Tools for identifying selected Australian aquatic oligochaetes (Clitellata: Annelida)





## **Adrian Pinder**

**TRIN Taxonomic Guide 2.** 



Taxonomy Research & Information Network building australian capacity





Department of Environment and Conservation

Our environment, our future 🗧

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**Adrian Pinder** 

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### Taxonomy Research and Information Network (TRIN)

## **TRIN Taxonomic Guide 2.**

Presented at the Taxonomic Workshop held at La Trobe University, Albury-Wodonga Campus, Wodonga, February 10-11<sup>th</sup> 2009.







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#### INTRODUCTION

This document provides:

- 1) a guide to the characters required for oligochaete identification
- 2) a key to major annelid groups,
- 3) a key to families and some distinctive lower taxa
- 4) a key to the Naidinae.

Since the guide to identification of Australian oligochaetes was produced by Pinder and Brinkhurst (1994) there have been significant changes in oligochaete systematics and improved knowledge of Australian oligochaete diversity. Molecular work has confirmed that Oligochaeta is a paraphyletic group without inclusion of the leeches and branchiobdellidans. Clitellata is now commonly used to describe the larger group (e.g. Erseus and Kallersjo, 2004), though Oligochaeta will no doubt continue to be used as a convenient term for the 'non-leech, non-branchiobdellidan Clitellata', as it is here. Moreover, it appears to the oligochaetes may lie within the polychaete clade (e.g. Rouset et al. 2007).

The other major change is that the former Naididae sensu strictu was synonymised with the Tubificidae, with the name Naididae having precedence (Erseus et al. 2002; Erseus et al. 2005; Erseus and Gustavsson, 2002; IUZN, 2007). Thus the Naididae s.s is now a subfamily, Naidinae, within a larger Naididae that includes all the former tubificid subfamilies

(Tubificinae, Rhyacodrilinae, Phallodrilinae, Limnodriloidinae and Telmatodrilinae). This refining of clitellate systematics is continuing with indications that Naidinae itself is not a monophyly. In Australia, the former Telmatodrilinae have been placed within new genera of Rhyacodrilinae, so the subfamily is no longer represented in Australia (Pinder and Brinkhurst 2000). Finally, phallodriline naidids have been recorded in Australian limnic waters for the first time (Pinder et al. 2006).

On top of these systematic changes, the number of limnic species known from Australia has grown from 91 to almost 200, though a large proportion remains undescribed. The number of marine oligochaetes known from Australia also continues to grow.

Family	limnic described	limnic known undescribed	terrestrial	marine/ estuarine	total
Naididae	76 (23)	36 (36)	0	135	247
Phreodrilidae	32 (28)	32 (32)	0	1	65
Lumbriculidae	2 (0)	0	0	0	2
Capilloventridae	3 (3)	0	0	0	3
Enchytraeidae	4 (2)	probably many	6	33	43
Haplotaxidae	4 (1)	1 (1)	0	0	5
TOTAL	121	69	6	169	365

# Species richness within the aquatic families and subfamilies of Australia (numbers in brackets are endemic).

#### CLASSIFICATION

#### **ANNELIDA**

#### POLYCHAETA

Aphanoneura: Aeolosomatidae (have been allied with oligochaetes in the past)

#### CLITELLATA

Branchiobdelida (ectocommensal on fw crayfish - not known from Australia)Euhirudinea (leeches and acanthobdellidans, phylogenetically within the former "Oligochaete" clade)

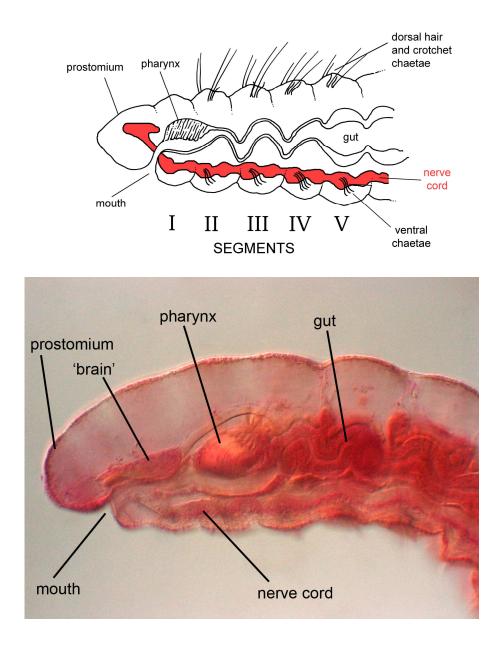
#### "Oligochaeta"

Several earthworm families	(some with aquatic or semi-aquatic representatives)	
Haplotaxidae	(freshwater/interstitial/profundal, stygal)	
Lumbriculidae	(2 introduced freshwater species)	
Naididae	(freshwater, marine, stygal)	
Tubificinae	(mostly freshwater, few marine)	
Phallodrilinae	(mostly marine, few stygal)	
Naidinae	(ex Naididae, mostly limnic, few stygal)	
Rhyacodrilinae	(marine and freshwater)	
Limnodriloidinae	(all marine)	
Phreodrilidae	(Gondwanan, almost entirely freshwater)	
Enchytraeidae	(marine, terrestrial and freshwater - poorly studied in Australia)	
Propappidae	(not known from Australia)	
Capilloventridae	(marine and freshwater)	
Opistocystidae	(not known from Australia)	
Narapidae	(not known from Australia)	
Randidrilidae	(not known from Australia)	

#### **EXPLANATION OF CHARACTERS**

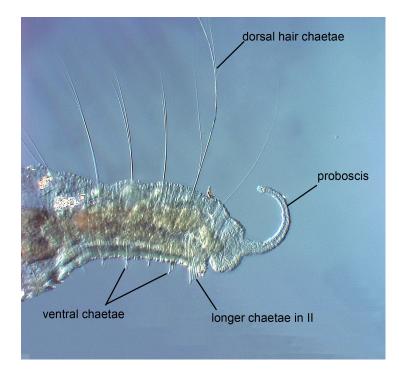
**Arrangement of the body.** Roman numerals are used to denote segments while Arabic numerals are used to denote septa between segments (e.g. 1/2, 4/5 ...). The head end is almost always broader than the anal end of a worm and has a ventral furrow (mouth) just behind the front of the worm. The most anterior part of the worm is the prostomium which is not counted as a segment. Segment I has the crescent-shaped ventral mouth but no chaetae. Chaetae generally start on segment II but dorsal chaetae may be absent on some anterior segments.

A useful orienting feature is the nerve cord which is normally easy to see and is always ventral. The nerve cord has a speckled appearance. The mouth is also ventral and normally long hairs (see below) are restricted to the dorsal side.

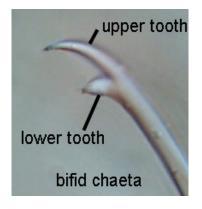


**Chaetae ( = setae of some authors)**: Chaetae occur in groups called **bundles** (although sometimes there will only be one chaeta/'bundle'). Most oligochaetes have 4 bundles per segment (2 ventro-lateral and 2 lateral to dorso-lateral). Chaetae are absent on the **prostomium** (head) and segment I but present on all or most segments thereafter, except for the anal segment (**pygidium**) and sometimes ventrally (rarely dorsally) on 1 or 2 segments containing the genitalia.

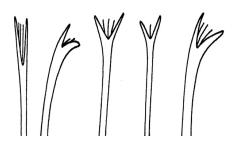
**Hair chaetae** (sometimes just called **hairs** or **capilliform** chaetae): long, very thin 'hair-like' chaetae, normally with a fine tapering tip. In most oligochaetes (except for the Capilloventridae, plus 2 naidids and 1 phreodrilid) these are restricted to dorsal bundles. Sometimes with annulations or serrations along the shaft or with secondary hairs giving a plumose appearance.



**Crotchet chaetae**: non-hair chaetae, often sigmoid in shape, usually with a swelling (**nodulus**) along the shaft, with tips either **single (=simple)** (a bluntly or sharply pointed tip), **bifid** (forked), **pectinate** (with comb-like teeth between the fork) or otherwise modified (e.g. paddle-shaped). In bifid and pectinate chaetae the **upper tooth** is on the convex side of the chaeta and the **lower tooth** is on the concave side. The relative length of these teeth is frequently used in keys.





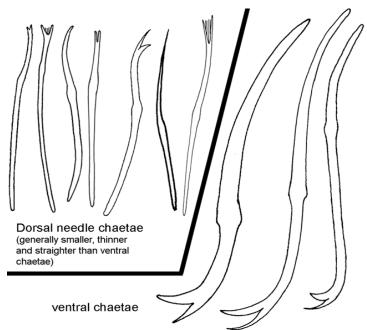


Tips of pectinate chaetae



Tips of single-pointed chaetae

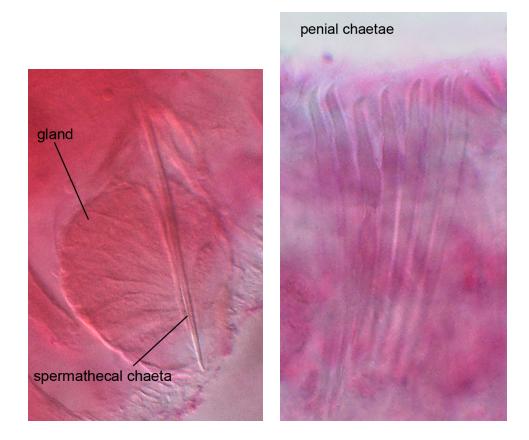
**Needle chaetae**: Most members of the subfamily Naidinae have short, fine, and usually fairly straight crotchet chaetae called **needles** (see previous page). These are generally <u>much</u> smaller than the bifid ventral chaetae and usually occur with hairs. High magnification is normally needed to see the form of the tips of needle chaetae.



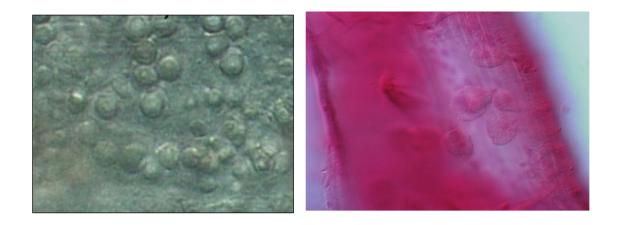
Naidine needle chaetae (left) compared to bifid ventral chaetae at same scale. Note nodulus (swelling part way along shaft).

**'Support' chaetae**: Most Phreodrilidae have a pair of small fine chaetae 'support' surrounding the base each hair or crotchet chaeta. These look very much like especially fine needle chaetae (when they can be seen) but they do not emerge from the chaetal sac within the body wall and are only visible on slide mounted specimens by looking through the epidermis. Some Naidinae, especially some *Pristina*, have such fine needles that they appear much like support chaetae but <u>do</u> emerge from the body wall.

**Genital chaetae**: The ventral chaetae of mature specimens are usually absent or modified on the segments bearing the genital pores. These are mostly of two forms, 1) long straight single chaetae with hollow distal halves and sharp blade-like tips, associated with a large gland and normally associated with one much smaller chaeta or 2) one or more chaetae, often in parallel or with the distal ends bunched together, with bifid or bluntly simple tips. Both types are largely hidden within the body but can be seen in stained and cleared specimens. Genital chaetae of capilloventrids are long thickened hairs.



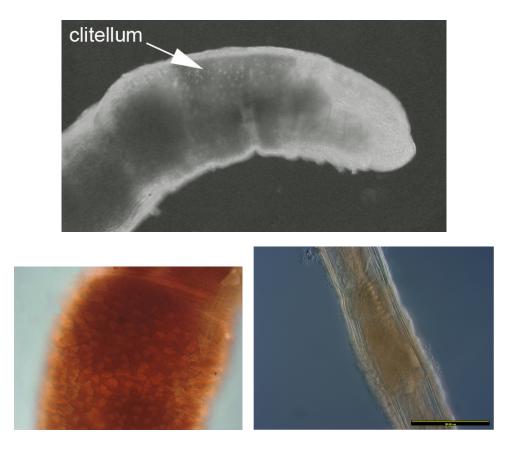
**Coelomocytes**: round to oval cells free floating in the coelomic cavity, in clusters or individually. Often very dense in naidine and rhyacodriline naidids. These can often be seen in glycerol mounted specimens.



**Genitalia:** Although most specimens can be identified to family using external features, the most reliable way to identify a specimen to family is to use the genitalia. Knowing the segmental position of the spermatheca and male genital ducts is generally sufficient to arrive at a family, though this requires mature specimens which are not always present in a sample. Oligochaetes are hermaphrodites so have male and female genitalia. Examination of the genitalia is normally required for genus or species level identifications, with the notable exception of the Naidinae.

**Clitellum**: The easiest way to determine whether a worm is mature is to look for the clitellum. This is a thickened and more opaque region of the body with a different texture resulting from development of a glandular layer of cells around 2 or more segments in the region of the genitalia. This cell layer secretes the cocoon (below right) into which the sperm and eggs are deposited.





The clitellum is located as follows for the different families and subfamilies:

Naidinae	two consecutive segments between IV and VIII
Other Naididae	X and XI
Enchytraeidae	XI and XII
Phreodrilidae	1/2 XII and all of XIII
Lumbriculidae	several segments from VIII or IX
Capilloventridae	1/2 XII to segment XIV
Haplotaxidae	over many segments from about X or XI

Genital organs can often be seen in glycerol mounted specimens if the worms are small, but its generally best to stain and clear the specimens (see below), especially for long term storage as glycerol will eventually over clear soft tissues.

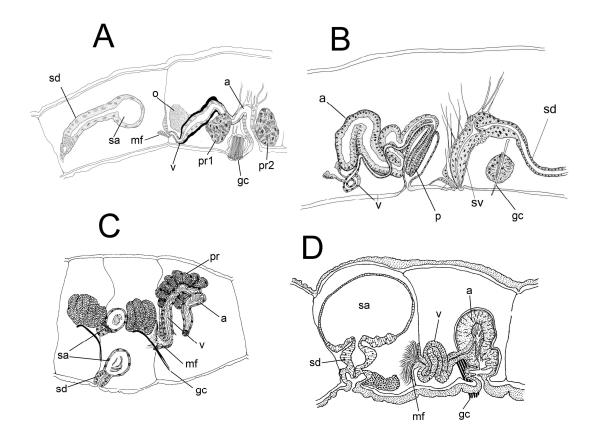
The **spermatheca** is an ovoid to elongate sac for storage of sperm after mating. There are normally two spermatheca present in one segment, usually with separate (and often indistinct) pores on the body wall of the same segment (normally ventro-lateral).

The **male ducts** transport sperm from the testes to the exterior. The normally paired male ducts start with **sperm funnels** on the posterior wall of the testes segment and in mature stained specimens this can be seen as a red mass looking like a dense tassel. The funnel feeds sperm into the vas deferens which leads to the male pores, usually via an **atrium** with associated glands (**prostate**) and a **penis** lying in an invagination of the body wall (**penis sac**), but there are many variations around this template. The pores are usually on the ventro-

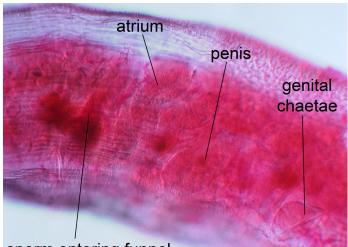
lateral body wall of the post-testes segment but in lumbriculids and earthworms pores can be 2 or more segments behind the testes.

Below are cross sections of the segments containing the genitalia of four species from different oligochaete groups, showing variation in position and form of the genital organs. A: *Pectinodrilus ningaloo* (Naididae: Phallodrilinae); B: *Insulodrilus linae* (Phreodrilidae); C: *Pristina osborni* (Naididae: Naidinae) and D: *Ainudrilus nharna* (Naididae: Tubificinae). The *Pristina osborni* illustration is from Erseus and Grimm (1998).

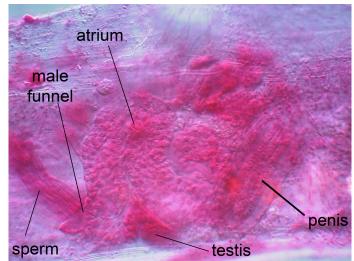
sa = spermatheca, sd = spermathecal duct, sv = spermathecal vestibule, mf = male funnel (=sperm funnel), p = penis, o = ovary, v = vas deferens, a = atrium, pr = prostate, gc = genital chaetae.



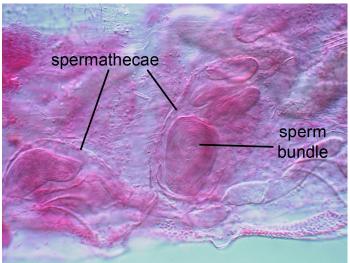
Images of genitalia on a stained and slide mounted specimen



sperm entering funnel



close up of male genitalia



spermathecae with sperm bundles

#### METHODS FOR EXAMINING WORMS

#### Fixation and preservation

Worms may be killed and preserved in 5 to 10% formalin (2 to 4% formaldehyde) or in other histological fixatives such as Bouins, then stored in alcohol, although alcohol preserved material is normally adequate.

#### **Examination of specimens**

Few features can be seen with a dissecting microscope. The head (prostomium) is usually larger and more rounded than the tail and has a slit (mouth) along the ventral side. The prostomium may bear a proboscis and the tail may have knobby or elongate gills. The chaetae can often be seen but, apart from seeing whether hair chaetae are present and how long they are compared to the body width, little of their form can be seen.

Where more than one species are present, a preliminary separation of specimens into species (or groups of similar species), based on preserved colour, size, degree of coiling and contraction and presence and number of hair chaetae, can be made in the Petri dish. However, multiple specimens of each such separated group should still be made as some species can look identical at this gross level and examination of multiple specimens is often required to see all features.

Temporary slide mounts can be made in glycerol. For ease of mounting, worms are best temporarily transferred to water then into glycerol. Try to mount the worm laterally, so that both dorsal and ventral chaetae can be seen. When a worm is lateral the mouth can be seen as a ventral indentation between the prostomium and segment I. Conveniently, many worms naturally lie this way after preservation. For bulk identifications about 5 specimens are easily mounted under each of 2 coverslips on a slide. Specimens should not be left in glycerol for more than a few days as they will over clear.

The procedure for examining a worm mounted on a slide is as follows.

Start at the head end. Check the prostomium for a proboscis, segment I for eye spots (usually one purple coloured cell cluster on each side where present) and the body itself for gills or papillae. Next, examine the chaetae; determine which segment has the first dorsal chaetae and establish the number and form of the ventral and dorsal chaeta from a number of anterior and posterior segments. Don't forget that ventral chaetae start on segment II. The nerve cord is often usually visible and as it is always ventral (below the gut) is a useful indicator of the ventral side. Determine the relative lengths of the teeth of any bifid or pectinate chaetae. Examine at least three bundles in those segments examined to make sure that both dorsal and ventral chaetae have been seen. Care should be taken to examine several chaetae from an exactly lateral aspect because slight deviations in angle of view can produce apparent distortion of the relative lengths of the teeth. Some compression of the worm may be required (but not too much as the genitalia can be damaged if required later.

Check for presence of a clitellum which will indicate maturity.

The worm should then be searched for genital characters, if required. Carefully check the genital segments (X-XI in tubificids for example, see below) to see if the ventral chaetae of

mature specimens are modified. Check the segment with male ducts to see if there are penis sheaths (cuticular coverings of the penes); these may be thin and inconspicuous but often visible through the body wall. Other components of the genital system, such as penes, atria and sperm and eggs in sacs may also be visible through the body wall almost always requires the specimen to be stained and cleared (see below). If a specimen is mature and cannot be identified without examining the genitalia, then the following staining procedure should be used.

- 1. Soak the worm in water or alcohol to remove glycerol if required (a couple of minutes).
- 2. Place in about 0.5ml carmine stain (see recipe below).
- 3. After 10-30 minutes depending on size of specimen (10 minutes usually suffices for most worms), add a single drop of concentrated hydrochloric acid.
- 4. After a few minutes, remove specimen from stain and place in a solution of 3% HCl in 70% alcohol. This removes excess stain. The worm may need placing in fresh solution once or twice before it becomes a light pink colour.
- 5. Remove to 70% alcohol for a few minutes to remove acid.
- 6. Place in 100% isopropanol (<u>not</u> ethanol). Enchytraeids sometimes shrink at this stage and may need some intermediate concentrations.
- 7. Place in a solution of 50% Histoclear/50% isopropanol (few minutes).
- 8. Place in 100% Histoclear (few minutes).
- 9. Mount in a resinous mountant such as Permount, Canada Balsam or Ultramount, trying to get the worm to lie exactly on its side (rather than dorsally or ventrally) so that both ventral and dorsal chaetae can be seen and the genitalia can be viewed laterally. If done properly the indentation between the prostomium and segment I (the mouth) will be visible and the body will be straight (not twisted) thereafter (or at least not twisted for the first 15 or so segments). On animals that are coiled, these anterior segments need to be cut off before mounting so they can be mounted without overlapping the rest of the body.

#### Recipe for Grenacher's borax carmine

Carmine (C.I. 75470) 3.0g Borax 4.0g 70% alcohol 100ml Distilled water 100ml

Dissolve borax in water and then add carmine, boil or leave to stand until the carmine is dissolved. Add 70% alcohol and allow to stand for 1 to 2 days before filtering.

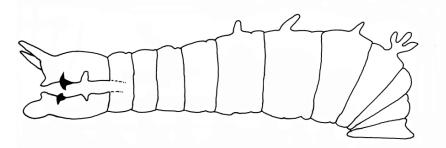
#### Examination of the genitalia

The location of the genital organs is fixed depending on the family or subfamily as follows:

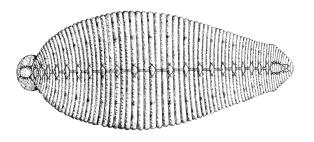
Enchytraeidae: spermathecae in V, testes in XI, male ducts in XII. Naidinae (except Naidinae): spermathecae and testes in X, ovaries and male ducts in XI Naididae (Naidinae): spermathecae and testes in V, ovaries and male ducts in VI. Phreodrilidae: testes in XI, ovaries and male ducts in XII, spermathecae in XIII The easiest to find genital structure is generally the sperm (or male) funnel (located at the rear of the testes segment) as it will have a mass of darkly stained sperm crowding into the funnel. From here the thin and usually transparent vas deferens can usually be followed to the atrium in the next segment. The atrium is usually the next most visible organ as it is often large, though may be obscured if large prostate glands are present. The spermathecae are also easy to find, especially in mated specimens as they will be filled with darkly stained sperm. It is necessary to focus up and down a lot to follow the often convoluted 3D paths of these ducts.

#### **KEY TO ANNELID GROUPS**

- 1 Chaetae absent; anterior and (usually) posterior attachment organs ('suckers') present 2
- Chaetae present (may be hard to see in some earthworms); attachment organs absent 3
- 2 Sometimes with short tentacles around the mouth and/or digitiform or multibranched dorsal appendages on some segments; protrusible proboscis absent; body consisting of head plus 11 body segments; ectosymbiotic on astacid crayfish and a few other crustacean Clitellata: Branchiobdellida (not known from Australia but present in Asia)

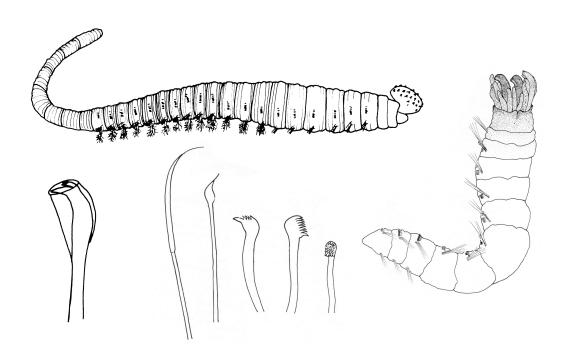


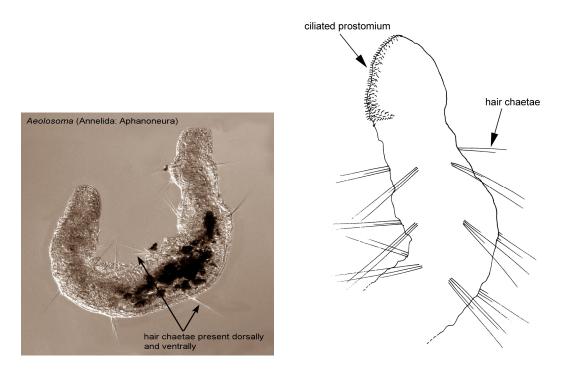
General body plan of a branchiobdellid

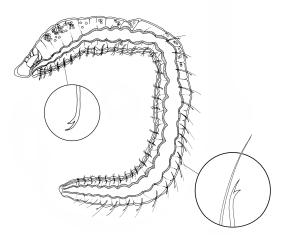


from Sawyer 1986

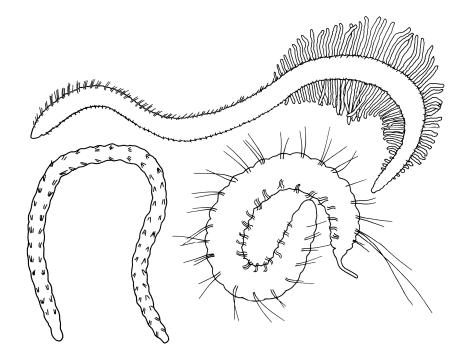
3. Head may bear multiple appendages; jaws may be present; body usually divided into distinct regions with different widths and types of chaetae; often separate male and females; chaetae often complex or hooded and usually borne on a pair of lateral outgrowths (parapodia); mostly marine but a few freshwater/athalassic saline species...
Polychaeta







Nais barbata (Naididae) showing hair chaetae in dorsal bundles only



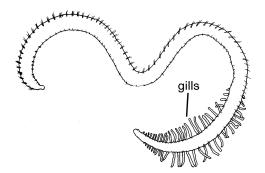
A variety of oligochaete body forms

#### KEY TO THE FAMILIES, SUBFAMILIES AND SOME ABERRANT SPECIES OF AQUATIC OLIGOCHAETES

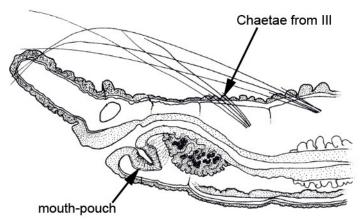
(mature specimens required for some groups)

1.	Hair chaetae present in ventral bundles	2
-	Hair chaetae absent in ventral bundles	5

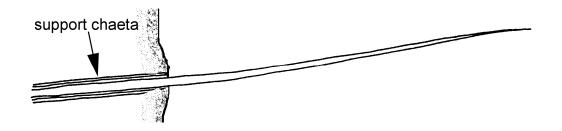
- Hair chaetae absent in anterior-most chaetal bundles; ventral chaetae present from II ....
   Naididae: Rhyacodrilinae: Breviatria papillatus (Tas only), see Pinder and Brinkhurst (2000).



- 4. Chaetae absent in segment II and sometimes III; ventral mouth-pouch present; mature specimens with ventral chaetae of XII modified as thickened hairs .... Capilloventridae: *Capilloventer*

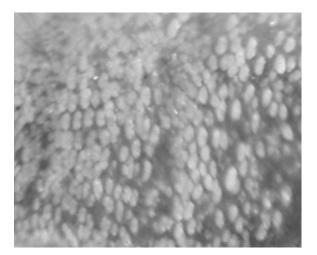


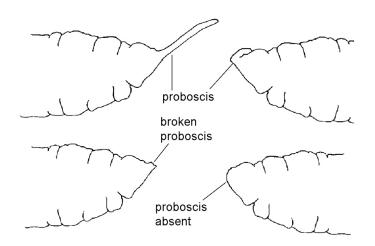
Chaetae present from II; ventral mouth-pouch absent; mature specimens with ventral chaetae of XI modified as hollow-tipped straight chaetae
 Naididae: Tubificinae: Antipodrilus n. sp. GABS4 (known only from Great Artesian Basin mound springs)

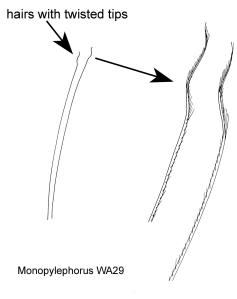


- Proboscis absent; dorsal chaetae (other than hairs) of similar size to ventral chaetae ..... Naididae (in part) 13
- 10. Proboscis absent; body densely covered by papillae with adhering foreign matter (below.

Naididae: Tubificinae: *Embolocephalus yamaguchii* (known only from Japan and South Australia)



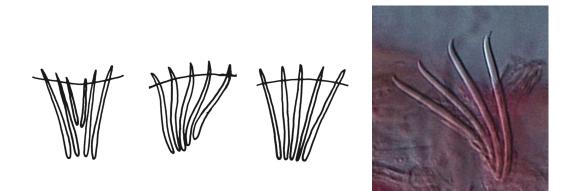




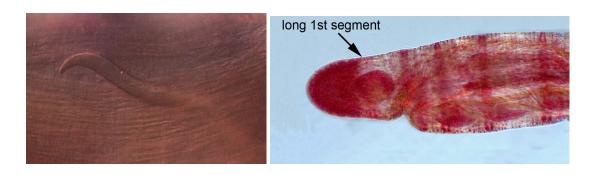




- Usually fewer than 6 chaetae per bundle; clitellum on XI and XII; spermathecae in V; ventral chaetae of IX or X not modified; widespread ...... Enchytraeidae (in part)







- Not as above ..... Other Haplotaxidae

#### **KEYS TO THE AUSTRALIAN NAIDINAE**

#### Characters and states used in the interactive key to Naidinae

First segment with dorsal chaetae

segment II ( = first segment with ventral chaetae) segment III segment IV segment V segment VI segment VII or more posterior

Form of single-pointed needles (no teeth present at tip) evenly tapering to a sharp point tapering but with a minutely swollen tip tapering to very fine whip-like tips with a blunt tip

Gross form of needle chaetae

with a bifid tip (forked, teeth sometimes minute) with a flared or spatulate tip with a pectinate tip (forked with fine intermediate teeth) with a single-pointed tip (sometimes with a swelling at the very tip)

Hair chaetae present or absent absent present

Needle chaetae fan-shaped or spatulate Needle chaetae with a fan-shaped tip Needle chaetae with a spatulate tip

Relative length of hair chaetae in some anterior segments Hairs absent in III or not much longer than those of other segments Some hairs of segmwent III much longer (up to 3x) than those of other segments (but these often missing) Some hairs of VI much longer than those of other segments

- Relative length of teeth on bifid and pectinate needles with teeth approximately equal in length and width with teeth unequal in length and width with teeth unequal in length but not greatly different in width
- type of non-hair dorsal chaetae needle chaetae, not of same general form as ventral chaetae, usually much smaller non-needle chaetae, of same general form as ventral chaetae
- Length of needle chaetae in anterior segments Not yet included in key
- Number of hair chaetae in anterior dorsal bundles Not yet included in key

Uniformity of ventral chaetae

Anterior-most chaetae (usually II to V or VI) clearly longer and sometimes a little thicker, but not greatly enlarged some chaetae in one or more segments from IV to X greatly enlarged Ventral chaetae all of about the same size Ventral chaetae of II to IV slightly shorter than the rest Presence and position of nodulus on sharply single-pointed needles Nodulus absent, needle shaft smoothly tapering Nodulus median Nodulus distal

Ventral chaetae in segment II

4-8 per bundle, less than 110 microns long, teeth curved5-20 per bundle, teeth long and strongly recurved6-13 per bundle, more than 140 microns long, teeth curved

U:L ratio of ventral chaetae of posterior chaetae Not yet included in key

U:L ratio of ventral chaetae of II-IV Not yet included in key

Divergence of teeth on bifid and pectinate needles Teeth almost parallel Teeth strongly divergent Teeth moderately divergent

Dorsal chaetae present/absent absent present

Serrations on hair chaetae present absent - hairs smooth

Length of longest tooth on bifid and pectinate needles Not yet included in key

Body wall papillae present, with adhered foreign matter absent

#### Symbiosis

endosymbiotic in ureters of frogs not symbiotic ectocommensal on snails (may be found free living)

Palps on posterior edge of gill chamber present absent

#### Gills

absent knob or finger-like within a chamber (fossa) on anal segment only long dorso-lateral gills on at least anterior segments, enclosing many dorsal chaetae

Number of pairs of gills on anal segment

5

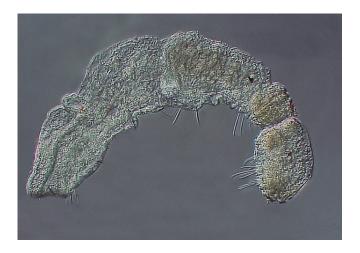
Eyes on peristomium (segment I) present

#### absent

Proboscis on prostomium present absent Raised ridge on body wall of each segment absent present

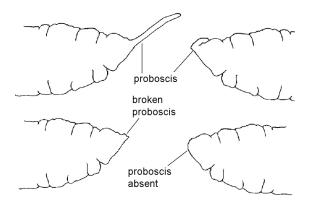
#### Dichotomous key to the Australian Naididae

- Body body short and broad, ventral chaetae of III-V missing; epigean .... Chaetogaster 3





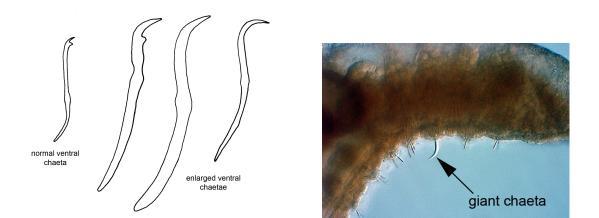
- 4. Ventral chaetae of II 140 to 350 um long ..... Chaetogaster diaphanus
- Ventral chaetae of II 70 to 110 um long ..... Chaetogaster diastrophus
- 5. Proboscis on prostomium (may be partly broken off) ... Stylaria and Pristina (in part) 6



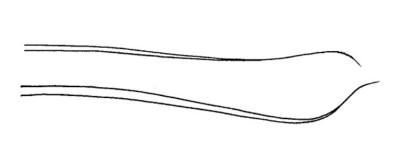




- Dorsal chaetae from II; ventral chaetae smoothly sigmoid ...... Pristina (in part) 7



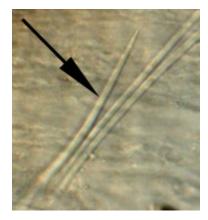




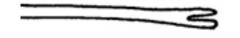




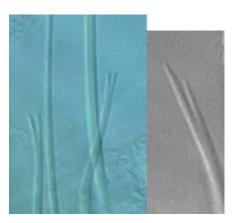
- 10. Dorsal chaetae from II ..... Bratislavia unidentata (in part), Pristina (in part) 11
- Dorsal chaetae from III ..... Bratislava 15
- 11. Needles with single sharp point ..... Bratislavia unidentata



- Needles bifid or pectinate..... Pristina (in part) 12
- 12. Needle chaetae bifid, teeth parallel or only slightly diverging (see below)...... 13
- Needle chaetae pectinate (sometimes bifid), outer teeth strongly diverging ...... 14
- 13. Needle chaetae teeth short (<2 um), parallel and of same length ...... Pristina bilobata



(?*Monopylephorus* WA2 [Rhyacodrilinae] will key out here if assumed to be a Naidinae because of the needle-like dorsal crotchet chaetae). This species has teeth on needles about 7  $\mu$ m but equal in length)

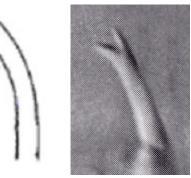


(Rodriguez (2002) Graelliana 58(2): 3-19, provide a good comparison of this species with *P. osborni*)



Photo Rodriguez (2002)

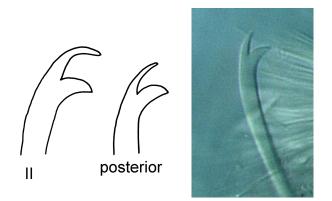
- Needle teeth subequal in length, 1.5 to 2 µm (1 to 3.3 in lit.) and width Pristina osborni



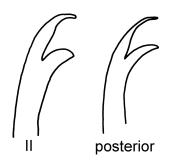
15. Needles with sharp simple tip ...... Bratislavia unidentata

-	Needles bifid	Bratislavia W	A1
16.	Hair chaetae absent	Paranais	17
-	Hair chaetae present		18

17. Ventral chaetae of II with upper teeth longer than lower (below left), ventral chaetae of some other anterior chaetae with upper teeth slightly longer than lower, all other dorsal and ventral chaetae with teeth subequal (below middle and right)...... *Paranais litoralis* 

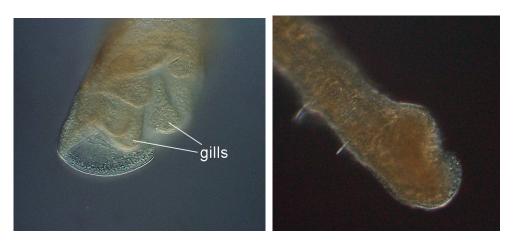


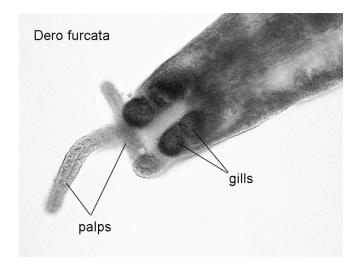
- All ventral chaetae of with upper teeth longer than lower ...... Paranais frici



18. Gills present as paired dorso-lateral processes on each segment from between IV to VII onwards, enclosing some of the hair chaetae ......Branchiodrilus hortensis



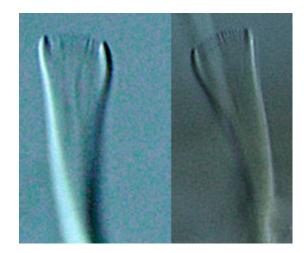




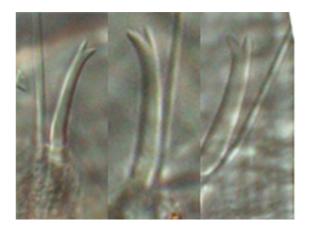




- Needle chaetae more moderately and symetrically expanded Dero (Aulophorus) vagus



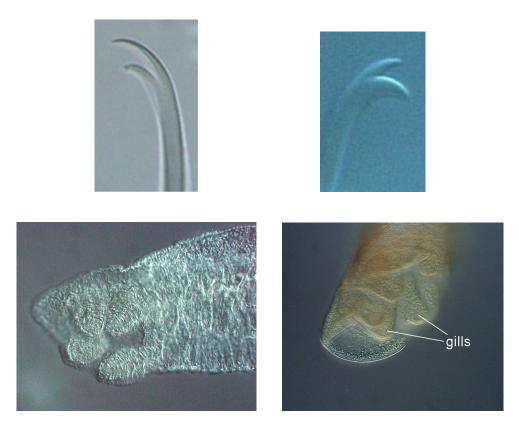
23. With 3 or 4 pairs of long thin gills; dorsal chaetae from V; needles with lower teeth slightly thicker, more curved and longer than upper ...... *Dero* (*Aulophorus*) *furcata* 



- With 3 pairs of short gills; dorsal chaetae from VI; needles with teeth subequal ...... Dero (Aulophorus) WA4

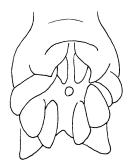


24. Ventral chaetae of first few segments (usually II to VI) longer and thinner than the rest and with teeth long and curved, the upper longer than the lower (below top left); posterior





- Not commensal; collected only from groundwater in the Pilbara; epidermis without raised ridges
   *Dero* (*Allodero*) sp. WA2

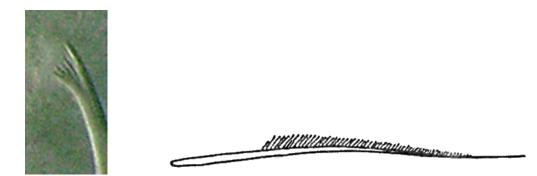


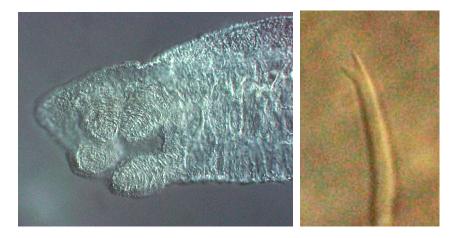
- Dorsal chaetae from V or VI; gill chamber with 2 to 4 pairs of gills ...... 27
- 27. Dorsal chaetae from V, with upper teeth shorter than lower (NOTE: easily confused with D. nivea) ...... Dero (Dero) cf. sawayai

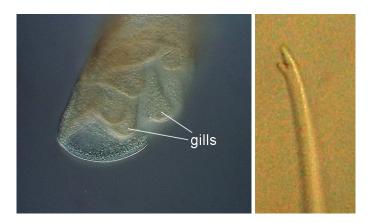


- Dorsal chaetae from VI, with teeth equal or the upper tooth longer ...... 28

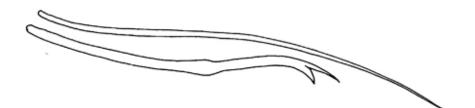
28. Hair chaetae plumose, needle chaetae pectinate ...... Dero (Dero) pectinata





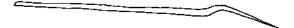


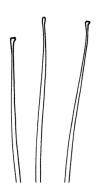
30. Dorsal bifid chaetae similar in size to ventral chaetae, with upper tooth twice as long as lower; dorsal chaetae starting on segments XVI to XXI ...... *Haemonais waldvogeli* 

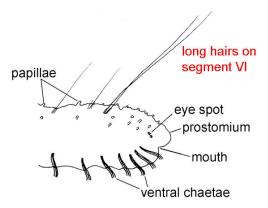


- Dorsal chaetae much finer than the ventral chaetae (dorsal chaetae are 'needle' chaetae) and present from IV to VI
   31



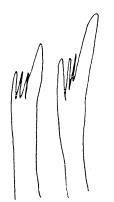








- Needles pectinate with pectinations finer than outer teeth ...... Allonais inequalis



37.	Needle chaetae pectinate	38
-	Needle chaetae bifid	39

38. Needle chaetae with short strongly divergent teeth ...... Allonais pectinata



- Needle chaetae with longer less strongly divergent teeth ...... Allonais ranauana



	<ol> <li>Some anterior ventral chaetae of segments VI onwards gree</li> <li>2 to 3 times longer than the lower</li> </ol>
	All ventral chaetae of normal size
Nais elinguis	0. Ventral chaetae all with upper teeth twice as long as lower
41	Posterior ventral chaetae with teeth subequal





## KNOWN OCCURRENCE OF NAIDINE SPECIES IN AUSTRALIA

	Species	sWA	NSW	VIC	TAS	SA	nWA	NT	QLD
Allonais	inaequalis (Steph.)							+	+
monuis	paraguayensis (Mich.)						+	+	+
	pectinata (Stephenson)						+		+
	ranauana (Mich. et Boldt)					+	+	+	•
Branchiodrilus	hortensis (Stephenson)		+			+	+	+	+
	unidentata (ex sp. NT1)			+		•	+	+	
127000500700	sp. WA1	+							
Chaetogaster	diaphanus (Gruithuisen)	+		+		+	+		
0	diastrophus (Gruithuisen)	+	+	+		+	+		
	<i>limnaei</i> von Baer	+	+	+		+			
Dero	digitata (Müll.)	+	+	+	+	+	+	+	+
	dorsalis Ferronniere								?
	litoria Pinder, Hill et Green								+
	nivea Aiyer	+	+	+	+	+	+	+	+
	? nr. cooperi Steph. (*)							?	
	nr. sawayai Marcus						+		
	pectinata Aiyer						+		+
	flabelliger (Steph.)		+	+		+	+	+	+
	furcata (Müll)	+	+	+		+	+	+	
	vagus (Leidy)						+	+	
	sp. WA1						+		
	sp. WA2						+		
	sp. WA4	+							
Nais	bretscheri Mich.	+		+		+			
	communis Piguet	+	+	+	+	+	+		+
	variabilis Piguet	+	+	+	+	+			+
	elinguis Mull.			+		+			+
	pseudobtusa Piguet								+
	barbata Muller		+						
Paranais	frici Hrabe			+					
	litoralis (Mull.)	+		+	+	+			
Pristina	aequiseta Bourne	+	+	+		+	+	+	+
	longiseta Ehrenberg	+	+	+		+	+	+	+
	leidyi Smith	+					+		
	sima (Marcus)	+					+		
	bilobata (Bretscher)			?					
	proboscidea Beddard	?		?		?	?	?	?
	jenkinae (Steph.)	+		+		+	+		
	osborni (Walton)	+		+		+		+	
	waldvogeli Bretscher		+				+		
Slavina	appendiculata d'Udekem		+	+		+			
	proceriseta Pinder et Brink.			+		+	+		
Stylaria	lacustris (Linnaeus)		+	+		+	+		

Australian Capitol Territory included with NSW and WA split into north (nWA) and south (sWA).

\* = tentatively id from North Qld by Brinkhurst (1971)

? = presence in Australia not confirmed

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