ZooKeys 814: 71–94 (2019)
doi: 10.3897/zookeys.814.24601
http://zookeys.pensoft.net

RESEARCH ARTICLE

A peer-reviewed open-access journal
ZooKeys
Launched to accelerate biodiversity research

A new species of *Acartia* (Copepoda, Calanoida) from the Philippines, based on morphological and molecular analyses

Khwanruan Srinui[1], Susumu Ohtsuka[2], Ephrime B. Metillo[3], Masahide Nishibori[2]

1 *Institute of Marine Science, Burapha University, Muang, Chon Buri 20131, Thailand* **2** *Graduate School of Biosphere Science, Hiroshima University, 1-4-4 Kagamiyama, Higashi-Hiroshima 739-8528, Japan* **3** *Department of Biological Sciences, Mindanao State University – Iligan Institute of Technology, Iligan City 9200, Philippines*

Corresponding author: *Khwanruan Srinui* (khwanruan@buu.ac.th)

Academic editor: *K.H. George* | Received 21 February 2018 | Accepted 12 November 2018 | Published 8 January 2019

http://zoobank.org/A0FA5FDC-2115-4C53-9927-550C0192E112

Citation: Srinui K, Ohtsuka S, Metillo EB, Nishibori M (2019) A new species of *Acartia* (Copepoda, Calanoida) from the Philippines, based on morphological and molecular analyses. ZooKeys 814: 71–94. https://doi.org/10.3897/zookeys.814.24601

Abstract

A new species of *Acartia* (*Odontacartia*), *A.* (*O.*) *edentata* **sp. n.**, was collected from Leyte Island in the Philippines. Morphologically, the new species resembles *A.* (*O.*) *pacifica* Steuer, 1915. The female of the new species differs from other species of the *A.* (*O.*) *erythraea* Giesbrecht, 1889 species group in the absence of a pair of sharp spines on the posterior border of the genital double-somite and absence of setules on the lateral margins of urosomites 1–3. Unlike other congeners of the species group, males of the new species lack fine setules along the posterior margin of the prosome. Comparison of the new species with *A.* (*O.*) *pacifica* by pairwise distance data for the 16S (282 bp) gene indicates that these species differ by 20–21%, while the COI gene (636 bp) indicates a difference of 16–17%. The new species seems to be a coastal, occurring in warm waters having a salinity of 33.5.

Keywords

Acartia, Calanoida, mitochondrial genes, Philippines, phylogeny

Introduction

The planktonic calanoid copepod genus *Acartia* Dana, 1846 so far comprises 64 species worldwide (Razouls et al. 2018). The genus consists of seven subgenera: *A.* (*Acanthacartia*) Steuer, 1915, *A.* (*Acartia*) Dana, 1846, *A.* (*Acartiura*) Steuer, 1915, *A.* (*Euacartia*) Steuer, 1915, *A.* (*Hypoacartia*) Steuer, 1915, *A.* (*Odontacartia*) Steuer, 1915, and *A.* (*Planktacartia*) Steuer, 1915 (Boxshall and Halsey 2004; Razouls et al. 2018). The subgenus *Odontacartia* is widely distributed in brackish to oceanic waters of the Indo-West Pacific and currently accommodates 12 species (Ueda and Bucklin 2006; Razouls et al. 2018). Generally, these can be distinguished by sexual dimorphic features of the posterior prosome, urosomites, antennules, and fifth legs.

The *centura* and *erythraea* species groups have so far accommodated 7 and 5 species, respectively, with the unassigned species *A.* (*O.*) *lilljeborgi* Giesbrecht, 1889. The *centrura* species group now accommodates the following seven species (Steuer 1923; Ueda 1986): *A.* (*O.*) *bowmani* Abraham, 1976; *A.* (*O.*) *centrura* Giesbrecht, 1889; *A.* (*O.*) *edentata* sp. n.; *A.* (*O.*) *mertoni* Steuer, 1917; *A.* (*O.*) *ohtsukai* Ueda & Bucklin, 2006; *A.* (*O.*) *pacifica*; and *A.* (*O.*) *spinicauda* Giesbrecht, 1889.

The common *A.* (*O.*) *pacifica* has a wide distribution on the coasts of the Indo-West Pacific and in East Asian continental waters. The population in the brackish Ariake Sea, western Japan, was identified as a different species, and, after detailed morphological and molecular analyses, was described as *A.* (*O.*) *ohtsukai* Ueda & Bucklin, 2006 and considered to be a continental relict (Ueda and Bucklin 2006).

During our copepod surveys in southeastern Asia, we found an undescribed species of *A.* (*Odontacartia*) from Leyte Island, the Philippines. It is closely related to *A.* (*O.*) *pacifica*, but unique in lacking paired posterodorsal pointed processes on the female genital double-somite. Following Ueda and Bucklin's (2006) methodologies, we are able to define the new species described herein. A key to species of the subgenus *Acartia* (*Odontacartia*) is also provided.

Material and methods

Morphology and sampling

The material examined was collected from three sites: Carigara Bay, Leyte Island, the Philippines (11°30'70"N; 124°69'01"E, depth 15 m) during the daytime on August 23, 2013 (local time 15:55); Ariake Bay, Seto Inland Sea, Pacific Ocean (34°18'40"N; 132°56'40"E, depth 12 m) on August 11, 2011 (local time 13:30); and South Korea (34°40'20"N; 127°48'24"E, depth 24 m) on August 30, 2011 (local time 15:00) (Stations 1–3, respectively, in Fig. 1). Water temperature and salinity were measured using a water quality meter, YSI model Pro2030. All specimens were obtained using a series of vertical tows from the bottom to the surface of the water with a conical plankton net (diameter: 30 cm, mesh size: 0.33 mm). Specimens for morphological examination

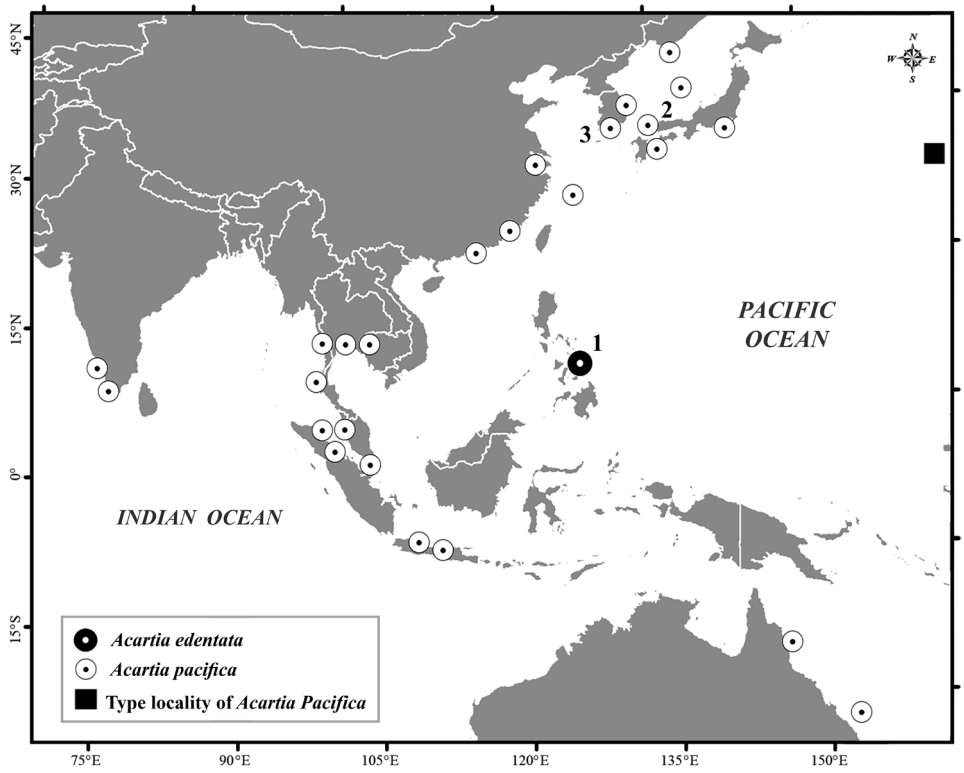

Figure 1. Distribution of *Acartia* (*Odontacartia*) *pacifica* and its sibling species based on data from this and previously published reports. In the present study, samples were obtained from three sites: Station 1, *Acartia* (*Odontacartia*) *edentata* sp. n., Leyte Island, the Philippines (black donut); Station 2, *A.* (*O.*) *pacifica*, Ariake Bay in the Seto Inland Sea, Japan (white donut); and Station 3, *A.* (*O.*) *pacifica*, South Korea Sea, Korea. Data from earlier studies indicate the distribution of *A.* (*O.*) *pacifica* in the Pacific Ocean (Steuer 1915; Tanaka 1965; Brodsky 1967; Ueda and Bucklin 2006); Korean waters (Kang 2011); the Yellow Sea, Chiekong River, Juilong Estuary, Changjiang Estuary, China (Shen and Lee 1963; Chen and Zhang 1965; Shang et al. 2007; Gao et al. 2008); eastern Indonesian waters of Java Sea, Bintulu coast, Indonesia (Früchtl 1923; Mulyadi 2004; Johan et al. 2013); the Gulf of Thailand (Pinkaew 2003); the Indian Ocean (Sewell 1933; Wellershaus 1969; Pillai 1971; Resai et al. 2004; Phukham 2008; Treeramaethee et al. 2013); water of the Great Barrier Reef, Moreton Bay, Australia (Farran 1936; Greenwood 1978); and type locality of *A.* (*O.*) *pacifica* (black square).

were preserved with a 4% neutral buffered formalin/seawater solution, while those for DNA analysis were fixed in 99.5% ethyl alcohol. Adult acartiids were sorted from the original samples under a stereomicroscope (Olympus SZX16, Olympus, Tokyo, Japan). Specimens were dissected using a stainless steel pin (no. 00), and transferred to a polyvinyl lactophenol solution. Morphological features were measured directly with an ocular micrometer, and were drawn using a camera lucida attached to a compound microscope (Olympus BX53, Olympus, Tokyo, Japan). Male and female urosomites of the new species were examined with a scanning electron microscope (JSM-6510LV,

Jeol Ltd, Tokyo, Japan). Terminology follows Huys and Boxshall (1991). Specimens of the species of *Acartia* and *A.* (*O.*) *pacifica* examined in the present study are deposited in the Institute of Marine Science, Burapha University (BIMS–Zoo–0266).

The structure of female and male antennules follows a pattern of the basically uniramous 28 segments. The antennules of both sexes are similar except for the geniculate right antennule in calanoid copepods as in the copepodid I (CI) setation pattern (Boxshall and Huys 1998). We followed Ueda and Bucklin (2006) because the new species more closely resembles *A.* (*O.*) *pacifica* in having equal antennule segment numbers, a similar setation pattern in right antennules of females and males, except segment 5 (XIII), and three rows of spinules ventrolaterally on the second somite of males.

Molecular analysis

In this study, we used adults of the undescribed species *Acartia* (*O.*) from the Philippines and *A.* (*O.*) *pacifica* individuals for genetic analysis of the mitochondrial cytochrome oxidase I (COI) and 16S rRNA (16S) genes. DNA for PCR amplification was prepared from individual males or females placed in microcentrifuge tubes with 50 µl chelex 5%, 1 µl Proteinase K (20 mg/ml). Tubes were heated to 65 °C for 1 hour, boiled at 100 °C for 8 min, and centrifuged at 10,000 *x g* for 8 min. PCR reagents included 5 µl of 10× buffer, 4 µl of 50 mM $MgCl_2$, 5 µl of 2 mM dNTPs, 0.25 µl of 10 µM primer solutions, 0.25 µl Taq DNA polymerase (Product no. PL1202, Vivantis, Malaysia) and 30.25 µl distilled water, following (Ueda and Bucklin 2006). We used the universal COI primers COI–LCO1490F: 5'– GGTCAACAAATCATAAAGATATTGG–3' and COI-HCO 2198R: 5'– TAAACTTCAGGGTGACCAAAAAATCA–3' according to (Folmer et al. 1994). Primers used for 16S amplification were 16S–167F: 5'– GACGAGAAGACCCTATGA/AG–3' and 16S–BR–HR: 5'–CCGGTTTGAACTCAGATCATGT–3' (Palumbi 1996; Bucklin et al. 1998). The PCR amplification protocol consisted of 40 cycles of denaturation at 94 °C for 1 min, annealing at 45 °C for 2 min, and extension at 72 °C for 3 min. PCR products were electrophoresed on a 1% agarose gel to confirm their size and quality, and then purified using the Hiyield™ Gel/PCR Fragments Extraction Kit (PG-913-12041, RBCBioscience, Taiwan). The purified PCR products were sequenced by Macrogen Inc. (Seoul, Korea) with an Automated Sequencer (model ABI 3730 XL, Applied Biosystems, USA).

DNA sequences were manually edited using Sequence Scanner version 1.0 (Applied Biosystems) and compared with the GenBank: *A.* (*O.*) *pacifica* (accession number DQ071177 for COI and DQ071175 for 16S); *A.* (*O.*) *ohtsukai* (accession no. DQ071177 for COI and DQ071176 for 16S); *Acartia* (*Acanthacartia*) *tsuensis* Itô, 1956 (accession no. KC287427 for COI); and *A.* (*O.*) *erythraea* (accession no. DQ320504 for 16S). Sequences and multiple alignments were constructed with BioEdit version 7.1 (Hall 1999). Pairwise distances were determined with MEGA 6 (Tamura et al. 2013) using the maximum likelihood (ML) and bootstrapping 1,000 times (Saitou and Nei 1987).

Systematics

Order Calanoida Sars, 1903
Family Acartiidae Sars, 1903
Genus *Acartia* Dana, 1846
Subgenus *Acartia* (*Odontacartia*) Steuer, 1915

***Acartia* (*Odontacartia*) *edentata* Srinui, Ohtsuka & Metillo, sp. n.**
http://zoobank.org/2EC6BED6-0611-465D-A0A3-643511DF8542
Figures 2–5

Material. Type locality: Carigara Bay, Off Leyte Island, the Philippines (11°30'70"N; 124°69'01"E) (Fig. 1), August 23, 2013 (10 ♀, 10 ♂).

Types. Holotype: ♀, dissected and mounted on 2 glass slides (BIMS–Zoo–0267); paratype (allotype): 1 ♂, dissected and mounted on 2 glass slides (BIMS–Zoo–0268); additional paratypes: 4 ♀, 3 ♂ partially dissected and mounted on 3 glass slides (BIMS–Zoo–0269).

Measurements. Female. Total length, 1.19–1.23 mm (mean ± SD = 1.21 ± 0.01 mm, *N* = 10; holotype, 1.19 mm); prosome length, 0.42–0.46 mm (0.44 ± 0.01 mm; holotype, 0.44 mm); prosome width, 0.24–0.29 mm (0.26 ± 0.01 mm; holotype, 0.25 mm). Male. Total length 1.08–1.15 mm (mean ± SD = 1.10 ± 0.02 mm, *N* = 10; allotype, 1.10 mm); prosome length, 0.39–0.41 mm (0.40 ± 0.00 mm; allotype, 0.40 mm); prosome width, 0.23–0.26 mm (0.24 ± 0.01 mm; allotype, 0.25 mm).

Descriptions. Female. Body (Fig. 2A, B) elongate; cephalosome completely separate from first pedigerous somite; anterior margin of cephalosome triangular in dorsal view; rostrum with pair of thick, strong and sharp filaments (Figs 4F, 5F); fourth and fifth pedigerous somites fused. Posterior prosome symmetrical with pair of acute processes on each side: large ventrolateral, pointed processes with pair of small prominences between and pair of smaller, pointed processes dorsally (Fig. 2A). Urosome composed of three free somites; genital double-somite symmetrical with ratio of width–length ratio approximately 1:1, lacking posterodorsal pointed processes (Figs 2A, 5A); second urosomite with pair of strong, posterodorsal, pointed processes; anal somite as wide as long, without lateral rows of fine setules. Proportional lengths of urosomites and caudal ramus 41:22:15:22 (= 100). Caudal rami with setules along lateral margin, and symmetrical with 6 plumose setae (II–VII). I absent, V longest, and VII inserted anterodorsally.

Antennule (Fig. 2C) reaching beyond posterior end of second urosomite, symmetrical, 17-segmented; segments II–VIII completely fused; segments II, IV, VII, VIII, XI, XV, and XVII with aesthetasc (ae). Fusion pattern and setal elements as follows (Roman numerals represent ancestral segments): I = 1, II–VIII= 7 + 2ae, IX = 1+ (1 spiniform), X–XI = 2 + (1 spiniform) + ae, XII = 1, XIII–XV = 3 + ae, XVI= 1 + ae, XVII–XVIII = 2 + (1 process) + ae, XIX = 1 + (1 process), XX = 1, XXI= 1 + (1 process) + ae, XXII = 1, XXIII = 1, XXIV = 2, XXV = 2 + ae, XXVI = 2, XXVII–XXVIII = 4 + ae.

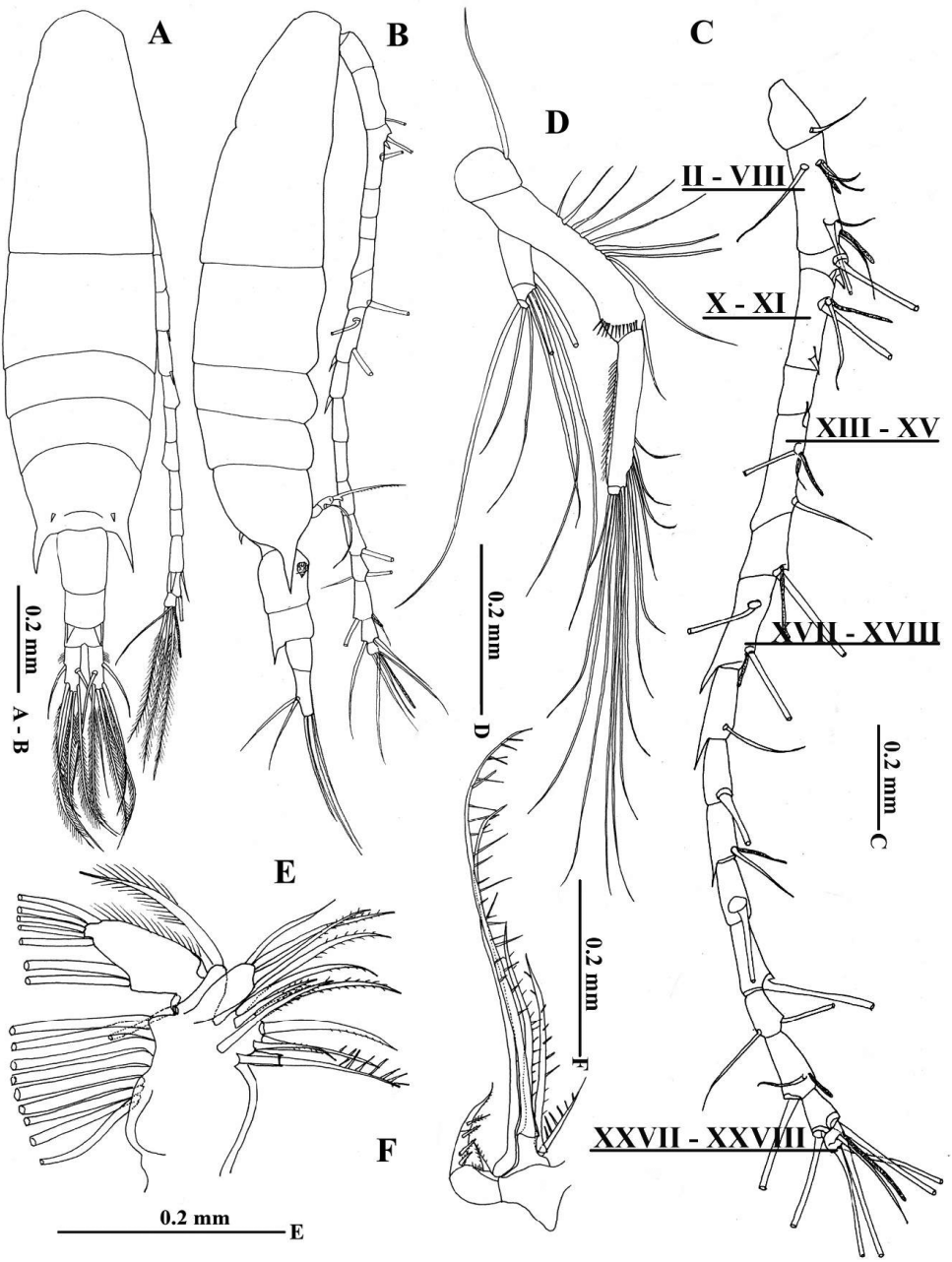

Figure 2. *Acartia* (*Odontacartia*) *edentata* sp. n. female (holotype) **A** Habitus, dorsal view **B** Habitus, lateral view **C** Antennule, Roman numerals denote segment numbers **D** Antenna **E** Maxilla **F** Maxilliped.

Antenna (Fig. 2D) coxa with single seta; basis fused to elongated first endopodal segment forming allobasis with eight setae on outer medial margin, and single lateral seta and transverse row of small spinules terminally; second segment with eight outer

setae and fine setules along inner margin; free terminal segment short with six setae. Exopod 3-segmented, setation formula 1, 4, 3.

Mandible (Fig. 3A) gnathobase having two sharp cuspid teeth, one blunt tooth, and three small sharp teeth bordered by small spinules at the proximal end; basis with fine setules on medial outer marginal, single seta distally, and patch of small spinules on surface at midlength; first endopodal segment short with two short setae, second segment with seven setae; exopod 4-segmented, first to fourth with setation formula 1, 1, 2, 2; first segment with row of small spinules.

Maxillule (Fig. 2E) with precoxal arthrite bearing nine strong spines; coxal endite with three terminal setae; coxal epipodite with one short and eight long setae; basal exite with one terminal seta and one proximal seta; exopod 1-segmented fused with basis and bearing two long medial setae, five setae terminally, with fine spinules along outer marginal; endopod absent.

Maxilla (Fig. 3B) with syncoxal endite bearing 5, 3, 3, 2 setae; basis with one long seta; endopod with four long, one medium, and one short setae.

Maxilliped (Fig. 2F) highly reduced; syncoxa with setation formula of two long, one medium and one short seta; basis with one short strong, one long setae and row of setules along inner margin; endopod 2-segmented, with four inner spines and terminal spiniform element.

Legs 1 to 4 (Fig. 3C–F) biramous, each with 2-segmented endopod and 3-segmented exopod; coxa unarmed; second endopodal segments of leg 1 and 4 and third exopodal segment of leg 1 with row of small spinules anteriorly. Seta and spine formula as shown in Table 1.

Leg 5 (Fig. 3G) symmetrical, coxae and intercoxal sclerite completely fused; basis longer than wide, outer margin with single lateral seta, slightly longer than terminal seta of exopod; exopod with knob-like projection basally, distal half spinulose.

Male. Body (Fig. 4A, B) similar to that of female; cephalosome anterior bluntly triangular in dorsal view; rostrum (Fig. 4E) with paired filaments (Figs 4E, 5E). Posterior prosome symmetrical with pair of short acute processes dorsolaterally and longer ventrolateral acute processes, with pair of small prominences between two dorsolateral processes. Posterior margin of prosome naked. Urosome composed of five somites, symmetrical in dorsal view; genital somite (= first urosomite) as long as wide, bearing 2 dorsolateral rows of small spinules; second urosomite with two pairs of strong posterior dorsolateral, processes (Figs 4A, B, 5 E), outer shorter than inner, and furnished with

Table 1. *Acartia* (*Odontacartia*) *edentata* sp. n. armature formula for legs 1–4, with spines and setae indicated by Roman and Arabic numerals, respectively, following Huys and Boxshall (1991).

	Coxa	Basis	Exopod segment			Endopod segment	
			1	2	3	1	2
Leg 1	0-0	0-0	1-1;	I-1;	2,I,4	0-1;	1,2,3
Leg 2	0-0	0-0	0-1;	0-1;	0,I,5	0-2;	1,2,4
Leg 3	0-0	0-0	0-1;	0-1;	0,I,5	0-2;	1,2,4
Leg 4	0-0	1-0	0-1;	0-1;	0,I,5	0-3;	1,2,3

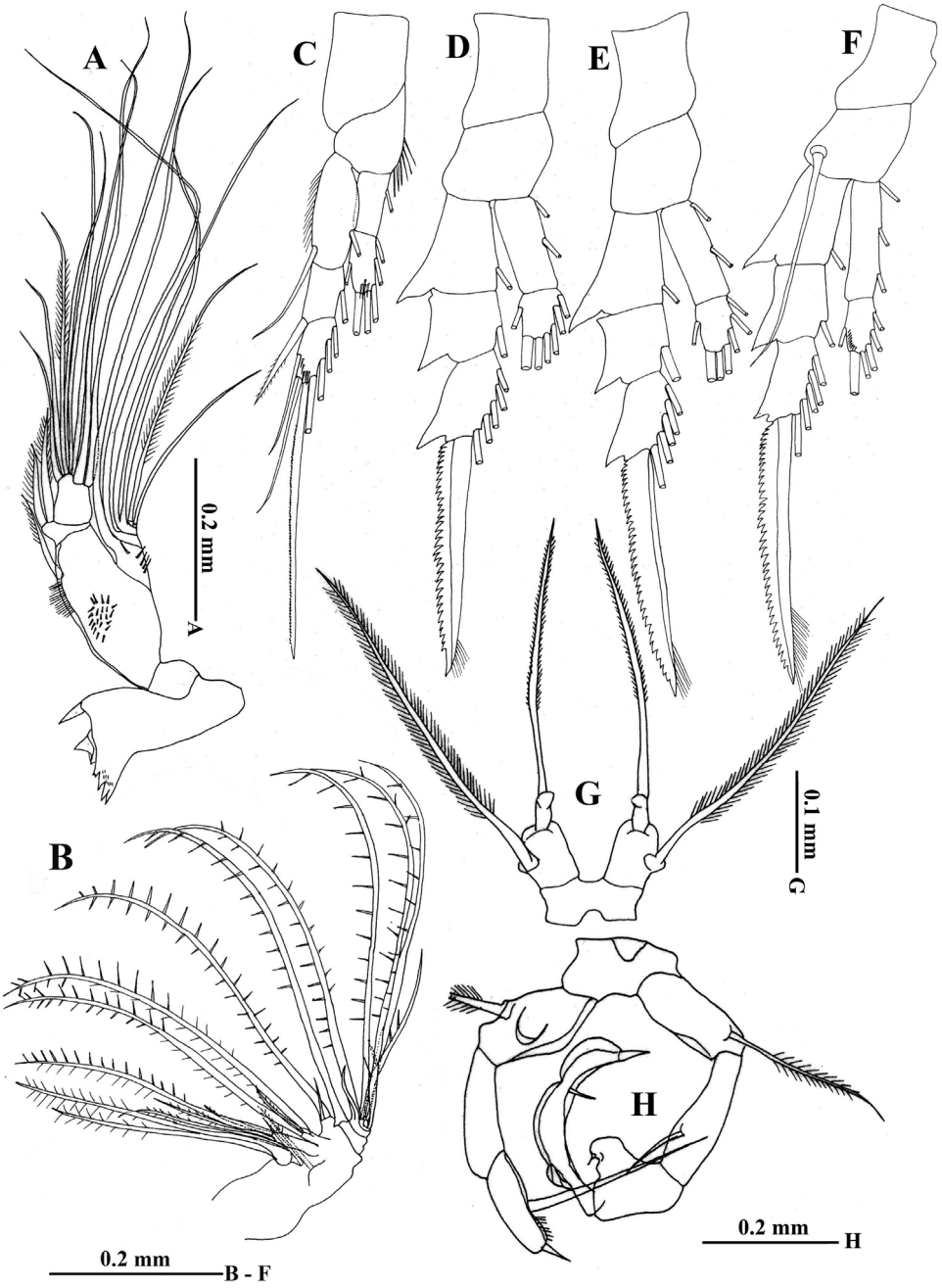

Figure 3. *Acartia* (*Odontacartia*) *edentata* sp. n. female (**A–G**) and male (**H**) (holotype) **A** Mandible **B** Maxilla **C** Leg 1 **D** Leg 2 **E** Leg 3 **F** Leg 4 **G** Legs 5 **H** Legs 5.

three rows of minute spinules ventrolaterally (Figs 4B, 5F); third urosomite with pair of strong acute processes dorsally (Figs 4 A, 5C, D); fourth urosomite 4.5 times shorter than wide and furnished with pair of small medium-sized acute processes dorsally; anal

somite with setules along outer margins. Caudal rami symmetrical, approximately 1.5 times as long as wide, having lateral setules along inner margin (Fig. 5D) and 6 plumose (II–VII) setae as in female.

Left antennule (Fig. 4C) incompletely 21-segmented; segments 2, 3 and 21 incompletely fused; armature elements and fusion pattern as follows (Roman numerals represent ancestral segments): I = 1, II–V = 3 + ae, XI–IX = 4 + ae, X= 1 + (1 spiniform), XI= 2 + ae, XII = 0, XIII = 0, XIV= 1 + (1 spiniform) + ae, XV = 1, XVI = 1 + ae, XVII = 1, XVIII = 1 + ae, XIX = 1, XX= 1, XXI= 1 + ae, XXII = 1, XXIII = 1, XXIV = 2, XXV = 2 + ae, XXVI = 2, XXVII–XXVIII = 4 + ae.

Right antennule (Fig. 4D) geniculate, incompletely 17–segmented, not reaching beyond posterior end of fifth pedigerous somite; segments 2 and 3 incompletely fused; segments 2, 3, 4, 7, 9, 15, 17 each with aesthetasc (ae); segments 4, 7 and 12 each with spiniform element. Armature elements and fusion pattern as follows (Roman numerals represent ancestral segments): I = 1, II–VI = 3 + (1 minute) + ae, VII–VIII = 2 + ae, IX–XI = 2 (2 spiniforms) + ae, XII = 0, XIII = 1 minute, XIV = 1 (1 spiniform) + ae, XV = 1, XVI = 1 + ae, XVII = 1, XVIII = 1, XIX = 1 + (1 spiniform), XX = 1 + longitudinal row teeth, XXI–XXIII = 3 + longitudinal row teeth, XXIV–XXV = 3+ ae, XXVI = 2, XXVII–XXVIII = 4 + ae.

Leg 5 (Fig. 3H) uniramous, coxae unarmed and completely fused with intercoxal sclerite; each side of basis with outer plumose seta subterminally, left basis approximately 2.5 times as long as wide, with concave inner margin. Right exopod 3-segmented, first segment approximately 3 times as long as wide with single seta subterminally, second segment with small subterminal spine at mid-length of inner irregularly triangular knob, third segment curved inward with small terminal spine and small inner spine midway. Left exopod 2-segmented, first segment about 2.5 times as long as wide, second segment with long inner seta and small terminal spine.

SEM observation of *Acartia* (*Odontacartia*) *edentata* sp. n. The absence of paired dorsal processes on the female genital double-somite and the thick rostrum were confirmed with scanning electron microscopy (Fig. 5 A, D). Paired genital slits are located at midlength and moderately separated (Fig. 5B, C). A row of setules is located along the anterior margin of each genital slit (Fig. 5C).

SEM observations of the male urosomite clearly showed fine ornamentation on the posterior border of first urosomite (Fig. 5E) and second segment furnished with three rows of minute spinules ventrolaterally (Fig. 5F). No fine setules were observed along the inner posterior margin of prosome, as described in the above descriptions of the type specimens.

Remarks. The subgenus *Acartia* (*Odontacartia*) is composed of two species groups, the *centrura* and *erythraea* species groups (Steuer 1923). *Acartia* (*O.*) *lilljeborgi* is regarded as an intermediate type between the *centrura* and *erythraea* species groups (Steuer 1923; Ueda 1986). The *centrura* species group is defined as follows: in the female, the genital double-somite with a pair of large processes, the first antennulary segment without a large spine, relatively long caudal rami, the exopod of leg 5 with a knob situated or extending to midlength; in the male, the third and fourth urosomites have a dorsal pair of large acute processes, the first exopodal segment of left leg 5 without an outer spine.

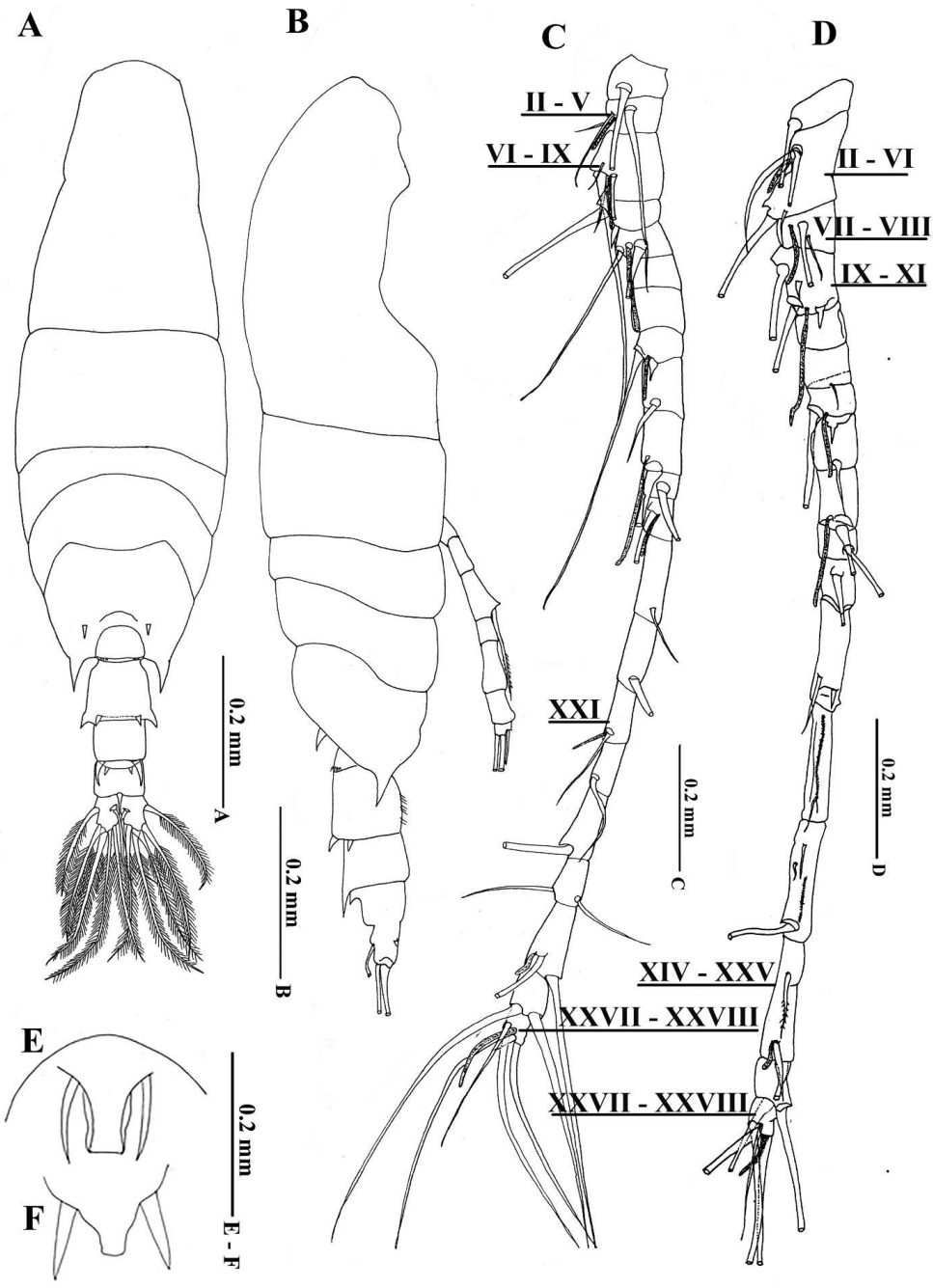

Figure 4. *Acartia* (*Odontacartia*) *edentata* sp. n., female (**F**) and male (**A–E**) (allotype) **A** Habitus, dorsal view **B** Habitus, lateral view **C** Antennule, Roman numerals denote segment numbers **D** Antennule, Roman numerals denote segment numbers **E** Rostrum of male **F** Rostrum of female.

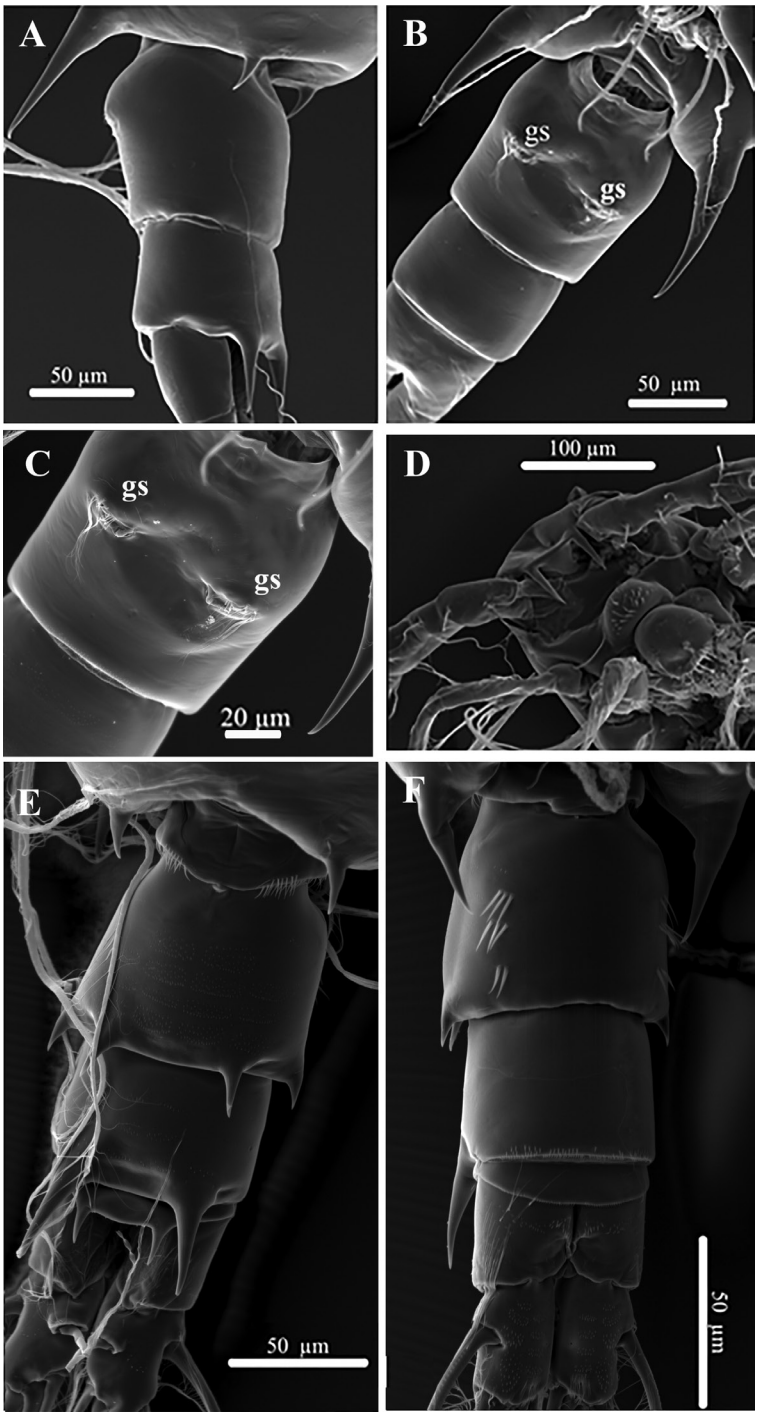

Figure 5. SEM micrographs of *Acartia* (*Odontacartia*) *edentata*, sp. n., female (**A–D**) and male (**E, F**) **A** The genital double-somite lateral view **B** Urosomite, ventral view **C** Genital double-somite, ventral view **D** Rostrum, ventral view **E** Urosomite, dorsal view **F** Urosomite, ventral view.

Among the *centrura* species, the female of *A.* (*O.*) *edentata* sp. n. is unique in lacking paired posterior dorsolateral processes on the genital double-somite unlike those of the closely related *A.* (*O.*) *pacifica* (Table 2). Such an absence can also be found in females of *A.* (*O.*) *bowmani* from India (Abraham 1976), but the morphology of posterior prosome and fifth legs of both sexes of *A.* (*O.*) *bowmani* differ from that of *A.* (*O.*) *edentata* sp. n.: (1) posterior prosomal border of female and male rounded with one pair of medium spines and one pair of small spines dorsally, (2) posterior margin of antennule with small spines on segments 4, 5, 10, 11 and 13 in female, (3) exopod in female fifth leg bulbous basally, (4) first segment of male urosomite bilobed and with fine setules on lateral margins, (5) second segment of right exopod of male fifth leg with quadrilateral shape of inner lobe, and (6) second segment of left exopod of male fifth leg with short seta and short segment. Irrespective of the presence or absence of the dorsal processes on the female genital double-somite, females of the new species and *A.* (*O.*) *pacifica* share the following features: (1) moderately long caudal rami (ca 2.7 times as long as wide), (2) the presence of a basal knob on the exopod of leg 5. Males of these species are also characterized together as follows: (1) the fine setules along posterior margin of first urosomite, (2) the presence of ventrolateral rows of minute spinules on the second urosomite laterally, and (3) dorsal processes on the third urosomite twice as long as those on the fourth urosomite.

Since "*A.* (*O.*) *pacifica*" s.l. morphologically and genetically consists of several cryptic species (Ueda and Bucklin 2006; Srinui et al. unpublished), we genetically compared specimens obtained from Japan (the Seto Inland Sea) and South Korea near the type localities to the new species (see "Molecular diversity" below). In conclusion, our Japanese and Korean specimens of *A.* (*O.*) *pacifica* clearly coincided with *A.* (*O.*) *pacifica* s.s. as morphologically/genetically redefined by Ueda and Bucklin (2006). Therefore, a comparison is made between the new species from the Philippines and *A.* (*O.*) *pacifica* s.s. obtained from Japan and South Korea in the present study. In addition to the absence of dorsal processes on the genital double-somite, females of the new species are distinguished from those of *A.* (*O.*) *pacifica* s.s. by: (1) segment 5 (VII) of right antennule with 1 seta (absent in *A.* (*O.*) *pacifica* s.s.), (2) dorsal processes on the second urosomite nearly reaching the posterior border of the anal somite (at most half the length of anal somite in *A.* (*O.*) *pacifica* s.s.), (3) length ratio of lateral seta of the basis to terminal seta of leg 5 is relatively short, about 1.3 (ca 2 in *A.* (*O.*) *pacifica* s.s.), (4) mandibular processes on gnathobase 1 blunt and 2 cuspidate (1 blunt and 6 cuspidate in *A.* (*O.*) *pacifica* s.s.) (Table 2). Males of the new species are differentiated from those of *A.* (*O.*) *pacifica s.s.* by: (1) Dorsal and lateral spines on the second somite are of medium-sized (longer in *A.* (*O.*) *pacifica* s.s.), (2) dorsal processes of the third urosomite are long enough to reach beyond those of the fourth urosomite (not reaching in *A.* (*O.*) *pacifica* s.s.), (3) terminal exopod segment of left leg 5 with an inner seta inserted midway (subterminally in *A.* (*O.*) *pacifica* s.s.), and (4) medial projection of second exopodal segment of right leg 5 with an inner irregularly triangular knob (rounded triangular in *A.* (*O.*) *pacifica* s.s. (Table 2).

Table 2. Differences in morphological characteristics among *Acartia* (*Odontacartia*) *edentata* sp. n., *A.* (*O.*) *pacifica* from Japan and Korea, and *A.* (*O.*) *ohtsukai*.

Features	***A.* (*O.*) *edentata* sp. n.**	***A.* (*O.*) *pacifica* (Japan and Korea)**	***A.* (*O.*) *ohtsukai****
Female			
Setae on segment (5) VII of right antennule	1 seta	Absent	1 seta
Paired posterior dorsolateral processes on the genital double-somite	Absent	Present	Present
Length of dorsal processes on the second urosomite relative to the posterior border of the anal somite	Reaching posterior border of anal somite	Half of length	Half of length
Length ratio of lateral to terminal setae of leg 5	1.3	2	1
Mandibular processes	1 blunt and 2 cuspidate	1 blunt covered with chitosan and 6 cuspidate	5 cuspidate
Male			
Length of dorsal and lateral spines on second somite	Medium	Long	Short
Presence of three rows of spinules ventrolaterally on second somite	Present	Present	Absent
Dorsal processes of third urosomite long enough to reach beyond those of fourth urosomite	Reaching	Not reaching	Not reaching
Insertion of inner seta on second exopod segment of left leg 5	Midway	Subterminal	Subterminal
Shape of medial projection on second exopodal segment of right leg 5	Irregular triangular	Rounded triangular	Quadrate

*According to Ueda and Bucklin, 2006.

Ueda and Bucklin (2006) described both left and right antenules of male *A.* (*O.*) *ohtsukai*, and we can compare setation in the new species as follows: Right: (3) VII–VIII–2 + ae (3 and 1 ae in *A.* (*O.*) *ohtsukai*), (4) IX–XI–2 (2 spiniform) + ae (plus 2 spinules in *A.* (*O.*) *ohtsukai*), (6) XII–1 minute (minute absent in *A.* (*O.*) *ohtsukai*), (7) XIV–1 (1 spiniform) + ae (plus 1 spinule in *A.* (*O.*) *ohtsukai*), (11) XVIII–1 (plus 1 ae in *A.* (*O.*) *ohtsukai*), (12) XIX–1 (spiniform) (plus longitudinal row teeth in *A.* (*O.*) *ohtsukai*), (14) XXI–XXIII–3 + longitudinal row teeth (longitudinal row teeth absent in *A.* (*O.*) *ohtsukai*), (15) XXIV–XXV–3 + ae (plus 1 spinule in *A.* (*O.*) *ohtsukai*). The segmentation and setation of the right antennule of *A.* (*O.*) *ohtsukai* are alternately interpreted as follows: 16-segmented with those of (1) I–1, (2) II–VI–4 + ae, (3) VII–VIII–3 + ae, (4) IX–XI–4 (2 spiniforms) + ae, (5) XII–unarmed, (6) XIII–unarmed, (7) XIV–2 (1 spiniform) + ae, (8) XV–1, (9) XVI–1 + ae, (10) XVII–1, (11) XVIII–1 + ae, (12) XIX–1 + process, (13) XX–1 (14) XXI–XIII–4, (15) XXIV–XXV–4 + ae, (16) XXVI–2 (17) XXVII–XXVIII–4 + ae. The segmentation and setation of the left antennule are similar in the *A.* (*O.*) *edentata* sp. n. and *A.* (*O.*) *ohtsukai*.

The taxonomy of the Indo-West Pacific *A.* (*O.*) *pacifica* should be revised, because the presence of several cryptic species has already been suggested by our study and oth-

ers. The above-mentioned sexual dimorphic features are species-specific, and should be carefully compared among *A.* (*O.*) *pacifica* s.l. to resolve the issue (see Discussion).

Etymology. The new species of *Acartia* was named *edentata* (Latin, meaning toothless) with reference to the absence of tooth-like processes on the posterodorsal border of the genital double-somite in females.

Genetic diversity. We obtained sequence data from mitochondrial 16S and COI genes for 14 individual specimens at three sites. A 282 bp fragment of the 16S gene was analyzed for five adult female specimens from the Philippines (*A.* (*O.*) *edentata* sp. n.), and a 162 bp 16S fragment was analyzed for *A.* (*O.*) *pacifica* specimens from Ariake Bay, the Seto Inland Sea, Japan and South Korea. A 636 bp fragment of the mitochondrial COI gene was analyzed in the new species and in specimens from Japan and Korea. GenBank sequences for *A.* (*O.*) *pacifica* (accession number DQ071175 for 16S and DQ071177 for COI) and two out group species of subgenus *A.* (*Odontacartia*), *A.* (*O.*) *ohtsukai* (accession number DQ071174 for 16S and DQ071176 for COI), *A.* (*O.*) *erythraea* (accession number DQ320504 for 16S) and *A.* (*O.*) *tsuensis* (accession number KC287427 for COI), were also used for comparison. The intraspecific variation in the 16S sequences from the five *A.* (*O.*) *edentata* sp. n. individuals was 0%, whereas *A.* (*O.*) *edentata* sp. n. sequences differ from those of *A.* (*O.*) *pacifica* from Japanese and Korean waters, *A.* (*O.*) *pacifica* based on GenBank, *A.* (*O.*) *ohtsukai*, and *A.* (*O.*) *erythraea* by 20–21%, 20–21%, 28%, and 31%, respectively. The COI sequences from *A.* (*O.*) *edentata* sp. n. individuals differ by only 0.02–0.08%; *A.* (*O.*) *pacifica* (from Japan, and Korea), *A.* (*O.*) *pacifica* (GenBank), *A.* (*O.*) *ohtsukai*, and *A.* (*O.*) *tsuensis* sequences differ from *A.* (*O.*) *edentata* sp. n. COI sequences by 16–18%, 16–17%, 16–17%, 22%, and 24%, respectively (Fig. 6; Table 3).

Ecology. Temperature and salinity appear to be important factors determining the distribution and abundance of copepods. In the Indo-West Pacific, *A.* (*O.*) *pacifica* occurs in the tropical and subtropical zones of the Pacific and Indian oceans. In the East China Sea (subtropical zone), *A.* (*O.*) *pacifica* was abundant in August (salinity 15.0) in the Changjiang (Yangtze River) Estuary, China (Gao et al. 2008), while in Korean waters, *A.* (*O.*) *pacifica* is strictly stenohaline, occurring waters of more than 32 in salinity (Moon et al. 2008). Kang (2011) also observed the *A.* (*O.*) *pacifica* and *A.* (*O.*) *erythraea* in Korean waters with temperature ranges of 18.0–27.2 °C and 14.6–26.4 °C and salinity ranges of 21.0–32.9 and 21.0–33.7, respectively. In Japanese waters *A.* (*O.*) *ohtsukai* was found in the estuary of the Rokkaku River, Ariake Bay in surface waters, where water temperature was 29.0 °C and salinity was 5.0, while *A.* (*O.*) *pacifica* was found in waters of 26.0 °C and 33.0 in the Seto Inland Sea, Japan (Ueda and Bucklin 2006). Furthermore, *A.* (*O.*) *pacifica* was dominant in Moreton Bay, Queensland waters with temperature ranges above 22.0 °C and salinities ranging from 34.0 to 36.5 (Greenwood 1981).

In the tropical zone, *A.* (*O.*) *edentata* sp. n. specimens were collected in the Philippines during the rainy season (August 2013), when water temperature and salinity were 30.2 °C and 33.5, respectively. In contrast, *A.* (*A.*) *tsuensis* represents the dominant species in brackish pond water from Panay Island in central Philippines during the dry season (November – April), with salinity ranging from 14.0 to 40.0 (Golez et al. 2002).

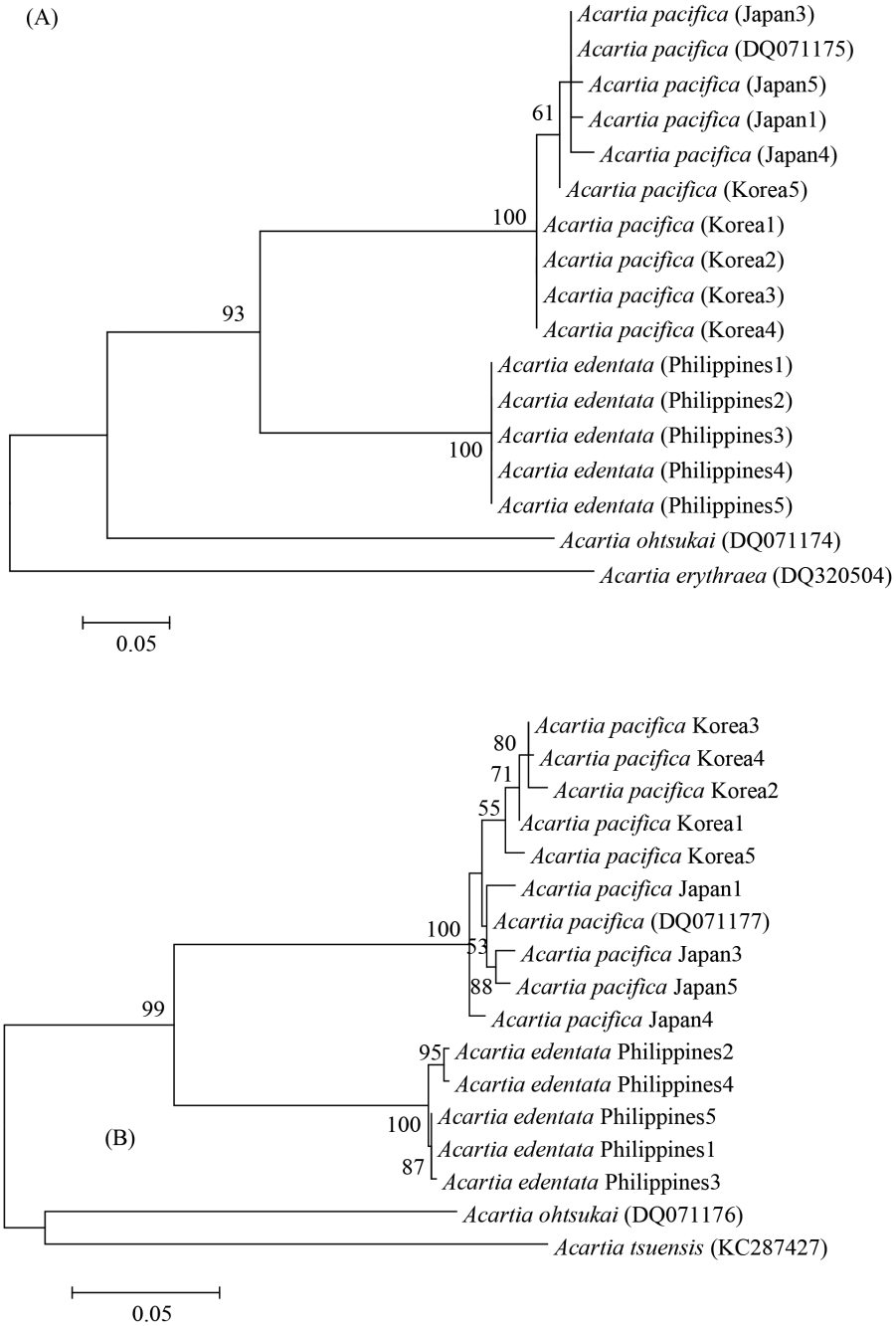

Figure 6. Neighbor-joining phylogenetic tree based on the mitochondrial 16S (**A**) and COI (**B**) genes of *Acartia* (*Odontacartia*) *edentata* sp. n., and *A.* (*O.*) *pacifica* from Ariake Bay in the Seto Inland Sea and Korean waters. *A.* (*O.*) *erythraea*, *A.* (*O.*) *ohtsukai* and *A.* (*O.*) *tseuensis* sequences from GenBank were used as outgroups. Bootstrap values (percentage) are shown for nodes with support > 50%. Supporting valves of each node obtained from 1,000 bootstrap replications.

Table 3. Pairwise differences for 16S and COI sequences between individual females of *Acartia* (*Odontacartia*) *edentata* sp. n. from Leyte Island, the Philippines; *A.* (*O.*) *pacifica* from Ariake Bay, Seto Inland Sea (GenBank accession no. DQ071175 for 16S and DQ071177 for COI); *A.* (*O.*) *pacifica* from South Korea and Japan (Seto Inland Sea); *A.* (*O.*) *ohtsukai* from the Rokkaku River Estuary, Ariake Bay (GenBank accession no. DQ071174 for 16S and DQ071176 for COI); *A.* (*O.*) *erythraea* (GenBank accession no. DQ320504 for 16S); and *A.* (*A.*) *tsuensis* (GenBank accession no. KC287427 for COI).

	16S	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
1	*Acartia edentata*																
2	*Acartia edentata*	0.000															
3	*Acartia edentata*	0.000	0.000														
4	*Acartia edentata*	0.000	0.000	0.000													
5	*Acartia edentata*	0.000	0.000	0.000	0.000												
6	*Acartia pacifica* (Japan1)	**0.206**	**0.206**	**0.206**	**0.206**	**0.206**											
7	*Acartia pacifica* (Japan3)	**0.206**	**0.206**	**0.206**	**0.206**	**0.206**	0.006										
8	*Acartia pacifica* (Japan4)	**0.219**	**0.219**	**0.219**	**0.219**	**0.219**	0.019	0.013									
9	*Acartia pacifica* (Japan 5)	**0.212**	**0.212**	**0.212**	**0.212**	**0.212**	0.013	0.006	0.019								
10	*Acartia pacifica* (Korea1)	**0.200**	**0.200**	**0.200**	**0.200**	**0.200**	0.025	0.019	0.031	0.025							
11	*Acartia pacifica* (Korea2)	**0.200**	**0.200**	**0.200**	**0.200**	**0.200**	0.025	0.019	0.031	0.025	0.000						
12	*Acartia pacifica*(Korea3)	**0.200**	**0.200**	**0.200**	**0.200**	**0.200**	0.025	0.019	0.031	0.025	0.000	0.000					
13	*Acartia pacifica* (Korea4)	**0.200**	**0.200**	**0.200**	**0.200**	**0.200**	0.025	0.019	0.031	0.025	0.000	0.000	0.000				
14	*Acartia pacifica* (Korea5)	**0.212**	**0.212**	**0.212**	**0.212**	**0.212**	0.013	0.006	0.019	0.013	0.013	0.013	0.013	0.013			
15	*Acartia pacifica* (DQ 071175)	0.206	0.206	0.206	0.206	0.206	0.006	0.000	0.013	0.006	0.019	0.019	0.019	0.019	0.006		
16	*Acartia ohtsukai* (DQ071176)	0.287	0.287	0.287	0.287	0.287	0.287	0.281	0.281	0.287	0.275	0.275	0.275	0.275	0.281	0.281	
17	*Acartia erythraea* (DQ320504)	0.313	0.313	0.313	0.313	0.313	0.356	0.350	0.338	0.350	0.344	0.344	0.344	0.344	0.350	0.350	0.331

Table 3. Continued.

	COI	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
1	*Acartia edentata*																
2	*Acartia edentata*	0.008															
3	*Acartia edentata*	0.002	0.009														
4	*Acartia edentata*	0.008	0.003	0.009													
5	*Acartia edentata*	0.000	0.008	0.002	0.008												
6	*Acartia pacifica* (Japan1)	**0.173**	**0.178**	**0.175**	**0.178**	**0.173**											
7	*Acartia pacifica* (Japan3)	**0.176**	**0.181**	**0.178**	**0.181**	**0.176**	0.019										
8	*Acartia pacifica* (Japan4)	**0.164**	**0.165**	**0.165**	**0.165**	**0.164**	0.017	0.020									
9	*Acartia pacifica* (Japan5)	**0.175**	**0.180**	**0.176**	**0.180**	**0.175**	0.017	0.011	0.019								
10	*Acartia pacifica* (Korea1)	**0.162**	**0.167**	**0.164**	**0.167**	**0.162**	0.024	0.024	0.022	0.022							
11	*Acartia pacifica* (Korea2)	**0.165**	**0.170**	**0.167**	**0.170**	**0.165**	0.030	0.030	0.028	0.028	0.009						
12	*Acartia pacifica*(Korea3)	**0.162**	**0.167**	**0.164**	**0.167**	**0.162**	0.024	0.024	0.022	0.003	0.022	0.006					
13	*Acartia pacifica* (Korea4)	**0.164**	**0.169**	**0.165**	**0.169**	**0.164**	0.025	0.025	0.024	0.024	0.005	0.008	0.002				
14	*Acartia pacifica* (Korea5)	**0.161**	**0.165**	**0.162**	**0.165**	**0.161**	0.025	0.025	0.024	0.024	0.011	0.020	0.014	0.016			
15	*Acartia pacifica* (DQ 071177)	0.167	0.172	0.169	0.172	0.167	0.009	0.009	0.011	0.008	0.014	0.020	0.014	0.016	0.016		
16	*Acartia ohtsukai* (DQ071176)	0.220	0.220	0.222	0.220	0.220	0.241	0.241	0.233	0.239	0.231	0.235	0.231	0.233	0.235	0.231	
17	*Acartia tsuensis* (KC287427).	0.243	0.244	0.244	0.244	0.243	0.247	0.252	0.244	0.249	0.250	0.249	0.247	0.249	0.252	0.243	0.247

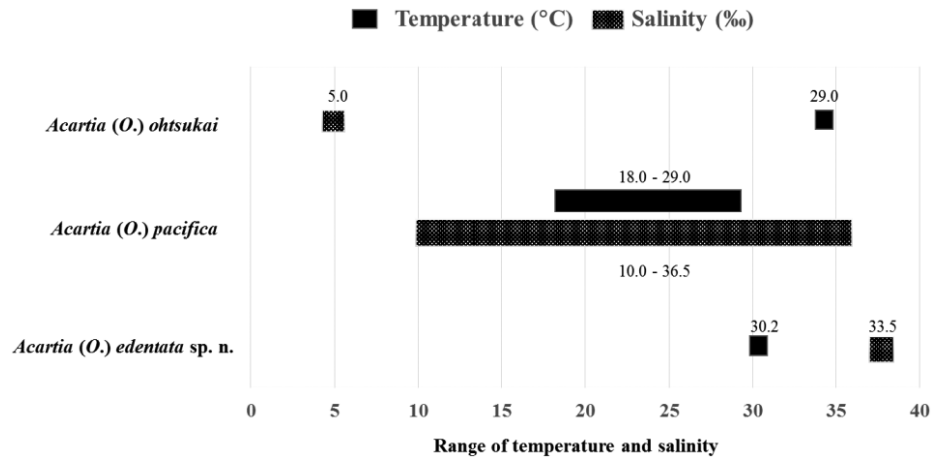

Figure 7. Ranges of salinity and temperature of three *Acartia* species occurring in the tropical and subtropical zones of the Pacific and Indian oceans.

In Bintulu, Sarawak, Malaysia, Johan et al. (2013) compared to *A.* (*O.*) *pacifica* in coastal waters with temperatures of 28.8–29.0 °C and high salinities (24.0–32.0). In the Indian waters, Wellershaus (1969) recorded the occurrence of female specimens of *A.* (*O.*) *pacifica* in waters with salinity ranging from 10.0 to 30.0, including in the Andaman Sea, Thailand (Surin Islands National Park, Phang Nga Province), *A.* (*O.*) *pacifica* were abundant in waters with temperatures of 29.7–31.0 °C and salinities ranging from 29.9 to 35.8 (Treeramaethee et al. 2013). However, we concluded that three species of *Acartia* appear to occupy water bodies differing in temperature and salinity of the tropical and subtropical zone of the Pacific and Indian oceans (Fig. 7).

Discussion

Prior to the current study, it was believed that *A.* (*O.*) *pacifica* was represented by a single species with a wide geographic range occupying the coastal brackish waters throughout the Western Pacific and Indian oceans (Früchtl 1923; Sewell 1933; Shen and Lee 1963; Chen and Zhang 1965; Tanaka 1965; Brodsky 1967; Wellershaus 1969; Pillai 1971; Pinkaew 2003; Mulyadi 2004; Ueda and Bucklin 2006; Shang et al. 2007; Gao et al. 2008; Lan et al. 2008; Moon et al. 2008; Phukham 2008; Kang 2011; Treeramaethee et al. 2013). From the findings of the present study, *A.* (*O.*) *edentata* sp. n. is described based on morphological features that permit its discrimination from *A.* (*O.*) *pacifica* in the West Pacific Ocean. Ueda and Bucklin (2006), suggested that the features provided strong evidence specific to the habitats occupied by species and could be used to discriminate *A.* (*O.*) *pacifica*, which occupies neritic waters, from *A.* (*O.*) *ohtsukai*, which occupies brackish waters.

Mitochondrial markers within the 16S and COI genes have proved to be of great utility in investigating the systematics of ecologically and geographically isolated popu-

lations of calanoid copepods (Bucklin et al. 2003). The molecular-based analyses using 16S and COI sequences in the current study lend good support to the morphology-based findings. The findings suggest two major clades that reflect the geographic distribution of *Acartia* in the Indo-West Pacific, i.e. *A.* (*O.*) *edentata* sp. n. and *A.* (*O.*) *pacifica* from the Seto Inland Sea and from Korean waters, and *A.* (*O.*) *ohtsukai* from Ariake Bay, Japan. Our sequencing results agree with those of Ueda and Bucklin (2006) and emphasize the wide divergence between *A.* (*O.*) *ohtsukai* from *A.* (*O.*) *pacifica*. The high level of sequence divergence observed in this study indicates that the Philippines Islands serve as a barrier limiting the spread of *A.* (*O.*) *pacifica* populations into the Philippine Archipelago. This supports the allopatric speciation hypothesis of Carpenter and Springer (2005) that an ecological vicariant seems to have blocked the migration of marine organisms in the Pleistocene from the West Pacific Ocean to the Indian Ocean. Srinui and Ohtsuka (2015) showed that the distribution patterns of 11 species of *Acartiella* could be separated into those inhabiting the West Pacific and those in the Indian Ocean. In the case of *A.* (*O.*) *pacifica* s.l., more studies on inter- and intra-specific molecular and morphological variation found in specimens collected from Asian waters are needed to further understand the distribution and evolution of sibling species in the West Pacific region.

Key to species of the subgenus *Acartia* (*Odontacartia*)

Thirteen species of the subgenus *Acartia* (*Odontacartia*), including *A.* (*O.*) *edentata* sp. n., have been described from the Indo-West Pacific (Razouls et al. 2018; present study), and are divided into three groups: the *centrura* and *erythraea* species groups and *A.* (*O.*) *lilljeborgi* (Steuer 1923; Ueda 1986). Key to species is provided below for both sexes of the subgenus *Acartia* (*Odontacartia*).

Female

1 Genital double-somite lacking posterodorsal sharp processes **2**
– Genital double-somite having paired posterodorsal processes **3**
2 Ventroposterior corners of prosome acutely pointed, reaching beyond half of genital double-somite..... ***A. (O.) edentata* sp. n.**
– Ventroposterior corners of prosome round with pair of acutely pointed processes not reaching beyond half of genital double-somite ... ***A. (O.) bowmani***
3 Second segment of antennule with strong curved processes posteriorly **4**
– Second segment of antennule without strong curved processes **5**
4 First antennule segment with two large processes terminally ..... ***A. (O.)bispinosa* Carl, 1907**
– First antennule segment lacking processes ..... ***A. (O.) spinicauda***
5 Exopod of leg 5 thickened proximally **6**
– Exopod of leg 5 not thickened proximally **8**

6 Exopod of leg 5 thickened proximally extending midway along exopod..... ***A. (O.) centrura***

– Exopod of leg 5 with thickened proximal part confined to base of exopod..**7**

7 Length ratio of outer basal setae to exopod of leg 5: ca 2..... ***A (O.) pacifica***

– Length ratio of outer basal setae to exopod, leg 5: ca 1..... ***A. (O.) ohtsukai***

8 Caudal ramus longer than wide by at most ca 2 times; second free urosomite with small spinules dorsally and posteriorly **9**

– Caudal ramus longer than wide by ca 3 times; second free urosomite lacking small dorsal spinules ..... ***A.(O.) mertoni***

9 Fifth to seventh antennule segments each with posterior hook; genital double-somite with two pairs of small processes dorsally..... ***A. (O.) lilljeborgi***

– Fifth to seventh antennule segments each lacking hook posteriorly; genital double-somite with pair of small processes dorsally.....**10**

10 First antennule segment with 2 or more strong processes distally **11**

– First antennule segment with single strong process distally.....**12**

11 Second antennule segment with single spinule posteriorly .... ***A. (O.) erythraea***

– Second antennule segment with 4 spinules posteriorly..... ***A. (O.) amboinensis* Carl, 1907**

12 Caudal ramus with 4–6 rows of minute spinules dorsally..... ***A. (O.) japonica* Mori, 1940**

– Caudal ramus lacking of dorsal rows of spinules..... ***A. (O.) australis* Farran, 1936**

Male

1 Urosomite 3 with large spine-like processes dorsally.....**2**

– Urosomite 3 without spine-like processes dorsally.....**9**

2 Dorsal processes of urosomite 3 long, reaching half-length of anal somite...**3**

– Dorsal processes of urosomite 3 short, reaching posterior-most border of urosomite 4.....**6**

3 Urosomite 4 with four spine-like processes between pair of dorsal processes.... ***A. (O.) spinicauda***

– Urosomite 4 lacking spine-like processes between pair of dorsal processes...**4**

4 Genital somite lacking spinular rows along posterodorsal border ..... ***A. (O.) pacifica***

– Genital somite with spinular rows along posterodorsal border.....**5**

5 Inner projection of first exopodal segment of right leg 5 quadrate..... ***A. (O.) mertoni***

– Inner projection of first exopodal segment of right leg 5 irregularly triangular.... ***A. (O.) edentata* sp. n.**

6 Urosomites 3 and 4 each with two prominences between pair of dorsal processes..... ***A. (O.) centrura***

– Urosomites 3 and 4 each lacking prominences between pair of dorsal processes **7**

7 Inner seta of terminal exopodal segment of left leg 5 longer than terminal segment ..... ***A. (O.) ohtsukai***
– Inner seta of terminal exopodal segment of left leg 5 nearly equal to terminal segment ..... ***A. (O.) bowmani***
8 Urosomite 4 without prominences dorsally ..... ***A. (O.) australis***
– Urosomite 4 with prominences dorsally **9**
9 Number of dorsal prominences on urosomite 4 fewer than five **10**
– Number of prominences on urosomite more than seven **12**
10 Terminal exopodal segment of left leg 5 with three elements ..... ***A. (O.) erythraea***
– Terminal exoposal segment of left leg 5 with single element **11**
11 Terminal element of left leg 5 spiniform ..... ***A. (O.) amboinensis***
– Terminal element of left leg 5 as fine seta ..... ***A. (O.) lilljeborgi***
12 Terminal elements of left leg 5 as three small prominence .... ***A. (O.) japonica***
– Terminal elements of left leg 5 as two spines ..... ***A. (O.) bispinosa***

Acknowledgements

We express our sincere thanks to Dr Mitsuyasu Moriya for his help with field sampling at Leyte Island in the Philippines. We are also grateful to Dr Wansuk Senanan of Burapha University for guidance on molecular techniques. Thanks are also due to Professor Ho Young Soh (Chonnan National University, Korea) who kindly provided us with specimens of *Acarita* (*Odontoacartia*) *pacifica* collected from Korea. This study was partially supported by grants from the Asian CORE-to-CORE and RONPAKU Programs of the Japan Society for the Promotion of Science (JSPS), a KAKEN grant from the JSPS (awarded to SO, 16K07825), and by a grant from Burapha University (to KS, 1/2559).

References

Abraham S (1976) A new calanoid copepod of the genus *Acartia* from the Cochin Backwaters, India, and a redescription of *Acartia centrura* Giesbrecht. Crustaceana 30(1): 73–81. https://doi.org/10.1163/156854076X00422

Boxshall GA, Huys R (1998) The ontogeny and phylogeny of copepod antennules. Philosophical Transactions of the Royal Society of London 353: 765–786. https://doi.org/10.1098/rstb.1998.0242

Boxshall GA, Halsey SH (2004) An Introduction to Copepod Diversity. The Ray Society, London, 421 pp.

Brodsky KA (1967) Calanoida of the Far Eastern Seas and Polar basin of the U.S.S.R. Keys to the Fauna of the USSR 35: 1–440.

Bucklin A, Bentley AM, Franzen SP (1998) Distribution and relative abundance of the copepod, *Pseudodiaptomus moultoni* and *P. newmani*, on Georges Bank based on molecular identification of sibling species. Marine Biology 132: 97–106. https://doi.org/10.1007/s002270050375

Bucklin A, Frost BW, Bradford-Grieve J, Allen LD, Copley NJ (2003) Molecular systematic and phylogenetic assessment of 34 calanoid copepod species of the Calanidae and Clausocalanidae. Marine Biology 142: 333–343. https://doi.org/10.1007/s00227-002-0943-1

Carl J (1907) Copépodes d'Amboine. Revue Suisse Zoologie 15: 7–18.

Carpenter KE, Springer VG (2005) The center of the center of marine shore fish biodiversity: the Philippine Islands. Environmental Biology of Fishes 72: 467–480. https://doi.org/10.1007/s10641-004-3154-4

Chen QC, Zhang SZ (1965) The planktonic copepods of the Yellow Sea and the East China Sea. I. Calanoida. Studia Marina Sinica 7: 20–131. [pl. 55]

Dana JD (1846) Notice of some genera of Cyclopacea. Annals and Magazine of National History 18: 181–185. https://doi.org/10.1080/037454809494408

Farran GP (1936) Copepoda. Great Barrier Reef Expedition 1928–29, Scientific Reports 5. British Museum, London, 73–142.

Folmer O, Black M, Hoen W, Lutz R, Vrijenhoek R (1994) DNA primers for amplication of mitochondrial cytochrome *c* oxidace subunit I from diverse metozone invertebrates Molecular Marine Biology Biotechnology 3: 294–299.

Früchtl F (1923) Cladocera und Copepoda der Aru-Inseln. (Vorläufige Mitteilung: Artenliste und kurze Diagnosen der neuen Formen). Abhandlungen Senckenbergische Naturforschende Gesellschaft 375(4): 449–457.

Gao Q, Xu Z, Zhuang P (2008) The relation between distribution of zooplankton and salinity in the Changjiang Estuary. Chinese Journal of Oceanology and Limnology 26(2): 178–185. https://doi.org/10.1007/s00343-008-0178-1

Giesbrecht W (1889) Elenco dei Copepodi pelagic raccolti dal tenente di vascello Gaetano Chierchia durante il viaggio della R. Corvette "Vettor Pisani" negli anni 1882–1885 e dal tenente dal vascello Francesco Orsini nel Mar Roso, nel 1884. Atti della Reale Accademia Nazionale dei Lincei (4) 5(2): 25.

Golez MSN, Ohno A, Toledo JD, Tanaka Y, Ishimaru T (2002) Population dynamics of the calanoid copepod, *Acartia tsuensis* in a brackish-water pond in the Philippines. Fisheries Science 68 (Supplement 1): 341–344. https://doi.org/10.2331/fishsci.68.sup1_341

Greenwood JG (1978) Calanoid copepods of Moretan Bay (Queenland) III. Families. Temoridae to Tortanidae, excluding Pontellidae. Proceedings of the Royal Society of Queensland 89: 1–21.

Greenwood JG (1981) Occurrences of congeneric pairs of *Acartia* and *Pseudodiaptomus* species (Copepoda, Calanoida) in Moreton Bay, Queenland. Estuarine, Coastal and Shelf Science 13: 591–596. https://doi.org/10.1016/S0302-3524(81)80060-6

Hall TA (1999) Bioedit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95–98.

Huys R, Boxshall GA (1991) Copepod Evolution. The Ray Society, London, 468 pp.

Itô T (1956) Three new copepods from brackish-water lakes of Japan. Pacific Science 10: 468–473.

Johan I, Hena MKA, Idris MH, Arshad A (2013) Taxonomic composition and abundance of zooplankton copepod in the coastal water of Bintulu, Sarawak, Malaysia. Journal of Fisheries and Aquatic Science 8(3): 472–479. https://doi.org/10.3923/jfas.2013.472.479

Kang JH (2011) The occurrence of *Acartia* species and their environmental characteristics at three ports in Korea. Ocean Science Journal 46(4): 219–237. https://doi.org/10.1007/s12601-011-0018-7

Lan YC, Lee MA, Liao CH, Chen WY, Lee DA, Liu DC, Su WC (2008) Copepod community changes in the southern east China sea between the early and late northeasterly monsoon. Zoological Studies 47(1): 61–74.

Moon SY, Ohtsuka S, Ueda H, Soh HY (2008) *Acartia* (*Odontacartia*) *ohtsukai* Ueda and Bucklin, 2006 (Copepoda, Calanoida, Acartiidae): first record of its occurrence in Korean waters and habitat segregation from its sibling species *A. pacifica* Steuer, 1915. Zootaxa 1841: 61–64.

Mori (1940) Two new copepods from Japanese waters. Dobutsugaku Zasshi 52(8): 328–330.

Mulyadi (2004) Calanoid copepods in Indonesian waters. Research Center for Biology, Indonesia Institute of Science, Bogor, Indonesia, 198 pp.

Palumbi SR (1996) Nucleic acids II: the polymerase chain reaction. In: Hillis DM, Moritz C, Mable BK (Eds) Molecular Systematics. Sinauer & Associates, Sunderland, Massachusetts, 205–247.

Pillai PP (1971) Studies in the estuarine copepod of India. Marine Biology Association of India 13(2): 162–172.

Pinkaew K (2003) Taxonomy of copepods in the Bangpakong river estuary and the Sriracha coast of Thailand. M.Sc. Thesis, Burapha University, Chon Buri, Thailand, 111 pp.

Phukham N (2008) Species diversity of calanoid copepods in Thai waters, Andaman Sea. M.Sc. Thesis, Kasetsart University, Bangkok, Thailand, 247 pp.

Razouls C, De BF, Kouwenberg J, Desreumaux N (2018) Diversity and geographic distribution of marine planktonic copepods. http://copepodes.obs-banyuls.fr/en [Accessed on: 2015-8-9]

Saitou N, Nei M (1987) A neigbour-joining method: a new method for reconstruction of phylogenetic trees. Molecular Ecology and Evolution 44(4): 406–425. https://doi.org/10.1093/oxfordjournals.molbev.a040454

Sars GO (1903) An Account of the Crustacea of Norway, with Short Descriptions and Figures of all the Species. Vol. IV. Copepoda, Calanoida. Bergen Museum Norway, Bergen, 171 pp.

Sewell RBS (1933) Notes on a small collection of marine copepod from the Malay States. The Raffles Bulletin of Zoology 8: 25–30.

Shang X, Wang G, Li S, Guo D (2007) Preliminary study of the retention mechanism of planktonic copepods in the Juilong estuary in China. ACTA Oceanologica Sinica 26(4): 156–163.

Shen CJ, Lee FS (1963) The estuarine Copepoda of Chiekong and Zaikong rivers, Kwangtung Province, China. Acta Zoologica Sinica 15: 571–596. [In Chinese with English abstract]

Srinui K, Ohtsuka S (2015) Supplementary description of three *Acartiella* species (Crustacea, Copepoda, Calanoida) from estuarine waters in Thailand. Species Diversity 20: 167–181. https://doi.org/10.12782/sd.20.2.167

Steuer A (1915) Revision der Gattung *Acartia* Dana. Zoologischer Anzeiger 45: 392–397.

Steuer A (1917) Über eine Lokalrasse der *Acartia* (*Odontacartia*) *pacifica* Steuer von den Aru-Inseln. Abhandlungen Senckenbergische Naturforschende Gesellschaft 35: 255–257.

Steuer A (1923) Bausteine zu einer Monographie der Copepodengattung *Acartia*. Arbeiten aus dem Zoologischen Institut der Universität Innsbruck 1(5): 1–148.

Tanaka O (1965) The pelagic copepods of the IZU region, middle Japan systematic account XIII Parapontellidae, Acartiidae and Tortanidae. Publications of the Seto Marine Biological Laboratory 12(5): 520–549. https://doi.org/10.5134/175378

Tamura K, Stecher G, Peterson D, Filipski A, Kumar S (2013) MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. Molecular Biology and Evolution 30: 2725–2729. https://doi.org/10.1093/molbev/mst197

Treeramaethee J, Phukham N, Patarajinda S (2013) Species diversity of marine calanoid copepod from Surin Islands National Park with a description of a new record to Thailand. Journal of Fisheries Technology 7(1): 46–59. [In Thai]

Ueda H (1986) Taxonomic reexamination and geographic distribution of copepods known as *Acartia clausi* in Japanese coastal and inlet waters. Journal of the Oceanographical Society of Japan 42: 134–138. https://doi.org/10.1007/BF02109100

Ueda H, Bucklin AC (2006) *Acartia* (*Odontacartia*) *ohtsukai*, a new brackish-water calanoid copepod from Ariake Bay, Japan, with a redescription of the closely related *A. pacifica* from the Seto Inland Sea. Hydrobiologia 560: 77–91. https://doi.org/10.1007/s10750-005-9513-0

Wellershaus S (1969) On the taxonomy of planktonic Copepoda in the Cochin Backwater (a South Indian estuary). VerÖffentlichungen des Instituts für Meeresforschung in Bremerhaven 11: 245–286.