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Sargassum species as hydrozoans substrates: Key patterns of association or just availability?

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ABSTRACT

The genus *Sargassum* comprises both benthic and holopelagic species, serving as basibionts for numerous marine organisms, including frequent epibiotic occurrences of hydrozoans (Cnidaria). Several aspects of the epibiotic interaction between hydrozoans and *Sargassum* species remain insufficiently explored. This includes patterns such as the hydrozoan community's composition and abundance across sections and structures of *Sargassum*. To address these gaps, we conducted an investigation aiming to uncover potential differences in the species composition and abundance of hydrozoan epibionts on benthic and holopelagic *Sargassum* species (BSS and HSS, respectively) from the Atlantic and Pacific Oceans of Mexico. Additionally, we sought to characterize hydrozoan epibiont species on *Sargassum*, belonging to Leptothecata (88.9%) and anthoathecate (11.1%), with one new register for *Sargassum* species. Each *Sargassum* species harbored a distinct hydrozoan on BSS exhibited less variability than HSS. The basal section of BSS hosted the highest number of species. Concerning *Sargassum* structures, a greater species richness was found on the axis and the blades. Our study also demonstrates that differences in hydrozoan species primarily depend on the geographical distribution of *Sargassum* species.

1. Introduction

The genus *Sargassum* (Phaeophyceae: Sargassaceae) comprises 360 accepted marine brown algae distributed globally in tropical and temperate waters (Guiry and Guiry, 2023). These algae can form massive benthic and holopelagic populations, creating complex ecosystems that offer substrate, food, and shelter for numerous organisms (Gower et al., 2013; Stiger-Pouvreau et al., 2023). Certain *Sargassum* species have applications in various human sectors such as animal feed, pharmacology, and cosmetics (Stiger-Pouvreau et al., 2023), while others are associated with ecological effects and impacts on human activities and health, such as the introduction of exotic *Sargassum* species, the occurrence of large holopelagic *Sargassum* accumulations in the Caribbean, and the presence of potential pathogens on *Sargassum* (Britton-Simmons, 2004; Van Tussenbroek et al., 2017; Gower and King,

2019; Rodríguez-Martínez et al., 2019; Mincer et al., 2023; Pérez-Posada et al., 2023).

Sargassum is a well-documented basibiont for various marine taxa, including Ascidiacea, Acarina, Cirripedia, Gastropoda, Gymnolaemata, Hydrozoa, Nematoda, and turbellarians (Weis, 1968; Abé et al., 2013; Alleyne et al., 2023). Hydrozoans (phylum Cnidaria) are ubiquitous components of marine food webs, associated with diverse organisms, including quite frequent epibiotic occurrences on Sargassum (Nishihira, 1965, 1968; Boero, 1984; Fitridge et al., 2012; Carral-Murrieta et al., 2023a). Furthermore, as the Sargassum ages, it tends to host a higher abundance of hydrozoans, which makes it less suitable for human utilization (Alleyne et al., 2023).

A recent global review listed 131 hydrozoan species associated with only 26 *Sargassum* species (Carral-Murrieta et al., 2023a). Hydrozoans are known as substrate generalists, colonizing various surfaces,

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Abbreviations: BSS, Benthic Sargassum species; HSS, Holopelagic Sargassum species.

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including natural and artificial substrates like floating docks, boats, vertebrates, aquatic plants, and macroalgae (Frick et al., 2003; Oliveira and Marques, 2007; Calder et al., 2014). Their colonization typically begins with swimming or drifting larvae, often in the form of a planula, which settles on a substrate and develops into a solitary polyp or a colony (Sommer, 1992). Efficient attachment to different substrates relies on a hydrorhiza, a net-like structure that extends onto the substrate, eventually giving rise to new stems or modules, such as hydranths (Mills et al., 2007).

Previous studies have suggested that the epibiotic interaction between *Sargassum* species and hydrozoans can be influenced by various factors, including morphological, behavioral, developmental, and ecological characteristics. It was found that hydrozoans possessing a chitin exoskeleton around the hydranth (= Leptothecata) are more prevalent than those lacking this morphological structure (i.e., the nonmonophyletic anthoathecate), as this structure usually allows Leptothecata to endure adverse environmental conditions (cf., Mendoza-Becerril et al., 2020a). Additionally, different hydrozoans tend to occupy specific areas on the algal thallus, which relates to larval settlement patterns and space competition (Nishihira, 1973; Ryland, 1974). Other studies have focused on seasonal patterns of hydroid cover (Cunha and Jacobucci, 2010) and their growth on specific sections or morphological structures along *Sargassum* thalli (Mendoza-Becerril et al., 2020a; Niermann, 1986; Nishihira, 1973; Rackley, 1974; Ryland, 1974). Furthermore, *Sargassum*'s morphological complexity and the presence of hydrozoans are directly related to the assemblage of other epibionts, such as bryozoan and caprellids (Ryland, 1974; Jackson, 1977; Carvalho et al., 2022).

Despite the accumulated knowledge, epibiosis patterns between hydrozoans and *Sargassum* species remain insufficiently explored due to the absence of comprehensive and representative sample collections, methodological heterogeneity in prior research, and even the inherent



Fig. 1. Collection sites of Sargassum thalli. A) Bahía de Todos Santos, Ensenada, Baja California; B) San Juan de La Costa, La Paz, Baja California Sur; C) Puerto Morelos, Quintana Roo.

biological complexity of both the basibionts and epibionts. Therefore, this study pursued two primary objectives aimed at elucidating basic patterns: (1) to investigate potential differences in the species composition and abundance of epibiont hydrozoans on benthic and holopelagic *Sargassum* from the Atlantic and Pacific Oceans of Mexico and (2) to examine whether hydrozoan epibionts exhibit preferences for specific sections and structures of *Sargassum*.

2. Material and methods

2.1. Sargassum samples

Sargassum species were selected based on their distribution along Mexican coasts, considering that holopelagic species are restricted to the Atlantic Ocean (Rocha-Ramírez and Siqueiros-Beltrones, 1990; Gower et al., 2013; Aguilar-Rosas et al., 2014). Living thalli of five Sargassum species were collected for hydrozoan epibiont analysis between July 2020 and October 2021. Three benthic species were collected in the Pacific Ocean, Sargassum horneri (Turner) C. Agardh and Sargassum muticum (Yendo) Fensholt in Bahía de Todos Santos (Baja California), and Sargassum horridum Setchell & N.L. Gardner in San Juan de la Costa (Baja California Sur, southwest of the Gulf of California). While in the Caribbean, two morphotypes of holopelagic Sargassum [Sargassum fluitans III (Børgesen) Børgesen; Sargassum natans VIII (Linnaeus) Gaillon] were collected in Puerto Morelos (Fig. 1, Fig. 2A, Table 1).

In each site, *Sargassum* collection was conducted following the methods outlined by Carral-Murrieta (2021). For benthic species, twenty thalli of approximately 30 cm long and exhibiting hydrozoan cover were collected by scuba diving and snorkeling approximately

40–50 m from the coast. The holopelagic species were collected by scoop net from floating aggregations approximately 45 m from the coast. The benthic *Sargassum* species (hereafter referred to as BSS) were identified based on the keys and descriptions provided by Aguilar-Rosas and Galindo (1990), Aguilar-Rosas et al. (2007), and Andrade-Sorcia et al. (2014), while the holopelagic *Sargassum* species (HSS) were identified following Parr (1939) and Schell et al. (2015). Each *Sargassum* thallus was examined under a stereomicroscope and a light microscope to separate and identify epibiont hydrozoans in the laboratory. The identification of hydrozoans was carried out to the lowest possible taxonomic level based on faunal and taxonomic review articles (Calder, 1988, 1991, 1997, 2020; Mendoza-Becerril et al., 2020b; Calder et al., 2021).

2.2. Hydrozoan cover on Sargassum

On each *Sargassum* thallus, epibiont hydrozoan species cover estimates were determined following the procedures described by Cunha and Jacobucci (2010) and Mendoza-Becerril et al. (2020a). For BSS, each thallus was divided into four equal sections (B = Basal, starting from the holdfast; LM = Lower Middle; UM = Upper Middle; A = Apical) (Fig. 2B). In thalli greater than 30 cm, the first 7.5 cm of section B, the middle 7.5 cm of sections LM and UM, and the last 7.5 cm of section A were analyzed. For HSS, sections were not designated since their morphology does not have a differentiation between basal and apical sections. Additionally, the epibiont hydrozoan species cover in each of the *Sargassum* morphological structures (aerocysts, axis, blades, holdfast, receptacles) was documented.



Fig. 2. A) *Sargassum* species sampled in this study. Benthic species: 1. *Sargassum horneri*; 2. *Sargassum muticum*; 3. *Sargassum horridum*. Holopelagic species: 4. *Sargassum fluitans* III; 5. *Sargassum natans* VIII. Scale = 1 cm. **B)** Hydrozoan cover estimation on both benthic and holopelagic *Sargassum* thalli. Thallus sections: B = Basal, LM = Lower Middle, UM = Upper Middle, A = Apical. Morphological structures: a = aerocysts, b = blade, h = holdfast, r = receptacles, x = axis. Hydrozoans are shown in purple.

Table 1

Sargassum species collected on Mexican coasts.

Sargassum species	3		Site				
	Sampling date	Depth		State	Climate	Salinity (%)	Water Temperature (°C)
Benthic							
S. horneri	October 2021	0.5–2 m	Bahía de Todos Santos	Baja California	Temperate and dry ^a	$33.3\% - 33.7\%^4$	11.0–22.5 °C ⁴
S. muticum	October 2021	0.5–2 m	Bahía de Todos Santos	Baja California	Temperate and dry ^a	$33.3\% - 33.7\%^4$	11.0-22.5 °C ⁴
S. horridum	July 2020	1–1.5 m	San Juan de la Costa	Baja California Sur	Hot dessert ²	$35.5\%^2$	22.0–27.0 °C ⁶
Holopelagic							
S. fluitans III	July 2021	Floating	Puerto Morelos	Quintana Roo	Tropical ³	35.7% ⁵	$24-32 {}^{\circ}C^3$
S. natans VIII	July 2021	Floating	Puerto Morelos	Quintana Roo	Tropical ³	35.7% ⁵	24–32 $^{\circ}C^{3}$

^a Simental, Martínez-Urtaza (2008), ²Obeso-Nieblas et al. (2004), ³Coronado et al. (2007), ⁴Mancilla, Martínez (1991), ⁵Merino, Otero (1991), ⁶Monteforte, Cariño (1992).

2.3. Data analysis

Data was analyzed using the R software libraries (R Development Core Team, 2023). A Venn diagram was constructed to identify exclusive hydrozoan species for each *Sargassum* species, employing the EULERR package (Larsson, 2022). Box plots were generated to visualize the percentages of epibiont hydrozoan cover for each *Sargassum* species, categorized under three cover categories: total, sections, and structures. To observe similarities in the community structure of hydrozoans among *Sargassum* species, their structures, sections, and total hydrozoan coverage on *Sargassum* thalli, we performed an nMDS using the Bray-Curtis method (Kruskal and Wish, 1978) with the prior transformation of the fourth root ($\sqrt[4]{}$) to reduce the influence of dominant taxa of the percentage coverages of each hydrozoan species (Clarke, 1993), using the PRIMER program version 7 (Clarke and Gorley, 2015).

To support the previous analyses, a Nonmetric Multidimensional Scaling (nMDS) ordering method was obtained using the Bray-Custis as a measure of similarity, and vectors of the sections and structures of the *Sargassum* were adjusted with the 'envfit' function with the 'vegan' package (Oksanen et al., 2007).

3. Results

We identified 18 hydrozoan epibiont species on Sargassum, with 16 belonging to the superorder Leptothecata (88.9%) and two to anthoathecate (11.1%). Obelia cf. dichotoma was found on all Sargassum species except S. muticum (Table 2). Among the BSS, we found 11 hydrozoan species, 10 Leptothecata, and one anthoathecate (Table 2). Six species were detected in Sargassum horneri and S. muticum, and three in S. horridum (Table 2, Fig. 3). The hydrozoans Hydrodendron sp. 1, Dynamena disticha, and Orthopyxis sp. 1 were shared between S. horneri and S. muticum. Two species were exclusive to S. horneri, while three and two species were found exclusively in S. muticum and S. horridum, respectively (Fig. 3). In HSS, we found eight hydrozoan species, seven Leptothecata, and one anthoathecate. Sargassum fluitans III had seven species, while S. natans VIII had eight, and all the hydrozoans were found on both Sargassum species except for Clytia cf. gracilis (Table 2, Fig. 3). Sargassum natans VIII had one exclusive hydrozoan species (Fig. 3). In contrast, S. fluitans III did not host any exclusive hydrozoans.

Some BSS thalli exhibited hydrozoan colonies with gonophores, viz. Orthopyxis sp. 1 and D. disticha on S. horneri, Orthopyxis sp. 1 on S. muticum, and O. cf. dichotoma and V. halecioides on S. horridum. For HSS, C. noliformis and C. cf. hemisphaerica with gonophores were found on S. fluitans III, and C. noliformis and O. cf. dichotoma colonies with gonophores were observed on S. natans VIII.

The total percentage coverage of hydrozoans on BSS showed less variability than on HSS. For example, *Sargassum natans* VIII exhibited cover values ranging from 0.05% to 100% (Fig. 4A). The hydrozoan with the highest mean percentage coverage of epibiosis for each *Sargassum* species were *Dynamena disticha* for *S. horneri* (0.6%), *Orthopyxis* sp. 1 for *S. muticum* (2.7%), *Ventromma halecioides* for *S. horridum* (11.0%), *Clytia* noliformis for *S. fluitans* III (31.5%), *Aglaophenia latecarinata* for *S. natans*

VIII (41.0%) (Table 2).

Regarding benthic species, *S. horneri* displayed low percentage coverages across all thalli sections. In contrast, *Sargassum muticum* exhibited the most significant variability in the basal area, while *S. horridum* showed the highest variation in the upper-middle section (Fig. 4B). Notably, the variation in hydrozoan cover on thalli structures was lower for BSS than for HSS (Fig. 4C). Within the BSS, the basal section hosted the highest number of hydrozoan species (*S. horneri* = 6; *S. muticum* = 6; *S. horridum* = 3). *Sargassum horneri* did not exhibit hydrozoans in the apical section. On the other hand, *S. muticum* displayed the lowest number of hydrozoan species (1) in the upper-middle, upper-middle, and apical sections (Table 2, SM 1).

The hydrozoans with the highest mean coverage percentages in each section were *D. disticha* in the basal section of *S. horneri* (1.4%), *Orthopyxis* sp. 1 in the basal section of *S. muticum* (9.0%), and *V. halecioides* in the upper-middle section of *S. horridum* (18.1%) (Table 2, SM 1). Regarding structures, the BSS *S. horneri* had the highest number of hydrozoan species on the blades (6 species). In comparison, *S. muticum* had five species on the axis, and *S. horridum* had three species on both the axis and blades. Among the HSS, *S. fluitans* III hosted six species on all its structures (Table 2, SM 1).

The hydrozoans with the highest mean percentages on structures were *D. disticha* on the holdfast (10.0%) of *S. horneri*, *Orthopyxis* sp. 1 on the axis (6.6%) of *S. muticum*, *V. halecioides* on the axis (16.8%) of *S. horridum*, *C. noliformis* on the axis (41.0%) of *S. fluitans* III, and *A. latecarinata* on the axis (41.0%) of *S. natans* VIII (Table 2, SM 1).

The nMDS ordination analyses of the composition of hydrozoan species in the sections and structures of different thalli of *Sargassum* species showed striking similarities, indicating no significant differentiation. However, the cluster for the total hydrozoan cover along *Sargassum* thalli evidenced a separation based on *Sargassum* species and geographic location (Fig. 5).

4. Discussion

Our results demonstrate that differences in hydrozoan species primarily depend on the geographical distribution of the *Sargassum* species. This observation aligns with a global trend where geographical regions exhibit similar hydrozoan-macrophyte associations, with the hydrozoan composition being secondarily influenced by the macrophyte lineages (Oliveira and Marques, 2011). On the other hand, the composition and abundance of epibiont hydrozoans on structures along the thalli of both benthic and holopelagic *Sargassum* species were highly variable without a pattern.

The number of hydrozoan epibionts found on BSS, between 3 and 6 species, was relatively low compared to the number reported for other BSS in previous studies, such as 14 hydrozoan species on Brazilian *Sargassum cymosum* (Cunha and Jacobucci, 2010). In contrast, the number of hydrozoan epibionts on HSS (seven for *S. fluitans* III and eight for *S. natans* VIII) was higher than previously reported for Mexican

Table 2

Mean percentage cover (\pm standard deviation) of hydrozoans according to *Sargassum* species, thalli sections (B=Basal, LM=Lower Middle, UM=Upper Middle, A=Apical), and structures (axis, blades, aerocysts, receptacles, holdfast). *Sargassum* sections and structures not observed or without hydrozoan cover are not included. * anthoathecate species, the rest of the species belong to the superorder Leptothecata. • New records of hydrozoans on *Sargassum* (species category).

		Thalli sections		Morphology		
Sargassum species	Hydrozoan species	Section	Mean % cover	Structure	Mean % cover	Mean % total
Sargassum horneri	Aglaophenia pinguis Fraser, 1938	В	0.19 ± 0.39	Axis	0.55 ± 1.46	0.08 ± 0.2
		LM	0.04 ± 0.12	Blades	0.26 ± 0.68	
	Dynamena disticha (Bosc, 1802)●	В	1.38 ± 4.13	Axis	$\textbf{2.86} \pm \textbf{5.98}$	0.60 ± 1.78
		LM	0.53 ± 2.05	Blades	0.41 ± 1.42	
		UM	0.1 ± 0.36	Holdfast	10 ± 30.78	
	Hydrodendron sp. 1	В	0.15 ± 0.53	Axis	$\textbf{0.80} \pm \textbf{2.62}$	$\textbf{0.09} \pm \textbf{0.30}$
		LM	0.04 ± 0.20	Blades	0.01 ± 0.04	
	Obelia cf. dichotoma (Linnaeus, 1758)●	В	0.26 ± 1.18	Axis	0.72 ± 03.22	0.15 ± 0.68
				Blades	0.07 ± 0.31	0.00 . 0.64
	Orthopyxis sp. 1	В	0.75 ± 1.43	Axis	4.23 ± 8.65	0.30 ± 0.61
		LIM	0.10 ± 0.27	Blades	0.12 ± 0.48	
	Diumularia on 1	P	0.01 0.05	Pladas	5 ± 22.30	0.002 ± 0.02
Saraassum muticum	Dumamana disticha (Bose, 1802)	B	0.01 ± 0.03 0.12 \pm 0.37	Avie	0.004 ± 0.02 0.21 \pm 0.60	0.003 ± 0.02
Surgussum muticum	Dynamena disticha (bosc, 1802)	IM	0.12 ± 0.37 0.08 ± 0.34	Blades	0.21 ± 0.00 0.01 ± 0.03	0.03 ± 0.13
	Fudendrium sp. 1 *	B	0.00 ± 0.04 0.51 ± 1.59	Avis	0.01 ± 0.03 0.19 ± 0.61	0.15 ± 0.31
	Educits tan 5p. 1	LM	0.37 ± 1.39	Blades	0.09 ± 0.01 0.09 ± 0.28	0.10 ± 0.01
	Haleciidae sp. 1	В	1.63 ± 6.31	Axis	0.37 ± 1.30	0.12 ± 0.39
	· · · · · · · · · · · · · · · · · · ·	А	0.04 ± 0.16	Blades	0.01 ± 0.04	
	Hydrodendron sp. 1	В	0.10 ± 0.46	Blades	0.02 ± 0.09	0.02 ± 0.08
	Orthoypxys sp.1	В	9.04 ± 13.82	Axis	6.56 ± 10.35	2.71 ± 4.25
		LM	4.50 ± 11.68	Blades	1.17 ± 2.36	
		UM	1.02 ± 2.65	Aerocysts	0.01 ± 0.05	
		Α	0.25 ± 1.10			
	Sertularellidae sp. 1	В	0.79 ± 3.53	Axis	0.30 ± 1.36	$\textbf{0.08} \pm \textbf{0.37}$
Sargassum horridum	Dynamena quadridentata (Ellis & Solander, 1786)	В	2.63 ± 11.16	Axis	$\textbf{2.25} \pm \textbf{8.86}$	$\textbf{0.89} \pm \textbf{3.82}$
		LM	0.91 ± 4.05	Blades	$\textbf{0.46} \pm \textbf{2.06}$	
		UM	0.14 ± 0.64			
	Obelia cf. dichotoma (Linnaeus, 1758)	В	0.17 ± 0.76	Axis	0.10 ± 0.46	0.22 ± 0.56
		A	1.13 ± 3.15	Blades	0.23 ± 0.72	
	Ventromma halecioides (Calder, 1859)	В	6.12 ± 5.55	Axis	16.78 ± 11.86	11.01 ± 7.11
		LM	8.39 ± 6.46	Blades	7.85 ± 4.98	
		UNI	18.17 ± 12.84	Aerocysts	1.96 ± 5.61	
		A	12.77 ± 15.50	Holdfoot	0.35 ± 1.14	
Samassum fluitans III	Aglaophania latecarinata Allmon, 1877	NA	NA	Avie	2.5 ± 11.10 0.04 \pm 0.17	0.1 ± 0.02
	Clutia of hemisphaerica (Lippaeus 1767)	NA	NΔ	Avis	0.04 ± 0.17 3.25 ± 0.71	0.1 ± 0.02 4 65 + 11 12
	Ciyiu ci. nemisphaericu (Linnacus, 1707)	1474	1471	Blades	3.25 ± 9.71 3.96 ± 9.72	4.05 ± 11.12
				Aerocysts	3.75 ± 8.54	
	Clytia noliformis (McCrady, 1859)	NA	NA	Axis	41.00 ± 19.15	31.52 ± 13.17
	5 5 5 5 5			Blades	28.60 ± 14.97	
				Aerocysts	21.31 ± 13.22	
	Halopteris sp. 1	NA	NA	Axis	0.05 ± 0.13	$\textbf{0.04} \pm \textbf{0.08}$
				Blades	$\textbf{0.02} \pm \textbf{0.08}$	
				Aerocysts	0.01 ± 0.05	
	Obelia cf. dichotoma (Linnaeus, 1758)	NA	NA	Blades	0.01 ± 0.03	$\textbf{0.01} \pm \textbf{0.03}$
	Plumularia strictocarpa Pictet, 1893	NA	NA	Blades	0.02 ± 0.08	0.03 ± 0.11
				Aerocysts	0.02 ± 0.09	
	Zanclea alba (Meyen, 1834)*●	NA	NA	Axis	0.32 ± 0.96	2.08 ± 6.55
				Blades	0.49 ± 1.78	
Carraceum natane VIII	Adaphania lateraringta Allmon, 1977	NA	NA	Aerocysts	0.49 ± 1.52	40.00 ± 25.26
	Aguophenia ialecarinata Alinian, 1877	INA	INA	Blader	40.38 ± 23.39 34.08 ± 10.05	40.90 ± 23.30
				Aerocysts	34.00 ± 19.03 20.20 \pm 26.33	
	Clytia of gracilis (Sars 1851)	NA	NA	Avis	0.02 ± 0.10	0.03 ± 0.15
	Gyna ci. gracius (Sais, 1031)	1474	1471	Blades	0.02 ± 0.10 0.01 ± 0.05	0.03 ± 0.13
				Aerocysts	0.01 ± 0.00 0.08 ± 0.38	
	Clytia cf. hemisphaerica (Linnaeus, 1767)	NA	NA	Axis	0.35 ± 1.05	0.33 ± 0.76
	- J J			Blades	0.19 ± 0.50	
				Aerocysts	0.46 ± 1.19	
	Clytia noliformis (McCrady, 1859)	NA	NA	Axis	$\textbf{8.12} \pm \textbf{19.15}$	$\textbf{9.22} \pm \textbf{19.91}$
				Blades	$\textbf{8.65} \pm \textbf{18.76}$	
				Aerocysts	$\textbf{6.61} \pm \textbf{14.13}$	
	Halopteris diaphana (Heller, 1868)	NA	NA	Axis	11.02 ± 17.23	$\textbf{0.14} \pm \textbf{0.33}$
				Blades	$\textbf{0.17} \pm \textbf{0.53}$	
				Aerocysts	$\textbf{0.08} \pm \textbf{0.23}$	
	Obelia cf. dichotoma (Linnaeus, 1758)●	NA	NA	Axis	$\textbf{0.88} \pm \textbf{1.40}$	$\textbf{0.49} \pm \textbf{1.12}$
				Blades	0.26 ± 0.77	
	Plant Internet Plant 1000 -	214	NTA	Aerocysts	0.56 ± 1.37	0.00 + 0.50
	Pumularia strictocarpa Pictet, 1893	NA	NA	AXIS	0.49 ± 1.24 0.13 ± 0.20	0.23 ± 0.58
				Diaues	0.13 ± 0.39	

(continued on next page)

Table 2 (continued)

		Thalli sections		Morphology		
Sargassum species	Hydrozoan species	Section	Mean % cover	Structure	Mean % cover	Mean % total
	Zanclea alba (Meyen, 1834)*			Aerocysts Axis Blades Aerocysts	$\begin{array}{c} 0.31 \pm 1.00 \\ 1.67 \pm 2.75 \\ 0.23 \pm 0.29 \\ 1.09 \pm 1.43 \end{array}$	0.59 ± 0.61



Fig. 3. Venn diagram illustrating shared hydrozoan species among *Sargassum* species.

waters (four on *S. fluitans* III and five on *S. natans* VIII; Mendoza-Becerril et al., 2020a). However, these differences need to be analyzed considering sampling at different times of the year, since temporal variation in hydrozoan species richness and abundance has been observed on *Sargassum* (i.e., Mendoza-Becerril et al., 2020a).

Each *Sargassum* species exhibited one hydrozoan epibiont species with a marked higher mean total coverage, a trend noted in other studies (Cunha and Jacobucci, 2010; Mendoza-Becerril et al., 2020a). For BSS, *D. disticha* on *S. horneri, Orthopyxis* sp. 1 on *S. muticum*, and *V. halecioides* on *S. horridum* had the highest mean coverage along the thallus. Among HSS, *C. noliformis* displayed the highest mean coverage on *S. fluitans* III,

while *A. latecarinata* presented that on *S. natans* VIII. The preference of *A. latecarinata* for *S. natans* VIII has been previously documented (Burkenroad in Parr, 1939; Govindarajan et al., 2019). However, *A. latecarinata* has also been found abundantly on *S. fluitans* III in some cases (Burkenroad in Parr, 1939; Govindarajan et al., 2019), a different pattern from our data. Space competition can be severe on algal surfaces (Oswald and Seed, 1986), and most likely, this is the case between *A. latecarinata* and *C. noliformis* in our *S. fluitans* III samples, in which the latter was the dominant species, as observed in previous studies (Mendoza-Becerril et al., 2020a; Alleyne et al., 2023).

Regarding the sections along BSS thalli, the basal section hosted the highest number of species. This is consistent with observations in other phaeophytes, where hydrozoans tend to attach to basal areas and grow upwards, decreasing abundance towards distal sections (Fraschetti et al., 2006). However, hydrozoans exhibited higher coverage in the upper-middle section of *S. horridum*, suggesting that they grew along with new algal growth, similar to other hydrozoan epibionts on *Sargassum* (Ryland, 1974). The number of hydrozoan epibiont species varied among *Sargassum* vegetative structures with more hydrozoan species on the axis of *Sargassum* have also been documented by Mendoza-Becerril et al. (2020a). In contrast, other marine organisms like bryozoans prefer aerocysts as settlement substrates (Niermann, 1986).

Regarding the reproductive characteristics of hydrozoans, we observed a prevalence of species with fixed gonophores (cf., Fraschetti et al., 2006; Cunha and Jacobucci, 2010). This suppression of the medusa stage can be considered a strategy to restrict the offspring within the same algal substrate, promoting population stability over dispersal (Ronowicz et al., 2008). Notably, some of the most abundant hydrozoan species bearing gonophores were only found on a few thalli. This pattern might be derived from asexual reproduction, which could efficiently sustain colony growth on brown algae (Gravier-Bonnet, Bourmaud, 2005).

According to the last revision of Carral-Murrieta et al. (2023a) about



Fig. 4. Percentage coverage (%) of hydrozoans on *Sargassum* species. A) Total percentage coverage. B) Percentage coverage of hydrozoans on different sections of benthic *Sargassum* (B = Basal, LM = Lower Middle, UP = Upper Middle, A = Apical). C) Percentage coverage of hydrozoans on different structures of *Sargassum* (a = aerocysts, b = blades, h = holdfast, r = receptacles, x = axis).



Fig. 5. Nonmetric Multidimensional Scaling showing the similitudes in the community structure of hydrozoans among *Sargassum* species. A) Hydrozoan coverage per structure on *Sargassum* thalli, B) Total hydrozoan coverage per structure on *Sargassum*, C) Hydrozoan coverage per section on *Sargassum* thalli, D) Total hydrozoan coverage per section on *Sargassum*.

the number of epibiont species of *Sargassum* in the world, only one species (*Clytia* cf. *gracilis*) is recorded for the first time as epibiont for HSS, growing on *S. natans* VIII. In addition, six species of hydroids are reported for the first time for *S. horneri*, four for *S. muticum*, two for *S. fluitans* III, and three for *S. natans* VIII. Additionally, we report species of *Hydrodendron* and *Eudendrium* for the first time on *Sargassum* in the Pacific Ocean (see Carral-Murrieta et al., 2023b).

The species O. cf. *dichotoma* exhibits a circumglobal distribution and was found on both BSS and HSS in the Caribbean and the Mexican Pacific (Lindner and Migotto, 2002). Also, it has been documented in the Atlantic, Indian, and Pacific Oceans (Millard, 1975). However, it is important to consider that this species exhibits recognized morphological variability and cryptic lineages (Cunha et al., 2017; Penney and Rawlings, 2021). Moreover, many hydrozoan species considered cosmopolitan may fall into a pseudo-cosmopolitan category, explained as a taxon comprised of more than one lineage but with a single binomial name with a false cosmopolitan status (Darling and Carlton, 2018; Calder, 2021).

Interestingly, the BSS *S. horneri* and *S. muticum*, collected in the same locality (Ensenada) simultaneously, only shared three hydrozoan

species, which can be attributed to differences in structural complexity, seasonal reproductive growth patterns, and life cycles of these two seaweeds (Cunha and Jacobucci, 2010). Conversely, the HSS shared all hydrozoan species except *C*. cf. *gracilis*, which has been reported on other BSS (e.g. Cunha and Jacobucci, 2010). This could be due to the opportunistic nature and low substrate selectivity typically exhibited by most hydrozoans (Oliveira and Marques, 2011), as well as the shared origin of *S. fluitans* III and *S. natans* VIII (Godínez-Ortega et al., 2021).

In conclusion, our findings suggest that differences in hydrozoan species primarily depend on geographical distribution of the *Sargassum* species. *Sargassum* species from different locations exhibit unique hydrozoan compositions and there is no pattern of epibiosis concerning the algal morphology (structures and sections) for each *Sargassum* species.

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SM 1. Nonmetric Multidimensional Scaling (nMDS) of sections and structures of hydrozoan epibionts on Sargassum. A-B) Hydrozoan cover per structure on Sargassum thalli, C-D) Hydrozoan cover per section on Sargassum thalli.

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CRediT authorship contribution statement

COCM: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Validation, Visualization, Writing - original draft; Writing - review & editing. **ESZ**: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Supervision, Validation, Visualization, Writing - original draft; Writing - review & editing. FRCR: Formal analysis, Methodology, Supervision, Visualization. ACM: Supervision, Validation, Writing - original draft; Writing - review & editing. MAMB: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Supervision, Validation, Visualization, Writing - original draft; Writing - review & editing. Writing - original draft; Writing - review & editing.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data Availability

Data will be made available on request.

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Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.aquabot.2023.103738.

References

- Abé, H., Komatsu, T., Kokubu, Y., Natheer, A., Rothausler, E.A., Shishido, H., Shizuha, Y., Ajisaka, T., 2013. Invertebrate fauna associated with floating *Sargassum horneri* (Fucales: Sargassaceae) in the East China Sea. Species Divers. 18 (1), 75–85. https:// doi.org/10.12782/sd.18.1.075.
- Aguilar-Rosas, L.E., Aguilar-Rosas, R., Kawai, H., Uwai, S., Valenzuela-Espinoza, E., 2007. New record of Sargassum filicinum Harvey (Fucales, Phaeophyceae) in Mexico's Pacific coast. Algae 22 (1), 17–21. https://doi.org/10.4490/ algae.2007.22.1.017.
- Aguilar-Rosas, L.E., Pedroche, F.F., Zertuche-González, J.A., 2014. Algas Marinas no nativas en la costa del Pacífico Mexicano. Especies Acuáticas Invasoras en México. Comisión Nacional para el Conocimiento y Uso de la Biodiversidad, Mexico, pp. 211–222.
- Aguilar-Rosas, R., Galindo, A.M., 1990. Ecological aspects of Sargassum muticum (Fucales, Phaeophyta) in Baja California, Mexico: reproductive phenology and epiphytes. Thirteenth International Seaweed Symposium: Proceedings of the Thirteenth International Seaweed Symposium held in Vancouver, Canada. Springer, Netherlands, pp. 185–190.
- Alleyne, K.S., Neat, F., Oxenford, H.A., 2023. A baseline assessment of the epiphytic community associated with pelagic *Sargassum* in the Tropical Atlantic. Aquat. Bot. 186, 103635 https://doi.org/10.1016/j.aquabot.2023.103635.
- Andrade-Sorcia, G., Riosmena-Rodriguez, R., Muniz-Salazar, R., Lopez-Vivas, J.M., Boo, G.H., Lee, K.M., Boo, S.M., 2014. Morphological reassessment and molecular assessment of *Sargassum* (Fucales: Phaeophyceae) species from the Gulf of California, Mexico. Phytotaxa 183 (4), 201–223. https://doi.org/10.11646/phytotaxa.183.4.1.
- Boero, F., 1984. The ecology of marine hydroids and effects of environmental factors: a review. Mar. Ecol. 5 (2), 93–118.
- Britton-Simmons, K.H., 2004. Direct and indirect effects of the introduced alga Sargassum muticum on benthic, subtidal communities of Washington State, USA. Mar. Ecol. Prog. Ser. 277, 61–78. https://doi.org/10.3354/meps277061.
- Calder, D.R., 1988. Shallow-water hydroids of Bermuda. The Athecatae. R. Ont. Mus. Life Sci. Contrib. 148, 1–107. https://doi.org/10.5962/bhl.title.52225.
- Calder, D.R., 1991. Shallow-water hydroids of Bermuda. The Thecatae, exclusive of Plumularioidea. Bull. Mar. Sci. 49 (3) https://doi.org/10.2307/1352274.
 Calder, D.R., 1997. Shallow-water hydroids of Bermuda: superfamily Plumularioidea
- (Vol. 3). R. Ont. Mus. Life Sci. Contrib. 161, 1-85. Calder, D.R., 2020. Some leptothecate hydroids (Cnidaria, Hydrozoa) from Hawaii,
- mostly from inshore and nearshore waters. Zootaxa 4830 (2), 201–246. https://doi. org/10.11646/zootaxa.4830.2.1.
- Calder, D.R., Choong, H.H., Carlton, J.T., Chapman, J.W., Miller, J.A., Geller, J., 2014. Hydroids (Cnidaria: Hydrozoa) from Japanese tsunami marine debris washing ashore in the northwestern United States. Aquat. Invasions 9 (4), 425–440. https:// doi.org/10.3391/ai.2014.9.4.02.
- Calder, D.R., Carlton, J.T., Keith, I., Larson, K., McCann, L., Geller, J., Wheelock, M., Choong, H.H., Ruiz, G.M., 2021. Additions to the hydroids (Cnidaria, Hydrozoa) of marine fouling communities on the mainland of Ecuador and in the Galapagos Islands. Aquat. Invasions 16 (2), 208–252.. https://doi.org/10.3391/ ai.2021.16.2.02.
- Carral-Murrieta, C.O., Serviere-Zaragoza, E., Mendoza-Becerril, M.A., 2023b. Sargassum species as hydrozoans substrates. Zenodo. https://doi.org/10.5281/ zenodo.8360133.
- Carral-Murrieta, C.O., 2021. Buenas prácticas de colecta de macroalgas. Medusozoa México. Infographic. https://medusozoamexico.com.mx/divulgaci%C3%B3n

/f/buenas-pr%C3%A1cticas-de-colecta-de-macroalgas-para-estudiar-hidroides?blogcategory=Infograf%C3%ADa.

- Carral-Murrieta, C.O., Marques, A.C., Serviere-Zaragoza, E., Estrada-González, M.C., Cunha, A.F., Fernandez, M.O., Mazariegos-Villarreal, A., León-Cisneros, K., Agüero, J., Mendoza-Becerril, M.A., 2023a. A survey of epibiont hydrozoans on Sargassum. PeerJ 11, e15423. https://doi.org/10.7717/peerj.15423.
- Carvalho, N.F., e Silva, R.C., Rosa Filho, J.S., Jacobucci, G.B., 2022. Which structural traits in *Sargassum* species really matter for caprellid assemblages? Estuar. Coast. Shelf Sci. 265, 107703 https://doi.org/10.1016/j.ecss.2021.107703.
- Clarke, K.R., 1993. Non-parametric multivariate analyses of changes in community structure. Aust. J. Ecol. 18, 117–143.
- Clarke, K.R., Gorley, R.N., 2015. Getting started with PRIMER v7. PRIMER-E: plymouth. Plymouth Mar. Lab. 20.
- Coronado, C., Candela, J., Iglesias-Prieto, R., Sheinbaum, J., López, M., Ocampo-Torres, F.J., 2007. On the circulation in the Puerto Morelos fringing reef lagoon. Coral Reefs 26, 149–163. https://doi.org/10.1007/s00338-006-0175-9.
- Cunha, A.F., Jacobucci, G.B., 2010. Seasonal variation of epiphytic hydroids (Cnidaria: Hydrozoa) associated to a subtropical Sargassum cymosum (Phaeophyta: Fucales) bed. Zool. (curitiba) 27, 945–955. https://doi.org/10.1590/S1984-46702010000600016.
- Cunha, A.F., Collins, A.G., Marques, A.C., 2017. Phylogenetic relationships of Proboscoida Broch, 1910 (Cnidaria, Hydrozoa): are traditional morphological diagnostic characters relevant for the delimitation of lineages at the species, genus, and family levels? Mol. Phylogenetics Evol. 106, 118–135. https://doi.org/10.1016/ i.ympey.2016.09.012.
- Darling, J.A., Carlton, J.T., 2018. A framework for understanding marine cosmopolitanism in the Anthropocene. Front. Mar. Sci. 5, 293 https://doi.org/ 10.3389/fmars.2018.00293.
- Fitridge, I., Dempster, T., Guenther, J., De Nys, R., 2012. The impact and control of biofouling in marine aquaculture: a review. Biofouling 28 (7), 649–669. https://doi. org/10.1080/08927014.2012.700478.
- Fraschetti, S., Terlizzi, A., Bevilacqua, S., Boero, F., 2006. The distribution of hydroids (Cnidaria, Hydrozoa) from micro-to macro-scale: spatial patterns on habitat-forming algae. J. Exp. Mar. Biol. Ecol. 339 (2), 148–158.
- Frick, M.G., Ross, A., Williams, K.L., Bolten, A.B., Bjorndal, K.A., Martins, H.R., 2003. Epibiotic associates of oceanic-stage loggerhead turtles from the southeastern North Atlantic. Mar. Turt. Newsl. 101, 18–20.
- Godínez-Ortega, J.L., Cuatlán-Cortés, J.V., López-Bautista, J.M., van Tussenbroek, B.I., 2021. A natural history of floating *Sargassum* species (Sargasso) from Mexico. Nat. Hist. Ecol. Mex. Cent. Am. 59–94. https://doi.org/10.5772/intechopen.97230.
- Govindarajan, A.F., Cooney, L., Whittaker, K., Bloch, D., Burdorf, R.M., Canning, S., Carter, C., Cella, S.M., Eriksson, F.A., Freyer, H., Huston, G., Hutchinson, S., McKeegan, K., Malpani, M., Merkle-Raymond, A., Ouellete, K., Petersen-Rockney, R., Schultz, M., Siuda, A.N.S., 2019. The distribution and mitochondrial genotype of the hydroid Aglaophenia latecarinata is correlated with its pelagic Sargassum substrate type in the tropical and subtropical western Atlantic Ocean. PeerJ 7, e7814. https:// doi.org/10.7717/peerj.7814.
- Gower, J., King, S., 2019. Seaweed, seaweed everywhere. Science 365 (6448). https:// doi.org/10.1126/science.aay0989.
- Gower, J., Young, E., King, S., 2013. Satellite images suggest a new Sargassum source region in 2011. Remote Sens. Lett. 4, 764–773.. https://doi.org/10.1080/ 2150704X.2013.79643.
- Gravier-Bonnet, N., Bourmaud, C., 2005. Cloning by releasing specialized frustules in a successful epiphytic zooxanthellate haleciid (Cnidaria, Hydrozoa, Haleciidae), with comments on stolonization and frustulation. Invertebr. Reprod. Dev. 48 (1–3), 63–69
- Guiry, M.D., Guiry, G.M., 2023. AlgaeBase. World-Wide Electronic Publication, National University of Ireland, Galway.
- Jackson, J.B.C., 1977. Competition on marine hard substrata: the adaptive significance of solitary and colonial strategies. Am. Nat. 111 (980), 743–767.
- Kruskal, J.B., Wish, M., 1978. Multidimensional Scaling. Sage Publications, Beverly Hills.
 J. Larsson _eulerr: Area-Proportional Euler and Venn Diagrams with Ellipses_. R package version 7.0.0 2022.https://CRAN.R-project.org/package=eulerr.
- Lindner, A., Migotto, A.E., 2002. The life cycle of *Clytia linearis* and *Clytia noliformis*: metagenic campanulariids (Cnidaria: Hydrozoa) with contrasting polyp and medusa stages. J. Mar. Biol. Assoc. U. Kingd. 82 (4), 541–553.. https://doi.org/10.1017/ S0025315402005866.
- Mancilla, P.M., Martínez, G.M., 1991. Variación estacional de temperatura, salinidad y oxígeno disuelto en la Bahía de Todos Santos, BC, México (marzo de 1986 a junio de 1987). Rev. De. Invest. Científica 2, 33–45.
- Mendoza-Becerril, M.A., Serviere-Zaragoza, E., Mazariegos-Villareal, A., Rivera-Pérez, C., Calder, D.R., Vazquez-Delfín, E.F., Freile-Pelegrín, Y., Agüero, J., Robledo, D., 2020a. Epibiont hydroids on beachcast *Sargassum* in the Mexican Caribbean. PeerJ 8, e9795 https//doi.10.7717/peerj.9795.
- Mendoza-Becerril, M.A., Estrada-Gonzalez, M.C., Mazariegos-Villarreal, A., Restrepo-Avendano, L., Villar-Beltran, R.D., Agüero, J., Cunha, A.F., 2020b. Taxonomy and diversity of Hydrozoa (Cnidaria, Medusozoa) of La Paz Bay, Gulf of California. Zootaxa 4808 (1), 1–37. https://doi.org/10.11646/zootaxa.4808.1.1.
- Merino, M., Otero, L., 1991. Atlas Ambiental Costero, Puerto Morelos-Quintana Roo. Centro de Investigaciones de Quintana Roo (CIQRO, Chetumal, México.
- Millard, N.A.H., 1975. Monograph on the hydroida of southern Africa. Ann. S. Afr. Mus. 68, 1–513.
- Mills, C.E., Marques, A.C., Migotto, A.E., Calder, D.R., Hand, C.A., Rees, J.T., Haddock, S., Dunn, C., Pugh, P.R., 2007. Hydrozoa: polyps, hydromedusae, and siphonophore. The Light and Smith manual: intertidal invertebrates from central California to Oregon. University of California Press, Berkeley, pp. 118–167.

Mincer, T.J., Bos, R.P., Zettler, E.R., Zhao, S., Asbun, A.A., Orsi, W.D., Vincent, S.G., Amaral-Zettler, L.A., 2023. Sargasso Sea Vibrio bacteria: underexplored potential pathovars in a perturbed habitat. Water Res, 120033. https://doi.org/10.1016/j. watres.2023.120033.

Monteforte, M., Cariño, M., 1992. Exploration and evaluation of natural stocks of pearl oysters *Pinctada mazatlanica* and *Pteria sterna* (Bivalvia: Pteriidae): La Paz Bay, Baja California Sur, Mexico. Ambio. 21 (4), 314–320.

- Niermann, U., 1986. Distribution of Sargassum natans and some of its epibionts in the Sargasso Sea. Helgoländer Meeresunters. 40, 343–353. https://doi.org/10.1007/ BF01983817.
- Nishihira, M., 1965. The association between Hydrozoa and their attachment substrata with special reference to algal substrata. Bull. Mar. Biol. Station Asamushi, Tohoku Univ. 12 (2–3), 75–92.
- Nishihira, M., 1968. Brief experiments on the effect of algal extracts in promoting the settlement of the larvae of *Coryne uchidai* Stechow (Hydrozoa). Bull. Mar. Biol. Station Asamushi, Tohoku Univ. 1, 91–101.
- Nishihira, M., 1973. Ecological distribution of epiphytic Hydrozoa with special reference to Sertularella miurensis. Publ. Seto Mar. Biol. Lab. 20, 401–418.
- Obeso-Nieblas, M., Shirasago, B., Sanchez-Velasco, L., Gaviño-Rodríguez, J.H., 2004. Hydrographic variability in Bahia De La Paz, BC S, Mexico, during the 1997–1998 El Niño. Deep Sea Res. Part II: Top. Stud. Oceanogr. 51 (6–9), 689–710.
- Oksanen, J., Simpson, G., Blanchet, F., Kindt, R., Legendre, P., Minchin, P., O'Hara, R., Solymos, P., Stevens, M., Szoecs, E., Wagner, H., Barbour, M., Bedward, M., Bolker, B., Borcard, D., Carvalho, G., Chirico, M., De Caceres, M., Durand, S., Evangelista, H., FitzJohn, R., Friendly, M., Furneaux, B., Hannigan, G., Hill, M., Lahti, L., McGlinn, D., Ouellette, M., Ribeiro Cunha, E., Smith, T., Stier, A., Ter Braak, C., Weedon, J. (2007). _vegan:Community Ecology Package_. R package version 2.6–2, https://CRAN.R project.org/package=vegan.
- Oliveira, O.M.P., Marques, A.C., 2007. Epiphytic hydroids (Hydrozoa: Anthoathecata and Leptothecata) of the world. Check List 3 (1), 21–38. https://doi.org/10.15560/ 3.1.21.
- Oliveira, O.M.P., Marques, A.C., 2011. Global and local patterns in the use of macrophytes as substrata by hydroids (Hydrozoa: Anthoathecata and Leptothecata). Mar. Biol. Res. 7 (8), 786–795. https://doi.org/10.1080/17451000.2011.578647.
- Oswald, R.C., Seed, R., 1986. Organisation and seasonal progression within the epifaunal communities of coastal macroalgae. Cah. De. Biol. Mar. 27 (1), 29–40.
- Parr, A.E., 1939. Quantitative observations on the pelagic Sargassum vegetation of the western North. Atl.: Prelim. Discuss. Morphol. Relatsh. Bull. Bingham Oceanogr. Collect. 6, 1–94.
- Penney, M.S., Rawlings, T.A., 2021. An examination of shallow-water hydroids (Cnidaria, Hydrozoa, Hydroidolina) in Cape Breton, Nova Scotia, using morphology and DNA barcoding. Northeast. Nat. 28 (m18), 1–38. https://doi.org/10.1656/ 045.028.m1801.

- Pérez-Posada, I., Cabanillas-Terán, N., Rosas-Luis, R., Hernández-Arana, H.A., Sánchez-Gonzalez, A., 2023. Isotopic niche shift in the sea urchins *Echinometra lucunter* and *E. viridis* after massive arrivals of *Sargassum* in the Mexican Caribbean. Reg. Stud. Mar. Sci. 103064 https://doi.org/10.1016/j.rsma.2023.103064.
- R Development Core Team, 2023. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna.
- Rackley, D.H., 1974. Hydroids of the pelagic Sargassum community of the Gulf Stream and Sargasso Sea. Thesis. https://doi.org/10.25773/v5-waa7-4e36.
 Rocha-Ramírez, V., Siqueiros-Beltrones, D.A., 1990. Review of the species of the genus
- Sargassa, C. Agardh recorded for Bahía de La Paz, BCS, México. Cienc. Mar. 16 (3), 15–26.
- Rodríguez-Martínez, R.E., Medina-Valmaseda, A.E., Blanchon, P., Monroy-Velázquez, L. V., Almazán-Becerril, A., Delgado-Pech, B., Vásquez-Yeomans, L., Francisco, V., García-Rivas, M.C., 2019. Faunal mortality associated with massive beaching and decomposition of pelagic Sargassum. Mar. Pollut. Bull. 146, 201–205. https://doi.org/10.1016/j.marpolbul.2019.06.015.
- Ronowicz, M., Włodarska-Kowalczuk, M., Kuklinski, P., 2008. Factors influencing hydroids (Cnidaria: Hydrozoa) biodiversity and distribution in Artic kelp forest. J. Mar. Biol. Assoc. U.K. 88 (8), 1567–1575. https://doi.org/10.1017/ S0025315408001495.
- Ryland, J., 1974. Observations on some epibionts of gulf-weed, Sargassum natans (L.) Meyen, J. Exp. Mar. Biol. Ecol. 14 (1), 17–25.
- Schell, J.M., Goodwin, D.S., Siuda, A.N.S., 2015. Recent Sargassum inundation events in the Caribbean: Shipboard observations reveal dominance of a previously rare form. Oceanography 28 (3), 8–10. https://doi.org/10.5670/oceanog.2015.70.
- Simental, L., Martínez-Urtaza, J., 2008. Climate patterns governing the presence and permanence of salmonellae in coastal areas of Bahia de Todos Santos, Mexico. Appl. Environ. Microbiol. 74 (19), 5918–5924.
- Sommer, C., 1992. Larval biology and dispersal of Eudendrium racemosum (Hydrozoa, Eudendriidae). Sci. Mar. 56 (2–3), 205–211.
- Stiger-Pouvreau, V., Mattio, L., De Ramon N'Yeurt, A., Uwai, S., Dominguez, H., Flórez-Fernández, N., Connan, S., Critchley, A.T., 2023. A concise review of the highly diverse genus *Sargassum* C. Agardh with wide industrial potential. J. Appl. Phycol. 35, 1453–1483. https://doi.org/10.1007/s10811-023-02959-4.
- Van Tussenbroek, B.I., Årana, H.A.H., Rodríguez-Martínez, R.E., Espinoza-Avalos, J., Canizales-Flores, H.M., González-Godoy, C.E., BarbaSantosa, M.G., Vega-Zepedab, A., Collado-Vides, L., 2017. Severe impacts of brown tides caused by *Sargassum* spp. on near-shore Caribbean seagrass communities. Mar. Pollut. Bull. 122 (1–2), 272–281. https://doi.org/10.1016/j.marpolbul.2017.06.057.
- Weis, J.S., 1968. Fauna associated with pelagic Sargassum in the Gulf Stream. Am. Midl. Nat. 80, 554–555.