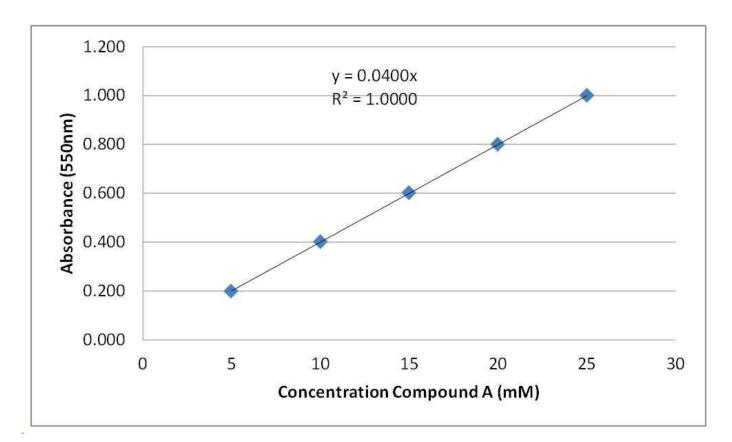
## MOLAR ABSORPTIVITY

DETERMINING MOLAR ABSORPTIVITY COEFFICIENTS.

Suppose that we made a standard curve where the data obeys Beer's Law and fits a straight line. We should be able to calculate the molar absorptivity coefficient ( $\epsilon$ ).

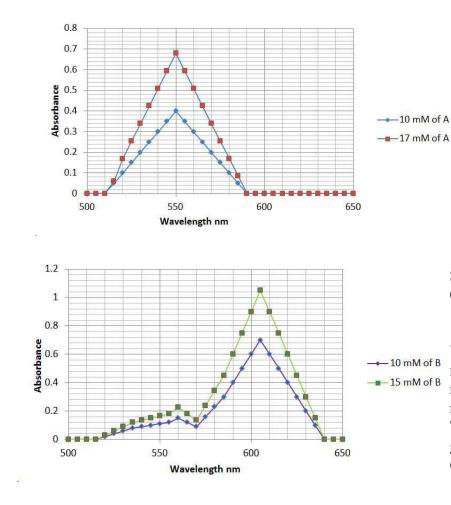


Since A=\varepsilon b and the graph is of A vs C (mM) A = (\varepsilon b) c the slope is therefore equal to (\varepsilon b). We get the slope from the y=0.400x. In the above: slope = 0.0400 mM<sup>-1</sup> = \varepsilon b where b is usually 1 cm (the pathlength of a cuvette) Therefore we can calculate the value of \varepsilon: \varepsilon = (slope / b) = (0.0400 mM<sup>-1</sup>/ 1 cm) = 0.0400 mM<sup>-1</sup> cm<sup>-1</sup> or 0.0400  $\frac{L}{mmoles cm}$ \varepsilon is most often written in M<sup>-1</sup> cm<sup>-1</sup> so  $\varepsilon = 0.0400 \frac{L}{mmoles cm} \times \frac{1000 mmoles}{mole} = 40.0 \frac{L}{moles cm} = 40.0 M^{-1} cm^{-1}$ 

## MOLAR ABSORPTIVITY

Two compounds absorbing at the same  $\boldsymbol{W}\!$  avelength.

Suppose we took a scan of a sample containing 10.0 mM Compound A (blue dots) and a scan of a sample containing 17.0 mM Compound A (red dots).



We can see that this compound has a maximum wavelength at 550 nm  $(\lambda_{max})$ . The higher the concentration, the greater the <sup>A</sup> absorbance at 550nm. It does not absorb light above 600 nm at either concentration.

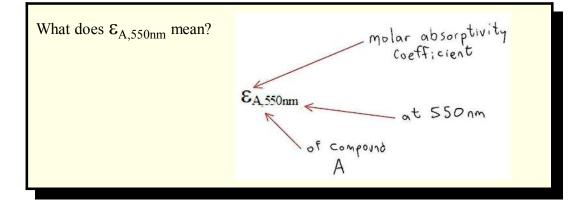
 $\begin{array}{ll} (\epsilon_{\text{A},550\text{nm}} = \ 0.0400 \ L \ \text{mmoles}^{-1} \ \text{cm}^{-1}) \\ (\epsilon_{\text{A},605\text{nm}} = \ 0.000 \ L \ \text{mmoles}^{-1} \ \text{cm}^{-1}) \end{array}$ 

Suppose we took a scan of 10.0 mM Compound B (blue) and a scan of 15.0 mM Compound B (green).

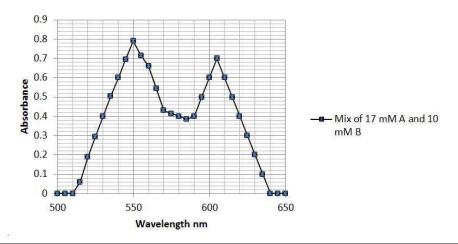
We can see that Compound B has a maximum wavelength  $(\lambda_{max})$  of 605 nm. It does absorb light below 600 nm, in the range of Compound A. The higher the concentration, the greater the absorbance at 550nm and 605nm.

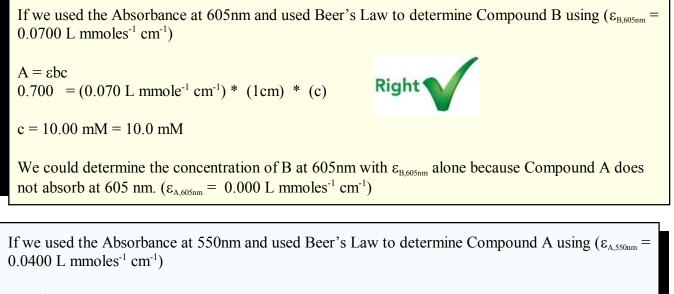
 $\begin{aligned} &(\epsilon_{\text{B},605\text{nm}} = \ 0.0700 \ \text{L mmoles}^{-1} \ \text{cm}^{-1}) \\ &(\epsilon_{\text{B},550\text{nm}} = \ 0.0110 \ \text{L mmoles}^{-1} \ \text{cm}^{-1}) \end{aligned}$ 

\*\* note usually a scan would have a curved appearance, the above scans are made up numbers for demonstration.



What happens if we have a mix of 17.0 mM A and 10.0 mM B in the same cuvette? How do we calculate concentration of A and B using Beer's Law?





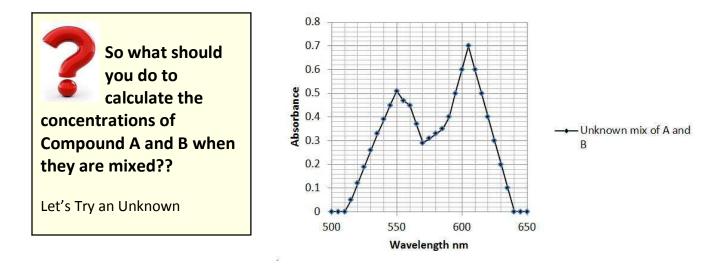
A =  $\epsilon bc$ 0.790 = (0.040 L mmole<sup>-1</sup> cm<sup>-1</sup>) \* (1cm) \* c c = 19.75 mM = 19.8 mM



19.8 mM would be our answer which is close to the actual, but we have overestimated the real value of 17.0 mM. The reason is because Compound B also absorbs at this wavelength. ( $\epsilon_{B,550nm} = 0.0110$  L mmoles<sup>-1</sup> cm<sup>-1</sup>).

In reality you must add Beer's Law together for compound A and compound B at 550nm.

$$A_{550nm} = \varepsilon_{A,550nm} bc + \varepsilon_{B,550nm} bc$$
  
= (0.0400 L mmole<sup>-1</sup> cm<sup>-1</sup> \* 1cm \* 17.0mM) + (0.0110 L mmole<sup>-1</sup> cm<sup>-1</sup> \* 1cm \* 10.0mM)  
= 0.680 + 0.110  
= 0.790



At 550 nm the mixed sample absorbs at 0.510. At 605 nm the mixed sample absorbs at 0.700. Since Compound A does not absorb at 605 nm ( $\epsilon_{A,605nm} = 0.000 \text{ L mmole}^{-1} \text{ cm}^{-1}$ ), we should determine B's concentration first.

 $\begin{array}{ll} A_{605nm} &= 0.700 &= \epsilon_{A,605nm} bc_A + \epsilon_{B,605nm} bc_B \\ &= 0.700 &= (0.000 \ L \ mmole^{-1} \ cm^{-1} \ * \ 1cm \ * \ c_A) + (0.0700 \ L \ mmole^{-1} \ cm^{-1} \ * \ 1cm \ * \ c_B) \\ &= 0.700 &= 0 + (0.0700 \ L \ mmole^{-1} \ cm^{-1} \ * \ 1cm \ * \ c_B) \\ &= 0.700/(0.0700 \ L \ mmole^{-1} \ cm^{-1} \ * \ 1cm) = \ c_B \end{array}$ 

 $c_B = 10.0 \text{ mM}$  therefore concentration of Compound B is 10.0 mM

Compound A's concentration  $(c_A)$  is a little more difficult to determine, but since we now know B's concentration  $(c_B)$  to be 10.0 mM from the above calculation, we can determine A's next.

 $\begin{array}{ll} A_{550nm} &= 0.510 &= ( \ \epsilon_{A,550nm} * b * c_{A}) + ( \ \epsilon_{B,550nm} * b * c_{B}) \\ &= 0.510 &= (0.0400 \ \text{L mmole}^{-1} \ \text{cm}^{-1} * 1 \ \text{cm}^{*} c_{A}) + (0.0110 \ \text{L mmole}^{-1} \ \text{cm}^{-1} * 1 \ \text{cm}^{*} 10.0 \ \text{mmole}) \\ &= 0.510 &= (0.0400 \ \text{L mmole}^{-1} \ \text{cm}^{-1} * 1 \ \text{cm}^{*} c_{A}) + (0.110) \\ &= 0.510 - 0.110 = (0.0400 \ \text{L mmole}^{-1} \ \text{cm}^{-1} * 1 \ \text{cm}^{*} c_{A}) \\ &= 0.400/(0.0400 \ \text{L mmole}^{-1} \ \text{cm}^{-1} * 1 \ \text{cm}) = c_{A} \end{array}$ 

 $C_A = 10.0$  mM therefore concentration of Compound A is 10.0 mM.

Using multiple wavelengths in a Biochemical assay is fairly common.

•An example of this is in determining the purity of DNA. Samples are measured at 260nm and 280nm.

•Another example is the AlamarBlue assay which is used to determine cellular proliferation. This takes measurements at 570nm and 600nm.