# Summary of Information on Aquatic Biota and Their Habitats in the Willamette Basin, Oregon, through 1995

By BOB ALTMAN, AVIFAUNA NORTHWEST; COLLEEN M. HENSON, U.S. FISH AND WILDLIFE SERVICE; AND IAN R. WAITE, U.S. GEOLOGICAL SURVEY

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# U.S. DEPARTMENT OF THE INTERIOR BRUCE BABBITT, Secretary

U.S. GEOLOGICAL SURVEY Gordon P. Eaton, Director

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For additional information write to:

E-mail: info-or@usgs.gov

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from:

Box 25286

District Chief U.S. Geological Survey 10615 South East Cherry Blossom Drive Portland, Oregon 97216

Denver, CO 80225-0286 Telephone: (303) 202-4210

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#### **FOREWORD**

The mission of the U.S. Geological Survey (USGS) is to assess the quantity and quality of the earth resources of the Nation and to provide information that will assist resource managers and policymakers at Federal, State, and local levels in making sound decisions. Assessment of water-quality conditions and trends is an important part of this overall mission.

One of the greatest challenges faced by waterresources scientists is acquiring reliable information that will guide the use and protection of the Nation's water resources. That challenge is being addressed by Federal, State, interstate, and local water-resource agencies and by many academic institutions. These organizations are collecting water-quality data for a host of purposes that include: compliance with permits and water-supply standards; development of remediation plans for specific contamination problems; operational decisions on industrial, wastewater, or watersupply facilities; and research on factors that affect water quality. An additional need for water-quality information is to provide a basis on which regionaland national-level policy decisions can be based. Wise decisions must be based on sound information. As a society we need to know whether certain types of water-quality problems are isolated or ubiquitous, whether there are significant differences in conditions among regions, whether the conditions are changing over time, and why these conditions change from place to place and over time. The information can be used to help determine the efficacy of existing waterquality policies and to help analysts determine the need for and likely consequences of new policies.

To address these needs, the U.S. Congress appropriated funds in 1986 for the USGS to begin a pilot program in seven project areas to develop and refine the National Water-Quality Assessment (NAWQA) Program. In 1991, the USGS began full implementation of the program. The NAWQA Program builds upon an existing base of water-quality studies of the USGS, as well as those of other Federal, State, and local agencies. The objectives of the NAWQA Program are to:

- Describe current water-quality conditions for a large part of the Nation's freshwater streams, rivers, and aquifers.
- Describe how water quality is changing over time.

 Improve understanding of the primary natural and human factors that affect water-quality conditions.

This information will help support the development and evaluation of management, regulatory, and monitoring decisions by other Federal, State, and local agencies to protect, use, and enhance water resources.

The goals of the NAWQA Program are being achieved through ongoing and proposed investigations of 60 of the Nation's most important river basins and aquifer systems, which are referred to as study units. These study units are distributed throughout the Nation and cover a diversity of hydrogeologic settings. More than two-thirds of the Nation's freshwater use occurs within the 60 study units and more than two-thirds of the people served by public water-supply systems live within their boundaries.

National synthesis of data analysis, based on aggregation of comparable information obtained from the study units, is a major component of the program. This effort focuses on selected water-quality topics using nationally consistent information. Comparative studies will explain differences and similarities in observed water-quality conditions among study areas and will identify changes and trends and their causes. The first topics addressed by the national synthesis are pesticides, nutrients, volatile organic compounds, and aquatic biology. Discussions on these and other water-quality topics will be published in periodic summaries of the quality of the Nation's ground and surface water as the information becomes available.

This report is an element of the comprehensive body of information developed as part of the NAWQA Program. The program depends heavily on the advice, cooperation, and information from many Federal, State, interstate, Tribal, and local agencies and the public. The assistance and suggestions of all are greatly appreciated.

Robert M. Hersch

Robert M. Hirsch Chief Hydrologist

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## CONVERSION FACTORS, ABBREVIATED WATER-QUALITY AND TOXICOLOGICAL UNITS, FREQUENTLY REFERENCED STREAMS AND LAKES, AND ACRONYMS AND ABBREVIATIONS

Multiply	Ву	To Obtain
acre	4,407	square meter
foot (ft)	0.3048	meter
inch (in.)	25.4	millimeter
mile (mi)	1.609	kilometer
square mile (mi <sup>2</sup> )	2.59	square kilometer

Temperature in degrees Fahrenheit (°F) can be converted to degrees Celsius (°C) as follows:

#### **Abbreviated Water-Quality and Toxicological Units**

mean
less than
greater than
alpha
beta
gamma
delta

μg/g micrograms per gram

nmol/mg-protein/min nanomoles per milligram protein per minute

pg/g picogram per gram

#### **Frequently Referenced Streams and Lakes**

Stream	Subbasin
Berry Creek	Luckiamute
Conser Slough	Willamette River
Johnson Creek	Willamette River
Oak Creek	Marys
Rock Creek	Tualatin

#### **Acronyms and Abbreviations**

AHH	Aryl hydrocarbon (benzo[a]pyrene) hydrolase
Al	Aluminum
As	Arsenic
Ba	Barium
Be	Beryllium
BLM	Bureau of Land Management
Cd	Cadmium
Cr	Chromium
Cu	Copper
DDD	Dichlorodiphenyldichloroethane
DDE	Dichlorodiphenyldichloroethylene
DDT	Dichlorodiphenyltrichloroethane
EPT	Ephemeroptera, Plecoptera, Trichoptera
EROD	Ethoxyresorufin O-deethylase
FDA	Food and Drug Administration
Fe	Iron
FEMAT	Forest Ecosystem Management Assessment Team
HCB	Hexachlorobenzene
HCBP	Hexachlorobiphenyl
HCH	Hexachlorocyclohexane
Hg	Mercury
HpCDD	Heptachlorodibenzo-p-dioxin
HpCDF	Heptachorodibenzofuran
HxCDD	Hexachlorodibenzo-p-dioxin

#### Acronyms and Abbreviations—Continued

HxCDF Hexachlorodibenzofuran

MgMagnesiumMnManganeseMoMolybdenum

NAWQA National Water-Quality Assessment

NCBP National Contaminant Biomonitoring Program

Ni Nickel

NMFS National Marine Fisheries Service

ODEQ Oregon Department of Environmental Quality
ODFW Oregon Department of Fish and Wildlife
ORIS Oregon Rivers Information System

OSU Oregon State University

OWRD Oregon Water Resources Department
PAH Polycyclic aromatic hydrocarbon

Pb Lead

PCA Pentachloroamisole
PCB Polychlorobiphenyl
PCBP Pentachlorbiphenyl

PCDD Polychlorodibenzo-p-dioxin
PCDF Polychlorodibenzofuran
PeCDD Pentachlorodibenzo-p-dioxin
PeCDF Pentachlorodibenzofuran

PHH Planar halogenated hydrocarbons

PSU Portland State University

RM River mile Se Selenium

TCBP Tetrachlorobiphenyl
TCDD Tetrachlorodibenzo-p-dioxin
TCDF Tetrachlorodibenzofuran

TEC Toxicity Equivalency Concentration

Tl Thallium

USACE U.S. Army Corps of Engineers

USEPA U.S. Environmental Protection Agency

USFS U.S. Forest Service

USFWS U.S. Fish and Wildlife Service USGS U.S. Geological Survey

WRBWQS Willamette River Basin Water Quality Study

WRTS Willamette River Toxics Study

Zn Zinc

# A Review of Aquatic Biological and Habitat Information in the Willamette Basin, Oregon, through 1995

By Bob Altman, Colleen M. Henson, and Ian R. Waite

#### **ABSTRACT**

Available information on aquatic biota of the Willamette Basin was reviewed and summarized to describe current and historical conditions as part of the U.S. Geological Survey's National Water-Quality Assessment Program. Biological parameters emphasized include the status, distribution, and trends of aquatic biota, particularly algae, macroinvertebrates, and fish; the condition of aquatic and riparian habitat in which these biota reside; and the response of these biota to natural and human-associated impacts, including the level, type, and effect of contaminants.

Considerable data are available on aquatic biota in the Willamette Basin, although the information is highly uneven relative to taxa and spatial scope. Extensive information exists for highprofile taxa, such as salmonid fishes, but less information is available for macroinvertebrates, and relatively little data exist for algae. Additionally, some areas such as the H.J. Andrews Experimental Forest and the main stem Willamette River have been extensively studied, whereas data are limited for many other areas.

The basin supports a diverse aquatic macroinvertebrate fauna. Available data indicate a relatively high diversity of taxa and a high richness of Ephemeroptera, Plecoptera, Trichoptera (EPT) taxa in the upper reaches of the basin. In the lower main stem reaches, macroinvertebrate assemblages are dominated by pollution tolerant organisms and those adapted to low dissolved oxygen levels. Most of the limited data on algae are from sampling in the main stem Willamette River. Diatoms and blue-green algae are the dominant algal forms.

Approximately 61 fish species occur in the basin, although nearly half are introduced. Species richness and distribution are highly correlated with elevation, stream gradient, and water temperature. High elevation, cold water, mountain streams are characterized by a few species of salmonids, sculpins, suckers, and whitefish. Low elevation, main stem reaches of major rivers and streams are dominated by warm water species, such as bass, catfish, and several species in the panfish group. The only species of fish listed as threatened or endangered is the Oregon chub (*Oregonichthys crameri*).

The effect of an expanding human presence in the Willamette Basin has substantially altered aquatic and riparian habitats, and the biota that use or reside in these habitats. Construction of dams, channelization and bank stabilization of rivers, species introductions, supplementations of fisheries through aquaculture, timber harvesting, agricultural activities, and urbanization have contributed to changes in aquatic habitats and biota from historical conditions.

Aquatic toxicological investigations in the basin have focused primarily on fish. These studies have addressed chlorinated pesticides, polychlorinated biphenyls (PCBs), dioxins and furans, polycyclic aromatic hydrocarbons (PAHs), and trace elements in aquatic tissue, as well as fish health assessments, skeletal abnormalities, and aquatic toxicological responses. Several pesticides exceeded U.S. Environmental Protection Agency and State water-quality criteria for the protection

of aquatic life. Elevated PCB, dioxin, and furan concentrations were associated with point sources, such as pulp and paper mills. Elevated concentrations of mercury in aquatic tissue were associated with several reservoirs. Fish health assessments and skeletal abnormality studies detected high levels of abnormalities in fish from the main stem Willamette River. Few investigations have examined aquatic toxicological responses, such as enzyme induction assays, growth assays, and biomarker studies.

#### INTRODUCTION

The National Water-Quality Assessment (NAWQA) Program was initiated in 1991 by the U.S. Geological Survey (USGS) to (1) describe the status and trends of water quality of a representative portion of the Nation's surface- and ground-water resources and (2) provide a sound, scientific understanding of the primary natural and human factors affecting the quality of these resources (Cohen and others, 1988; Hirsch and others, 1988; Leahy and others, 1990; Wentz and McKenzie, 1991). The program is designed to provide nationally consistent and technically sound water-quality information useful to water managers, local policy makers, and the general public.

The NAWQA Program incorporates standardized approaches and protocols for collection of data on physical, chemical, and biological components in a multidisciplinary, integrated assessment of water quality across a wide range of spatial scales. The principal study units are hydrological basins or aquifer systems that provide information at a regional scale, opportunities for comparisons among study units, and a mechanism to synthesize data for multiple study units on a national scale. The Willamette Basin, which includes the Willamette and Sandy River Basins (fig. 1), was selected as one of the first 20 NAWQA study units for full-scale implementation.

An important component of the NAWQA Program is a retrospective analysis that reviews and summarizes information on various constituents associated with water quality. The current report on aquatic biological information complements similar retrospective reports on physical and chemical constituents conducted as part of the assessment of water quality and aquatic ecosystem health in the Willamette Basin. This report will be useful in evaluating the NAWQA study

design in terms of selection of sampling locations and biological constituents most important for understanding water-quality conditions from a basinwide perspective. Gurtz (1993) summarizes the reasons for including biological components in the NAWQA Program.

## Rationale for a Review of Biological Information

Protection and enhancement of water quality and aquatic biota are considered to be critical long-term resource management issues in Oregon (Oregon Department of Environmental Quality, 1990). The Clean Water Act of 1972 is the regulatory driving force to "...restore and maintain the chemical, physical, and biological integrity of the Nation's waters...". The U.S. Environmental Protection Agency (USEPA) is the Federal agency and the Oregon Department of Environmental Quality (ODEQ) is the State agency responsible for administration of the act to ensure the availability of clean water for beneficial uses such as recreation (fishing, swimming, boating), drinking, navigation, hydroelectric power, agriculture, and fish and wildlife habitat. To address this responsibility, the ODEQ is required to prepare a report every 2 years assessing the status of water quality in the State. Specific information on the status and trends of water quality in the Willamette Basin is included in the most recent report (Oregon Department of Environmental Quality, 1994a).

The quality of surface water in the Willamette Basin is dependent upon numerous natural and human-associated factors. Changes that affect the physical, chemical, or biological processes in surface water can cause changes in the biological communities. Thus, measuring condition and change of biological communities provides an index of surface-water quality (Mulvey and others, 1992).

Development of biological criteria for stream habitats is a useful means of assessing water quality (Karr, 1991). Biological criteria are measurements of ecological and physiological characteristics of organisms and communities that can be used to assess the biological integrity of a stream relative to a "reference stream" that has been minimally impacted by human activities (Hughes and others, 1986; Plafkin and others, 1989). The use of biological criteria in bioassessments of aquatic ecosystems is integral to the USEPA Rapid Bioassessment Protocol (Plafkin and others, 1989) and

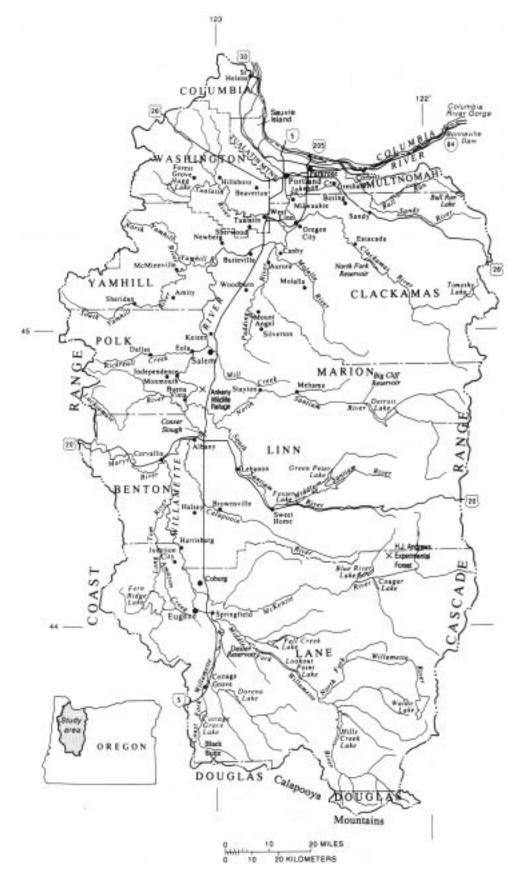


Figure 1. Location of the Willamette Basin, Oregon.

other programs, including those used by the ODEQ (Mulvey and others, 1992). Additionally, biological criteria have been used in protocols for monitoring of wadable streams in the Pacific Northwest (Hayslip, 1993).

Assessment of water quality using biological criteria is based on an analysis of multiple metrics. A metric is a characteristic of biota that changes in some predictable way with perturbations in human or natural influences (Barbour and others, 1995). The metrics used for macroinvertebrates in the Willamette River Basin Water Quality Study (WRBWQS) include measures of species and community richness, composition, tolerance, and trophic levels (Tetra Tech, Inc., 1992b; 1994). Examples include percent Ephemeroptera, Plecoptera, Trichoptera (EPT) taxa, number of Chironomidae taxa, percent Oligochaeta, and percent scrapers.

#### **Purpose and Scope**

This report reviews and summarizes available information on aquatic biological communities in the Willamette Basin through 1995. Specifically, the report describes (1) the distribution, abundance, and trends of three taxonomic groups—algae, macroinvertebrates, and fish—and, to a lesser degree, other selected semiaquatic taxa (i.e., taxa frequenting but not living wholly in water), including amphibians and reptiles, birds, and mammals, (2) the types of aquatic and riparian habitat that support and influence aquatic communities and their biotic constituents, (3) the natural and humanassociated impacts on that habitat and associated biological communities, and (4) the levels of environmental contaminants to which biological communities and specific biota are exposed. The information in this report is intended to aid in the identification of data gaps relative to taxa and geographic areas, and to stimulate collaboration and increase coordination in present and future ecological research in the Willamette Basin.

For consistency with other investigations conducted as part of the Willamette Basin NAWQA Program, biological data have been reviewed for the entire Sandy River Basin and for the Willamette River Basin upstream from river mile (RM) 12.8 (Morrison Street Bridge). The latter site, known as the Willamette River at Portland, is a long-term streamflow and waterquality data collection site sampled by the USGS and ODEQ (Bonn and others, 1995). Downstream from

RM 12.8 to the mouth of the Willamette River at Kelly Point Park, only data from the main stem have been considered. Thus, data from the Multnomah Channel and from the Columbia Slough are not included in this review.

#### **Sources of Information**

Numerous sources were contacted and documents reviewed for this report. Readily available technical reports and environmental documents from government agencies and nongovernmental organizations were a principal source of data. Additionally, an attempt was made to acquire in-house and unpublished agency reports and available consultant reports. To facilitate this effort, assistance was solicited via a letter request and phone calls to appropriate individuals on potential sources of information.

Another source of data included student theses and dissertations, and research reports of university faculty and staff from Portland State University (PSU), University of Oregon, and, particularly, Oregon State University (OSU). At OSU, researchers from several programs, including the Cooperative Wildlife and Fisheries Units of the U.S. Fish and Wildlife Service (USFWS), the Systematic Entomology Laboratory, the Water Resources Research Institute, and the multidisciplinary "Stream Team" have conducted studies throughout the Willamette Basin for a number of years.

Several reports were used that reviewed aquatic biota at different scales. These documents include a status and trends report for fauna of the Pacific Northwest (Smith and Collopy, in press), a review of biota in the Tualatin subbasin (Li and Gregory, 1993), bibliographies of research publications from the H.J. Andrews Experimental Forest (HJAEF) in the McKenzie subbasin, and compilations of invertebrate occurrences at the HJAEF (Anderson and others, 1982; Parsons and others, 1991) and Berry Creek in the Luckiamute subbasin (Anderson and Hansen, 1987).

Most of the information on fish of the Willamette Basin was obtained from research, monitoring, and investigative studies conducted by the Oregon Department of Fish and Wildlife (ODFW). This information was documented in reports from their Processed Report, Information Report, and Progress Report series, and from other ODFW publications.

Principal data bases used in this report include the Oregon Rivers Information System (ORIS) for fish species distribution (Oregon Department of Fish and Wildlife, 1994), a USFWS data base on threatened and endangered species, and the Oregon Natural Heritage Program data base, which houses information on the occurrence and distribution of rare, threatened, and endangered plants and animals in Oregon (Oregon Natural Heritage Program, 1995). Additionally, a literature search was conducted of appropriate scientific publications and several agency publication data bases.

#### **Background Studies**

Several large-scale studies have been conducted that provide information on the aquatic biological resources of the Willamette Basin. The Willamette River Environmental Survey was conducted in 1958 and 1959 by the Fish Commission of Oregon to determine environmental conditions detrimental to anadromous fish runs in the Willamette River system (Willis and others, 1960). This study provides a detailed account of the physical habitat features, pollution problems, obstructions to fish passage, and fish species present for 17 major river systems and their tributaries.

The Oregon State Game Commission's Basin Investigations Section conducted field work in the 1960s to define water problems and needs associated with Willamette Basin fish and wildlife resources. The results of these investigations were documented in three reports: Lower Willamette Basin (Hutchison and Aney, 1964), Middle Willamette Basin (Oregon State Game Commission, 1963), and Upper Willamette Basin (Hutchison and others, 1966a). Another comprehensive review of the aquatic resources within the basin was provided by the Willamette Basin Comprehensive Study (Willamette Basin Task Force, 1969).

The USGS conducted an extensive water-quality assessment program in the early 1970s "...to develop and document methods for evaluating basin-development alternatives in terms of potential impacts on water quality..." (Rickert and Hines, 1975). This study focused on (1) dissolved oxygen depletion, (2) algal problems, (3) trace element occurrence, and (4) the impact of land-use activity on erosion. Results of the study were published as USGS Circular 715 series (Chapters A-M) entitled "River-Quality Assessment of the Willamette River Basin, Oregon".

A recently completed Willamette River Toxics Study (WRTS) investigated the presence and effect of toxic pollutants in the Willamette River and selected tributaries (Oregon Department of Environmental Quality, 1994b). Data were collected on contaminant levels in sediment and fish from numerous sites and evaluated using bioassays and other aquatic-life toxicity testing methods.

An ongoing comprehensive study that complements the NAWQA Program is the WRBWQS. This 6year cooperative USGS and ODEQ study initiated in 1990 is evaluating ecological conditions, contaminants, and dissolved oxygen levels. The goal of the study is "... to develop a complete data base for the river basin coupled with water quality models that will enable Federal, State, and local agencies to cooperatively ensure the preservation and beneficial uses of the Willamette River Basin and its associated biota..." (Tetra Tech, Inc., 1993a). The study includes biological field investigations on benthic macroinvertebrates, fish, and algal communities. Data collected for the WRB-WQS will be used to assist in the development of biological criteria for monitoring water quality, and to develop predictive mathematical models for assessing water quality and ecological health of aquatic biota. A listing of reports generated by the WRBWQS is presented in Tetra Tech, Inc., (1995a).

Much of the information presented here for aquatic resources in forested ecosystems of the basin is a result of research conducted at the HJAEF (fig. 1), which has been designated as one of 17 Long Term Ecological Research sites in the United States by the National Science Foundation (Parsons and others, 1991). This approximately 16,000-acre site is located about 50 miles east of Eugene within the western Cascade Range of the Willamette Basin. Elevations range from 1,345 to 5,350 feet, and about 45 percent of the forest is old growth. The HJAEF was established in 1948, and early research efforts focused on efficiency of logging and road systems and on the success of forest regeneration (McKee and others, 1987). The research focus shifted in the 1960s to watershed studies and in the 1970s to ecosystem studies and community dynamics. Current research emphasis is shared between ecosystem and silvicultural studies. The HJAEF has become one of the most intensively studied forests in the world, as evidenced by more than 800 listings in a bibliography of research publications through 1987 (McKee and others, 1987; Blinn and others, 1988). Since 1977, the site has been jointly administered by the U.S. Forest Service (USFS) and OSU.

#### **Acknowledgments**

Numerous individuals assisted in the development and preparation of this document, and several people deserve recognition. Dennis Wentz, Chief of the USGS Willamette Basin NAWOA Program, provided guidance and oversight throughout the project, including technical input, review, and assistance in production. Carmen Thomas, USFWS, conducted the initial compilation of available information including literature searches, data base retrievals, and contacts with appropriate individuals and agencies. Gloria Bourne, ODFW Research Librarian, assisted in locating several reports. Ron Rhew, USFWS, assisted in use of the ORIS data base and reviewed some sections of the report. Dorie Brownell, Donita Parker, and Mark Uhrich, USGS, prepared the figures, Thelma Parks, USGS, formatted the report; and Ronnie Nelson, USFWS, assisted with the references. The report has benefitted greatly from the comments of several reviewers, including Jeremy Buck, Ron Garst, Carol Schuler, and Marv Yoshinaka, USFWS; Dave Ward, ODFW; Steve Lawrence, USGS; and Peter Bayley, OSU.

#### **ENVIRONMENTAL SETTING**

The Willamette Basin NAWQA Study Unit includes the Willamette and Sandy River Basins and comprises approximately 12,000 square miles of land between the crest of the Cascade and Coast Ranges in northwestern Oregon (Wentz and McKenzie, 1991; Bonn and others, 1995) (fig. 1). The basin contains between 9,000 and 10,000 miles of streams (Willamette Basin Task Force, 1969), and over 2,000 lakes, totaling more than 60,000 acres (Oregon Department of Environmental Quality, 1992).

The basin is roughly rectangular in shape, approximately 125 miles in length, and ranging from 50–100 miles in width. It includes the broad alluvial plain of the Willamette Valley floor (approximately 3,500 square miles), and is bounded by mountain slopes and foothills on three sides and by the Columbia River on the north. The Cascade Range accounts for more than 60 percent of the basin area (Rickert and others, 1977). Elevation ranges from slightly above sea level at the mouth of the Willamette River near Portland to approximately 11,500 feet in the Cascade Mountains (Shearman, 1976).

The drainage system of the Willamette Basin is dominated by the northward-flowing Willamette River and its 13 major tributaries (fig. 1), which combined account for 93 percent of the basin area. The headwaters of the Willamette River arise in two forks—the Coast Fork and the Middle Fork—which flow northward from the Calapooya and Cascade Mountains, respectively, to form the main stem Willamette River near Eugene. Major westward flowing tributaries from the Cascade Mountains include (from south to north) the McKenzie River, Calapooia River, Santiam River, Molalla River, and Clackamas River. These tributaries have relatively steep gradients and high base flows sustained by melting snows and ground-water discharge (U.S. Army Corps of Engineers, 1991a). Principal tributaries flowing eastward from the Coast Range include (from south to north) the Long Tom River, Marys River, Luckiamute River, Rickreall Creek, Yamhill River, and Tualatin River (fig. 1). These tributaries have steep gradients only in the upper reaches, a slow meandering character in the foothills and valley floor, and low base flows during the summer months (U.S. Army Corps of Engineers, 1991a).

The main stem Willamette River is the predominant hydrologic feature in the Willamette Basin. The river flows north from Eugene for approximately 187 river miles through the Willamette Valley before entering the Columbia River near Portland (Gleeson, 1972; Shearman, 1976; Hines and others, 1977). The Willamette River is the 13th largest river in the contiguous United States in terms of total discharge (Kammerer, 1990), the largest tributary to the Columbia River below the Snake River (Parkhurst and others, 1950; Galbreath, 1965), and the largest river in the country entirely within one state (Clady, 1971). Stream gradient is relatively gentle, averaging less than 2.5 feet per mile, including a single drop of about 45 feet at Willamette Falls near Oregon City (RM 26.5) (fig. 2).

Currently, flows in the Willamette River and its major tributaries are highly regulated by dams and reservoirs. There are 13 U.S. Army Corps of Engineers (USACE) reservoirs on the major tributaries of the Willamette River (table 1 and fig. 3). The only hydroelectric project on the main stem Willamette River is Portland General Electric's Sullivan Plant at Willamette Falls. On tributaries throughout the basin, there are numerous small projects that provide water for hydroelectric generation and irrigation.

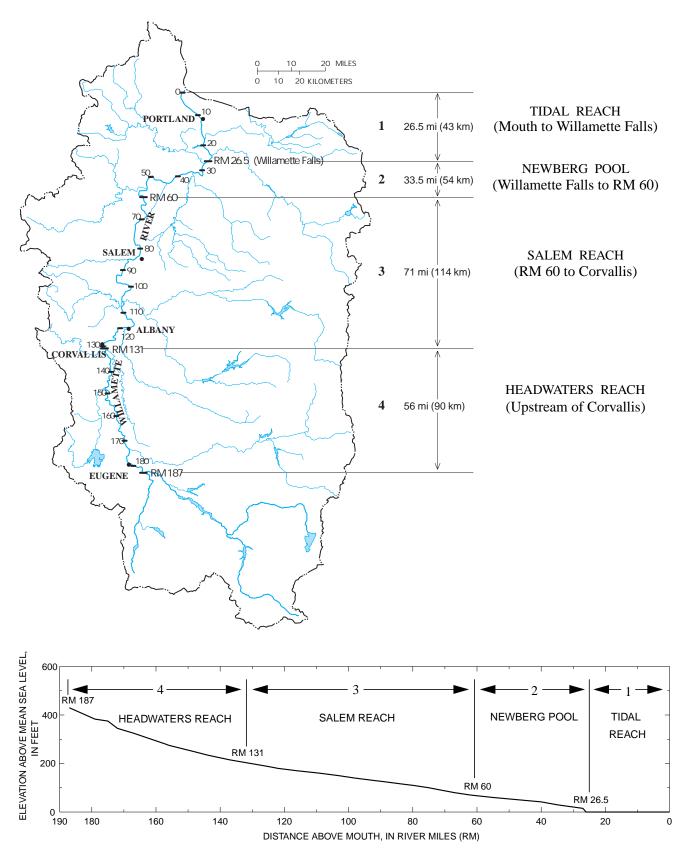


Figure 2. Profile and hydrologic reaches of the Willamette River, Oregon.

Table 1. U.S. Army Corps of Engineers reservoirs in the Willamette Basin, Oregon

[Sources: U.S. Army Corps of Engineers (1982, 1991b). Storage capacity is usable capacity for low-flow augmentation]

Name	Year completed	Storage capacity (acre-feet)	River	Subbasin
Fern Ridge Lake	1941	110,000	Long Tom River	Long Tom
Cottage Grove Lake	1942	30,060	Coast Fork Willamette River	Coast Fork
Dorena Lake	1949	70,500	Row River	Coast Fork
Dexter Reservoir	1954	4,800	Middle Fork Willamette River	Middle Fork
Lookout Point Lake	1953	349,400	Middle Fork Willamette River	Middle Fork
Fall Creek Lake	1962	115,000	Fall Creek	Middle Fork
Hills Creek Lake	1962	249,000	Middle Fork Willamette River	Middle Fork
Cougar Lake	1964	165,100	South Fork McKenzie River	McKenzie
Blue River Lake	1968	85,000	Blue River	McKenzie
Foster Lake	1966	33,600	Middle Santiam River	Santiam
Green Peter Lake	1966	333,000	South Santiam River	Santiam
Detroit Lake	1953	340,000	North Santiam River	Santiam
Big Cliff Reservoir	1953	2,430	North Santiam River	Santiam

#### **Physiographic Characterizations**

The Willamette Basin includes all of one physiographic province (Willamette Valley), and parts of three other provinces (Western Cascades, High Cascades, and Coast Range) (Franklin and Dyrness, 1973). It is commonly divided into three sections for reference; the Upper, Middle, and Lower Basins (Willamette Basin Task Force, 1969; Oregon Water Resources Department, 1992). The Upper Basin is bounded on the south by the Calapooya Mountains and on the north by the divide between the Calapooia/Santiam and McKenzie drainages east of the valley floor and the Long Tom and Marys River drainage divide west of the valley floor. The Middle Basin includes all lands that drain into the Willamette River between the Long Tom and Marys River drainage divide and Fish Eddy, a point three miles below the mouth of the Molalla River. The Lower Basin includes all lands that drain into the Willamette River from Fish Eddy to the mouth of the Willamette River.

The Willamette Basin has also been divided into ecoregions (Omernik and Gallant, 1986; Omernik, 1987) and subecoregions (Clarke and others, 1991). Ecoregions are defined on the basis of similarities of characteristics such as land use, potential vegetation, soils, land forms, precipitation, and biological communities. The Willamette Basin includes three ecoregions (Willamette Valley, Cascade Range, and Coast Range) and two subecoregions of the Willamette Valley (flat,

agricultural plains and the Coast Range and Cascade Range foothills) (Clarke and others, 1991) (fig. 4). Whittier and others (1988) identified similarities in streams within ecoregions in Oregon on the basis of data on physical habitat, water quality, and biological communities (fish, macroinvertebrates, and periphyton). Ecoregion divisions can be useful in water-quality assessment because they provide relatively distinct partitioning of areas with common climatic, hydrologic, geologic, and biologic features. Such partitioning is useful for evaluating the condition of aquatic biological communities, particularly if minimally impacted reference sites exist. These reference sites establish a baseline against which to compare sites where aquatic biological communities are potentially impacted. In Oregon, ecoregions have been used to describe geographic distribution of fish populations (Hughes and others, 1987).

Another physiographic delineation that is used extensively in this document is subbasins. These are based on hydrologic boundaries, and they correspond to the major tributaries of the Willamette River. Subbasins are useful for biological distinctions because streams within the defined geographic regions of watersheds or subbasins tend to be more similar to each other than those of streams within watersheds of a different geographic region. In the Willamette Basin, 15 major subbasins have been delineated on the basis of hydrologic boundaries (fig. 5). The area designated as "direct drainage to the Willamette River" includes

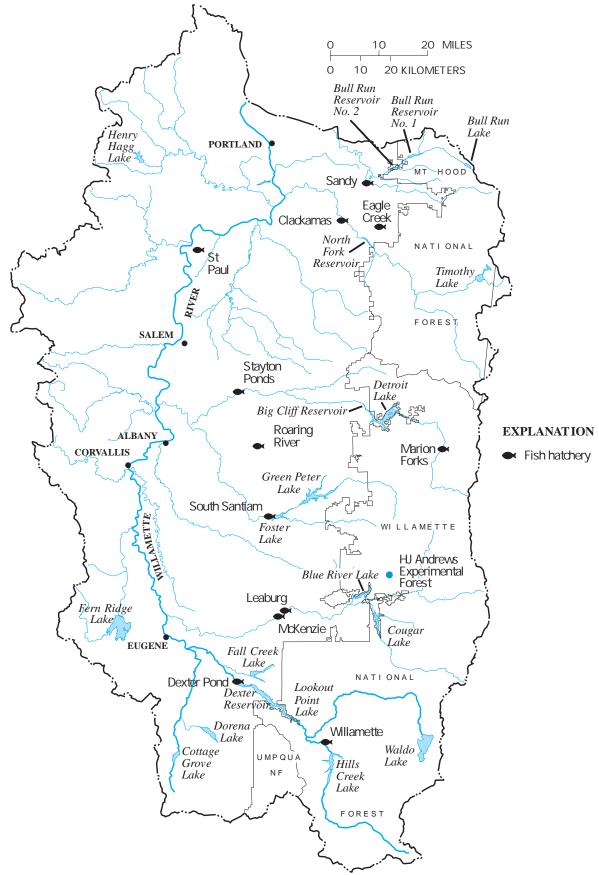


Figure 3. Fisheries and major lakes and reservoirs in the Willamette Basin, Oregon.

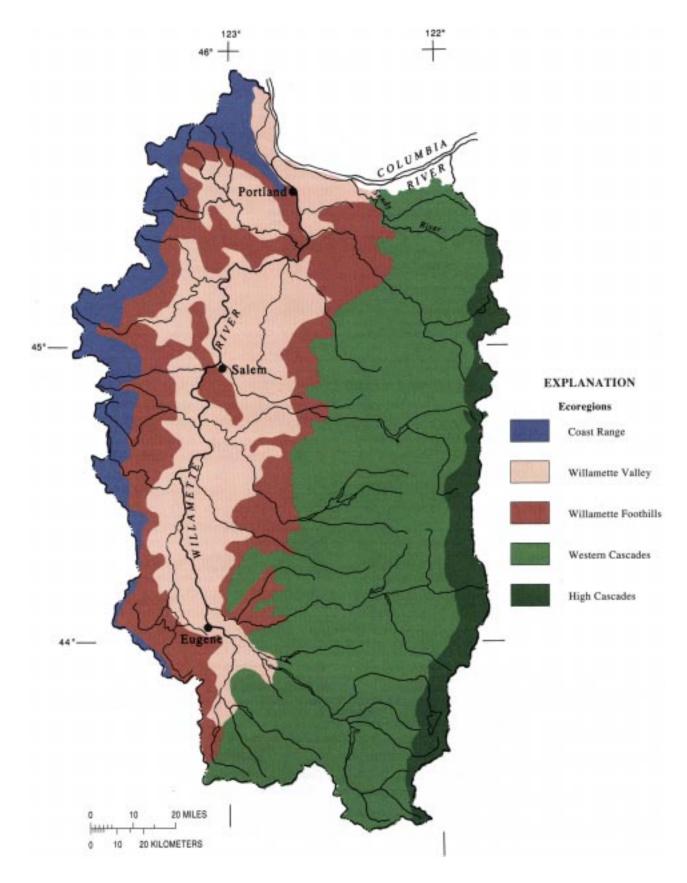
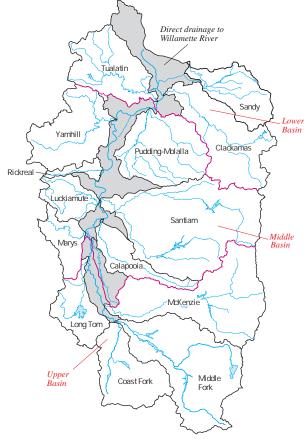


Figure 4. Ecoregions of the Willamette Basin, Oregon. (Modified from Clarke and others, 1991.)



**Figure 5**. Major subbasins of the Willamette Basin, Oregon.

the main stem Willamette River and numerous small and (or) unnamed tributaries, backwater sloughs, and abandoned channels. Physical descriptions of each subbasin are provided in OWRD (1991, 1992) and in each of the subbasin fish management plans (table 2).

The main stem Willamette River has also been characterized in terms of four reaches (Rickert and others, 1975; Gregory, 1993) based on channel characteristics. The reaches are the Headwaters Reach (just above Eugene [RM 187] to Corvallis [RM 131]), the Salem Reach (Corvallis [RM 131] to above Newberg [RM 60]), the Newberg Pool (above Newberg [RM 60] to Willamette Falls [RM 26.5]), and the Tidal Reach (Willamette Falls [RM 26.5] to the Columbia River) (fig. 2). Tetra Tech, Inc., (1992a) provides physical descriptions and biological characterizations of each reach.

#### Climate

The climate of the Willamette Basin is maritime temperate with cool, wet winters and warm, dry sum-

mers (Wentz and McKenzie, 1991); however, climatic conditions change with elevation. Precipitation ranges from approximately 35–40 inches of rainfall annually at lower elevations to approximately 175 inches (a high percentage as snowfall) in the mountains (Bonn and others, 1995). Rainfall decreases from north to south (Franklin and Dyrness, 1973), and approximately 90 percent of the average annual rainfall occurs between October and April. The seasonal dry period from May through September historically had an adverse impact on summer-early fall streamflow and water quality of the Willamette River.

#### **Land Use and Population**

The Willamette Basin is 70 percent forested (primarily in tributary subbasins), 22 percent agricultural (primarily on the valley floor), and 5 percent urbanized (Bonn and others, 1995). The basin includes 11 of the 12 largest cities in the State, including the five largest (Center for Population Research and Census, 1992), and approximately 2 million people or 70 percent of Oregon's population (Oregon Department of Environmental Quality, 1988; Bonn and others, 1995). Historically, the basin has supported most of Oregon's economic activity, including extensive timber, agricultural, industrial, and recreational economies (Shearman, 1976). Most of the agricultural activities occur in the midvalley counties of Linn, Benton, Polk, and Marion (fig. 1). The timber industry is an important part of the economy in Lane County in the southern part of the basin, and throughout the Cascade and Coast Range Mountains. Greater diversification in terms of trade, service, and manufacturing industries occurs in the northern part of the basin in Multnomah, Clackamas, and Washington Counties (U.S. Army Corps of Engineers, 1991a). Some sand and gravel mining activities occur adjacent to and within the main stem Willamette River and major tributaries, such as the Clackamas River.

The basin also has important fish and wildlife habitat (Shearman, 1976) and has historically been a favored hunting and sport fishing area (Willamette Basin Task Force, 1969). The main stem Willamette River near Portland provides recreational fishing to a major metropolitan population for resident fish such as black crappie (*Pomoxis nigromaculatus*), white crappie (*Pomoxis annularis*), smallmouth bass (*Micropterus dolomieui*), largemouth bass (*Micropterus salmoides*), and walleye (*Stizostedion vitreum*), and anadromous salmonids, including steelhead trout

**Table 2.** Fish management plans prepared by the Oregon Department of Fish and Wildlife (ODFW) for species and subbasins in the Willamette Basin, Oregon

[Coast Range Subbasin includes the Luckiamute, Marys, Rickreall, and Yamhill Subbasins]

Plan	Date	Reference
Basin/Subbasin Plans		
Clackamas Subbasin Fish Management Plan	January 1992	Murtagh and others (1992a)
Coast Fork Willamette Subbasin Fish Management Plan	December 1991	Connolly and others (1991)
Coast Range Subbasin Fish Management Plan	March 1992	Wevers and others (1992a)
Long Tom Subbasin Fish Management Plan	March 1992	Connolly and others (1992a)
Main stem Willamette Subbasin Fish Management Plan	March 1992	Rien and others (1992)
McKenzie Subbasin Fish Management Plan	April 1988	Howell and others (1988)
Middle Fork Willamette Subbasin Fish Management Plan	March 1992	Connolly and others (1992b)
Molalla and Pudding Subbasin Fish Management Plan	March 1992	Wevers and others (1992b)
North Fork of Middle Fork Willamette River Fish Management Plan	1979	ODFW (1979)
Santiam and Calapooia Subbasin Fish Management Plan	March 1992	Wevers and others (1992c)
Tualatin River Subbasin Fish Management Plan	January 1992	Murtagh and others (1992b)
Willamette Basin Fish Management Plan	June 1980	ODFW (1980)
Willamette Basin Fish Management Plan: Status and Progress 1979–85	October 1986	Howell (1986)
Willamette Basin Fish Management Plan (updated)	March 1988	ODFW (1988)
Willamette Basin Fish Management Plan (updated)	October 1991	ODFW (1991)
Species Plans		
Coho Salmon Plan	June 1982	ODFW (1982a)
Coho Salmon Plan Status Report	February 1985	ODFW (1985)
Comprehensive Plan for Production and Management of Oregon's Anadromous Salmon and Trout	June 1982	ODFW (1982c)
Implementation Plan for Spring Chinook Salmon	June 1993	ODFW (1993)
Steelhead Plan	July 1986	ODFW (1986)
Steelhead Plan (updated)	1995	ODFW (1995c)
Trout Plan	November 1987	ODFW (1987a)
Warmwater Game Fish Plan	August 1987	ODFW (1987b)

(Oncorhynchus mykiss gairdneri), coho salmon (Oncorhynchus kisutch), American shad (Alosa sapidissima), and white sturgeon (Acipenser transmontanus) (Farr and Ward, 1993). The main stem of the Willamette River, particularly below Willamette Falls, also provides the largest recreational spring chinook salmon (Oncorhynchus tshawytscha) fishery in the State (Oregon Department of Fish and Wildlife, 1990).

#### **AQUATIC AND RIPARIAN HABITAT**

The type, distribution, and quality of aquatic and riparian habitat in the Willamette Basin is highly vari-

able as a result of the diversity of environmental factors (topography, geomorphology, soils, climate, vegetation) and human-related factors (habitat perturbations, land use activities) that exist within the surrounding landscape. Since aquatic biological communities are affected not only by water quality but also by the physical features of aquatic and riparian habitat, condition of the physical habitat can be used as an indicator of the composition and condition of the biological community. A summary of the parameters used to evaluate aquatic and riparian habitat in the Pacific Northwest is presented in Tetra Tech, Inc., (1995b). The parameters include several measured variables of substrate, in-

stream cover, channel morphology, and riparian conditions.

Aquatic habitats may be broadly classified as running-water or slackwater systems (Holland, 1994). Running-water habitat in the main stem Willamette River differs substantially between the upper and lower reaches (Oregon Department of Fish and Wildlife, 1990). Gravel and cobble are the common substrate in the upper reaches, whereas the lower reaches are characterized by sand and finer sediment from the accumulated effects of sedimentation (Hughes and Gammon, 1987). Aquatic stream margin and floodplain slack-water areas, such as sloughs and backwater pools, are important for rearing juvenile fishes, invertebrate production, terrestrial organic input (leaf fall), and as a refuge during disturbances such as large floods (Moore, 1987; Naiman and others, 1988; Gregory and others, 1989; Sedell and others, 1990). Root masses of trees and emergent vegetation within slackwater aquatic habitat provides unique microhabitats for aquatic fauna (Holland, 1994).

Riparian habitat is the interface between aquatic and terrestrial ecosystems and is characterized by vegetation that is adapted to a high water table and periodic flooding (Gregory and others, 1991). Examples of important functions of riparian vegetation for stream ecosystems include shading, bank stabilization, uptake of nutrients, input of leaves and needles, retention of particulate organic matter during high flows, and contribution of large woody debris (Gregory and others, 1989). Riparian forests serve as buffers to adjacent habitats during floods (Holland, 1994) and also function to lessen the inflow of contaminant runoff into the aquatic community.

The riparian canopy in streams of the Cascade Mountains plays a dominant role in the abundance of most aquatic biota (Gregory, 1980; Murphy and Hall, 1981; Hawkins and others, 1983), often masking effects of substrate character (Murphy and others, 1981). The relationship between stream shading and biota in mountain streams of the basin has been investigated for fish (Aho, 1976; Murphy and others, 1981; Hawkins and others, 1983; Wilzbach, 1984), invertebrates (Grafius, 1977; Murphy and others, 1981; Hawkins and others, 1982), and salamanders (Hawkins and others, 1983). The results of these studies are discussed in a later section of this report (see "Forest Management").

Considerable work has been done in the basin on the ecological role of woody debris in streams

(Swanson and others, 1976), particularly in providing habitat for fish (Hall and Baker, 1982; Triska and others, 1982), and food resources for aquatic invertebrates (Swanson and others, 1982; Triska and others, 1982). The removal of woody debris from rivers in the basin for navigational and other purposes, and the fragmentation or elimination of riparian forests that provide woody debris sources, has rendered these aquatic habitats less complex and less suitable for some organisms, particularly salmonid fishes (Hicks and others, 1991).

As the human population has increased in the Willamette Basin, much of the aquatic and riparian habitat has been fragmented or eliminated. For example, the floodplain of the main stem Willamette River was once covered by dense woodland extending approximately 1–2 miles on either side of the river, but most of this forest has been cleared for farmland or timber (Sedell and Froggatt, 1984).

The earliest systematic and comprehensive attempt to evaluate aquatic and riparian habitat for anadromous fishes in the Willamette Basin was made between 1934 and 1942 as part of a program undertaken by the USFWS Bureau of Fisheries (now the National Marine Fisheries Service [NMFS]) to survey all tributaries in the Columbia River Basin (Rich, 1948). Data were collected on salmon and steelhead trout populations along with information on sources of pollution, impassable waterfalls, log and debris jams, and irrigation diversions. Another early assessment of stream conditions relative to the breeding, rearing, and migration of anadromous fishes was for the Sandy River and its tributaries (Craig and Suomela, 1940). Parkhurst and others (1950) reported the results of a several year survey of all the major tributaries of the Willamette River system. They described stream types and surrounding landforms, flows, barriers to fish, spawning habitat, and sources of pollution. Willis and others (1960) conducted an extensive evaluation of stream habitat characteristics, pollution problems, and fish passage problems for 17 river systems in the Willamette Basin. Another comprehensive assessment of stream habitat, spawning areas, and barriers to fish movement in the Willamette Basin was a cooperative effort by the Oregon State Game Commission, Fish Commission of Oregon, and USFWS Bureau of Commercial Fisheries (Thompson, 1965; Hutchison and others, 1966b).

The habitat surveys conducted by the USFWS Bureau of Fisheries between 1934 and 1942 (Rich, 1948) have been summarized in McIntosh and others

(1995). These surveys represent the earliest and most comprehensive documentation available on the condition and extent of anadromous fish habitat prior to hydropower development. Comparisons of historical riparian habitat conditions (as defined by these surveys) with present conditions have been the focus of recent research along the McKenzie River (Minear, 1994), and for 30 streams throughout the Willamette Basin (Bruce McIntosh, Oregon State University, oral commun., 1995). Another example of an attempt to compare historic and existing riparian habitat was a study along a section of the lower McKenzie River (EA Engineering, Science and Technology, Inc., 1991a).

State and Federal resource management and regulatory agencies, such as the USFS, ODFW, ODEQ, USEPA, and Bureau of Land Management (BLM), often conduct habitat surveys of streams to assess conditions relative to proposed or implemented management activities or specific project needs. These data are contained within project or program files or sometimes summarized in reports, such as Heller and Baker (1974) and Armantrout and Shula (1975). Some examples of project-related assessments of instream and riparian habitat of portions of the McKenzie River include Hawk and Zobel (1974), Hardin-Davis, Inc., (1988), and EA Engineering, Science and Technology, Inc., (1991a). Additionally, the majority of investigations of aquatic biota in the basin include some degree of instream and/or riparian habitat assessment.

Historically, there has been a lack of standardized methods and protocols on the type and level of aquatic and riparian habitat assessment. Several protocols have recently been developed to correct this deficiency. The USEPA has developed guidelines to evaluate the impacts of forest management on streams in the Pacific Northwest (MacDonald and others, 1991). The USFS Region 6 Level II protocol uses visual estimation methods established by Hankin and Reeves (1988) to estimate fish abundance and habitat area in small streams. The ODFW has developed standardized methodology to quantify the habitat condition of streams for its Aquatic Inventory Project (Oregon Department of Fish and Wildlife, 1995a). This methodology was developed in conjunction with other governmental and nongovernmental entities to be compatible with existing stream habitat assessment methodologies. The USEPA Index of Biotic Integrity protocol uses fish communities, and the Rapid Bioassessment Procedure uses stream habitat characteristics and macroinvertebrate communities, to evaluate ecological integrity of aquatic habitats by comparing study sites to an unimpaired or minimally disturbed site. The ODEQ has developed a protocol for monitoring nonpoint-source pollution using macroinvertebrates and habitat (Mulvey and others, 1992).

Protocols for habitat evaluation of large rivers, such as the main stem Willamette River and its major tributaries, are less developed owing to several factors, including difficulties in sampling deep flowing waters, high diversity in communities, and high temporal variability in environmental conditions (Bain, 1992). Tetra Tech, Inc., (1995b) used a combination of habitat parameters specified in other Federal and State protocols, such as those in Plafkin and others (1989), Mulvey and others (1992), Hayslip (1993), and Simonson and others (1994), for its habitat evaluation of the Willamette River as part of the WRBWQS. Because Tetra Tech, Inc.'s habitat evaluation was intended to support biological assessments, they used habitat parameters that emphasized the most biologically significant habitat features. They reported that the combination of metrics and scoring criteria selected were generally effective for determining biological condition. The NAWOA Program has also developed habitat protocols that can be used in small and large river systems (Meador and others, 1993).

#### **AQUATIC BIOLOGICAL COMMUNITIES**

Assessment of aquatic biological communities within a large watershed such as the Willamette Basin requires the recognition that physical and chemical changes to aquatic habitats result in changes in biological assemblages. From the high gradient, shallow headwater streams of the Cascades and Coast Range, to the low gradient, deep-water reaches of the lower Willamette River, there are extreme differences in environmental conditions (water flow, temperature, and dissolved oxygen content) and human-associated impacts. The occurrence and health of biota within these riverine aquatic communities is dependent upon, and varies as a result of, a combination of many sitespecific and landscape factors including both upstream and downstream phenomena (Vannote and others, 1980). Similarly, the biota of larger water bodies (natural lakes and reservoirs) in the basin varies, from those characteristic of nutrient-poor, low productivity, "oligotrophic" montane lakes of the Cascades, to those of the warmer, more productive, "eutrophic" lakes of

lower elevations, which typically support a high biomass of algae but a low diversity of aquatic fauna.

Most aquatic biological investigations in the Willamette Basin have focused on fish and particularly on salmonids because of their importance in sport and commercial fisheries. Information on macroinvertebrates is less extensive, and information on algae is considerably less extensive, particularly with regard to the historic occurrence of these taxa and their responses to physical, chemical, and biological impacts. Additionally, few aquatic macroinvertebrate and algal investigations have been spatially extensive (except for the WRBWQS), whereas several fish community studies have been spatially extensive, including Dimick and Merryfield (1945), Hughes and Gammon (1987), and Tetra Tech, Inc., (1993b).

Whittier and others (1988) provide a broad-scale characterization of aquatic biological communities in the Willamette Valley, relative to the seven other ecoregions in Oregon, based on assemblages of fish, invertebrates, and algae. In general, Willamette Valley streams had the greatest fish species richness and diversity, the most introduced species, and the fewest salmonids. Periphyton assemblages also had the greatest taxa richness and diversity, but macroinvertebrate assemblages had low richness and diversity, were lacking several common insect families, and had the highest proportion of noninsects.

#### **AQUATIC BIOTA**

The aquatic biota emphasized in this document are algae, macroinvertebrates, and fish. The importance of these taxa in the NAWQA Program is their role as indicators of water quality, and their use, particularly of fish, in contaminant analyses.

Algae are chlorophyll-containing photosynthetic organisms that range in size from microscopic single cells to long filamentous strands. They occur in rivers, lakes, and reservoirs, usually suspended in the water column (phytoplankton) or attached to a submerged substrate (periphyton). Algae, particularly diatoms, play an important role in aquatic ecosystems as the basis of production for aquatic food webs. Diatoms are considered to be of high food value for various aquatic fauna (Johnson and others, 1985).

Algal communities are useful in water-quality assessments because they are sensitive to changes in nutrient and dissolved oxygen concentrations, pH, and water temperature. Algae have short life cycles,

which makes them particularly responsive to environmental changes (Lowe and Pan, 1996) and have restricted mobility, which allows inferences to be drawn based on nearby sources of pollution (Tetra Tech, Inc., 1993a). The community composition and abundance of certain algae also provide a measure of trophic state or productivity in aquatic systems. Johnson and others (1985) list and describe algae used as indicators of aquatic conditions in Oregon lakes: *Anabaena* spp., *Aphanizomenon flos-aquae*, and *Stephanodiscus astraea* were found in eutrophic lakes, and *Chromulina* spp., *Cyclotella stelligera*, and *Sphaerocystis schroeteri* occurred in oligotrophic lakes.

Macroinvertebrates serve various functions in aquatic ecosystems, particularly as secondary consumers in many food chains (Healy, 1984) and as recyclers of organic matter (Merritt and others, 1984). They also are important organisms in the diet of fish, particularly trout and salmon. The macroinvertebrate community of streams, rivers, and lakes usually includes some or all of the following: insects, flatworms, crustaceans, and mollusks.

Macroinvertebrates are often used in assessments of the health of the aquatic community because they are relatively sessile, generally easily collected and identified, relatively abundant, and sensitive to physical and chemical changes in the water. Further, their responses to changing water conditions can be measured, and they often serve as the primary food source for many recreationally and commercially important fish (Plafkin and others, 1989; Mulvey and others, 1992). Macroinvertebrate communities also tend to have greater diversity than fish communities, and the natural integrity of the community is less compromised than that of fish communities, which are affected by fish stocking, sport fishing, and introduced species (Mulvey and others, 1992).

Several generalizations are recognized regarding the relationship between aquatic macroinvertebrates and water quality. High taxonomic diversity of aquatic macroinvertebrates is usually, but not always, indicative of healthy aquatic conditions. A metric commonly used to assess stream health is the ratio of EPT taxa (Orders Ephemeroptera, mayflies; Plecoptera, stoneflies; and Trichoptera, caddisflies) to chironomid midge larvae (Family Chironomidae). EPT taxa are generally classified as intolerant because of their sensitivity to degraded water conditions, and chironomids are generally considered tolerant for the opposite rea-

son. Organisms of the EPT orders generally require relatively large dissolved oxygen concentrations, minimal turbidity, and low water temperatures. Streams with high overall and high EPT taxa richness, a high EPT:chironomid ratio, and a lack of dominance by one or two taxa are considered to have good water quality (Smith and Collopy, in press).

The occurrence, abundance, and condition of fish species are frequently used to assess water quality, the health of the aquatic community, and the effects of land-use practices. Fish are used for these assessments because they are relatively easy to collect and identify, are widely distributed, and include representatives of many trophic levels. Additionally, species life histories are generally known, data are generally available from previous studies for temporal comparisons, and descriptive analyses of fish communities are relatively easy to understand (Karr and others, 1986). Because fish are consumed by humans, knowing the types and amount of contaminants accumulated is also important for assessing human health risks (Tetra Tech, Inc., 1995c).

Fish community composition depends on many factors, including habitat characteristics, water quality, and the availability of food sources. Kruse (1988) described the relationship between fish species distributions and assemblages in several Willamette Basin streams on the basis of gradients of habitat type (pools to riffles), cover (instream and riparian canopy cover), and stream discharge. One species assemblage (northern squawfish [Ptychocheilus oregonensis], largescale sucker [Catostomus macrocheilus], redside shiner [Richardsonius balteatus], and speckled dace [Rhinichthys osculus]) was most often found in pools, and another assemblage (longnose dace [Rhinichthys cataractae], and juvenile and adult torrent sculpin [Cottus rhotheus]) was found in riffles. Cutthroat trout (Oncorhynchus clarki) had a significant preference for instream and canopy cover, and longnose dace preferred areas with a lack of instream and canopy cover. The fish occurring in riffle habitats included longnose dace and various sculpin species, while northern squawfish and cutthroat trout were found in pools and runs.

Species interactions strongly affect fish community structure. Localized interactions among fishes using similar habitats results in shifts in microhabitat use (Li and others, 1987). For example, competitive dominance among salmonid species determines localized distribution patterns and microhabitat use. Coho

salmon are competitively dominant over steelhead trout, cutthroat trout, and chinook salmon respectively (Li and others, 1987). Similarly, the presence of torrent sculpin altered habitat use by the reticulate sculpin (*Cottus perplexus*) and Paiute sculpin (*Cottus beldingi*) in Marys River (Finger, 1982)

Fish community structure may also be affected by interspecific predation. Predation by the northern squawfish, the dominant piscivore in the basin, on juvenile anadromous salmonids was studied in free-flowing sections of the Willamette, Santiam, and McKenzie Rivers (Buchanan and others, 1981). Most of the fish preyed upon were sculpin, not salmonids, and predation was not as great as has been reported in lakes or immediately below dam tailraces or at hatchery release sites. Ward and others (1994) reported that only 12.3 percent of the 505 northern squawfish examined from the Portland Harbor contained juvenile salmonids. Beamsderfer and Reiman (1991) reported that northern squawfish are the principal predator of juvenile salmonids in the Columbia River system.

Information on selected semiaquatic amphibians and reptiles, birds, and mammals is also presented in this report to provide a thorough assessment of aquatic biota in the Willamette Basin. Although these taxa are not studied as part of the NAWQA Program, and they often are only semiaquatic, some of these organisms may be useful as indirect biological indicators of water quality, such as bald eagles (Haliaeetus leucocephalus) and osprey (Pandion haliaetus) for contaminant levels (Melancon, 1995; Blus, 1996), and American dipper (Cinclus mexicanus) for invertebrate abundance and community composition (John Loegering, Oregon State University, oral commun., 1995). These taxa are also often conspicuous and important biota in community ecology, often as top predators in food webs, and many species have important recreational (wildlife viewing) or economic value (waterfowl hunting). The presentation of information on these taxa focuses on selected species that are ecologically (as a foraging base) tied to NAWQA taxa, species used in contaminant studies, and/or species of management or research interest.

Considerable data are available on aquatic biota in the Willamette Basin, although our knowledge of the status of aquatic biota is highly uneven relative to taxa and spatial scope. Extensive information exists on high-profile taxa, such as anadromous salmonids, but relatively little information is available for many other aquatic taxa. Additionally, some areas have been studied extensively (e.g., the main stem Willamette River, the HJAEF and adjacent areas in the McKenzie subba-

sin, the Bull Run watershed and other sites in the Mt. Hood National Forest, and Oak Creek and Berry Creek near Corvallis on the eastern slope of the Coast Range), whereas many other areas in the basin have received very little attention.

The following sections enumerate or describe studies relating to the distribution, abundance, and trends of aquatic biota in the basin. Additional information on the spatial and temporal coverage of these studies is presented in appendix A. The algae and macroinvertebrate species lists compiled from available data (appendices B and C, respectively) do not represent a thorough assessment of taxon distribution or community diversity in the basin, but do provide a reference source for taxon occurrence as reported in studies reviewed for this document. This information is presented to assist in future studies by providing a baseline of existing information on distribution of algae and aquatic macroinvertebrates in the Willamette Basin.

#### **Algae**

Information on the abundance and distribution of algae in the Willamette Basin is limited in scope and is spatially uneven. Most of the algal sampling in the basin has been conducted in the main stem Willamette River. Some of the initial information was from USGS sampling (Rickert and others, 1977; Rinella and others, 1981), graduate student research (Wille, 1976), and water-quality monitoring (U.S. Public Health Service, 1964). Recent algal sampling was conducted to examine the effects of effluent discharge from a pulp and paper mill on aquatic biota (HMS Environmental, Inc., and Miller, 1988), and to examine impacts on dissolved oxygen concentrations as part of the WRBWQS (Gregory, 1993; and Tetra Tech, Inc., 1993c).

Results from sampling in 1973 and 1974 indicate that diatoms, particularly the genera *Melosira*, *Stephanodiscus*, *Cymbella*, *Achnanthes*, *Nitzschia*, and *Fragilaria*, dominated the taxa of the lower Willamette River between RMs 7 and 50 (Wille, 1976; Rickert and others, 1977). This dominance of diatoms in the lower river was consistent with results from sampling in 1963 (U.S. Public Health Service, 1964) despite a significant reduction in levels of organic pollution during the intervening time (Gleeson, 1972; Rickert and others, 1975). In a subsequent study, Rinella and others (1981) also reported diatoms, in both periphyton and phytoplankton samples, as the dominant algal form in the main

stem Willamette River. Below RM 50 (Newberg Pool and Tidal Reach; fig. 2), diatoms were primarily phytoplankton, whereas above RM 50, they were primarily periphyton. Rinella and others (1981) identified 86 species of algae throughout the main stem Willamette River and 54 species in the lower reaches of the Santiam River, with no major differences in abundance or diversity of algae from the previous studies of the U.S. Public Health Service (1964) and Rickert and others (1977). They also noted that phytoplankton abundance and diversity generally increased downstream. In the slower moving current of the Tidal Reach of the Willamette River (downstream from RM 26.5), the diatom *Stephanodiscus hantzschii* was the predominant alga.

As part of the WRBWQS, extensive algal sampling in the Willamette River was recently conducted by Gregory (1993). He identified 35 genera of algae from 23 sampling sites in the main stem Willamette River, 5 sites in the Coast Fork Willamette River, and 1 site each in the Middle Fork Willamette, McKenzie, Calapooia, Santiam, Yamhill, Molalla, and Tualatin Rivers near their junctions with the Willamette River. Blue-green algae accounted for more than 80 percent of the genera in the samples, and diatoms accounted for most of the remainder. The dominant genera of blue-green algae were *Anabaena*, *Aphanocapsa*, and *Chroococcus*. The dominance of blue-green algae decreased in the lower main stem of the Willamette River, and the proportion of diatoms increased.

Several other investigations of algal distribution and abundance have been done in streams of the Willamette Valley floor and foothills. The most abundant species of algae in the Willamette River near Halsey were the diatoms Gomphoneis herculeana and Fragilaria capucina (HMS Environmental, Inc., and Miller, 1988). They accounted for over 90 percent of the species composition of samples. Jackson (1973) sampled blue-green algae in the Willamette River and Middle Fork Willamette River, and Dever (1962) reported on algal composition of a controlled-flow section of Berry Creek. Carter (1975) sampled the middle course of the Tualatin River and reported that benthic forms were dominant in the upper river and planktonic forms were dominant in the lower river (downstream from Hillsboro). Algal sampling in the Tualatin River in 1976 indicated that benthic pennate diatoms were most common (Carter and others, 1976). The principal species were Melosira granulata, Stephanodiscus hantzschii, and Melosira distans. However, a shift in dominant taxa below RM 33 was observed, with the blue-green

alga *Aphanizomenon* spp. dominant in 1976 (Carter and others, 1976), and centric filamentous diatoms, such as *Melosira* spp. most abundant in 1987 (Li and Gregory, 1993). The USGS conducted algal sampling in four tributaries of the Molalla River during the drought year of 1977 (Miller, 1979): diatoms were the dominant algae, including the genera *Achnanthes*, *Gomphonema*, *Cymbella*, and *Cocconeis*.

Few studies have investigated algal composition of rivers of the Willamette Basin outside of the Willamette Valley floor and foothills. In streams of the HJAEF, Lyford and Gregory (1975) and Rounick and Gregory (1981) reported that open sites supported higher standing crops of periphyton than shaded sites. In six streams of the Bull Run watershed (Sandy River subbasin), the periphyton community was characterized by a high percentage of diatoms from June to October over a 6-year period, 1978–1983 (Clifton, 1985). The dominant periphyton were the diatom species Achnanthes minutissima and Achnanthes lanceolata. On the basis of a comparison with data from Hansmann and Phinney (1973) from Oregon coastal streams, Clifton (1985) suggested that the occurrence and/or abundance of several taxa, such as *Ulothrix* spp., *Chlamydomonas* spp., *Spirogyra* spp., Achnanthes spp., and Cocconeis placentula euglypta, may be useful in monitoring the impact of logging.

Algal studies in Willamette Basin reservoirs in the 1970s indicated that phytoplankton composition was similar to that of oligotrophic lakes in the Cascade Mountains (U.S. Army Corps of Engineers, 1991b). Diatoms such as Asterionella formosa, Fragilaria crotonensis, Synedra ulna, Stephanodiscus astraea, and Melosira granulata were dominant. The most common blue-green algae were Eudorina elegans, Straurastrum longiradiatum, and Sphaerocystis spp. The USACE also noted that phytoplankton blooms occurred regularly at some of the larger lakes, such as Lookout Point and Hills Creek.

In the Delta Ponds of Eugene adjacent to the Willamette River, the most common periphyton was the filamentous green alga *Rhizoclonium hieroglyphicum*, which is commonly associated with highly fertilized waters (Fetrow Engineering and Scientific Resources, 1989). Prescott (1923) and Lippert (1957) also provide information on species present in ponds in the floodplain of the Willamette River near Eugene.

Other investigations of algae in lakes and ponds include Burns (1993) for several high elevation mountain lakes in the Mt. Hood National Forest, Bullock and

others (1988) for Timothy Lake and Raymond (1983) for Bull Run Lake in the Mt. Hood National Forest, and Scheidt and Nichols (1976) for Hills Creek Lake in the Willamette National Forest. A series of county-based USGS reports from the mid-1970s includes information on the dominant algae in many of the lakes within the basin. The Willamette Basin counties covered by these reports were Columbia (Sanderson and others, 1973); Benton and Polk (Shulters, 1974); Multnomah, Washington, and Yamhill (Shulters, 1975); Clackamas (Shulters, 1976); and Marion (Rinella, 1977). Johnson and others (1985) provide similar information on algal composition of 41 lakes within the basin.

Chlorophyll *a* is an algal pigment used as an indicator of productivity through an estimation of algal biomass. As part of the WRBWQS, chlorophyll *a* was sampled throughout the Willamette River (Gregory, 1993). Concentrations tended to increase in a downstream direction, with higher concentrations in the Newberg Pool and Tidal Reach compared to the upstream sections of the river. Chlorophyll *a* concentrations have also been measured in mountain lakes (Sanderson and others, 1973; Shulters, 1974; 1975; 1976; Rinella, 1977; Johnson and others, 1985), an experimental stream section of Berry Creek (Reese, 1966), and streams of the western Cascades (Gregory, 1980; Murphy and Hall, 1981; Hawkins and Sedell, 1981; Rounick and Gregory, 1981).

#### **Macroinvertebrates**

The description of macroinvertebrate distributions and abundances in a systematic manner in the Willamette Basin is difficult because there is (1) unequal representation in sampling effort throughout the basin, (2) different sampling methodologies and protocols used in studies, and (3) varying taxonomic levels of identification. Thus, comparisons among studies are often precluded because of these inconsistencies.

The Willamette Basin supports a diverse aquatic macroinvertebrate fauna. In general, the Upper Basin within the Cascade Mountains is characterized by streams and rivers with a high diversity of taxa and a high richness of EPT taxa (Anderson, 1992; Whittier and others, 1988). The upper reach of the Willamette River (approximately equal to the Headwaters Reach) is also characterized by a high richness of EPT taxa (Johnson and others, 1989). Within the slow-current

reaches of the lower main stem Willamette River (Portland Harbor), the typical invertebrates are those that can tolerate low dissolved oxygen concentrations, such as oligochaetes (segmented worms), cladocerans (water fleas), amphipods (scuds), odonates (dragonflies and damselflies), and chironomid midges (Ward and others, 1988). The low gradient Tualatin River showed higher species diversity and greater richness of EPT taxa following installation of wastewater treatment plants (Li and Gregory, 1993).

Studies by researchers in the Entomology
Department at OSU often provide the most detailed
information on macroinvertebrate abundance and distribution in the Willamette Basin. Distribution and
abundance of macroinvertebrates in the Willamette
Basin also have been reported as part of studies on fish
communities. Unlike studies of algae, investigations of
macroinvertebrate communities have occurred in the
foothills and mountains of the basin as often as in the
main stem Willamette River and throughout the valley
floor.

The compilation of species lists from long-term research at particular sites provides excellent information on taxon occurrence. For example, Anderson and Hansen (1987) summarize the occurrence records of 325 taxa from over 25 years of research in Berry Creek, and Parsons and others (1991) provide an annotated list of invertebrate species (terrestrial and aquatic) that have been collected during 41 years of research at the HJAEF. Tetra Tech, Inc., (1993d) compiled a benthic macroinvertebrate species list for the Willamette River based on reports from four sampling efforts.

One of the most spatially extensive studies of aquatic macroinvertebrates in the Willamette Basin was conducted as part of the WRBWQS (Tetra Tech, Inc., 1994). Sampling occurred at 15 locations in the Willamette River between RMs 57 and 185, 2 locations in the McKenzie River near its confluence with the Willamette, and 1 location in the Tualatin River. The results indicated that water-quality degradation, rather than habitat degradation, appeared to account for biological impairment of downstream macroinvertebrate communities relative to upstream reference sites (Tetra Tech, Inc., 1994). Additionally, macroinvertebrate species composition was less diverse and less abundant in soft-bottom habitats than in riffle/run habitat. However, a comparison of macroinvertebrate community structure and composition within the two upper reaches of the Willamette River (Headwaters Reach and Salem Reach) did not reveal any significant differences based

on location by river mile within or between the two reaches.

Some macroinvertebrate studies in the Willamette River have been conducted to assess the relationship between pollution and macroinvertebrates. Deschamps (1952) conducted macroinvertebrate sampling throughout the entire main stem Willamette River and lower portions of several tributaries to assess the use of macroinvertebrates as biological indicators of pollution. The two most common macroinvertebrates in the Willamette River near Halsey (RM 142 to 150) during sampling above and below a pulp and paper effluent site in the summer of 1988 were a midge, Rheotanytarsus spp., and a caddisfly, Hydropysche spp. (HMS Environmental, Inc., and Miller, 1988). Both taxa are pollution tolerant organisms. However, at the same location, a total of 40 macroinvertebrate taxa were identified, and nearly half were EPT taxa, which indicates a high quality of water (Johnson and others, 1989).

Several agencies have reported on macroinvertebrate populations in streams in the lower and mid-elevations of the basin. The USGS conducted benthic macroinvertebrate sampling in four tributaries of the Molalla River during the drought year of 1977 (Miller, 1979). Caddisflies, mayflies, stoneflies, and midges were the dominant macroinvertebrates. Within the first three orders the dominant genera were *Cheumatop*syche (Trichoptera), Paraleptophlebia and Baetis (Ephemeroptera), and Nemoura (Plecoptera). The ODEQ conducted macroinvertebrate sampling in 1975 and 1976 as part of a biological assessment of the major tributaries of the Tualatin River system (Sutherland, 1976). The BLM has reported on macroinvertebrate sampling in nine streams of the McKenzie subbasin and one stream in the Middle Fork Willamette subbasin (Mangum, 1991a), and two streams in the Santiam subbasin (Mangum, 1991b). The USFS reported on macroinvertebrate sampling in Still Creek in the Mt. Hood National Forest (Mangum, 1990).

A comprehensive sampling program of macroinvertebrates in the lower McKenzie River was conducted to examine the influence of two hydroelectric projects that divert water from 13 miles of the river (EA Engineering, Science and Technology, Inc., 1990a). The results did not indicate any significant differences in taxon richness, EPT richness, or percent of the dominant taxon between diverted and undiverted reaches.

Several investigations of macroinvertebrates have been conducted in the high elevation, highgradient streams in and adjacent to the HJAEF. Hawkins and Sedell (1981) studied longitudinal and seasonal changes in macroinvertebrate communities. Lamberti and others (1991) and Anderson (1992) described the effects of a natural disturbance (debris torrent), and Wustenberg (1954) and Murphy and others (1981) evaluated the effects of logging on macroinvertebrate communities. Hawkins and others (1982) reported that streams without shading had higher abundances of invertebrates than shaded streams. Wilzbach and others (1986) studied the relationship between prey (macroinvertebrates) availability and cutthroat trout populations in logged and unlogged sites. Hawkins and Furnish (1987) discuss correlations of stream macroinvertebrate taxa with abundance of the snail Juga silicula. Other investigations that provide information on the distribution and abundance of macroinvertebrate taxa in and adjacent to the HJAEF include Anderson and others (1978) and Murphy and Hall (1981).

Macroinvertebrate sampling in four streams within the Bull Run watershed near Portland was conducted from 1978 to 1983 (Clifton, 1985). The dominant taxa were Chironomidae (midges), Hydracarina (water mites), and *Baetis* spp. (a mayfly). In November 1994, the most abundant taxon (over one-third of the composition) in a riffle of the Bull Run River was the plecopteran *Yoraperla brevis*; whereas, in pool habitat, plecopterans of the genus *Sweltsa* comprised nearly one-third of the individuals collected (TW Environmental, Inc., 1994). The amphipod *Hyalella azteca* was the most common macroinvertebrate collected in lakeshore substrate of Bull Run Lake in 1992 (Wisseman, 1992a).

Aquatic macroinvertebrate communities in lake/reservoir ecosystems within the Willamette Basin are less studied than those of riverine ecosystems. Investigations reporting on the presence of aquatic macroinvertebrates in Cascade lakes include Timothy Lake (Bullock and others, 1988), Bull Run Lake (Wisseman, 1992a), and Squaw Lakes (Wisseman, 1992b) in the Mt. Hood National Forest.

Several investigations have focused on the life history and ecology of specific macroinvertebrate taxa or groupings of similar taxa in streams of the Willamette Basin. Probably the most studied group is caddisflies (Trichoptera), which are well known because they are a principal food of trout and are imitated as flyfishing lures for trout. They have been a research focus of the Aquatic Entomology program at OSU, particularly at the HJAEF, where at least 99 species from 14 families have been recorded (Anderson and others, 1982), and in Benton County where 120 species have been recorded primarily from work in Berry and Oak Creeks (Anderson, 1976). The exhaustive work of Anderson (1976) summarizes information on the systematics, ecology, and distribution of this group of aquatic insects in Oregon.

Other taxon-specific studies have occurred at Oak Creek, where insect drift or seasonal occurrence have been studied for Trichoptera (Anderson and Wold, 1972; Anderson and Bourne, 1974), Epheme-roptera (Lehmkuhl, 1968, 1969; Lehmkuhl and Anderson, 1971), and Plecoptera (Ball, 1946; Kerst, 1969; Kerst and Anderson, 1974, 1975). Also in Oak Creek, Lehmkuhl (1968) reported on the life history of four species of Epeorus (Ephemeroptera), and Lehmkuhl and Anderson (1970) studied the biology of Cinygmula reticulata (Ephemeroptera). In Berry Creek, Azam (1969) studied the life history and production of Sialis californica (Megaloptera), and Grafius and Anderson (1979) studied the utilization of deciduous leaves as food by Lepidostoma quercina (Trichoptera). Studies of taxa associated with woody debris include craneflies of the genus Lipsothrix (Diptera) (Dudley and Anderson, 1987) in the Greasy Creek watershed of the Coast Range and the Quartzville Creek watershed of the Cascade Mountains, and the mayfly species Cinygma integrum in Berry Creek (Periera, 1980). Speir (1976) studied four blackfly (Diptera) species in Berry, Oak, and Soap Creeks near OSU. Steedman (1983) and Steedman and Anderson (1985) reported on the ecology of the aquatic beetle *Lara avara* (Coleoptera) in Berry and Yew Creeks. Taxon-specific studies on snails include the population dynamics of Juga plicifera in Oak and Berry Creeks (Diamond, 1982); growth, production, and distribution of Juga silicula in Oak Creek (Furnish, 1989); and production of Oxytrema silicula in Berry Creek (Earnest, 1967).

Some investigations have focused on macroinvertebrate composition of specific habitats. Aquatic macroinvertebrates associated with woody debris in forest streams of the basin was the focus of research by Anderson and others (1978). They reported that the three species most closely associated with woody debris were the aquatic beetle, *Lara avara*; a caddisfly, *Heteroplectron californicum*; and a snail, *Oxytrema silicula*. Based on additional work, Dudley and Anderson (1982) list 37 taxa of invertebrates closely associated

with woody debris in the Willamette Basin and 67 taxa as facultatively associated. Species composition of summer-dry headwater streams in the Oak Creek watershed included at least 27 species (Dieterich, 1992). Tew (1970) reported 58 species in a similar investigation of an intermittent stream in the Berry Creek watershed. Moore (1987) describes invertebrate assemblages associated with stream margins and backwaters of mountain streams. Hjort and others (1984) studied macroinvertebrate assemblages at revetments in the Willamette River and reported that the predominant taxa were organisms such as the polychaete worm *Manayunkia speciosa*, which attached to the substrate, or organisms such as the amphipod *Anisogammarus* spp., which were protected within interstitial spaces.

Freshwater clams, mussels, and snails are a conspicuous component of the aquatic macroinvertebrate fauna of the basin, and some were historically important as food items in the diet of Native Americans. Some clams also are harvested for bait and collected and sold by biological supply houses as classroom study specimens (Oregon Department of Fish and Wildlife, 1980). Thorough accounts of freshwater mollusc species were prepared for the Forest Ecosystem Management Assessment Team (FEMAT) report (Frest and Johannes, 1993). They reported 57 freshwater mollusc taxa within the range of the northern spotted owl (includes all forested parts of the Willamette Basin), many of which likely occur in the basin. Numerous other species not listed in the FEMAT report occur only in the Willamette Valley portion of the basin (Terrence Frest, Deixis Consultants, Seattle, Washington, written commun., 1995).

Crayfish are among the larger, more conspicuous aquatic macroinvertebrates in the basin. They are noteworthy because of their importance as fish forage, recreational use as bait, and commercial harvest for food in restaurants (Gladson, 1979; Oregon Department of Fish and Wildlife, 1980). Two species, *Pacifastacus leniusculus* and *Pacifastacus trowbridgii*, occur in the Willamette Basin (Gladson, 1979). The only study directed at crayfish within the basin occurred in Berry Creek (Mason, 1963).

Systematic long-term data collection at specific sites is lacking (except for the HJAEF, and Oak and Berry Creeks) to assess trends in macroinvertebrate community health in the basin. Biomonitoring programs for aquatic invertebrate communities have been recently implemented in several Cascade Mountain streams of the Mt. Hood National Forest (Wisseman,

1992b; 1995). The Xerces Society, in cooperation with several Federal and State agencies, has recently initiated an aquatic macroinvertebrate monitoring program designed to (1) assimilate and disseminate existing monitoring data, (2) evaluate the effectiveness of macroinvertebrate monitoring as a tool to determine watershed condition, and (3) produce a document describing monitoring programs and their effectiveness at assessing biotic integrity within watersheds (Sue Mauger, Xerces Society, Portland, Oregon, written commun., 1995).

#### Fish

Fish resources, particularly salmon and trout, have played a major cultural role in the lifestyle and economy of the Willamette Basin probably since Native American settlement of the area. Sport and commercial fisheries of salmon and trout historically sustained many local communities. The fisheries resource continues to be integral to Willamette Basin industry, recreation, and culture. On the basis of statewide estimates for 1980, sport fishing in the Willamette Basin generates approximately \$63 million in personal income annually (Howell, 1986).

The Willamette Basin supports a diverse and extensive fish community, which has changed since human occupation due to numerous factors, including habitat degradation, fish passage issues, aquaculture, and introductions of nonnative species. The ODFW (1988) listed 54 species of fish as being present within the Willamette Basin, and an additional 7 species have been reported from other sources (table 3). They include members of 16 families, including 9 anadromous species. Nearly half (48 percent) are introduced, nonnative species. T Hughes and others (1987) identified 15 fish species as characteristic of the western Cascades/Willamette River Basin ichthyogeographic region (table 3). Two of the species, Oregon chub (Oregonichthys crameri) and sand roller (Percopsis transmontana), are considered the most distinct fish species of this ichthyogeographic region, with little to no occurrence in other regions.

As a general rule, throughout the Willamette Basin and the Pacific Northwest, fish species richness tends to increase from the smaller, high elevation, steep gradient, cold water, headwater areas to the larger, low elevation, low gradient, warm water, main stem channels (Li and others, 1987; Beecher and others, 1988).

Table 3. Origin, trophic group, and relative tolerance to pollution for fish species occurring in the

Willamette Basin, Oregon [Sources: Friesen and Ward (1996); Hughes and Gammon (1987); Oregon Department of Fish and Wildlife (1988); Scott and Crossman (1973); Wydoski and Whitney (1979)]

Species	Scientific name	Origin	Trophic group <sup>1</sup>	Pollution tolerance
Bullhead catfishes	Ictaluridae		3 ***	
Black bullhead	Ameiurus melas	Introduced	Omnivore	Tolerant
Brown bullhead	Ameiurus nehulosus	Introduced	Omnivore	Tolerant
Yellow bullhead	Ameiurus natalis	Introduced	Omnivore	Tolerant
Channel catfish	Ictalurus punctatus	Introduced	Piscivore	Tolerant
White catfish	Ameiurus catus	Introduced	Omnivore	Tolerant
<u>Flounders</u>	<u>Pleuronectidae</u>			
Starry flounder <sup>2</sup>	Platichthys stellatus	Native	Piscivore	Tolerant
<u>Herrings</u>	<u>Clupeidae</u>			
American shad <sup>3</sup>	Alosa sapidissima	Introduced	Omnivore	Intermediate
<u>Lampreys</u>	<u>Petromyzontidae</u>			
River lamprey <sup>3</sup>	Lampetra ayresi	Native	Parasitic	Tolerant
Western brook lamprey <sup>4</sup>	Lampetra richardsoni	Native	( <sup>5</sup> )	Intermediate
Pacific lamprey <sup>3</sup>	Lampetra tridentata	Native	Parasitic	Intermediate
Livebearers	Poeciliidae			
Mosquitofish	Gambusia affinis	Introduced	Insectivore	Tolerant
Minnows	<u>Cyprinidae</u>			
Chiselmouth <sup>4</sup>	Acrocheilus alutaceus	Native	Herbivore	Intermediate
Goldfish	Carassius auratus	Introduced	Omnivore	Tolerant
Common carp	Cyprinus carpio	Introduced	Omnivore	Tolerant
Peamouth	Mylocheilus caurinus	Native	Insectivore	Intermediate
Oregon chub <sup>4,6</sup>	Oregonichthys crameri	Native	Insectivore	Intermediate
Fathead minnow	Pimephales promelas	Introduced	Omnivore	Tolerant
Northern squawfish <sup>4</sup>	Ptychocheilus oregonensis	Native	Piscivore	Tolerant
Longnose dace <sup>4</sup>	Rhinichthys cataractae	Native	Insectivore	Intermediate
Leopard dace	Rhinichthys falcatus	Native	Insectivore	Intermediate
Speckled dace	Rhinichthys osculus	Native	Insectivore	Intermediate
Redside shiner	Richardsonius balteatus	Native	Insectivore	Intermediate
Tench	Tinca tinca	Introduced	Insectivore	Tolerant
<u>Perches</u>	<u>Percidae</u>			
Yellow perch	Perca flavescens	Introduced	Insectivore	Intermediate
Walleye	Stizostedion vitreum	Introduced	Piscivore	Intermediate
<u>Sculpins</u>	<u>Cottidae</u>			
Prickly sculpin	Cottus asper	Native	Insectivore	Intermediate
Mottled sculpin <sup>4</sup>	Cottus bairdi	Native	Insectivore	Intolerant
Paiute sculpin <sup>4</sup>	Cottus beldingi	Native	Insectivore	Intolerant
Shorthead sculpin <sup>4</sup>	Cottus confuscus	Native	Insectivore	Intolerant
Riffle sculpin	Cottus gulosus	Native	Insectivore	Intolerant
Reticulate sculpin <sup>4</sup>	Cottus perplexus	Native	Insectivore	Tolerant
Torrent sculpin <sup>4</sup>	Cottus rhotheus	Native	Insectivore	Intolerant
Smelts	Osmeridae			
Eulachon	Thaleichthys pacificus	Native	( <sup>7</sup> )	

**Table 3**. Origin, trophic group, and relative tolerance to pollution for fish species occurring in the Willamette Basin, Oregon—Continued

Species	Scientific name	Origin	Trophic group <sup>1</sup>	Pollution tolerance
Sticklebacks	Gasterosteidae	Origin	group	tolerance
Threespine stickleback <sup>4</sup>	Gasterosteus aculeatus	Native	Insectivore	Intermediate
Sturgeons Steries Steries Startes	<u>Acipenseridae</u>			
White sturgeon	Acipenser transmontanus	Native	Omnivore	Intolerant
Suckers	Catostomidae	1 (441)	0111111 Y 011	11101011111
Largescale sucker	Catostomus macrocheilus	Native	Omnivore	Tolerant
Mountain sucker <sup>4</sup>	Catostomus platyrhynchus	Native	Herbivore	Intermediate
Oriental weatherfish	Misgurnus anguillicaudatus	Introduced	Omnivore	Tolerant
Sunfishes	Centrarchidae			
Green sunfish	Lepomis cyanellus	Introduced	Insectivore	Tolerant
Pumpkinseed	Lepomis gibbosus	Introduced	Insectivore	Tolerant
Warmouth	Lepomis gulosus	Introduced	Insectivore	Tolerant
Bluegill	Lepomis macrochirus	Introduced	Insectivore	Tolerant
Redear sunfish	Lepomis microlophus	Introduced	Insectivore	Tolerant
Smallmouth bass	Micropterus dolomieui	Introduced	Piscivore	Intermediate
Largemouth bass	Micropterus salmoides	Introduced	Piscivore	Tolerant
White crappie	Pomoxis annularis	Introduced	Insectivore	Tolerant
Black crappie	Pomoxis nigromaculatus	Introduced	Insectivore	Tolerant
<b>Topminnows</b>	<b>Fundulidae</b>			
Banded killifish	Fundulus diaphanus	Introduced	Insectivore	Tolerant
Trouts, Salmons, Whitefishes	<u>Salmonidae</u>			
Coho salmon <sup>3,4</sup>	Oncorhynchus kisutch	Native	Insectivore	Intolerant
Sockeye salmon <sup>3</sup>	Oncorhynchus nerka nerka	Introduced	Insectivore	Intolerant
Kokanee	Oncorhynchus nerka kennerlyi	Introduced	Insectivore	Intolerant
Chinook salmon <sup>3</sup>	Oncorhynchus tshawytscha	Native	Insectivore	Intolerant
Coastal cutthroat trout <sup>3</sup>	Oncorhynchus clarki clarki	Native	Insectivore	Intolerant
Steelhead (sea run rainbow) <sup>3</sup>	Oncorhynchus mykiss gairdneri	Native	Insectivore	Intolerant
Rainbow trout (resident)	Oncorhynchus mykiss	Native	Insectivore	Intolerant
Brown trout	Oncorhynchus trutta	Introduced	Insectivore	Intermediate
Mountain whitefish <sup>4</sup>	Prosopium williamsoni	Native	Insectivore	Intolerant
Brook trout	Salvelinus fontinalis	Introduced	Insectivore	Intolerant
Bull trout	Salvelinus confluentus	Native	Insectivore	Intolerant
Lake trout	Salvelinus namaycush	Introduced	Insectivore	Intolerant
Trout-perches	Percopsidae			
Sand roller <sup>4,6</sup>	Percopsis transmontana	Native	Insectivore	Intermediate

<sup>&</sup>lt;sup>1</sup>The principal foraging strategy of adults; does not include occasional opportunistic foraging.

 $<sup>^2\</sup>mathrm{Marine}$  species.

 $<sup>^3</sup>$ Anadromous.

<sup>&</sup>lt;sup>4</sup>Species characteristic of the West Cascades/Willamette River Basin Ichthyogeographic Region (Hughes and others, 1987).

<sup>&</sup>lt;sup>5</sup>Adults do not feed

<sup>&</sup>lt;sup>6</sup>One of two species most highly characteristic of the West Cascades/Willamette River Basin Ichthyogeographic Region (Hughes and others, 1987).

<sup>&</sup>lt;sup>7</sup>Does not feed in freshwater.

The fish of mid to high elevation tributaries and lakes in the basin tend to be dominated by a few cold water salmonid species, such as coho salmon and cutthroat trout, and a few species of suckers, minnows, and sculpins, and the mountain whitefish (*Prosopium williamsoni*). Species composition in low elevation reaches of the major rivers of the Willamette Valley and foothills includes numerous warm water fish such as bass (*Micropterus* spp.), catfish (*Ictalurus* spp.), and several species in the sunfish group. The fish fauna of the Willamette River is presently dominated by nonnative species, whereas in mountain streams, there is better representation of native species.

The transition from high elevation, cold water streams to low elevation, warm water streams and rivers also is characterized by ecological niche replacement among similar species. For example, mountain suckers (*Catastomus platyrhynchus*) are gradually replaced by largescale suckers as gradient decreases and water temperatures increase (Li and others, 1987). This type of change is also apparent in foraging guilds, which gradually change from mostly surface-insect feeders in the headwaters to large-invertebrate feeders in the low elevation tributaries and main stem Willamette River (Li and others, 1987).

The ORIS database (Oregon Department of Fish and Wildlife, 1994), which was used to develop table 4, includes information on fish species distribution in the Willamette Basin. This information is most valuable for a coarse assessment of stream conditions based on fish species composition and diversity, and in determining species of widespread distribution for use in comparative studies, particularly toxicological studies. The ORIS database also includes fish species distribution information at a much greater resolution (tributaries and subtributaries of the major rivers) than presented in table 4.

Willamette Basin fish species distribution and abundance have been described by numerous sources, including the Willamette Basin Task Force (1969) and Oregon Department of Fish and Wildlife (1990). Historical information on the distribution of fishes in the basin includes a letter by Abernethy (1886), and reports by Snyder (1908) and Rich and Holmes (1929). The first extensive sampling of fish distributions in the Willamette River below Willamette Falls was conducted in 1941 and 1942 by Craig and Townsend (1946) for the USACE. Dimick and Merryfield (1945) conducted the first extensive sampling throughout nearly the entire main stem Willamette River. They

reported 34 species of fish upstream from RM 15 on the basis of their sampling and the previous work of others. A report by the Oregon Game Commission in the late 1950s provides information on fish species occurring in the Willamette Basin at that time (Willis and others, 1960). A series of reports in the 1960s by the Oregon State Game Commission described fish resources for the entire basin (Thompson and others, 1966), Lower Willamette Basin (Hutchison and Aney, 1964), Middle Willamette Basin (Oregon State Game Commission, 1963), and Upper Willamette Basin (Hutchison and others, 1966a).

Two recent investigations throughout the entire main stem Willamette River (Hughes and Gammon, 1987; Tetra Tech, Inc., 1993b) provide information on changes in species assemblages based on comparisons with Dimick and Merryfield (1945). Hughes and Gammon (1987) reported more fish species, but fewer species tolerant of poor habitat than Dimick and Merryfield (1945). They attributed differences in fish assemblages between 1945 and 1986 primarily to changes in the physical habitat and improvements in water quality. They also characterized four distinct fish assemblages (Upper River, Middle River, Newberg Pool, and Portland Metro) corresponding to the major sections of the river.

Tetra Tech, Inc., (1993b) conducted fish sampling in 1992 with the same techniques and in most of the same locations as Hughes and Gammon (1987). They reported similar trends in fish assemblages throughout the river, and suggested that fish communities in the lower river may have become more robust (healthier) since 1983. They reported significant differences in fish communities between upstream (Eugene) and downstream (Portland) locations, although they could not statistically differentiate the two upstream communities (Upper River and Middle River) with regard to fish composition.

Friesen and Ward (1996) described fish assemblages in the lower Tualatin subbasin as part of a study to assess the impacts of urbanization on native fish populations. They suggested that native fish assemblages were moderately unhealthy on the basis of a high percentage of introduced species, a relatively low number of species intolerant to pollution and warm water, and a relatively large number of sites having a high proportion of fish with parasites or physical anomalies. Reticulate sculpin comprised nearly 70 percent of the individuals captured during sampling.

[ Sources: Dodge and Armantrout (1994); Hjort and others (1984); Hughes and Gammon (1987); Farr and Ward (1993); Friesen and Ward (1996); Li and Gregory (1993); Markle (1994a); Oregon Department of Fish and Wildlife (1994); W. Hunt (Oregon Department of Fish and Wildlife, written commun., 1995); S. Mamoyac (Oregon Department of Fish and Wildlife, written commun., 1995); T. Murtagh (Oregon Department of Fish and Wildlife, written commun., 1995); M. Wade (Oregon Department of Fish and Wildlife, written commun., 1995). Species listed include all documented occurrences regardless of size or distribution of the population within the river, or whether they are from wild or hatchery stocks; --, not known to occur; C.F., Coast Fork; M.F., Middle Fork; X, present]

	Major Rivers in the Willamette Basin															
Species	Cala- pooia	Clack- amas	C.F. Willa- mette	Long Tom	Luck-	Marys	Mc- Kenzie	M.F. Willa- mette	Molalla	Pud- ding	Rick- reall	Sandy	San- tiam	Tual- atin	Willa- mette	Yamhill
Bullhead Catfishes																
Black bullhead														X	X	
Brown bullhead	X	X	X	X	X	X	X	X		X	X		X	X	X	X
Yellow bullhead	X	X	X	X						X			X	X	X	
Channel catfish		X		X						X				X	X	X
White catfish														X		
<b>Flounders</b>																
Starry flounder		X												X	X	
<u>Herrings</u>																
American shad		X										X			X	
<b>Lampreys</b>																
Western brook lamprey		X		X		X	X	X	X	X				X	X	X
Pacific lamprey	X	X		X	X	X	X	X	X	X	X	$X^1$	X	X	X	X
River lamprey	X						X								X	
<u>Livebearers</u>																
Mosquitofish	X			X	X	X	X	X			X		X	X		X
Minnows																
Chiselmouth	X	X	X	X		X	X	X	X				X	X	X	
Common carp	X	X		X		X	X	X		X				X	X	X
Oregon chub						X		X					X			
Peamouth	X	X	X	X		X	X	X	X	X			X	X	X	
Fathead minnow														X		
Northern squawfish	X	X	X	X		X	X	X	X	X		X	X	X	X	
Goldfish		X								X				X	X	
Longnose dace		X	X	X				X				X	X	X	X	
Leopard dace	X		X	X				X					X		X	
Speckled dace	X	X	X			X	X	X	X	X	X	X	X	X	X	

Table 4. Distribution of fish species in major rivers of the Willamette Basin, Oregon—Continued

Major Rivers in the Willamette Basin																
	Cala-	Clack-	C.F. Willa-	Long	Luck-		Mc-	M.F. Willa-		Pud-	Rick-		San-	Tua-	Willa-	
Species	pooia	amas	mette	Tom	iamute	Marys	Kenzie	mette	Molalla	ding	reall	Sandy	tiam	latin	mette	Yamhill
Minnows—Continued																
Redside shiner	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
Tench															X	
<u>Perches</u>																
Yellow perch		$X^1$			X	X				X	X	$\mathbf{X}^{1}$		X	X	X
Walleye		$X^1$						X				$\mathbf{X}^{1}$		X	X	
<b>Sculpins</b>																
Prickly sculpin		X		X		X	X					X	X	X	X	
Mottled sculpin							X	X					X			
Paiute sculpin	X		X	X		X	X	X	X	X			X		X	
Shorthead sculpin							X	X					X			
Reticulate sculpin	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
Torrent sculpin	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	
Riffle sculpin	X			X											X	
Smelts																
Eulachon												X			X	
<b>Sticklebacks</b>																
Threespine stickleback	X	X				X	X	X					X	X	X	
<b>Sturgeons</b>																
White sturgeon		X					X	X				$\mathbf{X}^{1}$		X	X	
Suckers																
Largescale sucker	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
Mountain sucker		X	X	X		X	X	X	X	X			X	X	X	
Oriental weatherfish		X														
<b>Sunfishes</b>																
Green sunfish										X				X		
Pumpkinseed	X	X		X	X	X				X	X		X	X	X	X
Warmouth	X	X		X	X	X				X	X		X	X	X	X

Table 4. Distribution of fish species in major rivers of the Willamette Basin, Oregon—Continued

Species		Major Rivers in the Willamette Basin														
	Cala- pooia	Clack- amas	C.F. Willa- mette	Long Tom	Luck-	Marys	Mc- Kenzie	M.F. Willa- mette	Molalia	Pud- ding	Rick- reall	Sandy	San- tiam	Tua- latin	Willa- mette	Yamhill
Sunfishes—Continued																
Bluegill	X	X	X	X	X	X	X	X		X	X	X	X	X	X	X
Redear sunfish															X	
Smallmouth bass	X	X				X		X			X	X	X	X	X	X
Largemouth bass	X	X	X	X	X	X	X	X		X	X	X	X	X	X	X
White crappie	X	X		X	X	X	X	X		X	X	X	X	X	X	X
Black crappie	X	X		X	X	X		X		X	X	X	X	X	X	X
<b>Topminnows</b>																
Banded killifish		X												X	X	
<b>Trout and Salmon</b>																
Coho salmon		X			X	X	$X^1$		X	X	X	X	$X^1$	X	X	X
Sockeye salmon													$X^1$		X	
Kokanee		X											X			
Spring chinook salmon	X	X					X	X	X	X		X	X		X	
Fall chinook salmon		X	$X^1$				X	X	X	$X^1$	$\mathbf{X}^{1}$	X	X	$\mathbf{X}^{1}$	X	$X^1$
Mountain whitefish	X	X		X	X	X	X	X	X			X	X	X	X	
Cutthroat trout	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X
Summer steelhead trout		X					X	X	X			X	X		X	
Winter steelhead trout	X	X	$X^1$	X	X	X	$X^1$	X	X	X	X	X	X	X	X	X
Rainbow trout	X	X	X		X		X	X	X	X	X	X	X	X	X	X
Brown trout		X									X					
Brook trout		X					X	X				X	X			
Bull trout		$X^2$					X	X								
Lake trout <sup>3</sup>																
<b>Trout-perches</b>																
Sand roller	X	X	X	X	X	X	X	X	X	X	X		X	X	X	X

<sup>&</sup>lt;sup>1</sup> Rare; only a small population exists.

<sup>2</sup> Possibly extinct; no records since the early 1970s.

<sup>3</sup> Occurs in high elevation lakes only.

In the lower Willamette River, Ward and Nigro (1991) and Farr and Ward (1993) described fish assemblages in the Portland Harbor, from the confluence of the Willamette River with the Columbia River to RM 15. They found significant relationships between habitat and fish assemblages. Overall, northern squawfish were the dominant species, followed by black crappie, white crappie, largemouth bass, smallmouth bass, and walleye. Ward and others (1991) summarized information on the status and biology of white and black crappie in the lower Willamette River.

Anadromous salmonids are considered the most valuable fish in the Willamette Basin in terms of commercial and sport fisheries (Willamette Basin Task Force, 1969; U.S. Army Corps of Engineers, 1982). The historical value of the Willamette River to anadromous fish, particularly chinook salmon and steelhead trout, was primarily as a passageway to tributaries where spawning grounds were located (Parkhurst and others, 1950; Oregon State Game Commission, 1963; Willamette Basin Task Force, 1969). Spring chinook salmon and winter steelhead trout were able to negotiate Willamette Falls during high flows (Collins, 1968), but fall runs were likely absent or minimal above Willamette Falls due to low water conditions at that time of the year (Oregon Department of Fish and Wildlife, 1990). Below Willamette Falls, particularly in the Clackamas River, large runs of fall chinook salmon occurred prior to extermination caused by oxygen depletion in the water from pollution, particularly during the low flow period when these fish were migrating to spawning grounds (Holmes and Bell, 1960). Other historically common anadromous fish in the Willamette River were coho salmon and American shad (Hutchison and Aney, 1964).

The ODFW reports annually on the composition and abundance of anadromous fish passage at several locations. These have been summarized for Willamette Falls since the mid-1950s (Howell, 1986) and Leaburg Dam since 1970 (Downey and others, 1993). ODFW (1980) and Howell (1986) summarize, for each anadromous species, the sport catch, releases of hatchery stock in various rivers, and passage counts at Willamette Falls and other dams. Similar information is available in subbasin fish management plans (table 2). The ODFW also provides annual summaries of populations of various salmonid species in the Willamette River (Downey and others, 1993).

Most salmonid spawning in the basin occurs in tributaries of the Willamette River, except for the rees-

tablished runs of fall chinook salmon, which spawns from RM 50 (Newberg) through RM 187 (Springfield) of the main stem Willamette River (Oregon Department of Fish and Wildlife, 1990), and the mountain whitefish, which spawns throughout the river (Miller and others, 1991). It has also been reported that a few spring chinook salmon spawn in the Willamette River near Harrisburg (U.S. Army Corps of Engineers, 1982). Steelhead trout and chinook salmon also use the Willamette River for migration and juvenile rearing, and native cutthroat trout spawn in the upper tributaries and use it for juvenile rearing. Anadromous nonsalmonids such as white sturgeon and American shad spawn primarily in the main stem (Miller and others, 1991).

Despite the historic and current focus on salmonids and game fish, recent emphasis on investigations of entire aquatic ecosystems (Gregory and others, 1991; Reeves and Sedell, 1992) has resulted in more effort being focused on the effects of habitat loss and degradation on nonsalmonids and other nongame fish. Additionally, an increasing emphasis is being placed on aquatic biota and conservation efforts within the context of watersheds to account for the interrelated functions among aquatic, riparian, and upland habitats.

Because of the size and diversity of waters in the Willamette Basin, nearly all species of resident fish found in Oregon occur here (U.S. Army Corps of Engineers, 1982). In most cases, population size has not been adequately documented for resident fish species, although often these may be the most abundant fish in a particular area (U.S. Army Corps of Engineers, 1982). Angler effort and catch is most frequently used to provide an indication of the relative importance of resident fish. Most concern regarding nongame resident fish, such as suckers, northern squawfish, and common carp (*Cyprinus carpio*) has focused on their competition with desirable game species, and extensive efforts have been made to reduce or eliminate certain species where they compete with game fish.

Several investigations have reported on anadromous fish movements in major rivers and streams of the basin. Sams and Conover (1969) summarized data on the timing of migration of fall chinook and coho salmon in the lower Willamette River. Migratory characteristics of chinook salmon have also been reported in the lower Willamette River near Portland (Knutsen and Ward, 1991) and on the McKenzie River (EA Engineering, Science and Technology, Inc., 1991b). Timing of migration of Willamette River spring chinook salmon was summarized from tagging studies

and catch data by Galbreath (1965). Streamflows affected movement (upstream during periods of low flow and downstream during periods of high flow) and mortality rates (higher during summer and fall low flows) of cutthroat trout in a controlled flow section of Berry Creek (Nickelson, 1974). A variety of movement patterns were evident for coastal cutthroat trout during a tagging and recovery study (Moring and others, 1986). Radiotracking of winter steelhead on the Clackamas River has been conducted to monitor movement, distribution, and habitat use (Shibahara and Lumianski, 1995).

A few studies have investigated fish movements in and near impoundments. Massey (1965, 1967a, 1967b) captured juvenile salmon and steelhead near the industrial area at Willamette Falls and reported on the abundance, timing, and size of the downstream migrants. The size of juvenile coho salmon was related to their length of stay in North Fork Reservoir (Hreha, 1967). Larger individuals moved more quickly out of the reservoir after migration began in the spring than did smaller individuals. Zakel and Reed (1984) studied the timing of downstream migration of fish at Leaburg Dam on the McKenzie River. Hasselman and Garrison (1957) reported that northern squawfish moved from the main part of Lookout Point Lake to the upper end for spawning.

Numerous studies in the basin have been directed at specific taxa or groupings of similar taxa. These taxa include dace (*Rhinichthys* spp.) (Zirges, 1972; Dodge, 1994), sculpin (Bond, 1963; Finger, 1982), bull trout (Salvelinus confluentus) (Goetz, 1994), Oregon chub (Long, 1982; Markle and others, 1989; Pearsons, 1989; Markle and others, 1991; Scheerer and others, 1992, 1993, 1994, 1995), kokanee (Oncorhynchus nerka kennerlyi) (Wetherbee, 1965), cutthroat trout (Wustenberg, 1954; Wyatt, 1959; Warren and others, 1964; McIntyre, 1967; Nickelson, 1974; Aho, 1976; Wilzbach, 1984; Frissell and others, 1985; Wilzbach and others 1986: Moore, 1987; Moore and Gregory, 1989; House, 1995; Oregon Department of Fish and Wildlife, 1995b), chinook salmon (Mattson 1962), redside shiner (Rodnick, 1983), northern squawfish (Hasselman and Garrison, 1957; Buchanan and others, 1981), winter steelhead trout (Shibahara and Lumianski, 1995), and rainbow trout (Oncorhynchus mykiss) (Moore and Gregory, 1989).

Summaries of information on taxon distribution and abundance in the basin have also been compiled in status reports and literature reviews. These taxa

include cutthroat trout (Nicholas, 1978), chinook salmon (Mattson and Dimick, 1952; Mattson, 1963; Wevers, 1994), coho salmon in the Clackamas subbasin (Cramer and Cramer, 1994), steelhead trout in the Willamette River (Clady, 1971), and kokanee in Detroit Lake (Wetherbee, 1965).

A few investigations have been conducted on fisheries resources in lakes of the Willamette Basin. These studies include the effects of water withdrawal on fisheries resources in Bull Run Lake (Beak Consultants Inc., 1993), fish sampling in Detroit Lake (Wetherbee, 1962), and a study on the northern squawfish in Lookout Point Lake and Dexter Reservoir (Hasselman and Garrison, 1957).

Fish species assemblages associated with revetments (bank-stabilization structures) in the mid-Willamette River have been studied by Hjort and others (1984) and Li and others (1984). Hjort and others (1984) reported higher densities of fish at revetments than natural banks, but Li and others (1984) indicated that natural banks had higher densities of fish. The fish species that Hjort and others (1984) identified that benefit from the invertebrate and algae populations associated with revetments include prickly sculpin (*Cottus asper*), redside shiner, northern squawfish, largescale sucker, and chiselmouth (*Acrocheilus alutaceus*).

A few studies have reported on fish species distribution and abundance outside of the Willamette River. For example, in Cascade Mountain streams in and adjacent to the HJAEF, studies have been conducted by Hawkins and others (1983), Moore (1987), and Moore and Gregory (1989). Fish populations were the focus of an investigation by Everest and others (1985) in Fish Creek in the Mt. Hood National Forest. Fish species distribution and abundance in the lower elevations of the basin have been reported by Friesen and Ward (1996) for the lower Tualatin subbasin, and Baker and others (1995) for the Clackamas subbasin.

In the Willamette River near Halsey, three species of sculpin—prickly, torrent, and reticulate —were captured during sampling in the summers of 1988 and 1989 (HMS Environmental, Inc., and Miller, 1988; Johnson and others, 1989). Distribution of sculpin are of particular interest because this genus was used extensively by the Willamette NAWQA study unit for analysis of organochlorine compounds and trace elements in tissue during 1992–93 (Dennis Wentz, U.S. Geological Survey, oral commun., 1996).

In a field verification study of fish distribution and species composition within the Clackamas River

subbasin (Baker and others, 1995), sampling revealed a 45-percent overestimate (based on river miles in which a species occurred) of a "best guess" distribution based on available information. This finding exemplifies that, despite the extensive research that has been done on fish resources, existing data are still inadequate in some instances.

## **Fishery Plans**

Management of fish populations and habitat within the Willamette Basin is guided by the objectives and priorities initially set forth in the Willamette Basin Fish Management Plan (Oregon Department of Fish and Wildlife, 1980) and subsequent revisions (Oregon Department of Fish and Wildlife, 1988; 1991). The Willamette Basin Fish Management Plan: Status and Progress 1979–1985 (Howell, 1986) describes progress made on the objectives of the initial plan through 1985.

One of the high priorities of the initial plan was the preparation of a fish management plan for each subbasin. Ten subbasin plans have been completed (table 2). A fish management plan for the Sandy subbasin is being prepared (Tom Murtagh, Oregon Department of Fish and Wildlife, written commun., 1995). Separate plans have also been prepared for important reservoirs and lakes within the subbasins and for spring chinook salmon throughout the basin (Oregon Department of Fish and Wildlife, 1993). Additionally, the ODFW has completed statewide species management plans for coho salmon (Oregon Department of Fish and Wildlife, 1982a), steelhead trout (Oregon Department of Fish and Wildlife, 1986; 1995c), trout (Oregon Department of Fish and Wildlife, 1987a), and warm water game fish (Oregon Department of Fish and Wildlife, 1987b) (table 2). These plans were intended to guide the development of localized plans for river basins and subbasins.

In addition to fish management plans, production plans for anadromous fish have been prepared for the Willamette Basin and 11 subbasins: Clackamas, Coast Range, Coast Fork Willamette, Long Tom, McKenzie, Middle Fork Willamette, Molalla and Pudding, Sandy, Santiam and Calapooia, Tualatin, and main stem Willamette (Oregon Department of Fish and Wildlife, 1990). These plans provide the basis for salmon and steelhead production objectives and strategies in the Northwest Power Planning Council's Columbia River Fish and Wildlife Program. The plans include comprehensive information on fish resources in each of the

subbasins, including natural production, hatchery production, and harvest.

# **Semiaquatic Taxa**

The following sections enumerate or describe studies relating to the distribution, abundance, and trends of selected semiaquatic (i.e., taxa frequenting but not living wholly in water) amphibians and reptiles, birds, and mammals. A complete list of semiaquatic wildlife species occurring in the Willamette Basin was prepared for the WRBWQS (Tetra Tech, Inc., 1993d).

## **Amphibians and Reptiles**

Semiaquatic native amphibians and reptiles in the Willamette Basin include two species of turtle, and several species of frogs and salamanders. Most of the species use both aquatic and riparian habitats. Three salamanders, Pacific giant (*Dicamptodon tenebrosus*), Dunn's (*Plethodon dunni*), and northwestern (*Ambystoma gracile*), and one frog, tailed (*Ascaphus truei*), are considered riparian obligates (Anthony and others, 1987; Bury, 1988).

The most intensive inventory of amphibians and reptiles in the Willamette Valley was conducted in 1984–87 (St. John, 1987). The inventory included the valley floor and foothills of the Coast Range and Cascade Mountains, but not the upper elevations of the Willamette Basin. Amphibian and reptile inventories are also conducted on an opportunistic and periodic basis in the forests of the Willamette Basin by the USFS and BLM. Methods and protocols for inventorying amphibians and reptiles have been described by Applegarth (1994). Survey protocols also have been recently developed for five salamander species strongly associated with old-growth forests as part of the requirements of the Northwest Forest Plan (Olson, 1996).

Species-specific studies on the distribution and status of amphibians and reptiles in the basin have been reported for the spotted frog (*Rana pretiosa*) (Marshall, 1989; Hayes, 1994), Larch Mountain salamander (*Plethodon larselli*) (Kirk, 1983), rough-skinned newt (*Taricha granulosa*) (Kelley, 1951), and western pond turtle (*Clemmys marmorata marmorata*) (Holland, 1991; 1994). Blaustein and others (1995) provide detailed information on current status, ecology, behavior, and range of semiaquatic amphibians and reptiles inhabiting old-growth forests of the Pacific

Northwest. The occurrence of salamanders and frogs associated with streams in the Cascade Mountains has been reported by Hawkins and others (1983). The occurrence of amphibian species in the Mt. Hood National Forest is reported annually by volunteers participating in the Wetland Wildlife Watch program (Corkran, 1995).

The giant salamanders (*Dicamptodon* spp.) of the Willamette Basin include two species, Cope's giant salamander (*Dicamptodon copei*), and the Pacific giant salamander. Adult Pacific giant salamanders are relatively common in the moist coniferous forests of the basin, but are nocturnal and secretive (Blaustein and others, 1995). Only three adult Cope's giant salamanders have been described (Leonard and others, 1993), and Cope's giant salamander has only been reported from the Mt. Hood National Forest in the Willamette Basin (Marshall and others, 1996). Little is known about the larvae of either species, although they are apparently sensitive to land management practices (Corn and Bury, 1989).

Painted turtles (*Chrysemys picta*) inhabit shallow waters of ponds or small lakes, and slow-moving, backwater areas of streams and rivers in the Willamette Basin (Nussbaum and others, 1983). They prefer soft, muddy bottoms with considerable aquatic vegetation. The current status of painted turtle populations in the basin is unknown, although populations are likely declining due to unsuccessful recruitment (Gaddis and Corkran, 1985).

#### **Birds**

Semiaguatic birds in the Willamette Basin include numerous species of waterfowl, shorebirds, herons, and gulls, along with one passerine bird, American dipper, and two raptors, osprey and bald eagle. Principal breeding species of waterfowl in the Willamette Valley are mallard (Anas platyrhynchos) and wood duck (Aix sponsa). Small populations of nesting harlequin ducks (Histrionicus histrionicus), goldeneyes (Bucephala spp.), and mergansers (Mergus spp.) occur in higher elevations of the basin. Wintering waterfowl are extensive in the Willamette Valley, including mallard, pintail (Anas acuta), teal (Anas spp.), ring-necked duck (Aythya collaris), and several subspecies of Canada geese (Branta canadensis). In general, there has been a change in wintering waterfowl abundance because species adapted to feeding on agricultural crops, such as Canada geese, are now more abundant than aquatic plant and animal feeders (Puchy

and Marshall, 1993). Popular waterfowl hunting areas in the Willamette Valley include the Sauvie Island Wildlife Management Area and three National Wildlife Refuges (Finley, Baskett Slough, and Ankeny) in the mid-Willamette Valley.

The American dipper is a small resident bird that is closely associated with high gradient, montane streams in the Cascade and Coast Range Mountains. Dippers are significant components of the aquatic ecosystem because they forage within streams for aquatic insect larvae, particularly EPT taxa and Diptera (Mitchell, 1968), and some small fish, snails, and adult insects. Thus, they compete either directly or indirectly for food with fish and amphibians. Some researchers suggest they are important as bioindicators of stream quality because they are integrally tied to the aquatic invertebrate community, particularly EPT taxa (John Loegering, Oregon State University, oral commun., 1995). The only investigation of American dipper in the Willamette Basin was a winter time and energy budget study in the Cascade Range (Parsons, 1975). Ongoing research on the habitat selection and breeding season ecology of dippers in coastal streams of Oregon (John Loegering, Oregon State University, oral commun., 1995) will likely provide information applicable to dipper populations in the Willamette Basin.

The great blue heron (Ardea herodias) is a colonial nester (rookeries) in large trees along large streams, rivers, and lakes. Several studies have reported on population trends in the Willamette Basin. Henny and Bethers (1971) studied a colony near Albany and concluded that the population was stable on the basis of a comparison of productivity with that necessary to maintain a stable population. The ODFW has conducted basinwide inventories for nesting rookeries and populations of great blue heron (English, 1978; Ellingson, 1988). There were 40 more active nests counted in 1977 despite the fact that 17 more colonies were located in the 1988 census (Ellingson, 1988). Within the 24 rookeries active in both years, there was an 11 percent decrease in the number of nests. Colony fragmentation and/or a more comprehensive survey in 1988 were suggested as reasons for the differences.

The osprey is a fish-eating raptor that nests adjacent to or within a short distance of major rivers and reservoirs in the Willamette Valley. A long-term study of osprey populations along the Willamette River revealed that the number of nesting pairs in 1976 (13) increased to 78 pairs in 1993 (Henny and others, 1978;

Henny and Kaiser, 1996). It also revealed a change in nesting structures from live or dead trees (all 13 nests in 1976) to utility structures or nesting platforms (66 of 78 nests in 1993). Thus, the population nesting in the apparently small number of suitable nesting trees remained relatively unchanged (13 nests in 1976 and 12 in 1993). Factors suggested by Henny and Kaiser (1996) for the population increase include the learned response to use utility structures, a reduction in DDT-related reproductive problems, improved water conditions and fish populations in the Willamette River, and reduced shooting of adults.

### **Mammals**

Semiaquatic mammals in the Willamette Basin include beaver (*Castor canadensis*), river otter (*Lutra canadensis*), mink (*Mustela vison*), muskrat (*Ondatra zibethica*), nutria (*Myocastor coypu*), Steller sea lion (*Eumetopias jubatus*), and Pacific water shrew (*Sorex pacificus*). Most of the species use both aquatic and riparian habitats and several depend upon aquatic fauna as prey. For example, mink use both riparian and instream habitat, and primarily depend upon aquatic prey, such as crayfish and fish. The Pacific water shrew is a riparian obligate species (Anthony and others, 1987; Gomez, 1992; McComb and others, 1993) that is endemic to the coastal Pacific Northwest. It is mostly found in or near water (Christenson and Larrison, 1982).

Beaver are keystone species in aquatic and riparian habitats, and are responsible for natural disturbances to aquatic systems. Their dens and lodges are used as dens and rest sites for species such as river otter and provide habitat for other smaller species, including salamanders, mice, and voles. Pools created by beaver dams are also important habitat for a number of aquatic species.

The modern aquatic furbearer industry is small, particularly compared to the historical extent of the industry. Populations of aquatic furbearers have declined due to historic overexploitation from trapping and from habitat loss and degradation due to several factors associated with an expanding human population. Historical data on aquatic furbearer harvest is presented in the Basin Investigation Report for the upper Willamette Basin (Hutchison and others, 1966a). A study of the population status of the river otter in western Oregon included data from trapping conducted in the Willamette Basin during 1970–1972 (Tabor, 1974).

Population management of aquatic furbearers is based on harvest regulations set by the ODFW.

# **Introduced Species**

Numerous introduced (nonnative) species are present and have established populations in the Willamette Basin. Many introductions were intentional (several game fish), some species escaped from confinement (nutria, red-eared slider [*Pseudemys scripta elegans*]), and some species immigrated following introductions elsewhere (walleye). Additionally, the use of live bait for fishing has resulted in some fish introductions. Puchy and Marshall (1993) list suspected or known sources of introductions for fish in the basin.

The widest variety of introduced species occurs in lowland rivers, lakes, and ponds that support warm water ecosystems similar to the native habitats of most of these species (Bond and others, 1988). Additionally, the increase in slow-moving, deep-water habitat created by dam construction and bank revetments has likely contributed to the establishment and population increases for many of these species (Hjort and others, 1984; Farr and Ward, 1993).

An early history of fish introductions in the Willamette Basin is included in Lampman (1946). The timing of several fish introductions into the lower Willamette River is discussed in Farr and Ward (1993). Logan and others (in press) recently documented the occurrence of the nonnative aquarium fish, oriental weatherfish (*Misgurnus anguillicaudatus*), in the Clackamas subbasin. Within the Willamette River, Hughes and Gammon (1987) reported that the number of native fish species in the lower river was approximately half that in the upper river. Over half of the fish species recorded in the Portland Harbor were introduced to the Willamette River system (Farr and Ward, 1993).

Black crappie and white crappie are introduced warm water fishes occurring in lakes, impoundments, and relatively stagnant areas of rivers. Black crappie were estimated to be four times as abundant as white crappie in the lower Willamette River near Portland (Ward and others, 1991). Large individuals of both species have been reported to prey upon juvenile salmonids in the Willamette Basin (Grenfell, 1962; Ward and others, 1991), although the predation level is probably low (Ward and others, 1991).

The common carp is one of the most notorious introduced fish species and is widely regarded as a pest species that is difficult to eradicate or control. Common carp were introduced into the Pacific Northwest in the early 1880s as a food fish (Wydoski and Whitney, 1979). They occur throughout lowland aquatic habitats in the basin. The mosquito fish (*Gambusia affinis*) also has been introduced in lowland aquatic habitats, particularly urban and residential areas, for mosquito control.

The bullfrog (*Rana catesbeiana*) was introduced into the western United States to be farmed for sale in food markets, and spread rapidly in lowland aquatic habitats to where they are often the dominant species (Bury and Whelan, 1984). They have been directly or indirectly implicated in the decline or extirpation of a number of native amphibians and reptiles, particularly other *Rana* frogs (Bury and Whelan, 1984; Hayes and Jennings, 1986). The bull frog is believed to be the principal cause of extirpation of the spotted frog from the Willamette Valley (St. John, 1987; Marshall and others, 1996).

Since the introduction of escaped or fur-farm released nutria, this species has spread rapidly throughout the basin. They are semiaquatic and use the instream and shoreline habitat of lowland lakes, ponds, and slow-moving rivers in the basin. They are considered nuisance animals because of their ecological competition with beaver and muskrat, and the adverse physical effect (erosion) of their burrowing activities on streambanks. Peloquin (1969) studied growth and reproduction of nutria near Corvallis.

The Asiatic clam (*Corbicula fluminea*) was first introduced to the United States early this century, probably along the west coast in San Francisco Bay. Its hermaphroditic reproductive mode has allowed it to spread rapidly in most rivers throughout the west coast. Asiatic clams are found in the main stem Willamette River and in the lower sections of most tributaries. Where present, it is the recommended taxon for analysis of organochlorine compounds and trace elements in the NAWQA Program (Crawford and Luoma, 1993)

The aquatic macrophyte, Eurasian watermilfoil (*Myriophyllum spicatum*), is discussed here because it affects water quality and aquatic biota. The presence of this aggressive species is relatively recent to the Willamette Basin. Its occurrence in the Pacific Northwest dates from the late 1960s in British Columbia (Geiger, 1986). Within the Willamette Basin, the USACE (1982) documented the occurrence of the species in

the Delta Ponds near Eugene, and Halse and Dennis-Johnston (1981) reported its presence in the Coast Fork Willamette River near Eugene, at Fern Ridge Lake, and at Clear Lake. Its dominance in the Delta Ponds of Eugene was reported by Fetrow Engineering and Scientific Resources (1989). It is well established now in the Willamette River above Delta Ponds.

# **Special Status Species**

Special status species are defined here as species designated by the USFWS or ODFW as threatened or endangered, USFWS candidate species or species of concern, ODFW and USFS sensitive species, or species considered species of special concern by recognized experts, such as malacologists for mollusc species. A listing of special status aquatic fauna that are suspected or documented to occur in the Willamette Basin is presented in table 5. A separate listing of mollusc species of concern is presented in table 6.

There are no threatened or endangered species of aquatic macroinvertebrates in the Willamette Basin. However, 16 species of aquatic macroinvertebrates in the basin have special status, including 14 Federal species of concern (table 5). Twelve of the 16 species are caddisflies. This high number is partly due to the amount of information on caddisflies, but also due to their sensitivity to stream degradation. Other special status aquatic macroinvertebrate species include a stonefly, beetle, snail, and clam (table 5). In general, not enough is known about the status of aquatic macroinvertebrate species to adequately determine if they should be removed from the list or upgraded to candidate status.

Surveys have been conducted for special status caddisflies on the Mt. Hood National Forest (Wisseman, 1989) and Willamette National Forest (Wisseman, 1992c). Wisseman (1990) presents an overview of the ecology of several invertebrate special status species occurring on the Mt. Hood National Forest.

Frest and Johannes (1993) list 10 freshwater mollusc species of concern (8 snails and 2 clams) known or suspected to occur in the Willamette Basin table 6). Factors primarily responsible for the special status include impacts from dams/impoundments (such as alteration of flows), fluctuations in water temperatures, and degradation/loss of habitat; other factors are increases in siltation, nutrient enrichment, pollution, channelization and dredging, and land use practices

**Table 5.** Aquatic fauna with special status that occur in the Willamette Basin, Oregon [USFWS, U.S. Fish and Wildlife Service; USFS, U.S. Forest Service; ODFW, Oregon Department of Fish and Wildlife; --, No status or plans, trend unknown]

			Status			<u> </u>	
Species	Scientific Name	USFWS <sup>1</sup>	USFS <sup>2</sup>	ODFW <sup>3</sup>	Trend <sup>4</sup>	Plans <sup>5</sup>	Reasons for Status <sup>6</sup>
Mammals							
Steller sea lion	Eumetopias jubatus	T		V			Unknown; prey loss; disturbance
<u>Birds</u>							
Aleutian Canada goose	Branta canadensis leucopareia	T		E			
Bald eagle	Haliaeetus leucocephalus	T	X	T		yes	
Barrow's goldeneye	Bucephala islandica			U			Limited distribution; forestry practices
Bufflehead	Bucephala albeola			U			Limited distribution; forestry practices
Harlequin duck	Histrionicus histrionicus	SC	X	U	$U^7$		Limited distribution
<u>Fish</u>							
River lamprey	Lampetra ayresi	SC			D		
Pacific lamprey	Lampetra tridentata	SC		V	U		
Lower Columbia River coho salmon	Oncorhynchus kisutch	PT	X	С			Stream barriers; forestry practices; introductions; overutilizations
Coastal cutthroat trout	Oncorhynchus clarki clarki			С			Forestry practices; stream barriers; livestock grazing; urban development
Lower Columbia River fall chinook salmon	Oncorhynchus tshawytscha			С			Forestry practices; stream barriers
Oregon chub	Oregonichthys crameri	E	X	С		yes	Introductions; wetland draining; channelization; stream barriers
Bull trout	Salvelinus confluentus	С	X	С	D	(8)	Forestry practices; overutilizations; introductions; passage barriers; chemica treatment projects
Amphibians and Reptiles							
Western pond turtle	Clemmys marmorata marmorata	SC	X	С	U		Farming practices; wetland loss; introductions; urban development
Painted turtle	Chrysemys picta		X	C			Unknown; introductions; wetland loss
Clouded salamander	Aneides ferreus			U			Forestry practices
Cope's giant salamander	Dicamptodon copei		X	U			

Table 5. Aquatic fauna with special status that occur in the Willamette Basin, Oregon—Continued

			Status				
Species	Scientific Name	USFWS <sup>1</sup>	USFS <sup>2</sup>	ODFW <sup>3</sup>	Trend <sup>4</sup>	Plans <sup>5</sup>	Reasons for Status <sup>6</sup>
Amphibians and Reptiles—Continue	d						
Larch Mountain salamander	Plethodon larselli	SC	X	V			Limited distribution; threat from forestry practices
Oregon slender salamander	Batrachoseps wrighti			U			Forestry practices
Cascade torrent (seep) salamander	Rhyacotriton cascadae			V			Forestry practices
Columbia torrent (seep) salamander	Rhyacotriton kezeri			С			Forestry practices
Southern torrent (seep) salamander	Rhyacotriton variegatus	SC		С	D		Forestry practices
Western toad	Bufo boreas			V			
Tailed frog	Ascaphus truei	SC		V	D		Forestry practices
Northern red-legged frog	Rana aurora aurora	SC	X	U	U		Unknown
Foothill yellow-legged frog	Rana boylii	SC		V	D		Unknown
Cascades frog	Rana cascadae	SC		V	U		Drought; fish introductions: pathogens; habitat loss
Spotted frog	Rana pretiosa	C		C	U		Unknown; introductions
<u>Invertebrates</u>							
Beer's false water penny beetle	Acneus beeri	SC	X		U		
California floater	Anodonta californiensis	SC			D		Dams and impoundments
Columbia River pebblesnail or spire snail	Fluminicola columbianus	SC			U		Dams and impoundments
Cascades apatanian caddisfly	Apatania tavala	SC	X		U		<del></del>
Vertrees's ceraclean caddisfly	Ceraclea vertreesi	SC			U		
Mt. Hood primitive brachycentrid caddisfly	Eobrachycentrus gelidae	SC	X		U		
Mt. Hood farulan caddisfly	Farula jewetti	SC	X		U		
Tombstone Prairie farulan caddisfly	Farula reaperi	SC	X		U		
Fort Dick limnephilus caddisfly	Limnephilus atercus	SC	X		U		

**Table 5.** Aquatic fauna with special status that occur in the Willamette Basin. Oregon—Continued

			Status					
Species	Scientific Name	USFWS <sup>1</sup>	USFS <sup>2</sup>	ODFW <sup>3</sup>	Trend <sup>4</sup>	Plans <sup>5</sup>	Reasons for Status <sup>6</sup>	
<u>Invertebrates</u> —Continued								
Columbia Gorge neothremman caddisfly	Neothremma andersoni	SC	X		U			
Alsea ochrotrichian micro caddisfly	Ochrotrichia alsea		X					
Tombstone Prairie oliogophlebodes caddisfly	Oligophlebodes mostbento	SC	X		U			
Haddock's rhyacophilan caddisfly	Rhyacophila haddocki	SC	X		U			
One-spot rhyacophilan caddisfly	Rhyacophila unipunctata	SC	X		U			
Siskyou caddisfly	Tinodes siskiyou	SC	X		U			
Wahkeena Falls flightless stonefly	Zapada wahkeena		X		U			

<sup>1</sup> Sources: U.S. Fish and Wildlife Service (1994a, 1994b; G. Miller, U.S. Fish and Wildlife Service, written commun., 1996); E, Endangered; T, Threatened; PT, Proposed Threatened; C, Candidate; SC, Species of Concern.

<sup>&</sup>lt;sup>2</sup> Source: U.S. Forest Service (1989); X, Sensitive.

<sup>&</sup>lt;sup>3</sup> Source: C. Puchy (Oregon Department of Fish and Wildlife, written commun., 1995); E, Endangered; T, Threatened; C, Critical; V, Vulnerable; U, Undetermined.

<sup>&</sup>lt;sup>4</sup> Source: U.S. Fish and Wildlife Service (1994b); Throughout the species range, D, Declining; U, Unknown.

<sup>&</sup>lt;sup>5</sup> Includes recovery plans for threatened and endangered species, and management plans or conservation strategies/agreements for other species.

<sup>&</sup>lt;sup>6</sup> Sources: Most are from Marshall and others (1996) and Puchy and Marshall (1993); other sources include Ratliff and Howell (1992) for bull trout;

Frest and Johannes (1993) for California floater and Columbia River spire snail; Blaustein and others (1994) for foothill yellow-legged frog; and Scheerer and others (1995) for Oregon chub.

<sup>&</sup>lt;sup>7</sup>Cassirer and others (1993) indicated a stable to declining trend in western North America.

<sup>&</sup>lt;sup>8</sup>The McKenzie/Middle Fork Willamette bull trout working group is developing a conservation strategy (J. Ziller, Oregon Department of Fish and Wildlife, oral commun., 1995).

Table 6. Aquatic mollusc species of concern in the Willamette Basin, Oregon

[Source: Frest and Johannes (1993), except two Federal species of concern, Fluminicola columbianus and Anodonta californiensis, which are listed in table 8]

Common Name	Scientific Name	Distribution	Status
<u>Snails</u>			
Barren juga	Juga (Juga) hemphilli hemphilli	Multnomah County; east of Willamette River in Sandy Subbasin	Species of concern
None	Juga (Juga) hemphilli n. subsp.	Multnomah County; Mount Hood National Forest	Species of concern
Brown juga	Juga (Juga) n. sp. 1	Multnomah County; Mount Hood National Forest	Species of concern
Tall juga	Juga (Juga) n. sp. 3	Multnomah County: Mount Hood National Forest	Species of concern
Columbia duskysnail	Lyogyrus n. sp. 1	Multnomah County; Mount Hood National Forest	Species of concern
Rotund physa	Physella (Physella) columbiana	Near Columbia River in Columbia and Multnomah Counties	Species of concern <sup>2</sup>
Nerite rams-horn	Vorticifea neritoides	Near Columbia River in Columbia and Multnomah Counties	Species of concern
Clams			
Willamette floater	Anodonta wahlametensis	Lower Willamette and Columbia Rivers in Columbia, Multnomah, and Clackamas Counties	Species of concern <sup>1,2</sup>

<sup>&</sup>lt;sup>1</sup> Impacted by dams and impoundments.

such as grazing and logging. Frest and Johannes (1993) and Roth (1993) caution on the completeness of mollusc faunal lists because malacological research and knowledge is minimal, and it is likely that many new species and even genera remain to be discovered and described.

Currently, the only species of fish occurring in the Willamette Basin whose population is listed as threatened or endangered is Oregon chub (U.S. Fish and Wildlife Service, 1993) (table 5). Lower Columbia River coho salmon has been proposed for listing as a threatened species, and bull trout is a Candidate species (Gary Miller, U.S. Fish and Wildlife Service, written commun., 1996). Two species of fish are Federal species of concern; river lamprey (*Lampetra ayresi*) and Pacific lamprey (*Lampetra tridentata*). Two other species, coastal cutthroat trout and lower Columbia River fall chinook salmon, are listed as sensitive by the ODFW.

The decline of anadromous fish stocks throughout the Pacific Northwest, including the Willamette Basin, has been recognized for many years. The magnitude and extent of these declines were documented in an American Fisheries Society report by Nehlsen and others (1991). They identified over 100 stocks of anadromous salmon and trout as extinct, 102 stocks with a high risk of extinction, 58 with a moderate risk of extinction, and 54 of special concern due to low numbers and/or restricted distribution. For the Willamette Basin, two stocks were identified as already extinct, two at a high risk of extinction, two at a moderate risk of extinction, and two of special concern due to low numbers and/or restricted distribution (table 7). Declines in the Willamette Basin are further exemplified by a report that indicated that none of the 121 healthy native stocks of anadromous salmonids identified in the Pacific Northwest and California were within the Willamette Basin (Huntington and others, 1994). Documentation of declines of salmonids is problematic due to natural fluctuations in populations due to oceanic conditions (Lawson, 1993), and the lack of long-term data sets for many species/stocks.

The Oregon chub endemic to the Willamette Valley (Markle and others, 1991), was listed by the

<sup>&</sup>lt;sup>2</sup> May be extirpated from the Willamette and Columbia Rivers.

**Table 7.** Stocks of salmon and trout that are extinct or at risk of extinction in the Willamette Basin, Oregon [Source: Nehlsen and others (1991) for all except bull trout (Ratliff and Howell, 1992). Threat codes: 1, destruction, modification, and loss of habitat (includes passage and flow problems and predation in reservoirs); 2, overutilization for commercial, recreational, scientific, or educational purposes; 3, other factors, including hybridization, introduction of nonnative species, competition, and poor ocean survival conditions]

Species	Stock/Population	Risk	Threats
Chinook salmon	Willamette River, spring race	Special concern	1,2,3
	Willamette River, fall race	Extinct	?
	Sandy River, spring race	Extinct	1,3
	Sandy River, fall race	High	1,2
Coho salmon	Clackamas River	Moderate	1,2,3
	Sandy River	High	1,2,3
Steelhead trout	Clackamas River	Moderate	1,2,3
	Calapooia River	Special concern	1
Bull trout	Middle Fork Willamette River	High	1,2,3
	South Fork McKenzie River	Moderate	1,2
	McKenzie River, Anderson Creek	Moderate	1,2,3
	Trailbridge Reservoir	High	1,2,3
	Carmen Reservoir	Probably extinct	1,2,3
	North Santiam River	Probably extinct	1,2,3
	South Santiam River	Probably extinct	2,3
	Clackamas River	Probably extinct	1,2,3

USFWS as an endangered species in 1993 (U.S. Fish and Wildlife Service, 1993). Historically, it occurred in ponds and quiet waters of backwater reaches of the Willamette River and its tributaries. Oregon chub populations have been reduced drastically from former levels primarily due to predation by introduced species, particularly largemouth bass (Markle and others, 1989), and loss of habitat due to alteration of the hydrography of the Willamette River (Marshall and others, 1996). Water-quality degradation, habitat loss due to flood control, draining of wetlands, and channelization of the main stem Willamette River have also likely contributed to the species decline (Pearsons, 1989; Rien and others, 1992; Scheerer and others, 1995). Surveys have been conducted since 1990 by the ODFW throughout the Willamette Basin to quantify existing populations, search for unknown populations, and evaluate potential reintroduction sites (Scheerer and others, 1992, 1993, 1994, 1995). Current known localities include several areas in or adjacent to the Middle Fork Willamette River, Dry Muddy Creek in Linn County, the Finley National Wildlife Refuge, and a small section of the Santiam River (Marshall and others, 1996). Since 1970, no Oregon chub has been found in the main stem Willamette River (Rien and others, 1992). Other status and distributional studies include Long (1982), Bond and Long (1984), Pearsons (1989), and Markle and others (1989; 1991). A multiagency Conservation Agreement provides guidelines for management and reintroduction of the Oregon chub in the

Willamette Basin (Oregon Department of Fish and Wildlife, 1992).

Bull trout is listed as a Federal candidate species (U.S. Fish and Wildlife Service, 1994b). It spawns in the cold water of headwater tributaries of the basin and migrates downstream into larger tributaries in the same subbasin (Goetz, 1989). Historic and current distribution of the bull trout in the Willamette Basin has been described by Ratliff and Howell (1992) and Goetz (1994). The only known current locations of bull trout in the basin are three populations in the McKenzie subbasin and one population in the Middle Fork Willamette River. The population viability at all the sites is considered to be of moderate to high risk for extinction (Ratliff and Howell, 1992), and no individuals have been recorded from the Middle Fork Willamette River since 1992 (Mark Wade, Oregon Department of Fish and Wildlife, oral commun., 1995). Historical populations in the Clackamas River and the North and South Santiam Rivers are believed to be extinct. Factors contributing to the current population status include passage barriers, habitat degradation, overharvest, and hybridization and competition with brook trout (Salvelinus fontinalis) (Ratliff and Howell, 1992).

There are no threatened or endangered amphibians and reptiles in the Willamette Basin. However, 15 species of amphibians and reptiles have special status: 2 turtles, 5 frogs, 1 toad, and 6 salamanders (table 5). Of particular concern within the Willamette Basin are widespread declines of frogs of the genus *Rana* (Hayes

and Jennings, 1986), including four of the five special status frog species: spotted frog, a Federal candidate species; and Cascades frog (*Rana cascadae*), foothill yellow-legged frog (*Rana boylii*), and red-legged frog (*Rana aurora*), Federal species of concern.

Several semiaquatic amphibians and reptiles have experienced basinwide declines that mirror those experienced at larger scales, including globally (Blaustein and Wake, 1990). Additionally, some species have experienced range reductions with extirpations of populations from specific areas of the Pacific Northwest, including the Willamette Basin (Corn and Bury, 1989). For example, the northern red-legged frog has been extirpated from much of the Willamette Valley (Blaustein and others, 1994), and the spotted frog has apparently been completely extirpated from the Willamette Valley, in addition to most if not all of the Willamette Basin (Hayes, 1994). Within the Willamette Basin, these declines, range reductions, and population extirpations have likely resulted from a number of factors, particularly habitat loss, insecticides and pollution, and predation by introduced predators (St. John, 1987).

The spotted frog is listed as a Federal candidate species (U.S. Fish and Wildlife Service, 1994b). The spotted frog is an obligate aquatic species found in marshes near the edges of ponds and lakes (Nussbaum and others, 1983). Its historical range included all of the Willamette Basin, but it is believed to be extirpated from the Willamette Valley, and has not been found west of the Cascade Range in Oregon since the early 1970s (McAllister and others, 1993). Suggested causes for the decline include predation by the introduced bullfrog (Nussbaum and others, 1983; St. John, 1987), toxics (Kirk, 1988), introduced warm water fishes (Hayes and Jennings, 1986), and degradation and elimination of wetland habitats.

The western pond turtle is a Federal species of concern. It is absent from much of its former range in the Willamette Valley (Holland, 1991; Marshall and others, 1996; Bury and Holland, in press), and where present, there is often little evidence of reproduction. It inhabits ponds, sloughs, marshes, and slow-moving sections of rivers where basking sites (logs, exposed tree roots or rocks, vegetation mats) are present (Nussbaum and others, 1983). Because of the length of time to reach maturity (8–11 years), depleted populations rebound slowly. Factors likely contributing to declines include degradation and loss of habitat, predation by introduced species such as bullfrog and largemouth

bass, drought, and diseases (Marshall and others, 1996; Bury and Holland, in press). The most comprehensive and current information on the status and ecology of the western pond turtle in the Willamette Basin is provided in Holland (1994). Research has been conducted on the potential effects of improvements to the Beltline Highway in Eugene on western pond turtles (Fishman Environmental Services, 1994; CH<sub>2</sub>M Hill, 1994; Beak Consultants, Inc., 1994); and inventory, trapping, and movements of northwestern pond turtles at Fern Ridge Lake (Beal and Thaut, 1994).

Two semiaquatic bird species occurring in the Willamette Basin are federally listed as threatened—the bald eagle and the Aleutian Canada goose (*Branta canadensis leucopareia*)—and one species, the harlequin duck (*Histrionicus histrionicus*), is a Federal species of concern (table 5). Two other species, bufflehead (*Bucephala albeola*) and Barrow's goldeneye (*Bucephala islandica*), are listed as sensitive by the ODFW.

The harlequin duck is a Federal species of concern that nests along whitewater mountain streams of the Cascade Mountains and winters along the coast (Marshall and others, 1996). There is limited historical information on the species (Latta, 1992), but the distribution and abundance has likely always been relatively low based on the limited available habitat. Latta (1992) compiled historical and recent sightings in the Cascade Mountains, and Thompson and others (1993) reported on abundance, distribution, and habitat associations. Recent breeding has been confirmed at a few locations on the Mt. Hood National Forest (Marshall and others, 1996), and along the Molalla, Santiam, and McKenzie Rivers (Thompson and others, 1993; Marshall and others, 1996). A graduate student project initiated in 1994 is examining productivity and breeding season habitat use in the Cascade Mountains (Howard Bruner, Oregon State University, oral commun., 1995).

The only special status semiaquatic mammal occurring in the Willamette Basin is the Steller sea lion. It occurs in small numbers in the Willamette River, particularly below Willamette Falls, where it is attracted to runs of spring chinook salmon and winter steelhead trout.

## **IMPACTS ON AQUATIC COMMUNITIES**

Since European settlement of the Willamette Basin in the early 1800s, environmental changes have resulted in substantial changes to the aquatic communities of the basin. In general, as aquatic habitats disappeared or were degraded, aquatic biota dependent upon them declined or were extirpated, and there has been a general trend in reduction of biotic diversity (Holland, 1994).

Several recent aquatic ecosystem assessments describe the condition of aquatic communities in the Willamette Basin. The FEMAT (1993) report indicated that 95 percent of the streams surveyed in Oregon in 1988 were moderately to severely impaired. The ODEQ (1992) reported that only 4 percent of the 14,113 acres of lakes, and 32 percent of the 4,714 miles of streams and rivers surveyed in the Willamette Basin were listed as fully supporting potential beneficial uses. Recent work as part of the WRBWQS indicated that the Willamette River is "slightly to moderately impaired" compared to upstream locations on the basis of fish and invertebrate community composition (Tetra Tech, Inc., 1993b). The USFS (1993) compared current aquatic ecosystem conditions with the range of natural conditions for five rivers in the Willamette Basin (Clackamas, North Santiam, South Santiam, McKenzie, and Middle Fork Willamette) using two primary indicators, maximum water temperature and frequency of large pool habitat. They reported that most streams exhibit significant signs of degraded conditions, including being below the range of natural variability in pools per mile, and exceeding the natural range of maximum summer water temperatures.

Aquatic communities in the Willamette Basin are impacted by numerous natural (floods, fires, landslides, beaver activity) and human-related factors. The effects of these impacts are variable and must be assessed within the substantial spatial environmental variability (soils, slope, climate, vegetation) that exists within the basin. Multiple impacts can be cumulative, and many of the impacts also have secondary impacts, such as dams and introduced fish, and dams and fish disease. Additionally, the response of an individual organism to impacts that alter water quality and (or) the physical habitat in which it exists may be lethal or sublethal, such as effects on behavior, physiology, physical development, or reproduction. Thus, assessment of impacts on aquatic resources is complex and a determination of cause and effect can be difficult. In general, the cumulative effect of the many physical changes has been to simplify biological communities and increase the dominance of species most tolerant of altered conditions.

Although some sources of impact on aquatic communities are the result of naturally occurring events, the principal impacts in the Willamette Basin have resulted from human activities. Historically, one of the most extensive changes in aquatic/riparian habitat within the Willamette Valley occurred as a result of channelization and constrainment of the main stem Willamette River (Sedell and Froggatt, 1984). These changes were greatest in the southern half of the river, which historically was a braided system of numerous oxbows, sloughs, ponds, and small side-channels and a broad floodplain with extensive marshlands and riparian gallery forests. Two other human impacts on aquatic communities were massive clearing of riparian forests and draining and filling of wetland habitats. Extensive discussion on the impact of humans in these habitats within the Willamette Basin is presented in Holland (1994).

Impacts on aquatic communities also occur when land-use activities greatly accelerate natural processes of sedimentation and erosion, and when artificial elements, such as toxic chemicals or channelization, are introduced or alter the stream (Bottom and others, 1985). Several land use activities, such as irrigation, power generation, and municipal and industrial uses, also require water withdrawals, which have contributed to decreased streamflows and increased water temperatures in tributaries and upper reaches of the main stem Willamette River. Historically, municipal and industrial point source discharges were the principal impacts on water quality (Gleeson, 1972). Recently, most concern regarding water quality has focused on nonpoint source pollution caused by landuse activities such as agriculture, urbanization, logging, and road construction. A summary of land-use activities in the Willamette Basin, and their contributions as nonpoint sources of pollution, is provided in Tetra Tech, Inc., (1992c).

During the last 150 years, a variety of human impacts have seriously reduced the capacity of rivers and streams in the Pacific Northwest, including in the Willamette Basin, to support anadromous salmonids (Huntington and others, 1994). Responses of salmonid populations to these perturbations, particularly timber harvesting, have been investigated extensively (Hicks and others, 1991; Meehan, 1991). Bottom and others (1985) provides an overview of the impact of land-use practices on salmonid habitat and production in Oregon, and techniques to reduce these impacts. Hall and Baker (1982) also describe impacts on salmonid habi-

tat, and review methods to rehabilitate and enhance stream habitat.

Declining anadromous fish stocks in the Willamette Basin and elsewhere in the Pacific Northwest have been attributed to numerous factors, including loss and degradation of freshwater and riparian habitats; poor management and hatchery practices; introduction of nonnative fish species; construction and operation of dams and their affects on habitat, water flows, temperature, predation, mortality, and passage; and management of land uses, such as timber harvesting, grazing, and agriculture. Overfishing late in the 19th century also contributed to declines in anadromous fish runs (Willamette Basin Task Force, 1969), particularly for summer run chinook salmon (Li and others, 1987). Unlike resident fishes, anadromous salmonids are also subject to stresses encountered outside of the Willamette Basin, which have likely contributed to their declines (Lawson, 1993).

Impacts on anadromous salmonid populations in the Pacific Northwest have been estimated by several investigators. Approximately 16 million wild salmon and steelhead were produced annually in the Columbia Basin (including the Willamette Basin) 120 years ago (Wevers, 1994). This compares to the approximately 2 million produced today, about 80 percent of which are hatchery fish. The Northwest Power Planning Council (1986) further estimated that salmonid production in the Columbia Basin has declined 75-85 percent since settlement of the region by Europeans, with a reduction in wild fish production of about 95 percent. Similar reports of drastic declines have been reported for the Puget Sound (Bledsoe and others, 1989), Oregon coastal streams (Nickelson and others, 1992), California (Moyle, 1994), and northern California (Higgins and others, 1992).

The assessment of impacts on aquatic biota, particularly land-use impacts, can be complicated by natural variation in populations. Hall and Knight (1981) reported that year-to-year natural variation in salmonid population densities can be up to several orders of magnitude. House (1995) reported a substantial fluctuation (as much as two-fold between years) in populations of cutthroat trout populations over an 11-year period in the Pudding/Molalla subbasin, despite similar habitat conditions from year to year and an absence of management activities. He suggests caution regarding the development of conclusions on the effect of management activities based on short-term data collection. In addition, wide natural fluctuations in populations can

mask declines, and the increase in releases of hatchery fish complicates assessment of population status and trends of native stocks. Global and regional weather patterns, such as El Niño, can also significantly affect fish populations (Nickelson and Lichatowich, 1983; Mysak, 1986), particularly anadromous fish species, thus emphasizing the need to assess impacts in the context of long-term trends.

# SOURCES OF IMPACTS ON AQUATIC COMMUNITIES

#### **Natural Effects**

Some natural features within the Willamette Basin impact aquatic biota in regular and predictable ways. Willamette Falls at Oregon City on the main stem Willamette River served as a complete barrier to upstream migration of salmonids during the low flows of summer and fall, and a partial barrier at other times of the year, prior to improvements in fish ladders in the late 19th century (Holmes and Bell, 1960; Hutchison and Aney, 1964; Willamette Basin Task Force, 1969; Clady, 1971; Frazier, 1989). The first crude fish ladder to aid in the passage of salmonids at Willamette Falls was constructed from rock in 1885 (Holmes and Bell, 1960; Sams, 1977). It was followed by a more effective fish ladder in the mid-1890s and by others over the next 60 years as engineering and technological advancements occurred (Holmes and Bell, 1960; Oregon Department of Fish and Wildlife, 1982b). Holmes and Bell (1960) provide a history of the use of the falls to generate power and of the development and construction of fishways. Completion of the present fishway in 1971 has not only enhanced existing salmonid runs, but has allowed for the development of new summer and fall runs, including fall chinook salmon, coho salmon, and summer steelhead (U.S. Army Corps of Engineers, 1982).

Since 1946, the ODFW has reported annually on counts of fish passing through the fishway at Willamette Falls (Pulford, 1955; Holmes and Bell, 1960; Collins, 1968; Bennett, 1982; Frazier, 1989). Annual reports on counts of spring chinook salmon runs below Willamette Falls have also continued since 1946 (Bennett, 1985, 1995). A summary of the annual passage of spring chinook salmon, fall chinook salmon, coho salmon, sockeye salmon (*Oncorhynchus nerka nerka*), winter steelhead trout, and summer steelhead trout at

Willamette Falls from 1946–1981 is presented in USACE (1982).

Several studies have been conducted in the basin on the effects of natural disturbances on aquatic biota. The effects on biota from a catastrophic natural debris torrent caused by severe flooding in a Cascade Mountain stream near the HJAEF (Quartz Creek) were the complete, but relatively short-term, decimation of faunal populations (Lamberti and others, 1991; Anderson, 1992). Anderson (1992) noted the short-term elimination of insect fauna in a 300-meter reach of the stream. However, recovery was rapid, with emergence density and taxonomic richness similar to an upstream control site within one year, although effects on community structure persisted into the second year (Lamberti and others, 1991). Populations of cutthroat trout were decimated by the disturbance, but also recovered to predisturbance densities by the following year (Lamberti and others, 1991). Habitat surveys conducted in 1965 to determine the effect of floods on fish habitat in tributaries of the Clackamas River indicated that the greatest damage was loss of salmonid rearing habitat (Sams, 1965). Insect drift from fall floods in Oak Creek displaced large numbers of individuals, but overall biomass increased due to fall hatching and colonization from upstream areas (Anderson and Lehmkuhl, 1968; Lehmkuhl, 1969). They also noted that the disruption and temporary loss of habitat and allochthonous food may be more detrimental to aquatic insect populations than direct mortality caused by the floods.

When nutrient levels are excessive, nuisance algal blooms may develop in streams, ponds, lakes, and slackwater habitats. Thick mats of algae, particularly filamentous forms, can develop and adversely affect aquatic fauna by depleting oxygen in the water column (Johnson and others, 1985).

Beaver removal of trees along stream courses has various affects on aquatic communities and biota. Beaver dams modify stream hydrology, accumulate sediment, and increase wetted surface area. They are also important in creating habitat for coho salmon (Everest and Sedell, 1983), and the flooded areas create wetland habitat for many species of wildlife. Tree felling by beavers has a positive impact on pond turtle habitat by increasing the suitability of a given area for basking and providing tree trunks in the water for turtle basking, foraging, and refuge sites (Holland, 1994).

Steller sea lions prey upon migrating salmonids in the lower Willamette River, particularly below Willamette Falls. The ODFW is exploring nonlethal alternatives to control their impact on declining populations of spring chinook salmon and winter steelhead trout, including blocking access to the fish ladder, hazing techniques, and capture and removal.

#### **Overharvest**

The salmon fisheries of the Pacific Northwest have been heavily exploited since early settling of the area, including intensive fishing by native Americans at natural barriers (McKernan and others, 1950). A deleterious affect of overfishing for summer run chinook salmon throughout the Columbia Basin (includes the Willamette Basin) was documented in the late 1800s, even prior to commercial fishing (Li and others, 1987). An ecological impact from overharvest of anadromous salmonids is a substantial reduction in primary productivity in natal streams resulting from a reduction in the nutrients otherwise provided by carcass decomposition (Li and others, 1987). This decrease in primary productivity likely transfers to secondary and tertiary production.

Populations of other aquatic fauna in the basin have also been reduced from overharvest. Aquatic furbearer populations have been substantially reduced, initially due to overharvest. In fact, extensive trapping of beaver was probably the first form of nonnative human exploitation of aquatic resources in the basin, and was the initial reason for settlement of the area (Holland, 1994).

## Channelization and Bank Stabilization

Extensive channelization of the Willamette River since the late 1800s has reduced a historic river of meandering, braided channels with numerous sloughs and backwater areas and a broad floodplain (average width 1–2 miles wide) to essentially a single channel (Hjort and others, 1984; Sedell and Froggatt, 1984; Li and others, 1987). This channelization was partially done by closing off side channels with felled cottonwoods (*Populus* spp.) from the riparian zone. Channelization of the Willamette River was essentially complete by 1946, and it is estimated that 75 percent of the original shoreline has been lost to channelization (Sedell and Froggatt, 1984). The reasons for channelization were to facilitate river navigation, reduce land erosion, and increase land available for farming (Sedell and Froggatt, 1984; Pearsons, 1989).

The extensive channelization of the main stem Willamette River has resulted in a much simplified ecosystem and the loss of much of the original fish habitat. There has been a reduction in the number of side channels and off-channel refugia conducive for salmon and trout spawning and juvenile rearing. In 1854, the 15.6 mile distance between Harrisburg and the McKenzie River had over 156 miles of shoreline, but today there is less than 40 miles (Sedell and Froggatt, 1984). Dredging and channelization have also resulted in reduced organic material (leaf litterfall, downed trees) inputs (Sedell and Froggatt, 1984), increased water turbidity and bottom siltation, and removal of valuable spawning gravel (Willamette Basin Task Force, 1969).

Bank stabilization projects on the Willamette River and portions of most of the major tributaries were implemented to stabilize stream channels for navigation and flood control. Several types of bank stabilization techniques and materials have been tried since the first revetment on the Willamette River in 1888 (Thornber, 1965), although stone (rip-rap) has been the most extensively used revetment type. Well over 100 miles of stone revetments have been constructed in the Willamette Basin (Forbes and others, 1976), and 11 percent of the Willamette River shoreline is riprapped (Hughes and Gammon, 1987). Most construction of revetments within the basin has been conducted by the USACE since the 1930s as part of the Willamette River Basin Bank Protection Project (U.S. Army Corps of Engineers, 1975).

Stone revetments impact aquatic resources through changes in the physical environment of shoreline substrate, shoreline gradient, and water velocity (Hjort and others, 1984). The principal change in shoreline substrate is the reduction in riparian vegetation and large woody debris (Hjort and others, 1984; Bottom and others, 1985). Revetment construction also results in the loss of secondary side channels, backwater areas, and oxbows, which are important habitat for juvenile anadromous salmonids and the Oregon chub, a Federally endangered species (Li and others, 1987). Specific impacts from revetments have been the focus of several studies at PSU (Forbes and others, 1976), including investigations on birds (Perry, 1978) and mammals (Willis, 1981).

Fish assemblages at stone revetments on the Willamette River below Salem were characterized by lower species richness and diversity than at natural banks, but higher densities of smaller fish (Hjort and others, 1984). Five species positively associated with

revetments were prickly sculpin, redside shiner, northern squawfish, largescale sucker, and chiselmouth (Hjort and others, 1984; Li and others, 1984). Fish species associated positively with revetments are likely attracted by the high densities of invertebrate prey living in the interstices (Li and others, 1984). Higher densities of macroinvertebrates were found at revetments than at natural banks, particularly species adapted to exploit interstitial spaces between rocks as habitat or to cling to rock surfaces in fast water (Li and others, 1984). The stability of the bank and moderate water currents also likely reduce susceptibility of displacement and thus benefit macroinvertebrates at the revetments.

# **Dams and Impoundments**

Although dams have been constructed in the basin since the mid 1800s, extensive Federal government flood-control efforts began in the 1930s, particularly on the main stem Willamette River (Sedell and Froggatt, 1984). Most of the dams were constructed by the USACE between 1941 and 1968 (U.S. Army Corps of Engineers, 1982). Twenty-five major dams currently operate in the Willamette Basin (Oregon Water Resources Department, 1992). Eleven are single purpose hydroelectric projects operated by public and private utilities, one is a multipurpose project managed by the Tualatin Valley Irrigation District for the BLM, and 13 are multipurpose reservoirs operated by the USACE. The year completed, storage capacity, and subbasin for the 13 USACE dams/impoundments are presented in table 1; locations are given in figure 3.

The principal reason for construction of USACE dams was flood control (U.S. Army Corps of Engineers, 1991a). Dams and reservoirs are also operated for power generation, recreation, irrigation, public water supply, navigation, pollution abatement, and anadromous fish propagation. Summer water flows in the Willamette River are controlled primarily by releases from impoundments on the major tributaries. This results in higher summer flows and lower water temperatures than those occurring prior to construction of dams (Hines and others, 1977). When combined with passage improvements at Willamette Falls and hatchery inputs, this improved water quality has resulted in the establishment of anadromous runs of summer steelhead trout, coho salmon, fall chinook

salmon, and sockeye salmon above Willamette Falls, which historically did not support these runs.

The construction of dams has affected aquatic resources, particularly fish, upstream and downstream of the dam in both beneficial and harmful ways. Favorable effects include control of floods, which has reduced siltation, and augmentation of historic low flows in the summer with cooler water (Willamette Basin Task Force, 1969; U.S. Army Corps of Engineers, 1982). Additionally, impoundments have increased recreational fishing opportunities by providing lake or reservoir habitat for some species of fish that otherwise would not be present in these areas.

The principal negative impact of dams is the inundation of spawning areas and physical blockage of migration to upstream spawning areas (Li and others, 1987). Approximately 400 miles of previously important spawning and rearing habitat for salmon are no longer accessible (Foster, 1991). Other negative impacts include increased water temperature fluctuations and extremes, reduction of production and rearing habitat for some species of fish, alteration of natural hydrologic functions of seasonal flooding and recruitment of spawning gravel, and mortality in turbines at the dams (Buchanan and Wade, 1982; Bottom and others, 1985). Dams have also created conditions that may exacerbate disease problems, and the impoundments favor warm water introduced fishes that have proliferated, often at the expense of native fish (Li and others, 1987). Buchanan and others (1981) suggests that northern squawfish may concentrate at dams to feed on migrating juvenile salmonids.

Dams and the impoundments associated with them basically change a riverine ecosystem into a lake ecosystem. In general, dams in the Willamette Basin have impounded fast-moving, cold water rivers that were favorable to cold water fish into slow moving, warm water lakes that are favorable to warm water fish. Impoundments are favorable habitats for pond or lake species, such as most Centrarchidae—sunfish, crappie, and bass; some Ictaluridae—catfish and bullheads; and some Percidae—perch. Small impoundments tend to mimic natural riverine pool habitat; thus, they may act to increase the extent of pool-type habitat and increase the abundance and distribution of species associated with this habitat.

Numerous investigations have been conducted to evaluate the effects of dams (proposed or operating) in the Willamette Basin on aquatic habitat and biota, particularly fish. A description of these studies is

beyond the scope of this section, but a list of reports addressing the effects of the major dams and impoundments is presented in appendix D. In addition to the major dams and impoundments, numerous small dams and impoundments are present on tributaries of the major rivers within the basin. However, only a few investigators have examined their effect on fish resources. Korn and others (1967) reported on the effect of a small dam (North Fork) on the Clackamas River on the behavior of juvenile anadromous salmonids. They reported that movement of juvenile salmonids within the impoundment was correlated with floods and high flows in the winter and water temperatures in the spring. Li and others (1983) assessed the impact of small dams on the distribution of resident fishes in the Calapooia River. They suggest that dams located on the upstream reaches of the river may have less negative impacts on fish distribution because there are fewer species of fish and fewer migrating fishes, especially salmonids.

Anadromous fish have been most seriously affected by passage problems at dams on tributaries of the Willamette River. Fish passage has been a major problem at Foster and Green Peter Dams on the South Santiam River; Leaburg Dam and the Leaburg and Walterville diversions on the McKenzie River; and Fall Creek Dam on Fall Creek, a tributary of the Middle Fork Willamette River (Howell, 1986; Oregon Department of Fish and Wildlife, 1988).

Artificial spawning channels and fishways have been constructed to mitigate the effects of blocked passage of anadromous salmonids to natural spawning grounds. Both methods have only been partially successful. Spawning channels were popular in the 1950s and 1960s, but most were not successful in adequately replacing production lost by blockage of passage to natural spawning grounds (Smith, 1993). Fishways are structures that allow passage of anadromous fish over natural and human-made obstructions to natural spawning areas and hatcheries. However, some migrating fish continue to have difficulties passing dams with fish passage facilities (Howell, 1986).

In addition to passage problems, installation of turbines for power generation has created a situation for potential injuries and mortalities. Sources of mortality include physical injuries and changes in water pressure from the bottom of the reservoirs to the receiving waters. Mortality of salmon and steelhead smolts has been identified as the most serious problem at Willamette Falls turbines (Oregon Department of Fish and

Wildlife, 1980). In response to this problem, several studies have been conducted to assess mortalities and other injuries to downstream salmonid migrants at turbine installations near Willamette Falls (Oregon State Game Commission, 1960; 1961; Willis and others, 1960; Lichatowich, 1981; Williams, 1981). During 14 days of observation in the spring of 1981, 23 percent of the 5,202 steelhead trout examined were found to have sustained some bypass injury (Lichatowich, 1981). This is similar to the results of Williams (1981), who reported that 24 percent of steelhead trout examined sustained an abnormal amount of injury.

Mortality investigations at other fish passage facilities in the basin include those at North Fork Dam on the Clackamas River (Gunsolus and Eicher, 1970), Fall Creek Dam on Big Fall Creek (Smith and Korn, 1970), Cougar Dam on the South Fork of the McKenzie River (Ingram and Korn, 1969), Foster and Green Peter Dams on the South Santiam River (Wagner and Ingram, 1973), and Leaburg Dam on the McKenzie River (EA Engineering, Science and Technology, Inc., 1990b).

Dams and impoundments in the Willamette Basin have also been documented to adversely affect fish behavior and reproductive capacity. The presence of dams can delay migration of adult salmon and steel-head trout, particularly during the high flows of spring, when chinook salmon are reluctant to use fishways or have difficulty finding the entrance (Howell, 1986). Homolka and Downey (1995) also reported on the delay of migration for spring chinook salmon of the upper McKenzie River, apparently due to water temperature modifications caused by dams. The alteration of river flows below dams from drawdowns may negatively affect salmonid spawning habitat by exposing redds (Herb, 1972). Additionally, drawdowns have been reported to strand some fish (Herb, 1972).

#### **Fish Hatcheries**

Most fish hatcheries were built to mitigate for the loss of natural production of salmon due to habitat loss and degradation from construction of dams. Hatcheries were also constructed to increase adult returns to other stream areas where natural runs once thrived (Bennett, 1985). The first hatchery in the Willamette Basin was constructed on the Clackamas River in 1877 (Willamette Basin Task Force, 1969), and by 1900 all of the hatcheries in the Willamette Basin were under

Federal control (Oregon Department of Fish and Wildlife, 1982c). The location of fish hatcheries and holding/rearing ponds in the basin is shown in figure 3, and information on species reared is included as part of table 8. The only one of the 13 hatcheries/ponds in the basin not operated by the ODFW is Eagle Creek Fish Hatchery on the Clackamas River (operated by USFWS).

Protection and enhancement of wild fish stocks is given the highest priority in management of fish populations (Oregon Administrative Rule 635-07-525) (Bottom and others, 1985). Hatchery stocks are released where necessary to provide optimum benefits from the resource. Hatchery production and releases are relied upon to compensate for the loss of wild production, to provide additional fish to the fisheries, and to sustain production of introduced stocks of coho salmon and summer steelhead (Howell, 1986). Successes of hatchery programs in meeting these objectives is tempered by their high cost; problems in maintaining genetic diversity; and potential adverse affects on wild stocks, including disease transmission, competition, and interbreeding. Most of the recent production in runs of salmon and steelhead trout in the Willamette Basin has been from hatchery stocks, except for native winter steelhead trout (Howell, 1986).

A series of Oregon Fish Commission reports in the 1960s described and evaluated the operation and production of several salmon hatcheries in the Willamette Basin. These include Marion Forks (Wallis, 1963; DeCew, 1969), McKenzie (Wallis, 1961a), South Santiam (Wallis, 1961b), Sandy (Wallis, 1962a), and Willamette (Wallis, 1962b).

A history of the stocking of hatchery fish within the basin is provided in subbasin fish management plans and anadromous fish production plans for each subbasin, along with others, such as Willis and others (1960), Oregon State Game Commission (1963), Hutchison and Aney (1964), Hutchison and others (1966a), Koski (1971), and Collins (1974). The focus of these programs has been biological, such as migration studies, stock hardiness, and restoration of natural runs; and recreational, such as increased angling opportunities and development of new fisheries for anglers. The methods and results of these efforts have been documented in numerous ODFW reports, such as those on spring chinook salmon in the Willamette River (Smith, 1977, 1979), spawning, hatching, and rearing success of transplanted coho salmon (Pearson

Table 8. Fish hatcheries and holding/rearing ponds in the Willamette Basin, Oregon

[Sources: Oregon Department of Fish and Wildlife (1994); K. Bourne (Oregon Department of Fish and Wildlife, oral commun., 1995); A.G. Demaris (Oregon Department of Fish and Wildlife, oral commun., 1995); A. Smith (Oregon Department of Fish and Wildlife, oral commun., 1995). Fish species: cc, channel catfish; chf, fall chinook; chs, spring chinook; co, coho salmon; ct, cutthroat trout; hb, hybrid bass; lb, largemouth bass; rt, rainbow trout; sb, smallmouth bass; sf, sunfish; sts, summer steelhead; stw. winter steelhead]

Hatchery	Stream Name	Location	Fish Species	Subbasin
Eagle Creek <sup>1</sup>	Eagle Creek	Estacada	co,stw	Clackamas
Sandy	Cedar Creek	Sandy	co	Sandy
Clackamas	Clackamas River	McIver State Park	chs,stw	Clackamas
St. Paul Ponds	Mission Creek	St. Paul	cc,hb,lb,sb,sf	Pudding
Stayton Ponds	North Santiam River	Stayton	chf	Santiam
Marion Forks	Marion Creek	Idanha	chs,ct,stw	Santiam
Roaring River	Roaring River	Scio	rt,sts	Santiam
South Santiam	South Santiam River	Sweet Home	chs,sts	Santiam
McKenzie	McKenzie River	Leaburg	chs,sts	McKenzie
Leaburg	McKenzie River	Leaburg	ct,rt,sts	McKenzie
Dexter Pond	Middle Fork Willamette River	Lowell	chs,sts	Middle Fork
Willamette	Salmon Creek	Oakridge	chs,stw,rt	Middle Fork

<sup>&</sup>lt;sup>1</sup> Federal hatchery; all others are State hatcheries.

and others, 1967), releases of coho salmon in the Willamette River above Willamette Falls from 1952–1982 (Williams, 1983), and rainbow trout (Moring, 1976). The magnitude of hatchery releases in the Willamette Basin is exemplified by the numbers for 1988, which included 5.1 million fall (subyearling) chinook salmon, 1.1 million coho salmon, 700,000 summer steelhead trout, and 400,000 winter steelhead trout (Knutsen and Ward, 1991).

A cooperative program between the NMFS and ODFW was initiated in 1971 to develop natural runs of fall chinook and coho salmon, and winter and summer steelhead trout above Willamette Falls, historically a barrier to these fish prior to development of a fishway. The rationale and methods of the program are described in Sams (1973). Results of the program have been reported annually (Hansen, 1977; Hansen and Williams, 1979; Buchanan and Wade, 1982; Wade and Buchanan, 1983).

Hatchery fish may lower the fitness of native populations through interbreeding, competition, and social stress, and through population reductions due to increased angler effort and catch associated with stock-

ing programs (Nicholas and others, 1978). Hatchery salmonids may also usurp the territories of resident individuals and force them into less suitable habitat (Stein and others, 1972). Nicholas and others (1978) review the consequences of interbreeding and discuss four approaches to minimize the negative affects of interbreeding.

Hatchery fish are genetically and behaviorally distinct from their native progenitors (Nicholas and others, 1978; Li and others, 1987). Survival in the wild is lower for hatchery fish than wild fish (Reisenbichler and McIntyre, 1977; Chilcote and others, 1984; Nickelson and others, 1986). Moring (1982) evaluated three hatchery strains of rainbow trout used in stocking programs in the basin, and concluded that the Cape Cod variety yielded significantly better results in terms of higher returns to the angler at a lower cost to the State.

Another potential impact of fish hatcheries is the effect of hatchery effluent on water quality and biota in receiving waters. Discharged water from hatcheries may increase water temperature, pH, chemical oxygen demand, and concentrations of nutrients, ammonia, and suspended solids (Kendra, 1991). Additionally,

chemicals used to treat diseases and parasites which are discharged into receiving water may be harmful to aquatic biota. Kendra (1991) also reported that macroinvertebrate communities may respond to the organically enriched receiving waters by replacing sensitive taxa with more tolerant forms.

## **Pollution**

Aquatic resources in the Willamette Basin are affected by point-source pollution (resulting from a discharge at a specific location) and nonpoint-source pollution (resulting from diffuse runoff associated with land use activities). Historical pollution problems in the Willamette River were primarily due to pointsource pollution from municipal and industrial point source discharges (Merryfield and Wilmot, 1945; Gleeson, 1972). Current point sources include industrial, municipal, domestic, and agricultural discharge types. Tetra Tech, Inc., (1992d) lists 320 minor and 33 major National Pollutant Discharge Elimination System permittees that discharge pollutants from point sources into waters of the Willamette Basin. Approximately one-third of the minor permittees and two-thirds of the major permittees discharge into the main stem Willamette River. Thus, most concern regarding the effects of point-source pollution on aquatic biota is within the valley floor, including the Willamette River and the lower reaches of its tributaries.

Most current pollution problems in the Willamette Basin are from nonpoint sources (Tetra Tech, Inc., 1995a). These sources include areas having a variety of land-use activities, such as urban development, forest practices, and agriculture. On the basis of results of the nonpoint-source model developed for the Willamette River as part of the WRBWQS, agricultural land is considered to be the largest source of nonpoint-source pollution (Tetra Tech, Inc., 1995a). Tetra Tech, Inc., reported that most of the nonpoint-source pollution to the Willamette River is from the Pudding, Tualatin, Yamhill, and Long Tom subbasins.

The Willamette River has changed during the past 40 years from a river characterized as a conveyor of industrial and municipal sewage to a recreational and environmental asset. During the early to mid-1900s, the Willamette River, particularly near Portland, was "...in about as unsatisfactory a condition as a river could be..." (Gleeson, 1972). Studies documenting pollution levels in the Willamette River were made as

early as 1927 (U.S. Army Corps of Engineers, 1982), and numerous reports during this period documented the water quality of the river, including Langton and Rodgers (1929), Rodgers and others (1930), Gleeson (1936), Gleeson and Merryfield (1936), Craig and Townsend (1946), Fish and Rucker (1948), McKernan and Mattson (1950), Willis and others (1960), and Oregon State Game Commission (1963).

The most extensive documentation of the degree of pollution, particularly as it related to oxygen depletion and fish resources, was based on the work of Merryfield and Wilmot (1944) and Dimick and Merryfield (1945). They reported that the river contained high loadings of organic wastes, dense beds of algae, and floating and benthic sludge, which produced critically low dissolved oxygen concentrations that limited salmon migration. In some instances, the pollutant levels were lethal to local fish populations, including those of trout and salmon (Dimick and Merryfield, 1945; U.S. Army Corps of Engineers, 1982). Principal sources of pollution being discharged directly into the river were untreated sewage from municipalities and residences, and industrial wastes from canneries and paper product mills (Oregon State Game Commission, 1963; Hutchison and Aney, 1964). The discharge of sulphite pulp liquor from paper product mills was considered to be the most serious source of pollution affecting fishery resources because of its toxic effects (McKernan and Mattson, 1950; U.S. Army Corps of Engineers, 1982). In addition to the Willamette River, the lower portions of several tributaries, including Rickreall Creek and the Calapooia, Pudding, Tualatin, Yamhill, North and South Santiam, and Long Tom Rivers, also had high levels of pollution (Dimick and Merryfield, 1945; U.S. Army Corps of Engineers, 1982).

Gleeson (1972) provides an extensive discussion of the history of efforts to improve the water quality in the Willamette River, and their successes, from the late 1920s through the 1960s. Since the 1950s, water-quality improvement throughout the river has been documented, particularly in Portland Harbor. This improvement resulted from extensive efforts in sewage treatment, chemical recovery processes by industries, and increased low-flow augmentation (Gleeson, 1972; Huff and Klingeman; 1976; Hines and others, 1977). In 1972, the Willamette River became the largest river in the United States to have all known wastewater point sources under secondary treatment (Rinella and others, 1981).

The historic pollution load from domestic and industrial wastes discharged into the Willamette River was the most important factor contributing to the depletion of former great runs of anadromous fish (Fish and Rucker, 1948; Parkhurst and others, 1950; Oregon State Game Commission, 1963). In addition to the inherent toxic effects, the biochemical oxygen demand resulted in the lower reaches of the river being nearly devoid of oxygen. The dissolved oxygen requirements for salmonids in the lower Willamette River have been discussed by Sams and Conover (1969) and Alabaster (1988).

Several studies in the 1940–50s were conducted to ascertain the pollution status of the Willamette River and major tributaries by means of biological indicators. Noble (1952) assessed the sensitivity of fish to polluted habitat and reported that trout, salmon, whitefish, and sculpin were least tolerant of polluted conditions. Deschamps (1952) used the presence of benthic macroinvertebrates, along with certain physical and chemical conditions, as indicators of pollution at sites on the Willamette, McKenzie, South Fork Santiam, and Clackamas Rivers. Stoneflies, mayflies, and caddisflies were identified as least tolerant of pollution (Deschamps, 1952). Ziebell (1954) focused on invertebrate and fish communities at two sites on the South Fork Santiam River, and included the Order Odonata to the above list of least tolerant macroinvertebrates.

Some recent examinations of point-source pollution have been directed at specific effluent locations. Species richness and diversity of macroinvertebrates were similar, but the total number of individuals was significantly lower below the discharge of biologically treated effluent at a pulp and paper mill on the Willamette River near Halsey (HMS Environmental, Inc., and Miller, 1988). Species composition of periphyton was the same, but abundance and depth distribution were different below the effluent discharge. Hughes and Gammon (1987) found that point sources of pollution affected fish assemblages less than the gradual changes in water quality from the headwaters to the mouth of the river.

### **Land Use**

Diverse land uses, particularly agriculture, forest management, and urbanization, have substantially affected aquatic resources in the Willamette Basin. These land uses impact stream habitat quality

by reducing instream and riparian vegetation diversity and complexity, bank and channel structure and stability, the quality and quantity of spawning gravel, stream discharge and quality, and by exaggerating the natural processes of erosion and sedimentation (Bottom and others, 1985).

## **Agriculture**

As more land in the basin has been brought into intensive cultivation, there has been increased demand for irrigation water. Water withdrawals are principally for agricultural purposes, but in some urban/residential areas water is withdrawn for industrial and municipal needs. The withdrawal and diversion of large volumes of surface water for irrigation has resulted in changes in flow characteristics of streams, including complete elimination of flow during the summer in some streams. Irrigation accounts for more than 90 percent of the agricultural water use in the basin (Oregon Water Resources Department, 1992). In 1987, there were 285,000 irrigated acres within the Willamette Basin compared with 27,000 acres in 1940 (Oregon Water Resources Department, 1992). These water demands have contributed to reduced flows in many streams and caused erratic water levels; conditions unlike those under which native aquatic biota evolved. Additionally, naturally occurring low flows of summer are often exacerbated by withdrawal of water for irrigation.

Chemical contamination of aquatic ecosystems in the basin has resulted from runoff and leaching of chemicals resulting from farming practices, such as the use of pesticides, herbicides, and fertilizers (Anderson and others, 1996). These can cause impairments in general water quality and may be toxic to some aquatic biota. Principal types of farming in the basin that use chemicals are crop production (grass seed, nuts, fruits), nurseries (ornamental shrubs and trees), and animal production (dairy and beef cattle, poultry).

# **Forest Management**

Historic and ongoing logging and associated road construction throughout the Willamette Basin has had a substantial impact on aquatic and riparian habitat. Logging practices can change the basic community ecology of a stream by direct and indirect effects on the physical environment, which indirectly results in changes in the aquatic biota.

Extensive timber harvest in the Willamette Basin began in the late-19th century. The industry was essen-

tially unregulated, and this resulted in (1) harvesting in the riparian zone, which adversely affected water quality and salmonid fish habitat and (2) the accumulation of large amounts of instream debris, which blocked anadromous fish migration to spawning areas (McKernan and others, 1950; Willamette Basin Task Force, 1969; Delarm and others, 1989). Additionally, logs were transported by streams in huge rafts from upstream harvests sites to downstream mills, which resulted in blockage of streams, scoured streambeds, and ruination of spawning areas (U.S. Army Corps of Engineers, 1982).

Timber harvesting throughout watersheds and destruction of riparian cover along streams can have multiple effects, including rapid runoff and siltation due to erosion; fluctuations in stream flows, temperature, and dissolved oxygen content; loss of spawning habitat from scouring of gravel; changes in pool/riffle ratios; reduction of organic matter input from loss of trees adjacent to the stream; stimulation of primary production of algae, moss, or macrophytes as a result of increased nutrients and solar radiation reaching the stream; and destruction of food organisms. The effects of these impacts within a specific location is dependent upon numerous physical factors, including watershed geomorphology, climate, stream size and gradient, and the biotic composition of the stream (Murphy and Hall, 1981).

Much of the research in the Willamette Basin on the effects of forest management practices on aquatic ecosystems has focused on the western slope of the Cascade Mountains, particularly the HJAEF. Removal of forest canopy within the riparian corridor of small, cold water, high-gradient streams in the Cascade Mountains tends to increase stream productivity, but the increased sedimentation tends to degrade physical habitat (Murphy and others, 1981).

The increase in solar radiation reaching the stream after forest canopy removal apparently increases periphyton production and aquatic production at all trophic levels (Gregory, 1980; Hawkins and others, 1982; Hawkins, and others, 1983; Murphy and Hall, 1981; Murphy and others, 1981). Cutthroat trout populations increased in a stream flowing through a recent clear-cut compared to another section of the stream in undisturbed old-growth forest (Aho, 1976; Murphy and others, 1981; Hawkins and others, 1983; Wilzbach, 1984). Insect emergence (Grafius, 1977), annual primary production (Gregory, 1980; Murphy and Hall, 1981), and density of invertebrates (Murphy

and others, 1981; Hawkins and others, 1982) were also greater in clear-cut or open sections of streams. Wilzbach (1984) reported that, in spite of reduced cover, cutthroat trout had greater foraging success and growth rates in logged sections of the stream, where invertebrate drift in these unstable habitats provided a more reliable food source. However, Wilzbach and others (1986) cautions that any beneficial advantages of trout foraging efficiency in logged stream sections must be weighed against increased risk of mortality in these same stream sections from predation and physical disturbances due to reduction of shelter.

Conversely, other authors report on the degradation of physical habitat of streams after nearby timber harvesting, particularly through increased sedimentation from canopy removal during logging (Gibbons and Salo, 1973). In streams of the HJAEF, Wustenberg (1954) and Wyatt (1959) reported reduction or elimination of populations of cutthroat trout in some smaller tributaries and declines in aquatic insect populations for at least 1 year immediately following logging adjacent to the stream. In Minto Creek of the Santiam subbasin, Frissell and others (1985) reported a 40-percent reduction in trout density in a clear-cut segment of the stream compared to an unlogged segment. They also noted that the smallest size class of fish was absent, and large individuals were uncommon in the clear-cut section of the stream. They attributed the results to differences in the diversity of habitat types, particularly pool/ riffle habitat within the forested and clear-cut sections of the stream. The differences between the results of these studies and of those mentioned in the previous paragraph indicate that the response of fish populations to logging of riparian cover may be dependent upon several site-specific geomorphic features.

## Urbanization

Urbanization has affected water quality and aquatic biota particularly through domestic water use and discharge, and streamside development. Runoff and discharge of trace elements, bacteria, nutrients, and suspended solids are high in urban areas of the basin. The most extensive urbanization has occurred along the Willamette River (particularly metropolitan Portland), but urbanization has also occurred along most of the larger tributaries of the main stem Willamette, particularly the Tualatin, Clackamas, and McKenzie. In the lower Willamette River at Portland Harbor, natural shoreline and nearshore habitat have been substantially altered by the construction of wharfs, piers, boat

repair facilities, and the presence of rip-rapped shoreline. Rip-rap and pilings serve as current deflectors and create habitat that may affect the abundance and distribution of some sport fish, but also potentially increase predation on these fish by northern squawfish, who prefer areas of low velocity (Ward and Nigro, 1991).

The effects of urban development on juvenile salmonids in the lower Willamette River at Portland Harbor, as determined during a 4-year cooperative effort between the ODFW and the Port of Portland, were reported by Ward and others (1994). They identified few risks to juvenile salmonids from development in the harbor and did not detect significant changes in behavior at waterway developments. However, Farr and Ward (1993) suggested that development along the lower Willamette River may be adversely affecting populations of white sturgeon, a game fish.

The effects of urbanization on fish populations in the lower Tualatin subbasin were reported by Friesen and Ward (1996). Sites within the urban growth boundary near Portland were characterized as moderately unhealthy based on poor habitat quality and a large number of fish affected by parasites or physical anomalies. They also reported that species intolerant to pollution and relatively warm water temperatures occurred primarily at forested sections of streams that were unaffected by urban or agricultural influences.

# **Introduced Species**

The intentional or accidental introduction of species into aquatic systems can cause dramatic changes. Community ecology may be altered directly through predation and disease, and indirectly through increases in competitive interactions (Li and Moyle, 1981). In many cases, the biological consequences of these introductions are not known and cannot be accurately predicted, but interspecific competition with native species and introductions of diseases to aquatic biota are likely (Moyle, 1986).

In aquatic systems, most intentional fish enhancements have been implemented to improve sportfishing. This technique includes stocking of introduced species and the stocking of hatchery-reared indigenous fish species to reestablish or enhance populations. Much of the stocking in the Willamette Basin has occurred in major rivers and impoundments. Stocking of sport species, such as sunfish, crappie, and bass

has occurred throughout ponds and smaller impoundments in the basin.

The effect of introduced species on native biota is particularly pronounced in lowland Willamette Valley aquatic ecosystems, where warmer water temperatures are conducive to species introduced from southern ecosystems. Because of stocking of introduced species and dam-related habitat changes that favor warm water fishes, the overall pattern in the Pacific Northwest is that fish fauna assemblages often resemble those found in the Midwest (Li and others, 1987).

Introduced fish tend to dominate in highly disturbed habitats. In the Tualatin subbasin, introduced fish tend to be most numerous in low gradient, highly degraded (eroding banks) reaches, and in tributaries near urban and industrial areas where large ponds or marshes are present (Friesen and others, 1994).

The observance of declines of native fish following introduction of nonnative fish is widely reported. In the Willamette Basin, nonnative piscivorous fish, such as largemouth bass and bluegill (*Lepomis macrochirus*), have been implicated in the decline of the Oregon chub (Markle and others, 1989; Pearsons, 1989). Large black crappie and white crappie prey on small juvenile salmonids (Grenfell, 1962; Ward and others, 1991), although the predation level is unknown and probably low (Ward and others, 1991).

Other native aquatic biota in the basin, particularly amphibians, are also subject to high rates of predation because they did not evolve in the presence of the voracious predation of some introduced species, such as pumpkinseed (*Lepomis gibbosus*), largemouth bass, bluegill, and bullfrogs. For example, the bullfrog has been implicated in declines of the western pond turtle (Marshall and others, 1996) and spotted frog (Nussbaum and others, 1983). Additionally, diseases from introduced red-eared slider turtles have likely contributed to declines of the western pond turtle (Marshall and others, 1996).

Common carp can cause shallow waters of ponds, lakes, and marshes to become too turbid for good production of native plants important to waterfowl (Puchy and Marshall, 1993). Attempts to eliminate common carp and other unwanted fish with chemical treatments are expensive and often not successful (Johnson and others, 1985).

Stocking or escapement of introduced fish has also likely had negative effects on macroinvertebrate populations. However, the lack of historical or preintroduction data on macroinvertebrate populations often precludes documentation of these effects. Like other aquatic biota, many endemic macroinvertebrates are likely not equipped to deal with introduced fish predators. This may be particularly common in some high altitude, Cascade Mountain lakes, which were naturally without fish.

Little is known of the impact of the Asiatic clam on native mussel and clam populations, though some displacement of native fauna is probable. In some tributaries to the Willamette River, it can be found in densities greater than 600 individuals per square meter (Ian Waite, U.S. Geological Survey, unpubl. data, 1996), whereas native mussels and clams are rare at the same sites.

Eurasian watermilfoil impacts aquatic resources by reducing the diversity of fish habitat and interfering with the healthy development of fish populations (Geiger and others, 1983). It is also considered a recreational nuisance because it grows in dense masses and provides an obstacle to boaters. Only chemical control has been effective in controlling the occurrence and spread of this species.

# AQUATIC TOXICOLOGICAL INVESTIGATIONS

Chemical analysis of tissues can provide information on the occurrence and extent of contaminants in aquatic ecosystems because contaminants may be more concentrated in tissue than in surrounding water or sediment. Various studies addressing contaminant impacts on aquatic biota have been conducted in the Willamette Basin (table 9). Two comprehensive investigations include the WRBWQS (Tetra Tech, Inc., 1995d) and the WRTS (ODEQ, 1994b). The WRBWQS was initiated to develop the necessary technical and regulatory understanding and information base required to protect and enhance the water quality of the Willamette Basin. The study design includes the development of predictive water quality models and an assessment of various biological indices as measures of water quality. Modeling efforts focused on several water quality parameters, including dissolved oxygen, nutrients, chlorophyll, bacteria, toxic chemicals, and suspended sediment, and have addressed both point- and nonpoint-source categories. Biological indices were used to measure various ecological attributes of benthic invertebrates and fish assemblages as an assessment or bioindicator of water quality (Tetra Tech, Inc., 1995d).

The WRTS, conducted by the ODEQ in cooperation with OSU and the USEPA, recently completed a screening survey to investigate the presence and effect of toxic pollutants in the Willamette River and selected tributaries. The stated objectives of the study were to determine if bioaccumulative toxic pollutants were present in the sediments and fish tissue and to determine the possible effects of the pollutants present on the aquatic biota using bioassays and other aquatic-life toxicity testing methods (Oregon Department of Environmental Quality, 1994b). Information on contaminants in sediment and fish was gathered for the period 1988–91. Many of the sampling sites were used in previous toxics monitoring. They represented ambient (background) levels, effects of important industrial and municipal contaminant sources, and typical urban nonpoint source impacts.

The Willamette Basin has also been included in several national contaminant studies. The National Contaminant Biomonitoring Program (NCBP) was initiated in 1967 as part of the National Pesticide Monitoring Program under the auspices of the USFWS. The NCBP was established to document temporal and geographic trends in concentrations of persistent toxic chemicals that may threaten fish and wildlife resources. Since its inception, the program has expanded from an initial focus on organochlorine insecticides to include industrial chemicals, herbicides, and potentially toxic trace elements that accumulate in fish (Schmitt, 1990). A nationwide network of stations was established, one of which was located on the main stem Willamette River at Oregon City.

The USEPA also initiated a one-time screening investigation in 1986 to determine the prevalence of selected bioaccumulative pollutants in fish and to identify correlations with sources of these pollutants (U.S. Environmental Protection Agency, 1992a). This study, known as the National Study of Chemical Residues in Fish, was an outgrowth of the USEPA's National Dioxin Study, which detected elevated concentrations of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) in fish from major watersheds in the United States, including the Willamette Basin. Dioxins, furans, polychlorinated biphenyls (PCBs), chlorinated pesticides, and mercury were analyzed in fish from selected sites thought to be influenced by a variety of point and nonpoint sources. Sampling sites were established at four locations along the main stem Willamette River (Portland, Halsey, Newberg Pool, and Wilsonville) and two locations on the Tualatin River (Cherry Grove and Cook Park).

**Table 9.** Aquatic toxicology investigations in the Willamette Basin, Oregon [Subbasins (see fig. 5): CF, Coast Fork Willamette; LT, Long Tom; LU, Luckiamute; MC, McKenzie; MF, Middle Fork Willamette; SA, Santiam; TU, Tualatin; WR, Willamette River; --, no data; PAH, polyaromatic hydrocarbons; PCB, polychlorinated biphenyls; TCDD, tetrachlorodibenzo-p-dioxin; TCDF, tetrachlorodibenzofuran]

					Matrix studied					
Reference	Sub- basin	Temporal coverage	Spatial coverage	Торіс	Fish	Fauna	Water	Sedi- ment		
Allen and Curtis (1991)	CF	1989–90	Cottage Grove Lake	Environmental parameters affecting mercury dynamics and bioaccumulation in fish	X		X	X		
Curtis and others (1993)	WR	July, October 1990	Willamette River (RM 7–195)	Cytochrome P450–1A1 induction in fish as a biomarker for TCDD and TCDF; organochlorines, PAHs	X			X		
Curtis and Siddens (1995)	WR	Spring 1994	Newberg Pool (RM 56)	Teratogenic qualities of whole effluent	X		X			
Hart Crowser (1988)	WR	1980's	Portland	Remedial action plan for sediments; PCBs in crayfish and sculpin	X			X		
Henderson and others (1969)	WR	1967–68	Willamette River	Organochlorine insecticide residues in fish	X					
Henny and Bethers (1971)	WR	Spring 1970	Albany	Great blue heron nesting study		X				
Kuehl and others (1989)	MC,SA, WR	1983	Coburg, Jefferson, Portland, Salem,	2,3,7,8-TCDD contamination in fish	X					
Lowe and others (1985)	WR	1978–81	Oregon City	National study on trace element residues in fish	X					
Markle (1994b)	WR	1993	Newberg Pool, Corvallis	Fish hybridization and skeletal deformities	X					
May and McKinney (1981)	WR	1976–77	Oregon City	National study on trace element residues in fish	X					
Oregon Department of Environmental Quality (1994b)	WR	1988–91	Willamette River	Willamette River toxics study	X			X		
Pastorok and others (1994)	WR	1990–91	Portland, Wilsonville	Ecological risk assessment for river sediments contaminated by creosote	X			X		
Schmitt and others (1981)	WR	1970–74	Oregon City	National study on organochlorine residues in fish	X					

Table 9. Aquatic toxicology investigations in the Willamette Basin, Oregon—Continued

						Matrix	studied	
Reference	Sub- basin	Temporal coverage	Spatial coverage	Торіс	Fish	Fauna	Water	Sedi- ment
Schmitt and others (1983)	WR	1976–79	Oregon City	National study on organochlorine residues in fish	X			
Schmitt and others (1985)	WR	1980–81	Oregon City	National study on organochlorine residues in fish	X			
Schmitt and Brumbaugh (1990)	WR	1976–84	Oregon City	National study on trace element residues in fish	X			
Schmitt and others (1990)	WR	1976–84	Oregon City	National study on organochlorine residues in fish	X			
Tetra Tech, Inc. (1993b)	SA, WR	1992	Willamette River, Santiam River, Conser Slough	Fish health assessments and skeletal deformities	X			
Tetra Tech, Inc. (1995d)	LU, WR	1992–94	Willamette River, Luckiamute River	Skeletal deformities	X			
U.S. Army Corps of Engineers (1991b)	CF, MF, LT	1983	Cottage Grove, Dorena, and Fern Ridge Lakes, Dexter Reservoir	Potential contaminants in reservoirs	X		X	X
U.S. Environmental Protection Agency (1992a)	WR	1986	Willamette River, Tualatin River	National study of chemical residues in fish	X			
U.S. Fish and Wildlife Service (1994c)	TU	1994	Rock Creek near Sherwood	Chemical residues in fish	X		X	X
Walsh and others (1977)	WR	1971–73	Oregon City	National study on trace element residues in fish	X			
Worcester (1979)	CF	1974–75	Cottage Grove Lake	Mercury accumulation in fish	X			

A variety of contaminant guidelines and criteria have been established for the protection of aquatic life. Many of the cited investigations in this report compare tissue concentrations to water-quality criteria established by the USEPA and the State of Oregon for the protection of aquatic life (U.S. Environmental Protection Agency, 1986 and 1992b; Oregon Department of Environmental Quality, 1991). Action levels established by the Food and Drug Administration (FDA) for the protection of human health have also been used for comparison. An action level specifies the level below which the FDA exercises its discretion not to take enforcement action.

# **Synthetic Organic Compounds**

Sources of synthetic organic compounds in aquatic systems include atmospheric deposition, industrial and municipal effluent, and nonpoint-source runoff. Synthetic organic compounds commonly adsorb on suspended particles, which settle on the stream bottom where they may be ingested by bottom-dwelling organisms. Many of these compounds are highly soluble in lipids, are persistent in the environment, and tend to bioaccumulate in biota. Bioaccumulation of chlorinated pesticides may result in eggshell thinning, reduced productivity, and the decline of certain populations of wildlife (Porter and Wiemeyer, 1969; Anderson and Hickey, 1972; Wiemeyer and others, 1984; Hoffman and others, 1995). The toxicity of synthetic organic compounds varies by species, sex, and age, and may be influenced by stress, chemical formulations used, and numerous other factors (Hoffman and others, 1995).

The majority of information on synthetic organic compounds in aquatic tissue in the Willamette Basin comes from the WRTS undertaken by the ODEQ (1994b) between 1988 and 1991. In this study, chemical residue analyses were performed on the tissue of collected species of fish and crayfish. The following constituents were analyzed: chlorinated pesticides, PCBs, dioxins and furans, polycyclic aromatic hydrocarbons (PAHs), and trace elements. Whole body, edible flesh, and liver of common carp, crayfish, cutthroat trout, largemouth bass, largescale sucker, mountain whitefish, and northern squawfish were collected for analysis. Species and tissue type collected varied among stations and years. All concentrations were assessed by species and river mile and compared to

Federal and State criteria for the protection of aquatic life and human health. Several other studies also assessed synthetic organic compounds in fish from the Willamette Basin (table 9).

#### **Chlorinated Pesticides**

Chlorinated pesticides in aquatic tissue are reported in appendices E-1, E-2, and E-3. The WRTS (Oregon Department of Environmental Quality, 1994b) detected 18 pesticides in 66 samples collected from the main stem Willamette River; 8 pesticides were detected in 30 samples collected from tributaries to the Willamette. Heptachlor, p,p'-dichlorodiphenyldichloroethane (DDD), and p,p'-dichlorodiphenyldichlorethylene (DDE) from the main stem, and p,p'-DDE and p,p'-dichlorodiphenyltrichloroethane (DDT) from the tributaries were detected in greater than 20 percent of the samples collected. Maximum concentrations for heptachlor, p,p'-DDD, and p,p'-DDE from the main stem exceeded the USEPA water-quality criteria for the protection of aquatic life. The dieldrin concentration in one main stem sample (RM 28, upstream from Oregon City) was above the FDA action level. These action levels are specific to edible parts of fish and shellfish but are not directly comparable to concentrations in whole fish (U.S. Environmental Protection Agency, 1992b).

Crayfish collected from Johnson Creek contained concentrations of p,p'-DDD, p,p'-DDE, and p,p'-DDT above USEPA water quality criteria for the protection of aquatic life (appendix E-2). However, these concentrations were determined to be safe for human consumption.

Curtis and others (1993) assessed pesticide residues in fish at six sites along the main stem Willamette River. These sites (RMs 7, 72, 131, 148, 160, and 195) were exposed to different types of pollution such as a hydroelectric dam, bleached kraft pulp mill discharge, and sewage outfalls. Various pesticides were found in common carp, cutthroat trout, and northern squawfish (appendices E-1, E-2, and E-3). Detected pesticides included aldrin, α-hexachlorocyclohexane (HCH), β-HCH, DDD, DDE, dieldrin, endosulfan I, endrin, heptachlor, and heptachlor epoxide. No patterns were found in contaminant distribution, except that higher DDE concentrations were found in whole northern squawfish at RMs 72, 131, and 148, as compared with the two more upstream sites. No correlations were seen between organochlorine concentrations

in aquatic tissue and occurrences of lesions in liver, kidneys, spleen, gills, or gonads.

Concentrations of organic compounds in fish collected from Rock Creek near Sherwood (Tualatin subbasin) were below detection limits, with the exception of p,p'-DDE (U.S. Fish and Wildlife Service, 1994c). Sculpin and three-spine stickleback (*Gasterosteus aculeatus*) had contained p,p'-DDE concentrations ranging from 0.03 to 0.06 micrograms per gram ( $\mu g/g$ ), wet weight. These concentrations are below the geometric mean (0.19  $\mu g/g$ , wet weight) of p,p'-DDE found in fish sampled by the NCBP (Schmitt and others, 1990). The USEPA (1992a) also assessed organic compounds in crayfish, northern squawfish, and suckers from the main stem Willamette River and the Tualatin River. Results are reported in appendices E-1, E-2, and E-3.

In 1970, Henny and Bethers (1971) studied great blue herons nesting along the Willamette River near Albany. Two eggs from a single nest exhibited p,p'-DDE levels of 3.3 and 4.5  $\mu$ g/g, wet weight. These egg concentrations could be expected to impact production. Wiemeyer and others (1984) found that mean 5-year production for bald eagles was near normal for breeding areas where eggs contained  $<3.0 \mu g/g$ , wet weight, p,p'-DDE; production dropped markedly for breeding areas where eggs contained  $>5.1 \mu g/g$ , wet weight, p,p'-DDE; and nearly complete breeding failure occurred where egg levels exceeded 15 µg/g, wet weight, p,p'-DDE. DDE concentrations of 5 µg/g, wet weight, were also associated with 10-percent shell thinning for bald eagles (Wiemeyer and others, 1984). A recently deceased day-old great blue heron chick with a p,p'-DDE whole body concentration of 10.1  $\mu$ g/g, wet weight, was also collected from the Albany heronry. Prey species of fish found in the nests at the heronry included cutthroat trout, largescale sucker, northern squawfish, and white crappie. Previous studies on insecticide residues in fish from the Willamette River detected concentrations of DDT and its metabolites ranging from 0.29 µg/g (white crappie) to 2.65 µg/g (largescale sucker), and dieldrin levels ranging from 0.01 µg/g (white crappie and largescale sucker) to 0.03 µg/g (largescale sucker and northern squawfish) (Henderson and others [1969] as cited in Henny and Bethers [1971]).

Table 10 compares great blue heron eggshell thickness measurements between a Willamette River site (Henny and Bethers, 1971) and sites from throughout the Pacific Northwest (Anderson and Hickey,

**Table 10**. Eggshell thickness data for great blue herons in the Pacific Northwest

[Adapted from Henny and Bethers (1971). Thickness index =  $\{\text{shell weight (mg)}\}\ / \{\text{shell length (mm) x shell width (mm)}\}\$ from Ratcliffe (1967)]

Sample size	Thickness index	Reference
130	$2.02 \pm 0.02$	Anderson and Hickey (1972)
9	$1.83 \pm 0.09$	Anderson and Hickey (1972)
2	$1.98 \pm 0.54$	Henny and Bethers (1971)
	130 9	size     index       130 $2.02 \pm 0.02$ 9 $1.83 \pm 0.09$

1972). No significant difference in the eggshell thickness index (as defined by Ratcliffe [1967]) was detected between the two studies, although Anderson and Hickey (1972) reported a 9-percent decrease in eggshell thickness in eggs collected between 1956 and 1959 in the Pacific Northwest. Henny and Bethers (1971) concluded that despite documented elevated egg pesticide levels, great blue heron numbers were remaining fairly stable in western Oregon.

## **Polychlorinated Biphenyls**

Several studies have reported PCB concentrations in fish from the Willamette Basin (Hart Crowser, 1988; Oregon Department of Environmental Quality, 1994b; U.S. Fish and Wildlife Service, 1994c). The ODEQ (1994b) collected tissue samples from aquatic biota from the main stem Willamette River and its tributaries for analysis of co-planar PCBs and arochlor PCBs. Detected PCBs are reported in appendix F. Maximum concentrations for both co-planar and arochlor PCBs were below FDA action levels but above the USEPA water-quality criteria for the protection of aquatic life. Curtis and others (1993) found PCB 1260 in northern squawfish at a majority of sampling sites on the main stem Willamette River. Few PCB congeners were detected in common carp or cutthroat trout (appendix F). The presence of PCBs in fish collected from RM 195 may suggest point-source contamination from the electrical components of a hydroelectric dam near the site. Total PCBs were not detected in fish samples collected from Rock Creek near Sherwood by the USFWS (1994c).

PCB sampling has also been conducted at the site of a former steam-powered electricity generating plant on the Willamette River in Portland. The plant was in operation from the early 1900s through 1975.

PCB contamination of adjacent river sediments was discovered in the 1980s. Data were gathered on water, sediment, ground water, upland soils, and fish. PCB concentrations were assessed in 32 crayfish and 36 prickly sculpins collected from the river in the immediate vicinity of the plant (approximately RM 13) and 1 mile upriver (Hart Crowser, 1988). Arochlor 1260 concentrations in crayfish tissue at three sites were all less than the detection level of 0.04 µg/g, wet weight. Arochlor 1260 concentrations in prickly sculpin tissue ranged from 0.19 to 0.63 µg/g, wet weight, at the site and from 0.10 to 0.35 µg/g, wet weight, at the upriver reference site (appendix F). Mean concentrations were not statistically different between contaminated and reference sites. Arochlor 1260 concentrations in prickly sculpin tissue samples exceeded the predator protection criterion for total PCBs (0.1 µg/g) instituted by the International Joint Commission (1988) of the Great Lakes. The predator protection criterion is determined for whole-body fish residue and should not be exceeded to protect birds and mammals that consume fish.

#### **Dioxins and Furans**

The ODEQ (1994b) and USEPA (1992a) analyzed tissues of aquatic biota from the main stem Willamette River and its tributaries for dioxin and furan concentrations (tables 11a and 11b). In the ODEO study, 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) was detected in all but one sample, and 2,3,7,8-tetrachlorodibenzofuran (TCDF) was detected in all samples. All detected concentrations of TCDD were above the USEPA threshold value of 0.07 picogram per gram (pg/g), wet weight, and below the FDA action level, of 25 pg/g, wet weight, as listed by ODEQ (1994b). Mountain whitefish collected upstream of RM 147, which is in the vicinity of a bleached kraft pulp mill, had median TCDD and TCDF concentrations of 0.53 and 2.55 (pg/g), wet weight, respectively. Mountain whitefish collected downstream of this location contained median TCDD and TCDF concentrations of 2.7 and 13.0 pg/g, wet weight, respectively. Maximum concentrations of TCDD and TCDF differed in mountain whitefish collected in 1990 (7.9 pg/g, wet weight, TCDD; 30 pg/g, wet weight, TCDF) and 1991 (1.9 pg/ g, wet weight, TCDD; 6.6 pg/g, wet weight, TCDF). Concentrations of TCDD and TCDF also differed among species.

Tissue samples from aquatic biota were also collected from the McKenzie, Santiam, and main stem Willamette Rivers as part of the USEPA's National Dioxin Strategy (Kuehl and others, 1989; tables 11a and 11b). TCDD was not detected at 1.0 pg/g, wet weight, in the McKenzie and Santiam Rivers; concentrations in tissue of organisms from the main stem Willamette River ranged from <1.0 pg/g to 1.8 pg/g, wet weight. This national study determined that higher levels of TCDD in fish could be associated with the presence of pulp and paper manufacturing plants as compared to other sites.

Curtis and others (1993) collected common carp, cutthroat trout, and northern squawfish at six sites between RMs 7 and 195 on the main stem Willamette River for analysis of TCDD and TCDF concentrations. They found that whole-body TCDD and TCDF concentrations of northern squawfish were generally higher than for cutthroat trout. Differences were attributed to variations in prey base, body composition, or organochlorine elimination rates. Strong correlations (r>0.80) were observed between common carp muscle TCDD or TCDF and biomarker responses (hepatic microsomal ethoxyresorufin O-deethylase[EROD] and total cytochrome P450-1A1) for individuals in which both types of analyses were performed. Contamination in fish and sediments was heavier at RM 7, which is located in Portland Harbor, than at upstream sites. TCDF to TCDD concentration ratios were also significantly higher (p=0.05) at this industrial area than at other sampling sites, highlighting the residual effects of past chemical production and usage near the site. Pastorok and others (1994) found the range of polychlorinated dibenzo-p-dioxin (PCDD) and polychlorinated dibenzofuran (PCDF) concentrations in crayfish and largescale sucker from RM 7 to be similar to that reported by the USEPA (1992a) for industrialized portions of the Willamette River.

# **Polycyclic Aromatic Hydrocarbons**

Little information is available on PAHs in fish from the Willamette Basin. Between 1988 and 1991, the ODEQ (1994b) collected common carp, crayfish, cutthroat trout, largescale suckers, and northern squawfish from the main stem Willamette River (RMs 7 through 161) and major tributaries for analyses of PAHs. Benzo(*b*)fluoranthene, acenaphthene, benzo(*k*)fluoranthene, and benzo(*a*)pyrene were detected in two common carp from RMs 7 and 74 on the main stem Willamette River. Concentrations of these chemicals ranged from 0.5 to 0.8 µg/g, wetweight. Curtis and others (1993) found no detectable

**Table 11a.** Dioxins and furans in tissue of aquatic biota from the Willamette Basin, Oregon [River mile: --, not available; --, approximately. Tissue type: wb, whole body; f, fillet; --, not available. Number samples: c, composite sample. Concentration range: pg,/g, picograms per gram;

ND, not detected, --, not analyzed; <, less than. References: ODEO, Oregon Department of Environmental Quality; USEPA, U.S. Environmental Protection Agency

								Cond	centration rai	nge (pg/g, wet	weight)			
River	River mile	Year	Species	Tissue type	Number samples	2,3,7,8- TCDD	1,2,3,7,8- PeCDD	1,2,3,4,7,8- HxCDD	1,2,3,6,7,8- HxCDD	1,2,3,7,8,9- HxCDD	1,2,3,4,6,7, 8-HpCDD	2,3,7,8- TCDF	1,2,3,7,8- PeCDF	Refer- ence
Willamette	~7	1983	Bottom feeder	wb	1 or c	1.8								Kuehl and others (1989)
Willamette	~7	1983	Bottom feeder	f	1 or c	1.5								Kuehl and others (1989)
Willamette	~7	1987	Crayfish	wb		2.61	3.75	<2.64	10.05	1.42	34.42	48.14	54.32	USEPA (1992a)
Willamette	~7	1987	Largemouth bass	f	4c	.74	<1.41	<2.46	.82	<1.38	.43	1.09	<.84	USEPA (1992a)
Willamette	~7	1987	Sucker	wb	4c	2.25	3.31	1.10	4.06	.61	16.57	3.35	.91	USEPA (1992a)
Willamette, Halsey		1987	Mountain whitefish	f	5c	4.58	1.56	.35	1.79	<1.35	2.47	16.12	<.82	USEPA (1992a)
Willamette, Halsey		1987	Sucker	wb	4c	.76	.27	<2.46	<1.84	<1.37	.54	2.43	<.77	USEPA (1992a)
Willamette, Wilsonville		1987	Crayfish	wb		<1.11	<.99	<2.45	<1.84	<1.37	3.33	1.77	<.77	USEPA (1992a)
Willamette, Salem		1983	Bottom feeder	wb	1 or c	<1.0								Kuehl and others (1989)
Willamette	141	1991	Mountain whitefish		1	1.4	.28	.1	.22	.14	.46	4.6	.2	ODEQ (1994b)
Willamette	141	1991	Common carp		1	.45	<.24	.13	.34	<.093	.84	.45	.045	ODEQ (1994b)
Willamette	143	1990–91	Mountain whitefish		2	1.9– 7.9	.45- <1.8	.14– <3.5	.37- <3.1	.17- <.61	.54- <1.6	6.6– 30	.17- <.52	ODEQ (1994b)
Willamette	143	1991	Common carp		1	0.57	<0.23	0.14	0.5	<0.11	1.2	0.56	< 0.048	ODEQ (1994b)

Table 11a. Dioxins and furans in tissue of aquatic biota from the Willamette Basin, Oregon—Continued

								Con	centration rai	nge (pg/g, wet	weight)			
River	River mile	Year	Species	Tissue type	Number samples	2,3,7,8- TCDD	1,2,3,7,8- PeCDD	1,2,3,4,7,8- HxCDD	1,2,3,6,7,8- HxCDD	1,2,3,7,8,9- HxCDD	1,2,3,4,6,7, 8-HpCDD	2,3,7,8- TCDF	1,2,3,7,8- PeCDF	Refer- ence
Willamette	145	1990–91	Mountain whitefish		2	2.5– 2.7	.52- <.7	.19– <.5	.43- <.63	.19– <.52	.62- <1.3	8.3– 13	.12- <.35	ODEQ (1994b)
Willamette	145	1991	Common carp		1	.44	.36	.29	1	<.12	2	.54	.12	ODEQ (1994b)
Willamette	147	1990	Mountain whitefish		2	2.8– 4.6	1–1.6	<.85	<1.7	<.82	<1.8	13–22	.29– <.74	ODEQ (1994b)
Willamette	161	1991	Common carp		1	.41	.63	.47	1.7	.3	5.3	.41	.2	ODEQ (1994b)
Willamette	161	1991	Mountain whitefish		1	.27	.31	.3	.08	.12	.4	1.7	.09	ODEQ (1994b)
Willamette	176– 177	1990	Mountain whitefish		1	.87	1.1	<.62	1.3	<.23	37	4	<.25	ODEQ (1994b)
Middle Fork Willamette	5	1990	Mountain whitefish		1	.57	.63	<.36	.68	<.32	.97	2.7	<.19	ODEQ (1994b)
McKenzie	3	1990	Mountain whitefish		2	<.5- .57	<2	<2.5	<2.2	<1	<6.8	2.1– 2.4	<.74– .75	ODEQ (1994b)
McKenzie, Coburg		1983	Bottom feeder	wb	1 or c	<1.0								Kuehl and others (1989)
Santiam, Jefferson		1983	Bottom feeder	wb	1 or c	<1.0								Kuehl and others (1989)
Santiam, Jefferson		1983	Bottom feeder	f	1 or c	<1.0								Kuehl and others (1989)
Tualatin, Cherry Grove		1987	Crayfish	wb	1	<.99	<.92	<2.47	<1.85	<1.38	.49	<.57	<.78	USEPA (1992a)

**Table 11b.** Dioxins and furans in tissue of aquatic biota from the Willamette Basin, Oregon [River mile: --, not available; --, approximately. Tissue type: wb, whole body; f, fillet; --, not available. Number samples: c, composite sample. Concentration range: pg/g, picograms per gram; ND, not detected: --, not analyzed: <. less than. References: ODEO, Oregon Department of Environmental Quality: USEPA, U.S. Environmental Protection Agency]

Concentration range (pg/g, wet weight) Tis-River 2,3,4,7,8-1.2.3.6.7.8-1,2,3,7,8,9-2,3,4,6,7,8-1,2,3,4,6,7, Refersue Number 1,2,3,4,7,8-1,2,3,4,7,8, River Year PeCDF HxCDF HxCDF HxCDF HxCDF 8-HnCDF 9-HpCDF TEC mile Species tvpe samples ence Willamette ~7 1987 Cravfish 19.02 18.85 10.15 0.23 0.87 6.44 1.76 26.11 USEPA wh --(1992a) Willamette ~7 1987 Largemouth f 4c .34 < 2.83 < 2.84 < 2.77 <1.96 .24 < 2.61 1.11 USEPA bass (1992a) Sucker Willamette ~7 1987 2.27 3.02 < 2.79 < 2.72 2.66 < 2.56 6.61 USEPA wb 4c 1.16 (1992a) Willamette. 1987 Mountain f 5c .45 .60 < 2.79 < 2.72 .36 .40 < 2.56 7.54 USEPA Halsey whitefish (1992a) Willamette. 1987 Sucker 4c <.85 .40 < 2.84 < 2.77 .25 <1.44 1.21 USEPA wb < 2.61 Halsev (1992a) Willamette. 1987 Cravfish wb <.84 < 2.82 < 2.83 < 2.76 < 1.95 <1.44 < 2.60 .21 **USEPA** --Wilsonville (1992a) 1991 .09 .2 Willamette 141 Mountain .14 .06 .11 .06 .13 2.18 ODEO whitefish (1994b) Willamette 141 1991 Common 1 .14 <.037 <.025 <.052 .13 .17 <.046 .78 **ODEO** carp (1994b)1990– Mountain 2 .22-.09-.12-3.04-Willamette 143 .12-.13-.23-.17-**ODEO** whitefish <1.7 <.72 <.73 <.8 <4.3 <.53 <1.1 14.07 (1994b)Willamette 143 1991 Common 1 .14 <.045 <.028 <.025 .16 .17 <.056 .93 **ODEQ** carp (1994b)145 1990- Mountain 2 .2-.15-3.85-Willamette .13-.15-.11-.22-.15-ODEO

<.42

<.33

<.25

<1.5

<.88

<.43

<.51

5.07

(1994b)

91

whitefish

Table 11b. Dioxins and furans in tissue of aquatic biota from the Willamette Basin, Oregon—Continued

								Concer	ntration range	(pg/g, wet we	eight)			
River	River mile	Year	Species	Tis- sue type	Number samples	2,3,4,7,8- PeCDF	1,2,3,4,7,8- HxCDF	1,2,3,6,7,8- HxCDF	1,2,3,7,8,9- HxCDF	2,3,4,6,7,8- HxCDF	1,2,3,4,6,7, 8-HpCDF	1,2,3,4,7,8, 9-HpCDF	Refer- TEC ence	
Willamette	145	1991	Common		1	0.25	0.14	0.092	< 0.057	0.19	0.31	<0.061	1.02	ODEQ (1994b)
Willamette	147	1990	Mountain whitefish		2	.56- <.94	<.24	<.21	<.86	.22- <.39	<.46	<.31	5.21– 8.65	ODEQ (1994b)
Willamette	161	1991	Common carp		1	.46	.22	.18	<.032	.23	.44	<.039	1.39	ODEQ (1994b)
Willamette	161	1991	Mountain whitefish		1	.13	.08	.07	.08	.21	.07	.1	.77	ODEQ (1994b)
Willamette	176– 177	1990	Mountain whitefish		1	.56	<.22	<.18	<1.9	.33	<1.3	<.46	3.35	ODEQ (1994b)
Middle Fork Willamette	5	1990	Mountain whitefish		1	<.24	<.33	<.25	<1.1	<.11	<.24	<.32	1.62	ODEQ (1994b)
McKenzie	3	1990	Mountain whitefish		2	.72- <1.3	<.48	<.44	<5	<2.4	<.46	<.74	3.49– 3.93	ODEQ (1994b)
Tualatin, Cherry Grove		1987	Crayfish	wb	1	<.85	<2.84	<2.85	<2.78	<1.96	<1.45	<2.62	ND	USEPA (1992a)

concentrations of 17 different PAHs in common carp muscle, whole cutthroat trout, or whole-body northern squawfish from 6 sites between RMs 7 and 195 on the main stem Willamette River at a detection limit of  $0.030~\mu g/g$ . Pastorok and others (1994) reported slightly elevated concentrations of PAHs in crayfish and largescale suckers collected near a creosoting company at RM 7 on the Willamette River compared with reference area concentrations. They suggested that ongoing exposure of the fish to PAHs in water and sediments near the site may be indicated because PAHs are normally metabolized quickly in fish.

#### **Trace Elements**

Trace elements can be lethal to aquatic organisms and fish over a wide range of concentrations. Anthropogenic sources in the Willamette Basin that may elevate trace elements above background levels in surface water include electroplating, smelting, and mining industries; industrial and municipal discharges and sewage; atmospheric deposition from combustion of fossil fuels and solid wastes; road surface runoff; and fertilizers, some pesticides, and erosion from agricultural areas. Natural mercury sources include deposits of cinnabar (mercuric sulfide) related to geothermal and volcanic activity.

Tissue samples from aquatic biota from both the main stem Willamette River and its tributaries have been analyzed for 18 major and trace elements by several investigators (appendices G-1 and G-2). Concentrations in tissue did not exceed aquatic-life criteria listed by ODEQ (1994b) for beryllium, chromium, nickel, or thallium, but arsenic and mercury concentrations sometimes exceeded the listed criteria.

The USEPA (1993) has developed fish-advisory screening values for cadmium (10  $\mu g/g$ ), mercury (0.6  $\mu g/g$ ), and selenium (50  $\mu g/g$ ). These values are meant to serve as an indication to government agencies that fish from the particular body of water may be potentially hazardous for human consumption. Mercury concentrations listed in appendices G-1 and G-2 sometimes exceeded the suggested fish-advisory screening value, but cadmium and selenium concentrations did not.

The USACE investigated the potential for the existence of contaminants in Willamette Basin reservoirs in 1983 (U.S. Army Corps of Engineers, 1991b). Dexter Reservoir, Cottage Grove, Dorena, and Fern Ridge Lakes were identified as having levels of certain water-quality constituents that impaired project pur-

poses, violated Federal or State water quality standards, or threatened humans, fish, or wildlife. Naturally occurring arsenic and mercury were identified as potential contaminants at all four reservoirs. Elevated mercury concentrations in sediment at Cottage Grove Lake have been attributed to large cinnabar deposits in the Calapooya Mountains and the Black Butte area. These levels have been exacerbated by past mercury mining activities in the area. Studies indicate that fish in Cottage Grove Lake have elevated mercury concentrations when compared with fish from tributaries (Buhler and others [1973] as cited in Worcester [1979]; Worcester, 1979; Allen and Curtis, 1991; U.S. Army Corps of Engineers, 1991b). Worcester (1979) examined mercury concentrations in several fish species from Cottage Grove Lake between 1974 and 1976. The highest mercury concentrations were found in largemouth bass muscle collected in 1974 (0.15 to 1.44 µg/g; appendix G-2). Due to elevated mercury levels in fish, the Oregon Department of Health has issued a health advisory for Cottage Grove Lake. Analyses of tissue from aquatic biota done in 1982 at Dexter Reservoir indicated that arsenic concentrations in northern squawfish and suckers were above USEPA water-quality criteria (U.S. Army Corps of Engineers, 1991b).

The ODEQ (1996) recently completed a report on mercury in Oregon lakes. Sediments and tissues of aquatic biota were assessed for mercury concentrations in areas with both known and unknown watershed sources of mercury. Areas assessed in the Willamette Basin included Cottage Grove Lake (cinnabar deposits and previous mercury mining), Dorena Lake (cinnabar deposits and gold mining), Fern Ridge Lake (no potential mercury sources identified), Henry Hagg Lake (no potential mercury sources identified), Willamette River Coast Fork (downstream of Cottage Grove Lake), and the Row River (downstream of Dorena Lake). Results from the study (appendix G-2) confirmed the relationship between elevated mercury concentrations in fish with cinnabar geology or historical mercury mining activity within the watershed. Higher tissue concentrations were also noted in older fish, and piscivorous fish tended to have higher body burdens of mercury.

In response to elevated mercury levels in several lakes and reservoirs throughout Oregon, a mercury working group was formed to address mercury concerns. Headed by the ODEQ, the group consists of representatives from ODFW, Oregon Department of Geology and Mineral Industries, Oregon Department of Human Resources - Health Division, OSU, USACE,

BLM, USFS, USFWS, and USGS. Various studies are underway to document mercury concentrations in Oregon lakes.

In 1992, a nonviable egg from a bald eagle nest near Cottage Grove Lake was analyzed for trace elements. The nesting pair had previously produced young in 1989, 1990, and 1991. Although the majority of trace element concentrations were not at levels of concern, the mercury residues in the egg (2.9 µg/g, dry weight; 0.765 μg/g, wet weight; U.S. Fish and Wildlife Service, Oregon State Office, unpubl. data, 1992) surpassed national averages for both unsuccessful (0.15 µg/g, fresh weight) and successful bald eagle nests (0.11 µg/g, fresh weight) (Wiemeyer and others, 1984), as well as Columbia River averages (0.20 µg/g, wet weight) (Garrett and others, 1988). The mercury concentration in the egg also approached levels associated with reproductive impairment in other avian species (Heinz, 1979; Eisler, 1987).

## **Organism Health**

#### **Fish Health Assessments**

Few studies have examined fish health in the Willamette River. The ODEQ (1994b) collected 10 to 20 northern squawfish from each of 5 sites in 1988 and from each of 4 sites in 1989. Sites were located on the main stem Willamette River, Santiam River, and Conser Slough. Tetra Tech, Inc., (1993b) also collected 12 to 20 northern squawfish or largescale suckers at each of 6 sites on the main stem Willamette River (RM 1, 6.5, 25, 49, 128, and 185) in 1992 for a fish health assessment. Both studies examined external features, internal features, and blood parameters following a fish health/condition assessment system that was originally developed for salmonid fishes (Goede, 1988; 1991). Table 12 presents a summary of some of the indices examined and the percent abnormalities observed. Due to the movement of fish throughout the river and the unknown applicability of the assessment to nonsalmonids, it is difficult to assess the relative health status of different river regions on the basis of these studies.

For both studies, northern squawfish had at least 25 percent abnormalities and suckers had at least 35 percent abnormalities for one or more indices at each of the sampling locations. The ODEQ study noted that fish from main stem sites were higher in percent abnormalities than those from two tributary sites (Oregon Department of Environmental Quality, 1994b). Of the

eight organs examined, the gills, pseudobranchs, and liver had the highest percentage of abnormalities. Tetra Tech, Inc. (1993b) found that suckers collected from the two farthest upstream sites (RM 128 and 185) were markedly less healthy than suckers collected at the downstream sites (RM 1, 6.5, 25, and 49).

Blood parameters were only reported for the Tetra Tech, Inc., (1993b) study. Hematocrit, leucocrit, and plasma protein levels were difficult to assess because comparison values are unavailable for the species collected and analyzed. However, the coefficient of variation for hematocrit was relatively high (above 27 percent) at several sites for both northern squawfish and suckers. Levels above 15 percent indicate that some fish in the population may be unhealthy (Goede and Barton, 1990).

Curtis and others (1993) conducted a microscopic examination of common carp, cutthroat trout, and northern squawfish liver, gills, kidneys, spleen, stomach, and gonads. Fish were collected in 1990 from 6 sites between RMs 7 and 195 on the main stem Willamette River. No evidence of neoplasia, necrosis, or advanced organ failure was found. Mild degenerative changes, parasitism, and inflammation were detected, but these conditions varied randomly in degree among species and sites. No correlation was found between organochlorine concentrations in fish and the occurrence of liver, kidney, spleen, gill, or gonad lesions. Curtis and others (1993) noted that this lack of correlation suggests that existing organochlorine burdens in adult fish were not overtly toxic at any site.

Pastorok and others (1994) examined 249 large-scale sucker livers collected from 4 sites near a creosoting company at RM 7, a downstream site (near RM 6), and an upstream site (near Wilsonville). The presence of mononuclear cell infiltrates, which indicate mild liver inflammation, was the most commonly observed abnormal condition (66 percent of the fish). However, this condition was not significantly different between RM 7 and the upstream location. No serious lesions were observed in any of the livers examined. Mild liver abnormalities that were noted were mononuclear cell infiltration, focal necrosis in hepatocytes, serosal inflammation, parasite-associated inflammation, non-uniform vacuolation of hepatocytes, and fat infiltration.

## **Skeletal Abnormalities**

Studies of skeletal abnormalities have been conducted to a limited extent on the Willamette River.

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**Table 12.** Percent abnormal external and internal features in fish from the Willamette Basin, Oregon [--, not available; ODEQ, Oregon Department of Environmental Quality]

River	River mile	Years	Species	Number samples	Percent abnormalities										
					Eyes	Gills	Pseudo- branch	Thymus	Fins	Opercles	Spleen	Hindgut	Kidney	Liver	Reference
Willamette	1	1992	Northern	20	5	5	0	5	0	0	0	40	0	85	Tetra Tech, Inc.
Willamette	6.5	1992	squawfish Largescale sucker	20	0	0	5	20	25	5	15	5	0	45	(1993b, 1995d) Tetra Tech, Inc. (1993b, 1995d)
Willamette	7	1988	Northern squawfish	10 to 20	0	47	100	40			0	0	7	27	ODEQ (1994b)
Willamette	25	1992	Largescale sucker	20	5	0	5	5	15	20	0	20	0	35	Tetra Tech, Inc. (1993b, 1995d)
Willamette	38	1988 1989	Northern squawfish	10 to 20	0 0	47 0	100 0	40 20		 	0 0	12 67	7 0	27 20	ODEQ (1994b)
Willamette	47	1988 1989	Northern squawfish	10 to 20	8 0	69 41	100 0	46 53			8 6	0 0	8 24	69 12	ODEQ (1994b)
Willamette	49	1992	Northern squawfish	12	0	17	0	0	15	8	0	17	8	92	Tetra Tech, Inc. (1993b, 1995d)
Willamette	49	1992	Largescale sucker	12	0	8	0	25	17	8	25	17	0	42	Tetra Tech, Inc. (1993b, 1995d)
Willamette	74	1989	Northern squawfish	10 to 20	5	25	0	0			0	5	15	5	ODEQ (1994b)
Willamette	115	1989	Northern squawfish	10 to 20	5	50	5	15			0	5	20	35	ODEQ (1994b)
Willamette	128	1992	Largescale sucker	20	25	40	15	40	15	0	15	20	25	60	Tetra Tech, Inc. (1993b, 1995d)
Willamette	185	1992	Largescale sucker	20	15	80	5	35	20	0	30	25	20	55	Tetra Tech, Inc. (1993b, 1995d)
Santiam	0.5	1988	Northern squawfish	10 to 20	5	32	0	21			0	0	0	63	ODEQ (1994b)
Conser Slough <sup>1</sup>	0.1	1988	Northern squawfish	10 to 20	6	35	0	24			0	11	24	35	ODEQ (1994b)

<sup>&</sup>lt;sup>1</sup>Conser Slough is a tributary of the Willamette River near Albany, Oregon (Willamette River Subbasin).

impacts of environmental pollutants on fish (Mayer and others, 1992). From 1992–94, Tetra Tech, Inc., (1993b; 1995d) examined skeletal abnormalities in juvenile northern squawfish collected from 18 locations along the main stem Willamette River between RMs 3 and 185. The incidence of skeletal abnormalities at RM 3 (less than 2.7 percent) and between RMs 125 and 185 (mean = 2.6 percent) was consistently low and is within the range of 2–5 percent reported for unstressed natural fish populations and laboratory stocks (Gill and Fisk, 1966; Wells and Cowan [1982] as cited in Tetra Tech, Inc., [1995d]). Two sites were sampled between RMs 51 and 125.5. Elevated percentages of skeletal deformities occurred at RM 113 (22.2 percent) and RM 72 (21.7 percent) in 1994; these values are significantly higher than the values of upstream sampling locations. The highest incidence of deformities occurred within the Newberg Pool, which extends from RM 26.5 to RM 60 on the main stem Willamette River. Fish collected from the east bank of RM 49.7 exhibited 74 percent skeletal abnormalities in 1994. Studies of the Newberg Pool area in 1993 found a range of skeletal deformities from 22.6 percent to 52.0 percent, with values declining gradually in the downstream direction. Juvenile northern squawfish from a reference location on the Luckiamute River exhibited skeletal deformities at 1.6 percent.

These types of studies are useful for determining the

Overall, the results of the Tetra Tech, Inc., (1995d) study have shown that a background deformity rate of up to 3 percent in the main stem Willamette River is not uncommon. Although no specific cause for juvenile northern squawfish deformities in the Newberg Pool was identified, Tetra Tech, Inc., (1995d) cites a variety of potential causes, including genetic factors, nutritional deficiencies, parasitism, elevated water temperatures, low dissolved oxygen concentrations, trace elements, pesticides, PCBs, bleached kraft pulp and paper mill effluent, and ore smelter effluent.

The role of hybridization in causing skeletal deformities in the Newberg Pool area was assessed by Markle (1994b). The occurrence of hybrids between northern squawfish and chiselmouth have been documented in the Willamette River, and skeletal deformities could be associated with hybridization. Northern squawfish specimens previously collected from the Newberg Pool area (RM 49.7) that had high levels of deformities and from the Corvallis area (RM 125.5) that had low levels of deformities, were further exam-

ined to assess hybridization in the fish. Using multivariate analyses, Markle (1994b) suggested that hybridization contributed to, but was not the primary cause of, the observed pattern of deformities between the two sample sites. Markle (1994b) also noted that redside shiners, a nonhybrid species, from the Newberg Pool sample had high rates of deformities (26 percent of 76 fish). This information indicates that an area effect is present at the Newberg Pool, and lowers the probability that deformities were solely due to hybridization.

# **Aquatic Toxicological Responses**

## **Bioassays**

Tetra Tech, Inc., (1993b, 1995d) reported a higher incidence of skeletal deformities in juvenile northern squawfish from the Newberg Pool area (RM 26.5-60) on the main stem Willamette River than in northern squawfish from upstream or downstream sites. Effluents discharged near RM 56 were bioassayed by Curtis and Siddens (1995) to determine the teratogenicity of point sources of pollution. A fathead minnow (*Pimephales promelas*) embryo-larva survival and teratogenicity test was used for the assessment. The bioassay indicated that neither sewage-treatment-plant nor pulp-mill effluents were teratogenic. However, undiluted sewage treatment plant effluent was lethal to a high percentage (>90 percent) of embryos and larvae. The role of maternal transfer of contaminants to eggs or embryos and of male gamete damage was undetermined.

#### **Enzyme Induction Assays**

Various hydrocarbons have the potential to induce enzymatic activity in animals. Exposure to numerous aromatic compounds, including chlorinated organics such as dioxins, furans, and PCBs, induces cytochrome P450–1A1 activity in the liver. Induction of cytochrome P450–1A1, which catalyzes ethoxyresorufin O-deethylase (EROD) and aryl hydrocarbon (benzo[a]pyrene) hydrolase (AHH) activity, has been correlated with toxic potency of contaminants. Induction of cytochrome P450–1A1 may also be the most sensitive early indicator of exposure of organisms to toxic organic compounds. Determining the responses of these compounds (biomarkers) in animals that are sensitive to contaminant exposure allows a better estimate of exposure to chemicals or resultant effects and

an assessment of environmental degradation (Huggett and others, 1992).

In 1990, enzyme induction assays were performed on liver samples from mountain whitefish collected from the main stem Willamette River (RM 143, 145, 147, and 176) Middle Fork Willamette River (RM 8), and McKenzie River (RM 3); (Oregon Department of Environmental Quality, 1994b). Five liver samples were collected at each of the six sample sites. Liver samples were analyzed for both EROD and AHH activity. Although there were no significant differences among site means for EROD or AHH activity, ODEQ (1994b) notes that results may have been affected by sample degradation. Control or reference EROD values for mountain whitefish were not available or not reported by ODEQ, and it is unknown if induction of the enzyme activities at the levels reported would be indicative of exposure to chlorinated organic compounds.

Curtis and others (1993) attempted to determine the sensitivity of cytochrome P450–1A1 induction in fish as a biomarker for distribution of TCDD and TCDF in the Willamette River. This study found good correlations between hepatic microsomal EROD activity and total cytochrome P450-1A1 content (quantified by Western blotting) in both common carp and cutthroat trout, but no evidence for positive biomarker responses in northern squawfish. Strong correlations were demonstrated between carp muscle TCDD or TCDF concentrations and hepatic EROD activity or total cytochrome P450–1A1 content. Common carp collected from Portland Harbor near RM 7 contained elevated TCDF in muscle tissue and contained significantly more total cytochrome P450–1A1 in hepatic microsomes than in fish from upstream locations (Curtis and others, 1993). No significant seasonal effects were found in the hepatic biomarkers for the fish evaluated in July and October 1990. These results suggest that common carp may be better indicators than northern squawfish to document exposure to chlorinated organic compounds based on hepatic biomarker responses. Additionally, common carp at RM 7 are exposed to these contaminants to a greater extent than common carp in other areas of the Willamette River.

#### **Growth Assays**

The ODEQ (1994b) conducted a growth assay on sculpin to determine if growth varied between contaminated and reference sites. Twelve to 18 sculpins were collected from RM 7 on the main stem Willamette River (a contaminated area) and from RM 2 on the

Clackamas River (an uncontaminated reference area). Three groups of four to six individuals from each site were fed at "fast", "moderate", or "slow" growth rations for 21 days. The study found no significant difference in growth between sculpins collected from contaminated and reference sites.

#### **Ongoing Research**

A variety of research is currently being conducted in the Willamette Basin that will aid in the understanding of contaminant impacts on aquatic biota. As part of a national study on endocrine disrupters in fish, the USGS, Biological Resources Division (BRD) are assessing the effects of contaminants on common carp endocrine systems in the Willamette River Basin (Steve Goodbred, oral commun., 1995). Sampling occurred in 1994 and 1995 in an off-channel pond adjacent to the Middle Fork Willamette River near Springfield and on the main stem Willamette River at Portland (RM 6). Three biomarkers are being assessed: hormones (estrogen and testosterone levels), vitellogenin, and histopathology.

Dr. Charles J. Henny with the USGS (BRD) in Corvallis, Oregon, has been investigating population changes and productivity of osprey in the Willamette Basin (oral commun., 1995). As part of this investigation, 10 osprey eggs and fish samples from 16 pools along the main stem Willamette River have been collected for analysis of organochlorine pesticides, congener specific PCBs, and dioxins and furans.

Carmen Thomas, a cooperative education student at Oregon State University, has been funded by the USFWS to assess contaminants in great blue heron colonies. Both fish prey and eggs have been collected from heronries along the Willamette River. These samples will be analyzed for a variety of contaminants, including organochlorine pesticides, congener PCBs, dioxins and furans, and trace elements. Eggs will also be used in a bioassay to assess exposure to planar halogenated hydrocarbons (PHHs) and to determine relative levels of cytochrome P450–1A1 and induction of EROD activity.

#### **SUMMARY**

This report reviews and summarizes available aquatic biological data for the Willamette and Sandy River Basins (Willamette Basin), Oregon, as part of

the U.S. Geological Survey's National Water-Quality Assessment Program. This information will be used in conjunction with data on physical and chemical parameters in a multidisciplinary, integrated assessment of water quality to determine the status of aquatic environments and guide the design of future studies. Biological parameters emphasized include the status, distribution, and trends of aquatic biota; the condition of aquatic and riparian habitat, and the response of aquatic biota to natural and human-associated impacts, including the level, type, and effect of contaminants.

The aquatic biota emphasized are algae, macroinvertebrates, and fish because of their potential role as indicators of water quality and their potential role in contaminant analyses. Information on selected semiaquatic amphibians and reptiles, birds, and mammals is also presented to provide a more thorough assessment of aquatic biota in the Willamette Basin. Additional emphasis is placed on species designated as "special status species" by regulatory agencies.

The 12,000 square-mile Willamette Basin includes 15 major subbasins in 5 ecoregions. It also includes between 9,000 and 10,000 miles of streams and over 2,000 lakes. Elevations range from near sea level to approximately 11,500 feet. Land use is primarily forest and agriculture. The drainage system is dominated by the northward flowing Willamette River. Streamflow in the Willamette River and its major tributaries is highly regulated by dams and reservoirs that were constructed primarily for hydroelectric power generation.

The Willamette Basin contains a diversity of aquatic environments. High-elevation, headwater streams in the Cascade and Coast Range Mountains are high-gradient, fast-flowing, shallow, cold water streams. Streams and rivers of the lowlands are low-gradient, deep water habitats. Additionally, large water bodies, such as lakes and reservoirs, vary from nutrient-poor, low-productivity montane lakes to highly productive warm water lakes in the lowlands.

Considerable information is available on aquatic biota in the Willamette Basin, although the information is highly uneven relative to taxa and spatial scope. There is extensive information on high profile taxa such as salmonid fishes, but less information is available for macroinvertebrates, and relatively little data have been collected for algae. Additionally, some areas such as the H.J. Andrews Experimental Forest and the main stem Willamette River have been extensively

studied, whereas there are limited data available for most other areas.

Information on the abundance and distribution of algae in the Willamette Basin is limited primarily to the main stem Willamette River and a few sites in other Willamette Valley streams and rivers. Diatoms were the dominant algae in the Willamette River in the 1960s and 1970s, but recent sampling as part of the Willamette River Basin Water Quality Study indicate that blue-green algae are important.

The basin supports a diverse aquatic macroinvertebrate fauna. Available data indicate a relatively high diversity of taxa and a high richness of EPT (Ephemeroptera, Plecoptera, Trichoptera) taxa in the upper reaches of the basin. In the lower main stem reaches, invertebrate assemblages are dominated by pollution tolerant organisms and those adapted to low dissolved oxygen levels. Extensive long-term studies by Oregon State University researchers at H.J. Andrews Experimental Forest, Berry Creek, and Oak Creek provide the most thorough information on macroinvertebrate diversity and abundance in the basin.

Approximately 61 fish species occur in the basin, although nearly half are introduced. They include nine anadromous species (primarily salmonids) and members of 16 families. Several species have special Federal status, including Oregon chub (endangered), lower Columbia River coho salmon (proposed threatened), bull trout (candidate), and river lamprey and Pacific lamprey (species of concern). Two additional salmonids, fall chinook salmon and coastal cutthroat trout, are considered critical by the State of Oregon.

The occurrence, distribution, and abundance of fish in the Willamette Basin have changed since human occupation, primarily due to habitat degradation, dams and other fish passage issues, hatcheries, and introduced species. Fish species richness and distribution are highly correlated with elevation, stream gradient, and water temperature. High elevation, cold water, mountain streams are characterized by a few species of salmonids, sculpin, suckers, and whitefish. Low elevation, main stem reaches of major rivers and streams are dominated by warm water species, such as bass, catfish, and several species in the panfish group. Many of the lowland rivers and lakes are now dominated by introduced warm water species.

Semiaquatic wildlife in the basin include a few species of mammals; numerous birds, such as waterfowl, shorebirds, herons, and gulls; and several species of frogs, salamanders, and turtles. These taxa are conspicuous and important biota in aquatic communities, often as top predators, and some species, such as bald eagle and osprey, may be useful as indirect biological indicators of water quality. Twenty-one semiaquatic wildlife species have been designated as "special status species."

The effect of an expanding human presence in the Willamette Basin has substantially altered aquatic and riparian habitats, and the biota that use or reside in these habitats. Construction of dams, channelization and bank stabilization of rivers, species introductions, supplementations of fisheries through aquaculture, agricultural practices, timber harvest, and urbanization have contributed to changes in aquatic habitats and biota from historical conditions.

The extent of impacts on aquatic biota has been most apparent in declining populations of anadromous salmonids. Dam construction has resulted in the inundation and physical blockage of approximately 400 miles of salmonid spawning and rearing habitat. Development of fish passage facilities at dams and supplementation of native populations with hatchery fish have attempted to restore native runs. However, the successes have been minimal, and several species/stocks are extinct or at a moderate to high risk of extinction.

A variety of aquatic toxicological investigations, primarily focusing on fish, have been undertaken in the Willamette Basin. Two comprehensive investigations and several smaller studies have been conducted on the main stem Willamette River and selected tributaries. The Willamette River has also been included in several national contaminant studies administered by the U.S. Environmental Protection Agency and U.S. Fish and Wildlife Service. These studies have addressed chlorinated pesticides, polychlorinated biphenyls (PCBs), dioxins and furans, polycyclic aromatic hydrocarbons (PAHs), and trace elements in tissues of aquatic biota, as well as fish health assessments, skeletal abnormalities, and aquatic toxicological responses. Several pesticides exceeded USEPA and State water-quality criteria for the protection of aquatic life. Elevated PCB, dioxin, and furan concentrations were associated with point sources, such as pulp and paper mills. PAHs were seldom detected in fish. Elevated levels of mercury in fish tissue were associated with reservoirs in watersheds containing cinnabar deposits and in which there have been mercury and gold mining activities. Assessments of fish health indicated that abnormalities were higher in the main stem Willamette River than in its tributaries. Background skeletal deformity rates of

about 3 percent were not uncommon in the main stem Willamette River, with abnormalities reaching 74 percent in the Newberg Pool. Bioassays, enzyme induction assays, growth assays, and biomarker studies have generally produced mixed results, with no indication of substantial contaminant impacts.

Contaminant information on semiaquatic wild-life is generally lacking, with the exception of a few site-specific studies. Concentrations of *p,p*'-dichlorodiphenyldichloroethylene (DDE) at levels that could impair productivity have been reported in great blue heron eggs and a chick from Albany along the main stem Willamette River. Mercury concentrations that are associated with reproductive impairment have also been reported in a bald eagle egg from Cottage Grove Lake. Ongoing investigations include an examination of endocrine disruptors in fish, contaminants in osprey and great blue herons, and a summary of historical and current mercury concentrations in Oregon lakes.

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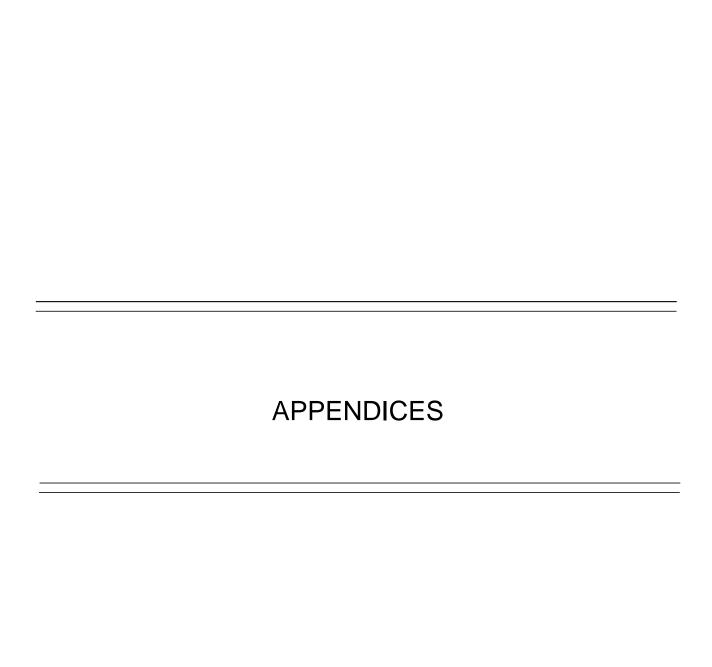
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[Includes references of field investigations that provide information on the distribution and abundance of algae, macroinvertebrates, and fish. Subbasins (see fig. 5): CA, Calapooia; CF, Coast Fork Willamette; CL, Clackamas; LT, Long Tom; LU, Luckiamute; MA, Marys; MC, McKenzie; MF, Middle Fork Willamette: PM, Pudding/Molalla; RI, Rickreall; SA, Santiam; SY, Sandy; TU, Tualatin; WR, Willamette River; WB, throughout the Willamette Basin; YH, Yamhill; RM, river mile; BLM, Bureau of Land Management; --, no data]

						Taxa	
Reference	Subbasin	Temporal coverage	Spatial coverage	Торіс	Algae	Inverte- brates	Fish
Aho (1976)	MC	1973–74	Mack Creek	Cutthroat trout populations in shaded and unshaded stream sections		X	X
Anderson (1992)	MC	1982—83 and 1986	Mack, Quartz, and Grasshopper Creeks	Influence of disturbance on insect fauna; riparian insect taxa		X	
Anderson and Lehmkuhl (1968)	MA	Fall 1965 and 1966	Oak Creek	Catastrophic drift of insects		X	
Anderson and Wold (1972)	MA	May 1968– Dec.1980	Oak Creek	Emergence trap collections of Trichoptera		X	
Anderson and Bourne (1974)	MA	Aug. 1970– July 1971	Oak Creek	Life history of 3 caddisflies; <i>Anagapetus bernea, Glossosoma penitum</i> , and <i>Agapetus bifidu</i> s		X	
Anderson and others (1978)	LU,MC	July 1976	Berry, Mack, Devils Club, and Lookout Creeks; McKenzie River	Role of invertebrates in wood processing in streams		X	
Azam (1969)	LU,MA	1966–68	Oak and Berry Creeks	Life history and production of <i>Sialis</i> californica and <i>Sialis rotunda</i>		X	
Ball (1946)	MA	unknown	Oak Creek	Seasonal succession of stoneflies		X	
Baker (1979)	MF	1976–78	Big Fall and Hehe Creeks	Impacts of logjam removal on fish populations and stream habitat			X
Baker and others (1995)	CL	Summer 1994	Collawash River watershed; other sites in subbasin	Field verification of fish distribution and composition			X
Beak Consultants, Inc. (1985)	SY	July 1984; March 1985	Sandy River between Bull Run River and Columbia River	Fish species occurrence and abundance			X
Beak Consultants, Inc. (1993)	SY	Oct. 1992	Bull Run Lake and tributaries	Effect of water withdrawal on fisheries resources		X	X
Buchanan and others (1981)	MC,SA,WR	April–June 1976–77	Willamette, Santiam, and McKenzie Rivers	Predation by squawfish on salmonids			X
Burns (1993)	CL	Summer 1991	10 Cascade Range lakes	Phytoplankton of mountain lakes	X		
Carter (1975)	TU	July 1972– Sept. 1973	Middle course of the Tualatin River	Algae occurrence and distribution	X		

						Taxa	
Reference	Subbasin	Temporal coverage	Spatial coverage	Topic	Algae	Inverte- brates	Fish
Carter and others (1976)	TU	June–Oct. 1976	Tualatin River	Algae occurrence and distribution	X		
Clifton (1985)	SY	July-Aug. 1977-81	Six streams in Bull Run watershed	Periphyton and invertebrate distribution and abundance	X	X	
Craig and Townsend (1946)	WR,MC,MF, PM,SA	1940–42	Molalla, Santiam, McKenzie, Willamette, and Middle Fork Willamette Rivers	Effects of Willamette Valley Project on fish			X
Deschamps (1952)	CL,MC,SA, WR	July–Nov. 1951	Willamette, McKenzie, Santiam, and Clackamas Rivers	Invertebrate use as bio-indices of pollution	-	X	-
Dever (1962)	LU	unknown	Berry Creek	Algae in a woodland stream	X	-	-
Diamond (1982)	LU, MA	June 1975-April 1976	Oak and Berry Creeks	Population dynamics of Juga plicifera	-	X	-
Dieterich (1992)	LU,MA	1987–88	MacDonald Forest	Insects of summer-dry headwater streams	-	X	-
Dimick and Merryfield (1945)	CA,CF,CL, LT,LU,MA, MC,MF,PM, RI,SA,TU, WR,YH	AugSept. 1944	Willamette River and tributaries	Fish distribution and abundance relative to pollution	-	X	X
Dodge (1994)	MC,MF	June-Aug. 1992	Middle Fork Willamette River and Lookout Creek	Habitat use by two species of dace	-	-	X
Dudley (1982)	MA,SA	1978	Marys River and sites in Santiam watershed	Ecology of <i>Lipsothrix</i> sp.	-	X	-
Dudley and Anderson (1982)	WB	1978 and 1979	100+ sites in WB (sites not listed)	Invertebrates associated with wood debris in aquatic habitats	-	X	-
Dudley and Anderson (1987)	MA,MC	Feb.1977–March 1979	Greasy Creek and Quartzville Creek watersheds	Wood inhabiting craneflies in streams	-	X	-
EA Engineering, Science and Technology, Inc. (1990a)	e MC	Oct. 1989	Lower McKenzie River between Springfield and Leaburg Dam	Aquatic insect distribution and abundance	-	X	-
Earnest (1967)	LU	Oct. 1963– Oct. 1965	Berry Creek	Production of Oxytrema silicula	-	X	-
Everest and others (1985)	CL	1982–85	Fish Creek	Salmon and steelhead abundance, behavior, and habitat	-	-	X
Farr and Ward (1993)	WR	MarNov. 1987-90	Portland Harbor, RM 0.3-27.0	Fish distribution	-	-	X
(1990a) Earnest (1967) Everest and others (1985)	CL	Oct. 1965 1982–85	Berry Creek Fish Creek	Salmon and steelhead abundance, behavior, and habitat	-		

						Taxa	
Reference	Subbasin	Temporal coverage	Spatial coverage	Торіс	Algae	Inverte- brates	Fish
Fetrow Engineering and Scientific Resources (1989)	WR	1988–89	Delta Ponds in Eugene	Phytoplankton distribution and abundance	X	-	-
Finger (1982)	MA	June 1977–Sept. 1978	Marys River	Segregation of three species of sculpin	-	-	X
Friesen and others (1994)	TU	Summer-Fall 1993	Tributaries of the Tualatin River	Distribution of fish and crayfish	-	X	X
Frissell and others (1985)	SA	Aug. 27, 1984	Minto Creek	Trout densities and habitat use in logged and forested stream sections			X
Furnish (1989)	MA	1982–85	Oak Creek	Growth, production, and distribution of <i>Juga silicula</i>	-	X	-
Goetz (1994)	MC,MF	1989–91	Anderson Creek, Trailbridge Reservoir, McKenzie River	Distribution and juvenile ecology of bull trout	-	-	X
Grafius (1977)	LU,MC	April 1974–Feb. 1975	Berry and Mack Creeks	Utilization and processing of leaves and needles by Trichoptera		X	
Grafius and Anderson (1979)	LU	Aug. 1974 –Feb. 1975	Berry Creek	Utilization of deciduous leaves as food by Lepidostoma quercina		X	
Gregory (1980)	MC	1974–75	Mack and Lookout Creeks	Effects of light, nutrients, and grazing on periphyton	X		
Gregory (1993)	CA,CF,CL, MC,MF,PM, SA,TU,WR, YH	Summer 1992	Main stem, Coast Fork, and Middle Fork Willamette, McKenzie, Calapooia, Santiam, Yamhill, Molalla, Clackamas, and Tualatin Rivers	Periphyton abundance and productivity	X		
Hasselman and Garrison (1957)	MF	Summer 1957	Lookout Point and Dexter Reservoirs	Squawfish reproduction and movements			X
Hawkins and Sedell (1981)	MC,MF	All seasons 1976	Devils Club, Mack, and Lookout Creeks; McKenzie River	Longitudinal and seasonal changes in invertebrate communities		X	
Hawkins and Furnish (1987)	MC,MF	1978–79	Mack, Mill, and Fawn Creeks	Correlations of stream taxa abundance with Juga silicula	X	X	-
Hawkins and others (1982)	MC	1978–79	Mack, Mill, Cougar, Fawn, and North Fork Wycof Creeks	Invertebrate community structure and abundance	X	X	
Hawkins and others (1983)	MC,MF	1978–79	Mack, Mill, and Fawn Creeks	Fish abundance relative to food sources and habitat features		X	X

						Taxa	
Reference	Subbasin	Temporal coverage	Spatial coverage	Topic	Algae	Inverte- brates	Fish
Hjort and others (1984)	WR	June-Aug. 1992	Willamette River below Salem, RM 58–66	Fish and invertebrates of revetments		X	X
HMS Environmental, Inc., and Miller (1988)	WR	Summer 1988	Willamette River near Halsey, RM 142.3 to 147.9	Effects of pulp mill discharge on aquatic organisms	X	X	X
House (1995)	PM	AugSept. 1981-91	Dead Horse Canyon Creek watershed	Cutthroat trout population variability			X
Hughes and Gammon (1987)	WR	August 1983	Throughout Willamette River	Longitudinal changes in fish assemblages			X
Hutchison and Aney (1964)	CL,SY,TU	Summer 1963	Lower Willamette Basin	Fish distribution			X
Hutchison, Thompson, and Fortune (1966)	CF,LT,MC, MF	1964–66	Upper Willamette Basin	Fish distribution			X
Johnson and others (1989)	WR	Summer 1989	Willamette River near Halsey, RM 142.3 to 150.5	Effects of pulp mill discharge on aquatic organisms	X	X	X
Kerst (1969), Kerst and Anderson (1974, 1975)	MA	June 1968–May 1969	Oak Creek	Occurrence and distribution of stoneflies		X	
Knutsen and Ward (1991)	WR	1987–90	Portland Harbor, RM 0-16.8	Juvenile salmonid migration behavior			X
Korn and others (1967)	CL	Feb. 1962–June 1965	North Fork Reservoir of the Clackamas River	Effect of small impoundments on juvenile anadromous salmonids			X
Kraft (1963)	LU	Dec. 1959–Dec. 1960	Berry Creek	Seasonal occurrence and distribution of aquatic insects		X	
Kruse (1988)	CA,MA,SA, YH	July-Sept. 1982; June-Sept. 1983	Calapooia River: Thomas, Rock, Turner, Griffith, and Greasy Creeks	Fish distribution and habitat			X
Lamberti and others (1991)	MC	Mar. 1986– Nov. 1988	Quartz Creek	Stream ecosystem recovery after a catastrophic debris flow	X	X	X
Lehmkuhl (1968)	MA	Oct. 1965–Sept. 1966	Oak Creek	Life history of four species of <i>Epeorus</i>		X	
Lehmkuhl (1969)	MA	June 1967– June 1968	Oak Creek	Biology and downstream drift of six species of Ephemeroptera		X	
Lehmkuhl and Anderson (1970)	MA	July 1967–June 1968	Oak Creek	Ecology of Cinygmula reticulata		X	
Lehmkuhl and Anderson (1971)	MA	July 1967– June 1968	Oak Creek	Biology and taxonomy of <i>Paraleptophlebia</i> sp.		X	

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						Taxa	
Reference	Subbasin	Temporal coverage	Spatial coverage	Topic	Algae	Inverte- brates	Fish
Li and others (1983)	MA,CA	1981	Greasy Creek; Calapooia River	Fish distribution and habitat above and below impoundments			X
Li and others (1984)	WR	Aug. 1983; April– June 1984	Willamette River, RM 118–142	Fish use of spur dikes and revetments			X
Logan and others (in press)	CL	Aug. 1991	Pond near Clackamas River	Capture of oriental weatherfish			X
Long (1982)	MF	Aug. 17–25, 1981	Lowell Ranger District, Willamette National Forest	Status and distribution of the Oregon chub			X
Mangum (1990)	SY	Oct. 1989 and May 1990	Still Creek	Macroinvertebrate sampling		X	
Mangum (1991a)	SA	Summer 1991	2 streams on Salem BLM District	Macroinvertebrate sampling		X	
Mangum (1991b)	MC,MF	Summer 1991	10 streams on Eugene BLM District	Macroinvertebrate sampling		X	
Markle and others (1989)	MF	July–Sept. 1987	Middle Fork Willamette River	Distribution of the Oregon chub			X
Markle and others (1991)	MF	July–Sept. 1987	Middle Fork Willamette River	Distribution of the Oregon chub		X	X
Mason (1963)	LU	1959–60	Berry Creek	Life history and production of the crayfish		X	
Massey (1967a;b)	WR	Aug. 1964–Dec. 1966	Industrial area near Willamette Falls	Juvenile fish abundance and timing of downstream migration			X
Mattson (1962)	PM,WR	Feb.1947– July 1951	Willamette River near Lake Oswego; Molalla River	Chinook salmon life history			X
McIntyre (1967)	LU	1964–66	Berry Creek	Food relations and production of cutthroat trout		X	X
Miller (1979)	PM	July and Sept. 1977	Four tributaries of the Molalla River	Periphyton and benthic invertebrate abundance and distribution	X	X	
Moore (1987)	MC	June 1982– Feb. 1984	Mack, Quartz, and Grasshopper Creeks	Species assemblages associated with stream margins	X	X	X
Moore and Gregory (1989)	MC	Summer, 1987	Lookout Creek	Riparian influences on distribution and abundance of salmonids			X
Moring and others (1986)	MC,SA,WR	April 1976–May 1979	Willamette, McKenzie, and Santiam Rivers	Movements of coastal cutthroat trout			X
Murphy and Hall (1981)	MC	July–Oct. 1976	33 sites in or near the H.J. Andrews Experimental Forest	Effects of clear-cut logging on predators and their habitat	X	X	X
Murphy and others (1981)	MC	June-Nov. 1978	Mack, Mill, Cougar, Fawn, and Wycof Creeks	Effects of canopy modification and accumulated sediment	X	X	X

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						Taxa	
Reference	Subbasin	Temporal coverage	Spatial coverage	Торіс	Algae	Inverte- brates	Fish
Nickelson (1974)	LU	Aug.1972– Aug.1973	Berry Creek	Population dynamics of coastal cutthroat trout			X
Noble (1952)	CL,MC,SA, WR	July–Nov. 1951	Willamette, Clackamas, Santiam, and McKenzie Rivers	Fish as biological indicators of pollution			X
Oregon Department of Fish and Wildlife (1995b)	MC	1992–94	Mohawk River	Relative abundance and timing of cutthroat trout movements			X
Oregon State Game Commission (1963)	LU,MA,RI, SA,YH	1961–62	Middle Willamette River Basin, Coast Range subbasins	Fish distribution			X
Pearsons (1989)	MF	May 1986–Sept. 1987	Shady Dell Pond; Buckhead Creek Slough	Occurrence and habitat of the Oregon chub	X	X	X
Pereira (1980)	LU,MC	July 1979– June 1980	Berry and Mack Creeks	Life history of Cinygma integrum		X	
Raymond (1983)	SY	Summer 1979	Bull Run Lake	Diatom and algae occurrence	X		
Reese (1966)	LU	Feb. 1964– Jan. 1965	Berry Creek	Structure of benthic communities	X		
Rickert and others (1977)	WR	June-Sept. 1973 and 1974	Willamette River; RMs 7.0,12.8, 21.2,35.0,50.0	Algal distribution and abundance	X		
Rinella and others (1981)	SA,WR	August 1978	Willamette and Santiam Rivers	Algal distribution and abundance	X		
Rodnick (1983)	MA	Aug. 1980- July 1981	Greasy Creek	Distribution and habitat selection of the redside shiner			X
Rounick and Gregory (1981)	MC	Oct. 1976–March 1977	Lookout, Mack, McCrae, and Arnold Creeks	Effects of light and discharge on periphyton	X		
Scheidt and Nichols (1976)	MF	May 1975- Feb. 1976	Hills Creek Reservoir	Algal abundance	X		
Scheerer and others (1992)	LT,MF,SA,	Oct. 1990; May– June 1991	Middle Fork Willamette and Santiam Rivers; Baskett Slough, Finley, and Ankeny National Wildlife Refuges	Status and habitat of the Oregon chub			X
Scheerer and others (1993)	CF,LU,MA, MF4.	May–June 1992	Coast Fork and Middle Fork Willamette River; Luckiamute River; Oak Creek	Status and habitat of the Oregon chub			X
Scheerer and others (1994)	CF,LT,MF, SA,	April–June 1993	Coast Fork and Middle Fork Willamette River; Santiam River; Finley and Ankeny National Wildlife Refuges	Status and habitat of the Oregon chub			X

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						Taxa	
Reference	Subbasin	Temporal coverage	Spatial coverage	Торіс	Algae	Inverte- brates	Fish
Scheerer and others (1995)	CF,MF,PM, SA,WR	April–Sept. 1994	Numerous sites in middle and upper Willamette Basin	Status and habitat of the Oregon chub			X
Shibahara and Lumianski (1995)	CL	May–June 1994	Lower Clackamas River	Movement, distribution and habitat use of winter steelhead			X
Smith and Korn (1970)	MF	June 1966–70	Big Fall Creek watershed and reservoir	Fish species composition			X
Speir (1976)	LU,MA	1971 and 1972	Berry, Oak, and Soap Creeks	Ecology and production of four black fly species		X	
Steedman (1983); Steedman and Anderson (1985)	LU,MA	Oct. 1980–82	Berry and Yew Creeks	Ecology of the beetle, Lara avara		X	
Sutherland (1976)	TU	Spring/summer 1975–76	13 tributaries of Tualatin River	Invertebrate composition and distribution		X	
Tetra Tech, Inc. (1993b)	MC,WR	AugSept. 1992	Willamette River and lower McKenzie River	Fish distribution and abundance			X
Tetra Tech, Inc. (1994)	MC,TU,WR	AugSept. 1992; Oct. 1993	Willamette, Tualatin, and McKenzie Rivers	Invertebrate distribution and abundance		X	
Tew (1970)	LU	1968–70	Tributary of Berry Creek	Insects of an intermittent stream		X	
Thompson (1965)	CF,MF	Summer 1964	Coast and Middle Fork Willamette River	Fish distribution and abundance			X
TW Environmental, Inc. (1994)	SY	Nov. 8, 1994.	Bull Run River	Invertebrate sampling		X	
Ward and Nigro (1991)	WR	MarJune 1988-90	Portland Harbor, RM 0-27	Fish assemblages and habitat characteristics			X
Ward and others (1988)	WR	Autumn 1986–87	Lower Willamette River	Migration, behavior, and survival of juvenile salmonids			X
Ward and others (1991)	WR	Mar.–June 1988–89; April–June 1990	Lower Willamette River; RM 0–16.8	Status and biology of black crappie and white crappie			X
Ward and others (1994)	WR	Mar.–June and Nov. 1987–90	Lower Willamette River up to RM 20	Migration behavior and habitat use of juvenile salmonids, and predation on them by northern squawfish			X
Wetherbee (1962)	SA	1953–61	Detroit Reservoir	Fish community sampling			X
Wille (1976)	WR	Summer 1974	Willamette River KM 11,21,35,56, and 80	Algal sampling	X		

						Taxa	
Literature citation	Subbasin	Temporal coverage	Spatial coverage	Topic	Algae	Inverte- brates	Fish
Willis and others (1960)	WB	1958–59	17 major rivers and tributaries	Environmental survey			X
Wilzbach (1984)	MC	1982–83	Grasshopper Creek	Effects of prey availability and cover on trout abundance and growth		X	X
Wilzbach and others (1986)	MC	June-Aug. 1984	Grasshopper Creek	Effects of habitat manipulations on cutthroat trout and invertebrate drift		X	X
Wisseman (1989)	SY	Aug. 1988	Timberline Lodge, Mt Hood National Forest	Occurrence and habitat of threatened/ endangered invertebrates		X	
Wisseman (1992a)	SY	June 11, 1992	Timberline Lodge, Mt. Hood National Forest	Inventory for sensitive caddisflies		X	
Wisseman (1992b)	CL	Sept. 1991	Mt. Hood National Forest, Squaw Lakes	Benthic invertebrate biomonitoring		X	
Wisseman (1992c)	MF	1990–91	Willamette National Forest, Rigdon Ranger District	UV light trap survey for sensitive caddisflies		X	
Wisseman (1995)	SY	Fall 1992–93	Mt. Hood National Forest, Zigzag Ranger District	Benthic invertebrate biomonitoring		X	
Wustenberg (1954)	MC	June–Oct. 1951 and 1952	Lookout Creek	Effects of logging on a trout stream		X	X
Wyatt (1959)	MC	June 1956–Sept. 1957	Lookout Creek	Movements and reproduction of cutthroat trout			X
Zakel and Reed (1984)	MC	Sept. 1980–Nov. 1983	Leaburg Dam on McKenzie River	Downstream migration of fish at Leaburg Dam			X
Ziebell (1954)	SA	Oct. 1952–Sept. 1953	South Santiam River below Lebanon	Biological evaluations of pollution conditions		X	X
Zirges (1972)	CF,LT,MA,RI	Spring/ Summer 1972	Rickreall Creek; Marys, Long Tom, and Coast Fork Willamette Rivers	Morphological study of blackside dace			X

## APPENDIX B. ALGAE REPORTED FROM AQUATIC ENVIRONMENTS IN THE WILLAMETTE BASIN, OREGON

[Subbasins (see fig. 5): CF, Coast Fork Willamette; CL, Clackamas; LU, Luckiamute; MC, McKenzie; MF, Middle Fork Willamette; PM, Pudding/Molalla; SA, Santiam; SY, Sandy; TU, Tualatin; WR, Willamette; YH, Yamhill. Sources: 1, Burns (1993); 2, Carter (1975); 3, Carter and others (1976); 4, Clifton (1985); 5, Fetrow Engineering and Scientific Resources (1989) (includes Delta Ponds as part of WR); 6, Gregory (1993); 7, HMS Environmental, Inc., and Miller (1988); 8, Johnson and others (1989); 9, Miller (1979); 10, Raymond (1983); 11, Reese (1966); 12, Rickert and others (1977); 13, Rinella and others (1981); 14, Scheidt and Nichols (1976); 15, Wille (1976)]

Family	Genus-Species	Subbasin	Source
	PHYLUM: Chlorophyta	a (Green Algae)	
Chaetophoraceae	Stigeoclonium sp.	CF,CL,LU,TU,WR,YH	2,6,11
Characiaceae	Characium sp.	WR	5
Chlamydomonadaceae	Chlamydomonas sp.	CF,CL,LU,SA,SY,TU,WR	1,3,4,6,11,13
Chlorococcaceae	Tetraedron sp.	CL,TU,WR	1,2,5,6
	Tetraedron caudatum	CL	1
	Tetraedron minimum	CL,WR	1,5
	Tetradron quadratum	MF	4,13
Cladophoraceae	Cladophora sp.	TU	3
Closteriaceae	Closterium sp.	TU	2,3
	Roya obtusa	TU	2
Coccomyxaceae	Elakatothrix gelatinosa	WR	5
Cosmariaceae	Micrasterias sp.	TU	2
	Staurastrum sp.	TU,WR	2,3,5,8,13
	Staurastrum gracile	CL,WR	1,5
	Staurastrum paradoxum	MF	4,13
Desmidiaceae	Cosmarium sp.	CL,LU,SA,TU,WR	1,2,5,11,13
Dictyosphaeriaceae	Botryococcus sp.	WR	5
	Botryococcus braunii	CL,MF	1,14
	Dictyosphaerium ehrenbergianum	CL,TU	1,2
Hyalothecaceae	Spondylosium sp.	TU	2
Hydrodictyaceae	Pediastrum sp.	TU,WR	2,3,12,15
	Pediastrum duplex	TU,WR	2,5
	Pediastrum tetras	CL,TU,WR	1,2,5,13
Micractiniaceae	Micractinium pusillum	TU	2,3
Mougeotiaceae	Mougeotia sp.	TU,WR	2,3,5
Oedogoniaceae	Oedogonium sp.	CF,PM,SA,TU,WR	2,3,6,13
Oocystaceae	Ankistrodesmus sp.	TU,WR	3,6
	Ankistrodesmus falcatus	CL,MF,SA,TU,WR	1,2,5,13,14
	Chlorella sp.	CF,CL,MC,MF,PM,TU,WR	3,6,13
	Closteriopsis longissima	WR	5
	Kirchneriella sp.	WR	13

# APPENDIX B. ALGAE REPORTED FROM AQUATIC ENVIRONMENTS IN THE WILLAMETTE BASIN, OREGON—Continued

Family	Genus-Species	Subbasin	Source
	PHYLUM: Chlorophyta (C	Green Algae)—Continued	
Oocystaceae—Continued	Kirchneriella lunaris	TU	2
	Nephrocytium sp.	WR	5
	Oocystis sp.	SA,WR	5,6,13
	Oocystis lacustris	CL	1
	Oocystis pusilla	CL,WR	1,5
	Planktosphaeria gelatinosa	WR	5
	Quadrigula closterioides	WR	5
	Quadrigula lacustris	CL	1
	Selenastrum sp.	CL,TU	1,2
	Selenastrum minutum	CL,WR	1,5,13
	Zoochlorella sp.	TU	2
Palmellaceae	Gloeocystis sp.	WR	5,13
	Sphaerocystis schroeteri	CL,WR	1,5
Scenedesmaceae	Actinastrum sp.	TU	2
	Actinastrum gracilimum	TU	2
	Coelastrum microporum	WR	5
	Crucigenia sp.	TU,WR	2,13
	Crucigenia crucifera	CL	1
	Crucigenia quadrata	CL,WR	1,13
	Crucigenia tetrapedia	WR	13
	Scenedesmus sp.	CL,PM,TU,WR	1,3,5,6,12,13,15
	Scenedesmus abundans	WR	5
	Scenedesmus bijuga	WR	5
	Scenedesmus denticulatus	CL,WR	1,5
	Scenedesmus obliquus	SA,WR	8,13
	Scenedesmus quadricauda	SA,TU,WR	2,5,13
Tetrasporaceae	Tetraspora sp.	LU	11
Jlotrichaceae	Stichococcus sp.	LU	11
	Ulothrix sp.	PM,SY,TU,WR	2,3,4,5,6,9,12
	Ulothrix aequalis	WR	13
	Ulothrix zonata	SY,WR	4,6
Volvocaceae	Eudorina sp.	TU	3
	Eudorina elegans	MF,TU,WR	2,13,14
	Gonium sp.	TU	2,3

# APPENDIX B. ALGAE REPORTED FROM AQUATIC ENVIRONMENTS IN THE WILLAMETTE BASIN, OREGON—Continued

Family	Genus-Species	Subbasin	Source
	PHYLUM: Chlorophyta (G	reen Algae)—Continued	
Volvocaceae—Continued	Pandorina sp.	TU	2,3
	Pandorina morum	MF,WR	13,14
	Volvox sp.	TU	2,3
	Volvox aureus	MF	14
Zygnemataceae	Mougeotiopsis calospora	TU	2
	Spirogyra sp.	SY,TU	2,3,4
	Spirogyra pseudo-floxidina	TU	2
	PHYLUM: Chrysophyta	(Golden-brown algae)	
Chrysococcaceae	Chrysococcus sp.	WR	5
	Chrysococcus rufescens	CL,WR	1,5
Dinobryaceae	Dinobryon sp.	CL,SY,TU,WR	1,2,3,5,10
	Dinobryon sertularia	CL,SA,WR	1,5,13
Ochromonadaceae	Ochromonas sp.	CF,CL,MC,PM,WR	1,5,6
Plagiotropidaceae	Plagiotropis sp.	WR	6
Prymnesiaceae	Chrysochromulina sp.	CL	1
Synuraceae	Chrysosphaerella sp.	CL	1
	Chrysosphaerella longispina	TU	2
	Mallomonas sp.	CL,MF,TU,WR	1,2,3,5,14
	Synura uvella	TU,WR	2,3,5
Vaucheriaceae	Vaucheria sp.	TU	2
Xanthopyceae	Tribonema sp.	LU	11
	PHYLUM: Bac (Diator	= -	
Achnanthaceae	Achnanthes sp.	CF,CL,LU,MC,MF,SA,SY, TU,WR	2,3,6,10,11,12,15
	Achnanthes brevipes	TU	2
	Achnanthes deflexa	WR	8
	Achnanthes hauckiana	CL,WR	1,5
	Achnanthes lewisiana	SA,SY,TU,WR	3,10,12,13,15
	Achnanthes linearis	PM,SA,SY,TU,WR	3,4,8,9,10,13
	Achnanthes minutissima	CL,PM,SA,SY,TU,WR	1,3,4,5,9,10,12,13,15
	Achnanthidium clevei	CL,SY	1,10
	Achnanthidium exigum	SY,WR	5,10
	Achnanthidium lanceolatum	CL,PM,SA,SY,TU,WR	1,3,4,5,8,9,10,12,13,1
	Achnanthidium minutissimum	CL,PM,SA,SY,TU,WR	1,3,4,5,9,10,12,13,15

# APPENDIX B. ALGAE REPORTED FROM AQUATIC ENVIRONMENTS IN THE WILLAMETTE BASIN, OREGON—Continued

Family	Genus-Species	Subbasin	Source
	PHYLUM: Baci (Diatoms)—Co	llariophyta ontinued	
Achnanthaceae—Continued	Cocconeis sp.	CF,LU,MC,MF,TU,WR	2,6,11,12,15
	Cocconeis pediculus	SA	13
	Cocconeis placentula	CL,SA,SY,TU,WR	1,3,5,10,13
	Cocconeis placentula-euglypta	PM,SA,SY,WR	4,8,9,13
	Cocconeis placentula lineata	PM	9
	Rhoicosphenia sp.	LU,WR	6,11
	Rhoicosphenia curvata	PM,SA,TU,WR	2,3,5,8,9,12,13,15
Amphipleuraceae	Amphipleura pellucida	WR	5
	Frustulia sp.	LU,SY	10,11
	Frustulia rhomboides	TU,SY	2,10
	Frustulia rhomboides saxonica	TU	3
Anomoeoneidaceae	Anomoeoneis cf. sphaerophora	WR	7
	Anomoeoneis serians	SY	10
	Anomoeoneis vitra	TU	3
Bacillariaceae	Denticula elegans	TU	3
	Hantzschia sp.	WR	6
	Hantzschia amphioxys	TU	3
Chromulinaceae	Chromulina sp.	CL	1
	Kephyrion sp.	CL	1
Coscinodiscaceae	Cyclotella sp.	TU,WR	2,8,12,15
	Cyclotella atomus	SY,WR	5,10
	Cyclotella comta	SY	10
	Cyclotella glomerata	SY	10
	Cyclotella kutzingiana	SY	10
	Cyclotella meneghiniana	SA,SY,TU,WR	3,5,10,12,13,15
	Cyclotella ocellata	SY	10
	Cyclotella pseudostelligera	SY,WR	10,13
	Cyclotella stelligera	CL,SA,SY,TU,WR	1,3,5,10,12,13,15
	Cyclotella striata	SY	10
	Melosira sp.	CL,LU,SA,SY,TU,WR	1,6,7,10,11,12,13,15
	Melosira ambigua	CL,SA,SY,WR	1,5,10,13
	Melosira distans	CL,SA,SY,TU,WR	1,2,3,10,12,13,15
	Melosira granulata	MF,TU,WR	2,3,12,14,15

Family	Genus-Species	Subbasin	Source
	PHYLUM: Bacill (Diatoms)—Co		
Coscinodiscaceae—Continued		SA,WR	13
	Melosira italica	CL,MF,SY,TU,WR	1,2,10,12,13,14,15
	Melosira varians	PM,SA,SY,TU,WR	2,3,4,5,8,9,12,13,15
	Stephanodiscus sp.	TU,WR	2,6,12,15
	Stephanodiscus astrea	SY,WR	10,12,13,15
	Stephanodiscus astrea minutula	MF,SA,WR	5,13,14
	Stephanodiscus dubius	SY	10
	Stephanodiscus hantzschii	SY,TU,WR	3,5,10,12,13,15
Cymbellaceae	Amphora sp.	CF,LU,MF,PM,SA,SY,WR	6,7,10,11
	Amphora ovalis	SY,WR	5,10,13
	Amphora perpusilla	PM,SA,WR	5,9,13
	Cymbella sp.	CF,LU,MC,MF,SY,TU,WR	2,3,6,10,11,12,15
	Cymbella affinis	PM,SA,TU,WR	3,5,9,13
	Cymbella angustata	SY,WR	5,10
	Cymbella aspera	TU	2
	Cymbella cesatii	SY	10
	Cymbella cistula	WR	5,7
	Cymbella cymbiformis	SA,WR	8,13
	Cymbella graecilis	TU	3
	Cymbella lanceolata	TU	3
	Cymbella lunata	SY	10
	Cymbella microcephala	CL,WR	1,5
	Cymbella minuta	CL,PM,SA,SY,TU,WR	1,3,4,5,8,9,10,13
	Cymbella perpusilla	TU	3
	Cymbella prostrata	TU	2,3
	Cymbella sinuata	PM,SA,SY,TU,WR	3,8,9,10,12,13,15
	Cymbella tumida	SA,TU,WR	2,3,12,13,15
	Cymbella turgidula	WR	8
	Cymbella ventricosa	TU,WR	2,12,15
Epithemiaceae	Epithemia sp.	TU	2,3
	Epithemia sorex	CL,SA,WR	1,5,13
	Epithemia turgida	WR	5
	Epithemia sp.	TU	2
	Rhopalodia sp.	TU	2

Family	Genus-Species	Subbasin	Source
	PHYLUM: Bacill (Diatoms)—Cor		
Epithemiaceae–Continued	Rhopalodia gibba	WR	5,13
Eunotiaceae	Eunotia sp.	LU,TU,WR	2,3,5,11,13
	Eunotia arcus	TU	2
	Eunotia curvata	SY	10
	Eunotia elegans	SY	10
	Eunotia incisa	WR	5
	Eunotia microcephala	SY	10
	Eunotia pectinalis	CL	1
	Eunotia rostellata	SY	10
	Eunotia tridentula var. perminuta	TU	2
	Eunotia tridentula var.persusilla	TU	2
	Eunotia vanheurckii	SY	10
Fragilariaceae	Asterionella formosa	CL,MF,SA,TU,WR	1,2,3,5,12,13,14,15
	Diatoma sp.	LU,WR	11,12,15
	Diatoma hiemale	SY,TU,WR	2,3,10,12,15
	Diatoma hiemale mesodon	PM,SY,WR	4,9,13
	Diatoma tenue elongatum	WR	5
	Diatoma vulgare	TU,WR	2,3,5,12,13,15
	Diatoma vulgare linearis	WR	8
	Fragilaria sp.	CF,LU,SY,TU,WR	2,6,8,10,11,12,13,15
	Fragilaria brevistriata	SY	10
	Fragilaria capucina	CL,MF,WR	1,5,7,14
	Fragilaria capucina mesolepta	WR	5
	Fragilaria construens	CL,SA,SY,TU,WR	1,3,5,10,13
	Fragilaria construens venter	CL,SY,WR	1,5,8,10
	Fragilaria crotonensis	MF,SA,TU,WR	2,5,12,13,14,15
	Fragilaria pinnata	CL,TU,WR	1,3,5
	Fragillaria vaucheria	SA,SY,WR	4,5,8,10,13
	Hannaea sp.	TU	2
	Hannaea arcus	PM,SY,TU,WR	2,3,4,9,10,12,13,15
	Meridion sp.	LU	11
	Meridion circulare	SY,TU	2,3,10
	Synedra sp.	LU,MF,SA,TU,WR	2,3,6,11,12,13,15

Family	Genus-Species	Subbasin	Source
	PHYLUM: Bacilla (Diatoms)—Con	riophyta tinued	
ragilariaceae—Continued	Synedra acus	SY	10
	Synedra cyclopum	CL	1
	Synedra Cunningtonii	WR	12,15
	Synedra delicatissima	TU,WR	3,5
	Synedra goulardi	WR	13
	Synedra mazamaensis	WR	8,12,13,15
	Synedra parasitica	SY,TU,WR	3,10,13
	Synedra radians	CL,WR	1,5
	Synedra rumpens	SA,SY,TU,WR	3,4,5,13
	Synedra tenera	TU	3
	Synedra ulna	PM,SA,SY,TU,WR	2,3,5,9,10,12,13,15
	Synedra ulna constricta	WR	8
	Synedra ulna contracta	SA,TU,WR	2,3,13
	Synedra ulna ulna	WR	8
	Tabellaria sp.	LU,TU	2,11
	Tabellaria fenestrata	TU	2,3
	Tabellaria flocculosa	SY	10
	Tetracyclus lacustris	SY	10
omphonemaceae	Gomphoneis sp.	WR	8,12,15
	Gomphoneis herculeana	SY,WR	7,13
	Gomphoneis herculeana robusta	WR	8
	Gomphonema sp.	CF,CL,LU,MF,PM,SA,SY, TU,WR	1,2,3,4,5,6,8,9,10,11 12,13,15
	Gomphonema acuminatum	TU,WR	3,5
	Gomphonema angustatum	CL,SA,SY,TU,WR	1,3,4,5,10,13
	Gomphonema constrictum	TU	3
	Gomphonema gracile	SY,TU	3,10
	Gomphonema olivaceum	WR	5
	Gomphonema parvulum	PM,SA,TU,WR	3,5,9,13
	Gomphonema simus	TU	3
	Gomphonema subclavatum	SA,WR	5,13
	Gomphonema tenellum	PM,SA,WR	9,13
	Gomphonema truncatum capitatum	WR	8
	Gomphonema ventricosum	WR	5

Family	Genus-Species	Subbasin	Source
	PHYLUM: Bacil (Diatoms)—Co		
Naviculaceae	Amphiprora sp.	LU	11
	Amphiprora ornata	TU	2,3
	Caloneis sp.	LU,SY	10,11
	Caloneis ventricosa	SY,TU	3,10
	Diatomella balfouriana	SY	10
	Diploneis elliptica	SY,TU	3,10
	Diploneis finnica	SY	10
	Diploneis oblongata	SY	10
	Gyrosigma sp.	TU	2
	Gyrosigma accuminatum	TU	3
	Mastogloia sp.	MF,WR	6
	Navicula sp.	CF,CL,LU,MC,MF,PM,SA, SY,TU,WR	1,2,3,5,6,8,10,11,12,11 15
	Navicula capitata	WR	13
	Navicula contenta biceps	WR	5
	Navicula cryptocephala	PM,SA,WR	5,9,13
	Navicula cryptocephala veneta	WR	5,13
	Navicula decussis	SA,WR	8,13
	Navicula disputans	SY	10
	Navicula exigua	SY,TU	3,10
	Navicula gregaria	TU	3
	Navicula inflexa	SY	10
	Navicula meniscula	PM	9
	Navicula menisculus upsaliensis	WR	5
	Navicula minima	CL,WR	1,5,8
	Navicula mutica	TU,WR	3,5,13
	Navicula pelliculosa	WR	5
	Navicula placenta	SY	10
	Navicula placentula	SY	10
	Navicula pupula	PM,SA,SY,TU,WR	3,5,9,10,13
	Navicula radiosa	WR	5
	Navicula radiosa tenella	WR	8
	Navicula rhynchocephala	WR	5

Family	Genus-Species	Subbasin	Source
	PHYLUM: Baci (Diatoms)—C		
Naviculaceae—Continued	Navicula salinarum	SA,WR	13
	Navicula tripunctata	SA,WR	5,13
	Navicula viridula	TU	3
	Neidium sp.	TU	3
	Neidium affine	SY	10
	Neidium dubium	SY	10
	Neidium iridis	WR	13
	Pinnularia sp.	LU,SY,TU,WR	2,3,5,10,11,13
	Pinnularia mesolepta	SY	10
	Pinnularia microstauron	TU	3
	Pinnularia nobilis	SY	10
	Pinnularia subcapitata	SY,TU	3,10
	Pleurosigma sp.	LU	11
	Stauroneis sp.	LU,SA,SY	10,11,13
	Stauroneis anceps	SY,TU	3,10
	Stauroneis phoenicentron	SY	10
Nitzschiaceae	Nitzschia sp.	CL,LU,MC,PM,SA,SY,TU, WR	1,2,3,5,6,8,9,10,11,12 13,15
	Nitzschia acicularis	CL,TU	1,2,3,10
	Nitzschia aricularis	CL,SA,TU,WR	1,3,5,13
	Nitzschia acuta	SA,TU,WR	2,3,13
	Nitzschia amphibia	SA,SY,WR	5,10,13
	Nitzschia capitellata	TU,WR	3,5
	Nitzschia dissipata	PM,SA,SY,TU,WR	3,5,8,9,10,13
	Nitzschia filiformis	TU	2
	Nitzschia frustulum	SA,TU,WR	3,5,13
	Nitzschia frustulum perpusilla	WR	8
	Nitzschia frustulum subsalina	SA,TU,WR	3,13
	Nitzschia holsatica	WR	8
	Nitzschia linearis	SY,TU,WR	3,5,10,13
	Nitzschia minima	WR	13
	Nitzschia oregona	WR	8
	Nitzschia palea	SA,SY,TU,WR	3,5,10,13
	Nitzschia palaceae	CL,SA,SY,WR	1,4,5,13

Family	Genus-Species	Subbasin	Source
	PHYLUM: Ba (Diatoms)—		
Nitzschiaceae—Continued	Nitzschia rectra	WR	5
	Nitzschia sigma	TU	2
	Nitzschia sigmoidea	TU	2
	Nitzschia sublinearis	SY	10
Surirellaceae	Cymatopleura solea	TU	2
	Surirella sp.	LU,SY,TU	2,10,11
	Surirella angusta	TU	3
	Surirella linearis constricta	TU	3
	Surirella oregonica	SY	10
	Surirella ovata	TU	2,3
	Surirella ovata salina	TU	3
	Surirella robusta	TU	3
Tabellariaceae	Tetracyclus lacustris	SY	10
	PHYLUM: C	ryptophyta	
Cryptochrysidaceae	Chroomonas sp.	CL,WR	1,5
	Rhodomonas minuta	CL,WR	5,11
Cryptomonadaceae	Cryptomonas sp.	WR	5,6
	Cryptomonas erosa	CL,WR	1,5
	Cryptomonas ovata	WR	5
	Cryptomonas obovoidea	MF	14
	PHYLUM: Cyanophy	ta (Blue-green algae)	
Chroococcaceae	Anacystis sp.	SY	4
	Anacystis marina	WR	5
	Aphanocapsa sp.	CF,CL,MC,MF,PM,SA,WR, YH	6
	Chroococcus sp.	CF,CL,LU,MC,PM,SA,SY, TU,WR	2,4,5,6,11,13
	Chroococcus minimus	WR	5
	Chroococcus minutas	PM	9
	Gloeocapsa sp.	MF,WR	6
	Microcystis sp.	SA,TU	2,3,13
	Microcystis aeruginosa	WR	5
Nostocaceae	Anabaena sp.	CF,CL,LU,MC,MF,PM,SA, TU,WR,YH	3,5,6,9,11,12,13,14,15
	Anabaena affinis	TU	2

Family	Genus-Species	Subbasin	Source
	PHYLUM: Cyanophyta (Blue	-green algae)—Continued	
Nostocaceae—Continued	Anabaena circinalis	MF,TU	2,14
	Anabaena flos-aquae	CL	1
	Aphanizomenon flos-aquae	TU	2
	Phormidium sp.	CF,TU,WR	2,6
Oscillatoriaceae	Lyngbya sp.	CF,MC,MF,SA,WR	6,13
	Lyngbya versicolor	WR	8
	Oscillatoria sp.	CF,CL,LU,MC,MF,SA,TU,W R	2,3,5,6,8,11,13
	Oscillatoria agardhii	PM	9
	Oscillatoria limnetica	WR	13
	Spirulina sp.	TU	2
Rivulariaceae	Amphithrix janthina	PM,SA,WR	9,13
	PHYLUM: Euglenop	hyta (Euglenoids)	
Euglenaceae	Euglena sp.	LU,TU,WR	2,3,5,6,11,13
	Euglena acus	TU	2
	Euglena tripleria	TU	2
	Phacus sp.	CL,TU	1,2,3
	Phacus birgei	TU	2
	Trachelomonas sp.	CL,SA,TU,WR	1,2,3,5,13
	Trachelomonas acanthostoma	WR	5
	Trachelomonas charkowensis	WR	5
	Trachelomonas hispida	WR	5
	Trachelomonas lacustris	WR	5
	Trachelomonas pulchella	WR	5
	Trachelomonas robusta	WR	5
	Trachelomonas rotunda	WR	5
	Trachelomonas volvocina	CL,WR	1,5
	PHYLUM: Pyrrhophy	ta (Dinoflagellates)	
Ceratiaceae	Ceratium sp.	TU	3
	Ceratium hirundiniella	CL,MF,WR	1,5,14
Glenodiniaceae	Glenodinium sp.	CL,WR	1,13
Gymnodiniaceae	Gymnodinium sp.	CL,WR	1,13
Peridiniaceae	Peridinium sp.	TU	2,3
	Peridinium cinctum	CL,WR	1,5

[This list is based on the references listed below and includes organisms identified to Order or a lower taxonomic level. It is not representative of taxon distribution due to disproportionate sampling effort throughout the Willamette Basin. The taxa are listed as identified to the highest taxonomic level from the original source (except for Anderson and Hansen (1987) and Parsons and others (1991) which are compilations from multiple sources), and misidentification or changes in taxonomy have not been addressed. Subbasin occurrence or source are not repeated for higher taxonomic levels (e.g., a source reporting *Baetis bicaudatus* is not repeated at the Family [Baetidae] or Order [Ephemeroptera] levels for that record). Genus and species are shown in italic. Subbasins (see fig. 5): CF, Coast Fork; CL, Clackamas; LT, Long Tom; LU, Luckiamute; MA, Marys; MC, McKenzie; MF, Middle Fork; PM, Pudding/Molalla; RI, Rickreall; SA, Santiam; SY, Sandy; TU, Tualatin; WR, Willamette River. Sources: 1, Aho (1976); 2, Anderson (1992); 3, Anderson and Hansen (1987); 4, Anderson and Wold (1972); 5, Ball (1946); 6, Clifton (1985); 7, EA Engineering, Science and Technology, Inc. (1990a); 8, J. Furnish (Bureau of Land Management, written commun., 1996); 9, Hawkins and others (1982); 10, Hawkins and Sedell (1981); 11, Hjort and others (1984); 12, HMS Environmental, Inc., and Miller (1988); 13, Johnson and others (1989); 14, Kerst (1969); 15, Lehmkuhl (1969); 16, Mangum (1990); 17, Mangum (1991a); 18, Mangum (1991b); 19, Miller (1979); 20, Moore (1987); 21, Parsons and others (1991); 22, Tetra Tech, Inc. (1994); 23, TW Environmental, Inc. (1994); 24, Wisseman (1995)]

Tax	on	Subbasin	Source
CLASS: HYDROZOA (Hydroids)			
ORDER: Hydroida (Hydroids)		WR	13
CLASS: TURBELLARIA (Flatworms	s)	SY	23,24
ORDER: Tricladida		MC,SY,TU,WR	7,22,24
FAMILY: Planariidae		PM,SY,WR	12,16,19
	Planaria sp.	MC,PM,SA	8,17,18
CLASS: NEMATODA (Nematodes)		MC,PM,SA,SY,WR	7,8,12,13,16,17,18,24
CLASS: POLYCHAETA (Marine wo	rms)		
FAMILY: Nereidae	Neris limnicola	WR	22
FAMILY: Sabellidae	Manayunkia speciosa	WR	11
CLASS: OLIGOCHAETA (Worms)		LU,MC,PM,TU,SA,SY, WR	6,7,10,11,12,13,16,17,18,19, 20,22,23,24
CLASS: HIRUDINEA (Leeches)		MC,WR	7,12,13
FAMILY: Erpobdellidae	Dina sp.	WR	11
FAMILY: Glossiphonidae	Helobdella sp.	WR	11
FAMILY: Hirudinidae		MC,WR	22
CLASS: GASTROPODA (Snails)		МС	1,7
FAMILY: Ancylidae		MC,PM	7,19
	Ferrissia sp.	WR	11
	Ferrissia rivularis	MC,TU,WR	22
	Juga sp.	MC,PM,WR	8,11,13,20
	Juga plicifera	MC,TU,WR	9,10,22
	Juga silicula	MC	7
Tax	on	Subbasin	Source

Taxon		Subbasin	Source
FAMILY: Pisauridae	Dolomedes sp.	MC,WR	22
	Piona sp.	WR	11
FAMILY: Pionidae	Forelia sp.	WR	11
FAMILY: Mideopsidae	Mideopsis sp.	WR	11
FAMILY: Lebertiidae	Lebertia sp.	WR	11
FAMILY: Hygrobatidae	Attractides sp.	WR	11
FAMILY: Eylaidae	Eylais sp.	LU	8
	Aturus sp.	WR	11
FAMILY: Aturidae		WR	11
FAMILY: Arrenuridae	Arrenurus sp.	WR	11
CLASS: ARACHNIDA (Arachnids)  ORDER: Hydracarina (Water-mites)		MC,MF,PM,SA,SY,TU,WR	6,7,8,10,16,17,18,19,20, 22,23,24
CLASS: TARDIGRADA (Water bears)		MC	20
		MC	7
FAMILY: Sphaeriidae		MC,PM,TU,WR	8,11,13,22
FAMILY: Margaritiferidae	Margaritifera sp.	WR	11
	Corbicula fluminea	TU,WR	22
FAMILY: Corbiculidae	Corbicula sp.	SY,WR	11,16
CLASS: BIVALVIA (Clams)		SA	18
FAMILY: Pleuroceridae	Goniobasis sp.	PM	19
	Vorticifex effusa	MC	22
	Vorticifex sp.	WR	11
	Heliosoma anceps anceps	TU,WR	22
FAMILY: Planorbidae		MC	7
FAMILY: Lymnaeidae	Lymnaea sp.	MC,SY	16,17
	Flumnicola virens	MC,TU,WR	22
FAMILY: Hydrobiidae	Fluminicola sp.	PM,SY,WR	8,11,19,24
	Physella propinqua	MC,WR	22
	Physella sp.	MC	7

Taxon		Subbasin	Source
	Deronectes griseostriatus	LU	3
	Agabus lutosus	LU	3
	Agabus lugens	LU	3
	Agabus confertus	LU	3
	Agabinus glabrellus	MC	21
	Acilius semisulcatus	MC	21
FAMILY: Dytiscidae		MC,SY	1,7,17,20,24
FAMILY: Carabidae		MC	17,18
ORDER: Coleoptera (Beetles)			
CLASS: INSECTA (Insects)			
ORDER: Ostracoda (Seed shrimp)		LU,MC,PM,SY,TU,WR	1,6,7,8,11,16,17,20,22,24
FAMILY: Ligiidae	Ligidium gracile	MC	21
FAMILY: Asellidae	Asellus sp.	TU,WR	11,22
ORDER: Isopoda (Sowbugs)		LU	8
	Pacifastacus leniusculus	MC,WR	7,10,11
FAMILY: Astacidae	Paciifastacus sp.	MC,PM	8,20
ORDER: Decapoda (Crayfish)			
FAMILY: Cyclopoda		SY	6
	Harpactacoida sp.	LU,MC,SY	6,8,20
	Cyclopoida sp.	LU	8
	Calanoida sp.	SY	6
ORDER: Copepoda (Copepods)		MC,PM,SY	1,6,7,8,10,16,17,24
FAMILY: Talitridae	Hyalella azteca	LU,WR	8,11
	Gammarus sp.	MC,TU,WR	22
	Anisogammarus sp.	WR	11
FAMILY: Gammaridae		SY	6
ORDER: Amphipoda (Scuds)		MC,SY,WR	1,7,12,13,16
CLASS: CRUSTACEA (Crustaceans)			
FAMILY: Unionicolidae	Unionicola sp.	WR	11
FAMILY: Sperchonidae	Sperchon sp.	WR	11

Taxon		Subbasin	Source
	Gyrinus plicifer	LU	3
FAMILY: Gyrinidae	Gyrinus pleuralis	LU	3
	Zaitzevia parvula	LU,MC,TU,WR	3,22
	Zaitzevia milleri	LU	3
	Zaitzevia sp.	MC,PM,SA,SY,WR	7,8,10,12,13,17,18,24
	Ordobrevia nubifera	MC	7
	Ordobrevia sp.	MC	10
	Optioservus quadrimaculatus	MC,TU,WR	22
	Optioservus sp.	LU,MC,PM,SY,WR	3,7,8,10,11,12,13,17,24
	Narpus concolor	LU,MC	3,7,22
	Narpus sp.	MC,SY,WR	10,13,17,24
	Lara avara	LU,MC,PM,SY	3,7,8,10,23,24
	Lara sp.	MC	17
	Heterlimnius koebeli	LU	3
	Heterlimnius sp.	MC,PM,SY	8,9,10,23,24
	Dubiraphia sp.	WR	11,13
	Cleptelmis ornata	MC,WR	22
	Cleptelmis sp.	LU,MC	3,10
	Ampumixis dispar	MC,SY	7,24
	Ampumixis sp.	MC	10
FAMILY: Elmidae		MC,MF,PM,SY	1,6.,8,16,17,19,20
	Oreodytes sp.	LU,MC,SY	3,6,9,10
	Laccophilus decipiens	LU	3
	Hydrovatus sp.	LU	3
	Hydroporus vilis	LU	3
	Hydroporus fortis	LU	3
	Hydroporus sp.	LU	3
	Dytiscus marginicollis	LU	3
	Dytiscus hatchi	LU	3
	Deronectes striatellus	LU	3

FAMILY: Haliplidae	Peltodytes callosus	LU	3
FAMILY: Helodidae	Cyphon brevicollis	MC	21
	Cyphon concinnus	LU,MC	3,21
	Elodes sp.	LU,MC	3,21
	Elodes angusta	MC	21
	Elodes apicalis	MC	21
FAMILY: Heteroceridae		SY	6
	Lanternarius brunneus	MC	21
FAMILY: Histeridae	Stictostix californicus	MC	21
FAMILY: Hydraenidae		SY	6
	Hydraena vandykei	LU	3
	Ochthebius rectus	LU	3
FAMILY: Hydrophilidae		MC,SY	1,24
	Ametor latus	LU	3
	Ametor scabrosus	MC	21
	Anacaena limbata	LU	3
	Crenitis sp.	MC	10
	Crentis rufiventris	MC	21
	Crenitis seriellus	LU	3
	Crenitis snoqualmie	MC	21
	Cymbiodyta dorsalis	MC	21
	Cymbiodyta imbellus	LU	3
	Cymbiodyta pacifica	LU	3
	Helophorus sp.	LU	3
	Hydrobius sp.	WR	22
	Hydrochus sp.	PM	8
	Laccobius californicus	LU	3
	Laccobius carri	MC	21
	Tropisternus sp.	LU	3
FAMILY: Psephenidae	Acneus sp.	LU,MC,PM	3,8,21
Tayon		Subbasin	Source

			<u> </u>
	Bezzia-Probezzia	SY,WR	11,16
	<i>Bezzia</i> sp.	MC,SA	17,18
	Atrichopogon epicautae	LU	3
	Atrichopogon sp.	MC,PM	8,21
FAMILY: Ceratopogonidae		MC,PM,SY	6,7,8,9,10,16,19,20,24
	Philorus californicus	MC	21
	Dioptopsis aylmeri	MC	21
	Dioptopsis sp.	SY	16
	Blepharicera ostensackeni	MC	21
	Blepharicera jordani	MC	21
	Blepharicera sp.	SY,WR	16,22
	Bibiocephala sp.	PM	8
	Agathon comstocki	LU,MC	3,21
	Agathon sp.	MC	2
FAMILY: Blephariceridae		MC,PM,SY	1,8,17,24
	Atherix sp.	LU,MC,SY	3,7,16,17
FAMILY: Athericidae (Rhagio	onidae)	SY	6
ORDER: Diptera (True flies)		MC	1
FAMILY: Sminthuridae		MC	20
FAMILY: Lophopodae	Pectinatella magnifica	TU	22
ORDER: Collembola (Springt	ails)	PM,SY	6,8
FAMILY: Tenebrionidae	Scaphidema pictum	MC	21
	Stenus maritimus	MC	21
	Neobisnius senilis	MC	21
	Dianous nitidulus	MC	21
FAMILY: Staphylinidae		SY	6
	Aegialia opaca	MC	21
	Aegialia mantanus	MC	21
	Aegialia lacustris	MC	21
FAMILY: Scarabaeidae	Aegialia blanchardi	MC	21

	Culicoides jamesi	LU	3
	Forcipomyia sp.	LU	3
	Johannsenomyia albibasis	LU	3
	Mallochohelea sybleae	LU	3
	Neurohelea nigra	LU	3
	Palpomyia sp.	WR	11
	Palpomyia aldrichi	LU	3
	Palpomyia flavipes	LU	3
	Serromyia barberi	LU	3
FAMILY: Chaoboridae		SY,WR	6,11
FAMILY: Chironomidae		MC,MF,PM,SA,SY,WR	1,6,7,8,9,11,12,16,17,18,19, 20,24
	Ablabesmyia sp.	WR	11
	Acricotopus sp.	LU	3
	Alotanypus venustus	MC	21
	Arctopelopia flavifrons	MC	21
	Boreochlus sp.	SY	24
	Boreochlus sinuaticornis	MC	21
	Boreohaptagyia sp.	SY	24
	Boreoheptagyia lurida	MC	21
	Brillia sp.	MC,SY,WR	2,11,24
	Brilla flavifrons	LU,MC	3,21
	Brilla retifinis	LU	3
	Brundiniella eumorpha	LU,MC	3,21
	Bryophaenocladius sp.	LU	3
	Cardocladius sp.	MC,TU,WR	21,22
	Chaetocladius sp.	LU,MC	3,21
	Chironomus sp.	WR	11
	Chironomus jucundus	LU	3
	Cladopelma sp.	WR	11
	Cladotanytarsus sp.	LU	3
Tayon		Subbasin	Source

Conchapelopia sp.	LU,MC	2,3
Conchapelopia currani	MC	21
Conchapelopia pallens	MC	21
Concgapelopia pilicaudata	MC	21
Corynoneura sp.	LU,MC,SY	3,21,24
Cricotopus sp.	WR	11
Cricotopus bicinctus	LU	3
Cricotopus nostocicola	MC	21
Cricotopus nostococladius	SY	24
Cricotopus tremulus	LU,MC	3,21
Cricotopus triannulatus	MC,TU,WR	22
Cryptochironomus sp.	LU,WR	3,11
Cryptotendipes sp.	MC	21
Diamesa sp.	MC	2
Diamesa chorea	MC	21
Diamesa garretti	MC	21
Diamesa greysoni	MC	21
Diamesa heteropus	LU,MC	3,21
Diamesa leoniella	MC	21
Diamesa sommermani	MC	21
Dicrotendipes sp.	TU,WR	11,22
Djalmabatista sp.	WR	11
Endochironomus sp.	TU,WR	11,22
Eukiefferiella sp.	MC,SY,WR	11,21,22,23,24
Eukiefferiella brevinervis	LU	3
Eukiefferiella brevicalcar	LU	3
Eukiefferiell claripennis	LU	3
Eukiefferiella coerulescens	LU	3
Eukiefferiella devonica	LU	3
Euryhapsis sp.	MC	21

Glyptotendipes sp.	TU,WR	22
Heleniella sp.	MC	2,21
Heleniella curtistila	LU	3
Heterotrissocladius sp.	MC	21
Heterotrissocladius marcidus	LU	3
Hydrobaenus sp.	MC	21
Krenosmittia sp.	MC	21
Krenosmittia boreoalpina	LU	3
Larsia pallens	LU	3
Larsia sequoiaensis	MC	21
Limnophyes sp.	LU,MC	3,21
Macropelopia sp.	LU,MC	3,21
Meropelopia flavifrons	LU	3
Metriocnemus sp.	MC	21
Metriocnemus aequalis	LU	3
Micropsectra sp.	MC,SY,WR	2,9,11,21,23,24
Micropsectra groenlandica	LU,MC	3,21
Micropsectra dives	LU	3
Micropsectra polita	LU	3
Microtendipes sp.	LU,SY,WR	3,22,24
Nanocladius sp.	WR	11
Nanocladius balticus	LU	3
Nanocladiusbrevinervis	LU	3
Natarsia sp.	LU	3
Nilotanypus sp.	LU	3
Orthocladius sp.	SY,LU,MC	3,21,24
Orthocladius appersoni	LU	3
Orthocladius curtiseta	LU,MC	3,21
Orthocladius dentifer	LU	3
	Subbasin	Source

Orthocladius dorenus	MC	21
Orthocladius frigidus	LU	3
Orthocladius lignicola	LU,MC	3,21
Orthocladius nigritus	MC	21
Orthocladius-Cricotopus complex	MC,SY,TU,WR	2,11,22,24
Pagastia sp.	MC,SY,WR	21,22,23,24
Pagastia partica	MC	21
Parachaetocladius hirtipectus	LU	3
Parachironomus sp.	WR	11
Paracladopelma sp.	WR	11
Paracricotopus sp,	MC	21
Parakiefferiella sp.	LU,MC	3,21
Paralauterborniella sp.	WR	11
Paramerina sp.	SY	24
Paramerina fragilis	LU	3
Parametriocnemus sp.	LU,MC,SY	2,3,21,24
Parametriocnemus lundbecki	LU	3
Paraorthocladius sp.	MC	21
Paraphaenocladius sp.	LU,SY	3,23,24
Paratanytarsus sp.	MC,SY,WR	11,21,22,24
Paratendipes sp.	MC	21
Paratendipes albimanus	LU	3
Paratrichocladius sp.	MC	21
Phaenopsectra sp.	LU,MC	3,21,22
Polypedilum sp.	LU,MC,SY,TU,WR	2,3,11,21,22,24
Polypedilum fuscipenne	LU	3
Polypedilum fallax	LU	3
Potthastia sp.	WR	11
Potthastia longimana	WR	22
Procladius sp.	LU,WR	3,11

Prodiamesa olivacea	LU	3
Psectrocladius sp.	MC,SY	9,24
Psectrotanypus dyari	MC	21
Pseudodiamesa sp.	MC	21
Pseudodiamesa diastena	LU	3
Psilometriocnemus sp.	MC	21
Psilometriocnemus triannulatus	LU	3
Radotanypus submarginella	LU	3
Rheocricotopus sp.	MC,SY,WR	11,21,24
Rheocricotopus effusus	LU	3
Rheotanytarsus sp.	LU,MC,SY,TU,WR	3,9,11,21,22,24
Stempellina sp.	LU,SY	3,24
Stempellinella sp.	LU,MC,SY,WR	3,11,21,24
Stempellinella brevis	LU	3
Stenochironomus sp.	WR	11
Stenochironomus colei	LU	3
Stilocladius sp.	LU,MC	3,21
Symposiocladius sp.	SY	24
Synorthocladius sp.	MC,SY,WR	11,21,24
Synorthocladius semivirens	LU	3
Tanytarsus sp.	MC,SY,WR	11,21,22,24
Tanytarsus eminulus	LU	3
Tanytarsus lugens	LU	3
Thienemanniella sp.	LU,MC,SY,TU,WR	3,9,11,22,24
Thienemannimyia sp.	LU,MC,SY	3,21,24
Tribelos protexus	LU	3
Tvetenia sp.	MC,SY,WR	21,22,24
Tvetenia bavarica	LU	3
Tvetenia calvescens	LU	3
Xenochironomus sp.	WR	11

	Zavrelia sp.	LU	3
	Zavrelimyia sp.	LU,MC	2,3,21
	Zavrelimyia thryptica	LU,MC	3,21
FAMILY: Culicidae		PM	19
	Aedes sierrensis	MC	21
	Culiseta sp.	SY	6
FAMILY: Deuterophlebiidae		MC	10
	Deuterophlebia inyoensis	MC	21
	Deuterophlebia coloradensis	MC	21
FAMILY: Dixidae		MC,SY	1,6
	Dixa sp.	MC,PM,SY	8,17,20,24
	Dixa arge	LU	3
	Dixa californica	LU	3
	Dixa johansenni	LU	3
	Dixa rhathyme	LU	3
	Meringodixa sp.	PM	8
FAMILY: Dolichopodidae		SY	6
	Argyra bimaculata	LU	3
	Campsicnemus claudicans	LU	3
	Campsicnemus degener	LU	3
	Dolichopus crenatus	LU	3
	Dolichopus duplicatus	LU	3
	Dolichopus grandis	LU	3
	Dolichopus nigricauda	LU	3
	Dolichopus renidescens	LU	3
	Dolichopus tenuipes	LU	3
FAMILY: Drosophilidae	Scaptomyza sp.	MC	21
FAMILY: Empididae		MC,SY	6,7,10,16,20
	Chelifera sp.	MC,PM,SY,TU,WR	7,8,17,22,23,24
	Clinocera sp.	MC,PM,SY	7,8,24
Taxon		Subbasin	Source

Taxo	on	Subbasin	Source
	Ptychoptera sp.	MC	9
	Bittacomorpha clavipes	MC	21
FAMILY: Ptychopteridae		SY	6
	Psychoda unbracola	MC	21
	Psychoda phalaenoides	MC	21
	Psychoda sp.	LU,SY	3,6
	Pericoma sp.	LU,MC,SY	3,10,16,17
	Maruina lanceolata	LU	3
FAMILY: Psychodidae	Maruina sp.	MC,PM,SY	8,10,17,24
	Glutops rossi	MC,MF,SY	16,17
	Glutops sp.	LU,PM,SY	3,8,23
FAMILY: Pelecorhynchidae		SY	23,24
FAMILY: Muscidae	Limnophora sp.	PM	8
FAMILY: Mycetophilidae	Symmerus sp.	LU	3
	Scatella paludum	MC	21
	Psilopa compta	MC	21
	Philygria opposita	MC	21
	Philygria nigrescens	MC	21
	Philygria debilis	MC	21
	Parydra sp.	MC	21
	Hydrellia griseola	MC	21
	Hydrellia sp.	LU	3
	Ditricophora argyrostoma	MC	21
FAMILY: Ephydridae		PM,WR	11,19
	Weidemannia sp.	MC	21
	Rhamphomyia sp.	MC	21
	Oreogeton sp.	PM,SY	8,24
	Hemerodromia sp.	LU,MC,SA,SY,WR	3,7,11,16,17,18,21,22
	Dolicocephala sp.	MC	21

Taxon		Subbasin	Source
	Chrysops noctifer pertinax	MC	21
	Chrysops excitans	MC	21
	Chrysops asbestos	MC	21
	Atolytus incisuralis	MC	21
FAMILY: Tabanidae		MC	10
	Xylota sp.	LU	3
FAMILY: Syrphidae	Pocota sp.	LU	3
	Twinnia nova	LU	3
	Simulium vittatum	LU	3
	Simulium tuberosum	LU	3
	Simulium pugetense	LU,MC	3,21
	Simulium piperi	LU	3
	Simulium canadense	LU	3
	Simulium arcticum	LU,MC	3,21
	Simulium sp.	MC,SY,WR	7,11,22,23,24
	Prosimulium fulvum	LU,MC	3,21
	Prosimulium esselbaughi	MC	21
	Prosimilium dicum	LU	3
	Prosimilium caudatum	LU	3
	Prosimilium sp.	MC,PM	7,8
	Parasimulium stonri	MC	21
	Parasimulium sp.	MC	21
	Cnephia minus	LU	3
FAMILY: Simuliidae		MC,MF,PM,SA,SY,WR	1,6,7,10,11,12,13,16,17, 18,19,24
	Pherbellia nana	MC	21
	Limnia sp.	MC	21
	Atrichomelina pubera	LU	3
FAMILY: Sciomyzidae		PM,SY	6,8
FAMILY: Sciaridae		SY	6
	Ptychoptera townesi	LU	3

Тахо	n	Subbasin	Source
	Dicranoptycha stenophallus	MC	21
	Dactylolabis sp.	MC	21
	Cladura macnabi	MC	21
	Chionea sp.	MC	21
	Austrolimnophila badia	LU,MC	3,21
	Antocha monticola	MC,SA,SY	16,17,18,21
	Antocha sp.	LU,MC,PM,SY,WR	3,6,7,8,10,11,12,13,17, 19,24
FAMILY: Tipulidae		MC,SY,WR	1,6,11,20,23,24
FAMILY: Thaumaleidae	Thaumalea sp.	MC,SY	21,24
FAMILY: Tanyderidae		MC	7
	Tabanus punctifer	MC	21
	Tabanus monoesis	MC	21
	Tabanus kesseli	MC	21
	Tabanus fratellus	MC	21
	Tabanus aegrotus	MC	21
	Tabanus sp.	MC	20
	Silvius gigantulus	MC	21
	Pilmas californica	MC	21
	Hybomitra zygota	MC	21
	Hybomitra sonomensis	MC	21
	Hybomitra sequax	MC	21
	Hybomitra rhombica	MC	21
	Hybomitra procyon	MC	21
	Hybomitra melanorhina	MC	21
	Hybomitra fulvilateralis	MC	21
	Hybomitra captonis	MC	21
	Hybomitra californica	MC	21
	Hybomitra atrobasis	MC	21
	Chrysops surdus	MC	21
	Chrysops proclivis	MC	21

	Subbasin	Source
Ulomorpha sp.	MC	21
Tipula fulvolineata	LU	3
Tipula aspersa	LU	3
Tipula sp.	LU,MC	2,3,21
Rhabdomastix sp.	LU,PM	3,8
Pilaria sp.	MC	21
Pedicia townesiana	MC	21
Pedicia bicomata	LU	3
Pedicia aperta	MC	21
Pedicia ampla	LU	3
Pedicia sp.	LU,MC	2,3,10
Paradelphomyia sp.	MC	21
Ormosia upsilon	LU	3
Molophilus sp.	LU	3
Lipsothrix nigrilinea	LU	3
Lipsothrix fenderi	LU,MC	3,21
Limonia sciophila	LU,MC	3,21
Limonia sp.	MC	21
Limnophila sp.	LU,MC,PM	3,8,21
Holorusia grandis	LU	3
Holorusia sp.	MC	2
Hexatoma sp.	LU,MC,MF,PM,SA,SY	3,6,7,8,10,16,17,18,20,24
Gnophomyia sp.	MC	21
Erioptera symplecta	MC	21
Erioptera oregonensis	LU	3
Erioptera cana	MC	21
Erioptera sp.	LU	3
Elliptera sp.	MC	21
Dicranota cayuga	LU	3
Dicranota sp.	LU,MC,PM,SA,SY,WR	3,6,8,10,12,13,16,17,18, 20,21,24

ORDER: Ephemeroptera (Mayflies)		MC	1
FAMILY: Baetidae		SY	6
	Baetis sp.	MC,MF,PM,SA,SY,WR	1,2,6,8,9,10,11,12,13,16,17, 18,19,20,22
	Baetis bicaudatus	LU,MA,MC,SY	3,15,21,23
	Baetis hageni	LU,MC	3,21
	Baetis insignificans	MC,WR	22
	Baetis parvus	MA,MC	9,15
	Baetis tricaudatus	LU,MA,MC,SY,TU,WR	3,7,15,21,22,24
	Centroptilum sp.	MC,SY,WR	7,9,11,22,24
	Centroptilum elsa	LU	3
	Diphetor hageni	MC,SY	21,24
	Pseudocleon sp.	MC,WR	7,11
FAMILY: Caenidae	Caenis sp.	WR	11
FAMILY: Ephemerellidae		PM,WR	8,13
	Attenella sp.	WR	12
	Attenella delantala	MC,SY	7,24
	Attenella margarita	MC,SY	21,24
	Caudatella sp.	SY	24
	Caudatella cascadia	MC,SY	21,23
	Caudatella edmundsi	MC,SY	21,24
	Caudatella heterocaudata	MC,SA	17,18,21
	Caudatella hystrix	MC,SY	7,17,21,24
	Drunella sp.	PM	8
	Drunella coloradensis	MC,SA	18,21
	Drunella coloradensis/ flavilinea	SY	23,24
	Drunella flavilinea	LU,MC	3,21
	Drunella doddsi	LU,MC,MF,SA,SY	3,7,17,18,21,23,24
	Drunella pelosa	MC	21
	Drunella spinifera	MC,SA,SY	7,18,21,23,24
Taxon		Subbasin	Source

Taxon		Subbasin	Source
	Cinygmula par	MC	21
	Cinygmula sp.	MC,PM,SA,SY	1,2,6,7,8,9,10,16,17,18, 19,20,21,23,24
	Cinygma integrum	LU,MC	3,21
	Cinygma dimicki	MC	21
	Cinygma sp.	MC,SY	6,10,16,20,22,23,24
FAMILY: Heptageniidae		MC,WR	7,12,13,20
FAMILY: Ephemeridae		SY	6
	Timpanoga hecuba	LU	3
	Serratella velmae	MC	21
	Serratella tibialis	LU,MC,MF,SA	3,17,18,21
	Serratella teresa	LU,MC	3,21
	Serratella sp.	MC,PM,WR	2,8,11,12,13
	Ephemerella tibialis	SY,WR	13,16
	Ephemerella teresa	SY	16
	Ephemerella spinifera	SY	16
	Ephemerella margarita	SY	16
	Ephemerella initera	SY	16
	Ephemerella infrequens	LU,MC	3,21
	Ephemerella inermis/ infrequens	SY	23,24
	Ephemerella inermis	MC,MF,SA,SY	7,16,17,18,24
	Ephemerella hystrix	SY	16
	Ephemerella heterocaudata	SY	16
	Ephemerella drunella	SY	6
	Ephemerella doddsi	MC,SY	10,16
	Ephemerella delantala	SY	16
	Ephemerella colorodensis	SY	16
	Ephemerella cascadia	SY	16
	Ephemerella aurivilli	MC,WR	22
	Ephemerella sp.	MC,PM,WR,SY	1,6,8,10,11,19,20,21

	Cinygmula ramaleyi	MC	21
	Cinygmula reticulata	LU,MA,MC	3,15,21
	Cinygmula uniformis	MC	21
	Epeorus sp.	MC,MF,PM,SA,SY,WR	1,2,7,8,10,12,13,16,17, 18,19,24
	Epeorus albertae	LU,MC,WR	3,22
	Epeorus deceptivus	LU,MC	3,21
	Epeorus grandis	MC,SY	7,21,24
	Epeorus hesperus	MC	21
	Epeorus iron	LU,SY	3,6
	Epeorus longimanus	LU,MC	3,21
	Heptagenia sp.	MC,MF,SA,SY,WR	7,9,11,16,17,18,24
	Ironodes sp.	MC,PM,SY	2,8,16,23,24
	Ironodes nitidus	LU,MC	3,21
	Leucrocuta sp.	WR	22
	Nixe sp.	MC,SY,WR	12,13,22,24
	Rhithrogena sp.	LU,MC,MF,PM,SY,WR	2,3,6,7,10,11,13,16,17, 19,23,24
	Rithrogenia morrisoni	LU,MC,WR	3,22
	Rithrogenia robusta	MC	21
	Stenonema sp.	MC,SY,WR	6,11,12,13,22
FAMILY: Leptophlebiidae		PM,SY	6,8
	Leptophlebia pacifica	LU	3
	Paraleptophlebia sp.	LU,MC,PM,SA,SY,WR	1,2,3,6,7,9,10,11,13,16, 17,18,19,20,23,24
	Paraleptophlebia aquilina	MC	21
	Paraleptophlebia bicornuta	LU,MC,SY,WR	3,7,9,22,24
	Paraleptophlebia debilis	LU,MA,MC	3,15,21,22
	Paraleptophlebia gregalis	LU,MC	3,21
	Paraleptophlebia heterone	MC	22
Taxon		Subbasin	Source

Taxon		Subbasin	Source
	Notonecta kirbyi	MC	21
FAMILY: Notonectidae	Notonecta sp.	LU	3
FAMILY: Mesoveliidae	Macrovelia horni	LU	3
	Limnoporus notabilis	MC	21
	Gerris remigis	LU,MC	3,21
	Gerris incognitus	LU	3
FAMILY: Gerridae	Gerris incurvatus	MC	21
FAMILY: Gelastocoridae	Gelastocoris oculatus	LU,MC	3,21
	Systelloderes grandes	MC	21
FAMILY: Enicocephalidae	Boreostolis americanus	MC	21
	Graptocorixa californica	LU	3
	Cenocorixa wileyae	MC	21
FAMILY: Corixidae	Calicorixa vulnerata	MC	21
ORDER: Hemiptera (True bugs)		PM	19
	Tricorythodes minutus	MC,SY,WR	16,22
FAMILY: Tricorythidae	Tricorythodes sp.	MC,WR	10,11,12,13
	Siphlonurus occidentalis	LU	3
	Ameletus vancouverensis	LU,MC	3,21
	Ameletus suffusus	LU,MC	3,21
	Ameletus sparsatus	MC	21
	Ameletus exquisitus	MC	21
	Ameletus connectus	LU	3
	Ameletus amador	MC	21
	Ameletus sp.	LU,MC,PM,SA,SY	1,2,3,7,8,9,10,16,17,18, 19,20,21,23,24
FAMILY: Siphlonuridae		SY	6
	Isonychia velma	MC,WR	22
FAMILY: Oligoneuriidae	Isonychia sp.	WR	13
	Paraleptophlebia vaciva	MC	21
	Paraleptophlebia temporalis	LU,MA,MC	3,15,21
	Paraleptophlebia sculleni	MC	21

Taxo	n	Subbasin	Source	
	Ischnura sp.	MC	21	
	Enallagma sp.	MC	21	
	Argia vivida	MC	21	
FAMILY: Coenagrionidae		WR	11	
	Aeshna interrupta	MC	21	
FAMILY: Aeshnidae		LU	3	
ORDER: Odonata (Dragonflies/Damselflies)				
FAMILY: Sisyridae	Climacia sp.	WR	11	
ORDER: Neuroptera (Spongilla-flies)				
	Sialis rotunda	LU	3	
	Sialis californicus	LU,MC	3,21	
FAMILY: Sialidae	Sialis sp.	MC,WR	1,7,10,11,12,13,22	
	Orohermes crepusculus	MC	7,21	
	Orohermes sp.	MC	17,20	
	Protochauliodes spenceri	LU	3	
FAMILY: Corydalidae	Dysmicohermes sp.	MC	10	
ORDER: Megaloptera (Dobsonflies)				
FAMILY: Pyralidae	Petrophila sp.	MC,PM,SY,TU,WR	7,8,11,13,16,22	
ORDER: Lepidoptera (Caterpillars)		MC,MF	17	
FAMILY: Sclaridae		SY	6	
FAMILY: Ichneumonidae	Sulcarius sp.	LU	8	
		SY	6	
ORDER: Hymenoptera (Wasps)				
	Microvelia paludicola	LU	3	
FAMILY: Veliidae	Microvelia californiensis	LU,MC	3,21	
	Saldula saltatoria	MC	21	
	Saldula pallipes	LU,MC	3,21	
	Saldula lattini	MC	21	
	Saldula comatula	MC	21	
FAMILY: Saldidae	Micracantha quadrimaculata	MC	21	

Taxon		Subbasin	Source
	Alloperla pallidula	MA	14
	Alloperla fraterna	MA,MC	5,14,21
	Alloperla fidelis	MA	5,14
	Alloperla delicata	MA,MC	5,14,21
	Alloperla coloradensis	MA	5,14
	Alloperla borealis	MA	5,14
	Alloperla sp.	MC,PM	2,9,19
FAMILY: Chloroperlidae		MC,PM,SA,SY	1,6,8,10,16,17,18,20,24
	Paracapnia oswegaptera	MC	21
	Mesocapnia projecta	MC	21
	Mesocapnia porrecta	MC	21
	Mesocapnia autumna	MC	21
	Isocapnia abbreviata	MA	5
	Eucanopsis brivicauda	LU,MA,MC	3,5,14,21
	Capnia tumida	MA	5
	Capnia promota	MA	5
	Capnia projecta	LU,MA	3,5
	Capnia pileata	LU	3
	Capnia porrecta	MA	14
	Capnia melia	MC	21
	Capnia excavata	MA,MC	5,21
FAMILY: Capniidae		MC,MF,PM,SY	7,8,10,16,17,20,23,24
ORDER: Plecoptera (Stoneflies)		MC	1
FAMILY: Petaluridae	Tanypteryx hageni	MC	21
FAMILY: Libellulidae	Sympetrum corruptum	MC	21
	Octogomphus specularis	LU,MC	3,21
	Octogomphus sp.	MC	7
	Gomphus sp.	WR	22
FAMILY: Gomphidae		MC,WR	7,12,13,17
FAMILY: Cordule-gastridae	Cordulegaster dorsalis	MC	21

	Alloperla signata	MA	5
	Hastoperla sp.	SY	6
	Hastaperla brevis	MA	5
	Hastaperla chilnualna	LU,MA	3,14
	Kathroperla sp.	PM	8,19
	Kathroperla perdita	LU,MA,MC,SY	3,14,21,24
	Paraperla sp.	PM,SY	8,24
	Paraperla frontalis	MC	21
	Plumiperla sp.	PM	8
	Plumiperla diversa	MC	21
	Suwallia sp.	PM,SA	8,18
	Suwallia autumna	MC	21
	Suwallia pallidula	MC	21
	Sweltsa sp.	MC,MF,PM,SA,SY	2,7,8,16,17,18,23,24
	Sweltsa borealis	LU,MC	3,21
	Sweltsa coloradensis	LU	3
	Sweltsa exquisita	MC	21
	Sweltsa fidelis	MC	21
	Sweltsa fraterna	LU	3
	Sweltsa oregonensis	MC	21
	Sweltsa revelstoki	MC	21
FAMILY: Leuctridae		MC,PM,SA,SY	1,8,10,16,17,18,20,24
	Despaxia sp.	MC,PM	2,8,9
	Despaxia augusta	LU,MC	3,21
	Leuctra augusta	MA	14
	Leuctra forcipata	MA	5,14
	Leuctra infuscata	MA	5,14
	Leuctra occidentalis	MA	5,14
	Leuctra sara	MA	14
	Megaleuctra complicata	MA	5
Taxon		Subbasin	Source

	Moselia sp.	MC,PM	2,8
	Moselia infuscata	MC,SY	21,23,24
	Paraleuctra sp.	MC	2
	Paraleuctra andersoni	MC	21
	Paraleuctra forcipata	LU,MC	3,21
	Paraleuctra jewetti	MC	21
	Paraleuctra occidentalis	MC	21
	Paraleuctra purcellana	MC	21
	Paraleuctra sara	LU	3
	Paraleuctra vershina	MC	21
	Perlomyia sp.	LU	3
	Perlomyia collaris	MA,MC	5,14,21
	Perlomyia utahensis	MA,MC	5,14,21
FAMILY: Nemouridae		MC,PM,SY	1,6,8,10,20,24
	Amphinemura sp.	SA,SY	16,18
	Malenka sp.	MC,PM,SY	2,7,8,9,24
	Malenka californica	LU,MC	3,21
	Malenka cornuta	LU,MC	3,21
	Neomoura sp.	PM	19
	Nemoura californica	MA	5,14
	Nemoura cinctipes	MA	5,14
	Nemoura cornuta	MA	14
	Nemoura dimicki	LU,MA	5,8
	Nemoura foersteri	MA	14
	Nemoura frigida	MA	14
	Nemoura interrupta	MA	5,14
	Nemoura obscura	LU,MA	5,8,14
	Nemoura oregonensis	LU,MA	5,8,14
	Nemoura producta	MA	14
	Ostrocerca dimicki	LU	3
Taxon		Subbasin	Source

	Ostracerca foersteri	MC	21
	Podmosta sp.	SY	16
	Podmosta obscura	LU	3
	Prostoia besametsa	MC	21
	Soyedina sp.	MC,PM	2,8
	Soyedina interrupta	LU,MC	3,21
	Soyedina producta	LU,MC	3,21
	Visoka sp.	PM,SY	8,16
	Visoka cataractae	MC,SY	21,23,24
	Zapada sp.	MC,PM,SY,WR	2,8,13,16,23,24
	Zapada cinctipes	LU,MC,MF,SA,SY	3,7,9,17,18,21,22,23,24
	Zapada columbiana	MC,SY	21,23,24
	Zapada frigida	MC,SY	21,24
	Zapada oregonensis	LU,MC,SA,SY	3,16,17,18,21,23,24
FAMILY: Peltoperlidae		MC,SY	1,6,20
	Peltoperla sp.	MC,PM	10,19
	Peltoperla brevis	MA	5,14
	Peltoperla quadrispinula	MA	14
	Soliperla sp.	PM,SY	8,24
	Soliperla campanula	MC	21
	Soliperla quadrispinula	LU	3
	Yoraperla sp.	MC,PM,SA,SY	2,8,16,17,18
	Yoraperla brevis	LU,MC,SY	3,21,23,24
	Yoraperla mariana	MC,SY	21,23
FAMILY: Perlidae		MC,PM,SA,SY,WR	1,7,8,13,16,18,20,24
	Acroneuria sp.	PM,SY	6,19
	Acroneuria californica	MA	5,14
	Acroneuria pacifica	MA	5,14
	Acroneuria theodora	MA	14
Taxon		Subbasin	Source

Calineuria sp.

MC,PM,SY

2,6,8,16

Taxon		Subbasin	Source
	Kogotus nonus	LU,MC,SY	3,16,21
	Kogotus sp.	SY	16
	Isoperla trictura	MA	5,14
	Isoperla sordida	MA	14
	Isoperla sobria	LU	3
	Isoperla mormona	LU,MA	3,5,14
	Isoperla marmorata	LU,MA	3,14
	Isoperla gravitans	MC	21
	Isoperla bifurcata	MC	21
	Isoperla ebria	MA	14
	Isoperla sp.	MC,MF,SA,PM,SY	6,7,8,16,17,18,19,23,24
	Isogenus nonus	MA	14
	Isogenus misnomus	MA	14
	Isogenus sp.	PM	19
	Frisonia picticeps	LU,MC	3,21
	Cultus sp.	LU,MC,SY	3,7,16,17
	Chernokrilus misnomus	LU,MC	3,21
	Cascadoperla trictura	LU	3
	Calliperla luctuosa	MA	14
	Arcynopteryx sp.	PM	19
FAMILY: Perlodidae		MC,PM,SY,WR	1,7,8,10,12,13,16,20,22, 24
	Hesperoperla pacifica	LU,MC,MF,SY,WR	3,7,10,16,17,21,22,24
	Hesperoperla sp.	SY,WR	12,16
	Doroneuria theodora	MC	10
	Doroneuria baumanni	MC,SY	16,21
	Doroneuria sp.	SY	16,23,24
	Classenia sabulosa	MC,WR	22
	Claasenia sp.	WR	13
	Calineuria californica	LU,MC,MF,SA,SY	3,7,9,10,17,18,21,24

Taxor	1	Subbasin	Source
FAMILY: Brachycentridae		MC,SY	1,6
	Parapsyche elsis	MC	21
	Parapsyche almota	MC	21
FAMILY: Arctopsychidae	Arctopsyche grandis	MC	21
ORDER: Trichoptera (Caddisflies)		MC	1
	Taeniopteryx maura	LU,MA	3,5
	Taenionema pallidum	MC	21
	Taenionema oregonensis	LU	3
	Taenionema nigripennis	LU,MC	3,21
	Taenionema sp.	MC,SY	2,7,16
	Doddsia occidentalis	MC	21
	Brachyptera pacifica	MA	5
	Brachyptera oregonensis	MA	5
	Brachyptera nigripennis	MA	5,14
FAMILY: Taeniopterygidae		MC,SY	1,24
	Pteronarcys princeps	LU,MA,MC,SY	3,5,14,21,23,24
	Pteronarcys californica	MC	7
	Pteronarcys sp.	MC,PM,SY,WR	2,8,10,13,19,24
	Pteronarcella regularis	LU,MA	3,5,14
FAMILY: Pteronarcyidae	Pteronarcella sp.	PM	8,19
	Skwala parallela	LU,MC,SY	3,16,21
	Skwala curvata	MC	21
	Skwala americana	MC,MF,SA	17,18
	Skwala sp.	MC,WR,SY	7,9,16,22,24
	Setvena tibialis	MC	21
	Rickera sorpta	MC	21
	Perlinodes aurea	MC,SY	21,24
	Perlinodes sp.	MC,SY	7,24
	Megarcys subtruncata	LU,MC	3,21
	Megarcys sp.	PM,SA,SY	8,16,18,23

Taxon		Subbasin	Source
	Protoptila coloma	MC	21
	Protoptila sp.	WR	11
	Glossosoma wenatchee	MC	21
	Glossosoma velona	MC	21
	Glossosoma traviatum	LU,MC	3,21
	Glossosoma pyroxum	MC	21
	Glossosoma pentium	LU,MA,MC	3,4,21
	Glossosoma oregonense	MC	21
	Glossosoma califica	MC	21
	Glossosoma sp.	MA,MC,MF,SA,SY,WR	2,4,6,7,9,11,12,13,16,17,18, 22,23,24
	Anagapetus bernea	MA,MC	4,21
	Anagapetus sp.	MC,PM	8,10
	Agapetus occidentis	MC	21
	Agapetus bifidus	LU,MA	3,4
	Agapetus sp.	MC,SY	10,24
FAMILY: Glossosomatidae		MC,PM	1,19
	Heteroplectron californicum	LU,MA,MC,SY	3,4,10,20,21,24
	Heteroplectron sp.	MC	17
FAMILY: Calamoceritidae		MC,SY	1,6
	Micrasema oregona	MC	21
	Micrasema onisca	MC	21
	Micrasema dimicki	LU,MA	3,4
	Micrasema bactro	MA,MC	4,21
	Micrasema sp.	MC,MF,PM,SA,SY	6,7,8,10,16,17,18,20,23, 24
	Brachycentrus occidentalis	MC	7
	Brachycentrus americanus	MC,SY,WR	7,21,22,24
	Brachycentrus sp.	MC,SY,WR	6,10,12,13,16,24
	Amiocentrus aspilus	MC	21
	Amiocentrus sp.	MC	10

Taxon		Subbasin	Source
	Paleagapetus nearcticus	MA,MC	4,21
	Paleagapetus sp.	PM,SY	8,16
	Ochotrichia sp.	MA,MC,SY	4,6,21
	Neotrichia okopa	MC	21
	Leucotrichia pictipes	TU,WR	22
	Leucotrichia sp.	WR	11
	Hydroptila arctia	MC	21
	Hydroptila sp.	LU,MC,SY,TU,WR	3,7,11,16,22,24
	Alisotrichia sp.	SY	16
	Agraylea saltesea	MC	9,21
	Agraylea multipunctata	MC	21
FAMILY: Hydroptilidae		MC,SY,WR	6,10,12,13
	Parapsyche elsis	MA,SY	4,16,23,24
	Parapsyche almota	LU	3
	Parapsyche sp.	MA,MC,PM,SY	4,8,10,16
	Hydropsycheoslari	LU,MC	3,21
	Hydropsyche centra	LU,MC	3,21
	Hydropsyche californica	LU	3
	Hydropsyche andersoni	MC	21
	Hydropsyche amblis	LU,MA,MC	3,4,21
	Hydropsyche sp.	MA,MC,PM,SA,SY,TU,W R	4,6,7,9,10,11,17,18,19, 22,24
	Homoplectra luchia	MC	21
	Cheumatopsyche campyla	LU	3
	Cheumatopsyche sp.	MA,MC,MF,PM,SY,TU,W R	4,6,11,12,17,19,22
	Arctopsyche grandis	MC,MF,SA,SY	7,10,17,18,22,24
	Arctopsyche sp.	PM,SY	6,16,19
FAMILY: Hydropsychidae		MC,PM,SY,WR	1,6,8,12,13,17
	Goeracea genota	MC	21
FAMILY: Goeridae	Goera archaon	MC	21
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Taxor	1	Subbasin	Source
	Dicosmoecus sp.	MC,PM,WR	10,13,19
	Cryptochia pilosa	MC	21
	Clostoeca disjuncta	MC	21
	Chyranda centralis	LU,MA	3,4
	Apatania sorex	MC	21
	Apatania sp.	MC,SY	6,20
	Allocosmoecus partitus	MC,SY	21,24
	Allocosmoecus sp.	MC	2
FAMILY: Limnephilidae		MC,SA,SY	1,6,7,16,18,24
	Triaenodes tarda	MC	21
	Oecetis inconspicua	MC	21
	Oecetis sp.	SY,WR	6,11
	Mystacides alafimbriata	MC,TU,WR	21,22
	Ceraclea cancellata	MC	21
	Ceraclea annulicornis	MC	21
	Ceraclea sp.	MC,WR	7,11,22
FAMILY: Leptoceridae		SY	6
	Lepidostoma veroda	MC	21
	Lepidostoma unicolor	LU,MA,MC	3,4,21
	Lepidostoma roafi	LU,MA,MC	3,4,21
	Lepidostoma recina	MC	21
	Lepidostoma rayneri	MC	21
	Lepidostoma quercina	LU,MA	3,4
	Lepidostoma podager	MC	21
	Lepidostoma jewetti	MC	21
	Lepidostoma hoodi	MA,MC	4,21
	Lepidostoma cinereum	LU	3
	Lepidostoma sp.	MC,PM,SA,SY,WR	2,7,8,9,10,12,13,17,18, 20,21,22,23,24
FAMILY: Lepidostomatidae		MC,SY	1,6
	Stactobiella delira	LU,MC	3,21

Dicosmoecus atripes	MC	21
Dicosmoecus gilvipes	LU,MC,WR	3,21,22
Ecclisocosmoecus sp.	PM	8
Ecclisocosmoecus scylla	MC,PM	8,21
Ecclisomyia sp.	MC,SY	6,10,20
Ecclisomyia conspersa	MC	21
Ecclisomyia maculosa	MC	21
Goera archaon	MC	7,10
Goeracea sp.	PM	8
Grammotaulius betteni	LU,MC	3,21
Halesochila taylori	LU,MC	3,21
Hesperophylax alaskensis	LU,MC	3,21
Hesperophylax incisus	LU	8
Homophylax sp.	MC,PM	8,21
Homophylax andax	MC	21
Hydatophylax hesperus	LU,MA,MC,SY	3,4,21,24
Lenarchus rho	LU,MC	3,21
Lenarchus vastus	MC	21
Limnephilus aretto	LU	3
Limnephilus externus	MC	21
Limnephilus fagus	MC	21
Limnephilus harrimani	LU,MA	3,4
Limnephilus lunonus	LU,MC	3,21
Limnephilus nogus	LU,MC	3,21
Limnephilus occidentalis	LU,MC	3,21
Limnephilus sitchensis	LU,MC	3,21
Neophylax sp.	MC,PM,SA,SY	2,8,10,16,18,24
Neophylax occidentis	MC,SY	21,24
Neophylax rickeri	LU,MA,MC	3,4,21
Neophylax splendens	LU,MA,MC,SY	3,4,21,24

Taxon Subbasin Source

	Oligophlebodes sp.	MC,SY	16,17,21,24
	Oligophlebodes mostbento	MC	21
	Onocosmoecus sp.	PM	19
	Onocosmoecus unicolor	LU,MA,MC,SY	3,4,21,24
	Pedomoecus sierra	MC	21
	Philocasca rivularis	MC	21
	Pseudostenophylax edwardsi	LU,MC	3,21
	Psychoglypha sp.	MC,PM	8,20
	Psychoglypha avigo	LU,MA,MC	3,4,21
	Psychoglypha bella	MA,MC	4,21
	Psychoglypha browni	MC	21
	Psychoglypha subborealis	LU,MC	3,21
FAMILY: Odontoceridae	Namamyia plutonis	MC	21
	Nerophilus californicus	MA	4
	Parthina linea	MC	21
FAMILY: Philopotamidae		MC,SY	1,6,10
	Dolophilodes sp.	MC,PM,SY	2,8,16,24
	Dolophilodes aequalis	MC	21
	Dolophilodes dorcus	MA,MC	4,21
	Dolophilodes novusamericanus	MC	21
	Dolophilodes pallidipes	LU,MC	3,21
	Dolophilodes sisko	MC	21
	Wormaldia sp.	PM,MC,SA,SY,WR	8,9,12,17,18,19,22,23
	Wormaldia anilla	LU,MA,MC	3,4,21
	Wormaldia gabriella	LU,MA,MC	3,4,21
	Wormaldia occidea	MC	21
FAMILY: Phryganeidae		SY	6
	Agrypnia improba	MC	21
	Ptilostomis ocellifera	LU	3
FAMILY: Poly-centropodidae		SY	6
Taxor	<u> </u>	Subbasin	Source

	Polycentropus sp.	MC,SY	9,10,16,20,24
	Polycentropus denningi	MC	21
	Polycentropus variegatus	LU,MC	3,21
FAMILY: Psychomyiidae		MC,SY	1,6,10
	Psychomyia sp.	MC,SY,WR	7,11,22,24
	Psychomyia lumina	LU,MA,MC	3,4,21
	Tinodes sp.	WR	22
	Tinodes cascadia	MC	21
FAMILY: Rhyacophilidae	Himalopsyche phryganea	MC	21
	Rhyacophila sp.	MC,PM,SY,WR	1,2,6,7,8,10,13,16,19,20,21, 23,24
	Rhyacophila acropedes	MF,SA,SY	16,17,18
	Rhyacophila angelita	LU,MC,SY	3,7,16,21
	Rhyacophila aranaudior arnaudi	MC,SY	7,21,24
	Rhyacophila betteni	MC,SY,WR	22,23,24
	Rhyacophila bifila	MC	21
	Rhyacophila blarina	LU,MC	3,21
	Rhyacophila brunnea	LU,MC,SY	3,7,21,24
	Rhyacophila colorandensis	MC	7
	Rhyacophila ecosa	MA,MC	4,21
	Rhyacophila fenderi	MA,MC	4,21
	Rhyacophila grandis	LU,MC	3,21
	Rhyacophila hyalinata	MC,SY	7,16,17,24
	Rhyacophila iranda	MA,MC,SY	4,21,23
	Rhyacophila jenniferae	MC	21
	Rhyacophila jewetti	MC	21
	Rhyacophila leechi	MC	21
	Rhyacophila malkini	MC	7
	Rhyacophila narvae	LU,MA,MC,SY	3,4,7,21,23,24
	Rhyacophila norcuta	LU,MC	3,21
Taxon		Subbasin	Source

	Rhyacophila oreta	MC	21
	Rhyacophila pellisa	MA	4
	Rhyacophila perda	MC	21
	Rhyacophilia rotunda	SY	16
	Rhyacophila sibirica	MC	7
	Rhyacophila tucula	MC	21
	Rhyacophila vaccua	LU,MA,MC,SY	3,4,16,21
	Rhyacophila vaefes	MC	21
	Rhyacophila vagrita	LU,MC,MF,SA,SY	3,16,17,18,21
	Rhyacophila valuma	MC	21
	Rhyacophila vao	LU,MA,MC	3,4,21
	Rhyacophila vedra	LU,MA,MC	3,4,21
	Rhyacophila vepulsa	SY	16
	Rhyacophila verrula	MC	2,21
	Rhyacophila vocala	LU,MC	3,21
	Rhyacophila vuzana	LU,MA,MC	3,4,21
	Rhyacophila willametta	MA,MC	4,21
FAMILY: Sericostomatidae	Gumaga sp.	MC,SY	7,16,21
FAMILY: Uenoidae	Farula malkini	MA	4
	Neothremma sp.	MC,PM,SY	6,8,10
	Neothremma didactyla	MC	21

## APPENDIX D. REFERENCES ON THE EFFECTS OF DAMS (PROPOSED OR OPERATING) ON AQUATIC HABITAT AND BIOTA, PARTICULARLY FISH, FOR THE WILLAMETTE BASIN, OREGON

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# APPENDIX D. REFERENCES ON THE EFFECTS OF DAMS (PROPOSED OR OPERATING) ON AQUATIC HABITAT AND BIOTA, PARTICULARLY FISH, FOR THE WILLAMETTE BASIN, OREGON—Continued

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[µg/g, microgram per gram. River mile: --, not available; ~, approximately. Tissue type: wb, whole-body; f, fillet. Number samples: c, composite sample; s, number of composite samples. Concentration range: ND, not detected; --, not analyzed; <, less than; quantification limit used for all USEPA (1992a) data. References: ODEQ, Oregon Department of Environmental Quality; USEPA, U.S. Environmental Protection Agency; USFWS, U.S. Fish and Wildlife Servicel

					-	Concentration range (μg/g, wet weight)									_
River	River mile	Year	Species	Tis- sue type	Number samples	α-НСН	β <b>-НСН</b>	∆-НСН	<b>Lindane</b> (γ-HCH)	Heptachlor	Heptachlor epoxide	Endo- sulfan I	Endo- sulfan II	Endosulfan sulfate	Reference
Willamette, Newberg Pool		1987	Northern squawfish	wb	3–5c	<0.0025			<0.0025	<0.0025	<0.0025				USEPA (1992a)
Willamette, Halsey		1987	Sucker	wb	4c	.00403			<.0025	<.0025	<.0025				USEPA (1992a)
Willamette	~7	1987	Sucker	wb	4c	.00717			.0186	<.0025	<.0025				USEPA (1992a)
Willamette	7	1990	Northern squawfish	wb	3	<.006	< 0.006	< 0.006	<.006	<.006	<.006	< 0.006	<0.006	<0.006	ODEQ (1994b)
Willamette	7	1990	Northern squawfish	wb	1	.004	<.002		<.002	<.002	<.002	<.002		<.002	Curtis and others (1993)
Willamette	7	1990	Common carp	f	3–5	<.002	<.002		<.002	.0023±.0004	<.002	<.002		<.002	Curtis and others (1993)
Willamette	7	1989– 90	Common carp	f	6	<.003	<.003	<.003	<.003	<.003	<.003	<.003	<.003	<.002026	ODEQ (1994b)
Willamette	7	1988	Common carp	f	5c,2s	<.003	<.003		<.003	<.003007	<.003				ODEQ (1994b)
Willamette	18	1989	Common carp	f	4c	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	ODEQ (1994b)
Willamette	18	1989	Sucker	f	3c	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	ODEQ (1994b)
Willamette	~26	1970	Common carp	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1970	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1970	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1971	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1971	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1971	Northern squawfish	wb	3–5c					.0					Schmitt and others (1981)

					-	Concentration range (μg/g, wet weight)									
River	River mile	Year	Species	Tis- sue type	Number samples	α-НСН	β <b>-НСН</b>	∆-НСН	<b>Lindane</b> <b>(</b> γ-HCH)	Heptachlor	Heptachlor epoxide	Endo- sulfan I	Endo- sulfan II	Endosulfan sulfate	Reference
Willamette	~26	1971	Northern squawfish	wb	3–5c					0.0					Schmitt and others (1981)
Willamette	~26	1972	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1972	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1972	Channel catfish	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1972	Northern squawfish	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1973	Common carp	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1973	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1973	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1973	Northern squawfish	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1974	Common carp	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1974	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1974	Largescale sucker	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1974	Northern squawfish	wb	3–5c					.0					Schmitt and others (1981)
Willamette	~26	1976	Smallmouth bass	wb	3–5c	0.01			0.02	.01					Schmitt and others (1983)
Willamette	~26	1976	Chiselmouth	wb	3–5c	.16			.02	.00					Schmitt and others (1983)
Willamette	~26	1976	Chiselmouth	wb	3–5c	.08			.02	.00					Schmitt and others (1983)

					-	Concentration range (μg/g, wet weight)										
River	River mile	Year	Species	Tis- sue type	Number samples	α-НСН	β-НСН	∆-НСН	Lindane (γ-HCH)	Heptachlor	Heptachlor epoxide	Endo- sulfan I	Endo- sulfan II	Endosulfan sulfate	Reference	
Willamette	~26	1978	Northern squawfish	wb	3–5c	0.00		-	0.00	0.00					Schmitt and others (1983)	
Willamette	~26	1978	Chiselmouth	wb	3–5c	.00			.01	.00					Schmitt and others (1983)	
Willamette	~26	1978	Chiselmouth	wb	3–5c	.00			.01	.00					Schmitt and others (1983)	
Willamette	~26	1980	Largescale sucker	wb	3–5c	.00			.00	.01					Schmitt and others (1985)	
Willamette	~26	1980	Largescale sucker	wb	3–5c	.00			.00	.00					Schmitt and others (1985)	
Willamette	~26	1980	Northern squawfish	wb	3–5c	.00			.00	.00					Schmitt and others (1985)	
Willamette	~26	1984	Northern squawfish	wb	3–5c	<.01			<.01	<.01					Schmitt and others (1990)	
Willamette	~26	1984	Peamouth	wb	3–5c	<.01			<.01	<.01					Schmitt and others (1990)	
Willamette	~26	1984	Peamouth	wb	3–5c	<.01			<.01	<.01					Schmitt and others (1990)	
Willamette	27	1988	Bass	f	3c	<.005	< 0.005		<.005	<.005	< 0.005				ODEQ (1994b)	
Willamette	27	1988	Common carp	f	3c,2s	<.003	<.003		<.003	<.003	<.003				ODEQ (1994b)	
Willamette	27	1988	Northern squawfish	f	3c	<.004	<.004		<.004	<.004	<.004				ODEQ (1994b)	
Willamette	28	1989	Common carp	f	3	.004	<.002	0.002005	<.002002	<.002005	<.002006	<0.002002	< 0.002	<0.002019	ODEQ (1994b)	
Willamette	28	1989	Common carp	liver	2	<.002	<.002	<.002	<.002	<.002068	<.002	<.002-0.148	<.002	<.002	ODEQ (1994b)	
Willamette	38	1989	Sucker	f	5c	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	ODEQ (1994b)	
Willamette	38	1988– 89	Northern squawfish	f	5c,2s	<.008	<.008	<.002	<.008	<.008	<.008	<.002	<.002	<.002	ODEQ (1994b)	
Willamette	38	1988	Common carp	f	3c	<.003	<.003		<.003	<.003	<.003				ODEQ (1994b	
Willamette	48	1988– 89	Common carp	f	3	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	ODEQ (1994b)	

						Concentration range (μg/g, wet weight)										
River	River mile	Year	Species	Tis- sue type	Number samples	α-НСН	<b>β-НСН</b>	Δ-HCH	<b>Lindane</b> (γ-HCH)	Heptachlor	Heptachlor epoxide	Endo- sulfan I	Endo- sulfan II	Endosulfan sulfate	Reference	
Willamette	48	1989	Common carp	liver	2	< 0.004	0.003-<.004	< 0.004	< 0.004	< 0.004	< 0.004	< 0.004	< 0.004	< 0.004	ODEQ (1994b)	
Willamette	48	1988	Northern squawfish	f	5c	<.003	<.003		<.003	<.003	<.003				ODEQ (1994b)	
Willamette	72	1990	Common carp	f	1	<.002	<.002		<.002	.002	<.002	<.002		<.002	Curtis and others (1993)	
Willamette	74	1990	Northern squawfish	wb	3	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	ODEQ (1994b)	
Willamette	74	1990	Common carp	f	6	<.003	<.003	<.003	<.003	.002-<.003	<.003	<.002004	<.003	<.003	ODEQ (1994b)	
Willamette	74	1990	Common carp	liver	3	<.003039	<.003006	<.003	<.003045	<.003031	<.003	<.003	<.003	<.003	ODEQ (1994b)	
Willamette	131	1990	Northern squawfish	wb	3	<.002	<.002006	<.003	<.002	<.002	<.002	<.002002	<.002	<.002	ODEQ (1994b)	
Willamette	131	1990	Common carp	f	3	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	ODEQ (1994b)	
Willamette	147	1990	Northern squawfish	wb	3	<.002	<.002002	<.002	<.002	<.002002	<.002	<.002	<.002	<.002	ODEQ (1994b)	
Willamette	147	1990	Cutthroat trout	wb	5	<.002	<.002	<.002	<.002	<.002005	<.002002	.002-<.003	<.002	<.002	ODEQ (1994b)	
Willamette	148	1990	Northern squawfish	wb	1	<.002	.002		<.002	<.002	<.002	<.002		<.002	Curtis and others (1993)	
Willamette	148	1990	Cutthroat trout	wb	1–5	<.002	<.002		<.002	.0033	<.002	<.002		<.002	Curtis and others (1993)	
Willamette	148	1990	Cutthroat trout	wb	1	<.002	<.002		<.002	<.002	.002	<.002		<.002	Curtis and others (1993)	
Willamette	148	1990	Cutthroat trout	wb	1–5	<.002	<.002		<.002	<.002	<.002	.0020±.0014		<.002	Curtis and others (1993)	
Willamette	160	1990	Cutthroat trout	wb	1–5	<.002	<.002		<.002	.0055±.0022	<.002	<.002		<.002	Curtis and others (1993)	
Willamette	160	1990	Cutthroat trout	wb	1	<.002	<.002		<.002	<.002	.002	<.002		<.002	Curtis and others (1993)	
Willamette	161	1990	Northern squawfish	wb	3	<.002	<.002	<.002	<.002	<.002	<.002	<.002002	<.002	<.002	ODEQ (1994b)	
Willamette	161	1990	Cutthroat trout	wb	5	<.002	<.002	<.002006	<.002	<.002008	<.002002	<.002	<.002	<.002	ODEQ (1994b)	

					_	Concentration range (μg/g, wet weight)									_
River	River mile	Year	Species	Tis- sue type	Number samples	α-НСН	β <b>-НСН</b>	∆-НСН	<b>Lindane</b> (γ-HCH)	Heptachlor	Heptachlor epoxide	Endo- sulfan I	Endo- sulfan II	Endosulfan sulfate	Reference
Willamette	195	1990	Cutthroat trout	wb	1–5	< 0.002	<0.002		<0.002	0.0055±.0022	<0.002	<0.002		<0.002	Curtis and others (1993)
Middle Fork Willamette	8	1990	Northern squawfish	wb	3	<.002	<.002	0.002	<.002	<.002	<.002	<.002	0.002	<.002	ODEQ (1994b)
Middle Fork Willamette	8	1990	Cutthroat trout	wb	3	<.002	<.002	.002	<.002	<.002007	<.002	<.002	.002	<.002	ODEQ (1994b)
Santiam	0.5	1988	Northern squawfish	f	1	<.004	<.004		<.004	<.004	<.004				ODEQ (1994b)
Tualatin	8	1989	Sucker	f	1	<.003	.005	<.003	<.003	<.003	<.003	<.003	<.003	<.003	ODEQ (1994b)
Tualatin, Cherry Grove		1987	Crayfish	wb	1	<.0025			<.0025	<.0025	<.0025				USEPA (1992a)
Tualatin, Cook Park		1987	Sucker	wb	3–5c	<.0025			.00934	<.0025	<.0025				USEPA (1992a)
Yamhill	5	1989	Sucker	f	1	<.002	<.002	.002	<.002	<.002	<.002	<.002	.002	<.002	ODEQ (1994b)
Conser Slough <sup>1</sup>	0.1	1989	Sucker	f	1	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	ODEQ (1994b)
Conser Slough	0.1	1989	Northern squawfish	f	2	<.003	<.003	<.003	<.003	<.003	<.003	<.003	<.003	<.003	ODEQ (1994b)
Conser Slough	0.1	1988	Bass	f	2c	<.003	<.003		<.003	.005	<.003				ODEQ (1994b)
Conser Slough	0.1	1988	Common carp	f	4c	<.003	<.003		<.003	.005	<.003				ODEQ (1994b)
Johnson Creek	( <sup>2</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	ODEQ (1994b)
Johnson Creek	( <sup>3</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	ODEQ (1994b)
Johnson Creek	6.1	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	ODEQ (1994b)
Johnson Creek	8.3	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	ODEQ (1994b)
Johnson Creek	( <sup>4</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	ODEQ (1994b)
Johnson Creek	16.9	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	ODEQ (1994b)
Johnson Creek	( <sup>5</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	ODEQ (1994b)

						Concentration range (μg/g, wet weight)									
River	River mile	Year	Species	Tis- sue type	Number samples	α-НСН	β <b>-НСН</b>	∆-НСН	Lindane (γ-HCH)	Heptachlor	Heptachlor epoxide	Endo- sulfan I	Endo- sulfan II	Endosulfan sulfate	Reference
Johnson Creek	-( <sup>6</sup> )	1991	Crayfish	wb	1	< 0.0025	< 0.0025	< 0.0025	< 0.0025	< 0.0025	< 0.0025	< 0.0025	< 0.0025	< 0.0025	ODEQ (1994b)
Rock Creek <sup>7</sup>	1.5	1994	Sculpin	wb	15-20c	<.01				<.01					USFWS (1994c)
Rock Creek	1.7– 2	1994	Three-spined stickleback	wb	9c	<.01				<.01					USFWS (1994c)
Rock Creek	1.7– 2	1994	Sculpin	wb	6c	<.01				<.01					USFWS (1994c)

<sup>&</sup>lt;sup>1</sup>Conser Slough is a tributary of the Willamette River near Albany, Oregon (Willamette River Subbasin).

<sup>&</sup>lt;sup>2</sup>McLoughlin Boulevard.

<sup>&</sup>lt;sup>3</sup>44th Avenue and Umatilla Street.

<sup>&</sup>lt;sup>4</sup>Jenne Road.

<sup>&</sup>lt;sup>5</sup>Orient Drive.

<sup>&</sup>lt;sup>6</sup>145th Avenue.

<sup>&</sup>lt;sup>7</sup>Rock Creek is a tributary of the Tualatin River near Sherwood, Oregon (Tualatin Subbasin).

[µg/g, microgram per gram. River mile: --, not available; ~, approximately. Tissue type: wb, whole-body; f, fillet. Number samples: c, composite sample; s, number of composite samples. Concentration range: ND, not detected; --, not analyzed; <, less than; DDE, dichlorodiphenyldichloroethylene; DDD, dichlorodiphenyldichloroethane; DDT, dichlorodiphenyltrichloroethane; quantification limit used for all USEPA (1992a) data. References: ODEQ, Oregon Department of Environmental Quality; USEPA, U.S. Environmental Protection Agency; USFWS, U.S. Fish and Wildlife Service]

#### Concentration range (µg/g, wet weight)

	River	River mile	Year	Species	Tissue type	Number samples	Aldrin	Dieldrin	Endrin	Endrin alde- hyde	p,p'-DDE	<i>p,p'</i> -DDD	p,p'-DDT	Methoxy- chlor	Chlor- dane	Toxa- phene	Reference
	Willamette Newberg Pool		1987	Northern squawfish	wb	3–5c		<0.0025	< 0.0025		0.0435			< 0.0025			USEPA (1992a)
	Willamette Halsey		1987	Sucker	wb	4c		<.0025	<.0025		.0358			.0056			USEPA (1992a)
	Willamette	~7	1987	Sucker	wb	4c		<.0025	<.0025		.0371			<.0025			USEPA (1992a)
	Willamette	7	1990	Northern squawfish	wb	3	< 0.006	<.006	<.006	< 0.006	<.002- .052	<0.002	< 0.002	<.002	< 0.075	< 0.075	ODEQ (1994b)
	Willamette	7	1989– 90	Common carp	f	6	<.003	<.003	<.003	<.003	<.002- .066	<.002- .063	<.002- .019	<.003	<.03		ODEQ (1994b)
2	Willamette	7	1988	Common carp	f	5c,2s	<.003	<.003	<.003		.012	.004009	<.003	<.003	<.003		ODEQ (1994b)
5	Willamette	18	1989	Common carp	f	4c	<.002	<.002	<.002	<.002	.028	.018	.009	<.002	<.025		ODEQ (1994b)
	Willamette	18	1989	Sucker	f	3c	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.025		ODEQ (1994b)
	Willamette	~26	1970	Common carp	wb	3–5c		.07	.0		.34	.35	.11				Schmitt and others (1981)
	Willamette	~26	1970	Largescale sucker	wb	3–5c		.04	.0		.57	.72	.81				Schmitt and others (1981)
	Willamette	~26	1970	Largescale sucker	wb	3–5c		.04	.0		.64	.77	.44				Schmitt and others (1981)
	Willamette	~26	1971	Largescale sucker	wb	3–5c		.01	.0		.25	.32	.21			.0	Schmitt and others (1981)
	Willamette	~26	1971	Largescale sucker	wb	3–5c		.02	.0		.25	.35	.18			.0	Schmitt and others (1981)
	Willamette	~26	1971	Northern squawfish	wb	3–5c		.01	.0		.37	.41	.14			.0	Schmitt and others (1981)
	Willamette	~26	1971	Northern squawfish	wb	3–5c		.01	.0		.33	.24	.21			.0	Schmitt and others (1981)

									Conce	entration ran	ige (μg/g, we	t weight)				
River	River mile	Year	Species	Tissue type	Number samples	Aldrin	Dieldrin	Endrin	Endrin alde- hyde	p,p'-DDE	p,p'-DDD	p,p'-DDT	Methoxy-	Chlor- dane	Toxa- phene	Reference
Willamette	~26	1972	Largescale sucker	wb	3–5c		0.02	0.0		0.40	0.16	0.00			0.0	Schmitt and others (1981)
Willamette	~26	1972	Largescale sucker	wb	3–5c		.00	.0		.50	.29	.51			.0	Schmitt and others (1981)
Willamette	~26	1972	Channel catfish	wb	3-5c		.06	.0		.57	.28	.15			.0	Schmitt and others (1981)
Willamette	~26	1972	Northern squawfish	wb	3–5c		.02	.0		.57	.13	.00			.0	Schmitt and others (1981)
Willamette	~26	1973	Common carp	wb	3–5c		.00	.0		.35	.00	.00			.0	Schmitt and others (1981)
Willamette	~26	1973	Largescale sucker	wb	3–5c		.00	.0		.31	.15	.00			.0	Schmitt and others (1981)
Willamette	~26	1973	Largescale sucker	wb	3–5c		.00	.0		.21	.11	.00			.0	Schmitt and others (1981)
Willamette	~26	1973	Northern squawfish	wb	3–5c		.00	.0		.53	.14	.00			.0	Schmitt and others (1981)
Willamette	~26	1974	Common carp	wb	3–5c		.03	.0		.1988	.33	.00			.0	Schmitt and others (1981)
Willamette	~26	1974	Largescale sucker	wb	3–5c		.00	.0		.15	.03	.02			.0	Schmitt and others (1981)
Willamette	~26	1974	Largescale sucker	wb	3–5c		.04	.0		.50	.15	.17			.0	Schmitt and others (1981)
Willamette	~26	1974	Northern squawfish	wb	3–5c		.03	.0		.19	.06	.00			.0	Schmitt and others (1981)
Willamette	~26	1976	Smallmouth bass	wb	3–5c		.04	.00		.06	.03	.02			.00	Schmitt and others (1983)
Willamette	~26	1976	Chisel- mouth	wb	3–5c		.02	.00		.07	.07	.00			.00	Schmitt and others (1983)
Willamette	~26	1976	Chisel- mouth	wb	3–5c		.02	.00		.12	.04	.00			.00	Schmitt and others (1983)

Concentration range (µg/g, wet weight)

											-9- (F-9-9,	<u> </u>				_
River	River mile	Year	Species	Tissue type	Number samples	Aldrin	Dieldrin	Endrin	Endrin alde- hyde	p,p'-DDE	p,p'-DDD	p,p'-DDT	Methoxy- chlor	Chlor- dane	Toxa- phene	Reference
Willamette	~26	1978	Northern squawfish	wb	3–5c		0.00	0.00		0.42	0.00	0.12			0.00	Schmitt and others (1983)
Willamette	~26	1978	Chisel- mouth	wb	3–5c		.02	.00		.09	.05	.00			.00	Schmitt and others (1983)
Willamette	~26	1978	Chisel- mouth	wb	3–5c		.02	.00		.09	.06	.00			.00	Schmitt and others (1983)
Willamette	~26	1980	Largescale sucker	wb	3–5c		.02	.00		.15	.05	.01	0.00		.0	Schmitt and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c		.01	.00		.21	.05	.02			.1	Schmitt and others (1985)
Willamette	~26	1980	Northern squawfish	wb	3–5c		.01	.00		.28	.03	.00	.00		.1	Schmitt and others (1985)
Willamette	~26	1984	Northern squawfish	wb	3–5c		<.01	<.01		.13	.02	.01			<.1	Schmitt and others (1990)
Willamette	~26	1984	Peamouth	wb	3–5c		.01	<.01		.03	.01	.01			<.1	Schmitt and others (1990)
Willamette	~26	1984	Peamouth	wb	3–5c		<.01	<.01		.03	.01	<.01			<.1	Schmitt and others (1990)
Willamette	27	1988	Bass	f	3c	< 0.005	<.005	<.005		<.005	<.005	<.005	<.005	< 0.005		ODEQ (1994b)
Willamette	27	1988	Common carp	f	3c,2s	<.003	<.003	<.003		.013- .073	<.003- .005	<.003	<.003	<.003		ODEQ (1994b)
Willamette	27	1988	Northern squawfish	f	3c	<.004	<.004	<.004		<.004	<.004	<.004	<.004	<.004		ODEQ (1994b)
Willamette	28	1989	Common carp	f	3	<.002	<.002- .01	<.002	<0.002- .025	.061- .102	.02- .05	.01- .018	<.002	<.025		ODEQ (1994b)
Willamette	28	1989	Common carp	liver	2	<.002	.086– .352	<.002- .061	<.002- .088	.127– .266	.141– .144	.092– .216	<.002- .832	<.025		ODEQ (1994b)
Willamette	38	1989	Sucker	f	5c	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	< 0.002	ODEQ (1994b)
Willamette	38	1988- 89	Northern squawfish	f	5c,2s	<.008	<.008	<.008	<.002	<.008	<.008	<.008	<.008	<.008	<.002	ODEQ (1994b)

									Conce	ntration rar	nge (μg/g, we	t weight)				
River	River mile	Year	Species	Tissue type	Number samples	Aldrin	Dieldrin	Endrin	Endrin alde- hyde	p,p'-DDE	<i>p,p'</i> -DDD	p,p'-DDT	Methoxy- chlor	Chlor- dane	Toxa- phene	Reference
Willamette	38	1988	Ccommon carp	f	3c	<0.003	<0.003	<0.003		< 0.007	<0.003	<0.003	< 0.003	<0.003		ODEQ (1994b)
Willamette	48	1988– 89	Common carp	f	3	<.002	<.002	<.002	<.002	<.002- .015	<.002	<.002	<.002	<.025		ODEQ (1994b)
Willamette	48	1989	Common carp	Liver	2	<.004	<.004	<.004	<.004	<.004- .063	<.004	<.004	<.004	<.04		ODEQ (1994b)
Willamette	48	1988	Northern squawfish	f	5c	<.003	<.003	<.003		.005	<.003	<.003	<.004	<.004		ODEQ (1994b)
Willamette	72	1990	Common carp	f	1	<.002	<.002	<.002	<.002	<.002	.010	<.002	<.002	<.025	<0.025	Curtis and others (1993)
Willamette	74	1990	Northern squawfish	wb	3	<.002	<.002	<.002	<.002	.022- .042	<.002	<.002	<.002	<.025	<.025	ODEQ (1994b)
Willamette	74	1990	Common carp	f	6	<.002- .02	<.003	<.003	<.003	.007– .047	<.002- .013	<.002- .01	<.003	<.003	<.025	ODEQ (1994b)
Willamette	74	1990	Common carp	Liver	3	<.002- .103	<.003	<.003	<.003- .109	<.003- .073	<.003- .055	<.003	<.003- .069	<.03		ODEQ (1994b)
Willamette	131	1990	Northern squawfish	wb	3	<.002	<.002	<.002	<.002	.015- .022	<.002- .008	<.002	<.002	<.025	<.025	ODEQ (1994b)
Willamette	131	1990	Northern squawfish	wb	1-5	<.002	<.002	<.002	<.002	<.002	.0043 ±.0023	<.002	<.002	<.025	<.025	Curtis and others (1993)
Willamette	131	1990	Common carp	f	3	<.002	<.002	<.002	<.002	<.002- .008	<.002	<.002	<.002	<.025	<.025	ODEQ (1994b)
Willamette	147	1990	Northern squawfish	wb	3	<.002- .004	<.002	<.002	<.002	.017- .044	<.002- .002	<.002	<.002	<.025- .025	<.025- .025	ODEQ (1994b)
Willamette	147	1990	Cutthroat trout	wb	5	<.002	<.002- .002	<.002- .002	<.002	<.002- .006	<.002- .002	<.002- .005	<.002	<.025	<.025	ODEQ (1994b)
Willamette	148	1990	Northern squawfish	wb	1	.004	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.025	<.025	Curtis and others (1993)
Willamette	148	1990	Northern squawfish	wb	1	<.002	<.002	<.002	<.002	<.002	.002	<.002	<.002	<.025	<.025	Curtis and others (1993)

									Conce	entration rar	nge (μg/g, we	t weight)				
River	River mile	Year	Species	Tissue type	Number samples	Aldrin	Dieldrin	Endrin	Endrin alde- hyde	p,p'-DDE	p,p'-DDD	p,p'-DDT	Methoxy- chlor	Chlor- dane	Toxa- phene	Reference
Willamette	148	1990	Cutthroat trout	wb	1	< 0.002	0.002	<0.002	< 0.002	0.022	<0.002	<0.002	< 0.002	<0.025	<0.025	Curtis and others (1993)
Willamette	160	1990	Cutthroat trout	wb	1–5	<.002	.0023 ±.0004	<.002	<.002	<.002	<.002	<.002	<.002	<.025	<.025	Curtis and others (1993)
Willamette	160	1990	Cutthroat trout	wb	1	<.002	<.002	<.002	<.002	.0022	<.002	<.002	<.002	<.025	<.025	Curtis and others (1993)
Willamette	161	1990	Northern squawfish	wb	3	<.002	<.002	<.002	<.002	<.002- .022	<.002	<.002	<.002	<.025	<.025	ODEQ (1994b)
Willamette	161	1990	Cutthroat trout	wb	5	<.002	<.002- .003	<.002- .002	<.002	<.002- .023	<.002- .003	<.002- .007	<.002	<.025	<.025	ODEQ (1994b)
Willamette	195	1990	Cutthroat trout	wb	1	<.002	<.002	.002	<.002	<.002	<.002	<.002	<.002	<.025	<.025	Curtis and others (1993)
Middle Fork Willamette	8	1990	Northern squawfish	wb	3	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.025	<.025	ODEQ (1994b)
Middle Fork Willamette	8	1990	Cutthroat trout	wb	3	<.002	<.002	<.002- .002	<.002	<.002	<.002	<.002	<.002	<.025	<.025	ODEQ (1994b)
Santiam	0.5	1988	Northern squawfish	f	1	<.004	<.004	<.004	<.004		<.004	<.004	<.004	<.004		ODEQ (1994b)
Tualatin, Che Grove	ггу	1987	Crayfish	wb	1		<.0025	<.0025		.00058			<.0025			USEPA (1992a)
Tualatin, Che Grove	ггу	1987	Sucker	wb	3–5c		.0373	<.0025		.463			<.0025			USEPA (1992a)
Tualatin	8	1989	Sucker	f	1	<.003	<.003	<.003	<.003	.009	.037	<.003	<.003	<.003		ODEQ (1994b)
Yamhill	5	1989	Sucker	f	1	<.002	<.002	<.002	<.002	.004	.006	<.002	<.002	<.025		ODEQ (1994b)
Conser Sloug	h <sup>1</sup> 0.1	1989	Sucker	f	1	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.002	<.025		ODEQ (1994b)
Conser Sloug	h 0.1	1989	Northern squawfish	f	2	<.003	<.003	<.003	<.003	<.003	<.003	<.003	<.003	<.03		ODEQ (1994b)
Conser Sloug	h 0.1	1988	Bass	f	2c	<.003	.004	.004		<.003	<.003	<.003	<.003	<.003		ODEQ (1994b)
Conser Sloug	h 0.1	1988	Common carp	f	4c	<.003	.004	.004		<.003	<.003	<.003	<.003	<.003		ODEQ (1994b)

										Conce	ntration ran	ıge (μg/g, we	t weight)				
	River	River mile	Year	Species	Tissue type	Number samples	Aldrin	Dieldrin	Endrin	Endrin alde- hyde	p,p'-DDE	p,p'-DDD	p,p'-DDT	Methoxy- chlor	Chlor- dane	Toxa- phene	Reference
	Johnson Creek	(2)	1991	Crayfish	wb	1	< 0.0025	< 0.0025	< 0.0025	< 0.0025	0.011	0.0025	0.0077	< 0.01	< 0.025	< 0.6	ODEQ (1994b)
	Johnson Creek	( <sup>3</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	.05	<.0025	.019	<.01	<.025	<.6	ODEQ (1994b)
	Johnson Creek	6.1	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	.014	.0051	<.0025	<.01	<.025	<.6	ODEQ (1994b)
	Johnson Creek	8.3	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	.045	<.0025	.007	<.01	<.025	<.6	ODEQ (1994b)
	Johnson Creek	( <sup>4</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	.086	.0042	.062	<.01	<.025	<.6	ODEQ (1994b)
	Johnson Creek	16.9	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	.16	.048	.22	<.01	<.025	<.6	ODEQ (1994b)
	Johnson Creek	( <sup>5</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	.10	<.0025	.016	<.01	<.025	<.6	ODEQ (1994b)
	Johnson Creek	( <sup>6</sup> )	1991	Crayfish	wb	1	<.0025	<.0025	<.0025	<.0025	.069	<.0025	.018	<.01	<.025	<.6	ODEQ (1994b)
7	Rock Creek <sup>7</sup>	1.5	1994	Sculpin	wb	15-20c			<.01		.0319	<.01				<.05	USFWS (1994c)
л	Rock Creek	1.7– 2	1994	Three- spined stickle-back	wb	9c			<.01		.0620	<.01				<.05	USFWS (1994c)
	Rock Creek	1.7- 2	1994	Sculpin	wb	6c			<.01		.0438	<.01				<.05	USFWS (1994c)

<sup>&</sup>lt;sup>1</sup>Conser Slough is a tributary of the Willamette River near Albany, Oregon (Willamette River Subbasin).

<sup>&</sup>lt;sup>2</sup>McLoughlin Boulevard.

<sup>&</sup>lt;sup>3</sup>44th Avenue and Umatilla Street.

<sup>&</sup>lt;sup>4</sup>Jenne Road.

<sup>&</sup>lt;sup>5</sup>Orient Drive.

<sup>&</sup>lt;sup>6</sup>145th Avenue.

<sup>&</sup>lt;sup>7</sup>Rock Creek is a tributary of the Tualatin River near Sherwood, Oregon (Tualatin Subbasin).

[µg/g, microgram per gram. River mile:--, not available; ~, approximately. Tissue type: wb, whole-body; f, fillet. Number samples: c, composite sample. Concentration range: HCB, hexachlorobenzene; PCA, pentachloroanisole; --, not analyzed; <, less than; quantification limit used for all USEPA (1992a) data. References: USEPA, U.S. Environmental Protection Agency; USFWS, U.S. Fish and Wildlife Service]

									Cor	ncentration	n range (μg/g,	wet weigl	nt)			
	River	River mile	Year	Species	Tissue type	Number samples	Dacthal	НСВ	cis-Chlor- dane	trans- Chlor- dane	<i>cis</i> -Nona- chlor	<i>trans</i> - Nona- chlor	Oxychlor- dane	Mirex	PCA	Reference
	Willamette Newberg Pool		1987	Northern squawfish	wb	3–5c		<0.0025	<0.0025	<0.0025	<0.0025	<0.0025	<0.0025	< 0.0025		USEPA (1992a)
	Willamette Halsey		1987	Sucker	wb	4c		<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025		USEPA (1992a)
	Willamette	~7	1987	Sucker	wb	4c		<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025		USEPA (1992a)
	Willamette	~26	1976	Smallmouth bass	wb	3–5c		.00	.04	.01	.00	.00				Schmitt and others (1983)
	Willamette	~26	1976	Chiselmouth	wb	3–5c		.00	.06	.02	.00	.00				Schmitt and others (1983)
156	Willamette	~26	1976	Chiselmouth	wb	3–5c		.01	.03	.01	.00	.02				Schmitt and others (1983)
	Willamette	~26	1978	Northern squawfish	wb	3–5c	0.00	.00	.04	.01	.03	.05	.00			Schmitt and others (1983)
	Willamette	~26	1978	Chiselmouth	wb	3–5c	.00	.00	.03	.01	.01	.02	.00			Schmitt and others (1983)
	Willamette	~26	1978	Chiselmouth	wb	3–5c	.00	.00	.03	.01	.01	.02	.00			Schmitt and others (1983)
	Willamette	~26	1980	Largescale sucker	wb	3–5c	.00	.01	.02	.01	.01	.03	.00	.00	0.05	Schmitt and others (1985)
	Willamette	~26	1980	Largescale sucker	wb	3–5c	.00	.00	.01	.01	.01	.02	.00	.01	.02	Schmitt and others (1985)
	Willamette	~26	1980	Northern squawfish	wb	3–5c	.00	.00	.01	.01	.01	.03	.00	.00	.02	Schmitt and others (1985)

	lamette ~26 1984 Norsqu  lamette ~26 1984 Pea  lamette ~26 1984 Pea  latin, Cherry 1987 Cra  ve  latin, 1987 Suc  sk Creek 1 1.5 1994 Scu  kk Creek 1.7- 1994 Thr							Con	centration	range (μg/g,	wet weigl	nt)			
River		Year	Species	Tissue type	Number samples	Dacthal	нсв	cis-Chlor- dane	<i>trans</i> - Chlor- dane	cis-Nona- chlor	trans- Nona- chlor	Oxychlor- dane	Mirex	PCA	Reference
Willamette	~26	1984	Northern squawfish	wb	3–5c	<0.01	<0.01	0.01	< 0.01	0.01	0.01	<0.01	< 0.01	0.01	Schmitt and others (1990)
Willamette	~26	1984	Peamouth	wb	3–5c	<.01	<.01	.01	<.01	<.01	.01	<.01	<.01	.01	Schmitt and others (1990)
Willamette	~26	1984	Peamouth	wb	3–5c	<.01	<.01	.01	<.01	<.01	.01	<.01	<.01	.01	Schmitt and others (1990)
Tualatin, Cherry Grove		1987	Crayfish	wb	1		<.0025	<.0025	<.0025	<.0025	<.0025	<.0025	<.0025		USEPA (1992a)
Tualatin, Cook Park		1987	Sucker	wb	3–5c		<.0025	.0181	.00653	<.0025	.0333	<.0025	<.0025		USEPA (1992a)
Rock Creek <sup>1</sup>	1.5	1994	Sculpin	wb	15–20c		<.01			<.01	<.01	<.01			USFWS (1994c)
Rock Creek		1994	Three-spined stickleback	wb	9c		<.01			<.01	<.01	<.01			USFWS (1994c)
Rock Creek	1.7– 2	1994	Sculpin	wb	6c		<.01			<.01	<.01	<.01			USFWS (1994c)

<sup>&</sup>lt;sup>1</sup>Rock Creek is a tributary of the Tualatin River near Sherwood, Oregon (Tualatin Subbasin).

[µg/g, microgram per gram. River mile: --, not available; --, not available; --, not available. Tissue type: wb, whole-body; f, fillet. Number samples: c, composite sample; s, number of composite samples. Concentration range: TCBP, tetrachlorobiphenyl; PCBP, pentachlorobiphenyl; HCBP, hexachlorobiphenyl; PCB, polychlorinated biphenyls; ND, not detected; --, not analyzed; <, less than. References: ODEQ, Oregon Department of Environmental Ouality; USEPA, U.S. Environmental Protection Agency]

									Conc	entration	range (	ւ <b>g/g, wet</b>	weight)				
River	River mile		Species	Tissue type	Number samples	3,3',4,4' TCBP	2,3,3',4,4' PCBP	3,3',4,4',5' PCBP	3,3',4,4',5,5' HxCBP	1221 PCB	1232 PCB	1242 PCB	1248 PCB	1254 PCB	1260 PCB	Total PCB	Refer- ence
Willamette, Newberg Pool		1987	Northern squawfish	wb	3–5c											0.10274 <sup>1</sup>	USEPA (1992a)
Willamette, Halsey		1987	Sucker	wb	3–5c											.00821	USEPA (1992a)
Willamette	~7	1987	Sucker	wb	3–5с											1.5369 <sup>1</sup>	USEPA (1992a)
Willamette	7	1990	Northern squawfish	wb	3	<0.002- .011	< 0.002	<0.002- .006	< 0.002	<0.125	< 0.05	< 0.025		<0.025	<0.025- .209	<.025- .209	ODEQ (1994b)
Willamette	7	1990	Northern squawfish	wb	3	.007	<.002	.006	<.002	<.125	<.050	<.025	<0.025	<.025	.127		Curtis and others (1993)
Willamette	7	1989– 90	Common carp	f	6	<.002- .037	<.002- .006	<.002- .021	<.002	<.15	<.06	<.03		<.025– .16	<.025- 1.403	<.025- 1.403	ODEQ (1994b)
Willamette	7	1990	Common carp	f	1	.037	.006	.021	<.002	<.125	<.050	<.025	<.025	<.025	1.400		Curtis and others (1993)
Willamette	7	1988– 89	Common carp	f	5c,2s					<.015	<.006	<.003		<.003- .16	.044– .066	.044– .16	ODEQ (1994b)
Willamette, Station #1	~13	1987	Crayfish	wb	3–4c/3s										<.04		Hart Crowser (1988)
Willamette, Station #2	~13	1987	Crayfish	wb	3c/2s										<.04		Hart Crowser (1988)
Willamette, Station #3	~13	1987	Crayfish	wb	3–4c/4s										<.04		Hart Crowser (1988)
Willamette	~13	1987	Prickly sculpin	wb	4c/4s										0.19– .63		Hart Crowser (1988)

									Conc	entration	range (	μ <b>g/g, wet</b>	weight)				
River	River mile	Years	Species	Tissue type	Number samples	3,3',4,4' TCBP	2,3,3',4,4' PCBP	3,3',4,4',5' PCBP	3,3',4,4',5,5' HxCBP	1221 PCB	1232 PCB	1242 PCB	1248 PCB	1254 PCB	1260 PCB	Total PCB	Refer- ence
Willamette	~14	1987	Prickly sculpin	wb	4c/5s										0.10- .35		Hart Crowser (1988)
Willamette	18	1989	Common carp	f	4c					<0.125	< 0.05	<0.025		0.36	<.025	0.36	ODEQ (1994b)
Willamette	18	1989	Sucker	f	3c					<.125	<.05	<.025		<.025	<.025	<.025	ODEQ (1994b)
Willamette	~26	1970	Common carp	wb	3–5c									1.25			Schmitt and others (1981)
Willamette	~26	1970	Largescale sucker	wb	3–5c									2.40			Schmitt and others (1981)
Willamette	~26	1970	Largescale sucker	wb	3–5c									4.58			Schmitt and others (1981)
Willamette	~26	1971	Largescale sucker	wb	3–5c									1.67			Schmitt and others (1981)
Willamette	~26	1971	Largescale sucker	wb	3–5c									1.35			Schmitt and others (1981)
Willamette	~26	1971	Northern squawfish	wb	3–5c									2.37			Schmitt and others (1981)
Willamette	~26	1971	Northern squawfish	wb	3–5c									2.60			Schmitt and others (1981)
Willamette	~26	1972	Largescale sucker	wb	3–5c									2.80			Schmitt and others (1981)
Willamette	~26	1972	Largescale sucker	wb	3–5c									5.40			Schmitt and others (1981)

									Conce	entration	n range (	μ <b>g/g, wet</b>	weight)				
River	River mile	Years	Species	Tissue type	Number samples	3,3',4,4' TCBP	2,3,3',4,4' PCBP	3,3',4,4',5' PCBP	3,3',4,4',5,5' HxCBP	1221 PCB	1232 PCB	1242 PCB	1248 PCB	1254 PCB	1260 PCB	Total PCB	Refer- ence
Willamette	~26	1972	Channel catfish	wb	3–5c									4.40			Schmitt and others (1981)
Willamette	~26	1972	Northern squawfish	wb	3–5c									3.00			Schmitt and others (1981)
Willamette	~26	1973	Common carp	wb	3–5c							0.0		.20	0.0		Schmitt and others (1981)
Willamette	~26	1973	Largescale sucker	wb	3–5c							.0		2.40	.0		Schmitt and others (1981)
Willamette	~26	1973	Largescale sucker	wb	3–5c							.0		1.60	.0		Schmitt and others (1981)
Willamette	~26	1973	Northern squawfish	wb	3–5c							.0		2.80	.0		Schmitt and others (1981)
Willamette	~26	1974	Common carp	wb	3–5c							.0		.00	.1		Schmitt and others (1981)
Willamette	~26	1974	Largescale sucker	wb	3–5c							.0		1.30	.0		Schmitt and others (1981)
Willamette	~26	1974	Largescale sucker	wb	3–5c							4.5		2.70	.0		Schmitt and others (1981)
Willamette	~26	1974	Northern squawfish	wb	3–5c							.0		2.30	.0		Schmitt and others (1981)
Willamette	~26	1976	Smallmouth bass	wb	3–5c							.0	0.0	.40	.2		Schmitt an others (1983)
Willamette	~26	1976	Chiselmouth	wb	3–5c							.0	.0	2.00	.3		Schmitt and others (1983)

									Conc	entration	range (	ւ <b>g/g, wet</b>	weight)				
River	River mile		Species	Tissue type	Number samples	3,3',4,4' TCBP	2,3,3',4,4' PCBP	3,3',4,4',5' PCBP	3,3',4,4',5,5' HxCBP	1221 PCB	1232 PCB	1242 PCB	1248 PCB	1254 PCB	1260 PCB	Total PCB	Refer- ence
Willamette	~26	1976	Chiselmouth	wb	3–5c							0.0	0.2	0.20	0.3		Schmitt and others (1983)
Willamette	~26	1978	Northern squawfish	wb	3–5c							.0	.0	.77	.6		Schmitt and others (1983)
Willamette	~26	1978	Chiselmouth	wb	3–5c							.0	.1	.30	.2		Schmitt and others (1983)
Willamette	~26	1978	Chiselmouth	wb	3–5c							.0	.2	.30	.1		Schmitt and others (1983)
Willamette	~26	1980	Largescale sucker	wb	3–5c								1	.3	.3		Schmitt and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c								.2	.3	.7		Schmitt and others (1985)
Willamette	~26	1980	Northern squawfish	wb	3–5c								.0	.2	.6		Schmitt and others (1985)
Willamette	~26	1984	Northern squawfish	wb	3–5c								<.1	.2	.1		Schmitt and others (1990)
Willamette	~26	1984	Peamouth	wb	3–5c								<.1	.1	.1		Schmitt and others (1990)
Willamette	~26	1984	Peamouth	wb	3–5c								<.1	.1	<.1		Schmitt and others (1990)
Willamette	27	1988	Bass	f	3c					< 0.025	< 0.01	<.005		<.005	<.005	<0.025	ODEQ (1994b)
Willamette	27	1988	Common carp	f	3c,2s					<.015	<.006- .0067	<.003		<.003- .205	<.003- .119	<.015- .324	

										Conce	entration	range (µ	ւ <b>g/g, wet</b>	weight)				
	River	River mile	Years	Species	Tissue type	Number samples	3,3',4,4' TCBP	2,3,3',4,4' PCBP	3,3',4,4',5' PCBP	3,3',4,4',5,5' HxCBP	1221 PCB	1232 PCB	1242 PCB	1248 PCB	1254 PCB	1260 PCB	Total PCB	Refer- ence
	Willamette	27	1988	Northern squawfish	f	3c					< 0.015	<0.006	<0.003		<0.003	<0.003	< 0.015	ODEQ (1994b)
	Willamette	28	1989	Common carp	f	3					<.125	<.05	<.025		<.025	<.025	<.025	ODEQ (1994b)
	Willamette	28	1989	Common carp	Liver	2					<.125	<.05	<.025		<.025	<.025	<.025	ODEQ (1994b)
	Willamette	38	1989	Sucker	f	5c					<.125	<.05	<.025		<.025	<.025	<.05	ODEQ (1994b)
	Willamette	38	1988– 89	Northern squawfish	f	5c,2s					<.125	<.05	<.025		<.025	<.025	<.05	ODEQ (1994b)
	Willamette	38	1988	Common carp	f	3c					<.015	<.006	<.003		<.003	.015	.015	ODEQ (1994b)
162	Willamette	48	1989	Common carp	f	2					<.125	<.05	<.025		<.025	<.025	<.125	ODEQ (1994b)
	Willamette	48	1989	Common carp	Liver	2					<.2	<.08	<.04		<.04	<.04	<.2	ODEQ (1994b)
	Willamette	48	1988	Common carp	f	5c					<.015	<.006	<.003		.109	.062	.171	ODEQ (1994b)
	Willamette	48	1988	Northern squawfish	f	5c					<.02	<.008	<.004		<.004	<.004	<.02	ODEQ (1994b)
	Willamette	72	1990	Northern squawfish	wb	3	0.003	<0.002	<0.002	<0.002	<.125	<.050	<.025	<0.025	<.025	.040		Curtis and others (1993)
	Willamette	72	1990	Common carp	f	1	.002	<.002	<.002	<.002	<.125	<.050	<.025	<.025	<.025	.040		Curtis and others (1993)
	Willamette	74	1990	Northern squawfish	wb	3	<.002- .005	<.002	<.002	<.002	<.125	<.05	<.025		<.025	.026– .058	.026– .058	ODEQ (1994b)
	Willamette	74	1989– 90	Common carp	f	6	<.002- .002	<.002	<.002	<.002	<.15	<.06	<.03		<.03	<.03	<.15	ODEQ (1994b)
	Willamette	74	1989	Common carp	Liver	3					<.15	<.06	<.03		<.03	<.03	<.15	ODEQ (1994b)

									Conce	entration	range (µ	ւ <b>g/g, wet</b>	weight)				
River	River mile	Years	Species	Tissue type	Number samples	3,3',4,4' TCBP	2,3,3',4,4' PCBP	3,3',4,4',5' PCBP	3,3',4,4',5,5' HxCBP	1221 PCB	1232 PCB	1242 PCB	1248 PCB	1254 PCB	1260 PCB	Total PCB	Refer- ence
Willamette	131	1990	Northern squawfish	wb	3	< 0.002	<0.002	< 0.002	<0.002	<0.125	< 0.05	< 0.025		<0.025	<0.025	<0.025	ODEQ (1994b)
Willamette	131	1990	Northern squawfish	wb	3	<.002	<.002	<.002	<.002	<.125	<.050	<.025	<0.025	<.025	<.025		Curtis and others (1993)
Willamette	131	1990	Common carp	f	1–5	.003 ±.001	<.002	<.002	<.002	<.125	<.050	<.025	<.025	<.025	<.025		Curtis and others (1993)
Willamette	131	1990	Common carp	f	3	<.002- .002	<.002	<.002	<.002	<.125	<.05	<.025		<.025	<.025	<.025	ODEQ (1994b)
Willamette	147	1990	Northern squawfish	wb	3	<.002- .003	<.002	<.002	<.002	<.125	<.05	<.025		<.025	<.025- .028	<.05- .028	ODEQ (1994b)
Willamette	147	1990	Cutthroat trout	wb	5	<.002- .003	<.002	<.002	<.002	<.125	<.05	<.025		<.025	<.025	<.05	ODEQ (1994b)
Willamette	148	1990	Northern squawfish	wb	3	.003	<.002	<.002	<.002	<.125	<.050	<.025	<.025	<.025	.028		Curtis and others (1993)
Willamette	148	1990	Cutthroat trout	wb	1–5	.0023 ±.0004	<.002	<.002	<.002	<.125	<.050	<.025	<.025	<.025	<.025		Curtis and others (1993)
Willamette	160	1990	Northern squawfish	wb	3	.003	<.002	.003	<.002	<.125	<.050	<.025	<.025	<.025	.055		Curtis and others (1993)
Willamette	160	1990	Cutthroat trout	wb	1	.003	<.002	<.002	<.002	<.125	<.050	<.025	<.025	<.025	<.025		Curtis and others (1993)
Willamette	161	1990	Northern squawfish	wb	3	<.002- .004	<.002	<.002- .005	<.002	<.125	<.050	<.025		<.025	.033– .085	.033- .085	ODEQ (1994b)
Willamette	161	1990	Cutthroat trout	wb	5	<.002- .003	<.002	<.002	<.002	<.125	<.050	<.025		<.025	<.025	<.025	ODEQ (1994b)
Willamette	195	1990	Cutthroat trout	wb	1	.003	<.002	<.002	<.002	<.125	<.050	<.025	<.025	<.025	<.025		Curtis and others (1993)

										Conce	entration	range (µ	ıg/g, wet	weight)				
Riv		River mile	Years	Species	Tissue type	Number samples	3,3',4,4' TCBP	2,3,3',4,4' PCBP	3,3',4,4',5' PCBP	3,3',4,4',5,5' HxCBP	1221 PCB	1232 PCB	1242 PCB	1248 PCB	1254 PCB	1260 PCB	Total PCB	Refer- ence
Willamet	ette	195	1990	Northern squawfish	wb	3	0.008	0.003	0.004	<0.002	<0.125	<0.050	<0.025	<0.025	<0.025	0.106		Curtis and others (1993)
Middle F Willamet		8	1990	Northern squawfish	wb	3	.005- .011	.002- .004	<.002- .007	<.002	<.125	<.050	<.025		<.025	.074– .131	0.074– .131	ODEQ (1994b)
Middle F Willamet		8	1990	Cutthroat trout	wb	3	<.002- .003	<.002	<.002	<.002	<.125	<.05	<.025		<.025	<.025	<.025	ODEQ (1994b)
Santiam		0.5	1988	Northern squawfish	f	1					<.02	<.008	<.004		<.004	<.004	<.02	ODEQ (1994b)
Tualatin, Cherry G	*		1987	Crayfish	wb	3–5c											ND	USEPA (1992a)
Tualatin, Cook Par			1987	Sucker	wb	3–5c											.72201	USEPA (1992a)
Tualatin		8	1989	Sucker	f	5c					<.15	<.06	<.03		<.03	<.03	<.15	ODEQ (1994b)
Yamhill		5	1989	Unknown	f	5c					<.125	<.05	<.025		<.025	<.025	<.025	ODEQ (1994b)
Conser S	Slough <sup>2</sup>	0.1	1989	Sucker	f	1					<.125	<.05	<.025		<.025	<.025	<.025	ODEQ (1994b)
Conser S	Slough	0.1	1989	Northern squawfish	f	3–4c,2s					<.15	<.06	<.03		<.03	<.03	<.15	ODEQ (1994b)
Conser S	Slough	0.1	1988	Bass	f	2c					<.15	<.06	<.225		<.003	<.003	<.225	ODEQ (1994b)
Conser S	Slough	0.1	1988	Common carp	f	4c					<.15	<.006	.242		.132	<.003	.374	ODEQ (1994b)

<sup>&</sup>lt;sup>1</sup>Total PCB refers to the sum of concentrations of compounds with 1 to 10 chlorines.

<sup>&</sup>lt;sup>2</sup>Conser Slough is a tributary of the Willamette River near Albany, Oregon (Willamette River Subbasin).

[µg/g, micrograms per gram. River mile: --, not available; ~, approximately. Tissue type: f, fillet; wb, whole-body. Number samples: c, composite sample; r, replicate sample; s, number of composite samples.

Concentration range: ND, not detected; --, not analyzed; <, less than; AL, aluminum; As, arsenic; Ba, barium; Be, beryllium; Cd, cadmium; Cr, chromium; Cu, copper; Fe, iron, Pb, lead; References: ODEQ, Oregon Department of Environmental Quality; USFWS, U.S. Fish and Wildlife Service]

		- (,						C	oncentrat	ion range (μ	g/g, wet we	ight)			
River	River mile	Years	Species	Tissue type	Number samples	AI	As	Ва	Ве	Cd	Cr	Cu	Fe	Pb	Refer- ence
Willamette	7	1989	Common carp	f	3		< 0.07			<0.01- .02	<0.03	0.56– .78		<0.03	ODEQ (1994b)
Willamette	7	1988– 89	Common carp	f	5c,2s		<.03			<.01	<.03	.16– .20		.03- <.05	ODEQ (1994b)
Willamette	18	1989	Common carp	f	4c		<.07			<.01	<.03	.30		<.03	ODEQ (1994b)
Willamette	18	1989	Sucker	f	3c		<.07			<.01	<.03	.27		<.03	ODEQ (1994b)
Willamette	~26	1971	Largescale sucker	wb	5c		.05			<.05				ND	Walsh and others (1977)
Willamette	~26	1971	Largescale sucker	wb	5c,r		.05			<.05				<.10	Walsh and others (1977)
Willamette	~26	1971	Northern squawfish	wb	3c		<.05			<.05				<.10	Walsh and others (1977)
Willamette	~26	1971	Northern squawfish	wb	3c,r		<.05			<.05				<.10	Walsh and others (1977)
Willamette	~26	1972	Channel catfish	wb	5c		<.05			<.05				.10	Walsh and others (1977)
Willamette	~26	1972	Northern squawfish	wb	5c		<.05			.13				.20	Walsh and others (1977)
Willamette	~26	1972	Largescale sucker	wb	5c		.14			<.05				.10	Walsh and others (1977)
Willamette	~26	1972	Largescale sucker	wb	5c,r		<.05			.02				.10	Walsh and others (1977)
Willamette	~26	1976– 77	Smallmouth	wb	3c		<.25			<.05				.12	May and McKinney (1981)

								C	oncentrati	on range (μ	g/g, wet wei	ght)			
River	River mile	Years	Species	Tissue type	Number samples	Al	As	Ва	Ве	Cd	Cr	Cu	Fe	Pb	Refer- ence
Willamette	~26	1976– 77	Chiselmouth	wb	5c		1.15			0.20				0.85	May and McKinney (1981)
Willamette	~26	1978	Northern squawfish	wb	3–5c		.05			.01		0.7		.10	Lowe and others (1985)
Willamette	~26	1978	Chiselmouth	wb	3–5c		.13			.03		1.2		.23	Lowe and others (1985)
Willamette	~26	1978	Chiselmouth	wb	3–5c		.16			.03		1.6		.54	Lowe and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c		.07			.01		.9		.15	Lowe and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c		.07			.02		1.0		.13	Lowe and others (1985)
Willamette	~26	1980	Northern squawfish	wb	3–5c		.06			.01		1.2		.10	Lowe and others (1985)
Willamette	~26	1984	Northern squawfish	wb	3–5c		.30			.00		.57		.03	Schmitt and Brumbaugh (1990)
Willamette	~26	1984	Peamouth	wb	3–5c		.7			.01		.50		.08	Schmitt and Brumbaugh (1990)
Willamette	~26	1984	Peamouth	wb	3–5c		.06			.01		.59		.05	Schmitt and Brumbaugh (1990)
Willamette	27	1988	Bass	f	3c		<.05			<.01	<.03	.23		<.03	ODEQ (1994b)
Willamette	27	1988	Common carp	f	3c,2s		<.03			<.01	<.03	.13– .16		<.03	ODEQ (1994b)
Willamette	27	1988	Northern squawfish	f	3c		<.03			<.01	<.03	.24		<.03	ODEQ (1994b)

								C	Concentrati	<b>ion range (</b> μ	g/g, wet we	ght)			
River	River mile	Years	Species	Tissue type	Number samples	Al	As	Ва	Ве	Cd	Cr	Cu	Fe	Pb	Refer- ence
Willamette	28	1989	Common carp	f	3		< 0.07			<0.01- .02	<0.03- .04	0.41- .54		< 0.03	ODEQ (1994b)
Willamette	38	1989	Sucker	f	5c		.06			<.01	<.03	.2		<.03	ODEQ (1994b)
Willamette	38	1988– 89	Northern squawfish	f	5c,2s		<.06			<.01	<.03	.24– .31		<.03	ODEQ (1994b)
Willamette	38	1988	Common carp	f	3c		<.03			<.01	<.03	.19		<.03	ODEQ (1994b)
Willamette	48	1989	Common carp	f	2		<.07 .1			<.01	<.03	.15– .33		<.03	ODEQ (1994b)
Willamette	48	1988	Common carp	f	5c		<.03			<.01	<.03	.22		<.03	ODEQ (1994b)
Willamette	48	1988	Northern squawfish	f	5c		<.03			<.01	<.03	.17		<.03	ODEQ (1994b)
Willamette	74	1989	Common carp	f	3		<.07			<.01	<.03	.36– .54		<.03	ODEQ (1994b)
Santiam	0.5	1988	Northern squawfish	f	1		<.03			.01	.06	.27		<.03	ODEQ (1994b)
Conser Slough <sup>1</sup>	0.1	1989	Sucker	f	5c		<.06			<.01	<.03	.28		<.03	ODEQ (1994b)
Conser Slough	0.1	1989	Northern squawfish	f	3c		<.06			<.01	<.03	.36		<.03	ODEQ (1994b)
Johnson Creek	1.5	1991	Crayfish	wb	1		<1.52	2.65	< 0.01	.02	.08	10.5		.18	ODEQ (1994b)
Johnson Creek	3	1991	Crayfish	wb	1		<1.52	2.72	<.01	.04	.06	15.47		.13	ODEQ (1994b)
Johnson Creek	6.1	1991	Crayfish	wb	1		<1.27	2.37	<.09	.02	<.03	12.17		.05	ODEQ (1994b)
Johnson Creek	8.3	1991	Crayfish	wb	1		<1.66	5.75	<.11	.02	<.04	19.89		.07	ODEQ (1994b)
Johnson Creek		1991	Crayfish	wb	1		<1.62	5.62	<.11	.02	.07	17.93		.09	ODEQ (1994b)
Johnson Creek	16.9	1991	Crayfish	wb	1		<1.75	7.22	<.12	.05	.05	18.17		<.07	ODEQ (1994b)

								C	oncentrati	on range (μ	g/g, wet we	ight)			
River	River mile	Years	Species	Tissue type	Number samples	Al	As	Ва	Ве	Cd	Cr	Cu	Fe	Pb	Refer- ence
Johnson Creek	21	1991	Crayfish	wb	1		<1.64	4.14	<0.11	0.06	< 0.04	14.17		<0.07	ODEQ (1994b)
Johnson Creek		1991	Crayfish	wb	1		<1.55	5.15	<.11	.02	.04	17.51		.08	ODEQ (1994b)
Rock Creek <sup>2</sup>	1.5	1994	Three-spined stickleback	wb	15– 20c	102		1.31		<.10	<.10	1.31	55.0	<.45	USFWS (1994c)
Rock Creek	1.5	1994	Largemouth bass	wb	2c	5.10		.43		<.10	1.71	.71	43.1	<.45	USFWS (1994c)
Rock Creek	1.7–2.0	1994	Three-spined stickleback	wb	9c	11.0		1.70		<.10	1.60	2.40	73.2	<.45	USFWS (1994c)
Rock Creek	1.7–2.0	1994	Sculpin	wb	6c	2.61		1.70		<.10	.58	2.10	32.1	<.45	USFWS (1994c)
Rock Creek	2	1994	Three-spined stickleback	wb	9c	11.0		2.00		<.10	.84	3.81	57.9	.84	USFWS (1994c)
Rock Creek	2	1994	Sculpin	wb	1	4.56		1.31		1.23	3.68	7.34	48.1	1.31	USFWS (1994c)
Rock Creek	2.9	1994	Three-spined stickleback	wb	1	5.39		2.90		<.48	.77	9.30	27.0	<2.16	USFWS (1994c)
Rock Creek	2.9	1994	Crappie	wb	1	2.20		1.10		.58	2.10	1.20	30.1	<.45	USFWS (1994c)

<sup>&</sup>lt;sup>1</sup>Conser Slough is a tributary of the Willamette River near Albany, Oregon (Willamette Subbasin).

<sup>&</sup>lt;sup>2</sup>Rock Creek is a tributary of the Tualatin River near Sherwood, Oregon (Tualatin Subbasin).

[µg/g, microgram per gram. River mile: --, not available; ~, approximately. Tissue type: wb, whole-body; f, fillet. Number samples: c, composite sample; r, replicate sample; s, number of composite samples. Concentration range: ND, not detected; --, not analyzed; <, less than; Mg, magnesium; Mn, manganese; Hg, mercury; Mo, molybdenum; Ni, nickel; Se, selenium; Sr, strontium; Tl, thallium; Zn, zinc. References: ODEQ, Oregon Department of Environmental Quality; USEPA, U.S. Environmental Protection Agency; USFWS, U.S. Fish and Wildlife Service]

								Cond	centration	n range (µ	ι <b>g/g, wet w</b>	eight)			
River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference
Willamette Newberg Pool		1987	Northern squawfish	wb	3–5c			0.11							USEPA (1992a)
Willamette Halsey		1987	Mountain whitefish	f	3–5c			.06							USEPA (1992a)
Willamette Halsey		1987	Sucker	wb	3–5c			.07							USEPA (1992a)
Willamette	~7	1987	Sucker	wb	3–5c			ND							USEPA (1992a)
Willamette	7	1989	Common carp	f	3			.11– .17						9.55–12.37	ODEQ (1994b)
Willamette	7	1989	Common carp	f	5c,2s			.17– .19						8.14–12.47	ODEQ (1994b)
Willamette	18	1989	Common carp	f	4c			.14						7.07	ODEQ (1994b)
Willamette	18	1989	Sucker	f	3c			.05						5.64	ODEQ (1994b)
Willamette	~26	1971	Largescale sucker	wb	5c			.28							Walsh and others (1977)
Willamette	~26	1971	Largescale sucker	wb	5c,r			.32							Walsh and others (1977)
Willamette	~26	1971	Northern squawfish	wb	3c			1.10							Walsh and others (1977)
Willamette	~26	1971	Northern squawfish	wb	3c,r			.99							Walsh and others (1977)
Willamette	~26	1972	Channel catfish	wb	5c			.29			0.06				Walsh and others (1977)
Willamette	~26	1972	Northern squawfish	wb	5c			.04			.04				Walsh and others (1977)
Willamette	~26	1972	Largescale sucker	wb	5c			.24			.12				Walsh and others (1977)
Willamette	~26	1972	Largescale sucker	wb	5c,r			.04			.09				Walsh and others (1977)

								Con	centration	n range (μ	ւ <b>g/g, wet w</b>	eight)			
River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference
Willamette	~26	1973	Largescale sucker	wb	5c			0.08			0.09				Walsh and others (1977)
Willamette	~26	1973	Largescale sucker	wb	5c,r			.20			.05				Walsh and others (1977)
Willamette	~26	1973	Northern squawfish	wb	5c			.65			<.05				Walsh and others (1977)
Willamette	~26	1973	Common carp	wb	5c			.15			.18				Walsh and others (1977)
Willamette	~26	1976– 77	Smallmouth bass	wb	3c			.13							May and McKinney (1981)
Willamette	~26	1976–77	Chiselmouth	wb	5c			<.02							May and McKinney (1981)
Willamette	~26	1978	Northern squawfish	wb	3–5c			.52			.13			23.2	Lowe and others (1985)
Willamette	~26	1978	Chiselmouth	wb	3–5c			.04			.17			31.9	Lowe and others (1985)
Willamette	~26	1978	Chiselmouth	wb	3–5c			.03			.14			42.2	Lowe and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c			.15			.20			22.4	Lowe and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c			.23			.23			22.6	Lowe and others (1985)
Willamette	~26	1980	Northern squawfish	wb	3–5c			.77			.45			17.6	Lowe and others (1985)
Willamette	~26	1984	Northern squawfish	wb	3–5c			.21			.25			16.35	Schmitt and Brumbaugh (1990)
Willamette	~26	1984	Peamouth	wb	3–5c			.05			.11			17.48	Schmitt and Brumbaugh (1990)
Willamette	~26	1984	Peamouth	wb	3–5c			.04			.13			17.55	Schmitt and Brumbaugh (1990)
Willamette	27	1988	Bass	f	3c			.1						5.8	ODEQ (1994b)
Willamette	27	1988	Common carp	f	3c,2s			.2- .46						4.85– 7.28	ODEQ (1994b)

					Concentration range (μg/g, wet weight)											
River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference	
Willamette	27	1988	Northern squawfish	f	3c			0.34						4.98	ODEQ (1994b)	
Willamette	28	1989	Common carp	f	3			.14– .16						6.7–1 4.56	ODEQ (1994b)	
Willamette	38	1989	Sucker	f	9c			.11						5.1	ODEQ (1994b)	
Willamette	38	1988–89	Northern squawfish	f	5c,2s			.14– .23						6.06– 7.68	ODEQ (1994b)	
Willamette	38	1988	Common carp	f	3c			.12						16.28	ODEQ (1994b)	
Willamette	48	1989	Common carp	f	2			.02- .1						4.97– 6.91	ODEQ (1994b)	
Willamette	48	1988	Common carp	f	5c			.16						9.9	ODEQ (1994b)	
Willamette	48	1988	Northern squawfish	f	5c			.44						4.65	ODEQ (1994b)	
Willamette	74	1989	Common carp	f	3			.12– .2						5.91– 9.11	ODEQ (1994b)	
Coast Fork Willamette		1994	Bluegill	f	1			.37							ODEQ (1996)	
Coast Fork Willamette		1994	Cutthroat trout	f	5			.24– .42							ODEQ (1996)	
Coast Fork Willamette		1994	Mountain whitefish	f	3			.06– .11							ODEQ (1996)	
Row		1994	Cutthroat trout	f	5			.09– .13							ODEQ (1996)	
Row		1994	Largemouth bass	f	5			.29– .58							ODEQ (1996)	
Santiam	0.5	1988	Northern squawfish	f	1			.10						8.11	ODEQ (1994b)	
Tualatin, Cherry Grove		1987	Cutthroat trout	f	3–5c			.07							USEPA (1992a)	
Tualatin, Cherry Grove		1987	Crayfish	wb	3–5c			ND							USEPA (1992a)	

								Con	centration	range (μ	g/g, wet w	eight)			
River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference
Tualatin, Cook Park		1987	Sucker	wb	3–5c			0.18							USEPA (1992a)
Conser Slough <sup>1</sup>	0.1	1989	Sucker	f	5c			.08						5.21	ODEQ (1994b)
Conser Slough	0.1	1989	Northern squawfish	f	3c			.49						5.57	ODEQ (1994b)
Johnson Creek	1.5	1991	Crayfish	wb	1			.03			<2.53		<5.05	17.98	ODEQ (1994b)
Johnson Creek	3	1991	Crayfish	wb	1			.09			<2.53		<5.05	19.02	ODEQ (1994b)
Johnson Creek	6.1	1991	Crayfish	wb	1			.14			<2.11		<4.23	15.04	ODEQ (1994b)
Johnson Creek	8.3	1991	Crayfish	wb	1			.12			<2.76		<5.53	18.12	ODEQ (1994b)
Johnson Creek		1991	Crayfish	wb	1			.11			<2.7		<5.4	19.87	ODEQ (1994b)
Johnson Creek	16.9	1991	Crayfish	wb	1			.08			<2.91		<5.83	18.41	ODEQ (1994b)
Johnson Creek	21	1991	Crayfish	wb	1			.05			<2.73		<5.45	17.44	ODEQ (1994b)
Johnson Creek		1991	Crayfish	wb	1			.09			<2.58		<5.5	17.72	ODEQ (1994b)
Rock Creek <sup>2</sup>	1.5	1994	Sculpin	wb	9c	320	7.41	.09	< 0.10	< 0.16		14.0		19.0	USFWS (1994c)
Rock Creek	1.5	1994	Largemouth bass	wb	2c	450	3.39	.08	<.10	.56		20.0		22.0	USFWS (1994c)
Rock Creek	1.7–2	1994	Three-spined stickleback	wb	9c	390	11.0	.11	<.10	.42		12.0		39.9	USFWS (1994c)
Rock Creek	1.7–2	1994	Sculpin	wb	6c	340	3.40	.12	<.10	.18		19.0		22.0	USFWS (1994c)
Rock Creek	2	1994	Three-spined stickleback	wb	9c	360	10.0	.13	.18	.28		12.0		35.1	USFWS (1994c)
Rock Creek	2	1994	Sculpin	wb	1	306	4.20	.10	.10	.86		15.7		23.6	USFWS (1994c)

								Con	centration	range (μ	g/g, wet v	veight)			
River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	ΤI	Zn	Reference
Rock Creek	2.9	1994	Three-spined stickleback	wb	1	500	12.0		<0.48	<0.77		15.0		29.0	USFWS (1994c)
Rock Creek	2.9	1994	Crappie	wb	1	430	2.10	< 0.05	<.10	.67		26.0		27.0	USFWS (1994c)
Cottage Grove Lake		1974	Brown bullhead	f	68			.17– .68							Worcester (1979)
Cottage Grove Lake		1974	Spring chinook salmon	f	122			.09– .82							Worcester (1979)
Cottage Grove Lake		1974	Cutthroat trout	f	33			.10– 1.36							Worcester (1979)
Cottage Grove Lake		1974	Rainbow trout	f	20			.05– .48							Worcester (1979)
Cottage Grove Lake		1974	Largemouth bass	f	35			.15– 1.44							Worcester (1979)
Cottage Grove Lake		1975	Brown bullhead	f	14			.08- .35							Worcester (1979)
Cottage Grove Lake		1975–76	Spring chinook salmon	f	84			.10– .96							Worcester (1979)
Cottage Grove Lake		1975	Cutthroat trout	f	12			.06– .35							Worcester (1979)
Cottage Grove Lake		1975	Rainbow trout	f	12			.07- .24							Worcester (1979)
Cottage Grove Lake		1990	Largemouth bass	f	5			.22– 1.79							Allen and Curtis (1991)
Cottage Grove Lake		1993	Black crappie	f	6			.38– .75							ODEQ (1996)
Cottage Grove Lake		1993–94	Bluegill	f	7			.46– 1.13							ODEQ (1996)
Cottage Grove Lake		1993–94	Bullhead	f	12			.26– .75							ODEQ (1996)
Cottage Grove Lake		1990–94	Largemouth bass	f	23			.22– 1.79							ODEQ (1996)
Dorena Lake		1993	Black crappie	f	8			0.16– .24							ODEQ (1996)

River		River mile Years						Cond	entration	n range (µ	ıg/g, wet v	veight)			
	River mile		Species	Tissue type	Number samples		Hg	Мо	Ni	Se	Sr	TI	Zn	Reference	
Dorena Lake		1993–94	Bluegill	f	12			.01– .355							ODEQ (1996)
Dorena Lake		1994	Bullhead	f	2			.25– .37							ODEQ (1996)
Dorena Lake		1993–94	Largemouth bass	f	39			.03- .94							ODEQ (1996)
Fern Ridge Lake		1993	Black crappie	f	2			.058– .068							ODEQ (1996)
Fern Ridge Lake		1993	Common carp	wb	2			.058– .108							ODEQ (1996)
Fern Ridge Lake		1993	Largemouth bass	f	1			.089							ODEQ (1996)
Henry Hagg Lake		1993	Largemouth bass	f	7			.069– .104							ODEQ (1996)

<sup>&</sup>lt;sup>1</sup>Conser Slough is a tributary of the Willamette River near Albany, Oregon (Willamette River Subbasin).

<sup>&</sup>lt;sup>2</sup>Rock Creek is a tributary of the Tualatin River near Sherwood, Oregon (Tualatin Subbasin).

[µg/g, microgram per gram. River mile: --, not available; ~, approximately. Tissue type: wb, whole-body; f, fillet. Number samples: c, composite sample; r, replicate sample; s, number of composite samples. Concentration range: ND, not detected; --, not analyzed; <, less than; Mg, magnesium; Mn, manganese; Hg, mercury; Mo, molybdenum; Ni, nickel; Se, selenium; Sr, strontium; Tl, thallium; Zn, zinc. References: ODEQ, Oregon Department of Environmental Quality; USEPA, U.S. Environmental Protection Agency; USFWS, U.S. Fish and Wildlife Service]

River	River mile		Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference
Willamette Newberg Pool		1987	Northern squawfish	wb	3–5c			0.11							USEPA (1992a)
Willamette Halsey		1987	Mountain whitefish	f	3–5c			.06							USEPA (1992a)
Willamette Halsey		1987	Sucker	wb	3–5c			.07							USEPA (1992a)
Willamette	~7	1987	Sucker	wb	3–5c			ND							USEPA (1992a)
Willamette	7	1989	Common carp	f	3			.11– .17						9.55–12.37	ODEQ (1994b)
Willamette	7	1989	Common carp	f	5c,2s			.17– .19						8.14–12.47	ODEQ (1994b)
Willamette	18	1989	Common carp	f	4c			.14						7.07	ODEQ (1994b)
Willamette	18	1989	Sucker	f	3c			.05						5.64	ODEQ (1994b)
Willamette	~26	1971	Largescale sucker	wb	5c			.28							Walsh and others (1977)
Willamette	~26	1971	Largescale sucker	wb	5c,r			.32							Walsh and others (1977)
Willamette	~26	1971	Northern squawfish	wb	3c			1.10							Walsh and others (1977)
Willamette	~26	1971	Northern squawfish	wb	3c,r			.99							Walsh and others (1977)
Willamette	~26	1972	Channel catfish	wb	5c			.29			0.06				Walsh and others (1977)
Willamette	~26	1972	Northern squawfish	wb	5c			.04			.04				Walsh and others (1977)
Willamette	~26	1972	Largescale sucker	wb	5c			.24			.12				Walsh and others (1977)
Willamette	~26	1972	Largescale sucker	wb	5c,r			.04			.09				Walsh and others (1977)

River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference
Willamette	~26	1973	Largescale sucker	wb	5c			0.08			0.09				Walsh and others (1977)
Willamette	~26	1973	Largescale sucker	wb	5c,r			.20			.05				Walsh and others (1977)
Willamette	~26	1973	Northern squawfish	wb	5c			.65			<.05				Walsh and others (1977)
Willamette	~26	1973	Common carp	wb	5c			.15			.18				Walsh and others (1977)
Willamette	~26	1976– 77	Smallmouth bass	wb	3c			.13							May and McKinney (1981)
Willamette	~26	1976–77	Chiselmouth	wb	5c			<.02							May and McKinney (1981)
Willamette	~26	1978	Northern squawfish	wb	3–5c			.52			.13			23.2	Lowe and others (1985)
Willamette	~26	1978	Chiselmouth	wb	3–5c			.04			.17			31.9	Lowe and others (1985)
Willamette	~26	1978	Chiselmouth	wb	3–5c			.03			.14			42.2	Lowe and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c			.15			.20			22.4	Lowe and others (1985)
Willamette	~26	1980	Largescale sucker	wb	3–5c			.23			.23			22.6	Lowe and others (1985)
Willamette	~26	1980	Northern squawfish	wb	3–5c			.77			.45			17.6	Lowe and others (1985)
Willamette	~26	1984	Northern squawfish	wb	3–5c			.21			.25			16.35	Schmitt and Brumbaugh (1990)
Willamette	~26	1984	Peamouth	wb	3–5c			.05			.11			17.48	Schmitt and Brumbaugh (1990)
Willamette	~26	1984	Peamouth	wb	3–5c			.04			.13			17.55	Schmitt and Brumbaugh (1990)
Willamette	27	1988	Bass	f	3c			.1						5.8	ODEQ (1994b)
Willamette	27	1988	Common carp	f	3c,2s			.2- .46						4.85– 7.28	ODEQ (1994b)

						Concentration range (μg/g, wet weight)									
River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference
Willamette	27	1988	Northern squawfish	f	3c			0.34						4.98	ODEQ (1994b)
Willamette	28	1989	Common carp	f	3			.14– .16						6.7–1 4.56	ODEQ (1994b)
Willamette	38	1989	Sucker	f	9c			.11						5.1	ODEQ (1994b)
Willamette	38	1988–89	Northern squawfish	f	5c,2s			.14– .23						6.06– 7.68	ODEQ (1994b)
Willamette	38	1988	Common carp	f	3c			.12						16.28	ODEQ (1994b)
Willamette	48	1989	Common carp	f	2			.02– .1						4.97– 6.91	ODEQ (1994b)
Willamette	48	1988	Common carp	f	5c			.16						9.9	ODEQ (1994b)
Willamette	48	1988	Northern squawfish	f	5c			.44						4.65	ODEQ (1994b)
Willamette	74	1989	Common carp	f	3			.12– .2						5.91– 9.11	ODEQ (1994b)
Coast Fork Willamette		1994	Bluegill	f	1			.37							ODEQ (1996)
Coast Fork Willamette		1994	Cutthroat trout	f	5			.24– .42							ODEQ (1996)
Coast Fork Willamette		1994	Mountain whitefish	f	3			.06– .11							ODEQ (1996)
Row		1994	Cutthroat trout	f	5			.09– .13							ODEQ (1996)
Row		1994	Largemouth bass	f	5			.29– .58							ODEQ (1996)
Santiam	0.5	1988	Northern squawfish	f	1			.10						8.11	ODEQ (1994b)
Tualatin, Cherry Grove		1987	Cutthroat trout	f	3–5c			.07							USEPA (1992a)
Tualatin, Cherry Grove		1987	Crayfish	wb	3–5c			ND							USEPA (1992a)

					Concentration range (μg/g, wet weight)										
River	River mile	Years	Species	Tissue type	Number samples	Mg	Mn	Hg	Мо	Ni	Se	Sr	TI	Zn	Reference
Tualatin, Cook Park		1987	Sucker	wb	3–5c			0.18							USEPA (1992a)
Conser Slough <sup>1</sup>	0.1	1989	Sucker	f	5c			.08						5.21	ODEQ (1994b)
Conser Slough	0.1	1989	Northern squawfish	f	3c			.49						5.57	ODEQ (1994b)
Johnson Creek	1.5	1991	Crayfish	wb	1			.03			<2.53		<5.05	17.98	ODEQ (1994b)
Johnson Creek	3	1991	Crayfish	wb	1			.09			<2.53		<5.05	19.02	ODEQ (1994b)
Johnson Creek	6.1	1991	Crayfish	wb	1			.14			<2.11		<4.23	15.04	ODEQ (1994b)
Johnson Creek	8.3	1991	Crayfish	wb	1			.12			<2.76		<5.53	18.12	ODEQ (1994b)
Johnson Creek		1991	Crayfish	wb	1			.11			<2.7		<5.4	19.87	ODEQ (1994b)
Johnson Creek	16.9	1991	Crayfish	wb	1			.08			<2.91		<5.83	18.41	ODEQ (1994b)
Johnson Creek	21	1991	Crayfish	wb	1			.05			<2.73		<5.45	17.44	ODEQ (1994b)
Johnson Creek		1991	Crayfish	wb	1			.09			<2.58		<5.5	17.72	ODEQ (1994b)
Rock Creek <sup>2</sup>	1.5	1994	Sculpin	wb	9c	320	7.41	.09	< 0.10	< 0.16		14.0		19.0	USFWS (1994c)
Rock Creek	1.5	1994	Largemouth bass	wb	2c	450	3.39	.08	<.10	.56		20.0		22.0	USFWS (1994c)
Rock Creek	1.7–2	1994	Three-spined stickleback	wb	9c	390	11.0	.11	<.10	.42		12.0		39.9	USFWS (1994c)
Rock Creek	1.7–2	1994	Sculpin	wb	6c	340	3.40	.12	<.10	.18		19.0		22.0	USFWS (1994c)
Rock Creek	2	1994	Three-spined stickleback	wb	9c	360	10.0	.13	.18	.28		12.0		35.1	USFWS (1994c)
Rock Creek	2	1994	Sculpin	wb	1	306	4.20	.10	.10	.86		15.7		23.6	USFWS (1994c)

						Concentration range (μg/g, wet weight)									
	River mile	Years	Species	Tissue type	Number samples		Sr	ΤI	Zn	Reference					
Rock Creek	2.9	1994	Three-spined stickleback	wb	1	500	12.0		<0.48	<0.77		15.0		29.0	USFWS (1994c)
Rock Creek	2.9	1994	Crappie	wb	1	430	2.10	< 0.05	<.10	.67		26.0		27.0	USFWS (1994c)
Cottage Grove Lake		1974	Brown bullhead	f	68			.17– .68							Worcester (1979)
Cottage Grove Lake		1974	Spring chinook salmon	f	122			.09– .82							Worcester (1979)
Cottage Grove Lake		1974	Cutthroat trout	f	33			.10– 1.36							Worcester (1979)
Cottage Grove Lake		1974	Rainbow trout	f	20			.05– .48							Worcester (1979)
Cottage Grove Lake		1974	Largemouth bass	f	35			.15– 1.44							Worcester (1979)
Cottage Grove Lake		1975	Brown bullhead	f	14			.08- .35							Worcester (1979)
Cottage Grove Lake		1975–76	Spring chinook salmon	f	84			.10– .96							Worcester (1979)
Cottage Grove Lake		1975	Cutthroat trout	f	12			.06– .35							Worcester (1979)
Cottage Grove Lake		1975	Rainbow trout	f	12			.07- .24							Worcester (1979)
Cottage Grove Lake		1990	Largemouth bass	f	5			.22– 1.79							Allen and Curtis (1991)
Cottage Grove Lake		1993	Black crappie	f	6			.38– .75							ODEQ (1996)
Cottage Grove Lake		1993–94	Bluegill	f	7			.46– 1.13							ODEQ (1996)
Cottage Grove Lake		1993–94	Bullhead	f	12			.26– .75							ODEQ (1996)
Cottage Grove Lake		1990–94	Largemouth bass	f	23			.22– 1.79							ODEQ (1996)
Dorena Lake		1993	Black crappie	f	8			0.16– .24							ODEQ (1996)

River		River mile Years						Cond	entration	n range (µ	ıg/g, wet v	veight)			
	River mile		Species	Tissue type	Number samples		Hg	Мо	Ni	Se	Sr	TI	Zn	Reference	
Dorena Lake		1993–94	Bluegill	f	12			.01– .355							ODEQ (1996)
Dorena Lake		1994	Bullhead	f	2			.25– .37							ODEQ (1996)
Dorena Lake		1993–94	Largemouth bass	f	39			.03- .94							ODEQ (1996)
Fern Ridge Lake		1993	Black crappie	f	2			.058– .068							ODEQ (1996)
Fern Ridge Lake		1993	Common carp	wb	2			.058– .108							ODEQ (1996)
Fern Ridge Lake		1993	Largemouth bass	f	1			.089							ODEQ (1996)
Henry Hagg Lake		1993	Largemouth bass	f	7			.069– .104							ODEQ (1996)

<sup>&</sup>lt;sup>1</sup>Conser Slough is a tributary of the Willamette River near Albany, Oregon (Willamette River Subbasin).

<sup>&</sup>lt;sup>2</sup>Rock Creek is a tributary of the Tualatin River near Sherwood, Oregon (Tualatin Subbasin).