

Morphological and genetic confirmation of *Jassa slatteryi* (Crustacea: Amphipoda) in a harbour of Argentina

CARLOS RUMBOLD, JUAN LANCIA, GUADALUPE VÁZQUEZ, MARIANO ALBANO, NAHUEL FARIAS, MARÍA PAZ SAL MOYANO, EDUARDO SPIVAK AND SANDRA OBENAT

Departamento de Biología e Instituto de Investigaciones Marinas y Costeras (IIMyC), Facultad de Ciencias Exactas y Naturales, Universidad Nacional de Mar del Plata, Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Casilla de Correo 1245, 7600 Mar del Plata, Argentina

Jassa slatteryi is reported for the first time in Mar del Plata harbour, Argentina. A morphological comparison with the original description and a genetic analysis are made to corroborate the identification of specimens. The report of range expansion along the South-western Atlantic coast is discussed.

Keywords: *Jassa slatteryi*, invasive species, harbour, Argentina

Submitted 26 November 2014; accepted 29 January 2015

INTRODUCTION

The genus *Jassa* Leach, 1814 includes approximately 36 species (Lowry, 2014) distributed from tropical to polar marine environments (Conlan, 1990; Jażdżewski *et al.*, 1991). These organisms are the main food source for many animals and generally constitute one of the most important components in peracarid assemblages (Jeong *et al.*, 2007; Beermann & Franke, 2012). The great similarity between species and the enormous intraspecific variation (i.e. strong sexual dimorphism and intra-male polymorphism) makes it difficult to characterize the taxonomy of the group (Conlan, 1990; Lim & Park, 2006; LeCroy, 2007), so many species have been erroneously determined. Up to this moment, three species of *Jassa* have been previously recorded along the coasts of Argentina: *Jassa alonsoae* Conlan, 1990, *Jassa marmorata* Holmes, 1905 and *Jassa falcata* (Montago, 1808) (López-Gappa *et al.*, 2006; Alonso, 2012).

Jassa slatteryi Conlan, 1990 is a small benthic amphipod that lives in self-constructed tubes and feeds principally on detritus and diatoms (Howard, 1982; Jeong *et al.*, 2007; Beermann & Franke, 2012). Its iteroparous life cycle, high fecundity, earlier maturity and presence in ballast water and ship fouling has favoured its invasion in numerous regions of the world; harbours constitute the initial area of settlement for this invasive species (Robinson *et al.*, 2005; Lim & Park, 2006; Jeong *et al.*, 2007; Flynn & Valério-Berardo, 2009; Ah Yong & Wilkens, 2011).

As part of an ongoing project for the monitoring and biological identification of native and exotic benthonic invertebrates in Mar del Plata harbour, Argentina, using DNA barcoding, in this paper we report, for the first time, the

occurrence of *Jassa slatteryi* using morphological and genetic analysis.

MATERIALS AND METHODS

The specimens of *Jassa slatteryi* were collected in December 2010, November 2011 and August 2012 in Mar del Plata harbour (Figure 1), one of the most important harbours in the country due to naval traffic, commercial trade and size (Rivero *et al.*, 2005; Albano *et al.*, 2013). The harbour presents the typical characteristics of a polluted environment: high levels of organic matter from sewage and biogenic sources, hydrocarbons from ships and factories, low salinity and pH and anoxic conditions due to low turbulence (Rivero *et al.*, 2005; Albano *et al.*, 2013). Despite these conditions, the docks serve as settlement sites for ascidians, algae and polychaete tubes that generate refuges for a great variety of fishes, molluscs, crustaceans, nematodes and annelids (Albano & Obenat, 2009; Albano *et al.*, 2013).

Polychaete tubes of *Phyllochaetopterus socialis* (Claparède, 1869) were scraped up with spatulas and samples were fixed in 98% alcohol. In the laboratory, amphipods were separated from the extracted material using a stereoscopic microscope and determined with taxonomic guides (Conlan, 1990; Lim & Park, 2006; Chapman, 2007). *Jassa slatteryi* specimens were measured, dissected and photographed. In order to confirm the species identification through genetic analysis, a tissue sample was obtained from two specimens and used for the molecular analysis of a partial fragment of the barcode gene cytochrome oxidase I (COI). DNA was extracted using an automated Glass Fibre protocol (Ivanova *et al.*, 2006). Amplification of the 5' barcode region of COI was first attempted using the primer pair LCO1490 (5'-GGTCAACAAATCATAAAGATATTGG-3') and HCO2198 (5'-TAAAC TTCAGGGTGACCAAAAAATCA-3') (Folmer *et al.*, 1994),

Corresponding author:

C.E. Rumbold

Email: c_rumbold@hotmail.com

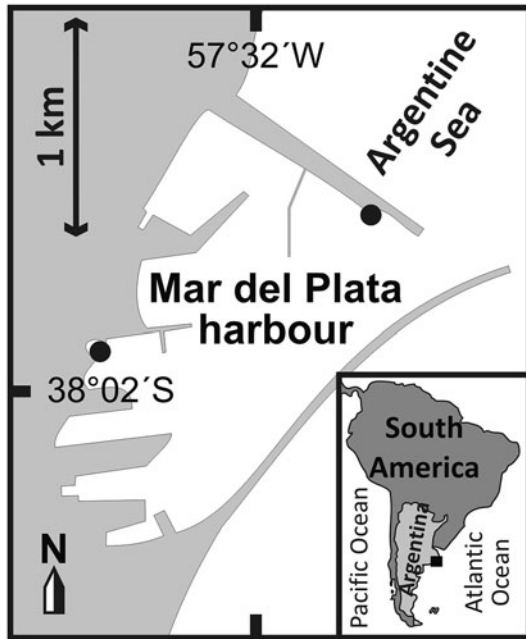


Fig. 1. Geographical localization of the study area and general view of Mar del Plata harbour, including the position of sampling sites where the species was found in the present study (●).

primers were appended with the M13F (5'-TTGTAA AACGACGGCCAGT-3') and M13R tails (5'-CAGGAAA CAGCTATGACC-3') to facilitate sequencing. The polymerase chain reaction (PCR) was performed in 12.5 μ L volume containing 2 μ L H₂O, 6.25 μ L 10% trehalose, 1.25 μ L 10 \times PCR buffer, 0.625 μ L MgCl₂ (50 mM), 0.0625 μ L dNTPs (10 mM), 0.06 μ L Platinum Taq polymerase (Invitrogen), 0.125 μ L of each primer (10 μ M) and 2 μ L DNA template. The PCR reaction profile consisted of 1 min at 95°C and 35 cycles of 30 s at 94°C, 40 s at 50°C and 1 min at 72°C, with a final extension at 72°C for 10 min. Additional details about laboratory protocols for each step are available from the Canadian Centre for DNA Barcoding (CCDB) website (<http://ccdb.ca/docs/>). The obtained COI sequences were compared against both the Barcode of Life (BOLD, <http://www.barcodeoflife.org>) and the GenBank (<http://www.ncbi.nlm.nih.gov/GenBank>) databases using BLAST. The BLAST algorithm finds regions of local similarity between the query



Fig. 2. *Jassa slatteryi* Conlan, 1990. Lateral view of adult male (total length: 6.45 mm) from Mar del Plata harbour, Argentina.

sequence and the sequences found in the above databases. BLAST provides a statistical value for each sequence, called the E-value, which describes the number of hits one can 'expect' to see by chance when searching a database. E-values closer to zero indicate a significant match.

A phylogenetic analysis based on the construction of a maximum-likelihood tree was conducted to validate the identity of our material and the *Jassa* spp. from GenBank. For this phylogenetic analysis, additional COI sequences of *Jassa* spp. were retrieved from GenBank (for species name and accession numbers, see Figure 3). The COI sequence of *Erichthonius punctatus* (= *Erichthonius brasiliensis*) (Bate, 1857) was used as an outgroup. The COI sequences were aligned using BioEdit v.7.08.0 software (Hall, 1999) through the Clustal W method (Thompson *et al.*, 1994) and were verified by sight. A phylogenetic tree was inferred in MEGA v6 (Tamura *et al.*, 2013; <http://www.megasoftware.net/>) and was evaluated with a bootstrap test with 1000 replications.

The COI sequences generated in the present study were deposited in GenBank (Accession numbers: KP252164, KP252165) as well as in the 'Native and exotic invertebrate of Mar del Plata harbour' project (code INVMP) on BOLD. Voucher specimens were deposited at the Estación Costera J.J. Nágera, Universidad Nacional de Mar del Plata, Argentina (catalogue number pending).

RESULTS

SYSTEMATICS

Order AMPHIPODA Latreille, 1816
 Suborder SENTICAUDATA Lowry & Myers, 2013
 Infraorder COROPHIIDA Leach, 1814
 Family ISCHYROCERIDAE Stebbing, 1899
 Subfamily ISCHYROCERINAE Stebbing, 1899
 Genus *Jassa* Leach, 1814
Jassa slatteryi Conlan, 1990
 (Figures 2 & 3)

Jassa slatteryi Conlan, 1990: 2038, Figures 8, 9, 10 & 20; Lim & Park, 2006: 300, Figures 1, 2 & 3; Chapman, 2007: 569, plate 268.

MORPHOLOGICAL ANALYSIS

MATERIAL EXAMINED

Mar del Plata harbour, Mar del Plata, Buenos Aires, Argentina, South-west Atlantic: eight males and ten females (four ovigerous), 38°02.48'S 57°32.27'W, water depth <1 m, December 2010.

Mar del Plata harbour, Mar del Plata, Buenos Aires, Argentina, South-west Atlantic: one male, 38°02.48'S 57°32.27'W, water depth <1 m, November 2011.

Mar del Plata harbour, Mar del Plata, Buenos Aires, Argentina, South-west Atlantic: one male, 38°02.18'S 57°32.5'W, water depth 3 m, August 2012.

DIAGNOSIS

Antenna 2: posterodistal margin of flagellum segment 1 and peduncular segment 5 in males with plumose setae. Mandible: segment 2 without setae on dorsal edge. Gnathopod 1: palm slightly concave, carpus with dorsal setae in anterodistal junction of the propodus and basis with

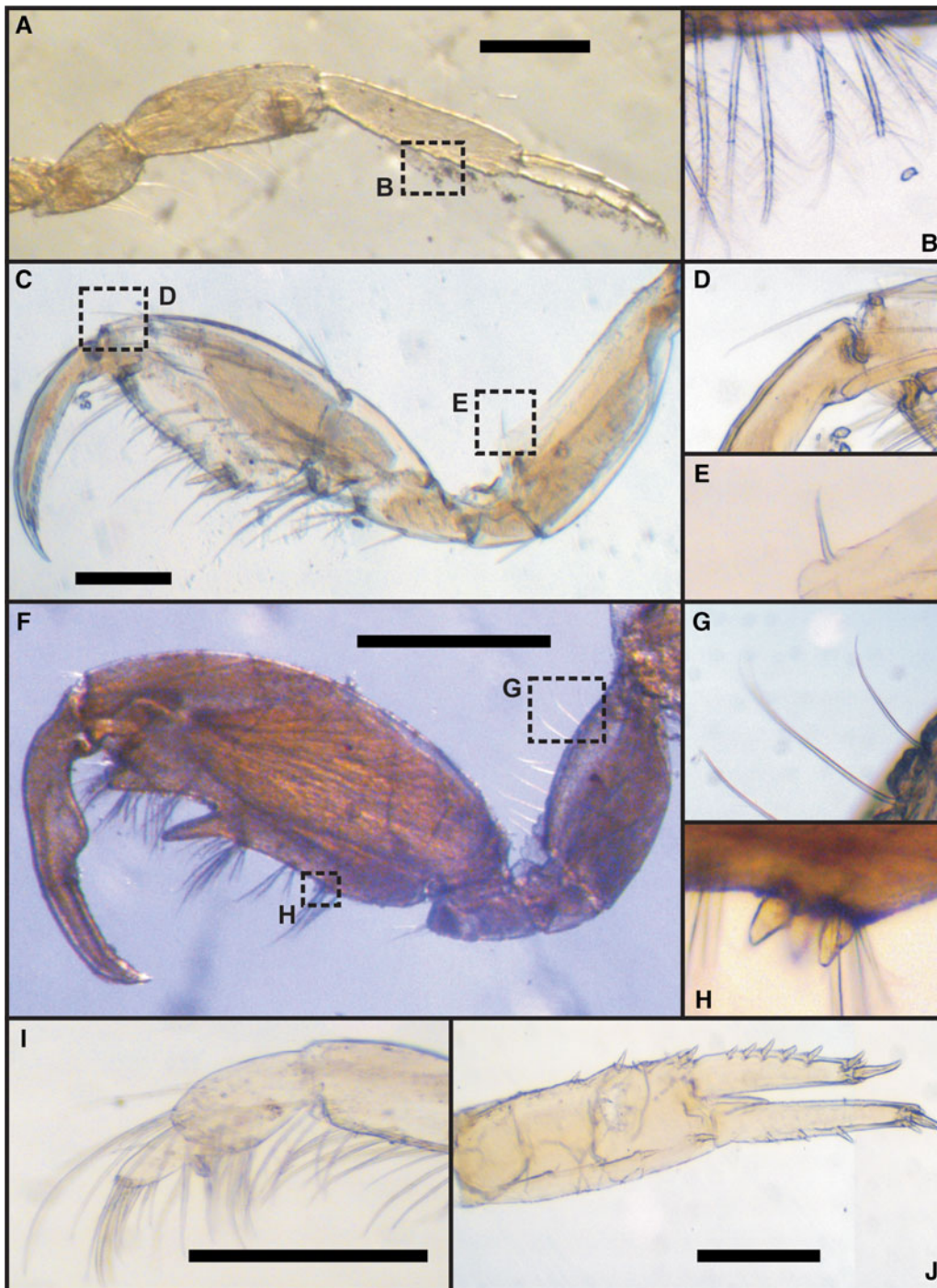


Fig. 3. *Jassa slatteryi* Conlan, 1990. (A) Antenna 2; (B) detail of plumose setae of flagellum of antenna 2; (C) gnathopod 1; (D) detail of dorsal setae in anterodistal edge of propodus; (E) detail of setae confined to anterodistal edge of basis; (F) gnathopod 2; (G) detail of scattered and simple setae of basis; (H) detail of small teeth next to origin of dactylus; (I) mandible; and (J) uropod 1. Scale bars: (A) and (F), 0.5 mm; (C), (I) and (J), 0.15 mm.

setae confined to the anterodistal edge. Gnathopod 2: palm concave without spines, propodus in males with small tooth next to origin of dactylus, basis and propodus with anterior marginal setae scattered and simple. Uropod 1: small spines occupying 50% the length of segment 2. Diagnosis from Conlan (1990), Lim & Park (2006) and Chapman (2007).

RECENT RECORDS

Jassa slatteryi has a wide distribution throughout the world: in Europe it is known in Spain (Mediterranean coast)

(Beermann, 2013), Ireland (Conlan, 1990), France (Mediterranean and Atlantic) and Croatia (Conlan, 1990); western Pacific coasts in Japan (Conlan, 1990; Sano *et al.*, 2003) and Korea (Conlan, 1990; Lim & Park, 2006; Jeong *et al.*, 2007); South Africa (Conlan, 1990; Robinson *et al.*, 2005) and southern coasts of Australia, Tasmania and New Zealand (Conlan, 1990; Ahyong & Wilkens, 2011). In the west coast of America it occurs from Alaska to Mexico (Conlan, 1990; Pilgrim & Darling, 2010), the Galapagos Islands and Chile (Conlan, 1990; González *et al.*, 2008), and

on the east coast of America it was only reported in Brazil (São Paulo) (Conlan, 1990; Flynn & Valério-Berardo, 2009). This is the first record of *J. slatteryi* in Argentina.

REMARKS

Body length ranged between 3.5 and 4.35 mm in females and between 3.35 and 6.45 mm in males. Some juveniles were recorded during surveys, but we could not determine if these specimens corresponded to *Jassa slatteryi* or *Jassa marmorata*. This new record is the most southerly for this species in South America.

GENETIC ANALYSIS

The 658 bp COI sequences obtained from the two specimens showed 98–99% similarity with a mitochondrial COI sequence of *Jassa slatteryi* in GenBank (accession numbers: EU243803, EU243804, EU243805, EU243806, EU243807, EU243808, EU243809, EU243810, EU243811, EU243812, EU243813, EU243814, EU243815, GU048095, GU048096) and 98.32–99.68% similarity with 14 specimens in BOLD from the Pacific coast of the United States, confirming the species identity (in all cases, E-value = 0).

The maximum-likelihood tree showed that specimens from Mar del Plata harbour were clustered with *J. slatteryi* specimens from the United States (Figure 4).

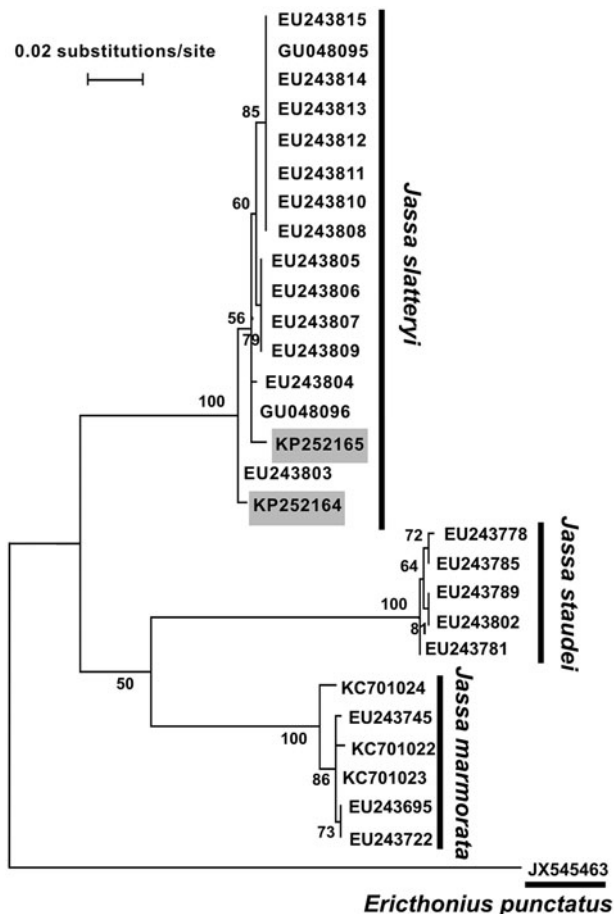


Fig. 4. Maximum likelihood phylogenetic tree of *Jassa* spp. cytochrome oxidase I (COI). Specimens of *Jassa slatteryi* collected in Mar del Plata harbour are marked with grey bars.

DISCUSSION

Harbours are important areas for the entrance, settlement and spread of exotic species. Monitoring projects that allow for the identification of potential invasive species are necessary for the prediction of possible sites of invasion and for determining their impact on the benthic community, such as a decrease in the density of native species, changes in biodiversity and alterations in interspecific interaction (Grabowski *et al.*, 2007; Piscart *et al.*, 2010; Galil *et al.*, 2011).

Here, we report the occurrence of *Jassa slatteryi* on the coast of Argentina for the first time and extend its range 1900 km southwards in the South-western Atlantic. The high reproductive potential of *J. slatteryi* and their tolerance to polluted habitats, such as harbours, make this organism a successful invasive species (Jeong *et al.*, 2007; Flynn & Valério-Berardo, 2009). Previous studies suggest that *J. slatteryi* is native from the western coast of North America (Pilgrim & Darling, 2010), invading different countries around the world through commercial vessels and recreational boats (Robinson *et al.*, 2005; Lim & Park, 2006; Flynn & Valério-Berardo, 2009; Ah Yong & Wilkens, 2011), which is the most likely explanation for understanding their presence in Mar del Plata harbour. On the other hand, it has recently been shown that one of the effects of global warming is the change in the geographical distribution of amphipods, related to the increase in seawater temperature and to species favouring the invasion of areas in which previously they could not survive or reproduce, so this hypothesis should not be discarded (Beermann & Franke, 2012).

The presence of adult males and ovigerous females in Mar del Plata harbour indicates that the population of *J. slatteryi* is well established. Further studies are necessary to analyse their current status and to assess their impact on the benthic community structure.

ACKNOWLEDGEMENTS

The authors would like to thank Club Náutico de Mar del Plata for facilitating access to the docks, E. Ocampo for his assistance with the phylogenetic analysis and Ana Faigón, of Fondo iBOL Argentina, for her help with the BOLD system. We are grateful for the support provided by the Government of Canada through Genome Canada and the Ontario Genomics Institute's International Barcode of Life Project. This funding enabled the Canadian Centre for DNA Barcoding (University of Guelph) to carry out the sequence analysis on our specimens. We also thank the Ontario Ministry of Economic Development and Innovation for funding the ongoing development of BOLD. Finally, we appreciate the important comments made by John Chapman and two anonymous referees.

REFERENCES

- Ah Yong S.T. and Wilkens S.L. (2011) Aliens in the antipodes: non-indigenous marine crustaceans of New Zealand and Australia. In Galil B.S., Clark P.F. and Carlton J.T. (eds) *In the wrong place – alien marine crustaceans: distribution, biology and impacts*. *Invasive Nature – Springer Series in Invasion Ecology*, Vol. 6. Dordrecht: Springer, pp. 451–485.

- Albano M.J., Lana P., Bremec C., Elías R., Martins C., Venturini N., Muniz P., Rivero S., Vallarino E.A. and Obenat S.** (2013) Macrobenthos and multi-molecular markers as indicators of environmental contamination in a South American port (Mar del Plata, Southwest Atlantic). *Marine Pollution Bulletin* 73, 102–114.
- Albano M.J. and Obenat S.M.** (2009) Assemblage of benthic macrofauna in the aggregates of the tubicolous worm *Phyllochaetopterus socialis* in the Mar del Plata harbour, Argentina. *Journal of the Marine Biological Association of the United Kingdom* 89, 1099–1108.
- Alonso G.M.** (2012) Amphipod crustaceans (Corophiidae and Gammaridea) associated with holdfasts of *Macrocystis pyrifera* from the Beagle Channel (Argentina) and additional records from the Southwestern Atlantic. *Journal of Natural History* 46, 1799–1894.
- Beermann J.** (2013) *Ecological differentiation among amphipod species in marine fouling communities: studies on sympatric species of the genus Jassa Leach, 1814 (Crustacea, Amphipoda)*. PhD thesis, Freien Universität Berlin, Berlin, Germany.
- Beermann J. and Franke H.** (2012) Differences in resource utilization and behavior between coexisting *Jassa* species (Crustacea, Amphipoda). *Marine Biology* 159, 951–957.
- Chapman J.W.** (2007) Gammaridea. In Carlton J.T. (ed.) *The light and Smith manual - intertidal invertebrates from central California to Oregon*. Richmond: The University of California Press, pp. 545–618.
- Conlan K.E.** (1990) Revision of the crustacean amphipod genus *Jassa* Leach (Corophioidea: Ischyroceridae). *Canadian Journal of Zoology* 68, 2031–2075.
- Flynn M.N. and Valério-Berardo M.T.** (2009) Depth-associated patterns in the development of Amphipoda (Crustacea) assemblages on artificial substrata in the São Sebastião Channel, Southeastern Brazil. *Nauplius* 17, 127–134.
- Folmer O., Black M., Hoeh W., Lutz R. and Vrijenhoek R.** (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology* 3, 294–299.
- Galil B.S., Clark P.F. and Carlton J.T.** (2011) *In the wrong place – alien marine crustaceans: distribution, biology and impacts*. *Invading Nature – Springer Series in Invasion Ecology*, Vol. 6, 1st edn. Dordrecht: Springer, 716 pp.
- González E.R., Hays P.A., Balandá M. and Thiel M.** (2008) Lista sistemática de especies de peracaridos de Chile (Crustacea, Eumalacostraca). *Gayana* 72, 157–177.
- Grabowski M., Bacela C. and Konopacka A.** (2007) How to be an invasive gammarid (Amphipoda: Gammaroidea) – comparison of life history traits. *Hydrobiologia* 590, 75–84.
- Hall T.A.** (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41, 95–98.
- Howard R.K.** (1982) Impact of feeding activities of epibenthic amphipods on surface-fouling of eelgrass leaves. *Aquatic Botany* 14, 91–97.
- Ivanova N.V., Dewaard J.R. and Hebert P.D.N.** (2006) An inexpensive, automation-friendly protocol for recovering high-quality DNA. *Molecular Ecology Notes* 6, 998–1002.
- Jażdźewski K., Broyer C., Teodorczyk W. and Konopacka A.** (1991) Survey and distributional patterns of the amphipod fauna of Admiralty Bay, King George Island, South Shetland Islands. *Polish Polar Research* 12, 461–472.
- Jeong S.J., Yu O.H. and Suh H.L.** (2007) Life history and reproduction of *Jassa slatteryi* (Amphipoda, Ischyroceridae) on a seagrass bed (*Zostera marina* L.) in Southern Korea. *Journal of Crustacean Biology* 27, 65–70.
- LeCroy S.E.** (2007) An illustrated identification guide to the nearshore marine and estuarine Gammaridean Amphipoda of Florida. Families: Anamixidae, Eusiridae, Hyaellidae, Hyalidae, Iphimediidae, Ischyroceridae, Lysianassidae, Megalurotopidae and Melphidippidae. *Tallahassee, Florida Department of Environmental Protection*, no. 4, 111 pp.
- Lim B.J. and Park J.Y.** (2006) Redescription of *Jassa slatteryi* (Crustacea: Amphipoda: Ischyroceridae). *Korean Journal of Environmental Biology* 24, 300–305.
- López-Gappa J., Alonso G.M. and Landoni N.A.** (2006) Biodiversity of benthic Amphipoda (Crustacea: Peracarida) in the Southwest Atlantic between 35°S and 56°S. *Zootaxa* 1342, 1–66.
- Lowry J.** (2014) *Jassa* Leach, 1814. In Horton T., Lowry J. and De Broyer C. (2013 onwards) World Amphipoda Database. *World Register of Marine Species*. Available at: <http://www.marinespecies.org/aphia.php?p=taxdetails&id=101571> (accessed 11 November 2014).
- Pilgrim E.M. and Darling J.A.** (2010) Genetic diversity in two introduced biofouling amphipods (*Amphitoe valida* & *Jassa marmorata*) along the Pacific North American coast: investigation into molecular identification and cryptic diversity. *Diversity and Distributions* 16, 827–839.
- Piscart C., Bergerot B., Laffaille P. and Marmonier P.** (2010) Are amphipod invaders a threat to regional biodiversity? *Biological Invasions* 12, 853–863.
- Rivero M.S., Elías R. and Vallarino E.A.** (2005) First survey of macroinfauna in the Mar del Plata Harbor (Argentina), and the use of polychaetes as pollution indicators. *Revista de Biología Marina y Oceanografía* 40, 101–108.
- Robinson T.A., Griffiths C.L., McQuaid C.D. and Rius M.** (2005) Marine alien species of South Africa – status and impacts. *African Journal of Marine Science* 27, 297–306.
- Sano M., Omori M. and Taniguchi K.** (2003) Predator-prey systems of drifting seaweed communities off the Tohoku coast, northern Japan, as determined by feeding habit analysis of phytal animals. *Fisheries Science* 69, 260–268.
- Tamura K., Stecher G., Peterson D., Filipksi A. and Kumar S.** (2013) MEGA6: Molecular Evolutionary Genetics Analysis Version 6.0. *Molecular Biology and Evolution* 30, 2725–2729.
- and
- Thompson J.D., Higgins D.G. and Gibson T.J.** (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research* 22, 4673–4680.

Correspondence should be addressed to:

C.E. Rumbold
 Departamento de Biología e Instituto de
 Investigaciones Marinas y Costeras (IIMyC)
 Facultad de Ciencias Exactas y Naturales, Universidad
 Nacional de Mar del Plata
 Consejo Nacional de Investigaciones Científicas y Técnicas
 (CONICET), Casilla de Correo 1245, 7600 Mar del Plata
 Argentina
 email: c_rumbold@hotmail.com