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Biological, Physical, And Chemical Data From Gulf of
Mexico Core PE0305-GC1

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Introduction

This paper presents benthic foraminiferal census data, and magnetic susceptibility, ^{210}Pb , radiocarbon, and geochemical measurements from gravity core PE0305-GC1 (=GC1). Core GC1 was collected from the Louisiana continental shelf as part of an initiative to investigate the geographic and temporal extent of hypoxia, low-oxygen water, in the Gulf of Mexico. Hypoxia (<1.4 ml/l or <2 ppm oxygen concentration) in Gulf of Mexico waters can eventually lead to death of marine species (Malakoff, 1998). The development of hypoxia off the Mississippi delta has increased steadily since routine and systematic measurements were begun in 1985 and has been linked to the use of fertilizer in the Mississippi basin (Rabalais, 2002). Benthic foraminifers provide a proxy to track the development of hypoxia prior to 1985 (Blackwelder et al., 1996; Sen Gupta et al., 1996). Previous work (Osterman, 2003) determined that the relative occurrence of three low-oxygen-tolerant species is highest in the hypoxia zone (Fig. 1). The cumulative percentage of these three species (% *Pseudonion atlanticum* + % *Epistominella vitrea*, + % *Buliminella morgani* = PEB index of hypoxia) was used to investigate fluctuation in paleohypoxia in four cores, including the upper 60 cm of GC1 (Osterman et al., 2005). In this report, we compile all available data from GC1 as the basis for further publications.

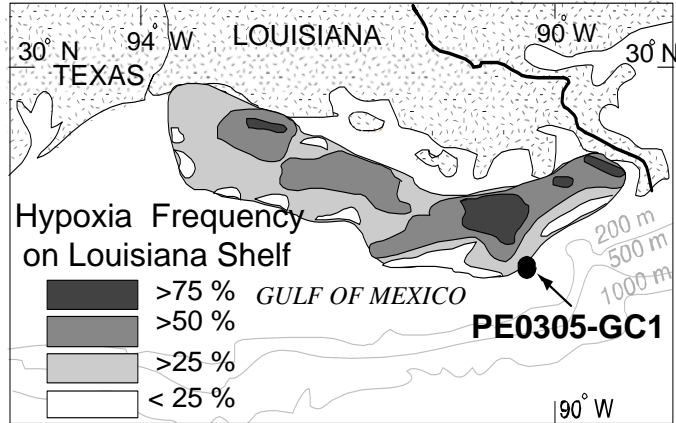


Figure 1. Location map of gravity core GC1 showing the frequency of hypoxia measured at a grid of stations since 1985 by N. Rabalais; taken from CENR (2000).

Methods and Results

Gravity core GC1 was collected at 47 mwd (meters water depth) on the Louisiana continental shelf during R/V *Pelican* cruise PE-03-05 (July 2002). The core was sectioned (at 60 and 120 cm), refrigerated, and transported to Reston, VA. X-radiographs were obtained on the whole core at the Maryland Geological Survey in Baltimore, MD. Digital scans of the x-radiographs are found in Appendix 1.

Whole-core magnetic susceptibility (MS) was measured on GC1 using the Bartington system with an 8-cm loop diameter. The meter was set on cgs (centimeters/gram/second), 0.1 (7 seconds/measurement) scale and later converted to SI (Système Internationale). Background measurements were taken before and after the measurement of each of the three core sections. Because of unreliable measurements near the ends of the core sections, the values from the top and bottom 4 cm of each section were discarded. The MS values are reported in Table 1.

Table 1. Measurement of magnetic susceptibility in Gravity core PE0305-GC1.

depth (cm)	MS value	depth (cm)	MS value
4	203.575	80	316.673
5	213.628	81	327.982
6	218.655	82	331.752
7	222.425	83	334.265
8	221.168	84	335.522
9	219.911	85	344.319
10	218.655	86	363.168
11	216.142	87	382.018
12	214.885	88	388.301
13	211.115	89	387.044
14	209.858	90	376.991
15	207.345	91	371.965
16	208.602	92	368.195
17	212.372	93	368.195
18	214.885	94	374.478
19	217.398	95	389.557
20	218.655	96	431.027
21	218.655	97	466.212
22	218.655	98	443.593
23	219.911	99	382.018
24	221.168	100	346.832
25	213.628	101	327.982
26	224.938	102	319.186
27	224.938	103	311.646
28	224.938	104	314.159
29	222.425	105	319.186
30	224.938	106	324.212
31	228.708	107	326.726
32	233.734	108	326.726
33	237.504	109	327.982
34	241.274	110	329.239
35	242.531	111	330.496
36	242.531	112	330.496
37	241.274	113	327.982
38	242.531	114	325.469
39	251.327	115	324.212
40	262.637	116	319.186
41	267.664	117	311.646
42	256.354	124	304.106
43	246.301	125	314.159
44	241.274	126	317.929
45	242.531	127	321.699
46	245.044	128	327.982
47	251.327	129	334.265
48	256.354	130	349.345
49	261.381	131	376.991
50	267.664	132	407.150
51	266.407	133	437.310
52	257.611	134	448.619
53	241.274	135	412.177
54	237.504	136	370.708
55	234.991	137	348.088
56	231.221	138	336.779
62	234.991	139	330.496
63	247.558	140	317.929
64	248.814	141	309.133
65	248.814	142	299.080
66	246.301	143	296.566
67	247.558	144	295.310
68	248.814	145	295.310
69	252.584	146	294.053
70	256.354	147	292.796
71	262.637	148	290.283
72	270.177	149	287.770
73	272.690	150	285.257
74	277.717	151	285.257
75	282.743	152	281.487
76	286.513	153	277.717
77	290.283	154	276.460
78	295.310	155	277.717
79	306.619	156	280.230
		157	281.487
		158	281.487
		159	282.743
		160	284.000
		161	281.487
		162	272.690

The core was split in half longitudinally. One half was sampled and processed for foraminifers in the U.S. Geological Survey (USGS) Foraminiferal Research Laboratory in Reston, VA, in the fall of 2002. The samples for faunal analyses were soaked in a dilute calgon solution and agitated for 30 minutes to assist in disaggregation, then wet sieved at 63 μm . The washed residue was oven dried at $\leq 50^\circ \text{C}$, then dry sieved at 125 μm . The samples were not dried prior to the wet-sieving process; the calculated dry weights were obtained by measuring the moisture content in a small subsample of each sample and are presented in Table 2.

Samples contained few to abundant benthic foraminifera (3 to 603 foraminifers/gram). When necessary, a representative subsample of approximately 300 specimens was obtained for faunal analysis with a microsplitter (Table 2). Benthic foraminifers were hand picked from the $>125\text{-}\mu\text{m}$ faunal split and placed on standard 60-square micropaleontological slides to be sorted by species. Identification of the benthic foraminifer species was made using standard literature, including the taxonomy of Loeblich and Tappan (1988, 1994), Phleger and Parker (1951), Parker (1954), Bandy (1954), Anderson (1961), Murray (1971, 2000), Bock et al. (1971), Poag (1981), and Morkhoven et al. (1986).

Table 2 records the number of specimens that were counted in each sample. Most specimens were identified to the species level. However, rare species may be grouped with other foraminifers of the same taxonomic level; by family (e.g., Nodosariidae spp.), or by genera (e.g., *Pyrgo* spp.). See Appendix 2 for more information on all identified species and on the groups of rare species. In addition, the number of planktonic foraminifers, ostracods, the *Ammonia* *Elphidium* (AE) Index (Sen Gupta et al., 1996), and the PEB Index (Osterman, 2003) are also reported in Table 2.

Samples for ^{210}Pb were collected from the upper 20 cm of the remaining core half in November 2003. Sediment geochronology of GC1 was derived using multiple radioactive tracers, including excess ^{210}Pb , ^{137}Cs , and $^{239,240}\text{Pu}$ isotopes (Table 3, Fig. 2). A detailed description of the methods utilized in developing a geochronological framework for the core can be found in Swarzenski et al. (2005). Very briefly, ^{210}Pb activities were determined either by alpha counting of its granddaughter, ^{210}Po (half life, $t_{1/2} = 138$ days) (Flynn, 1968), or by gamma-ray spectrometry. Polonium-210 reaches secular equilibrium with ^{210}Pb after approximately two years. Five-gram aliquots of dried and pulverized sediment were leached in a warm $\text{H}_2\text{O}_2/\text{HCl}$ solution, and a known amount of ^{209}Po was added to the pulverized sample as a yield tracer prior to digestion. Polonium was spontaneously electrodeposited onto silver planchets overnight on a warm stirring plate. The overall chemical yield was 83 - 95%, based on an introduced ^{209}Po yield tracer. The Ag planchets were subsequently counted under vacuum on a Si surface-barrier detector (300 mm^2) with an alpha-energy resolution of about 20 keV full-width-at-half maximum (FWHM). The average efficiency for the 5.3 MeV alpha-line of ^{210}Po was estimated at $\sim 20\%$. Supported ^{210}Pb activity was estimated from the mean ^{210}Pb activity within a deeper region of the cores, where total ^{210}Pb activity was relatively constant and assumed to be in secular equilibrium with ^{226}Ra . The mean value was subtracted from the total ^{210}Pb activities at each level to calculate excess ^{210}Pb activity. Supported ^{210}Pb activities for a subset of samples were also compared with ^{226}Ra activities determined by gamma-ray counting ^{214}Bi on a Ge well detector and were found to be within 10% of the sample values. For $^{239,240}\text{Pu}$ analyses, ~ 10 g of dried, powdered sample were leached repeatedly with hot 6 M HC, and the combined leachates were processed for Pu after the addition of a ^{242}Pu yield tracer. Pu was separated and purified using standard ion-exchange techniques (Baskaran et al., 1996). About 10-15 g of dried, powdered

sediment sample were placed in a gamma-counting vial, and specific concentrations of ^{226}Ra were determined using the 351 keV (^{214}Pb) and 609 keV (^{214}Bi) gamma lines for ^{226}Ra and the 661.6 keV line for ^{137}Cs . Either NIST radioactive spikes and material or radioactive standards that are calibrated with NIST standards were used for calibrating the counting equipment. Precision in the activities of ^{210}Pb , $^{239,240}\text{Pu}$, and ^{226}Ra was typically better than 5%.

Table 3. Lead-210 measurements.

Core ID	Depth (cm)	Mean Depth (cm)	Water Content	Loss On Ignition (% Dry Wt.)	Total Pb-210 Activity (dpm/g)	Total Pb-210 Activity Error (+/-)	ln Pb-210 xs (dpm/g)
PE03-05 GC-1	0-1	0.5	45.50	5.72	6.79	0.147	1.915881574
PE03-05 GC-1	1-2	1.5	46.44	5.97	6.22	0.128	1.827083617
PE03-05 GC-1	2-3	2.5	47.16	6.15	5.78	0.119	1.753927176
PE03-05 GC-1	3-4	3.5	46.93	5.83	5.78	0.127	1.755193036
PE03-05 GC-1	4-5	4.5	45.59	6.08	5.44	0.118	1.694164883
PE03-05 GC-1	5-6	5.5	45.52	6.09	5.94	0.127	1.782404
PE03-05 GC-1	6-7	6.5	45.24	5.95	4.92	0.125	1.592310586
PE03-05 GC-1	7-8	7.5	45.91	5.88	4.66	0.107	1.539364112
PE03-05 GC-1	8-9	8.5	45.88	5.60	4.61	0.103	1.527772096
PE03-05 GC-1	9-10	9.5	45.32	5.81	5.40	0.127	1.686187744
PE03-05 GC-1	10-11A	10.5	44.91	5.00	4.51	0.137	1.506789032
PE03-05 GC-1	10-11B	10.5	--	4.00	4.61	0.141	1.527978766
PE03-05 GC-1	10-11C	10.5	--	4.00	4.58	0.146	1.520694639
PE03-05 GC-1	11-12	11.5	45.37	4.40	2.90	0.071	1.065067487
PE03-05 GC-1	12-13	12.5	42.84	4.80	2.01	0.050	0.697804123
PE03-05 GC-1	13-14	13.5	47.81	5.07	1.92	0.053	0.653985353
PE03-05 GC-1	14-15	14.5	48.62	5.60	1.53	0.046	0.423845728
PE03-05 GC-1	15-16	15.5	45.85	5.00	1.71	0.044	0.535763038
PE03-05 GC-1	16-17	16.5	45.81	5.79	2.05	0.057	0.718325104
PE03-05 GC-1	17-18	17.5	46.81	5.50	3.25	0.077	1.178146698
PE03-05 GC-1	18-19	18.5	46.00	5.00	3.39	0.080	1.221843186
PE03-05 GC-1	19-20	19.5	46.49	5.20	3.82	0.094	1.341429152

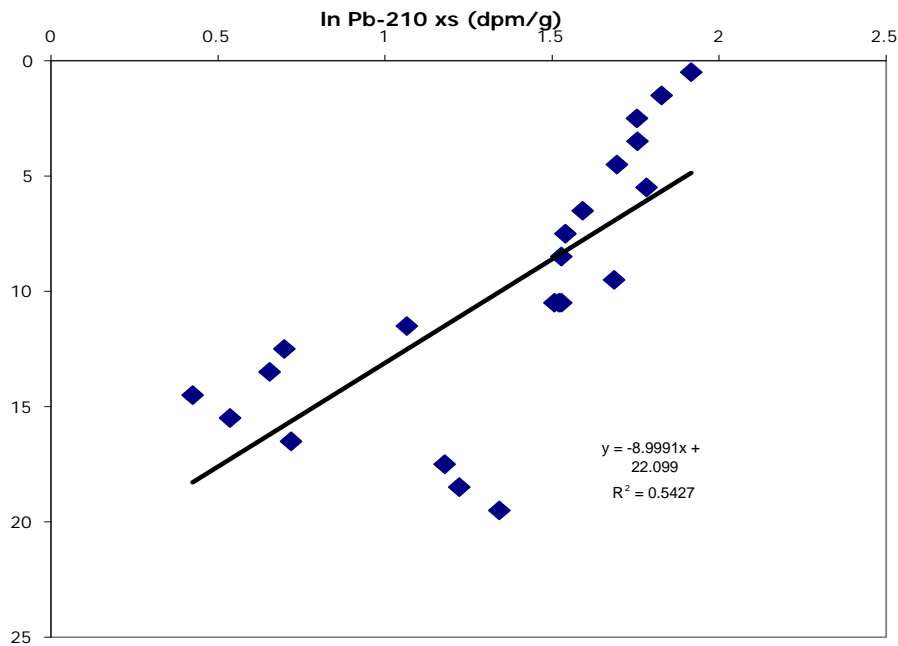


Figure 2. Lead-210 chronology for Gravity core PE0305-GC1.

A separate set of samples was taken in February 2005 from the remaining core half for geochemical analyses. Concentrations of total carbon and total inorganic carbon (IC) were determined using a UIC carbon coulometer (Table 4). A known quantity of each sediment sample (approximately 40 mg) was weighed and placed in a porcelain boat into the carbon-coulometer oven, which combusted the sample at 970°C in the presence of excess oxygen. All of the carbon was oxidized into carbon dioxide to determine the total carbon concentration. A known quantity of each sediment sample was weighed, placed into a glass flask, and acidified in a heated reaction vessel to measure, using 5 mL of 2 M perchloric acid to release all the inorganic carbon. Total organic carbon (TOC) was determined by calculating the difference between the total carbon (released by combustion) minus the total inorganic carbon (generated by acidification). Calcium-carbonate (CaCO_3 , wt. %) concentrations were determined by

dividing the inorganic carbon by the total weight of the sediment sample, and multiplying by a factor of 8.33 to account for carbonate stoichiometry.

Sediment samples for organic carbon and nitrogen stable-isotope analyses were dried at 50°C, powdered, and stored frozen. Powder sediments were acidified in 0.1N HCl to remove carbonate for analysis of the stable isotopic composition of organic carbon ($\delta^{13}\text{C}$) and organic nitrogen ($\delta^{15}\text{N}$). Small samples (>70 μg [micrograms] organic nitrogen) were loaded into tin boats using a microbalance capable of 1- μg resolution. Samples were combusted at 1050°C using a Carlo Erba 2500 series I Elemental Analyzer. The isotopic ratio of the combustion product, CO_2 and N_2 gases, was measured sequentially using a continuous-flow inlet system on a Finnigan Mat Delta Plus XL stable-isotope mass spectrometer. Carbon and nitrogen isotopic values are reported in conventional per mil (‰) notation relative to the VPDB and Air international standards, respectively. All carbon and nitrogen isotopic analyses were run in duplicate or triplicate, and the instrument was calibrated using multiple internal standards including urea, bovine liver, and/or spinach. Standard deviation of the isotopic measurements for organic carbon and nitrogen were better than 0.2‰ and 0.3‰, respectively (Table 4).

A single AMS ^{14}C date of 4195 \pm 35 years on mixed benthic foraminifers was obtained in the lower part of the core (152-157 cm). The graphite target for the date was processed at the ^{14}C laboratory of the (USGS) in Reston, VA, and the ^{14}C age was determined at the Center for Accelerator Mass Spectrometry (CAMS), Lawrence Livermore National Laboratory in Livermore, CA. The quoted age is in radiocarbon years (BP) using the Libby half-life of 5568 years, and no correction has been applied.

Table 4. Geochemical analyses

Sample	% Total C	Inorg. C, wt. %	CaCO ₃ , Wt. %	Org C, wt. %	d 15N, ‰ Air	d 13C, ‰ PDB
3-4	1.27	0.29	2.40	0.98	4.66	-23.24
4-5	1.20	0.34	2.81	0.87	4.13	-25.88
5-6	1.13	0.30	2.51	0.83	-0.98	-21.95
6-7	1.20	0.32	2.64	0.88	3.83	-25.23
7-8	1.13	0.32	2.69	0.81	3.72	-22.04
8-9	1.18	0.33	2.78	0.84	3.46	-22.77
9-10	1.22	0.34	2.79	0.89	0.27	-27.30
10-11	1.14	0.34	2.82	0.80	3.55	-25.55
11-12	1.11	0.28	2.36	0.82	-1.64	-28.92
12-13	1.11	0.27	2.26	0.84	2.84	-23.82
13-14	1.06	0.26	2.18	0.80	-4.96	-23.99
14-15	1.09	0.37	3.12	0.71	3.11	-24.11
15-16	1.24	0.28	2.33	0.96	3.85	-22.49
16-17	1.06	0.29	2.40	0.77	3.76	-26.10
17-18	1.11	0.34	2.85	0.77	4.11	-24.95
18-19	1.22	0.41	3.39	0.81	-2.31	-27.16
19-20	1.03	0.32	2.68	0.71	-2.36	-24.50
20-21	1.07	0.35	2.88	0.73	-5.41	-27.64
21-22	1.59	0.79	6.55	0.80	4.26	-23.84
22-23	1.30	0.55	4.55	0.76	4.69	-25.82
23-24	3.37	0.36	3.02	3.01	3.46	-26.21
24-25	2.29	0.54	4.52	1.74	-5.94	-30.66
25-26	0.99	0.23	1.92	0.76	3.69	-31.98
26-27	0.95	0.26	2.21	0.68	3.77	-24.10
27-28	0.90	0.22	1.86	0.68	3.60	-26.36
28-29	0.89	0.27	2.25	0.62	-3.80	-24.04
29-30	0.96	0.25	2.10	0.71	-0.97	-26.91
30-31	0.92	0.24	1.99	0.68	3.60	-25.48
31-32	0.87	0.27	2.21	0.60	3.82	-24.70
32-33	0.87	0.22	1.81	0.65	3.80	-23.77
33-34	0.83	0.21	1.79	0.62	2.80	-29.08
34-35	0.91	0.20	1.65	0.71	3.37	-28.03
35-36	0.82	0.22	1.87	0.60	3.52	-24.98
36-37	0.78	0.19	1.61	0.59	3.19	-31.20
37-38	0.87	0.25	2.09	0.62	3.81	-24.22
38-39	0.94	0.19	1.58	0.75	-1.72	-24.93
39-40	0.94	0.20	1.69	0.73	3.24	-22.90
40-41	0.83	0.22	1.79	0.61	0.42	-25.24
41-42	0.82	0.22	1.81	0.60	-2.41	-24.64
42-43	0.82	0.21	1.77	0.61	3.57	-27.37
43-44	0.86	0.19	1.61	0.67	3.03	-35.39
44-45	0.81	0.22	1.82	0.59	2.50	-31.17
45-46	0.86	0.19	1.61	0.66	3.62	-23.66
46-47	0.83	0.23	1.89	0.61	3.00	-25.31
47-48	0.88	0.21	1.77	0.67	-2.94	-24.57
48-49	0.88	0.19	1.60	0.68	2.66	-28.41
49-50	0.88	0.21	1.76	0.67	3.08	-29.69
50-51	0.79	0.21	1.72	0.58	0.45	-37.74
51-52	0.78	0.19	1.62	0.59	3.15	-27.05
52-53	0.88	0.18	1.49	0.70	3.59	-27.70
53-54	0.82	0.17	1.45	0.64	2.44	-27.97
54-55	0.87	0.19	1.60	0.68	3.36	-25.31
55-56	0.86	0.18	1.54	0.68	-2.56	-25.40
56-57	0.84	0.22	1.85	0.62	-3.37	-22.44
57-58	1.56	0.19	1.62	1.37	3.34	-30.30
58-59	0.85	0.24	2.01	0.61	3.89	-24.60
59-60	0.82	0.19	1.58	0.63		
60-61	0.84	0.21	1.78	0.63	2.97	-36.83
61-62	0.87	0.23	1.95	0.64	2.73	-25.52
62-63	0.87	0.22	1.87	0.64		
63-64	0.84	0.20	1.65	0.64	3.62	-25.25
64-65	0.90	0.24	1.97	0.66	-1.73	-28.17
65-66	0.85	0.23	1.93	0.61	0.31	-27.32
66-67	0.90	0.22	1.85	0.68	3.45	-24.70
67-68	0.86	0.22	1.86	0.63	-1.78	-26.38
68-69	0.82	0.21	1.75	0.61	3.24	-29.69
69-70	0.89	0.23	1.96	0.65	3.63	-28.16
70-71	0.91	0.22	1.84	0.69	3.63	-24.20
71-72	0.85	0.25	2.07	0.61	3.74	-23.29
72-73	0.86	0.25	2.07	0.61	3.73	-24.16
73-74	0.89	0.28	2.32	0.61	4.02	-23.58
74-75	0.91	0.26	2.15	0.65	2.50	-27.92

Table 4 continued. Geochemical analyses

Sample	% Total C	Inorg. C, wt. %	CaCO ₃ , Wt. %	Org C, wt. %	d 15N, ‰ Air	d 13C, ‰ PDB
75-76	0.89	0.25	2.07	0.64	3.50	-27.97
76-77	0.87	0.25	2.11	0.62	3.18	-26.19
77-78	0.87	0.25	2.06	0.63	3.42	-25.24
78-79	0.85	0.26	2.14	0.60	3.80	-24.62
79-80	0.90	0.23	1.94	0.66	3.28	-26.10
80-81	0.90	0.26	2.20	0.63	2.93	-33.58
81-82	0.91	0.25	2.07	0.67	3.44	-29.10
82-83	0.87	0.23	1.91	0.64	3.84	-23.32
83-84	0.83	0.23	1.88	0.60	-1.01	-25.31
84-85	0.82	0.23	1.95	0.58	1.47	-31.97
85-86	0.85	0.23	1.92	0.62	3.86	-36.46
86-87	0.79	0.20	1.69	0.59	1.95	-30.76
87-88	0.79	0.20	1.69	0.59	3.11	-27.00
88-89	0.83	0.20	1.67	0.63	2.86	-24.95
89-90	0.90	0.20	1.69	0.70		
90-91	0.87	0.21	1.79	0.66		
91-92	0.93	0.18	1.47	0.75	3.58	-25.30
92-93	0.97	0.19	1.61	0.77	3.51	-29.27
93-94	0.98	0.22	1.81	0.76	4.11	-24.05
94-95	0.99	0.26	2.20	0.73	2.51	-28.46
95-96	0.98	0.22	1.84	0.76	4.16	-26.07
96-97	1.05	0.21	1.74	0.84	3.35	-34.64
97-98	0.74	0.22	1.81	0.52	3.71	-25.06
98-99	0.86	0.20	1.63	0.67	3.67	-26.94
99-100	0.87	0.20	1.71	0.67	2.21	-26.55
100-101	0.93	0.20	1.66	0.73	2.86	-23.89
101-102	0.93	0.19	1.58	0.74	3.80	-26.42
102-103	0.87	0.20	1.65	0.67	2.11	-27.33
103-104	0.86	0.21	1.77	0.65	3.74	-23.46
104-105	0.88	0.21	1.74	0.67	3.54	-23.83
105-106	0.91	0.26	2.17	0.65	4.40	-28.46
106-107	0.90	0.28	2.32	0.63	3.66	-22.68
107-108	0.98	0.27	2.26	0.71	2.47	-25.29
108-109	0.94	0.29	2.41	0.65	3.96	-24.01
109-110	0.97	0.23	1.94	0.73	3.34	-24.52
110-111	1.01	0.31	2.54	0.70	3.57	-22.60
111-112	1.06	0.36	2.98	0.70	2.25	-22.71
112-113	0.97	0.40	3.32	0.57	3.60	-26.63
113-114	1.06	0.35	2.89	0.71	4.00	-23.28
114-115	1.06	0.38	3.17	0.68	4.24	-24.84
115-116	1.07	0.39	3.23	0.69	3.65	-24.92
116-117	1.04	0.40	3.30	0.65	4.19	-23.28
117-118	1.02	0.36	3.03	0.66	4.06	-21.53
118-119	1.07	0.41	3.44	0.66	2.74	-21.45
119-120	1.08	0.38	3.14	0.71	4.42	-22.14
120-121	1.05	0.42	3.47	0.64	-1.54	-26.89
121-122	1.10	0.39	3.29	0.70	3.60	-28.05
122-123	1.10	0.39	3.22	0.71	4.07	-23.72
123-124	1.08	0.37	3.06	0.71	5.47	-24.83
124-125	1.06	0.35	2.89	0.71		
125-126	1.05	0.38	3.18	0.67	3.96	-23.31
126-127	1.05	0.34	2.85	0.71	3.33	-26.98
127-128	1.14	0.38	3.20	0.75		
128-129	1.18	0.35	2.90	0.84	3.25	-23.62
129-130	1.28	0.40	3.36	0.87	2.95	-29.84
130-131	1.42	0.39	3.22	1.03	3.64	-22.89
131-132	1.17	0.36	2.98	0.81	2.40	-30.02
132-133	1.00	0.36	3.01	0.64	3.94	-31.03
133-134	0.93	0.33	2.74	0.60		
134-135	1.03	0.35	2.94	0.68	4.09	-23.67
135-136	1.04	0.34	2.87	0.70	2.31	-24.20
136-137	1.14	0.37	3.06	0.77	1.99	-25.56
137-138	1.07	0.26	2.18	0.81	3.75	-23.97
138-139	0.99	0.31	2.54	0.68	2.45	-24.20
139-140	0.96	0.31	2.56	0.65	2.93	-28.42
140-141	1.00	0.29	2.38	0.71	3.09	-24.84
141-142	0.97	0.24	2.02	0.73	4.10	-26.91
142-143	0.94	0.24	2.01	0.70		
143-144	1.04	0.25	2.05	0.79		

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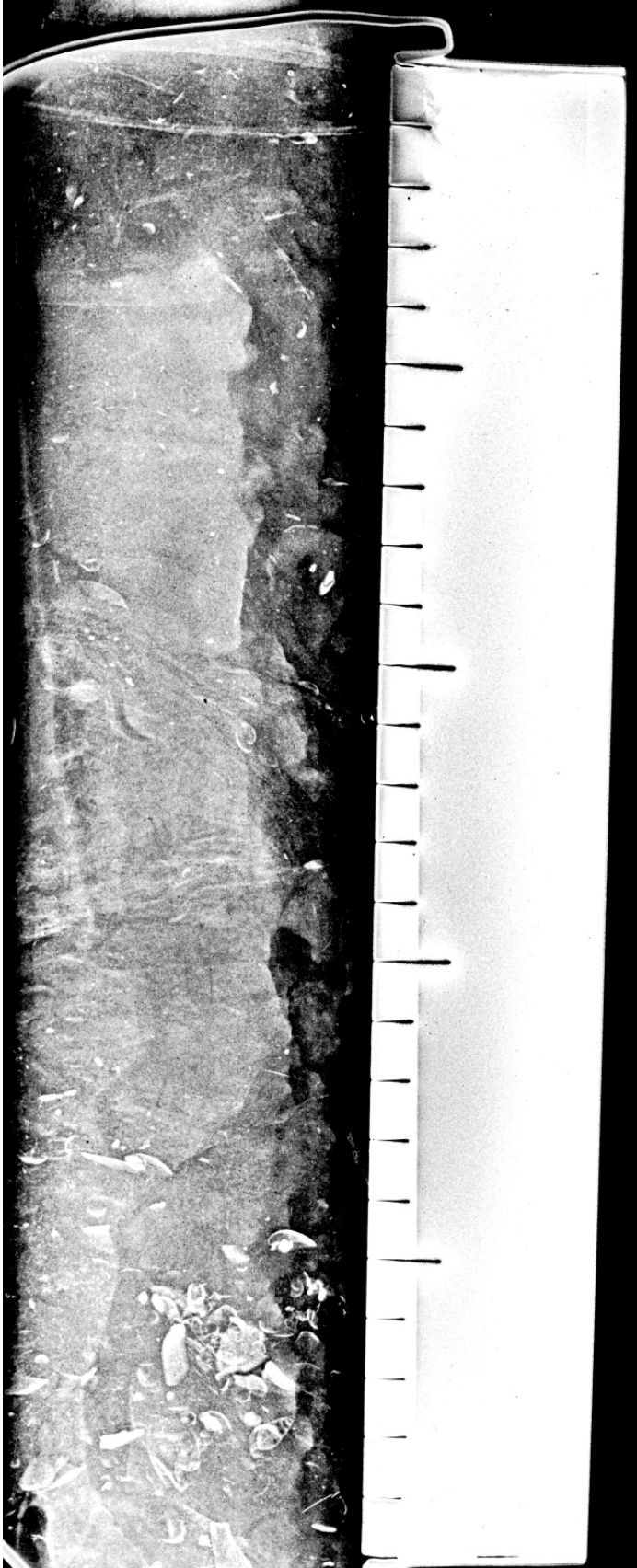
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Appendix 1

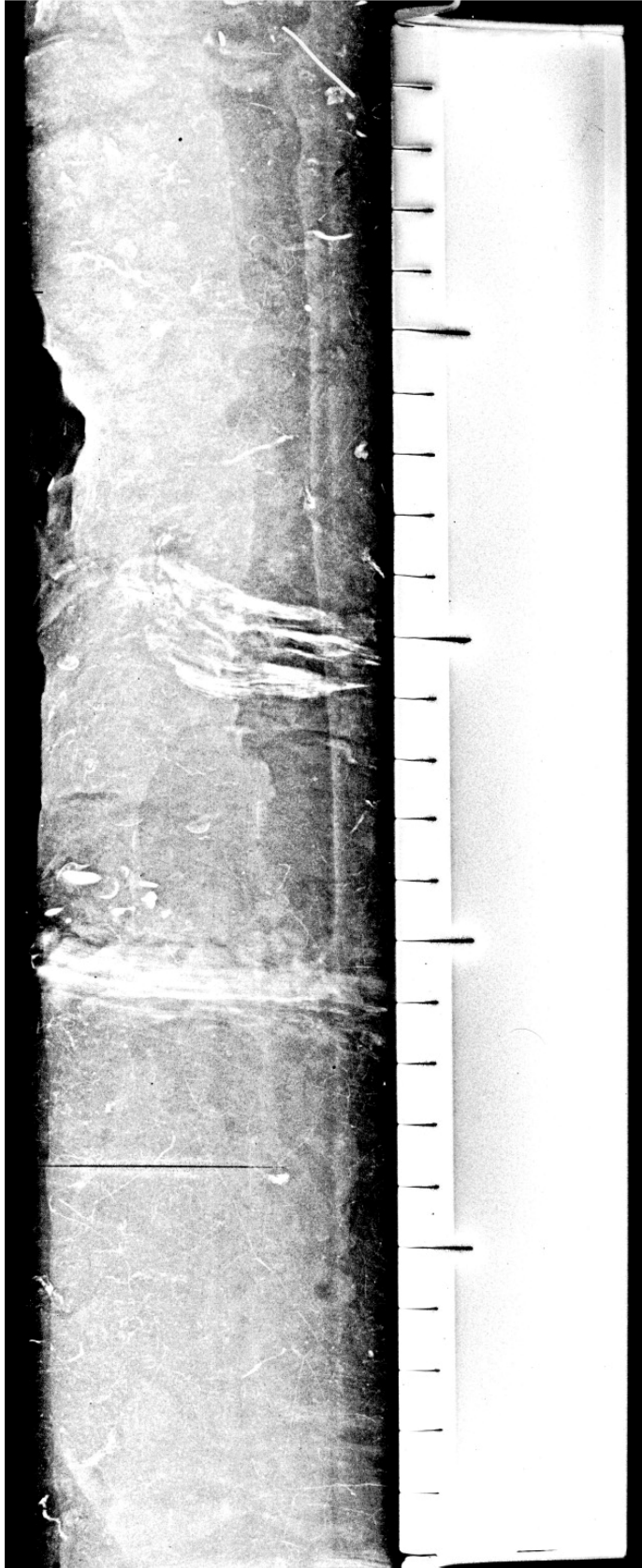
Core PE0305-GC1 X-rays



0 cm

PE03-05 GC1

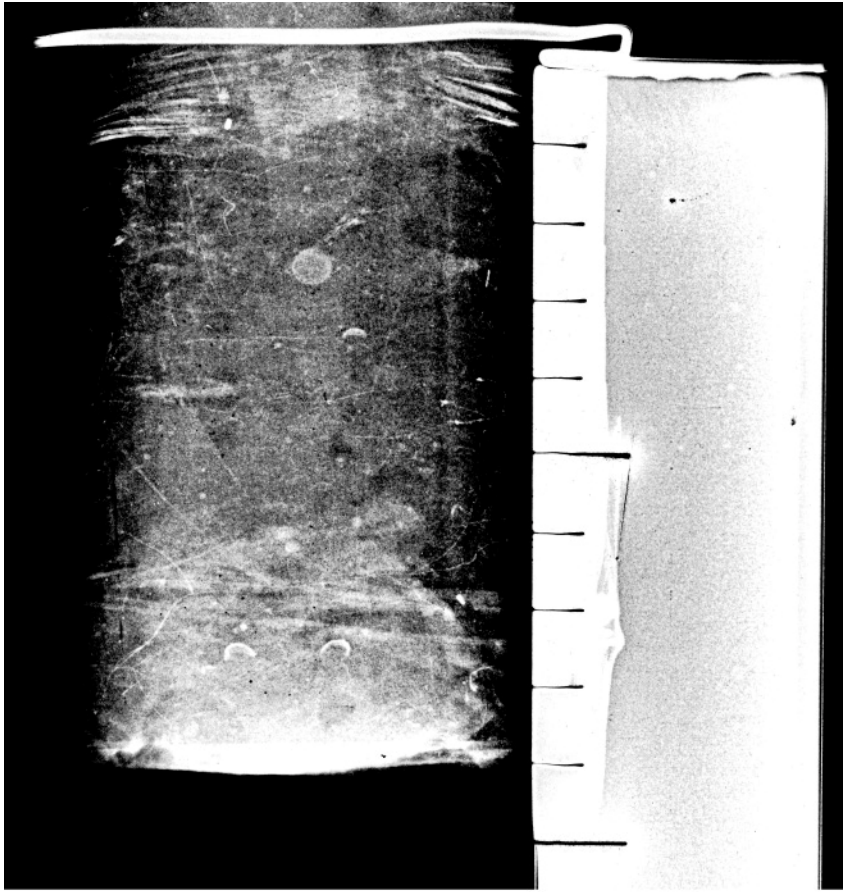
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25 cm

PE03-05 GC1

50 cm



50 cm

60 cm

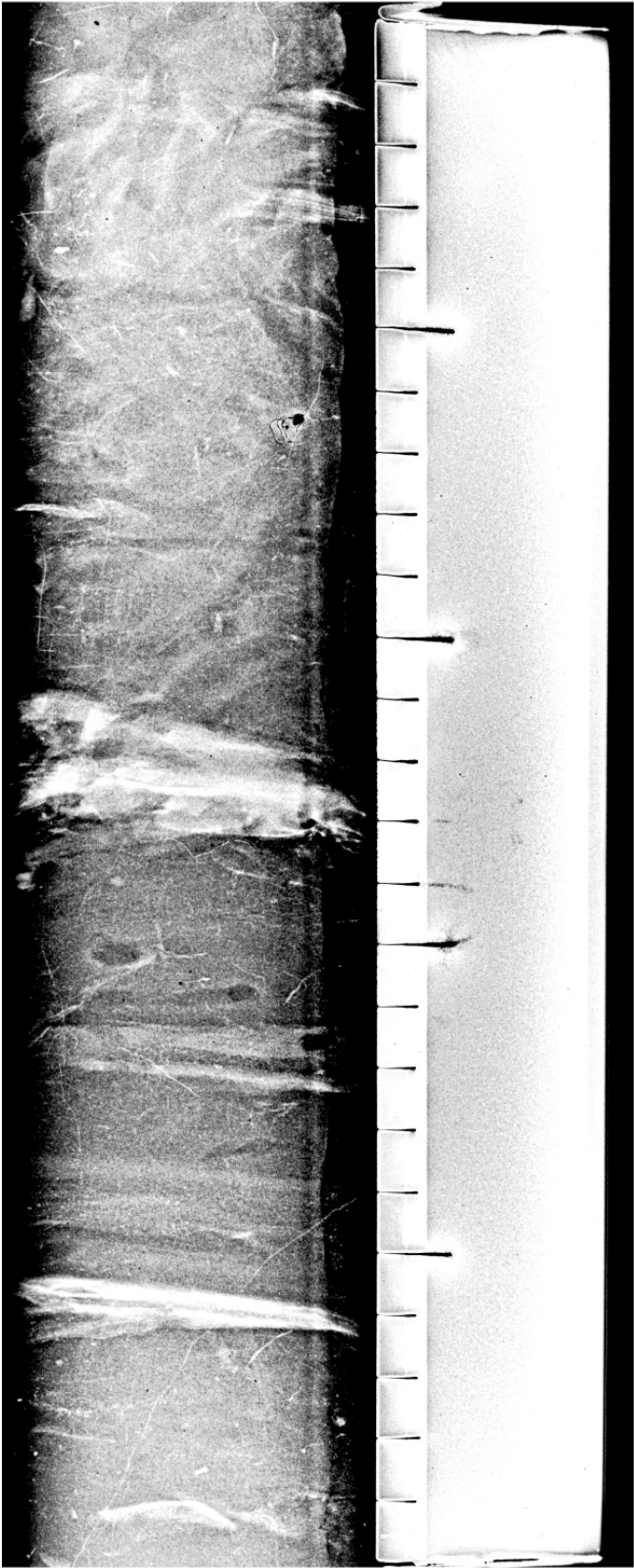
PE03-05 GC1



60 cm

PE03-05 GC1

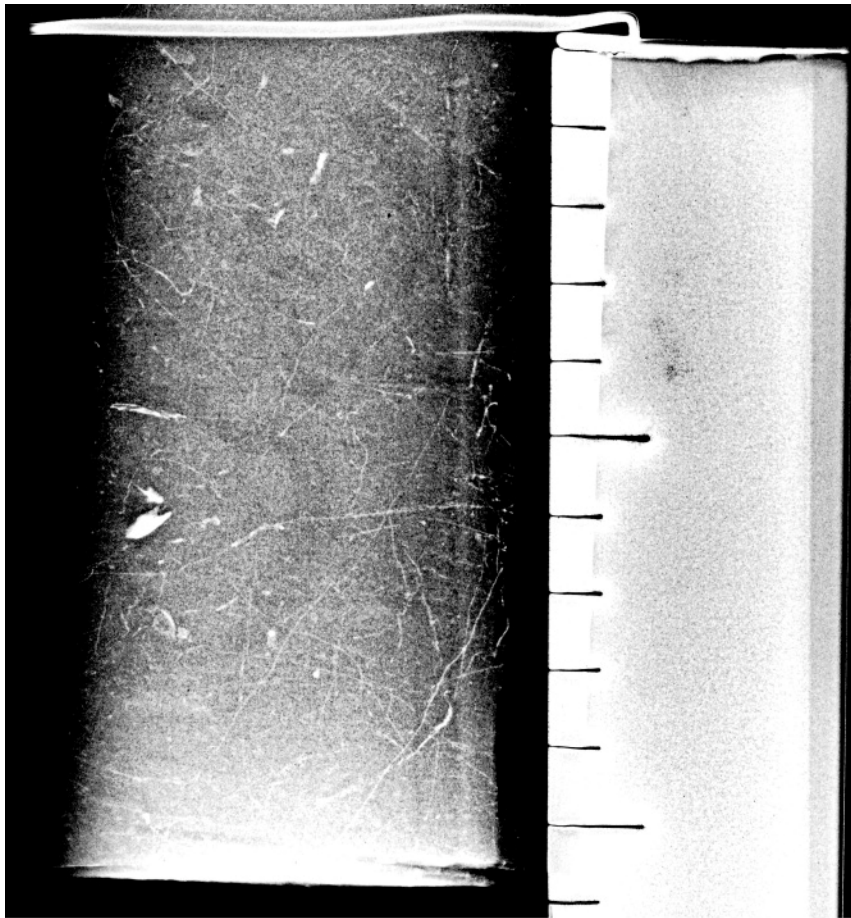
85 cm



85 cm

PE03-05 GC1

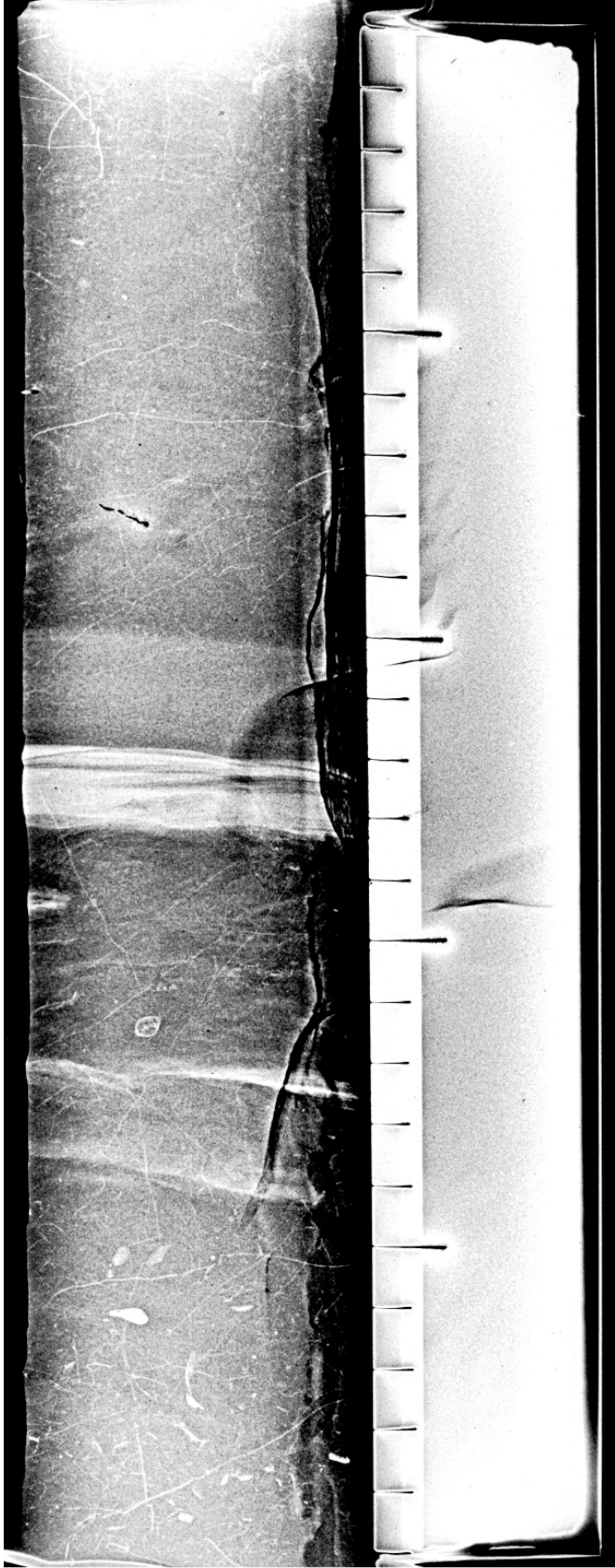
110 cm



110 cm

120 cm

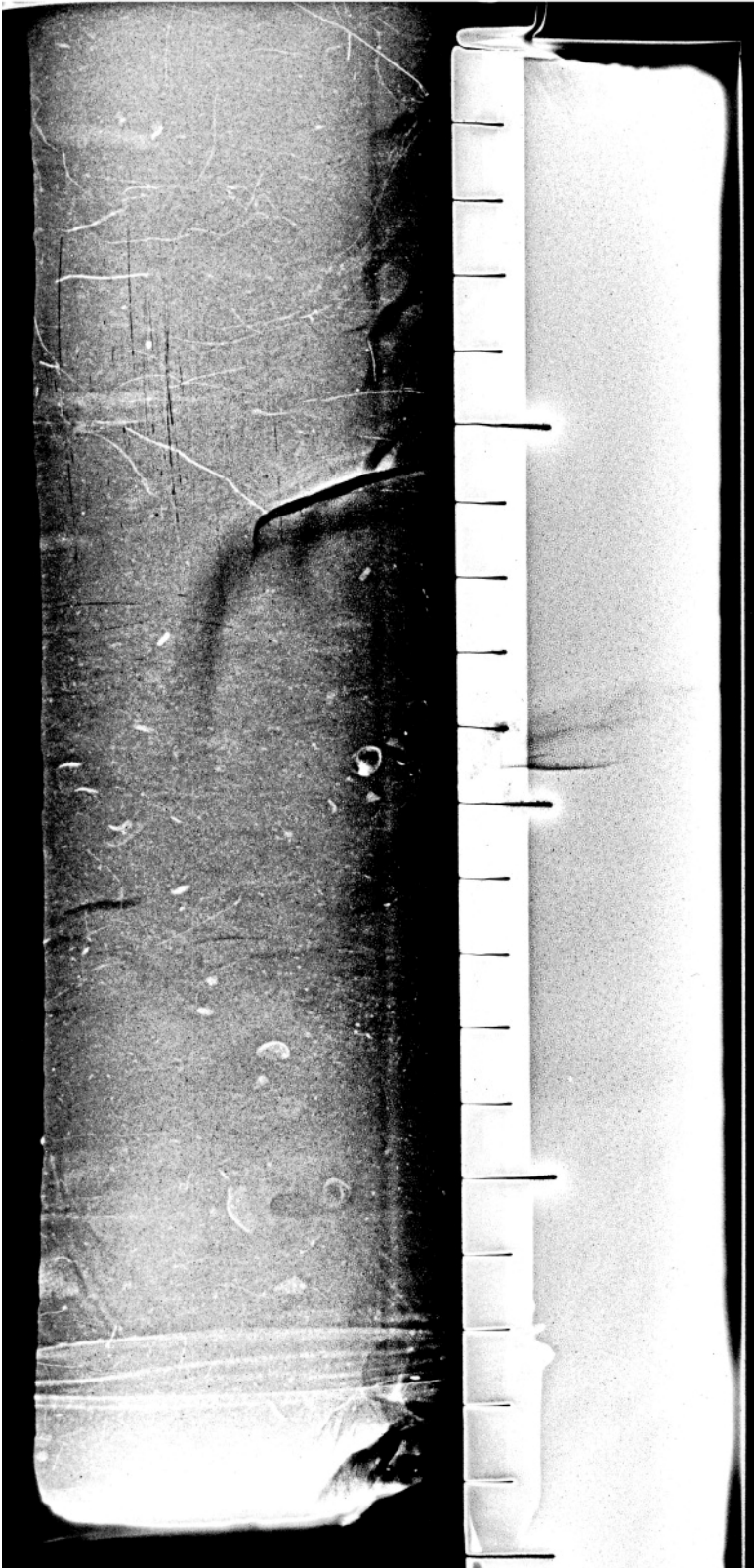
PE03-05 GC1



120 cm

PE03-05 GC1

145 cm



145 cm

PE03-05 GC1

164 cm

APPENDIX 2 GULF OF MEXICO FORAMINIFERS

AGGLUTINATED SPECIES

Ammotium salsum Cushman and Brönniman, 1948, Cush. Found. Foram. Res. Contr., v. 24, pt. 2, p. 39, pl. 7, fig. 9.

Bigeneria irregularis Phleger and Parker, 1951, Geol. Soc. Amer. Mem. no. 46, pt. II, p. 4, pl. 1, figs. 16-21.

Haplophragmoides bradyi (Robertson) = *Trochammina bradyi* Robertson, 1897, Ann. Mag. Nat. His., ser. 6, v. 7 p. 388. fine-grained trochamminid.

Lagenammina difflugiformis (Brady) = *Reophax difflugiformis* Brady, 1879, Notes on some of the Reticularian Rhizopoda of the *Challenger* Expedition: Quarterly J. of Micro. Sci., new series, v. 19, p. 51, pl. 4, figs. 3a,b.

Milliamina horrida (Cushman) = *Quinqueloculina horrida* Cushman, 1947, Contr. Cush. Lab. Foram. Res., v. 23, pt. 88, pl. 19, fig. 1.

Pseudoclavulina mexicana (Cushman) = *Clavulina humilis* Brady var. *mexicana* Cushman, 1922, Bull. 104, U.S. National Museum, pt. 3, p. 83, pl. 16, figs. 1-3. Note: initial part triserial with latter chambers uniserial, often fine grained

Reophanus oviculus (Brady) = *Reophax ovicula* Brady, 1879, Quarterly J. of Micro. Sci., new series, v. 19, p. 20-62.

Reophanus scorpiurus (Montfort) = *Reophax scorpiurus* Brady, 1884, Rept. Voy. *Challenger*, Zool., v. 9, p. 62, pl. 30, figs. 12, 14-17. Note coarse-to-medium grained multichambered.

Reophax spp. = other unidentified, rare forms.

Textularia candeina d'Orbigny, 1839, in Ramone de la Sagra, Hist. Fis. Pol. Nat. Cuba "Foraminiferes," p. 143, pl. 1, figs. 25-27. Note round form with typical textularia aperture

Textularia mayori Cushman, 1922, Pub. 311 Carnegie Inst. Washington, v. 17, p. 23, pl. 2, fig. 3. Note: spikey form found in shallow water.

CALCAREOUS SPECIES

Ammonia parkinsoniana (d'Orbigny) = *Rosalina parkinsoniana* d'Orbigny, 1939, in Ramone de la Sagra, Hist. Fis. Pol. Nat. Cuba "Foraminiferes," p. 99, pl. 4, figs. 25-27.

Amphicoryne hirsute (d'Orbigny) = *Nodosaria hirsute* d'Orbigny, 1826, Ann. Sci. Nat., v. 7, p. 252, n. 7.

Bolivina subspinensis Cushman, 1922, Bull. 104, U.S. National Museum, pt. 3, p. 48, pl. 7, fig. 5.

Brizalina barbata (Phleger and Parker) = *Bolivina barbata* Phleger and Parker, 1951, Geol. Soc. Amer. Mem., no. 46, pt. II, p. 13, pl. 6, figs. 12-13.

Brizalina striatula Cushman var. *spinata* Cushman = *Bolivina striatula* Cushman var. *spinata* Cushman, 1936, Spec. Pub. no. 6, Cush. Lab. Foram. Res., p. 59, pl. 8, figs. 9a,b.

Brizalina subaenariensis Cushman var. *mexicana* Cushman = *Bolivina subaenariensis* Cushman var. *mexicana* Cushman, 1922 Bull. 104, U.S. National Museum, pt. 3, pt. 47, pl. 8, fig. 1

Brizalina translucens (Phleger and Parker) = *Bolivina translucens* Phleger and Parker, 1951, Geol. Soc. Amer. Mem. no. 46, pt. II, p. 15, pl. 7, figs. 13, 14a,b.

Buccella hanni (Phleger and Parker) = *Eponides hanni* Phleger and Parker, 1951, Geol. Soc. Amer. Mem. no. 46, pt. 2, p. 21, pl. 11, figs. 1a,b, 2a,b.

Bulimina marginata d'Orbigny, 1826, Ann. Sci. Nat., v. 7, p. 269, no. 4, pl. 12, figs. 10-12.

Buliminella morgani Anderson, 1961, LA Geol. Bull. no. 35, pt. II, p. 87, pl. 19, fig. 10.

Cancris auriculus (Fichtel and Moll) = *Nautilus auricula* var. β Fichtel and Moll, 1798, p. 108, pl. 20, figs. a-c. NOTE: this is the *Cancris sagra* (d'Orbigny) of Anderson, 1961.

Cassidulina curvata Phleger and Parker, 1951, Geol. Soc. Amer. Mem. no. 46, pt. II, p. 26, pl. 14, figs. 5a,b.

Cassidulina reniforme (Nørvang) = *Cassidulina crassa* d'Orbigny var. *reniforme* Nørvang, 1945, The zoology of Iceland. *Foraminifera*, v. 2, no. 2

Chilostomella oolina Schwager, 1878, Bol. Com. Geol. Ital., v. 9, p. 527, pl. 1, fig. 16. NOTE: included in other calcareous species.

Cibicides spp. included in other calcareous species.

Cibicidoides mundulus (Brady, Parker, and Jones) = *Truncatulina mundula*, Brady, Parker, and Jones, 1888, Zool. Soc. London, Trans. v. 12, pt. 7, no. 1, p. 228, pl. 45, figs. 25a-c.

Cibicidoides pachyderma (Rzehak) = *Truncatulina pachyderma* Rzehak, 1886, Naturf. Ver. Brünn, Verh., Brünn, Bd. 24 (1885), p. 87, pl. 1, fig. 5a-c. (see Morkoven et al., 1986, p. 68-71). NOTE: *Planulina floridana* (Cushman) = *Truncatulina floridana* Cushman, 1919. U.S. Geol. Sur. Bull. 676, p. 62, pl. 19, fig. 2. (see Loeblich and Tappan 1994, p. 149, pl. 312, figs. 9-14)/ *Planulina floridana* (sublittoral) Poag 1981/ *Cibicides pseudoungerianus* (Cushman)/ *Cibicides cicatricosus* (Schwager) in Bock et al., 1971/ *Cibicides umbonatus* (Phleger and Parker, 1951. pl. 17, figs. 7a,b, 8a,b.)

Cornuspira planorbis Schultze, 1854, Organismus Polythal., p. 40, pl. 2, fig. 21.

Dentalina communis d'Orbigny in Brady, 1884, Rept. Voy. *Challenger*, Zool., v. 9, p. 130, pl. 62, figs. 21, 22. Note: most common species included in *Dentalina* spp.

Elphidium discoidale (d'Orbigny) = *Polystomella discoidalis* d'Orbigny, 1839, in Ramone de la Sagra, Hist. Fis. Pol. Nat. Cuba "Foraminiferes," p. 56, pl. 6, figs. 23, 24.

Elphidium excavatum (Terquem) = *Polystomella excavata* Terquem, 1876, Societé Dunquerqueoise, Memoires, v. 19 (1874-75), p. 429. NOTE: included in other *Elphidium* spp.

Elphidium poeyanum (d'Orbigny) = *Polystomella poeyana* d'Orbigny, 1839, in Ramone de la Sagra, Hist. Fis. Pol. Nat. Cuba "Foraminiferes," p. 55, pl. 6, figs. 25, 26. NOTE: included in other *Elphidium* spp.

Epistominella vitrea Parker, in Phleger, Parker, and Peirson, 1953, Cush. Found. Sp. Pub. no. 2, p. 9, pl. 4, figs. 34-36, 40, 41.

Eponides antillarum (d'Orbigny) = *Rotalina antillarum* d'Orbigny, 1839, in Ramone de la Sagra, Hist. Fis. Pol. Nat. Cuba "Foraminiferes," p. 75, pl. 5, figs. 4-6. NOTE: round.

Fursenkoina pontoni (Cushman) = *Virgulina pontoni* Cushman, 1932, Contr. Cush. Lab. Foram. Res., v. 8, pt. 1, p. 17, pl. 3, fig. 7.

Fursenkoina spp. other unidentified *Fursenkoina*.

Gavelinopsis translucens (Phleger and Parker) = "*Rotalia*" *translucens* Phleger and Parker, 1951, Geol. Soc. Amer. Mem. no. 46, pt. 2, p. 24, pl. 12, figs. 11a,b, 12a,b.

Glandulina laevigata (d'Orbigny) = *Nodosaria laevigata* d'Orbigny, 1826, Ann. Sci. Nat., ser. 1, v. 7, p. 252, pl. 10, figs. 1-3.

Globocassidulina subglobosa (Brady) = *Cassidulina subglobosa* Brady, 1881, Quart. J. Micr. Sci., v. 21, pl. 30 fig. 60.

Hanzawaia concentrica (Cushman) = *Truncatulina concentrica* Cushman, 1918, U.S. Geological Sur. Bull. 676, p. 64, pl. 21, fig. 3.

Lenticulina calcur (Linné) = *Nautilus calcur* Linné, 1767, Syst. Nat., 12th ed., p. 1162, no. 272
Note: most common species included in *Lenticulina* spp.

Marginulina marginulinoidea (Göes) = *Çristellaria aculeata* var. *marginulinoidea* Göes, 1896, Bull. Mus. Comp. Zool., v. 29, p. 56, pl. 5, figs. 15, 16. Note: most common form, included in *Marginulopsis* spp.

Miliolinella fichteliana (d'Orbigny) = *Triloculina fichteliana* d'Orbigny, 1839, in Ramone de la Sagra, Hist. Fis. Pol. Nat. Cuba "Foraminiferos," p. 171, pl. 9, figs. 8-10.

Neolenticulina peregrina (Schwager) = *Cristellaria peregrina* Schwager, 1866, *Novara* Expedition, Geol. Thiel, v. 2, p. 245, pl. 7, fig. 89. Note: more commonly referred to as *Lenticulina peregrina*.

Nodosaria albatrossi Cushman = *Nodosaria vertabralis* (Batsch), var. *albatrossi* Cushman, 1923, Bull. 104, U.S. National Mus., pt. 4, p. 87, pl. 15, fig. 1. NOTE: included in *Nodosaria* spp.

Nodosaria pyrula d'Orbigny, 1826, Ann. Sci. Nat., v. 7, no. 13, p. 253. Note: included in *Nodosaria* spp.

***Nodosariidae* spp.** includes assorted species of the genera *Lagena*, *Fissurina* and *Oolina*.

Nonion depressulum (Walker and Jacob) var. *matagordanum* Kornfeld, 1931, Stanford Univ., Dept. Geol., Contr., v. 1, no. 3, p. 87, pl. 13, figs. 2a,b.

Nonionella opima Cushman, 1947, Cont. Cush. Lab. Foram Res., v. 23, pt. 4, p. 90, pl. 20, figs. 1-3. NOTE: occurs rarely and included with *P. atlanticum*.

Oridorsalis umbonatus (Reuss) = *Rotalina umbonatus* Reuss, 1851, Zeitschrift der Deutschen Geologischen Gesellschaft, Berlin, v. 3, p. 75, pl. 5, figs. 35a-c. Note: *O. umbonatus* includes individuals identified as *Eponides tener* by other authors.

Other calcareous other unidentified and rare species.

Other miliolids other unidentified and rare species.

Planulina foveolata (Brady) = *Anomolina foveolata* Brady, 1884, Rept. Voy. Challenger, Zool., v. 9, p. 676, pl. 94, figs. 1a-c. NOTE: included with *Planulina* spp.

Pseudononion atlanticum (Cushman) = *Nonionella atlantica* Cushman, 1947, Cont. Cush. Lab. Foram. Res., v. 23, pt. 4, p. 11, pl. 5, figs. 21-23.

Pseudonodosaria comatula (Cushman) = *Nodosaria comatula* Cushman, 1923, Bull. 104, U.S. Nat. Museum, pt. 4, p. 83, pl. 14, fig. 5.

Pyrgo nasutus Cushman, 1935, Smithsonian Misc. Coll., v. 91, no. 21, p. 7, pl. 3, figs. 1-4.

Pyrgo murrhina (Schwager) = *Biloculina murrhina* Schwager 1866, *Novara* Expedition, Geol. Thiel, v. 2, p. 203, pl. 4, figs. 15a-c.

Pyrgo williamsoni (Silvestri) = *Biloculina williamsoni* Silvestri, 1923, Atti Accad. Pont. Romana Nuovi Lincei, v. 76 (1922-23), p. 73, pl. 6, figs. 169-170.

Quinqueloculina bicarinata d'Orbigny, 1826, Ann. Sci. Nat. v. 7, p. 302. NOTE: this is the triangular-shaped form with no striations.

Quinqueloculina compta Cushman, 1947, Contr. Cush. Lab. Foram. Res., v. 23, pt. 4, p. 87, pl. 19, fig. 2. Note: elongated form.

Quinqueloculina dimidiata Terquem, 1876, Essai sur le classement des animaux que vivent sur la plage et dans le environs le Dunkerque, p. 81, pl. 4, figs. 5a-c.

Rectobolivina advena (Cushman) = *Siphogenerina advena* Cushman, 1922, Carnegie Inst., Washington, Pub. 311, p. 35, pl. 5, fig. 2.

Reussella spinulosa (Reuss) = *Verneuilina spinulosa* Reuss, 1850, Denkschriften Kaiser. Akad. Der Wissensch. Math. - Natur. Classe. 1, 374. NOTE checked *Reussella spinulose* (Reuss) var. *atlantica* Cushman, 1947, Cont. Cush. Lab. Foram. Res., v. 23, pt. 4, p. 91, pl. 20, figs. 6-7, but I believe it to be the same.

Rosalina suezensis = *Rosalina bahamensis* Todd and Low, 1971, U.S. Geol. Sur., Prof. Paper 683-C, p. C14, pl. 3, fig. 2.

Saracenaria spp. assorted species of this genera.

Siphonina pulchra Cushman, 1919, Publ. 291, Carnegie Inst. Washington, p. 42, pl. 14, figs. 7a-c.

Spirolaccamina spp. assorted species of the genera.

Trifarina bella (Phleger and Parker) = *Angulogerina bella* Phleger and Parker, 1951, Geol. Soc. Amer. Mem. no. 46, pt. 2, p. 12, pl. 6, figs. 7, 8.

Triloculinella tegminis (Loeblich and Tappan) = *Scutuloris tegminus* Loeblich and Tappan, 1953, Studies of Arctic Foraminifera, Smithsonian Misc. Coll., v. 121, no. 7, p. 41, pl. 6, fig. 10; includes *Miliolinella chukchiensis* (Loeblich and Tappan) 1953, pl. 7, fig. 7 *Quinqueloculina subrotunda* (Montague) Todd and Low, 1967; and *Quinqueloculina subrotunda* (Montague?) Cushman, 1948.

Uvigerina peregrina Cushman, 1923, Bull. 104, U.S. Nat. Museum, pt. 4, p. 166, pl. 42, figs. 7-10.

Valvulina laevigata Phleger and Parker, 1951, Geol. Soc. Amer. Mem. no. 46, pt. 2, p. 25, pl. 13, figs. 11a,b, 12a,b.