

# **Benthic Studies in Alice Arm and Hastings Arm, B.C. in Relation to Mine Tailings Dispersal**

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1983

**Canadian Technical Report of  
Hydrography and Ocean Sciences  
No. 22**



## **Canadian Technical Report of Hydrography and Ocean Sciences**

These reports contain scientific and technical information of a type that represents a contribution to existing knowledge but which is not normally found in the primary literature. The subject matter is generally related to programs and interests of the Ocean Science and Surveys (OSS) sector of the Department of Fisheries and Oceans.

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Regional and headquarters establishments of Ocean Science and Surveys ceased publication of their various report series as of December 1981. A complete listing of these publications and the last number issued under each title are published in the *Canadian Journal of Fisheries and Aquatic Sciences*, Volume 38: Index to Publications 1981. The current series began with Report Number 1 in January 1982.

## **Rapport technique canadien sur l'hydrographie et les sciences océaniques**

Ces rapports contiennent des renseignements scientifiques et techniques qui constituent une contribution aux connaissances actuelles mais que l'on ne trouve pas normalement dans les revues scientifiques. Le sujet est généralement rattaché aux programmes et intérêts du service des Sciences et Levés océaniques (SLO) du ministère des Pêches et des Océans.

Les rapports techniques peuvent être considérés comme des publications à part entière. Le titre exact figure au-dessus du résumé du chaque rapport. Les résumés des rapports seront publiés dans la revue Résumés des sciences aquatiques et halieutiques et les titres figureront dans l'index annuel des publications scientifiques et techniques du Ministère.

Les rapports techniques sont produits à l'échelon régional mais sont numérotés et placés dans l'index à l'échelon national. Les demandes de rapports seront satisfaites par l'établissement auteur dont le nom figure sur la couverture et la page de titre. Les rapports épuisés seront fournis contre rétribution par des agents commerciaux.

Les établissements des Sciences et Levés océaniques dans les régions et à l'administration centrale ont cessé de publier leurs diverses séries de rapports depuis décembre 1981. Vous trouverez dans l'index des publications du volume 38 du *Journal canadien des sciences halieutiques et aquatiques*, la liste de ces publications ainsi que le dernier numéro paru dans chaque catégorie. La nouvelle série a commencé avec la publication du Rapport n° 1 en janvier 1982.

## ADDENDUM

After the manuscript was in press, a number of incorrect or outdated familial or generic assignments were discovered. The following classification and specific names, recognized by Austin (1983), should replace those presented in Appendices G, H, and I.

Porifera  
Hexactinellida  
Lyssacinosa  
Rosellidae

Coelenterata  
Anthozoa  
Pennatulacea  
Virgulariidae  
Virgularia cf. tuberculata Marshal

Platyhelminthes  
Turbellaria  
Polycladida

Nemertea  
Anopla  
Palaeonemertea

Heteronemertea  
Lineidae  
? Cerebratulus  
Micrura alaskensis? Coe

Enopla  
Hoploneurata

Nematoda

Sipunculoidea  
Golfingiidae  
Golfingia sp.

Echiuroidea

Annelida  
Oligochaeta  
Tubificida  
Tubificidae  
Limnodriloides sp.  
L. victoriensis Brinkhurst and Baker

Polychaeta  
(Errantiate)  
Polynoidae  
? Eunoe  
Gattyana treadwelli? Pettibone

Sigalionidae  
Pholoe minuta (Fabricius)

Phyllodocidae  
Eteone (Mysta) sp.  
E. columbiensis Kravitz and Jones  
Phyllodoce groenlandica Oersted

Pilargidae  
Ancistrosyllis sp.  
A. groenlandica McIntosh

Syllidae  
Exogone sp. 1  
Exogone sp. 2  
E. gemmifera Pagenstecher  
Eusyllis cf. blomstrandii Malmgren  
Syllis sp.  
S. alternata Moore

Nephtyidae  
Aglaophamus malmgreni (Theel)  
Nephtys cornuta cornuta Berkley and Berkley  
N. punctata Hartman

Sphaerodoridae  
Sphaerodoropsis sphaerulifer (Moore)

Goniadidae  
Goniada annulata Moore  
Glycinde armigera Moore

Lumbrineridae  
Lumbrineris sp.  
L. luti Berkley and Berkley  
Paraninoe simpla (Moore)

Dorvilleidae  
Schistomeringos sp.

Hesionidae  
Gyptis brevipalpa (Hartmann-Schroder)

(Sedentariate)  
Orbiniidae  
Leitoscoloplos pugettensis (Pettibone)

Paraonidae

Aricidea suecica Eliason

A. lopezi lopezi Berkley and Berkley

Cirrophorus branchiatus Ehlers

Levinsenia gracilis (Tauber)

Trochochaetidae

Trochochaeta multisetosa (Oersted)

Chaetopteridae

Spiochaetopterus costarum (Claparede)

Spionidae

Polydora sp.

Pseudopolydora kempii? (Southern)

Prionospio sp.

P. cirrifera Wiren

P. steenstrupi Malmgren

Spiophanes sp.

S. kroyeri Grube

Cirratulidae

Chaetozone setosa Malmgren

Cauleriella hamata (Hartman)

C. cf. hamata (Hartman)

Cossuridae

Cossura soyeri Laubier

Flabelligeridae

Brada sp.

B. villosa (Rathke)

Pherusa sp.

Opheliidae

Ophelina breviata (Ehlers)

Sternaspidae

Sternaspis scutata (Renier)

Capitellidae

Capitella capitata (Fabricius)

Decamastus sp.

Heteromastus sp.

H. filobranchus? Berkley and Berkley

Mediomastus sp.

Maldanidae

Maldane glebifex Grube

Rhodine sp.

Oweniidae  
Myriochele oculata Zachs  
Owenia fusiformis Chiaje

Amphictenidae  
Pectinaria sp.  
P. moorei Annenkova

Terebellidae  
Artacama conifera Moore  
Pista cristata (Müller)  
Polycirrus sp.

Trichobranchidae  
Terebellides sp.  
Terebellides stroemi Sars

Ampharetidae  
Amage anops (Johnson)  
Amphicteis sp.  
A. cf. scaphobranchiata Moore  
Anobothrus gracilis (Malmgren)  
Sosanopsis cf. hesslei Banse

Sabellidae  
Euchone sp.  
Jasmineira pacifica Annenkova

UID Polychaeta

Arthropoda  
Crustacea  
Cumacea  
Nannastacidae  
Campylaspis ?papillata Lomakina

Leuconidae  
Eudorella sp.  
Leucon sp.

Tanaidacea  
Leptognathiidae  
Leptognathia sp.

Amphipoda  
Corophiidae  
Corophium acherusicum Costa

Lysianassidae  
Cyclocaris challenger Stebbing  
Koroga megalops Holmes  
Orchomene obtusa (Sars)  
Pachynus barnardi Hurley

Phoxocephalidae  
Paraphoxus oculus Sars

Oedicerotidae  
Bathymedon pumilis Barnard  
Monoculodes cf. emarginatus Barnard

Eusiridae  
?Rhachotropis sp.

Hyperiididae  
Hyperia medusarum (Muller)  
Parathemisto sp.  
P. pacifica Stebbing

Mysidacea  
Mysidae  
Pseudomma truncatum S.I. Smith

Decapoda

Mollusca  
Aplacophora

Gastropoda  
Pyramidellida  
Pyramidellidae  
Turbonilla sp.

Cephalaspidea  
Cylichnidae  
Cylichna altonsa

Gastropteridae  
Gastropteron pacificum? Bergh

Thecosomata  
Limacinidae  
Limacina sp.

Neogastropoda  
Nucellidae

Buccinidae  
Buccinum sp.

Mesogastropoda  
Vitrinellidae  
?Vitrinella sp.

Scaphopoda  
Dentaliida  
Dentaliidae  
Rhabdus rectius Carpenter

Gadilida  
Siphonodentaliidae  
Polyschides californicus (Pilsbry and Sharp)

Bivalvia  
Veneroida  
Tellinidae  
Macoma sp.  
M. carlottensis Whiteaves  
M. eliminata Dunnill and Coan  
M. cf. nasuta (Conrad)

Cooperellidae  
Cooperella sp.

Veneridae  
Transenella tantilla (Gould)

Ungulinidae  
Diplodonta orbella Gould

Hiatellidae  
Hiatella arctica (Linnaeus)

Nuculoidea  
Nuculanidae  
Nuculana minuta Fabricius

Nuculidae  
Nucula tenuis (Montagu)

Yoldiidae  
Yoldia amygdalea Valenciennes  
Y. beringiana Dall  
Y. martyria Dall  
Y. myalis (Couthouy)

Echinodermata  
Asteroidea  
Paxillosida  
Goniopectinidae  
Ctenodiscus crispatus Retzius



Ophiuroidea  
Ophiurida  
Ophiuridae  
Ophiura sp.  
O. leptoctenia H.L. Clark  
O. sarsi Lutken

Holothuroidea  
Apodida  
Chiridotidae  
Chiridota albatrossi Ohshima

Molpadida  
Molpadiidae  
Molpadia intermedia (Ludwig)

Hemichordata  
Enteropneusta

Austin, W.C. 1983. An annotated checklist of marine invertebrates in the cold temperate Northeast Pacific. Khoyatan Marine Laboratory, Cowichan Bay, B.C. 621 pp.

Kathman, R.D., R.O. Brinkhurst, R.E. Woods, and D.C. Jeffries. 1983. Benthic studies in Alice Arm and Hastings Arm, B.C. in relation to mine tailings dispersal. Can. Tech. Rep. Hydrogr. Ocean Sci. 22: vii + 30 p.

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Minister of Supply and Services Canada 1983

Cat. No. Fs 97-18/22E      ISSN 0711-6764

Correct citation for this publication:

Kathman, R.D., R.O. Brinkhurst, R.E. Woods, and D.C. Jeffries. 1983. Benthic studies in Alice Arm and Hastings Arm, B.C. in relation to mine tailings dispersal. Can. Tech. Rep. Hydrogr. Ocean Sci. 22: vii + 30 p.

## TABLE OF CONTENTS

	<u>Page</u>
Abstract/Resume	vi
Acknowledgements	vii
SUMMARY	1
INTRODUCTION	2
TERMS OF REFERENCE	3
METHODS	4
SAMPLING	4
SORTING AND QUALITY CONTROL	4
IDENTIFICATION AND VERIFICATION	4
HIERARCHICAL ANALYSES	8
OTHER RELATED STUDIES	8
RESULTS	9
BENTHIC INVERTEBRATES - 0.25 mm SIEVE	9
BENTHIC INVERTEBRATES - 1.0 mm SIEVE	9
DISCUSSION	16
RECOMMENDATIONS	19
TAXONOMIC REFERENCES AND REFERENCES CITED	20

## APPENDICES

- APPENDIX A      AUXILIARY INFORMATION TAKEN DURING  
SAMPLE ACQUISITION.
- APPENDIX B      INFORMATION SHEET USED DURING SORTING  
OF 1.0 mm SIEVED SAMPLES.
- APPENDIX C      INFORMATION SHEET USED DURING SORTING  
OF 0.25 mm SIEVED SAMPLES.
- APPENDIX D      INFORMATION SHEET USED DURING QUALITY  
CONTROL OF 1.0 mm SAMPLES.
- APPENDIX E      PERTINENT STUDIES AND REPORTS.
- APPENDIX F-1     DENSITY AND DISTRIBUTION OF BENTHIC  
INVERTEBRATES FOR EACH SAMPLE  
(0.25 mm screen) FROM ALICE ARM.
- APPENDIX F-2     DENSITY AND DISTRIBUTION OF BENTHIC  
INVERTEBRATES FOR EACH STATION  
(0.25 mm screen) FROM ALICE ARM.
- APPENDIX F-3     DENSITY AND DISTRIBUTION OF BENTHIC  
INVERTEBRATES FOR EACH SAMPLE  
(0.25 mm screen) FROM HASTINGS ARM.
- APPENDIX F-4     DENSITY AND DISTRIBUTION OF BENTHIC  
INVERTEBRATES FOR EACH STATION  
(0.25 mm screen) FROM HASTINGS ARM.
- APPENDIX G      LIST OF ALL TAXA FOUND IN ALICE ARM  
AND HASTINGS ARM.
- APPENDIX H      DENSITY AND DISTRIBUTION FOR BENTHIC  
INVERTEBRATES (1.0 mm screen) FROM  
ALICE ARM.
- APPENDIX I      DENSITY AND DISTRIBUTION FOR BENTHIC  
INVERTEBRATES (1.0 mm screen) FROM  
HASTINGS ARM.

## LIST OF FIGURES

<u>Figure</u>		<u>Page</u>
1	Map showing general location of study area.	5
2	Station locations in Alice Arm and Hastings Arm, British Columbia.	6
3	Cluster analysis station groupings.	11
4	Tentative horizontal display of cluster analysis of station groupings derived from data in Fig. 3.	12
5	Longitudinal section of Alice Arm showing generalized sediment distribution, sampling locations, and tailings distribution.	13

## LIST OF TABLES

<u>Table</u>		<u>Page</u>
1	Quality control (QC) data for sorting benthic invertebrates from sediments sieved through a 1.0 mm screen.	7
2	Comparison of the results of 0.25 mm and 1.0 mm sieve screening of the same benthic sediment samples.	10
3	Summary of benthic invertebrate composition and density for Alice Arm and Hastings Arm.	15

## ABSTRACT

Kathman, R.D., R.O. Brinkhurst, R.E. Woods, and D.C. Jeffries. 1983. Benthic studies in Alice Arm and Hastings Arm, B.C. in relation to mine tailings dispersal. Can. Tech. Rep. Hydrogr. Ocean Sci. 22, 1-30.

Identification of benthic invertebrates was performed on samples collected in Alice Arm and Hastings Arm, British Columbia, in October 1982 to determine distribution of settled mine tailings and the effect on the benthos. Taxonomic and hierarchical analyses of these organisms indicated that mine tailings were affecting the benthic composition and density in the middle to lower section (towards the sill) of Alice Arm. Sediment data from other sources support these findings.

Key words: benthic invertebrates, mine tailings, sedimentation, fjords.

## RÉSUMÉ

Kathman, R.D., R.O. Brinkhurst, R.E. Woods, and D.C. Jeffries. 1983. Benthic studies in Alice Arm and Hastings Arm, B.C. in relation to mine tailings dispersal. Can. Tech. Rep. Hydrogr. Ocean Sci. 22, 1-30.

L'identification d'invertébrés benthiques a été faite à partir d'échantillons recueillis dans le bras Alice et le bras Hastings (Colombie-Britannique) en octobre 1982 pour déterminer la distribution de stériles miniers déposés et l'effet sur le benthos. L'analyse taxonomique de ces organismes et de leur hiérarchie montre que les stériles miniers ont une incidence sur la composition et la densité du benthos à partir du milieu jusqu'à la partie inférieure (vers le seuil) du bras Alice. Les données sur les sédiments provenant d'autres sources appuient ces conclusions.

Mots-cles: invertébrés benthiques, stériles miniers, sédimentation, fjords.

## ACKNOWLEDGEMENTS

E.V.S. Consultants would like to thank all the people who contributed to this project. Verifications of organisms were done by the following: Dr. Craig Staude - Amphipoda; Dr. Frank Bernard - Mollusca; Mr. Eugene Ruff - Polychaeta; Mr. Steve Stricker - Nemertea; and Dr. William Austin - Echinodermata and minor phyla. Mr. Jean Coustalin performed the initial taxonomic identifications. Several scientists provided additional information; we would especially like to thank Drs. Jeff Thompson and Rob Macdonald, and Mr. Darcy Goyette for their input. E.V.S. Consultants personnel included Dr. Peter Chapman, who reviewed the manuscript and added valuable comments; Mss. Marla Mees and Sarah Irwin, who prepared the report; Ms. Connie Priestley, who assisted with data and draft report preparation; and Mr. Paul Wheeler, who deserves special thanks for coordination and production of the report. Mss. Deedee Kathman and Roberta Woods were the Principal Investigators. Cluster analysis was done by Mr. Duncan Jeffries of Quantum Research. Dr. Ralph Brinkhurst assisted with report preparation.



## SUMMARY

Benthic invertebrate samples were collected during October 1982 in Alice Arm and Hastings Arm, British Columbia. Composition, density and distribution of the benthos were analyzed in relation to the distribution of settled mine tailings in Alice Arm as determined by corresponding studies.

Transects CC and DD in Alice Arm had very few species, indicating severe stress at these locations, particularly in the north and middle stations at CC and the middle station at DD. Transect EE had more species, although the deep middle station along this transect had reduced numbers, comparable to Transect DD. This indicates that mine tailings are probably beginning to affect the lower section of Alice Arm, and have progressed to the deep middle section of EE. Independent sedimentation data and sediment analyses from other sources agree with these findings.

North and south stations on Transect EE had similar numbers of species and individuals as the stations in Hastings Arm. However, the qualitative taxonomic composition of the fauna was quite different. Greater varieties and abundances of nemertean, polychaetes, gastropods, scaphopods, bivalves and echinoderms occurred in Hastings Arm, while nematodes, amphipods and holothuroids were more diverse and abundant at the north and south stations at EE. These differences were probably not related to the effects of tailings but rather to differences in physical, chemical, and biological factors between the two fjords.

Station FF nearest the sill in Alice Arm was dissimilar in composition and density from all other stations sampled and could not be used for comparisons. A heavy sediment load from Hans Creek which settled at Station FF, as well as large amounts of wood fibers in the substrate, contributed to the uniqueness of benthic invertebrates at this station.

## INTRODUCTION

The study of the effect of mine tailings deposition on marine benthic invertebrates in Alice Arm was carried out to indicate the extent and degree of dispersion of tailings along the deeper part of Alice Arm. This study formed one component of an overall study being conducted by the Canadian federal government into the effects of Kitsault mine tailings deposition on the Alice Arm aquatic ecosystem. Other concurrent surveys include zooplankton composition and distribution, sedimentation rates, trace metal concentrations in sediments, and hydrographical data collections.

Tailings discharge into Alice Arm began in April 1981 and ceased with the temporary shut-down of the mill in November 1982. During that period approximately four million tons of process effluent were discharged into Alice Arm (R. Hinder, pers. comm.). A variety of fauna may be affected by these tailings due to toxicity of heavy metal concentrations, increased turbidity limiting light penetration and primary production, and smothering of infaunal organisms.

Samples were collected from a series of transects in Alice Arm at increasing distances from the discharge site, and from adjacent Hastings Arm, an area unaffected by tailings discharge. Benthic invertebrates were sorted from the sediment and the infauna identified. The benthic infaunal data were used to assess possible effects, including extent and severity, of tailings deposition on benthic communities.

## TERMS OF REFERENCE

The overall objective of this study was to determine composition, density and distribution of the benthic invertebrate fauna found in relation to mine tailings distribution in Alice Arm, British Columbia. Specific objectives were:

1. Sort and identify benthic macroinvertebrates retained by a 1.0 mm mesh screen in samples collected from Alice Arm and Hastings Arm, British Columbia.
2. Count benthic invertebrates retained by a 0.25 mm mesh screen in samples collected from Alice Arm and Hastings Arm, British Columbia.
3. Verify all identified material from 1. above.
4. Perform statistical analyses on identification data from 1., allowing comparisons among stations and between the two basins.
5. Prepare a data report incorporating the above and other pertinent information with data on the distribution of infaunal benthic invertebrates in relation to settled mine tailings in Alice Arm.

## METHODS

### SAMPLING

Sampling in Alice Arm and Hastings Arm, British Columbia (Fig. 1), was conducted by Dobrocky Seatech Ltd. in October 1982. Two replicates were taken at each station with the exception of Station EEM on predetermined transect lines (Fig. 2), using a 0.1 m<sup>2</sup> Smith-McIntyre grab. Each sample was sieved through a 1.0 mm screen and preserved with seven percent formalin with phloxine B, an histological stain used to facilitate sorting. Residues from each 1.0 mm sieving were then screened at 0.25 mm and the contents preserved as previously described. Samples were transferred to the E.V.S. Consultants Taxonomy Centre on 12 and 18 November. Pertinent information recorded during the field survey is presented in Appendix A.

### SORTING AND QUALITY CONTROL

Upon receipt of the above samples by the E.V.S. Consultants Taxonomy Centre, each 1.0 mm-sieved sample was washed through a 1.0 mm sieve to remove formalin. Aliquots of each sample were examined using a Wild M3 or M5A stereomicroscope until the entire sample was sorted and all organisms removed. Organisms were counted and placed into separate containers filled with alcohol according to the following major taxonomic groups: Amphipoda, Other Crustacea, Polychaeta, Oligochaeta, Mollusca, Nematoda, and Others. Pertinent information for each sample was recorded using the format shown in Appendix B. Sample residues were represerved with seven percent formalin.

Each 0.25 mm-sieved sample was washed through a 0.25 mm sieve to remove formalin and examined as above. Organisms were counted according to major taxonomic groups (for example, Foraminifera, Nematoda, Polychaeta, Ostracoda, Harpacticoida). Only the Oligochaeta were removed, preserved in ethanol and given to the Scientific Authority. A separate information sheet was compiled for each sample using the format shown in Appendix C. Residues were re-preserved with seven percent formalin for possible future analyses.

To ensure quality control of sorting, thirty-two percent (10 samples) of the 1.0 mm residue samples were independently resorted following the procedure outlined above. If additional organisms were found, they were placed in the appropriately labelled vial and information on the sorting sheets was changed accordingly. A separate sheet for each quality control check was completed, in the format shown in Appendix D.

Quality control data for sorting of 1.0 mm screened samples are provided in Table 1. Percent error ranged from 0 to 4.2, well within the requisite limits for accuracy of sorting benthic invertebrate samples.

### IDENTIFICATION AND VERIFICATION

All organisms were identified to the lowest possible taxonomic level consistent with the presently-available literature. Identifications were accomplished using either a Wild M5A stereomicroscope or a Leitz Laborlux compound microscope. A list of the taxonomic references used for identifications and verifications is provided.

Fig. 1. Map showing general location of study area.

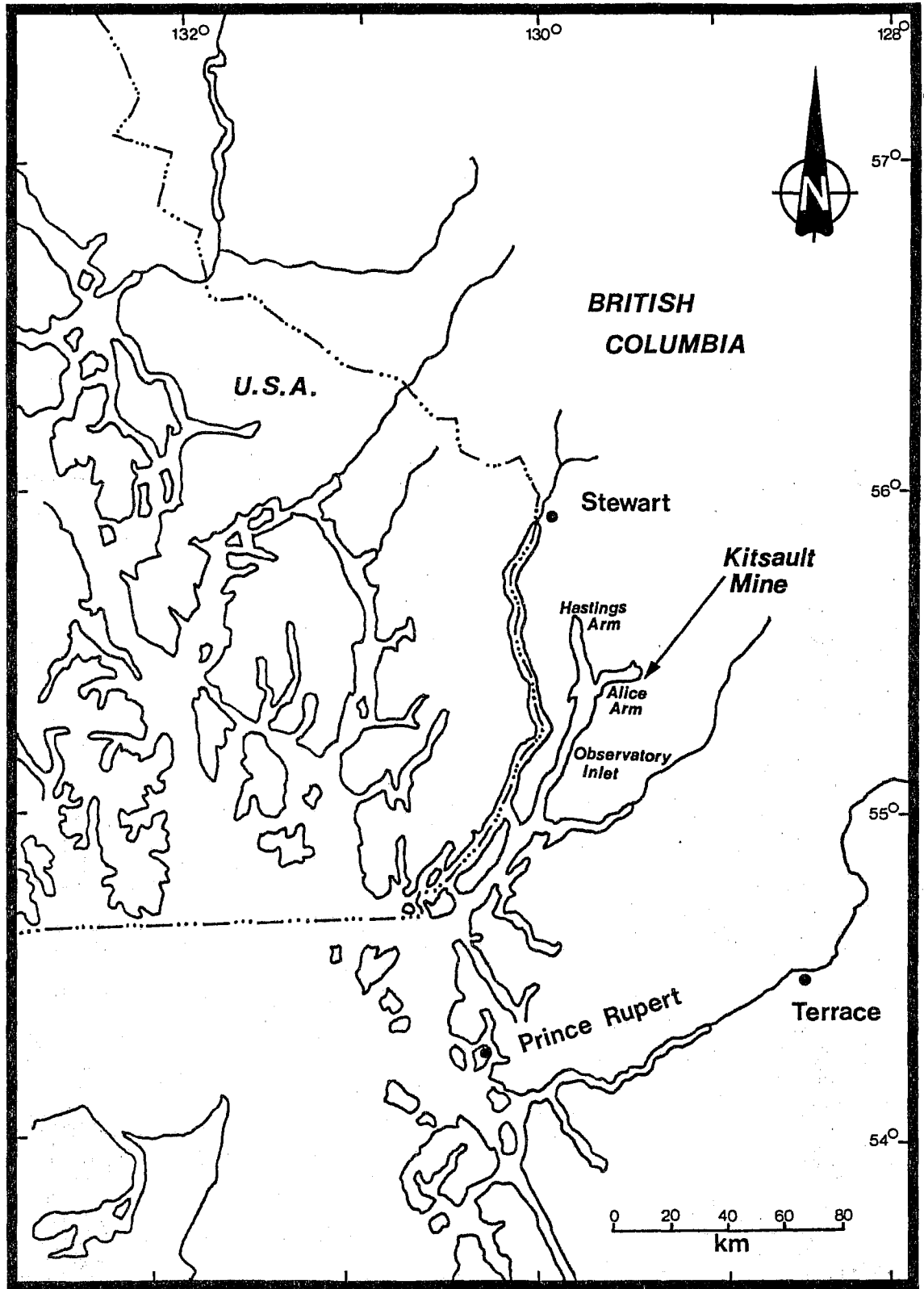


Fig. 2. Station locations in Alice Arm and Hastings Arm, British Columbia.

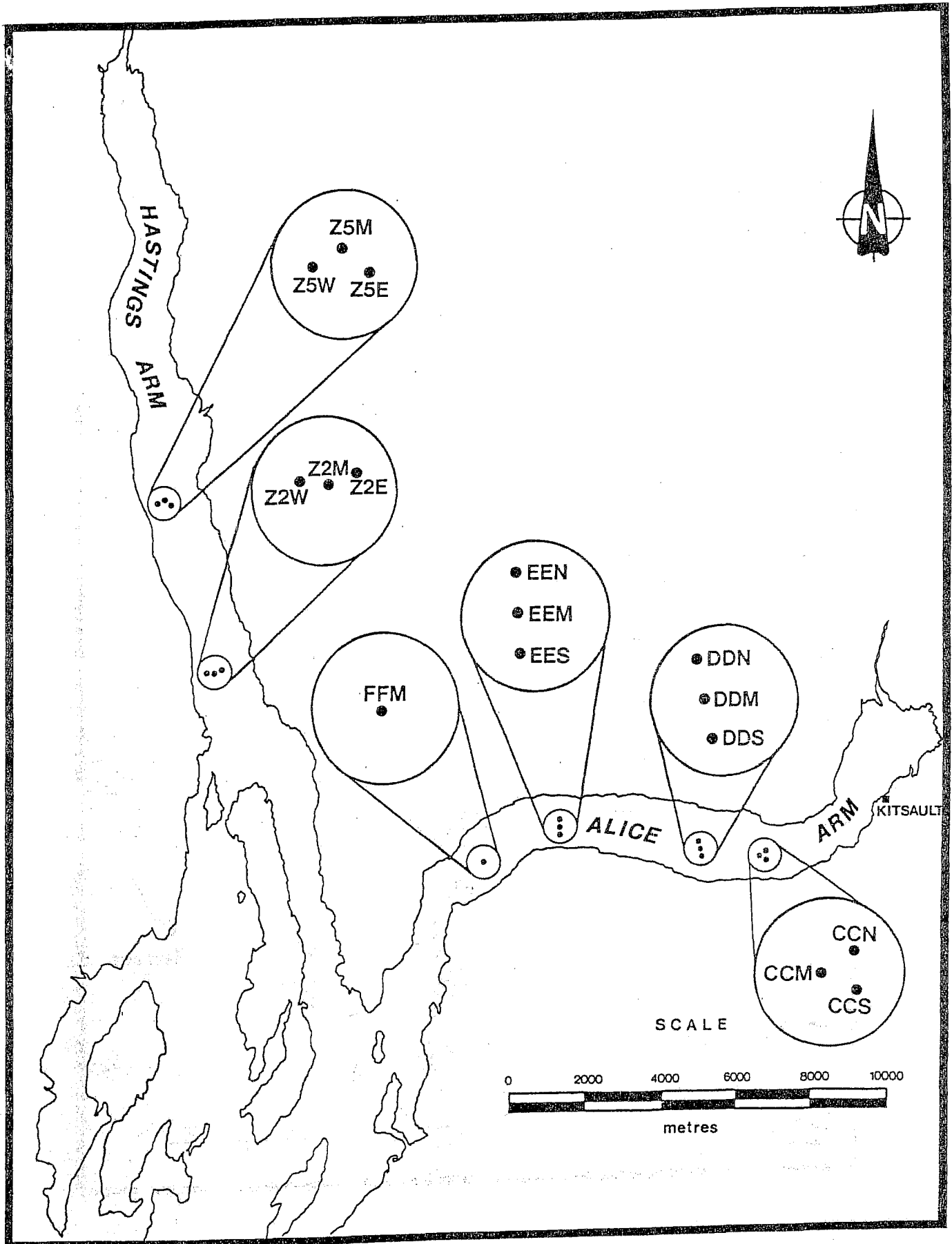


TABLE 1. Quality control (QC) for sorting benthic invertebrates from sediments sieved through a 1.0 mm screen.

Sample identification	Total number of individuals during sorting	Total number of individuals after QC	Percent error for sample
CCN-A	2	2	0
CCM-B	5	5	0
CCS-A	74	77	3.9
DDN-B	57	59	1.7
DDM-A	3	3	0
EEN-A	332	333	0.3
EEM-A	104	104	0
Z2W-B	114	119	4.2
Z5E-A	100	101	1.0
Z5W-B	208	210	1.0

Representative samples of organisms identified to the generic or specific level were sent to recognized taxonomic experts (approved by the Scientific Authority) for verification. If the original identification was incorrect, all samples of that genus or species were re-examined to ensure accuracy. In addition, all samples identified to the familial level were sent to experts for possible identification to a lower taxonomic unit. All necessary changes were incorporated into the original identification sheets and vial labels.

A representative sample of each genus/species was separated from the entire collection and presented to the Scientific Authority as a reference set for future use. The identity of all reference material was further verified by appropriate taxonomic experts.

### HIERARCHICAL ANALYSIS

Two-way cluster (hierarchical) analysis was performed on the data using the cluster analysis program "\*CGROUP", allowing comparison among species and among stations. This program also allowed comparison among species and between Alice Arm and Hastings Arm. The program uses the grouping algorithm as described by Ward (1963) and follows recognized and accepted techniques for analysis of benthic data. Replicate samples for each station were averaged in order to produce an appropriate data base for use in the cluster analysis. This also simplified the problem associated with the single sample for Station EEM. Data were also entered individually and are therefore on file and readily accessible for possible future analyses.

### OTHER RELATED STUDIES

Other agencies and individuals were contacted to identify pertinent past and current studies in Alice Arm and in other areas impacted by mine tailings (for example, Rupert Inlet). Respondents included Drs. D. Mackas, R. Macdonald, and J. Thompson and Mr. D. Stucchi (Institute of Ocean Sciences), Drs. J.-McInerney and D. Ellis (University of Victoria), Dr. S. Calvert (University of British Columbia), Dr. J. Patterson (Amax of Canada Ltd.), Mr. D. Goyette (Environmental Protection Service), and Mr. W. Drinnan (Dobrocky Seatech Ltd.). A partial listing of studies and reports is given in Appendix E. However, given the terms of reference for this study, a comprehensive review of all pertinent information was not possible.



## RESULTS

### BENTHIC INVERTEBRATES - 0.25 mm SIEVE SAMPLES

The results of simple scanning of the invertebrates retained by a 0.25 mm sieve are presented in Appendices F-1 through F-4. They are summarized for all taxa in Table 2. These samples were initially retained to ascertain if differences in benthic distribution and density in relation to tailings dispersal would be observed between these and the 1.0 mm sieved samples. Although the 1.0 mm sieved samples only accounted for approximately 23 percent of the total benthic invertebrates collected in Alice Arm and Hastings Arm, similar patterns between each set of samples among stations and transects can be observed. For example, for both sieve sizes fewer numbers of organisms were collected at the middle stations on Transects DD and EE than at the north and south stations, and the north and middle stations on Transect CC had fewer individuals than the south station.

Nematodes (which are not readily identifiable) comprised 70 percent of the small organisms found in Alice Arm. If the contribution of these organisms to overall numbers in samples of both sieve sizes is discounted, the patterns of similarity become apparent (see Table 2). The data indicate that benthic samples screened with 1.0 mm mesh will provide enough information to accurately assess trends and patterns in benthic composition and distribution.

### BENTHIC INVERTEBRATES - 1.0 mm SIEVE SAMPLES

A listing of all benthic invertebrates collected during this study is given in Appendix G. The number of organisms for each sample and the mean number of organisms for each station in Alice Arm and Hastings Arm are presented in Appendices H and I, respectively. The cluster analysis performed on these data clearly differentiates three groups of stations: Group 1 - Alice Arm not including Station FFM; Group 2 - Hastings Arm; and Group 3 - Station FFM in Alice Arm (see Figure 3). These groupings are based on the species, numbers of taxa and numbers of individuals collected. Three subgroups can be differentiated in Group 1. Stations CCN, DDM and CCM are very similar, having a very poor benthic infaunal composition. Visual observations made from Pisces IV and trace metal analyses of sediments in cores verify a more northern circulation of tailings at Transect CC (D. Goyette, pers. comm.). The next subgroup (1.2) comprises the remaining stations in Transects CC and DD which are also quite severely affected by tailings deposition. Stations on Transect EE join stations on CC and DD at Step 6 in the clustering. EEM is the most divergent station on Transect EE and most similar to stations on Transects CC and DD. Figure 4 illustrates a tentative horizontal display of the cluster analysis, showing the similarity among stations in relation to tailings distribution. All species collected at Station EEM were also found at EEN and EES, although approximately one-third as many individuals and one-half as many taxa occurred at EEM. This substantial decrease is indicated by the dashed lined in Figure 4. Sedimentation data substantiate settled tailings occurring at Transect EE as shown in Figure 5 (R. Macdonald, pers. comm.). Trace metal analyses indicate that surface sediments in 1981 at Station EEM contain much higher concentrations than at either EEN or EES, and are comparable to concentrations at Stations CCM and DDM (Goyette and Christie, 1982).

TABLE 2. Comparison of the results of 0.25 mm and 1.0 mm sieve screening of the same benthic sediment samples. All results are given in numbers per m<sup>2</sup> and are the mean of two samples (except EEM).

	0.25 mm			1.0 mm		
	No. of Individ. per Stat.	No. of Individ. per Trans.	No. of Individ. per Inlet	No. of Individ. per Stat.	No. of Individ. per Trans.	No. of Individ. per Inlet
CCN	675			20		
CCM	230	3,260		30	715	
CCS	2,355			665		
DDN	3,565			520		
DDM	775	7,235	31,200	25	1,630	9,950
DDS	2,895			1,085		
EEN	11,140			3,125		
EEM	1,755	20,705		1,040	7,605	
EES	7,810			3,440		
FFM*	64,380			12,565		
Z2E	11,725			1,835		
Z2M	6,415	22,455		1,255	5,175	
Z2W	4,315			2,085		
			56,355			10,305
Z5E	14,830			1,310		
Z5M	8,715	33,900		2,075	5,130	
Z5W	10,355			1,745		

\*Station FFM considered a separate and distinct area.

Fig. 3. Cluster analysis station groupings.

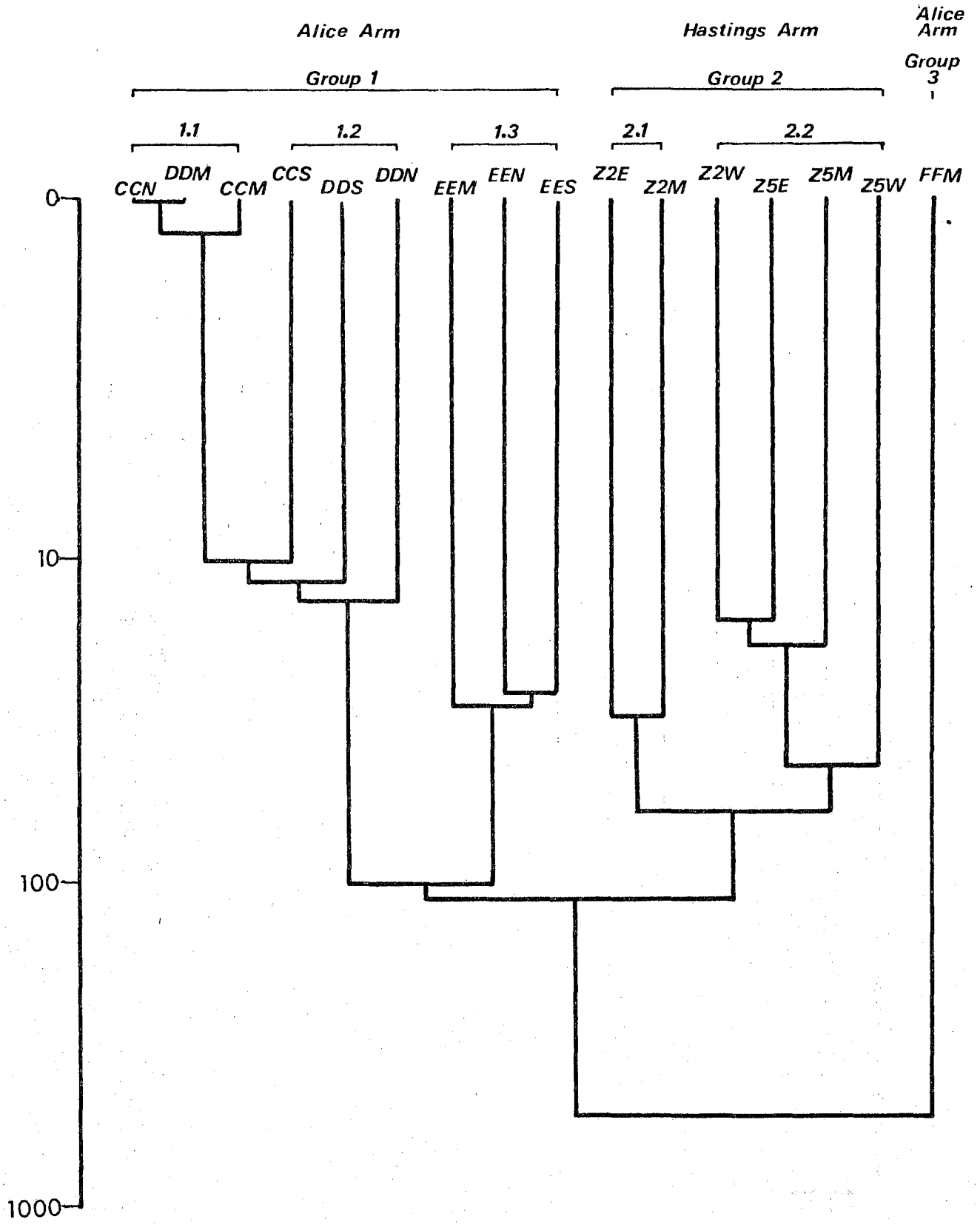


Fig. 4. Tentative horizontal display of cluster analysis of station groupings derived from data in Fig. 3.

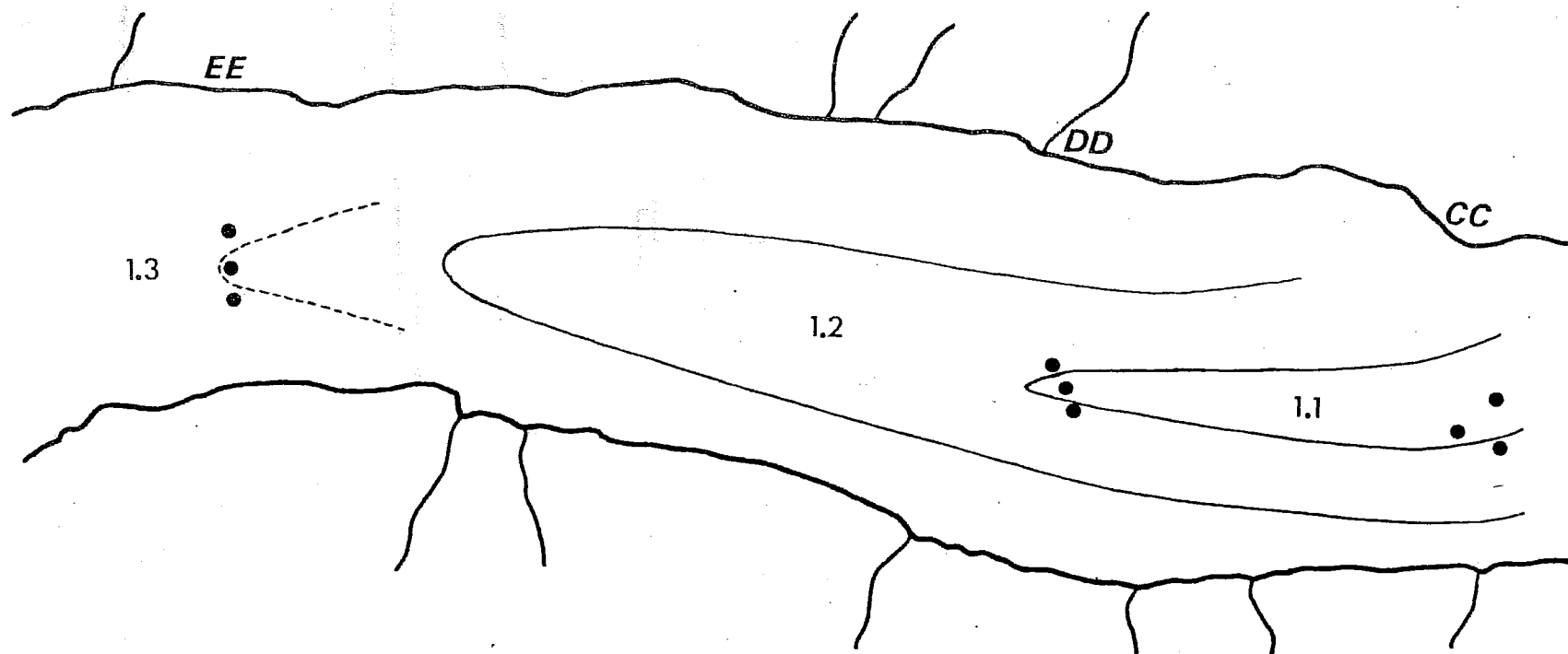
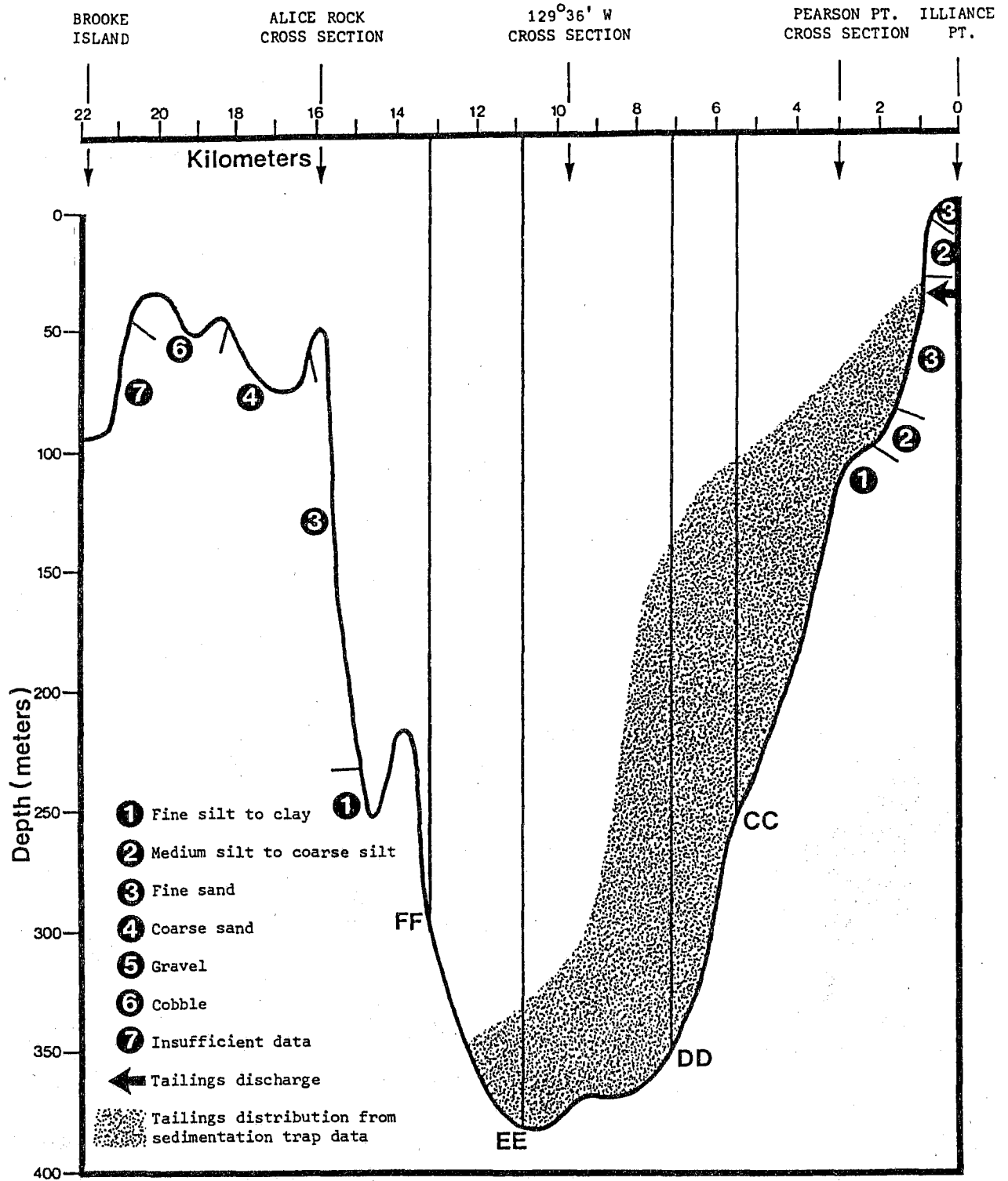


Fig. 5. Longitudinal section of Alice Arm showing generalized sediment distribution, sampling locations, and tailings distribution. Data from R.W. Macdonald and J.L. Littlepage.



While the cluster analysis delineates two subgroups of stations (2.1 and 2.2) within Hastings Arm, we feel the degree of dissimilarity is not a significant factor for comparison with Alice Arm. Group 2 stations are clearly differentiated from Group 1 stations at Step 14. Speciation and density of the benthos as seen in Table 3 and Appendices H and I are also distinct between the two inlets.

Station FFM in Alice Arm forms Group 3, which is widely divergent from all other stations. This station contained higher numbers of individuals and numbers of taxa than all other stations combined in either Alice Arm or Hastings Arm (Table 3). At least twice as many endemic taxa (organisms restricted to a particular station in this study) were found at this station compared to each other transect.

The number of individuals found in the northern and middle samples at CC, and the middle samples at DD and EE in Alice Arm were an order of magnitude lower than the number of individuals found in the equivalent southern samples at CC, and the equivalent northern and southern samples at DD and EE, as shown below.

Transect	<u>CC</u>			<u>DD</u>			<u>EE</u>		
	<u>N</u>	<u>M</u>	<u>S</u>	<u>N</u>	<u>M</u>	<u>S</u>	<u>N</u>	<u>M</u>	<u>S</u>
Number of individuals per m <sup>2</sup>	20	30	670	520	25	1,085	3,125	1,040	3,440

Polychaetes were the most abundant organisms at Transects CC, DD and FF, while polychaetes and molluscs were equally abundant at EE in Alice Arm (Table 3). Nephtys cornuta cornuta, Levinsenia gracilis, Spiophanes sp. and Myriochele oculata were the overall dominant polychaetes. Transenella tantilla and Diplodonta orbella were the most abundant bivalves. The fauna at Transects CC and DD, with the exception of five nemerteans at Station DDN, consisted of only four major groups: polychaetes, crustaceans, molluscs and echinoderms.

Polychaetes were the most abundant organisms in Hastings Arm, represented by large numbers of N. cornuta cornuta, Aricidea lopezi lopezi, L. gracilis, Chaetozone setosa, Cossura soyeri and Cirratulidae (Table 3). The bivalves T. tantilla, D. orbella and Nucula tenuis, and ophiuroids were also common.

TABLE 3. Summary of benthic invertebrate composition and density for Alice Arm and Hastings Arm.

Mean number of individuals per m <sup>2</sup>	Regions				
	Alice Arm			Hastings Arm	Alice Arm
	CC	DD	EE	Z2 + Z5	FF
All individuals	238	543	2,843	1,718	12,565
Polychaeta spp.	216	112	922	1,213	11,170
Mollusca spp.	8	73	1,648	306	335
Crustacea spp.	7	12	86	59	90
<u>Total number of taxa per transect</u>					
All taxa	27	25	52	69	93
Polychaeta spp.	18	12	22	35	61
Mollusca spp.	3	5	15	14	11
Crustacea spp.	3	6	9	12	7
Endemic taxa*	0	4	8	14	29
Endemic Polychaeta spp.	0	0	1	4	16
Endemic Mollusca spp.	0	0	2	4	4
Endemic Crustacea spp.	0	3	4	2	3
No. of samples	6	6	5	12	2

\*Species limited to a transect in this study.

## DISCUSSION

Visual examination of the data indicates a depressed number and variety of benthic infaunal species at all stations on Transects CC and DD in Alice Arm. Field observations (S. Byers, pers. comm.) suggest that deposits of mine tailings were present in the sample sediments from these stations. Data on the present areal distribution of mine tailings (Fig. 5; R. Macdonald, pers. comm.) indicate that both these transects are affected by settled tailings deposits. Cluster analysis of station groupings (Fig. 3) agrees with grain size distribution and trace metal concentrations (Goyette and Christie, 1982) and light transmission properties (D. Stucchi, pers. comm.). Suppressed benthic fauna at CCN and CCM, but slightly elevated benthic populations at CCS coincide with the initial northerly tailings plume track from Kitsault (D. Goyette, pers. comm. and D. Stucchi, pers. comm.). These tailings appear to be settling in the deep middle stations at Transects DD and EE, as shown in Figure 4 and evidenced by the reduced numbers of species and individuals at DDM and EEM. Equally high trace metal concentrations at CCN, CCM, DDM and EEM (Goyette and Christie, 1982) suggest that tailings have progressed down the inlet at least to EEM, but have not spread to the north or south sides of the channel in this area.

The benthic fauna at Station FFM was distinct from all other stations in both Alice Arm and Hastings Arm. Cluster analysis, visual observation and benthic composition indicate that this station is not representative of other areas in the inlet. This station is directly affected by Hans Creek with highly variable sediment and associated organic loadings. In October 1982, during the period of sample collection, large sediment inputs (three or four times average transport) were collected at this station location in sedimentation traps at subsurface and near-bottom locations (R. Macdonald, pers. comm.). The substrate also differed from that at other Alice Arm stations, with large amounts of wood fiber observed in the sediment during field sampling (S. Byers, pers. comm.). This station is also located close to the sill at the mouth of Alice Arm, which may produce different circulation patterns, making available more niches and more nutrients. Natural variability in this area would be expected to be high. Consequently, this station may not provide useful information related to effects of mine tailings dispersal in Alice Arm.

Faunal composition and density in Hastings Arm appeared to be distinct from that in Alice Arm, as evidenced in the cluster analysis (Fig. 3). The faunal density and diversity is similar between Hastings Arm Transects Z2 and Z5, and Alice Arm Stations EEN and EES, (Table 3; Appendices H and I). (The major difference between EEN and EES compared to Hastings Arm is the large number of the bivalves Transenella tantilla and Nucula tenuis found at EEN and EES.) Hastings Arm thus serves as a useful reference area for comparison of "normal" benthic composition, even though it differs in detail from Alice Arm. Although the two fjords are physically distinct and direct comparisons cannot be made between the benthic fauna, Hastings Arm serves as an important reference inlet on which to compare species groupings and diversity.

Changes in infaunal composition and density can be observed between the data from a 1977 benthic study and the 1982 data set. The mean numbers of taxa and mean numbers of individuals at Transect CC in Alice Arm (identified as Station G9.5 in 1977) have decreased substantially in the 1982 samples, as indicated below.



	Transect CC		Percent reduction
	1977	1982	
Mean number of taxa per sample	23	11	52
Mean number of individuals per m <sup>2</sup>	483	238	51

This decrease is even more noticeable if only CCN and CCM (1982) data, in which the mean number of taxa was four and the mean number of individuals was 25, are compared with the 1977 data. Station CCS, however, was similar to G9.5 (23 vs. 23 taxa; 665 vs. 483 individuals, respectively), suggesting that the north and middle stations have been dramatically affected by mine tailings.

Further comparisons between the 1977 and 1982 sampling data for Alice Arm are complicated by the fact that station locations differed. For instance, although the 1982 Transect EE was located in the same general area as the 1977 Station F15, the latter was approximately 20 m deeper and located farther from the tailings discharge. However, comparison of the two data sets indicated substantial differences not totally attributable to location. Of 18 species of polychaetes collected in 1977 and 23 species in 1982, only seven were common to both collections. The 1977 samples showed no clear dominance by particular organisms, while the 1982 benthos was dominated by the bivalve Transenella tantilla (38 percent) and the polychaete Nephtys cornuta cornuta (17 percent).

A number of factors in addition to increased mine tailings dispersal could be responsible for the marked differences between 1977 and 1982 collections. Sampling in 1977 took place during June whereas 1982 samples were collected in October and seasonal differences may be a factor. In addition, 1977 samples were not collected in the same locations as 1982 samples. The closest match between the two collection periods is provided by Transect CC (=Station G9.5) with only minor differences in location (latitude varied less than one minute and longitude less than 15 seconds), but the sampling records indicate that 1977 collection depths were greater than those recorded in 1982 by an average of over 24 meters. Other factors such as natural variability may also be implicated.

Overall, many species of polychaetes collected in 1977 were not found in 1982. But the most abundant sedentary polychaete found in 1982, Levinsenia gracilis, was not collected at all in 1977, indicating that both elimination and addition of species have occurred in Alice Arm. Of the 23 species of polychaetes identified in 1977 and the 28 species identified in 1982, only five were common to both sampling periods.

Previous sampling closer to the Alice Arm sill (Station I 17.5 in 1977) indicated that Station FFM was not representative of other areas during the October sampling. Taxa were approximately threefold higher and numbers of individuals tenfold higher in 1982 when the three sample average in 1977 is compared with the two sample average in 1982. Reasons for variabilities have been discussed previously.

Transects Z2 and Z5 in Hastings Arm were sampled in both 1982 and 1977, allowing comparisons to be made among numbers of taxa, numbers of individuals and species. Numbers of taxa and numbers of individuals were lower in 1977 than in 1982 as shown below.

Year	<u>1977</u>		<u>1982</u>	
	<u>Z2</u>	<u>Z5</u>	<u>Z2</u>	<u>Z5</u>
Mean number of taxa per sample	15	15	36	37
Mean number of individuals per m <sup>2</sup>	320	227	1725	1710

These changes in faunal composition and density are apparently typical of temporal changes observed in other fjords. For example, numerous studies related to tailings discharge from Island Copper Mine in Rupert Inlet indicate annual shifts in benthic composition and density (D. Ellis, pers. comm.). No individual taxon was clearly dominant in 1977, but polychaetes were dominant at both transects in 1982 (71 percent), followed by molluscs (18 percent). Nephtys cornuta cornuta, L. gracilis, A. lopezi lopezi and Cirratulidae were dominant polychaetes, and I. tantilla and N. tenuis were the dominant bivalve molluscs.

The 1982 data set indicated that benthic infaunal composition and density in Hastings Arm are representative of northern fjords. Undisturbed stations in Alice Arm (EEN, EES) have similar benthic density and diversity as stations in Hastings Arm, making the latter useful as a reference location for comparisons. The data for Alice Arm indicated that mine tailings in Alice Arm are progressing westward down the inlet and settling in the deepest part of the trough.

Elimination of most benthic invertebrates at Transects CC and DD and a large decrease at Station EEM coincides with sedimentation data on tailings distribution and trace metal concentrations of sediments. The present benthic infaunal data set also provides a basis for future comparisons to evaluate possible faunal changes and to relate these changes to substrate disturbances such as tailings sedimentation.

## RECOMMENDATIONS

1. An annual or biennial benthic program should be implemented to monitor the density and distribution of benthic invertebrates in relation to settled mine tailings in Alice Arm. This should include stations in both Alice Arm and Hastings Arm.
2. Additional transects should be established between DD and EE, and EE and FF to provide more detailed information on the dispersal of mine tailings and the effect on benthos.
3. Benthic sampling should be done prior to commencement of mill operations after a period of recovery has taken place, followed by sampling a sufficient time after operation to pinpoint and determine specific effects caused by tailings discharge.

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**APPENDIX A**  
**AUXILIARY INFORMATION TAKEN**  
**DURING SAMPLE ACQUISITION**

<u>Date</u> (October 1982)	<u>Station*</u>	<u>Avg. Depth</u> (m)	<u>North latitude</u> <u>West longitude</u>
23	CCN	264.5	55°26'40" 129°31'42"
26	CCM	277.5	55°26'36" 129°31'53"
26	CCS	256.0	55°26'33" 129°31'44"
25	DDN	349.5	55°26'48" 129°33'37"
26	DDM	351.0	55°26'44" 129°33'35"
26	DDS	356.0	55°26'41" 129°33'31"
25	EEN	370.0	55°27'12" 129°37'00"
25	EEM**	373.0	55°27'06" 129°37'00"
25	EES	369.0	55°27'00" 129°37'00"
26	FFM	303.0	55°26'30" 129°39'03"
24	Z2E	296.0	55°29'19" 129°45'37"
24	Z2M	296.0	55°29'17" 129°45'50"
24	Z2W	296.0	55°29'14" 129°45'57"
24	Z5E	294.0	55°31'41" 129°46'53"
24	Z5M	295.0	55°31'41" 129°47'02"
24	Z5W	295.0	55°31'38" 129°47'12"

\* Stations CCN to FFM are located in Alice Arm; Z2E to Z5W are located in Hastings Arm.

\*\* One sample taken at EEM; two samples at all other stations.

APPENDIX B

INFORMATION SHEET USED DURING  
SORTING OF 1.0 mm-SIEVED SAMPLES

Sample identification \_\_\_\_\_

Date of Collection \_\_\_\_\_ Sieve Size \_\_\_\_\_

Sorted by \_\_\_\_\_ Date sorted \_\_\_\_\_ No. Hours \_\_\_\_\_

<u>TAXA</u>	<u>NO. OF INDIVIDUALS</u>
Amphipoda	_____
Other Crustacea	_____
Polychaeta	_____
Oligochaeta	_____
Molluska	_____
Nematoda	_____
Others	_____

Substrate description  
(by percent) \_\_\_\_\_

\_\_\_\_\_

Observations/comments \_\_\_\_\_

\_\_\_\_\_

\_\_\_\_\_





APPENDIX D

INFORMATION SHEET USED DURING  
QUALITY CONTROL OF 1.0 mm SAMPLES

Sample identification \_\_\_\_\_

Date of Collection \_\_\_\_\_ Sieve Size \_\_\_\_\_

Sorted by \_\_\_\_\_ Date sorted \_\_\_\_\_ No. Hours \_\_\_\_\_

QC'd by \_\_\_\_\_ Date QC'd \_\_\_\_\_ No. Hours \_\_\_\_\_

<u>Organism</u>	<u>Number of indiv. for taxon found during QC</u>	<u>No. indiv. for taxon originally found</u>	<u>Percent error for taxon</u>	<u>Total no. of individuals in sample</u>	<u>Percent error for sample</u>
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Total number individuals missed \_\_\_\_\_

Percent error for total sample \_\_\_\_\_

APPENDIX E  
PERTINENT STUDIES AND REPORTS  
PARTIAL LISTING

- Amax of Canada Ltd. 1981. Kitsault mine effluent monitoring program, Int. Rep. prep. for Environ. Canada, EPS. 105 pp.
- Anderson, E. and D. Mackas. Short-term effects of tailings from the Amax Kitsault Mine on the survival, respiration, feeding and swimming behaviour of marine zooplankton. (In preparation)
- Anderson, E. and D. Mackas. Zooplankton community structure in Alice Arm, Hastings Arm and Observatory Inlet, August 1981 and June 1982, with reference to the discharge of tailings from the Amax Kitsault Mine. (In preparation)
- Goyette, D. and P. Christie. 1982. Environmental studies in Alice Arm and Hastings Arm, British Columbia. III. Initial production period - Amax/Kitsault Mine - Sediment and tissue trace metals, May - June and October 1981. Reg. Prog. Rep. 82-14, EPS, Pacific Region.
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- Macdonald, R.W. Sediment transport and sedimentation processes in Alice Arm.\*
- Nicoll, M. and D.J. Stucci. 1982. Alice Arm 1981 CTD data access guide. Can. Contract. Rep. Hydrogr. Ocean Sci. 5: 76 pp.
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- Retech Ltd. Sedimentation of mine tailings in a marine environment.\*
- Stucchi, D. Analysis of the Crash Program data (1981 CTD data set).\*
- Thompson, J.A.J. Heavy metal analyses of sediments and interstitial water from cores taken in Alice Arm and surrounding environs.\*

\*General title or description of ongoing study.

APPENDIX F-1

DENSITY AND DISTRIBUTION OF BENTHIC INVERTEBRATES  
FOR EACH SAMPLE (0.25 mm screen) FROM ALICE ARM.  
ALL RESULTS ARE GIVEN IN NUMBERS PER M<sup>2</sup>.

	CCN		CCM		CCS		DDN		DDM		DDS		EEN		EEM		EES		FFM		
	A	B	A	B	A	B	A	B	A	B	A	B	A	B*	A	B	A	B	A	B	
Coelenterata																					
Hydroidea																					
Platyhelminthes																					
Turbellaria																					
Nemertea	10						10				10	10	40				20			160	
Nematoda	240	10	150		940	1570	4060	640	40	900	170	1780	8260	8320	1450		1180	1870	50480	51080	
Annelida																					
Oligochaeta																					
Polychaeta	120		70	50	850	1100	260	870		140	650	840	2230	1320	560		4730	6040	14640	10160	
Arthropoda																					
Crustacea																					
Cumacea					10								10		10						
Tanaidacea																					
Amphipoda							80	10	10				130	90	70		60	40	40	160	160
Isopoda							10														
Ostracoda	20		10	20	50	40	250	20	40	60	220	60	710	510	790		60	590	160	120	
Harpacticoida	10				10	20	40	10	10	30	20	40	10		20		40	20	80	400	
Insecta																					
Collembola	10																				
Acarina							10							10							
Mollusca																					
Aplacophora							10														40
Gastropoda	770	70		110	60	30	410	10	120	110	30	100	170	20	280		10	20	160		
Pelecypoda	80	10	20	20	10	10	320	110	50	40		390	280	90	290		510	390	440	280	
Echinodermata																					
Ophiuroidea			10		10						140	1330	80		40		20	10			
Tardigrada																					40

■ No Sample taken.





APPENDIX F-4

DENSITY AND DISTRIBUTION OF BENTHIC INVERTEBRATES  
FOR EACH STATION (0.25 mm screen) FROM HASTINGS ARM.  
ALL RESULTS ARE GIVEN IN NUMBERS PER M<sup>2</sup>.

	Z2E	Z2M	Z2W	Z5E	Z5M	Z5W
Coelenterata Hydroidea	10				25	
Platyhelminthes Turbellaria						
Nemertea	15	5	15	85	20	90
Nematoda	1450	505	595	2005	3070	1015
Annelida Oligochaeta		5				
Polychaeta	9430	5445	3420	12280	5080	8910
Arthropoda Crustacea Cumacea	5	60	5	125	125	110
Tanaidacea		20	5	10	10	
Amphipoda	5	10	5	10	30	15
Isopoda						
Ostracoda	85	10	10	55	15	35
Harpacticoida	200	30	100	95	195	110
Insecta Collembola						
Acarina	5					
Mollusca Aplacophora	5			5		5
Gastropoda	110	25	25	25	45	10
Pelecypoda	405	300	135	135	85	40
Echinodermata Ophiuroidea					15	15
Tardigrada						

APPENDIX G

LIST OF ALL TAXA FOUND IN  
ALICE ARM AND HASTINGS ARM

Porifera

Hexactinellida  
Rosellidae

Coelenterata

Pennatulacea  
Virgulariidae  
Virgularia cf. tuberculata Marshal

Platyhelminthes

Turbellaria  
Polycladida

Nemertea

Anopla  
Paleonemertea

Heteronemertea

Lineidae  
? Cerebratulus  
Micrura alaskensis? Coe

Enopla

Hoplonemertea

Nematoda

Sipunculida

Golfingiidae  
Golfingia sp.

Echiura

Annelida

Oligochaeta  
Tubificida  
Tubificidae  
Limnodriloides sp.  
L. victoriensis Brinkhurst and Baker

Polychaeta

(Errantiate)  
Polynoidae

? Eunoe  
Gattyana treadwelli? Pettibone



Sigalionidae

Pholoe minuta (Fabricius)

Phyllodocidae

Eteone (Mysta) sp.

E. columbiensis Kravitz and Jones

Phyllodoce groenlandica Oersted

Pilargidae

Ancistrosyllis sp.

A. groenlandica McIntosh

Syllidae

Exogone sp. 1

Exogone sp. 2

E. gemmifera Pagenstecher

Eusyllis cf. blomstrandii Malmgren

Syllis sp.

S. alternata Moore

Nephtyidae

Aglaophamus malmgreni (Theel)

Nephtys cornuta cornuta Berkley and Berkley

N. punctata Hartman

Sphaerodoridae

Sphaerodoropsis sphaerulifer (Moore)

Goniadidae

Goniada annulata Moore

Glycinde armigera Moore

Lumbrineridae

Lumbrineris sp.

L. luti Berkley and Berkley

Paraninoe simpla (Moore)

Dorvilleidae

Schistomeringos sp.

Hesionidae

Gyptis brevipalpa (Hartmann-Schroder)

(Sedentariate)

Orbiniidae

Leitoscoloplos pugettensis (Pettibone)

Paraonidae

Aricidea suecica Eliason

A. lopezi lopezi Berkley and Berkley

Cirrophorus branchiatus Ehlers

Levinsenia gracilis (Tauber)

Trochochaetidae  
Trochochaeta multisetosa (Oersted)

Chaetopteridae  
Spiochaetopterus costarum (Claparede)

Spionidae  
Polydora sp.  
Pseudopolydora kempfi? (Southern)  
Prionospio sp.  
P. cirrifera Wiren  
P. steenstrupi Malmgren  
Spiophanes sp.  
S. kroyeri Grube

Cirratulidae  
Chaetozone setosa Malmgren  
Caulleriella hamata (Hartman)  
C. cf. hamata (Hartman)

Cossuridae  
Cossura soyeri Laubier

Flabelligeridae  
Brada sp.  
B. villosa (Rathke)  
Pherusa sp.

Opheliidae  
Ophelina breviata (Ehlers)

Sternaspidae  
Sternaspis scutata (Renier)

Capitellidae  
Capitella capitata (Fabricius)  
Decamastus sp.  
Heteromastus sp.  
H. filobranchus? Berkley and Berkley  
Mediomastus sp.

Maldanidae  
Maldane glebifex Grube  
Rhodine sp.

Oweniidae  
Myriochele oculata Zachs  
Owenia fusiformis Chiaje

Amphictenidae  
Pectinaria sp.  
P. moorei Annenkova

Terebellidae

Artacama conifera Moore  
Pista cristata (Muller)  
Polycirrus sp.

Trichobranchidae

Terebellides sp.  
Terebellides stroemi Sars

Ampharetidae

Amage anops (Johnson)  
Amphicteis sp.  
A. cf. scaphobranchiata Moore  
Anobothrus gracilis (Malmgren)  
Sosanopsis cf. hesslei Banse

Sabellidae

Euchone sp.  
Jasmineira pacifica Annenkova

UID Polychaeta

Arthropoda

Crustacea

Cumacea

Campylaspis ? papillata Lomakina  
Eudorella sp.  
Leucon sp.

Tanaidacea

Leptognathia sp.

Amphipoda

Corophiidae

Corophium acherusicum Costa

Lysianassidae

Cyphocaris challengeri Stebbing  
Koroga megalops Holmes  
Orchomene obtusa (Sars)  
Pachynus barnardi Hurley

Phoxocephalidae

Paraphoxus oculatus (Sars)

Oedicerotidae

Bathymedon pumilis Barnard  
Monoculodes cf. emarginatus Barnard

Eusiridae

?Rhacotropis sp.

Hyperiidæ  
Hyperia sedusarum Muller  
Parathemisto sp.  
P. pacificus Stebbing

Mysidacea  
Mysidæ  
Pseudomma truncatum S.I. Smith

Decapoda

Mollusca  
Aplacophora

Gastropoda  
Pyramidellida  
Pyramidellidæ  
Turbonilla sp.

Cephalospidea  
Cylichna altonsa  
Gastropteron pacificum? Bergh

Thecosomata  
Limacina sp.

Neogastropoda  
Thaididæ

Buccinidæ  
Buccinum sp.

Megogastropoda  
Vitrinellidæ  
?Vitrinella sp.

Scaphopoda  
Dentaloida  
Dentaliidæ  
Dentalium rectius Carpenter

Siphonodentaloida  
Cadulidæ  
Cadulus californicus (Pilsbury and Sharp)

Bivalvia  
Heterodonta  
Tellinidæ  
Macoma sp.  
M. carlottensis Whiteaves  
M. elimata Dunnill and Coan  
M. cf. nasuta (Conrad)

Cooperellidae  
Cooperella sp.

Veneridae  
Transenella tantilla (Gould)

Ungulidae  
Diplodonta orbella Gould

Hiatellidae  
Hiatella arctica (Linnaeus)  
Nuculana minuta Fabricius  
Nucula tenuis (Montagu)  
Yoldia amydalea Valenciennes  
Y. beringiana Dall  
Y. martyria Dall  
Y. myalis (Couthouy)

Echinodermata  
Asteroidea  
Paxilloridea  
Goniopectinidae  
Ctenodiscus crispatus Retzius

Ophiuroidea  
Ophiurida  
Ophiolepididae  
Ophiura sp.  
O. leptoctenia H.L. Clark  
O. sarsi Lutken

Holothuroidea  
Apodida  
Chirodotidae  
Chirodota albatrossi Ohshima

Molpadida  
Molpadiidae  
Molpadia intermedia (Ludwig)

Hemichordata  
Enteropneusta



	CCN			CCM			CCS			DDN			DDM			DDS			EEN			EEM	EES			FFM						
	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$		A	B	$\bar{x}$	A	B	$\bar{x}$				
Sigalionidae <u>Pholoe minuta</u>																		10	5					10	5	80	140	110				
Phyllodocidae <u>Eteone (Mysta) sp.</u> <u>E. columbiensis</u> <u>Phyllodoce groenlandica</u>																												10	5			
Pilargidae <u>Ancistrosyllis sp.</u> <u>A. groenlandica</u>							10	5										20	10	10												
Syllidae <u>Exogone sp. 1</u> <u>Exogone sp. 2</u> <u>E. gemmifera</u> <u>Eusyllis cf. blomstrandii</u> <u>Syllis sp.</u> <u>S. alternata</u>																								10	5	30	20	25				
Nephtyidae <u>Aglaophamus malmgreni</u> <u>Nephtys cornuta cornuta</u> <u>N. punctata</u>				10	5		130	70	100	10	5	90	10	5	650	440	545	330						430	690	560	80	530	305			
Sphaerodoridae <u>Sphaerodoropsis sphaerulifer</u>																													10	5		
Goniadidae <u>Goniada annulata</u> <u>Glycinde armigera</u>				10	10	10	30	20	25					10	5												50	20	35			
Lumbrineridae <u>Lumbrineris sp.</u> <u>L. luti</u> <u>Paraninoe simpla</u>				10	5													10	20	15				10								
Dorvilleidae <u>Schistomeringos sp.</u>																														10	5	
Hesionidae <u>Gyptis brevipalpa</u>																		30	40	35	20											
(Sedentariate) Orbinidae <u>Leitoscoloplos pugettensis</u>																														70	10	40









	CCN			CCM			CCS			DDN			DDM			DDS			EEN			EEM	EES			FFM		
	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$		A	B	$\bar{x}$	A	B	$\bar{x}$
Bivalvia																												
Heterodonta																												
Tellinidae																												
<i>Macoma</i> sp.																												
<i>M. carlottensis</i>																												
<i>M. climata</i>																												
<i>M. cf. nasuta</i>																												
Cooperellidae																												
<i>Cooperella</i> sp.																												
Veneridae																												
<i>Tranzenella tantilla</i>		10	5		10	5				30	170	100																
Ungulidae																												
<i>Diplodonta orbella</i>											50	150	100															
Hiatellidae																												
<i>Hiatella arctica</i>																												
<i>Nuculana minuta</i>																												
<i>Nucula tenuis</i>					10	5																						
<i>Yoldia amygdalea</i>																												
<i>Y. beringiana</i>																												
<i>Y. martyria</i>																												
<i>Y. myalis</i>																												
Echinodermata																												
Asteroidea																												
Paxilloridea																												
Goniopectinidae																												
<i>Ctenodiscus crispatus</i>																												
Ophiuroidea																												
Ophiurida																												
Ophiuridae																												
<i>Ophiura</i> sp.																												
<i>O. leptactenia</i>																												
<i>O. sarsi</i>																												
Holothuroidea																												
Apodida																												
Chirodotidae																												
<i>Chirodota albatrossi</i>																												
Molpadida																												
Molpadiidae																												
<i>Molpadia intermedia</i>																												
Hemichordata																												
Enteropneusta																												

\* Sample not taken.

APPENDIX I

DENSITY AND DISTRIBUTION FOR BENTHIC INVERTEBRATES (1.0 mm screen) FROM HASTINGS ARM.  
ALL RESULTS ARE GIVEN IN NUMBERS PER M<sup>2</sup>.

	Z2E			Z2M			Z2W			Z5E			Z5M			Z5W		
	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$
Porifera																		
Hexactinellida																		
Rosellidae																		
Coelenterata																		
Pennatulacea																		
Virgulariidae																		
<u>Virgularia cf. tuberculata</u>		20	10													10	5	
Platyhelminthes																		
Turbellaria																		
Polycladida								10	5									
Nemertea																		
Anopla	10		5	20	10		60	20	40	10	20	15	20	30	25	30	15	
Paleonemertea													20		10	20	10	
Heteronemertea																		
Lineidae		10	5	10	5					10		5						
? <u>Cerebratulus</u>																		
<u>Micrura alaskensis?</u>																		
Enopla																		
Hoplonemertea																		
Nematoda								10	5									
Sipunculida																		
Golfingiidae																		
<u>Golfingia sp.</u>																		
Echiura																		
Annelida																		
Oligochaeta																		
Tubificida																		
Tubificidae																		
<u>Limnodrioides sp.</u>																		
<u>L. victoriensis</u>																		
Polychaeta																		
(Errantiate)																		
Polynoidae																		
? <u>Eunoe</u>		10	5															
<u>Gattyana treadwelli?</u>				10	5					20	10	10		5				

	Z2E			Z2M			Z2W			Z5E			Z5M			Z5W		
	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$
Sigalionidae																		
<u>Phloe minuta</u>		10	5															
Phyllodocidae																		
<u>Eteone (Mysta) sp.</u>																		
<u>E. columbiensis</u>																		
<u>Phyllodoce groenlandica</u>																		
Pilargidae																		
<u>Ancistroyllis sp.</u>																		
<u>A. groenlandica</u>		10	5															
Syllidae																		
<u>Exogone sp. 1</u>																		
<u>Exogone sp. 2</u>																		
<u>E. gemmifera</u>																		
<u>Eusyllis cf. blomstrandii</u>																		
<u>Syllis sp.</u>																		
<u>S. alfernata</u>																		
Nephtyidae																		
<u>Aglaophamus malmgreni</u>	30	30	30	10		5	20	30	25				10		5	10		5
<u>Nephtys cornuta cornuta</u>	570	200	385	180	200	190	520	190	355	140	210	175	420	400	410	300	400	350
<u>N. punctata</u>					20	10				20		10						
Sphaerodoridae																		
<u>Sphaerodoropsis sphaerulifer</u>																		
Goniadidae																		
<u>Goniada annulata</u>	10	30	20	20	20	20	20	40	30	20	30	25	40	30	35		30	15
<u>Glycinde armigera</u>							10		5							20	10	15
Lumbrineridae																		
<u>Lumbrineris sp.</u>					10	5	10		5		20	10				30	20	25
<u>L. luti</u>													20		10		10	5
<u>Paraninoe simpla</u>	20	10	15										10		5			
Dorvilleidae																		
<u>Schistomeringos sp.</u>																		
Hesionidae																		
<u>Gyptis brevipalpa</u>		30	15	20	50	35	50	30	40	20	70	45	100	80	90	100	110	105
(Sedentariate)																		
Orbiniidae																		
<u>Leitoscoloplos pugettensis</u>							10		5				10		5			









	Z2E			Z2M			Z2W			Z5E			Z5M			Z5W		
	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$	A	B	$\bar{x}$
Bivalvia																		
Heterodonta																		
Tellinidae																		
Macoma sp.																10		5
<u>M. carlottensis</u>	30	20	25															
<u>M. climata</u>																	10	5
<u>M. cf. nasuta</u>																		
Cooperellidae																		
<u>Cooperella</u> sp.				70	80	75	70	60	65	10	10	10						
Veneridae																		
<u>Transenella tantilla</u>	50	30	40	80	80	80	140	120	130	170	120	145	20	190	105	40	10	25
Ungulidae																		
<u>Diplodonta orbella</u>	30	60	45	60	70	65		50	25	30	10	20	40	60	50	50	40	45
Hiatellidae																		
<u>Hiatella arctica</u>																		
<u>Nuculana minuta</u>																		
<u>Nucula tenuis</u>	50	90	70	100	190	145	120		60	10	40	25	20	20	20	10		5
<u>Yoldia amydalea</u>	10		5				40	30	35									
<u>Y. beringiana</u>	30	40	35	20	40	30				30	90	60	40	20	30	30		15
<u>Y. martyria</u>		10	5													10		5
<u>Y. myalis</u>																		
Echinodermata																		
Asteroidea																		
Paxilloridea																		
Goniopectinidae																		
<u>Ctenodiscus crispatus</u>	10		5	20		10	20		10	10	20	15		10	5			
Ophiuroidea				40		20												
Ophiurida	40		20		20	10	20		10	20	180	100	30	150	90	100	350	225
Ophiolepididae																		
<u>Ophiura</u> sp.										50	10	30		70	35			
<u>O. leptoctenia</u>							10		5		10	5				10	20	15
<u>O. sarsi</u>																10		5
Holothuroidea																		
Apodida																		
Chirodotidae																		
<u>Chirodota albatrossi</u>	20	10	15		30	15	20	20	20					10	5		20	10
Molpadida																		
Molpadiidae																		
<u>Molpadia intermedia</u>								10	5	10		5				10	10	10
Hemichordata																		
Enteropneusta														10	5			