# PALMS

Journal of the International Palm Society

Vol. 66(4) Dec. 2022



#### THE INTERNATIONAL PALM SOCIETY, INC.

#### **The International Palm Society**

#### Founder: Dent Smith

The International Palm Society is a nonprofit corporation engaged in the conservation, preservation, and study of palms. The society is international in scope with worldwide membership, and the formation of regional or local chapters affiliated with the IPS is encouraged. Please address all inquiries regarding membership or information about the society to The International Palm Society Inc., 1401 Lavaca St. #751, Austin, Texas 78701 USA, or to info@palms.org.

#### **OFFICERS**:

President: Robert Blenker, Robert.Blenker@gmail.com.

Vice-Presidents: Jeff Brusseau, Jbrusseau@newportnational.biz, 1-760-271-8003. Andy Hurwitz, ashcvs@yahoo.com, 1-310-702-0900.

Corresponding Secretary: Ray Hernandez, rayhernandez77@gmail.com, 1-813-832-3561.

Administrative Secretary: Larry Noblick, Inob@montgomerybotanical.org, 1-305-667-3800 ex 104.

Treasurer: Tom Jackson, jacksongrantgallery@outlook.com, 1-415-923-3299.

#### **Directors:**

#### 2022-2026:

2020-2024: Robert Blenker, FL, USA. Jeff Brusseau\*, CA, USA. Anthony Glomski, CA, USA. Gregg Hamann\*, CA, USA. Haresh\*, India. Susan Hubbell, FL, USA. Tom Jackson\*, CA, USA. Leland Lai\*, CA, USA. Michael Merritt, HI, USA. Romeo Montalvo, TX, USA. Larry Noblick, FL, USA. Lazaro Priegues, FL, USA. Grant Stephenson, TX, USA. Michael Valentine, CO, USA. Jill Menzel\*, Brazil. Colin Wilson, Australia. Scott Zona, NC, USA.

Cindy Adair, Puerto Rico. William J. Baker, UK. Norm Bezona, HI, USA. Julie Cleaver Malzoni, Brazil. John Dransfield, UK. Glenn Franklin, NC, USA. Len Geiger, CA, USA. Ray Hernandez, FL, USA. Horace Hobbs, FL, USA. Andy Hurwitz\*, CA, USA. Chad Husby, FL, USA. Mary Lock, HI, USA. Paco Martí Navarro, Spain. Fernando Roca, Peru. Tobias Spanner, Germany. Andrew Street, FL, USA. David Tanswell, Australia.

\* IPS Benefactors

Website: www.palms.org. The IPS website includes information on the history and organization of the society, past and future travel, publications and the IPS Bookstore, affiliate chapters, and the link to our popular on-line forum, PalmTalk.

@InternationalPalmSociety

@thepalmsociety

@IPS\_PalmSociety

#### **Palms** (formerly PRINCIPES)

Journal of The International Palm Society

Publication of PALMS is made possible by a generous gift from Drs. Kathleen Grant and Tom Jackson.

An illustrated, peer-reviewed quarterly devoted to information about palms and published in March, June, September and December by The International Palm Society Inc., 1401 Lavaca St. #751, Austin, Texas 78701 USA.

Editors: John Dransfield, j.dransfield@kew.org. Scott Zona, scott.zona@yahoo.com.

Associate Editor: William J. Baker, w.baker@kew.org.

Guidelines for authors are available on request from the Editors, or on-line at: http://palms.org/resources/palms-journal/

#### **Membership Levels:**

Free membership includes the monthly Newsletter delivered via email.

Electronic membership (US\$40 per year or \$100 for three years) includes the Newsletter and our journal, PALMS, in digital format.

Full membership (US\$60 per year or US\$150 for three years) includes the above plus a printed copy of PALMS. Dues include mailing of the journal by airlift service to addresses outside the USA

Benefactor and Lifetime memberships are also available. Dues may be paid on-line at www.palms.org. Subscription price is US\$55.00 per year for libraries and institutions.

Change of Address: Send change of address, phone number or e-mail to The International Palm Society Inc., 1401 Lavaca St. #751, Austin, Texas 78701 USA, or to info@palms.org.

Claims for Missing Issues: Claims for issues not received in the USA should be made within three months of the mailing date; claims for issues outside the USA should be made within four months of the mailing date.

Periodical postage paid at Austin, TX, USA and additional mailing offices. Postmaster: Send address changes to The International Palm Society Inc., The International Palm Society Inc., 56 Autumn Oaks Drive, The Hills, Texas 78738 USA.

PALMS (ISSN 1523-4495)

Mailed at Lawrence, Kansas 15 Dec. 2022 ©2022 The International Palm Society

The full text of PALMS is available on EBSCO Publishing's database.

This publication is printed on acid-free paper.

6

177

195

#### Kapawi: A Mega-Diverse Palm Community in the Eastern Amazon of Ecuador

T.L.P. COUVREUR, R. CARRILLO-FLORES, R. CÁMARA-LERET, J.C. COPETE & H. BALSLEV

A Review of Naturally Occurring Hybrids in Palms (Arecaceae) A. HENDERSON

#### Exploring for new Localities of *Tahina spectabilis* in Northeastern Madagascar

D. Rabehevitra



#### FRONT COVER

A hybrid between *Howea belmoreana* and *H. forsteriana* occurring naturally on Lord Howe Island. See article by A. Henderson, p. 177. Photo by W. Baker.

#### BACK COVER

A stand of *Syagrus sancona* in the Kapawi region of Ecuador. See article by Couvreur et al., p. 161. Photo by Thomas L.P. Couvreur.

#### Features

Palm News	160
Index to vol. 66	202
Patrons of the IPS	207



A garnet-red new leaf emerging on *Geonoma poeppigiana*. See article by Couvreur et al., p. 161. Photo by Thomas L.P. Courvreur.

## **PALM NEWS**

New research from Sidonie Bellot and colleagues finds that **more than half of the world's palm species are threatened**. The team used machine learning to assess extinction risk predictions for 1,381 palm species. Those predictions were integrated with published conservation assessments for 508 species to point to areas with the greatest need for conservation. Madagascar, New Guinea, the Philippines, Hawaii, Borneo, Jamaica, Vietnam, Vanuatu, New Caledonia and Sulawesi were identified as priority regions for palm conservation. The publication is at *Nature Ecology & Evolution* 6: 1710–1722 (2022).





Molecular tools are continuing to allow botanists to reveal the chemical pathways that control the growth and development of plants. One such study examined the **development in male and female flowers in the inflorescence of the betelnut palm**, *Areca catechu*. The work, published by G. Zhou et al. in *New Phytologist* (https://doi.org/10.1111/nph.18471), found that jasmonic acid, a common signaling chemical in plants, was expressed in female flowers at about ten times the concentration found in male flowers. They also found that in inflorescences bearing abnormal bisexual flowers, the

concentration of jasmonic acid was about twice that of normal male flowers. The author proposed a mechanism by which jasmonic acid promotes the development of female flower organs.



John Dowe and coauthors recently published "The botanical expedition of Hermann Wendland in Central America: a nomenclatural study and travel report" in *Boissiera* 73, a publication of the Conservatoire et Jardin Botaniques, Geneva, Switzerland. The work details Wendland's travels and botanizing in Central America, an expedition that resulted in 185 named taxa, many of which were palms. This publication examines the nomenclatural history and typification of those names

## Kapawi: A Mega-Diverse Palm Community in the Eastern Amazon of Ecuador

THOMAS L.P. COUVREUR<sup>1,2\*</sup>, ROBERTO CARRILLO-FLORES<sup>3</sup>, RODRIGO CÁMARA-LERET<sup>4</sup>, JUAN CARLOS COPETE<sup>5</sup> AND HENRIK BALSLEV<sup>6</sup>

The Achuar Kapawi community is located in the eastern Amazonian region of Ecuador. It is adjacent to the Kapawi river, a tributary of the large Pastaza river and is just 20 km from the Peruvian border. Because of its proximity to Peru, this region might harbor several species of palms not yet recorded in Ecuador. The present article is a personal account from a 5-day expedition to the region by the first author, and a preliminary synthesis of results from an expedition led by Henrik Balslev (ecology) and Rodrigo Cámara-Leret (ethnobotany) to investigate the diversity, abundance, and uses of palms in the region.

Ecuador is one of the richest countries in palm diversity of South America when related to its size. To date, around 140 species in 32 genera are recorded (Valencia et al. 2013). Yet, some

areas of the Amazon remain relatively little explored for palms (Borchsenius et al. 1998, Couvreur et al. 2008, 2021), especially those close to the Peruvian border. This is the case

<sup>1</sup> Institut de Recherche pour le Dévelopemment IRD, DIADE, CIRAD, Université de Montpellier, Montpellier, France. thomas.couvreur@ird.fr	<sup>4</sup> Department of Evolutionary Biology and Environmental Studies, University of Zurich, Zurich, Switzerland. rodrigo.camaraleret@ieu.uzh.ch
<ul> <li><sup>2</sup>Pontificia Universidad Católica del Ecuador, Facultad de Ciencias Exactas y Naturales, Av.</li> <li>12 de Octubre 1076 y Roca, Quito, Ecuador.</li> <li><sup>3</sup>Director de la Organización Turismo &amp; Conservación, Av. Machala N-54 225 y Jorge</li> </ul>	<sup>5</sup> Corporación Autónoma Regional del Centro de Antioquia, Cra. 65 #44A-32, Medellín, Antioquia, Colombia. juan_copete@corantioqia.gov.co <sup>6</sup> Department of Bioscience, Ecoinformatics and
Piedra, Quito, Ecuador. rcarrilloflores@turismoyconservacion.org	Biodiversity Group, Aarhus University, Aarhus, Denmark. henrik.balslev@bios.au.dk



1. A view of Kapawi Eco Lodge, deep in the Ecuadorian amazon, in the region of Pastaza, close to the Peru border. Notice *Astrocaryum chambira* (tall palm), and just under a juvenile *Iriartea deltoidea*. Photo by Thomas L.P. Couvreur.

of the Kapawi community of the Achuar, one of the most recent Indigenous ethnic groups to have been contacted.

The Achuar are one of the 13 Indigenous ethnic groups living in Ecuador. The word "Achuar" comes from the union of the words "shuar" or person and "achu" or *morete* palm (*Mauritia flexuosa*). The 'Achuar' are thus the people of the *morete* palm. The lifestyle and traditions of Achuar were famously described by the French anthropologist Phillipe Descola who visited Kapawi in the 1970s (Descola 1993). And yet, far too little is known about the Achuar people's knowledge of Amazonian biodiversity or about palms and their uses.

Kapawi, which refers to the name of a sort of flat fish in the Achuar-Shiwiar language (Descola 1993), is located deep in the Ecuadorian Amazon in the region of Pastaza, some 175 km southeast from the Amazonian town of Puyo and 20 km from the Peruvian border. This region is covered by a dense lowland rainforest drained by the large Pastaza river, which starts high in the Andes near Baños, and ends in the Mariño river in Peru (a tributary of the Amazon). Besides the relatively flat region close to Pastaza, there is a hilly area to the north, with peaks of about 350–400 m, providing some geological diversity to the region.

Kapawi was the focus of two palm expeditions that took place ten years apart, one in October 2011 led by Henrik Balslev (ecology team) and Rodrigo Cámara-Leret (ethnobotany team) and one in February 2021 led by Thomas Couvreur. The base for both expeditions was the Kapawi Eco Lodge (www.kapawi.com) (Figs. 1 & 2), a community-based project which aims to use ecotourism for the economic, social, and cultural development of local Achuar communities. The project, which is managed by the indigenous residents themselves, serves as an added source of income for the Kapawi, Wachirpas, Ishpingo, Kusutkau, Wayusentsa, Sharamensa and Suwa communities. Ecotourism revenues have served to increase local incomes. Importantly, ecotourism provides an added incentive for local communities to conserve the natural resources in the area.

To reach Kapawi, one needs to take a plane from the aerodrome located in the village



2. Example of the understory rain forest near Kapawi the common *Hyospathe elegans* subsp. *elegans* in the foreground. Photo by Thomas L.P. Couvreur.

called Shell, just north of Puyo. Once in the air, the small 3-passenger plane takes an eastern bound direction, leaving behind the mighty Andes. On a good day, one can see to the right the Sangay, one of the numerous active volcanoes in Ecuador spitting its fumaroles. As we progress into the Amazon, the impact of humans on the rainforest becomes less and less apparent. After a short 45 min flight, the plane makes an expert



3. Our Achuar guide, Jhonny Saant in the Kapawi rainforest. Photo by Thomas L.P. Couvreur.

landing on the short dirt runway of the Kapawi community. Several people come out of nowhere and check with the pilot if some relatives in Puyo sent them some food or a letter. After a short stop, the plane turns and takes off again.

One of our guides was Jhonny Saant, a 30year-old Achuar student in pedagogy and father of three (Fig. 3). After being a guide for the lodge for several years, he decided to resume higher education thanks to a government scholarship. Thomas was also accompanied by two other young Achuar students at the Kapawi community school.

The plan for Thomas's teamwas to walk as much as we could and collect as many palms as we saw. This general collecting approach allows one to cover more ground but with less detail. In contrast, the ecology team set up a much more thorough protocol to document palm diversity, results of which are partially presented here. A total of 11 transects measuring  $5 \times 500$  m each were installed, covering different lowland evergreen forest types: terra firme (4 transects), intermediate (1 transects, all palm individuals including seedlings, juveniles, sub-adults and adults are

meticulously counted, measured, identified and photographed. Finally, the ethnobotany team undertook several interviews to document to the importance of palms and their uses across the Achuar communities.

#### A mega-diverse palm community

After one week of painstaking work in October 2011, the ecology team (led by Henrik Balslev) documented 7505 palm individuals representing a total of 40 palm species in 21 genera across the 11 transects covering 2.75 ha (Table 1). That means there are 2729 palms per hectare and between 9 to 29 species per transect! In addition, four species were documented outside of the transects (Bactris gasipaes var. gasipaes (Fig. 4), Cocos nucifera, Syagrus sancona and S. smithii) leading to 44 palms known from Kapawi. Oenocarpus bataua was by far the most abundant palm in the region with 1529 individuals registered over the 11 transects. We can refer to such species as being super-abundant. The two other most abundant palms were the understory Geonoma macrostachys (953 individuals) and Geonoma stricta subsp. arundinacea (857 individuals reported). Both species are morphologically very variable and taxonomically hard to classify. Geonoma macrostachys is, in fact, so



4 (left). *Bactris gasipaes* var. *gasipaes*, or *uvi* in Achuar. Here one morphotype with red fruits (left) and one with white fruits (right) growing in Jhonny's *chacra* (community garden). Photo by Thomas L.P. Couvreur. 5 (right). *Wendlandiella gracilis* var. *simplicifrons* in flower (a new genus and species record for Ecuador). Photo by Henrik Balslev.

variable that it was impossible to define into different subspecies (Henderson 2011). Nevertheless, it is a spectacular understory palm with large (to 2 m tall) generally undivided leaves and a long erect generally unbranched inflorescence. It is common across the Ecuadorian Amazon growing sympatrically with many different morphotypes. Even though this morpho-diversity is a headache for taxonomists, it is a blessing for evolutionary biologists who are using the Geonoma macrostachys species complex model to unravel the impact of Amazonian environmental heterogeneity on understory plant speciation (Roncal 2006, Bacon et al. 2021).

In terms of species richness, the 2011 fieldwork showed that Kapawi is one of the most species rich palm communities across the whole of tropical America (Balslev et al. 2011). The terra firme evergreen forest was the most diverse with 29 species, while the intermediate lowland was the poorest, with 18 species. Even when we compare with similar habitats, Kapawi stands out as very species rich. For example, Kapawi is comparable to Yasuni, in lowland Ecuador Amazon and one of the most biodiverse places on earth. A recent survey documented between 30 and 33 palm species on terra firme over a surface of 2.5 ha (Balslev et al. 2011, although there might be different species concepts between studies). The region around Kapawi is thus a truly mega-diverse palm community in western Amazon.

Most of these species were already documented from Ecuador, although generally not reported for this region. Interestingly, and as we hypothesized before our expeditions, we documented the presence of two genera, one species and one subspecies as new records for Ecuador.

The two genera new to Ecuador are the monotypic genus *Wendlandiella* and *Iriartella* (two species). *Wendlandiella* represents a single polymorphic species, *W. gracilis*, distributed mainly in Peru and the State of Acre in Brazil (Eychenne et al. 2018). Kapawi harbors the most northern population of *W. gracilis* representing the variety var. *simplicifrons* (Fig. 5). It is a small understory palm, with entirebifid leaves and an erect once branched inflorescence with several rachillae. It can easily be confused with other understory bifidleaved palms such as *Chamaedorea pauciflora* 



6. *Iriartella stenocarpa* in fruit (another new genus and species record for Ecuador). Photo byThomas L.P. Couvreur.

(*yaun*) also present in the Kapawi region. This species is locally abundant, with 157 individuals all being recorded in a single transect (HB032) in a floodplain habitat.

*Iriartella stenocarpa* (Fig. 6) is a small understory palm up to 5 m tall with fishtail leaves and small stilt-roots, typical of the Iriarteinae subtribe (which also contains the genera *Dictyocaryum, Iriartea, Socratea, Wettinia*). One needs to be very careful with species of *Iriartella*, because, unlike the other genera, the leaf sheath is covered in small spines. If you inadvertently grab the leaf sheath, you will spend the rest of the day and night with tweezers plucking away at the spines like the first author experienced once in Brazil. Despite



7 (left). Wettinia drudei, a new species record for Ecuador. 8 (right). Geonoma maxima subsp. camptoneura, a new subspecies record for Ecuador. Photos by Thomas L.P. Couvreur.

being the first record of the genus for Ecuador, the species is quite common and can be present in dense populations.

The genus *Wettinia* is a mainly Andean centered genus (Pintaud et al. 2008), but one species occurs in the lowland rainforests of the Amazon, *Wettinia drudei* (Fig. 7). It is the smallest species of the genus, reaching to about 5 m, and is similar to *Iriartella stenocarpa* but without the nasty spines of the leaf sheath. In Kapawi, it is locally common, being quite abundant when present.

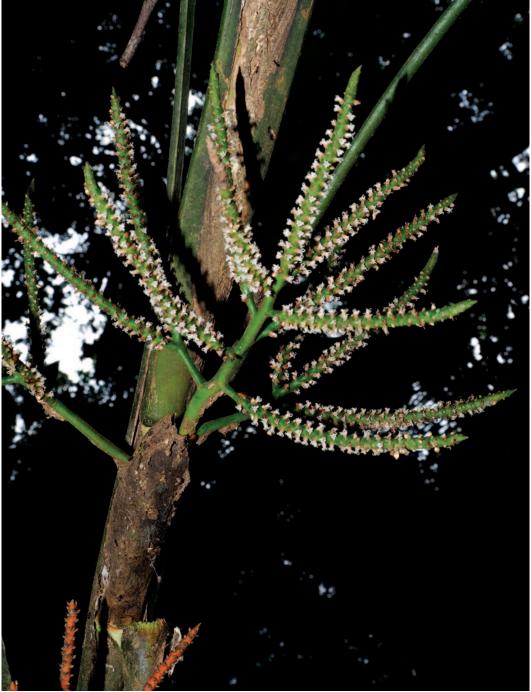
Finally, we also documented a new subspecies for Ecuador: Geonoma maxima subsp. camptoneura (Figs. 8 & 9). Geonoma maxima is a beautiful widespread Amazonian palm, reaching to about 5 meters. It is morphologically very variable in the number and arrangement of its pinnae. In the latest revision of the genus, Henderson described 10 subspecies. To date, only G. maxima subsp. multiramosa was documented from Ecuador, occurring in the northern part of the Amazon (Yasuni and Cuvabeno). The *camptoneura* subspecies has few (3 or 4) irregularly inserted and sized pinnae forming a sharp angle with the midrib. It differs from the numerous regularly inserted pinnae found in subsp. multiramosa (Henderson 2011). We were lucky

enough to have collected it in flower, during the female phase, revealing its bright white petals and stigmas.

Besides these new records for Ecuador, we also documented the presence of other interesting palms. One of them is the impressive *Elaeis* oleifera, the South American relative of the infamous African oil palm (*Elaeis guineensis*). Its documentation in Ecuador is fairly recent, dating to 1986 (Balslev & Henderson 1986, Montufar et al. 2018). This species grows in swampy areas and has a prostrate stem with only the terminal part erect and topped by several 6 m long leaves (Fig. 10). Its presence in the swamps brings a special feel to the forest. Another interesting species recorded for Kapawi is Bactris simplicifrons. Even though it is widely distributed and abundant (Henderson 2000), it is one of the few species of Bactris (together with B. schultesii and B. killipii) that have almost no noticeable spines, which is very unusual for this genus. One has to pass one's fingers along the margin of the leaves to feel them.

#### Palm uses

Besides species richness and abundance, diversity of uses is an important way to evaluate the dependence between a community and the environment. Palm uses



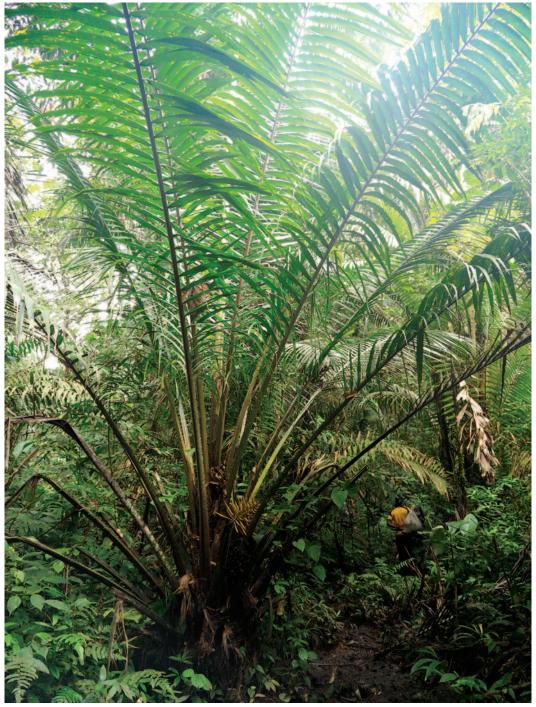
9. Flower (female phase) of Geonoma maxima subsp. camptoneura. Photo by Thomas L.P. Couvreur.

were documented in 2011 by the ethnobotany team (led by Rodrigo Cámara-Leret) with the assistance of Justo Saant by interviewing 65 informants of the Kapawi, Kusutkau and Wayusentsa communities, noting the common name of the species mentioned and the different associated uses. Uses were categorized into seven major groups: Animal food (a), Construction (b), Cultural uses (c), Human food (h), Medicinal and veterinary (m), Utensils and tools (u), and Other uses (o). In addition, in 2021 the first author had open and informal discussions with the guide Jhonny Saant. The 2011 survey noted a total of 36 species used by Achuar in Kapawi, with 1–24 uses reported for each species. Below and in Table 2, we present a summary of important palm species for the Achuar, based on the book co-authored with the Achuar (Cámara-Leret et al. 2018):

Iriartea deltoidea, tuntuam, was the most used species with 24 uses documented in five

categories. Out of the 65 informants, it was cited 542 times! This means that on average, tuntuam was mentioned eight times per informant. This is one of the most important Construction palms: the whole stems serve as house posts, the split stems are used as house

10. *Elaeis oleifera* in a swampy forest near the Kapawi community. Notice the young inflorescence at the base of the leaves. Photo by Thomas L.P. Couvreur.



floors and walls, the strips from the stem serve for the lower tapes of the roof and as support on which to weave leaves for thatch, while the rods from the stem are used to make chicken pens. Also, the split stem is used to make spears and blowguns and the strips from the stem are used to build fences on the river banks where plant poisons are used to fish. Beds are also made from boards obtained from the stem. The Achuar built the houses of the

11. *Muntish* (Achuar), grubs extracted from the rotting trunk of *Mauritia flexuosa* and ready to be eaten. Photo by Thomas L.P. Couvreur.





12. Uví (Bactris gasipaes var. gasipaes) fruits. The white ones are called Kuyu uví, and the red ones Kapuku uví. Photo by Thomas L.P. Couvreur.

Kapawi Ecolodge entirely with *tuntuam* and not a single iron nail was used in the building process!

In addition to being the most abundant palm species, *Oenocarpus bataua* or *kunkuk* is also one of the most used for the Achuar community, used in 16 different ways across five of the seven uses categorized. For instance, the leaves are used for thatching forest huts, the spear leaves for making brooms, the fibers of the leaf sheath are extracted to weave the Achuar headdress or *tewasan*, the raw palm heart is edible, the fruit is matured in water and eaten or drunk in juice, and its oil is applied directly on the hair as a pomade.

*Mauritia flexuosa*, or *achu*, was another well cited species in the surveys, and the second most used palm in terms of different categories. The fallen and rotting stems of achu are a well-known breeding ground for palm grubs called *mundish* (Fig. 11). After a full day walking in the forest with Jhonny one day, Thomas stopped near a fallen *Mauritia flexuosa*. Jhonny then pulled out his machete and began chopping away at the trunk. After a few minutes, he found several large white larvae,

with a black head. He swallowed them raw one after the other. Thomas tasted a raw one handed by Jhonny. As you chew, the head cracks under your teeth like a chip, before giving way to a warmish liquid. The taste is quite neutral, but on the greasy side, and is not unpleasant. Palm larvae are delicacies across the tropics. Jhonny explained the culinary variety of palm larvae depending on the palm species. "Each palm species has its own larva, with its own distinct taste," he said. That of the *puntish* Achu is described as being greasy with a sweet flavor, while the one of *tuntuam* (*Iriartea deltoidea*) is said to be less greasy with a more watery taste.

One important spot when visiting Amazonian communities is at the *chacra*, or the community garden. The most important palm is *uví* or *Bactris gasipaes* var. *gasipaes* (Fig. 4), the only fully domesticated palm of the Amazon. *Uví* was the only species in the survey to have been cited in all seven categories, underlining its central importance for Amazonian communities. The main product is the fruit which is eaten boiled or used to make the traditional *chicha* across the Amazon.



13. Harvesting of *chapi* (*Phytelephas tenuicaulis* leaves) for thatching of the Achuar houses. The living plant is in the background. Photo by Thomas L.P. Couvreur.

Jhonny's *chacra* contained three different morphotypes, mainly distinguished by the color and shape of the fruits: yellow, red and, the most weird one of all, white (Fig. 12). Every morphotype can be characterized by a different taste, from oily to floury, depending on what they use it for. After visiting the *chacra* we made a brief stopover in Jhonny's house. All seated around the fire, Jhonny's wife served us typical cassava beer, or *nijiamanch* (this brew was not yet fermented, luckily!). It is a creamy, whitish drink with a slightly acid taste. Jhonny lives in a typical Achuar house, which is oval with a high roof made of the leaves of *Phytelephas tenuicualis* (*chapi*) which can last 10 to 20 years (Fig. 13). The interesting thing about Achuar houses is the fact that they do not have walls, which gives an impression of openness and freedom.

#### Conclusion

These two expeditions underlined the importance of Kapawi for palm diversity and the Achuar's diverse palm knowledge. Kapawi's proximity to Peru allowed us to document several species and genera not previously recorded for Ecuador. As we flew back towards Shell (yes, our 3-passenger plane did come back to pick us up!), we waved goodbye to Jhonny and Kapawi Eco Lodge and hoped that ecotourism will protect this palm paradise for many generations to come.

#### Acknowledgments

The authors wish to thank Washington Santillian for assistance in the field, Jhonny Saant for sharing his knowledge of palm uses in Kapawi, and Justo Saant for support with translations and ethnobotanical research. We are also very grateful to the Kapawi Eco Lodge (https://www.kapawi.com/) for their help when receiving both teams and providing all the logistics needed. HB, RCL and JCM thank Renato Valencia, Hugo Navarrete and Mia Vedel for their valuable support with the 2011 fieldwork. TLPC is particularly grateful to Juan Carlos Garcia and Pascual Saant for the logistics. TLPC was supported by the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation program (grant agreement No. 865787, GLOBAL project). HB was supported by the Danish Council for Independent Research -Natural Sciences (grant no. 4181-00158) and the European Community (FP/2007-2013, ERC Agreement n. 212631).

#### LITERATURE CITED

- BACON, C.D. ET AL. 2021. Genomic and niche divergence in an Amazonian palm species complex. Botanical Journal of the Linnean Society 197: 498–512.
- BALSLEV, H. AND A. HENDERSON. 1986. *Elaeis* oleifera (Palmae) encontrada en el Ecualdor. Publicaciones Museo Ecuatoriano de Ciencias Naturales 5: 45–49.
- BALSLEV, H. ET AL. 2011. Species diversity and growth forms in tropical American palm communities. Botanical Review 77: 381–425.

- BORCHSENIUS, F., H.B. PEDERSEN AND H. BALSLEV. 1998. Manual to the Palms of Ecuador. AAU Reports 37, Aarhus, Denmark, 217 pp.
- CÁMARA-LERET, R. ET AL. 2018. Palmeras útiles Achuar: Amazonia ecuatoriana. 140 pg. ISBN: 978-9942-923-48-6.
- COUVREUR, T.L.P., M.L. JEANSON, J.E. GUEVARA AND A.J. HENDERSON. 2010. Palms of the south-west Cordillera Galeras, a remote premontane rain forest in eastern Ecuador. Palms 54: 94–103.
- COUVREUR, T.L.P., R. MONTÚFAR, J.N. ZAPATA, C. PERSSON AND Á.J. PÉREZ. 2022. Palms of the remote Cerro Plateado Biological Reserve, southeastern Ecuador. Palms 66: 5–19.
- DESCOLA, P. 1993. Les lances du crépuscule -Relations Jivaros, haute Amazonie. Paris: Pocket.
- EYCHENNE, J., N. ORTEGA, H. BALSLEV H., R. MUSCARELLA AND F.W. STAUFFER. 2018. Taxonomic revision, distribution and ecology of *Wendlandiella* (Arecaceae: Arecoideae: Chamaedoreeae). Webbia 73: 179–190.
- HENDERSON, A. 2000. Flora Neotropica Monograph 79: *Bactris* (Palmae). Bronx: The New York Botanical Garden.
- HENDERSON, A. 2011. A revision of *Geonoma* (Arecaceae). Phytotaxa. 17: 1–271.
- MONTÚFAR, R., C. LOUISE AND T. TRANBARGER. 2018. *Elaeis oleifera* (Kunth) Cortés: A neglected palm from the Ecuadorian Amazon. Revista Ecuatoriana de Medecina y Ciencias Biológicas. 39: 11–18.
- PINTAUD, J.-C. ET AL. 2008. The palms of South America: diversity, distribution and evolutionary history. Revista Peruviana Biologia 15: 7–29.
- RONCAL, J. 2006. Habitat differentiation of sympatric *Geonoma macrostachys* (Arecaceae) varieties in Peruvian lowland forests. Journal of Tropical Ecology 22: 483–486.
- VALENCIA, R., R. MONTÚFAR, H. NAVARRETE AND H. BALSLEV. 2013. Palmas ecuatorianas: biología y uso sostenible. Herbario QCA de la Pontificia Universidad Católica del Ecuador.

Table 1: Checklist of palm species recorded for Kapawi with total number of individuals recorded across 11 transects. HB: Henrik Balslev (vouchers deposited at AAU, QCA); TC: Thomas Couvreur (Vouchers deposited at QCA, WAG).

Species	Total individuals in 11 transects	Voucher
Aiphanes ulei (Dammer) Burret	20	HB8509
Aphandra natalia (Balslev & A.J.Hend.) Barfod	4	HB8520
Astrocaryum chambira Burret	32	HB8515
Astrocaryum urostachys Burret	237	HB8534
Attalea butyracea (Mutis ex L.f.) Wess. Boer	218	HB8517
Attalea maripa (Aubl.) Mart.	33	
Attalea phalerata Mart. ex Spreng.	6	
Bactris acanthocarpa Mart.	110	HB8527
Bactris corossilla H. Karst.	134	HB8531
Bactris gasipaes var. gasipaes Kunth	n.a.	
Bactris hirta Mart. var. hirta	13	HB8540; TC1367
Bactris maraja Mart. var. juruensis (Trail) A.J.H	end. 2	
Bactris maraja Mart. var. maraja	34	TC1337
Bactris schultesii (L.H.Bailey) Glassman	46	HB8526; TC1351
Bactris simplicifrons Mart.	54	TC1357
Chamaedorea pauciflora Mart.	43	HB8519; TC1372
Chamaedorea pinnatifrons (Jacq.) Oerst.	121	HB8513
Cocos nucifera L.	n.a.	
Desmoncus giganteus A.J.Hend	2	HB8530
Desmoncus mitis Mart. var. mitis	3	
Desmoncus polyacanthos Mart.	21	
Elaeis oleifera (Kunth) Cortés	72	HB8512; TC1365
Euterpe precatoria Mart.	190	
Geonoma brongniartii Mart.	6	HB8529
Geonoma camana Trail	2	
Geonoma longepedunculata Burret	161	HB8522
Geonoma macrostachys Mart.		
(var. <i>acaulis</i> (Mart.) Skov)	441	HB8536; TC1339
Geonoma macrostachys (var. macrostachys)	31	HB8511
Geonoma macrostachys		
(var. atrovirens Borchs. & Balslev)	13	
Geonoma macrostachys		
(as Geonoma supracostata Svenning)	468	HB8521

Table 1: Continued.		
Geonoma maxima Kunth		
subsp. multiramosa A.J.Hend.	79	HB8537; TC1369
Geonoma poeppigiana Mart.	59	HB8525
Geonoma multisecta (Burret) Burret		
(as Geonoma polyandra Skov)	49	
Geonoma stricta (Poit.) Kunth subsp. arundinacea		
(Mart.) A.J. Hend. (stricta var.		
piscicauda (Dammer) A.J.Hend.)	180	HB8510
Geonoma stricta subsp. arundinacea		
(stricta var. stricta)	630	HB8506
Geonoma stricta subsp. arundinacea		
(stricta var. trailii (Burret) A.J.Hend.)	47	HB8532; HB8538; TC1338
Hyospathe elegans Mart.	256	HB8524
Iriartea deltoidea Ruiz & Pav.	536	HB8514
Iriartella stenocarpa Burret	371	HB8528; HB8535; TC1350; TC1373
Mauritia flexuosa L. f.	11	
Oenocarpus bataua Mart.	1529	HB8516
Oenocarpus mapora H. Karst.	43	HB8523
Phytelephas tenuicaulis (Barfod) A.J.Hend.	558	HB8533
Prestoea schultzeana (Burret) H.E.Moore	37	
Socratea exorrhiza (Mart.) H.Wendl.	260	HB8518
Syagrus sancona H.Karst.		
Syagrus smithii (H.E.Moore) Glassman		
Wendlandiella gracilis Dammer var.		
simplicifrons (Burret) A.J.Hend.	157	HB8507; TC1356
Wettinia drudei (O.F.Cook & Doyle) A.J.Hend.	143	HB8539
Wettinia maynensis Spruce	43	HB8542

Table 2: Scientific and Achuar names of palms mentioned, different categories, number of uses mentioned and number of times the name was cited after 64 interviews. The table is ordered based on the decreasing number of citations. Animal food (a), Construction (b), Cultural uses (c), Human food (h), Medicinal and veterinary (m), Utensils and tools (u), and Other uses (o).

Species	Local names	Use categories	No. of uses	No. of citations
Iriartea deltoidea	Tuntuam	b, c, h, u, o	24	542
Oenocarpus bataua	Kunkuk	b, c, h, u, o	16	452
Astrocaryum chambira	Kumai	a, c, h, u, o	18	401

Table 2: Continued.				
	TT /		16	200
Bactris gasipaes var. gasipaes	Uví	a, b, c, h, m, u, o	16	398
Mauritia flexuosa	Achu	b, c, h, m, u, o	16	344
Attalea butyracea	Katira	b, h, o	10	342
Astrocaryum urostachys	Awan; Kurugurupish	b, c, h, u, o	9	286
Attalea maripa	Iñayoa; Tsentsal	x b, h, u, o	13	270
Aphandra natalia	Kintiuk	a, b, c, h, u, o	14	267
Attalea phalerata	Kuñua	b, h, u, o	11	259
Euterpe precatoria	Saké	b, c, h, o	11	244
Phytelephas tenuicaulis	Chapi	b, c, h, o	7	219
Oenocarpus minor	Shimpi	b, c, h, u, o	11	186
Socratea exorrhiza	Kupat	b, c, u, o	12	175
Wettinia maynensis	Teren	b, h, u, o	11	159
<i>Attalea</i> sp.	Kamacriña	b, h, u, o	10	148
Bactris corossilla	Murayá kamanchá	b, c, h, m, u	11	85
Syagrus sancona	Chuchuk	b, u, o	5	80
Bactris maraja	Pakaña kamanchá	b, c, h, m, u	9	79
Hyospathe elegans	Sapap	b, u	3	68
Iriartella stenocarpa	Kuuntas	b, u	4	66
Geonoma macrostachys var. acaulis	Turuji	b	1	65
Pholidostachys synanthera	Kampanak	b	1	63
Elaeis oleifera	Yunchik	a, h, u	5	62
Chamaedorea pauciflora	Yaun	С	1	61
Geonoma sp.1	Shushui turuji	b	1	61
Geonoma cf. deversa	Yunkup	b, h, u	3	57
Aiphanes ulei	Tuntuam janki	h, m, u	5	56
Syagrus smithii	Koemiank	b, u	2	48
Geonoma cf. longe- pedunculata	Murayá turuji	b	1	47
Desmoncus spp.	Makayai	b, m, u	5	45
Bactris concinna	Kamanchá	b, h, u	5	31
Geonoma stricta	Wapas turuji	b	1	16
Mauritiella armata	Achuku	b, u	4	15
Astrocaryum jauari	Wiriria	a, b, m	3	4
Geonoma sp.2	Pakaña turuji	b	1	3
Bactris riparia	Miririao	-	0	0

### A Review of Naturally Occurring Hybrids in Palms (Arecaceae)

ANDREW HENDERSON<sup>1</sup>

A review of the literature on naturally occurring hybrids in palms is presented. Approximately 114 instances of putative hybridization were found. Hybridization in palms appears to be relatively uncommon, although it may be under-reported. Within the family, there is a preponderance of records from two, taxonomically distant tribes: Trachycarpeae (Coryphoideae) and Cocoseae (Arecoideae). There does not appear to be any association between hybrids and habitat, region or morphology. Hybrids can be problematic in systematic studies, and their identification based on morphological data is challenging. There are very few polyploids in palms, and these do not appear to be a result of hybridization.

Hybrids are the result of sexual reproduction between two different taxa, usually congeneric species or subspecific taxa but sometimes between species from different genera. As such they may be isolated occurrences with no particular consequences. However, by repeated hybridization, alleles of one species may be incorporated into another (introgression). Sometimes large populations of hybrids, or hybrid zones, can form between two different, adjacent taxa. The processes of introgression and formation of hybrid zones have significant evolutionary consequences and have played a major role in the evolution of several plant families (Reiseberg & Wendel 1993).

Polyploidy, the heritable increase in genome copy number, commonly occurs in plants and may be derived from the same species (autopolyploidy), or, more often, from hybridization between different species (allopolyploidy). Wood et al. (2009) considered that most, or possibly all, plant species

<sup>1</sup>Institute of Systematic Botany New York Botanical Garden Bronx, NY 10458 USA ultimately had a polyploid ancestry. It is estimated that 15% of angiosperm speciation events were accompanied by ploidy increase (Soltis et al. 2007). It is also estimated that from one half to two-thirds of flowering plants are polyploids, including about 80% of the species in the grass family (Levy & Feldman, 2002, Soltis & Soltis 2009).

The goal of this paper is to review the literature on naturally occurring hybrids in palms and then to use the results to discuss several topics: the extent of hybridization in palms and the distribution of hybrids amongst subfamilies, tribes and subtribes; the association between hybrids and habitat, region or morphology; the implications of hybridization for palm systematics, both for revisionary and phylogenetic studies; and the role of hybrids in speciation of palms, particularly in ploidy levels.

#### Materials and Methods

A survey of the relevant literature on palms was carried out, and the results presented according to the most recent classification of the family (POWO 2022). In a few cases, personal observations from researchers were



1. Coccothrinax argentata × Leucothrinax morrisii on Big Pine Key, Florida. Photo by Paul Craft.



2. Coccothrinax macroglossa × C. pseudorigida in Camagüey, Cuba. Photo by D.R. Hodel.



3. Coccothrinax crinita subsp. brevicrinis (left), C. × angelae (center) and C. miraguama subsp. roseocarpa (right). Photo by Paul Craft.

included. Numbers of species per genus and nomenclature were taken from POWO (2022) or from the most recent monographs, and distributions were taken from Dransfield et al. (2008). References to cultivated palms were omitted because there are so many of them and they are not directly related to the subject. Many of the hybrids reported here are somewhat anecdotal and based on little data and are often referred to as possible hybrids or potential hybrids. All published reports are here assumed to represent actual hybrids regardless of their level of supporting data. Genera for which no natural hybrids are known are listed in Appendix 1.

#### Results

#### CALAMOIDEAE

#### CALAMEAE, METROXYLINAE

The subtribe comprises one genus (*Metroxylon*, 7 species) distributed in the Asian tropics. McClatchey (1998) reported that, on Samoa, two species (*M. paulcoxii* and *M. warburgii*) were sympatric and that one specimen might be a hybrid between the two. However, the sympatry of the two species was possibly an artifact of human activity.

#### CALAMEAE, CALAMINAE

The subtribe comprises one genus (*Calamus*, 411 species) widely distributed throughout the

Asian tropics with one species in Africa. Henderson (2020a) reported one possible naturally occurring hybrid, involving *C. bacularis* and *C. myriacanthus*, and a few, other potential cases (*C. javensis* and *C. tenompokensis*; *C. leloi* and an unknown species; *C. insignis* and *C. longiusculus*).

#### CORYPHOIDEAE

#### SABALEAE

The tribe comprises one genus (Sabal, 16 species) distributed in Central America and the Caribbean. In one of the most detailed studies of hybridization in palms, Goldman et al. (2011), using molecular data (AFLPs), considered that a population of Sabal in Texas was a hybrid between S. minor and S. palmetto  $(S. \times brazoriensis)$ . They postulated that the initial hybridization event could have taken place thousands of years ago when the geographic distribution of the parent species was different. They considered it possible that caulescent plants of S. minor as well as S. × brazoriensis could represent an extensive regional hybrid swarm between acaulescent S. minor and S. palmetto (see also Simpson 1988). Their data also suggested that a Mexican sample of S. minor showed evidence of introgression with S. mexicana.

#### CRYOSOPHILEAE

The tribe comprises 11 genera (*Schippia*, 1 species; *Trithrinax*, 3 species; *Zombia*, 1 species;



4 (left). *Copernicia* × *vespertilionum* in Sancti Spíritus, Cuba. 5 (right). *Copernicia* × *sueroana* in Las Tunas, Cuba. Photos by Paul Craft.

*Coccothrinax*, 56 species; *Hemithrinax*, 3 species; *Leucothrinax*, 1 species; *Thrinax*, 3 species; *Chelyocarpus*, 3 species; *Cryosophila*, 10 species; *Itaya*, 1 species; *Sabinaria*, 1 species) distributed throughout the Neotropics.

Naturally occurring intergeneric hybrids between *Coccothrinax* and *Thrinax* have been reported from the Florida Keys (Nauman 1989, 1990) (Fig. 1). These hybrids were found to be sterile.

In *Coccothrinax*, hybridization is common amongst cultivated species (particularly in botanic gardens), and there appear be several naturally occurring hybrids in Cuba (Craft 2017). Craft considered that *C. orientalis* and *C. pauciramosa* could hybridize, and also *C. macroglossa* and *C. pseudorigida* (Fig. 2). Suárez Oropesa (2015), based on morphological data, described a naturally occurring hybrid from Cuba between *C. crinita* and *C. miraguama* as *C. × angelae* (Fig. 3).

#### PHOENICEAE

The tribe comprises one genus (*Phoenix*, 13 species) widely occurring in Old World tropical and subtropical areas. The species are well-known to hybridize freely in cultivation (Dransfield et al. 2008, Gros-Balthazard 2013).

González-Peréz et al. (2004), using molecular data (RAPD markers) reported that on the Canary Islands, the naturally occurring *P. canariensis* hybridized with the introduced *P. dactylifera*. Pérez-Escobar et al. (2021) discussed the ancient hybrid origin of the date palm, *P. dactylifera*. They considered that its evolution was influenced by gene flow from two wild relatives, *P. theophrasti*, currently restricted to Crete and Turkey, and *P. sylvestris*, widespread from Bangladesh to the West Himalayas (see also Flowers et al. 2019).

#### TRACHYCARPEAE, LIVISTONINAE

The subtribe comprises six genera (*Livistona*, 28 species; *Licuala*, 167 species; *Lanonia* 13 species; *Johannesteijsmannia*, 4 species; *Pholidocarpus*, 6 species; *Saribus*, 9 species) distributed throughout the Asian tropics and subtropics and just reaching north-east Africa.

There are several anecdotal reports of hybridization in *Livistona* in Australia. Tucker (1975) reported a natural hybrid between *L. australis* and *L. humilis*. Dowe (2009) considered that the ranges of *L. australis* and *L. decora* overlapped in some areas and there could be hybridization between the two. Rodd (1998) mentioned possible hybrids between *L*.



6. Copernicia × shaferi in Camagüey, Cuba. Photo by D.R. Hodel.

*humilis* and *L. inermis* and considered it possible that *L. leichhardtii* hybridized with *L. lorophylla* and *L. mariae*. Henderson and Nguyen Quoc Dung (2019) considered that *L. jenkinsiana* and *L. saribus* formed hybrids in northern Vietnam.

Miyamoto et al. (2006) studied a population of *Licuala* at a site in lowland rainforest in Brunei, using morphological and molecular data. They considered that intermediate plants between *L. paludosa* and *L. bruneiana* could be hybrids between the two species.

Bacon et al. (2016a) found no evidence of hybridization between *Johannesteijsmannia* species, despite the species occurring in close proximity in Peninsular Malaysia.

#### UNPLACED GENERA OF TRACHYCARPEAE

This group comprises seven genera (*Acoelorraphe*, 1 species; *Serenoa*, 1 species; *Brahea*, 11 species; *Colpothrinax*, 3 species; *Copernicia*, 22 species; *Pritchardia*, 29 species; *Washingtonia*, 2 species) distributed in the New World tropical and subtropical regions, with the exception of the Pacific island *Pritchardia*.

Bacon et al. (2012a) considered that there was no evidence of hybridization in *Brahea*. Ramírez-Rodríguez et al. (2011, 2012), based on morphological and molecular data, showed evidence of hybridization between *B. dulcis* and *B. calcarea* in Mexico.

Six naturally occurring hybrids were reported in Copernicia from Cuba by Dahlgren and Glassman (1963): C. × vespertilionum (Fig. 4), C.  $\times$  sueroana (Fig. 5), C.  $\times$  burretiana, C.  $\times$ shaferi (Fig. 6), C. × textilis, and C. × occidentalis. Moya López et al. (2019) accepted a seventh species, C. × escarzana. The widely distributed C. hospita is a parent of five of these hybrids. Other species such as C. curbeloi, C. longiglossa and C. molinetii, may also be of hybrid origin, as well as C. oxycalyx (Craft 2017). Craft noted that most hybrids occurred in central and eastern Cuba, where species of Copernicia occurred in close proximity (see also Moya López & Mayotte 1996). In some places large numbers of hybrids occurred. Verdecia Pérez (2016), based on morphological data, described a natural hybrid, C. × dahlgreniana, in Cuba with intermediate characteristics of C. cowellii and C. macroglossa. Most species of Copernicia are endemic to Cuba and the common occurrence of hybridization has been a challenge for taxonomic revisions.

For *Pritchardia*, Hodel (1980) reported that species freely hybridized in cultivation, although Hodel (2007a) did not report any naturally occurring hybrids. He noted that species seldom occurred sympatrically. Bacon et al. (2012b) noted the difficulty of delimiting sympatric Hawaiian species because of their similarity due to recent divergence and their phenotypic plasticity. They suggested that hybridization had played an important role in the diversification of species from regions of sympatry, for example in Kauai (*P. flynnii, P. napaliensis, P. minor, P. waialealeana* and *P. viscosa*) and Oahu (*P. bakeri, P. kahukuensis* and *P. martii*).

#### CARYOTEAE

The tribe comprises three genera (*Arenga*, 24 species; *Caryota*, 14 species; *Wallichia*, 8 species) widely distributed in the Asian tropics.

Whitmore (1998) described "hybrid swarms" in Peninsular Malaysia between *Arenga westerhoutii* and *A. pinnata*, although he considered the latter to be introduced to the region.

Hahn (1993) suggested that *Caryota sympetala* was of hybrid origin, an idea not supported by Jeanson (2011). Hahn and Sytsma (1999) considered that there were several instances of hybridization in *Caryota*, including between sympatric species from the Philippines (*C. cumingii* and *C. rumphiana; C. cumingii* and *C. mitis*) and from Vietnam (*C. monostachya* and *C. maxima*).

Henderson (2007) proposed that one specimen of *Wallichia* could be a hybrid between *W. marianneae* and *W. caryotoides*, but this specimen was determined as *W. gracilis* by Jeanson (2011).

#### CEROXYLOIDEAE

#### CEROXYLEAE

The tribe comprises four genera (*Ceroxylon*, 12 species; *Juania*, 1 species; *Oraniopsis*, 1 species; *Ravenea* 22 species) distributed in South America, Madagascar, Australia, and Juan Fernandez.

María José Sanín (pers. comm.) reported isolated individuals of *Ceroxylon* in Colombia being morphologically intermediate between *C. quindiuense* and *C. parvifrons, C. quindiuense* and *C. alpinum,* and *C. quindiuense* and *C. vogelianum.* In Peru, a population of individuals appeared intermediate between *C. peruvianum* and *C. echinulatum.* 

#### PHYTELEPHEAE

The subtribe comprises three genera (*Ammandra*, 1 species; *Aphandra*, 1 species;

*Phytelephas*, 6 species) distributed in the Neotropics.

Barfod (1991) considered that *Phytelephas* seemannii subsp. brevipes from Panama was similar to *P. macrocarpa* subsp. schottii and could a hybrid between *P. seemannii* and *P. macrocarpa*.

#### ARECOIDEAE

#### IRIARTEEAE

The tribe comprises five genera (*Iriartella*, 2 species; *Dictyocaryum*, 3 species; *Iriartea*, 1 species; *Socratea*, 5 species; *Wettinia*, 22 species) widely distributed in the Neotropics.

Bacon et al. (2016b) considered that in Colombia there was high potential for hybridization between the widely distributed *Wettinia kalbreyeri* and the more narrowly distributed *W. disticha*. Galeano and Bernal (2010) considered that *W. oxycarpa* could be a natural hybrid between *W. quinaria* and *W. kalbreyeri*.

#### CHAMAEDOREEAE

The tribe comprises five genera (*Hyophorbe*, 5 species; *Wendlandiella*, 1 species; *Synechanthus*, 3 species; *Chamaedorea*, 104 species; *Gaussia*, 5 species) widely distributed in the Neotropics, with the exception of *Hyophorbe* from the Mascarene Islands. There are no records of natural hybrids in the tribe. Hybrids between species of *Hyophorbe* are common in cultivation (e.g., Hung & Pan 2014; see also Maunder et al. 2002). Hodel (1992) considered that natural hybrids probably did not occur in *Chamaedorea*, although several artificial hybrids are known from cultivation.

#### COCOSEAE, ATTALEINAE

The subtribe comprises 10 genera (*Beccariophoenix*, 3 species; *Jubaeopsis*, 1 species; *Voanioala*, 1 species; *Allagoptera*, 6 species; *Attalea*, 30 species; *Butia*, 22 species; *Cocos*, 1 species; *Jubaea*, 1 species; *Syagrus*, 67 species; *Parajubaea* 3 species) widely distributed in the Neotropics, southern Africa, and Madagascar.

Noblick (2017) reported a hybrid between *Butia odorata* and *Syagrus romanzoffiana*,  $\times$  *Butyagrus nabonnandii*. This cross was first made from cultivated plants in Europe in the 19<sup>th</sup> century but was later found to occur naturally in Uruguay.

Moraes (1996) considered that there might be hybrids between *Allagoptera campestris* and *A*.



Fig. 7. Attalea piassabossu in eastern Brazill. Photo by L. Noblick.

*leucocalyx* in areas where the two species overlapped, and from where intermediate specimens are known.

Henderson (2020b) considered that in at least half of the 30 species of *Attalea* there was evidence of hybridization, including single instances of hybrids as well as large scale hybrid zones. Hybrids greatly complicated the taxonomy of the genus. Some of the most detailed studies of hybrids in Attalea, based on morphological data, were carried out by Michael Balick and colleagues (e.g., Balick et al. 1987a, 1987b; see also Anderson & Balick 1988). The following hybrids have been postulated (Henderson 2020b; see also Pinheiro 1997): A. × minarum, A. × teixeriana, A. piassabossu (Fig. 7), A. × voeksii, A. camopiensis, A. degranvillei, (A. maripa × ?A. guianensis), A. salvadorensis, A. dahlgreniana, , (A. nucifera × A. butyracea) and (A. sagotii × A. barreirensis).

Gaiero et al. (2011) studied genetic diversity in four species of Butia in Uruguay, B. lallemantii, B. paraguayensis, B. yatay and B. capitata. They found that variability within populations was high, possibly due to gene flow, past hybridization or life history traits. In particular, they considered that current or recent hybridization between *B. lallemantii* and B. paraguayensis could explain the high number of admixed genotypes observed in both species. Brussa and Grela (2007) considered that B. lallemantii and Syagrus romanzoffiana formed hybrids in Uruguay. They also listed × Butyagrus nabonnandii as occurring naturally in riverine forests and hills in eastern Uruguay. Engels et al. (2021) described a new hybrid between B. eriospatha and S. romanzoffiana, which they called × Butyagrus paranaensis. It occurred in forests in the central-south region of Paraná state, Brazil. Soares et al. (2014) described × Butyagrus alegretensis from southern Brazil, a hybrid between B. lallemantii and S. romanzoffiana. Noblick (2014) suspected hybridization between *B. paraguayensis* and *B.* yatay along the Paraná River, which defines part of the border between Paraguay and Argentina.

Hodel (2011) discussed *Syagrus* hybrids in both cultivation and the wild. Noblick (2017) recognized 14 naturally occurring hybrids in *Syagrus*, mostly from eastern Brazil. He considered that hybrids had a low frequency of occurrence and were often characterized by low fruit production, low seed germination

and by hybrid vigor, making the hybrid more robust than either of its parents. Some species were remarkably promiscuous. Syagrus coronata formed hybrids with several other species, including S. vagans, S. microphylla, S. botryophora schizophylla, S. and S. romanzoffiana. Syagrus romanzoffiana formed hybrids with other species including S. coronata, S. flexuosa, S. oleracea and S. glaucescens. Noblick (2017) recognized the following hybrids: S. × lacerdamourae, S. × costae, S. × campos-portoana, S. × mirandana, S. × tostana, S. × matafome, S. × andrequiceana, S. × serroana, (S. loefgrenii × S. romanzoffiana), S. × teixeiriana, S. × altopalacioensis, (S. pleioclada × S. glaucescens), S. × cipoensis and (S. comosa × S. elata).

#### COCOSEAE, BACTRIDINAE

The subtribe comprises five genera (*Acrocomia*, 9 species; *Astrocaryum*, 39 species; *Aiphanes*, 38 species; *Bactris*, 79 species; *Desmoncus*, 24 species), widely distributed throughout the Neotropics.

Wessels Boer (1971) described a hybrid, *Bactris* × *moorei*, between *B. acanthocarpa* and *B. oligoclada* in Venezuela. Henderson (2000) reported several hybrids in *Bactris*: (*B. mexicana* × *B. gracilior*), (*B. acanthocarpa* × unknown species), (*B. barronis* × *B. gasipaes*), (*B. borongniartii* × *B. major*), (*B. caudata* × *B. coloradonis*), (*B. corossilla* × *B. maraja*), (*B. setulosa* × *B. major*) and (*B. setiflora* × *B. corossilla*). Couvreur et al. (2006) reported possible hybrids between wild and cultivated *B. gasipaes* in western Ecuador.

Henderson (2011a) reported that several, suspected hybrids were present amongst specimens of *Desmoncus*. Most of these involved *D. polyacanthos*, the most widespread and variable species, and either *D. pumilus*, *D. mitis* or *D. horridus*. All hybrids came from the central and western Amazon region, and none occurred in other regions: (*D. polyacanthos* × *D. horridus*), (*D. leptoclonos* × unknown species), (*D. polyacanthos* × *D. mitis*), (*D. parvulus* × unknown species) and (*D. polyacanthos* × *D. pumilus*).

#### EUTERPEAE

The tribe comprises five, Neotropical genera (*Hyospathe*, 6 species; *Euterpe*, 7 species; *Prestoea*, 10 species; *Neonicholsonia*, 1 species; *Oenocarpus*, 9 species).

Henderson (2004) reported that there were hybrids between subspecies of *Hyospathe* 

*elegans*, and a possible hybrid zone between *H. elegans* and *H. pittieri* along eastern Andean slopes.

Pereira et al. (2022) found two geographical genetic groups in *Euterpe edulis* from the Atlantic Coastal Forest of Brazil, a northern and a southern group, with a region of hybridization in the contact zone.

Balick (1991) described a hybrid between *Oenocarpus bacaba* and *O. minor, O.* × *andersonii,* from a single locality near Manaus in Brazil. Balick (1988) reported on hybrids between *O. bataua* and *O. bacaba* in Colombia, and Henderson et al. (1995) listed hybrids between *O. bataua* and *O. bacaba* and *O. bataua* 

#### GEONOMATEAE

The tribe comprises six, Neotropical genera (*Welfia*, 2 species; *Pholidostachys*, 8 species; *Calyptrogyne*, 18 species; *Calyptronoma*, 3 species; *Asterogyne*, 5 species; *Geonoma*, 68 species).

Henderson (2005) reported hybrids between *Calyptrogyne trichostachys* and *C. brachystachys*, and between *C. panamensis* and *C. fortunensis*, and a possible hybrid zone in Central America between *C. brachystachys* and *C. ghiesbreghtiana*.

There are numerous sympatric species of Geonoma in Neotropical forests. Knudsen (1999), Borchsenius (2002), and Borchsenius et al. (2016) have shown how differences in phenology and floral scent contribute to reproductive isolation in sympatric species or forms. Henderson (2011b) reported numerous hybrids in *Geonoma*, including three possible hybrid zones. The most convincing of these was a hybrid zone in the northeastern Amazon region involving G. deversa and G. leptospadix. He also reported numerous hybrids between subspecies within species. Henderson reported the following interspecific hybrids, four of which involved the widespread G. deversa: (G. deversa  $\times$  G. brongniartii), (G. deversa  $\times$  G. leptospadix), (G. deversa  $\times$  G. occidentalis), (G. deversa  $\times$  G. longivaginata), (G. poeppigiana  $\times$  G. brongniartii), (G. camana × G. macrostachys), (G. elegans  $\times$  G. pauciflora), (G. orbignyana  $\times$  G. undata), (G. interrupta  $\times$  G. pinnatifrons), (G. poeppigiana × unknown species), (G. pohliana  $\times$  G. pauciflora), (G. schottiana  $\times$  G. pauciflora) and (G. undata  $\times$  G. trigona).

#### ARECEAE, BASSELINIINAE

The subtribe comprises six genera (*Basselinia*, 14 species; *Burretiokentia*, 5 species;

*Cyphophoenix*, 4 species; *Cyphosperma*, 5 species; *Lepidorrhachis*, 1 species; *Physokentia*, 7 species), distributed in New Caledonia, Vanuatu, Fiji, the Solomon Islands, and the Bismarck Archipelago.

Pintaud (2006) reported on hybrids in Basselinia in disturbed forests in New Caledonia between *B. pancheri* and *B. gracilis* and between *B. pancheri* and *B. deplanchei*. In some areas *B. vestita* formed hybrids with *B. gracilis* and *B. pancheri*.

#### ARECEAE, LINOSPADICINAE

The subtribe comprises four genera (*Calyptrocalyx*, 27 species; *Linospadix*, 7 species; *Howea*, 2 species; *Laccospadix*, 1 species) distributed in the Moluccas, New Guinea, Australia, and Lord Howe Island.

Babik et al. (2009) and Hipperson et al. (2016) reported hybrids between *Howea belmoreana* and *H. forsteriana* on Lord Howe Island (Figs. 8 & Front Cover). However, they considered that gene flow between the two species was extremely limited.

#### ARECEAE, ONCOSPERMATINAE

The subtribe comprises four genera (*Oncosperma*, 5 species; *Deckenia*, 1 species; *Acanthophoenix*, 3 species; *Tectiphiala*, 1 species) widely distributed in the Asian tropics and Indian Ocean islands.

Adorador and Fernando (2017) considered that a population of *Oncosperma* on Samar Island in the Philippines could be of hybrid origin, although neither of the putative parents (*O. horridum* and *O. gracilipes*) occurs on Samar.

Ludwig (2006) reported that the allopatric distribution of *Acanthophoenix* meant that the possibility of hybridization in the wild was remote. However, she reported hybrids in plantations.

#### UNPLACED MEMBERS OF ARECEAE

The group comprises ten genera (*Bentinckia*, 2 species; *Clinostigma*, 11 species; *Cyrtostachys*, 7 species; *Dictyosperma*, 1 species; *Dransfieldia*, 1 species; *Heterospathe*, 39 species; *Hydriastele*, 39 species; *Iguanura*, 34 species; *Loxococcus*, 1 species; *Rhopaloblaste* 6 species) widely distributed throughout the Old World tropics.

Hodel (2007b) reported that a possible hybrid between *Clinostigma samoense* and *C. warburgii* occurred on Samoa.

#### Discussion

There are 180 currently recognized genera of palms. Naturally occurring hybrids have been



8. *Howea belmoreana* (front left), *H forsteriana* (front right) and putative hybrid (back left) on Lord Howe Island. Photo by W. Baker.

reported in 26 of these (14% of the total). However, of the 180 genera, 54 are monospecific and about 40 have few, allopatric species. This leaves about 90 genera having two or more species some of which are distributed sympatrically in which hybrids could potentially occur. Hybrids are reported in 29% of these genera.

There are about 2,533 currently recognized species of palms, and of these 26 are hybrid species (i.e., ones that have been officially described and currently recognized). These represent <1% of the total number of recognized species. About 114 instances of naturally occurring hybridization have been reviewed here. Because several of these have the same parent, only about 156 species (6% of the total number of recognized species) are involved in hybridization.

Hybrids are not evenly distributed amongst subfamilies, tribes and subtribes. At the subfamily level, most hybrids occur in the Arecoideae (73 hybrids, 64% of the total). However, there are many more species overall in the Arecoideae, about 50% of the total number of palm species. Fewer hybrids occur in the Coryphoideae (31 hybrids, 27% of the total), and fewer still in the Calamoideae (5 hybrids, 4% of total) and Ceroxyloideae (5 hybrids, 4% of total). At the tribal level, in the Arecoideae there are 48 hybrids reported for the Cocoseae, 16 for the Geonomateae, 4 for the Euterpeae, 4 for the Areceae, and 1 for Iriarteeae. The high number for the Cocoseae is based mostly on three genera in the subtribe Attaleinae (Attalea, Butia, Syagrus) and two in the subtribe Bactridinae (Bactris, Desmoncus). At the tribal level in the Coryphoideae, there are 20 hybrids reported for the Trachycarpeae, 4 each for the Cryosophileae and Caryoteae and 2 for the Sabaleae. The relatively high number for Trachycarpeae is based mostly on Livistona and Copernicia. Within genera it appears that the most widespread and variable species are more likely to form hybrids. Examples are Attalea maripa and A. speciosa, Syagrus coronata and S. romanzoffiana, Bactris acanthocarpa and B. major, Desmoncus polyacanthos, Oenocarpus bacaba, Geonoma deversa, Sabal palmetto, Livistona australis and Copernicia hospita.

Hybrids do not appear to be strongly connected with any particular habitat. Using the rather broad categorization of habitats of Henderson (2002)—lowland moist forests; montane moist forests; dry forests; grasslands, savannas and shrublands; and desert and xeric shrublands—most hybrids occur in lowland moist forests (55%), followed by dry forests (21%), and grasslands, savannas and shrublands (16%). Few hybrids are found in montane moist forests (6%) or desert and xeric shrublands (2%). These are approximately the same percentages of habitat distribution for all species of palm.

Hybrids do not appear linked to any particular continent, region, or island, although the majority (83%) are from the Neotropics. Two neotropical areas have high numbers of hybrids: Cuba and eastern Brazil.

Hybrids do not seem to be associated with any particular aspect of morphology, such as stem type, inflorescence type or germination type (Henderson 2002), nor with any particular type of pollination (although pollinators are not recorded for most parent species).

Most hybrids reviewed here are reported by systematists who are revising genera based on herbarium specimens. Different systematists have different propensities to recognize hybrids. Hybrids are anyway difficult to detect from specimens and may be under-reported. They are usually postulated based on sympatry of putative parents and morphological intermediacy of putative hybrids. Furthermore, the sample size of most species in herbaria is extremely small compared with population size, and thus few hybrid specimens may be present in samples. Hybrids appear to be a limited problem for systematists. Most reported hybrids (about 70%) appear to be isolated occurrences and may have little significance. About 20 hybrids are said to be relatively widespread, and seven hybrid zones have been reported. Some genera certainly appear to be more problematic than others for systematists, for example Attalea, Syagrus and *Copernicia*. As yet there are no studies that address the problem of palm hybrids in inferring phylogenies.

Most of the hybrids reviewed here are presumed to be contemporary or recent events, and most studies are based on morphological data. Only a few studies are based on molecular data, and of these some have addressed ancient hybrid events. Molecular data will no doubt become the norm for investigating hybrids and in studying the role of hybridization, if any, in the formation of species complexes.

Polyploidy, the duplication of whole genomes, is thought to have occurred in virtually all lineages of angiosperms (Alix et al. 2017). However, polyploidy appears to be uncommon in palms. Barrett et al. (2019) found evidence for a whole genome duplication event early in the evolution of the family. In general, they found that an ascending dysploid series of

chromosome numbers (i.e., 2n = 26, 28, 30,32, 34, 36) appeared to be the predominant mode of chromosomal change in palms, rather than repeated evolution of polyploidy. Dransfield et al. (2008), in a review of chromosome numbers in palms, found that only four cases of polyploidy had been reliably reported, presumably all autopolyploids, and thus not involving hybridization (compared with estimates of 60-75% polyploids in monocotyledons in general, Keeler 1998). Two of the palm polyploidys (Arenga caudata, Rhapis *humilis*) were tetraploids in otherwise diploid genera. The remaining two were polyploids in monotypic genera in the subtribe Attaleinae; Jubaeopsis from South Africa (2n = 160-200)and *Voanioala* from Madagascar (2n = ca. 600), the latter having the highest chromosome number amongst the monocotyledons. Though not a polyploid, Beccariophoenix, another monotypic member of the Attaleinae from Madagascar, was also distinct in having a diploid count of 2n = 36 as opposed to 2n =32 in the remaining species of the Attaleinae. Gunn et al. (2015) considered that genome size in the Attaleinae suggested that polyploidy also occurred in the neotropical Allagoptera caudescens. The unusual chromosome numbers in the subtribe Attaleinae are concentrated in the three genera (Beccariophoenix, Jubaeopsis, *Voanioala*) that occur outside the subtribe's principal distribution in the Neotropics. The remaining neotropical genera of the subtribe produce a high number of hybrids, particularly Attalea, Butia and Syagrus. The significance of this is unknown.

In summary, many of the records of naturally occurring hybrids appear somewhat anecdotal, and many seem to have been as a result of human disturbance. This review gives the general impression that congenerics can hybridize if brought together artificially but seldom do so naturally. However, it is certainly possible that hybrids are under-reported. Just over half of all reported hybrids are from two tribes: Trachycarpeae and Cocoseae.

#### Acknowledgments

I thank Christine Bacon and Adrian Hill of the University of Gothenburg for their help with an earlier draft of this paper, and William Baker, Paul Craft, Donald Hodel and Larry Noblick for supplying images.

#### LITERATURE CITED

ADORADOR, J.T. AND E.S. FERNANDO. 2017. Palms of Samar Island, Philippines. Palms 61: 161–195.

- ALIX, K., P.R. GÉRARD, T. SCHWARZACHER AND J.S. HESLOP-HARRISON. 2017. Polyploidy and interspecific hybridization: partners for adaptation, speciation and evolution in plants. Annals of Botany 120: 183–194.
- ANDERSON, A.B. AND M.J. BALICK, M. 1988. Taxonomy of the babassu complex (*Orbignya* spp.: Palmae). Systematic Botany 13: 32–50.
- BABIK, W., ET AL. 2009. How sympatric is speciation in the Howea palms of Lord Howe Island? Molecular Ecology 18: 3629–3638.
- BACON, C.D., W.J. BAKER AND M.P. SIMMONS. 2012a. Miocene dispersal drives island radiations in the palm tribe Trachycarpeae (Arecaceae). Systematic Biology. 61: 426–442.
- BACON, C.D., M.J. MCKENNA, M.P. SIMMONS AND W.L. WAGNER. 2012b. Evaluating multiple criteria for species delimitation: an empirical example using Hawaiian palms (Arecaceae: *Pritchardia*). BMC Evolutionary Biology 2012: 12–23.
- BACON, C.D., ET AL. 2016a. Species limits, geographical distribution and genetic diversity in *Johannesteijsmannia* (Arecaceae). Botanical Journal of the Linnean Society 182: 318–347.
- BACON, C.D., ET AL. 2016b. Phylogenetics of Iriarteeae (Arecaceae), cross-Andean disjunctions and convergence of clustered infructescence morphology in *Wettinia*. Botanical Journal of the Linnean Society 182: 272–286.
- BALICK, M.J. 1988. *Jessenia* and *Oenocarpus*: neotropical oil palms worthy of domestication. FAO Plant Production and Protection Paper 88. Rome.
- BALICK, M.J. 1991. A new hybrid palm from Amazonian Brazil, *Oenocarpus × andersonii*. Boletim do Museu Paraense Emílio Goeldi 7: 505–509.
- BALICK, M.J., A.B. ANDERSON AND J.T. MEDEIROS-COSTA. 1987a. Hybridization in the babassu palm complex. II. *Attalea compta × Orbignya oleifera* (Palmae). Brittonia 39: 26–36.
- BALICK, M.J., C.U.B. PINHEIRO AND A.B. ANDERSON. 1987b. Hybridization in the babassu palm complex: I. *Orbignya phalerata* × O. *eichleri*. American Journal of Botany 74: 1013–1032.
- BARFOD, A. 1991. A monographic study of the subfamily Phytelephantoideae (Arecaceae). Opera Botanica 105: 1–73.

- BARRETT, C., ET AL. 2019. Ancient polyploidy and genome evolution in palms. Genome Biology and Evolution 11: 1501–1511.
- BORCHSENIUS, F. 2002. Staggered flowering in four sympatric varieties of *Geonoma cuneata* (Palmae). Biotropica 34: 603–606.
- BORCHSENIUS, F., T. LOZADA AND J.T. KNUDSEN. 2016. Reproductive isolation of sympatric forms of the understorey palm *Geonoma macrostachys* in western Amazonia. Botanical Journal of the Linnean Society 182: 398– 410.
- BRUSSA, C.A. AND I. GRELA. 2007. Flora Arbórea del Uruguay. Con énfasis en las especies de Rivera y Tacuarembó. COFUSA.
- COUVREUR, T.L.P., ET AL. 2006. Close genetic proximity between cultivated and wild *Bactris gasipaes* Kunth revealed by microsatellite markers in Western Ecuador. Genetic Resources and Crop Evolution 53: 1361–1373.
- CRAFT, P. 2017. The Palms of Cuba. Palm Nut Pages, Florida.
- DAHLGREN, B.E. AND S.B. GLASSMAN. 1963. A revision of the genus *Copernicia*. 2. West Indian species. Gentes Herbarum 9: 43–232.
- Dowe, J.L. 2009. A taxonomic account of *Livistona* R.Br. (Arecaceae). Gardens' Bulletin Singapore 60: 185–344.
- DRANSFIELD, J., N.W. UHL, C.B. ASMUSSEN, W.J. BAKER, M.M. HARLEY AND C.E. LEWIS. 2008. Genera Palmarum. The Evolution and Classification of Palms. Kew Publishing, Royal Botanic Gardens, Kew, London, U.K.
- ENGELS, M.E., T.A. MEYER AND K.P. SOARES. 2021. A new  $\times$  *Butyagrus* (Arecaceae) from the Brazilian South Plateau. Hoehnea 48: e412020.
- FLOWERS, J.M., ET AL. 2019. Cross-species hybridization and the origin of North African date palms. PNAS 116: 1651–1658.
- GAIERO, P., C. MAZZELLA, G. AGOSTINI, S. BERTOLAZZI AND M. ROSSATO. 2011. Genetic diversity among endangered Uruguayan populations of *Butia* Becc. species based on ISSR. Plant Systematics and Evolution 292: 105–116.
- GALEANO, G. AND R. BERNAL. 2010. Palms de Colombia. Guía de campo. Universidad Nacional de Colombia, Bogotá.
- GOLDMAN, D.H., M.R. KLOOSTER, M.P. GRIFFITH, M.F. FAY AND M.W. CHASE. 2011. A

preliminary evaluation of the ancestry of a putative *Sabal* hybrid (Arecaceae: Coryphoideae), and the description of a new nothospecies, *Sabal* × *brazoriensis*. Phytotaxa 27: 8–25.

- GONZÁLEZ-PERÉZ, M.A., J. CAUJAPÉ-CASTELLS AND P.A. SOSA. 2004. Allozyme variation and structure of the Canarian endemic palm tree *Phoenix canariensis* (Arecaceae): implications for conservation. Heredity 93: 307–315.
- GROS-BALTHAZARD, M. 2013. Hybridization in the genus *Phoenix*. Emirates Journal of Food and Agriculture 25: 831–842.
- GUNN, B.F., ET AL. 2015. Ploidy and domestication are associated with genome size variation in palms. American Journal of Botany 102: 1625–1633.
- HAHN, W.J. 1993. Biosystematics and evolution of the genus *Caryota* (Palmae: Arecoideae). Ph.D. thesis, University of Wisconsin, Madison.
- HAHN, W.J. AND K.J. SYTSMA. 1999. Molecular systematics and biogeography of the southeast Asian genus *Caryota* (Palmae). Systematic Botany 24: 558–580.
- HENDERSON, A. 2000. *Bactris* (Palmae). Flora Neotropica Monograph 79: 1–181.
- HENDERSON, A. 2002. Evolution and ecology of palms. New York Botanical Garden Press.
- HENDERSON, A. 2004. A multivariate analysis of *Hyospathe* (Palmae). American Journal of Botany 91: 953–965.
- HENDERSON, A. 2005. A multivariate study of *Calyptrognye* (Palmae). Systematic Botany 30: 60–83.
- HENDERSON, A. 2007. A revision of *Wallichia* (Palmae). Taiwania 52: 1–11.
- HENDERSON, A. 2011a. A revision of *Desmoncus* (Arecaceae). Phytotaxa 35: 1–88.
- HENDERSON, A. 2011b. A revision of *Geonoma* (Arecaceae). Phytotaxa 17: 1–271.
- HENDERSON, A. 2020a. A revision of *Calamus* (Arecaceae, Calamoideae, Calameae, Calaminae). Phytotaxa 445: 1–656.
- HENDERSON, A. 2020b. A revision of *Attalea* (Arecaceae, Arecoideae, Cocoseae, Attaleinae). Phytotaxa 444: 1–76.
- HENDERSON, A. AND NGUYEN QUOC DUNG. 2019. Palms of Vietnam. New York Botanical Garden.

- HENDERSON, A., G. GALEANO AND R. BERNAL. 1995. A Field Guide to the Palms of the Americas. Princeton University Press.
- HIPPERSON, H., ET AL. 2016. Ecological speciation in sympatric palms: 2. Pre– and post–zygotic isolation. Journal of Evolutionary Biology 29: 2143–2156.
- HODEL, D.R. 1980. Notes on *Pritchardia*. Principes 24: 65–81.
- HODEL, D.R. 1992. *Chamaedorea* Palms. The International Palm Society and Allen Press, Lawrence, Kansas.
- HODEL, D.R. 2007a. A review of the genus *Pritchardia*. Palms 51(suppl.): 1–53.
- HODEL, D.R. 2007b. Unravelling *Clinostigma* in Samoa. Palms 51: 11–29.
- HODEL, D. 2011. Hybrids in the genus *Syagrus*. Palms 55: 141–154.
- HUNG, S.F. AND F.J. PAN. 2014. Identifying the natural hybrid between *Hyophorbe verschaffeltii* and *H. lagenicaulis* by RAPD markers. Journal of the Experimental Forest of National Taiwan University 28: 123–130.
- JEANSON, M. 2011. Systematique de la tribu des Caryoteae (Arecaceae). Ph.D. thesis, Muséum national d'Histoire naturelle, Paris.
- KEELER, K.H. 1998. Population biology of intraspecific polyploidy in grasses, pp. 183–206, in CHEPLIK, G.P. (ed.) Population Biology of Grasses. Cambridge, Cambridge University Press.
- KNUDSEN, J.T. 1999. Floral scent differentiation among coflowering, sympatric species of *Geonoma* (Arecaceae). Plant Species Biology 14: 137–142.
- LEVY, A.A. AND M. FELDMAN. 2002. The impact of polyploidy on grass genome evolution. Plant Physiology 130: 1587–1593.
- LUDWIG, N. 2006. *Acanthophoenix* in Réunion, Mascarene Islands. Palms 50: 82–98.
- MAUNDER, M., ET AL. 2002. The decline and conservation management of the threatened endemic palms of the Mascarene Islands. Oryx 36: 56–65.
- MCCLATCHEY, W. 1998. A new species of *Metroxylon* from western Samoa. Novon 8: 252–258.
- MIYAMOTO, J., M. NAKAYAMA, N.M. WATANABE AND E. SUZUKI. 2006. Genetic and morphological variation in *Licuala paludosa* Griff. and the

related taxa in the Tasek Merimbun Heritage Park, Borneo. Tropics 15: 237–243.

- MORAES R., M. 1996. *Allagoptera* (Palmae). Flora Neotropica Monograph 73: 1–34.
- MOYA LÓPEZ, C.E. AND P.T. MAYOTTE. 1996. "Paradiso Principium" – A palm paradise in Cuba. Principes 40: 152–155.
- MOYA LÓPEZ, C.E., M.T. ROSABAL AND Y.H. RIVERO. 2019. Novedades en la distribución conocida de *Copernicia* × *escarzana* (Arecaceae); implicaciones para la conservación. Monteverdia 12: 1–8.
- NAUMAN, C. 1989. Intergeneric hybridization between *Coccothrinax* Sargent and *Thrinax* Swartz (Palmae: Coryphoideae). American Journal of Botany 75: 195–196.
- NAUMAN, C. 1990. Intergeneric hybridization between *Coccothrinax* and *Thrinax* (Palmae: Coryphoideae). Principes 34: 191–198.
- NOBLICK, L.R. 2014. *Butia*: what we think we know about the genus. The Palm Journal 208: 5–23.
- NOBLICK, L.R. 2017. A revision of the genus *Syagrus* (Arecaceae). Phytotaxa 294: 1N262.
- PEREIRA, A.G., ET AL. 2022. Patterns of genetic diversity and structure of a threatened palm species (*Euterpe edulis* Arecaceae) from the Brazilian Atlantic Forest. Heredity 129:161–168.
- PÉREZ-ESCOBAR, O.A., ET AL. 2021. Molecular clocks and archeogenomics of a Late Period Egyptian date palm leaf reveal introgression from wild relatives and add timestamps on the domestication. Molecular Biology and Evolution 38: 4475–4492.
- PINHEIRO, C.U.B. 1997. Systematic and agroecological studies in the Attaleinae (Palmae). Ph.D. Thesis, City University of New York.
- PINTAUD, J.-C. 2006. The impact of forest disturbance on the palms of New Caledonia. Palms 50: 123–135.
- POWO 2022. Plants of the World Online. Facilitated by the Royal Botanic Gardens, Kew. Published on the Internet; http://www.plantsoftheworldonline.org/ Retrieved 25 September 2022.
- RAMÍREZ-RODRÍGUEZ, R., E. TOVAR-SÁNCHEZ, J. RAMÍREZ, K. FLORES AND V. RODRÍGUEZ. 2011. Introgressive hybridization between *Brahea dulcis* and *Brahea nitida* (Arecaceae) in

Mexico: evidence from morphological and PCR-RAPD patterns. Botany-Botanique 89: 545–557.

- RAMÍREZ-RODRÍGUEZ, R., P. MUSSALI-GALANTE, H. QUERO AND E. TOVAR-SÁNCHEZ. 2012. Management and its relation to hybridization, clonality and genetic structure of the Mexican palm *Brahea dulcis*. Forest Ecology and Management 285: 92–100.
- RIESEBERG, L.H. AND J.F. WENDEL. 1993. Introgression and its consequences in plants, pp. 70–109, in HARRISON, R.G. (ed). Hybrid Zones and the Evolutionary Process. Oxford University Press, UK.
- RODD, A.N. 1998. Revision of *Livistona* (Arecaceae) in Australia. Telopea 8: 49–153.
- SIMPSON, B.J. 1988. A field guide to Texas trees. Texas Monthly Press, Austin, Texas.
- SOARES, K.P., S.J. LONGHI, L. WITECK NETO AND L.C. ASIS. 2014. Palmeiras (Arecaceae) no Rio Grande do Sul, Brasil. Rodriguésia 65: 113–139.
- SOLTIS, P.S. AND D.E. SOLTIS. 2009. The role of hybridization in plant speciation. Annual Review of Plant Biology 60: 561–588.
- SOLTIS D.E., ET AL. 2007. Autopolyploidy in angiosperms: Have we grossly underestimated the number of species? Taxon 56: 13–30.
- SUÁREZ OROPESA, D. 2015. *Coccothrinax × angelae* (Arecaceae), nuevo híbrido natural del género para Cuba. Revista del Jardín Botánico Nacional, Universidad de la Habana 36: 9–14.

TUCKER, R. 1975. Letters. Principes 19: 115.

- VERDECIA PÉREZ, R. 2016. *Copernicia* × *dahlgreniana*, a new natural hybrid in the savannas of Camagüey, Cuba. Palms 60: 85–92.
- WESSELS BOER, J. 1971. *Bactris × moorei*, a hybrid in palms. Acta Botanica Neerlandica 20: 167–172.
- WHITMORE, T.C. 1998. Palms of Malaya. Second edition. White Lotus Press, Bangkok, Thailand.
- WOOD, T.E., N. TAKEBAYASHI, M.S. BARKER, I. MAYROSE, P.B. GREENSPOON AND L.H. RIESEBERG. 2009. The frequency of polyploid speciation in vascular plants. PNAS 106: 13875–13879.

Appendix 1. Taxa for which no natural h	
CALAMOIDEAE	BORASSEAE, HYPHAENINAE
EUGEISSONEAE	Bismarckia, 1 sp.
<i>Eugeissona</i> , 6 spp	Satranala, 1 sp.
LEPIDOCARYEAE, ANCISTROPHYLLINAE	Hyphaene, 8 spp.
Oncocalamus, 4 spp.	Medemia, 1 sp.
Eremospatha, 11 spp.	BORASSEAE, LATANIINAE
Laccosperma, 7 spp.	Latania, 3 spp.
LEPIDOCARYEAE, RAPHIINAE	Lodoicea, 1 sp.
Raphia, 22 spp.	Borassodendron, 2 spp.
LEPIDOCARYEAE, MAURITIINAE	Borassus, 5 spp.
Mauritia, 2 spp.	CEROXYLOIDEAE
Mauritiella, 4 spp.	CYCLOSPATHEAE
Lepidocaryum, 1 sp.	Pseudophoenix, 4 spp.
CALAMEAE, KORTHALSIINAE	ARECOIDEAE
Korthalsia, 28 spp.	PODOCOCCEAE
CALAMEAE, SALACCINAE	Podococcus, 2 spp.
Eleiodoxa, 1 sp.	ORANIEAE
Salacca, 23 spp.	Orania, 30 spp.
CALAMEAE, PIGAFETTINAE	SCLEROSPERMEAE
<i>Pigafetta</i> , 2 spp.	Sclerosperma, 3 spp.
CALAMEAE, PLECTOCOMIINAE	ROYSTONEEAE
Plectocomia, 14 spp.	Roystonea, 10 spp.
Myrialepis, 1 sp.	REINHARDTIEAE
Plectocomiopsis, 6 spp.	
NYPOIDEAE	<i>Reinhardtia</i> , 6 spp.
<i>Nypa</i> , 1 sp.	COCOSEAE, ELAEIDINAE
CORYPHOIDEAE	Barcella, 1 sp.
TRACHYCARPEAE, RHAPIDINAE	Elaeis, 2 spp.
Chamaerops, 1 sp.	MANICARIEAE
Guihaia, 4 spp.	<i>Manicaria</i> , 2 spp.
Trachycarpus, 10 spp.	LEOPOLDINIEAE
Rhapidophyllum, 1 sp.	Leopoldinia, 3 spp.
Maxburretia, 3 spp.	PELAGODOXEAE
Rhapis, 11 spp.	Pelagodoxa, 2 spp.
CHUNIOPHOENICEAE	Sommieria, 1 sp.
Chuniophoenix, 3 spp.	ARECEAE, ARCHONTOPHOENICINA
Kerriodoxa, 1 sp.	Actinorhytis, 1 sp.
Nannorrhops, 1 sp.	Archontophoenix, 6 spp.
Tahina, 1 sp.	Chambeyronia, 9 spp.
CORYPHEAE	
Corypha, 5 spp.	

Appendix 1. continued.	
ARECEAE, ARECINAE	Adonidia, 2 spp.
Areca, 47 spp.	Balaka, 10 spp.
Nenga, 5 spp.	Veitchia, 11 spp.
Pinanga, 143 spp.	Carpentaria, 1 sp.
ARECEAE, CARPOXYLINAE	Wodyetia, 1 sp.
Carpoxylon, 1 sp.	Drymophloeus, 3 spp.
Satakentia, 1 sp.	Normanbya, 1 sp.
Neoveitchia, 2 spp.	Brassiophoenix, 2 spp.
ARECEAE, CLINOSPERMATINAE	Ptychococcus, 2 spp.
Cyphokentia, 2 spp.	Jailoloa, 1 sp.
Clinosperma, 4 spp.	Manjekia, 1 sp.
ARECEAE, DYPSIDINAE	Wallaceodoxa, 1 sp.
Chrysalidocarpus, 54 spp.	ARECEAE, RHOPALOSTYLIDINAE
Dypsis, 106 spp.	Rhopalostylis, 2 spp.
Lemurophoenix, 2 spp.	Hedyscepe, 1 sp.
Marojejya, 2 spp.	ARECEAE, VERSCHAFFELTIINAE
Masoala, 2 spp.	Nephrosperma, 1 sp.
Vonitra, 10 spp.	Phoenicophorium, 1 sp.
ARECEAE, PTYCHOSPERMATINAE	Roscheria, 1 sp.
Ptychosperma, 29 spp.	Verschaffeltia, 1 sp.
Ponapea, 4 spp.	

MUNTED STATES Statement of Ownership, Management, and Circulation POSTAL SERVICE. (All Periodicals Publications Except Requester Publications)	Statement of Ownership, Management, and Circulation eriodicals Publications Except Requester Publications)	nt, and Circulation ster Publications)	Paims		Septem	September 2022	POSTAL STATES Statement of Ownership, Management, and Circulation POSTAL SERVICE (All Periodicals Publications Except Requester Publications)	p, management Except Reques	er Publications)
1. Publication Title 2. Pairris	2. Publication Number 3	3. Filing Date 09/28/2022	15. Extent and Nature of Circutation	e of Circutation	Average No. Copies Each Issue During Preceding 12 Months	No. Copies of Single Issue Published Nearest to Filing Date	16 Electronic Copy Circulation	Average No. Copies 1 Each Issue During 1 Preceding 12 Months 1	es No. Copies of Single Issue Published ths Nearest to Filing Date
4. Issue Frequency 5. Quanterly 4.	5. Number of Issues Published Annually 6	6. Annual Subscription Price \$55,00	a. Total Number o	a. Total Number of Copies (Net press run)	909	582	a. Pad Electronic Copies	•	0
<ol> <li>Complete Mailing Address of Known Office of Publication (Het printer) (Street. cdr. county, state, and 2(P+47) International Pain Society.</li> </ol>		Contact Person Scott Zona	ω	Matted Outside County Paid Subscriptions Stated on PS Form 3541 (Include paid distribution above normal rate, achverteet's proof copies, and exchange copies)	d 380	378	<ol> <li>Total Paid Print Copies (Line 15c) + Paid Electronic Copies (Line 16a)</li> </ol>	662	999
1401 Lavora St. #751 Auto, 17: 77700 Addise Auto, 17: 7700 Addise Ad		Telephone (Include area code) (512) 851-1411	b. Paid Circulation (2) (By Mail	Maited In-County Paid Subscriptions Stated on PS Form 3541 (Include paid distribution above nominal rate, advertater's proof copied, and exchange copied.	0	0	<ol> <li>Total Print Distribution (Line 151) + Paid Electronic Copies (Line 15a)</li> </ol>	\$63	569
International Pain Scorety MOI Lavora 8751	Name and Annual		and Outside the Mail [3]	Paid Distribution Outside the Mails Including Sales Through Dealers and Camlers, Steet Vendors, Counter Sales, and Other Paid Distribution Outside USPS®	20	8	d. Percent Paid (Both Pint & Electronic Copies) (16b divided by 16c $\times$ 100)	996	99.5
<ul> <li>Entrements and Complete Mailing Addresses of Publisher, Editor, and Managing Editor (Do not leave bland) Publisher (have and complete mailing address)</li> </ul>	ing Editor (Do not leave blank)		(4)	Paid Datrbution by Other Classes of Mail Through the USPS (e.g., First Class Mail <sup>1</sup> )	150	136	I certify that 50% of all my distributed copies (electronic and print) are paid above a nominal price.	nominal price.	
Jobn Chandleid, Hetbanum Royal Bobino Carden Kew, Redimnoud Saurey, TW9 JAB, UK			c. Total Part Dem	<ul> <li>Total Paid Destruction [Sum of 156 (1), (2), (3), and (4)]</li> </ul>	580	200	<ol> <li>Proceedings of constraints or constraints</li> <li>The publication is a general publication, publication of this statement is required. Will be printed</li> </ol>		Publication not required.
Editor (Name and complete mailing address) John Dronofeid Hernsteine			d. Free or (1) Nominal	Free or Nominal Rate Outside-County Copies included on PS Form 3541	-	-	in the December 2021 Issue of this publication.		
Royal Botanio Gardens Kew, Retimond, Surrey, TWB 3AE, UK			Rate Distribution (2)	Free or Nominal Rate in-County Copies Included on PS Form 3541	0	0	18. Signature and Title of Editor, Publisher, Business Manager, or Owner		Date
Managing Editor (Name and complete mailing address) Scott Zona 1807 Seminola Dr			and Outside (3) Pre Mail	Free or Nominal Rate Copies Mailed at Other Classes Through the USPS (e.g., First Class Mail)	0	•	X		09/29/2022
Historouch NC 27278-7967 10. Owner Do not leave blank. If the publication is owned by a corporation, give the name and address of the corporation immediately followed by the	the name and address of the corporation in	vmediately followed by the	(4)	Free or Nominal Rate Distribution Outside the Mail (Carriers or other means)	8	8	I certify that all information familyhed on this form is true and complete. I understand that anyone who fumilyhes takes or misiakading information on this form or who certify that all information measurated on the form may be subtlet to circlent and tencinonated and trutched on this form	who furnishes false or misler including fines and incrition	Sing information on this form
cares and additionate of all accordance some to charge) prevents from other host barrels and the charge of the barrels and the charge of the problem of the charge of the barrels and the charge of the barrels and the barrels are used in those of the additionation of the charge of the barrels and the barrels are used in those of the barrels and the problem of the problem of the barrels of the barrels and the barrels of the problem of the problem of the problem of the barrels of the barrels of the barrels of the barrels of the problem of the problem of the barrels of the barrels of the problem of the barrels of the problem of the barrels of the b	ore of the total amount of stock. If not owned, or other unincooppreated firm, give its name and alton, give its name and address.] Complete Mailing Address	by a coporation, give the d address as well as those of	e. Total Free or N	e. Total Pree or Nominal Rate Distribution (Sum of 154 (1), (2), (2) and (4))	e	n	(instants)		
International Palm Society	International Palm Society, 1401 Lavaca St., Austin, TX 76701	., Austin, TX 76701-1634	f. Total Distributio	Total Distribution (Sum of 15c and 15e)	583	569			
			g. Copies not Distr	Copies not Distributed (See Instructions to Publishers #4 (page #3))	23	ţ			
			h. Total (Sum of 15/ and g)	Strand g)	606	582			
			<ol> <li>Percent Paid (15c divided by 15f times 100)</li> </ol>	15f times 100)	9.66	99.5			
<ol> <li>Known Bondholden, Murguagees, and Offwer Security Holders Owning or Holding 1 Percent or More of Total Amount of Bunds, Hordiages, Other Securities if none check box.</li> </ol>	iding 1 Percent or More of Total Amount of	Bonds, Montgages, or	* If you are claiming el	" If you are claiming electronic copies, go to fire 16 on page 3. If you are not claiming electronic copies, why to fire 17 on page 3.	skip to line 17 on page 3.				
	Complete Mailing Address								
12. Tax Shara For completen by nonprofe councednoirs annound to mail all nonprofer ranks (Chried ove) The suppose, function, no propertit statistics of this organization and the exempt status for believal income kui purposes. The suppose theory Preceding 12 Montils of the Composed During Preceding 12 Montils. Probleme mult submit exploration of charge with this statement of the Composed During Preceding 12 Montils.	conprofit ratios) (Check one) rold statuta for federal income tax purposes: nation of change with this statement)								
PS Form 3525, July 2014 (Page 1 of 4 (see instructions page 4); PSN: 7530-01-000-9031 PREVACY NOTICE: See our privacy policy on surve upta com	TOT TOT DEVACY MOTICE: San 201							The second se	BRAIN OF STATES

# Exploring for New Localities of *Tahina spectabilis* in Northeastern Madagascar

DAVID RABEHEVITRA<sup>1</sup>

*Tahina spectabilis,* the most massive fan palm in Madagascar, was discovered in 2006 in a very remote locality northeast of Mahajanga. Recent field work has documented thirteen additional populations of this globally imperiled palm.

The plant was described as a new Coryphoid genus in 2008 (Dransfield et al. 2008a) with a statement on the restricted distribution of the species. The IUCN assessment categorized *Tahina spectabilis* as Critically Endangered (CR) (Rakotoarinivo et al. 2012), which triggered the hunt for new populations using the predictive distribution map published in the two first papers on the species (Dransfield et al. 2008a)

A second population of *Tahina spectabilis*, was discovered by researchers from Royal Botanic Gardens Kew (RBG Kew), Parc Botanique et Zoologique de Tsimbazaza (PBZT) and the University of the Sunshine Coast (USC) in 2017, after the first one where the species was described and published in 2008 (Dransfield et al. 2008b). The discovery was sparked off by the recognition of the species from a photo sent by a local guide to the Royal Botanic Gardens Kew office in Antananarivo, the Kew Madagascar Conservation Centre (KMCC). The team mounted an expedition, first visiting the original type population and then visiting the

<sup>1</sup>Kew Madagascar Conservation Centre Royal Botanic Gardens Kew Lot II J 131 B Ambodivoanjo – Ivandry 101 Antananarivo – Madagascar D.Rabehevitra@kew.org new site where the photo was taken. The plant photographed was checked with local guide communities, recognized as *Tahina spectabilis* (Fig. 1) and published (Gardiner et al. 2017). This new discovery, occurring outside the area predicted in the original paper (Dransfield et al. 2008a) suggested that the palm might be more widespread. Researchers from Royal Botanic Gardens visited a new area to explore for potential populations.

#### How it started

The trip to Amparihibe where the second population of *Tahina spectabilis* (Gardiner et al. 2017) was discovered, was an opportunity for RBG Kew team to launch a public awareness campaign with the local community on just how important the discovery of *T. spectabilis* is, and why the species is a natural legacy they have the obligation to take care of, for the present and for future generations. This campaign was followed by the distribution of a *T. spectabilis* poster and Tshirts several months later, with the emblem "let us protect *Tahina spectabilis*" in their own Malagasy dialect, emblazoned under a photo of *T. spectabilis* (Fig. 2).

#### Talk chains

As community members meet people wearing the *Tahina spectabilis* T-shirt and see the photo of the plant, conversations start with what the



1. Tahina spectabilis individual discovered in 2017.

plant is, why it is special and ending with "I know a place where there are some too." The site-based KMCC employee – Theophile Rajaonilaza – collected these "site names"

along with the names of individuals linked with the information. He visited them on a later date to talk about the species, check with them if possible or plan a later visit and



2. *Tahina specabilis* T-Shirt created to heighten awareness and initiate discussions about the palm.

distribute the T-shirts to expand the publicity campaign. These same people were the ones to guide the team to the sites when they were available.

#### Thirteen new localities

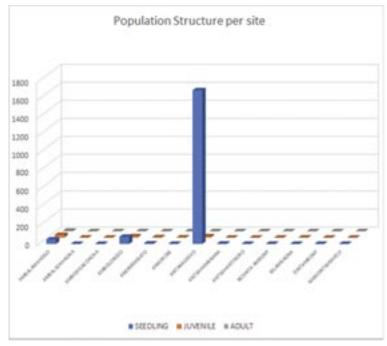
These talk chains amongst community members enabled the recording of thirteen new localities (Fig. 3) – twelve on the eastern belonging to the commune of side, Maromandia, and one, within the commune of Ankaramy on the western side of National Road No. 6, between Befotaka village in the south and Ankaramihely in the north. We took the road as a geographical reference as it is the most obvious landmark for the area. However, the distribution map shows a possible positive correlation with the Ano Malaza River, except for the Antanagnivo population on the western side of the road. These newly discovered localities cover a wide latitude range which will bring an important change in the species' Area of Occupancy and Extent of Occurrence.

#### Few adults and numerous seedlings

The population structure was categorized into seedlings (S), juveniles (J) and adults (A), with two subcategories each, based on their relative size compared to adult human height for S and J, and the relative height of the trunk for A. This population structure is marked by a very high number of seedlings, ca. 1970, and a low number of juveniles (74) and adults (18) for the thirteen new localities (Fig. 4). The most important concentration of seedlings was recorded in Antanagnivo on the western side

3. Distribution map of the new *Tahina spectabilis* populations. On the left is the map of Madagascar with insets showing the region in the northwest where the palm is found. On the right is enlarged map showing the locations of the 13 newly discovered populations.





4. A graph showing the population structure of *Tahina spectabilis* in the 13 newly documented populations. Size classes are indicated. The number of seedlings in Antanagnivo is striking. The cause of this disparity is not currently known.

of the National Road No. 6, which consists of over 1800 plants and more than 100 individuals per square meter (Fig. 5).

#### Seeds and Nursery

The RBG Kew team had the luck to observe a flowering *Tahina spectabilis* in the beginning

of 2018 in Manerinerina, which produced several hundred seeds (Fig. 6) that the team collected for nursery purposes in collaboration with the PBZT. Unfortunately, too many of the seeds collected dried out before they reached Antananarivo (Fig. 7), and germination was disappointingly low. Parc

5. The high density of seedlings in the population at Antanagnivo.





6. Flowering *Tahina spectabilis* in Manerinerina. The inset at the lower right shows a flower and a developing fruit.

Botanique et Zoologique de Tsimbazaza is now taking care of about 200 nursery seedlings, and about 50 seedlings are held in Theophile's village (Fig. 8).

#### Further comments

The high concentration of seedlings observed in Antanagnivo suggests a very prolific fruiting



7 (top). Germinating *Tahina spectabilis* seeds. 8 (bottom). *Tahina spectabilis* seedlings thriving in the nursery in Theophile's village.

event but a critical lack of natural dispersers, highlighting the importance of ecosystem interdependence for species survival. A community member living close to several *Tahina spectabilis* individuals in Ambolobozo mentioned seeing fruit bats visiting the fruiting palm and discarding the fleshless seeds, suggesting that bats may be the natural disperser of the palm.

It is also worth mentioning that more than ten site names are known to host *Tahina spectabilis* individuals, but some people are reluctant to allow the visit of strangers for different reasons that are not yet understood by the RBG Kew Team.

Finally, that Tahina spectabilis is a distinct palm different from the co-occurring fan palms Borassus spp. and Bismarckia nobilis is well understood in a few communities, and the local name, *bilambagna*, comes from the fact that their ancestors used the large leaf of the palm as a mat (*lambagna*) for eating during ceremonies. The KMCC Team is currently running a project which supports the community-led conservation of Tahina spectabilis near Maromandia through the promotion of yam cultivation, tree planting, well-drilling and classroom building within villages surrounding the best remaining habitat of the species, with an ultimate goal of implementing a community-managed Protected Area.

#### Conclusions

The second population of the Tahina spectabilis revealed in Amparihibe in 2016 has led researchers from Royal Botanic Gardens Kew to discover thirteen other localities, thanks to sustained collaboration with local communities and a continuous in-the field presence. These discoveries were the result of effective communication and public awareness undertaken – T-shirts and individual communication - to reveal people, localities and other key information. These newly discovered localities confirmed the possibility of a wider distribution of the species in this region and a wider AOO/EOO. Undertaking this research work enables the community to deliver their own knowledge of Tahina spectabilis and build a productive trust. These results open the door to further research in genetics and other fields such as history. Finally, researchers at RBG Kew hope to support the creation of a protected habitat for the species and reviewing the IUCN status of *Tahina spectabilis* soon.

#### Acknowledgments

We would like to thank the Fondation Franklinia and the International Palm Society's PalmTalk fund raising campaign for the financial support to these expeditions, which enabled the discovery of those new populations and in a better understanding of the critically endangered Tahina spectabilis. We would also like to acknowledge the local authorities of Amparihibe, Anketrakabe and Maromandia for allowing our work to be led within the best and the safest possible conditions. Finally, we would address our gratitude to Theophile Rajaonilaza and all Kew Madagascar Conservation Centre staff for the invaluable support regarding the Tahina spectabilis team.

#### LITERATURE CITED

- DRANSFIELD, J. ET AL. 2008a. A new Coryphoid palm genus from Madagascar. Botanical Journal of the Linnean Society 156: 79–91.
- DRANSFIELD, J., B. LEROY, X. METZ AND M. RAKOTOARINIVO. 2008b. *Tahina* A New Palm Genus from Madagascar. Palms 52: 31–39.
- RAKOTOARINIVO, M. AND J. DRANSFIELD. 2012. *Tahina spectabilis*. The IUCN Red List of Threatened Species 2012: e.T195893A2430024.
- GARDINER, L.M., D. RABEHEVITRA, R. LETSARA AND A. SHAPCOTT. 2017. *Tahina spectabilis*: An exciting new discovery in Madagascar ten years on. Palms 61: 69–82.

## **Index to Volume 66**

A new, large-flowered Licuala from New Guinea 69 A review of naturally occurring hybrids in palms (Arecaceae) 177 Acanthophoenix 186 Acoelorraphe 182 Acoelorraphe wrightii 116, 117 Acrocomía 185 Acrocomia aculeata 116 Actinorhytis 192 Addressing the monoculture system: Challenges in oil palm-based agroforestry 147 Adonidia 193 Aiphanes 24, 25, 185 Aiphanes bicornis 24, 26 Aiphanes chiribogensis 24 Aiphanes eggersii 24, 26 Aiphanes erinacea 24 Aiphanes gelatinosa 24 Aiphanes grandis 24, 25 Aiphanes hirsuta subsp. forsteriorum 24 Aiphanes macroloba 24, 26 Aiphanes multiplex 24 Aiphanes tricuspidata 24 Aiphanes ulei 174, 176 Allagoptera 183 Allagoptera campestris 183 Allagoptera caudescens 188 Allagoptera leucocalyx 185 Ammandra 183 Ancrenaz, M., as co-author 147 Aphandra 183 Aphandra natalia 174, 176 Archontophoenix 192 Areca 193 Areca catechu 160 Arenga 183 Arenga caudata 188 Arenga pinnata 183 Arenga undulatifolia 23 Arenga westerhoutii 183 Asterogyne 186 Asterogyne martiana 24, 27, 116 Asterogyne spicata 38 Astrocaryum 185 Astrocaryum aculeatum 119 Astrocaryum chambira 162, 174, 175 Astrocaryum jauari 176 Astrocaryum mexicanum 116 Astrocaryum murumuru 57 Astrocaryum rodriguesii 119 Astrocaryum standleyanum 24, 108 Astrocaryum urostachys 174, 176 Attalea 176, 183, 185, 187, 188 Attalea barreirensis 185 Attalea butyracea 174, 176, 185 Attalea camopiensis 185 Attalea cohuñe 116 Attalea colenda 24 Attalea cuatrecasana 24 Attalea dahlgreniana 185 Attalea degranvillei 185 Attalea guianensis 185 Attalea maripa 174, 176, 185, 187 Attalea × minarum 185 Attalea nucifera 185 Attalea phalerata 174, 176 Attalea piassabossu 184, 185 Attalea sagotii 185 Attalea salvadorensis 185

Attalea speciosa 187 Attalea × teixeriana 184 Attalea × voeksii 185 Auger-Micou, M, D. Roguet, A.A. Camara, P. Griffith, L. Noblick & F. Stauffer: Palm diversity and ethnobotany in Senegal, West Africa 127 Bacon, C.D., as co-author 57 Bacon, C.D., T. Clase, E. Fernández, X. Gratacos, A. Henderson & O. Montero: Dominican discoveries 81 Bactris 28, 167, 185, 187 Bactris acanthocarpa 174, 185, 187 Bactris barronis 185 Bactris brongniartii 185 Bactris caudata 185 Bactris coloradonis 24, 28, 185 Bactris concinna 176 Bactris corossilla 174, 176, 185 Bactris gasipaes 185 Bactris gasipaes var. chichagui 24, 28 Bactris gasipaes var. gasipaes 24, 28, 164, 165, 171, 174, 176 Bactris gracilior 185 Bactris hirta var. hirta 174 Bactris hondurensis 24, 29 Bactris killipii 167 Bactris major 185, 187 Bactris major var. major 116 Bactris maraja 24, 29, 176, 185 Bactris maraja var. juruensis 174 Bactris maraja var. maraja 174 Bactris mexicana 185 Bactris mexicana var. trichophylla 116 Bactris × moorei 185 Bactris obovata 38 Bactris oligoclada 185 Bactris pilosa 24 Bactris plumeriana 86, 87 Bactris riparia 176 Bactris schultesii 167, 174 Bactris setulosa 24, 28, 185 Bactris simplicifrons 167, 174 Baker, W.J., as co-author 69 Balaka 193 Balick, M.J. & A. Henderson: Field Guide to the Palms of Belize, reviewed 73 Balslev, H., as co-author 161 Barcella 192 Barfod, A.S. & W.J. Baker: A new, large-flowered Licuala from New Guinea 69 Basselinia 186 Basselinia deplanchei 186 Basselinia gracilis 186 Basselinia pancheri 186 Basselinia vestita 186 Beccariophoenix 183, 188 Bentinckia 186 Bismarckia 192 Bismarckia nobilis 23, 201 Borassodendron 192 Borassus 139, 192 Borassus aethiopum 131, 144 Borassus akeassii 128-130-145 Brahea 182 Brahea calcarea 182 Brahea dulcis 117, 182 Brassiophoenix 193 Burretiokentia 186 Butia 183, 187, 188 Butia capitata 108, 185 Butia eriospatha 185 Butia lallemantii 185 Butia odorata 108, 183

Index to vol. 66

Butia paraguayensis 185 Butia yatay 185 × Butyagrus alegretensis 185 × Butyagrus nabonnandii 183, 185 × Butyagrus paranaensis 185 Calamus 69, 180 Calamus bacularis 180 Calamus deerratus 129, 130, 139, 144 Calamus insignis 180 Calamus javensis 180 Calamus leloi 180 Calamus longiusculus 180 Calamus myriacanthus 180 Calamus tenompokensis 180 Calyptrocalyx 186 Calyptrogyne 38, 186 Calyptrogyne brachystachys 186 Calyptrogyne fortunensis 186 Calyptrogyne ghiesbreghtiana 186 Calyptrogyne ghiesbreghtiana subsp. spicigera 116 Calyptrogyne panamensis 186 Calyptrogyne trichostachys 186 Calyptronoma 186 Calyptronoma plumeriana 87 Calyptronoma rivalis 83, 87 Camara, A.A., as co-author 127 Cámara-Leret, R., as co-author 161 Carpentaria 193 Carpoxylon 193 Carpoxylon macrospermum 23 Carrillo-Flores, R., as co-author 161 Caryota 183 Caryota cumingii 183 Caryota maxima 183 Caryota mitis 183 Caryota monostachya 183 Carvota no 23 Caryota rumphiana 183 Caryota sympetala 183 Ceroxylon 10, 13, 23, 183 Ceroxylon alpinum 183 Ceroxylon amazonicum 23 Ceroxylon echinulatum 183 Ceroxylon parvifrons 8, 9, 10, 12, 19, 183 Ceroxylon peruvianum 183 Ceroxylon quindiuense 183 Ceroxylon vogelianum 183 Chamaedorea 41, 43, 73, 121, 183 Chamaedorea arenbergiana 116 Chamaedorea concolor 119 Chamaedorea deneversiana 24 Chamaedorea elatior 41–44, 49 Chamaedorea ernesti-augusti 33-36, 38, 39, 116 Chamaedorea frondosa 42 Chamaedorea geonomiformis 42, 116 Chamaedorea glaucifolia 41, 42 Chamaedorea graminifolia 119, 121 Chamaedorea klotzschiana 42 Chamaedorea linearis 24 Chamaedorea neurochlamys 116 Chamaedorea oblongata 42, 116 Chamaedorea pauciflora 165, 174, 176 Chamaedorea pinnatifrons 24, 174 Chamaedorea pochutlensis 42 Chamaedorea schiedeana 36 Chamaedorea schippii 109, 119, 120, 121, 122 Chamaedorea tacanensis 41, 42, 44-50 Chamaedorea tacanensis: A climbing species from Mexico and Guatemala 41 Chamaedorea tenella 42 Chamaerops 78, 192 Chambeyronia 192 Chelyocarpus 23, 181

Chrysalidocarpus 108, 193 Chuniophoenix 192 Clase, T., as co-author 81 Clinosperma 193 Clinostigma 186 Clinostigma samoense 186 Clinostigma warburgii 186 Coccothrinax 77, 81, 82, 84, 86, 87, 181 Coccothrinax alta 82 Coccothrinax × angelae 180, 181 Coccothrinax argentata 82, 177 Coccothrinax argentea 83, 85, 86, 87 Coccothrinax barbadensis 82 Coccothrinax borhidiana 82 Coccothrinax boschiana 82, 88 Coccothrinax concolor 82 Coccothrinax crinita 181 Coccothrinax crinita subsp. brevicrinis 180 Coccothrinax ekmanii 82 Coccothrinax fragrans 86 Coccothrinax gracilis 82, 86, 87 Coccothrinax gundlachii 82 Coccothrinax inaguensis 82 Coccothrinax jimenezii 82 Coccothrinax macroglossa 179, 181 Coccothrinax miraguama 181 Coccothrinax miraguama subsp. roseocarpa 180 Coccothrinax montana 82 Coccothrinax orientalis 181 Coccothrinax pauciramosa 82, 181 Coccothrinax pseudorigida 179, 181 Coccothrinax scoparia 82 Coccothrinax spirituana 82 Coccothrinax spissa 55, 82 Cocos 183 Cocos nucifera 129, 131, 164, 174 Colpothrinax 182 Copernicia 79, 182, 187, 188 Copernicia berteroana 86 Copernicia × burretiana 182 Copernicia cowellii 182 Copernicia curbeloi 182 Copernicia x dahlgreniana 182 Copernicia × escarzana 182 Copernicia hospita 187 Copernicia hospita 182 Copernicia longiglossa 182 Copernicia macroglossa 182 Copernicia molinetii 182 Copernicia × occidentalis 182 Copernicia oxycalyx 182 Copernicia × shaferi 182 Copernicia × sueroana 182 Copernicia × textilis 182 *Copernicia* × *vespertilionum* 182 Copete, J.C., as co-author 161 Corypha 192 Couell, M., R. Montúfar & T.L.P. Couvreur: Selva Chi: A unique integration of education and palm conservation 22 Couvreur, T.L.P., as co-author 22 Couvreur, T.L.P, R. Carrillo-Flores, R. Cámara-Leret, J.C. Copete & H. Balslev: Kapawi: A mega-diverse palm community in the eastern Amazon of Ecuador 161 Couvreur, T.L.P., R. Montúfar, N. Zapata, C. Persson & A.J. Pérez: Palms of the remote Cerro Plateado Biological Reserve, southeastern Ecuador 5 Couvreur, T.L.P. & T.C.H. Sunderland: Flore du Cameroun 44: Arecaceae - Palmae, reviewed 126 Cryosophila 181

Cryosophila stauracantha 116

Cryosophila warscewiczii 38

Cyphokentia 193 Cyphophoenix 186 Cyphosperma 186 Cyrtostachys 186 Deckenia 186 Desmoncus 176, 185, 187 Desmoncus chinantlensis 117 Desmoncus cirrhiferus 24, 31 Desmoncus giganteus 174 Desmoncus horridus 185 Desmoncus leptoclonos 185 Desmoncus mitis 185 Desmoncus mitis var. mitis 174 Desmoncus parvulus 185 Desmoncus polyacanthos 174, 185, 187 Desmoncus pumilus 185 Dictyocaryum 7, 13, 166, 183 Dictyocaryum lamarckianum 12, 14, 15, 19, 24 Dictyosperma 186 Djama, M., as co-author 147 Dominican discoveries 81 Dowe, J.L. & B.O. Schlumpberger: The uncertain history of an early illustration of Chamaedorea ernesti-augusti 33 Dowe, J.L. & E. Schweizer: *Livistona victoriae* and the North Austalian Exploring Expedition 1855–1856 90 Dowe, J.L. & H. Yoxall: William August Schipp (1891–1967): Commemorated in Schippia concolor and Chamaedorea schippii 109 Dransfield, J.: Palm Literature: Apprendre à Reconnaître les Palmiers Endémiques de Nouvelle-Calédonie. Version Numérique 66 Dransfield, J.: Palm Literature: Flore du Cameroun 44: Arecaceae – Palmae 126 Dransfieldia 186 Drymophloeus 193 Ducouret, E. & C. Henry: Apprendre à Reconnaître les Palmiers Endémiques de Nouvelle-Calédonie. Version Numérique, reviewed 66 Dypsis 108, 193 Elaeis 192 Elaeis guineensis 107, 129, 131, 133, 134, 136-139, 141–143, 145, 148, 167, 176 Elaeis oleifera 167, 169, 174, 176 Eleiodoxa 192 Emilio, T., as co-author 57 Eremospatha 143, 145, 192 Eremospatha dransfieldii 129 Eugeissona 192 Euterpe 7, 185 Euterpe edulis 186 Euterpe oleracea 24, 31, 57 Euterpe precatoria 31, 57, 174, 176 Euterpe precatoria var. longevaginata 12, 13, 19, 24 Exploring for new localities of Tahina spectabilis in northeastern Madagascar 195 Fernández, E., as co-author 81 Fountain, C.: The Poem Forest: Poet W.S. Merwin and the Palm Forest he Grew from Scratch, reviewed 72 Gaussia 183 Geonoma 11, 24, 38, 176, 186 Geonoma brongniartii 174, 186 Geonoma calyptrogynoidea 24 Geonoma camana 174, 186 Geonoma chococola subsp. awaensis 24, 26 Geonoma cuneata 24, 26 Geonoma cuneata subsp. cuneata 24, 26 Geonoma cuneata subsp. irena 3, 24, 26, 27 Geonoma cuneata subsp. linearis 24 Geonoma cuneata subsp. procumbens 24 Geonoma cuneata subsp. soderoi 24, 26

Geonoma deversa 24, 176, 186, 187 Geonoma deversa subsp. deversa 116 Geonoma elegans 186 Geonoma interrupta 24, 186 Geonoma lanata 24, 27 Geonoma leptospadix 186 Geonoma longepedunculata 174, 176 Geonoma longivaginata 186 Geonoma macrostachys 164, 165, 174, 186 Geonoma macrostachys var. acaulis 176 Geonoma maxima 167 Geonoma maxima subsp. camptoneura 167, 168 Geonoma maxima subsp. multiramosa 167, 175 Geonoma multisecta 175 Geonoma occidentalis 186 Geonoma orbignyana 186 Geonoma paradoxa 24, 27 Geonoma pauciflora 186 Geonoma pinnatifrons 186 Geonoma pinnatifrons subsp. binervia 116 Geonoma poeppigiana 159, 175, 186 Geonoma pohliana 186 Geonoma schottiana 186 Geonoma stricta 164, 176 Geonoma stricta subsp. arundinacea 164, 175 Geonoma tenuissima 3, 24, 25, 26 Geonoma trigona 186 Geonoma undata 3, 8, 11, 12, 13, 16, 19, 186 Geonoma undata var. skovii 8, 12, 17, 18, 19 Geonoma undata var. undata 10 Gratacos, X., as co-author 81 Griffith, P., as co-author 127 Guihaia 192 Guizol, P., as co-author 147 Gutiérrez-Ortega, J.S., as co-author 41 Hedyscepe 193 Hemithrinax 81, 181 Henderson, A., as co-author 81 Henderson, A.: A review of naturally occurring hybrids in palms (Arecaceae) 177 Henderson, A.: The history of a Hispaniolan Palm, Zombia antillarum 74 Heterospathe 186 Hodel, D.R., as co-author 41 Howea 186 Howea belmoreana 159, 186, 187 Howea fosteriana 159, 186, 187 Hydriastele 186 Hyophorbe 183 Hyospathe 185 Hyospathe elegans 24, 34, 38, 175, 176, 186 Hyospathe elegans subsp. elegans 163, 176 Hyospathe elegans subsp. sodiroi 31 Hyospathe pittieri 186 Hyphaene 192 Hyphaene thebaica 129, 130, 132, 134, 139, 141, 142, 144 Iguanura 186 Iriartea 166, 183 Iriartea deltoidea 24, 162, 169, 171, 175 Iriartella 165, 166, 183 Iriartella stenocarpa 166, 167, 175, 176 Itaya 181 Jailoloa 193 Johannesteijsmannia 181, 182 Johannesteijsmannia altifrons 23 Jubaea 183 Jubaeopsis 183, 188 Kapawi: A mega-diverse palm community in the eastern Amazon of Ecuador 161 Kerriodoxa 192 Korthalsia 192 Laccospadix 186

Laccosperma 143, 145, 192 Laccosperma secundiflorum, 130, 144 Lackman, I., as co-author 147 Lanonia 4. 181 Latania 192 Lemurophoenix 108, 193 Lemurophoenix halleuxii 23 Leopoldinia 23, 192 Lepidocaryum 62, 63, 65, 192 Lepidorrhachis 186 Leucothrinax 81, 181 Leucothrinax morrisii 178 Licuala 4, 69, 72, 181, 182 Licuala bruneiana 182 Licuala crassiflora 69 Licuala heatubunii 69, 70 Licuala longispadix 69 Licuala orbicularis 23 Licuala paludosa 182 Linospadix 186 *Livistona* 90, 96, 97, 101, 181, 187 *Livistona australis* 37, 181, 187 Livistona decora 181 *Livistona humilis* 93, 96, 181, 182 Livistona inermis 90, 93, 95, 96, 97, 98, 182 Livistona jenkinsiana 182 Livistona leichhardtii 93, 97, 98, 99, 182 Livistona lorophylla 182 Livistona mariae 182 Livistona saribus 182 Livistona victoriae 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100 Livistona victoriae and the North Austalian Exploring Expedition 1855-1856 90 Lock, M.: Palm Literature: The Poem Forest: Poet W.S. Merwin and the Palm Forest he Grew from Scratch 72 Lodoicea 192 Lodoicea maldivica 24 Loxococcus 186 Manicaria 192 Manicaria saccifera 24, 116 Manjekia 193 Marojejya 108, 193 Masoala 108, 193 Mauritia 62, 63, 65, 79, 192 Mauritia carana 64 Mauritia flexuosa 162, 170, 171, 175, 176 Mauritiella 23, 57, 59, 62, 63, 65, 192 Mauritiella aculeata 57 *Mauritiella armata* 57, 59, 64, 176 *Mauritiella disticha* 9, 55, 57–65 Mauritiella macroclada 57 Mauritiella pumila 57 Maxburretia 192 Medemia 192 Metroxylon 180 Metroxylon paulcoxii 180 Metroxylon vitiense 23 Metroxylon warburgii 180 Montero, O., as co-author 81 Montúfar, R., as co-author 5, 22 Myrialepis 192 Nannorrhops 192 Nenga 193 Neonicholsonia 185 Neoveitchia 193 Nephrosperma 193 Noblick, L., as co-author 127 Normanbya 193 Nypa 192 Nypa fruticans 23 Oenocarpus 185

Oenocarpus x andersonii 186 Oenocarpus bacaba 186, 187 Oenocarpus bataua 24, 31, 57, 164, 167, 171, 175, 186 Oenocarpus mapora 108, 175, 186 Oenocarpus minor 24, 30, 31, 176, 186 Oliveira, A.V.G. de, as co-author 57 Oncocalamus 192 Oncocalamus wrightianus 129 Oncosperma 186 Oncosperma gracilipes 186 Oncosperma horridum 186 Orania 192 Oraniopsis 183 Palm diversity and ethnobotany in Senegal, West Africa 127 Palm Literature 20, 66, 72, 73, 126 Palms of the remote Cerro Plateado Biological Reserve, southeastern Ecuador 5 Parajubaea 183 Pelagodoxa 192 Pérez, A.J., as co-author 5 Pérez-Farrera, M.A., J.S. Gutiérrez-Ortega & D.R. Hodel: Chamaedorea tacanensis: A climbing species from Mexico and Guatemala 41 Persson, C., as co-author 5 Phoenicophorium 193 Phoenix 181 Phoenix canariensis 181 Phoenix dactylifera 130, 181 Phoenix reclinata 130, 134, 144 Phoenix sylvestris 181 Phoenix theophrasti 181 Pholidocarpus 181 Pholidostachys 186 Pholidostachys dactyloides 24, 27 Pholidostachys pulchra 38 Pholidostachys synanthera 24, 176 Physokentia 186 Phytelephas 183 Phytelephas aequatorialis 23, 24 Phytelephas macrocarpa 183 Phytelephas tenuicaulis 172, 173, 175, 176 Phytelephas tumacana 24 Pigafetta 192 Pinanga 193 Plectocomia 192 Plectocomiopsis 192 Podococcus 192 Ponapea 193 Prata, E.M.B., M.F. Torres Jiménez, C.D. Bacon, A.V.G. de Oliveira & T. Emilio: The description of a charismatic new palm species in the Amazon 57 Prestoea 12, 13, 31, 185 Prestoea acuminata 10, 12, 16, 19, 24 Prestoea decurrens 24 Prestoea ensiformis 24 Prestoea montana 87 Prestoea schultzeana 175 Pritchardia 182 Pritchardia bakeri 183 Pritchardia flynnii 183 Pritchardia kahukuensis 183 Pritchardia martii 183 Pritchardia minor 183 Pritchardia napaliensis 183 Pritchardia viscosa 183 Pritchardia waialealeana 183 Pseudophoenix 86, 192 Ptychococcus 193 Ptychosperma 193 Rábehevitra, D.: Exploring for new localities of Tahina spectabilis in northeastern Madagascar 195

Raphia 143, 144, 192 Raphia farinifera 130 Raphia palma-pinus 130 Raphia sudanica 23, 130, 137, 140, 141, 142, 144 Ravenea 183 Ravenea glauca 23 Reinhardtia 23, 192 Reinhardtia gracilis 38 Reinhardtia gracilis var. gracilior 116 Reinhardtia latisecta 116 Rhapidophyllum 192 Rhapis 192 Rhapis humilis 188 Rhopaloblaste 186 Rhopalostylis 193 Rival, A., M. Ancrenaz, I. Lackman, M. Shafiq, P. Guizol & M. Djama: Addressing the monoculture system: Challenges in oil palm-based agroforestry 147 Rival, A.: Palm Literature: Planet Palm: How Palm Oil Ended Up in Everything — And Endangered the World 20 Roguet, D., as co-author 127 Roscheria 193 Roystonea 192 Roystonea boringuena 83, 86 Sabal 79, 180 Sabal × brazoriensis 180 Sabal causiarum 86 Sabal domingensis 86 Sabal mauritiiformis 116 Sabal mexicana 180 Sabal minor 180 Sabal palmetto 180, 187 Sabinaria 181 Salacca 192 Saribus 181 Satakentia 193 Satranala 192 Schippia 117, 121, 180 Schippia concolor 73, 107, 109, 111-117, 119, Schlumpberger, B.O., as co-author 33 Schweizer, E., as co-author 90 Sclerosperma 192 Selva Chi: A unique integration of education and palm conservation 22 Serenoa 182 Shafiq, M., as co-author 147 Socratea 7, 30, 38, 166, 183 Socratea exorrhiza 24, 30, 108, 175, 176 Socratea rostrata 24, 30 Sommieria 192 Stauffer, F., as co-author 127 Syagrus 183, 185, 187, 188 Syagrus × altopalacioensis 185 *Śyagrus* × andrequiceana 185 Syagrus botryophora 185 Syagrus × campos-portoana 185 Syagrus × cipoensis 185 Syagrus comosa 185 Syagrus coronata 185, 187 Syagrus × costae 185 Syagrus elata 185 Syagrus flexuosa 185 Syagrus glaucescens 185 Syagrus hoehnei 119 Syagrus insignis 119 Svagrus loefgrenii 185

Syagrus × lacerdamourae 185 Syagrus × matafome 185 Syagrus microphylla 185 Syagrus × mirandana 185 Syagrus oleracea 185 Syagrus pleioclada 185 Syagrus romanzoffiana 183, 185, 187 Syagrus sancona 24, 159, 164, 175, 176 Syagrus schizophylla 185 Syagrus × serroana 185 Syagrus smithii 164, 175, 176 Syagrus × teixeiriana 185 Syagrus × tostana 185 Syagrus vagans 185 Syagrus weddelliana 119 Synechanthus 183 Synechanthus fibrosus 38, 116 Synechanthus warscewiczianus 24 Tahina 192 Tahina spectabilis 195–201 Tectiphiala 186 The history of a Hispaniolan Palm, Zombia antillarum 74 The uncertain history of an early illustration of Chamaedorea ernesti-augusti 33 Thrinax 81, 119, 181 Thrinax radiata 86, 116 Torres Jiménez, M.F., as co-author 57 Trachycarpus 192 Trithrinax 180 Veitchia 193 Verschaffeltia 193 Voanioala 183, 188 Vonitra 108, 193 Wallaceodoxa 193 Wallichia 183 Wallichia caryotoides 183 Wallichia gracilis 183 Wallichia marianneae 183 Washingtonia 182 Welfia 186 Welfia regia 24, 27, 38 Wendlandiella 165, 183 Wendlandiella gracilis 165 Wendlandiella gracilis var. simplicifrons 165, 175 Wettinia 7, 9, 31, 166, 167, 183 Wettinia aequalis 24, 29 Wettinia drudei 167, 175 Wettinia kalbreyeri 24, 183 Wettinia maynensis 175, 176 Wettinia minima 7, 9, 10, 19 Wettinia oxycarpa 24, 183 Wettinia quinaria 24, 183 Wettinia verruculosa 24 William August Schipp (1891-1967): Commemorated in Schippia concolor and Chamaedorea schippii 109 Wodyetia 193 Yoxall, H., as co-author 109 Zapata, N., as co-author 5 Zombia 79, 81, 119, 180 Zombia antillarum 55, 74, 78, 79, 80 Zona, S: Palm Literature: Field Guide to the Palms of Belize 73 Zuckermann, J.C: Planet Palm: How Palm Oil Ended Up in Everything — And Endangered the World, reviewed 20

## The IPS is grateful for the support of the following patrons

## 2021

## Platinum (\$10,000 and up)

Dr. Thomas Jackson & Dr. Kathy Grant, California, USA, in support of the publication of PALMS Ms. Glenn Franklin, North Carolina, USA

### Gold (\$5000 to \$9999)

Mr. Leland Lai, California, USA Ms. Faith Bishock, Florida, USA

## Silver (\$1000 to \$4999)

Mr. Jeff Brusseau, California, USA Mr. Anthony Glomski, California, USA Dr. Andy Hurwitz, California, USA Ms. Mary Lock, Hawaii, USA Ms. Jill Menzel, Florida, USA Mr. Darold Petty, California, USA Dr. Laz Priegues, Florida, USA Mr. Haresh, Tamil Nadu, India Mr. Colin Wilson, New South Wales, Australia Ms. Libby Besse, Florida, USA

If you would like to make a gift, however large or small, please contact IPS President Robert Blenker, Robert.Blenker@gmail.com or any IPS Board member. To give on-line, visit www.palms.org.

