

Long-term changes in zooplankton community size structure: a global comparison

Hans M. Verheye (South Africa)

Elena Arashkevich (Russia), Sanae Chiba (Japan)

Ángel López-Urrutia (Spain), Todd O'Brien (USA), Mark Ohman (USA)

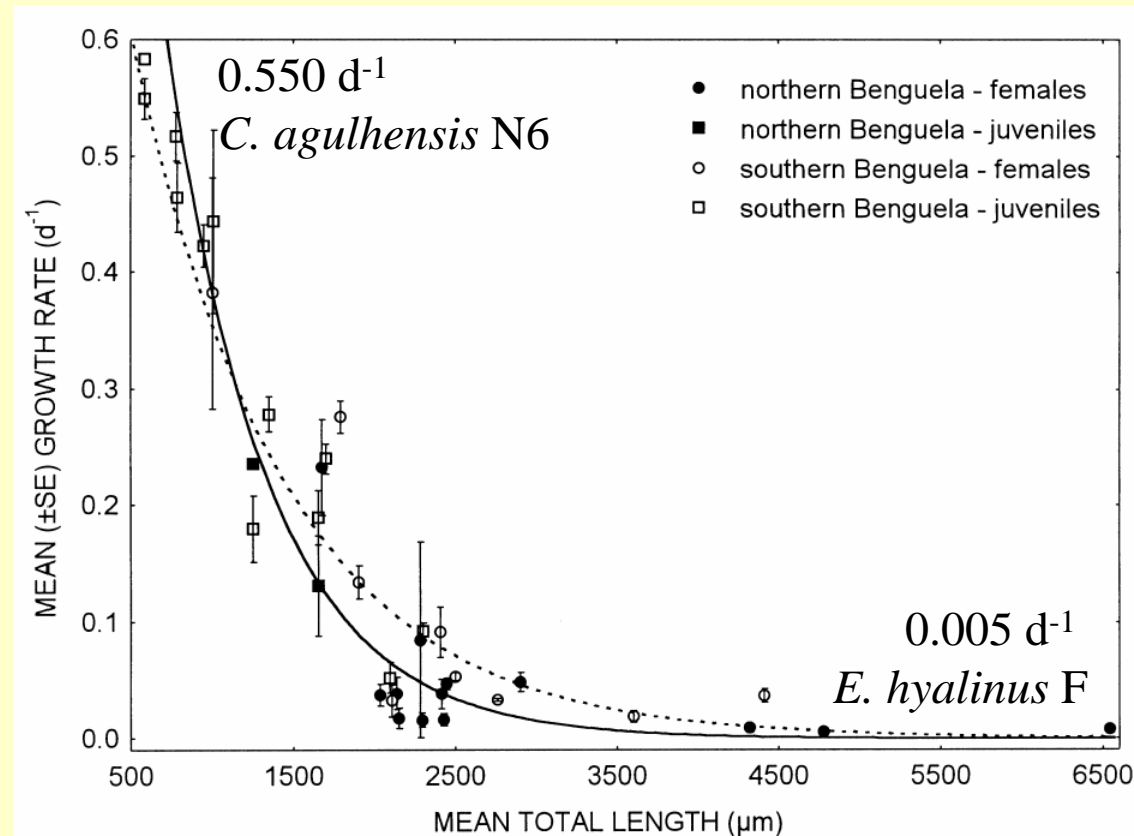
Anthony Richardson (Australia) and SCOR WG 125 Contributors

Importance of BODY SIZE in zooplankton:

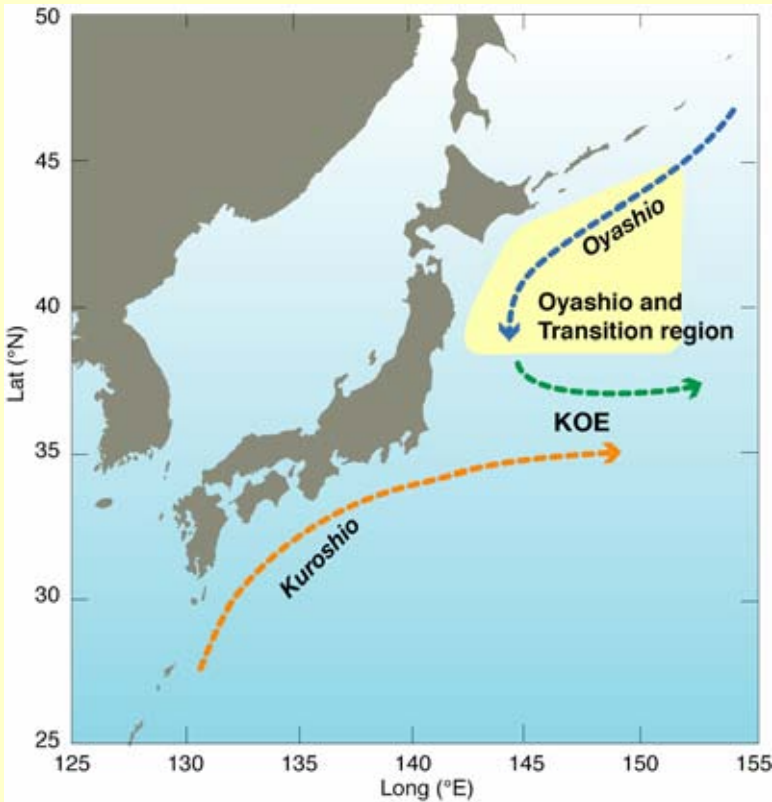
Allometric relationships describe many biological rate processes in the ocean;
e.g.

- **Predation:**
usually, large individuals eat smaller ones.

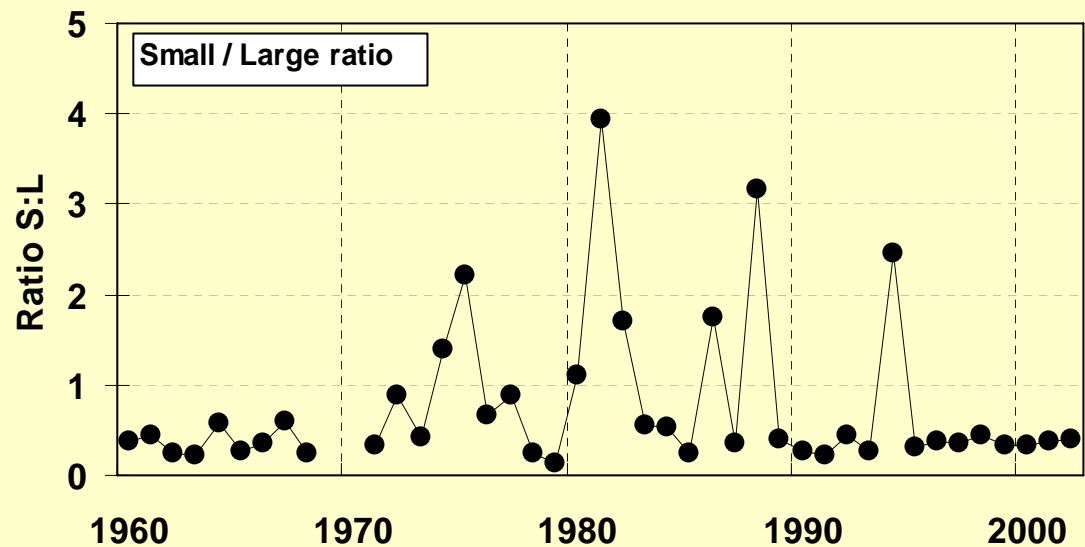
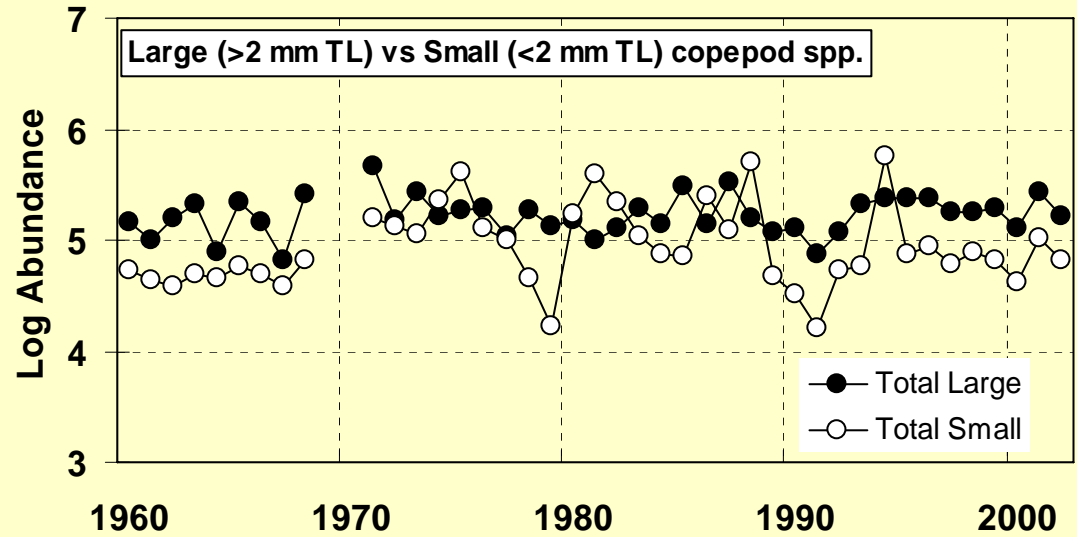
- **Growth:**
progressive decrease of somatic & reproductive copepod growth rate with increasing body size;



Oyashio, Japan 1960-2002



**59 dominant
copepod spp.**



Southern Benguela upwelling system

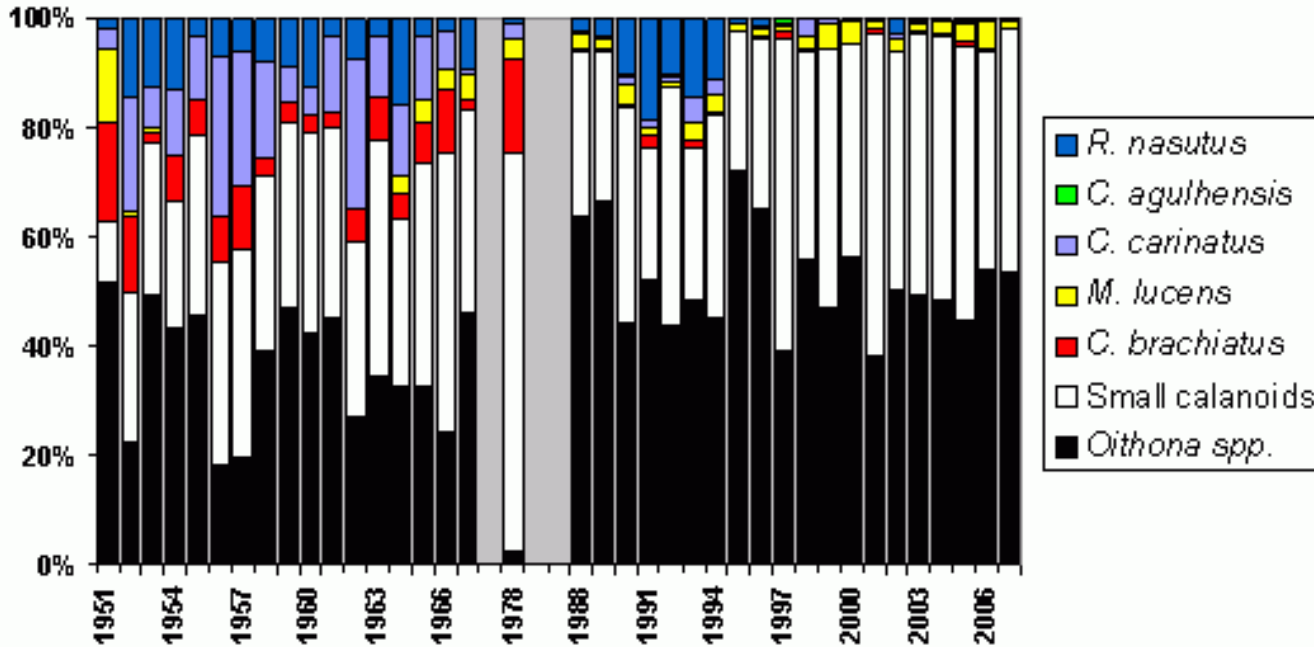
(St Helena Bay, South Africa)

1951-2007

Substantial changes through time in
species = size composition



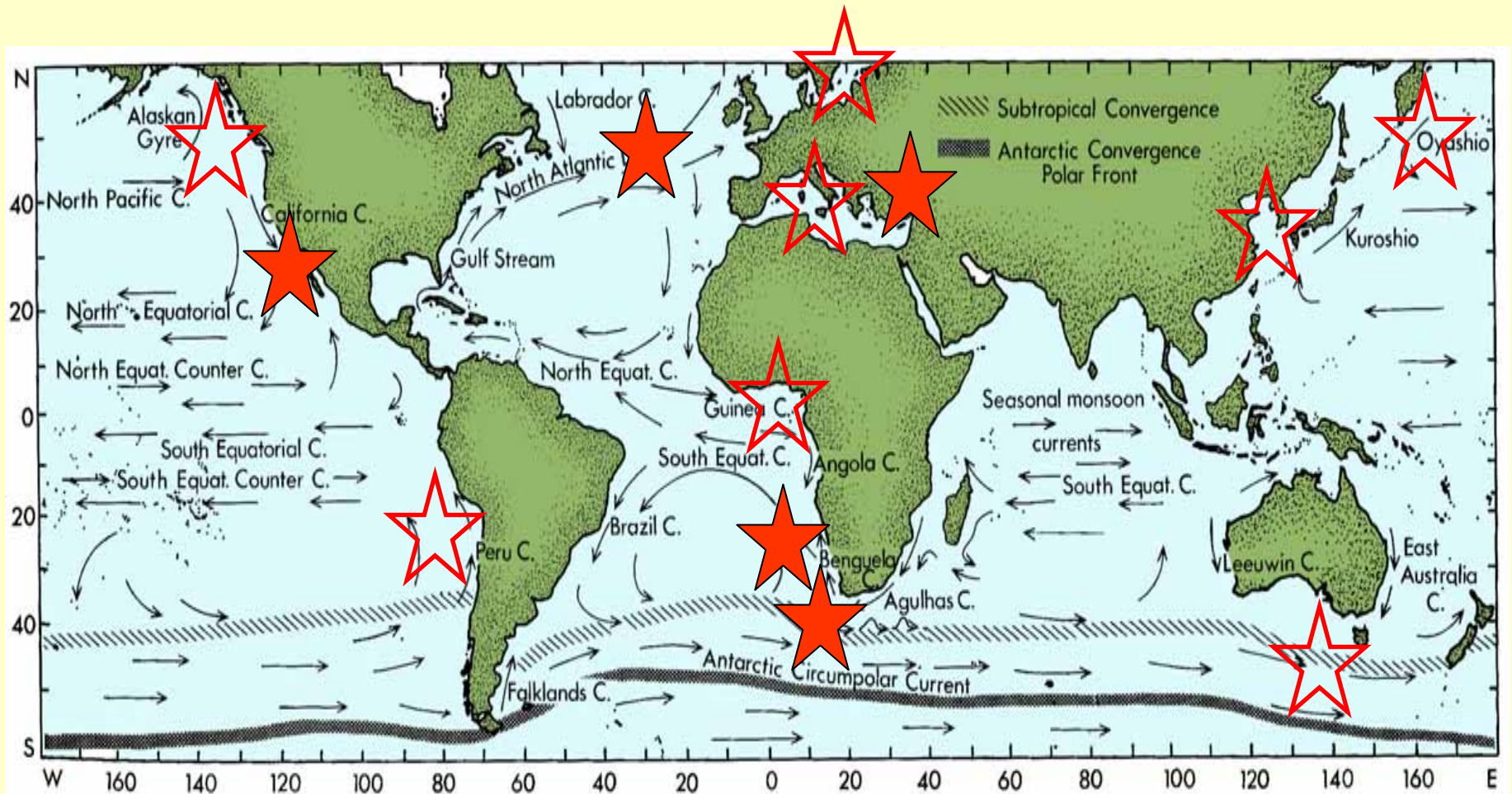
Copepod species composition



4.5
↑
Size (TL, mm)
0.7

Objectives:

To examine and compare long-term changes in the size structure of zooplankton communities from different regions of the world's oceans



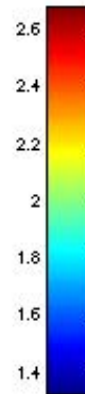
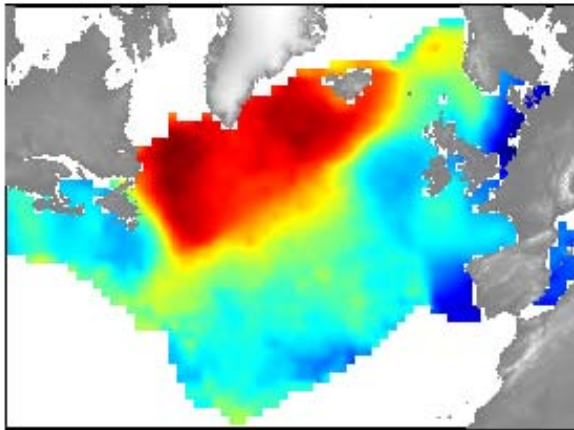
Hypothesis:

Warm-water zooplankton communities are often dominated by species of a smaller size (and more diverse) than their cold-water counterparts,

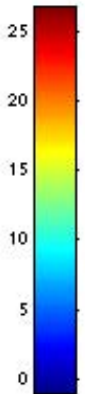
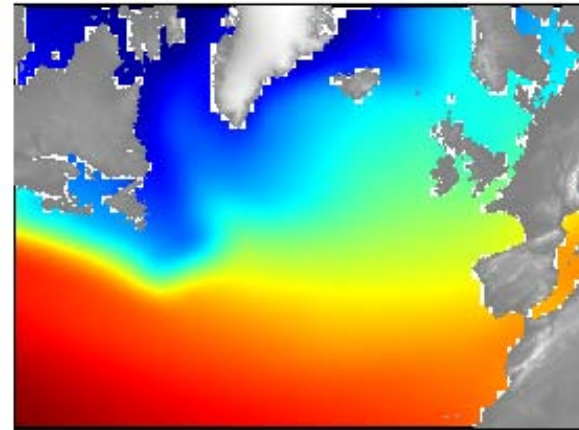
so that

a **shift to a smaller** average community size over time is indicative of **warming** of the ecosystem, while a **shift to a larger** average size indicates **cooling**.

Mean Calanoid Length



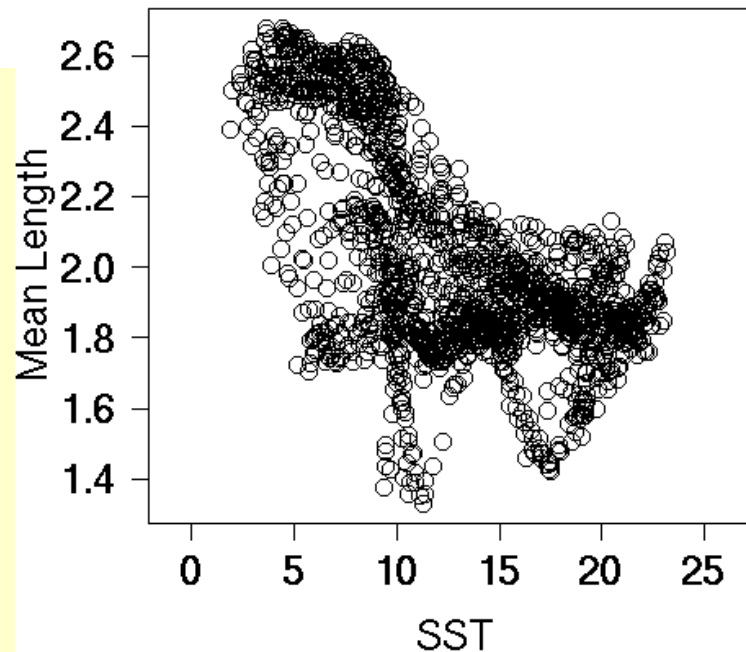
SST



CPR surveys

North Atlantic

1948-present



Ángel López-Urrutia,
Anthony Richardson

Approach:

To derive a **size-based index** of community structure and track its change over time.

- Focus on **substantial** changes in average community size based on changes in species composition.
- Focus on **copepod** community (most abundant; best resolved taxonomically in most sample analyses).
- Use **female** length (TL or PL) to represent the size of a species.

Calculation of average copepod community size \bar{S}

$$\bar{S} = \frac{\sum_{i=1}^N (L_i \times X_i)}{\sum_{i=1}^N X_i}$$

For each sample, multiply total length (L) of each species i (adult female) by its abundance (X_i), sum over all species (N), and divide by total abundance.

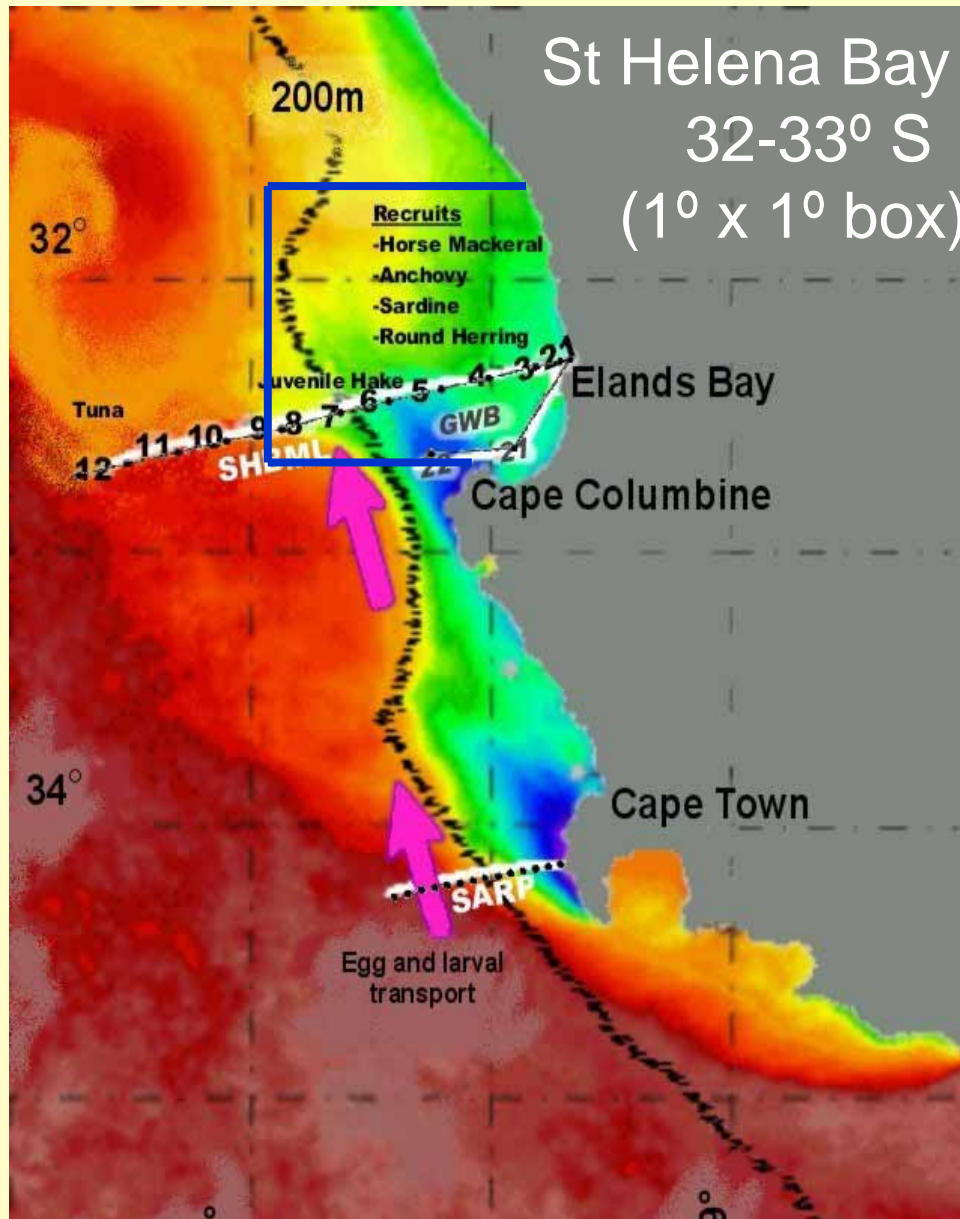
Assumptions:

- no change in mean body size of species over long time period
- no seasonal variations in mean body size

Southern Benguela Current

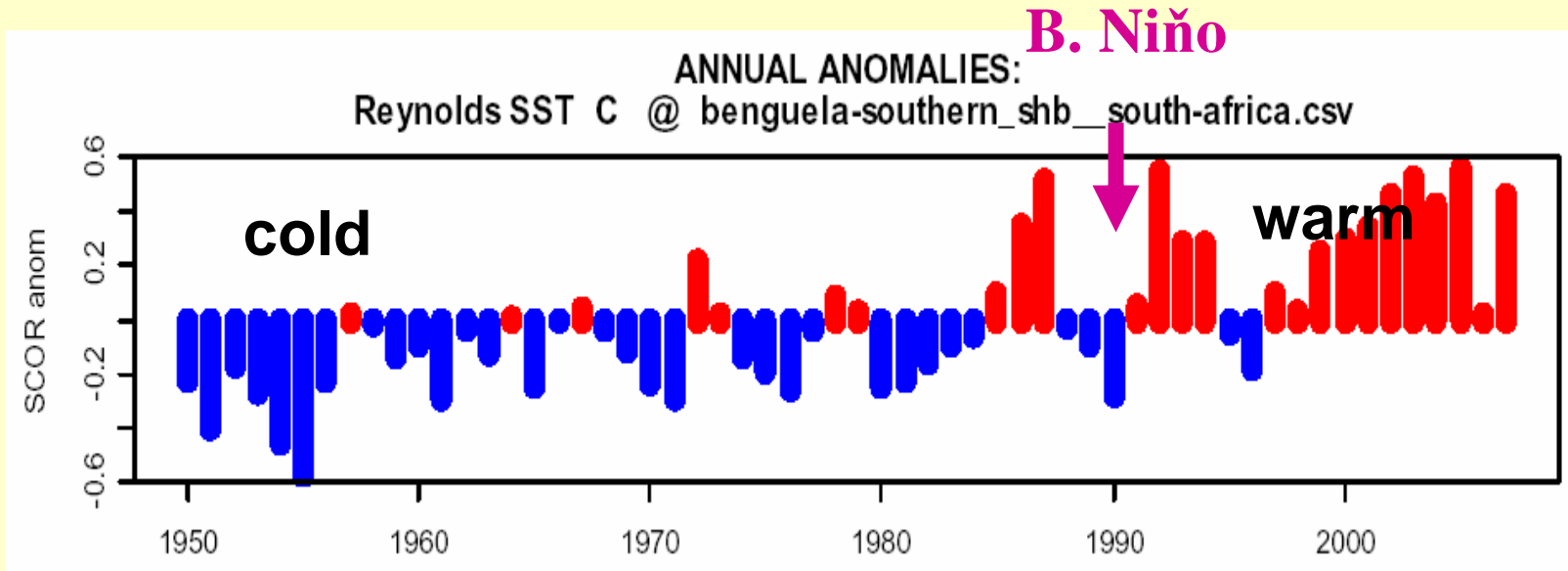
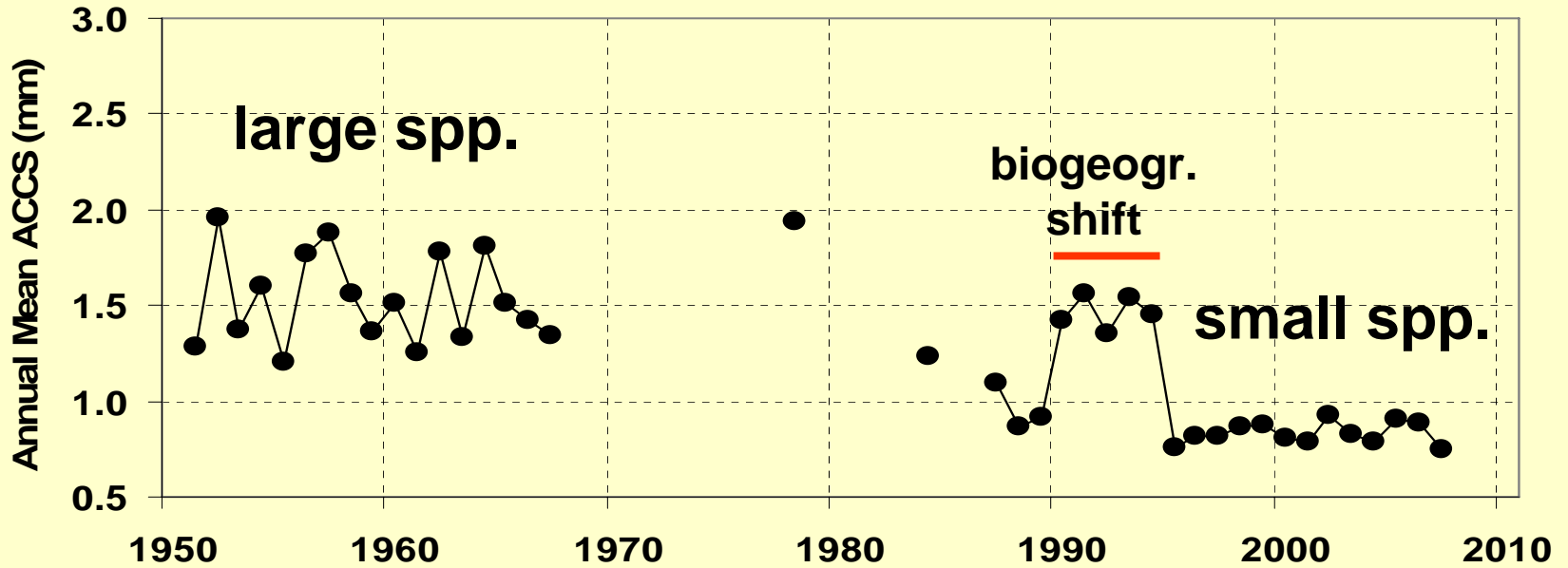
South Africa

1951-2007
(austral autumn)



Species	Fem min.TL (mm)
<i>Calanoides carinatus</i>	2.25
<i>Calanus agulahensis</i>	2.45
<i>Centropages brachiatus</i>	1.73
<i>Metridia lucens</i>	2.39
<i>Oithona</i> spp.	0.68
<i>Rhincalanus nasutus</i>	3.90

St Helena Bay transects (33°S), South Africa austral autumn (March-June)

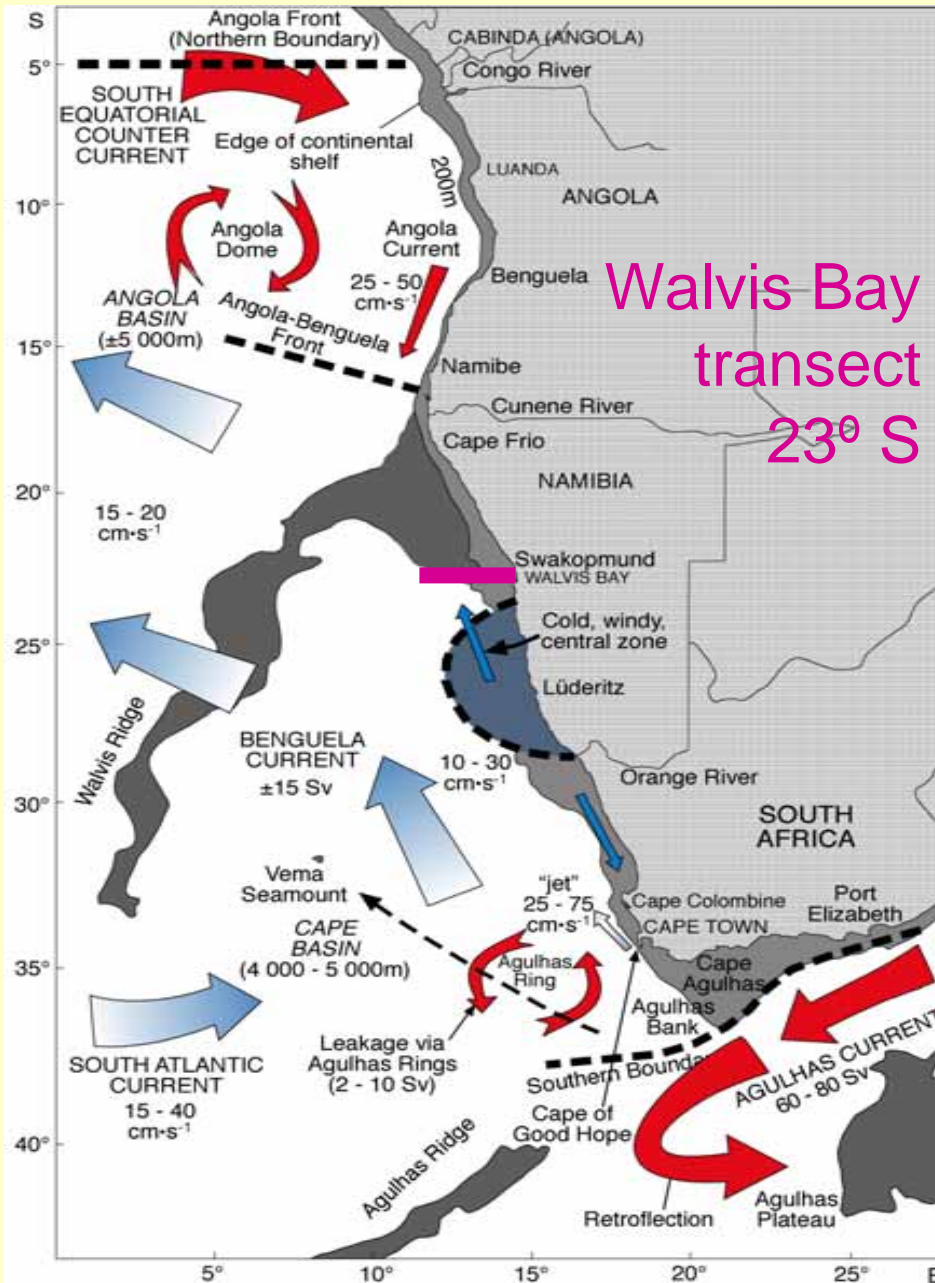


Northern Benguela Current

Namibia

1978-2006
(quasi-monthly)

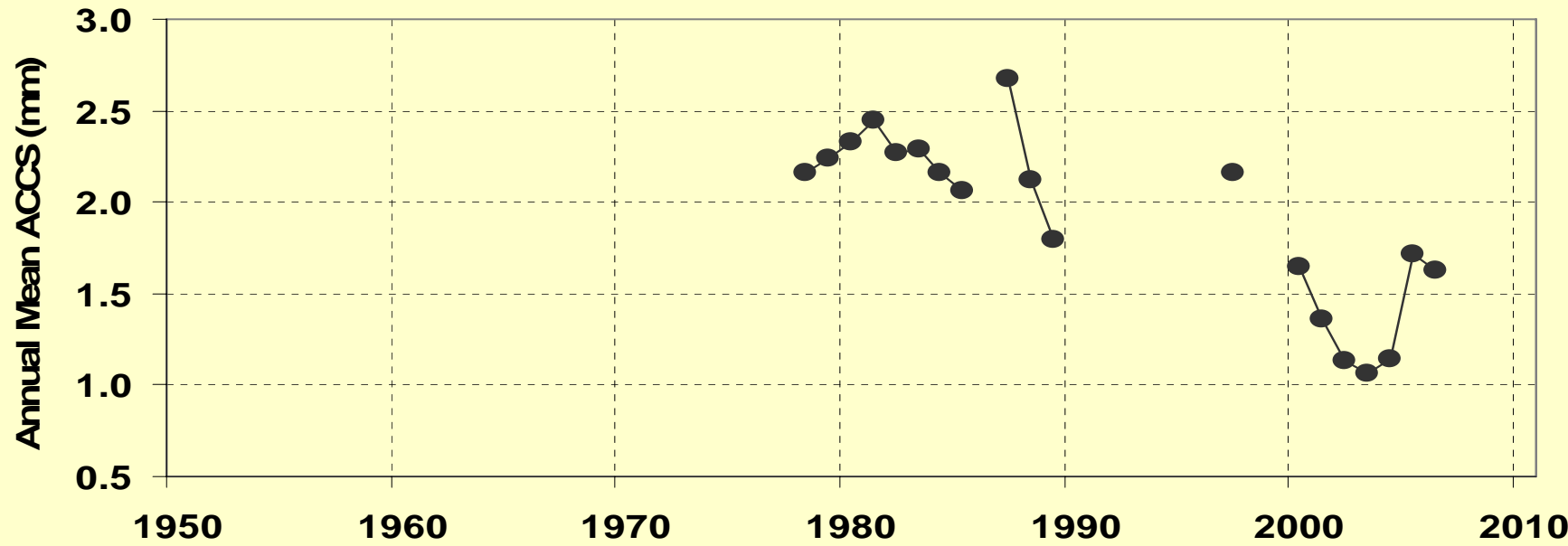
Walvis Bay
transect
23° S



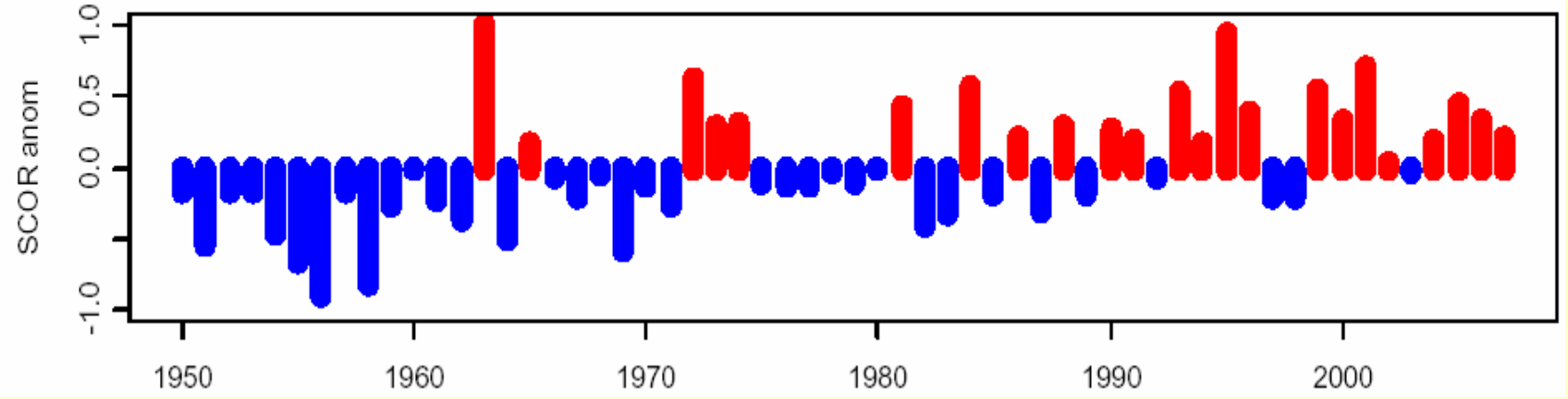
Species	Fem TL (mm)
<i>Calanoides carinatus</i>	2.445
<i>Calanus sp.</i>	2.760
<i>Centropages brachiatus</i>	1.677
<i>Metridia lucens</i>	2.294
<i>Oithona sp.</i>	0.850
<i>Pleuromamma sp.</i>	2.137
<i>Rhincalanus nasutus</i>	4.319

data: Fabienne Cazassus & Anja Kreiner

Walvis Bay transect (23°S), Namibia (quasi-)monthly sampling



ANNUAL ANOMALIES:
Reynolds SST C @ benguela-northern-wb_south-africa.csv

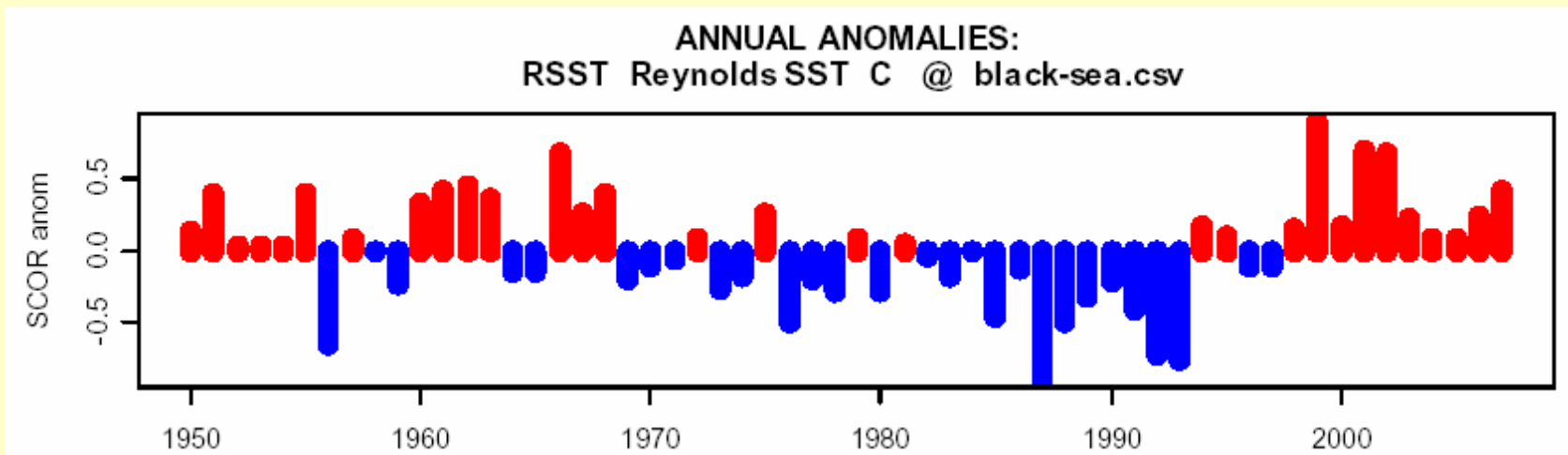
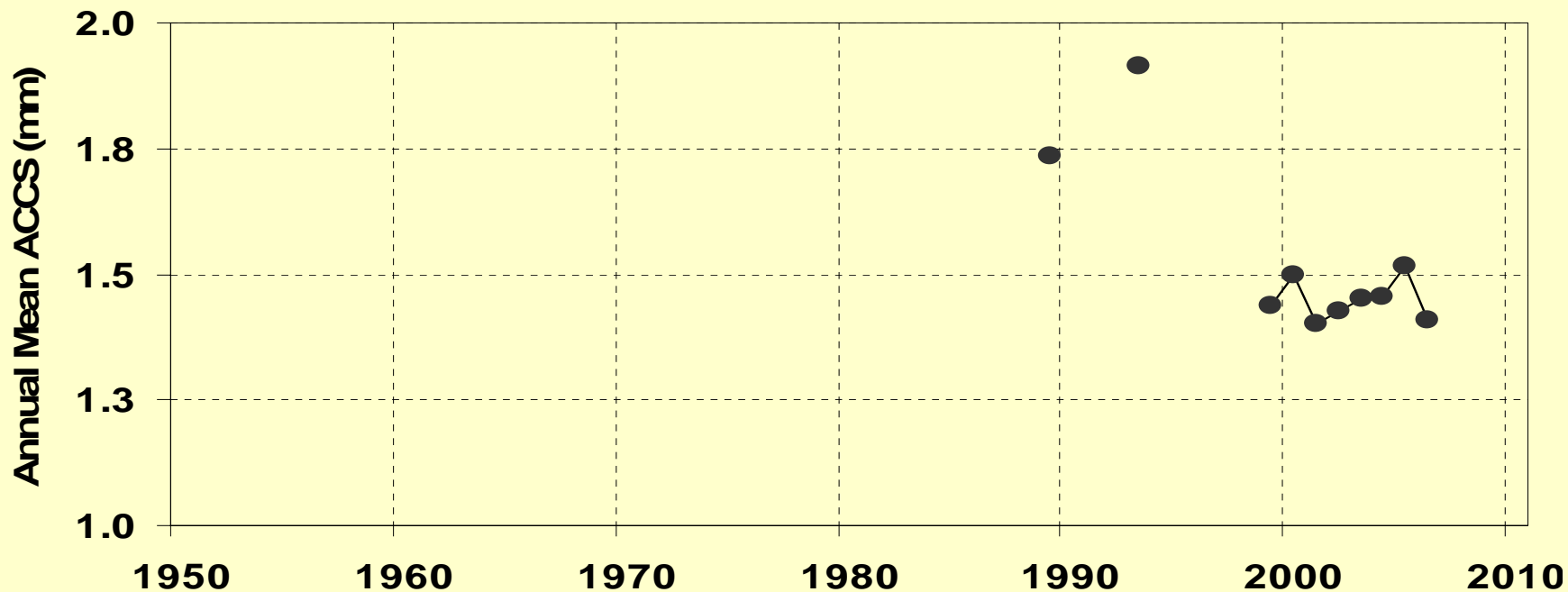


Black Sea:

1989-2006
(late autumn)

Species	Fem TL (mm)
<i>Acartia clausi</i>	1.50
<i>Calanus euxinus</i>	3.10
<i>Centropages ponticus</i>	1.70
<i>Oithona similis</i>	0.90
<i>Paracalanus parvus</i>	1.20
<i>Pseudocalanus elongatus</i>	1.60

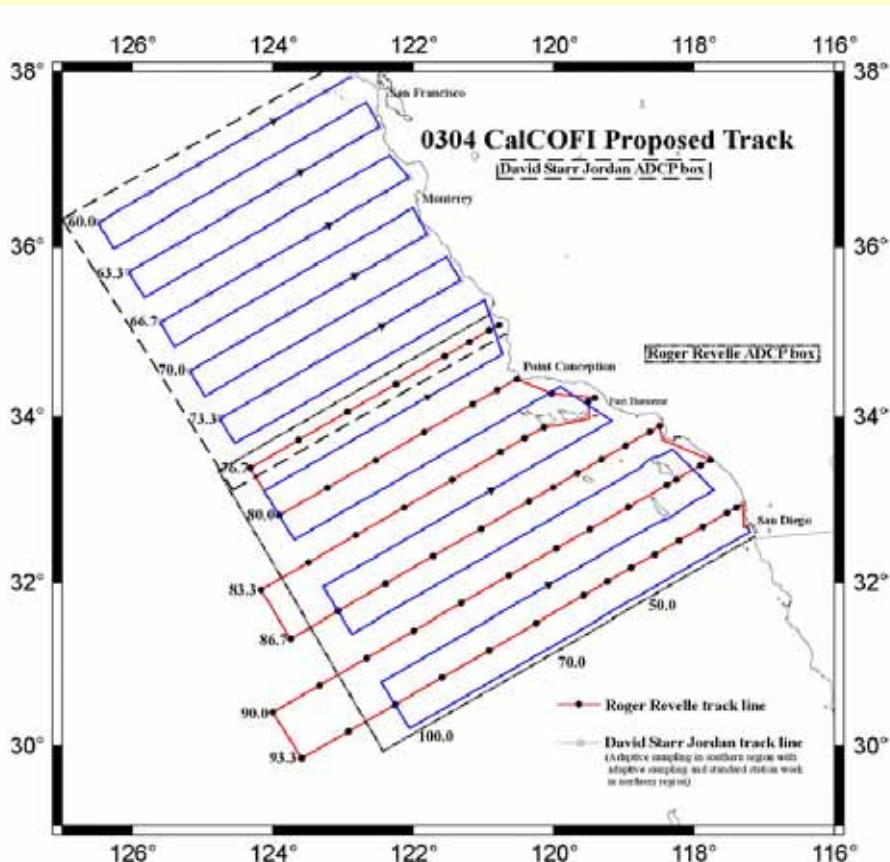
Black Sea - late autumn



California Current:

Southern California (CalCOFI)

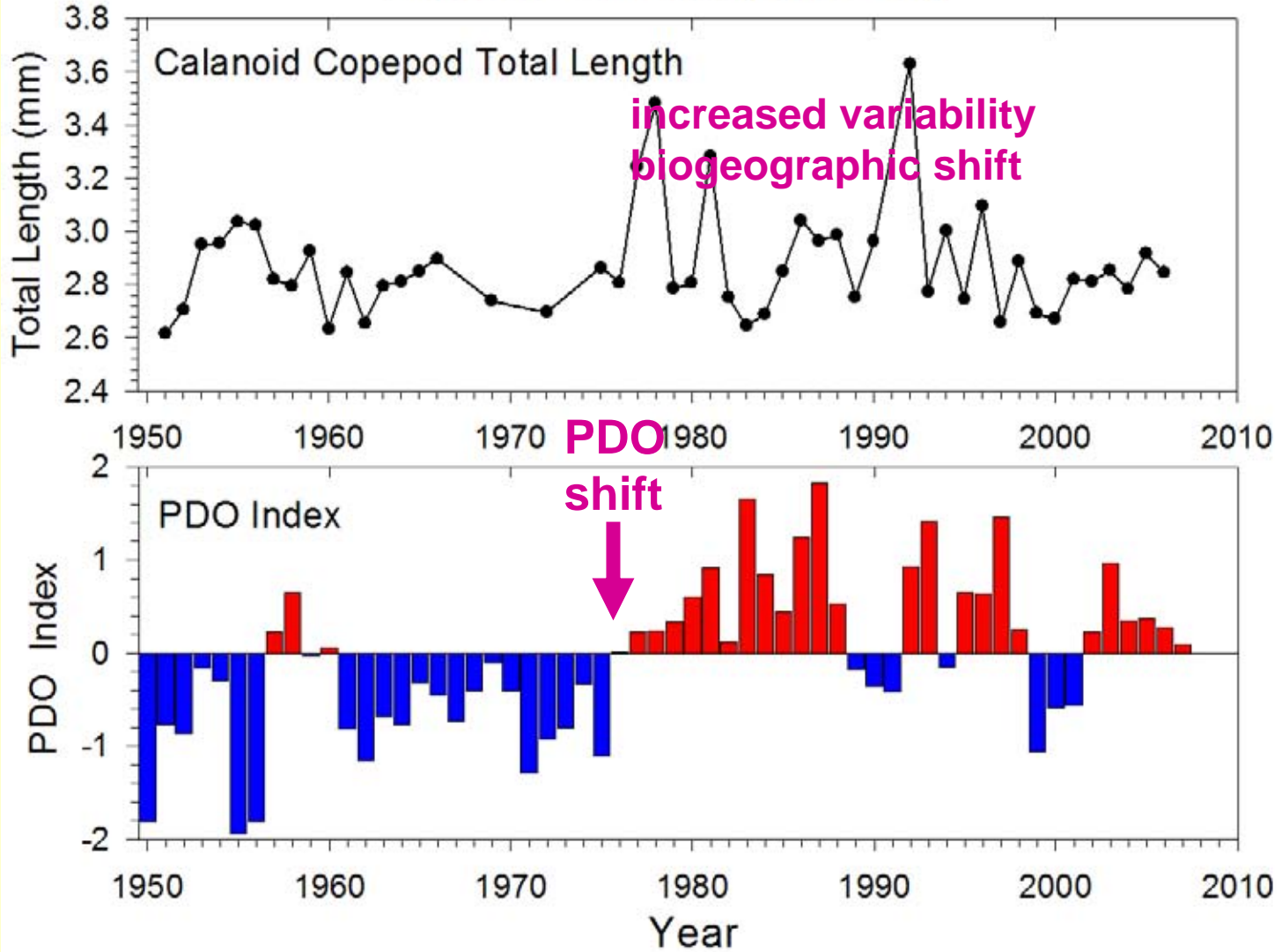
1951-2006 (spring)



Dominant 20 species	TL (mm)
<i>Aetideus bradyi</i>	1.59
<i>Calanus pacificus</i>	3.15
<i>Candacia bipinnata</i>	2.35
<i>Candacia curta</i>	2.26
<i>Eucalanus californicus</i>	6.21
<i>Euchaeta media</i>	3.53
<i>Euchirella pseudopulchra</i>	3.95
<i>Heterorhabdus papilliger</i>	2.03
<i>Labidocera trispinosa</i>	2.81
<i>Lucicutia flavicornis</i>	1.56
<i>Mesocalanus tenuicornis</i>	1.80
<i>Metridia pacifica</i>	2.37
<i>Nannocalanus minor</i>	2.03
<i>Pleuromamma abdominalis edentata</i>	3.05
<i>Pleuromamma abdominalis typica</i>	3.38
<i>Pleuromamma borealis</i>	2.07
<i>Pleuromamma gracilis</i>	2.08
<i>Pleuromamma piseki</i>	1.73
<i>Pleuromamma quadrungulata</i>	4.16
<i>Rhincalanus nasutus</i>	4.24

data: Mark Ohman

Southern California (CalCOFI)

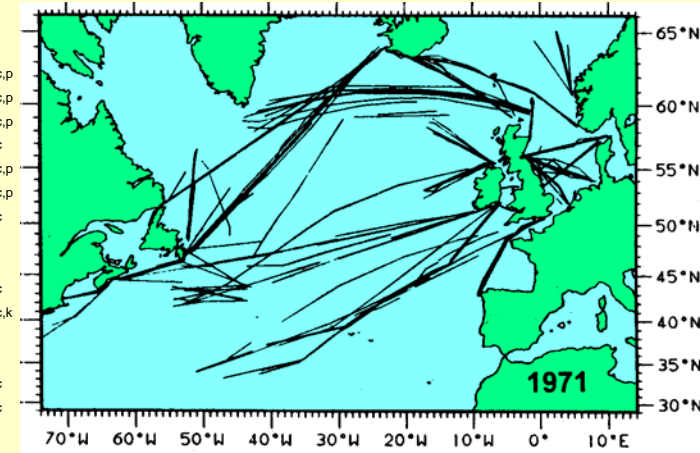


<i>Acartia danae</i>	1.08 ^{a,b,c}	<i>Heterorhabdus abyssalis</i>	2.40 ^c
<i>Acartia longiremis</i>	1.04 ^{a,c}	<i>Heterorhabdus clausi</i>	2.20 ^c
<i>Acartia negligens</i>	1.05 ^{b,c}	<i>Heterorhabdus norvegicus</i>	2.77 ^{b,c}
<i>Acartia</i> spp.	1.15 ^c	<i>Heterorhabdus papilliger</i>	1.76 ^{b,c}
<i>Aetideus armatus</i>	1.73 ^{b,c,d}	<i>Heterostylites longicornis</i>	3.00 ^c
<i>Anomalocera patersoni</i>	3.20 ^c	<i>Isias clavipes</i>	1.25 ^c
<i>Calanoides carinatus</i>	2.18 ^{b,c,e}	<i>Labidocera acutifrons</i>	3.00 ^c
<i>Calanus finmarchicus</i>	2.70 ^{c,e}	<i>Labidocera wollastoni</i>	2.20 ^c
<i>Calanus glacialis</i>	4.60 ^t	<i>Macrosetella gracilis</i>	1.40 ^c
<i>Calanus helgolandicus</i>	2.68 ^{b,c}	<i>Mecynocera clausi</i>	0.84 ^{c,k}
<i>Calanus hyperboreus</i>	6.95 ^{c,e}	<i>Metridia</i> I-IV	0.93 ^f
<i>Calanus</i> I-IV	1.65 ^f	<i>Metridia longa</i>	4.10 ^{c,o}
<i>Calanus tenuicornis</i>	1.74 ^{b,c,e}	<i>Metridia lucens</i>	2.27 ^{b,c,o}
<i>Calanus</i> V-VI	2.48 ^g	<i>Nannocalanus minor</i>	1.71 ^{c,e}
<i>Candacia armata</i>	2.18 ^{c,h}	<i>Neocalanus gracilis</i>	2.76 ^{b,c,e}
<i>Candacia bipinnata</i>	1.95 ^{b,c}	<i>Neocalanus robustior</i>	3.42 ^b
<i>Candacia ethiopica</i>	2.15 ^c	<i>Oithona</i> spp.	0.68 ⁱ
<i>Candacia longimana</i>	3.41 ^{b,c,h}	<i>Paracandacia bispinosa</i>	1.67 ^b
<i>Candacia norvegica</i>	2.75 ^{c,h}	<i>Paracandacia simplex</i>	1.75 ^b
<i>Candacia pachydactyla</i>	2.15 ^c	<i>Parapontella brevicornis</i>	1.37 ^c
<i>Candacia tenuimana</i>	2.14 ^{b,c,h}	<i>Parapseudocalanus</i> spp.	0.70 ⁱ
<i>Candacia varicans</i>	2.20 ^{b,c,h}	<i>Phaenna spinifera</i>	1.80 ^c
<i>Centropages bradyi</i>	1.87 ^{b,c,g}	<i>Pleuromamma abdominalis</i>	2.67 ^{b,c,p}
<i>Centropages hamatus</i>	1.30 ^c	<i>Pleuromamma borealis</i>	1.97 ^{b,c,p}
<i>Centropages typicus</i>	1.55 ^{c,g}	<i>Pleuromamma gracilis</i>	1.76 ^{b,c,p}
<i>Centropages violaceus</i>	1.80 ^{b,c,g}	<i>Pleuromamma piseki</i>	1.73 ^{b,c}
<i>Clausocalanus</i> spp.	1.15 ⁱ	<i>Pleuromamma robusta</i>	3.13 ^{b,c,p}
<i>Copepod nauplii</i>	0.48 ^u	<i>Pleuromamma xiphias</i>	4.13 ^{b,c,p}
<i>Corycaeus</i> spp.	1.57 ⁱ	<i>Pontellina plumata</i>	1.69 ^{b,c}
<i>Ctenocalanus vanus</i>	0.94 ^{b,c,j}	<i>Pontellopsis regalis</i>	4.00 ^c
<i>Diaixis hibernica</i>	1.20 ^c	<i>Pseudocalanus elongatus</i>	1.20 ^q
<i>Diaixis pygmoea</i>	0.95 ^c	<i>Rhincalanus cornutus</i>	3.21 ^{b,c}
<i>Euaetideus giesbrechti</i>	2.04 ^c	<i>Rhincalanus nasutus</i>	3.99 ^{b,c,k}
<i>Eucalanus attenuatus</i>	3.94 ^{b,c,k}	<i>Scaphocalanus echinatus</i>	1.92 ^c
<i>Eucalanus crassus</i>	2.85 ^{b,c,k}	<i>Scolecithricella</i> spp.	1.40 ^c
<i>Eucalanus elongatus</i>	4.69 ^{b,c,k}	<i>Scolecithrix bradyi</i>	1.16 ^{b,c}
<i>Eucalanus monachus</i>	2.13 ^c	<i>Scolecithrix danae</i>	2.05 ^{b,c}
<i>Euchaeta acuta</i>	3.84 ^{b,c}	<i>Scottocalanus persecans</i>	4.80 ^c
<i>Euchaeta gracilis</i>	6.60 ^{b,c}	<i>Scottocalanus securifrons</i>	4.30 ^{b,c}
<i>Euchaeta hebes</i>	2.80 ^{b,c}	<i>Temora longicornis</i>	1.00 ^c
<i>Euchaeta marina</i>	2.72 ^{b,c}	<i>Temora stylifera</i>	1.45 ^{b,c}
<i>Euchaeta media</i>	3.65 ^{b,c}	<i>Tortanus discaudatus</i>	2.00 ^f
<i>Euchaeta norvegica</i>	7.00 ^c	<i>Undeuchaeta major</i>	4.55 ^{b,c,s}
<i>Euchaeta pubera</i>	3.94 ^{b,c}	<i>Undeuchaeta plumosa</i>	3.18 ^{b,c,s}
<i>Euchaeta spinosa</i>	6.32 ^{b,c}	Unid. <i>Candacia</i> spp.	2.31
<i>Euchaeta tonsa</i>	6.50 ^c	Unid. <i>Centropages</i> spp.	1.63
<i>Euchirella brevis</i>	3.50 ^l	Unid. <i>Eucalanus</i> spp.	3.40
<i>Euchirella curticauda</i>	3.90 ^{b,c,l}	Unid. <i>Euchaeta</i> spp.	4.82
<i>Euchirella messinensis</i>	4.84 ^{b,c,l}	Unid. <i>Euchirella</i> spp.	4.24
<i>Euchirella pulchra</i>	3.00 ^l	Unid. <i>Heterorhabdus</i> spp.	2.49
<i>Euchirella rostrata</i>	2.95 ^{c,l}	Unid. <i>Labidocera</i> spp.	2.60
<i>Euterpina acutifrons</i>	0.50 ^c	Unid. <i>Paracandacia</i> spp.	1.71
<i>Gaetanus minor</i>	1.93 ^{c,m}	Unid. <i>Pleuromamma</i> spp.	2.56
<i>Gaidius tenuispinus</i>	3.10 ^{b,c,n}	Unid. <i>Scaphocalanus</i> spp.	1.92
<i>Halithalestris croni</i>	2.30 ^c	Unid. <i>Undeuchaeta</i> spp.	3.86
<i>Haloptilus acutifrons</i>	2.86 ^{b,c}	<i>Urocorycaeus</i> spp.	1.76 ^f
<i>Haloptilus longicornis</i>	1.96 ^{b,c}	<i>Xanthocalanus</i> spp.	5.80 ^c
<i>Haloptilus spiniceps</i>	4.14 ^{b,c}		

NE Atlantic

CPR surveys

1958-2005



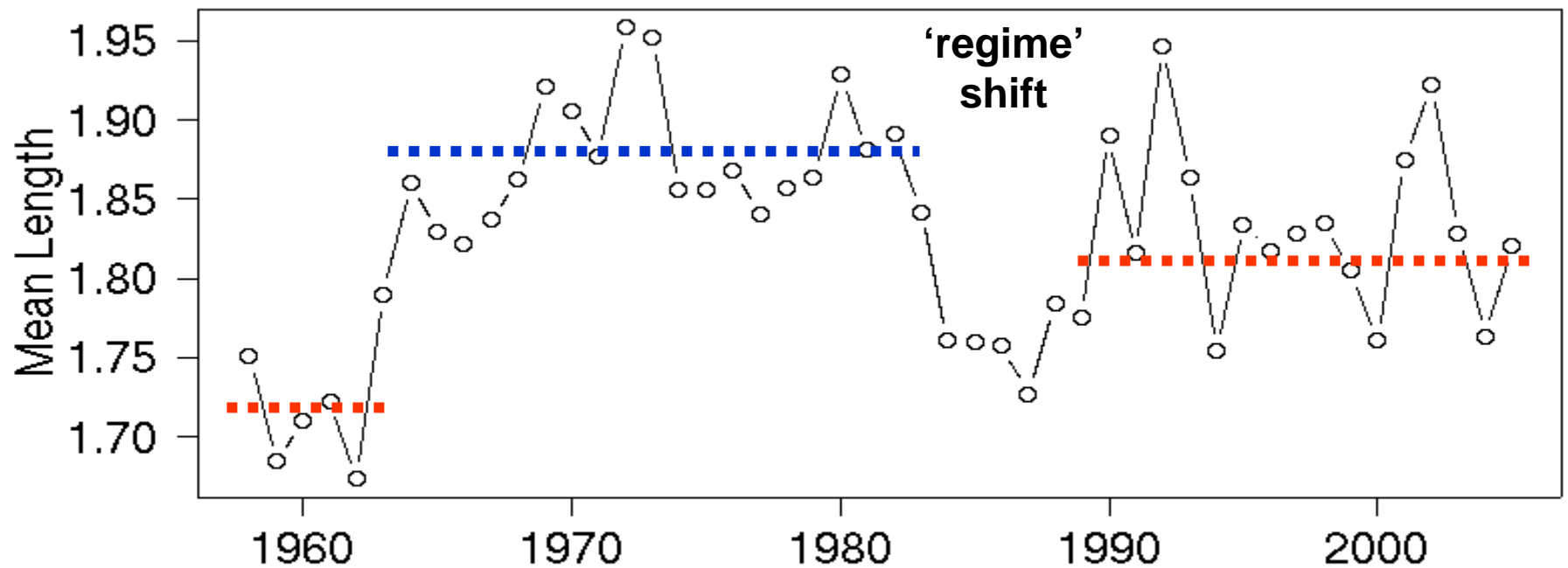
Calanoid copepods:

115 spp.

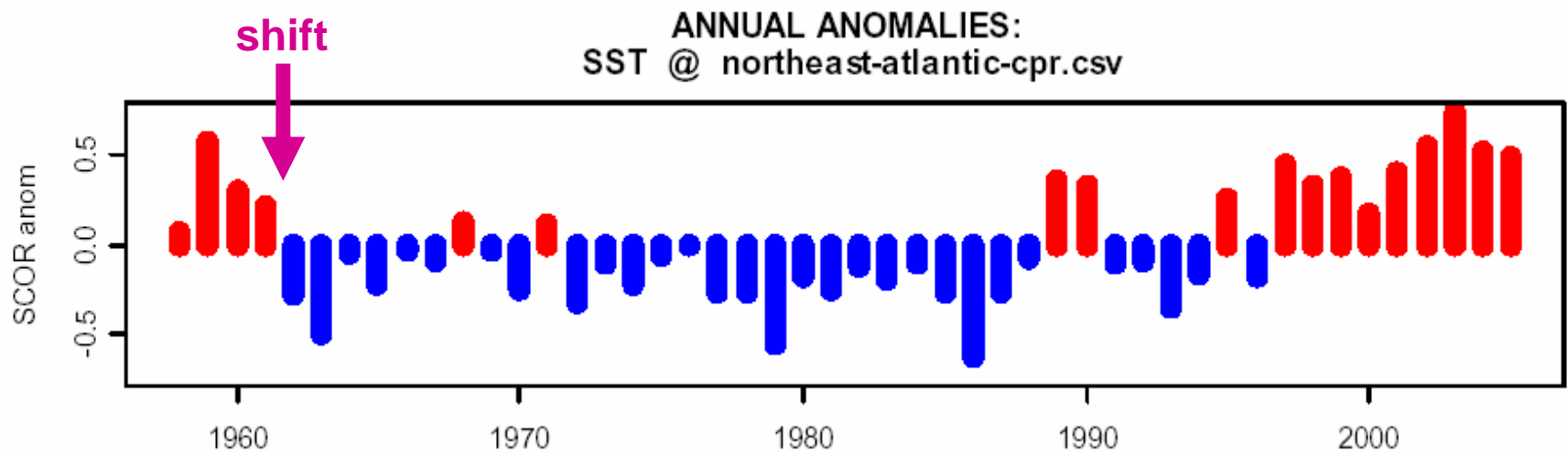
TL: 0.5 – 7.0 mm

data: Ángel López-Urrutia
& Anthony Richardson

Northeast Atlantic (CPR)



NAO
shift



Conclusions

- tracking changes over time of mean copepod communities enables detection of major changes in community structure as a consequence of hydro-climatic variability and shifts in biogeographical distribution
- correlation between **large**-species dominated communities and ocean **cooling**, & between **small**-species dominated communities and ocean **warming**, on regional and decadal scales
- shifts in zooplankton community size structure may have fundamental effects on biogeochemical processes and fisheries
- need access to other existing long-term datasets & application of appropriate statistical analysis techniques

Thank you...!

... and please, send your datasets (copepod female abundance + body size) to: hans.verheye@gmail.com
... or come visit me here...