

## Cladocera (Crustacea: Branchiopoda) from the Pantanal (Brazil)

### Ветвистоусые ракообразные Cladocera (Crustacea: Branchiopoda) Пантаналья (Бразилия)

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КЛЮЧЕВЫЕ СЛОВА: Cladocera, Аноморода, Стенопода, морфология, систематика, таксономия, фауна, распределение, Бразилия, Пантаналь.

ABSTRACT: In 1996 and 1999, a series of samples was collected from seven localities in the southern Pantanal (Mato Grosso do Sul, Brazil). Fifty species belonging to one family of the Ctenopoda Sars, 1865, and six families of the Anomopoda Sars, 1865 have been identified. They are typical for tropical fauna of Brazil. Previous records of Cladocera from Pantanal are summarised, and a check-list of species from there is composed, consisting of 69 taxa determined to species level and 19 taxa to genera though some of these records require confirmation. The species list of the nearby Paraná region consists of 40 species. The morphology of some ilyocryptids, macrothricids, and chydorids has been studied in detail using optical and scanning electron microscopy; these results are described and illustrated. Current taxonomic problems are discussed.

РЕЗЮМЕ: В течение 1996 и 1999 гг. серия планктонных и бентосных проб была отобрана в семи точках Южного Пантаналья (штат Мато Гроссо ду Сул, Бразилия). В них было найдено пятьдесят видов из одного семейства отряда Стенопода Сарс, 1865 и шести семейств отряда Аноморода Сарс, 1865, найденные животные типичны для тропической Бразилии. Проанализированы все предыдущие находки ветвистоусых ракообразных из Пантаналья, и составлен проверочный лист, включающий 69 таксона определенных до уровня вида и 19 — до уровня рода, некоторые из этих определений нуждаются в проверке. Для ближайшего района (Парана) отмечено 40 видов. Морфология некоторых представителей семейств Ilyocryptidae Smirnov, 1976 sensu Smirnov, 1992, Macrothricidae Norman & Brady, 1867 и Chydoridae Stebbing, 1902 была исследована под оптическим и сканирующим электронным микроскопом, эти результаты проиллюстрированы в данной статье. Обсуждены некоторые современные проблемы таксономии Cladocera.

## Introduction

Pantanal (Mato Grosso do Sul, Brazil) is an immense wetland in the heart of South America with an area of about 140 000 km<sup>2</sup>. Its most important environmental factor is the fluctuating level of rivers and ground waters which results in inundations of varying duration and subsequent drying up of large areas. It is the controlling factor of the structure and function of the inundation area due to the cycling of nutrients and disposability of water. The floodplain is a very productive habitat for flora and fauna. Until now there has been only a single investigation into the effects of the annual water level fluctuations on the zooplankton [Espíndola, 1996]. In a lake connected to the Paraguay River *Moina minuta* dominated during high water periods, while *Diaphanosoma birgei* dominated during low water. Zooplankton reacts differently when the water level rises and falls with fluctuating ground water level, as is the case of the Nhecolândia region. Here there are many lakes, some of which dry up while others are saline [Mourão, 1989; Sakamoto et al., 1999]. Cladocerans and some copepods can survive these harsh conditions by producing resting eggs that are still viable even after passing through the digestive system of higher animals.

Calheiros & de Oliveira [1999] considered only 6 papers on the limnology of the Pantanal region. The first author to mention the Pantanal was Daday [1905], in his massive volume about the freshwater microfauna of Paraguay. He examined many samples collected by Prof. J. D. Anisits mainly from Paraguay, including one from “Corumba, Matto Grosso, inundation pool of the River Paraguay”. From this sample he determined 17 cladoceran and two copepod species. We found 14 other papers with records of microcrustacean species: Matsumura-Tundisi [1986], Mourão [1989], Reid & Moreno [1990], Miranda [1992], Por [1995], Lima [1996], Espíndola et al. [1996], Reid (1997), Bonecker et al.

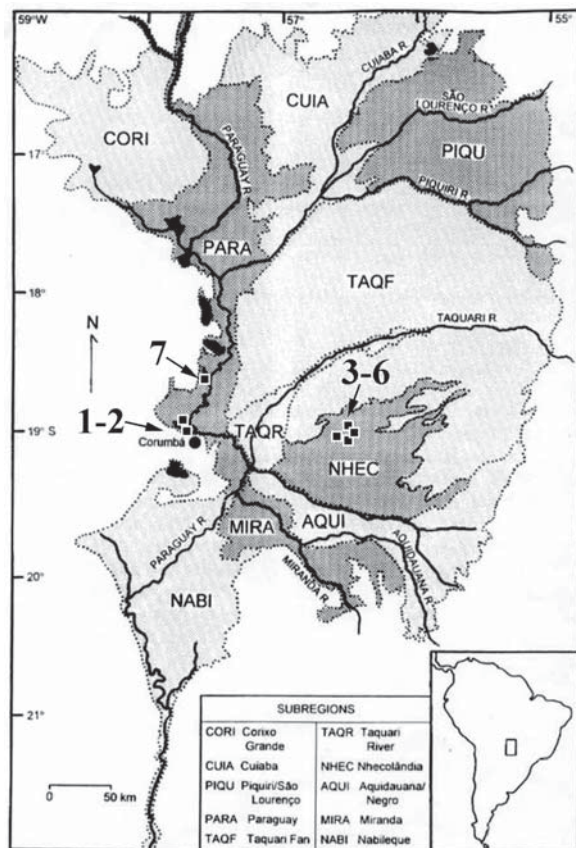


Fig. 1. Subregions of the Pantanal wetland (adopted from Hamilton et al. [1996] with kind permission of the E. Schweizerbart'sche Velagsbuchhandlung, Stuttgart).

Рис. 1. Подразделение Пантанала на подрайоны (по Хамильтону и др. [Hamilton et al., 1996], с любезного разрешения E. Schweizerbart'sche Velagsbuchhandlung, Stuttgart).

[1998], Rocha & Por [1998], Heckman [1998a, 1998b], Santos-Silva et al. [1999], and Bezerra et al. [1999]. Four of these papers, Matsumura-Tundisi [1986], Reid & Moreno [1990], Reid [1997], Santos-Silva et al. [1999], dealt only with copepods; Miranda [1992] and Bezerra et al. [1999] determined the animals only to generic level; Daday [1905], Por [1995] and Rocha & Por [1998] presented in their lists not only Cladocera and Copepoda, but also Conchostraca and Ostracoda.

The Pantanal is not a uniform area. Eberhard [2000] wrote that it is a complex of ecosystems. Adámoli [1982] differentiated several phytogeographical areas, but for our purpose we use the subregions of Hamilton et al. [1996], which are more appropriate for freshwater animals (Fig. 1). A series of papers refers to crustaceans from the following six subregions:

**Cuiabá River (CUIA):** Heckman [1998 a, b];

**Piquiri/São Lourenço Rivers (PIQU):** Matsumura-Tundisi [1986], Lima [1996], Santos-Silva [1999];

**Paraguay River (PARA):** Daday [1905], Reid & Moreno [1990], Por [1995], Espíndola et al. [1996];

**Nhecolândia (NHEC):** Mourão [1989], Reid & Moreno [1990], Por [1995], Reid [1997];

**Aquidauana/Negro Rivers (AQUI) and Miranda River (MIRA):** Miranda [1992], Por [1995], Bonecker et al. [1998], Rocha & Por [1998], Bezerra et al. [1999].

This paper examines original samples from two subregions – the Paraguay River and Nhecolândia – of the Pantanal, summarises previous cladoceran surveys to produce a check-list for the region, and comments on the taxonomic status of some species

## Study Area

Our samples were collected in the Paraguay River (samples 1–3, 7) and Nhecolândia subregions (samples 4–6) by colleagues of the Center of Agriculture Research in the Pantanal (EMBRAPA) (samples 1–2) and by one of the authors (GOB). These samples have already been studied by Koste [1999], who found 216 taxa of Rotifera.

Locality 1 (collected in 1996 and 1999) is Canal do Tamengo near the city of Corumbá, the outflow of the large floodplain lake Lagoa de Cáceres on the Bolivian side of the Paraguay River. The Canal do Tamengo is not a channel with strong currents but the waters flow slowly over a broad area. It has a mixture of waters from the Lagoa de Cáceres and waters from the Paraguay River which upstream leave the river channel and return through the Canal do Tamengo. The ionic content of the water is somewhat higher than that of the Paraguay River due to the release of solutes by weathering of carbonate rocks in the upland watersheds [Hamilton et al., 1999].

Locality 2 is the Paraguay River itself just above Corumbá. Water level fluctuations in 1996 were little more than two meters with high water in June/July. The hydrochemistry of this part of the river “represents a mixture of several upstream tributaries of diverse chemical composition” and “falls in the middle of the range of variation” of the Pantanal waters [Hamilton et al., 1999].

Locality 7 is Baía do Castelo, a floodplain lake on the right (west) side of the Paraguay River about 80 km north of Corumbá. The lake is intensively colonized by submerged and floating aquatic plants and receives water from many drainage channels from the surrounding higher lands. Oliveira & Calheiros [2000] studied the phytoplankton, which is dominated by the nanoplanktonic Cryptophyceae. Chemical data are given by Calheiros & Hamilton [1998], who reported that as the water of the Paraguay River flows over previously dry land there is a stimulation of decomposition rates of and leaching from aquatic materials. The resulting decrease of dissolved  $O_2$  and increase of  $CO_2$  causes fish die. How cladocerans are affected when this happens is not known. The sampling site “rompesaco” lies in the extreme northern part of the lake, which is intensively populated by floating vegetation and receives water through a channel from the Bolivian side.

Localities 4–6 are from the Nhecolândia subregion, which Hamilton et al. [1998] called the “Lake District”. There are numerous lakes called baias, most of which are found at an altitude of less than 100 m [Fernandes et al., 1999]. They lie in fluvial sand deposits, which in former

times were extensive dune fields. The crests of these ancient dunes, called cordilheiras, are covered by forests. The climate is hot with a pronounced rainy season from November to March. Evaporation is high, which results in an annual water deficit [Soriano, 1999]. Here there are many permanent and temporary freshwater ponds or “baías” and some saline lakes or “salinas” [Mourão, 1989; Sakamoto, 1999]. “The most dilute, and the most saline waters of the southern Pantanal are found” in the Nhecolândia subregion [Hamilton et al., 1999]. Mourão [1989] made an intensive study of the chemistry and the phytoplankton of two ponds and a saline lake. The “salina” exhibit an extremely high phytoplankton production but a very low diversity of zooplankton.

### Material and Methods

The material from the Paraguay River was collected by towing a plankton net (58 µm) for five minutes slowly behind a boat. All the other samples were taken by a series of vertical net hauls and by washing out the roots of some floating water plants. All samples were fixed in 5 % formaldehyde solution.

Specimens were isolated from the samples under a stereomicroscope, placed on slides in glycerol-formal-

dehyde mixture, and dissected under a stereoscopic microscope for the study of their appendages and post-abdomen. Drawings were prepared using a camera lucida attached to an Alphaphot compound microscope; optical photos were taken using Leitz-Dialux microscope with a system camera and interference optics. A few specimens of *Grimaldina brazzai* and *Onchobunops tuberculatus* were lyophilised, mounted on an aluminium stub, coated with gold, and examined under a scanning electron microscope (JEOL-840A).

### Results

#### General comments

Fifty species belonging to the orders Ctenopoda Sars, 1865 (1 family) and Anomopoda Sars, 1865 (6 families) were identified in our samples (Tab. 1). The greatest number of species (30) was found in the Baía do Castelo rompesaco, followed by Canal do Tamengo (19), Baía 38 (19) and Paraguay River (18). The most abundant species in the baías were: *Diaphanosoma brevireme*, *Ceriodaphnia cornuta*, *Moina micrura*, *M. reticulata*, *Alonella dadayi*, *Chydorus ventricosus*, *Euryalona orientalis*, and *Notoalona globulosa*. A few specimens could not be

Table 1. Distribution of cladoceran species in water bodies of the southern Pantanal. Highest abundance of the species in a sample: • (1–3 specimens) — single, •• (4–10) — few, ••• (11–25) — several, •••• (26–100) — many, ••••• (more than 100) — masses. Abbreviations: eph — ephippium, pa — postabdomen.

Таблица 1. Распространение ветвистоусых ракообразных в водоемах южного Пантанала. Обилие видов в пробе: • (1–3 экземпляра) — единично, •• (4–10) — немного, ••• (11–25) — среднее, •••• (26–100) — много, ••••• (более 100) — массовый вид. Условные обозначения: eph — эфиппиум, pa — постабдомен.

Species/ Localities:	1	2	3	4	5	6	7
	Canal do Tamengo, 28.3.1996, 29.4.1998	River Paraguay, 2–12.1996	Baía 38, 1.10.1997	Baía 85 Salina, 1.10.1997	Baía 69, 1.10.1997	Baía 75, 1.10.1997	Baía do Castelo rompesaco, 23.4.1998
ORDER CTENOPODA Sars, 1865							
<i>Sididae</i> Baird, 1850							
<i>Diaphanosoma brevireme</i> Kofínek, 1981		•••••	••				•••••
<i>Diaphanosoma spinulosum</i> Herbst 1975	•						
<i>Sarsilatona serricauda</i> Sars, 1901			•		•		•••
ORDER ANOMOPODA Sars, 1865							
<i>Daphniidae</i> Straus, 1820							
<i>Ceriodaphnia cornuta</i> Sars, 1885		•••••	•••••		eph		•••
<i>Ceriodaphnia reticulata</i> (Jurine, 1820)		•••••					
<i>Ceriodaphnia silvestrii</i> Daday, 1902						•	•
<i>Daphnia gessneri</i> Herbst, 1967		•••••	••			••	
<i>Simocephalus acutirostratus</i> (King, 1853)							•••
<i>Simocephalus latirostris</i> Stingelin, 1906		•••					•

Table 1 (continuing).  
Таблица 1 (продолжение).

Species/ Localities:	1	2	3	4	5	6	7
	Canal do Tamengo, 28.3.1996, 29.4.1998	River Paraguay, 2–12 1996	Baia 38, 1.10.1997	Baia 85 Salina, 1.10.1997	Baia 69, 1.10.1997	Baia 75, 1.10.1997	Baia do Castelo rompesaco, 23.4.1998
ORDER CTENOPODA Sars, 1865							
<u>Moinidae</u> Goulden, 1967							
<i>Moina micrura</i> Kurz, 1875	•		••••				••
<i>Moina minuta</i> Hansen, 1899		•••••			•		
<i>Moina reticulata</i> (Daday, 1905)		••	••				••••
<i>Moinodaphnia macleayi</i> (King, 1853)		••					
<u>Ilyocryptidae</u> Smirnov, 1976 sensu Smimov, 1992							
<i>Ilyocryptus sarsi</i> Stingelin, 1913						•	
<i>Ilyocryptus spinifer</i> Herrick, 1882	•	••••	•				
<u>Macrothricidae</u> Norman & Brady, 1867							
<i>Grimaldina brazzai</i> Richard, 1882		••					••
<i>Guernella raphaelis</i> Richard, 1882	•						
<i>Macrothrix elegans</i> Sars, 1901	•	••	•				••
<i>Macrothrix spinosa</i> King, 1853				••			•••
<i>Onchobunops tuberculatus</i> Fryer & Paggi, 1972							•
<u>Bosminidae</u> Sars, 1865							
<i>Bosmina hagmani</i> Stingelin, 1904		•••••					
<i>Bosmina tubicen</i> Brehm, 1953	•	••	•••		••	•	•
<i>Bosminopsis deitersi</i> Richard, 1895	•	•••••	•		•		•
<u>Chydoridae</u> Stebbing, 1902							
<i>Alona brasiliensis</i> (Bergamin, 1935)	••						••
<i>Alona guttata</i> Sars, 1862							•
<i>Alona iheringi</i> Sars, 1901	•						
<i>Alona monacantha</i> Sars, 1901			•				•
<i>Alona ossiani</i> Sinev, 1998			•		shell, pa	shell, pa	
<i>Alona rectangula</i> Sars, 1862						••	
<i>Alona verrucosa</i> Sars, 1901			•		shell		•
<i>Alonella dadayi</i> Birge, 1910		•	••••				
<i>Camptocercus dadayi</i> Stingelin, 1913							•
<i>Chydorus eurymotus</i> Sars, 1901		•					
<i>Chydorus nitidulus</i> (Sars, 1901)							••
<i>Chydorus pubescens</i> Sars, 1901			shell				
<i>Chydorus ventricosus</i> Daday, 1898	••						••••
<i>Chydorus</i> sp.			•	shell	••	shell	•
<i>Dunhevedia odontoplax</i> Sars, 1901	•	•					•
<i>Ephemeroporus hybridus</i> (Daday, 1905)			•		shell		••



Table 1 (continuing).  
Таблица 1 (продолжение).

Species/ Localities:	1	2	3	4	5	6	7
		Canal do Tamengo, 28.3.1996, 29.4.1998	River Paraguay, 2–12 1996	Baia 38, 1.10.1997	Baia 85 Salina, 1.10.1997	Baia 69, 1.10.1997	Baia 75, 1.10.1997
ORDER CTENOPODA Sars, 1865							
<i>Chydoridae</i> Stebbing, 1902							
<i>Ephemeropterus tridentatus</i> (Bergamin, 1939)						•	
<i>Euryalona orientalis</i> (Daday, 1898)	••		•				••••
<i>Karualona muelleri</i> (Richard, 1897)	••				•		••
<i>Kurzia polyspina</i> Hudec, 2000	•						••
<i>Leydigia</i> cf. <i>striata</i> Biraben, 1939		•					
<i>Leydigiopsis ornata</i> Daday, 1905	•		•				•
<i>Notoalona globulosa</i> (Daday, 1898)	••				•		••••
<i>Oxyurella longicaudis</i> Birge, 1910	••						
<i>Oxyurella</i> sp.	•						
<i>Pleuroxus</i> sp.						•	
<i>Pseudochydorus globosus</i> (Baird, 1848)	•						
NUMBER OF TAXA	19	18	19	2	11	9	29

identified as they consisted of solitary individuals or shells. In the samples from Paraguay River pelagic *Daphnia*, *Ceriodaphnia*, *Moina* and *Bosmina* were dominant. The majority of species we found (39) are typical representatives of the tropical fauna (see Figs. 2, 5–6 for some optical photos). A significant number (27) is Neotropical,

penetrating only the southern portion of Nearctic zone (Figs. 3–4, 7–9). About half of the species collected are new records for Pantanal (Tab. 2). To date the check-list of species from Pantanal, according to our and previously published data, consists of 68 taxa determined to species level, and a further 19 taxa to genera; however, some

Table 2. Occurrence of cladoceran species in the different subregions of the Pantanal (according to our and literature data), see comments below.

Таблица 2. Встречаемость ветвистоусых ракообразных в различных районах Пантанала (по нашим и литературным данным), см. комментарии ниже.

Species	PARA This paper	NHEC This paper	PARA Daday [1905], Espindola et al. [1996]	NHEC Mourao [1989]	AQUI Bonecker et al. [1998], Miranda [1992], Bezerra et al. [1999]	MIRA Bonecker et al. [1998], Miranda [1992], Rocha & Por [1998]	PIQU Lima [1996]	CUJA Heckman [1998a, b], Rocha & Por [1998]
CTENOPODA								
<i>Sididae</i>								
<i>Diaphanosoma birgei</i>			x				x	X
<i>Diaphanosoma brevirene</i>	x	x	x					
<i>Diaphanosoma spinulosum</i>	x							
<i>Diaphanosoma</i> sp.				x	x	x		

Table 2 (continuing).  
Таблица 2 (продолжение).

Species	PARA This paper	NHEC This paper	PARA Daday [1905], Espindola et al. [1996]	NHEC Mourao [1989]	AQUI Bonecker et al. [1998], Miranda [1992], Bezerra et al. [1999]	MIRA Bonecker et al. [1998], Miranda [1992], Rocha & Por [1998]	PIQU Lima [1996]	CUIA Heckman [1998a, b], Rocha & Por [1998]
CTENOPODA								
<u>Sididae</u>								
<i>Latonopsis fasciculata</i>			x					
<i>Pseudosida</i> sp.				x				
<i>Sarsilatona serricaudata</i>	x	x						
<i>Sida crystallina</i> (1)								X
ANOMOPODA								
<u>Daphniidae</u>								
<i>Ceriodaphnia cornuta</i>	x	x	x	x	x	x	X	X
<i>Ceriodaphnia reticulata</i>	x							
<i>Ceriodaphnia rigaudi</i>			x					
<i>Ceriodaphnia silvestrii</i> (2)	x	x						
<i>Ceriodaphnia</i> sp.					x	x		
<i>Daphnia gessneri</i>	x	x						
<i>Daphnia</i> sp.				x				
<i>Megafenestra aurita</i> (2)								X
<i>Scapholeberis mucronata</i> (2)			x					
<i>Scapholeberis</i> sp.						x		
<i>Simocephalus acutirostratus</i>	x							
<i>Simocephalus latirostris</i>	x							
<i>Simocephalus serrulatus</i>			x					X
<i>Simocephalus</i> sp.						x		
<u>Moinidae</u>								
<i>Moina micrura</i> (3)	x	x	x					X
<i>Moina minuta</i>	x	x	x	x				
<i>Moina reticulata</i>	x	x						
<i>Moina</i> sp.					x	x		
<i>Moinodaphnia macleayi</i>	x		x		x	x		
<i>Moinodaphnia</i> sp.						x		
<u>Ilyocryptidae</u>								
<i>Ilyocryptus sarsi</i>		x						
<i>Ilyocryptus sordidus</i> (2)						x		X
<i>Ilyocryptus spinifer</i>	x	x		x	x	x		X
<i>Ilyocryptus</i> sp.						x		
<u>Macrothricidae</u>								
<i>Grimaldina brazzai</i>	x							

Table 2 (continuing).  
Таблица 2 (продолжение).

Species	PARA This paper	NHEC This paper	PARA Daday [1905], Espindola et al. [1996]	NHEC Mourao [1989]	AQUI Bonecker et al. [1998], Miranda [1992], Bezerra et al. [1999]	MIRA Bonecker et al. [1998], Miranda [1992], Rocha & Por [1998]	PIQU Lima [1996]	CUJA Heckman [1998a, b], Rocha & Por [1998]
ANOMOPODA								
<u>Macrothricidae</u>								
<i>Guernella raphaelis</i>	x							
<i>Macrothrix elegans</i> (4)	x	x	x					
<i>Macrothrix latcornis</i> (2)			x				X	
<i>Macrothrix spinosa</i>	x	x			x	x		
<i>Macrothrix</i> sp.						x		
<i>Onchobunops tuberculatus</i>	x							
<u>Bosminidae</u>								
<i>Bosmina hagmanni</i>	x		x		x	x	x	
<i>Bosmina longirostris</i> (2)				x				x
<i>Bosmina macrostyla</i> (2)			x					
<i>Bosmina tenuirostris</i> (2)			x					
<i>Bosmina tubicen</i>	x	x					x	
<i>Bosmina</i> sp.					x	x		
<i>Bosminopsis deitersi</i> (5)	x	x	x					
<i>Bosminopsis</i> sp.				x	x	x		
<u>Chydoridae</u>								
<i>Acroperus</i> sp.						x		
<i>Alona brasiliensis</i> (6)	x							
<i>Alona costata</i>								x
<i>Alona dentifera</i> (6)			x					
<i>Alona guttata</i>	x						x	
<i>Alona iheringi</i> (7)	x							
<i>Alona intermedia</i>			x					
<i>Alona monacantha</i>	x	x	x				x	
<i>Alona ossiani</i>		x						
<i>Alona rectangula</i> -type		x						
<i>Alona verrucosa</i>	x	x						
<i>Alona</i> sp.				x	x	x		
<i>Alonella clathratula</i>			x					
<i>Alonella dadayi</i> (8)	x	x	x			x	x	
<i>Alonella</i> sp.					x	x		
<i>Camptocercus dadayi</i> (9)	x					x		
<i>Chydorus eurynotus</i>	x							
<i>Chydorus nitidulus</i>	x							

Table 2 (continuing).  
Таблица 2 (продолжение).

Species	PARA This paper	NHEC This paper	PARA Daday [1905], Espindola et al. [1996]	NHEC Mourao [1989]	AQUI Bonecker et al. [1998], Miranda [1992], Bezerra et al. [1999]	MIRA Bonecker et al. [1998], Miranda [1992], Rocha & Por [1998]	PIQU Lima [1996]	CUJA Heckman [1998a, b], Rocha & Por [1998]
ANOMOPODA								
<i>Chydoridae</i>								
<i>Chydorus pubescens</i>		x						
<i>Chydorus sphaericus</i>			x					
<i>Chydorus ventricosus</i>	x					x		
<i>Chydorus</i> sp.	x	x		x	x	x		x
<i>Dunhevedia odontoplax</i>	x					x		
<i>Ephemeroporus hybridus</i>	x	x						
<i>Ephemeroporus tridentatus</i>		x						
<i>Euryalona orientalis</i>	x	x				x		
<i>Karualona karua</i> (10)						x		
<i>Karualona muelleri</i> (10)	x	x						
<i>Kurzia polyspina</i>	x							
<i>Leptorhynchus dentifer</i> (2)			x					
<i>Leydigia striata</i>	x							
<i>Leydigiopsis curvirostris</i>						x		
<i>Leydigiopsis</i> cf. <i>megalops</i>						x		
<i>Leydigiopsis ornata</i>	x	x				x		
<i>Leydigiopsis</i> sp.						x		
<i>Notoalona globulosa</i> (11)	x	x				x		
<i>Oxyurella longicaudis</i>	x					x		
<i>Oxyurella</i> sp.	x					x		
<i>Pleuroxus scopuliferus</i>			x					
<i>Pleuroxus</i> sp.		x				x		
<i>Pseudochydorus globosus</i>	x							
ONYCHOPODA								
<i>Polyphemidae</i>								
<i>Polyphemus</i> sp. (1)						x		
Number of taxa	44	28	24	10	13	35	8	11

1) very dubious record;

2) record must be checked;

3) including *Moina micrura ciliata* Daday, 1905, which is regarded as a subspecies of *M. micrura* Kurz, 1875, but this must be checked;4) including records of *M. superaculeata* and *M. triserialis*, which are apparently misidentifications of *M. elegans*;5) *Bosminella anisitsi* Daday, 1903 is a jun. syn. of *Bosminopsis deitersi* Richard, 1895;6) recently transferred to the genus *Alona*, previously these were determined as *Alonella*;

7) see text;

8) *Phryxura dadayi* = *Alonella dadayi*;9) Apparently determinations of *Camptocercus australis* were dealing with *C. dadayi*;10) Recently separated into genus *Karualona* Dumont & Silva-Briano, 2000, previously these were determined as *Alona*, *Biapertura* or *Alonella*;11) Earlier it was placed into *Indialona* or *Alonella*.



Table 3. Occurrence of cladoceran species in the Pantanal and Paraná (Brazil). See comments to species below table 2. Distribution: C — cosmopolitan, N — Neotropical, penetrating southern portion of Nearctic zone, H — Holarctic, sometimes penetrating other zones, T — tropicopolitan, U — unknown.

Таблица 3. Встречаемость ветвистоусых ракообразных в Пантанале и Паране (Бразилия), см. также комментарии к Таб. 2. Распространение: С — космополитическое, N — неотропическое с проникновением в южную часть неарктической зоны, H — голарктическое, изредка с проникновением в другие зоны, T — циркумтропическое, U — неясно.

Species	Distribution	Pantanal (this paper)	Pantanal (previous papers, in total)	Parana [Lansac-Toha et al., 1997]
<b>CTENOPODA</b>				
<u>Sididae</u>				
<i>Diaphanosoma birgei</i>	N		x	x
<i>Diaphanosoma brevirene</i>	N	x	x	x
<i>Diaphanosoma fluviatile</i>	N			x
<i>Diaphanosoma spinulosum</i>	N	x		x
<i>Diaphanosoma</i> sp.			x	
<i>Latonopsis fasciculata</i>	N		x	
<i>Pseudosida</i> sp.			x	
<i>Sarsilatona serricauda</i>	N	x		x
<i>Sida crystallina</i>	H		x	
<b>ANOMOPODA</b>				
<u>Daphniidae</u>				
<i>Ceriodaphnia cornuta</i>	C	x	x	x
<i>Ceriodaphnia reticulata</i>	C	x		
<i>Ceriodaphnia rigaudi</i>	T		x	
<i>Ceriodaphnia silvestrii</i>	N	x		
<i>Ceriodaphnia</i> sp.			x	
<i>Daphnia gessneri</i>	N	x		x
<i>Daphnia</i> sp.			x	
<i>Megafenestra aurita</i>	P		x	
<i>Scapholeberis mucronata</i>	P		x	
<i>Scapholeberis</i> sp.			x	x
<i>Simocephalus acutirostratus</i>	T	x		
<i>Simocephalus latirostris</i>	N	x		
<i>Simocephalus serrulatus</i>	C		x	x
<i>Simocephalus vetulus</i>	C			x
<i>Simocephalus</i> sp.			x	
<u>Moinidae</u>				
<i>Moina micrura</i>	C	x	x	
<i>Moina minuta</i>	N	x	x	x
<i>Moina reticulata</i>	N	x		
<i>Moina</i> sp.			x	
<i>Moinodaphnia macleayi</i>	T	x	x	
<i>Moinodaphnia</i> sp.			x	
<u>Ilyocryptidae</u>				
<i>Ilyocryptus sarsi</i>	N	x		
<i>Ilyocryptus sordidus</i>	C		x	

Table 3 (continuing).  
Таблица 3 (продолжение).

Species	Distribution	Pantanal (this paper)	Pantanal (previous papers, in total)	Parana [Lansac-Toha et al., 1997]
ANOMOPODA				
<u>Ilyocryptidae</u>				
<i>Ilyocryptus spinifer</i>	T	x	x	x
<i>Ilyocryptus</i> sp.			x	
<u>Macrothricidae</u>				
<i>Grimaldina brazzai</i>	T	x		x
<i>Guernella raphaelis</i>	T	x		
<i>Macrothrix elegans</i>	N	x	x	x
<i>Macrothrix laticornis</i>	C		x	
<i>Macrothrix spinosa</i>	T	x	x	x
<i>Macrothrix</i> sp.			x	
<i>Onchobunops tuberculatus</i>	N	x		x
<u>Bosminidae</u>				
<i>Bosmina hagmanni</i>	N	x	x	x
<i>Bosmina longirostris</i>	C		x	
<i>Bosmina macrostyla</i>	N		x	
<i>Bosmina tenuirostris</i>	N		x	
<i>Bosmina tubicen</i>	N	x	x	x
<i>Bosmina</i> sp.			x	
<i>Bosminopsis deitersi</i>	C	x	x	x
<i>Bosminopsis</i> sp.			x	
<u>Chydoridae</u>				
<i>Acroperus</i> sp.			x	
<i>Acroperus harpae</i>	C			x
<i>Alona affinis</i>	C			x
<i>Alona brasiliensis</i>	N	x		
<i>Alona costata</i>	C		x	
<i>Alona dentifera</i>	N		x	x
<i>Alona</i> cf. <i>glabra</i>	U			x
<i>Alona guttata</i>	C	x	x	
<i>Alona iheringi</i>	N	x		
<i>Alona intermedia</i>	C		x	
<i>Alona monacantha</i>	U	x	x	x
<i>Alona ossiani</i>	N	x		
<i>Alona rectangula</i> -type	C	x		
<i>Alona verrucosa</i>	T	x		
<i>Alona</i> sp.			x	x
<i>Alonella clathratula</i>	T		x	
<i>Alonella dadayi</i>	N	x	x	x
<i>Alonella</i> sp.			x	
<i>Camptocercus dadayi</i>	N	x	x	x

Table 3 (continuing).  
Таблица 3 (продолжение).

Species	Distribution	Pantanal (this paper)	Pantanal (previous papers, in total)	Parana [Lansac-Toha et al., 1997]
ANOMOPODA				
<u>Chydoridae</u>				
<i>Chydorus eurynotus</i>	T	x		x
<i>Chydorus nitidulus</i>	N	x		
<i>Chydorus pubescens</i>	T	x		x
<i>Chydorus sphaericus</i>	C		x	
<i>Chydorus ventricosus</i>	T	x	x	
<i>Chydorus</i> sp.		x	x	
<i>Dunhevedia odontoplax</i>	N	x	x	x
<i>Ephemeroporus hybridus</i>	N	x		
<i>Ephemeroporus tridentatus</i>	N	x		
<i>Euryalona orientalis</i>	T	x	x	x
<i>Graptoleberis testudinaria</i>	C			x
<i>Karualona karua</i>	T		x	x
<i>Karualona muelleri</i>	N	x		
<i>Kurzia latissima</i>	C			x
<i>Kurzia polyspina</i>	N	x		
<i>Leptorhynchus dentifer</i>	N		x	
<i>Leydigia striata</i>	N	x		
<i>Leydigia</i> sp.				x
<i>Leydigiopsis brevirostris</i>	N			x
<i>Leydigiopsis curvirostris</i>	N		x	x
<i>Leydigiopsis</i> cf. <i>megalops</i>	N		x	
<i>Leydigiopsis ornata</i>	N	x	x	
<i>Leydigiopsis</i> sp.			x	
<i>Niscirmirovius eximius</i>	T			x
<i>Notoalona globulosa</i>	T	x	x	x
<i>Oxyurella</i> cf. <i>ciliata</i>	N			x
<i>Oxyurella longicaudis</i>	N	x	x	
<i>Oxyurella</i> sp.		x	x	
<i>Pleuroxus scopuliferus</i>	N		x	
<i>Pleuroxus</i> sp.		x	x	
<i>Pseudochydorus globosus</i>	C	x		
ONYCHOPODA				
<u>Polyphemidae</u>				
<i>Polyphemus</i> sp.	H		x	
Number of taxa		50	63	40

records are dubious while others must be confirmed by further investigations. Among the 40 species from the Paraná [Lansac-Tôha et al., 1997] there are 7 species that have not been found in the Pantanal (Tab. 3). We are sure

that some of them, for example *Niscirmirovius eximius*, *Graptoleberis testudinaria* and *Oxyurella ciliata*, will eventually be found there, whereas *Alona affinis* is, most probably, a misidentification of *A. ossiani*.

## Comments on selected species

A. Family ILYOCRYPTIDAE Smirnov, 1976 emend. Smirnov, 1992

### 1. *Ilyocryptus sarsi* Stingelin, 1913

COMMENTS: This species, which has recently been redescribed by Kotov et al. [2002: 208–219, Figs. 1–60] from material from the Pantanal (Baia 75), is one of the two most common *Ilyocryptus* species recorded from Brazil. It is unlike the European *I. sordidus* (Liévin, 1848) and most of the Brazilian records of “*I. sordidus*” (see Table 2 and 3) seem to be dealing with *I. sarsi* (also see Kotov et al. [2002]). Daday [1905] recorded *I. sordidus* in his Paraguayan samples but this is in fact *I. paranaensis* Paggi, 1989 (P. Štifter, personal communication based on study of Daday’s original material). We did not find *I. paranaensis* in our material.

### 2. *Ilyocryptus spinifer* Herrick, 1882

COMMENTS: Kotov & Williams [2000: 68–79, Figs. 1–91] selected a neotype and redescribed *I. spinifer* s. str. from the North American continent. Kotov & Dumont [2000: 88–100, Figs. 1–149] analyzed many populations from different countries (see intensive synonymy in these publications) including material from Pantanal (Baia 38 and Baia do Castelo). This is one of the most common anomopods in tropical water bodies of different types. It is well-differentiated from other species.

B. Family MACROTHRICIDAE Norman & Brady, 1867 emend. Smirnov, 1992

### 3. *Grimaldina brazzai* Richard, 1892 Figs. 10–40.

Sars, 1901: 28–31, Pl. 5; Daday, 1905: 192–193; Martínez de Ferrato, 1966: 403, Pl. 2: Fig. 5; Fryer, 1974: 236–238, Figs. 128–129; Smirnov, 1976: 155–156, Figs. 137–140; Brandorff et al., 1982: 103, Fig. 79; Smirnov, 1992: 107–109, Figs. 461–468; Paggi, 1995: Figs. 87–88; Elmoor-Loureiro, 1998: 24; Silva-Briano, 1998: 149–151, Figs. 1–10.

Type locality: “Mayoumba et Caca Mueca”, Congo, Africa

Parthenogenetic female: Body subquadrangular to ovoid, relatively high and significantly compressed laterally. Border between head and valves as a very shallow depression, dorsal margin of valves straight or even concave, postero-dorsal angle distinct. Dorsal margin finely serrated; this serration is connected with specific reticulations and looks like fish scales. On lateral surface of head and valves reticulation appears as a system of dorso-ventral striae. Head triangular-rounded, with short rostrum; ventral margin generally straight, with a small outgrowth like a transversal plate posterior to antenna I, a minute frontal head pore near distal extremity of rostrum, dorsal head pore completely absent. Labrum in general subquadrangular, with high, setulated projection at antero-ventral angle, naked projection at postero-ventral angle, and large, setulated distal labral plate. Compound eye small, ocellus somewhat smaller than eye, located near rostrum. Valves large, with numerous marginal setae; in anterior half of ventral margin these setae are thicker and shorter, while in posterior half longer and thinner. Postero-ventral region with series of short, setulated spines, a solitary seta located submarginally on inner surface dorsally to the series of spines. Posterior margin with fine setules. Postabdomen

large, significantly compressed laterally, with longitudinal keel along entire preanal margin. Preanal margin with fine spines, subdivided into two lobes, a bunch of fine setules near the boundary between the lobes; postanal and anal margins with series of plumose setae, a solitary seta at posterior edge of anus. Postabdominal setae longer than postabdomen, with distal segment half the length of the basal segment. Postabdominal claw relatively robust, with a single basal spine, two rows of setules along dorsal margin and a series of fine setules along ventral margin. Antenna I long, thin, “rod-like” in terms of Smirnov [1992], with a sensory seta arising at distance  $\frac{1}{4}$  of its length from base, setules along its posterior margin, a row of robust denticles around its distal end, and 9 aesthetascs of unequal size. Antenna II long, two sensory setae of similar size at coxal part, basal segment cylindrical, all branch segments elongated, with concentric rows of small denticles. Antennal formula: setae 0-0-1-3/1-1-3, spines 0-1-0/0-0-1. Swimming setae not too long (shorter than basal segment plus rami), the largest seta (basal lateral swimming seta of endopod) asymmetrically armed with short setules distally, all other lateral and apical setae with bilaterally setulated basal segments, and asymmetrically armed distal segments, chitinous insertions within each distal segment near joint with basal segment. All spines thin, about three times shorter than segments from which they arise. Limb I with ODL bearing a single, large seta, IDL with three setae, and gnathobase I with two setae of unequal size. Other limbs as described by Silva-Briano [1998]. Size 590–670  $\mu\text{m}$  in our material, up to 820  $\mu\text{m}$  according to Sars [1901].

Ephippial female and male: See Sars [1901].

COMMENTS: This taxon seems to be tropicopolitan, but must be revised. The most detailed revision of African and South American populations by Silva-Briano [1998] did not reveal any significant differences in their morphology, but only a very limited number of parthenogenetic females was studied.

### 4. *Macrothrix elegans* Sars, 1901

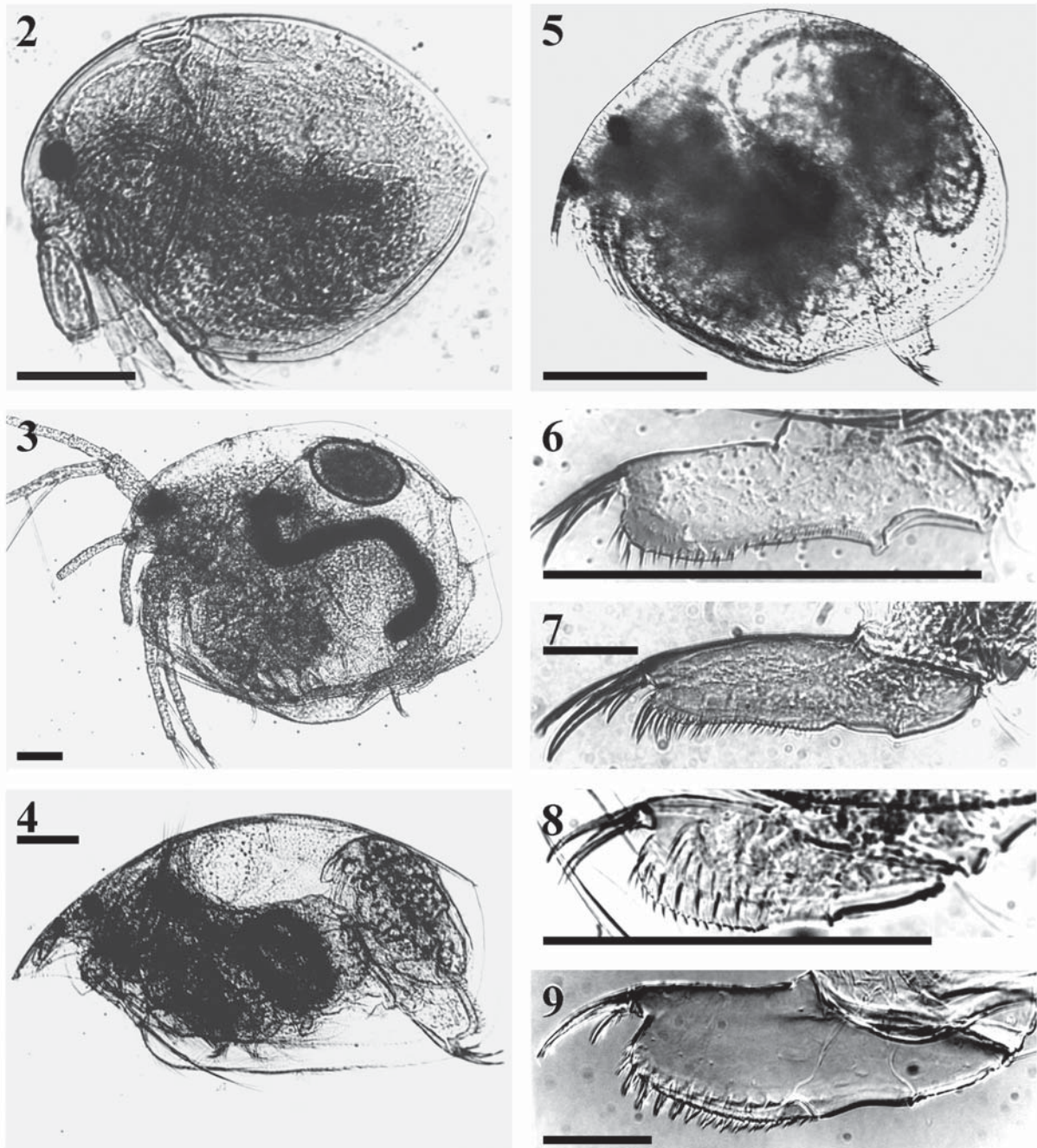
COMMENTS: This species, described from Brazil, was for a long time confused with Afro-Asiatic *M. triserialis* Brady, 1886. Dumont et al. [2002] pointed out intercontinental differences of members from the *triserialis*-group, but decided that *M. superaculeata* Smirnov, 1982 is a junior synonym of *M. elegans*. More recently Kotov et al. [2003a] studied many Neotropical populations of *M. elegans* (see intensive synonymy here), including material from Baia do Castelo, and concluded that *M. superaculeata* is a valid, but relatively rare species. We did not find it in our Pantanal samples, but it will probably be found in the future. All previous determinations of *M. superaculeata* must be re-examined, because Smirnov’s [1992] key, which determines both species by the presence or absence of large additional spines on segments of the 4-segmented antennal branch, is not correct, as both species have these spines.

### 5. *Macrothrix spinosa* King, 1853 Figs. 41–55.

Richard, 1897: 286–287 (*laticornis*); Sars, 1901: 36–37; Pl. 6: Figs. 10–12 (*squamosa*); Daday, 1905: 194–195 (*laticornis*); Harding, 1955: 339, Figs. 40–44 (*laticornis*); Smirnov, 1976: 72–79, Figs. 40–47; Smirnov, 1992: 29–35, Figs. 60–93; Elmoor-Loureiro, 1998: 25; Silva-Briano, 1998: 346–348, Figs. 1–45.

Type locality: “A pond between Sydney and Liverpool, now destroyed by urbanization” [Smirnov, 1992], New South Wales, Australia.



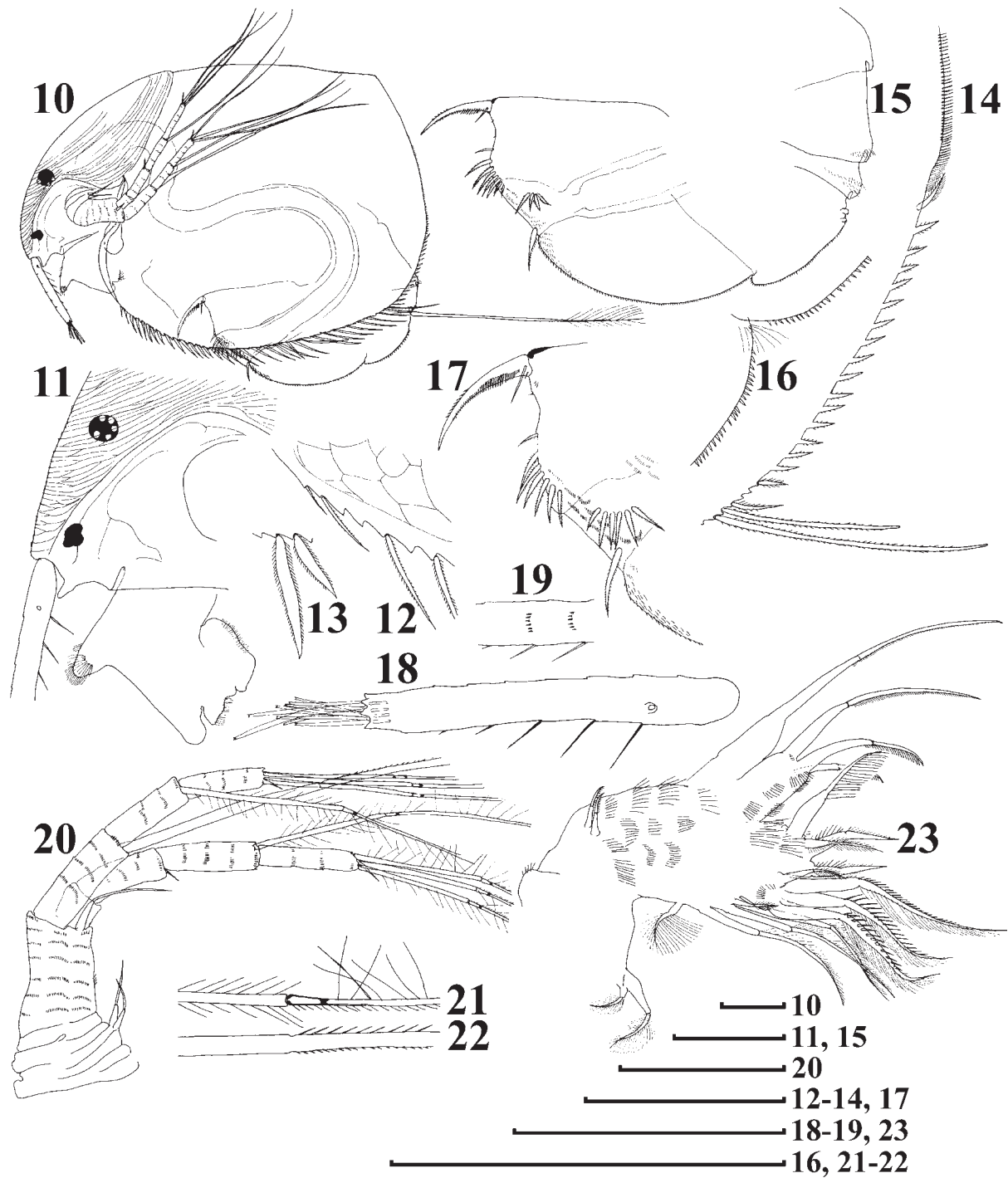


Figs. 2–9. Selected anomopods from Pantanal, Mato Grosso do Sul, Brazil: 2 — *Guernella raphaelis* from Canal do Tamengo; 3 — *Onchobunops tuberculatus* from Baia do Castelo; 4 — *Alona ossiani* from Baia 38 in Nhecolândia; 5 — *Chydorus ventricosus* from Baia do Castelo; 6 — its postabdomen; 7 — postabdomen of *Oxyurella longicaudis* from Canal do Tamengo; 8 — *Karualona muelleri* from Canal do Tamengo; 9 — postabdomen of *Alona ossiani* from Baia 38. Scales: 100  $\mu$ m.

Рис. 2–9. Некоторые представители Аномопода из Пантанала, Мато Гроссо ду Сул, Бразилия. 2 — *Guernella raphaelis* из Canal do Tamengo; 3 — *Onchobunops tuberculatus* из Baia do Castelo; 4 — *Alona ossiani* из Baia 38; 5 — *Chydorus ventricosus* из Baia do Castelo; 6 — его постабдомен; 7 — постабдомен *Oxyurella longicaudis* из Canal do Tamengo; 8 — *Karualona muelleri* из Canal do Tamengo; 9 — постабдомен *Alona ossiani* из Baia 38. Масштаб: 100  $\mu$ m.

Parthenogenetic female: Body ovoid in lateral view, somewhat compressed laterally, with well-developed dorsal keel. Dorsal margin finely serrated along entire length, regularly curved from tip of rostrum to rounded postero-dorsal angle. Reticulation well-expressed on head, valves, and postabdomen. Head relatively large, with relatively small, ovoid dorsal

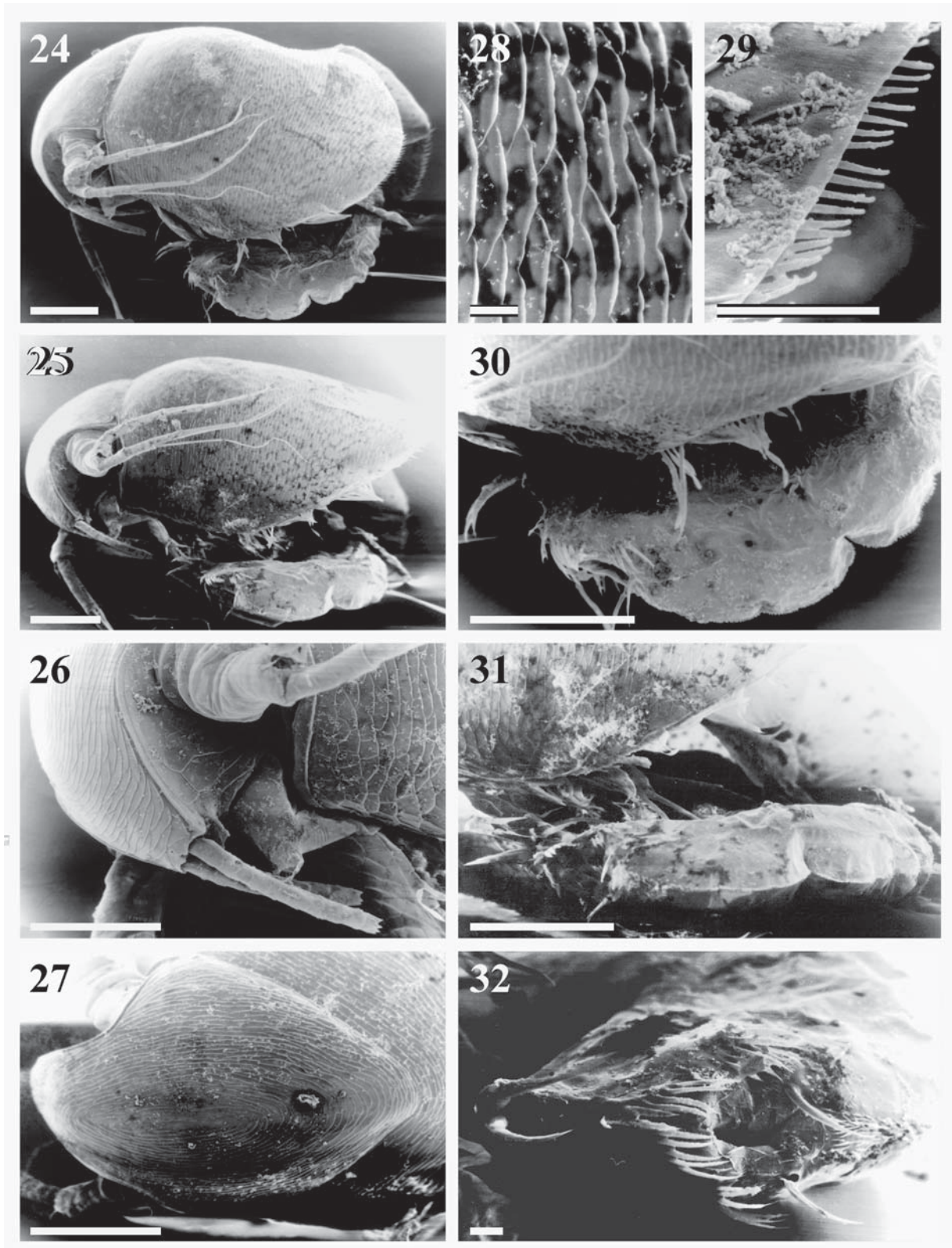
head pore; in lateral view head triangular-round, with short rostrum, minute frontal head pore near its tip, ventral margin of head concave. Eye large, ocellus small, located closer to tip of rostrum than eye. Labrum subquadrangular, with a small projection at postero-ventral angle and relatively large distal labral plate. Valves ovoid, with serrated free margin due to



Figs. 10–23. *Grimaldina brazzai*, parthenogenetic female from Baia do Castelo: 10–11 — adult in lateral view and its head; 12–13 — setae at anterior portion and middle of ventral margin; 14 — postero-ventral portion of valve; 15–17 — postabdomen, its preanal margin and distal portion; 18–19 — antenna I and its medial portion; 20 — antenna II; 21–22 — its apical and largest (basal) lateral swimming seta; 23 — thoracic limb I. Scales: 100  $\mu$ m.

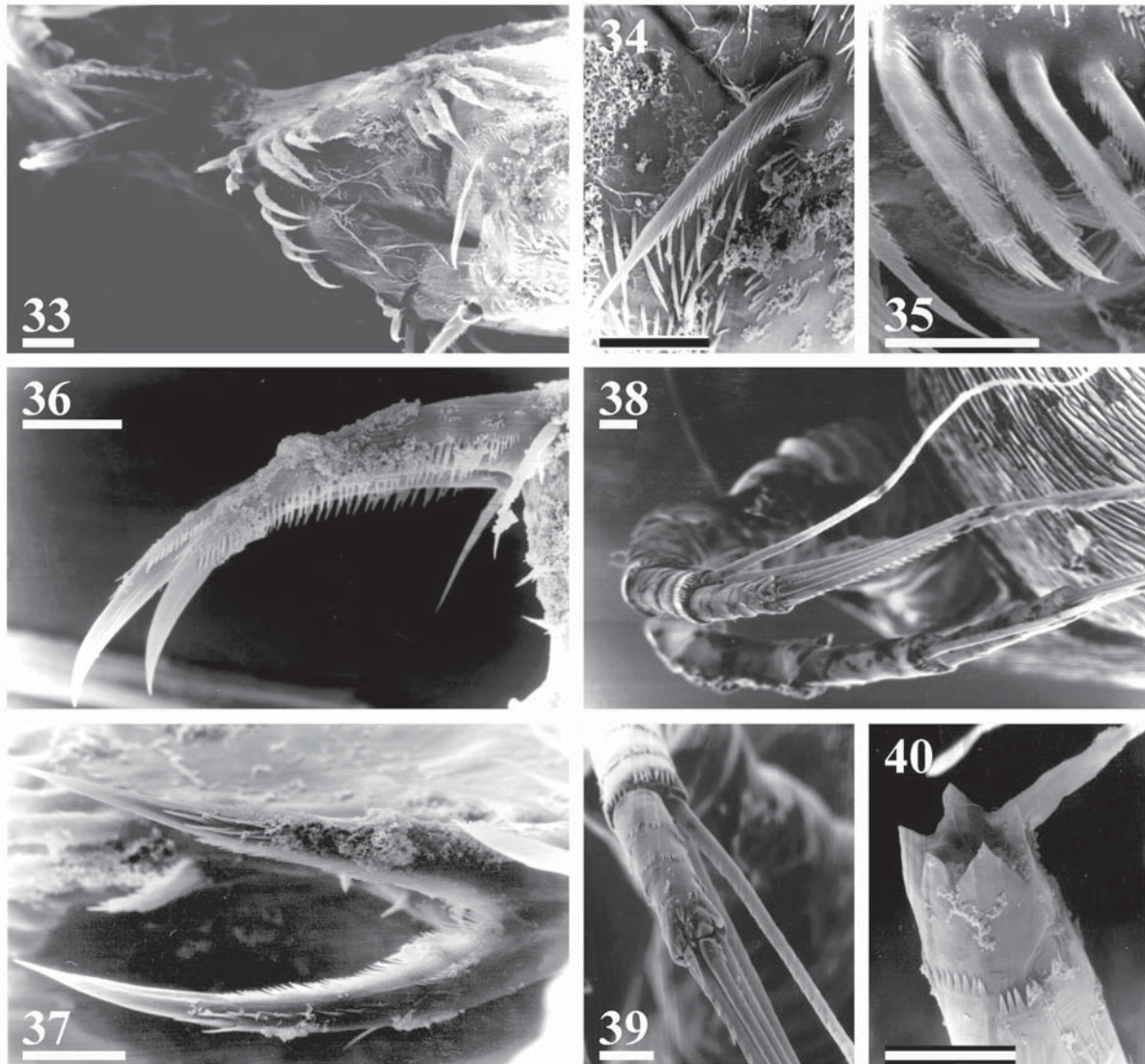
Рис. 10–23. *Grimaldina brazzai*, парthenогенетическая самка из Baia do Castelo: 10–11 — вид сбоку и голова; 12–13 — щетинки в передней части и середине брюшного края; 14 — задне-брюшная часть створки; 15–17 — постабдомен, его преанальный край и дистальная часть; 18–19 — антенна I и ее средняя часть; 20 — антенна II; 21–22 — ее апикальная и наибольшая латеральная плавательная щетинка; 23 — нога I. Масштаб: 100  $\mu$ m.





Figs. 24–32. *Grimaldina brazzai*, parthenogenetic female from Baia do Castelo: 24–25 — adult in lateral and latero-ventral view; 26–27 — head in latero-ventral and dorsal view; 28 — reticulation of valves; 29 — armature of posterior margin of valve; 30–32 — postabdomen in lateral and dorsal view, and its distal portion in distal view. Scales: 100  $\mu\text{m}$  for 24–27, 30–31; 10  $\mu\text{m}$  for 28–29, 32.

Рис. 24–32. *Grimaldina brazzai*, партеногенетическая самка из Baia do Castelo: 24–25 — вид сбоку и латеро-вентрально; 26–27 — голова латеро-вентрально и со спины; 28 — скульптура створки; 29 — вооружение заднего края створки; 30–32 — постабдомен, вид сбоку и со спины, и его дистальная часть. Масштаб: 100  $\mu\text{m}$  для 24–27, 30–31; 10  $\mu\text{m}$  для 28–29, 32.



Figs. 33–40. *Grimaldina brazzai*, parthenogenetic female from Baia do Castelo: 33–35 — distal portion of postabdomen in dorsal view, and its setae; 36–37 — postabdominal claws in lateral and ventral view; 38–39 — antenna II and its distal segment; 40 — distal portion of antenna I. Scales: 10  $\mu$ m.

Рис. 33–40. *Grimaldina brazzai*, партеногенетическая самка из Baia do Castelo: 33–35 — дистальная часть постабдомена, вид со спины, и щетинки на ней; 36–37 — постабдоминальные когти, вид сбоку и с брюшной стороны; 38–39 — антенна II и ее дистальный членик; 40 — дистальная часть антенны I. Масштаб: 10  $\mu$ m.

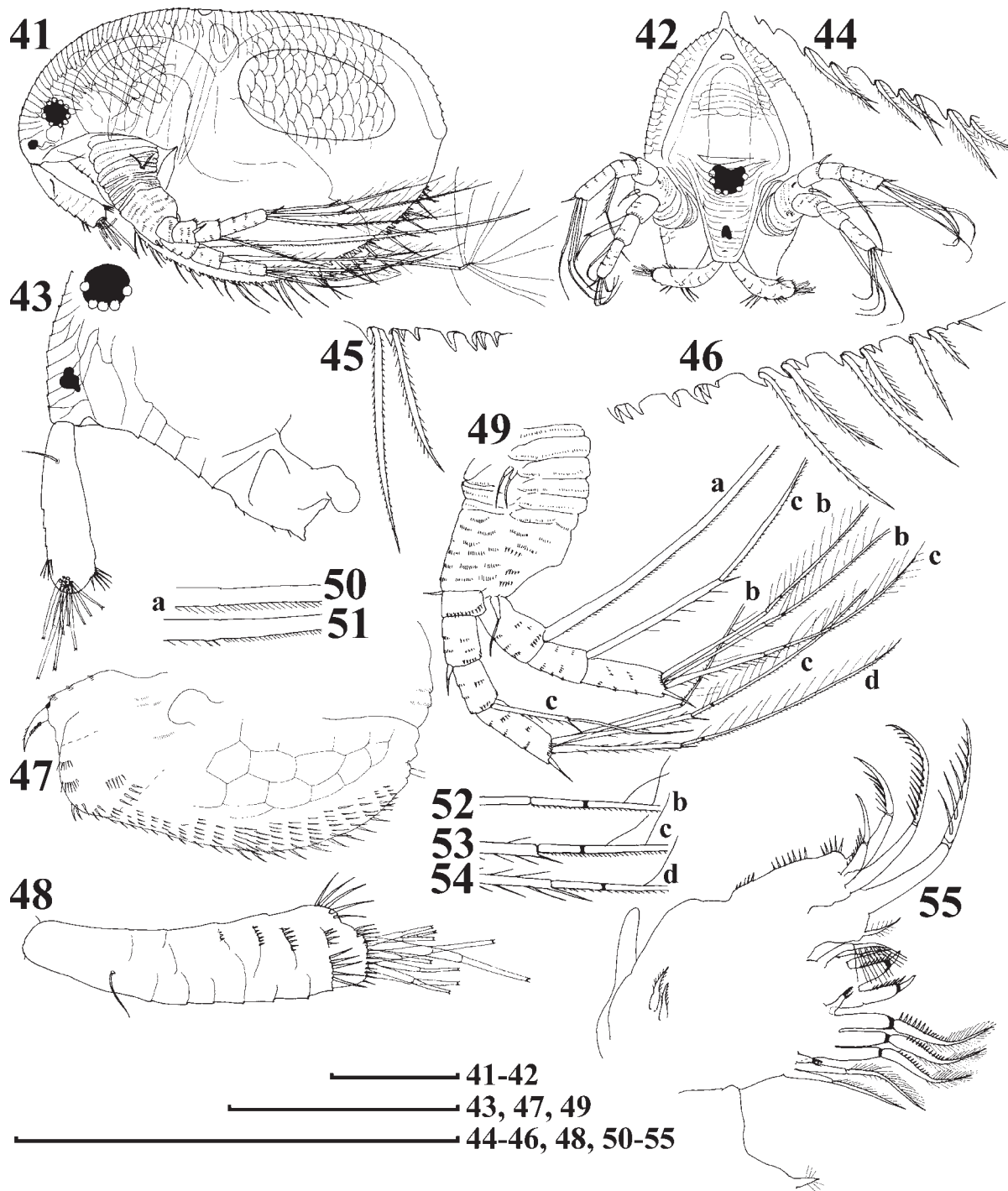
presence of marginal triangular teeth here and marginal setae articulated exactly to these teeth. Postabdomen sub-quadrangular, not bilobed, truncated distally, without a “heel” basally. Transverse rows of setules on preanal margin, postanal margin very small. Postabdominal setae short, as long as postabdomen or shorter, with very short distal segment, armed with a bunch of long setules. Postabdominal claws small, armed with rows of denticles along dorsal and ventral margins. Antenna I relatively short and robust for the genus, significantly widened distally, with a short sensory seta arising at distance of  $\frac{1}{4}$  of antenna length from its base, with robust setules distally, row of setules around distal end, nine aesthetascs of significantly different sizes. Antenna II relatively large, with two sensory setae on coxal part, robust basal segment and elongated branches with rows of denticles. Antennal formula: setae 0-0-1-3/1-1-3, spines 0-1-0-1/0-0-1. Largest (basal lateral) seta significantly longer than the rest,

unilaterally setulated, other setae armed in different manner, as is shown in Figs 52–54. Limb I with ODL supplied with a large and a rudimentary seta, IDL with 3 setae of different size and series of setules, a very short seta on gnathobase I. Size 320–450  $\mu$ m in our material, up to 540  $\mu$ m according to Smirnov [1992].

Ephippial female: was not described from South America. Male: see Smirnov [1992].

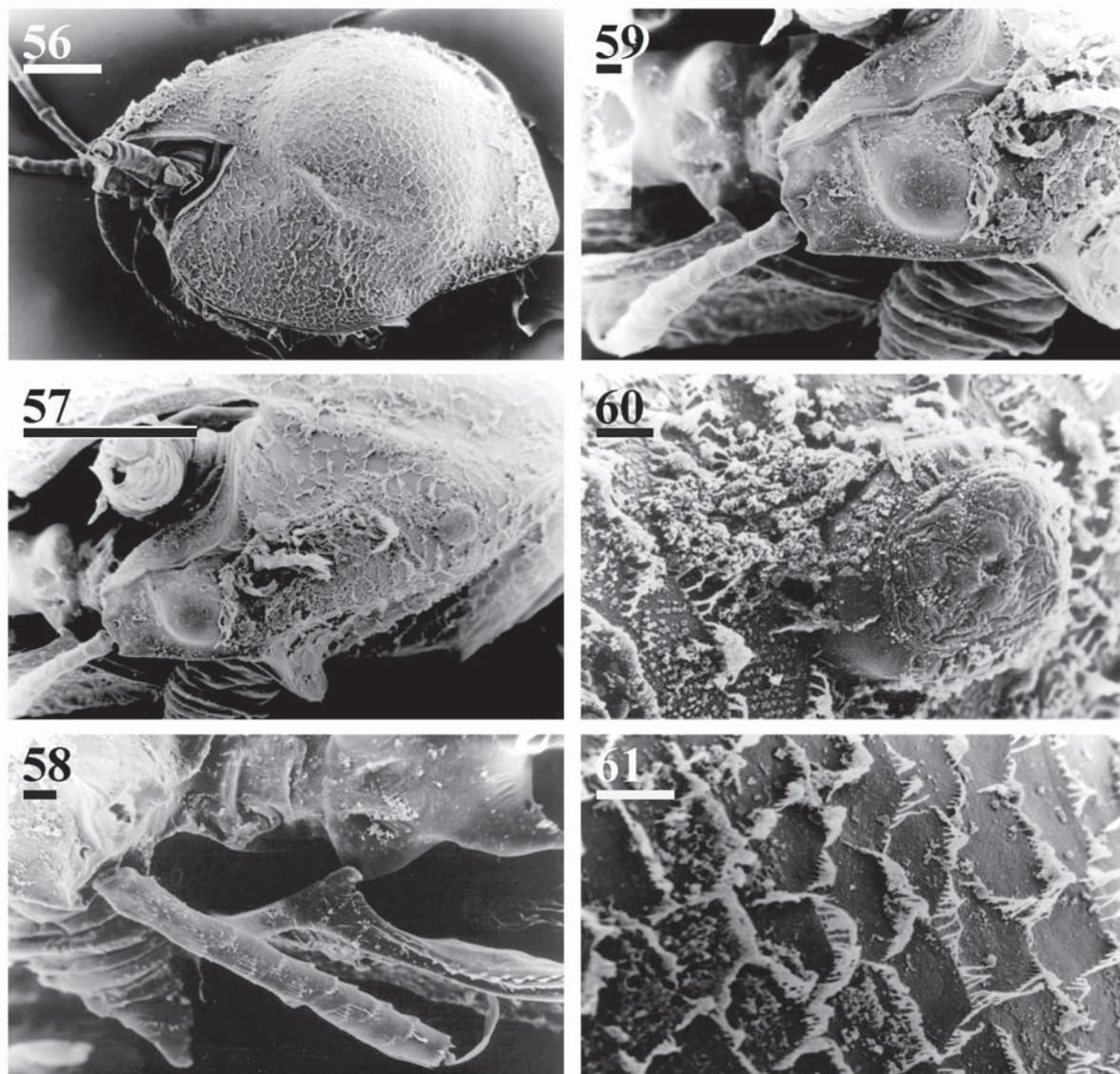
COMMENTS. Most probably, the *M. laticornis* reported by Richard [1897], Daday [1905] and Harding [1955] is *M. spinosa*, a very common species from South America. The difference between “serrated” and “squamosa” dorsal outline of the valve — the main distinctive trait in all old and recently used keys — is quite subjective, and is not supported by any morphometric data for these conditions. This confusion accounts for many of the misidentifications from different regions of the world. Real differences between *laticornis* and





Figs. 41–55. *Macrothrix spinosa*, parthenogenetic female from Baia do Castelo: 41–42 — adult in lateral and anterior view; 43 — head; 44–46 — setae at antero-ventral, middle, and postero-ventral part of valve; 47 — postabdomen; 48 — antenna I; 49 — antenna II; 50–51 — largest basal (lateral) seta in two specimens; 52–54 — swimming setae of three types in antenna II; 55 — thoracic limb I. Scales: 100  $\mu$ m.

Рис. 41–55. *Macrothrix spinosa*, партеногенетическая самка из Baia do Castelo: 41–42 — вид сбоку и спереди; 43 — голова; 44–46 — щетки в передне-брюшной, средней и задне-брюшной части створки; 47 — постабдомен; 48 — антенна I; 49 — антенна II; 50–51 — наибольшая латеральная щетинка двух самок; 52–54 — плавательные щетинки трех типов на антенне II; 55 — нога I. Масштаб: 100  $\mu$ m.



Figs. 56–61. *Onchobunops tuberculatus*, parthenogenetic female from Baia do Castelo: 56 — adult in lateral view; 57–58 — head in dorsal and latero-ventral view; 59 — anterior head portion with frontal head pore and supra-ocular dome; 60 — dorsal head pore; 61 — sculpture of valves. Scales: 100  $\mu\text{m}$  for 56–57, 10  $\mu\text{m}$  for 58–61.

Рис. 56–61. *Onchobunops tuberculatus*, партеногенетическая самка из Baia do Castelo: 56 — вид сбоку; 57–58 — голова сбоку и латеро-вентрально; 59 — передняя часть головы с фронтальной головной порой и глазным куполом; 60 — спинная “головная пора”; 61 — скульптура створок. Масштаб: 100  $\mu\text{m}$  для 56–57, 10  $\mu\text{m}$  для 58–61.

*spinosa* groups concern the thoracic limbs [Silva-Briano, 1998; Silva-Briano et al., 1999]. Sars [1901] discussed differences between his *M. squamosa* and *M. laticornis*, but said nothing about *M. spinosa*, perhaps because he did not know King’s species. Only a new, more detailed revision can reveal the true status of South American *spinosa*-like populations. Smirnov [1992] discussed the possibility of *M. goeldi* Richard, 1897 as a junior synonym of *M. spinosa*, but populations from the non-tropical regions of the southern South American continent (“*M. goeldi*”) must be checked.

#### 6. *Onchobunops tuberculatus* Fryer & Paggi, 1972 Figs. 3, 56–61.

Fryer & Paggi, 1972: 255–261, Figs. 1–13; Fryer, 1974: 204–211, Figs. 78–90; Smirnov, 1976: 148, Fig. 133; 1988: 76, Fig. 35;

1992: 106–107, Figs. 454–458; Paggi, 1995: Figs. 77–78; Silva-Briano, 1998: 136–138, Figs. 31–40; Silva-Briano & Dumont, 2001: 25–27, Figs. 31–40 (*Bunops*); Elías-Gutiérrez et al., 2001: 46, Fig. 13.

? Daday, 1905: 195, pl. 12: Figs. 15–17 (*Macrothrix gibbera*).  
Type locality: “Madrejón Don Felipe also known as Laguna Ramirez — a lagoon lying between the Rio Colastine and the Rio Santa Fe, between the towns of Santa Fe and Paraná” [Fryer & Paggi, 1972], Argentina.

Parthenogenetic female: Body high, almost globular, with slight dorsal keel. Dorsal margin arched from tip of rostrum to broadly round postero-dorsal angle, border between head and valves not expressed. Reticulation on valves and head, rows of setules arising from striae of reticulation, due to this dorsal margin squamose. Head small, with straight ventral margin and convex dorsal margin, massive supra-ocular dome and a relatively large projection for dorsal head pore elevated under



the latter, rostrum short, with frontal head pore. Valves in general ovoid, a projection on ventral margin subdivides it into two seta-fringed flanges (see Fryer & Paggi [1972]); at anterior margin of valve there is an inwardly directed hook-like projection. Postabdomen bilobed, postabdominal seta short with a relatively long distal segment. Postabdominal claws relatively large. Antenna I, II and thoracic limbs described in detail by Silva-Briano [1998] and Silva-Briano & Dumont [2001]. Size 570–900 µm in our material, but up to 1100 µm according to Fryer & Paggi [1972].

Ephippial female and male unknown.

COMMENTS. Silva-Briano & Dumont [2001] have serious doubts in the validity of the genus *Onchobunops* Fryer & Paggi, 1972, and their argument based on the similarity of all thoracic limbs in *O. tuberculatus* and *Bunops serricaudata* (Daday, 1884) is quite convincing. Recently the system of “macrothricid-like anomopods” (in terms of Silva-Briano [1998]) has been under reconstruction (see Dumont & Silva-Briano [1998]), and we are not sure about the necessity of immediate rejection of “conventional” generic names. There are problems with the species name also: previous specimens described as *Macrothrix gibbera* Daday, 1905 are most probably this taxon, and “*gibbera*” is a pretender to “*tuberculatus*” keeping in mind the impossibility of replacing of the major synonym in this situation, according to paragraph 23.9.1. of ICZN [2000]. Unfortunately, Daday’s type specimen (DAD II/ p-467) is in a very poor condition [Smirnov, 1992], and cannot be used to clarify this difficult taxonomic situation. The situation could be resolved by an investigation of animals from the exact type locality as reported by Daday [1905], but, most probably, conservation of “*tuberculatus*” must be confirmed in particular by the International Commission on Zoological Nomenclature.

#### C. Family CHYDORIDAE Stebbing, 1902 emend. Dumont & Silva-Briano, 1998

##### a. Subfamily *Aloninae* Dybowski & Grochowski, 1894 emend. Frey, 1967

###### 7. “*Alona iheringi* Sars, 1901”

Sars, 1901: 49–51, Pl. 9: Figs. 2a–c (*Alona iheringi* n.sp.); Sinev, 2001: 113–119, Figs. 1–33.

Not *Alona davidi* var. *iheringi* Richard, 1897: 294–296, Figs. 42–43.

COMMENTS: This species of the *Alona costata*-group, which is closely related to *A. rustica*, was redescribed by Sinev [2001]. He examined Sars’ type material from São Paulo and additional material from Boa Vista. We found only one female in the Canal do Tamengo, with external morphology that agrees with the characteristic features of this species. *A. iheringi* Sars, 1901 is an unacceptable taxon, a junior synonym of *A. iheringi* Richard, 1897, which was not mentioned by either Sars [1901] or Sinev [2001]. This taxon must be renamed, and, most probably, should be described as a new species.

###### 8. *Alona ossiani* Sinev, 1998 Figs. 4, 9.

Sars, 1901: 48–49, Pl. 9: Figs. 1a–c (*affinis*); Daday, 1905: 171 (*affinis*); Stingelin, 1913: 621; Birabén, 1939: 656–658, Figs. 7–8 (*affinis*); Elmoor-Loureiro, 1998: 33 (*Biapertura affinis*); Sinev, 1998: 105–110, Figs. 1–4.

COMMENTS: *Alona ossiani* is a newly described Neotropical species from the *A. affinis* group, which was described based on Sars’ specimens from São Paulo [Sinev, 1998]. We found a single specimen as well as several shell remains that could definitely be identified by the well-developed posterodorsal corner of the carapace and the setules at the posteroventral corner. The position of the head pores as well as the carapace covered with fine striae are further characteristics of *Alona ossiani*.

###### 9. *Karualona muelleri* (Richard, 1897) Fig. 8.

COMMENTS: Dumont & Silva-Briano [2000] established a new genus *Karualona*, and recently the South American species *Alona muelleri* Richard, 1897 has been re-described and translocated into this genus [Sinev & Hollwedel, 2003].

###### 10. *Leydigia* cf. *striata* Birabén, 1939

Daday, 1905: 185, Pl. 11: Fig. 19 (*acanthocercoides*); Birabén, 1939: 661–663, Figs. 14–16; Harding, 1955: 242–243, Figs. 50–60 (*ciliata*); Smirnov, 1971: 454, 458 (Neotropical records).

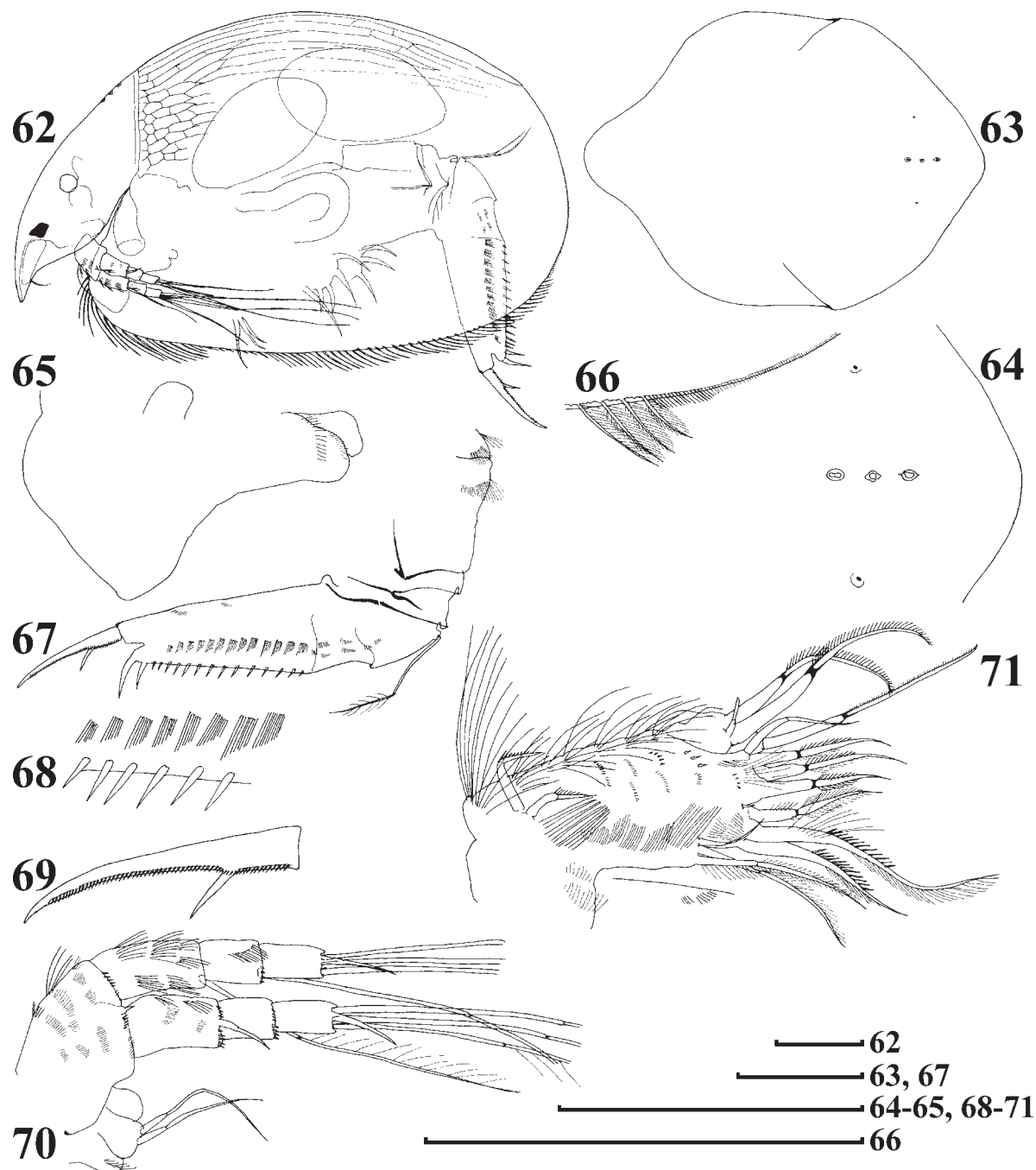
COMMENTS: Harding [1955] considered this species a junior synonym of *L. ciliata* Gauthier, 1939, but recently Kotov et al. [2003b] demonstrated differences between these two species.

###### 11. *Oxyurella longicaudis* (Birge, 1910) Figs. 7, 62–71.

Daday, 1905: 178–180, Pl. 11: Figs. 12–13 (*Euryalona tenuicaudis*); Birge, 1910: 1045–1048, Pl. 71: Figs. 3–4, 7 (*Odontalona*); Brehm & Thomsen, 1936: 216 (*Odontalona*); Brehm, 1938: 100, Fig. 5 (*Odontalona*); Bergamin, 1941: 163, Fig. 2; Martínez de Ferrato, 1966: 400–401, Pl. 2: Figs. 3–4; Smirnov, 1971: 496, Figs. 627–628; Elmoor-Loureiro, 1998: 37.

Type locality: Not clearly indicated: “Lake Charles, La.,” from U.S.A., and “Demerara, South America”, from unknown country [Birge, 1910: 1046]. Also, this author referred to Daday’s [1905] description of *Euryalona tenuicaudis* from Paraguay as *O. longicaudis*, so Daday’s specimens are included as members of the Birge’s type series according to ICZN [2000]! The type locality must be determined by means of selection of the lectotype (from materials of Birge or Daday).

Parthenogenetic female: Body ovoid, elongated, compressed laterally. Dorsal margin regularly arched from tip of rostrum to practically completely smooth postero-dorsal angle, posterior margin regularly convex. Reticulation absent on head shield, while well-developed longitudinal striation present on valves. Head with relatively long rostrum, head shield somewhat elongated, with three major head pores without connection, postpore distance somewhat more than interpore distance, ocellus somewhat smaller than eye. Lateral head pores at distance similar to interpore distance from middle line of head shield. Labral keel triangular-round, large, with expressed apex, rows of setules in posterior portion of labrum, distal labral plate small. Valves ovoid, with numerous ventral marginal setae, fine setules submarginally on inner face of valve in region of postero-ventral angle and whole posterior margin. Postabdomen elongated, narrowing somewhat distally, with anal margin somewhat shorter than preanal margin, thin teeth along all postanal margin, only two distal-most teeth significantly larger than the rest, lateral fascicles of setules decreasing in size distally, and absent near largest



Figs. 62–71. *Oxyurella longicaudis*, parthenogenetic female from Canal do Tamengo: 62 — adult in lateral view; 63–64 — head shield and head pores; 65 — labrum; 66 — setae at postero-ventral portion of valve; 67–69 — postabdomen, its postanal margin, and postabdominal claws; 70 — antenna II; 71 — thoracic limb I. Scales: 100  $\mu$ m.

Рис. 62–71. *Oxyurella longicaudis*, партеногенетическая самка из Canal do Tamengo: 62 — вид сбоку; 63–64 — головной щит и головные поры; 65 — лярбрум; 66 — щетинки в задне-брюшной части створки; 67–69 — постабдомен, его постанальный край и постабдоминальный коготь; 70 — антенна II; 71 — нога I. Масштаб: 100  $\mu$ m.

teeth, a deep embayment under base of claws. Postabdominal setae short, postabdominal claws relatively thin, curved distally, with a large basal spine located relatively far from base of claw. Antenna I not reaching tip of rostrum, with 9 aesthetascs. Antenna II relatively short, but thin, two sensory setae on coxal portion long, somewhat different in size, basal segment robust, with numerous rows of setules, branches

relatively thin. Antennal formula: setae 0-0-3/1-1-3, spines 1-0-1/0-0-1, all spines relatively long. Limb I with ODL bearing a single long seta, and IDL with 2 setae and a rudiment of a third seta, no gnathobase on limb I. Size 520–635  $\mu$ m in our material, maximal size more than that reported by Smirnov [1971].

Ephippial female: Unknown.



Male: Described poorly by Brehm [1938].

COMMENTS. This species is well-defined, and South and North American populations seem to belong to the same species. They are easily distinguished from other Neotropical species, *O. ciliata* Bergamin, 1939 (see Rey & Vasquez [1986]; Dimas Flores [2002]) in lacking setules on the labral keel. In contrast to *O. tenuicaudis* (Sars, 1862) and *O. brevicaudis* Michael & Frey, 1983, *O. longicaudis* has two especially robust teeth in distal portion of postabdomen and third seta on IDL.

#### b. Subfamily Chydorinae Stebbing, 1902

##### 12. *Alonella dadayi* (Birge, 1910)

Figs. 72–84.

Daday, 1905: 168–169, Pl. 10: Figs. 18–23 (*Leptorhynchus dentifer*); Stingelin, 1909: 644, Fig. 2 (*Leptorhynchus dentifer*); Birge, 1910: 1052; Brehm & Thomsen, 1936: 217, Fig. 9; Martínez de Ferrato, 1966: 398–399, Pl. 1: Figs. 5–7; Smirnov, 1971: 277, Fig. 290 (*Disparalona*); Green, 1972: 221; Paggi, 1995: Fig. 127 (*Phryxura*); Smirnov, 1996: 88–90, Figs. 328–331; Elmoor-Loureiro, 1998: 29 (*Disparalona*); Dimas Flores, 2002: 77, Pl. 1: Figs. A–F.

Type locality: Not reported accurately in the first description, only marked as “Louisiana, Tex.” in a table on p. 1053. Also Birge [1910] referred to *Leptorhynchus dentifer* Daday, 1905, so Daday’s material can be regarded a part of the Birge’s type series according to ICZN [2000]! It is necessary to select a lectotype of this species also.

Parthenogenetic female: Body ovoid, moderately elongated, compressed laterally. Dorsal margin regularly arched from tip of rostrum to distinct postero-dorsal angle, posterior margin truncated, postero-ventral angle rounded, with series of small denticles, setules between them. Whole shell richly sculptured with longitudinal striation in dorsal portion and head shield, and polygonal reticulation in posterior portion of valves, parallel strokes within reticulation cells. Head large, with remarkably long, somewhat curved backwards rostrum, ocellus smaller than eye. Head pores on special narrow, non-reticulated plate, each of two major head pores with thin ring, postpore distance only 1/4 interpore distance, two minute pores on mid-line, somewhat closer to anterior major pore. Labral keel large, more or less triangular-round, with wavy anterior margin and apex directed posteriorly, distal labral plate relatively large. Postabdomen robust, high, narrowing distally, preanal margin much longer than anal margin, preanal angle inflated, obtuse or right angled, a series of especially long setules in this region; anal margin significantly concave, with small, numerous teeth. Postanal margin shorter than preanal margin, with 6–8 teeth. Postabdominal seta shorter than postabdomen, its distal segment shorter than basal one, with relatively long setules; postabdominal claws relatively short and thick, with robust basal spine at a distance from claw base, and a small additional spine closer to claw base. Antenna I thick, with fine, short sensory seta and 9 aesthetascs of different size. Antenna II relatively short, but elongated, antennal formula: setae 0-0-3/1-1-3, spines 1-0-1/0-0-1. Each swimming seta with distal segment armed with setules, and chitinous insertions near joint with basal segment. ODL I with a single naked seta, IDL with 3 setae, smallest naked. Size 240–270  $\mu\text{m}$  in our material, up to 450  $\mu\text{m}$  according to Smirnov [1996].

Ephippial female: unknown.

Male: Smirnov [1996] only gave the size as 200  $\mu\text{m}$ .

COMMENTS. This is a very typical neotropical species of *Alonella*, remarkably different from others (see Smirnov [1996]).

##### 13. *Ephemeroporus hybridus* (Daday, 1905)

Figs. 85–96.

Daday, 1905: 158–159, Pl. 10: Figs. 5–7 (*Chydorus*); Stingelin, 1913: 629–630, Fig. 31 (*Chydorus*); Martínez de Ferrato, 1967: 329–330, Pl. 1: Fig. 11 (*Chydorus*); Smirnov, 1971: 306, Fig. 336 (*Chydorus*); Green, 1972: 220 (*Chydorus*); Frey, 1982: 238–243, Pl. 2: Figs. 1–8, Pl. III: Figs. 1–18, Pl. 4: Figs. 1–20; Rey & Vasquez, 1986: 152: Pl. 7: Figs. 1–12; Smirnov, 1996: 155–156, Figs. 643–648; Elmoor-Loureiro, 1998: 29.

Type locality: there is only a note “Paraguay” on labels of Daday’s types in HNHM. Also, in his text, the author did not select the locality from the series of water bodies investigated, so the exact type locality is not clear [Smirnov, 1996].

Parthenogenetic female: Body globular, high and thick, dorsal margin regularly arched from tip of rostrum to a rounded, but visible, postero-dorsal angle, posterior margin straight or concave. Postero-ventral angle widely rounded, with single denticle, but in many females from Pantanal this denticle significantly or completely reduced, and with specialized short setae dorsally to the tooth. Head with well-developed rostrum directed downwards, ocellus somewhat smaller than eye. No head pores in adult state. Labral keel projected ventrally, with well-developed apex and single tooth at anterior margin (in a single female this tooth was greatly reduced). Valves ovoid, with polygonal reticulation, numerous short lines within cells of reticulation, setae in anterior portion of ventral edge marginal, in the middle absent, in posterior portion submarginal, densely setulated. Postabdomen elongated, narrowing somewhat distally, dorso-distal angle rounded, distal end truncated. Preanal margin long, preanal angle prominent, postanal margin with large teeth, 3 proximal anal teeth especially long. Postabdominal seta shorter than postabdomen, postabdominal claw regularly curved, with a long basal spine and a minute additional basal spine. Antennal formula: setae 0-0-3/0-1-3, spines 1-0-1/0-0-1, all spines very short. Limb I with IDL bearing a single seta, IDL with 3 seta of different size, gnathobase I with a single seta and setulated projection. Size 220–270  $\mu\text{m}$  in our material, up to 320  $\mu\text{m}$  according to Rey & Vasquez [1986].

Ephippial female: See Frey [1982].

Male: still unknown [Smirnov, 1996]

COMMENTS. Our *E. hybridus* was collected close to the presumed type locality. Our main conclusion is that it is necessary to be very careful with presence-absence of denticles at postero-ventral angle of valves as a key character for the discrimination of species within the genus *Ephemeroporus*, because in Pantanal populations these were reduced in many specimens. A similar situation was observed with another diagnostic feature, a tooth on labrum: in one specimen from Pantanal the tooth on the labrum was strongly reduced in size. In addition, Frey [1982] noted that there are several species in the world, not yet described, allied to *E. hybridus*.

##### 14. *Pseudochydorus globosus* (Baird, 1843)

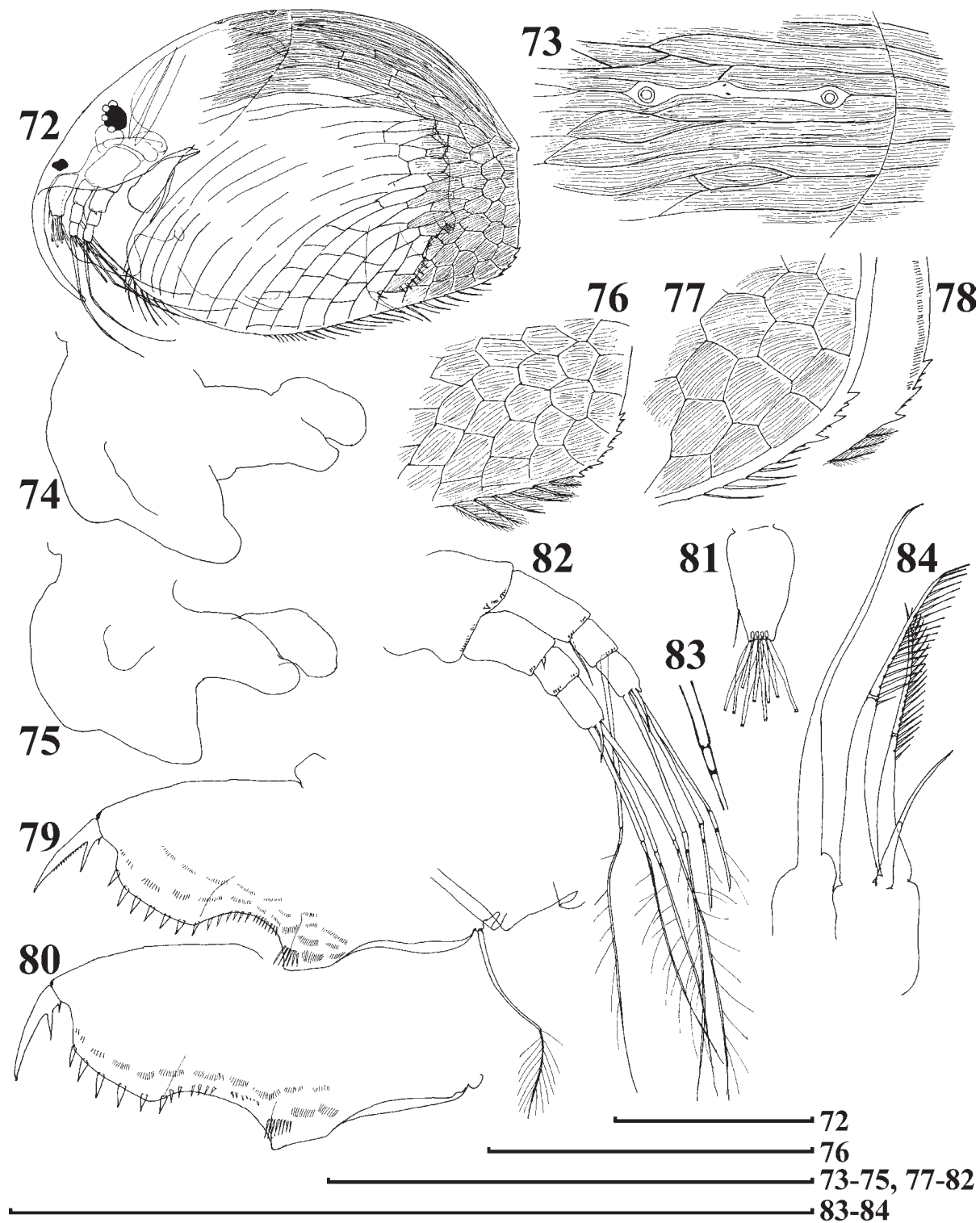
Figs. 97–108.

Bergamin, 1940: 99, Fig. 18 (*Chydorus*); Martínez de Ferrato, 1967: 328–329, Pl. 1: Figs. 6–7; Paggi, 1995: Figs. 104–106; Elmoor-Loureiro, 1998: 31.

Not Rey, 1993: 259, Figs. 26–27.

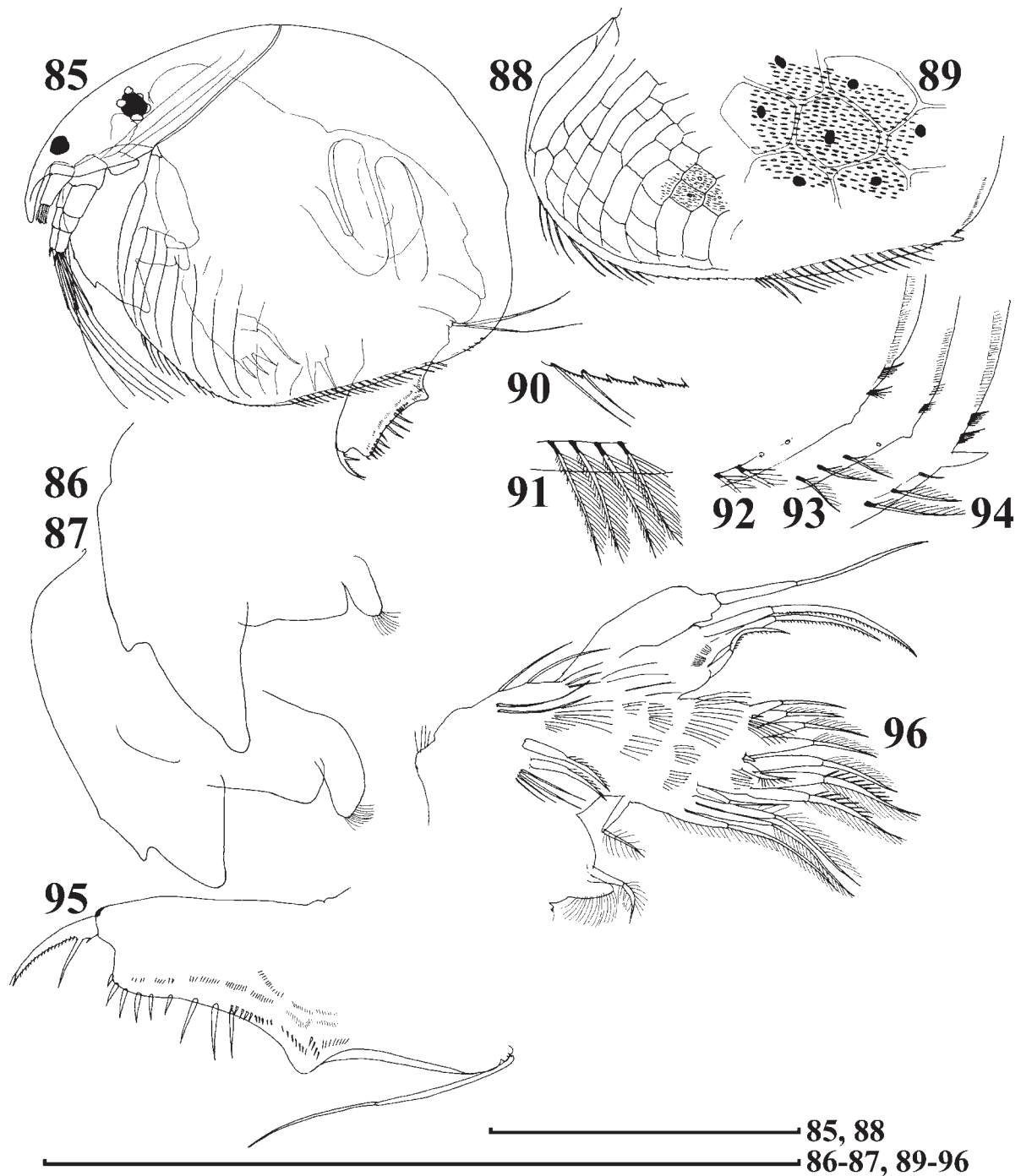
Type locality: “Ditch near Richmond; pond near Isleworth”, United Kingdom, Europe.

Parthenogenetic female: Body subglobular to globular, thick, dorsal margin regularly arched from tip of rostrum to posterior margin, sometimes there is a slight prominence



Figs. 72–84. *Alonella dadayi*, parthenogenetic female from Baia 38: 72 — adult in lateral view; 73 — head pores; 74–75 — labrum; 76–77 — postero-ventral portion of valve in external view; 78 — the same in internal view; 79–80 — postabdomen; 81 — antenna I; 82–83 — antenna II and joint part of its swimming seta; 84 — distal portion of thoracic limb I. Scales: 100  $\mu$ m.

Рис. 72–84. *Alonella dadayi*, партеногенетическая самка из Baia 38: 72 — вид сбоку; 73 — головные поры; 74–75 — лябрум; 76–77 — задне-брюшная часть створку снаружи; 78 — изнутри; 79–80 — постабдомен; 81 — антенна I; 82–83 — антенна II и сочленение дистального и базального члеников плавательной щетинки; 84 — дистальная часть ноги I. Масштаб: 100  $\mu$ m.

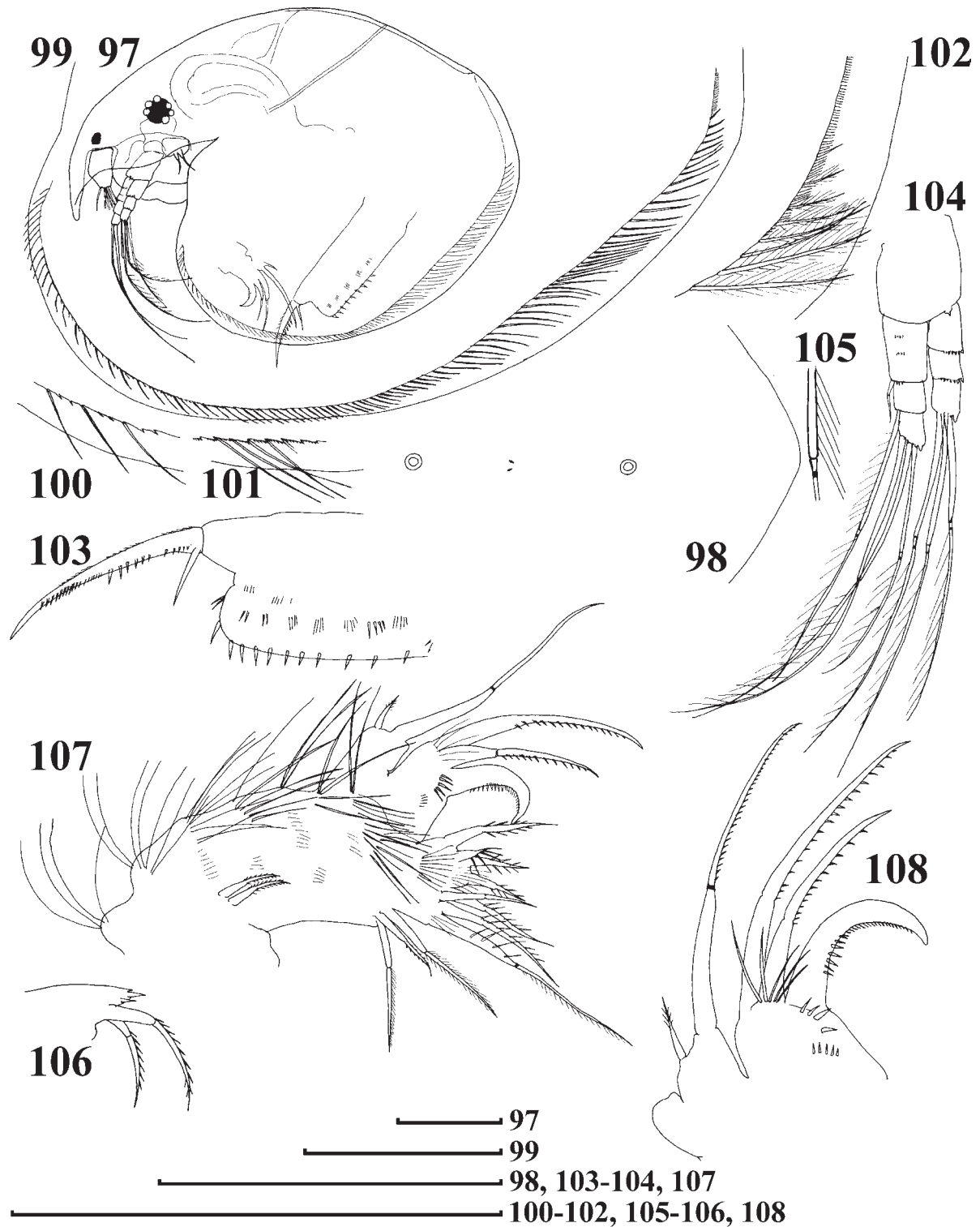


Figs. 85–96. *Ephemeroporus hybridus*, parthenogenetic female from Baia 38: 85 — adult in lateral view; 86–87 — labrum; 88–89 — valve and its sculpture; 90–91 — setae at middle and posterior portions of valve ventral margin; 92–94 — armature of postero-ventral angle in 3 valves; 95 — postabdomen; 96 — thoracic limb I. Scales: 100  $\mu$ m.

Рис. 85–96. *Ephemeroporus hybridus*, партеногенетическая самка из Baia 38: 85 — вид сбоку; 86–87 — ляррум; 88–89 — створка и ее скульптура; 90–91 — щетинки в середине и задней части брюшного края; 92–94 — вооружение задне-брюшного угла трех разных створок; 95 — постабдомен; 96 — нога I. Масштаб: 100  $\mu$ m.

under compound eye, postero-dorsal angle smooth. Reticulation not visible. Head with well-developed rostrum, two major head pores in mid-line of head shield, interpore distance greater than postpore one, small pores in mid-line also, between two major pores. Labral keel reduced. Valves ovoid, with setae submarginally on inner face along entire

ventral margin. Postabdomen elongated, with parallel dorsal and ventral margins, preanal margin longer than anal margin, preanal angle somewhat prominent, postanal margin with small teeth, distal angle rounded. Basis of postabdominal claws remarkably prominent distally, postabdominal claw relatively long and thin, with a single basal spine immediately



Figs. 97–108. *Pseudochydorus globosus*, parthenogenetic female from Canal do Tamengo: 97 — adult in lateral view; 98 — head pores; 99–102 — armature of valve, and its antero-ventral, ventral and postero-ventral portions; 103 — distal portion of postabdomen; 104–105 — antenna II and joint portion of its swimming seta; 106 — maxilla I; 107–108 — thoracic limb I and its distal portion. Scales: 100  $\mu\text{m}$ .

Рис. 97–108. *Pseudochydorus globosus*, партеногенетическая самка из Canal do Tamengo: 97 — вид сбоку; 98 — головные поры; 99–102 — вооружение створки, ее передне-брюшной, брюшной и задне-брюшной части; 103 — дистальная часть постабдомена; 104–105 — антенна II и сочленение дистального и базального члеников ее плавательной щетинки; 106 — максилла; 107–108 — нога I и ее дистальная часть. Масштаб: 100  $\mu\text{m}$ .



at its base, and spaced denticles along its dorsal margin. Antenna I short, robust, with 9 aesthetascs of different size. Antenna II relatively small, basal segment and segments of branches robust. Antennal formula: setae 0-0-3/1-1-3 (seta at basal endopod segment very short), spines 0-0-1/0-0-1, apical spines minute. Distal segments of swimming setae setulated, with chitinous insertions near joints with basal segments. No filter setae on any limbs or maxillae I. Limb I with ODL bearing a long and a short seta, IDL with three setae, shortest seta hook-like, and a bunch of robust setules. Other limbs not different from those described by Smirnov [1971]. Size 400-450  $\mu\text{m}$  in our material.

Ephippial female, male: Unknown from the Neotropics.

COMMENTS. The size of the labral keel, which is used in all keys for the discrimination of *Chydorus* and *Pseudochydorus*, is in fact not a major difference between these genera; structure of limbs is a far more important trait; in Pantanal females all limbs were clearly non-filtering. We did not find any significant differences in the structure of thoracic limbs in Pantanal and European females, but our ideas on the latter were based on pictures of Fryer [1968] and Smirnov [1971], published around 30 years ago, that do not comply with recent standards of anomopod descriptions. The status of South American populations should be checked after a redescription of European *P. globosus* s. str. In contrast, Rey [1993: 259, Figs 26–27] most probably described *Chydorus*, not *Pseudochydorus*, from Lake Titicaca: the labral keel in her animal was small but existing, with expressed apex, and the postabdomen was not of the *Pseudochydorus*-type.

## Discussion

The Pantanal fauna is quite typical of Brazil and Neotropical regions. The similarity of our species list with ones already published for these regions is quite obvious, especially bearing in mind that some differences can be explained by the changing taxonomic status of some species. Although no new species are presented in this paper, we have indicated many areas of confusion with European forms (see above) that require clarification, but they are not endemics of the Pantanal. Large South American rivers such as the Rio Paraguay flow through thousands of kilometres, and are important arteries for dispersion of freshwater species; as are the myriads of aquatic birds associated with the river banks that participate in dispersion of anomopod ephippia.

The definitive list of Brazilian Cladocera is far from complete, with new species being described all the time [Smirnov & Santos-Silva, 1995; Sinev & Hollwedel, 2002]. Brazilian investigators have published many lists of Cladocera from different regions, including the Pantanal [Lima et al., 1996, 1998] but detailed investigations including descriptions of some species have only been made for a few regions. These regions include the vicinity of São Paulo, Itatiba and Ipiranga (São Paulo State) [Sars, 1901], the city of São Paulo [Bergamin, 1940, 1941], the region of Rio Nhamundá (border of Pará and Amazonas) [Brandorff et al., 1982], Lagoa dos Patos (Rio Grande do Sul) [Montú & Gloeden, 1986], the region of Boa Vista (Roraima) [Smirnov & Santos-Silva, 1995], and Lençóis Maranhenses dune field (Maranhão) [Van Damme, in preparation]. Most of these

surveys only examined a limited number of water bodies — about 10 or even fewer. Thus the majority of Brazil is waiting for thorough investigation of its cladoceran fauna.

Many previously established species, European as well as tropical, must be redescribed according to the new standards [Korovchinsky, 1996]. Not only rare but many very common Brazilian, other South American, and Australian species of the Anomopoda are described very poorly, and this has resulted in continual misidentifications.

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