

***Dawsonia* Nicholson: linguliform brachiopods, crustacean tail-pieces
and a problematicum rather than graptolite ovarian vesicles**

A reassessment of *Dawsonia* Nicholson

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ABSTRACT: Though little is known of the graptoloid reproductive mechanism, graptolites with putatively sac-like appendages, supposedly ovarian vesicles, have been known from the Moffat Shales Group, Southern Uplands, Scotland, for over 150 years. Locally, these co-occur with isolated, two-dimensional, discoidal or ovato-triangular fossils. In the 1870s, Nicholson interpreted these isolated fossils as being graptoloid ‘egg-sacs’, detached from their parent and existing as free-swimming bodies. He assigned them to the genus “*Dawsonia*”, though the name was preoccupied by a trilobite, and named four species: “*D.*” *campanulata*, “*D.*” *acuminata*, “*D.*” *rotunda* (*sic.*) and “*D.*” *tenuistriata*. A reassessment of Nicholson’s type material from the Silurian of Moffatdale, Scotland, and the Ordovician Lévis Formation of Quebec, Canada, shows that *Dawsonia* Nicholson comprises the inarticulate brachiopods *Acrosaccus? rotundus*, *Paterula? tenuistriata* and *Discotreta cf. levisensis*, the tail-piece of the crustacean *Caryocaris acuminata* and the problematic fossil “*D.*” *campanulata*. Though “*D.*” *campanulata* resembles sac-like graptolite appendages, morphometric analysis reveals the similarity to be superficial and the systematic position of this taxon remains uncertain. There is no definite evidence of either “*D.*” *campanulata* or sac-like graptoloid appendages having had a reproductive function.

KEYWORDS: Arenig, Llandovery, Canada, graptolites, Henry Nicholson, reproduction, Scotland, Silurian, taxonomy.

Our knowledge of reproductive structures and strategies in the animal fossil record is sparse. Though reproductive organs are occasionally found, they usually require exceptional preservation for their true nature to be discerned (e.g. Siveter *et al.* 2003; Dunlop *et al.* 2004). Whilst a certain amount is known of the reproductive strategies and mechanisms of the pterobranch hemichordates (Gilchrist 1915; Stebbing 1970; Dilly 1973; Hutt 1991), the extant sister group of the graptolites (Kozłowski 1947, 1948; Towe & Urbanek 1972; Crowther 1978; Cameron 2005), little is known about reproduction in the graptolites themselves (e.g. Urbanek & Jaanusson 1974). Likewise, as little is known of the buoyancy mechanism employed by the graptoloids (Bates 1987), any putatively vesicular graptoloid tissues (e.g. Fig. 1d, f) tend to attract interest and debate (e.g. Underwood 1993; Rickards *et al.* 1994).

There are many gaps in our knowledge of the earliest developmental stages of the graptolites, especially surrounding the events prior to the dispersal of their prosiculae (cone-shaped larvae). Working on well-preserved material from the Tremadoc of Poland, Kozłowski (1948) showed clutches of eggs and embryos in the autothecae of benthonic graptolites; similar structures have also been found in *Reticulograptus tuberosus*, a bushy tuboid graptolite from Götland, Sweden (Bulman & Rickards 1966). Unlike the graptoloids, benthonic graptolites have two types of thecae. These differentiated thecae have been interpreted as sexual dimorphs, with the smaller bithecae housing the male zooid and the larger autothecae housing the female (Kozłowski 1948), though this has yet to be confirmed. Nevertheless, the loss of bithecae in the graptoloids may indicate that their reproductive strategy altered as they colonised the plankton (Hutt 1991), and neither eggs nor embryos have been found in graptoloid thecae. Some workers have suggested that the graptolite synrhabdosome may represent short-lived congregations of several colonies in sexual congress (e.g.

Zalasiewicz 1984) or asexually developing supercolonies (Ruedemann 1947; Bulman 1970). However, the nature of synrhabdosomes remains enigmatic (cf. Rigby 1993; Underwood 1993; Gutiérrez-Marco & Lenz 1998).

In order to assess whether putatively vesicular graptoloid tissues played a role in reproduction, we have undertaken a thorough re-evaluation of sac-like graptolite appendages (Fig. 1d, f) and also of “*Dawsonia*” Nicholson 1873 (Figs 1, 2 & 3), which was originally interpreted as being an ovarian vesicle of a graptoloid detached from its parent colony. Though this genus has a long history of research, it has “caused confusion ever since it was first described” (Williams, 1981). We initially establish the context in which the fossils were originally interpreted and how they have been subsequently reinterpreted, prior to re-evaluating them based on the original material and on new specimens.

1. History of research

Prior to Kozłowski’s seminal monograph of 1948, the zoological affinities of the graptolites attracted much debate. They were initially thought to be a moss (Von Bromell 1727), to be artefacts (Linnaeus 1768), or even, as Nimmo (1847) suggested, the tail spines of the Indian Ocean ray *Raja pastinaca*, though Nimmo had probably never seen a graptolite (Elles & Wood 1901-1918, p. xiii). Eventually, graptolites were recognised as colonies consisting of a series of cup-shaped orifices (thecae) and they were variously assigned to the Cnidaria and Bryozoa (see summaries in Elles & Wood 1901-1918; Kozłowski 1948; Crowther 1978). As the reproductive strategies employed in these groups differ considerably, no small part of the discussion of their

systematic position focussed on the interpretation of rare, attached, putatively sac-like appendages (e.g. Nicholson 1872; Ulrich & Ruedemann 1931).

1.1 1850-1870: sac-like appendages and ‘graptogonophores’

Graptolites bearing sac-like appendages have been known since the 1850s, but due to “various [unspecified] accidental difficulties” (Logan *in* Hall 1865 and references therein) illustrations were not published until the next decade. Hall (1865) proposed that these appendages were reproductive bodies similar to hydrozoan gonothecae, and suggested that graptolites were closely related to the sertularians. It seems Hall’s work captured the imagination of the young H. Alleyne Nicholson: Hall’s monograph was likely used by Nicholson to aid the identification of the graptolites he collected in the Southern Uplands. Indeed, Nicholson (1866, 1872, 1873) regularly referred to Hall’s work on graptolite reproduction.

Whilst collecting the Silurian strata of the Southern Uplands in the summer of 1866, Nicholson discovered a variety of discoidal and ovato-triangular fossils associated with graptolites (e.g. Fig 2). Though not found attached to graptolites themselves, these fossils resemble the supposed reproductive organs described by Hall (1865), and Nicholson (1866) argued that they represented graptolite ovarian vesicles which had detached from their parent colony, and called them ‘graptogonophores’. He supported Hall’s argument for a hydroid affinity for the graptolites, suggesting that the concentrically ‘ribbed’ discoidal specimens represented vertical compressions of a sertularian-like graptogonophore, and that the ovato-triangular specimens were preserved in profile. Nicholson supported his interpretation by illustrating several examples where the discoidal and ovato-triangular specimens were closely associated with graptoloids (e.g. Fig 1c).

However, this work was controversial, drawing a vociferous reply from William Carruthers who argued that the associations of the supposed graptogonophores with graptolites were no more than fortuitous juxtapositions, and that the discoidal specimens most likely represented the brachiopod *Siphonotreta micula* (Carruthers 1867a). This precipitated a lengthy correspondence, with Nicholson (1867a, 1867b, 1867c, 1868b) arguing that graptolites were hydrozoans, largely on the basis of their reproductive strategy, whilst the more vehement Carruthers (1867b, 1867c, 1868a, 1868b) stated that considerations of zoological position should be based on ‘normal’ characters such as colony construction rather than on rare and ambiguous evidence. The latter felt that whilst graptolites were closely allied to the Hydrozoa, they also shared characters with the Polyzoa. As much by perseverance as by any tendency to provide new information, Nicholson’s view that graptolites were extinct hydrozoans became more widely accepted. Nicholson’s work subsequently focussed more on corals than graptolites (see Benton 1979); and, at this time, graptolite research itself also moved away from more theoretical discussions of their affinity with workers such as Lapworth focussing on the more practical concerns of taxonomy and biostratigraphy (e.g. Elles & Wood 1901-1918; Oldroyd 1990; Rushton 2001).

1.2 1870-1900: Graptolite reproduction and “*Dawsonia*” Nicholson

Nicholson’s theory of graptolite reproduction supposed that once sufficiently mature, the ‘graptogonophore’ detached itself from its connection with the parent colony and became a free-swimming zooid (Nicholson 1868a). This assertion was supported in part by evidence for the co-occurrence of ‘graptogonophores’ and graptolites furnished with ramifying fibres (e.g. Fig. 1f) in the same strata (Nicholson 1872).

Nicholson (1873) noted that there were several distinct types of graptogonophore in the Ordovician Lévis Fm. at Point Lévis, Quebec. As it would be almost impossible to relate these back to the individual graptolite species they came from, he referred them to the form genus "*Dawsonia*", much in the manner that one names ichnotaxa independently of the animal that constructed them. Nicholson (1873) named four species: "*D.*" *acuminata*, "*D.*" *rotunda*, "*D.*" *tenuistriata*, and "*D.*" *campanulata*. Gurley (1896), also working in Point Lévis, added two further species: "*D.*" *monodon* and "*D.*" *tridens*.

These species have disparate temporal and geographical ranges. "*D.*" *campanulata* is only known from Early Silurian strata of the British Isles (Nicholson 1873; Lapworth 1876, 1876-7; Marr & Nicholson 1888; Peach & Horne 1899; Williams 1981, 1996), though it occurs in both Laurentia (in Moffatdale, Scotland, and Coalpit Bay, Donaghadee, Northern Ireland) and Avalonia (the English Lake District), which were on either sides of the Early Palaeozoic Iapetus Ocean at this time. "*D.*" *rotunda* and "*D.*" *tenuistriata* are only found in the Ordovician Quebec Group at Point Lévis, Canada (Nicholson 1873), and Gurley's species have only been recorded in the Ordovician of North America at Point Lévis, Quebec, and the Deep Kill, near Melrose, New York (Ruedemann 1904, 1934; Vannier *et al.* 2003). However, "*D.*" *acuminata* appears to be more cosmopolitan, with Nicholson (1873) stating that it occurs at both Point Lévis, Canada, and in northern England. Nicholson (1873) noted that his concept of "*D.*" *acuminata* was similar to "*D.*" *campanulata* in both stratigraphic range and form, though he acknowledged that its outline was notably more triangular and its mucro (proximal termination) less sharply delineated.

Nicholson (1873) believed that his localities in the UK and Canada were contemporaneous and of similar age to graptolites bearing sac-like appendages (Hall

1865, Nicholson 1872), though graptolite biostratigraphy now reveals this to be untrue (cf. section 6 & refs in Fig 5a). However, in the 1870s the age of the strata in the Southern Uplands sections was somewhat of an enigma (Oldroyd 1990; Rushton 2001), making correlation with North American sections problematic.

1.3 Twentieth century work on graptolite reproduction

The 20th century saw graptolite reproduction become a less prevalent area of research, and since a pterobranch affinity for graptolites has been clearly demonstrated (Kozłowski 1947, 1948; Towe & Urbanek 1972; Crowther 1978), Nicholson's work has become largely overlooked. However, biserial graptoloids with sac-like appendages unquestionably attached to their rhabdosome continued to be described as reproductive structures in the early twentieth century (e.g. Elles 1940). Likewise, Ulrich and Ruedemann (1931) reported dendroid graptolites with swollen, oval appendages purportedly homologous with bryozoan ooecia. However, these correspond to bithecae in terms of position and arrangement, and the specimens are too poorly preserved to discern their precise nature (Kozłowski 1948).

The discovery of eggs and embryos inside the autothecae of benthic graptolites (Kozłowski 1948; Bulman & Rickards 1966) led to a reinterpretation of sac-like appendages in graptolites. Kozłowski (1948) considered Hall's supposed 'egg sacs' to be chitinous envelopes associated with the zooids, though he did not speculate on their function, whilst Bulman (1964) figured several similar specimens in an early discussion of graptolite hydrodynamics. Similarly, more recent discussions of graptolite reproduction have overlooked these and other supposed reproductive organs (e.g. Crowther 1978; Hutt 1991; Underwood 1993). For example, the branching appendages described in Ruedemann (1936) were thought to represent epibionts

(Kozłowski 1948). The swollen, oval appendages documented by Ulrich and Ruedemann (1931) might also be epibionts: Kozłowski (1965) showed that *Cephalocystis graptolithifilius*, a similar structure found on other graptolites, is in fact a cephalopod egg capsule comparable to those of the recent *Sepia officinalis* which encrusts the sea grass *Zostera*. Similarly, Underwood (1993) suggested that the putative cases of connection between graptogonophores and graptolites, as illustrated by Nicholson (1866), could plausibly represent parasitic outgrowths or epizoans colonising a graptolite ‘benthic island’ *sensu* Kaufmann (1978).

However, these and other examples of sac-like graptoloid appendages (e.g. Fig 1d, f) are certainly distinct from the unambiguous parasites figured by Bates and Loydell (2000), but are superficially similar in form and preservation to “*D.*” *campanulata* and “*D.*” *acuminata*. As such they require re-examination. Since Kozłowski’s influential work ‘graptogonophores’ have generally been described as sac-like or vane-like appendages with little comment as to their function (e.g. Bulman 1964; Koren’ & Rickards 1997).

1.4 A pterobranch-like model for graptolite reproduction?

Since scanning electron microscopy has been employed for studies of graptolite ultrastructure (e.g. Towe & Urbanek 1970; Crowther 1978, 1981), an affinity for graptolites with the pterobranch hemichordates has become widely accepted (e.g. Dilly 1993; Cameron 2005; Maletz *et al.* 2005). As such, recent discussions of reproduction in graptolites (e.g. Hutt 1991) have been premised on the belief that graptolites adopt pterobranch-like mechanisms (cf. Gilchrist 1915; Stebbing 1970). The pterobranchs *Rhabdopleura* and *Cephalodiscus* reproduce both sexually and asexually (Hutt 1991). Though most zooids in *R. compacta* are neuter or sexually

immature, certain zooids may metamorphose and develop either an ovum or testis (Stebbing 1970). Whilst the sexes are separate in *Rhabdopleura*, certain species of *Cephalodiscus* colonies may be hermaphroditic, with certain zooids bearing both male and female reproductive organs (Horst 1939, Bulman 1970). Though its colonies are sessile and its zooids have limited movement, *Rhabdopleura* undergoes internal fertilisation, with its oviduct serving only as a conduit for sperm to reach the ova (Stebbing 1970). In both *Rhabdopleura* and *Cephalodiscus*, clutches of embryos remain in the creeping tube until they mature as ciliated, lecithotrophic larvae (Dilly 1973; Lester 1988a). The larva leaves the creeping tube as a free-swimming individual which secretes a collagenous, dome-shaped prosiculum (Dilly 1973; Dilly & Ryland 1985). Later, the larva metamorphoses under the prosiculum and emerges as a juvenile (Dilly & Ryland 1985; Lester 1988b) that settles on the substrate and later asexually buds to form a colony (Stebbing 1970; Dilly 1973).

Though there is little direct evidence of reproduction in the graptolite fossil record (cf. Hutt 1991), eggs and embryos have been reported in certain specimens (Kozłowski 1948; Bulman & Rickards 1966). This would be consistent with graptolites producing a free-swimming lecithotrophic larva, that later secretes a prosicula (equivalent to the pterobranch prosiculum) in the plankton before maturing (cf. Williams & Clarke 1999) and budding to form a colony.

1.5 Historical interpretations of “*Dawsonia*” Nicholson

1.5.1 “*D.*” *monodon*, “*D.*” *tridens*, “*D.*” *acuminata*, and “*D.*” *campanulata*:

crustaceans, molluscs or algae? These mucronate species have received more attention than the other species, which are dealt with below. “*D.*” *monodon* and “*D.*” *tridens* were originally described by Gurley (1896), but have long been considered to

be tail-pieces of the crustacean *Caryocaris* (Ruedemann 1934; Rolfe in Theokritoff 1964). Rolfe (1969, p. 316) stated that “*Dawsonia*” is a junior synonym of *Caryocaris* Salter (1863), but did not re-examine Nicholson’s material (Ian Rolfe, pers. comm.). Though “*D.*” *acuminata* has all but vanished from the literature, “*D.*” *campanulata* is often used.

Lapworth (1876-77) considered “*D.*” *campanulata* to be a member of the crustaceans in his catalogue of fossils from western Scotland, perhaps due to its similarity and common co-occurrence with the putative crustaceans *Aptychopsis* Barrande (1872), *Peltocaris* Salter (1863), and *Discinocaris* Woodward (1866) (Lapworth 1876, 1876-77; Marr & Nicholson 1888; Peach & Horne 1899). These putative crustaceans look similar to certain “*Dawsonia*” species: disarticulated valves of *Peltocaris* and *Aptychopsis* are similar to “*D.*” *campanulata* in outline, and *Discinocaris* has an ornament similar to that of “*D.*” *rotunda* and “*D.*” *tenuistriata*. However, none of the “*Dawsonia*” material examined in this study bears the characteristic dovetail symmetry that characterises complete specimens of these other taxa. Gürich (1928) also compared “*D.*” *campanulata* to *Peltocaris*, which he considered to be the covers of a hyolithid or chiton-like organism. However, this work offered no firm conclusions as to the affinities of “*Dawsonia*” and we have found no evidence of either hyolithids or chitons co-occurring in the same strata as it.

The affinities of *Aptychopsis*, *Peltocaris*, and *Discinocaris* remain uncertain. Rolfe (1969, p. 328) noted that “they have been compared and confused with graptolite ‘swim bladders’ and ‘gonangia’, eurypterid metastomata, hyolith opercula, polyplacophoran plates, bivalves, arthodire dermal plates, and branchiopod carapaces,” and he noted there was no evidence to support an affinity for either *Aptychopsis*, *Peltocaris*, or *Discinocaris* with the phyllocarids. Rolfe (1969, pp. 328-

329) suggested that these taxa may perhaps represent the aptychi of soft-bodied cephalopods rather than being crustacean carapaces. However, he did not go as far as synonymising *Aptychopsis*, *Peltocaris*, and *Discinocaris* with the aptychus morpho-genus *Sidetes* Giebel *sensu* Moore & Sylvester-Bradley (1957).

There is no good reason to group “*D.*” *campanulata* with these supposed aptychi. Indeed, neither Gürich’s (1928) work nor a crustacean affinity gained serious consideration in the most recent re-examination of “*D.*” *campanulata*, which tentatively reinterpreted it as an alga (S.H. Williams 1981).

1.5.2 “*D.*” *rotunda* and “*D.*” *tenuistriata*: possible brachiopods? Neither “*D.*” *rotunda* nor “*D.*” *tenuistriata* are mucronate: together they form a group of small, subcircular shelly-fossils. Though Nicholson (1873) stated that “*D.*” *rotunda* and “*D.*” *tenuistriata* appeared too variable in form and appearance to be inarticulate brachiopods, this assertion was questioned from the outset (Carruthers 1867a; Ruedemann 1904, 1934). More recently, Benton (1979) noted that some of the Nicholson’s type material may be small brachiopods.

1.5.3 Misdiagnoses. Several incompatible forms have been erroneously assigned to “*Dawsonia*” Nicholson principally because little or no reference was made to the type specimens. As Benton (1979) noted, the trace fossil *Lockeia* U.P. James was misdiagnosed as “*Dawsonia*” by U.P. James’s son, J.F. James (1885, 1892). As “*Dawsonia*” is preserved as a body fossil, this name clearly cannot be applied to a trace fossil (Häntzschel 1965, 1975; Osgood 1970). However, the name “*Dawsonia cycla*” is still used for another fossil from the Cincinnati area which consists of small, black, shiny discs that are found encrusting the surfaces of nautiloid conchs. Though

Frey (1989) thought that these discs may represent the attachment sites of the dendroid graptolite *Mastigograptus*, they are now thought to represent the epibiont *Sphenothallus* (Neal & Hannibal 2000).

1.6 The current status of “*Dawsonia*” Nicholson

The name “*Dawsonia*” is still widely used by graptolite workers (e.g. Williams 1996), though now it is almost exclusively used as shorthand for “*D.*” *campanulata*, which is its type species (Miller 1889). No consensus as to its taxonomic status or systematic position has yet emerged. Though the genus “*Dawsonia*” was conceived to describe the egg sacs of a sertularian-like hydroid, this name has been applied to unrelated fossils from all of the major divisions of the bilateria. Its type species, “*D.*” *campanulata* was most recently interpreted as an alga (Williams 1981). *Dawsonia* Nicholson is junior homonym of the trilobite *Dawsonia* Hartt in Dawson (1868). With all the above in mind, Nicholson’s genus is in need of taxonomic revision.

2. Material and methods used in this study

2.1 Nicholson’s types and comparative material

Much of Nicholson’s type and figured material is housed in the Natural History Museum [NHM], London, which purchased a collection of 1400 graptolites from Nicholson in 1883 (Benton 1979). The unfigured portion of Nicholson’s collection remains in the Aberdeen University Geology Department, and is catalogued in Benton and Trewin (1978). Nicholson’s material from the Lake District lies in the Harkness and Marr collections of the Sedgwick Museum, Cambridge [SM].

The type material of “*Dawsonia*” Nicholson, as recognised by Benton (1979) is in the G.J. Hinde collection of the NHM. It has been re-examined and re-accessioned as part of this study. Nicholson did not identify any specimens from UK strata in this collection as “*D.*” *acuminata*, despite mentioning its occurrence in northern England (Nicholson 1873). Given that Nicholson’s illustrations are often idealised woodcuts taking features from several specimens (Benton & Trewin 1978), it has been impossible to precisely determine his type specimens. However, as Nicholson’s diagnoses can be recognised from his distinctive handwriting on the manuscript specimen labels (Fig 2n,o), we have assigned lectotypes for each of his four species. “*D.*” *campanulata* remains the type species of the genus (secondary diagnosis, Miller 1889 *contra* Ruedemann 1904, 1934). Other comparative material is housed in the British Geological Survey [BGS] collections at Keyworth, near Nottingham, the Ulster Museum, Belfast [BEL] and in the Lapworth Museum, University of Birmingham [BU].

In order to compare “*Dawsonia*” with the sac-like appendages of graptolites, we undertook an extensive search of museum holdings and appropriate literature. “*D.*” *campanulata*-bearing localities in Moffatdale, southern Scotland, and the Lake District of England were also recollected to provide an unbiased sample of this species. We were unable to collect field specimens of graptolites bearing sac-like appendages, perhaps due to their relative rarity, and we rely entirely on museum collections for such graptolites.

In addition to the occurrences of “*Dawsonia*” noted in section 1.2 and above, Ruedemann (1904, p. 734) commented that “[“*D.*” *campanulata*] is very common in the Trenton (Normanskill) graptolite shales of New York and Canada.” However, we have not been able to identify this fossil amongst Ruedemann’s original collections,

although there are plenty of graptolites bearing ‘graptogonophores’ in his material. In addition, Ruedemann (1908) reported that in 1889, H.M. Ami named three new species of “*Dawsonia*” from graptolitic strata in the St Lawrence region of Canada. However, we have been unable to find any trace of Ami’s “*Dawsonia*” species in either the literature or in museum collections. Likewise, we have been unable to find Gurley’s type specimens of “*D.*” *monodon* and “*D.*” *tridens*. Though they were once held in the collections of the New York State Museum, Albany, NY [NYSM] (Ruedemann 1934 & references therein), they are no longer in its possession. When Ruedemann (1904) illustrated “*D.*” *monodon* and “*D.*” *tridens* specimens from NYSM collections, he chose examples from the Quarry at the Deep Kill, near Melrose, New York, only copying Gurley’s drawings of the Point Lévis material. This suggests that they were not in the NYSM at that time either. It may be that the specimens went missing at the very end of the 1800s when a long-term budget deficit led James Hall to sell many specimens to keep the Geological Survey and State Museum afloat (Ed Landing, pers. comm. 2004).

Though Ruedemann’s (1904, 1934) material has been re-examined for comparative purposes, neither those specimens nor Nicholson’s Point Lévis specimens clearly preserve the carapace. Given that *Caryocaris* taxonomy is primarily based on carapace morphology, we are unable to determine whether Ruedemann’s specimens are truly synonymous with Gurley’s species. As such, this paper focuses on clarifying Nicholson’s concept of the “*Dawsonia*” species, rather than entering the more nebulous realm of phyllopod systematics.

2.2 Methods used and terminology employed

All fossils have been studied under reflected light microscopy. Additionally, uncoated specimens were examined at 15KV in backscatter mode in Hitachi S-3600N and LEO 435VP SEMs, with phases identified using energy dispersive X-ray analyses (EDS) using Oxford Instruments INCA and ISIS software, respectively. The electron microscope techniques used closely follow those described in Martill *et al.* (1992) and Orr *et al.* (2002). Illustrated images have been digitally enhanced to increase the contrast between fossil and matrix.

Details of repositories and specimen numbers are listed with the appropriate figures and in the systematic section; details of the criteria used in the morphometric analysis are given in Figure 5. As the brachiopod taxa were often incomplete, morphological measurements were taken on well-preserved growth-lines as well as on outlines, though in each case these are clearly distinguished in the appropriate figure caption. All measurements were made on camera lucida drawings of x40 or x50 optical magnification, and recorded to an accuracy of greater than one percent.

Morphological terms used in systematic descriptions are as employed in Holmer & Popov (2000) for the brachiopod species, in Rachebouef *et al.* (2000) and references therein for the *Caryocaris* tail-pieces, and defined in Figure 5 for “*D.*” *campanulata*. Because Nicholson’s Point Lévis material consists entirely of disarticulated specimens, we have used morphological criteria to assess which forms could plausibly conjoin based on the present understanding of inarticulate brachiopods and *Caryocaris* in order to avoid unnecessary taxonomic inflation. Abbreviations used in the synonymy lists are those of Matthews (1973) and the qualifiers used in open nomenclature may be found in Bengtson (1988).

3. The nature of “*Dawsonia*” Nicholson

It is clear that “*Dawsonia*” is polyphyletic. The lectotype of “*D.*” *acuminata* is a furcal ramus from the tail-piece of the crustacean *Caryocaris acuminata* (Fig. 4). Other fossils within the fauna include telsons and carapace fragments which are considered conspecific given the present understanding of *Caryocaris* morphology (Fig. 4d). The lectotypes of “*D.*” *rotunda* and “*D.*” *tenuistriata* are linguliform brachiopods (Fig. 3c,d) and have been tentatively re-assigned to the genera *Acrosaccus* and *Paterula* respectively, and Nicholson’s type collection also contains a form provisionally identified as *Discotreta* cf. *levisensis* (Fig. 3f). As no articulated specimens are present we cannot unambiguously determine which shells articulated in life. However, two of the four discrete shell morphotypes shown in Figure 5c have indistinguishable outlines (with W/L ~1) and probably represent an unequivocal species. The other two shell morphotypes could not plausibly co-join (cf. Fig 5c), consistent with there being three species present in the collection. The systematic palaeontology of these taxa is dealt with in section 6.

The style of preservation of the above listed dawsoniids is different from the graptolites which co-occur in the Point Lévis fauna, suggesting that they were originally composed of non-graptolitic material. The graptolites are preserved as dull, black compressions, whereas the dawsoniids are generally in relief, having a horny texture and some having a bronze, pyritous sheen. EDS analyses of the Point Lévis dawsoniids specimens reveals that they are preserved as phosphate with some associated pyrite (Fig. 6f,g). This composition is consistent with these taxa being linguliform brachiopods and caryocarid arthropods rather than graptolites, which are carbonaceous. Therefore, these species of “*Dawsonia*” are reassigned to their

appropriate clades and can be discounted from any consideration of graptolite reproduction.

Though Nicholson (1873) mentioned the occurrence of “*D.*” *acuminata* in English strata, we have been unable to identify it in UK collections. Morphometric analysis reveals that there is some overlap between “*D.*” *campanulata* and *C. acuminata* (Fig. 5b, d). However, even the most slender “*D.*” *campanulata* can be clearly distinguished from *C. acuminata* by the presence of a delineated mucro, its rounded latero-distal margin (Fig. 3), and its composition (Fig. 6a-d). It therefore seems most likely that Nicholson was either referring to slender “*D.*” *campanulata* morphotypes as “*D.*” *acuminata*, or he was perhaps confusing the tail-pieces of *Caryocaris wrighti* which occur in strata of the British Isles (Rushton & Williams 1996; Vannier *et al.* 2003) with *C. acuminata*. To avoid unnecessary confusion, we have included Nicholson’s so-called “*D.*” *acuminata* from the British Isles within our amended definition of “*D.*” *campanulata*, with *C. acuminata* only referring to his Canadian material.

There is little similarity between “*D.*” *campanulata* and *Caryocaris* tail-pieces (cf. Figs 1 & 4), or indeed with the Point Lévis dawsoniids, the most notable differences being in its composition and outline. It is preserved as an organic compression (Fig 6a-d) unlike *Caryocaris*, which is preserved in phosphate (Fig 6g). It is more symmetrical than either the carapace or furcal ramus of a *Caryocaris*, and notably more ovate than the *Caryocaris* telson. Its mucro is too centrally positioned to represent either an anterior horn or a postero-dorsal spine of the *Caryocaris* carapace, and it differs from the marginal spinules of the *Caryocaris* ramus in terms of size and position. Unlike a furcal ramus, the “body” of “*D.*” *campanulata* is ovato-triangular rather than ovato-parallelogrammic, and it lacks a serrated lateral margin. Though

lacking a mucro, the grossly teardrop form of the *Caryocaris* telson is similar in shape to the “body” of “*D.*” *campanulata*. However, morphometric analysis (Fig. 5d) reveals no overlap between *Caryocaris* telsons (where $D/L < 0.2$) and “*D.*” *campanulata* (where $0.2 < D/L < 0.6$).

Though “*D.*” *campanulata* shares a similar preservation style to the sac-like appendages seen in graptolites (Fig. 6a-e), there is no evidence to support a homology. Whilst both are found as silvery organic films in the black shales of the Southern Uplands, “*Dawsonia*” cannot be recognised as a graptolite (Bulman 1970). Morphometric analysis reveals that the similarity between the two is superficial, with the graptolite appendages having a consistently more distal centroid (Fig. 5a). They are also more asymmetrical and more variable in their form than “*D.*” *campanulata*, and there is no discrete transition between their connecting rods and their lobate distal part, which is quite unlike the transition between the mucro and the lobate “body” in “*D.*” *campanulata*. Though “*D.*” *campanulata* may be found in close association with graptolites, it is not attached to them; instead, they may be merely juxtaposed (e.g. Fig 1c). Neither is there any good evidence to suggest that “*D.*” *campanulata* is a sac-like appendage broken from a graptolite. For one thing, its mucro tapers to a narrow point, rather than having a blunt or irregular end. Indeed, the connecting rod and margins of the sac-like graptolite appendage seem unlikely to break readily. They are well-defined in most specimens, having a similar mode of preservation to the nema. This suggests they are recalcitrant tissues and may have possessed a noticeable elasticity: Crowther (1978) noted that the nema displays a certain ‘springiness’ in acid-isolated specimens. In summary, there is no evidence for “*D.*” *campanulata* being a graptolite egg-sac, or for it being related to graptolites at all.

A concentric, raised, nipple-like structure occurs in several specimens of “*D.*” *campanulata* (e.g. Fig. 2f) and has previously been interpreted as evidence for it having originally had a hollow body (Williams 1981). Nicholson (1872, 1873) believed that this ‘nipple’ represented compression of a hollow three-dimensional egg-sac onto its more rigid mucro. However this does not appear to be the case, as many specimens reveal both a mucro and a ‘nipple’ (e.g. Fig 1c), and some specimens show that “*Dawsonia*” may only partially overlap a ‘nipple’ (Fig . 6e). Instead, SEM investigation reveals the nipples to be composed of diagenetic pyrite adopting a rounded and concentric habit (cf. Allison 1988; Underwood & Bottrell 1994). As such, the ‘nipple’ is best considered to be a product of compression of “*Dawsonia*” onto pyrite formed in early (?pre-compaction) diagenesis, rather than an intrinsic part of the fossil.

Detailed examination of the sac-like appendages of *Hallograptus bimucronatus* reveals that concentric lines are also present in them (Fig. 1f), cross-bracing better preserved margins. However, they are consistent with being the remnants of fusellar structures like those seen in the *Orthoretiolites hami* scopula (Bates & Kirk 1991). Such a mode of fabrication would deny the possibility that these structures formed a housing from which an ‘egg-sac’ could easily detach as Nicholson (1868a, 1872) suggested. Nicholson believed that “*D.*” *campanulata* represented a graptolite egg-sac that became a free-swimming entity, supposing that it was hollow and filled with eggs whilst housed in a cup of ramifying fibres connected to the graptolite. He proposed that this ‘ovarian vesicle’ slid out once it was able to swim freely. However, if *Hallograptus* constructed its appendages in a manner comparable to the scopula of *O. hami*, it would represent a plate-like, rather than cup-like, structure (cf. Bates & Kirk 1991).

Indeed, it is doubtful whether sac-like graptolite appendages represent egg-sacs. The preponderance of these features in scalariform preservation suggests that they originated from the interthecal wall rather than connecting the thecae *per se*, so there is no direct evidence for their intimate connection with the zooid itself. Moreover, their regularity of form is inconsistent with what one would expect of an unambiguously vesicular structure such as the *Climacograptus wilsoni* vesicle (Williams 1994). Given that these structures are only known in the biserial graptolites, it seems unlikely that they are related to graptolite reproduction.

4. Discussion

Nicholson remains one of the great early graptolite workers, despite being wrong in his views of graptolite reproduction (Nicholson 1866, 1872, 1873, etc.). It was not until Clupáč (1970) discovered well-preserved caryocarids in limestone nodules from the Ordovician of Bohemia that the morphology of their tail-piece was fully understood; hence, it is understandable that Nicholson (1873) did not recognise “*D.*” *acuminata* as such, despite recognising *Caryocaris* carapaces in the Point Lévis fauna. Nicholson’s assertion that “*D.*” *rotunda* and “*D.*” *tenuistriata* were not brachiopods appears at odds with his (1867a) claim that “it is impossible that any palaeontologist, possessed of ordinary powers of observation, should fall into an error so gross [as to fail to recognise an inarticulate brachiopod]”.

Noting the variability of form within “*D.*” *tenuistriata*, for example, Nicholson (1873, p. 142) argued that describing the species as egg-sacs allowed for greater morphological plasticity, otherwise “we should have to believe there were four

or five distinct species of brachiopods in these beds which is very unlikely” (the information in Benton (1979) confirms that Nicholson was not accustomed to such faunal diversity in UK sections). It appears that he conflated the beak of the brachiopods with the variably positioned “nipple” of “*D.*” *campanulata* (e.g. Fig. 2), a false homology that underpinned his “*Dawsonia*” concept. So, in an age before taphonomy and palaeoenvironment were generally considered, when many species were only known from disarticulated fragments, Nicholson explained the vagaries of variable preservation and differing morphology in a strikingly diverse fauna by appealing to his theory of graptolite reproduction.

5. Conclusions

There is no evidence to support the notion that “*Dawsonia*” is related to graptolite reproduction. Likewise, there is no strong case for sac-like appendages on graptolites having a reproductive function, given the discovery of eggs and embryos in the thecae of benthonic graptolites, and our knowledge of reproduction in the pterobranchs (Kozłowski 1948; Bulman & Rickards 1966; Stebbing 1970; Dilly 1973; Hutt 1991). As such, the function of these sac-like appendages remains enigmatic, and these, along with the function of synrhabdosomes, should be re-examined in order to assess what, if any role either plays with regard to graptolite reproduction.

All known species of “*Dawsonia*” have been reassigned to valid genera except “*D.*” *campanulata*, which is best considered a problematicum. “*D.*” *acuminata* Nicholson, “*D.*” *tridens* Gurley and “*D.*” *monodon* Gurley represent the tail-pieces of *Caryocaris acuminata* (Nicholson 1873). We suggest that *C. monodon*

(Gurley) should not apply to specimens from Point Lévis (*contra* Ruedemann, 1934). “*D.*” *rotunda* Nicholson is tentatively reassigned to the brachiopod genus *Acrosaccus*, and “*D.*” *tenuistriata* Nicholson is accommodated by the brachiopods *?Paterula tenuistriata* and *Discotreta cf. levisensis* (Walcott 1908). The trace fossil misdiagnosed as *Dawsonia* Nicholson by J.F. James (1885, 1892) has long been known to represent the trace fossil *Lockeia* U. P. James (1879) (see Benton 1979), whilst “*Dawsonia cyclo*” most likely represents the epibiont *Sphaenothallus* (Frey 1989; Neal & Hannibal 2000).

“*D.*” *campanulata* is a problematicum, currently being investigated by the authors. Though *Dawsonia* Nicholson is an invalid generic name, it would be premature to formally re-describe it until further information pertaining to the affinity of “*D.*” *campanulata* is available. That nobody has provided a more definite idea of what “*D.*” *campanulata* may represent in the hundred years since Nicholson’s early death can be taken as a minor tribute to the man who clearly recognised its uniqueness.

6. Systematic Palaeontology

Phylum *Arthropoda*, von Siebold & Stannius, 1845

Superclass *Crustacea* Pennant, 1777

Class *Malacostraca* Latrielle, 1806

Subclass *Phyllocarida* Packard, 1879

Order *Archaeostraca* Claus, 1888

Family *Caryocarididae* Racheboeuf, Vannier & Ortega, 2000

Genus *Caryocaris* Salter 1863

- * 1863 *Caryocaris* n. gen. Salter, p. 139.
- non 1868 *Dawsonia* Hartt in Dawson, p 655.
- p. 1873 *Dawsonia* Nicholson, pp. 139-140 *pars*.
- 1896 *Dawsonia* Nicholson; Gurley, p. 88.
- 1904 *Caryocaris* Salter; Ruedemann, pp. 738-742.
- 1969 *Caryocaris* Salter; Rolfe in Moore, p. 316.
- 2000 *Caryocaris* Salter; Racheboeuf, Vannier & Ortega, pp. 322-323.

Remarks. The synonymy above is in addition to the detailed list in Racheboeuf, Vannier & Ortega (2000). In the absence of a carapace, a tail-piece consisting of elongate, leaf-shaped furcal rami and a shorter, narrow triangular telson is sufficient to diagnose the genus (Racheboeuf, Vannier & Ortega, 2000, p. 328).

Caryocaris acuminata (Nicholson 1873)

(Figures 4 & 6g)

- vp. 1873 *Dawsonia acuminata* n. gen. et n. sp. Nicholson, pp. 140-141, Figs 3a-a' *pars*.
- v. 1873 *Caryocaris* sp. Nicholson, p. 143.
- . 1896 *Caryocaris oblongus* n. sp. Gurley, p. 87, Pl. 4, Fig. 2.
- p. 1896 *Caryocarus* [sic] *curvilatus* n. sp. Gurley, pp. 87-88 *pars*, ?Pl. 4, Fig. 3, ?Pl. 5, Fig. 3.
- . 1896 *Dawsonia monodon* n. sp. Gurley, p. 88, Pl. 5, Fig. 4.

- . 1896 *Dawsonia tridens* n. sp. Gurley, p. 88, Pl. 5, Fig 5.
- non* 1904 *Caryocaris* cf. *curvilineatus* [sic] Gurley; Ruedemann, p. 738, Pl. 17, Fig. 17.
- non* 1904 *Caryocaris* cf. *oblongus* Gurley; Ruedemann, p. 738, Pl. 17, Figs 14-16.
- p. 1904 *Dawsonia tridens* Gurley; Ruedemann, p. 741 *pars*, ?Pl. 17, Fig. 18, *non* Pl. 17, Figs. 19-20 [= *C. monodon*].
- p. 1904 *Dawsonia monodon* Gurley; Ruedemann, pp. 741-742 *pars*, Fig 105, ?Pl. 17, Figs 21-23, *non* Pl. 17 Figs 24-26 [= *C. monodon*].
- non* 1934 *Caryocaris curvilata* Gurley; Ruedemann, p. 92, Pl. 22, Figs 1-9.
- p. 1934 *Caryocaris monodon* (Gurley); Ruedemann, p. 93-95 *pars*, *non* Pl. 22, Figs 10-14.

Type material. NHM P1985.3 lectotype (furcal ramus).

Additional material. Syntypes NHM P1977, P1982.1-3, P1984.1-2, P1985.3-5, P1988.3: 0 complete carapaces; 8 incomplete carapace fragments; 0 articulated tail-pieces; 11 well-preserved furcal rami; 7 telsons; 15 poorly-preserved or fragmentary furcal rami, telsons and indeterminate fragments.

Type locality. Lévis Shale, Point Lévis, Quebec, Canada. Lévis Formation, Ordovician (Arenig).

Diagnosis. Carapace outline indeterminate; linear corrugated ornament on fragments. Tail-piece with narrow triangulate telson lacking ridge or carina; furcal rami elongate, leaf-shaped, ca. 1.5 times longer than telson on average, with acuminate distal margin, bearing large, triangular, posteriorly-directed spines along their outer margin; distinctive narrow ridge and furrow adjacent to its proximal inner margin along its proximal third. Telson ranges from 1.8-2.9 mm in length and 0.9-1.5

mm in width. Furcal ramus ranges from 2.7-6.1 mm in length and 1.3-2.5 mm in width; smaller specimens may have two marginal spines (e.g. Fig. 4b), larger specimens are stouter and more asymmetrical and have three marginal spines (e.g. Fig. 4a, 6g)

Remarks. Until the morphology of its carapace is better known, *C. acuminata* should remain a species separate from *C. monodon* and other caryocarids. *C. acuminata* refers exclusively to caryocarids from Point Lévis and *C. monodon* refers to caryocarids from the exposure at the Deep Kill at Melrose, as laid out in the synonymy above. Though the outline of the tail piece is similar in both localities, the morphology of the tail-piece alone is not well enough placed in the hierarchy of characters to determine synonymy at a species level (Racheboeuf, Vannier & Ortega 2000, p. 328). The variation in number of marginal spines may represent allometric growth (cf. Rushton & Williams 1996); however, small spines may not necessarily be apparent on poorly preserved specimens (see Fig. 4b).

As the type specimens of *C. oblongus* Gurley, “*D.*” *monodon* Gurley and “*D.*” *tridens* Gurley are presumed lost, Nicholson’s material assumes topotype status. Therefore, these species can be suppressed as junior synonyms. This is supported by comparison with Gurley’s original descriptions and illustrations: “*D.*” *tridens* corresponds exactly with our observations on the furcal ramus of *C. acuminata*, whilst “*D.*” *monodon* most likely represents an articulated *Caryocaris* tail-piece preserved in lateral view. *C. oblongus* presumably represents the fragments of a carapace. Nicholson (1873) also noted *Caryocaris* carapace fragments in the Point Lévis fauna. Similarly, *Caryocarus* [sic] *curvilatus*, described as an aberrant graptolite in Gurley (1896), is most likely an articulated abdomen and tail-piece. Likewise, we wholeheartedly concur with Ruedemann (1904, 1934) that “*D.*”

monodon and “*D.*” *tridens* represent parts of a crustacean rather than being unusual graptolites.

Though no articulated specimens are present in Nicholson’s collection, it seems more parsimonious to describe the disarticulated parts as one species rather than several. Nicholson described “*D.*” *acuminata* prior to mentioning the specimens which he referred to *Caryocaris* sp. As such, there seems little controversy in retaining the specific name *acuminata*, which refers to the pointed end of the furcal ramus.

Phylum Brachiopoda Duméril, 1806

Subphylum Linguliformea Williams *et al.*, 1996

Class Lingulata Gorjansky & Popov, 1985

Remarks. We place the three species of brachiopods from Nicholson’s Point Lévis material within the Lingulata on the basis of their organophosphatic composition, rudimentary articulation and larval shells. As noted above, many authors have considered them to be brachiopods, though they have not been formally assigned to the phylum until now.

Order Lingulida Waagen, 1885

Superfamily Linguloidea Menke, 1828

Family Paterulidae Cooper, 1956

Genus *Paterula* Barrande, 1879

non 1868 *Dawsonia* Hartt in Dawson, p. 655.

- p. 1873 *Dawsonia* Nicholson, p. 139-140 *pars*.
 * 1879 *Paterula* n. gen. Barrande, Pl. 110.
 2000 *Paterula* Barrande; Holmer & Popov, 2000, p 75.

Paterula? tenuistriata (Nicholson 1873)

(Figure 3c,e)

- vp. 1873 *Dawsonia tenuistriata* Nicholson, pp. 141-142 *pars*, Figs 3 c-d'.

Type material. Lectotype: NHM P1984.3 (part), P1984.2 (counterpart).

Additional material. NHM P1984.1-3, P1985.3 (5 valves).

Type locality. Lévis Shale, Point Lévis, Quebec, Canada. Lévis Formation, Ordovician (Arenig).

Description. Shell with elongate oval outline, convex. Apex and limbus submarginal to subcentral. Anterior-posterior valve length 1.4->3.4 mm, valve breadth 1.2-3.7 mm, typical specimen breadth >2 mm; length-width ratio 1.2-1.5, typically 1.35; maximum breadth at anterior-posterior midpoint. Growth lines continuous and fine, equally prominent, regular 0.04-0.1 mm spacing throughout the valve.

Remarks. Though this genus is typically unequivalved, only a single valve is present in Nicholson's collections. As *Dawsonia* Nicholson is an invalid taxon there is no conflict of names. While there is some similarity between this form and the younger taxon *P. cf. portlocki* Geinitz (1852) as illustrated by Henningsmoen *in* Waern *et al.* (1948), the material described herein is too poorly preserved to properly compare the taxa. As such, Nicholson's collections need to be supplemented with

additional material exhibiting the shell's internal view before this taxon can be precisely placed. Therefore, we have kept the taxon in open nomenclature.

Superfamily Discinoidea Gray, 1840

Family Discinidae Gray, 1840

Genus *Acrosaccus*? Willard, 1928

- non* 1868 *Dawsonia* Hartt in Dawson, p. 655.
 p. 1873 *Dawsonia* Nicholson, p. 139-140 *pars*.
 * 1928 *Acrosaccus* n. gen. Willard, p. 258.
 2000 *Acrosaccus* Willard; Holmer & Popov, 2000, p. 86.

Acrosaccus? rotundus (Nicholson 1873)

(Figure 3a-b,d)

- v. 1873 *Dawsonia rotunda* [*sic.*] Nicholson, pp. 141-142, Figs 3c-3d'.

Type material. Lectotype NHM P1982.1.

Additional material. Syntypes: NHM P1984.2-3, P1985.3 (13 valves: 2 dorsal, 5 pedicle, 6 indet.)

Type locality. Lévis Shale, Point Lévis, Quebec, Canada. Lévis Formation, Ordovician (Arenig).

Description. Shell unequivalved with subcircular outline, equally biconvex. Beak slightly submarginal on one valve and submarginal to subcentral on the other. Anterior-posterior valve length 2.1-2.9 mm, valve breadth 2.1-3.0

mm; length-breadth ratio 0.95-1.1, typically slightly elongate. Growth lines continuous, some more prominent, regular 0.05-0.1 mm spacing, growth lines more clearly defined towards the anterior margin, particularly in valve with submarginal to subcentral beak.

Remarks. Though no articulated specimen is known, the two valves can be inferred as belonging to a single species as their outlines are indistinguishable, suggesting they once did meet. By comparison with the type species, *A. schuleri* Willard (1928), the valve with the more marginal beak is assumed to be the dorsal valve, and the valve with the more central beak being the pedicle valve.

As *Dawsonia* Nicholson is an invalid taxon there is no conflict of names. However, Nicholson's collections need to be supplemented with additional material displaying conjoined valves and internal views for the generic assignment to be confirmed. Until then the taxon should remain in open nomenclature.

We have corrected Nicholson's use of the name 'rotunda' to 'rotundus' as the latter is the neuter gender singular of the nominative case.

Superfamily? *Acrotheloidea* Walcott & Schuchert *in* Walcott, 1908

Family? *Acrothelidae* Walcott & Schuchert *in* Walcott, 1908

Subfamily? *Conodiscinae* Rowell, 1965

Genus *Discotreta* Ulrich & Cooper (1936)

non 1858 *Dawsonia* Hartt *in* Dawson, p. 655.

p. 1873 *Dawsonia* Nicholson, p. 139-140 *pars*.

* 1936 *Discotreta* n. gen. Ulrich & Cooper, 1936, p. 619.

2000 ?*Discotreta* Ulrich & Cooper, 1936; Holmer & Popov, 2000, p. 94-95.

Remarks. There appears to be some doubt as to the affinity of the genus, with Rowell (1965) considering it *Incertae Familiae* and Holmer & Popov (2000) expressing a degree of uncertainty in its systematic position. These specimens do not preserve sufficient characters to contribute to the debate. There is no doubt, however, in the status of the generic name, as the invalidity of the name *Dawsonia* Nicholson avoids conflict.

Discotreta cf. *levisensis* (Walcott, 1908)

(Figure 3f)

- p. 1873 *Dawsonia tenuistriata* Nicholson; p. 141-142 *pars, non* Figs 3 c-d'.
- * 1908 *Acrothele levisensis* Walcott, 1908, p. 85, pl. 8, fig. 13.
- 1936 *Discotreta levisensis* (Walcott, 1908); Ulrich & Cooper, p. 619.
- 1938 *Discotreta levisensis* (Walcott, 1908); Ulrich & Cooper, pl. 6a.
- 1965 *Discotreta levisensis* (Walcott, 1908); Rowell, p. 281, fig 176.
- 2000 *Discotreta levisensis* (Walcott, 1908); Holmer & Popov, Fig. 47, 2a-d.

Type material. Lectotype GSC 8230, paratypes GSC 8230a, b; housed in the Geological Survey of Canada collections.

Material. NHM P1984.1-3, P1985.3-5 (9 valves).

Type locality. Lévis Shale, Ordovician (Arenig); Point Lévis, Quebec, Canada.

Diagnosis. As Ulrich & Cooper (1936).

Description. Shell unequivalved with transversely suboval outline, equally biconvex. Apex submarginal to subcentral and posteriorly positioned, seemingly more submarginal in one valve than the other. Anterior-posterior length 1.0-2.6 mm, valve breadths 1.2-3.4 mm, typical breadth around 3 mm; length-breadth ratio 0.65-0.9, typically 0.8; maximum breadth at anterior posterior midpoint. Growth lines continuous, more clearly defined away from the apex, regular 0.06-0.11 mm spacing throughout the valve.

Remarks. The quality of preservation, especially the lack of internal features, precludes precise assignment. The valve with the most submarginal apex is most likely the ventral valve by comparison with the specimens of *Di. levisensis* illustrated in Holmer & Popov (2000, Fig. 47, 2a-d).

This species was originally accommodated in Nicholson's (1873) concept of "*D.*" *tenuistriata* which allowed for considerable variation in the position of the apex by comparison with the variably positioned 'nipples' (actually diagenetic pyrite) in "*D.*" *campanulata*. However, as Nicholson's description is of an elongate oval fossil, it seems best to remove this form from "*D.*" *tenuistriata* and compare it with *Di. levisensis*. As it is unknown whether Nicholson's Point Lévis material was collected from the precise locality and horizon of Walcott (1908), this material should not be assigned topotype status.

Phylum, Class, Order & Family uncertain

Genus "*Dawsonia*" Nicholson

non 1858 *Dawsonia* Hartt in Dawson, p. 655.

- p. 1873 *Dawsonia* Nicholson, pp. 139-140, *pars*.
- p. 1889 *Dawsonia* Nicholson; Miller, p. 184.
- non* 1904 *Caryocaris* Salter; Ruedemann pp. 738-742.
- non* 1969 *Caryocaris* Salter; Rolfe *in* Moore, p. 316.
- non* 1970 *Lockeia* James; Osgood, pp. 308-312.
- p. 1981 *Dawsonia* Nicholson; Williams, p. 55.
- non* 1989 “*Dawsonia*”; Frey, Fig 7.

Type species. “*Dawsonia*” *campanulata* Nicholson; secondary diagnosis, Miller (1889).

Diagnosis. Ovato-triangular carbonaceous fossil consisting of a flat, tapering lobate body and a sharply-delineated, narrow triangular mucro. Specimens range in size from 3-12 mm length and 1-4mm width, with the mucro itself being typically less than 0.5 mm in length, and seemingly isometric growth.

“*Dawsonia*” *campanulata* Nicholson

(Figures 1a-c, 6a-c)

- non* 1837 *Prionotus pristis* Hisinger, p. 114, pl. 35, fig. 5.
- non* 1843 *Graptolithus (Prionotus) Sedgewickii [sic]* Portlock, p. 318, Pl. 19. Fig. 1.
- p. 1866 *Graptolites sedgewickii* (Portlock) Pl. 17, Fig. 3 *pars*.
- v. 1867 *Diplograpsus pristis* (Hisinger); Nicholson, pp. 111-113, Pl. 7, Figs 21-21b.
- v.* 1873 *Dawsonia campanulata* Nicholson, pp. 142-143, Fig. 3e-f.
- p. 1873 *Dawsonia acuminata* Nicholson, pp. 142-143, *pars*.

- v. 1877 *Dawsonia* sp.; Lapworth, p.7, Pl. 7, Figs 23a-d.
- . 1981 *Dawsonia campanulata* Nicholson; Williams p. 55, Pl. 6, Figs 1-15, Pl. 7, Fig. 6.
- . 1995 *Dawsonia* sp.; Williams, p. 196, Pl. 36, Fig. 16.

Type Material. Lectotype NHM 1976.

Additional Material. Topotypes in Nicholson's collection NHM P1976; material measured in Fig. 5: BGS GSM 105814-9, GSE 10800-1, 3366, PHW 501-553, 18E 73,81,90,94-5,99,102-4,112-3,117 and SM A38754; additional material: SM A20905a-c, A20906; BEL K681; NHM 55641.1-2, 55647.

Type locality. Dob's Linn, near Moffat, Scotland. Birkhill Shale Formation, Silurian: Llandovery: Rhuddanian.

Range & horizons. Rhuddanian to Aeronian (Llandovery, Silurian) of the British Isles. Birkhill Shale Formation (Moffat Shale Group) in Dob's Linn, Garpol Linn, Plewlands Burn and Duffkinnel Burn, Southern Uplands, Scotland, and in Coalpit Bay, Donaghadee, Northern Ireland; Skelgill Formation (Stockdale Group) in Spengill, nr Sedbergh, Howgill Fells, and Hol Beck, Skelgill, English Lake District.

Description. As genus.

Remarks. "*D.*" *campanulata* cannot be easily accommodated in any higher taxonomic group. It is clearly unrelated to graptolite scopulae, and bears little similarity to either phyllocarids or algae. Although recent works have tried to accommodate it in these groups (cf. Rolfe 1969 and Williams 1981, respectively), neither assignment is entirely convincing. Meanwhile, Underwood (1993, Fig 4e) illustrated a carbonaceous fossil that looks conspicuously "*Dawsonia*"-like as a faecal pellet. In the most detailed recent study of "*Dawsonia*", Williams (1981) argued that

it represented a spore-carrying alga. He stated that “*D.*” *campanulata* had an open, flared “posterior margin, giving the [hollow] body a ‘crocus flower’ type of appearance”. However, it is an order of magnitude larger than such spore-carrying alga in the modern oceans (Tappan 1980) and there is no evidence to suggest it had significant three-dimensionality in life. The ‘nipples’ seen associated with “*D.*” *campanulata* superficially suggest a three dimensionality, but, as noted in section 3, they actually represent compression of the fossil on to diagenetic pyrite in the sediment. This pyrite notably differs in fabric from the pyrite infill of hollow cavities (cf. Allison 1988; Underwood & Bottrell 1994). Moreover, “*D.*” *campanulata* lacks the morphological variation seen when unambiguously hollow tissues such as the *Climacograptus wilsoni* vesicle are found flattened in these shales (cf. Williams 1994). And, although the distal margin of “*D.*” *campanulata*’s lobate body may be fragmented (e.g. Fig. 1b), and, at times, less-well delineated than the proximal end and mucro (e.g. Fig. 6b), there are many examples showing a well-defined, rounded distal margin (e.g. Figs 1a,c & 6a,c), suggesting that this represents variability in preservation rather than a crocus-flower-like morphology. With this in mind, “*Dawsonia*” is best considered to be a flat problematicum rather than a hollow alga.

7. Acknowledgments

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Figure captions

Fig. 1. (a-c) “*D.*” *campanulata* Nicholson 1873 and (d-f) graptolites with appendages, Dobb’s Linn, Birkhill Shale Formation. (a) lectotype, NHM P1976; (b) syntype, NHM P1976; (c) exhibiting a prominent “nipple” and juxtaposed to an indet. monograptid, though preserved on a different sedimentary lamina, NHM P1976; (d) *Dittograptus?* sp. (Elles 1940) with well-preserved scopulae (‘graptogonophores’), SM A13731; (e) nemal vane of ?*Pribylograptus incommodus* (Törnquist 1899) cf. Crowther (1978) overlain by indet. graptolite, see also Fig. 2h, NHM P1981; (f) detail of poorly-preserved scopula (‘graptogonophore’) of *Hallograptus bimucronatus* (Nicholson 1869), the scopula originates from the graptolite to the right, the concentric lines represent progressive growth increments constrained by the scopula’s better margin, BU 1420. All specimens photographed under reflected light. Nicholson’s specimens (a-c, e). Scale bar = 1 mm (a-c, e-f), 500 µm (d).

Fig. 2. Nicholson’s illustrations of supposed graptolite ovarian capsules, including (a-f) his “*Dawsonia*” type specimens (Nicholson 1873). (a) “*D.*” *acuminata*, (b) “*D.*” *rotunda*, (c-d) “*D.*” *tenuistriata*, (e-f) “*D.*” *campanulata*; (g-m) in Nicholson (1872); (n-o) Nicholson’s handwritten specimen labels, NHM.

Fig. 3. (a-f) Linguliform brachiopods with camera lucida interpretations and (g) graptolite, from the Lévis Shale, Point Lévis, Quebec, Canada. (a-b, d) *Acrosaccus?* *rotunda* (Nicholson 1873): (a) pedicle valve, NHM P1985.3, (b) dorsal valve, NHM P1985.3, (d) lectotype, NHM P1982.1; (c, e) *Paterula?* *tenuistriata* (Nicholson 1873): (c) lectotype, NHM P1984.3, (e) counterpart NHM P1984.2; (f) *Discotreta* cf.

levisensis (Walcott 1908), lectotype NHM P1984.2; (g) *Clonograptus* sp., NHM P1982. All specimens photographed under reflected light and are from Nicholson's material. All scale bars = 500 μ m.

Fig 4. *Caryocaris acuminata* (Nicholson 1873), from the Lévis Shale, Point Lévis, Quebec, Canada. (a) Furcal ramus, lectotype NHM P1985.3; (b) furcal ramus, NHM 1977, (c) telson, NHM P1984.3; (d) a reconstruction of the tail-piece and the whole animal (note schematic carapace). All specimens photographed under reflected light. Scale bars = 500 μ m.

Fig. 5. Morphometric analyses. “*D.*” *campanulata* (open circles) compared with (a) graptolite scopulae (closed squares), and (b) the furcal rami and telsons of *Caryocaris acuminata* (combined, filled diamonds). (c) Morphometrics of brachiopod outlines and growth lines (combined) showing distinct populations corresponding to *Di.* cf. *levisensis* (open triangles), *A?* *rotunda* (filled circles) and *P?* *tenuistriata* (open squares). (d) Absolute sizes (mm) of all of the above specimens (brachiopod outlines shown as closed circles, others as before). Morphometric criteria as illustrated: L = anterior-posterior length; W = lateral width; D = distance from blunt margin to centroid; C = distance from anterior margin to growth centre of brachiopod. “*D.*” *campanulata* specimens are those listed under additional material in Section 6; brachiopod and *Caryocaris* specimens are Nicholson's specimens from the Lévis Shale Fm., Point Lévis, Quebec, with specimen numbers listed in Section 6. Graptolite scopulae measurements based on those specimens illustrated in Ruedemann (1908), Elles (1940), Bates & Kirk (1991), Štorch (1994) and Koren' & Rickards (1997).

Fig 6. High contrast BS SEM images illustrating the preservation and morphology of (a-d) “*D.*” *campanulata*, (e) graptolite scopula, (f) brachiopod, and (g) *Caryocaris*.

The low brightness of “*Dawsonia*” and the scopula indicate preservation as organic compressions; the high brightness of the brachiopod and *Caryocaris* reflect their primary phosphatic compositions; white areas are accessory diagenetic minerals and weathering products. (a) petal-shaped morph, BGS GSM 105817; (b) bell-shaped morph, BGS GSM 105816; (c) partially overlying and imprinting a diagenetic pyrite to produce a well-developed "nipple" (outlined), SM A20905a; d) close-up of a "nipple" showing its concentric structure defined by diagenetic pyrites, SM A 20905a; (e) holdfast and proximal body of scopula (outlined) attached to graptolite illustrated in Fig. 1d, SM A13731; (f) *A. rotunda*, syntype NHM P1985.2; g) furcal ramus of *C. acuminata*, NHM P1985. Specimens from the Birkhill Shale Fm. of Duffkinnel Burn (a-b), Coalpit Bay (c, d) and Dobb's Linn (e); and, Lévis Shale Fm., Point Lévis (f-g). Scale bar = 1mm (a-c, f-g), 500 μ m (d-e).

ALEX PAGE*

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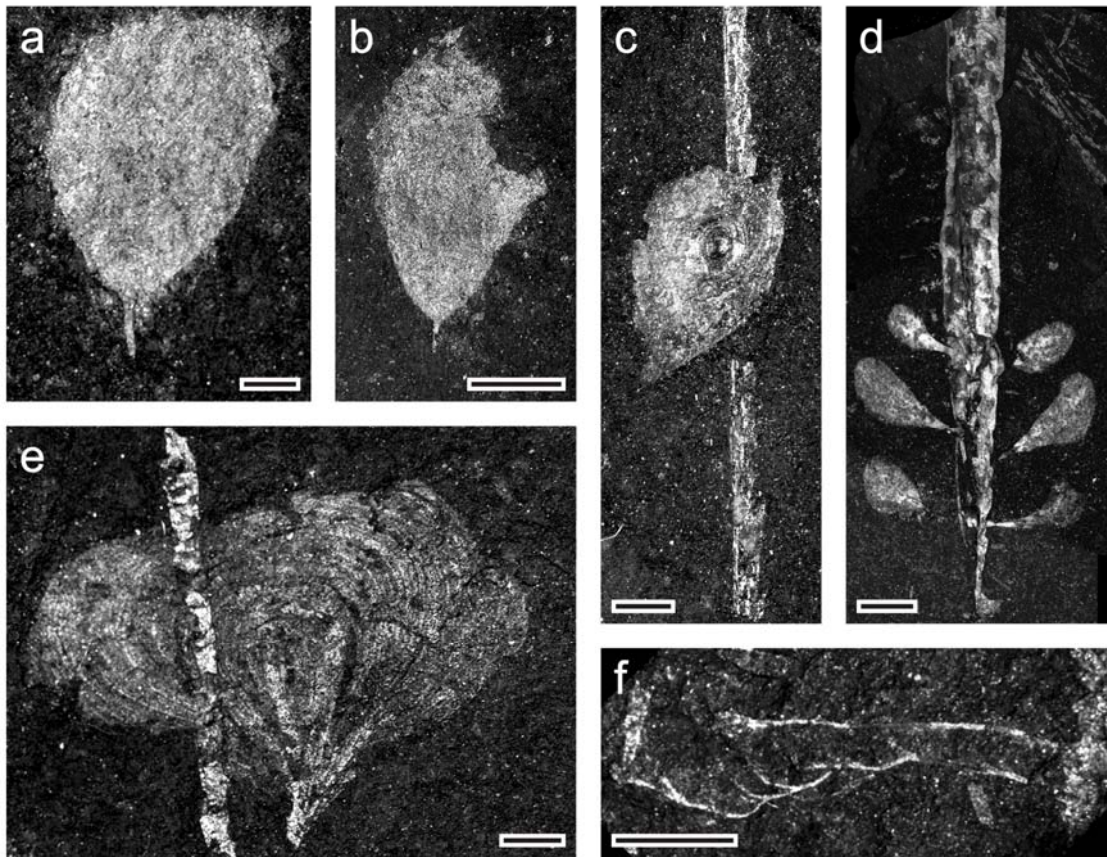


Figure 1.

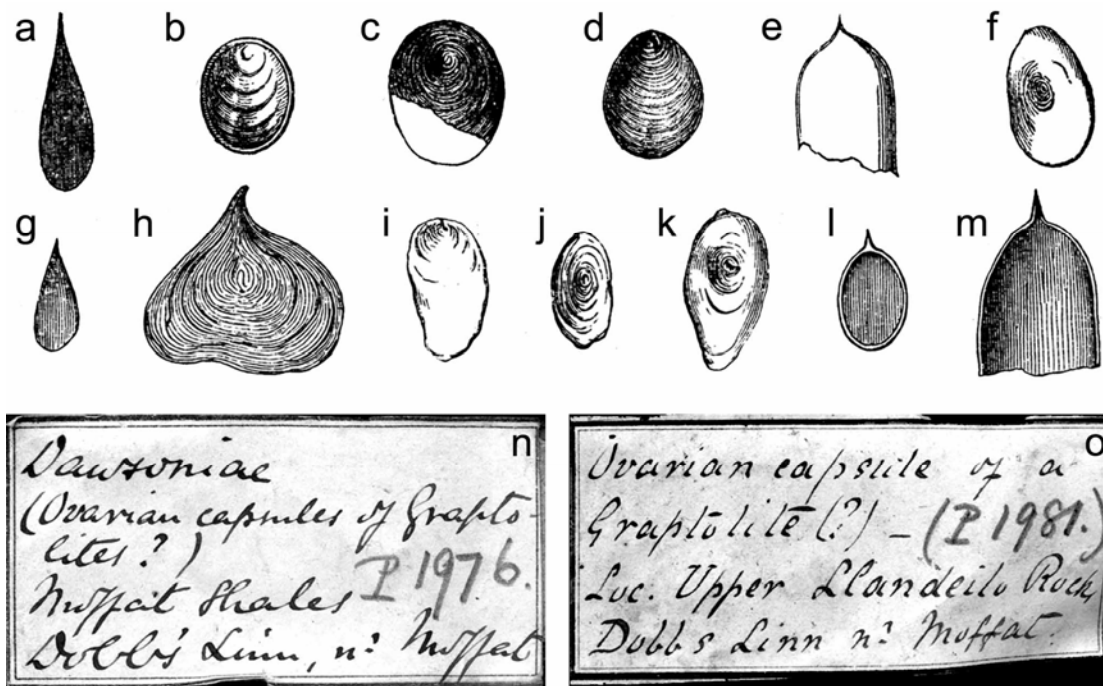


Figure 2.

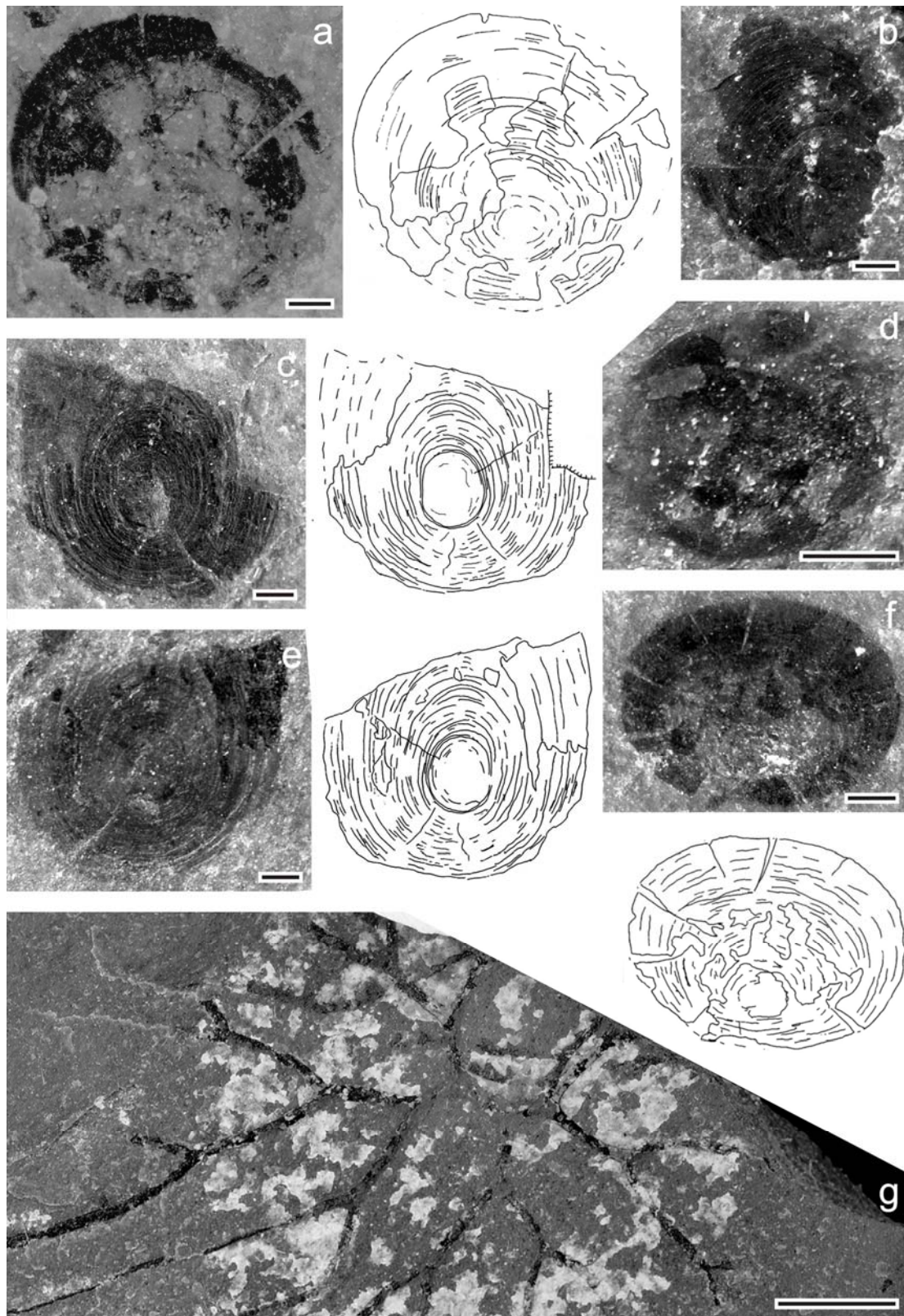


Figure 3.

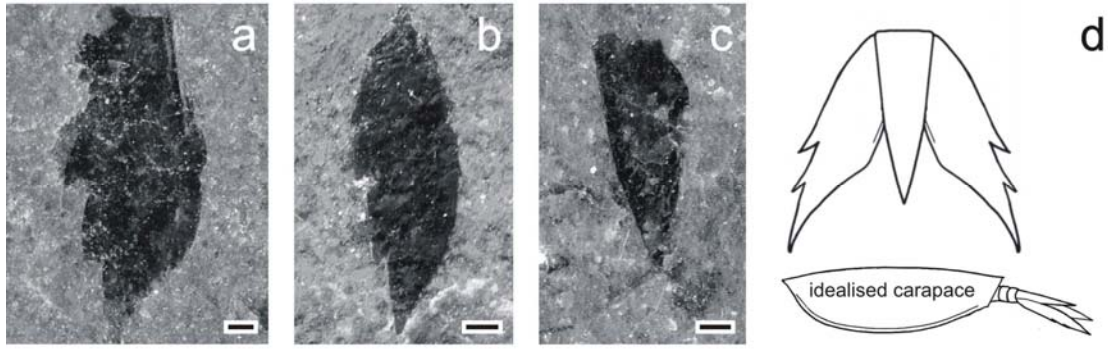


Figure 4.

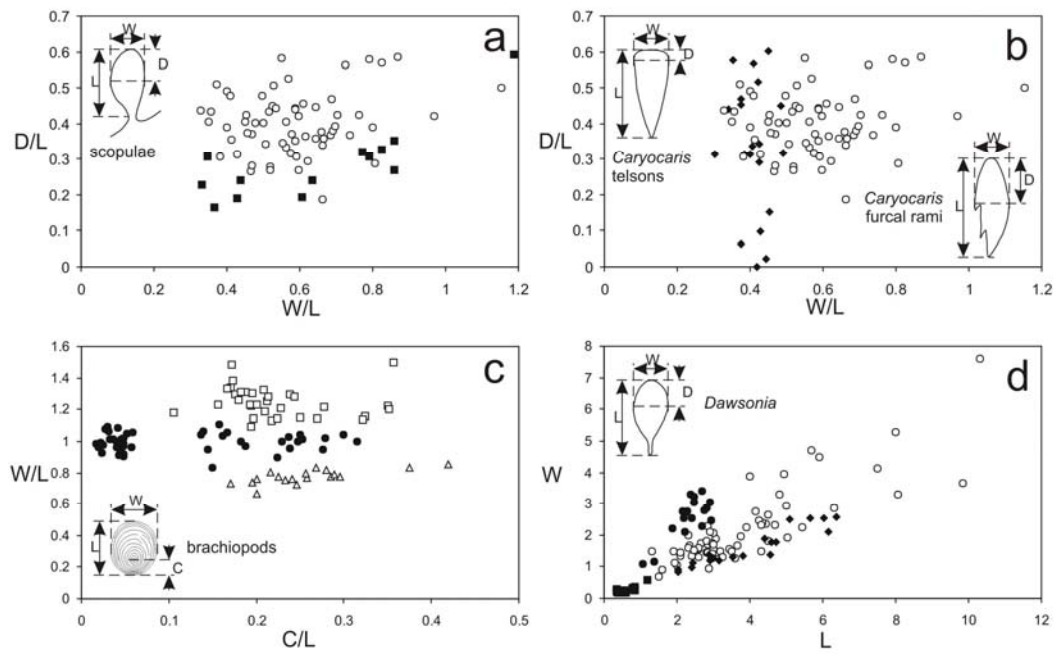


Figure 5.

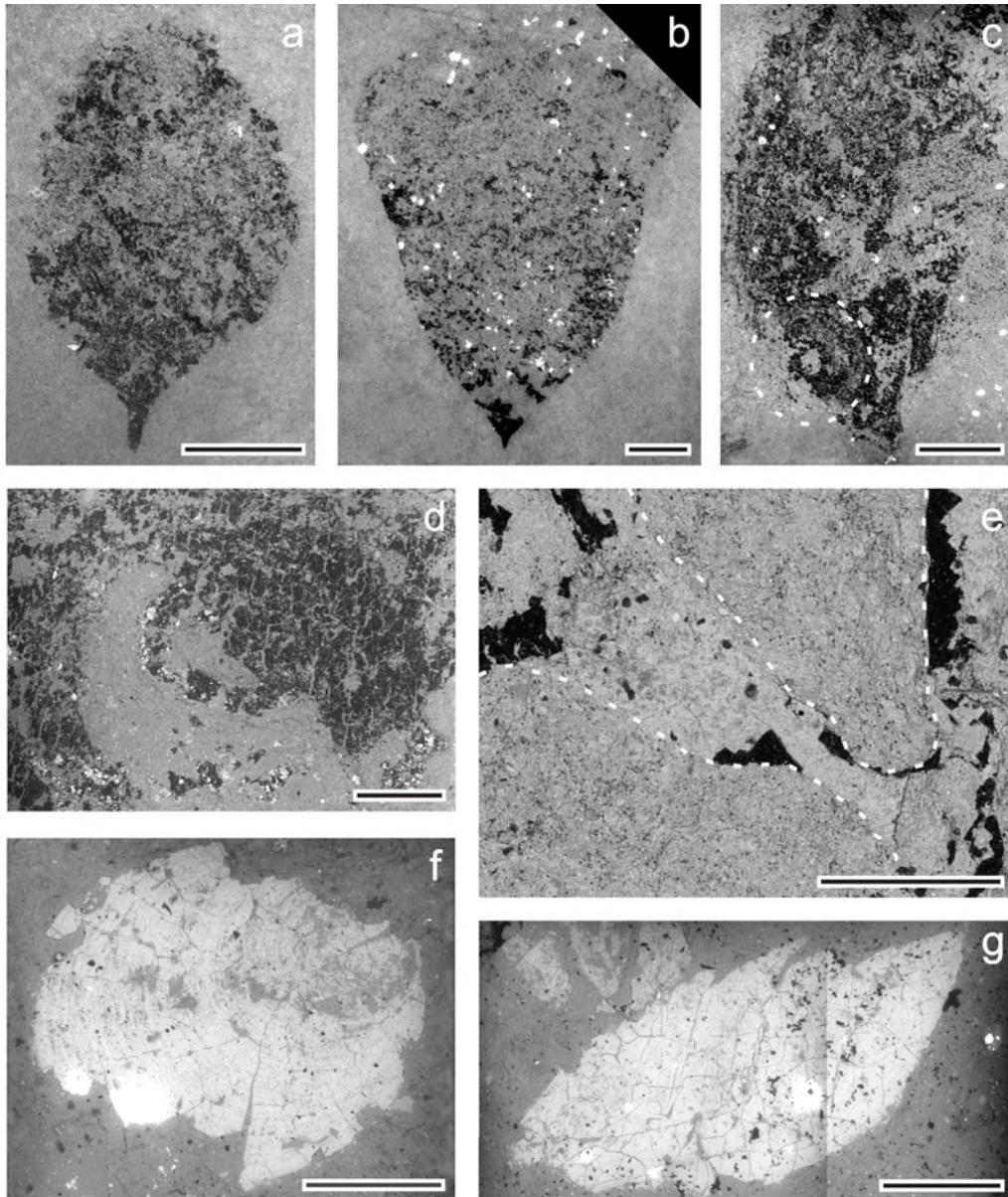


Figure 6.