

Article

Isomalabaricane Triterpenes from the Marine Sponge *Rhabdastrella* sp.

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Abstract: The marine sponge of the genus *Geodia*, *Jaspis*, *Rhabdastrella*, and *Stelletta* are characterized chemically by a variety of isomalabaricane triterpenes. This class of compounds drew spotlights in marine lead discovery due to their profound anti-proliferative properties. Further research on exploring its chemical diversity led to the identifications of two new isomalabaricane-type triterpenes rhabdastin H (**1**) and rhabdastin I (**2**). Their structures were unraveled using a series of spectroscopic approaches. These isolates were found to exhibit unique structural features with the only reported tetrahydrofuran functionality among all marine-derived isomalabaricanes. Both compounds **1** and **2** showed activities against K562 (IC₅₀ 11.7 and 9.8 μM) and Molt4 (IC₅₀ 16.5 and 11.0 μM) leukemic cells in MTT cell proliferative assay.

Keywords: isomalabaricane; tetrahydrofuran moiety; anti-leukemia

1. Introduction

Marine sponges continue to act as a fruitful source of bioactive and unusual metabolites. Among these metabolites are the sponge-derived isomalabaricanes. They were reported to exhibit potent anti-proliferative properties against a series of cancer cells, including colorectal carcinoma [1], ovarian carcinoma [2], acute promyelocytic leukemia [3], acute lymphoblastic leukemia [4], prostate carcinoma [3], gastric adenocarcinoma [3], ductal carcinoma [3], hepatocellular carcinoma [3], cervix carcinoma [3], papillomavirus-related endocervical adenocarcinoma [5], and malignant melanoma [4]. The isomalabaricane-type triterpenoids are characterized by an α -methyl group at C-8, owing to their *trans-syn-trans* tricyclic ring junction instead of *trans-anti-trans* of their isomers, malabaricanes [1]. Isomalabaricanes are only found in sponges and are considered as chemotaxonomic markers of *Rhabdastrella* sp. [1] Despite the significant difficulties in the isolation and characterization of isomalabaricanes due to light induced 13-*E/Z* isomerization [6], their significant cytotoxic activity prompted extensive research on this class of compounds aiming to find new drug leads.

The current study aimed to explore novel isomalabaricanes from marine sources. We isolated two unusual compounds with tetrahydrofuran functional group, the moiety that

has been reported previously in malabaricane-type triterpenoids but without any cytotoxic assessments [7,8]. Then, the isolated compounds were evaluated against several cancer cell lines using MTT assay to further interpret their anti-proliferative properties.

2. Results

The freeze-dried specimen of the wild-type sponge *Rhabdastrella* sp. (Figure 1) was extracted with a 1:1 mixture of methanol (MeOH) and dichloromethane (CH₂Cl₂) (1:1) to provide the crude extract. The obtained crude extract was further fractionated and purified using normal and reversed-phase column chromatography yielding two isomalabaricanes, rhabdastin H (1) and rhabdastin I (2). These isolates demonstrated unique structural features with the presence of the first identified tetrahydrofuran moiety in this class of compounds.

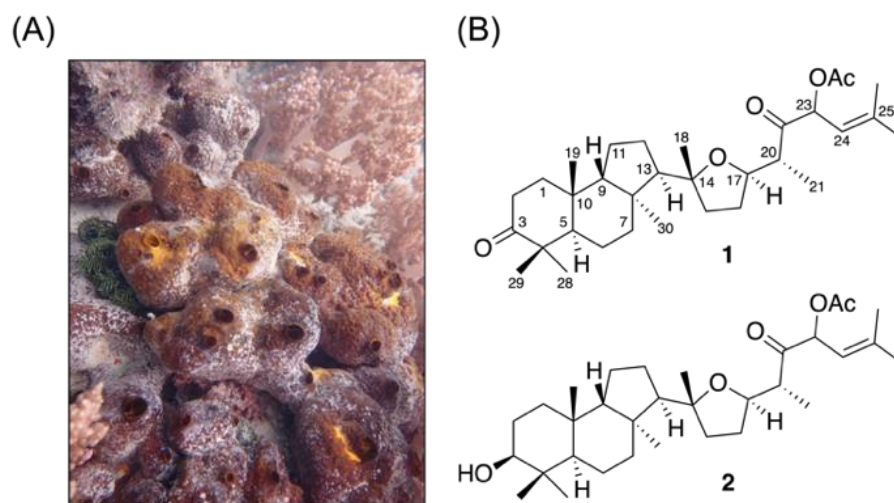


Figure 1. (A) Aquatic ecology of the sponge *Rhabdastrella* sp. and (B) the isolated isomalabaricane triterpenes.

The molecular formula of **1** was suggested as C₃₂H₅₀O₅ based on ¹³C NMR and HRESIMS data that showed a molecular ion peak at *m/z* 537.3539 [M + Na]⁺ implying eight degrees of unsaturation. The IR spectrum of **1** revealed the presence of carbonyl groups from absorptions at 1742, 1726, and 1703 cm⁻¹. Its ¹³C NMR spectrum of **1** (Table 1), measured in CDCl₃, showed the presence of thirty-two carbon signals, which were assigned by the assistance of DEPT data to nine methyl groups, eight sp³ methylenes, six sp³ methines (including two oxymethines), four sp³ quaternary carbons, one sp² methine and four sp² quaternary carbons (including two ketone carbonyl). The NMR signals at δ_C 169.8 (C) and 20.9 (CH₃) and δ_H 2.12 (3H, s) and the IR absorption at 1742 cm⁻¹ suggested the presence of an acetoxy group. Carbon signals of the eight methyl groups (δ_C 31.8, 29.3, 26.0, 24.5, 24.0, 19.3, 19.0 and 12.1), one oxygen-bearing methine carbon (δ_C 79.8), and one oxygenated quaternary carbon (δ_C 86.2), one trisubstituted carbon-carbon double bonds (δ_C 116.9, CH; 142.6, C), two ketones (δ_C 220.3 and 207.4) were also assigned. The resonances of one olefinic proton (δ_H 5.11, d, *J* = 10.0 Hz) and two oxygenated methines (δ_H 6.09, d, *J* = 10.0 Hz; δ_H 3.76, ddd, *J* = 14.0, 8.5, 4.5 Hz) were observed from the ¹H NMR spectroscopic data of **1** (Table 1).

Based on the above results and with the assistance of ¹H-¹H COSY and HMBC experiments (Figure 2), the planar structure of **1** was determined. To establish the proton sequences in **1**, the ¹H-¹H COSY spectrum analysis established five proton sequences from H₂-1 to H₂-2, H-5 to H₂-7, and H₂-11 to H₂-12, H₂-15 to H-21, and H-23 to H-24. These data, together with the HMBC correlations (Figure 2) from H₂-1 and H₂-2 to C-3, H₂-11 and H₂-12 to C-9, H₂-12 to C-8 and C-13, H₃-19 to C-1, C-5, C-9 and C-10, H₃-28 and H₃-29 to C-3, C-4 and C-5 and H₃-30 to C-7, C-8, C-9 and C-13 established the connectivity within the 6-membered (A), 6-membered (B), and 5-membered (C) rings.

Table 1. ^1H , ^{13}C , ^1H - ^1H COSY, and HMBC NMR data of **1**.

Position	δ_{H} (J in Hz) ^a	δ_{C} (Mult.) ^b	^1H - ^1H COSY	HMBC
1	2.08 m; 1.46 m	31.9 (CH ₂) ^d	H-2	C-3
2	2.66 m; 2.30 m	33.7 (CH ₂)		C-3
3		220.3 (C)		
4		46.8 (C)		
5	2.23 m	45.7 (CH)	H-6	
6	1.45 m; 1.34 m	20.0 (CH ₂)	H-5, H-7	
7	2.15 m; 1.35 m	32.3 (CH ₂)		C-5, C-9
8		42.2 (C)		
9	1.44 m	52.9 (CH)	H-11	
10		35.0 (C)		
11	1.52 m; 1.30 m	21.4 (CH ₂)	H-9, H-12	C-9
12	1.85 m; 1.55 m	25.4 (CH ₂)	H-11, H-13	C-8, C-9, C-13
13	1.78 m	59.8 (CH)	H-12	
14		86.2 (C)		
15	1.80 m	38.6 (CH ₂)	H-16	
16	1.97 m; 1.58 m	31.0 (CH ₂)	H-17	
17	3.76 ddd (14.0, 8.5, 4.5) ^c	79.8 (CH)	H-20	
18	1.17 s	24.5 (CH ₃)		C-13, C-14, C-15
19	0.79 s	24.0 (CH ₃)		C-1, C-5, C-9, C-10
20	2.70 m	47.6 (CH)	H-17, H-21	C-16, C-17
21	0.95 d (7.0)	12.1 (CH ₃)	H-20	C-17, C-20, C-22
22		207.4 (C)		
23	6.09 d (10.0)	77.0 (CH)	H-24	OAc
24	5.11 d (10.0)	116.9 (CH)	H-23	
25		142.6 (C)		
26	1.81 s	26.0 (CH ₃)		C-24, C-25, C-27
27	1.89 s	19.0 (CH ₃)		C-24, C-25, C-26
28	1.02 s	19.3 (CH ₃)		C-3, C-4, C-5, C-29
29	1.06 s	29.3 (CH ₃)		C-3, C-4, C-5, C-28
30	1.09 s	31.8 (CH ₃)		C-7, C-8, C-9, C-13
OAc	2.12 s	20.9 (CH ₃) 169.8 (C)		

^a 500 MHz in CDCl₃; ^b 125 MHz in CDCl₃; ^c *J* values (Hz) are given in parentheses; ^d Numbers of the attached protons were deduced by DEPT experiments.

Ring A of **1** was found to possess one ketone at C-3 and two methyl groups (C-28 and C-29), one methyl group (C-19), and one methyl groups (C-30) attached at C-4, C-8, and C-10, respectively. The key HMBC correlations suggested the connection of H₃-18 to C-13, C-14, and C-15, H-20 to C-16 and C-17; H₃-21 to C-17, C-20, and C-22; H-23 to C-22; H₃-26 and H₃-27 to C-24 and C-25. Thus, the side chain C-20 to C-27 was found to possess one double bond at C-24/C-25, one ketone group at C-22, two methyl groups at C-25, and one methyl group at C-20. One acetoxy group at C-23 was confirmed by the HMBC correlation between an oxymethine proton resonating at δ_{H} 6.09 (H-23) and the protons of an acetate methyl (δ_{H} 2.12) to the ester carbonyl carbon at δ_{C} 169.8. An ether linkage was proposed between C-14 and C-17 forming a tetrahydrofuran ring based on the degrees of unsaturation and molecular formula. Based on the above analysis, the gross structure of **1** was established unambiguously and named rhabdastin H following up the previous investigation of cytotoxic isomalabaricanes with oxygenated and olefinic side chains from the sponge *Rhabdastrella globostellata* [9].

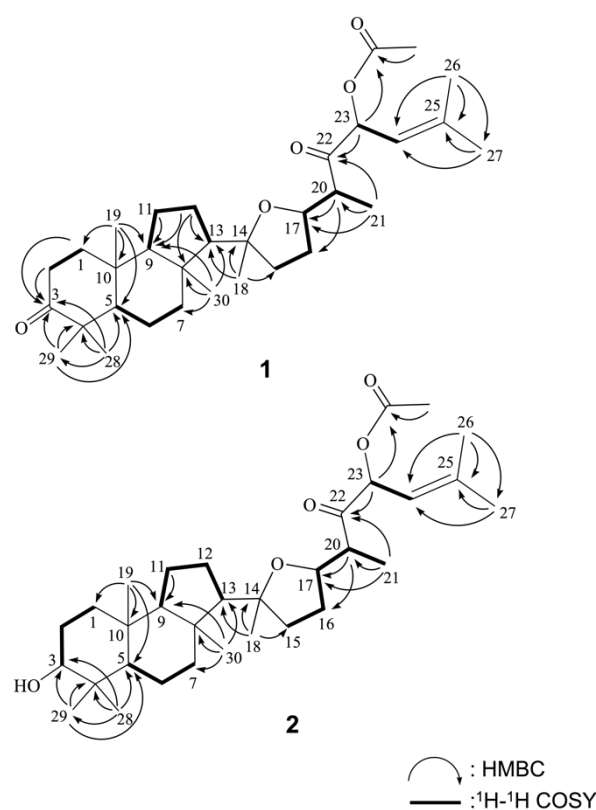


Figure 2. Selected ^1H - ^1H COSY and HMBC correlations of **1** and **2**.

The relative configurations of the eight chiral centers at C-5, C-8, C-9, C-10, C-13, C-14, C-17, and C-20 in **1** were elucidated by the following NOE analysis (Figure 3). It was found that H₃-19 (δ_{H} 0.79, s) showed NOE interaction with H-9 (δ_{H} 1.44, m) and H-9 with H₃-18.

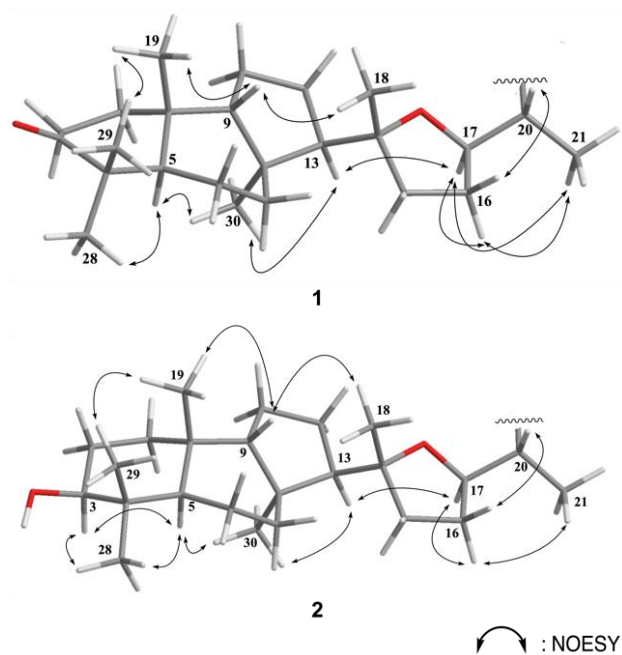


Figure 3. Selected NOESY correlations for **1** and **2**.

Since all naturally occurring isomalabaricanes displayed that H-5 is *trans* to Me-19, we assumed the β -orientation of H₃-19. Thus, H₃-18 and H-9 were suggested to be positioned on the β -face. One of the methyl groups (H₃-29) at C-4, which showed a NOE

correlation with H₃-19, was β -oriented and the other one (H₃-28) was α -oriented. The NOE correlations observed between H₃-28/H-5, H-5/H₃-30, H₃-30/H-13, H-13/H-17, H-17/H₃-21, and H-17/H-16 suggested α -orientation of H-5, H-13, H-17, H₃-21, and H₃-30. One of the methylene protons at C-16 (δ_{H} 1.97), which showed a NOE correlation with H-17 and H₃-21, was assigned as H-16 α and the other one (δ_{H} 1.58) as H-16 β . Moreover, the detection of large proton coupling constants at H-17 ($J = 13.5$ Hz) and H-20 ($J = 13.5$ Hz) further supported their *trans* conformation in between. The observed NOE correlation between H-20 and H-16 β suggested a β -orientation of H-20.

The HR-ESI-MS of rhabdastin I (**2**) showed a molecular ion peak at (m/z 539.3696 [M + Na]⁺) and the molecular formula C₃₂H₅₂O₅ was suggested based on the HRESIMS and ¹³C NMR data. The IR spectrum of **2** showed the absorption of carbonyl groups (1745 and 1725 cm⁻¹) and a hydroxy group (3436 cm⁻¹). The ¹H and ¹³C NMR spectroscopic data of **2** (Table 2) and **1** (Table 1) indicated similarity in structure except that the ketocarbonyl carbon (C-3) in **1** was replaced by a hydroxy group-bearing methine carbon in **2**. In the ¹³C NMR spectrum, the signal at δ_{C} 220.3 was replaced by a signal at δ_{C} 79.5, and in the ¹H NMR spectrum, a signal at δ_{H} 3.24 (dd, $J = 11.5, 6.5$ Hz) was attributed to a hydroxy-bearing methine at C-3. H-3 showed an NOE correlation with H-5 (δ_{H} 1.54, m) suggesting a β -orientation of the hydroxy group at C-3. Therefore, **2** was identified as the 2*S*-hydroxy derivative of **1**.

Table 2. ¹H, ¹³C, ¹H-¹H COSY, and HMBC NMR data of **2**.

Position	δ_{H} (J in Hz) ^a	δ_{C} (Mult.) ^b	¹ H- ¹ H COSY	HMBC
1	1.48 m; 1.41 m	33.9 (CH ₂) ^d	H-2	
2	1.73 m; 1.61 m	29.2 (CH ₂)	H-3	
3	3.24 dd (11.5,6.5)	79.5 (CH)		
4		39.0 (C)		
5	1.54 m	46.6 (CH)	H-6	
6	1.64 m; 1.38 m	18.5 (CH ₂)	H-5, H-7	
7	2.04 m; 1.35 m	32.9 (CH ₂)		
8		42.0 (C)		
9	1.42 m	55.0 (CH)	H-11	
10		35.6 (C)		
11	1.49 m; 1.30 m	21.2 (CH ₂)	H-9, H-12	C-9
12	1.82 m; 1.54 m	25.0 (CH ₂)	H-11, H-13	
13	1.78 m	59.9 (CH)	H-12	
14		86.2 (C)		
15	1.80 m; 1.75 m	38.8 (CH ₂)	H-16	
16	1.98 m; 1.60 m	31.1 (CH ₂)	H-17	
17	3.75 ddd (13.5, 8.5, 4.5) ^c	79.5 (CH)	H-20	
18	1.16 s	24.5 (CH ₃)		C-13, C-14, C-15
19	0.95 s	23.1 (CH ₃)		C-1, C-5, C-9, C-10
20	2.69 ddd (13.5, 8.5, 1.5)	47.5 (CH)	H-17, H-21	C-16, C-17
21	0.94 d (6.0)	12.1 (CH ₃)	H-20	C-17, C-20, C-22
22		207.4 (C)		
23	6.10 d (10.0)	78.7 (CH)	H-24	C-22, C-24, OAc
24	5.11 d (10.0)	116.9 (CH)	H-23	
25		142.6 (C)		
26	1.81 s	26.0 (CH ₃)		C-24, C-25, C-27
27	1.89 s	19.0 (CH ₃)		C-24, C-25, C-26
28	0.98 s	15.9 (CH ₃)		C-3, C-4, C-5, C-29
29	0.78 s	29.1 (CH ₃)		C-3, C-4, C-5, C-28
30	1.04 s	31.6 (CH ₃)		C-7, C-8, C-9, C-14
OAc	2.10 s	20.9 (CH ₃) 169.8 (C)		

^a 500 MHz in CDCl₃; ^b 125 MHz in CDCl₃; ^c J values (Hz) are given in parentheses; ^d Numbers of the attached protons were deduced by DEPT experiments.

The plausible biosynthesis route of the obtained isomalabaricane presented in Figure 4. The main isomalabaricane 6/6/5 carbocyclic skeleton might be derived from 2,3*S*-oxidosqualene through hydroxylation and electrocyclizations. The attached tetrahydrofuran moiety and the subsequent side chain were suggested to go through a hydroxylation, an electrocyclization, dehydration, oxidation, and acetylation, forming the first identified isomalabaricane with tetrahydrofuran functionality.

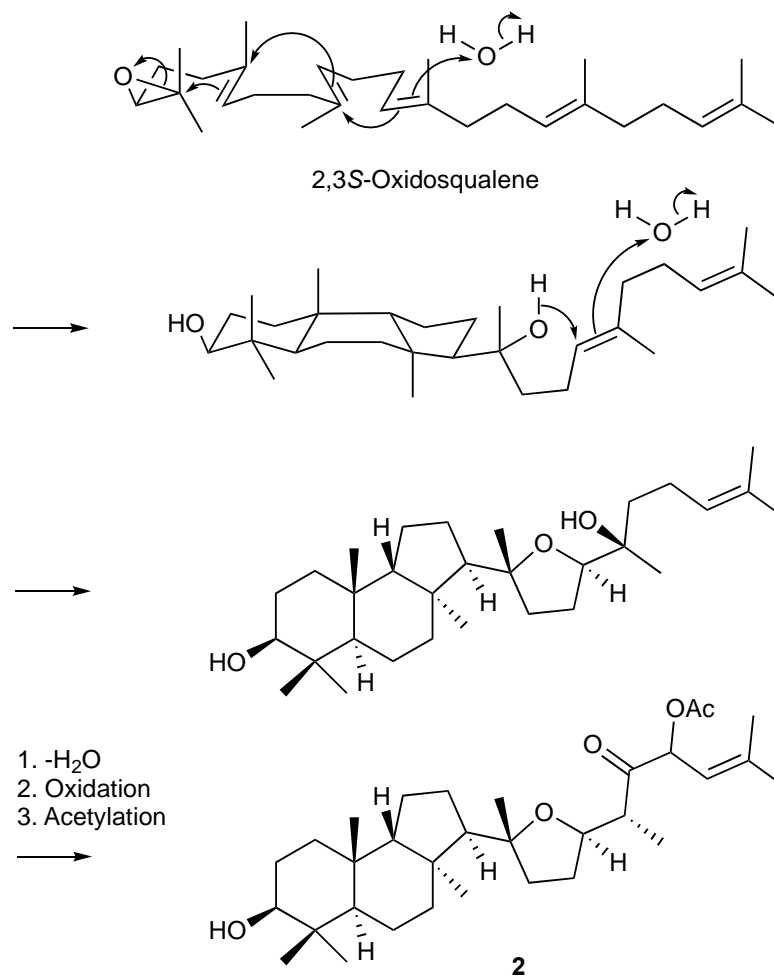


Figure 4. The plausible biosynthetic pathway of **2**.

To further clarify the anti-proliferative potential of the isolated compounds, four cancer cell lines (DLD-1: colorectal adenocarcinoma; T-47D: ductal carcinoma; Molt4: acute lymphoblastic leukemia; K562: chronic myelogenous leukemia) were used for MTT screening (Table 3). Compounds **1** and **2** were found to exhibit anti-proliferative activities against Molt4 and K562 leukemic cells with the IC₅₀ value ranging from 9.81 to 16.54 μM.

Table 3. Anti-proliferative activities of compounds **1** and **2**.

Compounds	Cell Lines (IC ₅₀ μM)			
	DLD-1	T-47D	Molt4	K562
1	– ^a	– ^a	16.54	11.71
2	– ^a	17.48	11.03	9.81
Doxorubicin ^b	0.42	0.18	0.02	0.28

^a IC₅₀ > 20 μM; ^b positive control.

3. Materials and Methods

3.1. General Experimental Procedures

Optical rotation spectra were recorded on a JASCO P-1010 polarimeter (JASCO, Tokyo, Japan). UV spectra were analyzed using JASCO UV-530 ultraviolet spectrophotometers. IR spectra were obtained on a Fourier-transform IR spectrophotometer Varian Digilab FTS 1000 (Varian Inc., Palo Alto, CA, USA). NMR spectra were obtained on a JEOL ECZ 400S or an ECZ 600R NMR (JEOL, Tokyo, Japan). HRESIMS data were collected on a Bruker APEX II instrument (Bruker Daltonik, Bremen, Germany). TLC was performed on Kieselgel 60 F₂₅₄ (0.25 mm, Merck, Darmstadt, Germany) and/or RP-18 F₂₅₄ (0.25 mm) coated plates and then visualized by spraying with 50% H₂SO₄ and heating on a hot plate. Silica gel 60 (Merck, 40–63 µm and 63–200 µm) were used for column chromatography. A Rheodyne 7725 injection port, a Hitachi L-2455 Photodiode Array Detector, and a Hitachi L-7100 pump (Hitachi, Tokyo, Japan), as well as a column Supelco Ascentis® C-18 Cat #: 581343-U, were applied for HPLC chromatography. All methods were carried out following the relevant guidelines and regulations.

3.2. Animal Material

The specimen of the wild-type sponge *Rhabdastrella* sp. was collected by scuba diving at a depth about 3–5 m from Kenting, Pingtung, Taiwan in December 2017. The voucher specimen was deposited at –20 °C at the National Museum of Marine Biology and Aquarium, Taiwan (specimen No. 2017-1221-SP). Taxonomic identification was performed by Dr. Hsing-Hui Li using 18S DNA sequence and morphology determination.

3.3. Extraction and Isolation

Rhabdastrella sp. (500 g fresh weight) was collected and freeze-dried. The freeze-dried material (75 g, dry weight) was minced and extracted three times with a 1:1 mixture of methanol (MeOH) and dichloromethane (CH₂Cl₂). The crude extract was evaporated under reduced pressure to afford a residue (8 g), and the residue was subjected to a normal phase column chromatography on silica gel (70–230 mesh), using *n*-hexane, increasing polarity of *n*-hexane:EtOAc mixtures, and acetone to yield 13 fractions: L1 (eluted by *n*-hexane), L2 (eluted by *n*-hexane:EtOAc, 100:1), L3 (50:1), L4 (30:1), L5 (20:1), L6 (10:1), L7 (5:1), L8 (3:1), L9 (2:1), L10 (1:1), L11 (1:2), L12 (eluted by EtOAc) and L13 (eluted by acetone). L10 was further separated with silica gel (*n*-hexane:acetone 6:1) using normal phase HPLC to afford ten subfractions (L10-1 to L10-10). Subfraction L10-4 was then subjected to a reversed-phase HPLC (RP-HPLC) (MeOH:H₂O, 85:15), yielding **1** (3.1 mg). Similarly, the subfraction L10-6 was purified by RP-HPLC (MeOH:H₂O, 80:20) to provide **2** (8.5 mg).

Rhabdastin H (**1**): Colorless oil; $[\alpha]_D^{25} -47.1$ (*c* 0.03, CHCl₃); IR (ATR, CHCl₃) ν_{\max} 1742 and 1726 cm⁻¹; ¹H NMR data, see Table 1; HRESIMS *m/z* 537.3539 [M + Na]⁺ (calcd. 537.3550, see supplementary materials).

Rhabdastin I (**2**): Colorless oil; $[\alpha]_D^{25} -137.5$ (*c* 0.02, CHCl₃); IR (ATR, CHCl₃) ν_{\max} 3436, 1745, and 1725 cm⁻¹; ¹H NMR data, see Table 2; ESIMS *m/z* 539.3696 [M + Na]⁺ (calcd. 539.3707, see supplementary materials).

3.4. MTT Cell Proliferation Assay

MTT assay was used to examine the cellular proliferation of DLD-1 (colorectal adenocarcinoma), T-47D (ductal carcinoma), Molt4 (acute lymphoblastic leukemia), and K562 (chronic myelogenous leukemia) after 1 and 2 treatments. American Type Culture Collection (ATCC, Manassas, VA, USA) was the source for all cell lines. Briefly, cells at 1 × 10⁵ cells/mL were seeded at 96-well plates (150 µL/well) and incubated with several concentrations of compounds **1** and **2** for 24 h. After adding 50 µL MTT solution (1 mg/mL in PBS), the culture was incubated at 37 °C for 4 h, following which 200 µL DMSO was added to dissolve the formazan. The plate was then read on an ELISA microplate reader at 595 nm absorbance.

4. Conclusions

The current study highlighted the discovery of the first isomalabaricane derivatives with tetrahydrofuran moiety. Although the identified functionalities did not lead to a dramatic increase in the anti-proliferative properties, the chemical diversity of this class of triterpenes was enriched by these compounds with such unique structures.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/md19040206/s1>, ESIMS, HRESIMS, IR, 1D, 2D, and DEPT NMR spectra of 1–2.

Author Contributions: K.-H.L., W.-C.W. and J.-H.S. conceived and designed the experiments; Z.-H.H., K.-H.L. and J.-H.S. performed the sample collections, extraction, isolation, structures determination, and qualitative HPLC analysis; the pharmacological experiments were carried out by W.-C.W.; K.-H.L., W.-C.W. and J.-H.S. contributed reagents and analysis tools; B.-R.P., K.-H.L., M.E.-S., W.-C.W. and J.-H.S. participated in data interpretation, wrote the manuscript and revised the paper. All authors have read and agreed to the published version of the manuscript.

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Institutional Review Board Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

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