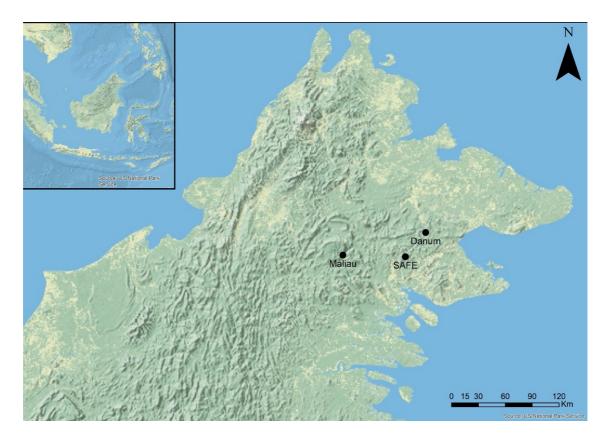
Supplementary information 1: field work



Locations of study sites in Sabah, Malaysia.

We sampled bats using six harp traps per night at three lowland tropical rainforest sites within Sabah, Malaysia, each <500m above sea level and with largely unseasonal climate. Two sites (Danum Valley and Maliau Basin) are primary rainforest, and the final site (the Stability of Altered Forest Ecosystems Project) have been subject to substantial anthropogenic disturbance.

With the exception of pregnant, lactating or juvenile individuals (which were released immediately after capture), bats were retained for up to 12 hours, usually around 10 hours, to allow sufficient time for defecation following the morning/evening's feeding activity. Sample sites were often substantial distances from the research camp, and so bats were placed in individual bags and transported to camp so they could be kept in quiet, humid conditions to minimise stress.

The Danum Valley Conservation Area (hereafter 'Danum') is a 438 km² region of undisturbed primary rainforest in Sabah (Reynolds, Payne, Sinun, Mosigil, & Walsh, 2011), where there has been little logging or hunting in historic times. It

is designated as a fully protected forest reserve. All of my sampling was in the forest <1.2 km from Danum Valley Field Centre. Traps were erected in 2016 for ten nights in a 21 night period and 2017 for ten nights in a 12 night period.

- The Maliau Basin Conservation Area (hereafter 'Maliau') is a 588 km² fully protected forest reserve made up of lowland and hill forest, most of which has neither been logged nor inhabited in historical times. WE sampled at Maliau Basin on the Seraya trail, an area of primary lowland rainforest. Traps were erected in 2016 and 2017 for ten nights in a 16 night period.
- The Stability of Altered Forest Ecosystems Project (hereafter 'SAFE') is a large ecological experiment where an area of degraded forest is being converted to oil palm plantation, with fragments of forest and riverine buffers being retained for scientific study (Ewers et al., 2011). The site has been subjected to disturbance since approximately March 2015, with areas of forest outside of designated fragments being salvage-logged. At the time that WE completed my sampling in July 2017, the forest was still in the process of conversion; thus, the study area was composed of experimental forest areas located within a matrix of highly degraded forest and shrubby clearings. WE sampled in the blocks 'LFE', 'B' and 'C', within the Ulu Segama Forest Reserve and Kalabakan area, during 2015, 2016 and 2017. Each of the blocks was sampled for a 5-night period, and then resampled at least 5 weeks later.

	Number captured	Number of samples obtained	Rate
Balionycteris maculata	5	1	0.2
Emballonura monticola	2	1	0.5
Hesperoptenus blanfordi	19	10	0.526
Hipposideros ater	5	1	0.2
Hipposideros bicolor	10	6	0.6
Hipposideros cervinus	1147	493	0.43
Hipposideros diadema	44	24	0.545
Hipposideros dyacorum	57	25	0.439
Hipposideros galeritus	1	1	1
Hipposideros ridleyi	39	24	0.615
Kerivoula hardwickii	99	33	0.333
Kerivoula intermedia	273	97	0.355
Kerivoula lenis	3	3	1
Kerivoula minuta	12	4	0.333
Kerivoula papillosa	103	40	0.388
Kerivoula pellucida	29	11	0.379
Megaderma spasma	5	3	0.6
Murina aenea	1	1	1
Murina cyclotis	4	2	0.5
Murina peninsularis	4	4	1
Murina rozendaali	5	3	0.6
Murina suilla	16	9	0.562
Myotis muricola	3	3	1
Myotis ridleyi	1	1	1
Nycteris tragata	7	2	0.286
Phoniscus atrox	3	1	0.333

Total number of captures and samples collected

The number of captures and samples obtained for the bat species used to make the ecological networks, and the defecation rate per species.

	Number captured	Number of samples obtained	Rate
Hipposideros cervinus	1147	493	0.43
Hipposideros diadema	44	24	0.545
Hipposideros dyacorum	57	25	0.439
Kerivoula hardwickii	99	33	0.333
Kerivoula intermedia	273	97	0.355
Rhinolophus borneensis	103	40	0.388
Rhinolophus sedulus	58	37	0.638
Rhinolophus trifoliatus	81	34	0.42

Supplementary information 2: analysis

Laboratory work

WE undertook DNA extraction and PCR using the protocol outlined in Chapter 2. Quality-control on amplicons was then performed using a DNA D1000 TapeStation (Agilent Technologies), and quantified using a QuBit dsDNA HS Assay Kit (Invitrogen, Life Technologies). Finally, sequencing took place on three 96-well plates, which were run on an Illumina MiSeq at the Genome Centre, London (UK). Sequencing was performed bi-directionally with Fluidigm indexes following manufacturer's instructions.

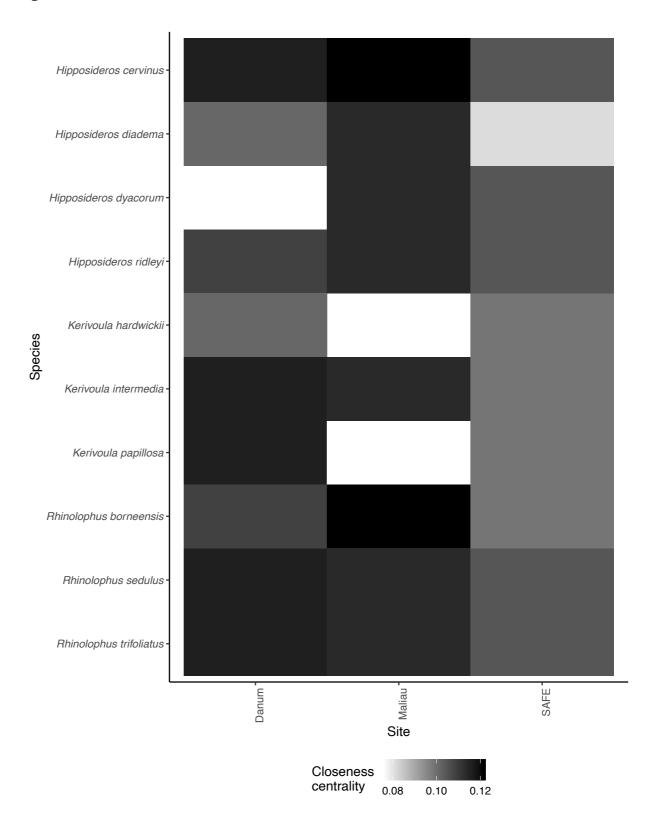
Bioinformatics pipeline

WE assembled forward and reverse sequence reads into contigs using mothur (Schloss et al., 2009), and then removed forward and reverse primers using the galaxy web platform on the public server at usegalaxy.org (Afgan et al., 2016), as per the protocol in Chapter 2. A custom python script was used to remove any haplotypes either represented by a single sequence or falling outside of a length of 155-159bp. WE then generated subsequent datasets using MOTU clustering thresholds at ranges 91-98% similarity, using the Uclust algorithm (Edgar, 2010) as implemented in the QIIME platform (Caporaso, Kuczynski, Stombaugh, Bittinger, & Bushman, 2010). Representative sequences for each MOTU per clustering level were then compared to one another using BLAST+ (Camacho et al., 2009), with the resulting data being reduced in LULU (Frøslev et al., 2017) for quality control. All resulting bat-MOTU adjacency lists were then transformed into adjacency matrices using a custom perl script. These matrices were then split into multiple binary adjacency matrices by site, where *aij* denotes the consumption of MOTU *j* by bat individual *i*. Networks were created by pooling samples from multiple years. All bioinformatic statistical recorded and steps are at https://github.com/hemprichbennett/bat-diet.

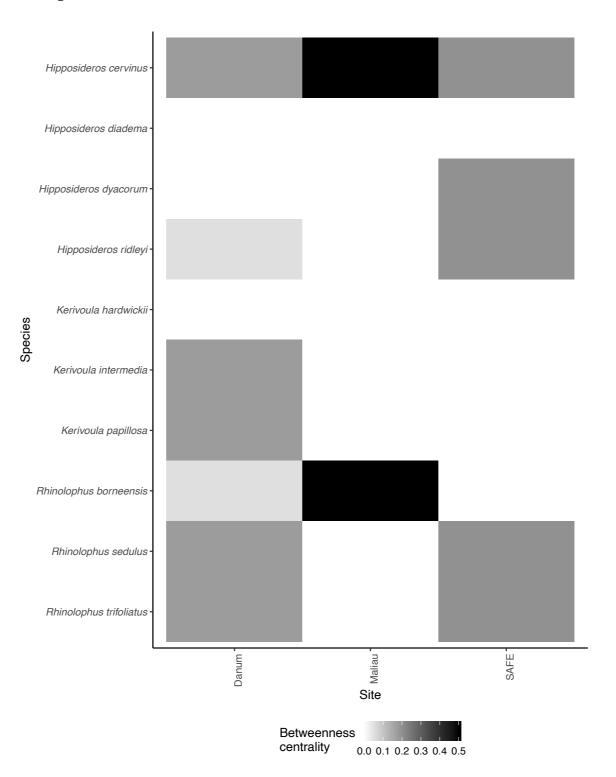
Supplementary information 3. The number of MOTUs consumed by bats at each site, as observed at each MOTU clustering level.

Clustering level	Danum	Maliau	SAFE
91	1329	920	1205
92	1373	884	1243
93	1369	875	1237
94	1378	876	1203
95	1388	842	1207
96	1401	837	1224
97	1449	841	1256
98	1544	873	1301

Supplementary information 4. Closeness centrality of each bat species studied

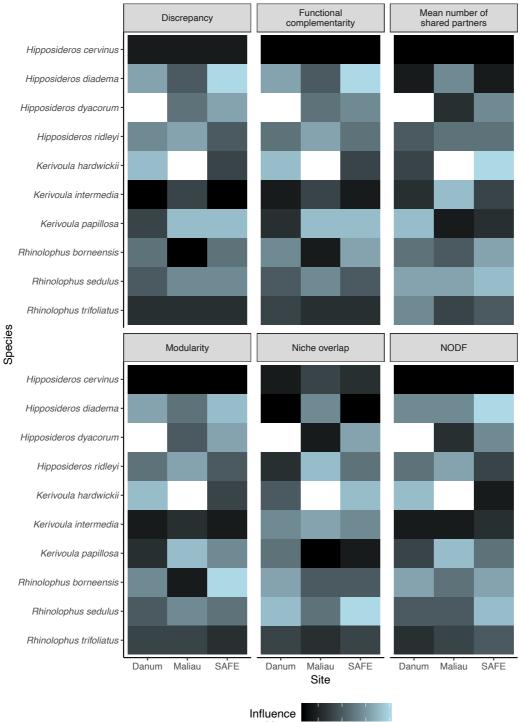


Supplementary information 5. Betweenness centrality of each bat species studied



Supplementary information 6. Rankings of the influence of each bat species on a given network-level metric.

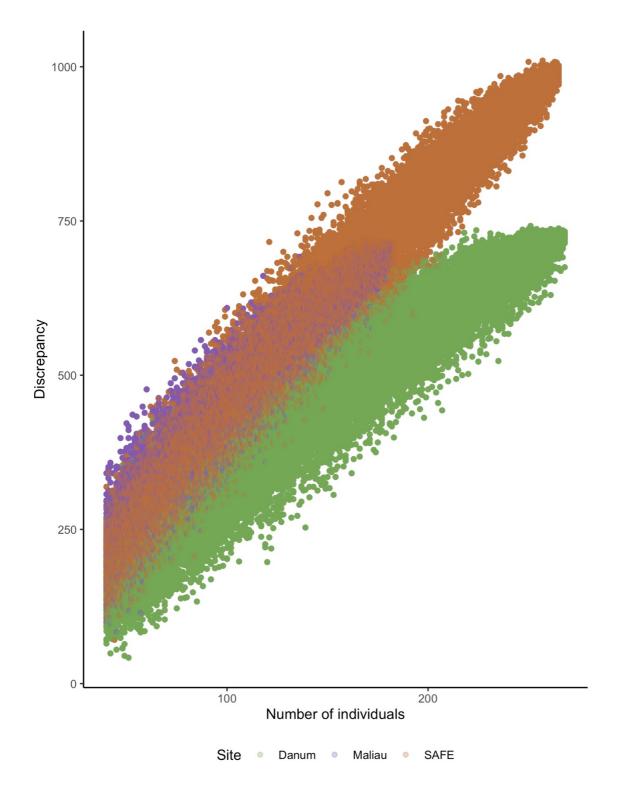
1 is the ranking of highest influence, 10 is the lowest. White tiles denote a bat species not included in the network for that site, due to lack of captures.

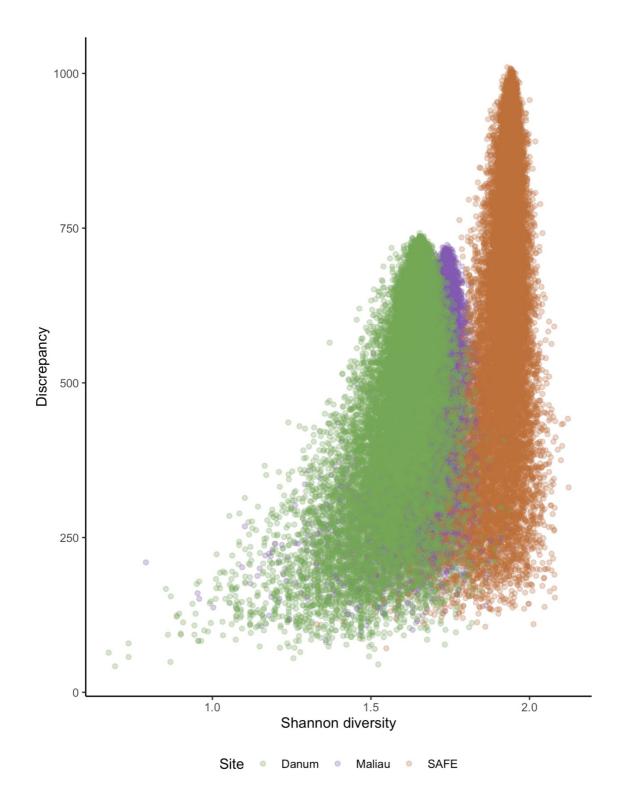


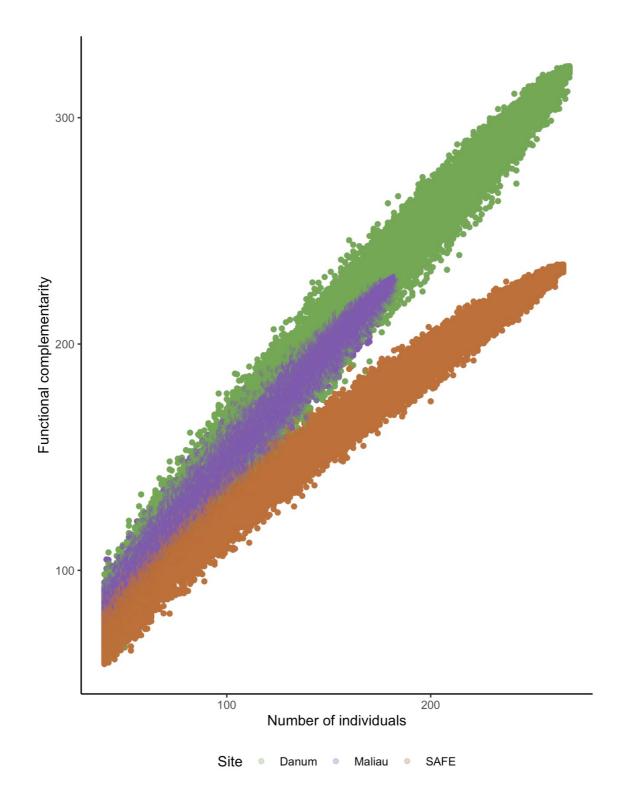
ranking 2.5

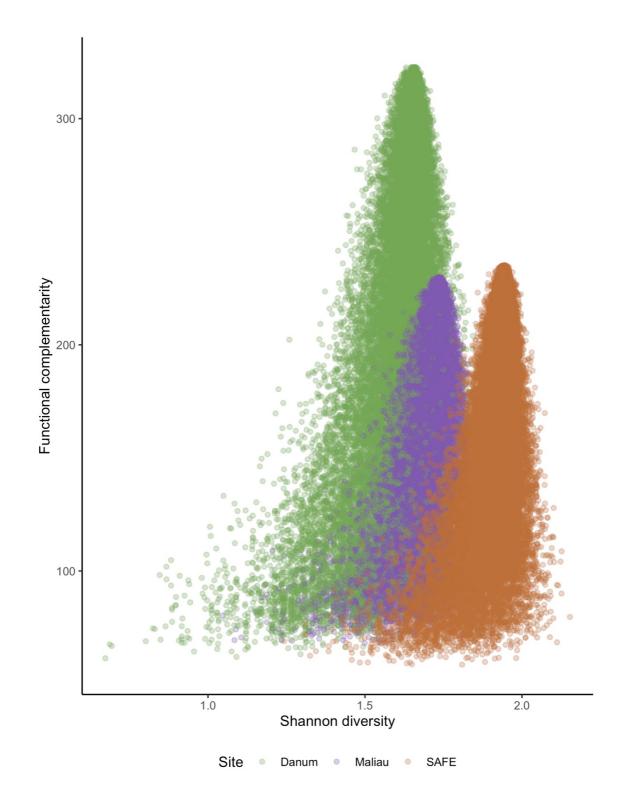
2.5 5.0 7.5 10.0

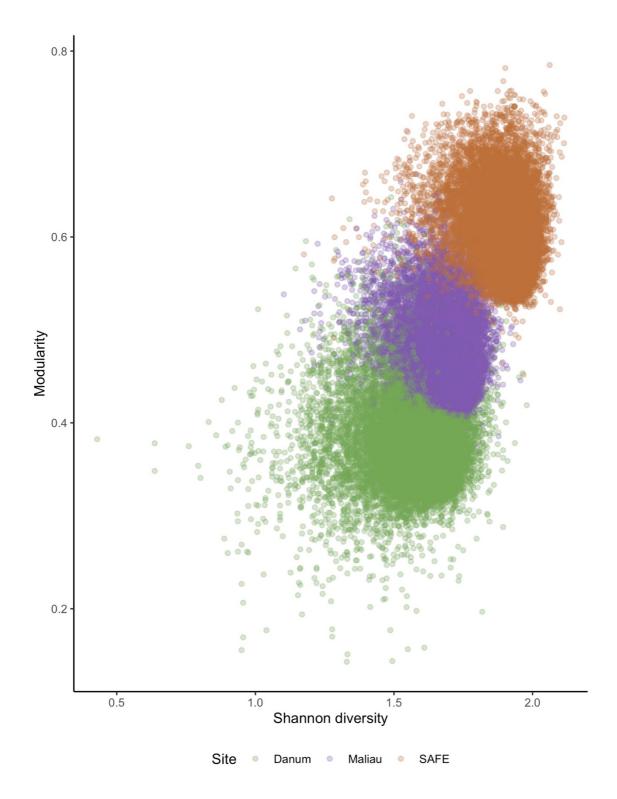
Supplementary information 7. Alterations in observed metric values when rarefying networks

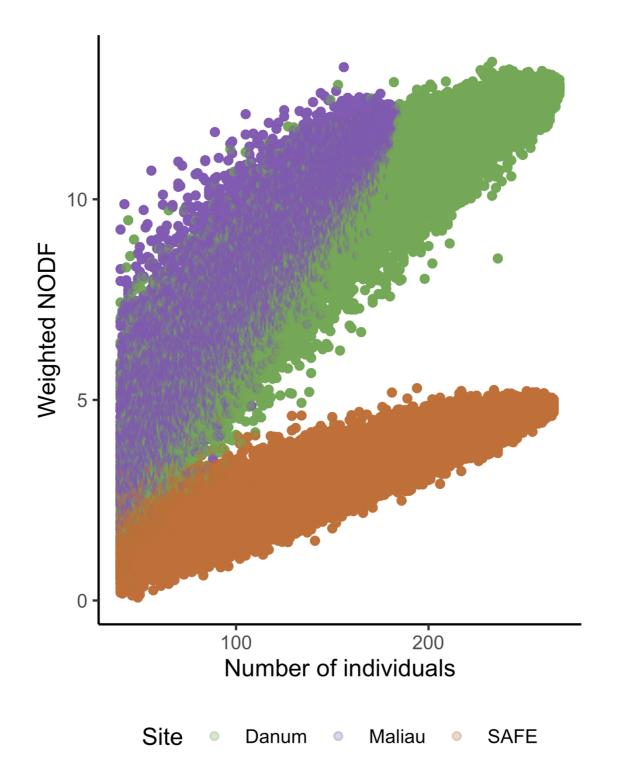


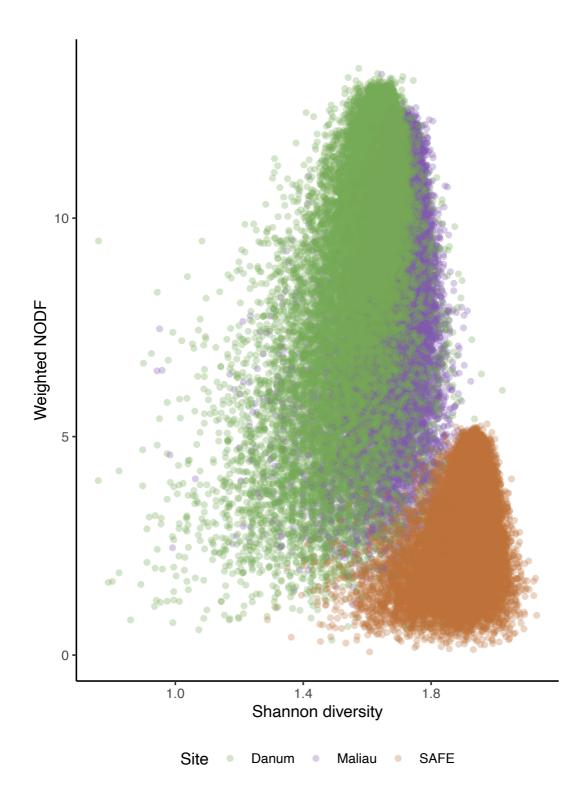


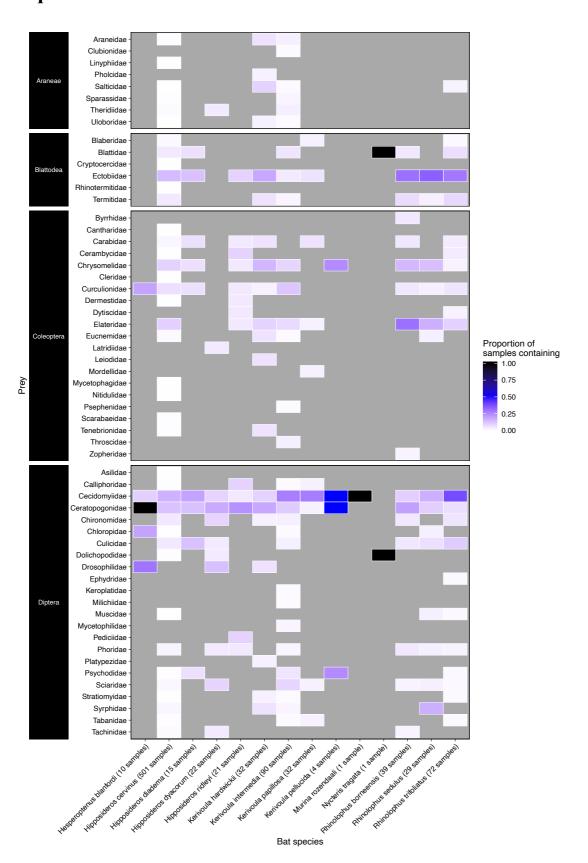




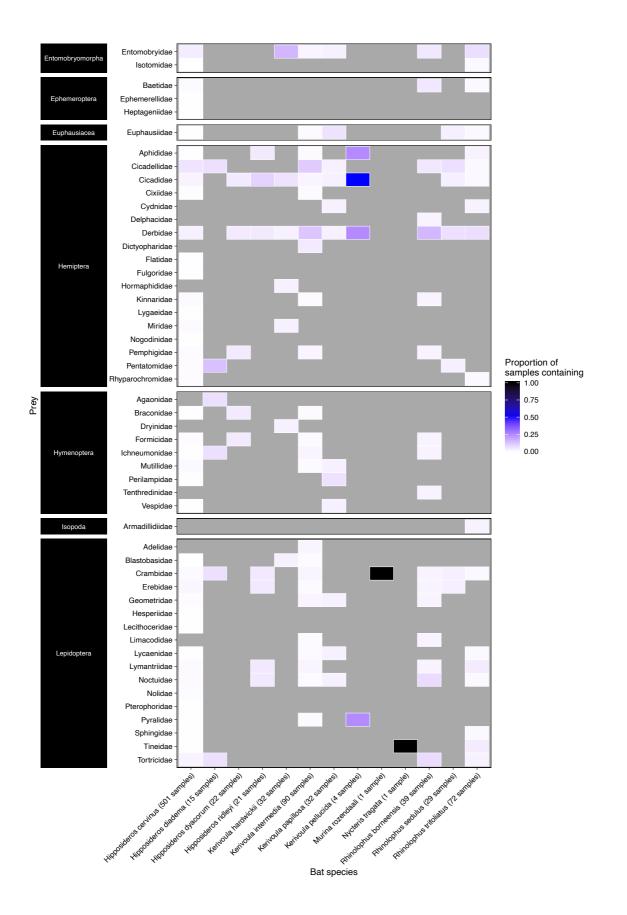


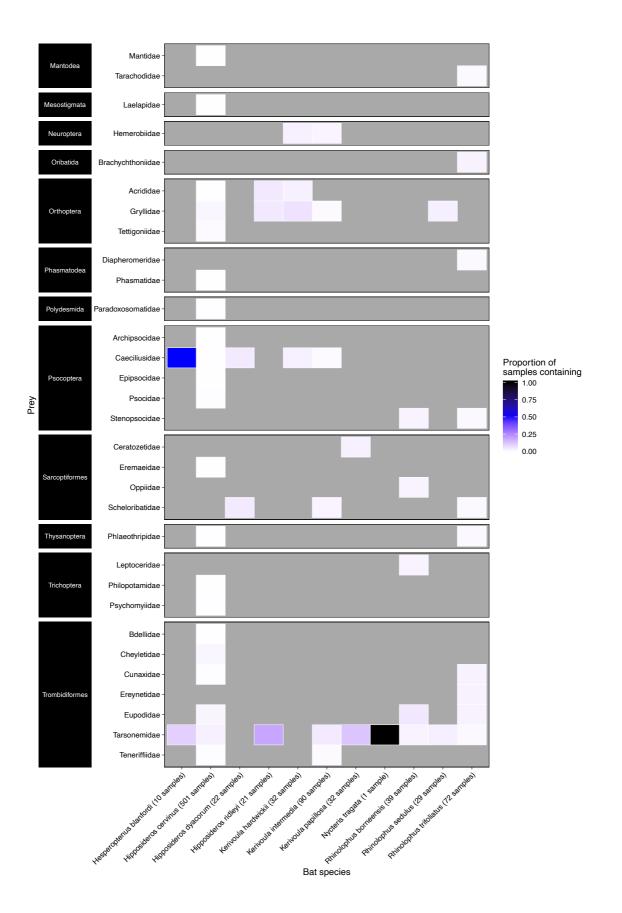






Supplementary information 8. Family-level diets of all bats sequenced





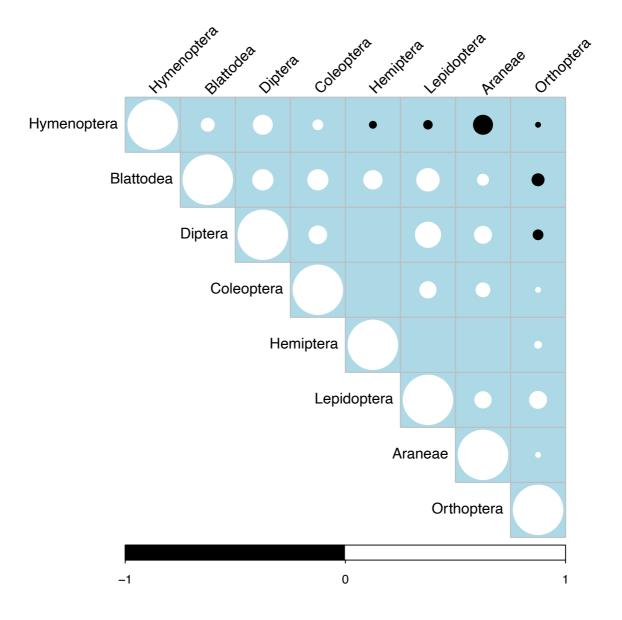
Supplementary information 9 Dietary correlations

Methods

To identify if there are patterns of taxonomic Orders co-occurring within the diet of individual bats (e.g. bat individuals that feed on Coleoptera may be more likely to feed on Blattodea), we analysed the co-occurrence between taxonomic Orders within the diets of bats using Pearson's correlation coefficient, with each individual bat acting as a sample. This took place to identify potential significant correlations of prey consumption, and potential primer biases as the ZBJ primers used here have previously been shown to preferentially amplify some taxa (Alberdi, Aizpurua, Gilbert, & Bohmann, 2018). The analysis was restricted to bat individuals that had consumed more than five taxonomically-matched MOTUs, and to Orders of prey that had been consumed by over 20 bats; this approach reduced the biases arising from zero-values. The resulting data was visualised using the R package 'corrplot' (Wei & Simko, 2017).

Results:

Significant positive and negative correlations occurred between many taxonomic Orders (see Supplementary Information), with Hymenoptera and Orthoptera being the only Orders negatively correlated with multiple Orders, though correlations were weak. Despite reported primer bias towards Lepidoptera and Diptera (Alberdi et al., 2018), these Orders were each only negatively correlated with a single other Order, and these correlations were also weak.



Positive and negative correlations of orders consumed by all bats sequenced. Only bats consuming more than 5 taxonomically identified MOTUs, and orders consumed by more than 20 bats were included. Circle size is proportional to the strength of the correlation, black circles are negative correlations, white circles are positive correlations. Empty locations within the grid show non-significant (p>0.05) correlations.

Supplementary information 10: licenses and permits for fieldwork

licenses JKM/MBS.1000- 2/2 (374), JKM/MBS.1000-2/2 (access JLD.4 (23), JKM/MBS.1000-2/2 JLD.4 (45), JKM/MBS.1000-2/2 JLD.4 (41), Access licenses JKM/MBS.1000-2/2 JLD.4 (46), JKM/MBS.1000-2/2 JLD.5 (123) and JKM/MBS.1000-2/2 JLD.5 (153), export licenses JKM/MBS.1000-2/3 JLD.2(55), JKM/MBS.1000-2/3 JLD.2 (95) JKM/MBS.1000-2/3 JLD.3 Danum Valley and (31). access permits (refs YS/DVMC/2015/221, YS/DVMC/2016/11, YS/DVMC/2015/222, YS/DVMC/2016/13, YS/DVMC/2017/42, YS/DVMC/2017/41) Maliau Basin access permits (refs YS/MBMC/2015/186, YS/MBMC/2016/23, YS/MBMC/2015/187, YS/MBMC/2016/25, YS/MBMC/2017/67 and YS/MBMC/2017/66).

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