Zoological Journal of the Linnean Society, 2023, **198**, 310–325. With 3 figures.

Endemic lineages of spiny frogs demonstrate the biogeographic importance and conservational needs of the Hindu Kush–Himalaya region

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Received 2 August 2022; revised 26 December 2022; accepted for publication 27 December 2022

The relict, endemic taxa *Allopaa* and *Chrysopaa* are key elements of the Hindu Kush–Himalayan amphibian fauna and potentially share a similar biogeographic evolution, making them important proxies for the reconstruction of the palaeoenvironmental and palaeotopographic history of the Himalaya–Tibet–Orogen. However, little is known about the taxonomy, phylogeography, genetic diversity and distribution of these taxa. We here provide new molecular data on Himalayan spiny frogs and species distribution models (SDMs) for *A. hazarensis* and *C. sternosignata*. The results reveal a better resolved phylogeny of these frogs compared to previous trees and strongly support the placement of *A. hazarensis* in the genus *Nanorana*. We further identify a so far unknown clade from the western Himalayas in *Nanorana*, apart from the subgroups *Chaparana*, *Paa* and the nominal *Nanorana*. In *A. hazarensis*, genetic diversity is relatively low. The results strengthen support for the recently proposed out-of-Tibet-into-the-Himalayan-exile hypothesis and a trans-Tibet dispersal of ancestral spiny frogs during the Palaeogene. Moreover, SDMs provide the first detailed distribution maps of *A. hazarensis* and *C. sternosignata* and strong evidence for distinct niche divergence among the two taxa. Our findings contribute to the knowledge about the distribution of these species and provide basic information for guiding future conservation management of them.

KEYWORDS: *Allopaa* – *Chrysopaa* – *Nanorana* – niche divergence – phylogeny.

INTRODUCTION

Understanding the palaeoenvironmental and topographic history of the northwestern Himalaya, also known as Kashmir or Indus Himalaya, and the adjacent Karakoram and Hindu Kush Mountain ranges is crucial for the reconstruction of the biogeographic history of species groups endemic to these parts of the Himalaya–Tibet–Orogen (HTO) (Schmidt *et al.*, 2012; Hofmann *et al.*, 2017; Jablonski *et al.*, 2021). For different groups of organisms, e.g. ground beetles, lazy toads and spiny frogs, the presence of

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ancestral lineages in the northwestern Himalaya has been shown (Schmidt *et al.*, 2012; Hofmann *et al.*, 2017, 2019, 2021a). These lineages are most closely related to representatives in the eastern Himalaya or eastern central Himalaya, but they have no relatives across wide regions of the West and Central Himalaya. Such a paradoxical distributional pattern has been explained by a palaeo-Tibetan origin of Himalayan lineages and/or a trans-Tibet dispersal of ancestral lineages across central Tibet during the Late Palaeogene or Early Neogene when the Plateau was at significant lower elevation (Schmidt *et al.*, 2012; Hofmann *et al.*, 2017, 2019, 2021a).

In this context, the phylogenetic placement of the two westernmost dicroglossid frogs that occur in the HTO, *Allopaa hazarensis* (Dubois & Khan, 1979) from the Kashmir Himalaya and *Chrysopaa sternosignata* (Murray, 1885) from the Hindu Kush, have recently been addressed for the first time (Hofmann *et al.*, 2021a). While the basal position of the monotypic *Chrysopaa* Ohler & Dubois, 2006 relative to *Nanorana* Günther, 1896 and *Allopaa* Ohler & Dubois, 2006 was strongly supported, the placement of *A. hazarensis* from Pakistan in the genus *Nanorana* seemed reasonable, but remained less supported, presumably either due to a gap in taxon sampling and/or because it represents a lineage that diverged in an early stage of the evolutionary history of *Nanorana*. Both taxa, *Chrysopaa* and *Allopaa*, are endemic to the Hindu Kush–Himalayan area and were previously considered a single genus (*Paa*) (Khan, 2006). To the best of our knowledge, sympatric or syntopic records do not exist, although they might be possible and misidentification of the two frogs cannot be excluded. The two taxa are key elements of the regional thermophile fauna and potentially share a similar biogeographic origin, making them important proxies for the reconstruction of the palaeoenvironmental and palaeoelevational evolution of the Tibetan Plateau (Hofmann *et al.*, 2021a) and the evaluation of respective modern scenarios of the development of the HTO (Xiong *et al.*, 2022). Apart from molecular data, information about the current distribution and ecological niches of these relict taxa are crucial to assess their biogeographic history in the context of the evolution of the HTO.

Allopaa hazarensis can be found in boulder-rich streams (Dubois, 1975) or clear pools with flowing water (Khan *et al.*, 2008), often surrounded by subtropical or warm-temperate forest at elevations above 1000 m a.s.l. (own observation; Ahmed *et al.*, 2020). The species was described from near Datta (~34.30°N, 73.26°E), northern Pakistan (Manshera District, Hazera Division, about 1200 m a.s.l.; for molecular data from the type locality, see: Hofmann *et al.*, 2021a) and is known to occur in the Khyber Pakhtunkhwa Province of Pakistan, and in adjacent Kashmir, India (see: Frost, 2022). Apart from its still not fully resolved taxonomic position, little is known about the life history, ecology and distribution of *A. hazarensis*, and far less data are available about its genetic diversity across the known distribution range of the species.

On the other hand, *Chrysopaa sternosignata* has been originally described from Sindh Province in Pakistan (Murray, 1885). The syntypes, presumably at the Karachi Museum, are apparently lost (Frost, 2022); two further syntypes are noted in the BMNH (1947.2.1.21–22; Boulenger, 1920) with the type localities 'Mulleer (= Malir) near Kurrachee (= Karachi); Zandra and Quetta, in South Afghanistan' (now all localities in Pakistan). However, we consider the occurrence of the species at Malir as unlikely (own data; see also: Mertens, 1969) since the environmental conditions in the south-eastern edge of Pakistan (Sindh Province) are remarkably different compared to those of places where the species has its major distribution range (Balochistan, Afghanistan; Supporting Information, Fig. S1). *Chrysopaa* seems to be exclusively aquatic and has been reported to be common in pools and water-channels in the Quetta and Pishin districts of Balochistan Province in Pakistan from 1500 up to 1800 m a.s.l. (Boulenger, 1920), as well as in other areas of southern, western and central Afghanistan (Wagner *et al.*, 2016). Noteworthy, the species has never been recorded in Iran. It occupies predominantly regions in the colline zone with warm-temperate, arid climate. Similar to *A. hazaranesis*, there are virtually no studies on the ecology of the species and only a few molecular datasets for *C. sternosignata* are available. However, such information is essential not only for species conservation, but also for biogeography and molecular taxonomy.

Therefore, we aim (1) to place new samples of both species and additional *Nanorana* specimens onto the phylogeny of closely related spiny frogs in order to enhance phylogenetic resolution and to assess previous biogeographic hypotheses, (2) to provide information on the genetic diversity of *Allopaa hazarensis* and (3) to model the habitat suitability and potential distribution of the two taxa, which is vital for understanding the historical processes that shaped the disjunction of species and are probably important in guiding conservation planning. Our data represent the most detailed evaluation of molecular and occurrence data of these enigmatic frogs. The results encourage discussion on our understanding of their biogeographical history and might be important for conservation genetics.

MATERIAL AND METHODS

Sampling, laboratory protocols and data acquisition

We used sequence data of the *16S* ribosomal RNA (rRNA), mitochondrial cytochrome c oxidase I (*COI*)

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and nuclear recombination activating gene 1 (*Rag1*) region of spiny frogs compiled in our previous studies (Hofmann *et al.*, 2019, 2021a). We complemented these data with newly generated sequences from 22 specimens of *Allopaa hazarensis*, two *Chrysopaa* and four *Nanorana* museum samples from the West Himalaya (for details see Supporting Information, Table S1). The additional *Nanorana* samples could not be identified at species level; they represent taxa apparently adapted to warm-temperate climatic conditions, because the specimens were collected between 1400 and 2100 m a.s.l., in the lower cloud forest zone of the West Himalaya (Miehe, 1991). We further considered sequences of *A. hazarensis* and *Nanorana* taxa that have recently been uploaded to GenBank (mainly *16S*). Sampling was performed under the permit of the Pakistan Museum of Natural History, Islamabad, Pakistan (No. PMNH/EST-1[89]/05), according to the regulations for the protection of terrestrial wild animals. We followed the laboratory procedure using primers and polymerase chain reaction (PCR) conditions, as previously described (Hofmann *et al.*, 2019). Briefly, DNA was isolated from ethanol tissues using the DNeasy Blood & Tissue Kit (Qiagen, Venlo, Netherlands). Approximately 570 bp of the 16S, 539 bp of the *COI* and 1207 bp of the *Rag1* gene were amplified; PCR products were purified using the ExoSAP-IT enzymatic clean-up (USB Europe GmbH, Staufen, Germany) and the mi-PCR Purification Kit (Metabion, Planegg, Germany) and sequenced by Macrogen Inc. (Amsterdam, The Netherlands).

Sequence alignment and phylogenetic reconstruction

The rRNA *16S* sequences were aligned based on their secondary structures using RNAsalsa v.0.8.1 (Stocsits *et al.*, 2009) and the ribosomal structure model of *Bos taurus* Linnaeus, 1758, provided with the RNAsalsa package. Protein-coding genes were aligned with MUSCLE (Edgar, 2004) using default settings in MEGA X (Kumar *et al.*, 2018). We found no ambiguities, such as deletions, insertions or stop codons, neither in the nucleotide-based nor amino acids alignments.

We performed phylogenetic analysis based on two datasets: (1) the *16S* sequence data, comprising a total of 88 samples and 572 nucleotide positions and (2) the concatenated mtDNA + nuDNA sequence alignment containing 2316 bp and 58 samples for which sequence data of at least two of the three loci were available. To reduce computational time, we considered only one sample of *Allopaa hazarensis* per locality in the concatenated sequence dataset (Supporting Information, Table S2).

We inferred a Bayesian inference (BI) tree for each of the two datasets using MrBayes v.3.2.6 (Ronquist *et al.*, 2012) considering stem and loop regions (*16S*) as partitions, as well as genes and codons. To provide secondary structure information for MrBayes we converted the RNAsalsa consensus structure output (a dot-bracket structure string) into a list of paired and unpaired positions and implemented them in the MrBayes input file. We assigned the doublet model (16 × 16; Schoniger & von Haeseler, 1999) to the rRNA stem pairs, and the standard 4 × 4 option with a Generalised time-reversible (GTR) evolutionary model to the remaining nucleotide positions. The site-specific rates were set variable. MrBayes was run for 10 million generations, sampling trees every 1000th generation. Inspection of the standard deviation of split frequencies, as well as an effective sample size value > 200 of the traces using TRACER v.1.7.1 (Rambaut *et al.*, 2018), indicated convergence of Markov chains. In all analyses, we used four parallel Markov chain Monte Carlo simulations with four chains and discarded the first 25% of the samples of each run as burn-in.

Based on the concatenated dataset we also inferred a maximum likelihood (ML) tree using RAxML-NG v.1.1.0 (Kozlov *et al.*, 2019). We used PartitionFinder v.1.1.1 (Lanfear *et al.*, 2012) to optimize the partition scheme with the following setting: branch lengths linked, corrected Akaike information criterion (AICc), greedy search algorithm, and the substitution models implemented in RAxML. RAxML-NG was then run with 20 random and 20 parsimony starting trees, 10 000 bootstrap replicates, and specifying the Felsenstein's bootstrap (FBP), as well as the recently introduced transfer bootstrap expectation (TBE; Lemoine *et al.*, 2018), as branch support metrics. Trees were visualized with FigTree v.1.4.3 (Drummond & Rambaut, 2007).

We also constructed minimum-spanning haplotype networks for *Allopaa hazarensis* for *16S* and *COI* using the software PopArt (http://popart.otago.ac.nz), and calculated genetic distances per locus between taxa with MEGA X. Sequences with > 25% missing data were excluded from haplotype networks. Nuclear heterozygote positions could not be phased because most populations were represented by only a few or single individuals which did not allow a robust statistical inference of haplotypes. Therefore, network reconstruction was not applied to *Rag1* sequence data.

Molecular dating

Using BEAST2 v.2.6.7 we reconstructed a dated phylogeny, based on the concatenated dataset, partitioned by genes and codons, and our previous calibration approach (Hofmann *et al.*, 2021a). Briefly, we imposed the following age constraints derived from fossil-calibrated divergence estimates (Hofmann *et al.*,

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2019): most recent common ancestor (MRCA) of Paini 38.10 Mya, 28.70–47.50 (normal distribution, sigma: 4.80); split of Tibetan *Nanorana* and Himalayan *Paa* 12.59 Mya, 7.93–17.30 (normal distribution, sigma: 2.38); separation of the Plateau frog *Nanorana parkeri* (Stejneger, 1927) and *Nanorana ventripunctata* Fei & Huang, 1985 + *Nanorana pleskei* Günther, 1896 *c*. 6.35 Mya, 3.54–9.16 (normal distribution, sigma: 1.44). We performed three runs, each with a chain length of 50 million, a thinning range of 5000, a log-normal relaxed clock model, a birth–death tree prior, a random starting tree and the site models selected by the bModelTest package (Bouckaert & Drummond, 2017) implemented in BEAST2. Runs were then combined with BEAST2 LogCombiner v.2.6.7 by resampling trees from the posterior distributions at a lower frequency, resulting in 12 500 trees. Stationary levels and convergence of the runs were verified with TRACER based on the average standard deviation of split frequencies and ESS values > 200. The final tree was obtained with TreeAnnotator v.2.6.7 and visualized with FigTree.

Occurrence data and species distribution modelling (SDM) computation

We performed SDM to assess the geographical distribution of suitable climatic conditions for both taxa separately. A total of 38 individual records of *Allopaa hazarensis* and 43 of *Chrysopaa sternosignata* were available to us (Supporting Information, Table S2), including our own observations, data from literature (Ohler & Dubois, 2006; Akram *et al.*, 2022) and databases like VertNet (http://vertnet.org) and GBIF (https://gbif.org). Grids of 19 standard bioclimatic variables for the current climate (WorldClim v.2.1 climate data for 1970–2000) and elevation were downloaded at a resolution of 30 arc-seconds (~1 km grid cells at the equator) from http://www.worldclim.org (accessed on 24 July 2022; Fick & Hijmans, 2017). All layers were projected to WGS84 and clipped to the spatial modelling extent, covering the West Himalaya, Hindu Kush and adjacent regions. Prior to the modelling approach we carried out a principal components analysis (PCA) with the SDMtoolbox v.2.5 (Brown, 2014) available for ArcGIS v.10.8, reducing the clipped layers to three orthogonal principal components describing the majority (> 99%) of the variability in climate. We then used these three components to assess the climatic heterogeneity across the area of interest. To eliminate spatial clusters of species localities we filtered our presence data by Euclidian distances (min. 1 km, max. 5 km; three distance classes) according to climate heterogeneity, using the rarefying module in SDMtoolbox, resulting in the exclusion of 13 sites for the *A. hazarensis* and five for the *C. sternosignata* occurrence dataset. Because climatic variables are often highly correlated, we explored all climate variables and the elevation data for potential multicollinearity by calculating squared Pearson's correlation coefficients (r^2) using the respective python script in the SDMtoolbox and removed highly correlated variables. The remaining variables, namely BIO2 = mean diurnal temperature range, BIO6 = minimal temperature of coldest month, BIO8 = mean temperature of wettest quarter, BIO9 = mean temperature of driest quarter, BIO12 = annual precipitation and elevation had a mean intercorrelation of $r^2 = 0.27$ (range 0.012–0.811, SD = 0.295; Supporting Information, Table S3), and were than included in the model. We restricted background selection with a buffered minimum-convex polygons based on the known occurrences and using a buffer size of 300 km. Modelling was performed with MaxEnt v.3.4.3 (Phillips *et al.*, 2004, 2006), which implements the maximum entropy algorithm that is appropriate for analysing presence-only data and has proven high predictive accuracy compared with other modelling approaches (Elith *et al.*, 2006) and robustness for small sample sizes (Pearson *et al.*, 2006). We set different regularization multipliers (1; 1.5; 2; 2.5) to optimize model performance. Each model was generated using 80% of the species records for model training and 20% for model evaluation applying a bootstrap approach, specifying five spatial groups, and using the 'equal sensitivity and specificity threshold' as the minimum threshold above which the species is considered to be present. We also tested a minimum training presence (MTP) threshold to classify average continuous probabilities of the MaxEnt model into binary maps. The MTP value is frequently used in biomod2 approaches (see below); it represents the lowest value observed in the continuous prediction map at a presence location for a specific species.

Model performance and the importance of the environmental variables to the model were assessed using the mean area under the curve (AUC) of the receiver operating characteristics (ROC; Hanley & McNeil, 1982), which is a common measure of model accuracy (Swets, 1988), and jack-knife testing. Models with AUC values above 0.7 are considered potentially informative, good between 0.8 and 0.9, and excellent for AUC between 0.9 and 1 (Swets, 1988; Elith *et al.*, 2006; Préau *et al.*, 2018).

For reasons of comparison we also performed SDM modelling using an ensemble approach in biomod2 (Thuiller *et al.*, 2013, 2021) using the following algorithms: generalized linear models (GLM; McCullagh & Nelder, 1989), generalized additive models (GAM; Hastie & Tibishirani, 1990), generalized boosted models (GBM; Ridgeway, 1999), classification tree analysis (CTA, Breiman *et al.*, 1984) and artificial neural networks (ANN; Ripley, 1996). Model performance was internally tested in biomod2 via a ten-fold data-splitting approach (80% training/20% test)

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and quantified using AUC (Hanley & McNeil, 1982), Cohen's Kappa (Monserud & Leemans, 1992) and the true skill statistic (TSS; Allouche *et al.*, 2006). The relative performance of the models was compared to derive consensus predictions based on all models with both AUC and TSS > 0.7, weighted by their proportional predictive performance on test data. The extrapolation areas were masked in the final maps and they were rescaled to 0–1, applying the MTP as threshold.

To test for environmental niche divergence among the two species we performed the niche identity test implemented in ENMTOOLS v.1.4.4 (Warren *et al.*, 2010), based on Schoener's *D* (Schoener, 1968), as recommended by Rödder & Engler (2011), and a measure derived from Hellinger distance called *I* (Warren *et al.*, 2008). These metrics measure the similarity of two distribution models as an indicator of niche overlap, ranging from zero (no overlap) to one (complete overlap). We tested against the null hypothesis of SDMs being identical by randomly generating a distribution of niche overlap values with unknown species identities to which the observed overlap of Schoener's *D* and *I* is compared (Miller & Franklin, 2002; McIntyre, 2012). The null hypothesis was rejected if the observed value of niche overlap between two taxa falls outside the 95% confidence limits ($P < 0.05$) of the simulated values. Statistical significance against the null hypothesis was determined by 100 pseudoreplicates, and the number of background points were set to 10 000. For niche quantification, only the MAXENT models were used.

RESULTS

PHYLOGENY AND GENETIC DIVERSITY

The BI and ML gene trees for the partitioned concatenated dataset (mtDNA + nuDNA) were well resolved with almost identical topologies, while in the *16S* tree most clades remained weakly supported (Fig. 1; Supporting Information, Fig. S2). However, consistent with previous findings, *Chrysopaa* was placed basally relative to the genus *Nanorana* and *Allopaa hazarensis*. All analysis recovered a placement of *A. hazarensis* within *Nanorana*, rendering that genus in the sense of Frost *et al.* (2022) paraphyletic. In fact, *A. hazarensis* formed a highly supported monophyletic clade that constitutes the sister-position to the *Chaparana* subgroup (concatenated analysis) or, based on *16S* and only weakly supported, to *Chaparana* + *Nanorana liebigii* (Günther, 1860) (*Paa*) + the clade with the unidentified specimens from the West Himalaya. The so far unidentified specimens from the West Himalaya formed a separate clade within *Nanorana*, beside *Chaparana*, *Paa* and the nominal *Nanorana* (Fig. 1). Noteworthy, species from the eastern Himalaya [*Nanorana chayuensis* (Ye, 1977), *Nanorana conaensis* (Fei & Huang, 1981), *Nanorana maculosa* (Liu *et al.*, 1960) and *Nanorana medogenesis* (Fei & Ye, 1999)] clustered together with lineages from the West Himalaya, except for the unidentified specimens, while all lineages from the central Himalaya formed an own distinct clade. Moreover, it appears that several undescribed species may exist within Himalayan *Paa*, and some taxa branched in multiple subclades, e.g. *N. liebigii*, *Nanorana vicina* (Stoliczka, 1872), indicating high intraspecific variation or potential cryptic diversity (Fig. 1). Also, our *16S* phylogeny (Supporting Information, Fig. S2), as well as the estimates of genetic distances (Supporting Information, Table S4), showed the taxonomical misclassification of the species *Nanorana arunachalensis* (Saikia, Sinha & Kharkongor, 2017) within *Nanorana*.

The average genetic distances between *Allopaa hazarensis* or *Chrysopaa sternosignata* and *Nanorana* species ranged between ~5–8% (*16S*), ~13–21% (*COI*) and ~2–4% (*Rag1*), except for *N. arunachalensis*, which was more than 14% (*16S*) distant from all taxa (Supporting Information, Tables S4, S5a, b). Within *A. hazarensis*, genetic distances were present but relatively low (Supporting Information, Table S6a–c), ranging up to 1.5% (*16S*), 0.8% (*COI*) and 0.3% (*Rag1*). Similarly, overall nucleotide variability was small. Haplotype networks for the three genes were geographically less structured, with eight (*16S*) and six (*COI*) haplotypes (Fig. 2). Populations east and west of the Indus did not share any *COI* haplotype. However, in *16S* the haplotype h7 was present on both sides of the Indus River (Fig. 2), indicating at least some exchange between localities. Most *COI*-haplotypes were divergent by one or two mutations, indicating a more recent origin and no major divergence among these sequences.

The four distinct phylogenetic groups (*Allopaa*, *Chaparana*, *Nanorana* and *Paa*) were close to each other genetically with genetic distances between them of similar magnitudes (14–17%, *COI*; 2.3–3.6% *Rag1*; Supporting Information, Table S7a, b). In contrast, the pairwise distances between the *Nanorana* clades (including *Allopaa*) and *Chrysopaa* were slightly higher in *COI*, ranging between 17 and 21%, but not in the nuclear *Rag1* (2.1–3.1%). Since main clades were less supported in the *16S* tree, we did not estimate genetic distances for this locus.

DIVERGENCE TIME

Separation of *Quasipaa* Dubois, 1992 (southern China, South-East Asia) and *Chrysopaa* from the other Paini lineages of the HTO occurred in the Mid-Oligocene between *c.* 28 Mya (20.7–35.5 Mya) and 30 Mya (22.2–37.6 Mya) (Supporting Information, Fig. S3). The warm-temperate sister-clades *Allopaa* (Kashmir Himalaya) and *Chaparana* (eastern margin of the

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Figure 1. Bayesian inference (BI; left) and maximum likelihood tree (ML; right) based on concatenated mtDNA and nDNA sequence data (*16S* + *COI* + *Rag1*) of the tribe Paini. Numbers at branch nodes refer to posterior probabilities ≥ 0.9 (BI tree), as well as Felsenstein's bootstrap values ≥ 70% and transfer bootstrap expectation ≥ 0.9 (ML tree). Branches of *Allopaa hazarensis* are indicated red, while *Chrysopaa sternosignata* is highlighted green. Species names are followed by voucher number (if available). Coloured shaded boxes indicate subgroups of *Nanorana* and the new clade (in yellow) with so far unidentified specimens.

HTO) split from the Himalayan *Paa* and Tibetan *Nanorana* in the Early Miocene, around 19 Mya (14.1–23.7 Mya), and diverged only slightly later *c.* 16 Mya (11.4–20.4 Mya). A similar age (17 Mya; 13.9–21.5 Mya) was estimated for the newly discovered clade from the West Himalaya, while the ancestral Plateau lineage (*Nanorana*) appeared around 14 Mya (11.1–17.8 Mya), although that lineage diversified only in the Late Miocene and Pliocene (7–4 Mya). The clade comprising lineages from western and eastern Himalaya (*Paa*) split from the central Himalayan clade around 11 Mya (7.7–13.4 Mya). Diversification in both of these clades had then taken place continuously during the whole Late Cenozoic. Our estimated divergence times (Supporting Information, Fig. S3) were consistent with previous results (Hofmann *et al.*, 2019, 2021a).

Species distribution model

All records are located in montane regions between *c.* 700 and 2100 m a.s.l. (*Allopaa hazarensis*), and between *c.* 1000 and 3000 m a.s.l. (*Chrysopaa sternosignata*), respectively. These regions are characterized by a heterogeneous climate (Supporting Information, Fig. S4), particularly those along the Himalaya. The average performance of our MaxEnt models was considered significantly better than random (mean $AUC_{Allopaa}$ = 0.978; mean $AUC_{Chrysopaa}$ = 0.802; Supporting Information, Table S8). The highest

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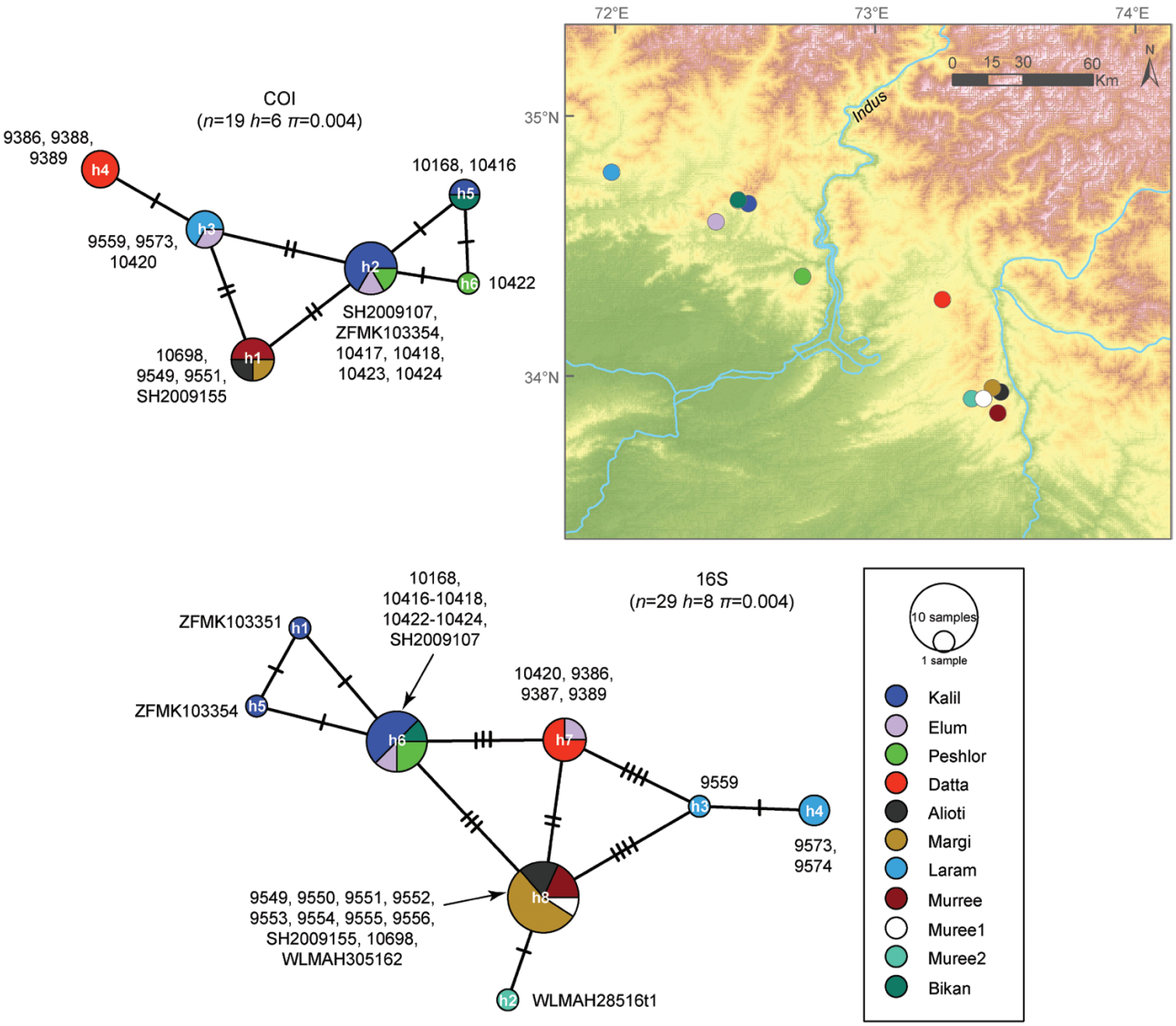

Figure 2. Minimum-spanning haplotype networks of *Allopaa hazarensis* generated for *16S* and *COI* sequence data with the number of used sequences, detected haplotypes, and the level of nucleotide variability. Symbol sizes reflect haplotype frequencies, and a small black line between two haplotypes corresponds to one mutation step. Sequence-IDs are indicated for each haplotype (h1–h8). Map shows the localities from where the respective haplotypes originate.

probability for the distribution of *A. hazarensis* is indicated for the southern foothills of the northwestern Himalaya across northern Pakistan, India and Nepal, with a warm-temperate climate (Fig. 3A), while *C. sternosignata* occurs in arid regions of the Hindu Kush and its southern and western extensions, e.g. the Paropamisus Mountains and Sulaiman mountain ranges (Fig. 3B). According to the MAXENT model, a distribution of *A. hazarensis* in the southwestern area of the Hindu Kush is also plausible, e.g. along the Spīn Ghar mountain range. The potential distribution range of *Chrysopaa* connects to the Iranian plateau and Zagros Mountains, where the species may find favourable conditions (note: the species has never been recorded from there). When applying the MTP threshold to the MAXENT results, the potential distribution range of *Chrysopaa* also encompasses parts of the Karakoram Range, and the Tibetan Plateau (Supporting Information, Fig. S5).

The performance of the biomod2 ensembles was likewise excellent, showing high-performance values, as well as sensitivity and specificity ($AUC_{Allopaa}$ = 0.998; mean $AUC_{Chrysopaa}$ = 0.977; Supporting Information, Tables S9, S10). Compared to the MAXENT models, the ensembles predicted a basically similar potential distribution for *C. sternosignata* and a more restricted range

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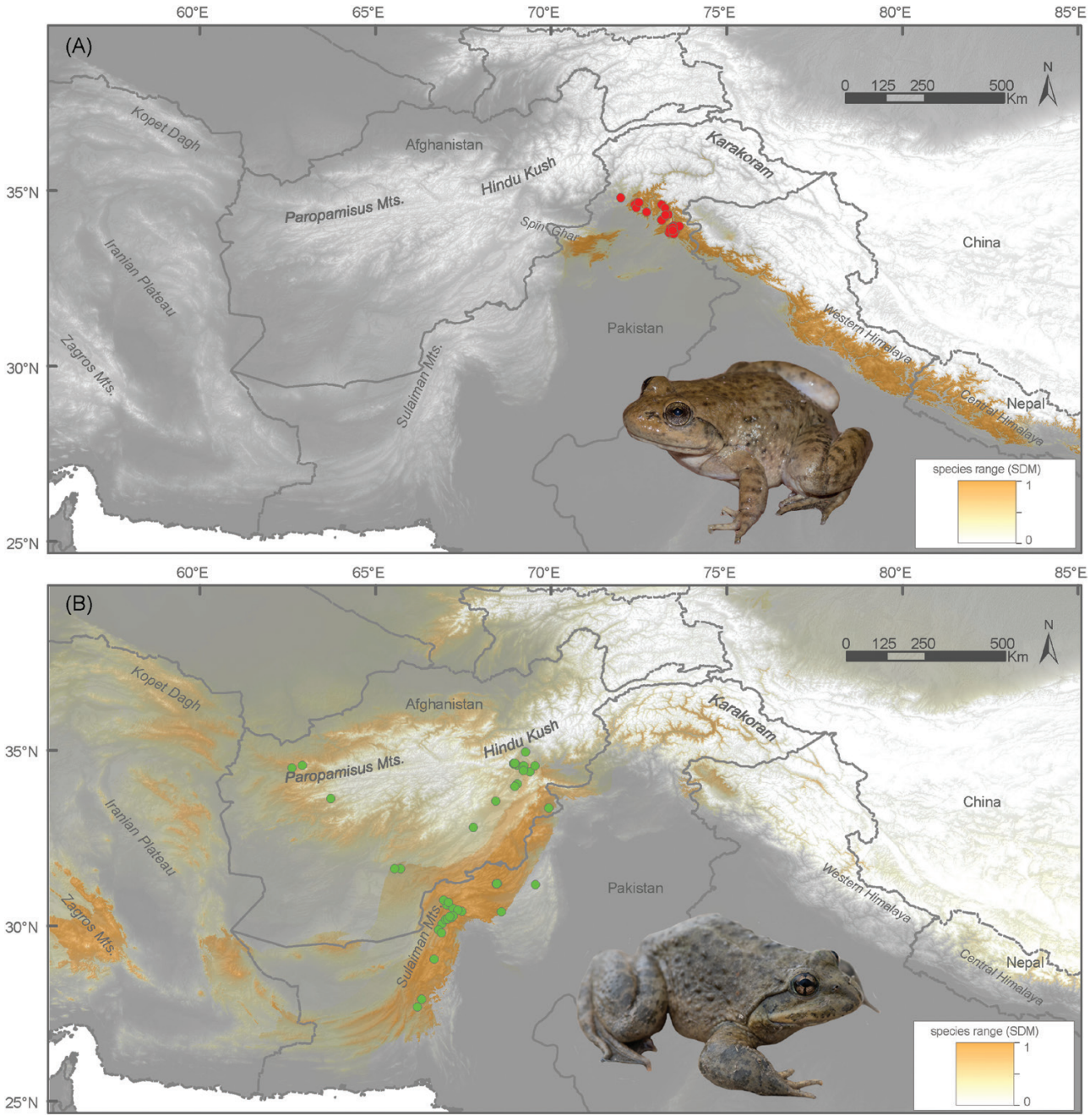

Figure 3. Distribution map for *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (B) derived from species distribution model (SDM) using MAXENT, including known records of the species (red = *A. hazarensis*, green = *C. sternosignata*). Photo credit: D. Jablonski.

with high distribution probability of *A. hazarensis* in northern Pakistan (Supporting Information, Fig. S6).

The evaluation of the variable contribution in the MAXENT models implies that for both *Allopaa hazarensis* and *Chrysopaa sternosignata*, BIO8 (mean temperature of wettest quarter) was most important to the MAXENT model (Supporting Information, Table S8). Raster values for the original occurrence points were highly significant between the two species (two-tailed test $P < 0.001$, d.f. = 138; Supporting Information, Fig. S7).

Biomod2 results showed that for *Allopaa hazarensis* BIO12 (annual precipitation) followed by BIO6 (minimal temperature of coldest month), and for *Chrysopaa sternosignata* elevation were most important to the ensemble model (Supporting Information, Table

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S10). This is well in concert with the results from the MAXENT modelling, where these variables had the most useful information by themselves, since they had the highest gain when used in isolation.

Overall, the species did not show equivalent niche patterns under current climatic conditions in the western HTO ($D = 0.028$, $I = 0.094$). The niche identity tests revealed significant environmental divergence in the SDM comparison, since the observed values of overlap were smaller than the null distributions of background divergence, indicating that the niches of *A. hazarensis* and *C. sternosignata* are different ($P < 0.05$; Supporting Information, Table S11; Fig. S8).

DISCUSSION

HISTORICAL BIOGEOGRAPHY OF HINDU KUSH–HIMALAYAN SPINY FROGS

Consistent with former studies (Che *et al.*, 2010; Hofmann *et al.*, 2019), the South-East Asian *Quasipaa* is sister to all other spiny frogs. Our results also confirm the basal phylogenetic placement of *Chrysopaa* from the Hindu Kush mountains relative to *Allopaa* and *Nanorana* (Hofmann *et al.*, 2021a), and strongly support *A. hazarensis* nested in *Nanorana*, rendering *Nanorana* paraphyletic. In previous work, the paraphyletic nature of *Nanorana* has been already indicated, although its monophyly could not be rejected (Akram *et al.*, 2021; Hofmann *et al.*, 2021b). Phylogenetic resolution and support in weakly supported parts of the tree can be increased by adding more data for a single taxon without adding more characters/genes (San Mauro *et al.*, 2012). Here, we included for the first time a number of additional *A. hazarensis* and *Nanorana* samples from the West Himalaya in the tree, which enhanced support for previously less robust internal branches.

So far, three subgroups are distinguished in *Nanorana*, namely *Chaparana* from montane regions of the southeastern margin of the Tibet Plateau (TP) and mountains of north-eastern China, *Paa* from montane to high-montane regions of the West, Central and East Himalaya, and nominal *Nanorana* from high-montane and alpine regions of the TP and its eastern margin. The strongly supported finding that *Allopaa* is phylogenetically most closely related to *Chaparana*, which occurs at the diametrically opposite end of the HTO near the ancestral area of spiny frogs (Che *et al.*, 2010; Hofmann *et al.*, 2019), strengthens support for the recently proposed hypothesis of a trans-Tibet dispersal of ancestral lineages during the Palaeogene (Hofmann *et al.*, 2021a). Accordingly, it is assumed that the ancestor of *Allopaa* (and *Chrysopaa*) appeared elsewhere near the eastern margin of the developing mountains during the Late Oligocene–Early Miocene (between 28 and 19 Mya; Supporting Information, Fig. S3) and expanded their range up to the western margin of the Himalayan–Tibetan orogenic system. This movement must have been facilitated by a moderately elevated corridor in the Late Oligocene–Early Miocene 'Tibet' (this area should not be perceived like the alpine plateau today) with subtropical and warm-temperate climates and associated sufficient humidity. The current climatic niche differentiation of *A. hazarensis* and *Chrysopaa* fits to this hypothesis. Our scenario is also consistent with subtropical to warm-temperate fossil floras in significant parts of Tibet's interior between 26 and 19 Mya (Ding *et al.*, 2014; Sun *et al.*, 2014; Ai *et al.*, 2019), and with modern geoscientific models of the HTO (Spicer *et al.*, 2021; Xiong *et al.*, 2022). The position and deep divergence of the newly discovered clade with some unidentified specimens endemic to a small area in the West Himalaya (Fig. 1; Supporting Information, Fig. S9) further supports the out-of-Tibet-into-the-Himalayan-exile hypothesis (Schmidt *et al.*, 2012; Hofmann *et al.*, 2019). Because species of this clade are adapted to the warm-temperate zone, as is also observed for *Allopaa*, the existence of large-scale, warm-temperate environments north of the greater Himalaya during the early Neogene has to be assumed (~17 Mya, Supporting Information, Fig. S3; see also: Hofmann *et al.*, 2019, 2021a). The well-supported sister-group position of the newly discovered clade to the Tibetan *Nanorana* and Himalayan *Paa* indicates that this group evolved in the southern parts of palaeo-Tibet and subsequently diversified in the West Himalaya using transverse valleys as immigration corridors. This scenario is similar to the distributional history of wingless ground beetle lineages, which share habitat preferences and distributional patterns, as seen in spiny frogs (Schmidt *et al.*, 2012). Due to the extreme topographic dynamics of the Greater Himalaya and low dispersal capacity of both wingless ground beetles and spiny frogs, these organisms are unable to disperse paralleling this mountain chain. Therefore, most lineages remain endemic to, and diversify within, restricted parts of the Himalaya. The new clade also confirms our previous expectation of the existence of additional, so far unknown lineages endemic to the Kashmir and West Himalaya, which may contribute to resolve the evolution of the HTO (Hofmann *et al.*, 2019).

Similarly, in the *Paa* subgroup, the western Himalayan lineages, including *Nanorana vicina* (Stoliczka, 1872), cluster together with species from the eastern parts of the Himalaya and Transhimalaya [*Nanorana chayunesis* (Ye, 1977), *Nanorana conaensis* (Fei & Huang, 1981), *Nanorana maculosa* (Liu *et al.*, 1960), *Nanorana medogensis* (Fei & Ye, 1999); MRCA 10.5 Mya], but there are no relatives in the vast intermediate area covered by the Central Himalaya

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(Supporting Information, Fig. S9). Such a paradoxical distributional pattern can be most parsimoniously explained by the scenario described above, that is, by the evolution of ancestral lineages in palaeo-Tibet during the Miocene, and dispersal and diversification of descendant lineages in the course of the uplifting Greater Himalaya. Based on geoscientific studies, there is high uncertainty with respect to the time of uplift of certain parts of the Greater Himalaya, ranging between *c*. 15 Mya (Gébelin *et al.*, 2013) and relatively recent (Wang *et al.*, 2006). Timing of the Himalayan uplift is one of the most controversially debated aspects in the geosciences of the area and needs to be unravelled (Mulch & Chamberlain, 2006). Based on our phylogeny and previous work (Hofmann *et al*., 2017, 2019, 2021a), the Himalaya is a young geological feature.

Genetic diversity of *Allopaa hazarensis*

In contrast to the deeply divergent clades in the phylogeny of the spiny frogs, genetic diversity of *Allopaa hazarensis* across its known range is small and without a clear distribution pattern of haplotypes, according to their geographic origin. Major haplotypes are present at multiple localities. However, the Indus seems to act as physical barrier, limiting gene flow between populations. The relatively high haplotype and low nucleotide diversity suggest a recent expansion of the species. This is supported by the higher number of unique haplotypes in relation to all haplotypes (Slatkin & Hudson, 1991; Fu, 1997), although this is influenced by sample sizes. Most likely, all of these haplotypes originated from the respective predominant ancestral haplotypes after the expected population expansion. Such a pattern and interpretation has been reported before for other amphibians (Garcia-Gonzalez *et al.*, 2012; Greenwald *et al.*, 2020). Overall, neither genetic distances, nor haplotype networks and nucleotide variability indicate potential cryptic diversity within *A. hazarensis*. In general, the networks suffer from the low number of sampling sites and, therefore, we consider these results only as preliminary. Because amphibians are predominantly site-loyal and of low vagility (Vences & Wake, 2007; da Fonte *et al.*, 2019), and since *Allopaa* is highly adapted for a semi-aquatic lifestyle in mountain streams, we assume that colonization by *A. hazarensis* is mainly facilitated through (small) aquatic corridors (e.g. by rafting), not via terrestrial dispersal routes. Movement of *A. hazarensis* in natural habitats has recently been addressed using radio transmitters, suggesting almost no overland dispersal (Akram *et al.*, 2022) and movement distances of only a few meters. However, in this study data collection was limited to only eight days in September, and almost 40% of the transmitters were lost during that time, rendering the results less conclusive.

Noteworthy, according to some authors [e.g. Frost (2022) and references therein], the genus *Allopaa* consists of two species, *A. hazarensis* and *Allopaa barmoachensis* (Khan & Tasnim, 1989), the latter originally described as *Rana barmoachensis* Khan & Tasnim, 1989. Based on morphological examination of the holotype, Dubois (1992) and repeatedly Ohler and Dubois (2006) considered this taxon as a junior synonym of *hazarensis* (as *Paa*). Given these data, as well as the geographical proximity of the type locality of *A. barmoachensis* to the main distribution range of *A. hazarensis* (and an even similar elevation of the species records), we tentatively agree with the opinion that *A. barmoachensis* is a synonym of *A. hazarensis*. However, molecular and additional morphological data are required to verify that conclusion.

Taxonomic placement of *Nanorana arunachalensis*

Besides, our *16S* phylogeny and estimates of genetic distances revealed the misclassification of *Nanorana arunachalensis*. This species had been described as *Odorrana arunachalensis* (Saikia *et al.*, 2017) but was recently reassigned to *Nanorana* based on morphological considerations (Qi *et al.*, 2019). We here used the *16S* sequence data of two vouchers (ZSIS-M37: MN496464 and ZSIS-M40: MN636773) from the type locality that were uploaded to GenBank (https://www.ncbi.nlm.nih.gov/genbank/) in September 2019 by Saikia and colleagues, probably in response to the reassignment proposed by Qi *et al.* (2019). Based on our data, the species should be reclassified into *Odorrana* Fei *et al.*, 1990 as long as there are no further molecular and morphological data that support a classification into another genus.

Niche differentiation confirms phylogeny

Our species distribution models, for the first time, present the potential geographic distribution range of the relict taxa *Allopaa hazarensis* and *Chrysopaa sternosignata*, which are geographically separated with an allopatric distribution pattern. Although our MaxEnt model predicted the occurrence of *A. hazarensis* across the West and Central Himalaya (Fig. 3A), this must be rejected. Suitable habitats for the species might be available there, but there is no evidence about the presence of the genus in these areas; especially since the biomod2 ensemble model does not predict this either (Supporting Information, Fig. S6A). On the other hand, such environmental suitability supports our scenario described above and the hypothesis of a trans-Tibet dispersal by ancestral lineages of *A. hazarensis* (and *Chrysopaa*) during the Miocene/Oligocene from the eastern into

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the northwestern regions of the HTO. Consequently, a colonization of suitable habitats in the western and central parts of the Himalaya by those lineages must be excluded. In fact, there are no records of *A. hazarensis* in other than the northwestern part of the Himalaya, although that taxon is easy to detect if present due to its strictly semi-aquatic natural history. However, it remains possible that *A. hazarensis* and *Chrysopaa* may occur in other untapped areas, where suitable habitats are present, i.e. the Afghan part of the Hindu Kush and eastern Iran, respectively. Both areas are zoologically least-investigated, thus, the presence of the two taxa there cannot be excluded.

Note that there was a trend for over-estimating suitable areas when using the less restrictive MTP threshold in the MAXENT model to produce the binary habitat map, in particular for *Chrysopaa sternosignata* (Supporting Information, Fig. S5B). It is well known that SDM predictions, particularly those that are based on small occurrence data and thresholded by MTP values, can result in higher estimates (Vale *et al.*, 2013; Kass *et al.*, 2021).

CONSERVATIONAL NEEDS OF RELICT HINDU KUSH–HIMALAYA LINEAGES

Amphibians are highly vulnerable to changes in thermal and hydric environments due to their ectothermic physiology, unshelled eggs, highly permeable skin and biphasic life-cycles [Araujo *et al.* (2006) and references therein]. Thus, temperature and precipitation are among the most important factors that determine the geographical distribution and abundance of amphibian species (Carey & Alexander, 2003). In both *Allopaa hazarensis* and *Chrysopaa sternosignata*, BIO8 (temperature of the wettest month) was important to the model and was significantly different between the localities of the two species (Supporting Information, Fig. S7), probably simply because the precipitation in northern Pakistan is highest during July/August, while in western Pakistan and adjacent Afghanistan most of the annual precipitation occurs between December and March. The niche identity test analyses revealed a clear divergence of the environmental niches of the two species and indirectly confirmed the different habitat requirements of *A. hazarensis* and *Chrysopaa*. While *A. hazarensis* grows in warm-temperate and wetter places in the foothills to lower montane zone (comparable to those of its sister group *Chaparana*; Ohler *et al.*, 2000; Che *et al.*, 2010), *Chrysopaa* occurs in higher, warmer and drier environments. Reproduction of *Chrysopaa* starts in April, after the rain season; for *A. hazarensis* breeding starts probably later in June with the first summer rain (Dubois & Khan, 1979).

According to the IUCN Red List, both *Allopaa hazarensis* and *Chrysopaa sternosignata* are considered as 'least concern' (Khan *et al.*, 2004, 2008). The two species are flagged with a stable population trend: except from prolonged drought, there are supposedly no threats to *A. hazarensis*, while *Chrysopaa* faces threats from aquatic pollution and over-harvesting for science teaching and research in schools and colleges in Pakistan. However, endemic species inhabiting mountain systems are particularly exposed to the effects of global climate change [Cordier *et al.* (2019) and references therein]. Given their topographic characteristics, the environmental gradients of mountain systems are especially heterogeneous (see also Supporting Information, Fig. S4). Increasing temperatures and decreasing precipitation along these gradients could lead to a significant upward shift of suitable climatic conditions for amphibians and certain other organisms. Logically, this displacement affects species in higher elevations, like *A. hazarensis* and *C. sternosignata*, more negatively because of the reduction of their geographically explicit climatic niche (Blaustein *et al.*, 2010). Consequently, if these species are not able to adapt, they successively will become locally lost by extinction. In fact, glaciers in the northwestern Himalayan region of Jammu and Kashmir have been reported to be retreating at higher rates as compared to other parts of the Himalayan arc [Rashid *et al.* (2021) and references therein], potentially impacting severely the stream-flow regimes. Therefore, we encourage future studies related to the status, trends and fragmentation of *A. hazarensis* and, particularly, *Chrysopaa* populations, in order to determine the appropriate conservation status for these species. Compared to the geographic distribution maps in the most recent status assessment for the IUCN Red List from 2008, our predicted potential distribution of the two frogs differs from these data by a significantly better resolution and higher precision. Our results contribute to the knowledge about the distribution of these species and may provide basic information for guiding future management of them, e.g. by surveying and protecting suitable habitats.

CONCLUSIONS

The Hindu Kush–Himalaya region represents a unique area with a biodiversity that has a potentially high informative value for phylogeographic research against the context of the evolution of the HTO. However, until now, its endemic faunal (and floristic) diversity has remained least-investigated (Jablonski *et al.*, 2021). Our phylogenetic results demonstrate the close relationship of northwestern Himalayan spiny frogs to representatives in the eastern parts of the

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HTO, supporting the recently proposed out-of-Tibet-into-the-Himalayan-exile hypothesis and a trans-Tibet dispersal of ancestral spiny frogs during the Palaeogene. Given the current taxonomy (Fig. 1) and a distribution strictly linked to the Himalaya (see also: Hofmann *et al.*, 2019), the genus *Allopaa* [type species *Rana* (*Paa*) *hazarensis* Dubois & Khan, 1979] could be treated under *Nanorana* (type species *Nanorana pleskei* Günther, 1896) rather than as distinct genus. If so, the genus *Allopaa* should be synonymized with *Nanorana*.

Alternatively, *Chaparana* could be elevated to genus level (and consequently also *Paa*), maintaining *Allopaa* at genus level and resolving the paraphyletic nature of *Nanorana*. However, this would imply that each monophyletic Himalayan or Hindu Kush lineage that is identified in the future (what to expect) might be erected to genus level, potentially resulting in a significant taxonomic inflation across *Nanorana*. Considering the shared biogeographic history of these spiny frogs, the present and the new distinct clades to be discovered in the future should all be assigned to *Nanorana s.l.*, until a systematic review of this group is available. Changes in supraspecific classification (and especially at the genus level) should only be carried out on the basis of highly stable phylogenies and if clearly supported by additional information (Vences *et al.*, 2013), e.g. morphological, ecological or behavioural data. Similarly, the new *Nanorana* clade from the West Himalaya with the so far unidentified specimens has to be validated by morphological and additional molecular data. We further suggest excluding *Nanorana arunachalensis* from *Nanorana* and reassigning that species to the genus *Odorrana*.

SDM findings show distinct environmental niches and non-overlapping distribution ranges of *Allopaa hazarensis* and *Chrysopaa sternosignata* and provide crucial information for guiding potential future management of the two species. In view of the rapid global change of today, it might be expected that the species track their niches toward higher elevations or become locally extinct. Currently, understanding climate effects and consequences of other potential threats on *Allopaa* and *Chrysopaa* is hindered by limitations of existing data. Our study sets a baseline to the completion of the overall picture of the species distribution patterns and may aid our understanding to target conservation actions.

ACKNOWLEDGEMENTS

We are grateful to Svetlana D. Vershinina (†2021) for assistance in the field. We thank Jana Poláková for her work in the lab. We also thank the anonymous referees and the editors Jinzhong Fu and Maarten Christenhusz for their constructive comments. This work was supported by the German Research Foundation (DFG, grant no. HO 3792/8-1) to SH. DJ was supported by the Slovak Research and Development Agency under the contract no. APVV-19-0076. LJB was supported by the Zoological Institute within the state assignment of the Ministry of Science and Higher Education, Russian Federation, No. 122031100282-2.

DATA AVAILABILITY

All new sequences were uploaded to GenBank; details on individual samples and accession numbers are available in the Supporting Information, Tables S1 and S2.

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SUPPORTING INFORMATION

Additional supporting information may be found in the online version of this article on the publisher's website.

Table S1. GenBank accession numbers of sequences used in this study. Taxa with newly obtained sequences are indicated by an asterisk.

Table S2. Locations (Loc) of the two species (Sp) *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (C) as shown in Figure 3. Some coordinates and elevations are approximations from descriptions in the original publications. Lat(itude) and Long(itude) are given in decimal degree. Ctr = country (Pakistan [PK], Afghanistan [AFG]), Alt = elevation. Samples CUHC10278 and CUHC11352 could not be georeferenced unambiguously due to lack of information.

Table S3. Pairwise correlation of variables used for species distribution modelling: Pearson's r^2 (upper triangular) and Spearman's ρ (lower triangular).

Table S4. Uncorrected genetic distances (%; lower left matrix), including standard error estimates (upper right matrix) between *16S* sequences of taxa as shown in the Supporting Information, Figure S2. A = *Allopaa hazarensis*, C = *Chrysopaa sternosignata*. *Nanorana* species names were abbreviated after five characters and are as follows: *phrynoides*, *sichuanensis*, *zhaoermii*, *arunachalensis*, *vicina* (prefix WLMNV), *xuelinensis*, *aenea*, cf. *blanfordii*, cf. *polunini*, *rarica*, cf. *rostandi*, *chayuensis*, *conaensis*, *liebigii*, *maculosa*, *medogensis*, *pleskei*, *quadranus*, sp. (2Bahn_RAS), cf. *vicina* (2Pul_RAS, 782_RAS), sp. (A1966/13_NME), *taihangnica*, *unculuanus*, *ventripunctata*, *yunnanensis*, cf. *ercepea*, *parkeri*, *Quasipaa boulengeri*, sp. (VV5.1/8.1/11.1_RAS), cf. *vicina* (VV9.1_RAS). A star after the abbreviated species name refers to 'cf. '. Grey shaded cells indicate the large genetic distance between *N. arunachalensis* and other spiny frogs, including *Nanorana* species, showing the taxonomic misclassification of that species. Values in coloured cells are distances < 2.5%.

Tables S5. a,b Uncorrected genetic distances (%; lower left matrix), including standard error estimates (upper right matrix), between taxa used in this study for *COI* (a) and *Rag1* (b). A star after the species name refers to 'cf. '. Bold values in grey shaded cells highlight distances lower than 5% (*COI*) and lower than 0.1% (*Rag1*). A = *Allopaa hazarensis*, C = *Chrysopaa sternosignata*, N = *Nanorana*, Q = *Quasipaa boulengeri*.

Table S6. a–c Uncorrected pairwise genetic distances between sequences of *Allopaa hazarensis* for *16S* (a), *COI* (b) and *Rag1* (c). Distances ≥ 1% (*16S*) and ≥ 0.5% (*COI*) and ≥ 0.2% (*Rag1*) are highlighted bold. [#]CUHC = Comenius University Herpetological Collection; *=prefix 'WLM:AH'; [$]=ZFMK(SH20) (Museum Koenig, Bonn).

Table S7. a,b Uncorrected genetic distances (%; lower left matrix) with standard error (upper right matrix) between *Allopaa hazarensis* (A), *Chrysopaa* (C), *Quasipaa* (Q) and subgenera of *Nanorana* (*Chaparana* [Ch], *Nanorana* [N], *Paa* [P]), including the unknown clade from the Western Himalaya (WH) for *COI* (a) and *Rag1* (b).

Table S8. SDM performance and evaluation for *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (C). Information on the model performance and evaluation, and the variable contribution are given. Test AUC values indicate performance as follows: > 0.9 excellent, > 0.8 good, and > 0.7 useful discrimination ability of the model. High values in variable contribution are highlighted bold.

Table S9. Performance evaluation scores for the final ensemble classifier combining biomod2 algorithms.

Table S10. Statistics for single models specified in biomod2; (a) for *Allopaa hazarensis* and (b) *Chrysopaa sternosignata*.

Table S11. Observed niche overlap values and results of niche identity test. Empirical overlap values smaller than the null distribution support niche divergence. Asterisk denote significance at $^{*}P < 0.05$.

Figure S1. PCA of 19 WorldClim v.2.1 variables, showing climate space: the more similar the colours the more similar values. Records of *Chrysopaa sternosignata* are indicated as green filled circles (for details see Supporting

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Information, Table S1). The arrow points to Malir, Pakistan, that is noted as type locality [Mulleer (= Malir) near Kurrachee (= Karachi)] for the syntypes BMNH 1947.2.1.21, 1947.2.1.22 of *C. sternosignata*. Malir has apparently substantially different climate conditions compared to the known distribution range of the species. Thus, we consider that type locality as erroneously defined.

Figure S2. Bayesian inference tree based on *16S* rRNA sequence data. Numbers at branch nodes refer to posterior probabilities ≥ 0.9. The clade of *Allopaa hazarensis* is indicated red, while *Chrysopaa sternosignata* is highlighted green. Species names are followed by voucher number (if available).

Figure S3. Ultametric time-calibrated phylogeny generated with BEAST2 based on the concatenated sequence data of spiny frogs. Grey bars specify the 95% HPD for the respective nodes; ages are shown for nodes that are supported by Bayesian posterior probability ≥ 0.95.

Figure S4. Climate heterogeneity raster based on recent WorldClim v.2.1 data; warm colours depict high areas of climatic heterogeneity.

Figure S5. Distribution map for *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (B) derived from species distribution model (SDM) using MAXENT and a minimum training presence threshold. Maps include known records of the species (red = *A. hazarensis*, green = *C. sternosignata*).

Figure S6. Distribution map for *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (B) derived from species distribution model (SDM) using biomod2, including known records of the species (red = *A. hazarensis*, green = *C. sternosignata*).

Figure S7. Environmental variable BIO8 (mean temperature of wettest quarter) across the modelled area based on WorldClim v.2.1 climate data for 1970–2000. Records of *Allopaa hazarensis* and *Chrysopaa sternosignata* are indicated by red and green circles, respectively.

Figure S8. Sample output from ENMTOOLS v.1.4.4 for identity test for all three implemented niche overlap metrics. Histograms represent the distribution of overlaps for each metric from the null distribution, while the dashed vertical line represents the overlap between the models built using the empirical data.

Figure S9. Simplified Bayesian inference tree with the main (sub)genera mapped to High Asia with the Indus and Brahmaputra River systems. Topology and colour codes of the clades match Figure 1.

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Supporting Information

Endemic lineages of spiny frogs demonstrate the biogeographic importance and conservational needs of the Hindu Kush Himalaya region

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Table S1. GenBank accession numbers of sequences used in this study. Taxa with newly obtained sequences are indicated by an asterisk.

Taxon	SampleID/voucher	*16S*	*COI*	*Rag1*
Allopaa hazarensis	CUHC 9386	MW598397	MW603002	MW598465
Allopaa hazarensis	CUHC 9389	MW598398	MW603003	MW598466
Allopaa hazarensis	CUHC 9549	MW598393	MW603004	MW598461
Allopaa hazarensis	CUHC 9551	MW598394	MW603005	MW598462
Allopaa hazarensis	CUHC 9559	MW598395	MW603006	MW598463
Allopaa hazarensis	CUHC 9573	MW598396	MW603007	MW598464
Allopaa hazarensis	WLM:AH28516t1	MW898163		
Allopaa hazarensis	WLM:AH305162	MW898162		
Allopaa hazarensis	ZFMK 103351	MW723172	MW723177	MW728951
Allopaa hazarensis	ZFMK 103354	MW723173	MW723178	MW728952
Chrysopaa sternosignata	USNM Herp 589843	MG700154		
Chrysopaa sternosignata	USNM Herp 589844	MG700155	MG699938	
Chrysopaa sternosignata	USNM Herp 589845	MG700153	MG699937	
Nanorana aenea		EU979830	KR087830	HM163609
Nanorana arunachalensis	ZSIS-M40	MN636773		
Nanorana cf. *blanfordii*	JS040529_NME	MN012067		MN032491
Nanorana cf. *blanfordii*	JS040534_NME	MN012071		MN032495
Nanorana cf. ercepeae	A1_12_NME	MN012077	MN012213	MN032501
Nanorana cf. *polunini*	SH070507_NME	MN012084	MN012219	MN032508
Nanorana cf. *rarica*	A2015/13_NME	MN012204	MN012324	MN032607
Nanorana cf. *rostandi*	R11_12_NME	MN012092	MN012227	MN032516
Nanorana chayuensis	SCUM050410CHX	EU979838		HM163587
Nanorana chayuensis	67X		JN700888	
Nanorana conaensis	KIZ-YP152	EU979834		HM163589
Nanorana liebigii	A17_12_NME	MN012104	MN012237	MN032528
Nanorana liebigii	SH070515_NME	MN012106		MN032530
Nanorana liebigii	SH080506_NME	MN012108		MN032532
Nanorana liebigii	JS040512_NME	MN012119	MN012246	MN032542
Nanorana liebigii	JS040513_NME	MN012120	MN012247	MN032543
Nanorana liebigii	KIZ-RDXZL1	DQ118499	KJ810987	HM163607
Nanorana maculosa	YNU-HU2002308	EU979835		HM163588
Nanorana medogensis	SYNU-XZ35	DQ118506		HM163590
Nanorana parkeri	JS0507B01_NME	MN012143	MN012265	MN032556
Nanorana phrynoides	CIBYN2008053001	KU139986		
Nanorana pleskei	KQ1_14_NME	MN012157	MN012279	MN032563
Nanorana quadranus	SCUM20045195CJ	DQ118514		HM163591
Nanorana sichuanensis	CIBYN2008053013	KU140028		
Nanorana rarica	KQ2_12_NME	MN012171	MN012292	MN032574
Nanorana sp.	A1966/13_NME	MN012198	MN012318	MN032599
Nanorana cf. *vicina*	2Bhan_RAS	MN012199	MN012319	
Nanorana vicina	2Pul_RAS	MN012200		MN032604
Nanorana vicina	782_RAS	MN012201		MN032605
Nanorana vicina	WLM:NV28917	MW898173		
Nanorana vicina	WLM:NV289171	MW898174		
Nanorana vicina	WLM:NV299172	MW898175		

Nanorana vicina	WLM:NV1310171	MW898178		
Nanorana taihangnica		KF199146	KF199146	HM163608
Nanorana unculuanus	YNUHU2002502601	DQ118491		HM163595
Nanorana ventripunctata	SH050538_NME	MN012208	MN012328	MN032610
Nanorana xuelinensis	KIZL2019014	MZ410626		
Nanorana yunnanensis		KF199150	KF199150	HM163593
Nanorana zhaoermii	SYNU-1706049	MH315956		
Quasipaa boulengeri	YNU-HU20025106	KX645665	KX645665	HM163604
Quasipaa delacouri	FMNH 255623	EU979810	EU979664	HM163600
Quasipaa exilispinosa	KF199151	KF199151	KF199151	HM163610
Quasipaa jiulongensis	KF199149	KF199149	KF199149	HM163603
Quasipaa shini	KF199148	KF199148	KF199148	HM163602
Quasipaa spinosa		NC_013270	NC_013270	HM163606
Quasipaa verrucospinosa		KF199147	KF199147	HM163599
Quasipaa yei	HNNU0908I061	KJ842105	KJ842105	HM163596
Fejervarya cancrivora		EU652694	EU652694	HM163581
Hoplobatrachus rugulosus		NC_019615	NC_019615	HM163612
Limnonectes fragilis	ZNAC11006	AY899241	AY899241	HM163611
*Allopaa hazarensis**	CUHC 10168	OP173761	OP601386	OP204875
*Allopaa hazarensis**	CUHC 10278	OP601360	OP601387	
*Allopaa hazarensis**	CUHC 10416	OP173762	OP601388	OP204876
*Allopaa hazarensis**	CUHC 10417	OP173763	OP601389	OP617248
*Allopaa hazarensis**	CUHC 10418	OP173764	OP601390	OP204877
*Allopaa hazarensis**	CUHC 10420	OP173765	OP601391	OP204878
*Allopaa hazarensis**	CUHC 10422	OP173766	OP601392	OP204879
*Allopaa hazarensis**	CUHC 10423	OP601361	OP601393	
*Allopaa hazarensis**	CUHC 10424	OP601362	OP601394	
*Allopaa hazarensis**	CUHC 10698	OP173767	OP601395	OP617249
*Allopaa hazarensis**	CUHC 11352		OP601396	
*Allopaa hazarensis**	CUHC 9387	OP173768		OP204880
*Allopaa hazarensis**	CUHC 9388		OP174420	OP204881
*Allopaa hazarensis**	CUHC 9550	OP173769		
*Allopaa hazarensis**	CUHC 9552	OP173770		OP617242
*Allopaa hazarensis**	CUHC 9553	OP173771		OP617243
*Allopaa hazarensis**	CUHC 9554	OP173772		OP617244
*Allopaa hazarensis**	CUHC 9555	OP173773		OP617245
*Allopaa hazarensis**	CUHC 9556	OP173774		OP617246
*Allopaa hazarensis**	CUHC 9574	OP173775		OP617247
*Allopaa hazarensis**	ZFMK SH2009107	OP173776	OP174421	OP204882
*Allopaa hazarensis**	ZFMK 103387	OP173777	OP174422	OP204883
*Chrysopaa sternosignata**	CUHC 9561	OP173778	OP601397	OP617250
*Chrysopaa sternosignata**	CUHC 9562	OP173779	OP601398	OP617251
Nanorana sp.*	RAS VV5.1	OP173780	OP174423	OP204884
Nanorana sp.*	RAS VV8.1	OP173781	OP174424	OP204885
Nanorana sp.*	RAS VV9.1	OP173782	OP174425	OP204886
Nanorana sp.*	RAS VV11.1	OP173783	OP174426	OP204887

Museum/Institution abbreviations of voucher specimens from which sequences were newly obtained are as follows: CUHC = Comenius University Herpetological Collection; RAS = Russian Academy of Science; ZFMK = Zoological Research Museum Alexander Koenig.

Table S2. Locations (Loc) of the two species (Sp) *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (C) as shown in Fig. 3. Some coordinates and elevations are approximations from descriptions in the original publications. Lat(idute) and Long(itude) are given in decimal degree. Ctr=country (Pakistan [PK], Afghanistan [AFG]), Alt=altitude. Samples used for phylogenetic analysis (concatenated sequence data set) are indicated bold. Samples CUHC10278 and CUHC11352 could not be georeferenced unambiguously due to lack of information.

ID	*Sp*	*Loc*	*Ctr*	*Lat*	*Long*	*Alt*	*Source*
WLM:AH252171	A	Murree	PK	33.7970	73.3767	986	[1]
WLM:AH265161,11	A	Murree	PK	33.8848	73.5004	1784	[1]
WLM:AH265163	A	Murree	PK	33.8432	73.4694	1686	[1]
WLM:AH28516t1	A	Murree	PK	33.9143	73.3939	2018	[1]
WLM:AH289173	A	Kotli Sattian	PK	33.8137	73.5011	1647	[1]
WLM:AH289174	A	Kotli Sattian	PK	33.8229	73.5289	1437	[1]
WLM:AH289175	A	Kotli Sattian	PK	33.8137	73.5011	1647	[1]
WLM:AH296161	A	Murree	PK	33.8146	73.3688	957	[1]
WLM:AH296162	A	Murree	PK	33.7994	73.3538	1187	[1]
WLM:AH296163	A	Murree	PK	33.9878	73.4928	1184	[1]
WLM:AH299171	A	Murree	PK	33.8483	73.4294	1295	[1]
WLM:AH299173,175	A	Kotli Sattian	PK	33.7521	73.4919	1128	[1]
WLM:AH299174	A	Murree	PK	33.8432	73.4696	1688	[1]
WLM:AH305162	A	Murree	PK	33.9142	73.4167	2100	[1]
WLM:AH305164	A	Murree	PK	33.8687	73.4672	1886	[1]
WLM:AH305165	A	Murree	PK	33.9142	73.3939	2017	[1]
WLM:AH305167	A	Murree	PK	33.9012	73.4338	1923	[1]
WLM:AH305168	A	Murree	PK	33.8687	73.4672	1886	[1]
WLM:AH461611	A	Murree	PK	33.8609	73.3795	1359	[1]
MNHN RA 1978.3063	A	Abbottabad	PK	34.2500	73.2500	1307	GBIF
NHMUK ZOO 1978.795	A	near Datta, Manshera, Hazara	PK	34.2914	73.2566	1200	GBIF
UF Herp 79622	A	Battal, NW Frontier	PK	34.5868	73.1497	1497	GBIF
UF Herp 79623	A	Lassan, NW Frontier	PK	34.3076	73.3366	1542	GBIF
UF Herp 79639	A	Ichchrian [Icherrian], NW Frontier	PK	34.4918	73.2485	1026	GBIF
UF Herp 82422	A	6.8 km W. Abbottabad on road to Sherwin	PK	34.1682	73.1476	1377	GBIF
UF Herp 82735	A	6.2 km W. Abbottabad	PK	34.1514	73.1529	1385	GBIF
UF Herp 82861	A	Arja, Azad Kashmir	PK	33.9743	73.6569	833	GBIF
CUHC **9386**-9389	A	Datta	PK	34.2951	73.2576	1279	[2], this study
CUHC **9549**, 9550	A	Alioti	PK	33.9505	73.4679	1537	[2], this study
CUHC **9551**-9556	A	Margi	PK	33.9396	73.4652	1641	[2], this study
CUHC **9559**, 9573, 9574	A	Laram Qilla, Lower Dir	PK	34.7849	71.9856	1411	[2], this study
ZFMK 103351	A	Kalil, Tehsil Gadezai, Buner	PK	34.6401	72.4716	932	[2]
ZFMK 103354, ZFMK **SH2009107**	A	Kalil, Tehsil Gadezai, Buner	PK	34.6564	72.4961	1519	[2]
CUHC **10416**,10418,10424	A	Kalil, Tehsil Gadezai, Buner	PK	34.6563	72.4960	1524	this study
CUHC **10168**	A	Bikan Kalay, Buner	PK	34.6564	72.4591	1551	this study
CUHC 10417,**10420**	A	Malakpur Elum Side, Tehsil Gadezai, Buner	PK	34.5938	72.3872	1625	this study
CUHC **10422**,10423	A	Peshlor,Tehsil chagharzai, Buner	PK	34.3846	72.7200	1244	this study
CUHC 10698, ZFMK **103387**	A	SE of Murree	PK	33.8596	73.4701	1632	this study
MNHNP 1978.3056	A	Datta	PK	34.2950	73.2580	1294	Type locality
	C	Naal	PK	27.6820	66.1830	1178	[3]
	C	Baghunna	PK	27.9070	66.2980	1605	[3]
USNM:Herp:589843-45	C	Parvan, Bagram Air Force Base	AFG	34.9461	69.2650	1473	ncbi
BMNH 1940.3.1.6-9	C	Arbarp [Arbab]	AFG	34.5636	69.0150	2100	[4]
MNHN 1994.4505	C	Sinjiri [Sinjiri, Kandahar]	AFG	31.6206	65.5317	981	[4]
NMW 18608	C	Karokh	AFG	34.4911	62.6031	1342	[4]
SMF 67971, 67972	C	Ganzi [Ghanzi]	AFG	33.5451	68.4174	2180	[4]
SMF 68292	C	Kabul	AFG	34.5553	69.2075	1793	[4]

ZMK R.07288	C	Mukur [Moqor]	AFG	32.8061	67.7735	1989	[4]
AMNH 68382-68385	C	Baleli [Balochistan]	PK	30.0465	66.8737	2303	[4]
AMNH 75190-75193	C	near Mastung	PK	29.8825	66.7844	1599	[4]
AMNH 75194; AMNH 57971-57976; BMNH 1891.4.14.21-22; SMF 62835-62836; SMF 65888	C	near Quetta	PK	30.1798	66.9750	1660	[4]
AMNH 75198-75202.A-D, AMNH 68386-68389, AMNH 75195-75197	C	near Pishin	PK	30.5897	67.0107	1556	[4]
SMF 47763	C	Yaseen [Yansin valley], 14 mi. nördl. Pishin;	PK	30.7291	66.9233	1522	[4]
SMF 62833-62834	C	Dhobi Ghat, Quetta	PK	30.3916	68.5771	1461	[4]
SMF 62837	C	Hanna valley [Shal Tungi]	PK	30.1812	67.0331	1730	[4]
CUHC 9561	C	Paghman	AFG	34.6143	68.9127	2636	this study
CUHC 9562	C	Paghman	AFG	34.6143	68.9127	2636	this study
SMF 62833	C	Dhobi Ghat, Quetta	PK	30.3916	68.5771	1461	this study
UF 85699-85702	C	Mastung, Kharez Biani	PK	29.7901	66.8718	1728	VertNet.org
UF 85703-85705	C	16.6 km NE Quetta & 4 km NE Spin Karez	PK	30.2780	67.2030	2212	VertNet.org
UF 85706-85707, 85717, 85725-85739	C	Hannah Lake	PK	30.2567	67.0970	1904	VertNet.org
UF 85708	C	16.4 km W. Kach	PK	30.4934	67.1944	1740	VertNet.org
UF 85709-85710	C	Spin Karez	PK	30.2220	67.1434	1981	VertNet.org
UF 85711-85716	C	23.1 km NE Quetta, in mouth of Karez system just before water left underground channel to enter irrigation system	PK	30.4460	67.3290	1934	VertNet.org
UF 85718-85719	C	Band Khushdil Khan	PK	30.6650	67.0650	1540	VertNet.org
UF 85720-85724	C	Mastung, Kharez Biani	PK	29.7901	66.8718	1728	VertNet.org
CAS 115917-115918; FMNH 161221, 161224	C	Paghman	AFG	34.6000	68.9333	2611	[5]
CAS 133828	C	Kargha stream, near Kabul	AFG	34.6332	68.9264	3117	[5]
CAS 151216-1511219	C	Kabul She Carte	AFG	34.5315	69.1695	1800	[5]
CAS 151223-151226	C	Kurdkabul	AFG	34.3865	69.3851	2155	[5]
CAS 92330-92336	C	Logar River, 7-8 min from Kabul	AFG	34.5575	69.5267	1400	[5]
CAS 96171	C	Khost [=Khoast]	AFG	33.3489	69.9216	1200	[5]
FMNH 161279	C	Kandahar	AFG	31.6253	65.7028	1021	[5]
MNHN 1985.300	C	Sinjui [?=Band-e Sinju]	AFG	33.6261	63.7120	2133	[5]
MVZ 237438	C	ca. 4 km above Paghman	AFG	34.6284	68.9562	2423	[5]
USNM 194591-194594	C	Culangor	AFG	34.0386	69.0283	1888	[5]
USNM 194964-194965, USNM 194967-194970	C	Baraki Barak	AFG	33.9667	68.9495	1940	[5]
ZFMK 18981	C	Kabul	AFG	34.5553	69.2075	1793	[5]
BMNH 1947.2.1.22	C	Zandra	PK	30.4026	67.4394	2368	[5]
NHMW 62988	C	38km NE Herat	AFG	34.5710	62.8973	2335	this study
PMNH 1340-1341	C	Logar River, Logar Province	AFG	34.4190	69.1938	1806	this study
PMNH 1906-1932, PMNH 1959-1963	C	Zeba Nala, Nawab Dam, Tanishpa, Killa Saifullah	PK	31.1986	68.4415	2446	this study
PMNH 495-501	C	Killa Saiffulah, Tanishpa	PK	31.1978	68.4256	2645	this study
PMNH 614-615	C	Zhob, Raghey	PK	31.1722	69.5431	1788	this study
PMNH 631-638	C	Kalat	PK	29.0420	66.6595	2096	this study
PMNH 720-23	C	Killa Saiffulah, Tanishpa	PK	31.2064	68.4656	2370	this study

Museum abbreviations are as follows: AMNH = American Museum of Natural History; BMNH = British Museum of Natural History; CAS = Chinese Academy of Science; CUHC = Comenius University Herpetological Collection; FMNH = Field Museum of Natural History; MNHN(P) = Muséum national d'Histoire Naturelle Paris; MVZ = Museum of Vertebrate Zoology; NHMUK = Natural History Museum London; NMW/NHMW = Naturhistorisches Museum Wien; PMNH = Pakistan Museum of Natural History; SMF = Senckenberg Research Institute and Natural History Museum Frankfurt/M.; UF = University of Florida; USNM = Smithsonian National Museum of Natural History; ZFMK = Zoological Research Museum Alexander Koenig; ZMK = Zoological Museum of the CAU University Kiel.

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Table S3. Pairwise correlation of variables used for species distribution modeling: Pearson's r^2 (upper triangular), and Spearman's ρ (lower triangular).

	alt	bio12	bio2	bio6	bio8	bio9
alt		0.012	0.114	0.811	0.484	0.746
bio12	0.016		0.237	0.071	0.047	0.006
bio2	0.026	0.228		0.038	0.039	0.107
bio6	0.741	0.069	0.007		0.532	0.705
bio8	0.608	0.003	0.024	0.616		0.134
bio9	0.497	0.000	0.088	0.479	0.118	

Table S4. Uncorrected genetic distances (%; lower left matrix), including standard error estimates (upper right matrix) between *16S* sequences of taxa as shown in Figure S2. A=*Allopaa hazarensis*, C=*Chrysopaa sternosignata*. *Nanorana* species names were abbreviated after five characters and are as follows: *phrynoides*, *sichuanensis*, *zhaoermii*, *arunachalensis*, *vicina* [prefix WLMNV], *xuelinensis*, *aenea*, cf. *blanfordii*, cf. *polunini*, *rarica*, cf. *rostandi*, *chayuensis*, *conaensis*, *liebigii*, *maculosa*, *medogensis*, *pleskei*, *quadranus*, sp. [2Bahn_RAS], cf. *vicina* [2Pul_RAS, 782_RAS], sp. [A1966/13_NME], *taihangnica*, *unculuanus*, *ventripunctata*, *yunnanensis*, cf. *ercepea*, *parkeri*, *Quasipaa boulengeri*, sp. [VV5.1/8.1/11.1_RAS], cf. *vicina* [VV9.1_RAS]. A star after the abbreviated species name refers to 'cf. '. Gray shaded cells indicate the large genetic distance between *N. arunachalensis* and other spiny frogs, including *Nanorana* species, showing the taxonomic misclassification of that species. Values in colored cells are distances < 2.5%.

	A	C	phry	sichu	zhao	arun	vici	xueli	aene	blan*	polu*	rari	ros*	chay	cona	liebi	mac	med	plesk	quad	sp.	vici*	sp.	taiha	uncu	ventr	yunn	erce*	park	Q	sp.	vici*
A		1.2	1.1	1.2	1.0	1.5	1.2	1.2	1.1	1.0	1.1	1.0	1.1	1.1	1.1	1.1	1.0	1.1	1.0	1.2	1.1	1.1	1.1	1.0	1.2	1.0	1.1	1.0	1.1	1.2	1.1	1.0
C	8.3		1.1	1.1	1.0	1.5	1.2	1.2	1.1	1.0	1.0	0.9	1.1	1.1	1.0	1.0	1.0	1.1	1.1	1.2	1.2	1.1	1.0	1.0	1.2	1.1	1.0	1.0	1.1	1.0	1.1	1.0
phry	6.6	6.9		0.6	0.9	1.5	1.0	1.0	1.0	0.8	0.9	0.8	0.9	1.0	0.8	0.9	1.0	1.0	0.8	1.0	0.9	1.0	1.0	0.9	1.0	0.8	0.5	0.9	0.9	1.2	1.0	0.9
sichu	7.4	6.5	2.1		0.9	1.5	1.1	1.0	0.9	0.8	0.9	0.8	0.9	1.0	0.8	0.8	1.0	1.0	0.9	0.9	1.0	1.0	0.9	0.8	1.0	0.8	0.7	0.9	0.8	1.2	1.0	0.9
zhao	5.8	5.6	4.3	4.5		1.5	0.8	0.9	0.9	0.6	0.8	0.6	0.7	0.7	0.7	0.8	0.6	0.7	0.7	0.8	0.8	0.7	0.7	0.8	1.0	0.7	0.8	0.7	0.7	1.1	0.9	0.6
arun	15.8	15.2	15.4	15.4	15.0		1.6	1.5	1.5	1.4	1.5	1.5	1.5	1.6	1.5	1.5	1.6	1.5	1.5	1.6	1.5	1.5	1.5	1.5	1.6	1.5	1.5	1.5	1.5	1.5	1.6	1.5
vici	7.9	7.6	5.8	6.0	3.8	16.3		1.1	1.1	0.8	0.9	0.9	0.8	0.8	0.9	0.9	0.8	0.9	0.9	1.1	0.4	0.5	0.9	0.9	1.1	0.8	0.9	0.8	0.9	1.3	1.0	0.9
xueli	8.3	8.2	5.9	5.3	5.0	14.9	6.8		1.0	0.9	0.9	0.9	0.9	1.0	0.9	1.0	0.9	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.1	0.9	1.0	0.9	1.0	1.1	1.0	0.9
aene	7.3	7.6	5.2	4.6	5.1	15.7	6.9	6.4		0.9	1.0	0.9	0.9	0.9	0.9	0.9	0.9	1.0	0.9	1.1	1.0	1.0	0.9	0.9	1.0	0.9	0.9	0.9	0.9	1.2	0.9	0.9
blanf*	6.8	6.7	4.1	3.9	2.4	14.7	4.9	5.6	5.3		0.8	0.6	0.6	0.7	0.7	0.8	0.7	0.7	0.7	0.8	0.8	0.7	0.7	0.8	0.9	0.6	0.7	0.7	0.7	1.1	0.9	0.5
polun*	6.4	6.1	5.2	4.7	3.6	14.7	5.1	5.4	5.7	4.4		0.8	0.7	0.9	0.8	0.8	0.8	0.8	0.9	1.0	0.9	0.8	0.7	0.8	1.1	0.8	0.9	0.7	0.8	1.1	0.9	0.7
rari	6.0	5.4	4.1	4.1	2.3	14.7	4.5	5.0	5.0	3.0	3.7		0.7	0.7	0.7	0.8	0.6	0.7	0.7	0.9	0.8	0.8	0.7	0.8	1.0	0.7	0.8	0.7	0.7	1.0	0.9	0.6
ros*	5.9	6.5	4.1	4.3	2.7	14.8	3.8	4.8	5.1	2.7	3.2	2.9		0.8	0.8	0.7	0.6	0.7	0.7	0.9	0.8	0.8	0.7	0.8	1.0	0.7	0.8	0.3	0.7	1.1	0.8	0.5
chay	6.5	6.8	4.8	5.0	2.3	15.6	4.0	5.9	5.4	3.4	4.4	3.0	3.2		0.8	0.8	0.4	0.7	0.9	1.0	0.9	0.9	0.8	0.9	1.0	0.8	0.9	0.7	0.8	1.2	1.0	0.8
cona	6.8	5.7	4.0	3.4	3.1	14.1	4.5	5.3	5.5	3.9	3.6	2.9	3.8	3.6		0.9	0.9	0.9	0.8	1.0	0.8	0.8	0.8	0.8	1.0	0.8	0.8	0.8	0.8	1.0	0.9	0.8
liebi	6.8	7.6	5.0	4.5	4.4	15.3	5.6	6.5	5.6	4.8	4.3	4.5	3.7	4.3	5.4		0.8	0.8	0.8	1.0	0.9	0.9	0.8	0.8	1.0	0.8	0.8	0.7	0.8	1.1	0.9	0.8
mac	5.5	6.4	5.5	5.3	2.2	15.7	3.8	5.3	4.9	3.5	3.8	2.9	2.4	1.1	4.2	4.1		0.6	0.8	1.0	0.9	0.8	0.8	0.8	1.0	0.8	0.9	0.6	0.7	1.1	0.9	0.6
med	6.5	7.1	5.5	5.3	2.6	15.3	4.3	5.5	5.9	3.5	4.0	3.3	3.1	2.7	4.5	4.6	2.2		0.8	0.9	0.9	0.8	0.8	0.9	1.0	0.8	0.9	0.8	0.8	1.1	1.0	0.7
plesk	5.9	7.4	4.1	4.3	2.9	15.9	4.1	5.7	4.9	3.4	4.6	3.6	3.0	4.4	4.4	4.4	3.8	4.0		1.0	0.8	0.8	0.8	0.8	1.1	0.6	0.9	0.7	0.7	1.1	0.8	0.7
quad	8.4	7.9	4.7	4.5	3.6	17.3	7.2	5.9	7.1	4.2	6.3	4.7	4.6	5.2	5.2	6.9	5.5	5.2	5.7		1.1	1.0	1.0	1.0	1.1	0.9	1.0	0.9	0.9	1.2	1.1	0.8
sp.	7.1	7.2	5.1	5.6	3.7	14.8	0.8	6.0	6.3	4.7	4.5	4.1	3.6	4.0	4.0	5.2	4.2	4.8	3.9	6.5		0.5	0.9	0.9	1.1	0.8	0.9	0.8	0.9	1.2	1.0	0.9
vici*	7.6	7.2	5.3	5.1	3.4	15.1	1.6	5.9	6.3	4.3	4.7	4.2	3.5	4.2	4.3	5.2	3.7	4.0	3.8	6.5	1.5		0.9	0.8	1.1	0.8	0.9	0.8	0.8	1.1	0.9	0.8
sp.	7.2	5.9	5.0	4.8	2.9	15.9	4.5	6.3	5.3	3.7	3.6	3.6	3.2	3.6	4.0	4.6	3.4	4.4	4.1	5.9	4.7	4.6	0.0	0.8	1.1	0.8	0.9	0.7	0.8	1.1	0.9	0.8
taiha	6.1	6.4	4.3	3.5	3.9	15.2	4.7	5.2	5.1	4.3	4.3	3.8	3.2	4.6	4.2	3.9	4.0	4.9	3.4	6.3	4.5	4.1	3.9		1.1	0.8	0.8	0.8	0.8	1.1	0.9	0.8
uncu	8.1	9.1	5.3	5.5	6.4	16.2	7.8	7.3	5.8	5.9	7.5	6.7	6.6	6.5	6.6	6.9	6.6	6.9	6.7	6.9	7.1	7.5	6.9	6.6		1.0	1.0	1.1	1.0	1.3	1.1	1.0
ventr	5.6	7.3	3.9	3.3	2.7	14.8	4.0	5.4	5.3	3.0	4.1	3.7	3.2	3.4	3.6	4.5	3.4	3.6	2.0	4.6	3.9	3.8	4.3	4.3	6.0		0.8	0.8	0.5	1.1	0.9	0.6
yunn	6.7	6.3	1.6	2.3	3.9	15.2	4.9	5.6	4.9	3.9	4.8	3.9	3.8	4.8	4.2	4.9	4.5	4.9	4.3	5.5	4.9	4.4	4.6	3.9	5.6	3.9		0.8	0.8	1.1	0.9	0.8
erce*	5.6	6.1	4.3	4.5	2.7	15.2	3.8	5.6	4.6	3.2	3.2	2.9	0.7	2.8	3.4	3.7	2.0	3.5	3.0	5.3	3.6	3.6	2.8	3.2	6.9	3.6	3.9		0.8	1.1	0.8	0.5
park	6.0	6.9	4.3	3.7	2.9	14.7	4.5	5.4	5.1	3.3	3.9	3.0	3.0	3.6	3.3	4.4	3.1	3.6	3.0	4.8	4.7	4.3	3.6	3.6	6.4	1.8	4.1	3.2		1.0	0.8	0.7
Q	9.0	6.3	7.6	7.6	6.8	14.8	9.5	7.9	8.8	7.3	7.1	6.5	6.8	8.2	6.5	8.6	7.6	7.5	7.1	8.8	9.0	8.4	7.8	7.9	10.1	7.3	7.9	7.0	6.8		1.2	1.0
sp.	7.1	7.3	5.9	5.9	4.8	16.3	5.6	6.7	5.3	5.3	5.4	4.9	4.2	6.1	5.1	5.7	5.1	5.8	4.2	7.2	5.9	5.0	5.4	4.7	7.0	4.9	5.5	3.8	4.3	8.3		0.8
vici*	5.8	6.1	4.5	4.3	2.1	14.8	4.5	5.2	5.1	2.2	3.2	2.3	1.8	3.2	3.6	4.3	2.5	3.1	2.8	4.2	4.3	3.9	3.4	3.9	6.5	2.7	3.9	1.8	2.9	6.4	3.7	

Tables S5 a,b. Uncorrected genetic distances (%; lower left matrix), including standard error estimates (upper right matrix), between taxa used in this study for *COI* (**a**) and *Rag1* (**b**). A star after the species name refers to 'cf. '. Bold values in gray shaded cells highlight distances lower than 5% (*COI*) and lower than 0.1% (*Rag1*). A=*Allopaa hazarensis*, C=*Chrysopaa sternosignata*, N=*Nanorana*, Q=*Quasipaa boulengeri*.

(a)	A	C	*aenea*	*blanfordii**	*polunini**	*rarica*	*ercepeae**	*rostandi**	*liebigii*	*parkeri*	*pleskei*	sp. (2Bahn)	*vicina** (A1966)	*taihangnica*	*ventripunctata*	*yunnanensis*	Q	sp. (VV5, 8,11)	sp. (VV9.1)	*chayuensis*
A		1.68	1.58	1.48	1.57	1.50	1.57	1.54	1.41	1.64	1.56	1.67	1.57	1.66	1.61	1.69	1.63	1.63	1.52	1.51
C	20.00		1.67	1.69	1.60	1.63	1.74	1.67	1.55	1.68	1.67	1.70	1.71	1.66	1.72	1.78	1.70	1.76	1.67	1.62
N. aenea	14.59	19.16		1.42	1.50	1.45	1.59	1.54	1.39	1.61	1.61	1.67	1.48	1.49	1.58	1.57	1.71	1.60	1.55	1.50
*N. blanfordii**	13.26	19.71	13.17		1.31	1.19	1.36	1.37	1.21	1.46	1.44	1.48	1.33	1.60	1.42	1.55	1.66	1.58	1.36	1.26
*N. polunini**	15.19	17.49	14.29	11.32		1.19	1.36	1.37	1.17	1.41	1.39	1.45	1.30	1.54	1.42	1.55	1.70	1.58	1.43	1.34
N. rarica	15.75	19.50	14.94	10.20	10.20		1.15	1.04	1.07	1.45	1.39	1.34	1.05	1.55	1.36	1.46	1.69	1.49	1.30	1.19
*N. ercepeae**	15.08	19.29	15.21	11.69	11.69	8.63		0.87	1.17	1.51	1.57	1.58	1.23	1.59	1.58	1.55	1.52	1.49	1.37	1.40
*N. rostandi**	15.87	18.37	15.40	11.32	11.50	7.24	**4.27**		1.19	1.49	1.54	1.59	1.26	1.68	1.48	1.52	1.60	1.59	1.33	1.30
N. liebigii	15.06	18.69	14.33	11.41	10.99	10.39	10.11	10.71		1.40	1.37	1.37	1.17	1.45	1.35	1.45	1.49	1.38	1.23	1.26
N. parkeri	16.83	20.08	16.51	12.80	11.69	14.10	14.10	13.73	14.10		1.42	1.69	1.52	1.55	1.32	1.69	1.71	1.59	1.57	1.50
N. pleskei	16.58	21.01	16.88	13.54	12.06	13.36	15.03	15.03	14.42	12.99		1.55	1.45	1.62	1.21	1.59	1.72	1.54	1.53	1.42
N. sp. (2Bahn)	16.37	17.24	15.51	11.95	11.53	11.53	14.05	13.84	13.26	14.68	13.42		1.50	1.69	1.60	1.68	1.89	1.67	1.42	1.36
N. sp. (A1966)	16.25	19.71	13.91	11.87	10.58	8.26	8.72	9.09	11.04	14.10	13.17	12.79		1.64	1.50	1.56	1.69	1.54	1.35	1.33
N. taihangnica	16.76	19.71	12.43	15.21	15.96	16.33	15.77	17.81	15.86	15.21	17.63	15.72	16.51		1.62	1.69	1.67	1.55	1.61	1.62
N. ventripunctata	17.63	20.64	15.96	12.99	13.36	12.89	15.58	14.10	14.24	11.69	8.72	15.51	15.03	16.70		1.59	1.71	1.62	1.57	1.41
N. yunnanensis	17.74	20.55	15.77	15.03	15.40	14.19	14.66	14.29	15.21	18.37	15.96	16.14	15.58	18.00	15.96			1.66	1.52	1.58
Q	18.07	18.51	19.11	17.63	19.85	20.04	15.58	17.44	17.39	19.85	20.41	20.75	18.55	18.37	20.41	19.11		1.56	1.61	1.63
N. sp. (VV5,8,11)	17.56	20.52	15.89	15.65	16.20	15.83	14.22	16.64	14.59	16.33	15.52	16.21	15.40	15.89	16.76	17.50	16.45		1.54	1.57
*N. vicina** (VV9.1)	15.09	18.97	14.47	11.69	12.80	11.22	10.95	10.39	11.73	15.40	14.66	10.90	10.76	15.77	15.40	13.91	16.70	14.66		1.36
N. chayuensis	14.61	19.06	13.54	9.46	11.13	9.83	11.50	10.39	11.83	13.54	12.24	10.27	10.95	15.77	12.62	15.03	18.00	15.28	11.13	

(b)	A	*aenea*	*blanfordii**	*polunini**	*rarica*	*ercepeae**	*rostandi**	*chayuensis*	*conaensis*	*liebigii*	*maculosa*	*medogensis*	*parkeri*	*pleskei*	*quadranus*	*vicina*	sp. A1966	*taihangnica*	*unculuanus*	*ventripunctata*	*yunnanensis*	sp. VV5, 8,11	*vicina** VV9.1	C	Q
A		0.00	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.01	0.00	0.01	0.01	0.00	0.00	0.00	0.00	0.01	0.01	0.00	0.00
N. aenea	2.07		0.00	0.00	0.01	0.01	0.01	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00
*N. blanfordii**	3.58	3.02		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
*N. polunini**	3.39	2.83	0.33		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
N. rarica	3.89	3.63	0.86	0.73		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.01	0.01	0.00	0.00	0.01	0.01	0.01	0.01	0.01	0.00	0.01	0.01
*N. ercepeae**	3.80	3.87	0.82	0.60	0.60		0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.01	0.01	0.00	0.00	0.01	0.01	0.01	0.01	0.01	0.00	0.01	0.01
*N. rostandi**	3.80	3.87	0.82	0.60	0.60	**0.00**		0.00	0.00	0.00	0.00	0.00	0.01	0.01	0.01	0.00	0.00	0.01	0.01	0.01	0.01	0.01	0.00	0.01	0.01
N. chayuensis	3.52	2.98	0.54	0.50	0.89	0.74	0.75		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
N. conaensis	3.58	3.07	0.70	0.75	1.16	1.04	1.04	0.58		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.01	0.00	0.00	0.00
N. liebigii	3.63	3.22	0.52	0.49	1.03	1.04	1.04	0.54	0.80		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.01	0.00	0.00	0.00
N. maculosa	3.43	2.90	0.54	0.50	0.89	0.74	0.75	0.17	0.58	0.54		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
N. medogensis	3.43	2.90	0.46	0.42	0.85	0.74	0.75	**0.08**	0.50	0.48	**0.08**		0.00	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
N. parkeri	3.27	2.65	2.57	2.58	3.12	3.27	3.28	2.57	2.57	2.83	2.49	2.49		0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.01	0.00	0.01	0.00
N. pleskei	3.08	2.49	2.40	2.41	3.11	3.42	3.43	2.40	2.40	2.72	2.32	2.32	0.83		0.00	0.00	0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
N. quadranus	2.12	1.49	2.53	2.33	3.15	3.42	3.43	2.49	2.57	2.72	2.40	2.40	2.49	2.32		0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
N. vicina	3.42	2.86	0.41	0.37	0.86	0.82	0.82	0.37	0.46	0.59	0.37	0.29	2.44	2.28	2.36		0.00	0.00	0.01	0.00	0.00	0.00	0.00	0.00	0.00
N. sp. (A1966)	3.58	2.98	0.54	0.33	0.81	0.74	0.75	0.66	0.75	0.59	0.66	0.58	2.57	2.40	2.49	0.37		0.00	0.01	0.00	0.00	0.01	0.00	0.00	0.00
N. taihangnica	2.11	1.49	2.69	2.50	3.31	3.57	3.58	2.82	2.90	2.89	2.73	2.74	2.65	2.49	0.99	2.69	2.65		0.00	0.00	0.00	0.00	0.00	0.00	0.00
N. unculuanus	2.62	0.83	3.36	3.16	4.02	4.32	4.32	3.32	3.40	3.57	3.23	3.24	3.15	2.99	1.82	3.19	3.32	1.66		0.00	0.00	0.01	0.01	0.00	0.00
N. ventripunctata	2.93	2.32	2.40	2.25	3.04	3.27	3.28	2.40	2.40	2.69	2.32	2.32	0.83	0.33	2.15	2.28	2.40	2.32	2.82		0.00	0.00	0.00	0.00	0.00
N. yunnanensis	2.39	1.16	2.82	2.66	3.47	3.72	3.73	2.82	2.82	3.06	2.82	2.74	2.73	2.57	1.66	2.69	2.82	1.66	1.49	2.40		0.00	0.00	0.00	0.00
N. sp. (VV5,8,11)	3.65	3.01	2.97	2.86	3.76	4.02	4.02	3.09	3.18	3.15	3.09	3.01	3.01	2.84	2.87	2.97	3.09	2.84	3.34	2.68	2.68		0.00	0.01	0.00
*N. vicina** (VV9.1)	3.37	2.82	0.37	0.33	0.88	0.89	0.89	0.33	0.58	0.57	0.33	0.25	2.40	2.24	2.32	0.21	0.50	2.65	3.15	2.24	2.65	2.93		0.00	0.00
Chrysopaa	3.07	2.54	2.75	2.55	3.30	3.80	3.81	2.71	2.79	2.89	2.71	2.63	3.21	3.04	2.38	2.58	2.54	2.38	2.88	2.88	2.38	3.24	2.54		0.00
Quasipaa	2.67	1.91	2.78	2.58	3.49	4.02	4.02	2.73	2.82	3.04	2.73	2.65	2.73	2.57	1.91	2.61	2.73	1.74	2.40	2.40	1.91	2.76	2.57	2.13	

Tables S6 a-c. Uncorrected pairwise genetic distances between sequences of *Allopaa hazarensis* for *16S* (**a**), *COI* (**b**), and *Rag1* (**c**). Distances ≥1% (*16S*) and ≥0.5% (*COI*), and ≥0.2% (*Rag1*) are highlighted bold. #CUHC = Comenius University Herpetological Collection; *=prefix "WLM:AH"; $=ZFMK(SH20) (Museum Koenig, Bonn).

(a)	10 168	10 278	10 416	10 417	10 418	10 420	10 422	10 423	10 424	10 698	93 86	93 87	93 89	95 49	95 50	95 51	95 52	95 53	95 54	95 55	95 56	95 59	95 73	95 74	09 107	103 351	103 354	103 387	305 162
10278#	0.56																												
10416#	0.00	0.56																											
10417#	0.00	0.56	0.00																										
10418#	0.00	0.56	0.00	0.00																									
10420#	0.56	0.38	0.56	0.56	0.56																								
10422#	0.00	0.56	0.00	0.00	0.00	0.56																							
10423#	0.00	0.56	0.00	0.00	0.00	0.56	0.00																						
10424#	0.00	0.56	0.00	0.00	0.00	0.56	0.00	0.00																					
10698#	0.56	0.00	0.56	0.56	0.56	0.38	0.56	0.56	0.56																				
9386#	0.56	0.38	0.56	0.56	0.56	0.00	0.56	0.56	0.56	0.38																			
9387#	0.56	0.38	0.56	0.56	0.56	0.00	0.56	0.56	0.56	0.38	0.00																		
9389#	0.56	0.38	0.56	0.56	0.56	0.00	0.56	0.56	0.56	0.38	0.00	0.00																	
9549#	0.56	0.00	0.56	0.56	0.56	0.37	0.56	0.56	0.56	0.00	0.37	0.37	0.37																
9550#	0.56	0.00	0.56	0.56	0.56	0.37	0.56	0.56	0.56	0.00	0.37	0.37	0.37	0.00															
9551#	0.56	0.00	0.56	0.56	0.56	0.37	0.56	0.56	0.56	0.00	0.37	0.37	0.37	0.00	0.00														
9552#	0.56	0.00	0.56	0.56	0.56	0.37	0.56	0.56	0.56	0.00	0.37	0.37	0.37	0.00	0.00	0.00													
9553#	0.56	0.00	0.56	0.56	0.56	0.37	0.56	0.56	0.56	0.00	0.37	0.37	0.37	0.00	0.00	0.00	0.00												
9554#	0.56	0.00	0.56	0.56	0.56	0.38	0.56	0.56	0.56	0.00	0.37	0.37	0.37	0.00	0.00	0.00	0.00	0.00											
9555#	0.58	0.00	0.58	0.58	0.58	0.38	0.58	0.57	0.57	0.00	0.38	0.38	0.38	0.00	0.00	0.00	0.00	0.00	0.00										
9556#	0.56	0.00	0.56	0.56	0.56	0.37	0.56	0.56	0.56	0.00	0.37	0.37	0.37	0.00	0.00	0.00	0.00	0.00	0.00	0.00									
9559#	0.94	0.75	0.94	0.94	0.94	0.75	0.94	0.93	0.93	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.76	0.75								
9573#	**1.13**	0.94	**1.13**	**1.13**	**1.13**	0.94	**1.13**	**1.12**	**1.12**	0.94	0.93	0.93	0.93	0.93	0.93	0.93	0.93	0.93	0.93	**0.96**	0.93	0.19							
9574#	**1.12**	0.94	**1.12**	**1.12**	**1.12**	0.94	**1.12**	**1.12**	**1.12**	0.94	0.93	0.93	0.93	0.93	0.93	0.93	0.93	0.93	0.93	**0.95**	0.93	0.19	0.00						
SH2009107$	0.00	0.56	0.00	0.00	0.00	0.56	0.00	0.00	0.00	0.56	0.56	0.56	0.56	0.56	0.56	0.56	0.56	0.56	0.56	0.57	0.56	0.93	**1.12**	**1.12**					
103351$	0.19	0.75	0.19	0.19	0.19	0.75	0.19	0.19	0.19	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.75	0.77	0.75	**1.13**	**1.32**	**1.31**	0.19				
103354$	0.37	0.94	0.37	0.37	0.37	0.94	0.37	0.37	0.37	0.94	0.94	0.94	0.94	0.94	0.94	0.94	0.94	0.94	0.94	**0.96**	0.94	**1.32**	**1.50**	**1.50**	0.37	0.37			
103387$	0.56	0.00	0.56	0.56	0.56	0.37	0.56	0.56	0.56	0.00	0.37	0.37	0.37	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.75	0.93	0.93	0.53	0.75	0.94		
305162*	0.57	0.00	0.57	0.57	0.57	0.38	0.57	0.57	0.57	0.00	0.38	0.38	0.38	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.76	**0.95**	**0.95**	0.54	0.76	0.76	0.00	
28516t1*	0.75	0.19	0.75	0.75	0.75	0.56	0.75	0.75	0.75	0.19	0.56	0.56	0.56	0.19	0.19	0.19	0.19	0.19	0.19	0.19	0.19	0.93	**1.12**	**1.12**	0.72	0.94	1.13	0.18	0.18

(b)	9386	9388	9389	9549	9551	9559	9573	SH20 09107	103 354	103 387	10 168	10 278	10 416	10 417	10 418	10 420	10 422	10 423	10 424	10 698
9388#	0.00																			
9389#	0.00	0.00																		
9549#	**0.56**	**0.56**	**0.56**																	
9551#	**0.56**	**0.56**	**0.56**	0.00																
9559#	0.19	0.19	0.19	0.37	0.37															
9573#	0.19	0.19	0.19	0.37	0.37	0.00														
SH2009107$	**0.61**	**0.61**	**0.61**	0.41	0.41	0.41	0.41													
103354$	**0.63**	**0.63**	**0.63**	0.42	0.42	0.42	0.42	0.00												
103387$	**0.61**	**0.61**	**0.61**	0.00	0.00	0.41	0.41	0.41	0.42											
10168#	**0.74**	**0.74**	**0.74**	**0.56**	**0.56**	**0.56**	**0.56**	0.20	0.21	**0.61**										
10278#	**0.56**	**0.56**	**0.56**	0.00	0.00	0.37	0.37	0.41	0.42	0.00	**0.56**									
10416#	**0.74**	**0.74**	**0.74**	**0.56**	**0.56**	**0.56**	**0.56**	0.20	0.21	**0.61**	0.00	**0.56**								
10417#	**0.56**	**0.56**	**0.56**	0.37	0.37	0.37	0.37	0.00	0.00	0.41	0.19	0.37	0.19							
10418#	**0.56**	**0.56**	**0.56**	0.37	0.37	0.37	0.37	0.00	0.00	0.41	0.19	0.37	0.19	0.00						
10420#	0.19	0.19	0.19	0.37	0.37	0.00	0.00	0.41	0.42	0.41	**0.56**	0.37	**0.56**	0.37	0.37					
10422#	**0.56**	**0.56**	**0.56**	0.37	0.37	0.37	0.37	0.00	0.00	0.41	0.00	0.37	0.00	0.00	0.00	0.37				
10423#	**0.56**	**0.56**	**0.56**	0.37	0.37	0.37	0.37	0.00	0.00	0.41	0.19	0.37	0.19	0.00	0.00	0.37	0.00			
10424#	**0.56**	**0.56**	**0.56**	0.37	0.37	0.37	0.37	0.00	0.00	0.41	0.19	0.37	0.19	0.00	0.00	0.37	0.00	0.00		
10698#	**0.56**	**0.56**	**0.56**	0.00	0.00	0.37	0.37	0.41	0.42	0.00	**0.56**	0.00	**0.56**	0.37	0.37	0.37	0.37	0.37	0.37	
11352#	**0.56**	**0.56**	**0.56**	0.37	0.37	0.37	0.37	0.00	0.00	0.41	0.19	0.37	0.19	0.00	0.00	0.37	0.00	0.00	0.00	0.37

(c)	10168	10416	10418	10420	10422	9386	9387	9388	9389	9549	9551	9559	9573	SH2009107$	103351$	103354$	103387$	9552	9553	9554	9555	9556	9574	10417	10698
10416#	0.00																								
10418#	0.00	0.00																							
10420#	0.00	0.00	0.00																						
10422#	0.10	0.10	0.00	0.00																					
9386#	0.00	0.00	0.00	0.00	0.10																				
9387#	0.00	0.00	0.00	0.00	0.10	0.00																			
9388#	0.00	0.00	0.00	0.00	0.10	0.00	0.00																		
9389#	0.00	0.00	0.00	0.00	0.10	0.00	0.00	0.00																	
9549#	0.12	0.13	0.14	0.14	**0.26**	**0.23**	0.13	0.12	0.12																
9551#	0.00	0.00	0.00	0.00	0.10	0.09	0.00	0.00	0.00	0.11															
9559#	0.00	0.00	0.00	0.00	0.10	0.00	0.00	0.00	0.00	0.11	0.00														
9573#	0.00	0.00	0.00	0.00	0.10	0.09	0.00	0.00	0.00	0.00	0.00	0.00													
SH2009107$	0.00	0.00	0.00	0.00	0.10	0.09	0.00	0.00	0.00	0.11	0.00	0.00	0.00												
103351$	0.00	0.00	0.00	0.00	0.10	0.00	0.00	0.00	0.00	0.13	0.00	0.00	0.00	0.00											
103354$	0.00	0.00	0.00	0.00	0.10	0.00	0.00	0.00	0.00	0.12	0.00	0.00	0.00	0.00	0.00										
103387$	0.00	0.00	0.00	0.00	0.10	0.09	0.00	0.00	0.00	0.11	0.00	0.00	0.00	0.00	0.00	0.00									
9552#	0.00	0.00	0.00	0.00	0.13	0.11	0.00	0.00	0.00	0.11	0.00	0.00	0.00	0.00	0.00	0.00	0.00								
9553#	0.00	0.00	0.00	0.00	0.11	0.10	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00							
9554#	0.00	0.00	0.00	0.00	0.11	0.10	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00						
9555#	0.10	0.11	0.12	0.12	**0.22**	0.19	0.11	0.10	0.10	0.00	0.09	0.10	0.00	0.09	0.11	0.10	0.09	0.10	0.00	0.00					
9556#	0.10	0.11	0.12	0.12	**0.22**	0.19	0.11	0.10	0.10	0.00	0.09	0.10	0.00	0.09	0.10	0.10	0.09	0.10	0.00	0.00	0.00				
9574#	0.00	0.00	0.00	0.00	0.11	0.10	0.00	0.00	0.00	0.11	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.09	0.09			
10417#	0.00	0.00	0.00	0.00	0.11	0.10	0.00	0.00	0.00	0.11	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.09	0.09	0.00		
10698#	0.10	0.11	0.12	0.12	**0.23**	**0.20**	0.11	0.11	0.11	0.00	0.10	0.10	0.00	0.10	0.11	0.10	0.10	0.11	0.00	0.00	0.00	0.00	0.10	0.10	

Tables S7 a,b. Uncorrected genetic distances (%; lower left matrix) with standard error (upper right matrix) between *Allopaa hazarensis* (A), *Chrysopaa* (C), *Quasipaa* (Q), and subgenera of *Nanorana* (*Chaparana* [Ch], *Nanorana* [N], *Paa* [P]), including the unknown clade from the West Himalaya (WH) for *COI* (**a**) and *Rag1* (**b**).

(a)	A	C	Ch	P	N	Q	WH
A		1.68	1.37	1.28	1.41	1.32	1.63
C	20.00		1.46	1.43	1.53	1.31	1.76
Ch	16.36	19.81		1.10	1.23	1.28	1.33
P	15.25	18.83	15.16		1.07	1.18	1.27
N	17.01	20.58	16.57	13.94		1.24	1.40
Q	17.66	17.80	18.34	16.93	17.88		1.33
WH	17.56	20.52	16.43	15.31	16.20	16.92	

(b)	A	Ch	P	N	WH	C	Q
A		0.37	0.53	0.50	0.51	0.48	0.43
Ch	2.26		0.42	0.39	0.38	0.39	0.28
P	3.61	3.07		0.42	0.34	0.45	0.40
N	3.09	2.57	2.68		0.39	0.48	0.39
WH	3.57	2.88	2.27	2.67		0.44	0.36
C	3.07	2.51	2.92	3.04	3.01		0.36
Q	2.78	2.12	2.88	2.75	2.77	2.20	

Table S8. SDM performance and evaluation for *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (C). Information on the model performance and evaluation, and the variable contribution are given. Test AUC values indicate performance as follows: > 0.9 very good, > 0.8 good, and > 0.7 useful discrimination ability of the model. High values in variable contribution are highlighted bold.

	Model performance			Variable percent contribution					
species	Train. AUC	Test AUC	AUC SD	BIO2	BIO6	BIO8	BIO9	BIO12	altitude
A	0.978	0.978	0.01	2.2	9.0	**38.0**	0.0	28.2	22.6
C	0.862	0.802	0.05	1.4	15.6	**39.9**	33.8	6.9	2.4

Table S9. Performance evaluation scores for the final ensemble classifier combining biomod2 algorithms.

		KAPPA	TSS	ROC
Allopaa hazarensis	Test	0.639	0.992	0.998
	Cutoff	862	699	706.5
	Sensitivity	86.842	100	100
	Specificity	99.67	99.17	99.24
Chrysopaa sternosignata	Test	0.257	0.886	0.977
	Cutoff	842	547	552.5
	Sensitivity	53.488	97.674	97.674
	Specificity	98.87	90.88	90.98

Table S10. Statistics for single models specified in biomod2; **a)** for *Allopaa hazarensis*, and **b)** *Chrysopaa sternosignata*.

a)	*Alllopaa hazarensis*															
	Variable Importances						Teststatistics									
names(predictors)	alt	bio_12	bio_2	bio_6	bio_8	bio_9	KAPPA	TSS	ROC	FAR	SR	ACCURACY	BIAS	POD	CSI	ETS
GBM.RUN1.PA1	0.008	0.606	0.016	0.543	0.524	0.15	0.665	0.872	0.938	0.545	0.545	0.997	0.625	1	0.5	0.498
GLM.RUN1.PA1	0.245	0.873	0	0	0	0	0.112	0.911	0.957	0.065	0.065	0.996	0	1	0.063	0.059
GAM.RUN1.PA1	0.71	0.913	0.617	1	0.896	0.903	0.464	0.868	0.934	0.333	0.333	0.996	0	0.875	0.304	0.302
CTA.RUN1.PA1	0	0.831	0	0	0.727	0	0.082	0.804	0.902	0.047	0.047	0.996	0	0.875	0.046	0.043
ANN.RUN1.PA1	0.274	0.737	0.42	0.997	0.69	0.886	0.287	0.859	0.921	0.175	0.175	0.996	0	1	0.171	0.167
GBM.RUN2.PA1	0.026	0.604	0.075	0.678	0.325	0.135	0.536	0.993	0.996	0.389	0.389	0.996	0.125	1	0.368	0.366
GLM.RUN2.PA1	0.901	0.815	0	0	0.098	0.512	0.069	0.891	0.938	0.041	0.041	0.996	0	1	0.04	0.036
GAM.RUN2.PA1	0.691	0.661	0.617	0.859	0.88	0.891	0.477	0.867	0.933	0.353	0.353	0.996	0	0.875	0.316	0.313
CTA.RUN2.PA1	0	0.997	0	0	0	0	0.029	0.788	0.894	0.018	0.018	0.996	0	1	0.018	0.015
ANN.RUN2.PA1	0.338	0.61	0.107	0.859	0.103	0.332	0.174	0.964	0.982	0.099	0.099	0.996	0	1	0.099	0.095
GBM.RUN3.PA1	0.018	0.634	0.066	0.588	0.28	0.078	0.452	0.859	0.873	0.4	0.4	0.996	0.75	0.875	0.294	0.292
GLM.RUN3.PA1	0.48	0.813	0	0	0.024	0	0.058	0.882	0.929	0.036	0.036	0.996	0	1	0.034	0.03
GAM.RUN3.PA1	0.839	0.691	0.621	0.887	0.897	0.849	0.425	0.743	0.872	0.3	0.3	0.996	0	0.75	0.273	0.27
CTA.RUN3.PA1	0	0.614	0	0.796	0	0.62	0.209	0.729	0.865	0.125	0.125	0.996	0	0.75	0.12	0.117
ANN.RUN3.PA1	0.772	0.775	0.077	0.726	0.545	0.77	0.14	0.478	0.773	0.085	0.085	0.996	0	1	0.078	0.075
GBM.RUN4.PA1	0.033	0.625	0.012	0.678	0.509	0.295	0.613	0.995	0.998	0.444	0.444	0.996	0	1	0.444	0.442
GLM.RUN4.PA1	0.252	0.867	0	0	0	0	0.113	0.867	0.942	0.066	0.066	0.996	0	1	0.063	0.06
GAM.RUN4.PA1	0.728	0.633	0.487	0.905	0.947	0.579	0.761	0.998	0.998	0.615	0.615	0.998	0.5	1	0.615	0.614
CTA.RUN4.PA1	0	0.825	0	0.706	0	0	0.091	0.926	0.963	0.051	0.051	0.996	0	1	0.051	0.047
ANN.RUN4.PA1	0.526	0.722	0.231	0.997	0.574	0.806	0.351	0.986	0.993	0.216	0.216	0.996	0	1	0.216	0.213
GBM.RUN5.PA1	0.006	0.642	0.042	0.578	0.247	0.094	0.452	0.833	0.932	0.357	0.357	0.996	0.25	1	0.294	0.292
GLM.RUN5.PA1	0.226	0.885	0	0	0	0	0.1	0.825	0.941	0.058	0.058	0.996	0	1	0.056	0.053
GAM.RUN5.PA1	0.868	0.744	0.673	0.976	0.754	0.483	0.414	0.74	0.871	0.312	0.312	0.996	0	0.75	0.263	0.261
CTA.RUN5.PA1	0	0.834	0	0.698	0	0	0.072	0.794	0.897	0.042	0.042	0.996	0	0.875	0.041	0.038
ANN.RUN5.PA1	0.799	0.805	0.345	0.335	0.499	0.575	0.183	0.648	0.953	0.111	0.111	0.996	0	1	0.104	0.101
GBM.RUN6.PA1	0.022	0.603	0.041	0.634	0.314	0.211	0.516	0.98	0.995	0.368	0.368	0.996	0.375	1	0.35	0.348
GLM.RUN6.PA1	0.229	0.881	0	0	0	0	0.116	0.909	0.955	0.068	0.068	0.996	0	1	0.065	0.061
GAM.RUN6.PA1	0.698	0.633	0.625	0.974	0.862	0.768	0.421	0.866	0.933	0.312	0.312	0.996	0	0.875	0.269	0.266
CTA.RUN6.PA1	0	0.996	0	0	0	0	0.029	0.792	0.896	0.019	0.019	0.996	0	1	0.019	0.015
ANN.RUN6.PA1	0.753	0.66	0.604	0.867	0.475	0.445	0.27	0.857	0.948	0.163	0.163	0.996	0	1	0.159	0.156
GBM.RUN7.PA1	0.011	0.642	0.069	0.366	0.621	0.078	0.319	0.715	0.928	0.273	0.273	0.996	0.625	1	0.192	0.189
GLM.RUN7.PA1	0.443	0.824	0	0	0.008	0	0.063	0.859	0.927	0.037	0.037	0.996	0	1	0.036	0.032
GAM.RUN7.PA1	0.843	0.708	0.485	0.876	0.607	0.752	0.432	0.62	0.809	0.333	0.333	0.996	0.5	0.625	0.278	0.275
CTA.RUN7.PA1	0	0.747	0	0.577	0.828	0	0.208	0.608	0.804	0.128	0.128	0.996	0	0.625	0.119	0.116
ANN.RUN7.PA1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
GBM.RUN8.PA1	0.018	0.608	0.088	0.693	0.341	0.134	0.558	0.992	0.996	0.429	0.429	0.996	0.25	1	0.389	0.387
GLM.RUN8.PA1	0.469	0.814	0	0	0.017	0	0.083	0.9	0.944	0.049	0.049	0.996	0	1	0.047	0.043
GAM.RUN8.PA1	0.678	0.653	0.615	0.919	0.859	0.756	0.497	0.992	0.995	0.333	0.333	0.996	0	1	0.333	0.331
CTA.RUN8.PA1	0	0.828	0	0.707	0	0	0.094	0.929	0.965	0.053	0.053	0.996	0	1	0.053	0.05
ANN.RUN8.PA1	0.364	0.707	0.325	0.764	0.685	0.732	0.464	0.989	0.997	0.318	0.318	0.996	0	1	0.304	0.302
GBM.RUN9.PA1	0.01	0.618	0.044	0.754	0.336	0.158	0.581	0.99	0.998	0.438	0.438	0.996	0.25	1	0.412	0.41
GLM.RUN9.PA1	0.417	0.808	0	0	0	0	0.061	0.87	0.933	0.037	0.037	0.996	0	1	0.035	0.031
GAM.RUN9.PA1	0.702	0.634	0.25	0.847	0.918	0.773	0.421	0.866	0.934	0.28	0.28	0.996	0	0.875	0.269	0.266
CTA.RUN9.PA1	0	0.862	0	0.702	0	0	0.095	0.814	0.913	0.054	0.054	0.996	0	1	0.054	0.05

ANN.RUN9.PA1	0.449	0.803	0.239	0.669	0.509	0.754	0.241	0.976	0.985	0.14	0.14	0.996	0	1	0.14	0.137
GBM.RUN10.PA1	0.016	0.624	0.024	0.76	0.445	0.167	0.635	0.994	0.997	0.5	0.5	0.996	0.5	1	0.467	0.465
GLM.RUN10.PA1	0.421	0.806	0	0	0	0	0.063	0.865	0.933	0.038	0.038	0.996	0	1	0.036	0.033
GAM.RUN10.PA1	0.649	0.795	0.529	0.888	0.911	0.5	0.694	0.997	0.998	0.538	0.538	0.997	0.375	1	0.533	0.531
CTA.RUN10.PA1	0	0.864	0	0.698	0	0	0.093	0.812	0.913	0.053	0.053	0.996	0	1	0.053	0.049
ANN.RUN10.PA1	0.562	0.708	0.297	0.97	0.288	0.457	0.337	0.974	0.992	0.212	0.212	0.996	0	1	0.206	0.203

b) *Chrysopaa sternosignata*

	Variable Importances						Teststatistics									
names(predictors)	alt	bio_12	bio_2	bio_6	bio_8	bio_9	KAPPA	TSS	ROC	FAR	SR	ACCURACY	BIAS	POD	CSI	ETS
GBM.RUN1.PA1	0.684	0.263	0.163	0.194	0.124	0.056	0.186	0.642	0.813	0.167	0.167	0.996	0.667	1	0.105	0.103
GLM.RUN1.PA1	0	0	0.249	0	0.653	0.549	0.043	0.636	0.846	0.027	0.027	0.996	1	1	0.026	0.022
GAM.RUN1.PA1	0.848	0.53	0.433	0.63	0.546	0.354	0.086	0.473	0.762	0.052	0.052	0.996	0	0.667	0.049	0.045
CTA.RUN1.PA1	0.766	0.335	0.157	0.176	0	0	0.022	0.559	0.779	0.016	0.016	0.996	0	0.778	0.016	0.011
ANN.RUN1.PA1	0.934	0.224	0.015	0.471	0.387	0.376	0.028	0.412	0.725	0.02	0.02	0.996	0	0.778	0.018	0.014
GBM.RUN2.PA1	0.676	0.204	0.168	0.136	0.092	0.118	0.205	0.826	0.919	0.143	0.143	0.996	0	1	0.118	0.114
GLM.RUN2.PA1	0	0	0	0	0	0.996	0.028	0.593	0.784	0.019	0.019	0.996	0.889	1	0.018	0.014
GAM.RUN2.PA1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
CTA.RUN2.PA1	0.833	0.424	0.302	0	0	0.186	0.032	0.351	0.672	0.021	0.021	0.996	0.111	0.444	0.02	0.016
ANN.RUN2.PA1	0.838	0.43	0.15	0.304	0.499	0.549	0.064	0.786	0.897	0.037	0.037	0.996	0	1	0.037	0.033
GBM.RUN3.PA1	0.589	0.273	0.137	0.146	0.16	0.105	0.131	0.785	0.878	0.082	0.082	0.996	0.556	1	0.074	0.07
GLM.RUN3.PA1	0.56	0	0	0	0.161	0.322	0.039	0.589	0.831	0.024	0.024	0.996	0	1	0.024	0.02
GAM.RUN3.PA1	0.623	0.577	0.253	0.603	0.587	0.417	0.137	0.637	0.834	0.091	0.091	0.996	0	0.778	0.077	0.073
CTA.RUN3.PA1	0.744	0.507	0	0.284	0	0	0.032	0.612	0.787	0.021	0.021	0.996	0	0.889	0.021	0.016
ANN.RUN3.PA1	0.703	0.073	0.029	0.044	0.177	0.105	0.015	0.491	0.789	0.012	0.012	0.996	0	1	0.012	0.008
GBM.RUN4.PA1	0.68	0.208	0.147	0.264	0.109	0.108	0.115	0.564	0.868	0.083	0.083	0.996	0.667	1	0.065	0.061
GLM.RUN4.PA1	0	0	0	0.341	0.737	0.581	0.036	0.613	0.791	0.024	0.024	0.996	0	1	0.023	0.018
GAM.RUN4.PA1	0.756	0.492	0.306	0.638	0.389	0.389	0.089	0.645	0.83	0.053	0.053	0.996	0	0.778	0.051	0.046
CTA.RUN4.PA1	0.952	0	0	0.308	0	0	0.019	0.454	0.743	0.014	0.014	0.996	0	1	0.014	0.009
ANN.RUN4.PA1	0.664	0.218	0.021	0.071	0.276	0.141	0.062	0.53	0.812	0.041	0.041	0.996	1	1	0.036	0.032
GBM.RUN5.PA1	0.642	0.288	0.156	0.241	0.1	0.081	0.119	0.726	0.903	0.087	0.087	0.996	0.667	1	0.067	0.063
GLM.RUN5.PA1	0.483	0	0	0	0.169	0.315	0.036	0.755	0.867	0.023	0.023	0.996	0	1	0.023	0.018
GAM.RUN5.PA1	0.715	0.494	0.272	0.665	0.446	0.356	0.194	0.811	0.914	0.118	0.118	0.996	0	0.889	0.111	0.107
CTA.RUN5.PA1	0.754	0.447	0.254	0.184	0	0	0.031	0.608	0.801	0.02	0.02	0.996	0.444	0.778	0.02	0.016
ANN.RUN5.PA1	0.421	0.315	0.382	0.53	0.856	0.661	0.064	0.44	0.703	0.043	0.043	0.996	0	0.889	0.037	0.033
GBM.RUN6.PA1	0.662	0.208	0.202	0.209	0.183	0.049	0.108	0.76	0.857	0.083	0.083	0.996	0.667	1	0.061	0.057
GLM.RUN6.PA1	0.996	0	0	0	0	0	0	0	0.48	0.003	0.003	0.996	0	0.111	0.003	0
GAM.RUN6.PA1	0.78	0.504	0.317	0.613	0.537	0.48	0.056	0.696	0.862	0.034	0.034	0.996	0	0.889	0.033	0.029
CTA.RUN6.PA1	0.753	0.382	0.261	0.184	0.058	0.108	0.016	0.216	0.606	0.013	0.013	0.996	0	0.333	0.012	0.008
ANN.RUN6.PA1	0.864	0.155	0.028	0.129	0.229	0.15	0.034	0.601	0.857	0.022	0.022	0.996	0	1	0.022	0.017
GBM.RUN7.PA1	0.583	0.162	0.286	0.216	0.106	0.248	0.082	0.431	0.751	0.056	0.056	0.996	0.667	1	0.047	0.043
GLM.RUN7.PA1	0.749	0.333	0.354	0.298	0.157	0.336	0.047	0.488	0.792	0.03	0.03	0.996	0.444	1	0.028	0.024
GAM.RUN7.PA1	0.628	0.386	0.379	0.591	0.658	0.602	0.054	0.344	0.674	0.042	0.042	0.996	0	0.444	0.032	0.028
CTA.RUN7.PA1	0.75	0	0.447	0.249	0	0.483	0.028	0.341	0.667	0.019	0.019	0.996	0	0.444	0.019	0.014
ANN.RUN7.PA1	0.39	0.666	0.126	0.248	0.496	0.617	0.055	0.485	0.792	0.034	0.034	0.996	0	1	0.032	0.028
GBM.RUN8.PA1	0.648	0.355	0.169	0.256	0.114	0.039	0.218	0.625	0.876	0.167	0.167	0.996	0.667	1	0.125	0.122
GLM.RUN8.PA1	0.996	0	0	0	0	0	0.048	0.6	0.867	0.029	0.029	0.996	0	1	0.029	0.024
GAM.RUN8.PA1	0.767	0.47	0.347	0.71	0.434	0.326	0.118	0.701	0.871	0.073	0.073	0.996	0	0.889	0.067	0.063
CTA.RUN8.PA1	0.761	0.387	0.186	0.21	0	0	0.026	0.499	0.73	0.018	0.018	0.996	0	0.889	0.017	0.013
ANN.RUN8.PA1	0.786	0.215	0.221	0.477	0.951	0.497	0.217	0.671	0.924	0.139	0.139	0.996	0	1	0.125	0.121
GBM.RUN9.PA1	0.646	0.231	0.148	0.294	0.121	0.13	0.195	0.818	0.939	0.143	0.143	0.996	0	1	0.111	0.108
GLM.RUN9.PA1	0.862	0	0	0	0	0.157	0.048	0.627	0.843	0.032	0.032	0.996	0.889	1	0.029	0.025
GAM.RUN9.PA1	0.74	0.48	0.339	0.613	0.493	0.408	0.084	0.793	0.92	0.05	0.05	0.996	0	1	0.048	0.044
CTA.RUN9.PA1	0.758	0.344	0.163	0.182	0	0	0.03	0.688	0.855	0.02	0.02	0.996	0	1	0.019	0.015
ANN.RUN9.PA1	0.697	0.212	0.263	0.357	0.633	0.535	0.1	0.611	0.89	0.064	0.064	0.996	0.889	1	0.057	0.053
GBM.RUN10.PA1	0.527	0.236	0.225	0.267	0.096	0.247	0.188	0.755	0.848	0.136	0.136	0.996	0.889	1	0.107	0.104

GLMRUN10.PA1	0	0	0.118	0	0.554	0.752	0.027	0.632	0.8	0.019	0.019	0.996	0.222	1	0.018	0.014
GAMRUN10.PA1	0.688	0.491	0.447	0.584	0.612	0.522	0.125	0.614	0.837	0.075	0.075	0.996	0	0.778	0.07	0.066
CTARUN10.PA1	0.674	0	0.25	0.174	0	0.448	0.028	0.681	0.84	0.019	0.019	0.996	0.111	0.889	0.019	0.014
ANNRUN10.PA1	0.268	0.06	0.26	0.628	0.173	0.658	0.019	0.46	0.784	0.014	0.014	0.996	0	1	0.014	0.01

Table S11. Observed niche overlap values and results of niche identity test. Empirical overlap values smaller than the null distribution support niche divergence. Asterisk denote significance at *P < 0.05.

	D	*I*	Spearman rank correlation coefficient
Empirical values	0.028*	0.094	-0.207
Permuted critical value	0.658	0.887	0.658

Figure S1. PCA of 19 WorldClim v. 2.1 variables, showing climate space: the more similar the colors the more similar values. Records of *Chrysopaa sternosignata* are indicated as green filled circles (for details see Table S2). The arrow points to Malir, Pakistan, that is noted as type locality (Mulleer [= Malir] near Kurrachee [= Karachi]) for the syntypes BMNH 1947.2.1.21, 1947.2.1.22 of *C. sternosignata*. Malir has apparently substantially different climate conditions compared to the known distribution range of the species. Thus, we consider that type locality as erroneously defined.

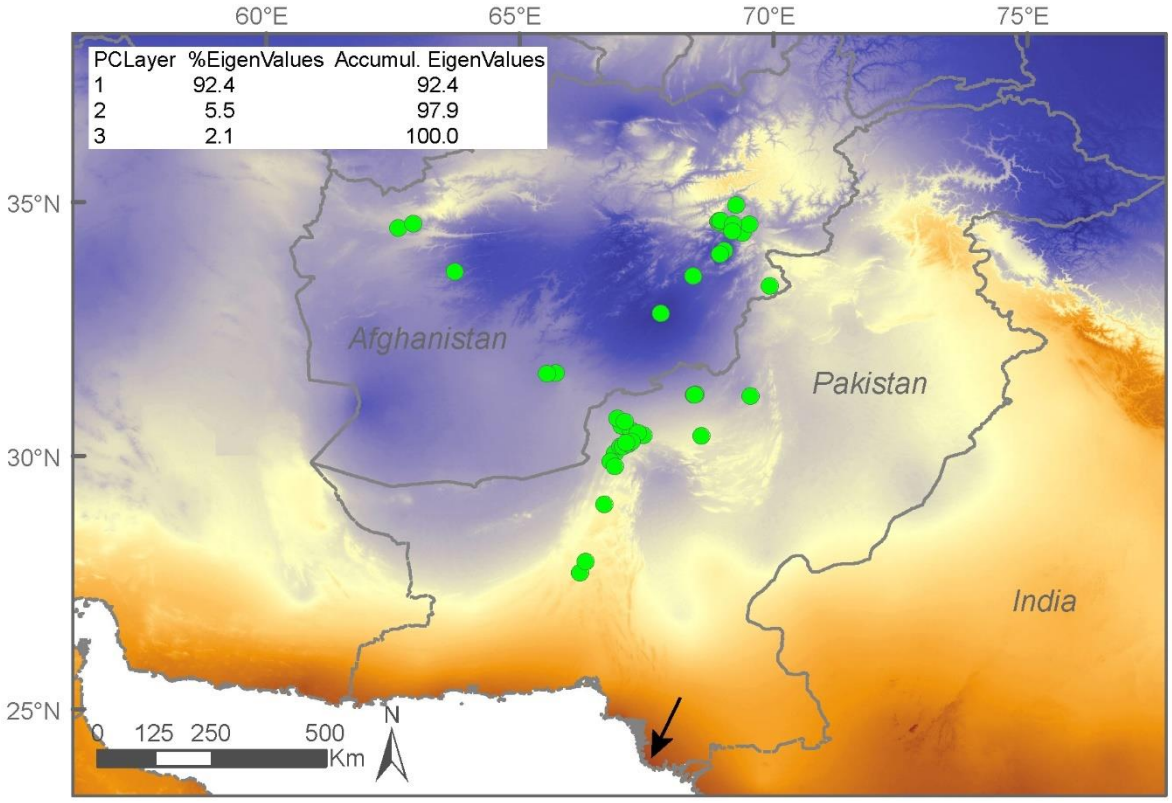

Figure S2. Bayesian inference tree based on 16S rRNA sequence data. Numbers at branch nodes refer to posterior probabilities ≥ 0.9. The clade of *Allopaa hazarensis* is indicated red, while *Chrysopaa sternosignata* is highlighted green. Species names are followed by voucher number (if available).

Figure S3. Ultametric time-calibrated phylogeny generated with BEAST2 based on the concatenated sequence data of spiny frogs. Grey bars specify the 95% HPD for the respective nodes; ages are shown for nodes that are supported by Bayesian posterior probability ≥ 0.95.

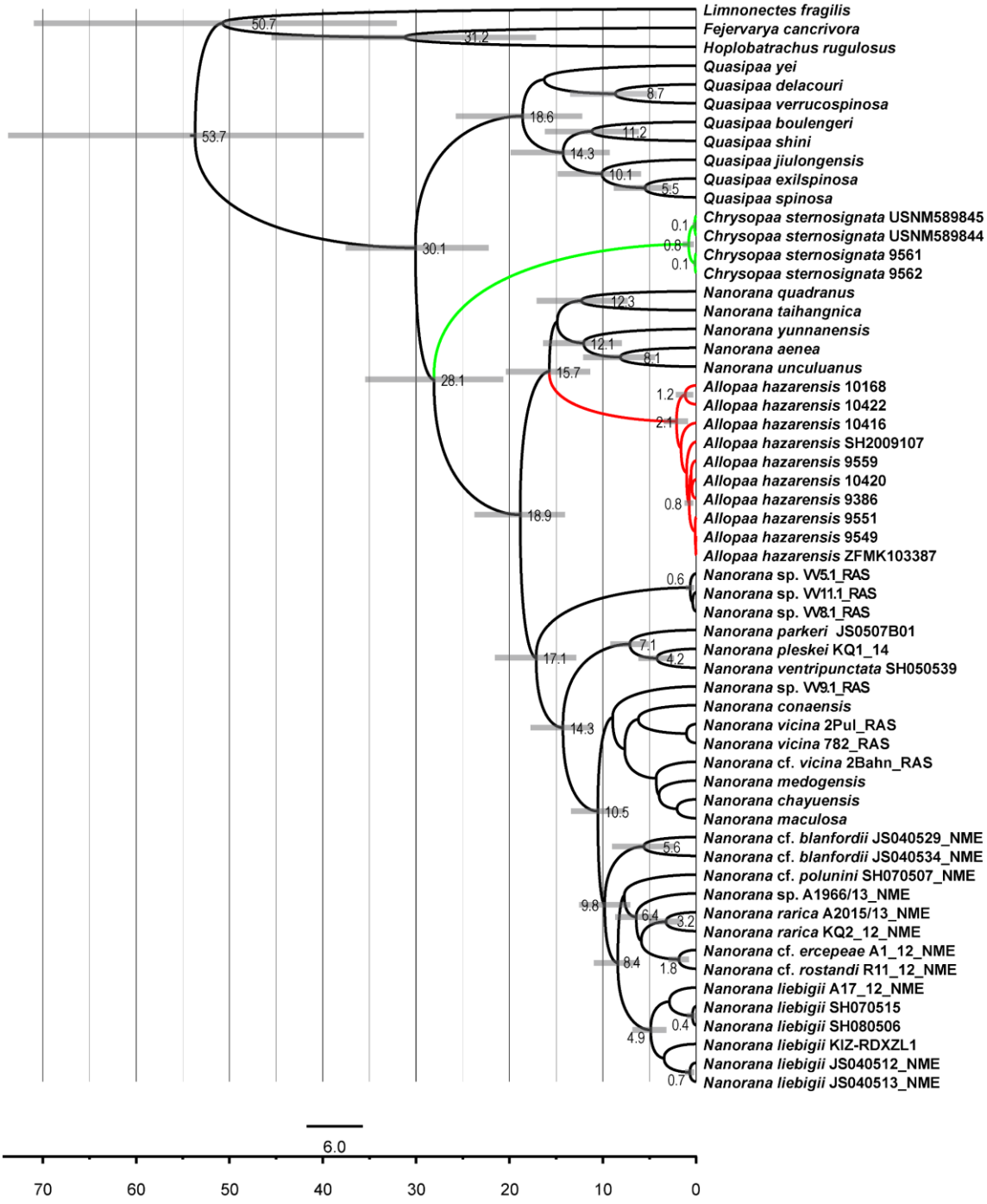

Figure S4. Climate heterogeneity raster based on recent WorldClim v.2.1 data; warm colors depict high areas of climatic heterogeneity.

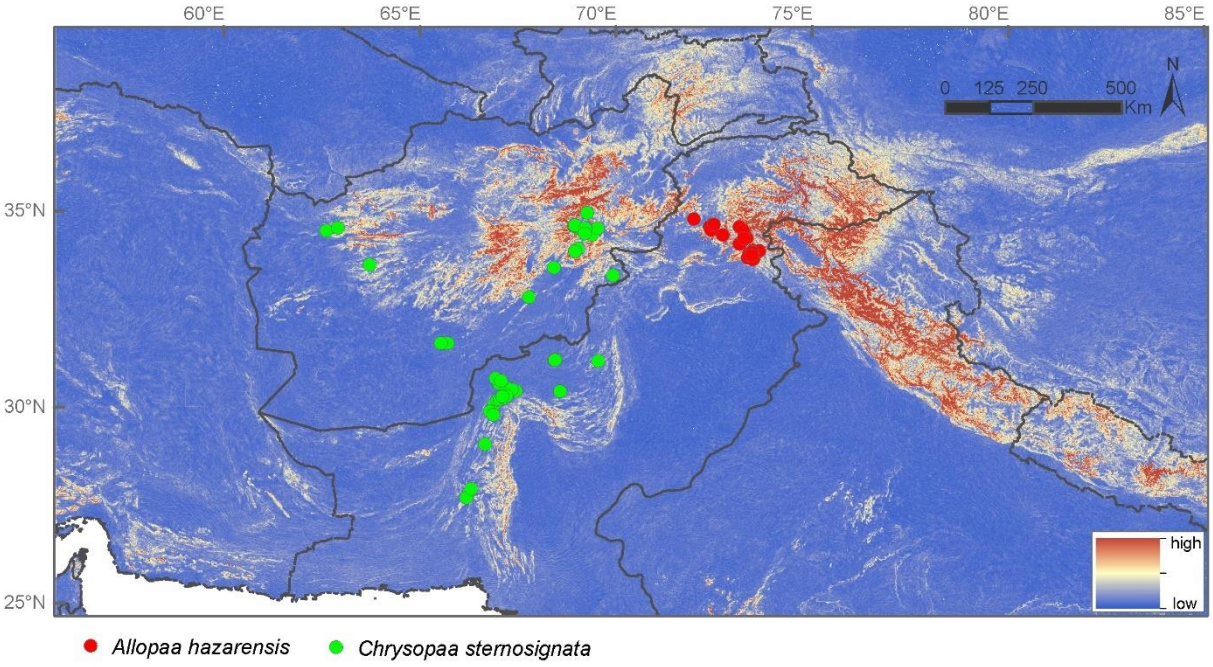

Figure S5. Distribution map for *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (B) derived from Species Distribution Model (SDM) using MaxEnt and a minimum training presence threshold. Maps include known records of the species (red = *A. hazarensis*, green = *C. sternosignata*).

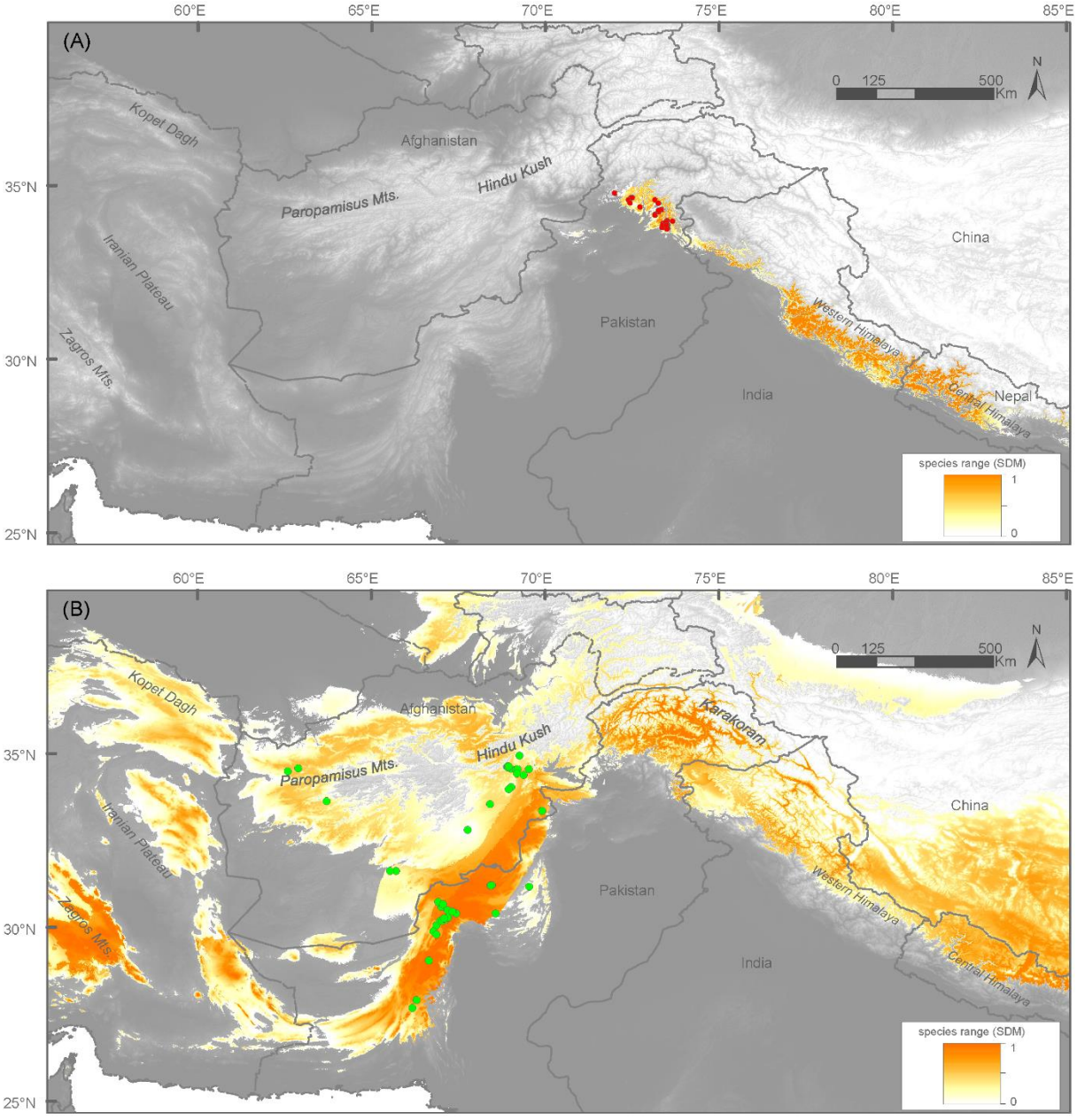

Figure S6. Distribution map for *Allopaa hazarensis* (A) and *Chrysopaa sternosignata* (B) derived from Species Distribution Model (SDM) using biomod2, including known records of the species (red = *A. hazarensis*, green = *C. sternosignata*).

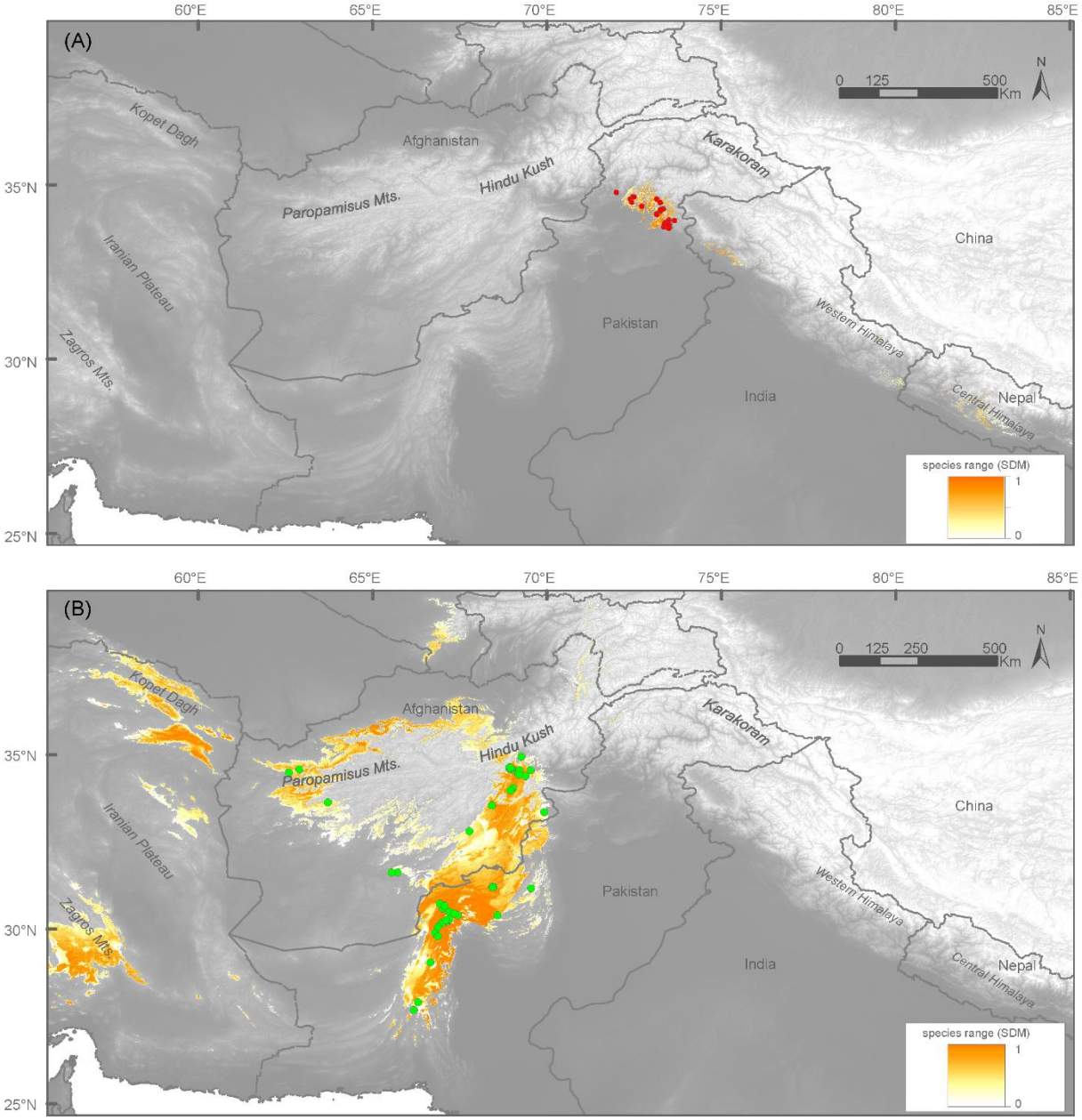

Figure S7. Environmental variable BIO8 (mean temperature of wettest quarter) across the modeled area based on WorldClim version 2.1 climate data for 1970-2000. Records of *Allopaa hazarensis* and *Chrysopaa sternosignata* are indicated by red and green circles, respectively.

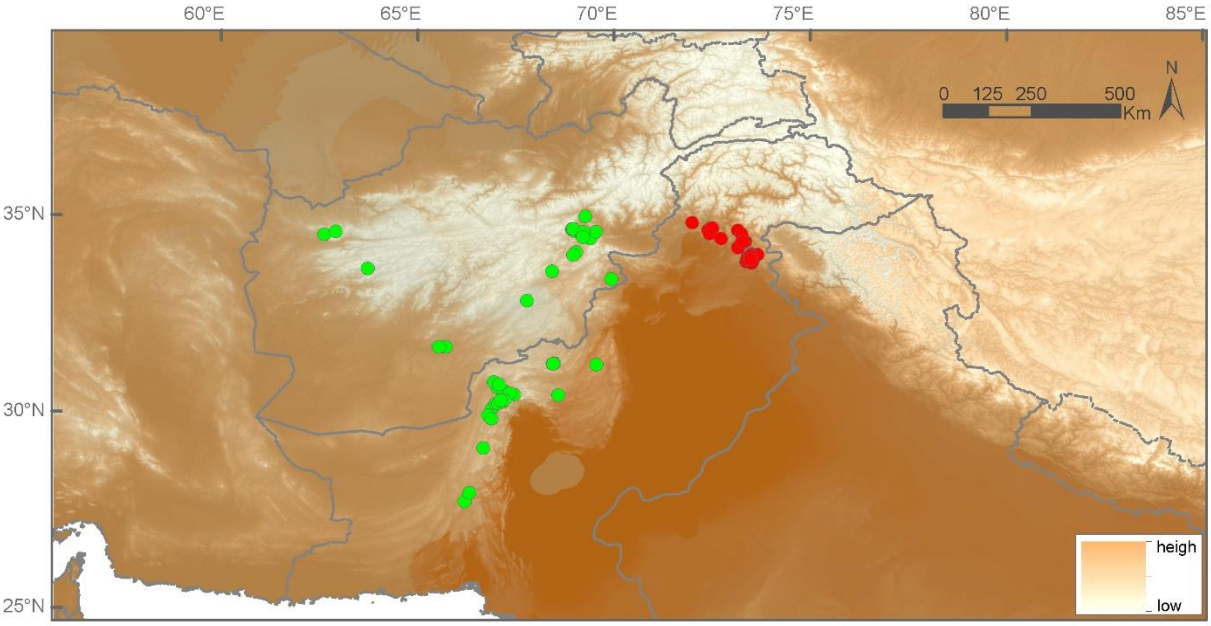

Fig. S8. Sample output from ENMTools v.1.4.4 for identity test for all three implemented niche overlap metrics. Histograms represent the distribution of overlaps for each metric from the null distribution, while the dashed vertical line represents the overlap between the models built using the empirical data.

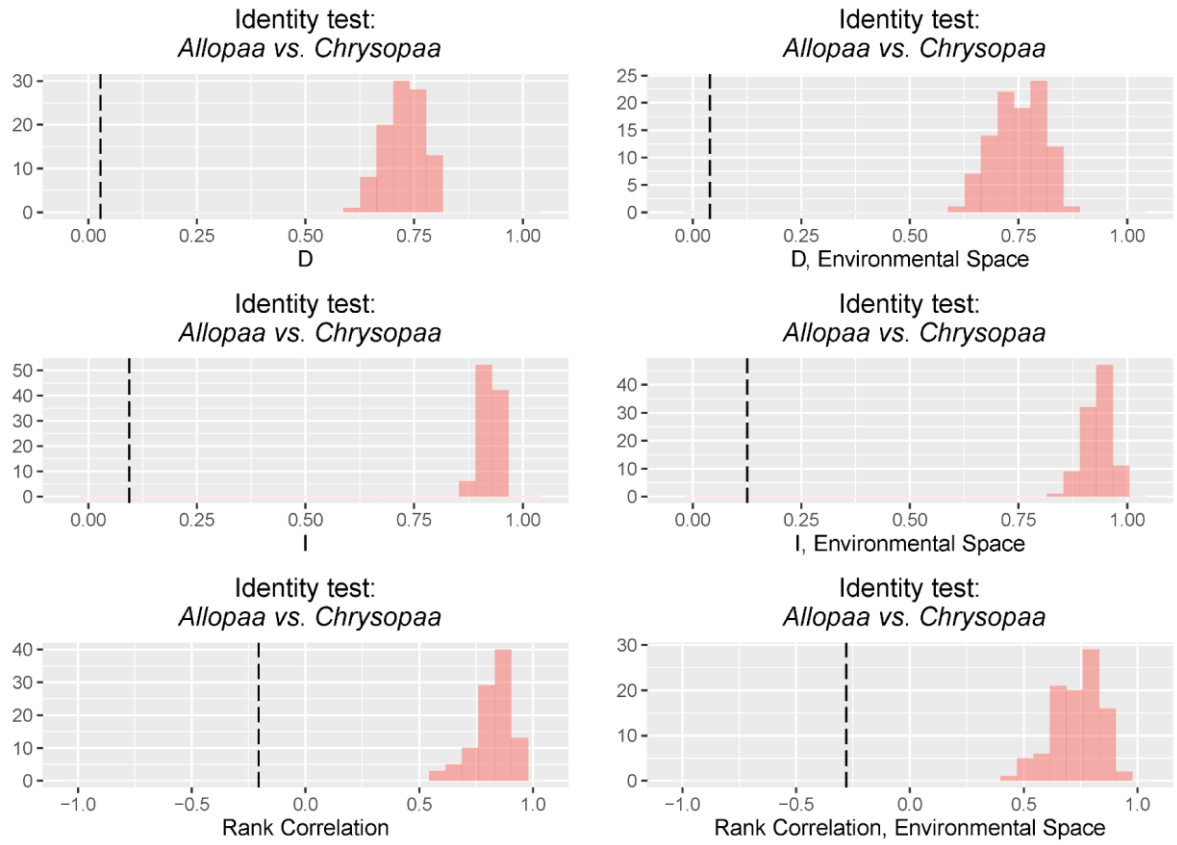
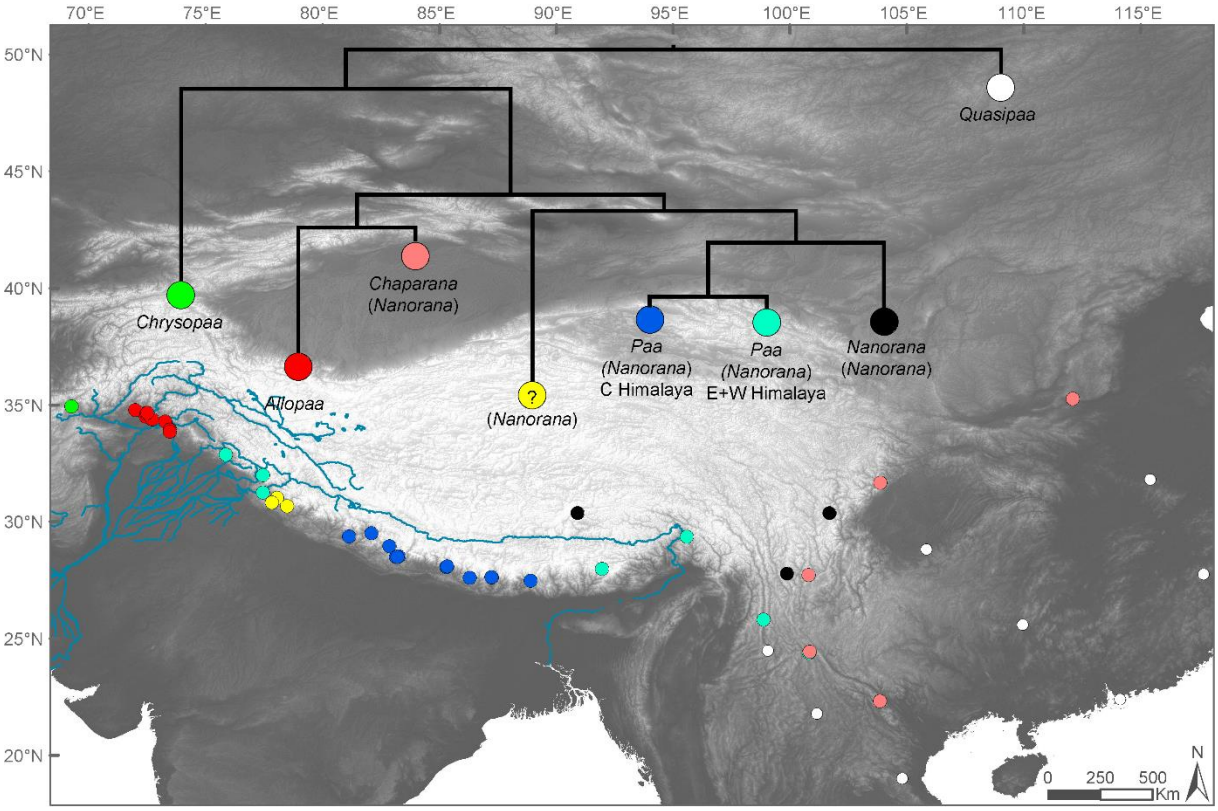
Figure S9. Simplified Bayesian inference tree with the main (sub)genera mapped to High Asia with the Indus and Brahmaputra River systems. Topology and color codes of the clades match Figure 1.