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Stink bugs (Hemiptera: Heteroptera: Pentatomidae) of Colombia: An annotated checklist of species

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Table of Contents

Abstract	5
Introduction	5
Materials and methods	6
Results	6
List of species	7
Subfamily Asopinae	7
<i>Alcaeorrhynchus</i> Bergroth, 1891	7
<i>Andrallus</i> Bergroth, 1905	7
<i>Apateticus</i> Dallas, 1851	7
<i>Coryzorhaphis</i> Spinola, 1837	8
<i>Discocera</i> Laporte, 1833	8
<i>Euthyrhynchus</i> Dallas, 1851	9
<i>Heteroscelis</i> Latreille, 1829	9
<i>Marmessulus</i> Bergroth, 1891	9
<i>Oplonus</i> Spinola, 1837	10
<i>Podisus</i> Herrich-Shaeffer, 1851	10
<i>Stiretrus</i> Laporte, 1833	13
<i>Supputius</i> Distant, 1889	14
<i>Tylospilus</i> Stål, 1870	14
<i>Tynacantha</i> Dallas, 1851	15
<i>Tyrannocoris</i> Thomas, 1992	15
Subfamily Cyrtocorinae	15
<i>Ceratozygum</i> Horváth, 1916	15
<i>Cyrtocoris</i> White, 1842	15
Subfamily Discocephalinae	16
Tribe Discocephalini	16
<i>Ablaptus</i> Stål, 1872	16
<i>Alveostethus</i> Ruckes, 1966	17
<i>Agaclitus</i> Stål, 1864	17
<i>Antiteuchus</i> Dallas, 1851	17
<i>Coriplatus</i> White, 1842	18
<i>Colpocarena</i> Stål, 1868	19
<i>Dinocoris</i> Burmeister, 1835	19
<i>Discocephalessa</i> Kirkaldy, 1909	20
<i>Dryptocephala</i> Laporte, 1832	20
<i>Eurystethus</i> Mayr, 1864	21
<i>Paralcippus</i> Becker & Grazia, 1986	23
<i>Patronatus</i> Ruckes, 1965	23
<i>Pelidnocoris</i> Stål, 1867	23
<i>Platycarenum</i> Fieber, 1861	24
<i>Phoeacia</i> Stål, 1862	24
Tribe Ochlerini	24
<i>Coranda</i> Rolston, 1992	24
<i>Eritrachys</i> Ruckes, 1959	24
<i>Herrichella</i> Distant, 1911	26
<i>Lincus</i> Stål, 1867	26
<i>Macropygium</i> Spinola, 1837	27
<i>Melambyrsus</i> Breddin, 1912	28
<i>Neoadoxoplatys</i> Kormilev, 1956	28
<i>Ochlerus</i> Spinola, 1837	28
<i>Orbatina</i> Ruckes, 1961	29
<i>Paralincus</i> Distant, 1911	29
<i>Phereclis</i> Stål, 1862	29
<i>Schraderiellus</i> Rider, 1998	30
<i>Stalius</i> Rolston, 1992	30
<i>Stapecolis</i> Garbelotto & Campos, 2016	31
<i>Xynocoris</i> Garbelotto & Campos, 2014	31
Subfamily Edessinae	31
<i>Brachystethus</i> Laporte, 1832	31
<i>Doesburgedessa</i> Fernandes, 2010	32
<i>Edessa</i> Fabricius, 1803	32
<i>Grammedessa</i> Correia & Fernandes, 2016	34
<i>Hypoxys</i> Amyot & Serville, 1843	34

<i>Lopadusa</i> Stål, 1860	34
<i>Paraedessa</i> Silva & Fernandes, 2013	36
<i>Peromatus</i> Amyot & Serville, 1843	36
Subfamily Pentatominae	36
Tribe Carpocorini	36
<i>Agroecus</i> Dallas, 1851	36
<i>Amauromelpia</i> Fernandes & Grazia, 1998	37
<i>Berecynthus</i> Stål, 1862	37
<i>Cosmopepla</i> Stål, 1867	38
<i>Diceraeus</i> Dallas, 1851	38
<i>Dichelops</i> Spinola, 1837	38
<i>Euschistus</i> Dallas, 1851	39
<i>Galedanta</i> Amyot & Serville, 1843	40
<i>Hypatropis</i> Fernandes & Grazia, 1996	41
<i>Lattinidea</i> Rider & Eger, 2008	41
<i>Lattinellica</i> Rider & Eger, 2008	41
<i>Mitripus</i> Rolston, 1978	41
<i>Mormidea</i> Amyot & Serville, 1843	42
<i>Oebalus</i> Stål, 1862	44
<i>Padaeus</i> Stål, 1862	45
<i>Paratibraca</i> Campos & Grazia, 1995	47
<i>Poriptus</i> Stål, 1861	47
<i>Proxys</i> Spinola, 1837	47
<i>Sibaria</i> Stål, 1872	48
<i>Spinalanx</i> Rolston & Rider, 1988	49
<i>Stysiana</i> Grazia, Fernandes & Schwertner, 1999	49
<i>Tibraca</i> Stål, 1860	49
Tribe Catacanthini	49
<i>Arocera</i> Spinola, 1837	49
<i>Rhysocephala</i> Rider, 1992	51
<i>Runibia</i> Stål, 1861	52
<i>Vulsirea</i> Spinola, 1837	52
Tribe Chlorocorini	53
<i>Chlorocoris</i> Spinola, 1837	53
<i>Loxa</i> Amyot & Serville, 1843	55
<i>Mayrinia</i> Horváth, 1925	56
<i>Rhyncholepta</i> Bergroth, 1911	56
Tribe Menidini	56
<i>Elanela</i> Rolston, 1980	56
<i>Rio</i> Kirkaldy, 1909	57
Tribe Nezarini	57
<i>Chinavia</i> Orian, 1965	57
<i>Cyptocephala</i> Berg, 1883	60
<i>Nezara</i> Amyot & Serville, 1843	60
<i>Roferta</i> Rolston, 1981	60
<i>Thyanta</i> Stål, 1862	61
Tribe Pentatomini	62
<i>Arvelius</i> Spinola, 1837	62
<i>Banasa</i> Stål, 1860	63
<i>Boea</i> Walker, 1867	65
<i>Elsiella</i> Froeschner, 1981	66
<i>Neotibilis</i> Grazia & Barcellos, 1994	66
<i>Pellaea</i> Stål, 1872	66
<i>Phalaeus</i> Stål, 1862	66
<i>Pharypia</i> Stål, 1861	67
<i>Placocoris</i> Mayr, 1864	67
<i>Serdia</i> Stål, 1860	69
<i>Taurocerus</i> Amyot & Serville, 1843	69
Tribe Piezodorini	69
<i>Piezodorus</i> Fieber, 1860	69
Tribe Procliticini	70
<i>Odmalea</i> Bergroth, 1914	70
<i>Pseudobaeus</i> Distant, 1911	70
Tribe Strachiini	71
<i>Murgantia</i> Stål, 1862	71

Unplaced	71
<i>Patanius</i> Rolston, 1987	71
Discussion	72
Acknowledgements	80
References	80

Abstract

A checklist of 5 subfamilies, 108 genera, and 246 species of stink bugs (Hemiptera, Pentatomidae) from Colombia is provided. Three genera [*Andrallus* Bergroth, *Placocoris* Mayr, and *Pseudobebaeus* Distant], and eight species [*Arocera spectabilis* (Drury), *Andrallus spinidens* (Fabricius), *Banasa excavata* Thomas, *Banasa saileri* Thomas, *Banasa varians* Stål, *Chinavia scutellata* (Distant), *Pelidnocoris haglundii* Ruckes, and *Pseudobebaeus truncatus* (Fallou)] are recorded for the first time from Colombia. Images of dorsal external habitus for all genera that occur in Colombia are offered. This is the first time a comprehensive list of the stink bugs from Colombia is presented, providing a baseline to develop further studies in this group of true bugs.

Key words: species list, stink bugs, Neotropical region, Heteroptera, Northwestern South America

Introduction

Stink bugs (Hemiptera: Pentatomidae) are a worldwide distributed group with approximately 900 genera and 5,000 species known, of which about 260 genera and nearly 1,600 species have been recorded in the Neotropical region (Grazia *et al.* 2015; Rider *et al.* 2018). Pentatomidae is the third largest family of Heteroptera in the world, and second in the Neotropics (Panizzi & Grazia 2015; Henry 2017). Most stink bugs are phytophagous, feeding on the sap of plants directly from the vascular system, and from seeds or fruits in development, including important crop plants. For this reason, many species are recognized as agricultural pests (Grazia & Schwertner 2017). Predatory feeding habits occur in species of the subfamily Asopinae, which have been considered as potential biological control agents (De Clercq 2008). Pentatomidae is strongly supported as a monophyletic group based on morphological and molecular evidence, with 9 subfamilies: Aphyliinae, Asopinae, Discocephalinae, Edessinae, Pentatominae, Phyllocephalinae, Podopinae, Serbaninae, and the exclusively Neotropical Strotarsinae (Grazia *et al.* 2008; Rider *et al.* 2018). Recent molecular phylogeny found Cyrtocorinae as an independent lineage from Pentatomidae, reinstating the status of family, despite the very low branch support values (Roca-Cusachs *et al.* 2022).

Colombia is one of the top megadiverse countries in the world, and more than 10% of the global biodiversity is recorded within its borders (Andrade-C 2011; Arbeláez-Cortés 2013b; SIB Colombia 2020). Due to its geographic location and diverse habitat conditions, with great altitudinal and climatic variation, several taxa present high species richness in Colombia. However, several groups of animals, including most groups of insects, are still understudied (e.g., Arbeláez-Cortés 2013a; Romero-Ortiz *et al.* 2019; Cárdenas-Bautista *et al.* 2020). Regional taxonomic species lists and information on geographic distribution are fundamental for understanding the evolution of the Neotropical fauna, thus highlighting the importance in generating diversity data for poorly known groups.

In recent years, taxonomic studies on Colombian stink bugs have been lacking. Information available is mostly outdated and scattered in the literature, e.g., the last published world catalogue of the group (Kirkaldy 1909), or modern descriptive or revisionary work at the subfamily (Thomas 1992; Packauskas & Schaefer 1998), tribal (Rolston 1992; Rider 1994), or generic levels (e.g., Rolston 1974; Grazia 1978; Barcellos & Grazia 2003; Schwertner & Grazia 2006; Fernandes 2010; Garbelotto *et al.* 2014). More recently, regional oriented studies include distributional records of 36 genera currently included in the subfamily Pentatominae (Torres 2004), and new records for the Colombian fauna (Castro-Huertas *et al.* 2015). No other work assembling taxonomic information about this important bug family in Colombia is available, and an accurate identification of the Pentatomidae local fauna is still hard to accomplish.

Considering the economic importance of this group, and the scarce knowledge about its diversity in Colombia, we compile the first checklist of genera and species of stink bugs recorded for the country. The list includes information on the general distribution, Departmental records, identification remarks and other general comments, and a complete set of references for each species. Habitus images of one species for each recorded genus in Colombia are also provided.

Materials and methods

Examined specimens. The checklist was elaborated based on Pentatomidae literature from the Neotropical region (catalogues, taxonomic revisions, new species descriptions, agronomic literature, etc.) containing records from Colombia. Additionally, we examined and identified specimens deposited in the following Colombian entomological collections: Instituto de Investigación de Recursos Biológicos Alexander von Humboldt, Villa de Leiva, Colombia (IAvH-E); Instituto de Ciencias Naturales of the Universidad Nacional de Colombia, Bogotá, Colombia (ICN), Museo Entomológico Facultad de Agronomía, Universidad Nacional de Colombia, Bogotá, Colombia (UNAB); Museo de la Universidad del Valle, Cali, Colombia (MUSENUV); and Museo Javeriano de Historia Natural, Pontificia Universidad Javeriana, Bogotá, Colombia (MPUJ_ENT). We also identified specimens from Colombia deposited at the Entomological Collection of the Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil (UFRG).

Structure of the checklist. Classification of subfamilies and tribes follows Rider *et al.* (2018). The list is arranged in alphabetical order (subfamily to species) and includes information on the general distribution (country records), distribution in Colombia (Department records), remarks including morphological characters useful for the recognition of each genus and species within the more inclusive taxa, and a list of references on taxonomy for each species. Subgenera are not considered in our list. The distribution in Colombia is based on the literature and information retrieved from the labels of the examined specimens. Examined material is listed in the main text to document the new records. For comparison purposes, information on Pentatomidae distribution from other countries was compiled from the literature, revised, and updated to the current classification.

Specimen imaging. We provide dorsal habitus photographs for all genera recorded from the country. Seventy-nine photographs are of specimens collected in Colombia and deposited in the entomological collections studied for this project. We used a dissecting scope Nikon SMZ1270 equipped with a Nikon D5300 digital camera, with image stacking and editing in Adobe Photoshop CS6 v13.1.2. The images of *Tyrannocoris nigriceps* and *Paralcippus dimidiatus* are courtesy of Jhon César Neita (IAvH-E). For the twenty-eight remaining photographs, we use specimens collected in other countries of Central and South America, as follow (see below for authors names): *Ab-laptus brevirostrum* [Costa Rica, holotype American Museum of Natural History AMNH, courtesy R. Salas], *Alveostethus politus* [Argentina, David A. Rider personal collection DAR], *Coriplatus depressus* [indeterminate locality, courtesy T. Garbelotto], *Patronatus binotatus* [Panama, DAR], *Platycarenum umbractulatus* [indeterminate locality, courtesy T. Garbelotto], *Phoeacia* sp. [Peru, MPUJ, original image], *Coranda picipes* [Peru, DAR], *Herrichella thoracica* [Colombia, holotype British Museum of Natural History BMNH, courtesy T. Roell], *Lincus rufospilotus* [French Guiana, DAR], *Melambyrsus hoplita* [Colombia, holotype Naturmuseum Senckenberg, Frankfurt am Main SMFD, image published in Campos & Roell (2017), courtesy of the authors], *Neodoxoplatys longirostra* [Mexico, AMNH], *Orbatina fuliginia* [Bolivia, DAR], *Phereclis pluto* [Colombia, holotype Swedish Museum of Natural History NHRS, image by Gunvi Lindberg, courtesy L. Campos], *Stalius tartareus* [Mexico, DAR], *Stapecolis bimaculatus* [Colombia, holotype AMNH, courtesy T. Garbelotto], *Doesburgedessa elongatispina* [Colombia, Museu de Zoologia da Universidade de São Paulo MZUSP, courtesy R. Carrenho], *Grammedessa stillativentris* [Colombia, UFRG, courtesy A. Correia], *Hypoxys* sp. [Brazil, MZUSP, courtesy R. Carrenho], *Amauromelpia miri* [Peru, Paratype UFRG, courtesy F. Bianchi], *Diceraeus melacanthus* [Brazil, MZUSP, courtesy L. Fonzaghi], *Hypatropis rolstoni* [Panama, UFRG, courtesy L. Barros], *Lattinellica decora* [Colombia, UFRG, courtesy L. Barros], *Paratibraca infuscata* [Surinam, UFRG, courtesy F. Bianchi], *Tibraca limbativentris* [Brazil, MZUSP], *Vulsirea violacea* [Brazil, MZUSP], *Nezara viridula* [Brazil, MZUSP], *Pseudobebaeus truncatus* [Brazil, UFRG], and *Patanius vitatus* [Brazil, MZUSP]. For the genus *Padaeus*, recorded from Colombia but without species identified, we included *Padaeus viduus* (Vollenhoven, 1868) as genus representative [Mexico, holotype DAR, photographer V. Carabajal, courtesy L. Barros]

Results

A total of 246 species in 108 genera and 5 subfamilies of Pentatomidae are recorded from Colombia, representing about 5% of the world fauna and 15% of the Neotropical fauna (Table 1). Colombia is among the six most diverse countries in the Neotropical region, with similar species richness to Costa Rica (267 spp.), Argentina (279 spp.), and Panamá (226 spp.) (Table 2). Seven taxa could be identified only at the genus level, as they did not fit any of

the known species and may represent new taxa. The genera *Boea* Walker, *Marmessus* Bergroth, *Padaeus* Stål, *Peromatus* Amyot & Serville, *Phoecia* Stål, *Phalaeus* Stål, and *Placocoris* Mayr all represent new records for the Colombian fauna and do not have recent revisionary work published (except *Padaeus*, Carabajal *et al.* in prep.).

Regarding the distribution pattern of the Colombian stink bug fauna throughout the Neotropical region, we recognize five major groups (Table 3). More than half of the species recorded for the country are only known from Northern South America. Eighteen species are endemics from Colombia, representing about 7,4% of the stink bug fauna in the country.

List of species

Subfamily Asopinae

Alcaeorrhynchus Bergroth, 1891

The genus can be identified by having the peritreme not elevated and terminating in a short or long sulcus that is not surrounded by a shagreened area of the cuticle, and the humeral angles produced and bidentate (Thomas 1992).

Alcaeorrhynchus grandis (Dallas, 1851) [Fig. 1]

Distribution. USA, Mexico, Cuba, Jamaica, Honduras, Trinidad and Tobago, Colombia, French Guiana, Ecuador, Peru, Brazil, Bolivia, Paraguay, Uruguay, and Argentina.

Distribution in Colombia. Chocó, Cundinamarca, Magdalena, Meta, and Valle del Cauca.

Remarks. *Alcaeorrhynchus grandis* has the humeral angles bifid and directed forward. It is the only species recorded widely in the Neotropical region. *Alcaeorrhynchus phymatophorus* (Palisot de Beauvois, 1805) is only known from the Caribbean, including the Florida Keys.

References. Thomas 1992.

Examined material. COLOMBIA: 1♂, **Chocó**, Cuevita, 01 vi 1950, L. Richter / ICN_055397 (ICN); 1♀, **Cundinamarca**, Bogotá, Barrio Galerías, 31 vi 2010, J. Avendaño (ICN); 1♂, **Magdalena**, Santa Marta, PNN Tayrona, 8 xii 1977, M. Romero / ICN_055398 (ICN); 1♀, **Meta**, Villavicencio, Bosque Bavaria, 600m, M. Rodríguez, 10 iv 2006 / MPUJ_ENT 0018128 (MPUJ_ENT); 1♂, 2♀, **Valle del Cauca**, Buenaventura, vii 1962 / MPUJ_ENT 0010679 (MPUJ_ENT); Cali, 1000 m, 20 vi 1980, 22792 (MUSENUV); Rio Maya, Playa el Ají, 0 m, 28 iv 1984, Yesid Solarte, 22791 (MUSENUV).

Andrallus Bergroth, 1905

The genus can be recognized by the humeral spine with a postapical tooth, and the profemur usually with a small anteapical tooth (Thomas 1992).

Andrallus spinidens (Fabricius, 1787) [Fig. 2]

Distribution. USA, Mexico, Nicaragua, Costa Rica, and Cuba. New record for Colombia.

Distribution in Colombia. Caldas, Cundinamarca, Meta, and Valle del Cauca.

Remarks. *Andrallus* is a monotypic genus.

References. Thomas 1992.

Examined material. COLOMBIA: 1♂, **Caldas**, Dorada, L. Loaiza, 15 viii 1992 (UNAB); 1♂, **Cundinamarca**, Girardot, 6 ix 1992 [handwriting, illegible] (UNAB); 2♀, **Meta**, Villavicencio, Corpoica La Libertad, 31 v 2000, Santana N, Mazorra M, Espinosa J (UNAB); 1♀, **Valle del Cauca**, Cali, v 1970, L. Angel / ICN_055393 (ICN).

Apateticus Dallas, 1851

The genus can be recognized by the mandibular plates distinctly convergent, and the humeral angles rounded (Thomas 1992).

Apateticus lineolatus (Herrich-Shaeffer, 1840) [Fig. 3]

Distribution. USA, Mexico, Costa Rica, Panama, El Salvador, Honduras, Colombia, and Venezuela.

Distribution in Colombia. Boyacá and Meta.

Remarks. *Apateticus lineolatus* has the basal abdominal spine robust and reaching the metacoxa, and the posterior angle of the pronotum without a tooth.

References. Thomas 1992.

Examined material. COLOMBIA, 1♂, 1♀, **Boyacá**, Paipa, Fuente termal, 14 vi 1970, G. Aguirre / ICN_055457 (ICN); Somondoco, Vda. Boyasegundo, Fca. Bellavista, 4°58'N, 73°27'W, 1500 m, 24 iii 1997, D. Forero / IAvH-E 05092 (IAvH-E); 2♂, **Meta**, Guayabetal, 12 ix 1976, N. Pinzón / ICN_055454 (ICN); Villavicencio—Restrepo Km 8, 500 m, 11 ix 1976, N. Pinzón / ICN_055456 (ICN).

Coryzorhaphis Spinola, 1837

The genus has the profemur unarmed, and the protibia at least somewhat dilated (Thomas 1992).

Coryzorhaphis carneolus Erichson, 1848 [Fig. 4]

Distribution. Colombia, Guyana, French Guiana, Brazil, Ecuador, Peru, and Bolivia.

Distribution in Colombia. Meta.

Remarks. *Coryzorhaphis carneolus* have the humeral angles acute, the apex of the clypeus whitish; the head, thorax, scutellum and corium with black spots, the head with a medial and broad whitish spot, and the apex of humeral angles black.

References. Thomas 1992; Castro-Huertas *et al.* 2015.

Coryzorhaphis cruciata Stål, 1870

Distribution. Mexico, Belize, Honduras, Panama, Colombia, Venezuela, and Brazil.

Distribution in Colombia. Boyacá.

Remarks. *Coryzorhaphis cruciata* resembles *C. carneolus*, but the coloration pattern is different in which the head has a longitudinal whitish band, and the apex of the humeral angles are orange.

References. Thomas 1992.

Examined material. COLOMBIA, 1♂, **Boyacá**, Puerto Boyacá, Puerto Romero, Hda. Puracé, 350 m, 26 iv 2001 (ICN).

Coryzorhaphis superba Breddin, 1906

Distribution. Colombia, Ecuador, Peru, Bolivia, and Uruguay.

Distribution in Colombia. Meta.

Remarks. *Coryzorhaphis superba* has the humeral angles projected as cylindrical spines, the corium has black spots, and the profemur is not entirely dark brown.

References. Thomas 1992; Dellapé *et al.* 2003; Castro-Huertas *et al.* 2015.

Discocera Laporte, 1833

The genus can be recognized by the antennal segment IV dilated, body dorsally strongly convex, and ventrally depressed (Thomas 1992).

Discocera coccinea (Fabricius, 1798) [Fig. 5]

Distribution. Brazil, Peru, Colombia, Ecuador, Surinam, Paraguay, and Bolivia.

Distribution in Colombia. Amazonas and Guainía.

Remarks. *Discocera coccinea* has the base of abdomen with a short tubercle, the humeral angles produced in a short spine, and the protibia not dilated.

References. Thomas 1992.

Examined material. COLOMBIA: 2♀, **Amazonas**, PNN Amacayacu, Tierra Firme, Jameo, 5 ix 1997, F. Fernández (ICN); Leticia, Km 10 vía Tarapacá, 150 m, 20 xi 2002, Est. Sist. Animal (ICN); 1 nymph, **Guainía**, Santa Rosa, Caño Bocón, 3°40'46"N, 68°2'26"W, 100 m, 4-13 xi 1996, S.A. Amézquita (ICN).

***Euthyrhyncus* Dallas, 1851**

The genus can be recognized by the elevated peritreme surrounded by a shagreened area of the cuticle, the protibia expanded, and the profemur unarmed (Thomas 1992).

Euthyrhyncus floridanus (Linnaeus, 1767) [Fig. 6]

Distribution. USA, Mexico, Guatemala, El Salvador, Honduras, Costa Rica, Panama, Colombia, Brazil, Ecuador, and Bolivia.

Distribution in Colombia. Boyacá, Cundinamarca, Santander, and Valle del Cauca.

Remarks. *Euthyrhyncus* is a monotypic genus.

References. Thomas 1992; Castro-Huertas *et al.* 2015.

Examined material. COLOMBIA: 2♀, **Valle del Cauca**, C.H. Anchicayá, 400 m, 1978, 1938 (MUSENUV); San Antonio, La Horqueta, 2100 m, 14 iii 1994, O. Grijalva, 1939 (MUSENUV).

***Heteroscelis* Latreille, 1829**

The genus can be recognized by having the base of the abdomen with an anteriorly directed spine or tubercle, the humeral angles cleft or bidentate apically, and the anterolateral prothorax convex (Thomas 1992).

Heteroscelis lepida (Stål, 1862) [Fig. 7]

Distribution. USA; Mexico, Guatemala, Panama, Colombia, Venezuela, French Guiana, and Ecuador.

Distribution in Colombia. Bolívar, Chocó, and Santander.

Remarks. *Heteroscelis lepida* has the protibia expanded, the mandibular plates as long as the clypeus, the profemur without anteapical spine, and the base of scutellum entirely orange.

References. Thomas 1992.

Examined material. COLOMBIA: 1♂, **Bolívar**, Zambrano, Hda. Monterrey, 9°37'48"N, 74°54'44"W, 70 m, F. Fernández & G. Ulloa / IAvH-E 87327 (IAvH-E); 1♂, **Chocó**, Riosucio, La Balsa, 7°2'26"N, 77°20'16"W, 65 m, Malaise, 29 xii 1993, F. Fernández / IAvH-E 87702 (IAvH-E); 1♂, **Santander**, Charalá, Virolin, 01 iii 1981 / ICN_055894 (ICN).

Heteroscelis servillei Laporte, 1833

Distribution. Colombia, Venezuela, Guyana, French Guiana, Surinam, Trinidad and Tobago, Brazil, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Chocó.

Remarks. *Heteroscelis servillei* has the protibia expanded and the mandibular plates longer than the clypeus.

References. Thomas 1992.

Examined material. COLOMBIA: 1♀, **Chocó**, Riosucio, La Balsa, 7°2'26"N, 77°20'16"W, 65 m, F. Fernández / IAvH-E 87582 (IAvH-E).

***Marmessus* Bergroth, 1891**

The genus has the base of the abdomen without spine or tubercle, and the profemur is unarmed (Thomas 1992).

Marmessus sp. [Fig. 8]

Distribution of the genus. Colombia, Brazil, Argentina, and Uruguay.

Distribution in Colombia. Meta.

Remarks. The genus was recorded for Colombia by Castro-Huertas *et al.* (2015) and noted the impossibility of the identification of the species. The specimen examined resembles, but it is not, *M. nigricornis* Stål, 1865, whose distribution is restricted to Southern South America (Brazil, Uruguay, and Argentina).

References. Thomas 1992; Castro-Huertas *et al.* 2015.

Oplomus Spinola, 1837

The genus can be recognized by the profemur with an antepical spine or tubercle, the protibia usually dilated, and the frenal margin of the scutellum shorter than the postfrenal margin (Thomas 1992).

Oplomus festivus Dallas, 1851 [Fig. 9]

Distribution. Panama, Colombia, Venezuela, Surinam, Ecuador, Brazil, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Casanare, Chocó, Cundinamarca, Meta, and Santander.

Remarks. *Oplomus festivus* has the paramere triangular, and the anterolateral pronotal margin bead not reflexed.

References. Thomas 1992; Rider & Rolston 1995.

Examined material. COLOMBIA, 1♂, **Casanare**, Mochuelo, Caserío indígena, Selva de galería, 500 m, 1 vii 1978, F. Ortiz / ICN_077198 (ICN); 1♂, **Chocó**, Riosucio, Tilupo, 5 iv 1978, H. Echeverri / ICN_077200 (ICN); 1♀, **Cundinamarca**, Chinauta, 1640 m, 27 x 1991, L. Rubiano / ICN_077185 (ICN); 1♂, **Meta**, Restrepo, Salinao (UPIN), 910 m, 2 v 1988 / ICN_077196 (ICN); 1♀, **Santander**, Carare-Opón, Campo Capote, Bosque margen, 31 vii 1968, H. Sturm / ICN_077184 (ICN).

Oplomus salamandra (Burmeister, 1835)

Distribution. Colombia, Venezuela, Surinam, Ecuador, Bolivia, Peru, and Argentina.

Distribution in Colombia. Casanare, Chocó, Cundinamarca, Meta, and Valle del Cauca.

Remarks. *Oplomus salamandra* has the paramere narrow and recurved, with the anterolateral pronotal margins reflexed.

References. Thomas 1992.

Examined material. COLOMBIA: 1♂, **Casanare**, Cusiana, Pozo M1, 480 m, malaise, 26 ix 1995, F. Fernández / ICN_077199 (ICN); 1♂, **Chocó**, Riosucio, Sautatá, 24 vii 1978, H. Echeverri / ICN_077197 (ICN); 1♂, **Cundinamarca**, Guayabetal, Susumuco, 950 m, 5 vi 1979, D.N. Pinto / ICN_077183 (ICN); 1♂, **Meta**, Villavicencio, Bosque Bavaria, 600 m, 10 iv 2005, Sistemática (ICN); 1♂, **Valle del Cauca**, Calima III, 14 iv 1994, G. Morante, 1927 (MUSENUV).

Podisus Herrich-Shaeffer, 1851

The genus can be recognized by the spine on the abdominal base usually short, not surpassing the metacoxa, the posterior tibia usually sulcate, the dorsal surface of the head usually with dense and dark punctate, and with the mandibular plates parallel (Thomas 1992).

Podisus aenescens (Stål, 1860) [Fig. 10]

Distribution. Mexico, Guatemala, Costa Rica, British West Indies, Panama, Colombia, French Guyana, Ecuador, Bolivia, Peru, Brazil, Paraguay, and Argentina.

Distribution in Colombia. Chocó, Cundinamarca, Meta, Nariño, Santander, and Valle del Cauca.

Remarks. *Podisus aenescens* has the anterolateral pronotal margin pale, calloused and smooth.

References. Thomas 1992.

Examined material. COLOMBIA: 1♂, 2♀, **Chocó**, Riosucio, Sautatá, 26 vii 1978, H. Echeverri / ICN_055931 / ICN_055951 (ICN); Peyé, 18 xi 1978, H. Echeverri / ICN_055929 (ICN); 1♂, **Cundinamarca**, Medina, Mesacura, 20 x 1989, G. Bernal / ICN_055956 (ICN); 1♀, **Meta**, Villavicencio, Bosque Bavaria. nr río Guatiquía 4°10.657'N, 73°38.852'W, 1684 ft, 3-5 vii 2013, J.E. Eger & A.A. Calixto (ICN); 1♀, **Nariño**, Tumaco, Vereda Imbili, margen

izquierdo río Mira, 8 vii 1984, A. Guerra / ICN_055952 (ICN); 1♀, **Santander**, Alto río Opón, 1050 m, 26 viii 1948, L. Richter / ICN_055955 (ICN); 2♀, **Valle del Cauca**, Restrepo, Campo Alegre, 1100 m, 10 ii 1984, O. Cepeda / ICN_055928 (ICN); Camp. Río bravo, 900 m, 6 ii 1984, B. Mongui / ICN_055943 (ICN).

Podisus cornutus (Dallas, 1851)

Distribution. Colombia and Brazil.

Distribution in Colombia. Unknown. This species was recorded from Colombia by Thomas (1992), but without specific locality.

Remarks. *Podisus cornutus* has the humeral angles produced, elongated and slender, and curving peritreme, reaching two thirds of the distance to the pleural edge.

References. Thomas 1992.

Podisus crassimargo (Stål, 1860)

Distribution. Panama, Colombia, Venezuela, Ecuador, Brazil, Chile, and Argentina.

Distribution in Colombia. Unknown. This species was recorded from Colombia by Thomas (1992), but without specific locality.

Remarks. *Podisus crassimargo* has the anterolateral margin of the pronotum not emarginated and rugulose, and a well developed, spinose humeri; the peritreme does not reach the middle of the metapleuron, and the abdominal tubercle is short, barely reaching the metacoxa, and the femora are immaculate.

References. Thomas 1992; Dellapé *et al.* 2015.

Podisus fuscescens (Dallas, 1851)

Distribution. Mexico, Honduras, Panama, Colombia, Venezuela, French Guiana, Brazil, Ecuador, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Caldas, Caquetá, Cundinamarca, Meta, Putumayo, Tolima, and Valle del Cauca.

Remarks. *Podisus fuscescens* has the anterolateral margins of the pronotum concolor with disc, and crenulated; the humeral angles are acute and laterally produced, with a well-developed posterior tooth (frequently as appearing a bifid humeral angle).

References. Brugnera *et al.* 2020b.

Podisus nigrispinus (Dallas, 1851)

Distribution. Costa Rica, Panama, Surinam, Colombia, Guyana, Ecuador, Brazil, Peru, Bolivia, Paraguay, Uruguay, and Argentina.

Distribution in Colombia. Cundinamarca, Meta, Quindío, Tolima, and Valle del Cauca.

Remarks. *Podisus nigrispinus* has the anterolateral pronotal margin paler than disk and the humeral angles bifid and dark.

References. Thomas 1992, Dellapé *et al.* 2003.

Examined material. **COLOMBIA:** 2♂, 1♀, **Cundinamarca**, La Vega, 26 iv 1974, A.B. Lotero / ICN_055942 (ICN); Tolemaida, 1 xi 1969, D. Castro / ICN_055953 (ICN); Villeta, 11 xii 1972, N. Bastidas / ICN_055941 (ICN); 1♀, **Meta**, Cumaral, Vía Veracruz, Vda. Laguna brava, Fca. El Campín, 4°9'N, 73°20'W, 400 m, 23 x 1994, M. Ospina / IAVH-E_05100 (IAVH-E); 1♂, **Quindío**, Salento, 1 viii 1990, G. Medina & C. Rodríguez / ICN_055944 (ICN); 1♀, **Tolima**, Icononzo, Vereda el Palmar, 1710 m, 20 iii 1978, R. Restrepo / ICN_055945 (ICN); 1♂, **Valle del Cauca**, Restrepo Vereda río bravo, 900 m, 9 ii 1984, I. de Arévalo / ICN_055954 (ICN).

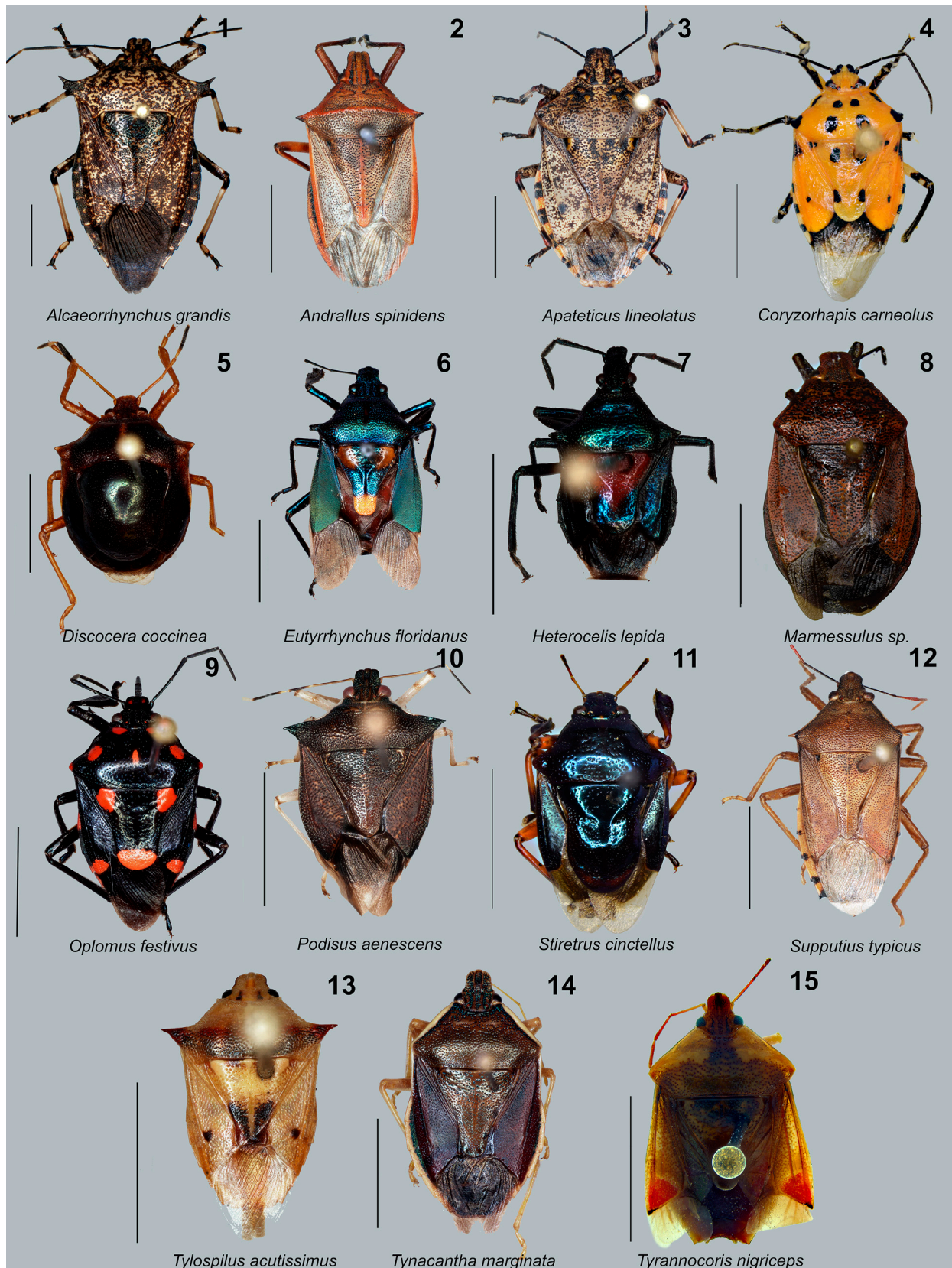
Podisus nigriventris Distant, 1880

Distribution. Mexico, Guatemala, Nicaragua, Costa Rica, Panama, and Colombia.

Distribution in Colombia. Unknown. This species was recorded from Colombia by Thomas (1992), but without specific locality.

Remarks. *Podisus nigriventris* has the abdomen ventrally dark, or at least with a row of dark spots on the midline.

References. Thomas 1992.



FIGURES 1–15. Asopinae genera in Colombia. 1. *Alcaeorrhynchus grandis* (MPUJ). 2. *Andrallus spinidens* (UNAB). 3. *Apateticus lineolatus* (MPUJ). 4. *Coryzorhaphis carneolus* (ICN). 5. *Discocera coccinea* (MPUJ). 6. *Eutyrrhynchus floridanus* (MPUJ). 7. *Heterocelis lepida* (MPUJ). 8. *Marmessus* sp. (MPUJ). 9. *Oplomus festivus* (MPUJ). 10. *Podisus aenescens* (MPUJ). 11. *Stiretrus cinctellus* (ICN). 12. *Supputius typicus* (MPUJ). 13. *Tylospilus acutissimus* (MPUJ). 14. *Tynacantha marginata* (MPUJ). 15. *Tyrannocoris nigriceps* (IAVH). Scale bar: 5 mm.

Podisus sagitta (Fabricius, 1794)

Distribution. USA, Mexico, Guatemala, El Salvador, Honduras, Nicaragua, Jamaica, Grenada, Dominican Republic, Haiti, Puerto Rico, Trinidad and Tobago, Curacao, Cuba, Bahamas, Costa Rica, Panama, Colombia, and Venezuela.

Distribution in Colombia. Valle del Cauca.

Remarks. *Podisus sagitta* has the anterolateral pronotal margin concolorous with the disk, and the females have a black spot on the lateral area of the abdominal sternite VII.

References. Thomas 1992; Castro-Huertas *et al.* 2015.

Examined material. COLOMBIA: 1♂, Valle del Cauca, P. Wilches, 1 x 1996, O. Grijalva, 15046 (MUSENUV).

Podisus tinctus (Dallas, 1851)

Distribution. Colombia and Ecuador.

Distribution in Colombia. Cundinamarca (Bogotá?).

Remarks. *Podisus tinctus* has the abdominal spine long, reaching the mesocoxae; and the humeral angles thick and acute.

References. Thomas 1992.

Podisus thomasi Lemaître, Roell & Brugnera, 2021

Distribution. Panama, Colombia, Venezuela, and Ecuador.

Distribution in Colombia. Boyacá, Cundinamarca, and Tolima.

Remarks. *Podisus ventralis* has the pronotum without spots, the anterolateral pronotal margins are not calloused, crenulated, and paler than pronotal disc; the humeral angles are angular and elongated, without a posterior tooth, and laterally directed; and the apex of the scutellum is paler than the disc.

References. Roell *et al.* 2021.

Podisus ventralis (Dallas, 1851)

Distribution. Colombia, Venezuela, Ecuador, Brazil, Peru, and Argentina.

Distribution in Colombia. Boyacá, Cundinamarca, and Risaralda.

Remarks. *Podisus ventralis* has the pronotum without spots, the anterolateral pronotal margins are calloused, crenulated, and paler than the pronotal disc; the humeral angles are angular, without a posterior tooth and laterally directed; and the apex of the scutellum is concolorous with the disc.

References. Roell *et al.* 2021.

Stiretrus Laporte, 1833

The genus can be recognized by the scutellum greatly enlarged, covering most of the abdomen, the apex surpassing the apex of the corium and nearly attaining apex of abdomen, antennal segment IV cylindrical, not dilated, body moderately convex dorsally and ventrally (Thomas 1992).

Stiretrus anchorago (Fabricius, 1775)

Distribution. USA, Mexico, Nicaragua, Guatemala, El Salvador, Costa Rica, Honduras, Panama, and Colombia.

Distribution in Colombia. Chocó.

Remarks. *Stiretrus anchorago* has the anterolateral pronotal margin strongly sinuated in dorsal view and the peritreme long, reaching at least halfway to pleural margin.

References. Thomas 1992; Castro-Huertas *et al.* 2015.

Stiretrus bifrenatus Breddin, 1906

Distribution. Colombia, Ecuador, Brazil, and Peru.

Distribution in Colombia. Unknown. This species was recorded from Colombia by Thomas (1992), but without specific locality.

Remarks. *Stiretrus bifrenatus* has the coloration pattern uniform metallic blue, and the tooth on the anterolateral pronotal angles is produced laterad of the eyes by about half the width of an eye.

References. Thomas 1992.

Stiretrus cinctellus Germar, 1839 [Fig. 11]

Distribution. Colombia, Brazil, and Argentina.

Distribution in Colombia. Chocó.

Remarks. *Stiretrus cinctellus* has the protibia expanded, and the basal abdominal spine short, not surpassing the metacoxa.

References. Thomas 1992; Castro-Huertas *et al.* 2015.

***Supputius* Distant, 1889**

The genus can be recognized by the short and simple humeral spine, and the unarmed profemur (Thomas 1992).

Supputius typicus Distant, 1889 [Fig. 12]

Distribution. Mexico, Costa Rica, Panama, Colombia, Venezuela, Ecuador, and Brazil.

Distribution in Colombia. Boyacá, Meta, and Valle del Cauca.

Remarks. *Supputius typicus* has the base of the abdomen without a tubercle.

References. Thomas 1992.

Examined material. COLOMBIA: 1♀, **Boyacá**, Villa de Leiva, 1 xi 1998, Líquido 1115, D. Forero / IAvH-E_151829 (IAvH-ENT); 2♀, **Meta**, Guayabetal, Manzanares, 1100 m, 1 vii 1940, L. Richter / ICN_055907 / ICN_055908 (ICN); 3♀, **Valle del Cauca**, Cali, Ca. Cali, 1000 m, 1972, 1959 (MUSENUV); Cali, Tierra cult., 1000 m, 1973, 1958 (MUSENUV); Univalle, en Samanea saman, 3°22'62.7"N, 76°32'14.9"W, 970 m, 14 ii 2005, Entomophilo (MUSENUV).

***Tylospilus* Stål, 1870**

The genus can be recognized by the long spine on abdominal base extending to the mesocoxa, the metatibia cylindrical or slightly tapering, most or all of the dorsal surface of the head with pale colorless punctation; peritreme auriculate short, reaching less than halfway to the pleural edge, and the anterolateral pronotal margin from rugulose to smooth (Thomas 1992).

Tylospilus acutissimus Stål, 1870 [Fig. 13]

Distribution. USA, Mexico, Nicaragua, “Antilles”, and Colombia.

Distribution in Colombia. Meta.

Remarks. *Tylospilus acutissimus* has the base of the scutellum with indistinct calli at the basal angles.

References. Thomas 1992; Brugnera *et al.* 2020a.

Tylospilus cloelia (Stål, 1862)

Distribution. Mexico, Panama, Colombia, Bolivia, Brazil, Paraguay, and Argentina.

Distribution in Colombia. Unknown. This species was recorded from Colombia by Thomas (1992), but without specific locality.

Remarks. *Tylospilus cloelia* has the anterolateral pronotal margin smooth, pale and calloused.

References. Thomas 1992; Brugnera *et al.* 2020a.

Tylospilus peruvianus Horváth, 1911

Distribution. Colombia, Surinam, Brazil, Peru, Bolivia, and Argentina.

Distribution in Colombia. Meta.

Remarks. *Tylospilus peruvianus* has the humeral angles acute but without a spiniform process, and the clypeus pale,

concolorous with the mandibular plates.

References. Thomas 1992; Castro-Huertas *et al.* 2015; Brugnera *et al.* 2020a; Dellapé 2021a.

***Tynacantha* Dallas, 1851**

The genus can be recognized by the anterolateral pronotal margins and outer connexivum calloused and impunctate (Thomas 1992).

Tynacantha marginata Dallas, 1851 [Fig. 14]

Distribution. Colombia, Venezuela, Ecuador, Brazil, Peru, Bolivia, Paraguay, Uruguay, and Argentina.

Distribution in Colombia. Antioquia and Cundinamarca.

Remarks. *Tynacantha marginata* has the scutellum concolorous dark.

References. Thomas 1992; Brugnera *et al.* 2019.

***Tyrannocoris* Thomas, 1992**

The genus can be recognized by the anterolateral pronotal margins and connexivum punctate to edge, not calloused (Thomas 1992).

Tyrannocoris nigriceps Thomas, 1992 [Fig. 15]

Distribution. Colombia, Brazil, and Argentina.

Distribution in Colombia. Bolívar.

Remarks. *Tyrannocoris nigriceps* has the dorsal region of head dark, connexivum unicolorous, and the anterior region of the pronotum pale, interrupted by a medial dark brown band.

References. Thomas 1992; Castro-Huertas *et al.* 2015.

Subfamily Cyrtocorinae

***Ceratozygum* Horváth, 1916**

The genus can be recognized by the scutellar process, measured from the abdominal dorsum longer than length of the scutellum, notched apically, with two large basal anterior tubercles; and the humeral processes strongly expanded apically, nearly two times wider apically than at base (Packauskas & Schaefer 1998).

Ceratozygum horridum (Germar, 1839) [Fig. 16]

Distribution. Panama, Colombia, Venezuela, Brazil, and Peru.

Distribution in Colombia. Amazonas and Casanare.

Remarks. *Ceratozygum* is a monotypic genus.

References. Packauskas & Schaefer 1998; Castro-Huertas *et al.* 2015.

Examined material. COLOMBIA, Casanare, municipio de Trinidad, reserva natural La Palmita, 05.4202°N 71.5997°W, 182m, 18-23 viii 2019 / colecta manual, bosque de galería inundable, sobre vegetación / A. Fajardo & L. Castiblanco, 4 adults / MPUJ_ENT 0068053 - MPUJ_ENT 0068056; *idem*, C. Salazar & L. Pineda 5 adults / MPUJ_ENT 0068590 - MPUJ_ENT 0068594, 2 nymphs / MPUJ_ENT 0068595 - MPUJ_ENT 0068596, 13 nymphs in 96% ETOH / MPUJ_ENT 0068671 (MPUJ_ENT).

***Cyrtocoris* White, 1842**

The genus can be recognized by the scutellar process measured from abdominal dorsum shorter than the length of the scutellum, the anterior angles of the pronotum consisting of teeth, and shorter than half the length of the humeral processes (Packauskas & Schaefer 1998).

Cyrtocoris egeris Packauskas & Schaefer, 1998 [Fig. 17]

Distribution. Mexico, Honduras, Costa Rica, Panama, Colombia, Trinidad and Tobago, Venezuela, Ecuador, Bolivia, Brazil, and Argentina.

Distribution in Colombia. Casanare, Chocó, Magdalena, Meta, Putumayo, and Santander.

Remarks. *Cyrtocoris egeris* has the labium not extending beyond the anterior margin of the metasternum, rarely reaching the anterior margin of the metacoxa.

References. Packauskas & Schaefer 1998.

Examined material. COLOMBIA: 1♂, **Casanare**, Cusiana, Pozo M1, 480 m, Malaise, 28 ix 1995, F. Fernández / IAvH-E_88081 (IAvH-E); 1♂, **Chocó**, PNN Los katíos, Centro Administrativo, Sautatá, Fuera del bosque, 7°51'N, 77°8'W, 30 m, Malaise, 6 v-13 vi 2003, Líquido 3758, P. López / IAvH-E_151786 (IAvH_ENT); 1♂, 1♀, **Magdalena**, PNN Tayrona, Cañaveral, barridas, 8 xi 1977, C. Kugler (IAvH-E); 1♀, **Meta**, Villavicencio, Vía Puerto Colombia, 1h, 23 ix 1997, E. González / IAvH-E 88080 (IAvH-E).

Cyrtocoris gibbus (Fabricius, 1803)

Distribution. Guatemala, Costa Rica, Colombia, Venezuela, Brazil, and Argentina.

Distribution in Colombia. Valle del Cauca.

Remarks. *Cyrtocoris gibbus* has the humeral expansions nearly reaching the level of the anterior angles of the pronotum, strongly surpassing a line drawn through the bases of the larger anterior angled spines.

References. Packauskas & Schaefer 1998.

Examined material. COLOMBIA: 2♂, 1♀, **Valle del Cauca**, Ca. Buenaventura, 385 m, 24 v 2008, Mx. Urrutia, 21615 (MUSENUV); Dagua, Centella, 3°39'37"N, 76°41'34"W, Jama, 3 xi 2007, Doris Canacuan, 22965 (MUSENUV); Mateguadua, 4°1'27.1"N, 76°9'47.4"W, 1104 m, 29 ix 2007, Andrés Escobar, 22884 (MUSENUV).

Cyrtocoris trigonus (Germar, 1839)

Distribution. Mexico, Guatemala, Costa Rica, Panama, Colombia, Brazil, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Putumayo and Valle del Cauca.

Remarks. *Cyrtocoris trigonus* has the labium not extending beyond anterior margin of the metasternum, never reaching the metacoxa, the scutellum is produced as a low ridge, the humeral expansions not surpassing the level of the bases of the anterior angles of the pronotum.

References. Packauskas & Schaefer 1998.

Subfamily Discocephalinae

Tribe Discocephalini

Ablaptus Stål, 1872

The genus can be recognized by the interocular region is shorter than the length of the head, the apex of the mandibular plates is overlapping, the antennae has five segments, the first antennal segment is nearly reaching the anterior margin of the head, the bucculae is weakly produced anteriorly, the mesosternum is stoutly carinate medially, the humeral angles slightly projected but without a process (spine or tubercle), and the male with a transversal subapical sulcus on the hemelytra (Rolston 1988, Becker & Grazia 1989).

Ablaptus brevirostrum Rolston, 1988 [Fig. 18]

Distribution. Costa Rica and Colombia.

Distribution in Colombia. Valle del Cauca.

Remarks. *Ablaptus brevirostrum* has the lateral projections of the pygophore wider and apically acute and curved in the male, and the posterior margin of the gonocoxite VIII convex uniformly in the female.

References. Rolston 1988; Becker & Grazia 1989.

Alveostethus Ruckes, 1966

The genus can be recognized by the oval body, the impunctate metasternum with its lateral margins thickened, elevated, longitudinally furrowed, or deeply sulcate to form a canal in which the terminal segments of the labium lie; margin of the body from the apex of the head to the base of the hemelytron forming more or less an even, continuous arc (Ruckes 1966b).

Alveostethus politus (Signoret, 1851) [Fig. 19]

Distribution. Colombia, Venezuela, Brazil, Peru, and Argentina.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Ruckes (1966b), but without specific locality.

Remarks. *Alveostethus politus* has the eyes sloping posteriorly over the humeral angles of the pronotum; ocelli in line with mid region of eyes; scutellum very smooth, with a large, piceous, discal spot and conspicuous pale brown apex.

References. Ruckes 1966b, Dellapé *et al.* 2015.

Agaclitus Stål, 1864

This genus can be recognized by having the mandibular plates surpassing the clypeus, the apical margins of the mandibular plates are adjacent; the humeral angles with a spine, and the scutellum with a conspicuous basal tubercle (Becker & Grazia 1992).

Agaclitus dromedarius Stål, 1864 [Fig. 20]

Distribution. Colombia, French Guiana, Brazil, Peru, and Bolivia.

Distribution in Colombia. Amazonas.

Remarks. *Agaclitus dromedarius* has a conspicuous tubercle on the scutellum, nearly as long as wide.

References. Becker & Grazia 1992; Castro-Huertas *et al.* 2015.

Antiteuchus Dallas, 1851

This genus can be recognized by the oval, shiny, and smooth body; the head nearly as wide as long before the eyes, the labium reaching the abdominal segment V, connexivum visible and usually with a coloration pattern mixed in pale and dark areas (Fernandes & Grazia 2006).

Antiteuchus graziae Engleman, 1983

Distribution. Colombia, Brazil, Ecuador, Peru, and Bolivia.

Distribution in Colombia. Vaupés.

Remarks. *Antiteuchus graziae* has the median process of the urotergite VII absent, with a concavity, the laterodorsal process of the pygophore conspicuous and projected, and the dorsal margin of the paramere dorsally with several carinas.

References. Fernandes & Grazia 2006.

Antiteuchus cuspidatus Ruckes, 1964

Distribution. Panama and Colombia.

Distribution in Colombia. Cundinamarca.

Remarks. *Antiteuchus cuspidatus* has the apex of the median process of the urotergite VII expanded, with its posterior margin slightly sulcate, the posterolateral angle of the pygophore with a conspicuous bicuspid process.

References. Fernandes & Grazia 2006.

Antiteuchus macrasis (Perty, 1834)

Distribution. Costa Rica, Panama, Colombia, Venezuela, Surinam, French Guiana, Brazil, and Peru.

Distribution in Colombia. Huila.

Remarks. *Antiteuchus macrasis* has the apex of the median process of the urotergite VII slightly wider, and the posterolateral region of the abdominal segment X projected posteriorly.

References. Fernandes & Grazia 2006.

Antiteuchus melanoleucus (Westwood, 1837)

Distribution. Colombia, Venezuela, Guyana, Surinam, Brazil, Peru, Bolivia, and Argentina.

Distribution in Colombia. Meta.

Remarks. *Antiteuchus melanoleucus* has the body dorsally with brown or dark brown spots, and the margin of the urotergite VII with several setae.

References. Fernandes & Grazia 2006; Castro-Huertas *et al.* 2015.

Antiteuchus rolstoni Engleman, 1976

Distribution. Colombia and Ecuador.

Distribution in Colombia. Amazonas.

Remarks. *Antiteuchus rolstoni* has the posterolateral angle of the pygophore narrow, and the paramere dorsally wide in posterior view.

References. Fernandes & Grazia 2006.

Antiteuchus nigricans Ruckes, 1964

Distribution. Colombia and Ecuador.

Distribution in Colombia. Antioquia, Huila, and Valle del Cauca.

Remarks. *Antiteuchus nigricans* has the body ventrally with a yellow to pale brown with dark brown spots coloration pattern, and the posterolateral angle of the pygophore nearly as long as the sclerotized area of the abdominal segment X.

References. Fernandes & Grazia 2006.

Antiteuchus sepulcralis (Fabricius, 1803)

Distribution. Trinidad and Tobago, Colombia, Venezuela, Surinam, Brazil, Bolivia, and Argentina.

Distribution in Colombia. Amazonas and Meta.

Remarks. *Antiteuchus sepulcralis* has the median process of the urotergite VII long, with the apex wide, and the ventral surface of the pygophore glabrous.

References. Fernandes & Grazia 2006; Castro-Huertas *et al.* 2015.

Antiteuchus tripterus (Fabricius, 1787)

Distribution. Panama, Trinidad and Tobago, Colombia, Venezuela, Brazil, Ecuador, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Meta and Valle del Cauca.

Remarks. *Antiteuchus tripterus* has the abdominal segment X globose.

References. Fernandes & Grazia 2006.

***Coriplatus* White, 1842**

The genus can be recognized by the head being longer than the length of the pronotum, the anterolateral margin of the pronotum terminating posteriorly in a laterally directed, slightly curved, narrowly triangular spine behind which is a shallow parahumeral curve, and the margin of the postfrenal scutellar lobe reflexed midway along its length (Ruckes & Becker 1970).

Coriplatus depressus White, 1842 [Fig. 22]

Distribution. Colombia and Guyana.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Ruckes & Becker (1970), but without specific locality.

Remarks. *Coriplatus* is a monotypic genus.

References. Ruckes & Becker 1970.

Colpocarena Stål, 1868

The genus can be recognized by the subrectangular pronotum, its margin as wide as the head across the eyes, the margins of the mandibular plates distinctly sinuate before eyes, the scutellum is attaining the apex of the abdominal tergite V, the postfrenal lobe is narrowed, and the external apical angle of the corium is acute (Becker 1977).

Colpocarena sp. [Fig. 23]

Distribution. Colombia, Venezuela, Surinam, and Brazil.

Distribution in Colombia. Chocó, Risaralda, and Valle del Cauca.

Remarks. *Colpocarena complanata* (Burmeister, 1835) is the only species of this monotypic genus, but with at least an undescribed species (Rider 2018). Dallas (1851) recorded *C. complanata* for Colombia, but the specimen does not have a specific locality. The colombian specimens seem to be different from the known species.

References. Dallas 1851; Becker 1977.

Examined material. COLOMBIA: 1♀, **Chocó**, Acandí, Capurganá, 260m, 7 x 2007, Bosque Girasoles, 16:00 pm, manual, Arciniegas, J, *et al.* / MPUJ_ENT 0002267 (MPUJ); 1♂, Bosque, 330 m, 8 x 2017, manual. 16h, Arciniegas *et al.* (MPUJ); 1♀, Jardín Botánico del Darién, rastrojo, 90 m, 28 iii 2009, manual, N. Novoa (MPUJ); 1♂, **Risaralda**, Mun. de Pueblo Rico, Corregimiento de Santa Cecilia, área Amurrapá, □1.1 km WSW de Santa Cecilia, 05.33783 °N 76.15532 °W, 402 m, 19-23 ii 2018, J. Bolívar / Bosque húmedo tropical, margen del río, colecta manual / MPUJ_ENT 0060437 (MPUJ); 1♀, Pueblo Rico, Montezuma Ecolodge, 5.9 km WbN de Pueblo Rico, 05.23552 °N 76.08327 °W, 1456 m, 12-15 ix 2017, M.C. Moreno, Camino 2 / MPUJ_ENT 0062287 (MPUJ); 1♀, **Valle del Cauca**, Buga, laguna del Sonso, 03.88340 °N 76.34908 °W, 965m, 1-2 iii 2017, C. Castillo / MPUJ_ENT 0003509 (MPUJ); 1♀, M. Peña / MPUJ_ENT 0050288 (MPUJ).

Dinocoris Burmeister, 1835

The genus can be recognized by the antenna usually having four segments, the first antennal segment is the widest, and it is surpassing the mandibular plates; the pronotum is trapezoid, the humeral angles not projected, the mesosternum is elevated, the metasternum is carinated, the scutellum is triangular and reaching or surpassing the abdominal segment VI; and with the coloration pattern usually yellow to pale brown with several dark brown spots dorsally (Becker & Grazia 1985).

Dinocoris gibbosus (Fallou, 1889)

Distribution. Panama, Colombia, Venezuela, and Brazil.

Distribution in Colombia. Meta.

Remarks. *Dinocoris gibbosus* has the tarsus yellow to pale brown, with the posterior portion of the last tarsal segment dark brown.

References. Becker & Grazia 1985; Castro-Huertas *et al.* 2015.

Dinocoris maculatus Laporte, 1832

Distribution. Colombia, Venezuela, Brazil, and Peru.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Becker & Grazia (1985), but the specimen does not have a specific locality.

Remarks. *Dinocoris maculatus* has a testaceous corium with a spot on the radial vein.

References. Becker & Grazia 1985.

Dinocoris nigroantennatus Becker & Grazia, 1985

Distribution. Colombia and Peru.

Distribution in Colombia. Cundinamarca.

Remarks. *Dinocoris nigroantennatus* has the antennae entirely dark brown, and the corium with transversal reddish bands.

References. Becker & Grazia 1985.

Dinocoris rufitarsus Ruckes, 1958

Distribution. Honduras, Nicaragua, Panama, Colombia, and Brazil.

Distribution in Colombia. Chocó, Cundinamarca, Santander, and Valle del Cauca.

Remarks. *Dinocoris rufitarsus* has the tarsus entirely reddish, and sometimes the apex of the tarsal segment III dark brown.

References. Becker & Grazia 1985.

Examined material. COLOMBIA: 1♀, **Chocó**, Riosucio, Sautatá, 24 iv 1978, H. Echeverri / PK 00783 (ICN); 1♀, **Meta**, Macarena, Río Güejar, 380-420 m, 1 i 1951, L. Richter (ICN); 1♂, 1♀, **Santander**, Alto río Opón, 800 m, 1 i 1950, L. Richter (ICN).

Dinocoris variolosus (Linnaeus, 1758)

Distribution. Trinidad and Tobago, Panama, Colombia, Venezuela, and French Guiana.

Distribution in Colombia. Meta.

Remarks. *Dinocoris variolosus* has the abdomen entirely yellow, and both the pronotum and the corium with sparse spots without stripes.

References. Becker & Grazia 1985; Castro-Huertas *et al.* 2015.

***Discocephalessa* Kirkaldy, 1909**

This genus can be recognized by having the mesosternum with a tumid portion of on each side of medial sulcus, the medial sulcus largely smooth and impunctate; the antennal segment II slightly longer than I, and the anterolateral pronotal margins entire (Ruckes 1966b).

Discocephalessa humilis (Herrich-Schaeffer, 1843) [Fig. 25]

Distribution. Colombia.

Distribution in Colombia. Bolívar.

Remarks. *Discocephalessa humilis* has less than 10 mm in length, the pronotum has a shallow transverse furrow across the disc, and the scutellum has three pale points across the basal margin.

References. Ruckes 1966b.

Examined material. COLOMBIA: 1♂, **Bolívar**, Tierrabomba, Bosque seco, 10°21'36"N, 75°34'30"W, 100 m, manual, F. Escobar / IAvH-E_05055 (IAvH-E).

Discocephalessa terminalis (Walker, 1867)

Distribution. Colombia, Brazil, and Bolivia.

Distribution in Colombia. Amazonas.

Remarks. *Discocephalessa terminalis* has the pronotum transversely subrectangular, with a shallow furrow across the disc and with six irregular points along the anterior margin of the furrow; the apex of the scutellum is yellowish, and the corium has a pale discal spot.

References. Ruckes 1966b; Castro-Huertas *et al.* 2015.

***Dryptocephala* Laporte, 1832**

This genus can be recognized by having the first labial segment just reaching the anterior margin of the prosternum,

a cylindrical metatibia, with a narrow dorsal groove or impressed line, and the scutellum not surpassing the abdominal tergite V (Ruckes 1966c).

Dryptocephala crenata Ruckes, 1966

Distribution. Colombia and Peru.

Distribution in Colombia. Caquetá.

Remarks. *Dryptocephala crenata* is dorsally testaceous, the head and the pronotum are orange-red, the anterolateral pronotal margins are deeply dentated with the posterior angles rounded, and each connexival segment with a conspicuous orange-red spot.

References. Ruckes 1966c; Castro-Huertas *et al.* 2015.

Dryptocephala dentifrons (Latreille, 1811) [Fig. 26]

Distribution. Colombia and Peru.

Distribution in Colombia. Valle del Cauca.

Remarks. *Dryptocephala dentifrons* has the inner margins of the mandibular plates separated, with the apical incisure more evident, the apical angles of the connexival segments subacute, slightly produced.

References. Ruckes 1966c; Castro-Huertas *et al.* 2015.

Dryptocephala lurida Erichson, 1848

Distribution. Colombia, Guyana, Brazil, and Argentina.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Ruckes (1966c), but the specimen examined is without specific locality.

Remarks. *Dryptocephala lurida* has the apex of the antecular process not reaching the middle of the margin of the mandibular plates, the anterior apical pronotal angles are sub-obtuse, barely reaching a line drawn through the eyes.

References. Ruckes 1966c.

Dryptocephala obtusiceps Stål, 1872

Distribution. Panama, Colombia, Ecuador, Brazil, and Peru.

Distribution in Colombia. Cundinamarca.

Remarks. *Dryptocephala obtusiceps* has the internal margins of the mandibular plates contiguous for almost their entire length, and the apical angles of the connexival segments rectilinear, not produced.

References. Ruckes 1966c.

***Eurystethus* Mayr, 1864**

This genus can be recognized by having body depressed oval to elliptical, the head is usually slightly shorter than the median length of the pronotum, the mesosternum is hexagonal and wider than long, the scutellum does not reach the apex of abdomen, the labial segments I and II with an intercalary unit between them, the surface of the pronotum and scutellum has tubercles or processes, and the basal region of the scutellum with a pair of large tubercles (Ruckes 1966d).

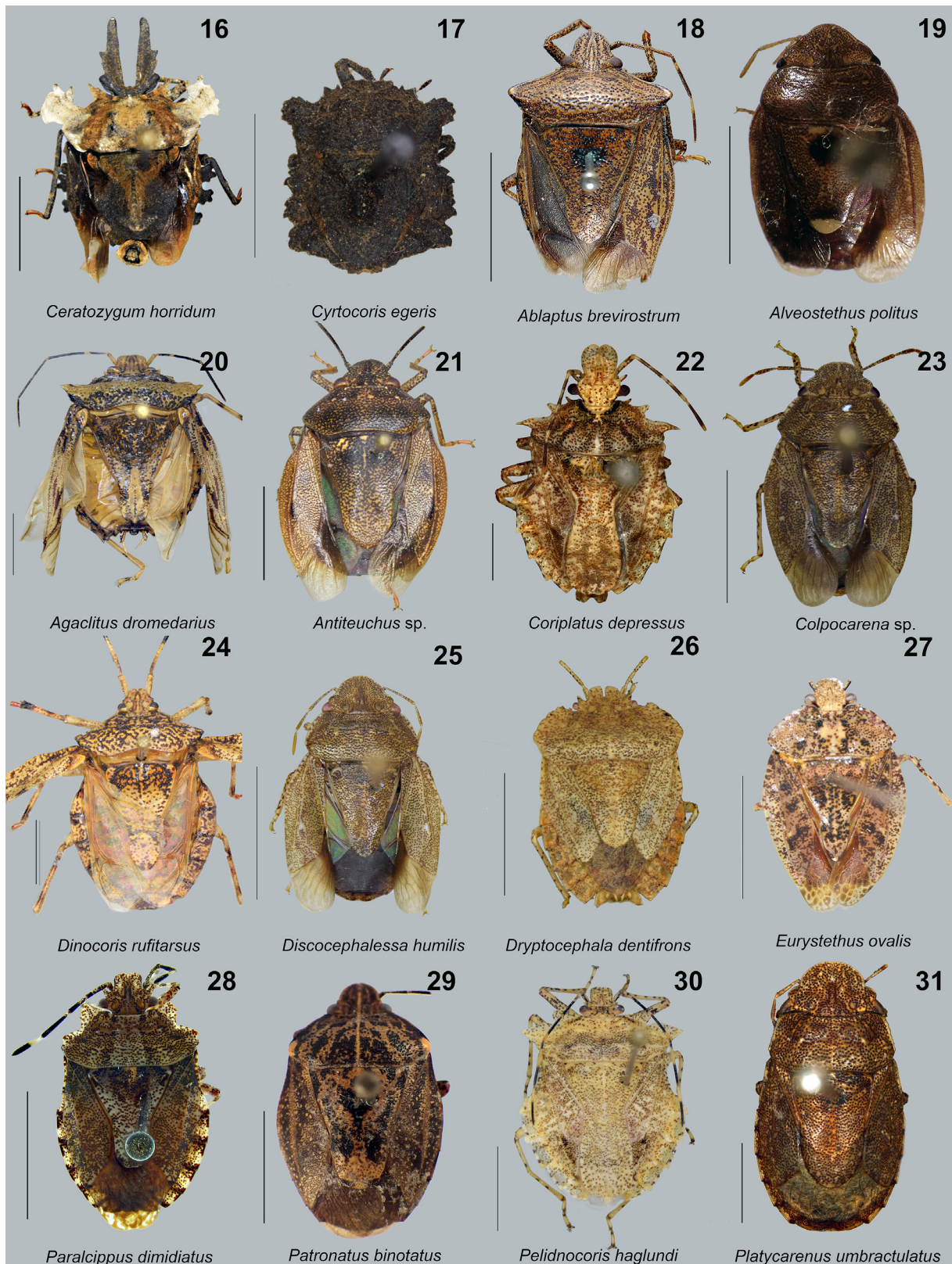
Eurystethus ovalis Ruckes, 1966 [Fig. 27]

Distribution. Colombia, Peru, and Bolivia.

Distribution in Colombia. Santander.

Remarks. *Eurystethus ovalis* has the apex of the head narrowly rounded, subangular, its margins reflexed, the disc concave, the antennal segment II shorter than I, subequal to III, and the margins of the scutellar apex reflexed.

References. Ruckes 1966d; Castro-Huertas *et al.* 2015.



FIGURES 16–31. Cyrtocorinae and Discocephalinae genera in Colombia. Cyrtocorinae: 16. *Ceratozygum horridum* (MPUJ). 17. *Cyrtocoris egeris* (ICN). Discocephalinae: 18. *Ablaptus brevirostrum* (AMNH). 19. *Alveostethus politus* (DAR). 20. *Agaclytus dromedarius* (MPUJ). 21. *Antiteuchus* sp. (MPUJ). 22. *Coriplatus depressus* (Indet.). 23. *Colpocarena complanata* (MPUJ). 24. *Dinocoris* sp. (MPUJ). 25. *Discocephalessa humilis* (ICN). 26. *Dryptocephala dentifrons* (MPUJ). 27. *Eurystethus ovalis* (MPUJ). 28. *Paralcippus dimidiatus* (IAVH). 29. *Patronatus binotatus* (DAR). 30. *Pelidnocoris haglundi* (MUSENUV). 31. *Platycarenum umbractulatus* (Indet.). Scale bar: 5 mm.

***Paralcippus* Becker & Grazia, 1986**

This genus can be recognized by having the head as long as the median length of the pronotum, the antecular region has a prominent lingulate process, the mandibular plates are wider, its apex overlapping in front of the clypeus, the antenniferous tubercles visible from above, the antennae has five segments, the antennal segment I not reaching the anterior margin of the head, the antennal segment II is longer than III, the labium reaching the abdominal segment IV, and the apex of the humeral angle with a tubercle (Becker & Grazia 1986).

Paralcippus dimidiatus (Ruckes, 1959) [Fig. 28]

Distribution. Colombia and Ecuador.

Distribution in Colombia. Risaralda.

Remarks. *Paralcippus* is a monotypic genus.

References. Becker & Grazia 1986; Castro-Huertas *et al.* 2015.

***Patronatus* Ruckes, 1965**

The genus can be recognized by the pronotum with a longitudinal pale brown band, and the labium not exceeding the middle of the abdominal disc (Ruckes 1965).

Patronatus binotatus Ruckes, 1965 [Fig. 29]

Distribution. Panama and Colombia

Distribution in Colombia. “Santo Domingo”. This species was recorded for Colombia by Ruckes (1965), but the specimen deposited in the AMNH could not be examined, and the specific locality is ambiguous as there are several places in Colombia with that name.

Remarks. *Patronatus binotatus* has the humeral angles calloused and yellowish.

References. Ruckes 1965.

***Pelidnocoris* Stål, 1867**

The genus can be recognized by the overlapping mandibular plates, the antecular region with a process, and the lateral margin of postfrenal lobe of the scutellum not elevated (Ruckes 1966a).

Pelidnocoris stalii Haglund, 1868

Distribution. Mexico, Costa Rica, Panama, and Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Ruckes (1966a), but without specific locality.

Remarks. *Pelidnocoris stalii* has the apical angle of the posterior lobe of the pronotal margin acute, and the anterior lobe of the pronotal margin acutely triangular.

References. Ruckes 1966a.

Pelidnocoris haglundi Ruckes, 1966 [Fig. 30]

Distribution. Brazil. New record for Colombia.

Distribution in Colombia. Cauca.

Remarks. *Pelidnocoris haglundi* can be recognized by the short antecular processes, acutely triangular, not surpassing middle of the eyes; apex of the posterior lobe of the pronotal margin acutely angled; and the pronotal margin with a distinct notch posteriorly to the humeral angles.

References. Ruckes 1966a.

Examined material. COLOMBIA: 1♂, Cauca, Caloto, vereda Morales, 2°1'59.8" N, 76°24'42.3" W, 129 m, manual, 4 x 2009, J. Mendivil (MUSENUV).

***Platycarenum* Fieber, 1861**

The genus can be recognized by the antennal segment II being not more than one-third of length of the segment III, the anterolateral pronotal margins finely serratulate, and the ostiolar peritreme is not more than 1 mm in length (Ruckes 1966b).

Platycarenum umbractulatus (Fabricius, 1803) [Fig. 31]

Distribution. Panama, Colombia, Venezuela, Guyana, Surinam, French Guiana, Bahamas, Ecuador, Brazil, Peru, Argentina.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Ruckes (1966b), but without specific locality.

Remarks. *Platycarenum* is a monotypic genus.

References. Ruckes 1966b, Melo *et al.* 2017.

***Phoeacia* Stål, 1862**

The genus can be recognized by the apex of the scutellum reaching or projecting beyond an imaginary line connecting the anterolateral connexival angles of the last abdominal segment, and the antenna has four segments (Rolston 1990).

Phoeacia sp. [Fig. 32]

Distribution of the genus. Guatemala, Panama, Colombia, and Brazil.

Distribution in Colombia. Huila.

Remarks. The genus was recorded for Colombia by Castro-Huertas *et al.* (2015), who indicated the uncertainty of the identification at the species level.

References. Kirkaldy 1909; Rolston 1990, Castro-Huertas *et al.* 2015.

Tribe Ochlerini

***Coranda* Rolston, 1992**

The genus can be recognized by having the anterolateral angles of the pronotum acute, surpassing the posterior margin of the eyes in dorsal view; the anterolateral margins of the pronotum are reflexed; the labium reaching the last abdominal segment; the labial segment II reaching the metacoxae; the antecular region without a process; the clypeus is slightly longer than the mandibular plates; the antennae is five segmented; the antennal segment I surpassing the apex of the head; and the metasternum clearly carinate mesially on its entire length (Rolston 1992).

Coranda picipes (Stål, 1872) [Fig. 33]

Distribution. Colombia and Peru.

Distribution in Colombia. Cundinamarca.

Remarks. *Coranda picipes* can be distinguished from *C. castana* by the shape of the pygophore and its fuscous coloration.

References. Rolston 1992.

***Eritrachys* Ruckes, 1959**

The genus can be recognized by the shape of the head with pedunculate and prominent eyes exceeding the width of the anterior pronotal margin, which are separated from the pronotum by more than half of the diameter of one eye (Ruckes 1959a; Campos & Roell 2018).



FIGURES 32–47. Discocephalinae genera in Colombia. 32. *Phoeacia* sp. (MPUJ). 33. *Coranda picipes* (DAR). 34. *Eritrachys antennata* (MPUJ). 35. *Herrichella thoracica* (Holotype, BMNH). 36. *Lincus rufospilotus* (DAR). 37. *Macropygium reticulare* (MPUJ). 38. *Melambyrsus hoplita* (SMFD). 39. *Neoadoxoplatys longirostra* (holotype, AMNH). 40. *Ochlerus* sp. (ICN). 41. *Orbatina fuliginia* (DAR). 42. *Paralincus bimaculatus* (MPUJ). 43. *Phereclus pluto* (NHRS). 44. *Schraderiellus cinctus* (ICN). 45. *Stalius tartareus* (DAR). 46. *Stapecolis bimaculatus* (AMNH). 47. *Xynocoris recavus* (ICN). Scale bar: 5 mm.

Eritrachys antennata (Distant, 1911) [Fig. 34]

Distribution. Costa Rica, Panama, Colombia, and Ecuador.

Distribution in Colombia. Cauca, Risaralda, and Valle del Cauca.

Remarks. *Eritrachys antennata* was recently transferred from the genus *Phereclus* and it is considered a senior synonym of *E. bituberculata*. This species has the clypeus longer than the mandibular plates, and the pronotum not elevated.

References. Ruckes 1959a; Ortega-León & Thomas 2010; Castro-Huertas *et al.* 2015; Campos & Roell 2018.

Examined material. COLOMBIA: Risaralda, Pueblo Rico Área Amurrapá, aproximadamente 1.1 km WSW de Santa Cecilia, 05.33783° N, 76.15532° W, 402 m, sobre corteza de árbol, Bosque húmedo tropical, 19-23 ii 2018, L. Ángel / MPUJ_ENT0060591 (MPUJ_ENT); *idem*, V. Sánchez, sobre vegetación (MPUJ_ENT).

***Herrichella* Distant, 1911**

The genus can be recognized by the space between each eye and pronotum less than one-half the diameter of an eye, the antennae is five segmented, the antennal segment I surpassing slightly the apex of the head, the mandibular plates being as long as or longer than the clypeus, the anteocular region has a short and obtuse process, the labium is insert behind of the imaginary plane bisecting the head at the anterior margin of the eyes; and reaching the abdominal sternite III, the labial segment II reaching the anterior margin of the mesocoxae, and the metasternum is mesially carinate (Rolston 1992).

Herrichella thoracica Distant, 1911. [Fig. 35]

Distribution. Colombia.

Distribution in Colombia. “Dagua”, probably Valle del Cauca.

Remarks. *Herrichella thoracica* has the pronotum largely spotted with yellowish maculae not forming a transverse line between the humeral angles, the connexivum and the legs are brownish-black, the femora do not have elliptical brownish spots, and the labium does not surpass the abdominal sternite IV.

References. Rolston 1992; Campos & Roell 2018.

***Lincus* Stål, 1867**

The genus can be recognized by having the labium inserted behind the imaginary plane bisecting the head at the anterior margin of the of the e, and its apex reaching or surpass the penultimate abdominal sternite, the labium segment II surpassing slightly the mesocoxae, the antennae is five segmented, the antennal segment I reaching or surpass the apex of the head. The mandibular plates are as long as or longer than the clypeus, the corium surpassing the apex of the scutellum, and the metasternum flat or slightly tectiform, sometimes weakly carinate in part of its length (Rolston 1992).

Lincus lamelliger Breddin, 1908

Distribution. Colombia, French Guiana, and Surinam.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1983b), but without specific locality.

Remarks. *Lincus lamelliger* has the length of the pronotal lobes from the base of the incision to the apex of the lobe subequal to width of the lobes at the base.

References. Rolston 1983b, 1992.

Lincus punctatus (Dallas, 1851)

Distribution. Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia in the original description, and by Campos & Roell (2018) but without specific locality.

Remarks. *Lincus punctatus* can be distinguished by the large pronotal lobes; having the ventral rim of the pygo-

phore deeply and broadly emarginate; a large proctiger, well protruded beyond the posterior pygophoral margin, and the paramere not visible within the pygophore.

References. Campos & Roell 2018.

Lincus rufospilotus (Westwood, 1837) [Fig. 36]

Distribution. Colombia, Surinam, Guyana, Brazil, and Peru.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1983b), but without specific locality.

Remarks. *Lincus rufospilotus* has the pronotal lobe extending no further laterad of an eye than 0.2 times the width of an eye, apically narrowly round and often reflexed; the incision between the posterior margin of the lobes and the anterolateral pronotal margins does not reach as far mesad as the middle of the eyes, the antennal segments II and III are subequal in length, the segments IV and V are lighter in coloration than the basal three segments, neither of them annulated; and the mandibular plates are projecting beyond the clypeus.

References. Rolston 1983b.

Lincus styliger Breddin, 1908

Distribution. Colombia and Peru.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1983b), but without specific locality.

Remarks. *Lincus styliger* has the pronotal lobes elongated, projecting laterad from eyes by about half the width of an eye.

References. Rolston 1983b.

Lincus substyliger Rolston, 1983b

Distribution. Colombia.

Distribution in Colombia. Between Cundinamarca and Meta (Road Bogotá – Villavicencio).

Remarks. *Lincus substyliger* resembles *L. styliger* but can be differentiated by male genitalic characters.

References. Rolston 1983b.

Lincus subuliger Breddin, 1908

Distribution. Colombia and Venezuela.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1983b), but without specific locality.

Remarks. *Lincus subuliger* has the pronotal lobes extending laterad of the eyes by 0.2–0.4 times the width of an eye, are longer than wide at base, the mandibular plates surpass the apex of the clypeus, their lateral margins are subparallel anteriorly to the concavity above the antennifers; and the antennal segments I and II are subequal in length.

References. Rolston 1983b.

***Macropygium* Spinola, 1837**

The genus can be recognized by the oval body, brown or dark brown coloration with black irregular spots, the anterolateral margins of the pronotum are sometimes yellowish; the head is triangular; the mandibular plates are surpassing the apex of the clypeus; four or five antennal segments, the antepenultimate antennal segment wider in the males than the females; the pronotum is slightly sloping anteriorly to the humeri; the scutellum is shorter than the corium; and the apex of the labium is surpassing the urosternite III (Da Silva & Campos 2021).

Macropygium reticulare (Fabricius, 1803) [Fig. 37]

Distribution. Mexico, Guatemala, Costa Rica, Panama, Colombia, Venezuela, Brazil, Ecuador, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Nariño and Vaupés.

Remarks. *Macropygium reticulare* has the antennae five-segmented, the first antennal segment longer than the second; the conical projection of the ventral rim of the pygophore with apical setae; and the abdominal segment X oval, tapering toward the apex.

References. Rolston 1992; Campos & Grazia 2006; Da Silva & Campos 2021.

Macropygium spinolae Stål, 1860

Distribution. Mexico, Guatemala, Honduras, Nicaragua, Costa Rica, Panama, Colombia, Venezuela, Brazil, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Unknown. This species was recorded from Colombia in the taxonomic revision of the genus by Da Silva & Campos (2021), but without specific locality. The geographic coordinates recorded in the examined material suggest Vichada as a possible locality.

Remarks. *Macropygium spinolae* has the antennae five-segmented, the first and second antennal segments subequal in length; the segment X of pygophore is wider than long, the apex is rounded with abundant setae; the lateral angle of the laterotergite VIII is projected well beyond the line connecting the mesial trichobothria of the urosternite VII, and covering completely the base of the laterotergite IX.

References. Da Silva & Campos 2021.

***Melambyrsus* Breddin, 1912**

The genus can be recognized by having the triangular head and large scutellum, the head and the pronotum are of equal length, and the anterior margin of the bucculae is rounded (Campos & Roell 2017).

Melambyrsus hoplita Breddin, 1912 [Fig. 38]

Distribution. Colombia.

Distribution in Colombia. Cauca.

Remarks. *Melambyrsus* is a monotypic genus.

References. Campos & Roell 2017.

***Neadoxoplatys* Kormilev, 1956**

The genus can be recognized by the metasternum broadly and shallowly sulcate, without the median carina but sometimes tectiform; the inferior surface of the femora with one or two pairs of preapical spiniform processes; the superior surface of tarsal segment III of the hind legs is flattened in the females, and sometimes it is excavated, the abdominal basal segments flat, the labium is inserted before of the imaginary plane bisecting the head at the anterior margin of the eyes, and its apex reaching the abdominal sternite 5, the labial segment II reaching the mesocoxae, and it has an intercalary segment I the base; the antennae is five segmented, the mandibular plates are longer than or subequal to the clypeus, and the corium surpassing the apex of the scutellum (Rolston 1992).

Neadoxoplatys longirostra Ruckes, 1959 [Fig. 39]

Distribution. Colombia.

Distribution in Colombia. Córdoba (“Amaya Cispatá bay”).

Remarks. *Neadoxoplatys longirostra* has the mandibular plates and the clypeus subequal in length, the anterior lateral angles of the pronotum are lobulate, and the labium reach the abdominal sternite VI.

References. Rolston 1992; Cervantes-Peredo & Ortega-León 2014.

***Ochlerus* Spinola, 1837 [Fig. 40]**

The genus can be recognized by having the eyes prominent, the labium inserted before of the imaginary plane bisecting the head at anterior margin of the eyes; the apex of the labium reaching the abdominal sternites V, VI or VII,

the labial segment II reaching the mesocoxae, the antennae is five segmented, the antennal segment I surpassing the apex of the head, the mandibular plates and the clypeus are subequal in length; the mandibular plates are unarmed, its apex is subacute or obliquely truncate; the anterolateral angles of the pronotum have a small tubercle, usually directed laterad; the scutellum is not constricted near the posterior end of the frenum, and the metasternum has a thin and weak carina (Rolston 1992).

Ochlerus coriaceus Herrich-Schäffer, 1844

Distribution. Colombia, Trinidad and Tobago, and Venezuela.

Distribution in Colombia. Unknown. This species was recorded from Colombia in the original description, and by Simões & Campos (2014), but without specific locality.

Remarks. *Ochlerus coriaceus* has a small yellow spot at the base of the scutellum and paired smaller yellow spots close to the fovea; the legs and the entire ventral surface are yellow with brown spots; the anterolateral angles of the pronotum are slightly produced laterad, and the connexivum is partially emarginate, with yellow spots in the middle of each segment.

References. Rolston 1992; Simões & Campos 2014.

***Orbatina* Ruckes, 1961**

The genus can be recognized by having the humeral angles subacute, produced laterad of the hemelytra by a distance subequal to one-half of the width of an eye, the labium is inserted before of the imaginary plane bisecting the head at anterior margin of the eyes and reaching the abdominal sternite V, the labial segment II reaching the mesocoxae, the antennae is five segmented, the antennal segment I surpassing the apex of the head, the clypeus and mandibular plates subequal in length, and the elevated margin of each evaporative area is sigmoid (Rolston 1992).

Orbatina fuliginia Ruckes, 1961 [Fig. 41]

Distribution. Panama, Colombia, and Bolivia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1992), but without specific locality.

Remarks. *Orbatina* is a monotypic genus.

References. Rolston 1992.

***Paralincus* Distant, 1911**

The genus can be recognized by having the inferior surface of the pro- and mesofemur armed distally with small peg-like tubercles, these reduced or absent on the metafemur, and the space between the eye and the pronotum about 0.5 times of the diameter of an eye (Ruckes 1958; Rolston 1983c).

Paralincus bimaculatus (Ruckes, 1958) [Fig. 42]

Distribution. Colombia and Guyana.

Distribution in Colombia. Amazonas.

Remarks. *Paralincus bimaculatus* has the paramere laminate, and the antennal segments II, IV and V with equal length, longer than the antennal segment III.

References. Ruckes 1958; Rolston 1983c; Castro-Huertas *et al.* 2015.

***Phereclus* Stål, 1862**

The genus can be recognized by having the eye separated from the pronotum by a distance equal to 0.3 diameter of an eye; the labium inserted before an imaginary plane bisecting the head at anterior margin of the eyes; the labial segment II reaching the mesocoxae; the anterior margin of the pronotum moderately emarginated; the antennae is

five segmented; the antennal segment I surpassing the apex of the head; the antennal segment II is about one-half as long as III; and the clypeus and mandibular plates are subequal in length (Rolston 1992). The genus included three species until recently (Rolston 1992). Campos and Roell (2018) transferred *Phereclus antennatus* to *Eritrachys*, and *P. punctatus* to *Lincus*, thus, the genus *Phereclus* is currently monotypic.

Phereclus pluto Stål, 1862 [Fig. 43]

Distribution. Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1992), but without specific locality.

Remarks. The genus is monotypic.

References. Kirkaldy 1909; Rolston 1992; Campos & Roell 2018.

***Schraderiellus* Rider, 1998**

The genus can be recognized by having an anteapical tooth on the lateral margins of each mandibular plate; the ventral rim of the pygophore is not visible in dorsal view, and its base of the posterolateral angles has a pair of projections with tuft of setae, and the posterolateral angles of the pygophore have a basal dorsal excavation (Roell & Campos 2018).

Schraderiellus cinctus (Ruckes, 1959) [Fig. 44]

Distribution. Nicaragua, Costa Rica, Panama, and Colombia.

Distribution in Colombia. Valle del Cauca.

Remarks. *Schraderiellus cinctus* is dark brown with the anterolateral margin of the pronotum, the anterolateral margin of hemelytra, and the lateral margin of the connexivum orange or yellowish.

References. Ruckes 1959a; Castro-Huertas *et al.* 2015; Roell & Campos 2018.

Schraderiellus hughesae (Ruckes, 1959)

Distribution. Costa Rica, Panama, and Colombia.

Distribution in Colombia. Valle del Cauca.

Remarks. *Schraderiellus hughesae* is dark brown with small yellow spots adjacent to the cicatrices on the pronotum, on the scutellum and hemelytra, and the anterolateral margin of the pronotum and hemelytra are dark brown or yellowish.

References. Roell & Campos 2018.

Schraderiellus luteipedis Roell & Campos, 2018

Distribution. Colombia and Ecuador.

Distribution in Colombia. Cauca and Valle del Cauca.

Remarks. *Schraderiellus luteipedis* is dark brown with the connexivum not punctured, and the anterolateral margins of the pronotum are yellowish.

References. Roell & Campos 2018.

***Stalius* Rolston, 1992**

The genus can be recognized by having the anterolateral angles of the pronotum anteriorly projecting surpassing the posterior margin of the eyes, nearly reaching the middle of the eyes; the scutellum usually slightly constricted near the posterior margin of the frenum; and the metasternum flat, tectiform, or thinly and weakly carinate (Rolston 1992).

Stalius tartareus (Stål, 1862) [Fig. 45]

Distribution. Mexico, Panama, and Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1992), but without specific locality.

Remarks. *Stalius tartareus* has the metasternum with a slight carina, and the scutellum short with the apex attaining the penultimate abdominal segment.

References. Kirkaldy 1909; Rolston 1992.

***Stapecolis* Garbelotto & Campos, 2016**

The genus can be recognized by having the anterolateral margins of the pronotum crenulated, the dorsal rim of the pygophore with paired expansions laterad to the abdominal segment X, the apex of the foliaceous expansions of the inferior layer of the ventral rim with a small spinose carina; absence of the lobular expansions on the superior layer of the ventral rim, the subrectangular expansions below the ventral opening of the ventral rim, and the ventral opening of the pygophore is subrectangular (Garbelotto *et al.* 2016).

Stapecolis bimaculatus Garbelotto & Campos, 2016 [Fig. 46]

Distribution. Colombia.

Distribution in Colombia. Valle del Cauca.

Remarks. *Stapecolis bimaculatus* has a yellow spot on the apex of the radial vein of the corium with equal diameter to an eye, and the scutellum without spots on the fovea.

References. Garbelotto *et al.* 2016.

***Xynocoris* Garbelotto & Campos, 2014**

The genus can be recognized by having a long, projected, and acute anteocular process, and the apex of the posterolateral angles of the pygophore tumid (Garbelotto *et al.* 2014).

Xynocoris recavus Garbelotto & Campos, 2014 [Fig. 47]

Distribution. Colombia, Venezuela, and Brazil.

Distribution in Colombia. Meta.

Remarks. *Xynocoris recavus* has the labium reaching the posterior margin of abdominal sternite III, and the dorsal rim of the pygophore deeply concave.

References. Garbelotto *et al.* 2014; Castro-Huertas *et al.* 2015.

Subfamily Edessinae

***Brachystethus* Laporte, 1832**

The genus can be recognized by having a yellow callosity on the pronotal margin and corium, and the metasternal process is anteriorly truncated (Barcellos & Grazia 2003).

Brachystethus cribrus (Fabricius, 1781)

Distribution. Colombia, Venezuela, Guyana, Surinam, Brazil, Ecuador, Peru, and Bolivia.

Distribution in Colombia. Cundinamarca, Meta, and Valle del Cauca.

Remarks. *Brachystethus cribrus* has the abdominal sternites with longitudinal series of yellowish spots, the antennal segment IV is medially swollen, and the processes of the pygophore are conspicuously visible dorsally.

References. Barcellos & Grazia 2003; Castro-Huertas *et al.* 2015.

Examined material. COLOMBIA: 1♂, Valle del Cauca, Yotoco, 1600m, 21 x 2005, C. Solís / 22762 (MUSENUV).

Brachystethus improvisus Breddin, 1905 [Fig. 48]

Distribution. Guatemala, Costa Rica, Panama, Colombia, Brazil, and Bolivia.

Distribution in Colombia. Cundinamarca and Valle del Cauca.

Remarks. *Brachystethus improvisus* has the abdominal sternites uniformly brown to dark brown, the antennal segment IV is cylindrical, and the lobes of the dorsal rim of the pygophore are projected and well developed.

References. Barcellos & Grazia 2003.

Examined material. COLOMBIA: 2♀, **Valle del Cauca**, El Queremal, Antigua vía al Mar, 3°33'49.1" N, 76°45'31.1" W, 107.2 m, 8 ix 2007, Diana M. Montoya (MUSENUV); Reserva Forestal de Yotoco, 3°51'48"N, 76°23'9"W, 609 m, manual, 5 xii 2009, Marcela González (MUSENUV).

Brachystethus tricolor Bolívar, 1879

Distribution. Colombia, Ecuador, and Peru.

Distribution in Colombia. Huila and Quindío.

Remarks. *Brachystethus tricolor* has the dorsal surface of the body black, with red spots on the hemelytra, in variable extension.

References. Barcellos & Grazia 2003; Castro-Huertas *et al.* 2015.

***Doesburgedessa* Fernandes, 2010**

The genus can be recognized by having the metasternal process with a single conical anterior projection (Fernandes 2010).

Doesburgedessa elongatispina Fernandes, 2010 [Fig. 49]

Distribution. Colombia and Brazil.

Distribution in Colombia. Meta.

Remarks. *Doesburgedessa elongatispina* has the humeral angles strongly developed and laterally produced, the anterior projection of the metasternal process is long, the posterolateral angles of the pygophore are developed and rounded, and the parameres are straight.

References. Fernandes 2010; Castro-Huertas *et al.* 2015.

***Edessa* Fabricius, 1803**

This genus is very difficult to diagnose, but several species can be recognized by having the head dorsally flat, the margins rounded and barely emarginated, the first labial segment inserted in the buccula, and some species have a dark spot on the humeral angles equal to or longer than head width (Silva *et al.* 2017; Nunes *et al.* 2019).

Edessa antilope (Fabricius, 1798)

Distribution. Mexico, Dominican Republic, Nicaragua, Costa Rica, Panama, Colombia, Venezuela, Guyana, Surinam, and French Guiana.

Distribution in Colombia. Cundinamarca (?) ("Bogotá").

Remarks. *Edessa antilope* has the dorsal surface green mainly, the humeral angles are black, the corium is dark brown, the ventral surface is yellow mainly, the evaporatorium is dark brown, the ventral surface of the humeral angles is gently excavated, the scutellum is long, with the apex acute, the superior process of the genital cup is straight, the paramere is falcate, and the distal margin of the gonocoxite VIII is arcuate, projected over bases of the laterotergite IX.

References. Silva *et al.* 2017.

Edessa corallipes Erichson, 1848

Distribution. Colombia, Venezuela, Surinam, Guyana, French Guiana, Brazil, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Antioquia, Cundinamarca, and Meta.

Remarks. *Edessa corallipes* has the posterior margin of the corium oblique, and the apex of the paramere with two processes.

References. Silva *et al.* 2006.

Edessa elaphus Breddin, 1905

Distribution. Colombia, Venezuela, Surinam, Brazil, Ecuador, Peru, and Bolivia.

Distribution in Colombia. Putumayo.

Remarks. *Edessa elaphus* has the paramere wide with a narrow tip and superior processes, and the gonocoxite 8 with the mesial concavity wider.

References. Fernandes *et al.* 2000.

Edessa meditabunda (Fabricius, 1794)

Distribution. Cuba, Colombia, French Guiana, Trinidad and Tobago, Bolivia, Brazil, Paraguay, Uruguay, and Argentina.

Distribution in Colombia. Unknown. This species was recorded from Colombia by Posada-Ochoa (1989), but without specific locality.

Remarks. *Edessa meditabunda* has the body mostly green contrasting with brown corium (including yellowish veins of corium), the hemelytra are narrow, the abdominal dorsolateral tergites are green, broadly visible, the apex of the scutellum is subacute, the mesosternum is distinctly carinate, and the osteolar sulcus is long, terminating in a very short ridge.

References. Barber & Bruner 1932; Posada-Ochoa 1989.

Edessa ovina Dallas, 1851

Distribution. Trinidad and Tobago, Colombia, Guyana, Brazil, Paraguay, and Argentina.

Distribution in Colombia. Unknown. This species was recorded from Colombia in the “*Edessa ovina* group” study by Fernandes and Silva (2021), but without specific locality. The geographic coordinates recorded in the examined material suggest Cundinamarca as a possible locality.

Remarks. *Edessa ovina* has the dorsal surface green; the humeral angles with the apices rounded, tumid and black; the scutellum is short, the apex is not reaching the level of the apex of the corium; the ventral surface is dark yellow; the thorax is irregularly punctate; the peritreme is long; the superior process of the genital cup is claw-like; and the apex of the paramere is falcate.

References. Fernandes & Silva 2021.

Edessa rufomarginata (De Geer, 1773) [Fig. 50]

Distribution. Mexico to Argentina.

Distribution in Colombia. Amazonas, Boyacá, Caldas, Casanare, Chocó, Cundinamarca, Meta, Tolima, and Valle del Cauca.

Remarks. *Edessa rufomarginata* has the first labial segment longer than the bucculae.

References. Silva *et al.* 2006.

Edessa splendens Fernandes & Campos, 2011

Distribution. Colombia.

Distribution in Colombia. Antioquia and Valle del Cauca.

Remarks. *Edessa splendens* has the head without iridescent coloration; the scutellum and the ventral surface are as in the other species of the “*Edessa metallica* group”, but the diagnostic characters are mostly in the male and female genitalia.

References. Fernandes & Campos 2011.

Edessa virididorsata Silva, Fernandes & Grazia 2004

Distribution. Colombia, Venezuela, Guyana, Surinam, French Guiana, Brazil, Paraguay, and Argentina.

Distribution in Colombia. Magdalena.

Remarks. *Edessa virididorsata* has the processes of the pygophore reduced and tab-shape.

References. Silva *et al.* 2006.

***Grammedessa* Correia & Fernandes, 2016**

The genus can be recognized by having four black punctured longitudinal stripes on the dorsal surface of the head; the humeral angles are laterally projected and slightly flattened dorsoventrally; the metasternal process with narrow, long and apically acute branches of the anterior bifurcation, the abdominal trichobothria placed laterally, not aligned with the spiracles; the gonocoxites VIII and laterotergites VIII are punctured, and the gonocoxites VIII large, subequal in length to laterotergites IX (Correia & Fernandes 2016).

Grammedessa stillativentris (Breddin, 1905) [Fig. 51]

Distribution. Colombia and Venezuela.

Distribution in Colombia. Unknown. This species was recorded from Colombia by Correia & Fernandes (2016), but without specific locality.

Remarks. *Grammedessa stillativentris* has the dorsal surface of the body green with dark spots associated to punctures uniformly distributed, the ventral surface greenish yellow with spots associated with punctures brown to black, and the metasternal process is spotted only in the branches of the anterior bifurcation.

References. Correia & Fernandes 2016.

***Hypoxys* Amyot & Serville, 1843 [Fig. 52]**

The genus is very similar to *Edessa*, but *Hypoxys* has the antennal segment II longer than antennal segment III; the prothorax has the posterior angles acute but flattened and a little salient; and the abdomen is dorsally yellow, sometimes the lateral margin is black (Nunes *et al.* 2020).

Hypoxys quadridens (Fabricius, 1803)

Distribution. Colombia, Venezuela, Surinam, French Guiana, and Brazil.

Distribution in Colombia. Cundinamarca.

Remarks. *Hypoxys quadridens* has the pygophore with an expansion of the ventral rim acuminate and flat, the proctiger has a tuft of setae separated by a conspicuous tumid ridge, and the sutural margin of the gonocoxites 8 has V-shaped.

References. Nunes *et al.* 2020.

Hypoxys oxyacanthus (Breddin, 1904)

Distribution. Colombia, Surinam, French Guiana, Ecuador, Peru, Brazil, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Meta.

Remarks. *Hypoxys oxyacanthus* has the head five times wider than the humeral angle, and the abdomen ventrally has intersegmental dark stripes.

References. Nunes *et al.* 2020.

***Lopadusa* Stål, 1860**

The genus can be recognized by having the metasternal process not truncated, and the abdominal sternites pale with dark spots (Becker & Grazia 1970).

Lopadusa fuscopunctata (Distant, 1880) [Fig. 53]

Distribution. Panama, Colombia, Venezuela, Guyana, Brazil, Ecuador, Peru, and Bolivia.

Distribution in Colombia. Chocó and Valle del Cauca.

Remarks. *Lopadusa fuscopunctata* has the pronotum and the scutellum with dark brown spots, and the humeral angles rounded.

References. Becker & Grazia 1970; Castro-Huertas *et al.* 2015.

Examined material. COLOMBIA: 1 ♀, **Valle del Cauca**, Cali, 1000 m, 1 xii 1991 / 1951 (MUSENUV).



FIGURES 48–63. Edessinae and Pentatominae genera in Colombia. 48. *Brachystethus improvisus* (MPUJ). 49. *Doesburgedessa elongatispina* (MZUSP). 50. *Edessa rufomarginata* (MPUJ). 51. *Grammedessa stillativentris* (UFRG). 52. *Hypoxys* sp. (MZUSP). 53. *Lopadusa fuscopunctata* (ICN). 54. *Paraedessa subrectangulata* (ICN). 55. *Peromatus* sp. (MPUJ). 56. *Agroecus griseus* (ICN). 57. *Amauromelpia miri* (UFRG). 58. *Berecynthus hastator* (MPUJ). 59. *Cosmopepla cruciaria* (MPUJ). 60. *Diceraeus melacanthus* (MZUSP). 61. *Dichelops* sp. (MPUJ). 62. *Euschistus carbonerus* (MPUJ). 63. *Galedanta myops* (MPUJ). Scale bar: 5 mm.

***Paraedessa* Silva & Fernandes, 2013**

The genus is similar to *Edessa*, but the male of *Paraedessa* has the pygophore with a lateral expansion on the proctiger, and the female has the gonocoxite VIII reduced, and the gonapophyses VIII exposed and sclerotized (Silva & Fernandes 2013).

Paraedessa ecuadoriensis Silva & Fernandes 2013

Distribution. Colombia and Ecuador.

Distribution in Colombia. Meta.

Remarks. *Paraedessa ecuadoriensis* has an ogival genital cup process, apex of paramere bifid, lateral expansion of proctiger trapezoidal or cordiform; and gonocoxite VIII subtriangular and reduced.

References. Silva *et al.* 2013.

Paraedessa subretangulata Silva & Fernandes, 2013 [Fig. 54]

Distribution. Nicaragua, Costa Rica, Colombia, and Venezuela.

Distribution in Colombia. Tolima.

Remarks. *Paraedessa subretangulata* has a small genital cup process, the lateral expansion of the proctiger is subrectangular; and the posterolateral angle of the gonocoxite VIII is dentiform, reaching or surpassing the posterior margin of the gonapophyse VIII.

References. Silva *et al.* 2013.

Paraedessa verhoeffi (Breddin, 1904)

Distribution. Colombia, Venezuela, Brazil, and Peru.

Distribution in Colombia. Amazonas.

Remarks. *Paraedessa verhoeffi* has an elongated genital cup process, the paramere is triangular, and the dorsal face of the proctiger is very short, and nearly four times as wide as long.

References. Silva *et al.* 2013.

***Peromatus* Amyot & Serville, 1843**

The genus can be recognized by having the antennae with 4 segments, the first labial segment surpassing the bucculae, and the peritreme is short (Grazia *et al.* 2015).

Peromatus sp. [Fig. 55]

Distribution of the genus. Guatemala, Costa Rica, Panama, Colombia, Belize, Brazil, and Argentina.

Distribution in Colombia. Amazonas.

Remarks. The genus was recorded for Colombia by Castro-Huertas *et al.* (2015) and noted the impossibility of the identification to species level.

References. Kirkaldy 1909; Castro-Huertas *et al.* 2015.

Subfamily Pentatominae

Tribe Carpocorini

***Agroecus* Dallas, 1851**

The genus can be recognized by having the abdominal sternite III without a process (mesial tubercle or spine), the peritreme is short and auriculate, the femora have two parallel rows of short tubercles, the corium has several longitudinal, impunctate bands, the anterolateral margin of the pronotum is denticulate, and the basal plates are conspicuously convex in lateral view (Rider & Rolston 1987).

Agroecus griseus Dallas, 1851 [Fig. 56]

Distribution. South of Panama to northern Argentina.

Distribution in Colombia. Cesar and Valle del Cauca.

Remarks. *Agroecus griseus* has the anterolateral pronotal margin nearly straight.

References. Rider & Rolston 1987.

Examined material. COLOMBIA: 1♂, **Valle del Cauca**, Restrepo, Camp. Río Bravo, 900 m, 5 ii 1984, O. Cepeda / ICN_055451 (ICN).

***Amauromelpia* Fernandes & Grazia, 1998**

The genus is similar to *Hypatropis* and only can be distinguished by genital characters (Fernandes & Grazia 1998).

Amauromelpia miri Fernandes & Grazia, 1998 [Fig. 57]

Distribution. Colombia and Peru.

Distribution in Colombia. Amazonas.

Remarks. *Amauromelpia miri* has the anterolateral margins of the pronotum serrulate and slightly concave.

References. Fernandes & Grazia 1998.

***Berecynthus* Stål, 1862**

The genus can be recognized by having the mesosternum with a carina, the anterior margin of the clypeus is acute, and longer than the mandibular plates; and the phallotecal processes are long and parallel (Grazia & Hildebrand 1982).

Berecynthus hastator (Fabricius, 1798) [Fig. 58]

Distribution. Mexico, Costa Rica, Panama, Colombia, Venezuela, French Guiana, Surinam, Brazil, Peru, and Bolivia.

Distribution in Colombia. Amazonas, Caldas, Caquetá, Casanare, Cesar, Cundinamarca, Meta, Tolima, and Valle del Cauca.

Remarks. *Berecynthus* is a monotypic genus. Grazia & Hildebrand (1982) cited in the examined material six specimens deposited in AMNH (3 males and 3 females) from “Magdalena” but the specific localities (Valledupar, Curumani, and Becerril) correspond to the Cesar department.

References. Grazia & Hildebrand 1982; Silva *et al.* 2018; Dellapé 2021b.

Examined material. COLOMBIA: 1♀, **Caldas**, La Dorada, 23 iii 1970, C. Riveros / ICN_055825 (ICN); 1♀, **Caquetá**, Florencia, 26 i 1969, J. Norato / ICN_055778 (ICN); 1♂, **Casanare**, Orocué, 7 v 1974, M.L. Bueno / ICN_055779 (ICN); 4♂, 4♀, **Cundinamarca**, Fusagasugá, 10 vi 1974, J. Cogua / ICN_055780 (ICN); Guaduas, 3 xii 1972, G. Emira / ICN_055769 (ICN); Sasaima, 6 iii 1976, C. Córdoba / ICN_055758 (ICN); Tocaima, 30 vi 1972, Jimenez Ronetta / ICN_055759 (ICN); 24 x 1970, Rubby G.M. / ICN_055785 / ICN_055831 (ICN); Tolemaida, La Naranjala, 28 v 1968, G. Aguirre / ICN_055823 (ICN); Villeta, 799 m, 16 vi 1976, R.M. Gómez / ICN_055822 (ICN); 6♂, 8♀, **Meta**, Puerto López, Alto Menegua, 288 m, 10 iv 1984, E. Aya / ICN_055774 / ICN_055776 (ICN); Vereda Menegua, 280 m, 12 iv 1984, R. Restrepo / ICN_055757 / ICN_055782 (ICN); Vda. Menegua, 288 m, 17 iv 1984, R. Restrepo / ICN_055775 (ICN); 290 m, 1 iv 1984, M. Fergusson / ICN_055773 (ICN); 16 iv 1984, E. Cárdenas / ICN_055770 (ICN); P. Lasso / ICN_055772 (ICN); 17 iv 1984, L. Salazar / ICN_055768 (ICN); R. Pardo / ICN_055781 (ICN); Vereda la Balsa, Hda. Mozambique. 230 m, 29 vii 1983, A. Becerra / ICN_055783 (ICN); Villavicencio, 11 x 1970, C. Princo / ICN_055824 (ICN); 27 xi 1970, P. Rodríguez / ICN_055784 (ICN); Km 8 entre Villavicencio y Restrepo, 510 m, 11 ix 1976, N. Pinzón / ICN_055777 (ICN); 1♂, **Tolima**, Nilo, 22 vii 1972, G. Hurtado / ICN_055767 (ICN).

***Cosmoepepla* Stål, 1867**

The genus can be recognized by the oval body, entirely black with yellow or orange markings, or reddish spots; the clypeus and the mandibular plates equal in length, the labium reaching the metasternum, and the orifice of the peritreme elevated as a tubercle (McDonald 1986).

Cosmoepepla cruciaria Stål, 1867 [Fig. 59]

Distribution. Colombia and Ecuador.

Distribution in Colombia. Cundinamarca (?) (“Bogotá”).

Remarks. *Cosmoepepla cruciaria* has the pronotum with a conspicuous cross on the disc.

References. McDonald 1986.

Cosmoepepla coeruleata Montandón, 1893

Distribution. USA, Mexico, El Salvador, Costa Rica, Colombia, and Venezuela.

Distribution in Colombia. Cundinamarca.

Remarks. *Cosmoepepla coeruleata* has the frenum nearly 0.3 times the length of the scutellum, the body is broadly oval, and the apex of the paramere has a thumb-like process.

References. McDonald 1986.

***Diceraeus* Dallas, 1851**

The genus can be recognized by having body dorsally brown, the humeral angles are concolorous with the body or black, the mandibular plates are longer than clypeus, the anterolateral margins of the pronotum are serrated or crenulated, the posterolateral margins of the pronotum are crenulated, the humeral angles are produced as a spine, and the scutellum is apically calloused (Barão *et al.* 2020).

Diceraeus melacanthus (Dallas, 1851) [Fig. 60]

Distribution. Colombia, Venezuela, Peru, Brazil, Bolivia, Paraguay, Uruguay, and Argentina.

Distribution in Colombia. Caquetá and Cundinamarca.

Remarks. *Diceraeus melacanthus* has the mandibular plates acute, the ventral margin of the pygophore is medially bisinuate, and the gonocoxite VIII is flat, not inflated.

References. Grazia 1978; Barão *et al.* 2020

***Dichelops* Spinola, 1837 [Fig. 61]**

The genus can be recognized by having the mandibular plates outlined in black, and longer than the clypeus; the apex of the scutellum is smooth; the posterolateral angles of the pygophore have bristle tufts, the gonocoxite VIII is swollen along sutural margin, the posterior margin of gonocoxites VIII is concave; and the laterotergites VIII are angulate (Barão *et al.* 2020).

Dichelops divisus (Walker, 1867)

Distribution. Colombia and Brazil.

Distribution in Colombia. Amazonas.

Remarks. *Dichelops divisus* has the humeral angles with cylindrical processes, laterally projected and without spots.

References. Grazia 1978; Castro-Huertas *et al.* 2015.

Dichelops nigrum Bergroth, 1914

Distribution. Colombia, Venezuela, and Brazil.

Distribution in Colombia. Meta.

Remarks. *Dichelops nigrum* has the humeral angles with a conspicuous acute process, laterally projected; the posterolateral margin of the connexivum is slightly projected, and the medial region of each segment of the connexivum with several dark brown spots.

References. Grazia 1978; Castro-Huertas *et al.* 2015.

Euschistus Dallas, 1851

The genus can be recognized by having the metasternum not produced, the peritreme reaching less than 0.4 times the distance from the ostiole to the lateral margin of the segment, the femora is unarmed, the eyes are contiguous to pronotum, the superior face of the tibiae is sulcate, the mandibular plates and the clypeus or both are apically rounded and usually subequal in length, the bucculae is evanescent or truncate at base of the head, and the antennae with five segments (Cioato *et al.* 2015).

Euschistus agudus Rolston, 1974

Distribution. Panama, Colombia, Venezuela, and Trinidad and Tobago.

Distribution in Colombia. Cundinamarca (?) (“Bogotá”).

Remarks. *Euschistus agudus* has the humeral angles conspicuously projected upward, the mandibular plates slightly surpass the clypeus, and the dorsal surface does not have calloused spots.

References. Rolston 1974.

Euschistus atrox (Westwood, 1837)

Distribution. Nicaragua, Panama, Colombia, and Guyana.

Distribution in Colombia. Bolívar, Chocó, Cundinamarca, Meta, and Tolima.

Remarks. *Euschistus atrox* has the anterior disk of the pronotum inclined in lateral view, and the posterior margin of the pygophore is trisinate.

References. Kirkaldy 1909; Rolston 1984; Maes 1994.

Examined material. **COLOMBIA:** 1♂, **Bolívar**, Zambrano, Hda. Monterrey, 9°37'48" N, 74°54'44" W, 70 m, Malaise, 24 xii 1996, F. Fernández & G. Ulloa / IAvH-E_05077 (IAvH-E); 1♂, 1♀, **Chocó**, Riosucio, Cacarica, 10 viii 1978, H.E.E. (IAvH-E), 3 viii 1978, H.E.E. (IAvH-E); 1♂, 1♀, **Cundinamarca**, Medina, Mesacura, 20 x 1989, G. Bernal (IAvH-E); Nilo, 29 xi 1972, M.C. Isaza (IAvH-E); 1♂, 1♀, **Meta**, Río Guayuriba, 450 m, 1 xi 1944, L. Richter (IAvH-E); Puerto López, Vereda Menegua, 300 m, 13 iv 1984, R. Restrepo (IAvH-E); 2♀, **Tolima**, Melgar, Club de suboficiales, 4 i 1969, I. de Arévalo (IAvH-E); Valencia, 8 i 1993, J. Villanueva (IAvH-E).

Euschistus bifibulus (Palisot de Beauvois, 1805)

Distribution. USA, Mexico, Cuba, Dominican Republic, Jamaica, Guatemala, Honduras, Nicaragua, Costa Rica, Panama, and Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Kirkaldy (1909), but without specific locality.

Remarks. *Euschistus bifibulus* has the posterior margin of the pygophore smoothly concave.

References. Kirkaldy 1909; Rolston 1984; Maes 1994.

Euschistus carbonerus Rolston, 1984 [Fig. 62]

Distribution. Colombia and Brazil.

Distribution in Colombia. Valle del Cauca.

Remarks. *Euschistus carbonerus* has the two last antennal segments dark brown with a basal pale brown transversal stripe, the evaporatorial area is unicolored, and the pronotal denticles are black.

References. Kirkaldy 1909; Rolston 1984; Maes 1994, Castro-Huertas *et al.* 2015.

Euschistus crenator (Fabricius, 1794)

Distribution. USA, Mexico, Jamaica, Hispaniola, Cuba, St. Vincent and the Grenadines, Guatemala, Costa Rica, Honduras, Panama, Colombia, Venezuela, Trinidad and Tobago, and Brazil.

Distribution in Colombia. Antioquia, Bolívar, Chocó, Cundinamarca, Meta, Quindío, Risaralda, Tolima, and Valle del Cauca.

Remarks. *Euschistus crenator* has the pronotal denticles with punctures crowded on the margin.

References. Kirkaldy 1909; Rolston 1984; Maes 1994.

Examined material. COLOMBIA: **Antioquia**, San Luis, Reserva Natural Cañon del Río Claro, 440m, 1 iii 1994, A. Quintero / MPUJ_ENT 0010756 / MPUJ_ENT 0010757 (MPUJ_ENT); 1♀, **Bolívar**, Zambrano, Hda. Monterrey, 9°37'48" N, 74°54'44" W; 70 m, 15 xii 1993, F. Fernández & G. Ulloa (IAvH-E); 1♂, **Chocó**, Riosucio, Saucataá, 26 vii 1978, H. Echeverri (IAvH-E); 2♂, 1♀, **Cundinamarca**, Cáqueza, Vda. Ubatoque, 1746 m, 14 viii 1989, L.P. Moreno (IAvH-E); 2 x 1983, G. Andrade (IAvH-E); Villeta, 22 vii 1972, G. Hurtado (IAvH-E); 1♂, **Meta**, Puerto Limón, 300 m, 25 iii 1983, Camilo C. (IAvH-E); 1♀, **Quindío**, Salento, 1 viii 1990, G. Medina & C. Rodríguez (IAvH-E); **Risaralda**, Pereira, S.F.F. Otún Quimbaya, Est. La Suiza, 1995 m, 4.732977° N, 75.589199° W, Gui-Ger, viii 1992 / MPUJ_ENT 0010758 (MPUJ_ENT); 1♀, **Tolima**, Espinal, 29 xii 1980, O. Jimenez (IAvH-E); Melgar, Centro Cafam, 450m, Eraso *et al.* / MPUJ_ENT 0010755 (MPUJ_ENT); 1♀, **Valle del Cauca**, Restrepo, Camp. Madroñal, 1300 m, 5 ii 1984, J. Adama (IAvH-E); Tuluá, Mateguadua, Jardín Botánico Juan María Céspedes, 1100m, 04.02913° N, 76.166981° W, 24-31 viii 1996, P. Lattig / MPUJ_ENT 0010753 (MPUJ_ENT); E. Amat / MPUJ_ENT 0010754 (MPUJ_ENT).

Euschistus emoorei Rolston, 1972

Distribution. Mexico to Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1974), but without specific locality.

Remarks. *Euschistus emoorei* has the posterior margin of the pygophore with a broad, sinuous to subrectangular emargination in ventral view.

References. Rolston 1974.

Euschistus rohus Rolston, 1984

Distribution. Panama, Colombia, and Ecuador.

Distribution in Colombia. Valle del Cauca.

Remarks. *Euschistus rohus* has the medial protuberance on the posterior margin of the pygophore minutely notched in caudal view.

References. Kirkaldy 1909; Rolston 1984; Maes 1994.

Euschistus rufimanus Stål, 1872

Distribution. Colombia.

Distribution in Colombia. Meta.

Remarks. *Euschistus rufimanus* has the apex of the clypeus usually acute, and the pronotal denticles widely spaced.

References. Kirkaldy 1909; Rolston 1984; Maes 1994; Agudelo *et al.* 2008.

Galedanta Amyot & Serville, 1843

The genus can be recognized by having body yellowish, reddish to brown, entirely covered by black punctures; the clypeus is shorter than the mandibular plates, and they are contiguous; the antennal segments II and III are nearly equal length, and the anterolateral margins of the pronotum dentated (Grazia 1967).

Galedanta myops (Fabricius, 1803) [Fig. 63]

Distribution. Costa Rica, Colombia, Guyana, and Brazil.

Distribution in Colombia. Tolima.

Remarks. *Galedanta myops* has the basal angles of the scutellum conspicuously punctured.

References. Kirkaldy 1909; Grazia 1967, 1981.

Examined material. COLOMBIA: 1♀, Tolima, Bosque Municipal Mariquita, 620 m, 16 iv 2000, Álvarez, M.B. leg (MPUJ_ENT).

***Hypatropis* Fernandes & Grazia, 1996**

The genus can be recognized by having the first labial segment not surpassing the bucculae, the mandibular plates longer or as long as the clypeus, the external margins of the mandibular plates above the level of the clypeus in lateral view, and the paramere is absent (Fernandes & Grazia 1996; Barros *et al.* 2021).

Hypatropis rolstoni Fernandes & Grazia, 1996 [Fig. 64]

Distribution. Honduras, Nicaragua, Costa Rica, Panama, Colombia, and Venezuela.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Fernandes & Grazia (1996), but without specific locality.

Remarks. *Hypatropis rolstoni* is recognized by internal genital characters.

References. Fernandes & Grazia 1996.

***Lattinidea* Rider & Eger, 2008**

The genus can be recognized by having the labium surpassing the abdominal sternite III, and the posterior margin of the pygophore broadly and deeply emarginated in ventral view (Rider & Eger 2008).

Lattinidea geografica (Fabricius, 1803) [Fig. 65]

Distribution. Costa Rica, Panama, Colombia, Venezuela, French Guiana, Ecuador, Brazil, and Peru.

Distribution in Colombia. Amazonas.

Remarks. *Lattinidea* is a monotypic genus.

References. Rider & Eger 2008.

***Lattinellica* Rider & Eger, 2008**

The genus can be recognized by having the dorsal surface of the pro, meso- and metatibia sulcate (distally at least), the pronotum, scutellum and corium usually with metallic blue or green to black coloration pattern, and the head is yellowish (Rider & Eger 2008).

Lattinellica decora (Walker, 1867) [Fig. 66]

Distribution. Colombia, Ecuador, Brazil, and Peru.

Distribution in Colombia. Amazonas.

Remarks. *Lattinellica* is a monotypic genus.

References. Rider & Eger 2008.

***Mitripus* Rolston, 1978**

The genus can be recognized by having the profemur unarmed, the labium not surpassing the metacoxa, the mandibular plates usually as long as the clypeus, and the apex of the radial vein with a pale brown spot (Bianchi *et al.* 2017).

Mitripus acutus (Dallas, 1851) [Fig. 67]

Distribution. Trinidad and Tobago, Peru, Brazil, Bolivia, and Argentina.

Distribution in Colombia. Amazonas, Boyacá, Guaviare, and Meta.

Remarks. *Mitripus acutus* has the humeral angles acute, and punctuation on the scutellum not conspicuously dense.

References. Rolston 1978a; Grazia 1987; Bianchi *et al.* 2017; Dellapé 2021a.

Examined material. COLOMBIA: 2♀, Amazonas, Leticia, Comunidad Monifue Amena, 70 m, [4.14166 S–

69.923256 W], 30 iii 2005, G. / MPUJ_ENT 0010872 (MPUJ_ENT); 2 x 2003, varzea (MPUJ_ENT); 5♂, 3♀, **Boyacá**, Santa María, sector de La Almenara, ~1.7 km NNE de Santa María, 4.87486 N, 73.25508 W, 1.123 m, 13-17 iii 2016, P. Erazo *et al.* / Borde de bosque, sobre vegetación, manual / MPUJ_ENT 0051146 / MPUJ_ENT 0051122 / MPUJ_ENT 0051136 / MPUJ_ENT 0051120 / MPUJ_ENT 0050045 (MPUJ_ENT); J. Benavides *et al.* / MPUJ_ENT 0052474 (MPUJ_ENT); Sendero Hycá Quye, ~5.5 km NW de Santa María, 4.89811 N, 73.29344 W, 900 m, 15-19 viii 2016, J. Peña / Borde de bosque, margen de camino, sobre vegetación baja, red entomológica / MPUJ_ENT 0047082 (MPUJ_ENT); Cadena, Suaza & Zapata / MPUJ_ENT 0047273 (MPUJ_ENT); 3♀, **Guaviare**, San José, Margen del río Platanal, 120 m, J.M. Ardila, malaise, x 2003 (MPUJ_ENT); Puerto Colombia, 170 m, malaise, 12 viii 2005, J. Ardila (MPUJ_ENT); 1♂, 1♀, **Meta**, San Martín, Reserva Natural El Caduceo, km 4.5 vía San Francisco, 3.671389 N, 73.659444 W, 30 ix-5 x 2013, P. Duarte / Trampa de luz, pastizal (MPUJ_ENT); Villavicencio, Bosque Bavaria, 600 m, 8 iii 2006, M. López (MPUJ_ENT); La Libertad, 366 m, Est. Int (MPUJ_ENT); G. E. La Libertad, 326 m, manual (MPUJ_ENT).

***Mormidea* Amyot & Serville, 1843**

The genus can be recognized by having the antennal segment I not surpassing the anterior margin of the head, the antennal segments II to V longer than the antennal segment I, the clypeus is slightly longer than the mandibular plates, the first labial segment surpassing the bucculae, and the posterior angles of the pronotum are unarmed (Rolston 1978b).

Mormidea bridarolli Pirán, 1963

Distribution. Colombia, Ecuador, Peru, and Bolivia.

Distribution in Colombia. Putumayo.

Remarks. *Mormidea bridarolli* has the tibiae with several small black spots.

References. Rolston 1978b.

Mormidea bovilla (Distant, 1887)

Distribution. Colombia, Ecuador, Surinam, and Brazil.

Distribution in Colombia. Meta.

Remarks. *Mormidea bovilla* has the humeral angles laterally produced, and usually with a subapical tubercle on the posterior margin.

References. Rolston 1978b; Castro-Huertas *et al.* 2015.

Mormidea collaris Dallas, 1851

Distribution. Mexico, Panama, and Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1978b), but without specific locality.

Remarks. *Mormidea collaris* has a tiny mark dark on the posterior margin of the supracoxal clefts.

References. Rolston 1978b.

Mormidea cubrosa (Dallas, 1851)

Distribution. USA, Antillas, West Indies, and Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1978b), but without specific locality.

Remarks. *Mormidea cubrosa* has the abdominal sternites entirely dark brown except on the lateral margins, and a sub-spiracular pale brown spot on each side of some or all abdominal sternites.

References. Rolston 1978b.

Mormidea fusca Stål, 1872

Distribution. Colombia and Venezuela.

Distribution in Colombia. Antioquia, Chocó, Cundinamarca, Magdalena, Meta, and Quindío.

Remarks. *Mormidea fusca* has the anterolateral margins of the pronotum fuscus or black, and the pygophore has the superior ridge with a deep mesial notch.

References. Rolston 1978b.

Examined material. COLOMBIA: 1♀, **Meta**, Restrepo, 4 viii 1979, J.L. Pinto (ICN); **Quindío**, Filandia, Estación Bremen (C.R.Q), 1900 m, iv 1998 / MPUJ_ENT 0010732 (MPUJ_ENT).

Mormidea maculata Dallas, 1851

Distribution. Colombia, Brazil, Peru, Bolivia, Argentina.

Distribution in Colombia. Amazonas, Chocó, Cundinamarca, and Meta.

Remarks. *Mormidea maculata* has the dorsal margin of the pygophore deeply emarginated in caudal view, and the lateral margins of the abdominal sternite II in the female has brown punctuation weaker than the adjacent region of the metathorax.

References. Rolston 1978b; Dellapé *et al.* 2015.

Examined material. COLOMBIA: 1♂, 3♀, **Amazonas**, Leticia, Vda. San Agustín, w.a. Los Lagos, 22 iii 1992, J. Parada & A. Sáenz / ICN_076935 (ICN); Vda. San José, 5 v 1992, J. Parada / ICN_076931 (ICN); Puerto Nariño, Vda. Boyavazú, 21 v 1992, J. Parada & A. Sáenz / ICN_076934 (ICN); J. Parada / ICN_076930 (ICN); 1♀, **Chocó**, Riosucio, Sautatá, 26 vi 1976, H. Echeverri / ICN_076917 (ICN); 1♀, **Cundinamarca**, Medina, Vda. Choopal, 540 m, 29 vii 1986, Sist. Avanz. / ICN_076920 (ICN); 1♂, 1♀, **Meta**, Macarena, Los Micos, 28 vii 1971, M. Smith Robles / ICN_076915 (ICN); Puerto Limón, 300 m, 24 iii 1983, Camilo C. / ICN_076918 (ICN).

Mormidea metallica Stål, 1872

Distribution. Panama and Colombia.

Distribution in Colombia. “Bogotá”, probably Cundinamarca.

Remarks. *Mormidea metallica* has the abdominal sternite IV usually without a medial pale spot (*mesial vitta*).

References. Rolston 1978b.

Mormidea notulata (Herrich-Schäffer, 1844)

Distribution. Mexico, Colombia, and Peru.

Distribution in Colombia. Magdalena, Quindío, and Risaralda.

Remarks. *Mormidea notulata* has the lateral margins of the connexivum entirely pale brown, and the abdomen with a medial pale spot (*mesial vitta*).

References. Rolston 1978b.

Examined material. COLOMBIA: **Quindío**, Filandia, Estación Bremen(C.R.Q), 1900 m, 14-20 iv 1998, manual, J. Bastidas / MPUJ_ENT 0010730 (MPUJ_ENT); **Risaralda**, Pereira, S.F.F. Otún Quimbaya, Est. La Suiza, 1850 m, 4.732977° N, 75.589199° W, C. Aristizabal, 25 iv-4 v 1997 / MPUJ_ENT 0010727 / MPUJ_ENT 0010728 / MPUJ_ENT 0010729 (MPUJ_ENT).

Mormidea pama Rolston, 1978

Distribution. USA, Jamaica, Cuba, Mexico, and Colombia.

Distribution in Colombia. Antioquia and Meta.

Remarks. *Mormidea pama* has the posterior margin of the pygophore with a conspicuous medial notch in ventral view.

References. Rolston 1978b.

Mormidea pictiventris Stål, 1862

Distribution. USA, Mexico, Colombia, and Venezuela.

Distribution in Colombia. Magdalena and Meta.

Remarks. *Mormidea pictiventris* has the posterodorsal margin of the pygophore with a small sharp tooth on the beginning of the deflexed carina.

References. Rolston 1978b.

Examined material. COLOMBIA, Meta, Puerto Gaitán, Hacienda Yamato, 105 m, 4.524944° N, 71.811889° W, 2-9 xi 1996, D. Forero / MPUJ_ENT 0010726 (MPUJ_ENT); Villavicencio, La Libertad. I.C.A. Palma Africana, 8 i 1983, R. Restrepo / ICN_076923 (ICN).

Mormidea triangularis (Walker, 1867)

Distribution. Colombia, Ecuador, and Peru.

Distribution in Colombia. Meta and Nariño.

Remarks. *Mormidea triangularis* has the body entirely black; the lateral margin of the mandibular plates, the anterolateral and posterolateral margins of the pronotum, the lateral margins of the scutellum, and the costal margin of the coria are yellowish; the femur and tibia are yellowish with black bands. Rolston (1985) described *Mormidea kirkaldyi* based in specimens identified as *Mormidea montandoni* Kirkaldy in Rolston (1978). Thomas (1990) synonymized *M. kirkaldyi* with *M. triangularis*.

References. Kirkaldy 1909; Rolston 1985; Thomas 1990; Castro-Huertas *et al.* 2015.

Mormidea ypsilon (Linnaeus, 1758) [Fig. 68]

Distribution. Mexico to northern Argentina.

Distribution in Colombia. Amazonas, Meta, and Nariño.

Remarks. *Mormidea ypsilon* has a pale brown Y-shaped mark on the scutellum, which is calloused in the basal angles and the posterior margin of the frenum.

References. Rolston 1978b.

Examined material. COLOMBIA: 2♂, 2♀, **Amazonas**, Leticia, Km 2 w.a. Tarapacá, J. Parada & A. Sáenz / ICN_076932 (ICN); Km 1 w.a. Tarapacá, 10 vi 1992, J. Parada & A. Sáenz / ICN_076928 (ICN); Puerto Nariño, Vda. Naranjales, 22 v 1992, J. Parada / ICN_076929 (ICN); Vda. Boyavazú, 21 v 1992, A. Sáenz / ICN_076933 (ICN); 1♂, 4♀, **Meta**, San Martín, R. El Caduceo, 3°39'54.9" N, 73°39'23.9" W, 393 m, 7 xii 2006, F. Fandiño (ICN); Villavicencio, Guanaviche Estadero, río Guatiquía, 4°10,506' N, 73°38,233' W, 1465 ft, 3-5 vii 2013, J.E. Eger & A.A. Calixto (ICN); 2♀, **Nariño**, Tumaco, Vda. Imbili, margen izq. Río Mira, 14 vii 1984, A. Guerra / ICN_076921 / ICN_076919 (ICN).

Oebalus Stål, 1862

The genus can be recognized by having the mandibular plates not surpassing the clypeus, first labial segment between the bucculae, the proctiger is wedge-shaped, and its lateral margins are sloping from the median line, and the apex of the subgenital plates in the females are slightly acuminate (Sailer 1944).

Oebalus insularis Stål, 1872

Distribution. USA, Mexico, Cuba, Haiti, Puerto Rico, Honduras, Panama, and Colombia.

Distribution in Colombia. Boyacá, Caldas, Chocó, Cundinamarca, Santander, and Tolima.

Remarks. *Oebalus insularis* has the genital structure similar to *O. poecilus*, but *O. insularis* can be recognized by a small tooth on the inner rim of the lateral angle of the pygophore; and the paramere is regularly concave in lateral view, and its apex is linear in dorsal view.

References. Sailer 1944.

Examined material. COLOMBIA: 1♂, 1♀, **Boyacá**, Chiquinquirá, 19 ii 1970, L.M. Caballero / ICN_077062 (ICN); Sogamoso, 29 xi 1970, L.M. Caballero / ICN_076953 (ICN); 1♂, 1♀, **Caldas**, La Dorada, 23 iii 1970, C. Riveros / ICN_077050 / ICN_076992 (ICN); 1♂, 1♀, **Chocó**, Golfo de Tribugá, cultivo de arroz, 1 iv 1998 / IAvH-E_151847 / IAvH-E 151848 (IAvH-E); 16♂, 10♀ **Cundinamarca**, Guaduas, 3 xii 1972, Emira G. / ICN_077041 (ICN); La Capilla, 24 x 1970, H.A. Palma / ICN_077068 (ICN); La Mesa, maíz, 22 v 1982, A. Murillo / ICN_077031 / ICN_077038 (ICN); Mesitas, suelo, 26 v 1982, A. Murillo / ICN_076995 (ICN); Sasaima, 1 x 1965, N. Becerra / ICN_076954 (ICN); Villeta, 11 vii 1972, N. Bastidas / ICN_077039 (ICN); Tocaima, 30 vi 1972, Jimenez Pianetta / ICN_077004 (ICN); Tocaima, 30 vi 1972, J.A. Rodríguez / ICN_077151 (ICN); Tolemaida, La Naranjala, 19 v 1968, J.M. Rincón / ICN_077122 (ICN); 1♂, **Santander**, Vélez, 17 xi 1970, H.A. Palma / ICN_077140 (ICN); 4♂, 2♀, **Tolima**, Armero, maíz, 12 xi 1972 / ICN_077125 (ICN); Alvarado, Cultivo de arroz, 28 iii 1974, G. Márquez / ICN_076955 (ICN); Melgar, 29 xi 1970, JVR-JEMS / ICN_077063 / ICN_077046 / ICN_076975 / ICN_056007 / ICN_056011 / ICN_056012 / ICN_077152 / ICN_077064 / ICN_077127 / ICN_077136 / ICN_077072 / ICN_077070 / ICN_077073 (ICN); Melgar, 29 ii 1970, L.M. Caballero / ICN_077044 / ICN_077137 / ICN_077128 (ICN); Miravalle, 18 iv 1970, G. Guillot / ICN_076983 (ICN); Miravalle, 15 iii 1970, G. Guillot / ICN_077069 (ICN); Prado, 6 km de Prado, Chanchito, 26 iii 1986 (ICN).

Oebalus ornatus Stål, 1872

Distribution. Puerto Rico, Hispaniola, Colombia, and Brazil.

Distribution in Colombia. Valle del Cauca.

Remarks. *Oebalus ornatus* has the apical external projection of the paramere (ectal arm) flattened at right angles to the apical margin. This species was recorded from Colombia by Daza (1991), Pantoja *et al.* (1995, 2005), and Torres (2004) but we have not found specimens of this species in the entomology collections visited.

References. Sailer 1944, 1957; Daza 1991; Pantoja *et al.* 1995, 2005; Torres 2004.

Oebalus poecilus (Dallas, 1851)

Distribution. Colombia, Trinidad and Tobago, Guyana, French Guiana, Brazil, Ecuador, Bolivia, Paraguay, Uruguay, and Argentina.

Distribution in Colombia. Vichada.

Remarks. *Oebalus poecilus* has the apex of the paramere not linear in dorsal view, and not regularly concave in lateral view; the apical external projection of the paramere (ectal arm) slightly bent upward.

References. Sailer 1944.

Examined material. COLOMBIA: 1♂, 1♀, **Vichada**, Amanavén, 1 v 1952, L. Richter, ICN_077154 / ICN_077067 (ICN).

Oebalus pugnax (Fabricius, 1775)

Distribution. USA, Mexico, Nicaragua, Honduras, West Indies, Panama, and Colombia.

Distribution in Colombia. Antioquia, Caldas, Cundinamarca, and Tolima.

Remarks. *Oebalus pugnax* has the antennal segment II longer than the antennal segment I, the scutellum is longer than the basal width, the humeral angles are acute and anteriorly projected; and the lateral margins of the pronotum on the anterior region to the humeral angles are calloused.

References. Maes 1994; Froeschner 1999; Thomas 2000; Arismendi & Thomas 2003; Perez-Gelabert & Thomas 2005; Castro-Huertas *et al.* 2015.

Oebalus ypsilongriseus (DeGeer, 1773) [Fig. 69]

Distribution. Colombia, Venezuela, Guyana, Surinam, Brazil, Peru, Paraguay, Uruguay, and Argentina.

Distribution in Colombia. Amazonas, Bolívar, Boyacá, Caquetá, Cundinamarca, Meta, and Tolima.

Remarks. *Oebalus ypsilongriseus* has the humeral angles with an acute process, the antennae with four segments, the antennal segment II is shorter than the antennal segment III, and both antennal segments are fused.

References. Sailer 1944, 1957; Becker & Grazia 1971; Castro-Huertas *et al.* 2015.

***Padaeus* Stål, 1862 [Fig. 70]**

The genus can be recognized by having body oval and convex ventrally, black or dark brown coloration, the head is triangular, the clypeus is rounded and slightly surpassing the mandibular plates, the posterior region of the frenum is less or equal width to the half to the basal region width of the scutellum; and the anterolateral margins of the pronotum rounded, with its surface striated or with sparse denticles (Brailovsky 1987; Torres 2004).

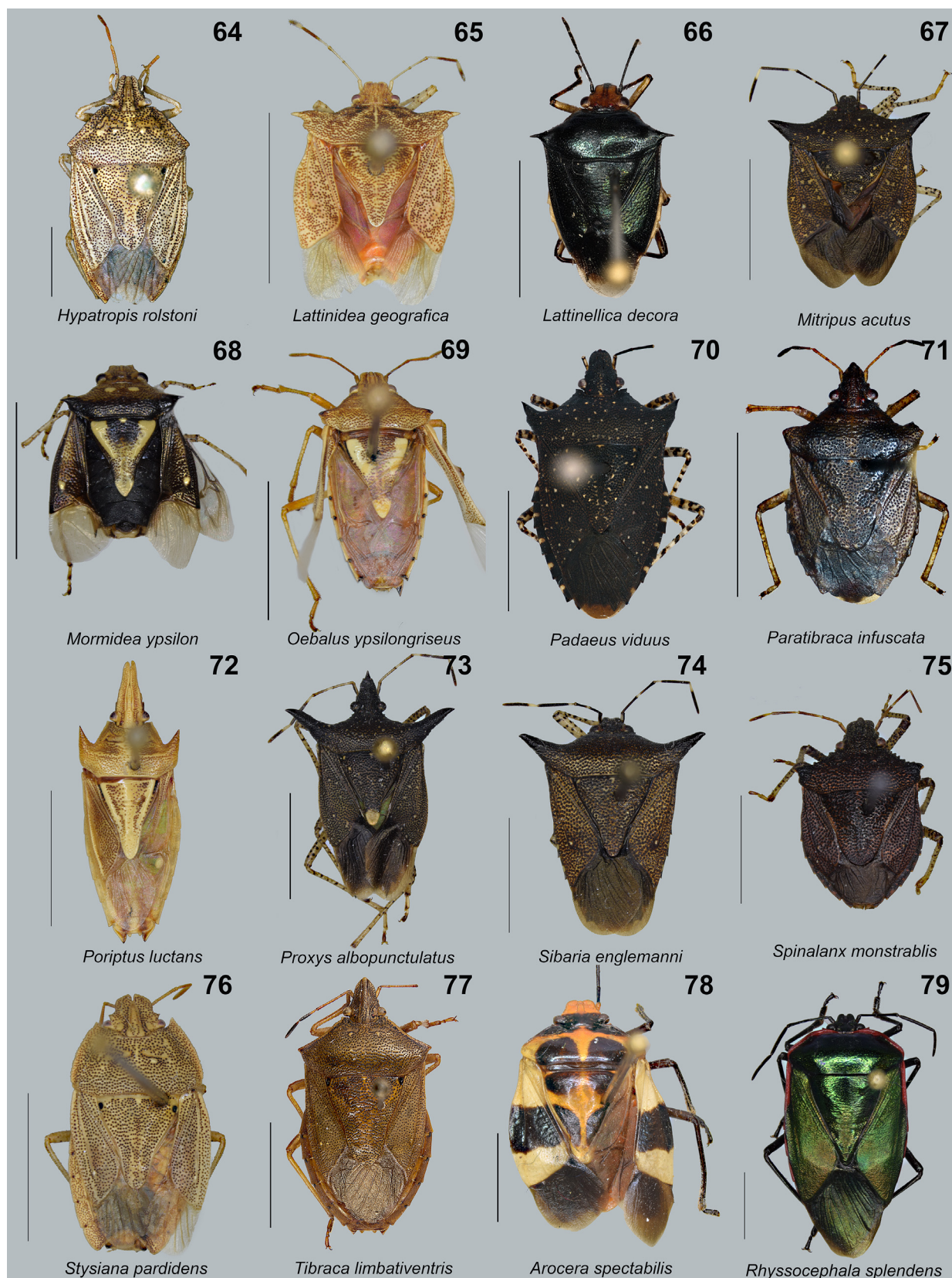
Padaeus sp.

Distribution of the genus. Mexico, Costa Rica, Guatemala, Colombia, and Brazil.

Distribution in Colombia. Valle del Cauca.

Remarks. *Padaeus* was recorded from Colombia by Torres (2004), but this study did not provide the species identification. The image of the dorsal habit of *Padaeus* offered by Torres (2004) does not allow confirmation to species level.

References. Kirkaldy 1909; Rolston & McDonald 1984; Torres 2004.



FIGURES 64–79. Pentatominae genera in Colombia. 64. *Hypatropis rolstoni* (UFRG). 65. *Lattinidea geografica* (MPUJ). 66. *Lattinellica decora* (UFRG). 67. *Mitripus acutus* (MPUJ). 68. *Mormidea ypsilon* (MPUJ). 69. *Oebalus ypsilon* (MPUJ). 70. *Padaeus viduus* (DAR). 71. *Paratibraca infuscata* (UFRG). 72. *Poriptus luctans* (MPUJ). 73. *Proxys albopunctulatus* (MPUJ). 74. *Sibaria englemanni* (MPUJ). 75. *Spinalanx monstrabilis* (MPUJ). 76. *Stysiana pardidens* (MPUJ). 77. *Tibraca limbiventris* (MZUSP). 78. *Arocera spectabilis* (MPUJ). 79. *Rhysocephala splendens* (MPUJ). Scale bar: 5 mm.

***Paratibraca* Campos & Grazia, 1995**

The genus can be recognized by having the head wider than long, the antecular region is nearly as wide as long, the antennal segment 5 is the longest, and the anterolateral margins of the pronotum are concave (Campos & Grazia 1995).

Paratibraca infuscata Campos & Grazia, 1995 [Fig. 71]

Distribution. Panama, Colombia, Trinidad and Tobago, Brazil, Peru, and Bolivia.

Distribution in Colombia. Meta, Santander, Tolima, and Valle del Cauca.

Remarks. *Paratibraca* is a monotypic genus.

References. Campos & Grazia 1995.

***Poriptus* Stål, 1861**

The genus can be recognized by having the head triangular and longer than the pronotum; the mandibular plates conspicuously surpassing the clypeus, the humeral angles have spines directed anteriorly, the peritreme is very short, and the pronotum and scutellum has calloused areas (Barcellos & Grazia 2008).

Poriptus luctans Stål, 1861 [Fig. 72]

Distribution. Colombia, Surinam, Brazil, Bolivia, and Argentina.

Distribution in Colombia. Meta.

Remarks. *Poriptus luctans* has the antennal segment III not surpassing the apex of the head, the humeral angles forming an acute angle with the anterolateral margins of the pronotum, the paramere and the lateral processes of the dorsal rim of the pygophore are bordered in black, and the sutural margins of the gonocoxite VIII are parallel.

References. Torres 2004; Barcellos & Grazia 2008.

***Proxys* Spinola, 1837**

The genus can be recognized by the distance between eye to pronotum, approximately half to the diameter of eye, with a striated region between eye and pronotum; and the clypeus is acute and surpass the mandibular plates (Rolston & McDonald 1984; Brailovsky 1987; Torres 2004)

Proxys albopunctulatus (Palisot de Beauvois, 1805) [Fig. 73]

Distribution. USA, Mexico, Honduras, Grenada, St. Vincent and Grenadines, Panama, Colombia, Venezuela, Brazil, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Boyacá, Casanare, Cundinamarca, Huila, and Meta.

Remarks. *Proxys albopunctulatus* has the metafemur lacking the apical black bands, the ventral margin of the pygophore is distinctly inflated, produced posteroventrally; and the lateral wall of the pygophore has 20 or more longitudinal ridges.

References. Kirkaldy 1909; Pirán 1948, 1956; Becker & Grazia 1971a; Thomas 2000; Arismendi & Thomas 2003; Rider 2009; Castro-Huertas *et al.* 2015.

Proxys obtusicornis Stål, 1872

Distribution. Colombia.

Distribution in Colombia. Boyacá, Casanare, Cundinamarca, Huila, Meta, Santander, and Tolima.

Remarks. *Proxys obtusicornis* has the humeral angles obtusely rounded or somewhat truncate with the posterior margin produced in a short, caudoventrally directed spine.

References. Kirkaldy 1909; Rider 2009.

Examined material. COLOMBIA: 1♂, 1♀, Boyacá, Maripi, 27 iv 1979, I. de Arévalo / ICN_077371 (ICN); Tunja, 1 vi 1991, S. Arpi / ICN_077379 (ICN); 1♀, Casanare, Cusiana, Pozo 1, 480 m, malaise, 28 ix 1995, F.

Fernández (ICN); 1♂, 4♀, **Cundinamarca**, El Colegio, 15 vi 1974, E. Zerda / ICN_077320 (ICN); El Colegio, 15 vi 1974, D. Bernal / ICN_077343 (ICN); Mosquera, 15 iv 1968, J.M. Rincón / ICN_077334 (ICN); Sasaima, 1 v 1972, J.A. Rodríguez / ICN_077335 (ICN); Silvania, 13 ii 1974, L.E. Forero / ICN_077318 (ICN); 1♀, **Huila**, Neiva, 6 iv 1971, Medina / ICN_077349 (ICN); 1♂, 1♀, **Meta**, Villavicencio, Bellavista, 1120 m, 10 ix 1976, L. Orozco / ICN_077375 (ICN); Bosque Bavaria, directa, 14 v 2005, L. Torres (ICN); 1♀, **Santander**, Virolín, margen río Luisito, 14 iii 1981 / ICN_077346 (ICN); 3♂, 1♀, **Tolima**, Alvarado, Ajonjolí, 22 xii 1971, A. Jaramillo / ICN_077331 (ICN); Espinal, al vuelo, 29 xii 1980, O. Jimenez / ICN_077350 (ICN); Espinal, en tomate, día claro, 8 ix 1974, A. Sánchez / ICN_077377 (ICN); Ibagué, 4°26' N, 75°13' W, 1250 m, 17 iii 2001, C. Torres & D. Forero / ICN_077380 (ICN).

Proxys punctulatus (Palisot de Beauvois, 1805)

Distribution. USA, Mexico, Guatemala, Nicaragua, Cuba, Dominican Republic, Honduras, Costa Rica, Colombia, Venezuela, and French Guiana.

Distribution in Colombia. Bolívar, Caldas, Chocó, Cundinamarca, and Tolima.

Remarks. *Proxys punctulatus* has the bucculae entirely black; the pro-, meso-, and metafemur usually with a black apical band.

References. Kirkaldy 1909; Becker & Grazia 1971a; Rider 2009; Castro-Huertas *et al.* 2015.

Proxys victor Fabricius, 1775

Distribution. Mexico, Guatemala, Nicaragua, Antilles, Panama, Colombia, Trinidad and Tobago, and Brazil.

Distribution in Colombia. Casanare, Cundinamarca, and Meta.

Remarks. *Proxys victor* has the evaporatorium usually unicolorous fuscous to black; the subdorsal ridge on the lateral wall of the pygopore is prominent, produced in a prominent ridge or tubercle.

References. Kirkaldy 1909; Rider 2009.

Examined material. COLOMBIA: 1♂, **Casanare**, Caserío Indígena Mochuelo, selva de galería, 100 m, 1 v 1976, F. Ortiz / ICN_077372 (ICN); 1♂, **Cundinamarca**, Medina, 540 m, 29 vii 1986, Sist. Avanz. / ICN_077324 (ICN); 1♀, **Meta**, Villavicencio, km 8 entre Villavicencio y Restrepo, 510 m, 11 ix 1976, N. Pinzón / ICN_077319 (ICN).

Sibaria Stål, 1872

The genus has the profemur armed with a pair of apical spines, the apex of the labium is reaching or surpassing the posterior margins of the mesocoxae, the metepisternum extending to the lateral margins of the thorax, and the capsula seminalis has finger-like processes (Rolston 1975; Bianchi *et al.* 2017; Krein *et al.* 2020).

Sibaria armata (Dallas, 1851)

Distribution. Colombia, Venezuela, Trinidad and Tobago, Guyana, Surinam, French Guiana, Brazil, Ecuador, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Amazonas, Boyacá, and Meta.

Remarks. *Sibaria armata* has the median emargination on the posterior margin of the pygophore broad, subquadrate, and with a tubercle on each side projecting caudad beyond the posterolateral angles of the pygophore.

References. Rolston 1975; Castro-Huertas *et al.* 2015; Melo *et al.* 2017; Krein *et al.* 2020.

Sibaria englemani Rolston, 1975 [Fig. 74]

Distribution. Mexico to Colombia.

Distribution in Colombia. Antioquia, Boyacá, Chocó, Magdalena, and Valle del Cauca.

Remarks. *Sibaria englemani* has the median emargination on the posterior margin of the pygophore wider than the proctiger, rounded, and with a shallower bisinuate emargination on each side.

References. Rolston 1975; Krein *et al.* 2020

Examined material. COLOMBIA: 1♂, **Boyacá**, Pto. Romero, Vda. La Cristalina, Q. La Cristalina, 400 m, 24 ix 1997, A. Ospina / ICN_MHN He 00409 (ICN).

***Spinalanx* Rolston & Rider, 1988**

The genus can be recognized by having the abdominal sternite III without a medial spine or tubercle; the anterolateral margin of the pronotum has denticles; the pro-, meso- and metafemur with several tubercles with setae, except on the distal region; the area between the posterior margin of the pygophore and the inferior ridge broad is nearly flat; and the basal plates are irregularly convex, with their posterior margins tuberculated (Rolston & Rider 1988).

Spinalanx monstrabilis Rolston & Rider, 1988 [Fig. 75]

Distribution. Colombia and Peru.

Distribution in Colombia. Putumayo.

Remarks. *Spinalax monstrabilis* has the gonocoxite VIII weakly convex with tubercles on the posterior margin and the lateral margin of the emargination.

References. Rolston & Rider 1988.

***Stysiana* Grazia, Fernandes & Schwertner, 1999**

The genus can be recognized by having the body dorsally depressed; the mandibular plates in a higher level than the clypeus in lateral view; the labial segment II almost as long as III and IV together; and the inferior layer of the ventral rim of the pygophore with a pair of tooth-like projections (Grazia *et al.* 1999).

Stysiana pardidens Grazia, Fernandes & Schwertner, 1999 [Fig. 76]

Distribution. Colombia, Guyana, Brazil, and Peru.

Distribution in Colombia. Meta.

Remarks. *Stysiana pardidens* has the mesosternum with a carina, and the anterolateral margins with a long and forward directed tooth.

References. Grazia *et al.* 1999.

***Tibraca* Stål, 1860**

The genus can be recognized by having the head longer than wide, the anteocular region longer than wide, the mandibular plates and the clypeus nearly equal in length, the apex of the mandibular plates are not contiguous, the labial segment I is between the bucculae, the antennal segments I and II equal in length, the antennal segment II is the longest, the anterolateral margins of the pronotum are rectilinear, and the frenum is distally wider than the half basal of the scutellum (Fernandes & Grazia 1998b, Torres 2004).

Tibraca limbativentris Stål, 1860 [Fig. 77]

Distribution. Dominican Republic, Costa Rica, Colombia, Venezuela, Brazil, Peru, Bolivia, and Argentina.

Distribution in Colombia. Valle del Cauca.

Remarks. *Tibraca limbativentris* is usually large (equal or longer than 13 mm), and the legs are brown, concolor with the body.

References. Fernandes & Grazia 1998b; Dellapé *et al.* 2020.

Tribe Catacanthini

***Arocera* Spinola, 1837**

The genus can be recognized by having the coloration pattern usually with yellow and red combined with black; the dorsal punctuation is minuscule and sparse, without wrinkles on the scutellum; the mandibular plates and the clypeus usually equal in length; the antennal segment I nearly reaching the apex of the head; the apex of the labium

reaching or surpassing the metacoxa; the anterolateral margins of the pronotum are reflexed; the metathoracic sulcus is elongated and upward; the humeral angles are rounded, the abdominal segment III is unarmed; and the tarsi is three segmented (Rider 1992).

Arocera aequinoxialis (Westwood, 1837)

Distribution. Guatemala, Costa Rica, Panama, Colombia, Guyana, Surinam, French Guiana, Ecuador, Brazil, Peru, and Bolivia.

Distribution in Colombia. Amazonas, Antioquia, and Valle del Cauca.

Remarks. *Arocera aequinoxialis* has the dorsal surface of the body is pale yellow with a large U-shaped black marking, and the femora are fuscous to black on the distal one-half, the basal one-half is yellow with the margin between yellow and black areas sharp.

References. Rider 1992.

Arocera apta (Walker, 1867)

Distribution. Costa Rica, Panama, Colombia, Venezuela, Guyana, French Guiana, Ecuador, Peru, Bolivia, Brazil, and Argentina.

Distribution in Colombia. Amazonas, Antioquia, Arauca, Atlántico, Caquetá, Cundinamarca, Meta, Putumayo, Santander, and Valle del Cauca.

Remarks. *Arocera apta* has the humeral angles yellow, the connexivum uniformly pale yellow, and the abdominal sternites are uniformly pale yellow except for some black markings on the gonocoxite VIII.

References. Rider 1992.

Examined material. COLOMBIA: 1♀, **Arauca**, La Conquista, Fca. Macaguán, 120 m, v 1976, F. Ortíz / ICN_055434 (ICN); 1♀, **Cundinamarca**, Yacopí, Vda. La Oscura, Qda. Caipala, 400 m, 11 iii 2000, M. Rocha & Est. (ICN); **Santander**, Bucaramanga, 22 i 1974, Lozada / ICN_055428 (ICN); **Putumayo**, Puerto Asis, 26 ii 1972 / MPUJ_ENT 0010745 (MPUJ_ENT).

Arocera elongata Showalter, 1929

Distribution. Panama, Colombia, Venezuela, Ecuador, Peru, Brazil, and Bolivia.

Distribution in Colombia. Antioquia, Boyacá, Meta, Putumayo, and Valle del Cauca.

Remarks. *Arocera elongata* has the dorsal surface of the body is mostly black with six orange spots, one on the head, one on the basal third of each corium, one on the posterolateral angle of each corium, and one on the apex of the scutellum; the spot on the apex of the scutellum sometimes continued cephalad as a thin orange line.

References. Rider 1992.

Examined material. COLOMBIA: 2♀, **Boyacá**, San Luis de Gaceno, Horizontes Buena Vista, 15 x 2005, N. Ocampo (ICN); Santa María, Sendero Ecológico, 1200 m, iv 1997, G. Amat / ICN_055441 (ICN); **Putumayo**, Orito, 1 iii 1972 / MPUJ_ENT 0010742 (MPUJ_ENT); 28 ii 1972 / MPUJ_ENT 0010743 (MPUJ_ENT); 2♂, **Valle del Cauca**, B. Anchicayá, 400 m, H. Delgado / 1940 (MUSENUV); C.H. Anchicayá, 400 m, 1 vii 1976 / 1941 (MUSENUV).

Arocera nigrorubra (Dallas, 1851)

Distribution. Colombia.

Distribution in Colombia. Cundinamarca and Valle del Cauca.

Remarks. *Arocera nigrorubra* has the antennal segment I fuscous or black, concolorous with remaining of the antennal segments, and the posterolateral angles of the pygophore are armed with one to three minute, medially directed teeth.

References. Rider 1992.

Examined material. COLOMBIA: 2♂, **Valle del Cauca**, Buenaventura-Anchicayá, 300 m, 1 vii 1970 / 1925 (MUSENUV); Peñas Blancas, 1500 m, 1 x 1981 / 1926 (MUSENUV).

Arocera placens (Walker, 1867)

Distribution. Mexico to Argentina.

Distribution in Colombia. Magdalena, Putumayo, and Valle del Cauca.

Remarks. *Arocera placens* has the posteroventral surface of the pygophore with a shallow, semicircular depression, and an obtuse carina on the ventral margin.

References. Rider 1992.

Arocera rufifrons (Dallas, 1851)

Distribution. Colombia, Ecuador, Peru, and Brazil.

Distribution in Colombia. Valle del Cauca.

Remarks. *Arocera rufifrons* has the ocelli very small, the distance from the ocellus to the adjacent eye is at least three or four times the diameter of an ocellus, and the dorsal margin of the paramere is concave in lateral and medial views.

References. Rider 1992.

Arocera spectabilis (Drury, 1782) [Fig. 78]

Distribution. Ecuador, Peru, and Brazil. New record for Colombia.

Distribution in Colombia. Antioquia.

Remarks. *Arocera spectabilis* has the humeral angles black, the connexivum is alternately fuscous and pale yellow, and the abdominal sternites usually have black spots.

References. Rider 1992.

Examined material. COLOMBIA: 1♂, **Antioquia**, Río Claro. 440m. 7 ix 1994. D. Arias.leg / MPUJ 13.349 / No 1542 (MPUJ).

***Rhysocephala* Rider, 1991**

The genus can be recognized by having the body black, dark brown, and metallic green or blue, with some yellow, orange, or red spots or bands; the dorsal punctuation usually is very fine, dense and forming wrinkles on the scutellum; the mandibular plates are dorsally striated; the labium reaching or surpassing the metacoxae, the metathoracic sulcus is elongated; the abdomen base is flat, and the tarsi are three segmented (Rider 1991).

Rhysocephala immaculata (Pirán, 1963)

Distribution. Colombia, Peru, and Bolivia.

Distribution in Colombia. Boyacá, Cundinamarca, and Meta.

Remarks. *Rhysocephala immaculata* has the posterior wall of the pygophore with several black spicules, except on the medial glabrous depressed area and on the submarginal band along the posterior margin of the pygophore.

References. Rider 1991.

Rhysocephala macdonaldi Rider, 1991

Distribution. Guatemala to northwestern South America. Record in Jamaica doubtful.

Distribution in Colombia. Amazonas, Antioquia, Cundinamarca, Magdalena, Putumayo, and Tolima.

Remarks. *Rhysocephala macdonaldi* has a reddish band along anterolateral pronotal margins usually lacking infuscated areas, basal plates fuscous to black, becoming reddish basally, ventral margin of pygophore in ventral view with a large, subtriangular, medial emargination, and posterior wall of pygophore with small black spicules laterally.

References. Rider 1991.

Examined material. COLOMBIA: 1♀, **Amazonas**, PNN Amacayacu, Boca Caño Matamata, 6 x 1988, F. Fernández / IAvH-E 05036 (IAvH-E); 1♀, **Cundinamarca**, Nariño, 300 m, luz bombillo, 22 iv 1988, R. Jaramillo / ICN_077422 (ICN); **Tolima**, Ibagué, 28 ii 1973, J. Rojas / ICN_077424 / ICN_077421 (ICN).

Rhysocephala principalis (Stål, 1855)

Distribution. Colombia and Venezuela. Record in Mexico doubtful.

Distribution in Colombia. Boyacá and Cundinamarca,

Remarks. *Rhysocephala principalis* has the dorsal punctuation of the body minute, relatively dense, forming weak

rugulose lines on the pronotum and the scutellum; and the posterior wall of the pygophore is lacking the black spicules.

References. Rider 1991.

Rhysocephala splendens (Blanchard, 1840) [Fig. 79]

Distribution. Mexico to northwestern South America.

Distribution in Colombia. Boyacá, Cundinamarca, Huila, Magdalena, Risaralda, and Valle del Cauca.

Remarks. *Rhysocephala splendens* has the dorsal surface of the head bicolorous, metallic green basally becoming orange to red distally at least on the apices of the mandibular plates.

References. Rider 1991.

Examined material. COLOMBIA: 1♂, 1♀, **Cundinamarca**, San Agustín, Chaguani, 10 iv 1974, H. León & C. Barbosa / ICN_077410 (ICN); Vía a Girardot, km 108, Centro vacacional “El Abanico” Asmedas, 330 m, viii 1991, R.O. / MPUJ_ENT 0018131 (MPUJ_ENT); 1♀, **Risaralda**, La Suiza, 1910 m, 23 iii 1992, Aguilar & Guerrero / MPUJ_ENT 0018130 (MPUJ_ENT); 1♀, **Valle del Cauca**, Tuluá, Mateguadua, Jardín Botánico Juan María Céspedes, 1100 m, 04.02913° N, 76.166981° W, 24-31 viii 1996, D. Forero / MPUJ_ENT 0010701 (MPUJ_ENT); L. Barragán / MPUJ_ENT 0018132 (MPUJ_ENT).

***Runibia* Stål, 1861**

The genus can be recognized by having the body oval and slightly convex dorsoventrally; the coloration pattern is yellowish to reddish, the dorsal surface with several dark brown spots, the humeral angles are not projected, and the mesosternum has a carina (Zwetsch & Grazia 2001).

Runibia decorata (Dallas, 1851) [Fig. 80]

Distribution. Colombia, Venezuela, Ecuador, Brazil, Peru, and Bolivia.

Distribution in Colombia. Meta.

Remarks. *Runibia decorata* has the anterolateral margins of the pronotum slightly sinuated and convex, and the distal portion of the hemelytra has a pair of dark spots.

References. Zwetsch & Grazia 2001.

Runibia discoidea (Fabricius, 1787)

Distribution. Colombia, French Guiana, Brazil, and Peru.

Distribution in Colombia. Amazonas.

Remarks. *Runibia discoidea* has the dorsal surface of the body yellowish to reddish, with a conspicuous black spot on the pronotum, the scutellum and the hemelytra.

References. Zwetsch & Grazia 2001.

***Vulsirea* Spinola, 1837**

The genus can be recognized by having the body conspicuously convex, the antennal segment I surpassing the apex of the head; the antennal segments IV and V very long; the humeral angles are not projected; the mesosternum with a carina; and the basal region of the abdomen with a medial tubercle (Rolston & McDonald 1980; Torres 2004).

Vulsirea violacea Fabricius, 1803 [Fig. 81]

Distribution. Mexico, Cuba, Jamaica, Costa Rica, Panama, Colombia, Brazil, and Argentina.

Distribution in Colombia. Antioquia and Tolima.

Remarks. The genus was recorded for Colombia by Torres (2004). We confirm the identification of the species as *V. violacea*, which can be recognized by the coloration metallic dark blue with red maculae distributed along the body, which present high polymorphism within populations.

References. Kirkaldy 1909; Kormilev 1951, 1956; Rolston & McDonald 1980; Torres 2004; Lopez & Cervantes 2010.

Tribe Chlorocorini

Chlorocoris Spinola, 1837

The genus can be recognized by having the body yellow or green and dorsoventrally flattened; the anterolateral margin of the pronotum is serrate; the antennal segment I reaching the apex of the mandibular plates; the mandibular plates slightly longer than the clypeus; the femora are unarmed; the metasternum is not elevated; and the basal abdominal spine is lacking (Thomas 1985).

Chlorocoris depressus (Fabricius, 1803)

Distribution. Colombia, Venezuela, Surinam, Trinidad and Tobago, and Brazil.

Distribution in Colombia. Cundinamarca, Meta, and Valle del Cauca.

Remarks. *Chlorocoris depressus* has the humeral angles with short spines, its abruptly produced; the margins of the mandibular plates are concolorous with disc; and the body, particularly the abdomen, extremely dorsoventrally compressed.

References. Thomas 1985.

Examined material. COLOMBIA: 1♀, **Cundinamarca**, Medina, Granja Experimental, 520 m, 29 vii 1986, Sist. Avanz. / ICN_055892 (ICN); 1♀, **Meta**, Puerto López, 6 vi, L.E. Aguirre / ICN_055880 (ICN); 1♂, 1♀, **Valle del Cauca**, Buenaventura, Río Yurumanguí. Manglar, 2 vii 1997, M. Reyes / ICN_055891 (ICN); vii 1970, L. Ángel / ICN_055879 (ICN).

Chlorocoris distinctus Signoret, 1851

Distribution. Colombia, Venezuela, Surinam, Trinidad and Tobago, and Brazil.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas (1985), but without specific locality.

Remarks. *Chlorocoris distinctus* has the posterior margin of the connexivum bordered with a thin black line, and the posterior surface of the pygophore with a broad and oval excavation.

References. Thomas 1985.

Chlorocoris isthmus Thomas, 1985

Distribution. Costa Rica, Panama, Colombia, and Ecuador.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas (1985), but without specific locality.

Remarks. *Chlorocoris isthmus* has the humeral angles with a V-shaped marking, and the pygophore with a deep and narrow emargination in ventral view.

References. Thomas 1985.

Chlorocoris sororis Thomas, 1985 [Fig. 82]

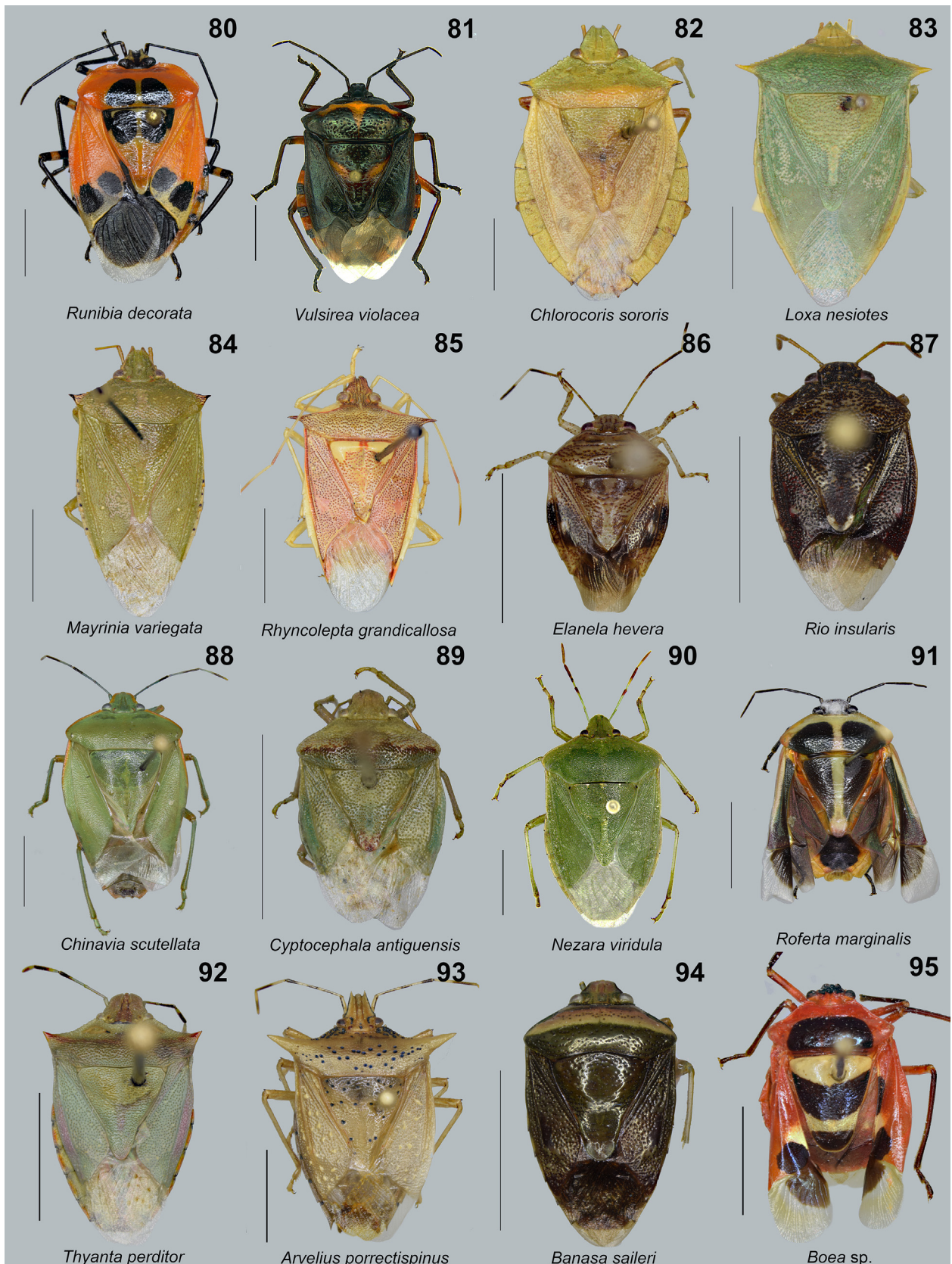
Distribution. Colombia.

Distribution in Colombia. Caldas and Valle del Cauca.

Remarks. *Chlorocoris sororis* has the posterior margin of the connexivum bordered by a thin black line, and the posterior surface of the pygophore has a narrow, deep, and triangular excavation.

References. Thomas 1985.

Examined material. COLOMBIA: 1♂, **Caldas**, Ca. Norcasia, 230 m, 30 xii 1984, Martha. C. Alvarado / 1977 (MUSENUV); 4♂, 1♀, **Valle del Cauca**, Anchicayá, 620 m, 16 ix 1993, F. López / 1956 (MUSENUV); Cali, Tierra cult., 1000 m, 1 v 1970 / 1955 (MUSENUV); Cali, Univalle, 1000 m, 2 ii 2002, Catalina Gutierrez / 21609 (MUSENUV); Ca. Cali, 1000 m, 1972 / 1957 (MUSENUV); Bajo Anchicayá, 430 m, 1 x 1985, Martha L. Baena / 1978 (MUSENUV); Palmira, Tenjo, 3°32' N, 76°10' W, 1550 m, manual, 28 ix 2013, David Rivera / 28122 (MUSENUV).



FIGURES 80–95. Pentatominae genera in Colombia. 80. *Runibia decorata* (MPUJ). 81. *Vulsirea violacea* (MZUSP). 82. *Chlorocoris sororis* (MPUJ). 83. *Loxa nesiotis* (MPUJ). 84. *Mayrinia variegata* (MPUJ). 85. *Rhyncolepta grandicallosa* (ICN). 86. *Elanela hevera* (MPUJ). 87. *Rio insularis* (MPUJ). 88. *Chinavia scutellata* (MUSENUV). 89. *Cyptocephala antiguensis* (ICN). 90. *Nezara viridula* (MZUSP). 91. *Roferta marginalis* (MPUJ). 92. *Thyanta perditor* (MPUJ). 93. *Arvelius porrectispinus* (MPUJ). 94. *Banasa saileri* (MPUJ). 95. *Boea* sp. (MPUJ). Scale bar: 5 mm.

Loxa Amyot & Serville, 1843

The genus can be recognized by having the osteolar sulcus extending less of the half distance between the medial margin of the peritreme and the metapleura; the apex of the femora have a spine, and the hemelytron without a callosity (Eger 1978).

Loxa nesiotus Horváth, 1925 [Fig. 83]

Distribution. Lesser Antilles, Curaçao, Panama, Colombia, Venezuela, and Guyana.

Distribution in Colombia. Caldas.

Remarks. *Loxa nesiotus* has the humeral angles with spines angled slightly caudad at most, usually projected laterally, and sometimes curving cephalad.

References. Eger 1978.

Examined material. COLOMBIA: Caldas, Dorada, Guarinocito, 480m, 17 ix 1989; Arango *et al.* / MPUJ_ENT 0010695 (MPUJ_ENT).

Loxa virescens Amyot & Serville, 1843

Distribution. Southern of Mexico to Argentina.

Distribution in Colombia. Casanare, Caldas, Cundinamarca, Magdalena, Meta, Risaralda, Santander, Tolima, and Valle del Cauca.

Remarks. *Loxa virescens* has the paramere process projected laterally, and the female has the pronotum between the humeral angles with a transverse fascia of the rugae, and the corium has a discoidal pale spot.

References. Eger 1978; Dellapé *et al.* 2020.

Examined material. COLOMBIA: 1♀, **Casanare**, Mochuelo, caserío indígena. selva de galería, 100 m, vii 1972, E. Ortiz / ICN_077280 (ICN); 1♂, **Caldas**, Samacá, Norcasia, Campamento de la CHEC, 500 m, 7 xi 1988, I.S.A / ICN_077302 (ICN); 2♀, **Cundinamarca**, Cáqueza, 29 xii 1982, P. Moreno / ICN_077215 (ICN); Ubalá, San Pedro de Jagua, Puerto Soya B, 4°42'48" N, 3°18'6" W, 550 m, manual, 2 vii 1998, A. Varón & E. Flórez (ICN); 1♂, 1♀, **Magdalena**, Santa Marta, Minca, Hda. La Victoria, Casa Vieja, 11°7'7.2" N, 54°5'21.6" W, 1027 m, con luz, 13-16 x 2007, F. Fernández & Est. Sist. (ICN); Sierra Nevada de Sta. Marta, Minca, iv 1977, R. Restrepo / ICN_077214 (ICN); 3♂, 3♀, **Meta**, Macarena, 500 m, xii 1949, L. Richter / ICN_077272 (ICN); Macarena, 500-650 m, ii 1950, L. Richter / ICN_077254 (ICN); Puerto López, 6 vi, L.E. Aguirre / ICN_077279 (ICN); Restrepo, Sede CREAD, 523 m, jameo, 5 iv 1988, C. Novoa / ICN_077304 (ICN); San Martín, Fca. El Caduceo, cerca al río Camoa, 400 m, trampa de luz, 18 v 2006, D. Ochoa (ICN); Villavicencio, Vda. La Vanguardia, Pozo azul, trampa de luz, 6 iv 2005, Curso Sist. (ICN); Risaralda, Pereira, PRN Ucumari, P. Nevados, 19 viii 1992, Matuk-Ochoa / MPUJ_ENT 0010673 (MPUJ_ENT); 1♂, **Santander**, Carare, Pwd. Campo Capote, v 1970, I. Cabrera / ICN_077211 (ICN); 2♂, 4♀, **Tolima**, Alvarado, Miravalle, 19 iv 1974, M.L. Baena / ICN_077257 (ICN); Armero, Vda. San Felipe, 15 i 1983, G. Hilda / ICN_077305 (ICN); Chaparral, 11 iii 1973, L. Quiñones / ICN_077260 (ICN); Ibagué, Vda. El Totumo 4°26,701' N, 75°11,950' W, 1100 m, 6-9 vii 2013, J.E. Eger & A.A. Calixo (ICN); Mariquita, 29 viii 1987, R. Arce / MPUJ_ENT 0010676 (MPUJ_ENT); Mariquita, 495 m, 29 viii 1987, S. Niño *et al.* / MPUJ_ENT 0010677 (MPUJ_ENT); 1♂, 3♀, **Valle del Cauca**, Buenaventura, Bajo Calima, 70 m, 22 iii 1995, CFT / MPUJ_ENT 0010678 (MPUJ_ENT); Buenaventura, Río Raposo, manglar, 17 vii 1997, C. Medina / ICN_077256 (ICN); Río Anchicaya, 400 m, 20 Oct 1970, R. Cortés / MPUJ_ENT 0010674 / MPUJ_ENT 0010675 (MPUJ_ENT); Sevilla, 11 iv 1974, J. Aguirre / ICN_077278 (ICN).

Loxa viridis (Palisot de Beauvois, 1805)

Distribution. Southern of USA to southern of Brazil and Argentina.

Distribution in Colombia. Antioquia, Caldas, Chocó, Cundinamarca, Huila, Meta, Santander, Tolima, and Valle del Cauca.

Remarks. *Loxa viridis* has the paramere process with the apex rounded and inclined dorsally in caudal view, and the female has the lateral margins of the corium strongly sinuous, and the lateral angles produced.

References. Eger 1978.

Examined material. COLOMBIA: **Antioquia**, San Luis, Reserva Natural Cañon del Río Claro, 440m, 27 ii 1994, A. Quintero / MPUJ_ENT 0010696 (MPUJ_ENT); 3♂, 2♀, **Caldas**, La Dorada, 23 iii 1970, C. Riveros / ICN_077303

/ ICN_077276 / ICN_077274 (ICN); La Dorada, 19 iv 1974, A. B. Lotero / ICN_077273 / ICN_077301 (ICN); 2♀, **Chocó**, Ríosucio, Cacarica, 9 viii 1978, H.E.E. / ICN_077229 / ICN_077255 (ICN); 3♀, 2♂, **Cundinamarca**, Agua de Dios, 2 x 970, C. Princo / ICN_077212 (ICN); Cachipay, 30 xii 1979, J. Rojas / ICN_077226 (ICN); El Ocaso, Casa Vacacional, 1 i 1978, H. Alarcón / ICN_077251 (ICN); Girardot, viii 1968, J. Hernández / ICN_077253 (ICN); Guaduas, 16 vii 1972, M.I. Urrego / ICN_077271 (ICN); Nilo, 29 vi 1972, L. de Peñaranda / ICN_077275 (ICN); Tocaima, 2 vii 1972, G. Cortés / ICN_077227 (ICN); 2♂, **Huila**, Neiva, 6 iv 1971, Medina ICN_077228 (ICN); Neiva, 11 vii 1974, Albagos / ICN_077259 (ICN); 3♀, **Meta**, Puerto Gaitán, 1 xi 1977, A. Jimenez / ICN_077258 (ICN); Río Ocoa, 350 m, v 1945, L. Richter / ICN_077252 (ICN); Villavicencio, Bosque de Bavaria, 15 v 2005, P. Restrepo (ICN); 1♀, **Santander**, Landaxuri, 7 ii 1949, L. Richter / ICN_077230 (ICN); 2♀, **Tolima**, Km 6 de Prado, Chanchito, 26 iii 1986, A. Molano (ICN); Melgar, La Naranjala, xii 1969, Fajardo / ICN_077213 (ICN); 3♀, **Valle del Cauca**, Buenaventura, Bajo Calima, 70 m, 21 iii 1995, Irle *et al.* / MPUJ_ENT 0010697 (MPUJ_ENT); Caralmer, vi 1970, L. Ángel / ICN_077277 (ICN); Cartago, 7 xii 1969, Fajardo / ICN_077316 (ICN).

***Mayrinia* Horváth, 1925**

The genus can be recognized by having the oval body, slightly convex, with green coloration with dark green puncture; the head and the pronotum together are triangular; the mandibular plates longer than the clypeus; the external margins of the mandibular plates from convex to sinuate; the humeral angles with a spine, anterior or laterally projected; the hemelytron with pale callosity on the mesocorium; the osteolar sulcus extending beyond of the half distance between the medial margin of the peritreme and the metapleura; and the apex of the femora with a short spine (Grazia 1972).

Mayrinia variegata Distant, 1880 [Fig. 84]

Distribution. Nicaragua, Costa Rica, Colombia, Venezuela, Guyana, Peru, Brazil, and Argentina.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Grazia (1972), but without specific locality.

Remarks. *Mayrinia variegata* has the humeral spines projected upward with the margins black.

References. Grazia 1972; Dellapé 2021a.

***Rhyncholepta* Bergroth, 1911**

The genus can be recognized by having the body elongated, typically reddish brown with pale yellowish green areas; the head, although relatively flat dorsally and subtriangular, does not have the apices of the mandibular plates acute or spinose, but are narrowly rounded; and the humeral angles are projected with acute spines laterally directed (Kment *et al.* 2018).

Rhyncholepta grandicallosa Bergroth, 1911 [Fig. 85]

Distribution. Panama, Colombia, Venezuela, French Guiana, and Brazil.

Distribution in Colombia. Chocó.

Remarks. *Rhyncholepta grandicallosa* has a pale callosity on the anterolateral margins of the pronotum, and on the basal and distal portions of the scutellum.

References. Becker & Grazia 1971b; Castro-Huertas *et al.* 2015; Kment *et al.* 2018.

Examined material. COLOMBIA: 1♀, **Chocó**, Riosucio, Sautatá, 10 iii 1978, H. Echeverri / ICN_077425 (ICN).

Tribe Menidini

***Elanela* Rolston, 1980**

The genus can be recognized by having the body castaneous with brown punctuations; the lateral margins of the

mandibular plates outlined in black; the distal half of the antennal segments IV and V are black; the scutellum has small yellow calluses on the basal angle and on the basal margin medially; the disc of scutellum is black, reddish, or ivory, and calloused; the head is wider than long, strongly sloping ventrally; the apex of the mandibular plates not surpassing the clypeus; the margins of the mandibular plates are strongly concave before the eyes; the posterior margin of the buccula is rounded; the first labial segment surpassing the bucculae; the pronotum is trapezoidal; the anterolateral margins of the pronotum are straight; the humeral angles are rounded; the mesosternum is carinate; the metasternum is produced, the peritreme is spout-shaped; the evaporatorium is covering the most of the mesopleura and metepisternum; and the posterolateral angle of the connexivum is acute (Grazia *et al.* 2016).

Elanela colombiana Barros, Barão & Grazia, 2020

Distribution. Colombia.

Distribution in Colombia. Valle del Cauca.

Remarks. *Elanela colombiana* has the disk of scutellum callused, callus pale brown, pro-, meso- and metafemur yellowish or light, abdominal sternite VII brown medially, gonocoxite VIII wider than long, and gonocoxite IX swollen.

References. Barros *et al.* 2020.

Elanela diamphidiosa Grazia, Barros & Barão, 2016

Distribution. Colombia and Ecuador.

Distribution in Colombia. Amazonas.

Remarks. *Elanela diamphidiosa* has the pygophore with acute marginal processes on each side of the dorsal rim medially, each projected posteriorly and inwards in the genital cup; the parameres are apically spatulate, embedded in the abdominal segment X basally; the abdominal segment X is elongate, surpassing the posterior margin of the pygophore, carinate dorsally, sinuous distally, and blunt apically; the gonocoxites IX are strongly concave; and the anterior and posterior margins of the gonocoxites VIII are sinuate, with a notch medially.

References. Grazia *et al.* 2016.

Elanela hevera Rolston, 1980 [Fig. 86]

Distribution. Colombia and Peru.

Distribution in Colombia. Amazonas.

Remarks. *Elanela hevera* has the pygophore globose, ventral rim rounded laterally, apex of paramere not juxtaposed medially with the other one, and posterior margin of gonocoxite IX sinuate, and with a medial notch.

References. Rolston *et al.* 1980; Torres 2004; Grazia *et al.* 2016; Barros *et al.* 2020.

Rio Kirkaldy, 1909

The genus can be recognized by having the oval body, entirely pale to dark brown with dark brown punctured on the head, pronotum, scutellum and hemelytra; the humeral angles are rounded; the pronotum and scutellum sometimes with yellow calluses; the connexivum with yellowish spots; and the pro-, meso-, and metatibiae are sulcate (Grazia & Fortes 1995).

Rio insularis Ruckes, 1960 [Fig. 87]

Distribution. Panama, Colombia, and Venezuela.

Distribution in Colombia. Valle del Cauca.

Remarks. *Rio insularis* has the abdominal sternites brown-yellowish, and the dorsal margin of the pygophore is excavated on each side.

References. Grazia & Fortes 1995; Castro-Huertas *et al.* 2015.

Tribe Nezarini

***Chinavia* Orian, 1965**

The genus can be recognized by having the body entirely green, the pro-, meso- and metatibiae surface with longitudinal striation, and the first labial segment is between the bucculae (Schwertner & Grazia 2007).

Chinavia dallasi (Distant, 1900)

Distribution. Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1983a), but without specific locality.

Remarks. *Chinavia dallasi* has the body green with a reddish band on the lateral margins, the pronotum has two pairs of black spots, the basal angles of the scutellum have a pair of black spots, the spiracles are black, and a callosity at the side to spiracles.

References. Rolston 1983a; Schwertner & Grazia 2007.

Chinavia gravis (Walker, 1867)

Distribution. Colombia, Brazil, and Peru.

Distribution in Colombia. Probably Amazonas. This species was recorded for “Amazon region” by Rolston (1983a), and for Colombia by Schwertner & Grazia (2007), but without specific locality.

Remarks. *Chinavia gravis* has the body dark green with a yellowish medial longitudinal band on the pronotum and scutellum; the margins of the mandibular plates, the pronotum, the basal portion of the hemelytron and connexivum, and the legs are green with dark brown spots.

References. Rolston 1983a; Schwertner & Grazia 2007.

Chinavia impicticornis (Stål, 1872)

Distribution. Haiti, Dominican Republic, Colombia, Venezuela, French Guiana, Surinam, Ecuador, Brazil, Peru, Bolivia, Peru, Paraguay, and Argentina.

Distribution in Colombia. Meta and Valle del Cauca.

Remarks. *Chinavia impicticornis* has the body green with the lateral margins of the mandibular plates, the pronotum, the basal portion of the hemelytron and connexivum yellowish; the basal angles of the scutellum have a pair of black spots; and the spiracles are green.

References. Rolston 1983a; Schwertner & Grazia 2007; Servino & Schwertner 2020.

Examined material. COLOMBIA: 1♀, **Meta**, Macarena, iv 1951, L. Richter / ICN_055868 (ICN); **Valle del Cauca**, Tuluá, Mateguadua, Jardín Botánico Juan María Céspedes, 1100 m, 04.02913° N, 76.166981° W, 24-31 viii 1996, A. Fajardo, MPUJ_ENT 0010704 (MPUJ_ENT).

Chinavia marginata (Palisot de Beauvois, 1817)

Distribution. USA, Mexico, Dominican Republic, Puerto Rico, Jamaica, Honduras, Nicaragua, Costa Rica, Panama, Colombia, Venezuela, and Peru.

Distribution in Colombia. Antioquia, Meta, and Valle del Cauca.

Remarks. *Chinavia marginata* has the humeral angles slightly produced laterally; the abdominal tubercle is compressed, reaching the middle of the metacoxa; the posterolateral angles of the connexival segments, including the corner of the laterotergites and sternites, with black spots.

References. Rolston 1983a; Arismendi & Thomas 2003; Segarra-Carmona *et al.* 2016; Servino & Schwertner 2020.

Examined material. COLOMBIA: 1♀, **Antioquia**, San Luis, Reserva Natural Cañon del Río Claro, 440 m, 27 ii 1994, MPUJ_ENT 0010702 (MPUJ_ENT); 2♀, **Meta**, Km 91, Carretera Bogotá—Villavicencio, Pipiralito, 1000 m, 14 vi 1980, C. Bohorquez & C. Hernández / ICN_055870 (ICN); Villavicencio, Vda. Guanaviche, Estadero, río Guataquí, 4°10.506' N, 73°38.233' W, 1465 ft, 3-5 vii 2013, J.E. Eger & A.A. Calixto (ICN); 1♂, **Valle del Cauca**, Restrepo, Camino Hda. El Pital, 900 m, 6 ii 1984, D. Morales / ICN_055855 (ICN).

Chinavia plaumanni (Rolston, 1983)

Distribution. Colombia, Ecuador, and Peru.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Schwertner & Grazia (2007), but without specific locality.

Remarks. *Chinavia plaumanni* has the connexivum with a narrow dark band, and the abdominal spine inconspicuous forming a tubercle.

References. Rolston 1983a; Schwertner & Grazia 2007; Servino & Schwertner 2020.

Chinavia runaspis (Dallas, 1851)

Distribution. Colombia, Venezuela, Surinam, French Guiana, Brazil, Ecuador, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Antioquia, Chocó, Valle del Cauca, and Vaupés.

Remarks. *Chinavia runaspis* has the body green with the margins of the mandibular plates, the pronotum, the basal portion of the hemelytron and the connexivum with a reddish band; and the spiracles are green.

References. Rolston 1983a; Schwertner & Grazia 2007; Servino & Schwertner 2020.

Examined material. COLOMBIA: **Antioquia**, San Luis, Reserva Natural Cañon del Río Claro, 440 m, 4 ix 1994, E.F.A / MPUJ_ENT 0010703 (MPUJ_ENT); 1♀, **Chocó**, B. González / ICN_055867 (ICN); 2♀, **Valle del Cauca**, Buenaventura, río Raposo, manglar, 27 vii 1997, C. Medina / ICN_055876 (ICN); Bugalagrande, 8 viii 1969, M. Chaparro / ICN_055866 (ICN); 1♂, **Vaupés**, río Guayabero, 190 m, vi 1968 / ICN_055877 (ICN).

Chinavia schuhi Schwertner & Grazia, 2006

Distribution. Colombia, Peru, and Brazil.

Distribution in Colombia. Bolívar.

Remarks. *Chinavia schuhi* has the body green with the margins of the head, the pronotum, the hemelytron and the connexivum reddish; the basal angles of the scutellum are green without spots; and the labium surpassing the metacoxae.

References. Schwertner & Grazia 2006, 2007.

Chinavia scutellata (Distant, 1890) [Fig. 88]

Distribution. Mexico to Panama. New record for Colombia.

Distribution in Colombia. Valle del Cauca.

Remarks. *Chinavia scutellata* has the connexivum with large black macules, each bisected by a transverse connexival suture, continuing onto the margin of the venter; the humeral angles are rounded, not produced laterally; the femora are distally and tibiae are basally usually reddish; the abdominal spine is compressed, surpassing the metacoxae, sometimes reaching the mesocoxae; and the spiracles are yellowish.

References. Rolston 1983a.

Examined material. COLOMBIA: 2♀, **Valle del Cauca**, Cali, Univalle, en *Guazuma ulmifolia*, 3°22'63.8" N 76°31'86.4" W, 970m, 17 viii 2005, Entomophilo (MUSENUV).

Chinavia teretis (Rolston, 1983)

Distribution. Colombia.

Distribution in Colombia. Magdalena.

Remarks. *Chinavia teretis* has the basal angles of the scutellum immaculate and the abdominal tubercle nearly reaching the anterior limit of the metacoxae.

References. Rolston 1983a.

Chinavia ubica (Rolston, 1983)

Distribution. Haiti, Dominican Republic, Bahamas, Granada Island, Panama, Colombia, Surinam, Guyana, Venezuela, Ecuador, Brazil, and Bolivia.

Distribution in Colombia. Cauca, Cundinamarca, Meta, and Valle del Cauca.

Remarks. *Chinavia ubica* has the body green with the lateral margin of the head, the pronotum, the hemelytron, and the connexivum reddish; and the anterolateral angles of the connexivum have dark spots.

References. Rolston 1983a; Schwertner & Grazia 2007; Servino & Schwertner 2020.

Examined material. COLOMBIA: 1♂, **Cauca**, Timbiu, Robles, en *Morus indica*, 1750 m, 19 iv 1990, S. Hurtado / 1973 (MUSENUV); **Cundinamarca**, Fusagasugá, al vuelo, 28 iv 1974, A. Casanova / ICN_055878 (ICN); *idem*, 10 v 1974, O. Rangel / ICN_055854 (ICN); **Meta**, S. Macarena, S. Baja Las Dantas, 23 ix 1987, F.F. / ICN_055869

(ICN); 4♀, **Valle del Cauca**, Cali, 1000 m, 12 i 1991, E. Gaitan / 1965 (MUSENUV); *idem*, Ciudad Univ., 1 x 1977, R. Torres / 1945 (MUSENUV); Chicoral, vegetación, 1900 m, 15 xi 1990 / 1966 (MUSENUV); Tuluá, Mateguadua, 4°1'29.5" N, 76°9'45.4" W, 1127 m, manual, 19 ix 2013, A. F. Gutierrez / 28125 (MUSENUV).

***Cyptocephala* Berg, 1883**

The genus can be recognized by having the mandibular plates surpassing slightly the clypeus; first labial segment entirely between the bucculae; the labium reaching or surpassing the abdominal base; the anterolateral margins of the pronotum angular dorsoventrally, at least posteriorly; the femora are unarmed; the tibiae are sulcate; the abdomen without spiniform processes; and the paramere is bilobed with denticles between the lobes (Rolston 1986).

Cyptocephala antiguensis (Westwood, 1837) [Fig. 89]

Distribution. Southern USA to northern of Peru.

Distribution in Colombia. Meta and Tolima.

Remarks. *Cyptocephala antiguensis* has the gonocoxite VIII conspicuously swollen.

References. Rolston 1986.

Examined material. COLOMBIA; 2♂, **Meta**, Remolinos, Cafam Llanos, 250 m, 29 iii 1996 (MPUJ_ENT); San Martín, Reserva Natural El Caduceo, Km 4.5 a San Francisco, 309 m, 3.665533° N, 73.658316° W, 10-14 iii 2014, C. Oviedo, trampa de luz, manual (MPUJ_ENT); 1♂, **Tolima**, Armero, Bosque Santo Tomás, 350 m, 12 xi, G. Gallego / MPUJ_ENT 0010933 (MPUJ_ENT).

***Nezara* Amyot & Serville, 1843**

The genus can be recognized by having the antennal segment I not surpassing the apex of the head; the abdomen has a medial tubercle, it is reaching the posterior margin of the metacoxae; the spiracles without callose area; the metathoracic peritreme is short (Rolston & McDonald 1980; Torres 2004).

Nezara viridula (Linnaeus, 1758) [Fig. 90]

Distribution. Cosmopolitan.

Distribution in Colombia. Boyacá, Cesar, Guajira, and Magdalena.

Remarks. *Nezara viridula* is the only species of *Nezara* that occurs in America.

References. Rolston & McDonald 1980; Torres 2004.

Examined material. COLOMBIA: 1♀, **Boyacá**, Somondoco, Vda. Boyasegundo, Fca. Bellavista, 4°58' N, 73°27' W, 1500 m, 24 iii 1997, D. Forero / IAvH-E 05084 (IAvH-E).

***Roferta* Rolston, 1981**

The genus can be recognized by having the median tubercle on the abdominal sternite III short, obtuse, scarcely reaching the metacoxae; the scutellum is strongly convex; the antennal segment I not reaching the apex of the head; the bucculae is arcuate anteriorly, weakly produced caudad of arch, evanescent at base of the head; the first rostral segment lying entirely between the bucculae; the mesosternum mildly tumescent on each side of the meson; the median carina is moderately developed, extending to the full length of the mesosternum; the femora is unarmed, and the paramere has a tubercle on the cephalic margin (Rolston & McDonald 1980).

Roferta marginalis (Herrich-Schäffer, 1836) [Fig. 91]

Distribution. Cuba, Honduras, Panama, Colombia, Belize, Trinidad and Tobago, Brazil, and Argentina.

Distribution in Colombia. Meta.

Remarks. *Roferta* is a monotypic genus.

References. Kirkaldy 1909; Barber & Bruner 1946; Rolston & McDonald 1980; Arismendi & Thomas 2003; Grazia & Schwertner 2008; Castro-Huertas *et al.* 2015.

Thyanta Stål, 1862

The genus can be recognized by having the abdominal segment III without medial spiniform process; the peritreme have an elongate sulcus; the mandibular plates and the clypeus nearly equal in length; labium reaching at least the metacoxae; the femora are unarmed; the superior surface of the tibiae is sulcate; and the paramere is rounded and narrow, its apex is acute, sometimes with a lateral spiniform lobe (Rider & Chapin 1991).

Thyanta excavata Rider, 1991

Distribution. Colombia and Venezuela.

Distribution in Colombia. Magdalena.

Remarks. *Thyanta excavata* has the gonocoxite VIII distinctly excavated with the concavity wider than long, and the surface of the gonocoxite VIII is weakly rugose.

References. Rider & Chapin 1991.

Thyanta hamulata Rider, 1991

Distribution. Colombia and Peru.

Distribution in Colombia. Valle del Cauca.

Remarks. *Thyanta hamulata* has the paramere curved forming a hook and with an acute triangular lateral lobe.

References. Rider & Chapin 1991.

Thyanta obtusa Rider, 1991

Distribution. Colombia and Venezuela.

Distribution in Colombia. Magdalena.

Remarks. *Thyanta obtusa* has the posterior margin of the pygophore broadly with U-shaped in caudal view, and the apex of the paramere is rounded.

References. Rider & Chapin 1991.

Thyanta perditor (Fabricius, 1794) [Fig. 92]

Distribution. Southern USA to northern Argentina.

Distribution in Colombia. Antioquia, Cundinamarca, Huila, Magdalena, Meta, Quindío, Risaralda, Tolima, and Valle del Cauca.

Remarks. *Thyanta perditor* has the humeral angles with longer spines, protruding the adjacent corium, and the pronotal reticulation is reduced and restricted to the half nearest of the head.

References. Rider & Chapin 1991.

Examined material. **COLOMBIA:** 1♀, **Huila**, Neiva, 7 ii 1974, AI Bagos (IAvH-E); 1♀, **Meta**, Puerto Gaitán, 21 iii 1993, C. González (IAvH-E); **Quindío**, Filandia, Estación Bremen(C.R.Q), 1900 m, iv 1998 / MPUJ_ENT 0010708 / MPUJ_ENT 0010709 (MPUJ_ENT); **Risaralda**, Pereira, S.F.F. Otún Quimbaya, Est. La Suiza, 1910 m, 4.732977° N, 75.589199° W, 23 iii 1992, Est. Bio / MPUJ_ENT 0010707 (MPUJ_ENT); *idem*, 1900 m, 19 viii 1992 / MPUJ_ENT 0010710–MPUJ_ENT 0010712 (MPUJ_ENT).

Thyanta sinuata Rider, 1991

Distribution. Colombia and Venezuela.

Distribution in Colombia. Magdalena.

Remarks. *Thyanta sinuata* has the posterior margin of the pygophore broadly and sinuously V-shaped in caudal view.

References. Rider & Chapin 1991.

Thyanta straminea Rider, 1991

Distribution. Colombia and Ecuador.

Distribution in Colombia. Valle del Cauca.

Remarks. *Thyanta straminea* has the anterior disk of the pronotum pale yellowish, contrasting with the posterior disk green to brown; and the humeral angles are nearly spinose.

References. Rider & Chapin 1991.

Thyanta testacea (Dallas, 1851)

Distribution. Lesser Antilles and northern South America.

Distribution in Colombia. Chocó, Cundinamarca, Magdalena, and Tolima.

Remarks. *Thyanta testacea* has the apex of the paramere curving gently dorsally and with acute processes.

References. Rider & Chapin 1991.

Examined material. COLOMBIA: 1♂, Chocó, Ríosucio, Cacarica, 3 viii 1978, H.E.E. / ICN_055922 (ICN).

Tribe Pentatomini

Arvelius Spinola, 1837

The genus can be recognized by having the mandibular plates longer than clypeus; the apex of the mandibular plates is acute; the antennal segment I is wide and short; the antennal segment IV is the longest; the pronotum is trapezoidal, with the anterior portion inclined; the anterolateral margins of the pronotum are tuberculated; the humeral angles with acute processes laterally projected; the femora have a distal short spine; the tibiae are sulcate; the mesosternum has a prominent medial carina; and the abdomen ventrally with a medial tubercle (Brailovsky 1981).

Arvelius acutispinus Breddin, 1909

Distribution. Colombia, Venezuela, Peru, Brazil, Bolivia, and Paraguay.

Distribution in Colombia. Amazonas.

Remarks. *Arvelius acutispinus* has the humeral spines conspicuously long and thin, the labium reaching the abdominal sternite IV, and the basal protuberance of the paramere is triangular.

References. Brailovsky 1981.

Arvelius albopunctatus (De Geer, 1773)

Distribution. Southern USA to Argentina.

Distribution in Colombia. Magdalena.

Remarks. *Arvelius albopunctatus* has the paramere bilobate and the antennal segments without a reddish or brown continuous line.

References. Brailovsky 1981.

Arvelius confusus Brailovsky, 1981

Distribution. Colombia and Brazil.

Distribution in Colombia. Santander.

Remarks. *Arvelius confusus* has the humeral spines long and robust, the ventral margin of the pygophore is tetralobate, and the superior margin of the paramere has a medial protuberance.

References. Brailovsky 1981.

Arvelius peruanus Brailovsky, 1981

Distribution. Colombia and Peru.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Brailovsky (1981), but without specific locality.

Remarks. *Arvelius peruanus* has the apex of the scutellum rounded, and the basal region of the paramere with a conspicuous protuberance.

References. Brailovsky 1981.

Arvelius porrectispinus Breddin, 1909 [Fig. 93]

Distribution. Antilles, Colombia, Guyana, and Brazil.

Distribution in Colombia. Chocó, Cundinamarca, Tolima, and Valle del Cauca.

Remarks. *Arvelius porrectispinus* has the humeral angles spines long and acute, the ventral margin of the pygophore not tetralobate, and the superior margin of the paramere with a slightly protuberance.

References. Brailovsky 1981.

Examined material. COLOMBIA: 1♀, **Chocó**, Ríosucio, Cacarica, 3 viii 1978, H.E.E. / ICN_055420 (ICN); 1♂, 1♀, **Tolima**, Ibagué, Vda. El Totumo, 4°23,701' N, 75°11,950' W, 1100 m, 6-9 vii 2013, J.E. Eger & A.A. Calixo (ICN).

Banasa Stål, 1860

The genus can be recognized by having the tubercle on the base of the abdomen in apposition to a notch in the posterior face of the metasternum; the metasternum is flat or sulcate; the mesosternum has a low, obtuse, mesial carina; the first labial segment is between the bucculae; the mandibular plates and clypeus are equal in length; the antennal segment I not reaching the apex of the head; the femora is unarmed; the tarsi is three segmented; and the frenal margin of the scutellum is longer than the apical portion (Thomas & Yonke 1990).

Banasa angulobata Thomas, 1990

Distribution. Colombia, Venezuela, Ecuador, Brazil, Peru, Paraguay, and Argentina.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas & Yonke (1990), but without specific locality.

Remarks. *Banasa angulobata* has the posterolateral angles of the pygophore lobate.

References. Thomas & Yonke 1990.

Banasa bochica Thomas, 1990

Distribution. Colombia.

Distribution in Colombia. Cauca and Valle del Cauca.

Remarks. *Banasa bochica* has the inferior ridge of the pygophore with a pair of robust processes.

References. Thomas & Yonke 1990.

Examined material. COLOMBIA: 1♀, **Valle del Cauca**, Buenaventura, vii 1962, J. Jaillier / MPUJ_ENT 0010722 (MPUJ_ENT).

Banasa centralis Sailer, 1959

Distribution. Mexico, Belize, Guatemala, Costa Rica, Panama, and Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas & Yonke (1988), but without specific locality.

Remarks. *Banasa centralis* has the body entirely dark greenish brown, with a distinctly bicolored pattern on the pronotum, and the proctiger is bilobate.

References. Thomas & Yonke 1988.

Banasa discolor (Dallas, 1851)

Distribution. Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas & Yonke (1990) as the type-locality, but without specific locality information.

Remarks. *Banasa discolor* has dark punctation on the pleura.

References. Thomas & Yonke 1990.

Banasa expallescens Bergroth, 1891

Distribution. Colombia and Venezuela.

Distribution in Colombia. Boyacá and Cundinamarca.

Remarks. *Banasa expallescens* has the apex of the paramere acute. Castro-Huertas *et al.* (2015) cited this species as new record, but it is erroneously referenced because the type-locality is Colombia.

References. Thomas & Yonke 1990.

Examined material. COLOMBIA: 1♀, **Boyacá**, Villa de Leyva, Quebrada La Colorada, 2200 m, 13 ix 1996, M. Galvis / IAvH-E 05104 (IAvH-E); 1♀, **Cundinamarca**, Cáqueza, 29 xii 1982, P. Moreno / ICN_055734 (ICN).

Banasa excavata Thomas, 1988

Distribution. Guatemala, El Salvador, Costa Rica, and Panama. New record for Colombia.

Distribution in Colombia. Caquetá.

Remarks. *Banasa excavata* has the body longer than 10 mm, and the gonocoxite VIII is weakly emarginate.

Examined material. COLOMBIA: 1♀, **Caquetá**, PNN Los Picachos, 2°47'51" N 74°51'18" W, 1,560m. Trampa de luz, 20 xi 1997, F. Escobar.leg / IAvH-05089 (IAvH).

References. Thomas & Yonke 1988.

Banasa fulgida Thomas, 1990

Distribution. Colombia, Venezuela, and Brazil.

Distribution in Colombia. Magdalena.

Remarks. *Banasa fulgida* has the mandibular plates impunctate or with very few punctations, and clypeus with at most one or two punctations.

References. Thomas & Yonke 1990; Castro-Huertas *et al.* 2015.

Banasa gynamagna Thomas, 1990

Distribution. Colombia and Venezuela.

Distribution in Colombia. Cauca and Putumayo.

Remarks. *Banasa gynamagna* has the ventral rim of the pygophore quadrate in outline and the gonocoxite VIII is unusual elongate.

References. Thomas & Yonke 1990.

Examined material. COLOMBIA: 1♂, **Putumayo**, Sibundoy, Quebrada la Hidráulica, 1°12' N, 76°55' W, 2100 m, 17 iv 1994, F. Fernández / IAvH-E 05101 (IAvH-E).

Banasa panamensis Sailer, 1957

Distribution. Central America, Colombia, Venezuela, Guyana, French Guiana, and Peru.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas & Yonke (1990), but without specific locality.

Remarks. *Banasa panamensis* has small size; the labium is long, reaching the posterior margin of the tubercle-bearing sternite in repose; the pronotal margin is rectilinear in dorsal view, not impressed dorsally; the humeral angles are not produced; and the abdominal tubercle is a broad protuberance, not conical.

References. Thomas & Yonke 1990.

Banasa paraexpallescens Thomas, 1990

Distribution. Colombia and Venezuela.

Distribution in Colombia. Boyacá, Chocó, Cundinamarca, and Magdalena.

Remarks. *Banasa paraexpallescens* is similar to *B. expallescens*, but *B. paraexpallescens* has the apex of the paramere obtuse, not acute.

References. Thomas & Yonke 1990, Castro-Huertas *et al.* 2015.

Examined material. COLOMBIA: 1♀, **Cundinamarca**, Albán, 1700 m, 13 xi 1981, Schmidh, MPUJ_ENT 0010720 (MPUJ_ENT).

Banasa peruana Thomas, 1990

Distribution. Colombia, Ecuador, Brazil, Peru, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Boyacá.

Remarks. *Banasa peruana* has a body longer than 10 mm, usually with a discolored yellow band running through the apex of the hemelytra and scutellum, without stigmatose spots on the pleura, the anterior and posterior portions of the pronotum are obscurely intergrading, and the gonocoxite VIII is emarginate.

References. Thomas & Yonke 1990; Dellapé 2021a.

Examined material. COLOMBIA: 1♂, **Boyacá**, Villa de Leyva, 5°38' N, 73°31' W, manual, 10 ix 1998, F. Fernández / IAvH-E 05087 (IAvH-E).

Banasa saileri Thomas, 1990 [Fig. 94]

Distribution. Venezuela. New record for Colombia.

Distribution in Colombia. Valle del Cauca.

Remarks. *Banasa saileri* has the pronotum not distinctly bicolored and the abdominal margin is uniformly pale.

References. Thomas & Yonke 1990.

Examined material. COLOMBIA: 1 ♀, **Valle del Cauca**, Buenaventura, Bajo Calima, 70m. 23 iii 1995, Adriana Marlene / MUJ 1684 (MPUJ_ENT).

Banasa salvini Distant, 1911

Distribution. Mexico, Guatemala, Belize, Honduras, Costa Rica, Panama, Colombia, and Venezuela.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas & Yonke (1988, 1990), but without specific locality

Remarks. *Banasa salvini* has large size, the corium is green, and the head is dorsally pink.

References. Thomas & Yonke 1988, 1990.

Banasa subrufescens Stål, 1860

Distribution. Colombia, Venezuela, Surinam, Guyana, Brazil, and Argentina.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas & Yonke (1990), but without specific locality.

Remarks. *Banasa subrufescens* has small size, the pleura dark punctate, and the anterolateral pronotal margins are dorsally impressed.

References. Thomas & Yonke 1990; Dellapé *et al.* 2020.

Banasa varians Stål, 1872

Distribution. Brazil and Argentina. New record for Colombia.

Distribution in Colombia. Boyacá.

Remarks. *Banasa varians* has size small, the labium not reaching the abdominal tubercle in repose, the pygophore on each side with an erect, digitlike tooth on the posterior margin.

References. Thomas & Yonke 1990; Dellapé *et al.* 2020.

Examined material. COLOMBIA: 1 ♂, **Boyacá**, Pajarito, Comisoque, 2000 m, 10 ii 1980, A Fajardo (ICN).

Banasa zeteki Sailer, 1959

Distribution. Cuba, Mexico, Belize, Dominican Republic, Guatemala, Honduras, Costa Rica, Nicaragua, Panama, and Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Thomas & Yonke (1988, 1990), but without specific locality

Remarks. *Banasa zeteki* has the propleura dark punctate, the paramere is bent angularly, and the gonocoxite VIII is emarginate.

References. Thomas & Yonke 1988, 1990.

***Boea* Walker, 1867**

The genus can be recognized by having the lateral margins of the mandibular plates strongly reflexed, the antennae are four segmented, and the labium reaching the metacoxae (Rolston & McDonald 1984).

Boea sp. [Fig. 95]

Distribution of the genus. Costa Rica, Colombia, and Brazil.

Distribution in Colombia. Valle del Cauca.

Remarks. *Boea* was recorded for Colombia by Castro-Huertas *et al.* (2015) and noted the impossibility of the identification to species level. The specimen examined by them is very similar to *B. costaricensis* Distant, 1890.

References. Kirkaldy 1909; Rolston & McDonald 1984; Castro-Huertas *et al.* 2015.

***Elsiella* Froeschner, 1981**

The genus can be recognized by having the elevated metasternum; the peritreme in a spout; the median spine on the abdominal segment III has a right or obtuse angle; the ventral rim of the pygophore is dorsally projected; the superior ridge of the ventral rim of the pygophore with a pair of projections; and the paramere is thin with a long and curved projection (Simões *et al.* 2012).

Elsiella plana (Walker, 1867) [Fig. 96]

Distribution. Colombia and Ecuador.

Distribution in Colombia. Cauca.

Remarks. *Elsiella* is a monotypic genus.

References. Simões *et al.* 2012.

***Neotibilis* Grazia & Barcellos, 1994**

The genus can be recognized by the protuberant eyes; the labium reaching the mesocoxa; the abdominal spine reaching the procoxa, and the body is rounded (Grazia & Barcellos 1994).

Neotibilis parva (Distant, 1893) [Fig. 97]

Distribution. Panama, Colombia, Venezuela, and Brazil.

Distribution in Colombia. Magdalena.

Remarks. *Neotibilis parva* has the apex of the scutellum without callous and the ventral margin of the pygophore without foldings.

References. Grazia & Barcellos 1994.

***Pellaea* Stål, 1872**

The genus can be recognized by having the oval body, slightly convex; the pronotum with punctations in transversal, vermiform lines, and separated by smooth and callosity regions; the first antennal segment not reaching the apex of the head; the clypeus is nearly equal or slightly longer than the mandibular plates; and the abdominal medial tubercle not reaching the procoxae, sometimes reduced (Brailovsky 1987; Rolston & McDonald 1980).

Pellaea stictica Dallas, 1851 [Fig. 98]

Distribution. USA, Mexico, Honduras, Nicaragua, Costa Rica, Panama, Colombia, Venezuela, Guyana, Ecuador, Peru, Brazil, Chile, Paraguay, and Argentina.

Distribution in Colombia. Valle del Cauca.

Remarks. *Pellaea stictica* has the pygophore with the concavity on each side interrupted by an obtuse and diagonal carina, and the paramere with processes.

References. Kirkaldy 1909; Rolston & McDonald 1980; Rolston 1984; Silva *et al.* 2018; Dellapé 2021a.

Examined material. COLOMBIA: 4♂, 3♀, **Valle del Cauca**, Cali, 1000 m, 1982, Naranjo / 1932 (MUSENUV); *idem*, 1979 / 1931 (MUSENUV); *idem*, Tierra cult., 1 x 1970 / 1952 (MUSENUV);

Ca. Cali, árbol, 27 xii 1991, S.Villamizar / 1935 (MUSENUV); Palmira, 1001 m, 17 iii 2001, C. Vargas / 20970 (MUSENUV); Tuluá, Mateguadua, 1 xi 1985, Murillo / 1930 (MUSENUV); *idem*, maleza, 1000 m, 1 xi 1991, S.

Mosquera / 1933 (MUSENUV).

***Phalaecus* Stål, 1862**

The genus can be recognized by having the first labial segment is between the bucculae; the pro-, meso- and metasternum are elevated; the mesosternal carina is anteriorly narrow and reaching the procoxae; the metasternum

is hexagonal, its anterolateral margins are straight, and its posterior margin is conspicuously concave; the medial abdominal tubercle is insert in the posterior margin of the metasternum; the apex of the metafemur without spine; the tarsi are bi-segmented; and the body has a colorful coloration pattern (Grazia 1983)

Phalaeus sp. [Fig. 99]

Distribution of the genus. Guyana, French Guiana, Surinam, and Brazil.

Distribution in Colombia. Amazonas.

Remarks. *Phalaeus* was recorded for Colombia by Castro-Huertas *et al.* (2015), and noted the impossibility of the identification to species level. The examined specimen did not fit any known species of the genus.

References. Grazia 1983; Castro-Huertas *et al.* 2015.

***Pharypia* Stål, 1861**

The genus can be recognized by having large size and typical coloration pattern; the mandibular plates are shorter or equal than clypeus; the first labial segment surpassing the bucculae; the mesosternal carina reaching the procoxae; the labium reaching or surpassing the abdominal segment III; and the abdominal spine is developed, concealed by the posterior margin of the metasternum (Brailovsky 1987; Rolston *et al.* 1980; De los Santos *et al.* 2016).

Pharypia nitidiventris (Stål, 1861)

Distribution. Mexico, Guatemala, Honduras, Nicaragua, Costa Rica, Panama, Colombia, Venezuela, and Brazil.

Distribution in Colombia. Unknown.

Remarks. *Pharypia nitidiventris* has the anterolateral pronotal margins reflected and the particular coloration pattern orange, white, and black.

References. Kirkaldy 1909; Rolston *et al.* 1980; Cervantes-Peredo, 2013.

Pharypia pulchella (Drury, 1782) [Fig. 100]

Distribution. Mexico, Guatemala, Dominican Republic, Puerto Rico, Honduras, Costa Rica, Panama, Colombia, Venezuela, and Brazil.

Distribution in Colombia. Amazonas.

Remarks. *Pharypia pulchella* has a unique coloration pattern: blue greenish metallic, with red and black maculae. Some specimens have a black and yellow pattern, resembling the genus *Arocera*, but the species of *Pharypia* can be identified by the metasternum and the abdominal spine developed.

References. Kirkaldy 1909; Rolston *et al.* 1980; De los Santos *et al.* 2016.

Examined material. COLOMBIA: 2♀, Amazonas, PNN Amacayacu, río Cotuhe, Caño Lorena, 6 viii 1990, F. Fernández / IAvH-E 05032 (IAvH-E); *idem*, 1 vi 1990, F. Fernández / IAvH-E 05031 (IAvH-E).

***Placocoris* Mayr, 1864**

The genus can be recognized by having the body dorsoventrally depressed, and at least the metafemur with two rows of spines on its inferior surface (Rolston *et al.* 1980).

Placocoris sp. [Fig. 101]

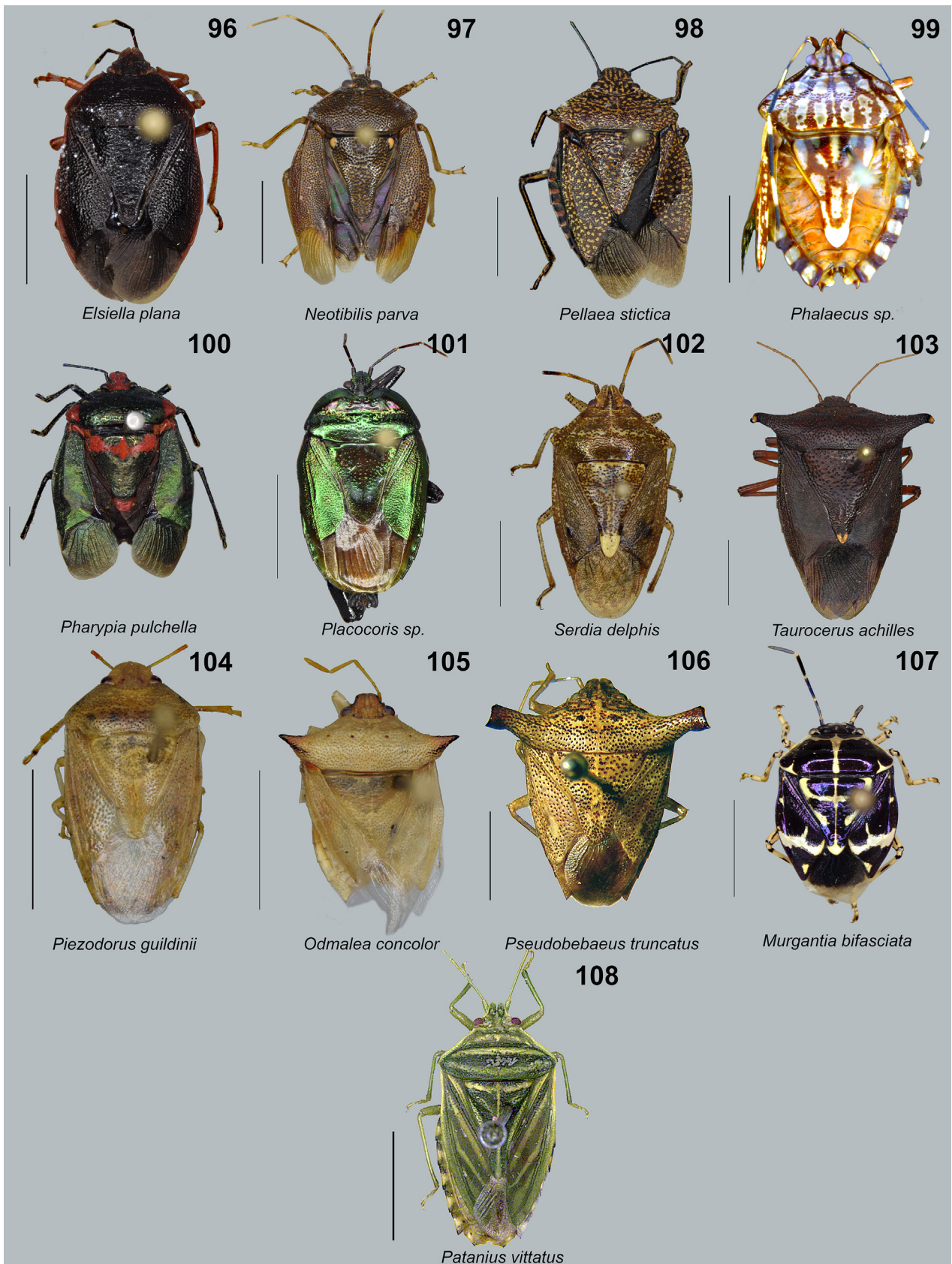
Distribution of the genus. Brazil, Peru, Paraguay, and Argentina. New record for Colombia.

Distribution in Colombia. Caquetá.

Remarks. The specimens examined did not fit any known species of the genus. Thomas (2021) includes this genus in Ochlerini (Discocephalinae) based on the trichobothrial arrangement but without testing these (and other) characters in a cladistic analysis. Because of the uncertain position of *Placocoris*, we decide to maintain it within Pentatomini in this study.

References. Walker 1867; Pirán 1948; Rolston *et al.* 1980; Grazia & Schwertner 2008; Thomas 2021.

Examined material. COLOMBIA: 1♂, 1♀, 2 nymphs, Caquetá, Centro de Investigación Macagual, 01.5014000° N 75.6632833° W, 246 m, 13-17 vii 2013, D. Forero / Ex. *Guadua* cf. *angustifolia* Kunth (MPUJ_ENT).



FIGURES 96–108. Pentatominae genera in Colombia. 96. *Elsiella plana* (MPUJ). 97. *Neotibilis parva* (MPUJ). 98. *Pellaea stictica* (MPUJ). 99. *Phalaeus* sp. (IAVH). 100. *Pharypia pulchella* (MPUJ). 101. *Placocoris* sp. (MPUJ). 102. *Serdia delphis* (MPUJ). 103. *Taurocerus achilles* (MPUJ). 104. *Piezodorus guildinii* (MPUJ). 105. *Odmalea concolor* (MPUJ). 106. *Pseudobebaeus truncatus* (UFRG). 107. *Murgantia bifasciata* (ICN). 108. *Patanius vittatus* (MZUSP). Scale bar: 5 mm.

***Serdia* Stål, 1860**

The genus can be recognized by having the first labial segment slightly surpassing the bucculae; the mesosternum is carinate, which usually is projected reaching the procoxae; the labium reaching the mesocoxae, and the metasternal region is elevated, with the posterior margin opposed to the abdominal tubercle (Rolston *et al.* 1980).

Serdia concolor Ruckes, 1958

Distribution. Colombia, Venezuela, Brazil, Bolivia, Paraguay, and Argentina.

Distribution in Colombia. Risaralda.

Remarks. *Serdia concolor* has the pygophore with a medial small tooth-shape process in ventral view.

References. Fortes & Grazia 2005; Castro-Huertas *et al.* 2015.

Serdia delphis Thomas & Rolston, 1985 [Fig. 102]

Distribution. Colombia, Peru, Ecuador, and Bolivia.

Distribution in Colombia. Santander and Valle del Cauca.

Remarks. *Serdia delphis* has the paramere with two processes, a lateral process lobate and an apical long and curve; and the posterior margin of the gonocoxite IX is convex.

References. Fortes & Grazia 2005.

Examined material. COLOMBIA: 1♀, Valle del Cauca, Palmira, 965 m, 7 xi 2005, Abadia (MUSENUV).

***Taurocerus* Amyot & Serville, 1843**

The genus can be recognized by having the first antennal segment not surpassing the apex of the head; the mandibular plates shorter than the clypeus; the labium reaching the abdominal sternite III to IV; the first labial segment surpassing the bucculae; the bucculae is anteriorly truncate and posteriorly lobed; the humeral angles are projected in strong spines with the apex curve and backward; the femora distally with short spines, one dorsally and a pair laterally; the tibiae is dorsally sulcate; the mesosternum is slightly more elevated than the metasternum; the abdominal sternites have a medial, longitudinal kneel; and the abdominal sternite III has a medial tubercle opposed to posterior margin of the metasternum (Grazia & Barcellos 2005).

Taurocerus achilles Stål, 1862 [Fig. 103]

Distribution. Colombia and Brazil. The record in Mexico is doubtful.

Distribution in Colombia. Amazonas.

Remarks. *Taurocerus achilles* has the body brown, and the humeral angles with the apex black and somewhat curved directed anterolaterally.

References. Thomas 2000; Grazia & Barcellos 2005; Castro-Huertas *et al.* 2015.

Taurocerus edessoides (Spinola, 1837)

Distribution. Mexico, Guatemala, Costa Rica, Panama, Colombia, Guyana, Ecuador, Brazil, Peru, Bolivia, and Argentina.

Distribution in Colombia. Amazonas and Meta.

Remarks. *Taurocerus edessoides* has the apex of the humeral angles yellowish, the ventral rim of the pygophore with a conspicuous and median tubercle, the lateromedial processes of the pygophore are conical, and the paramere is strongly curved ventrally.

References. Grazia & Barcellos 2005; Dellapé *et al.* 2015.

Tribe Piezodorini

***Piezodorus* Fieber, 1860**

The genus can be recognized by having the first antennal segment not surpassing the apex of the head; the meso-

ternum has a conspicuous carina, flattened in its lateral margins; and the abdominal venter with a medial tubercle reaching the metacoxae (Rolston & McDonald 1980).

Piezodorus guildinii (Westwood, 1837) [Fig. 104]

Distribution. USA, Mexico, Cuba, Haiti, Jamaica, Santo Domingo, Trinidad and Tobago, Guatemala, Honduras, Nicaragua, Costa Rica, Panama, Colombia, St. Vincent, Grenada, Brazil, Paraguay, Uruguay, and Argentina

Distribution in Colombia. Antioquia, Caldas, Caquetá, Cundinamarca, Magdalena, Meta, Tolima, and Valle del Cauca.

Remarks. *Piezodorus guildinii* is the only species of genus that occur in America.

References. Kirkaldy 1909; Torres 2004; Dellapé 2021a.

Tribe Procliticini

Odmalea Bergroth, 1914

The genus can be recognized by having the head wider across the eyes than long; the lateral margins of the head are sigmoid and tapering to apex; the mandibular plates are usually separate at apex; the first antennal segment not surpassing the apex of the head; the antennal segment I and II shorter than III to V; the labium reaching the metacoxae; the humeral angles are produced in a spine; the prosternum and metasternum are flat or nearly so; the mesosternum is weakly tumescent behind the procoxae; the opening of scent efferent system straight or slightly curved, and the abdominal ventral spine reaching the metacoxae or sometimes the mesocoxae (Rolston 1978c; Rolston & McDonald 1980).

Odmalea concolor (Walker, 1867) [Fig. 105]

Distribution. Honduras, Panama, Colombia, French Guiana, Trinidad and Tobago, and Brazil.

Distribution in Colombia. Meta.

Remarks. *Odmalea concolor* has the propleura almost uniformly pale colored, and the paramere has digitiform processes on the apex.

References. Rolston 1978c; Arismendi & Thomas 2003; Castro-Huertas *et al.* 2015.

Odmalea vega Rolston, 1978

Distribution. Colombia.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Rolston (1978c) as the type-locality, but without specific locality information.

Remarks. *Odmalea vega* has the propleura bicolored, and the margin of the genital cup of the pygophore depressed directly caudad from the paramere.

References. Rolston 1978c.

Pseudobebaeus Distant, 1911

The genus can be recognized by having the humeral angles with prominent processes, and the costal angle of the corium is extending caudally, surpassing the apex of the scutellum (Rolston & McDonald 1980).

Pseudobebaeus truncatus (Fallou, 1888) [Fig. 106]

Distribution. Brazil and Peru. New record for Colombia.

Distribution in Colombia. Meta.

Remarks. *Pseudobebaeus truncatus* has the body brown yellowish, irregularly with dark brown to black punctate, rather thickly so on dorsum.

References. Rolston & McDonald 1980; Rider 1994; Rider & Fischer 1998.

Examined material. COLOMBIA: 1♀, Meta, Río Ocoa, 350m, vii 1948, Silv. / ICN He 00408 (UFRG).

Tribe Strachiini

Murgantia Stål, 1862

The genus can be recognized by the apex of the first labial segment surpassing the bucculae by a distance subequal to the labial width at posterior margin of the bucculae; and the body with brightly coloration (Brailovsky & Barrera 1989).

Murgantia bifasciata (Herrich-Schaeffer, 1836) [Fig. 107]

Distribution. Costa Rica, Colombia, Brazil, and Argentina.

Distribution in Colombia. Risaralda and Valle del Cauca.

Remarks. *Murgantia bifasciata* has the anterior margin of scutellar disk with a transversal band yellow, and the lateral margins of the pygophore are excavated.

References. Kirkaldy 1909; Jensen-Haarup, 1926; Rolston & McDonald 1984; Brailovsky & Barrera 1989; Castro-Huertas *et al.* 2015.

Murgantia varicolor (Westwood, 1837)

Distribution. USA, Mexico, Panama, Colombia, and Brazil.

Distribution in Colombia. Unknown. This species was recorded for Colombia by Kirkaldy (1909), but without specific locality.

Remarks. *Murgantia varicolor* has the head dark dorsally with five pale spots, a pair oval on the mandibular plates, and a pair elongated on the anteocular region, and one longitudinal medial; the pronotum is dark with the lateral margins pale, a longitudinal stripe pale and a pair of the transversal stripes pale on anterior and posterior portion respectively.

References. Kirkaldy 1909; Brailovsky & Barrera 1989.

Murgantia violascens (Westwood, 1837)

Distribution. Jamaica, Colombia, and Brazil.

Distribution in Colombia. Bolívar.

Remarks. *Murgantia violascens* has the membrane of hemelytron brownish with blue-green metallic reflexes; the gonocoxite VIII is conspicuously expanded almost covering the gonoplac; and the lateral margins of the pygophore is excavated.

References. Kirkaldy 1909; Brailovsky & Barrera 1989; Castro-Huertas *et al.* 2015.

Unplaced

Patanius Rolston, 1987

The genus can be recognized by having the mandibular plates surpassing clypeus, the antennae is four segmented; the antennal segment I surpassing the apex of the head; the first labial segment surpassing the bucculae; the labium surpassing the procoxae but not attaining the mesocoxae; the superior surface of the pro-, meso- and metafemur unarmed apically; the prosternum is nearly flat; the mesosternum somewhat tumid and scarcely carinate medially; the base of the abdomen without processes; and the parameres are absent (Rolston 1987).

Patanius vittatus Rolston, 1987 [Fig. 108]

Distribution. Colombia and Brazil.

Distribution in Colombia. Meta.

Remarks. Monotypic genus.

References. Rolston 1987; Castro-Huertas *et al.* 2015.

Discussion

Colombia as a key area to understand diversity pattern of the Neotropical stink bug fauna.

With the data presented in this paper, Colombia represent the fifth most diverse country in the Neotropical region with respect to stink bug diversity, with about 15% of the Pentatomidae species found in the Neotropics (Table 1). The present survey also shows that at least 7% of the species recorded from Colombia are endemic, and similar or higher percentages are recorded in other biological groups such as plants, butterflies, mammals, or birds (e.g., Andrade 2011). Although endemism estimation depends on unique criteria for each taxonomic group, the consistent values of endemic species percentages may suggest a key role the area has played in the diversification, in this case on the Pentatomidae fauna in the Neotropical region. The current biota composition in Colombia, including the stink bug fauna, could be explained considering the interaction of several factors, including the geographical position and the geological history (in particular during the Paleogene-Pleistocene), that have allowed the dispersal, exchange, and speciation of the biota (Rangel-Ch 2015). The interaction of these biotic and abiotic factors in the Neotropics have been documented and implicated in the diversification of several taxa (e.g., Kattan *et al.* 2004; Cadena *et al.* 2016; Torres & Rojas 2021). In order to understand the biogeographical patterns found in the Neotropics, the taxonomic and distributional data from Colombia plays a critical role. In the following, we briefly outline the main factors that had probably played a role in its configuration (e.g., Hoorn *et al.* 2010; Rull 2011; Antonelli *et al.* 2018), all of which should be thoroughly explored and tested in future investigations including the Pentatomidae fauna.

TABLE 1. Comparative genera and species richness for the World, Neotropics and Colombia, and proportion of Colombian fauna*. G, number of genera; S, number of species. Subfamilies in bold.

Subfamily	World		Neotropics		Colombia		% spp.	
	G	S	G	S	G	S	W	Neot
Asopinae	63	303	23	113	15	32	9,9%	26,5%
Cyrtocorinae	4	11	4	11	2	4	36,4%	36,4%
Discocephalinae	81	325	81	325	31	55	16,6%	16,6%
Discocephalini	46	195	46	195	15	31	15,9%	15,9%
Ochlerini	35	130	35	130	15	23	17,7%	17,7%
Edessinae	12	338	15	337	8	20	4,7%	4,7%
Pentatominae	660	3484	136	816	53	136	3,9%	16,7%
Aelini	4	28	1	1	0	0	0,0%	0,0%
Agonoscelidini	1	26	1	1	0	0	0,0%	0,0%
Bathycoelini	1	32	1	2	0	0	0,0%	0,0%
Cappaeini	24	151	1	1	0	0	0,0%	0,0%
Carpocorini	127	503	53	277	22	50	9,7%	17,7%
Catacanthini	11	63	5	36	4	14	22,2%	38,9%
Chlorocorini	8	77	8	75	4	9	11,7%	12,0%
Halyini	91	430	2	24	0	0	0,0%	0,0%
Mecideini	1	16	1	3	0	0	0,0%	0,0%
Menidini	28	164	2	23	2	4	1,8%	13,0%
Nezarini	26	272	8	127	5	20	7,4%	15,7%
Pentatomini	56	316	32	187	11	33	10,4%	17,6%
Piezodorini	5	21	1	1	1	1	4,8%	100,0%
Procliticini	11	36	11	35	2	3	8,3%	8,6%
Sciocorini	18	126	3	5	0	0	0,0%	0,0%
Strachiini	20	142	3	14	1	3	2,1%	21,4%
Unplaced	29	108	3	4	1	1	0,9%	25,0%
Podopinae	64	269	3	3	0	0	0,0%	0,0%
TOTAL	940	4.949	259	1.602	108	246	4,9%	15,3%

*Data compiled from Rider 2018 (World) and Grazia *et al.* 2015 (Neotropics).

TABLE 2. Comparative number of genera and species of Pentatomidae in the countries of Neotropical region with available data, in descending order of species diversity*. G, number of genera; S, number of species. Subfamilies in bold.

Subfamily	Brazil		Mexico		Costa Rica		Argentina		Panama		Honduras		Venezuela		Ecuador		Uruguay		Hispaniola		Nicaragua		Puerto Rico		Chile			
	G	S	G	S	G	S	G	S	G	S	G	S	G	S	G	S	G	S	G	S	G	S	G	S	G	S		
Asopinae	18	46	15	34	11	26	13	30	11	22	13	24	7	10	8	11	9	15	6	8	5	5	6	6	7	6	8	
Cyrtocorinae	4	9	2	2	1	3	1	5	2	3	0	0	2	3	1	1	0	0	0	0	0	0	0	0	0	0	0	
Discocephalinae	49	161	13	20	20	31	14	21	34	48	12	20	12	21	13	14	4	4	1	1	1	5	6	1	1	0	0	
Discocephalini	32	119	8	13	12	21	10	16	19	30	7	12	11	20	9	10	3	3	0	0	3	3	0	0	0	0	0	
Ochlerini	17	42	5	7	8	10	4	5	15	18	5	8	1	1	4	4	1	1	1	1	2	3	1	1	1	0	0	
Edessinae	6	135	5	52	5	90	6	28	8	47	5	28	5	9	3	32	2	3	2	9	2	9	2	2	2	0	0	
Pentatominae	85	327	51	208	46	116	60	158	45	106	43	108	46	107	28	43	36	72	25	57	30	51	24	45	13	31		
Aelini	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Agonoscelidini	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	
Bathycelini	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Cappaeni	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	
Carpocorini	35	122	13	71	12	37	25	76	12	30	10	39	16	27	7	14	18	37	7	19	7	24	6	11	4	11		
Catacanthini	4	14	4	7	4	10	4	7	3	9	3	8	4	9	1	3	3	3	2	2	2	3	3	3	3	0	0	
Chlorocorini	5	33	3	13	3	8	3	7	5	11	4	8	5	10	2	3	3	5	2	6	3	4	3	4	3	4	1	1
Halyini	0	0	2	19	1	1	0	0	0	0	2	2	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	
Mecideini	0	0	1	2	1	2	1	1	0	0	1	2	1	1	0	0	1	1	1	1	1	1	2	1	1	0	0	
Memidini	2	13	1	3	1	2	1	1	1	1	1	3	1	5	0	0	0	0	0	0	0	0	0	0	0	0	0	
Nezarini	5	43	6	30	5	11	5	25	5	12	5	8	3	17	2	10	5	14	5	13	4	6	4	8	3	12		
Pentatomini	24	84	13	41	16	40	13	29	13	32	13	32	13	35	5	6	3	5	3	10	7	7	7	4	5	1	1	
Piezodorini	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	
Prolecticini	5	12	2	11	1	2	6	9	1	1	1	2	1	1	0	0	2	5	1	2	1	1	1	1	1	1	1	
Scitocorini	0	0	1	1	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	0	0	0	0	0	1	3	
Strachini	1	2	2	7	1	2	1	2	1	1	1	2	1	1	0	0	0	0	0	1	1	1	1	0	0	1	1	
Unplaced	2	2	1	1	0	0	0	0	1	2	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
Podopinae	0	0	2	2	1	1	1	1	0	0	0	0	0	0	0	0	1	1	1	1	1	0	0	0	0	1	1	
TOTAL	162	678	88	318	84	267	105	279	100	226	73	180	72	150	53	101	52	95	35	76	42	71	33	55	20	40		

*The following references were used to compile the data by country: Brazil—Grazia *et al.* 2020; Mexico—Thomas 2000; Costa Rica—Lewis and Schwertner (unpublished); Argentina—Coscarón 2017, Dellapé *et al.* 2020, Dellapé 2021a; Panama—Froeschner 1999, Cambra *et al.* 2018; Honduras—Arimendi & Thomas 2003; Venezuela—Becker and Grazia 1971, Grazia 1984, Schwertner (unpublished); Ecuador—Froeschner 1981; Uruguay—Casini and Grazia 1973, Schwertner (unpublished); Haiti and Dominican Republic—Pérez-Gelabert and Thomas 2005, Pérez-Gelabert 2008; Nicaragua—Maes 1994; Puerto Rico—Segarra-Carmona *et al.* 2016; Chile—Prado 2008, Faundez & Carvajal 2011, Faundez & Rider 2014.

1) Geographic complexity. Colombia has a particular geological history that has resulted in a diverse and complex configuration of the landscape. The main salient feature is the presence of the Andes, which in Colombia is divided into three cordilleras, forming two large inter-Andean valleys (Ocampo-Pérez *et al.* 2007; Antonelli *et al.* 2009). The altitude variation of the landscape allowed for the consolidation of different climatic zones, to which the biota responded differently, having thus contrasting ecosystems such as paramo, tropical dry forests, and tropical humid forests. This complexity offers diverse habitats and adequate conditions for different biological groups. Such is the ecosystem variation that Colombia is cataloged as a megadiverse country, being one of the top 10 most biodiverse countries in the world (Ocampo-Pérez *et al.* 2007; Andrade-C 2011). Because of this geographic complexity, Colombia is usually divided into five main regions: Amazon, Andes, Caribbean, Orinoco, and Pacific (Rangel-Ch 2015; but see also Morrone 2017).

2) Localization. Colombia is located in the northwestern portion of South America, which after the closure of the Panamanian land bridge allowed a biotic exchange between Central and South America (Marshall *et al.* 1982; Cody *et al.* 2010). Although the details of the geological timing and sequence leading to this biotic exchange are still controversial (Montes *et al.* 2015; O’Dea *et al.* 2016; Jaramillo *et al.* 2017), it is clear that this geological event affected profoundly the insect faunal composition present nowadays in South America (McCafferty 1998; Wilson *et al.* 2014). Different groups of organisms are known to have restricted distributions in this area, such as those only present in Panama and Colombia or restricted to the Caribbean coast of lower Central America and South America (Morrone 2017); but many other patterns probably lack adequate documentation.

3) Ensemble of endemic areas. Areas of endemism are the result of particular geological and biological interactions, processes that have been particularly important in the tropical Andes (Hazzi *et al.* 2018). The study of Ferrari *et al.* (2010) on areas of endemism for Neotropical Pentatomidae shows that at least seven of the areas of endemism identified by them partially include areas in Colombia, several of which are shared with other areas in Central and South America. This ensemble of different endemic areas in Colombia are related to the two aforementioned factors (geographic complexity and localization), thus the high species diversity found in the country resulted from both local speciation processes and dispersal events from other regions or areas. Future research including the fauna of stink bugs from Colombia will greatly improve our understanding of the evolution of Neotropical areas.

Neotropical biogeographic patterns of the Pentatomidae fauna in Colombia.

This study does not pretend to be a formal biogeographic analysis of the Pentatomidae fauna of Colombia. Nonetheless, the distribution of species recorded in Colombia offers valuable information about biogeographical patterns in the Neotropical region. The Colombian stink bug fauna can be organized into five major groups: 1) widespread in the Neotropics (distributed from Mexico to Argentina); 2) Northern Neotropics (distributed from Mexico to northern of South America, including the Caribbean); 3) Northern South America (distributed in Colombia and at least one of the following countries: Bolivia, Brazil [northern States], Ecuador, French Guiana, Guyana, Panama, Peru, Surinam, and Venezuela); 4) widespread in South America (distributed in Colombia, northern countries, and at least one of the following southern countries: Argentina, Brazil [southern States], Paraguay, and Uruguay; and 5) Endemics (species recorded only from Colombia). These distribution patterns are congruent with previous and current regionalization of the Neotropical region into biogeographic units (i.e., Morrone 2017), also demonstrated in Ferrari *et al.* (2010) for the stink bug fauna of the Neotropics. Interestingly, approximately half of the species recorded for Colombia (45%) are only known from Northern South America, and nearly 25% distributed in the Northern Neotropics. Widespread species, found in both the entire Neotropics or in all South America, represent 23% of the species (Table 3).

Stink bugs of agricultural importance.

Species of some genera such as *Antiteuchus*, *Chinavia*, *Edessa*, *Euschistus*, *Lincus*, *Diceraeus*, *Nezara* and *Piezodorus* are recognized pests of agricultural importance in different countries of the region (Grazia *et al.* 2015; Grazia & Schwertner 2017). Asopines species of *Alcaeorrhynchus*, *Oplonus*, *Podisus* and *Supputius* are successfully used as biological control agents in different crops (De Clercq 2008; Pires *et al.* 2015). Several other species around the world, both phytophagous and predators, are considered of potential of economic importance, and more recently invasive species are spreading in different countries in the Americas (McPherson 2018). In Colombia, stink bugs pests attack important crops such as beans, cocoa, corn, fruits, rice, soybean, tobacco and vegetables (e.g., Posada-Ochoa 1989). Nevertheless, our results show that knowledge on this import group of insects is still limited in

TABLE 3. Distribution patterns in the Neotropical region of the Colombian stink bug fauna. Widespread Neotropics = Mexico to Argentina; Northern Neotropics = Mexico to northern South America; Northern South America = distributed in Colombia and at least one of the following countries: Bolivia, Brazil (northern States), Ecuador, French Guiana, Guyana, Peru, Surinam, Venezuela; Widespread South America = distributed in Colombia and at least one of the following countries: Argentina, Brazil (southern States), Paraguay, Uruguay; Endemics = recorded only in Colombia.

Widespread (Neotropics) [30 spp.]	Northern Neotropics [59 spp.]	Northern South America [104 spp.]	Widespread (South America) [22 spp.]	Endemics [18 spp.]
<i>Alcaeorrhynchus grandis</i>	<i>Andrallus spinidens</i>	<i>Coryzorhaphis carneolus</i>	<i>Discocera coccinea</i>	<i>Herrichella thoracica</i>
<i>Podisus aenescens</i>	<i>Apateticus lineolatus</i>	<i>Coryzorhaphis superba</i>	<i>Heterocelis servillei</i>	<i>Linacus punctatus</i>
<i>Podisus fuscescens</i>	<i>Coryzorhaphis cruciare</i>	<i>Oplonus salamandra</i>	<i>Oplonus festivus</i>	<i>Linacus substyliger</i>
<i>Tylospilus cloelia</i>	<i>Euthyrhynchus floridanus</i>	<i>Podisus cornutus</i>	<i>Podisus nigripinus</i>	<i>Melambyrsus hoplita</i>
<i>Cyrtocoris egeris</i>	<i>Heteroscelis lepida</i>	<i>Podisus crassimargo</i>	<i>Stiretrus cinctellus</i>	<i>Neadoxoplays longirostra</i>
<i>Cyrtocoris gibbus</i>	<i>Podisus nigriventris</i>	<i>Podisus tinctus</i>	<i>Tynacantha marginata</i>	<i>Phereclius pluto</i>
<i>Cyrtocoris trigonus</i>	<i>Podisus sagitta</i>	<i>Stiretrus bifrenatus</i>	<i>Antiteuchus tripterus</i>	<i>Stapecolis bimaculatus</i>
<i>Antiteuchus macraspis</i>	<i>Stiretrus anchorago</i>	<i>Tylospilus peruvianus</i>	<i>Dryptocephala lurida</i>	<i>Edessa splendens</i>
<i>Discocephalessa humilis</i>	<i>Supputius typicus</i>	<i>Tyrannocoris nigriceps</i>	<i>Edessa corallipes</i>	<i>Euschistus rufimanus</i>
<i>Platycaenus umbractulatus</i>	<i>Tylospilus acutissimus</i>	<i>Ceratozygum horridum</i>	<i>Edessa viridiorata</i>	<i>Proxys obtusicornis</i>
<i>Macropygium reticulare</i>	<i>Ablaptus brevisrostrum</i>	<i>Alveostethus politus</i>	<i>Hypoxys oxyacanthus</i>	<i>Arocera nigrorubra</i>
<i>Brachystethus improvisus</i>	<i>Dinocoris rufitarsus</i>	<i>Agacilitus dromedarius</i>	<i>Agroecus griseus</i>	<i>Chlorocoris sororis</i>
<i>Edessa rufomarginata</i>	<i>Pelidnocoris stalii</i>	<i>Antiteuchus graziae</i>	<i>Diceraeus melacanthus</i>	<i>Elanella colombiana</i>
<i>Berecynthus hastator</i>	<i>Alathetus rufitarsus</i>	<i>Antiteuchus cuspidatus</i>	<i>Oebalus poecilus</i>	<i>Chinavia dallasi</i>
<i>Galedanta myops</i>	<i>Eritrachys antennata</i>	<i>Antiteuchus melanoleucus</i>	<i>Oebalus ypsilongriseus</i>	<i>Chinavia teretis</i>
<i>Mormidea ypsilon</i>	<i>Schraderiellus cinctus</i>	<i>Antiteuchus rolstoni</i>	<i>Poriptus luctans</i>	<i>Banasa bochica</i>
<i>Proxys albopunctulatus</i>	<i>Schraderiellus hughesae</i>	<i>Antiteuchus nigricans</i>	<i>Chinavia impicticornis</i>	<i>Banasa discolor</i>
<i>Proxys victor</i>	<i>Stalius tartareus</i>	<i>Antiteuchus sepulcralis</i>	<i>Chinavia runaspis</i>	<i>Odmalea vega</i>
<i>Tibraca limbiventris</i>	<i>Paraedessa subretangulata</i>	<i>Coriplatus depressus</i>	<i>Arvelius acutispinus</i>	
<i>Arocera placens</i>	<i>Cosmopepla coeruleata</i>	<i>Colpocarena complanata</i>	<i>Banasa angulobata</i>	
<i>Vulsirea violacea</i>	<i>Euschistus bifibulus</i>	<i>Dinocoris gibbosus</i>	<i>Serdia concolor</i>	
<i>Loxa virescens</i>	<i>Euschistus crenator</i>	<i>Dinocoris maculatus</i>	<i>Oebalus ornatus</i>	
<i>Loxa viridis</i>	<i>Euschistus emoorei</i>	<i>Dinocoris nigroantennatus</i>		
<i>Chinavia ubica</i>	<i>Hypatropis rolstoni</i>	<i>Dinocoris variolosus</i>		
<i>Nezara viridula</i>	<i>Lattinidea geographica</i>	<i>Discocephalessa terminalis</i>		

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TABLE 3. (Continued)

Widespread (Neotropics) [30 spp.]	Northern Neotropics [59 spp.]	Northern South America [104 spp.]	Widespread (South America) [22 spp.]	Endemics [18 spp.]
<i>Roferta marginalis</i>	<i>Mormidea collaris</i>	<i>Dryptocephala crenata</i>		
<i>Thyanta perditor</i>	<i>Mormidea cubrosa</i>	<i>Dryptocephala dentifrons</i>		
<i>Arvelius albopunctatus</i>	<i>Mormidea fusca</i>	<i>Dryptocephala obtusiceps</i>		
<i>Pellaea stictica</i>	<i>Mormidea maculata</i>	<i>Eurystethus ovalis</i>		
<i>Piezodorus guildinii</i>	<i>Mormidea metallica</i>	<i>Paralcippus dimidiatus</i>		
	<i>Mormidea notulata</i>	<i>Patronatus binotatus</i>		
	<i>Mormidea pama</i>	<i>Pelidnocoris haglundii</i>		
	<i>Mormidea pictiventris</i>	<i>Coranda picipes</i>		
	<i>Oebalus insularis</i>	<i>Lincus lamelliger</i>		
	<i>Oebalus pugnax</i>	<i>Lincus rufospilotus</i>		
	<i>Proxys punctulatus</i>	<i>Lincus styliger</i>		
	<i>Sibaria armata</i>	<i>Lincus subuliger</i>		
	<i>Sibaria englemanni</i>	<i>Ochlerus cortiaceus</i>		
	<i>Arocera aequinoxialis</i>	<i>Orbatina fuliginia</i>		
	<i>Arocera apta</i>	<i>Paralincus bimaculatus</i>		
	<i>Rhyssoccephala splendens</i>	<i>Schraderiellus luteipedis</i>		
	<i>Rhyssoccephala macdonaldi</i>	<i>Xynocoris recavus</i>		
	<i>Chlorocoris isthmus</i>	<i>Brachystethus cribrus</i>		
	<i>Mayrinia variegata</i>	<i>Brachystethus tricolor</i>		
	<i>Chinavia marginata</i>	<i>Doesburguedessa elongatispina</i>		
	<i>Cyptocephala antiquensis</i>	<i>Edessa elaphus</i>		
	<i>Chinavia scutellata</i>	<i>Hypoxys quadridens</i>		
	<i>Banasa centralis</i>	<i>Lopadusa fuscopunctata</i>		
	<i>Banasa excavata</i>	<i>Paraedessa ecuadoriensis</i>		
	<i>Banasa panamensis</i>	<i>Paraedessa verhoeffi</i>		
	<i>Banasa salvini</i>	<i>Amauromelpia miri</i>		
	<i>Banasa zeteki</i>	<i>Cosmopepla cruciaria</i>		

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TABLE 3. (Continued)

Widespread (Neotropics) [30 spp.]	Northern Neotropics [59 spp.]	Northern South America [104 spp.]	Widespread (South America) [22 spp.]	Endemics [18 spp.]
	<i>Pharypia nitidiventris</i>	<i>Dichelops divisus</i>		
	<i>Pharypia pulchella</i>	<i>Dichelops nigrum</i>		
	<i>Taurocerus edessoides</i>	<i>Euschistus agudus</i>		
	<i>Odmalea concolor</i>	<i>Euschistus atrox</i>		
	<i>Murgantia violascens</i>	<i>Euschistus carboneris</i>		
	<i>Murgantia bifasciata</i>	<i>Euschistus rohus</i>		
	<i>Murgantia varicolor</i>	<i>Lattinellica decora</i>		
		<i>Mitripus acutus</i>		
		<i>Mormidea bridarolli</i>		
		<i>Mormidea boyilla</i>		
		<i>Mormidea triangularis</i>		
		<i>Paratibraca infuscata</i>		
		<i>Spinalanx monstrabilis</i>		
		<i>Stysiana pardidens</i>		
		<i>Arocera elongata</i>		
		<i>Arocera rufifrons</i>		
		<i>Arocera spectabilis</i>		
		<i>Rhysocephala immaculata</i>		
		<i>Rhysocephala principalis</i>		
		<i>Rumibia decorata</i>		
		<i>Rumibia discoidea</i>		
		<i>Chlorocoris depressus</i>		
		<i>Chlorocoris distinctus</i>		
		<i>Loxa nesiotis</i>		
		<i>Rhyncolepta grandicallosa</i>		
		<i>Elanella hevera</i>		
		<i>Rio insularis</i>		

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TABLE 3. (Continued)

Widespread (Neotropics) [30 spp.]	Northern Neotropics [59 spp.]	Northern South America [104 spp.]	Widespread (South America) [22 spp.]	Endemics [18 spp.]
		<i>Chinavia gravis</i>		
		<i>Chinavia plaumanni</i>		
		<i>Chinavia schuhi</i>		
		<i>Thyanta excavata</i>		
		<i>Thyanta hamulata</i>		
		<i>Thyanta obtusa</i>		
		<i>Thyanta sinuata</i>		
		<i>Thyanta straminea</i>		
		<i>Thyanta testacea</i>		
		<i>Arvelius confusus</i>		
		<i>Arvelius peruanus</i>		
		<i>Arvelius porrectispinus</i>		
		<i>Banasa expallescens</i>		
		<i>Banasa fulgida</i>		
		<i>Banasa gynamagna</i>		
		<i>Banasa paraexpallescens</i>		
		<i>Banasa peruana</i>		
		<i>Banasa saileri</i>		
		<i>Banasa subrufescens</i>		
		<i>Banasa varians</i>		
		<i>Elsiella plana</i>		
		<i>Neotibilis parva</i>		
		<i>Serdia delphis</i>		
		<i>Taurocerus achilles</i>		
		<i>Patanius vittatus</i>		

several cases. For instance, the distribution of important economic species such as *Antiteuchus tripterus*, *Chinavia marginata*, *Euschistus rufimanus*, *Podisus nigrispinus* and *Tibraca limbativentris* is still poorly known, with only a few records in Colombia. Even widespread and easily recognized species, such as *Nezara viridula*, *Piezodorus guildinii* and *Thyanta perditor* records include few known localities. Furthermore, updated information about host plants, life history and biology in Colombia, as well as the impact on crops in the country, are scarce and available only for a few species of economic importance (e.g., Waldbauer 1977; Vidal & Segura 1981; Osorio *et al.* 1988; Daza 1991; Hallman *et al.* 1992; Pantoja *et al.* 1995; Pantoja *et al.* 2005; Pantoja *et al.* 2007; Agudelo *et al.* 2008). Taxonomic issues still need to be clarified in some cases, e.g., records of *Tibraca obscurata* (cited in Pantoja *et al.* 1995, Pantoja *et al.* 2005) were considered *incertae sedis* in Fernandes & Grazia (1998), and in at least two cases (*Edessa meditabunda* in tobacco and *Oebalus ornatus* in rice), we were not able to study specimens to confirm its records from Colombia.

Limitations and future research directions.

This study constitutes the first effort to assemble accurate information about the diversity of stink bugs in Colombia. Besides offering a checklist and new taxa records for the country, it provides a starting point for further analyses on the Neotropical pentatomid fauna. The data presented in the species list, including the new records, places Colombia as one of the most speciose countries in the Neotropics regarding the stink bug fauna, although this should be viewed as an underestimate of its real diversity. There are several factors that confound the real number of species of Pentatomidae in the country, and in the following we list what may be considered the most important in our opinion, including future steps to overcome these limitations.

1) Lack of researchers and focused entomological collections. Currently, in Colombia there are no taxonomic researchers actively focusing on pentatomoid diversity. As a consequence, there are little field efforts to document pentatomoids in natural areas, thus, local identification of specimens relies mostly on reference specimens collected and identified years ago by non-specialists. The lack of specialized taxonomic research means not only a lag in biodiversity knowledge, but also that the representation of Pentatomidae in biological collections nationwide is insufficient, and that their natural history is being inadequately documented. As another consequence, most of the specimens in collections are not adequately curated, negatively impacting other scientific areas, such as in agricultural production, where several Pentatomidae species might be of concern. Therefore, we propose strengthening collaborative networks among international colleagues, in particular between Colombia and Brazil, where there are many active Pentatomidae researchers.

2) Collections. The specimens examined for this study came from five of the most representative national entomological collections. These collections have a geographic representation that covers most of the Colombian territory. The examined entomological collections are in areas mostly located in the North-Andean region, therefore, the adjacent areas are better documented than other places in the country. Nonetheless, we are sure that the total species number is going to be higher if additional entomological collections are studied, in particular regional collections. Right now, there are several new regional, smaller entomological collections in Colombia (<http://rnc.humboldt.org.co/wp/colecciones/>) that are extremely important to document local fauna, which is difficult to sample by other researchers. Only a coordinated effort between researchers and collections, to facilitate the loan and study of these specimens, will further advance our knowledge of the Pentatomidae fauna from Colombia.

3) Collecting gaps. From the data gathered here, we recorded pentatomid species from 27 of the 32 departments of Colombia. We have no information for the Caribbean or Pacific Oceanic Insular Territories (Archipelago of San Andrés y Providencia, and Isla de Malpelo), and the Arid Peri-Caribbean Belt (particularly Córdoba, La Guajira, and Sucre). This result does not mean absence of pentatomids in these areas, but likely insufficient collecting efforts in these regions or limited access to regional collections as mentioned above.

4) Poorly known groups. Although some stink bug groups have recent taxonomic revisions that offer comprehensive descriptions and keys that allow recognizing genera and species with relative easiness, not all Pentatomidae groups are easily identified. Some Pentatomidae groups still lack adequate taxonomic studies, negatively affecting the capacity to properly identify them. For instance, the subfamilies Discocephalinae and Edessinae, particularly the tribe Ochlerini (Discocephalinae), are two of the most taxonomically difficult groups, with scarce taxonomic revisions and with just one Neotropical active researcher each, Luiz A. Campos (UFRGS) and José A. M. Fernandes (UFPA). Most Discocephalinae species are not commonly collected, probably because of their specialized life history traits, negatively impacting their taxonomic knowledge (Grazia *et al.* 2015; Schuh & Weirauch 2020). Edessi-

nae, especially the genus *Edessa*, is a very speciose group, which is easily collected and thus abundantly represented in the Colombian entomological collections. Nonetheless, very little is known about Edessinae in Colombia due to their complicated taxonomic status and identification (Fernandes & Doesburg 2000a, b; Silva *et al.* 2013; Fernandes *et al.* 2018; Nunes *et al.* 2019). Although one of the most diverse groups of Neotropical stink bugs, Edessinae in our list represented less than 5% of the known Colombian fauna. We strongly suggest focusing taxonomic efforts in these poorly known groups, taking advantage of the material accumulated in Colombian entomological collections. Future studies should also explore species and genera with agricultural importance in Colombia, as well as endemic and invasive species, which in turn have a potential impact on the economy of the country.

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