Comparison of hydroponic and conventional grafting techniques in Black pepper (*Piper nigrum* L.)

3 Abstract

4 The most devastating disease affecting black pepper (*Piper nigrum* L.) is 5 Phytophthora foot rot against which none of the varieties is resistant. This difficulty has been 6 overcome by the horticultural technique of grafting *P. nigrum* on the resistant root stock viz. 7 P. colubrinum- a marshy wild relative of black pepper. The conventional technique of 8 grafting of P. nigrum on P. colubrinum takes nine months to one year for developing a graft 9 ready to be planted in the field and is labour and irrigation intensive. Hydroponic soil-less 10 methods based on the marshy adaptation of *P. colubrinum* and which require lesser labour and water have been explored to address the issues with the conventional method. An experiment 11 12 in complete randomised design with five grafting methods and six replications was 13 conducted at Pepper Research Station, Panniyoor, Kannur, Kerala, India during 2019-20 to 14 compare the conventional grafting technique with hydroponic techniques. Instant grafting of 15 the scion portion on the non - rooted root stock portion and rooting of the root stock 16 hydroponically once the grafting procedure is completed has been found to produce field 17 ready grafts in 95 days with the lowest graft loss of 6.8 per cent and water use efficiency of

18 117.7 per cent.

19 Key words: Black pepper, graft, soil-less, *Piper colubrinum*, hydroponic

20 **1.Introduction**

Black pepper (*Piper nigrum* L.), the historic 'King of Spices' has been used as spice since 4th century B.C. (Jibat and Alo 2021).The most devastating disease affecting black pepper is *Phytophthora* foot rot against which no variety has been found to be resistant (Hao 24 et al. 2016). This difficulty has been overcome by the horticultural technique of grafting P. 25 nigrum on root stocks of the wild related marshy species *Piper colubrinum* L. which is 26 resistant not only to *Phytophthora* but also to nematodes causing root knot in black pepper 27 (Krishnamoorthy and Parthasarathy 2009). The technique of grafting of P. nigrum on P. 28 colubrinum has been very successful and has been well standardized and well known (Vanaja 29 et al. 2007). In the conventional method, two to three node cuttings of P. colubrinum of 30 approximately 15 cm length are planted in poly bags of appropriate size to get them rooted. It 31 takes approximately six to nine months for the cuttings of P. colubrinum to get rooted 32 properly and reach a height of around 65 cm and appropriate girth of around 1 cm diameter at 33 45 cm height where cleft grafting is done in the popular method (Fig.1). Grafting at a height 34 of around 50 cm is inevitable to avoid splashing of soil and debris that contain spores of 35 Phytophthora and also to get more aerial roots growing from the root stock to the ground 36 (Krishnamoorthy and Parthasarathy 2009).

37 Once grafting is completed, it takes another three months for the grafts to grow well with 5-6 38 well developed leaves so that they can be planted in the field. In total, it takes nine months to 39 one year for developing a graft with a graft union height of 45 cm or higher which is ready to 40 be planted in the field. The conventional method is a slow process which requires greater 41 quantities of irrigation water and is labour intensive. It is required to find out faster and 42 efficient alternatives to the conventional method with lesser irrigation water requirement. 43 Besides this, the potting medium in poly bags often harbour pathogenic fungal spores and 44 nematodes which unintentionally invade farmers' fields once these grafts are planted in their 45 field. Transportation of grafted plants is cumbersome and costly due to the bulky nature of 46 potting medium. Adaptation of *P. colubrinum* to marshy habitat which indicates its ability to 47 root and thrive in water has never been exploited in graft production (Fig.2). Hence a trial

- 48 was undertaken to compare the performance of soil- less hydroponic methods of grafting with
- 49 the conventional and other possible alternatives.

50 **2.Material and methods**

51 The trial was undertaken at Pepper Research Station, Panniyoor, Kannur, Kerala, India during

52 2019-20. The experiment consisted of 5 treatments (Table 1).

53 Table 1.Treatments

| Sl.No. | Treatment | Description of treatment | Figure No. |
|--------|-----------|---|------------|
| 1 | T1 | 45 cm long <i>Piper colubrinum</i> cuttings planted in poly bags for rooting and further grafting | 3 |
| 2 | T2 | <i>P.nigrum</i> grafted on 45 cm long <i>P. colubrinum</i> and the basal nodes of the root stock submerged in water | 4 |
| 3 | T3 | Two node cuttings of <i>P.colubrinum</i> planted in poly bags (conventional method - control) | 5 |
| 4 | T4 | 45 cm long <i>P.colubrinum</i> cuttings put in water for further grafting | 6 |
| 5 | T5 | <i>P.nigrum</i> grafted on 45 cm long <i>P.colubrinum</i> and planted in potting medium in poly bag | 7 |

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The experimental design was completely randomised design (CRD) with 6 replications. Only
orthotropic shoots of *P.colubrinum* were used as root stock and runners (basal adventitious

58 shoots) of *P.nigrum* variety 'Panniyur 1' were used as scion in the experiment. The size of 59 the poly bags used was 20 cm x 15 cm and potting medium was 1:1:1 mixture of sand, soil 60 and dry farm yard manure. Irrigation water required to bring the potting medium in poly bags 61 to saturation point was calculated as per Imakumbili 2019. Two hundred millilitres of water 62 once daily was applied to each poly bag to bring it to saturation. In hydroponic treatments, 63 500 ml clean tap water was taken in 1 litre containers and only the basal two nodes of the root 64 stock were submerged in water. Water in the container was replaced with fresh tap water 65 once in 10 days. All the treatments were kept in a closed poly house nursery, under natural 66 day light. The temperature inside the poly house seasonally varied from 28°C to 35°C and 67 relative humidity varied from 85% to 98%. Data was recorded on days to bud activation on 68 root stock, days to root initiation, number of roots on root stock 60 days after 69 planting/submergence, length of roots on root stock 60 days after planting/submergence (cm), 70 days taken for grafting on root stock, days to get field- ready grafted plants, water 71 requirement per field- ready graft (L), per cent loss of grafted plants and water use efficiency. 72 Days taken for grafting on rootstock was recorded as the number of days taken by 73 *P.colubrinum* cutting to grow and reach a height of around 65 cm and appropriate girth of 74 around 1 cm diameter at 45 cm height where cleft grafting could be done properly .Grafts 75 with minimum graft union height of 45 cm and with 5-6 well developed leaves on the scion 76 were considered to be ready to be planted in the field. Water requirement per graft was 77 calculated by dividing the total volume of irrigation water applied to the treatments by the 78 number of replications in case of treatments where root stocks were planted in poly bags. 79 Water requirement per graft in case of hydroponic treatments was calculated as the total 80 volume of water to be kept in containers for keeping the lower two nodes of root stock 81 submerged for all the replicates divided by the number of replications. For calculating per 82 cent loss of grafted plants, wilted and dried up scion was taken as the indication for loss of the graft even though the rootstock portion was intact. Water use efficiency (%) was calculated as the total number of successful grafts recovered per litre of irrigation water applied. Statistical analysis was done as per Gomez and Gomez, 1984.

- 86 **3.Results and discussion**
- 87 Days to bud activation on root stock ranged from 7.5 to 15 (Table 2).
- 88 Table 2.Growth parameters and recovery of grafts under different systems of grafting in black
- 89 pepper

| Tr | Days | Days | Numbe | Length | Days | Days to | Days | Water | Per cent | Water |
|----|------------------|-------------------|------------------|-------------------|-------------------|-------------------|--------------------|----------|------------------|-----------|
| ea | to bud | to | r of | of | taken | bud | to | requirem | loss of | use |
| tm | activat | root | roots | roots | for | activati | field | ent per | grafts | efficienc |
| en | ion on | initiat | on root | on root | grafti | on on | ready | field | | у |
| ts | root | ion | stock | stock | ng | scion | grafte | ready | | (%) |
| | stock | | 60 days | 60 | | after | d | graft(L) | | |
| | | | after | days | | graftin | plants | | | |
| | | | plantin | after | | g | | | | |
| | | | g/subm | plantin | | | | | | |
| | | | ergence | g/subm | | | | | | |
| | | | | ergenc | | | | | | |
| | | | | e(cm) | | | | | | |
| | | | | | | | | 36.9 | 22.8(28 | 12.6 |
| Т1 | 7 5° | 13 7 ^a | $7 0^{a}$ | 7 3 [°] | 85 8 ^b | 17 5 ^b | 184 5 ^b | 50.7 | 6)* ^b | 12.0 |
| 11 | 1.5 | 13.7 | 7.0 | 1.5 | 05.0 | 17.3 | 104.5 | | 0)* | |
| T2 | 9.3 ^c | 5.7° | 7.3 ^a | 16.1 ^ª | - | 12.5 ^c | 95.0 ^d | 4.75 | 6.8(14.9 | 117.7 |
| | | | | | | | | | | |

| | | | | | | | | |) ^d | |
|-----|-------------------|--------------------|--------------------|-------------------|-------------------|-------------------|--------------------|-------|-----------------|------|
| | | | | | 206.7 | | | 57.46 | 15.3(22. | 8.8 |
| Т3 | 9.5° | 13.3 ^{ab} | 3.5° | 7.7 ^c | а | 17.7 ^b | 287.3 ^a | | 9) ^c | |
| | | | | | | | | | | |
| | | | | | | | | 6.21 | 12.2(20. | 84.8 |
| T4 | 11.8 ^b | 11.8 ^b | 5.8 ^b | 12.3 ^b | 49.2 ^c | 13.8 ^c | 124.2 ^c | | 4) ^c | |
| | | | | | | | | | | |
| | | | | | | | | - | 94.0 | - |
| Т5 | 15 ^a | 14.5 ^a | 7.0^{a} | 4.6 ^d | - | 19.8 ^a | - | | $(76.4)^{a}$ | |
| | | | | | | | | | | |
| С | | | | | | | | - | | - |
| D(| | | | | | | | | | |
| 5 | | | | | | | | | | |
| % | | | | | | | | | | |
| , o | 2.2 | 1.6 | 1.1 | 0.7 | 12.0 | 1.0 | <i>.</i> | | 2.4 | |
|) | 2.2 | 1.6 | 1.1 | 0.7 | 12.8 | 1.9 | 6 | | 3.4 | |
| С | | | | | | | | - | | - |
| v | 17.5 | 11.5 | 14.4 | 6.3 | 9.1 | 10.1 | 2.9 | | 8.9 | |
| • | 17.5 | 11.5 | 1 | 0.5 | 7.1 | 10.1 | 2.9 | | 0.9 | |
| | | 1 | 1 | 1 | 1 | 1 | 1 | | 1 | 1 |

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*Figures in parantheses indicate transformed values

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The earliest bud activation on root stock was recorded for T2 (*P.nigrum* grafted on 45 cm long *P. colubrinum* and the root stock put in water) which was on par with T1 and T3 and the last to record bud activation was T5 (*P.nigrum* grafted on 45 cm long *P.colubrinum* and planted in poly bag).T2 recorded significant earliness for root initiation on root stock while the last to root was T5 which was on par with T1.Root stocks planted in potting mixture generally took more time for root initiation. The highest number and length of roots 60 days

98 after planting/submergence was recorded for T2 (Fig.8 and 9). This trait can be supposed to 99 impart greater anchorage once planted in soil and greater nutrient absorption ability to T2 100 during the early months of graft union. Fourty five centimetre long *P.colubrinum* cutting put 101 in water (T4) became ready for grafting around one month earlier than T1 even though the 102 earliest bud initiation on root stock was observed in T1. This could be attributed to the early 103 root initiation and better root length in T4. Control (T3) took the highest number of days 104 (206.7) for producing sufficient growth for getting 45 cm long root stock. Instant grafting was 105 possible for T2 and T5 only. Bud activation on scion after grafting was the earliest in T2 106 which was on par with T4.T2 produced field- ready grafted plants in 95 days while 107 T3(control) took 287.3 days and T4 took 124.2 days. Irrigation water requirement per field-108 ready graft was the lowest for T2 (4.75L) and the highest for the conventional method (57.46 109 L). Though instant grafting was possible in T2 and T5, percent loss of grafts was the highest 110 and the most significant in T5. This could be due to the failure of non-rooted root stock to 111 supply water to the scion and consequent wilting of scion. Per cent loss of grafts was the least 112 for T2 (6.8%).George and Sujatha (2019) has reported a maximum per cent survival of 113 93.33% among Piper species. In general, early bud activation on scion and lesser per cent 114 loss of grafts were observed when the basal nodes of root stocks were put in water rather than 115 anchored in potting mixture. Water use efficiency was the highest for T2 (117.7%) and the 116 least for the conventional method (8.8%).

Soil -less method of grafting *P.nigrum* on *P.colubrinum* and keeping the lower nodes of root stock in water was found to produce the highest number of field- ready grafted plants in the minimum time. This method has the additional advantages of high water use efficiency, freedom from daily irrigation and, thus saving irrigation water and labour. Inadequate water availability and labour scarcity have been indicated as two constraints faced by black pepper farmers (Krishnamoorthy and Parthasarathy, 2009) and this soil- less grafting method hasgreat relevance under such challenges.

124 **4.**Conclusion

125 Instant grafting of *P.nigrum* on *P.colubrinum* root stock of sufficient length followed by 126 submerging the basal two nodes of the root stock in water was found to produce the highest 127 number of grafts in much shorter time than in the conventional method. This soil-less, water 128 use efficient method also saves irrigation water and labour .

129 **5.**Acknowledgement

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132 **6.***References*

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- 151
- 152 Figures
- 153







157 Fig.2 Profuse root growth in P.colubrinum under water



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159 Fig.3 T1



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161 Fig.4 T2





163 Fig.5 T3



165 Fig.6 T4







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169 Fig.8 Treatments 60 days after planting or submergence



171 Fig.9 T2 Sixty days after grafting