F. Polymorphidae)—5 Male with less than eight cement glands; proboscis sheath sac-like, with two muscular layers
	Male with eight cement glands, not definitely paired; proboscis sheath a single-walled sac, not stout; proboscis hooks in 5–6 transverse circles, very powerful, roots in first (apical) row of hooks bifurcated; male 12 mm long, female 400 mm long; eggs oval; accidental in waterfowl
3.	Males with six cement glands, of highly variable form (Fig. 1.1); sometimes with lateral or anteriorly directed processes from main root of proboscis hooks (Fig. 1.2); accidental in waterfowl, normally in fish or amphibians
4.	Males with three very long narrow cement glands
	oval
5.	Anterior trunk spines in two horizontal zones, separated by small gap (Fig. 1.5) Andracantha Anterior trunk spines in one zone
6.	Spines not limited to anterior region of trunk, some (genital) occurring at posterior end; anterior trunk spines often extending more posteriad on ventral surface (Fig. 1.6)
	Spines limited to anterior, none at posterior end
7.	Hooks on midventral surface of proboscis often distinctly larger and heavier than other hooks at corresponding level on dorsal surface; proboscis usually enlarged (swollen) near center, spindle-shaped (Fig. 1.7)
8.	Hooks of dorsal and ventral sides of proboscis similar in size (Fig. 1.8) Arhythmorhynchus 8
0,	Body markedly sexually dimorphic; proboscis of female spherical, with reduced hooks in star-shaped radiating rows at tip (Fig. 1.10; actually on proboscis portion of sphere formed from inflated proboscis and anterior portion of neck), sphere usually lying outside intestinal wall due to perforation, with body of worm still inside intestine; proboscis of male nearly orbicular, with 18 longitudinal rows of 10–11 hooks each, 21–31 µm long, anterior with roots; 6 compact reniform cement glands
	Proboscis spherical to cylindrical, similar in both sexes, row of hooks extend entire length of proboscis, proboscis entirely within intestine of host
9.	Four cement glands in male, elongate, narrow, lying parallel in one bundle; proboscis spherical to cylindrical, constriction in body usually evident
	Six cement glands in male, intestine-shaped, in two groups of three each (Fig. 1.11); proboscis ovoid, with eleven longitudinal rows of hooks; constriction on trunk slight (females fall into <i>Polymorphus</i> key)
0.	Eggs oval; proboscis spherical, rarely slightly ovoid (Fig. 1.12)
	Eggs elongate, middle shell thick, extended and narrowed at poles; proboscis ovoid, pyriform, or cylindrical (Fig. 1.9)

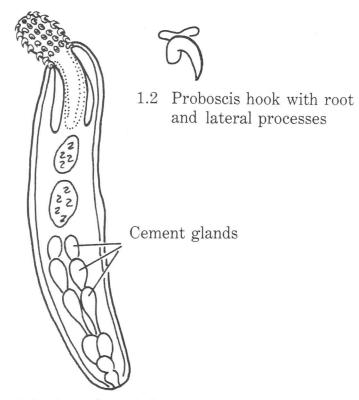
Guide to Identification of Species

(Acanthocephala Reported in Waterfowl)

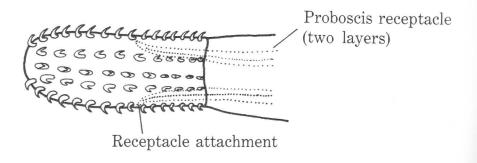
Note: Includes all genera reported in waterfowl in recent literature.

Genus	Species reported in waterfowl
$\overline{Acanthocephalus}$	3 species in waterfowl, of perhaps 35 in genus; all accidental. See key to species.
Andracantha	2 species in waterfowl, of 3 in genus. See key to species.
Arhythmorhynchus	3 species, of perhaps 25 in genus. See key to species.
Centrorhynchus	2 species, of perhaps 60 in genus. See key to species.
Corynosoma	8 species, of perhaps 30 in genus. See key to species.
Falsifilicollis	Synonym of Profilicollis.
Filicollis	2 species, all in genus. See key to species.
Hexaglandula	H. paucihamatus, of 4 species in genus. Perhaps accidental, type host a gull. Genus
0	perhaps belongs in Polymorphus; see Petrochenko 1958 or 1971b for description.
Neoncicola	N. avicula, of 2 in genus; rare; South America; see Petrochenko 1958 or 1971b for
	description, under Prosthenorchis.
Parafilicollis	Synonym of Profilicollis.
Polymorphus	24 species, of about 26 in genus. See key to species.
Profilicollis	5 species, of 8 in genus. See key to species.
Plagiorhynchus	2 species, of about 22 in genus. See key to species.

Fig. 1. Acanthocephalan Genera



1.1 Acanthocephalus



1.3 Centrorhynchus

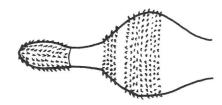
Proboscis receptacle (one layer)



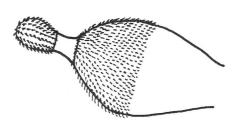
1.4 Plagiorhynchus



1.5 Posterior end, with hooks



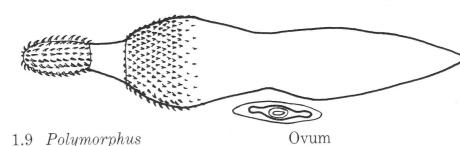
1.6 Andracantha



1.7 Corynosoma



1.8 Arhythmorhynchus



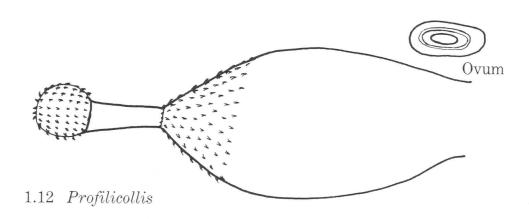
1.9 Polymorphus



1.10 Filicollis proboscis



1.11 Hexaglandula



Keys to Species

${\tt Genus}\ A can tho cephalus$

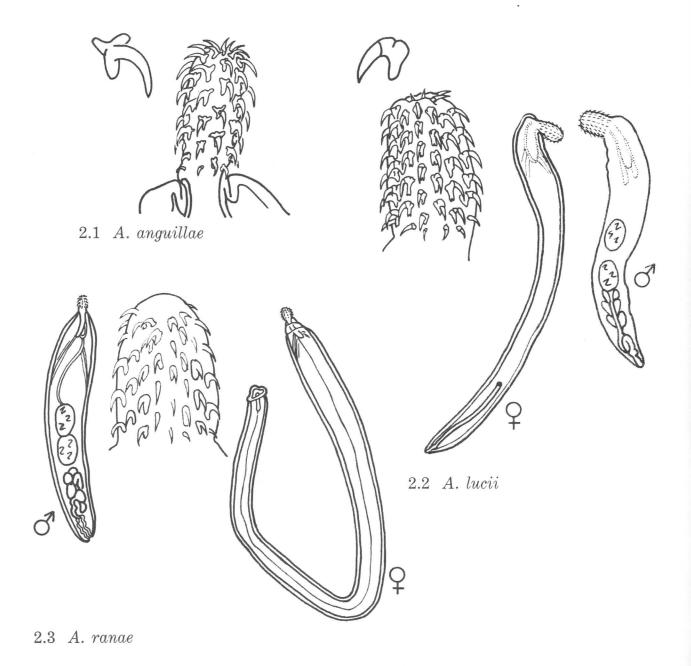
Note: Infections obtained by ingestion of freshwater isopod Crustacea.

1.	Proboscis hooks with two lateral processes at top of roots; hooks in 10 longitudinal rows, with
	6-7 hooks in each row; proboscis club-shaped, width about 1/3 of length (Fig. 2.1); accidental,
	normally in fish; North America, Eurasia
	Proboscis hooks without lateral processes on roots

6	Fomolo	9 17 mm lang mala 5 10 mm lang li 1 i 1 a 1 a 1 a 1 a 1 a 1 a 1 a 1 a 1
4	2. remaie	8–17 mm long, male 5–12 mm long, cylindrical; proboscis hooks in 12–16 longitudinal rows
	of 7-	hooks each; proboscis 500-700 μm long, 250-300 μm wide; lemnisci about 1,600 μm long
	(Fig.	2.2); accidental, normally in fish; Europe
	Female	20-60 mm long, male 5-12 mm long, cylindrical; proboscis hooks mostly in 16 (12-20)
	longi	udinal rows, with 4-6 hooks in each row; proboscis 450-500 µm long, 300 µm wide; lem-
	nisci	600-800 µm long (Fig. 2.3); accidental, normally in Amphibia; North America, Europe
		A range

Descriptions: Petrochenko 1956, 1971a.

Fig. 2. Genus Acanthocephalus

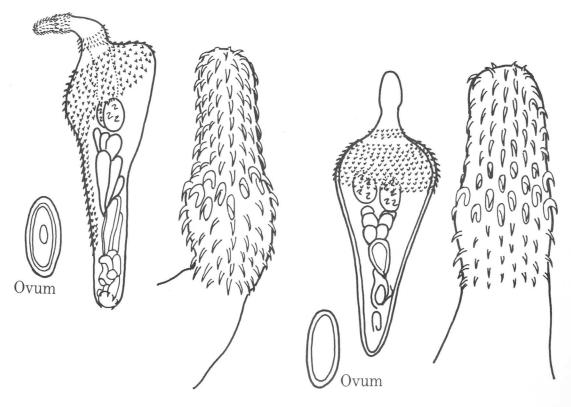


Genus Andracantha

Note: Source of infections unknown; probably marine.

Descriptions: Schmidt 1975.

Fig. 3. Genus Andracantha



3.1 A. phalacrocoracis

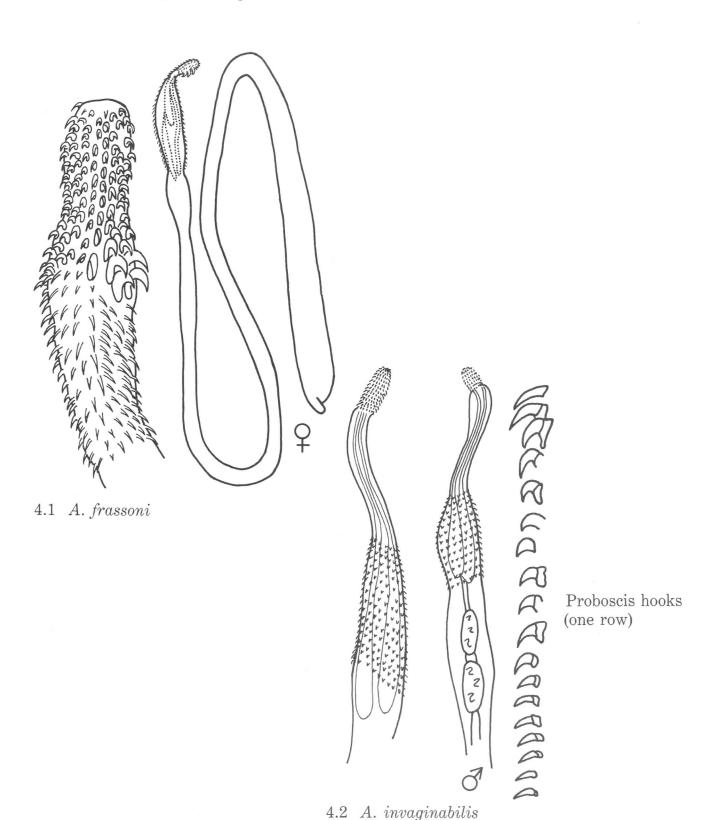
3.2 A. mergi

Genus Arhythmorhynchus

Note: One species obtained by ingestion of marine decapod Crustacea; fish serve as transport hosts.

Descriptions: Petrochenko 1958, 1971b.

Fig. 4. Genus Arhythmorhynchus

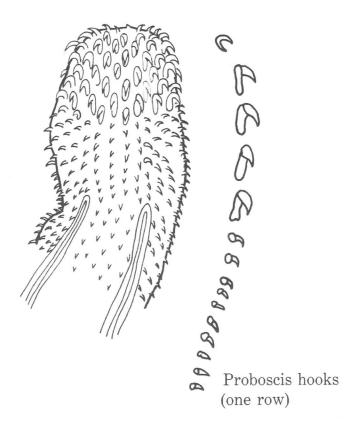


Genus Centrorhynchus

Note: Source of infections unknown; genus mostly in terrestrial birds. *Corynosoma aluconis* in snakes and amphibians as transport hosts.

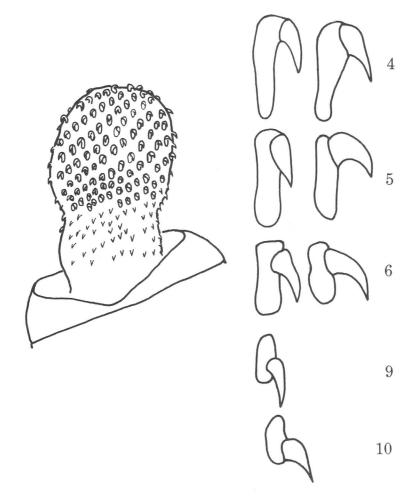
Descriptions: Petrochenko 1958, 1971b.

Fig. 5. Genus Centrorhynchus



5.1 C. aluconis





5.2 C. teres

Genus Corynosoma

Note: Infections obtained by ingestion of freshwater (one species) or marine (two species) amphipod and isopod Crustacea, or by ingestion of marine fish serving as transport hosts (two species).

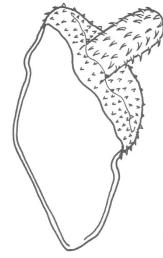
1.	Triangular shape, showing spines facing 1 mm, on slant	Z
	Anterior end slanted to ventral side, to middle or beyond	

- 6. Forebody swollen, strong constriction below forebody; body 2.28–6.0 mm long; proboscis cylindrical, swollen below middle, largest hooks at about swollen area, 41–47 μ m long, other hooks 30–41 μ m long at tip and 35–41 μ m long at base of proboscis; testes oval, overlapping about 1/3; eggs 80–108 μ m long by 12–16 μ m wide (Fig. 6.5); common in ducks; North America . .

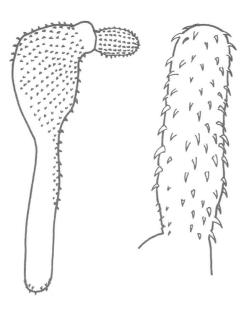
Body elongate, broadening anteriorly, slight constriction; body 8–11 mm long; body club-shaped, widened forward, anterior part with strong conical spines; males with spines present sparingly at posterior end; proboscis 340–430 μm long, with 14–18 longitudinal rows of 12–14 hooks each; largest hooks number 4; lemnisci cylindrical, about twice as long as proboscis sheath; eggs 64–68 μm long by 17–18 μm wide; rare, in rosybills; South America (Brazil, Argentina)

Descriptions: Petrochenko 1958, 1971b; Belopol'skaya 1959; Machado Filho 1961a, 1961b, 1962; Schmidt 1965b; Oshmarin 1963; Bourgeois and Threlfall 1982.

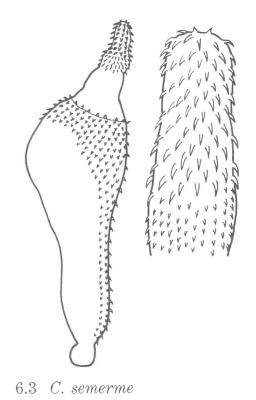
Fig. 6. Genus Corynosoma

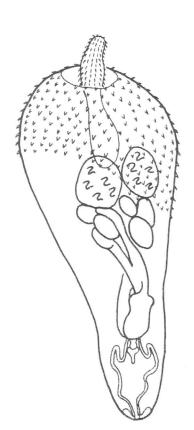


6.1 C. tunitae

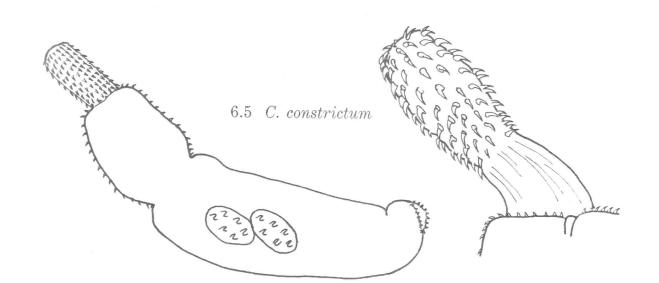


6.2 C. strumosum



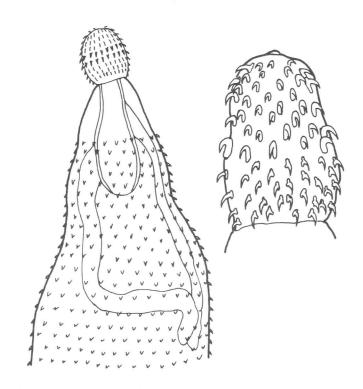


6.4 C. sudsuche





6.6 C. anatarium



6.7 C. enriettii

Family Polymorphidae

Note: Infections obtained by ingestion of freshwater isopod Crustacea. Salamanders serve experimentally as transport hosts in one species (*F. trophimenkoi*).

Proboscis larger, hooks up to 48 µm long in male, reduced in female; body 6–7.4 mm long in male, 14.3–21 mm long in female (reported 2/3 as large as F. anatis); eggs 84–118 µm by 25.6–39.3 µm, with surface covered by fine fibrils (reported 1.5 times larger than F. anatis), middle shell weakly projecting at poles; mature female flat–ovate, tablet-shaped with bulb on ventral side posterior of anterior end (bulb 2.275–3.212 mm diameter; Fig. 7.2); infrequent, in long-tailed ducks, greater scaups, and eiders; Asia (Chukotka) F. trophimenkoi

Descriptions: Petrochenko 1958, 1971b; Atrashkevich 1982.

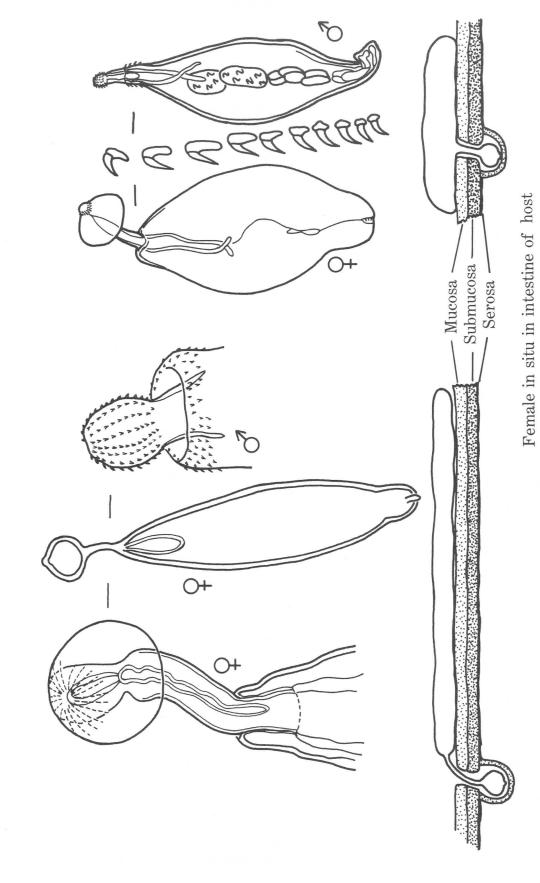


Fig. 7. Genus Filicollis

Genus Polymorphus

Family Polymorphidae

Note: Infections obtained by ingestion of freshwater (seven species) or marine (one species) amphipod Crustacea. One species also uses fish as transport hosts.

1.	Proboscis markedly sexually dimorphic, strongly inflated pear-shaped in female, ovoid without
	inflation in male; in female posterior of proboscis 380-560 μm long, 310-490 μm in diameter,
	anterior end nipplelike, 100–175 μm in diameter; in male proboscis 385–560 μm long, 210 μm
	in diameter, elliptic; with 20 longitudinal rows of hooks, alternately 11 and 12 hooks each, largest
	46-53 μm long (numbers 3 and 4; Fig. 8.1); infrequent or accidental in puddle ducks, common
	and normal in American coots; North America
	Proboscis not sexually dimorphic, same in both sexes
2	Force wide evel inner shell without elements as least the
4.	Eggs wide-oval, inner shell without elongate polar prolongations
	Eggs elongate, middle shell particularly spindle-shaped, with polar prolongations often enlarged
	at tips (not mentioned further) 5

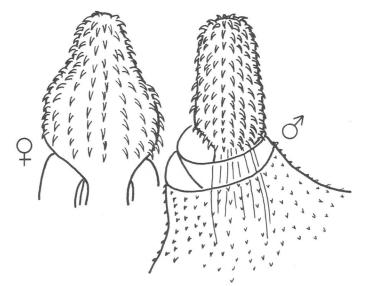
	Proboscis almost as broad as long, 300 μ m long by 300 μ m wide in male, 400 μ m long by 300 μ m wide in female; with 16–17 longitudinal rows of hooks; 6 hooks in each, largest hooks 63–69 μ m long (number 3); lemnisci club-shaped; body elongate, 7–14 mm long; eggs 96–108 μ m long by 18 μ m wide; rare, in mallards and tufted ducks; Europe
8.	Body less than 6 mm long
9.	Each longitudinal row of proboscis hooks with 7 (to 8 or 9) hooks
10.	Proboscis ovoid, slightly swollen, with 20 longitudinal rows of hooks (occasionally 18 or 22 rows), with 7 or 8 hooks in each row; largest hooks 40 μ m long (number 6), root 52 μ m long; first 4 hooks large, with long roots, remainder small; testes round to oval, opposite; body 1.83–3.8 mm long; eggs 105–112 μ m long by 19–20 μ m wide, surface covered by network of fine fibrils in loops (Fig. 8.6); frequent in ducks, particularly in northern pintails; Arctic Asia
11.	Body short, plump
12.	Body 5.5–10.5 mm long; constriction rather evident, in middle of body; neck conical, twice as long as proboscis; proboscis ellipsoid, with 17–18 (rarely 16) longitudinal rows of hooks, 9 (rarely 8) hooks in each row; largest hooks 63–86 μm long (number 4); lemnisci about twice as long as proboscis sheath; testes oval, opposite or oblique; eggs 102–123 μm long by 19–21 μm wide (Fig. 8.8); infrequent in ducks, normally in aquatic rodents; North America (Canada)
	Body 2–12.5 mm long, constriction evident; proboscis ovoid, slightly swollen; with 16 (sometimes 18, 20, or 22) longitudinal rows of hooks, 8–10 hooks in each row; largest hooks 67–79 μm long (number 4); first 4 with strong hooks and roots, remainder much reduced; lemnisci longer than proboscis sheath; testes oval, oblique to each other; eggs 73–149 μm long by 17–23 μm wide (Fig. 8.9); characteristic and common in Eurasia, infrequent in North America <i>P. minutus</i>
13.	Body 3.85-4.33 mm long, spindle-shaped, length of sexes identical; proboscis long-oval, swollen, with 12 longitudinal rows of hooks, 9-10 hooks in each row; largest hooks 53-71 μm long (number 5); long roots on first hooks, last four with reduced roots; lemnisci equal length of proboscis sheath; testes round, opposite; eggs 125 μm long by 17.8 μm wide (Fig. 8.10); infrequent, in puddle ducks and sea ducks; Asia

round, oblique to each other; eggs 107-124 µm long by 21-22 µm wide (Fig. 8.11); experimental

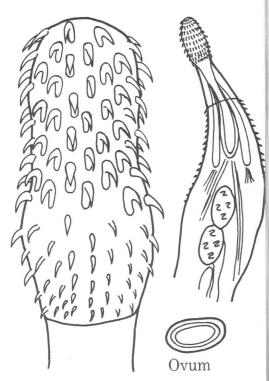
14.	Body short, swollen at anterior, 3.5–5.5 mm long, pear-shaped; spines on forebody extend 3 times as far ventrally as dorsally; proboscis wide cylindric, with 16–18 longitudinal rows of hooks, 9–11 hooks in each row; largest hooks 40–52 µm long, about middle of proboscis on swollen area; lemnisci about the length of proboscis sheath; testes round, opposite; eggs 102–111 µm long by 18–21 µm wide (Fig. 8.12); infrequent, in sea ducks; Eurasia
15.	Body elongate, forebody long, swollen, spiny; body 4.75 mm long (female); constriction below forebody at 1/3 of the length; proboscis cylindrical, inflated 2/3 from anterior end; with 18 longitudinal rows of hooks, 10–11 hooks in each row; longest hooks 60 μm long (numbers 5–6), last 3 hooks without roots; lemnisci club-shaped, slightly longer than proboscis sheath; eggs 78–86 μm long by 15–16 μm wide (Fig. 8.13); infrequent, in common goldeneyes; North America (Alaska)
16.	Each longitudinal row of proboscis hooks with 11 (and 12) alternating or total present17
17.	Body 2.63–4.46 mm long; anterior swollen, no constriction, tapering to rear; with 20 longitudinal rows of hooks on proboscis, 11 and 12 hooks alternating in rows, largest hooks 49 μm long (numbers 7 or 8), first 7 or 8 hooks well developed, with roots, remaining 4 without roots; testes round, opposite; lemnisci shorter than proboscis sheath; eggs 102–109 μm long by 17.8–23 μm wide (Fig. 8.15); frequent, in puddle ducks and sea ducks; Arctic Asia
18.	Body with evident constriction
19.	Forebody spiny, with constriction before end of spiny area; body 10.3 mm long, uniform width; proboscis long-oval, with 16 (sometimes up to 18) longitudinal rows of hooks, with 7 (sometimes 6) hooks in each row; rare, in <i>Anas</i> sp.; Magellan Straits, South America
20.	Body 6–15 mm long, swollen in forebody, tapered toward rear; proboscis with 16 longitudinal rows of hooks, with 12–16 hooks in each row; largest hooks in middle of rows; eggs 95–99 μ m long by 27 μ m wide (Fig. 8.18); frequent, in ducks and swans; Europe
21.	Body 10–13 mm long; proboscis ovoid, swollen below middle; with 22 longitudinal rows of hooks, 12 hooks in each row; largest hooks at tip, 73–85 μm long; lemnisci twice as long as proboscis sheath; testes long-oval, tandem, slightly overlapping (Fig. 8.19); rare, in hooded mergansers:

Descriptions: Petrochenko 1958, 1971b; Johnston and Edmonds 1948; Bezubik 1957a, 1957b; Connell and Corner 1957; Schmidt 1965a; Khokhlova 1966a, 1966b, 1971, 1977; Denny 1969; Podesta and Holmes 1970; Bourgeois and Threlfall 1982.

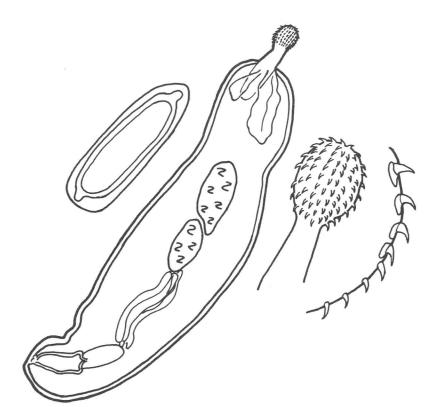
Fig. 8. Genus Polymorphus



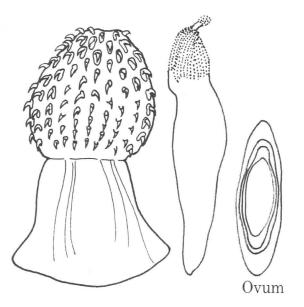
8.1 P. trochus



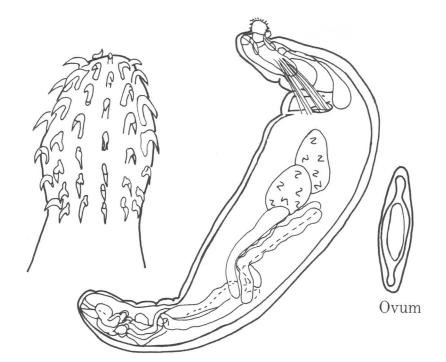
8.2 P. biziurae



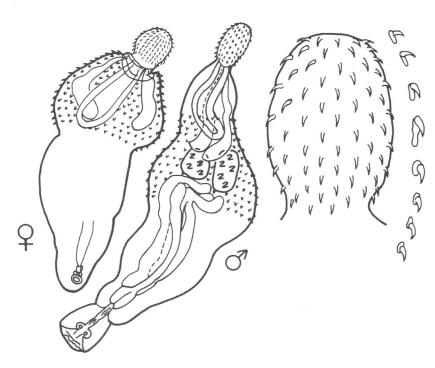
8.3 P. obtusus



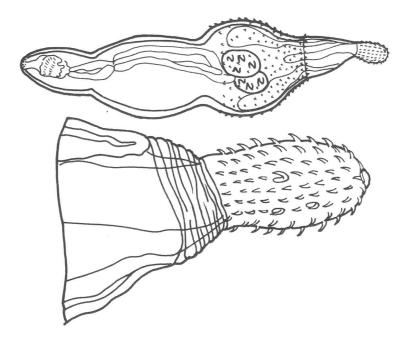
8.4 *P. pupa*



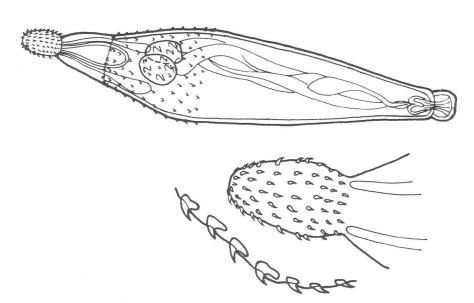
8.5 Hexaglandula paucihamatus



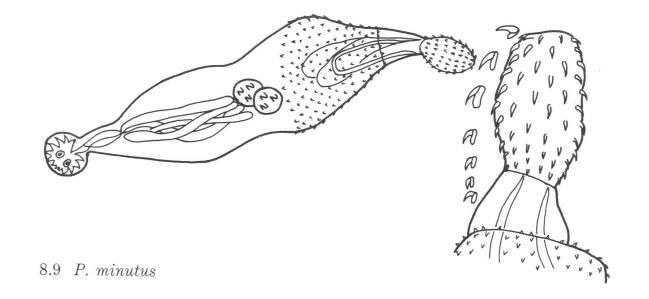
8.6 P. sp. (P. trochus of Khokhlova)



8.7 P. diploinflatus

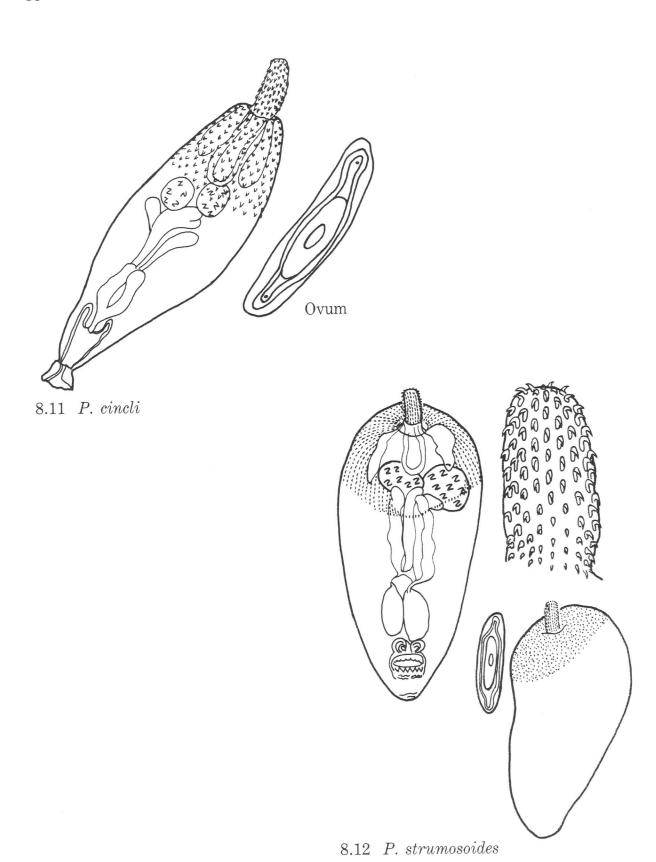


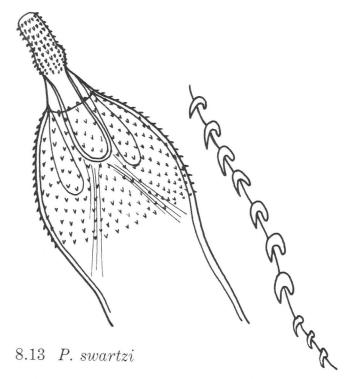
8.8 P. paradoxus

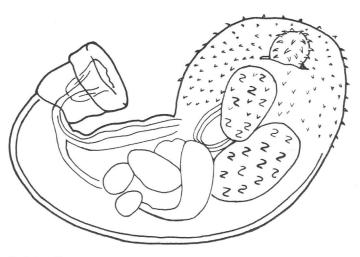




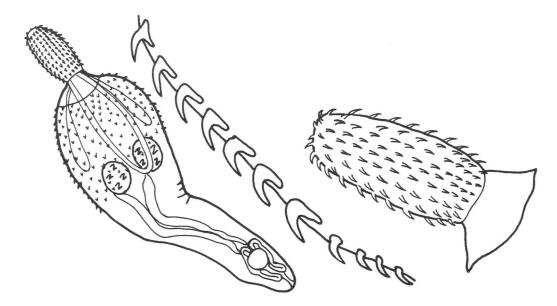
8.10 P. kostylewi



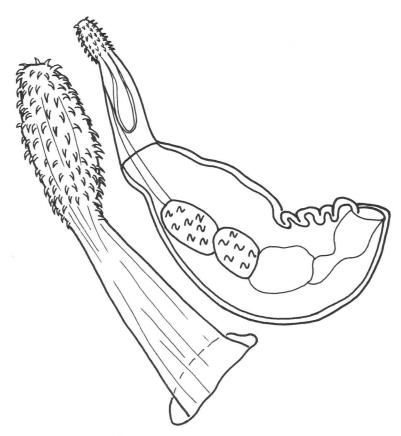




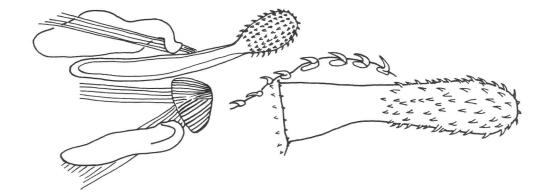
8.14 P. corynoides



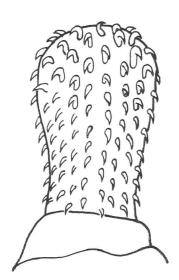
8.15 P. mathevossianae



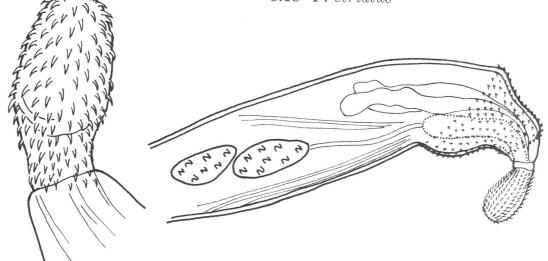
8.16 P. acutis



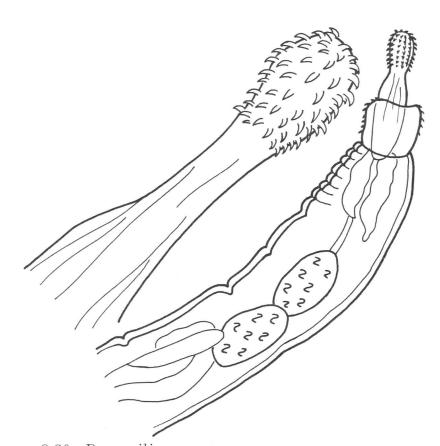
8.17 P. actuganensis



8.18 P. striatus



8.19 P. cucullatus



8.20 P. marilis

Genus Profilicollis

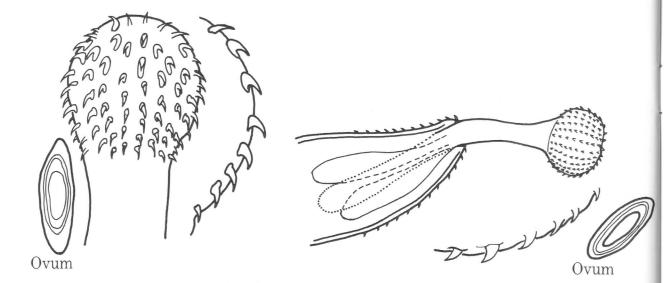
Note: Infections obtained by ingestion of decapod Crustacea, one in a freshwater species and three in marine forms.

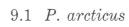
See also *Polymorphus biziurae*, a species whose proboscis is ovate or cylindrical, but whose eggs are oval, without polar prolongations of the middle membrane, and whose intermediate host may be a freshwater decapod crustacean.

P. altmani

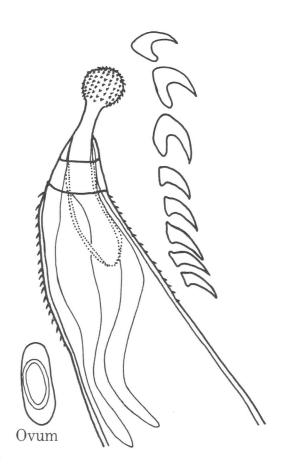
Descriptions: Petrochenko 1958, 1971b; Schmidt and Kuntz 1967; Khokhlova 1977; Bourgeois and Threlfall 1982.

Fig. 9. Genus Profilicollis

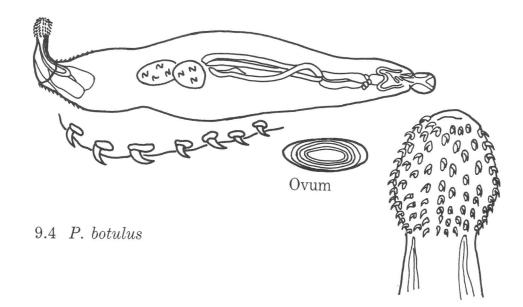


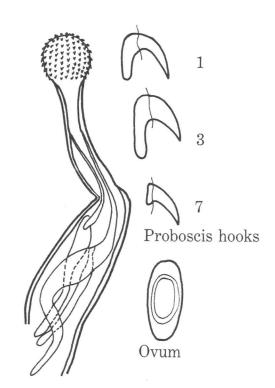


9.2 P. altmani



9.3 P. formosus





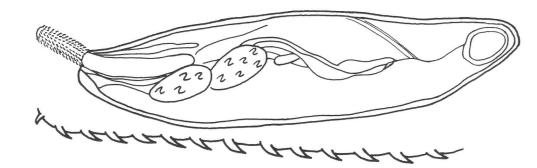
9.5 P. major

Genus Plagiorhynchus

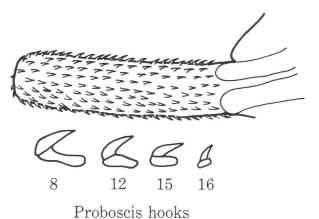
Note: Infections obtained in one species by ingestion of terrestrial isopod crustacea.

Descriptions: Petrochenko 1958a, 1971b.

Fig. 10. Genus Plagiorhynchus



10.1 P. cylindraceus



110000015 11001

10.2 P. gracilis

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This key to Acanthocephala is the third in a series describing helminths found in waterfowl from throughout the world. The format includes a checklist of 52 species, a general key to families and genera, a guide to identification of species, and keys to each genus that has more than one species reported in waterfowl.

Key words: Acanthocephala, coot, Corynosoma, helminth, Polymorphus, waterfowl.

McDonald, Malcolm E. 1988. Key to Acanthocephala Reported in Waterfowl. U.S. Fish Wildl. Serv., Resour. Publ. 173, 45 pp.

This key to Acanthocephala is the third in a series describing helminths found in waterfowl from throughout the world. The format includes a checklist of 52 species, a general key to families and genera, a guide to identification of species, and keys to each genus that has more than one species reported in waterfowl.

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A list of current Resource Publications follows.

- 157. The Breeding Bird Survey: Its First Fifteen Years, 1965–1979, by Chandler S. Robbins, Danny Bystrak, and Paul H. Geissler. 1986. 196 pp.
- 158. Techniques for Studying Nest Success of Ducks in Upland Habitats in the Prairie Pothole Region, by Albert T. Klett, Harold F. Duebbert, Craig A. Faanes, and Kenneth F. Higgins. 1986. 24 pp.
- 159. Research and Development Series: An Annotated Bibliography, 1889–1985, compiled by Thomas J. Cortese and Barbara A. Groshek. 1987.
- 160. Manual of Acute Toxicity: Interpretation and Data Base for 410 Chemicals and 66 Species of Freshwater Animals, by Foster L. Mayer and Mark R. Ellersieck. 1986. 579 pp.
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- 166. Checklist of Vertebrates of the United States, the U.S. Territories, and Canada, by Richard C. Banks, Roy W. McDiarmid, and Alfred L. Gardner. 1987. 79 pp.
- 167. Field Guide to Wildlife Diseases. Vol. 1. General Field Procedures and Diseases of Migratory Birds, by Milton Friend, Cynthia J. Laitman, and Randy Stothard Kampen. 1987. 225 pp.
- Mourning Dove Nesting: Seasonal Patterns and Effects of September Hunting, by Paul H. Geissler, David D. Dolton, Rebecca Field, Richard A. Coon, H. Franklin Percival, Don W. Hayne, Lawrence D. Soileau, Ronnie R. George, James H. Dunks, and S. Dwight Bunnell. 1987. 33 pp.
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- 172. Satellite Telemetry: A New Tool for Wildlife Research and Management, by Steven G. Fancy, Larry F. Pank, David C. Douglas, Catherine H. Curby, Gerald W. Garner, Steven C. Amstrup, and Wayne L. Regelin. 1988. 54 pp.