

# Historical changes and inventory of macroalgae from Königshafen Bay in the northern Wadden Sea

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Dedicated to P. Kornmann (†) and P.-H. Sahling

**ABSTRACT:** Knowledge on the distribution, abundance and species richness of intertidal macroalgae occurring on sandy and muddy flats of the German Wadden Sea is still incomplete. We summarize published and unpublished information available on the presence of macroalgae on the tidal flats of Königshafen Bay (island of Sylt, North Sea), one of the more extensively studied areas of the Wadden Sea. A total of 46 green algal species, 36 brown algal species and 26 red algal species has been recorded within the last 120 years on soft and hard substrata of Königshafen Bay (disregarding species found unattached or drifting). Several of these species were only temporarily resident on the tidal flats. Today, at least 35 green, 15 brown and 12 red algal species occur within or close to Königshafen Bay. Significant long-term changes in species abundances have occurred in all three major groups of algae: Since the late 1970s, dense green algal mats dominated by *Enteromorpha flexuosa*, *E. radiata* and *E. prolifera* have occurred regularly on the intertidal flats, whereas a general decrease of brown and red algae has been documented. Two red algal species, *Gracilaria verrucosa* and its epiphyte *Callithamnion corymbosum*, were conspicuous members of the macroflora until the middle of this century. Although still present in the 1980s, they have now disappeared completely. On the other hand, the brown alga *Sargassum muticum* has begun to colonize mussel beds. The causal background of long-term changes in the macroalgal flora of Königshafen Bay is discussed. Owing to substantial nomenclatural changes during the last 120 years, a revised species list with authors' names and synonyms is included.

## INTRODUCTION

Towards the turn of last century, Reinke (1889a, b, 1892), Kuckuck (1897, 1900, 1902) and several other botanists (see Mollenhauer & Lüning, 1988) investigated the algal flora of the German Baltic Sea and the island of Helgoland. They all showed comparatively little interest in the rare macroalgal vegetation of the intertidal mud- and sandflats of the Wadden Sea, an ecosystem which appeared as "a unique plantless desert continuing in the subtidal" (modified from Reinke, 1889c). Therefore, it is not surprising that the first information on Wadden Sea macroalgae was obtained as a mere byproduct of the North Sea Pommerania-expedition in 1872 (Magnus, 1874).

In contrast, the intertidal flats of Königshafen Bay are the only Wadden Sea areas in Germany where marine botanists have worked repeatedly, although not continually. Ac-

cordingly, somewhat more detailed descriptions of the macroalgal vegetation and its changes over time are available from this region.

Magnus (1874) and Reinbold (1892) both spent only a few days in Königshafen Bay, giving incomplete accounts of the algal species present in the area. Kuckuck was the first marine botanist to undertake several excursions to the intertidal flats of the North Frisian Wadden Sea. Unfortunately, his collection of herbarium material and most of his unpublished botanical diaries were lost in the Second World War. Only a diary about the Wadden Sea, written between 1896 and 1903, still exists in the Department of Marine Botany of the Biologische Anstalt Helgoland (hereafter BAH). It contains the earliest detailed recordings of the algal vegetation at the island of Sylt with special emphasis on Königshafen Bay.

More than 20 years later, Nienburg (1927) investigated the upper eulittoral region of Königshafen. He concentrated on the development of the *Enteromorpha*-zone and the presence of *Fucus vesiculosus* f. *mytili* on mussel beds, but he did not produce a complete check-list of macroalgae. Except for Hagmeier's (1941) description of the macroflora associated with the North Frisian oyster beds, there were no botanical activities in Königshafen Bay after Nienburg until the 1950s.

As a consequence of the destruction of the BAH on Helgoland in the Second World War, the botanist Kornmann and his assistant Sahling worked on the island of Sylt from 1946 to 1958 where they studied the macroalgae of Königshafen Bay in detail. Although only a minor portion of this work was published by Kornmann at that time (1952), he made available to us many unpublished data. With the reconstruction of the BAH on Helgoland and the subsequent departure of Kornmann and Sahling from the island of Sylt, all botanical activities ended at once, so that there were no more detailed investigations of macroalgae in Königshafen Bay for several decades. In the late 1970s mass development of ephemeral green algae (Reise, 1983) indicated dramatic changes in the macrophytobenthos (Reise, 1994) and initiated recent phycological studies.

We reinvestigated the macrophytobenthos of Königshafen Bay with particular reference to historical changes. The species lists presented here summarize published and unpublished data. They are a preliminary approach towards an estimate of the macroalgal diversity of Königshafen Bay and adjacent Wadden Sea areas and the lists do not claim to be comprehensive. We offer information about the potential background for changes over time, as well as some details on the historical changes in single species' occurrences.

Ecological data on factors influencing abundance and distribution of green algae in Königshafen Bay are given by Schories & Reise (1993), Lotze (1994) and Schories (1995 a, b), whereas Albrecht (1995) worked on macroalgal communities associated with mussel beds (*Mytilus edulis*).

## STUDY AREA

Königshafen Bay is a shallow tidal bay at the northern tip of the island of Sylt (North Sea, Germany) (Fig. 1). The hydrography and macrofauna of the area have been described by Wohlenberg (1937), Reise (1985) and Reise et al. (1994a). Historical changes in the biological structural entities (e.g. mussel beds, seagrass meadows and green algal mats) have been summarized by Reise et al. (1989). A sedimentological study of the area was

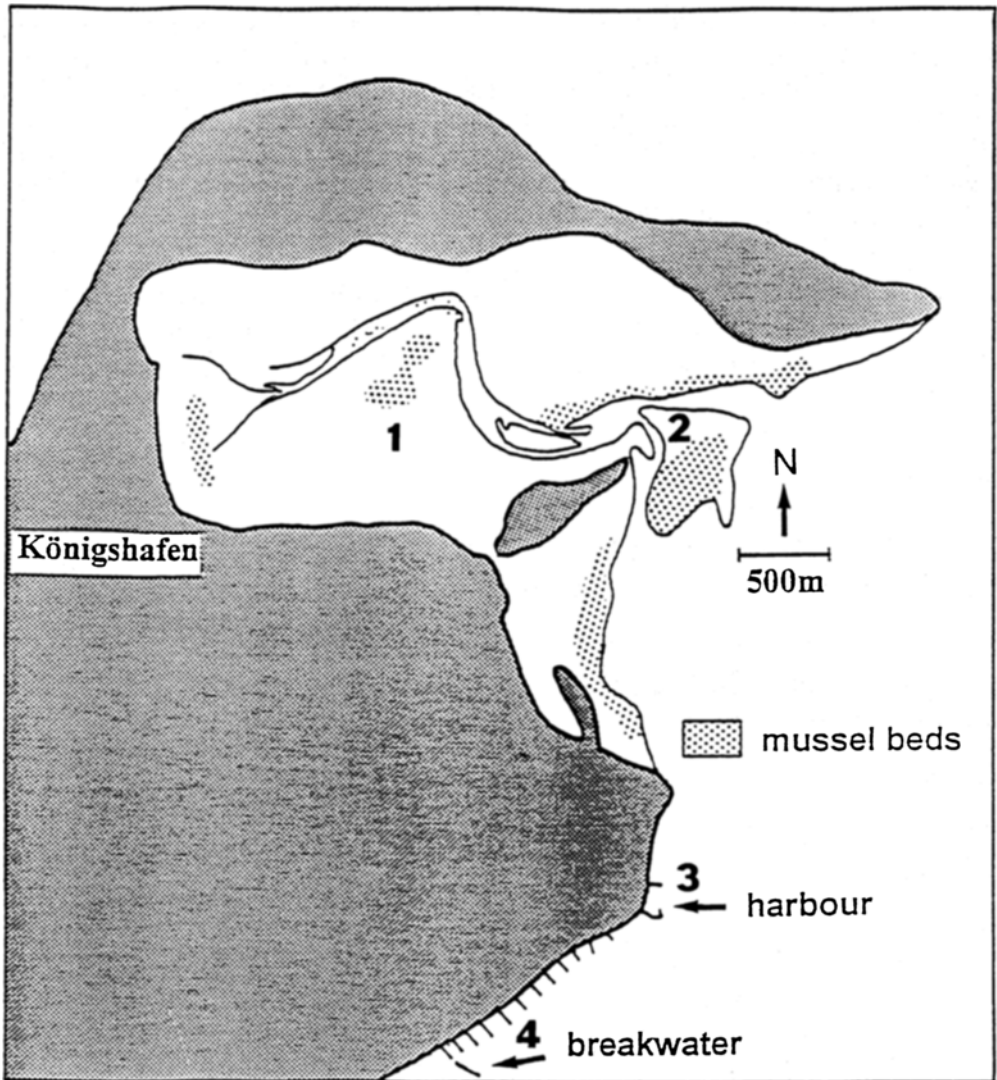


Fig. 1. Diagram of the Wadden Sea area near the island of Sylt as found at present, showing inner Königshafen Bay (1), outer Königshafen Bay (2), List harbour (3) and a stony breakwater, south of the village of List (4)

carried out by Austen (1992). The aeolic input of dune-sands from the west and the formation of a sand spit system in the south of the bay have caused substantial morphological changes of Königshafen Bay through to the present (Bayerl & Higelke, 1994). Austen (1994) showed that surface sediments have changed over the last 50 years, resulting in an areal decline in mudflats and an increasing consolidation of the mud in the period up to 1981.

## MATERIALS, METHODS AND NOMENCLATURE

Since the binomials of many species have been subject to major changes, a revised list of the species with authors' names and synonyms from the reference material is included (Table 1). In most cases, it was not possible to countercheck the taxonomic information given in the older literature because of missing herbarium material. This might have led to some errors in genera as *Cladophora* or *Enteromorpha*, in which detailed

Table 1. List of species with authors, and synonyms. – Within this century, many changes have been made to the Latin binomials. This is a revised species list with authors' names and synonyms used in the cited literature (see Table 2) and in the unpublished data material. The names are arranged alphabetically within the classes: Rhodophyceae, Fucophyceae (Phaeophyceae) and Chlorophyceae, with the valid names printed in **bold italic letters** and synonyms in *italics*. For more detailed information about further synonyms used in earlier literature see Nielsen et al. (1995) and Pankow (1971, 1990)

RHODOPHYCEAE	
<i>Acrochaetium secundatum</i> (Lyngb.) Dixon	<i>Chantransia secundata</i> (Lyngb.) Thuret in Le Jolis
<i>Antithamnion plumula</i>	see <b><i>Pterothamnion plumula</i></b>
<b><i>Bangia atropurpurea</i></b> (Roth) C. A. Ag.	<i>Bangia fuscopurpurea</i> (Dillw.) Lyngb.
<i>Bangia fuscopurpurea</i>	see <b><i>Bangia atropurpurea</i></b>
<b><i>Callithamnion corymbosum</i></b> (Smith in Engl. Bot.) Lyngb.	
<b><i>Ceramium nodulosum</i></b> (Lightf.) Ducluz.	<i>Ceramium rubrum</i> (Huds.) C. A. Ag.
<i>Ceramium rubrum</i>	see <b><i>Ceramium nodulosum</i></b>
<i>Chantransia daviesii</i>	see <b><i>Colaconema daviesii</i></b>
<i>Chantransia secundata</i>	see <b><i>Acrochaetium secundatum</i></b>
<i>Chantransia thuretii</i>	see <b><i>Colaconema nemalionis</i></b>
<b><i>Chondrus crispus</i></b> Stackh.	
<b><i>Colaconema daviesii</i></b> (Dillw.) Stegenga	<i>Chantransia daviesii</i> (Dillw.) Thuret
<b><i>Colaconema nemalionis</i></b> (De Not. ex Dufour)	<i>Chantransia thuretii</i> (Bornet) Kylin
<b><i>Dumontia contorta</i></b> (Gmel.) Rupr.	<i>Dumontia filiformis</i> (O. F. Müller in Flor. dan.) Grev.
	<i>Dumontia incrassata</i> (O. F. Müller in Flor. dan.) Lamour.
<i>Dumontia filiformis</i>	see <b><i>Dumontia contorta</i></b>
<i>Dumontia incrassata</i>	see <b><i>Dumontia contorta</i></b>
<b><i>Erythrotrichia carnea</i></b> (Dillw.)	<i>Erythrotrichia ceramicola</i> (Lyngb.) Aresch.
<i>Erythrotrichia ceramicola</i>	see <b><i>Erythrotrichia carnea</i></b>
<b><i>Erythrotrichia reflexa</i></b> (Crouan et Crouan) De Toni sensu Rosenv.	see <b><i>Stylonema alsidii</i></b>
<i>Goniotrichum elegans</i>	see <b><i>Gracilaria verrucosa</i></b>
<i>Gracilaria confervoides</i>	<i>Gracilaria confervoides</i> (L.) Grev.
<b><i>Gracilaria verrucosa</i></b> (Huds.) Papenf.	see <b><i>Hildenbrandia rubra</i></b>
<i>Hildenbrandia prototypus</i>	<i>Hildenbrandia rosea</i> Kütz.
<b><i>Hildenbrandia rubra</i></b> (Sommerf.) Menegh.	<i>Hildenbrandia prototypus</i> Nardo
<i>Hildenbrandia rosea</i>	see <b><i>Hildenbrandia rubra</i></b>
<b><i>Lithothamnion sonderi</i></b> Hauck	
<i>Melobesia lejolisii</i>	see <b><i>Pneophyllum lejolisii</i></b>
<b><i>Melobesia membranacea</i></b> Lamour. non Esper	
<b><i>Nemalion multifidum</i></b> (Weber & Mohr) Endl.	

Table 1 (Continued)

<i>Pneophyllum lejolisii</i> (Rosanoff) Chamberlain	<i>Melobesia lejolisii</i> Rosanoff
<i>Polysiphonia atrorubescens</i>	see <i>Polysiphonia nigra</i>
<i>Polysiphonia elongata</i> (Huds.) Sprengel	
<i>Polysiphonia fibrillosa</i> (Dillw.) Sprengel	<i>Polysiphonia violacea</i> (Roth) Grev. in Hooker
<i>Polysiphonia fucoides</i> (Huds.) Grev.	<i>Polysiphonia nigrescens</i> (Huds.) Griff. ex Harv.
<i>Polysiphonia nigra</i> (Huds.) Batters	<i>Polysiphonia atrorubescens</i> (Dillw.) Grev.
<i>Polysiphonia nigrescens</i>	see <i>Polysiphonia fucoides</i>
<i>Polysiphonia stricta</i> (Dillw.) Grev.	<i>Polysiphonia urceolata</i> (Lightf. ex Dillw.) Grev.
<i>Polysiphonia urceolata</i>	see <i>Polysiphonia stricta</i>
<i>Polysiphonia violacea</i>	see <i>Polysiphonia fibrillosa</i>
<i>Porphyra laciniata</i>	see <i>Porphyra umbilicalis</i>
<i>Porphyra umbilicalis</i> (L.) Kütz.	<i>Porphyra laciniata</i> (Lightf. 1792) J. G. Ag. 1882/83
<i>Pterothamnion plumula</i> (Ellis) Näg. in Näg. et Cramer	<i>Antithamnion plumula</i> (Ellis) Thuret in Le Jolis
<i>Rhodomela confervoides</i> (Huds.) Silva	<i>Rhodomela subfusca</i> (Woodw.) C. A. Ag.
<i>Rhodomela subfusca</i>	see <i>Rhodomela confervoides</i>
<i>Stylonema alsidii</i> (Zanard.) Drew	<i>Goniotrichum elegans</i> (Chauv.) Le Jolis
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FUCOPHYCEAE (PHAEOPHYCEAE)	
<i>Acinetospora crinita</i> (Carm. ex Harvey) Kornmann	
<i>Ascocyclus orbicularis</i>	see <i>Myrionema magnusii</i>
<i>Botrytella micromora</i> Bory	<i>Sorocarpus uvaeformis</i> Pringsh.
<i>Chorda filum</i> (L.) Stackh.	
<i>Chordaria flagelliformis</i> (O. F. Müller in Flor. dan.) C. A. Ag.	
<i>Desmotrichum balticum</i> Kütz.	
<i>Desmotrichum undulatum</i>	see <i>Punctaria tenuissima</i>
<i>Dictyosiphon foeniculaceus</i> (Huds.) Grev.	
<i>Dictyota dichotoma</i> (Huds.) Lamour.	
<i>Dumontia filiformis</i>	see <i>Dictyosiphon chordaria</i>
<i>Ectocarpus confervoides</i>	see <i>Ectocarpus siliculosus</i>
<i>Ectocarpus dasycarpus</i>	see <i>Ectocarpus siliculosus</i>
<i>Ectocarpus fuscatus</i>	see <i>Hincksia ovata</i>
<i>Ectocarpus granulatus</i>	see <i>Hincksia granulosa</i>
<i>Ectocarpus sandrianus</i>	see <i>Hincksia sandriana</i>
<i>Ectocarpus siliculosus</i> (Dillw.) Lyngb.	<i>Ectocarpus confervoides</i> (Roth) Kjellm.
	<i>Ectocarpus dasycarpus</i> Kuckuck
	see <i>Spongonema tomentosum</i>
<i>Ectocarpus tomentosus</i>	
<i>Elachista fucicola</i> (Vellay) Aresch.	
<i>Fucus mytili</i>	see <i>Fucus vesiculosus</i> f. <i>mytili</i>
<i>Fucus platycarpus</i>	see <i>Fucus spiralis</i> var. <i>platycarpus</i>
<i>Fucus spiralis</i> L. var. <i>platycarpus</i> (Thur.) Batt.	<i>Fucus platycarpus</i> Thur.
<i>Fucus vesiculosus</i> L. f. <i>mytili</i> (Nienburg) Nienhuis	
<i>Fucus vesiculosus</i> L.	
<i>Hincksia granulosa</i> (Smith) Silva	<i>Ectocarpus granulatus</i> (Smith) C. A. Ag.
<i>Hincksia ovata</i> (Kjellm.) Silva	<i>Ectocarpus fuscatus</i> Zanard. ex Menegh.
<i>Hincksia sandriana</i> (Zanard.) Silva	<i>Ectocarpus sandrianus</i> Zanard.
<i>Laminaria digitata</i> (Huds.) Lamour.	

Table 1 (Continued)

<b>Laminaria saccharina</b> (L.) Lamour.	
<b>Leathesia difformis</b> (L.) Aresch.	
<i>Leptonema fasciculatum</i>	see <b>Leptonematella fasciculata</b>
<b>Leptonematella fasciculata</b> (Reinke) Silva	<i>Leptonema fasciculatum</i> Reinke
<i>Mesogloia divaricata</i>	see <b>Sphaerotrichia divaricata</b>
<b>Microsyphar porphyrae</b> Kuckuck	
<b>Myrionema magnusii</b> (Sauv.) Lois.	<i>Ascocyclus orbicularis</i> (J. G. Ag.) De Toni
<b>Myrionema strangulans</b> Carm. mscr. in Grev.	<i>Myrionema vulgare</i> Thuret in Le Jolis
<i>Myrionema vulgare</i>	see <b>Myrionema strangulans</b>
<b>Myriotrichia clavaeformis</b> Harv.	<i>Myriotrichia repens</i> Hauck
<i>Myriotrichia repens</i>	see <b>Myriotrichia clavaeformis</b>
<b>Petalonia fascia</b> (Müller in Flor. dan.) Kuntze	<i>Phyllitis fascia</i> (Müller in Flor. dan.) Kütz.
<b>Petalonia zosterifolia</b> (Reinke) Hamel	<i>Phyllitis zosterifolia</i> (Reinke) Kuntze
<i>Phyllitis fascia</i>	see <b>Petalonia fascia</b>
<i>Phyllitis zosterifolia</i>	see <b>Petalonia zosterifolia</b>
<b>Pilayella littoralis</b> (L.) Kjellm.	
<b>Punctaria latifolia</b> Grev.	
<b>Punctaria plantaginea</b> (Roth) Grev.	
<b>Punctaria tenuissima</b> (J. G. Ag.) Grev.	<i>Desmotrichum undulatum</i> (J. G. Ag.) Reinke
<b>Ralfsia verrucosa</b> (Aresch.) J. G. Ag.	
<b>Sargassum muticum</b> (Yendo) Fensholt	
<b>Scytosiphon lomentaria</b> (Lyngb.) Endl.	
<i>Sorocarpus uvaeformis</i>	see <b>Botrytella micromora</b>
<b>Sphacelaria cirrosa</b> (Roth) C. A. Ag.	
<b>Sphaerotrichia divaricata</b> (C. A. Ag.) Kylin	<i>Mesogloia divaricata</i> (C. A. Ag.) Kütz.
<b>Spongonema tomentosus</b> (Huds.) Kütz.	<i>Ectocarpus tomentosus</i> (Huds.) Lyngb.
<i>Stictyosiphon</i> sp.	
<b>Ulonema rhizophorum</b> (Foslie) Sauvag.	
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<b>CHLOROPHYCEAE</b>	
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<b>Acrochaete repens</b> Pringsh.	
<b>Acrochaete wittrockii</b> (Wille) Nielsen	<i>Endoderma wittrockii</i> (Wille) Lagerh.
	<i>Ectochaete wittrockii</i> (Wille) Kylin
<b>Blidingia marginata</b> (J. G. Ag.) Dang.	
<b>Blidingia minima</b> (Nägeli ex Kütz.) Kylin	<i>Enteromorpha minima</i> Nägeli ex Kütz.
<i>Bryopsis hypnoides</i>	see <b>Bryopsis plumosa</b>
<b>Bryopsis plumosa</b> (Huds.) C. A. Ag.	<i>Bryopsis hypnoides</i> Lamour.
<b>Capsosiphon fulvescens</b> (C. A. Ag.) Setch. et Gardner	
<b>Chaetomorpha aerea</b> (Gooden ex Dillw.) Kütz.	
<b>Chaetomorpha linum</b> (O. F. Müller in Flor. dan.) Kütz.	
<b>Chaetomorpha sutoria</b> (Berk.) Kormmann	
<i>Cladophora glaucescens</i>	see <b>Cladophora sericea</b>
<i>Cladophora flexuosa</i>	see <b>Cladophora sericea</b>
<b>Cladophora fracta</b> (O. F. Müller ex Vahl in Flor. dan.) Kütz.	
<i>Cladophora hirta</i>	see <b>Cladophora sericea</b>
<b>Cladophora hutchinsiae</b> (Dillw.) Kütz.	
<i>Cladophora lanosa</i>	see <b>Spongomorpha aeruginosa</b>
<b>Cladophora lehmaniana</b> (Lindenberg) Kütz.	<i>Cladophora utriculosa</i> Kütz.
<b>Cladophora sericea</b> (Huds.) Kütz. sensu van den Hoek	
	<i>Cladophora glaucescens</i> (Giff. ex. Harv.) Harv.

Table 1 (Continued)

<i>Cladophora utriculosa</i>	<i>Cladophora hirta</i> Müller ex Vahl
<b><i>Codium fragile</i></b> (Suringar) Hariot	<i>Cladophora flexuosa</i> (Griff.) Harv.
<b><i>Codium tomentosum</i></b> Stackh.	see <b><i>Cladophora lehmaniana</i></b>
<i>Diplonema percursum</i>	see <b><i>Percursaria percursa</i></b>
<i>Ectochaete wittrockii</i>	see <b><i>Acrochaete wittrockii</i></b>
<i>Endoderma wittrockii</i>	see <b><i>Acrochaete wittrockii</i></b>
<b><i>Enteromorpha ahlneri</i></b> Bliding	
<b><i>Enteromorpha clathrata</i></b> (Roth) Grev.	
<b><i>Enteromorpha compressa</i></b> (L.) Grev.	
<b><i>Enteromorpha crinita</i></b> J. G. Ag.	
<b><i>Enteromorpha flexuosa</i></b> (Wulfen ex Roth) J. G. Ag.	<i>Enteromorpha lingulata</i> J. G. Ag.
	<i>Enteromorpha tubulosa</i> Kütz.
<b><i>Enteromorpha flexuosa</i></b> (Wulfen) J. G. Ag. <b>subsp. paradoxa</b> (C. A. Ag.) Bliding	<i>Enteromorpha plumosa</i> Kütz.
	<i>Enteromorpha paradoxa</i> (Dillw.) Kütz.
<b><i>Enteromorpha intestinalis</i></b> (L.) Link	
<i>Enteromorpha lingulata</i>	see <b><i>Enteromorpha flexuosa</i></b>
<b><i>Enteromorpha linza</i></b> (L.) J. G. Ag.	
<b><i>Enteromorpha linziformis</i></b> Bliding	
<i>Enteromorpha minima</i>	see <b><i>Blidingia minima</i></b>
<i>Enteromorpha paradoxa</i>	see <b><i>Enteromorpha flexuosa</i></b> subsp. <b><i>paradoxa</i></b>
<b><i>Enteromorpha pilifera</i></b> Kütz.	
<i>Enteromorpha plumosa</i>	see <b><i>Enteromorpha flexuosa</i></b> subsp. <b><i>paradoxa</i></b>
<b><i>Enteromorpha prolifera</i></b> (O. F. Müller in Flor. dan.) J. G. Ag.	
<b><i>Enteromorpha radiata</i></b> J. G. Ag.	
<b><i>Enteromorpha ralfsii</i></b> Harvey	
<b><i>Enteromorpha ramulosa</i></b> (J. E. Smith in Engl. Bot.) Hooker	
<b><i>Enteromorpha simplex</i></b> (Vinogradova) Koeman	
<b><i>Enteromorpha torta</i></b> Reinb.	
<i>Enteromorpha tubulosa</i>	see <b><i>Enteromorpha flexuosa</i></b>
<b><i>Epicladia perforans</i></b> (Huber) Nielsen	
<b><i>Eugomontia sacculata</i></b> Kornmann	
<b><i>Gomontia polyrhiza</i></b> (Lagerh.) Bornet et Flahault	
<i>Monostroma grevillei</i>	see <b><i>Ulvopsis grevillei</i></b>
<b><i>Percursaria percursa</i></b> (C. A. Ag.) Bory in Dup.	<i>Diplonema percursum</i> (C. A. Ag.) Kjellm.
<b><i>Prasiola stipitata</i></b> Suhr in Jessen	
<i>Rhizoclonium implexum</i>	see <b><i>Rhizoclonium riparium</i></b>
<i>Rhizoclonium kochianum</i>	see <b><i>Rhizoclonium riparium</i></b>
<b><i>Rhizoclonium riparium</i></b> (Roth) Harvey	<i>Rhizoclonium implexum</i> (Dillw.) Kütz.
	<i>Rhizoclonium kochianum</i> Kütz.
<b><i>Spongomorpha aeruginosa</i></b> (L.) van den Hoek	<i>Cladophora lanosa</i> (Roth) Kütz.
<b><i>Ulothrix flacca</i></b> (Dillw.) Thuret in Le Jolis	
<b><i>Ulothrix implexa</i></b> (Kütz.) Kütz.	
<b><i>Ulva lactuca</i></b> L.	<i>Ulva latissima</i> Harv.
<i>Ulva latissima</i>	see <b><i>Ulva lactuca</i></b>
<b><i>Ulva rigida</i></b> C. A. Ag.	
<b><i>Ulva scandinavica</i></b> Bliding	
<b><i>Ulva tenera</i></b> Kornmann	
<b><i>Ulvopsis grevillei</i></b> (Thuret) Gayral	<i>Monostroma grevillei</i> (Thuret) Wittr.
<b><i>Urospora penicilliformis</i></b> (Roth) Aresch.	

taxonomic studies were first carried out in the 1960s (Bliding, 1963; Söderström, 1963; van den Hoek, 1963). Abundant herbarium material and slides exist from the period 1946 to 1958, when Kornmann and Sahling worked at the Wadden Sea Station of the BAH. Macroalgae which were encountered only as drifting specimens are not considered here. Data sources, both published literature and unpublished data integrated in this study, are given in Table 2 a.

Our own investigations, carried out between 1990 and 1995, were largely restricted to Königshafen Bay, although all available information from adjacent areas has been added to the check-list. SCUBA diving was used to determine the lower depth limit of the macroalgal vegetation growing on artificial hard substrata in direct vicinity of Königshafen Bay (i.e. on the walls of List harbour).

## RESULTS

### Historical changes

A total of 46 green algal species, 36 brown algal species and 26 red algal species has been recorded on the soft and hard substrata of Königshafen Bay or in adjacent areas over the last 120 years (Table 2 b). However, several of these species have been detected only infrequently on the tidal flats. For example, there are only single recordings of *Dictyota dichotoma* and *Codium tomentosum* (Hagmeier, 1941).

Table 2 a. Literature and unpublished data used for the survey, and explanations of the symbols

<p><b>Origin of data:</b></p> <ol style="list-style-type: none"> <li>1 Magnus (1874)</li> <li>2 Reinbold (1892)</li> <li>3 Kuckuck (1896-1903, unpubl. data)</li> <li>4 Nienburg (1927)</li> <li>5 Hagmeier (1941)</li> <li>6 Kornmann (1952)</li> <li>7 Kornmann (1953)</li> <li>8 Kornmann (1956)</li> <li>9 Kornmann (1959)</li> <li>10 Kornmann (1960)</li> <li>11 Kornmann (1972)</li> <li>12 Kornmann &amp; Sahling (1946-1958, unpubl. data)</li> <li>13 Kessler (1976-1980, unpubl. data)</li> <li>14 Lotze (1994)</li> <li>15 Kornmann &amp; Sahling (1994)</li> <li>16 Schories &amp; Albrecht (1995)</li> <li>17 this study</li> </ol> <p><b>Symbols:</b></p> <p><b>K</b> Königshafen Bay</p> <p><b>O</b> Oyster beds around the island of Sylt</p> <p><b>A</b> Intertidal and low subtidal areas in the surroundings of Königshafen Bay (including anthropogenic hard substrata such as dikes and walls)</p> <p><b>B</b> Beach region of the village of List</p>
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Table 2 b. Check-list of multicellular benthic macroalgae settling in Königshafen Bay or in adjacent tidal areas during the last 120 years. Due to taxonomic difficulties individuals were not always determined to species level; therefore, individuals marked as "sp" or as "spp" do not necessarily represent additions to the listed members of the same genus. The taxonomic classification of the *Ulva*-, the *Enteromorpha*- and the *Cladophora*-group is uncertain in all data material before 1990. The *Porphyra*-group was revised by Kornmann & Sahling (1992), but this could not be taken into consideration in this table. Recently M. Steentoft (unpubl.) separated individuals of *Gracilaria verrucosa* (Huds.) Papenf. found in Königshafen Bay by Kornmann & Sahling [No. 12 in Table 2 a] and Kessler [No. 13] into the species *Gracilaria gracilis* (Stackh.) Steentoft, L. Irvine & Farnham and *Gracilariopsis longissima* (S. G. Gmelin) Steentoft, L. Irvine & Farnham (see Steentoft et al. [1995] for detailed information). However, we maintain the name *Gracilaria verrucosa* (Huds.) Papenf. in this study.

	1872-1927	1932-1958	1976-1980	1990-1994
<b>RHODOPHYCEAE</b>				
<i>Acrochaetium secundatum</i>	3 K	6 A	-	-
<i>Bangia atropurpurea</i>	3 A	6 A	-	-
<i>Callithamnion corymbosum</i>	3 K	6 K; 12 A, K	13 K	-
<i>Ceramium nodulosum</i>	1B; 3A, K; 4 K	5 O; 6 K; 7 A; 12 A, K	13 A, K	17 A, K
<i>Chondrus crispus</i>	3 A	6 K; 12 A, K	13 K	17 A, K
<i>Colaconema</i> sp.	3 A	-	6 A	-
<i>Colaconema daviesii</i>	-	12 A	-	17 K
<i>Colaconema nemalionis</i>	-	12 K	-	-
<i>Dumontia contorta</i>	3 A, K	6 K; 12 A, K	13 K	17 A, K
<i>Erythrotrichia carnea</i>	1 B	6 A, K; 12 A, K	-	17 K
<i>Erythrotrichia reflexa</i>	-	12 K	-	17 K
<i>Gracilaria verrucosa</i>	1 B; 3 K	5 O; 6 K; 12 A, K	13 K	-
<i>Hildenbrandia rubra</i>	1 B; 3 A	12 K	-	-
<i>Lithothamnion</i> sp.	-	5 O, 6 K	-	-
<i>Lithothamnion sonderi</i>	3 A	12 A, K	-	-
<i>Melobesia membranacea</i>	1 B	-	-	-
<i>Nemalion multifidum</i>	3 A	6 A; 12 A	-	-
<i>Pneophyllum lejolisii</i>	4 K	-	-	-
<i>Polysiphonia elongata</i>	2 A; 3 A, K	5 O; 6 K; 7 A; 12 A, K	13 K	7 A, K
<i>Polysiphonia fibrillosa</i>	3 A, K; 4 K	6 A; 12 A, K	13 K	17 K
<i>Polysiphonia fucoides</i>	1 B; 3 A, K	5 O; 6 A, K; 7 A; 12 A, K	13 A, K	17 A, K
<i>Polysiphonia nigra</i>	3 K; 4 K	6 A, K; 7 A; 12 A, K	13 K	-
<i>Polysiphonia stricta</i>	4 K	6 A	-	-
<i>Porphyra</i> spp.	-	12 A, K	13 A, K	17 A, K
<i>Porphyra umbilicalis</i>	3 K	6 A, K; 12 A, K	-	17 A, K
<i>Pterothamnion plumula</i>	-	-	13 K	-
<i>Rhodomela confervoides</i>	3 A, K	-	-	-
<i>Stylonema alsidii</i>	3 A	6 K; 12 A, K	-	17 K
<b>FUCOPHYCEAE (PHAEOPHYCEAE)</b>				
<i>Acinetospora crinita</i>	-	7 A; 12 K	-	-
<i>Botrytella micromora</i>	-	12 A	-	-
<i>Chorda filum</i>	1 B; 3 A	5 O; 6 A; 12 A, K	-	17 K
<i>Chordaria flagelliformis</i>	3 A	6 A	-	-
<i>Desmotrichum balticum</i>	-	6 A; 12 K	-	-
<i>Dictyosiphon foeniculaceus</i>	3 A	-	-	-
<i>Dictyota dichotoma</i>	-	5 O	-	-
<i>Ectocarpus siliculosus</i>	3 A, K; 4 K	6 A; 7 A; 12 A, K	13 K	17 A, K

Table 2 b (Continued)

	1872-1927	1932-1958	1976-1980	1990-1994
<i>Elachista fucicola</i>	3 K	6 K; 12 A, K	13 K	17 A, K
<i>Fucus spiralis</i> var. <i>platycarpus</i>	-	6 A; 12 A	-	17 A, K
<i>Fucus vesiculosus</i> f. <i>mytili</i>	4 K	5 O; 6 K; 12 A, K	13 K	17 A, K
<i>Fucus vesiculosus</i>	1 B; 3 A, K, 4 K	6 A; 12 A	-	17 A, K
<i>Hincksia granulosa</i>	-	6 A; 7 A; 12 A, K	13 K	17 K
<i>Hincksia ovata</i>	-	12 K	-	-
<i>Hincksia sandriana</i>	-	12 A, K	-	-
<i>Laminaria digitata</i>	-	12 A	-	-
<i>Laminaria saccharina</i>	-	6 A; 12 A	13 A	17 A
<i>Leathesia difformis</i>	3 K	-	-	-
<i>Leptonematella fasciculata</i>	-	12 K	-	-
<i>Microsyphar porphyrae</i>	3 K	6 K; 12 K	-	-
<i>Myrionema magnusii</i>	3 K	-	-	-
<i>Myrionema strangulans</i>	3 A, K	6 K; 12 A, K	-	17 K
<i>Myriotrichia clavaeformis</i>	-	6 A; 12 A, K	-	-
<i>Petalonia fascia</i>	3 A	6 A; 12 A, K	13 A, K	17 A, K
<i>Petalonia zosterifolia</i>	3 A	-	-	-
<i>Pilayella littoralis</i>	3 A, K	6 A; 12 A, K	-	17 A, K
<i>Punctaria latifolia</i>	-	6 A	-	-
<i>Punctaria plantaginea</i>	-	12 K	-	17 A
<i>Punctaria tenuissima</i>	-	12 K	-	-
<i>Ralfsia verrucosa</i>	3 K	6 K; 12 A, K	-	17 A, K
<i>Sargassum muticum</i>	-	-	-	16 A, K; 17 A, K
<i>Scytosiphon lomentaria</i>	3 A, K	6 A, K; 12 A, K	13 K	17 A, K
<i>Sphaelaria cirrosa</i>	2 A, 3 K	-	-	-
<i>Sphaerotrichia divaricata</i>	-	12 K	-	-
<i>Spongonema tomentosus</i>	-	12 K	-	-
<i>Stictyosiphon</i> sp.	-	12 K	-	-
<i>Ulonema rhizophorum</i>	-	12 K	-	-
<b>CHLOROPHYCEAE</b>				
<i>Acrochaete repens</i>	-	12 K	-	-
<i>Acrochaete wittrockii</i>	-	12 K	-	-
<i>Blidingia marginata</i>	-	-	-	17 K
<i>Blidingia minima</i>	-	6 A, 12 A	-	17 K
<i>Bryopsis plumosa</i>	-	6 A, 12 A, K	13 A, K	17 A, K
<i>Capsosiphon fulvescens</i>	-	6 A, 12 K	-	-
<i>Chaetomorpha aerea</i>	-	5 O; 6 K, 12 A, K	-	17 A, K
<i>Chaetomorpha linum</i>	2 A; 3 K, 4 K	6 K, 7 A; 12 A, K	13 A, K	17 A, K
<i>Chaetomorpha sutoria</i>	-	11 K	13 K	17 K
<i>Cladophora</i> spp.	3 A	-	13 A, K	17 A, K
<i>Cladophora fracta</i>	-	12 A	-	-
<i>Cladophora hutchinsiae</i>	-	6 A, 12 K	13 K	-
<i>Cladophora lehmaniana</i>	-	12 A	-	-
<i>Cladophora sericea</i>	2 A; 3 K	5 O; 6 A; 12 A, K	13 K	17 A, K
<i>Codium fragile</i>	-	-	13 K	17 A
<i>Codium tomentosum</i>	-	5 O	-	-
<i>Enteromorpha</i> spp.	-	-	13 A, K	17 A, K

Table 2 b (Continued)

	1872–1927	1932–1958	1976–1980	1990–1994
<i>Enteromorpha ahlneriana</i>	–	–	–	17 K
<i>Enteromorpha clathrata</i>	3 K, 4 K	12 A, K	–	14 K; 17 A, K
<i>Enteromorpha compressa</i>	3 A	5 O; 6 K; 12 A, K	13 K	14 K; 17 A, K
<i>Enteromorpha crinita</i>	4 K	12 A, K	–	–
<i>Enteromorpha flexuosa</i>	–	6 A; 12 A, K	–	14 K; 17 A, K
<i>Enteromorpha flexuosa</i> subsp. <i>paradoxa</i>	3 K; 4 K	–	–	–
<i>Enteromorpha intestinalis</i>	–	6 K; 12 A, K	–	14 K; 17 A, K
<i>Enteromorpha linza</i>	3 A, 4 K	5 O; 6 A; 12 A, K	13 A	14 K; 17 A, K
<i>Enteromorpha linziformis</i>	–	–	–	14 K; 17 K
<i>Enteromorpha pilifera</i>	–	–	–	14 K; 17 K
<i>Enteromorpha prolifera</i>	–	–	–	14 K; 17 K
<i>Enteromorpha radiata</i>	–	–	–	14 K; 17 A, K
<i>Enteromorpha ralfsii</i>	–	–	–	14 K; 17 K
<i>Enteromorpha ramulosa</i>	4 K	–	–	–
<i>Enteromorpha simplex</i>	–	–	–	14 K; 17 A, K
<i>Enteromorpha torta</i>	–	6 K; 7 A; 12 K; 8 K	–	14 K; 17 A, K
<i>Epicladia perforans</i>	–	–	–	17 K
<i>Gomontia polyrhiza</i>	2 A	–	9 K	–
<i>Eungomontia sacculata</i>	–	10 K	–	17 A, K
<i>Percursaria percursa</i>	–	6 K; 7 A; 12 A, K; 8 A, K	–	17 K
<i>Prasiola stipitata</i>	–	–	–	17 K
<i>Rhizoclonium riparium</i>	4 K	7 A; 12 A, K	–	17 K
<i>Spongomorpha aeruginosa</i>	–	6 A	–	–
<i>Ulothrix flacca</i>	3 A	6 A; 12 A, K	–	17 K
<i>Ulothrix implexa</i>	–	7 A; 6 A; 12 A, K	–	17 K
<i>Ulva lactuca</i>	2 A; 4 K	6 A; 12 A, K	13 A, K	14 K, 17 A, K
<i>Ulva rigida</i>	–	–	–	14 K; 17 K
<i>Ulva scandinavica</i>	–	–	–	14 K, 17 K
<i>Ulva tenera</i>	–	15 K	–	–
<i>Ulvopsis grevillei</i>	–	6 A; 12 A, K	13 A, K	17 A, K
<i>Urospora penicilliformis</i>	–	6 A; 12 A	–	17 K

It is assumed that the data sets referred to in this paper (with the exception of two sets, i. e. Kornman & Sahling, unpublished data [No. 12] and this present study covering Königshafen Bay [No. 17]) do not comprise the total species abundance during the respective time of investigation

From the often selective emphasis of workers on certain groups of algae, it is not feasible to assess the relative abundance of single species which are listed in Table 2 b. Neither does the fragmentary and infrequent information represent a comprehensive and useful basis for estimating absolute abundance of each species over time. Nevertheless, the benthic macroflora recorded in the period between 1870 and 1946 differs in two major respects from that which occurs at present:

(A) Before the late 1970s, dense green algal mats were not mentioned in any of the references to Königshafen Bay (see Table 2 a), although the genus *Enteromorpha*, which is at present the most important contributor to the algal mats (Fig. 2), was already a conspicuous member of the macrophytobenthos (Nienburg, 1927).



Fig. 2. A dense *Enteromorpha* spp. mat covering a bed of *Zostera marina* in the western part of Königshafen Bay in 1992

(B) Our measurements at the List harbour walls show a maximum vegetation depth of 3 m below mean high water level (MHWL). Only crustose forms of brown algae (e.g. *Ralfsia verrucosa*) were found in greater depths, on mobile biogenic substrata such as shells of hermit crabs (*Eupagurus bernhardus*), but not on firm substrata. In the earlier part of the century the lower vegetation depth limit must have been several metres below that of the present vegetation as Hagmeier (1941) reported the growth of macroalgae on subtidal oysterbeds down to ca 8 m.

Currently, at least 12 red algal species, 13 brown algal species and 34 green algal species are present in Königshafen Bay. The green alga *Codium fragile* and the brown algae *Laminaria saccharina* and *Punctaria plantaginea* were found only outside Königshafen. In relation to the findings of Kornmann (1952) and Kornmann & Sahling (1946–1958, unpublished data) in Königshafen, the number of red algal species has decreased from 16 to 12, and brown algal species from 23 to 12. The abundance of green algal species increased from 23 to 34. The latter is related to high numbers of species in the two genera *Enteromorpha* and *Ulva* which were mainly found on sand- and mudflats in our survey.

The complete disappearance of the red algae *Gracilaria verrucosa* and its epiphyte *Callithamnion corymbosum* is particularly interesting. Both species, first mentioned by Magnus (1874), were still conspicuous members of the macroflora of the outer part of Königshafen Bay in the 1950s, but became rare in the 1980s. Also, *Polysiphonia nigra*, *Hildenbrandia rubra* and *Lithothamnion sonderi* have not been found in the recent survey. The two crustose species never occurred very frequently in Königshafen, whereas *Polysiphonia nigra* was regularly present in the outer part of the bay, growing attached to mussel shells and pebbles.

#### Present situation

Three *Enteromorpha* species were regularly represented in the algal mats of Königshafen Bay (*E. flexuosa*, *E. prolifera*, *E. radiata*). *E. ralfsii* was only of local importance in the north east of Königshafen, where it grew attached to tubes of the polychaete *Lanice conchilega*. *E. clathrata* and *E. torta* were found commonly, though often with low biomass. *Chaetomorpha sutoria* was scarce on the tidal flats in Königshafen in 1990 and between 1992 and 1994, but formed mats measuring up to 20 cm deep in 1991 (Fig. 3). We very rarely found juveniles of *Chaetomorpha* and *Cladophora* in Königshafen Bay, whereas juvenile stages of *Enteromorpha* occurred in high abundances (hundreds m<sup>-2</sup>) attached to mudsnail shells in spring (Schories & Reise, 1993; Schories, 1995 a, b). *Ulva* spp., on the other hand, are transported by currents into the bay from other areas as adults and continue to grow without reproducing in protected localities. Thus, we recorded a maximum length of ca 3 m for a single plant of *Ulva scandinavica* in 1992. Direct development of *Ulva* germlings into adult plants is very rare in Königshafen Bay.

On mussel beds, the bladder wrack *Fucus vesiculosus* f. *mytili* is the most conspicuous macroalga. Additionally, extensive growth of the red alga *Dumontia contorta* and the brown alga *Petalonia fascia* can be observed in spring in depressions of individual mussel beds. In summer months, large individual plants of *Porphyra* spp. may grow up to 50 cm in length when grazing pressure is low. The red sheets cover parts of some mussel bed surfaces completely. This blanketing was observed under natural conditions in the outer part of Königshafen Bay in 1993, and in grazer exclusion experiments carried out in 1993 and 1994 by Albrecht (1995).

The occurrence of the invasive Japanese brown alga *Sargassum muticum* is a new phenomenon in Königshafen Bay (Schories & Albrecht, 1995). In this location *S. muticum* grows attached to mussel beds (*Mytilus edulis*), and this represents the first record for attachment to a biogenic hard substratum in the Frisian Wadden Sea. Prior to this, unattached specimens had been found for several years following storm events along the beaches of the island of Sylt. In March 1993, *S. muticum* was discovered attached to the substratum almost simultaneously in two locations: on mussel beds in the outer part of

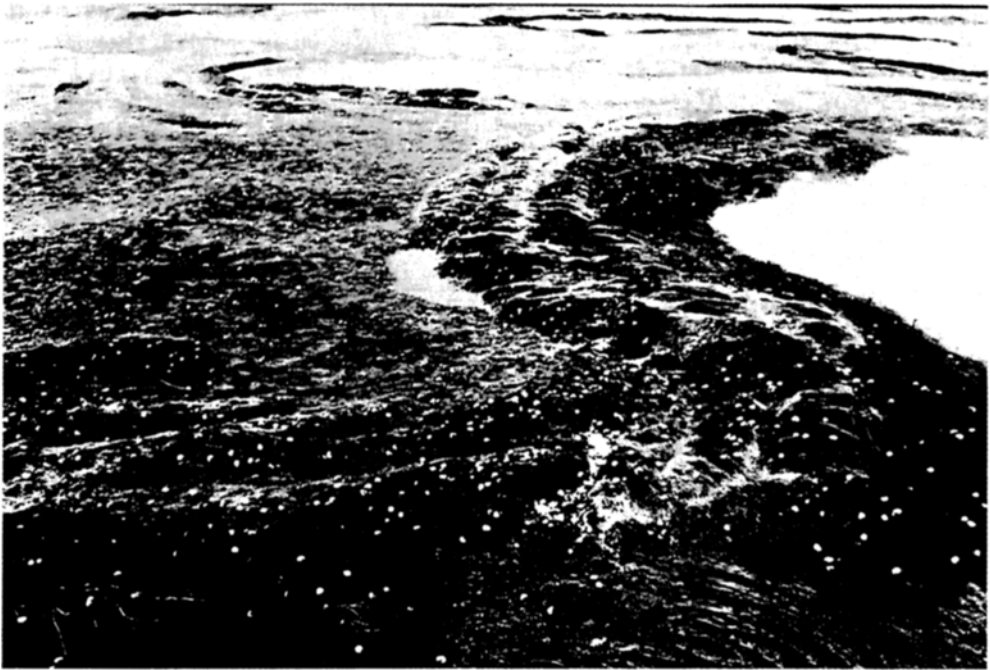


Fig. 3. *Chaetomorpha sutoria* growing in mats in the western part of Königshafen Bay in 1991. Littorinid snails (*Littorina littorea*) were abundant in decaying parts of the algal mats (below)

Königshafen Bay close to the subtidal zone and outside the bay in the low subtidal zone on a breakwater south of the village of List. During summer, the plants grow to a length of up to 2 m before degeneration processes lead to thallus losses in autumn (Fig. 4). At present, *S. muticum* is still part of the macroflora in these two areas.

Another invasive species recorded for the first time in the vicinity of Königshafen Bay is the green alga *Codium fragile*. It occurs regularly on the breakwater where *S. muticum* was found.

#### DISCUSSION

The macrophytobenthos of Königshafen Bay has undergone a change more dramatic than that reflected in our check-list (Table 2). Since the beginning of the century, two opposing trends are evident. Some red and brown algal species have become rare or have disappeared completely from the intertidal zone (e.g. *Callithamnion corymbosum*, *Gracilaria verrucosa*, *Hildenbrandia rubra*). In contrast, the abundances and biomasses of several green algal genera (*Chaetomorpha*, *Cladophora*, *Enteromorpha* and *Ulva*) have exhibited massive increases in the upper eulittoral zone. The presence in the intertidal zone of species such as *Ceramium nodulosum*, *Polysiphonia elongata*, *P. fucooides* and *Porphyra* spp. seems not to have changed in the long term.

When trying to explain these historical developments, it is important to realize that environmental conditions which influence the occurrence of wild organisms have changed through both natural and anthropogenic effects. It has become obvious that human



Fig. 4. Specimen of *Sargassum muticum* found attached to mussel beds between two tidal inlets in the outer part of Königshafen Bay; July, 1994

impact is of great significance for often dramatic changes in the environment (Agger, 1989), particularly in an ecosystem such as the Wadden Sea which has been structured by human activity for a thousand years. However, in Königshafen Bay natural fluctuations of environmental parameters interact with anthropogenic changes and are at least partly responsible for the observed changes. Therefore, natural as well as human-induced aspects of different factors affecting macroalgal growth will be discussed together.

#### Substratum availability

During the late 1980s, De Jonge & Peletier (1992) found a dense cover of *Ulva* spp. in the western Dutch Wadden Sea, apparently related to the presence of hard substrata supplied by blue mussel cultures (*Mytilus edulis*). After *M. edulis* became rare, because of insufficient spatfall, *Ulva* became unimportant in the western Dutch Wadden Sea. In this case, the availability of a hard substratum seems to have been the limiting factor for the development of *Ulva*. On the other hand, mussel beds have always been present in Königshafen Bay in this century (Nienburg, 1927; Wohlenberg, 1937), but they have also shown high fluctuations in terms of abundance and spatial distribution (Reise et al., 1989). In contrast to the observations of De Jonge & Peletier (1992) in the Netherlands, the

growth of *Ulva* on mussel beds in Königshafen Bay has never been of any importance. High biomass of *Ulva* only occurs when plants drift into the bay and begin vegetative growth, so that this species' abundance is probably regulated by other factors in our study area (see below).

However, the availability of a suitable substratum is also of relevance to the occurrence of epiphytic macroalgae: seagrass, which was abundant throughout the Wadden Sea, decreased dramatically in the 1930s. Almost the entire population of *Zostera marina*, growing at, or just below, the low water mark was affected in the Dutch (den Hartog, 1994) as well as in the German Wadden Sea. Wohlenberg (1935) reported a strong disease-related decline of *Zostera* in a subtidal bed east of Königshafen Bay during 1933–34 (for further information see Reise et al., 1989). This bed disappeared completely. The loss of seagrass beds in the Dutch and German Wadden Sea more than 60 years ago is thought to have altered the abundance of macroalgae, especially of the red algal epiphytes associated with them. The work of van Goor (1921) on the Dutch Wadden Sea gives a lasting impression of the potential species richness of macroalgae in seagrass beds.

Additionally, the construction of artificial hard substratum – in Königshafen Bay this refers particularly to the building of a new dike in the early 1930s – might have enhanced the growth of some species which had previously been limited in their distribution by lack of suitable attachment surfaces. In particular, the high eulittoral species *Blidingia minima*, *B. marginata* and *Prasiola stipitata* have been found recently growing on the dike, thus contributing to the increase in green algal species compared to earlier investigations.

### Grazing

Not all substrata serving as potential attachment surfaces are colonized by macroalgae otherwise abundant in the area. This is often attributable to grazing effects.

Geertz-Hansen et al. (1993) showed that, in a eutrophic Danish estuary, the development of nuisance macroalgae like *Ulva lactuca* can be controlled by grazing isopods (*Idotea* sp.). Albrecht (1995) demonstrated the influence of littorinid snails on the presence of several macroalgal species on mussel beds in Königshafen Bay. Experiments using exclusion cages to reduce *Littorina littorea* densities showed that, within a short time in summer, opportunistic species such as *Porphyra* spp., *Ulva lactuca* and *Enteromorpha* spp. developed on the mussel shells. Therefore, changes in abundance of ephemeral green and red algae might often have been related to grazing pressure in Königshafen Bay. Unfortunately, no simultaneous investigations of grazers and macroalgae were carried out in the bay before 1992 (Wilhelmsen & Reise, 1994; Albrecht, 1995)

### Eutrophication

Eutrophication has been the predominant factor of change over the last 50 years. Proceeding from the estuaries, enhanced nitrogen and phosphorus loads have increased the nutrient concentrations in intertidal waters of the North Frisian Wadden Sea during winter (Martens, 1989 a, b, 1992; Hickel et al., 1993). Several authors (cf. Buttermoore, 1977; Montgomery & Soulsby, 1981; Kolbe et al., 1994; Reise & Siebert, 1994) assumed a positive correlation between eutrophication and green algal biomass. Thus, there are many regions in the world where mass development of *Ulva* to nuisance proportions is strongly



exacerbated by high nutrient inputs into the system (Sawyer, 1965; Sfriso et al., 1987; Piriou & Menesguen, 1992).

Although more and more global records substantiate these positive correlations, the evidence for a direct link is by no means conclusive. In our view, the general relationship between the potential for green algal mass development and the environmental parameters involved is often too complex to be demonstrated in single case studies. Grazing pressure (Geertz-Hansen et al., 1993; Wilhelmsen & Reise, 1994), availability of substrata and anchorage (Schories & Reise, 1993), rainfall (Piriou & Menesguen, 1992), desiccation (Townsend & Lawson, 1972), competition between macroalgae, microphytobenthos and phytoplankton (Fong et al., 1993) are only a few examples indicating the complexity of factors which may influence the development of green algal mats in eutrophicated areas. However, Raffaelli et al. (1989) found a positive correlation between nutrient loads (especially nitrogen) and biomass of *Enteromorpha* sp. in the Ythan River (Scotland). In Königshafen Bay large quantities (in terms of abundance and biomass) of opportunistic green algae (e.g. *Cladophora* spp., *Chaetomorpha sutoria*, *Enteromorpha* spp. and *Ulva* spp.) have occurred regularly on muddy and sandy tidal flats since 1979 (Reise, 1983; Schories, 1995a), covering sediments and seagrass beds (*Zostera marina* and *Zostera noltii*) as thick mats (Fig. 2). However, maximum standing crop of these green algae has been extremely variable over the years (Reise, unpubl. data), making it almost impossible to predict their abundance and/or distribution in Königshafen Bay. Nevertheless, our investigations (in concordance with Reise et al., 1994 b) suggest that, although diverse factors interact, mass development of *Enteromorpha* is, overall, a consequence of high nutrient availability and, as such, a man-made problem.

In contrast, the higher number of species of *Enteromorpha* and *Ulva* nowadays is probably an artifact of the taxonomic revision of these genera since Kornmann's (1952) and Kornmann & Sahling's (1946–1958, unpublished data) work. In the 1960s and 1980s, Bliding (1963, 1968) and Koeman & van den Hoek (1982 a, b; 1984) introduced substantial taxonomic changes to *Enteromorpha* and *Ulva*, elevating several subspecies to species.

The red alga *Gracilaria verrucosa*, which disappeared from Königshafen in the 1980s, is viewed widely as a warm-temperate species, but it frequently occurred, together with its epiphyte *Callithamnion corymbosum*, in the outer part of Königshafen Bay between 1870 and 1958. We do not believe that eutrophication, expressed as increased nitrogen and phosphorus concentrations in the intertidal waters of the North Frisian Wadden Sea during winter (Martens, 1989 a, b, 1992; Hickel et al., 1993), is likely to be responsible for the disappearance of *Gracilaria* and *Callithamnion*. In the Lagoon of Venice, which is much more eutrophicated than Königshafen, *Gracilaria verrucosa* is still an abundant member of the macroalgal flora (Sfriso et al., 1992). Indeed, some evidence exists that *Gracilaria*, known for its fast growth- and high nitrogen uptake rates (Ryther et al., 1981; Thomas & Harrison, 1987), has been suppressed by a natural phenomenon.

Nevertheless, comparing the Lagoon of Venice with Königshafen in terms of macroalgal vegetation, the same long-term trends can be observed in both areas: a decrease in red algal species richness, an increase in abundance of green algal species and a tendency for an upward movement of the lower vertical vegetation depth limits (Sfriso, 1987). Further information about long-term trends in marine macroalgae of three polluted estuaries in north-east England are given by Hardy et al. (1993).

### Turbidity

Reduced transparency of the tidal waters has probably affected macroalgal distribution, especially in the subtidal zone. Submerged algae have temporarily and locally decreased in the vicinity of Königshafen Bay. The shallower vegetation depth limit in the sublittoral at present must have resulted in a remarkable loss of macroalgae between 3 m and 8 m below MHWL as was concluded by Reise et al. (1989) after reinvestigating Hagmeier's (1941) study sites. Light limitation through water turbidity might have resulted from excessive phytoplankton blooms (e. g. *Phaeocystis globosa*) as a consequence of eutrophication, or from an increase in current velocity accompanied by an increase in seston concentration. The latter process is thought to have originated from the extensive coastal embankments during recent decades. Straightening of the seaward dike lines has diminished numerous previously stable sedimentation fields, thus reducing intertidal areas, speeding up current velocities, intensifying erosion processes and increasing tidal level. For instance, the constructions of the Hindenburg causeway, which has connected the island of Sylt with the mainland since 1927, and of the Rømø causeway (finished in 1949), resulted in a substantial increase of muddy flats along the structures themselves (Wohlenberg, 1953), indicating that elsewhere in the Sylt-Rømø area a substantial amount of mud must have been eroded.

Within Königshafen Bay, on the other hand, a pronounced compaction of sediment appears to have taken place over the last 50 years (Austen, 1994), but is of no importance for the vegetation depth because there are no important subtidal areas present in the inner part of the bay other than the main tidal channel.

### Introduction of new species

The anthropogenic introduction of marine organisms to intertidal areas is well documented for the Frisian Wadden Sea (Reise, 1990). Its increase over recent decades is partly due to higher human mobility and trade activity at sea which facilitates the transport of species across previously existing natural boundaries in terms of space and time. Thus, the green alga *Codium fragile*, which invaded European coasts in the present century, but which is very scarce in the vicinity of Königshafen Bay, and the brown alga *Sargassum muticum*, which was first discovered in the area in March 1993 (Schories & Albrecht, 1995) have both made their way around the world with oyster cultures and are only two examples of a series of introduced species now established in the bay. The further extension of *S. muticum* on hard substrata remains currently unpredictable. However, there is some evidence that its distribution will be restricted to the lower intertidal zone, where it might compete with the native *Fucus vesiculosus* f. *mytili*, because both species grow attached to mussel beds.

Both examples show that the ecosystem of the Wadden Sea is dynamic and flexible enough to accommodate new species, so that further changes in the macroalgal flora due to introductions from elsewhere remain possible.

### CONCLUSIONS

The macroalgal flora of Königshafen and its adjacent areas has changed during the last century. The decrease of intertidal red and brown algae is obvious. If the decline of the macrophytobenthos vegetation depth is indeed attributable to eutrophication and tur-

bidity, prospects for the persistence of rare algal species and species richness are rather dim in the adjacent subtidal areas to Königshafen Bay, because the underlying processes still continue. Also, in the short term, there is no evidence that the mass development of green algae will cease.

We conclude that, with respect to Wadden Sea monitoring projects, Königshafen Bay is an appropriate area for phycological studies because of its overall species richness and the availability of abiotic and biotic background data. It meets the requirements for a successful monitoring program (Reise, 1989).

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