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First record of *Perinereis macropus* and *Perinereis cultrifera* (Annelida, Polychaeta) from rocky shores east Algeria, (SW Mediterranean sea)

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Abstract

Polychaetous from Algeria are little studied, despite their importance, they comprise diverse, and ecologically significant species, in the present study individuals of two Nereididea species, *Perinereis macropus* and *Perinereis cultrifera* were sampled on the rocky shores of Collo and Skikda Coasts, and were described for the first time with biometric measurements, morphological variations and reproduction aspects, in attempt to provide general information about this group from Algeria, necessary for other future works, especially those on marine biodiversity; we marked the absence of *P. macropus* at the second site, affected by pollution, as well as the difference between algae associated with each species; weight and size variables show differences intersites and among species; oogenesis is asynrochronous, mature oocytes occur during July and May for *P. macropus* and *P. cultrifera* respectively, and only adults of *P. macropus* show epitokal modifications, the semelparity character is observed in both sexes of the two species.

Keywords: Nereididae, Ecology, Biodiversity, Anatomy, Reproduction, Algeria

1. Introduction

At present, marine biodiversity studies are a matter of high interest to scientists and decision makers focusing on the current impacts generated by human activities on ecosystem processes and climate change ^[1], unlimited number of recent ones confirms that data of the actual situation of marine biodiversity must have a high priority, as a baseline for the indication of actual ecosystem's health, and the evaluation of future changes ^[2], in order to protect, and valorize high quality ecosystems, and if possible rehabilitate and restore degraded ones ^[3].

In this context, the marine benthic macrofauna is particularly suitable for monitoring environmental long term changes at the ecosystem level ^[4]; among this benthic groups, ^[5] confirm that any long term changes assessment of the benthos should be reflected in the polychaete community; as they play a key role in ecosystems they colonize, they are widely used in the environmental quality assessments, using indicators that vary from biomarkers measured at different levels of biological organization or at the whole organism ^[6-8], to benthic indices used and developed to evaluate the ecological quality into different degrees of disturbance, based on the community diversity, and the classification of taxa to ecological, or trophic groups ^[9, 10].

In Algeria, compared with other countries of the Western Mediterranean sea, such growing interest to environmental quality assessment, especially coastal zones confront scarce information on marine biodiversity, which contributes with 21% of the Mediterranean Sea global biodiversity, and does not reflect the real state [11].

Among the 85 families of polychaeta known to occur worldwide [12], Nereididae [13] is a significant component of communities occupying diverse habitats, from the intertidal to the deep sea, and represents a large family of approximately 44 genus and 677 species [14]; one of these diverse genera is *Perineris* [15] represented by around 74 species worldwide [14], but only 04 species *Perinereis cultrifera*, *Perinereis macropus*, *Perinereis marionii*, and *Perinereis oliveirae* have been recorded from Algerian waters [16, 17], view of recent literature from Algerian coasts conducted especially on this genus, shows an interest about the value of a single species *Perinereis cultrifera* as bioindicator of anthropogenic impact, and a few studies focusing on its biology, but highlights like said above, the lack of documented information in terms of inventory, ecology, description and illustration of this group from Algeria, necessary

to provide a valuable interpretation of numerous study fields that can be conducted in the future on this genus.

Perinereis cultrifera ^[18] is known to have a wide geographical distribution, and has a promoting role in economic sector as fishing baits ^[19], as well as feed in aquaculture ^[20], and belong to a species complex ^[21]. While for *Perinereis macropus* ^[22] information about its geographic distribution, biology and ecology still poorly documented, in the Mediterranean Sea, this species was also described by ^[23] (Naples, Monaco).

The current study presents a description in terms of biometric measurements and illustration of the two polychaetes species *P. macropus* and *P. cultrifera* from Algeria, and attempts to increase our current knowledge about their distribution,

habitat, identification and biology in general.

2. Materials and Methods

2.1 Sampling

Examined individuals were sampled at three occasions July 2013, May and June 2014, at low tide in the intertidal zone of a rocky shore from two stations north east Algeria, the first is located on the eastern part of Collo coast, whereas the second on the western part of Skikda coast (Figure 1). Worms were forced out from hard substrates by using bleaching liquid (10% in sea water) and collected gently by a pincer to avoid damaging them.



Fig 1: Location of the sampling sites.

2.2 Lab procedures

Worms were fixed in 4% formaldehyde, sorted under a stereomicroscope then preserved in 70% ethanol; all lab procedures were performed on collected individuals of the two species *P. macropus* and *P. cultrifera*.

Identification and description were basically focused on paragnaths' count and shape from all areas of everted pharynx, so worms were forced to evert their pharynx by generating a little pressure at the third chaetegerous toward the head; in addition of microscopical observations of jaws, parapodia and chaeta after performing sections.

Wet weight (g), and four parts of the animal's body were measured (mm), namely total length of the worm, length of the three first segments prostomium, peristomium and first chaetigerous (L₃), length of the head and ten first chaetigerous (H+10), and width at chaetigerous 10 (excluding parapodia), by using an ocular micrometer mounted on a stereo microscope; beside total length, the number of chaetegerous was also counted only in complete worms.

In order to determine the sex and describe the reproductive products of collected worms, each individual was opened approximately at the $10^{\rm th}$ – $20^{\rm th}$ segment, the coelomic fluid contents were released and deposited on a glass slide, then examined under an optic microscope, for females the mean oocytes size was estimated using the average value of the diameter from thirty oocytes, using an eyepiece graticule.

3. Results and Discussion

3.1 Material examined

Perinereis macropus (Claparède, 1870) Fig. 2 (A-E) Nereis (Lipephile) macropus Claparède 1870, p 80, Pl VIII, Fig. 1 (A-F).

Specimens of *P. macropus* were sampled among Rhodophyta corallinaceae Lithophyllum byssoides and vermetid reef formed by an association between Rhodophyta corallinaceae and Mollusca vermetidae Dendropoma petraeum; station 01: In July, 2013 individuals occur with posteriorly epitokal modification, (modification starts at 16-18 chaetigerous), two Heteronereis males largest complete 0.0872 g, 25 mm long, L₃ 3.4 mm, H+10 7 mm long and 2 mm wide, with 90 chaetigerous; seven heteronereis females largest complete 0.0941 g, 28 mm long, L₃ 3 mm, H+10 7 mm long and 2 mm wide, with 96 chaetigerous. In May 2014, three males, largest complete 0,2688 g, 58 mm long, L₃ 4 mm, H+10 9 mm long and 2.4 mm wide, with 100 chaetigerous; three females largest incomplete 0.2423 g, L₃ 3.6 mm, H+10 9 mm long and 3.4 mm wide. In June 2014, eleven males, largest complete $0.4240~g,\ 72~mm$ long, $L_3\ 5~mm,\ H+10\ 10~mm$ long and 2.2mm wide, with 97chaetigerous; one specimen regenerated posteriorly 0.1556 g, 49 mm long, L₃ 3.4 mm, H+10 8 mm long and 2 mm wide, with 70 chaetigerous (last 16 chaetigerous regenerated); four females, largest complete 0.4380 g, 90 mm long, L₃ 4.2 mm, H+10 10 mm long and 3 mm wide, with 95 chaetigerous. Station 02: we have noticed the absence of *P. macropus* during the period of sampling that might be a consequence of the very restricted area of vermetid reef, and the total absence of Lithophyllum byssoides, in addition of pollution known to affect this site, that may have a consequence on habitat and related species biodiversity.

Perinereis cultrifera (Grube, 1840) Fig. 2 (F-I)

Nereis cultrifera Grube 1840, p 74, Fig. 6

Specimens of *P. cultrifera* were collected among rhodophyta corallinaceae *jania rubens* and different species of the genus *Coralina* present at the site sampling; station 1: *In May 2014*,

three males, largest complete 0.4171 g, 52 mm long, L_3 5.2 mm, H+10 10 mm long and 3.2 mm wide, with 55 chaetigerous; one female largest incomplete 0.4929 g, L_3 6 mm, H+10 14 mm long and 3 mm wide. In June 2014, four males largest complete, regenerated posteriorly 0.5152 g, 70 mm long, L_3 6.8 mm, H+10 12 mm long and 4 mm wide, with 56 chaetigerous (last 10 chaetigerous regenerated); another specimen regenerated posteriorly 0.2123 g, 44 mm long, L_3 4.6 mm, H+10 12 mm long and 3 mm wide, with 47 chaetigerous (last 4 chaetigerous regenerated); four females, largest incomplete 0.4151 g, L_3 7.1 mm, H+10 12 mm long and 4 mm wide.

Station 2: In May 2014, three males, largest complete 0.3258 g, 60 mm long, L_3 5 mm, H+10 11 mm long and 2.4 mm wide,, with 68 chaetigerous; four females largest complete 0.2192 g, 46 mm long, L_3 5.4 mm, H+10 8 mm long and 3.2 mm wide, with 56 chaetigerous.

In June 2014, two males, largest complete 0.5190g, 60 mm long, L₃ 5.8 mm, H+10 11.2mm long and 3mm wide, with 64

chaetigerous; four females, largest incomplete L_3 6 mm, H+1012 mm long and 3 mm wide. *In July 2013*, individuals were absent at the two stations both, as a result of the semelparous characters of these species, adults died in days after reproduction.

3.2 Description and identification traits

The two species are recognized by their elongated and multi segmented form, and by the presence simultaneously of two antennas, two biarticulated palps, four pairs of tentacular cirri, four eyes, and a pair of dark or light brown of serrated jaws, with 4-6 teeth on an eversible pharynx, these characters agree with the description of Nereididae by ^[24]; thus the general form is the same with segments along the body all alike, except at the anterior end where there is the head formed by prostomium and peristomium, and at the posterior extremity the pygidium, with two anal cirri that vary in length (Figure 2).

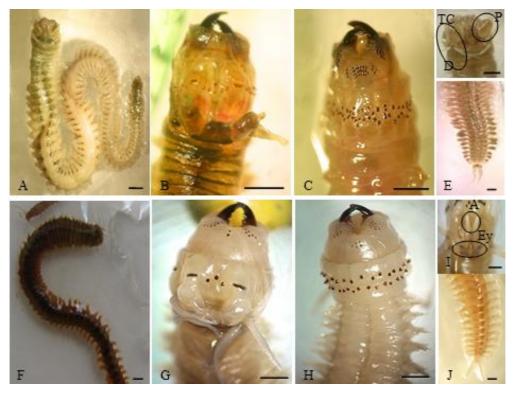


Fig 2: *P. macropus* (A) Dorsal view of whole animal; (B) Dorsal view of anterior region; (C) Ventral view of anterior region; (D) Dorsal view of details of the head; (E) Ventral view of pygidium; *P. cultrifera* (F) Dorsal view of anterior and middle region; (G) Dorsal view of anterior region; (H) Ventral view of anterior region; (I) Dorsal view of details of the head; (J) Ventral view of pygidium, (TC) Tentacular cirri, (P) Biarticulated palp, (Ey) Four eyes, (bar scale: 1mm).

Thus all nereids were easy to separate from other Polychaetes, but the difficulty was that *P. macropus* and *P. cultrifera* coexist with two other closer nereids *Nereis falsa* and *Platynereis dumerilii*, so we had followed the usual method for description, based on the number and shape of paragnaths on maxillary and oral ring of pharynx, divided into areas from

I to VIII ^[25], so first, we had been interested in area VI key element for description of the genus *Perinereis*, represented by a large transverse smooth bar, then identification to species level was conducted according to ^[24, 26] as summarized in Table 01.

Table 1: Number and shape of paragnaths on maxillary and oral rings on everted pharynx according to [24, 26].

| | Areas | P. macropus | P. cultrifera |
|-------------------|----------|--|---|
| Maxillary ring | I | 3-7 cones rarely 2 | 1-3 cones in longitudinal line |
| | II | each side triangular clusters of cones | each side cluster of cones in oblique rows |
| | III | rectangular cluster of cones flanked with 3-5 ones | rectangular cluster of cones in 2 rows |
| | IV | each side triangular clusters of cones | each side triangular clusters of cones |
| oral ring | V | 1 large and 5-12 (usually 10) sub equal in irregular line, or in a group | 3 cones in a triangle or 1 single (floridana) |
| | VI | each side 1 large transverse bar-shaped | each side 1 large transverse bar-shaped |
| | VII-VIII | 4-5 transverse rows of sub equal cones | 2 transverse rows of equal cones |

Morphological characters of parapodia, and chaetae structure are also important, and were examined for both species after performing cross sections at different region of the body; parapodia of the two first chaetegerous are uniramous with absence of notochaeta, whereas the rest are biramous, having two parts, the first is the notopodium at the dorsal side with a dorsal cirrus, and two conical ligules, between which we found notochaetae and a notoaciculum, the second is the neuropodium at the ventral one, with a ventral cirrus, one conical ligule, well developed tuft of neurochaetae and a neuroaciculum; chaetae are compounds homogomph spinigers, heterogomph spinigers and heterogomph falcigers, also in heteronereis species we noticed the natatory chaetae (Figure 5: J, K, L), (Figure 6: G).

3.3 Morphological variations

With more practice, the two species were first separated on the basis of color which is bright green and green bronze for *P. macropus*, and *P. cultrifera* respectively; also in preserved worms, we can notice a pigmentation pattern present dorsally in *P. macropus* (Figure 2: A, B), that varies from light to dark green bands, while for *P. cultrifera* it can be noticed on the prostomium and palps, and it appears to be easy to fade very quickly after preservation (Figure 4: D, E), this character is

useful for identifying criptic nereidid species [27].

In general P. macropus worms were clearly more elongated, they were longer with a higher chaetegerous number, compared with P. cultrifera worms, characterized by a longer L₃ and larger width size, beside longer tentacular cirri extending to the 5th-6th chaetegerous, while for P. macropus the longest ones can reach the 3rd-4th chaetegerous; the maximum number of chaetegerous of the two species is lower than that cited in [24], in Polychaetes, chaetegerous' number can vary considerably, in fact they can easily lose the posterior part of the body by autotomy, and regenerate it by addition of new segments from the posterior end of the body, called the posterior growth zone [28], In the present study worms with regenerating few or many chaetegerous are observed, from either the middle or posterior part of the body; thus the number of lost and replaced chaetigerous, affect even the total length, that sometimes does not correlate with other biometric measurements; also biometry of individuals P. cultrifera vary according to geographic location [29].

Parapodia from *P. cultrifera* shows no differences along the body, while for *P. macropus* we clearly notice a modified form of parapodia, that began from the posterior region and extend to the pygidium, with elongated and enlarged notopodial ligule and short dorsal cirrus (Figure 5: A-E, H, I).

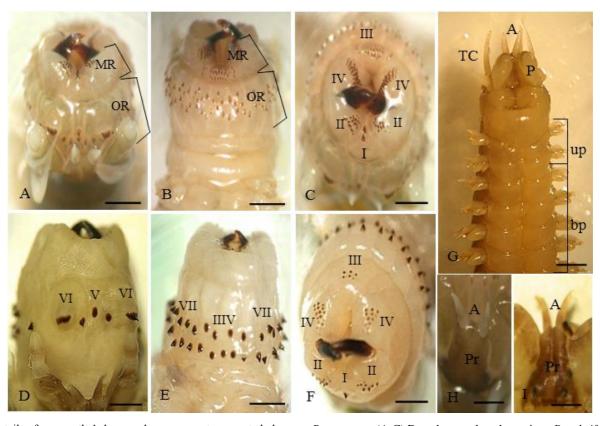


Fig 3: Details of paragnaths' shape and arrangement on everted pharynx; *P. macropus* (A-C) Dorsal, ventral, and top view; *P. cultrifera* (D-F) Dorsal, ventral, and top view; (G) General aspect of the anterior ventral part of a worm showing devagenated pharynx and parapodia; Prostomium and antenna for (H) *P. macropus*, and (I) *P. cultrifera*; (MR) Maxillary ring, (OR) Oral ring, (A) Antenna, (P) Biarticulated palp, (TC) Tentacular cirri, (up) Uniramous parapodia, (bp) Biramous parapodia, (Pr) Prostomium, (bar scale: 1mm).

The shape and distribution of paragnaths in the two species show rectangular-base paragnaths (bar-shaped), with pointed or rounded apex named Shield-shaped bars, (Figure 3: A, D) for area VI; this type is present in *Perinereis* species having a single area VI paragnaths, and *Pseudonereis* species [25]; the remaining areas are characterized by uniform-base paragnaths, with circular base, and are tapered towards an apex named conical paragnaths (Figure 3: B, C, E, F); size height and color of paragnaths in *P. cultrifera*, present

differences, first on the oral ring, are particularly larger, higher and brown darker, while those on the maxillary one are comparatively small and faint, second in contrast with *P. macropus* where paragnaths are almost similar in size, smaller and less darker, except those on the maxillary ring that are more slender and acutely pointed; unlike these types we found another type of the uniform-base paragnaths in *P. dumerelii*, with a circular base, which are long and slender, called Rodlike paragnaths.

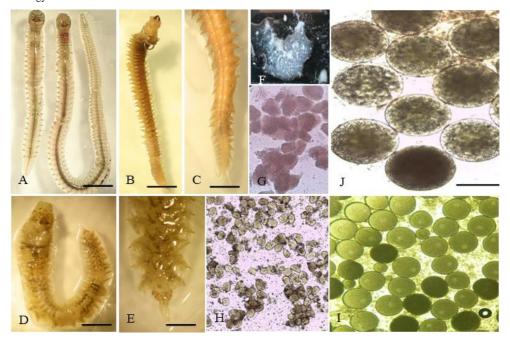


Fig 4: (A) Two individuals *P. macropus* showing differences in total length caused by autotomy, despite regenerating lost chaetigerous; (B) Regenerated chaetigerous beginning from the middle region of the body; (C) Close up of many regenerating chaetigerous; (D) Individual *P. cultrifera* showing a regenerating posterior region and a pigmentation pattern; (E) Close up of few regenerating chaetigerous; (F) Normal aspect of coelomic fluid containing sperm aggregates; (G-H) Sperm aggregates (I) Asynchronous oocytes; (J) Mature oocytes, (bar scale: 1mm, J: 200 μm), (G: x 100), (H, I, J: x 40).

3.4 Reproduction

The two species are gonochoric, males present sperm aggregates within coelomic cavity, whereas females present spherical or oval free oocytes within parapodia also, when females were completely packed with mature ones, this finding is similar to literature on oogenesis in nereids, gonads had never been localized, and oocytes grow free in the coelomic cavity [30], observations show that oocytes maturation is asynchronous in the two species (Figure 4: I), this agree with earlier studies where a clear heterogeneous size of oocytes occurs [31, 32], unlike in *Platynereis dumerelii* [33] and *Nereis falsa* [34]. where oocytes maturation is

synchronous; for *P. macropus* mature oocytes can reach a maximum diameter of 268.76±11.68 μm in July; for *P. cultrifera* maximum oocyte diameter showed a similar value 334.69±19.55 μm and 345.33±13.82 μm in May at both stations Collo and Skikda respectively, these values are similar of those mentioned in ^[29, 31, 32-35] suggested that oocyte growth follows an initial phase of very slow growth, a phase of rapid growth, and a final phase of egg differentiation with little growth, likewise ^[30]. Reported that oocytes proliferation and growth occur over an extended period of time, where smaller oocytes catch up the larger ones, to reach a uniform size, when spawning is imminent ^[36]. (Figure 4: J).

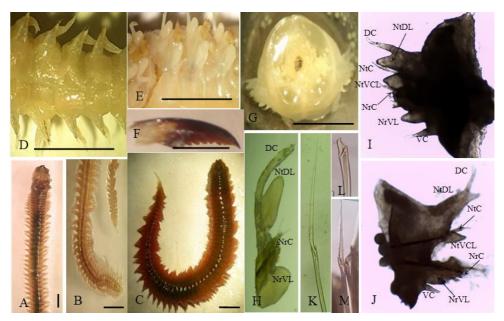


Fig 5: (A, B) *P. macropus* showing a modified form of parapodia of the posterior part; *P. cultrifera* with unmodified form along the whole body (C); Close up of parapodia from posterior region (D) *P. macropus*, (E) *P. cultrifera*; (F) Jaw; (G) Cross section of an anterior segment; (H) Uniramos parapodium; (I) Biramous parapodium from anterior region; (J) Biramous parapodium from posterior region; (K) Homogomph spiniger chaeta; (L) Heterogomph falciger chaeta; (M) Heterogomph spiniger chaeta; (DC) Dorsal cirrus, (NtDL) Notopodial dorsal ligulle, (NtC) Notochaeta, (NtVL) Notopodial ventral ligulle, (NrC) Neurochaeta, (NrVL) Neropodial ventral ligulle, (VC), (ventral cirrus), (bar scale: 3 mm, bar scale F: 0.5mm), (G, H, I: x 10), (J, K, L: x 40).

In Nereididea, the sexual reproduction is accompanied or not by morphological changes, called epitoky or atoky respectively, in the present study these changes was recorded only for *P. macropus*, the heteronereis stage is fully developed with strong epitokous modifications, greatly enlarged black eyes, flattened posterior parapodia as a result of changes in the parapodial lobes' morphology, with natatory chaetae and paddle for swimming, a change of the body coloration, and a reduction in worms' length and width, resulting from histolysis of correlated segments' muscles (Figure 6), this metamorphosis is a special reproductive form in Nereididae, during which in sexually maturing worms, new tissues differentiate, while old ones degenerate or

transdifferentiate ^[37], These observations agree with literature on other closer nereids species, where morphological, behavioral and physiological changes correlate with metabolic and biochemical ones ^[38], epitokous worms are prepared for a short pelagic existence during swarming, where they swim and release gametes on the water surface, days after spawning individual of the two sexes die, this agree with our observations, the two species are known to be strictly semelparous, they breed once in their life cycle; literature on *P. cultrifera* along the Algerian coast, reported two reproductive patterns, with atokous or epitokous type, according to geographic location ^[39].

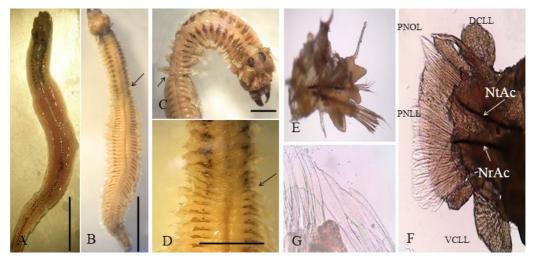


Fig 6: (A) Heteronereis specimen showing modification of the body color; (B) Another one showing morphological modifications of epitoky; Enlarged eyes, and the beginning of a modified region; (D) Close up of flatned parapodia, arrows indicate start of the first modified parapodium; (E) Parapodium from anterior region; (F) Parapodium from posterior region; (G) Natatory chaeta, (DCLL) Dorsal cirrus lamella, (PNOL) Post-chaetal neuropodial lamella, (NtAc) Notoaciculum, (PNLL) Post-chaetal neropodial lamella, (NrAc) Neuroaciculum, (VCLL) Ventral cirrus lamella, (bar scale: 5 mm); (E, F: x 10), (G: x 40).

4. Conclusion

Individuals of P. macropus and P. cultrifera were sampled at two stations Collo and Skikda, from hard substrates, to investigate their ecology, biology and reproduction in general, and also to give a description based on these aspects; our observations had clarified a difference between the associated algae species that might explain the absence of *P. macropus*, in Skikda, results of biometric measurements vary between the two species and among sites, some clear morphological variations can be detected such as color, paragnathes' shape and arrangement, and parapodia forms along the body, others require several macro and microscopic observations; oocytes' diameter values and their aspects indicate the maturation's degree of females, where we had easily noticed an heterogamous diameter in maturing females, indicating that oogenesis is asynchronous; concerning epitokal modifications adults of P. cultrifera show an atokous character of reproduction, and the two species are semelparous.

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6. References

- 1. Loreau M, De Mazancourt C. Biodiversity and ecosystem stability: a synthesis of underlying mechanisms Ecology Letters 2013; 16:106-115.
- 2. Coll M, Piroddi C, Albouy C, Ben Rais Lasram F, Cheung WWL, Christensen V *et al.* The Mediterranean Sea under siege: spatial overlap between marine biodiversity, cumulative threats and marine reserves.

- Global Ecology and Biogeography 2012; 21:465-480.
- 3. Lotze HK, Coll M, Magera AM, Ward-Paige C, Airoldi L. Recovery of marine animal populations and ecosystems, Trends in Ecology and Evolution 2011; 26(11):595-605.
- 4. Kroncke I. Long-term changes in North Sea benthos. Senckenbergiana Maritima 1995; 26(1/2):73-80.
- Papageorgiou N, Arvanitidis C, Eleftheriou A. Multicausal environmental severity: a flexible framework for microtidal sandy beaches and the role of polychaetes as an indicator taxon. Estuarine, Coastal and Shelf Science 2006; 70:643-653.
- 6. Deschênesa J, Desrosiersa G, Ferronb J, Cloutierb R, Stora G. Environmental influence on activity levels and behavioural allocation in the polychaete *Nereis virens* (Sars), Journal of Experimental Marine Biology and Ecology. 2005; 317:203-212.
- 7. Mouneyrac C, Perrein-Ettajani H, Amiard-Triquet C. Influence of anthropogenic stress on fitness and behaviour of a key-species of estuarine ecosystems, the ragworm Nereis diversicolor Environmental Pollution 2010; 158:121-128.
- 8. Bouraoui Z, Ghedira J, Banni M, Boussetta H. Acute effects of cadmium and copper on cytochemical responses in the polychaete Hediste diversicolor, International Journal of Environmental Research. 2016; 10(1):131-138.
- 9. Bakalem A, Ruellet T, Dauvin JC. Benthic indices and ecological quality of shallow Algeria fine sand community ecological indicators 2009; 9:395-408.

- 10. Dauvina JC, Andraded H, de-la-Ossa-Carreteroe JA, Del-Pilar-Rusoe Y, Riera R. Polychaete/amphipod ratios: An approach to validating simplebenthic indicators Ecological Indicators 2016; 63:89-99.
- 11. Grimes S. Biodiversité marine en Algérie: Crise des taxonomistes et enjeux de l'activité en réseau. Numéro spécial du bulletin du Centre National de Développement des Ressources Biologiques, 2011, 9-17.
- 12. Verdonschot PFM. Introduction to Annelida and the Class Polychaeta In: Thorp and Covich's Freshwater Invertebrates (Fourth Edition) Ecology and General Biology 2015; 1:509-528.
- 13. Blainville H. Mémoire sur la classe des Sétipodes, partie des Vers à sang rouge de M. Cuvier, et des Annélides de M. de Lamarck. Bulletin des Sciences, par la Société Philomatique de Paris, 1818, 78-85.
- Read G, Fauchald K. World Polychaeta database.
 Accessed through: Costello MJ, Bouchet P, Boxshall G,
 Arvantidis C, Appeltans W, 2016. European Register of
 Marine Species at http://www.marbef.org. 2016-03-29
- 15. Kinberg JGH, Annulata nova. Öfversigt af Königlich Vetenskapsakademiens förhandlingar, Stockholm 1865; 22(2):167-179.
- 16. Madani LN, Grimes S, Douga A, Oubraham Arezki S. Diversité des peuplements de polychètes de la baie d'Alger, Numéro spécial du bulletin du Centre National de Développement des Ressources Biologiques 2011, 81-87
- 17. Meghlaoui Z, Daas T, Snani M, Daas-Maamcha O, Scaps P. Annual variations of the presence of Nereididae (Annelida: Polychaeta) from intertidal rocky shores along the east coast of Algeria Check List 2015; 11(6):1-9.
- 18. Grube E. Actinien, Echinodermen und Würmer des Adriatischen und Mittlemeers. Könisberg, 1840, 92.
- Younsi M, Daas T, Daas O, Scaps P. Polychaetes of commercial interest from the Mediterranean east coast of Algeria. Mediterranean Marine Science 2010; 11:185-189.
- 20. Cardinaletti G, Mosconi G, Salvatori R, Lanari D, Tomassoni D, Carnevali O *et al.* Effect of dietary supplements of mussel and polychaetes on spawning performance of captive sole, *Solea solea* (Linnaeus, 1758). Anim Reprod Sci, 2008, 36-46.
- 21. Scaps P, Rouabah A, Leprêtre A. Morphological and biochemical evidence that Perinereis cultrifera (Polychaeta: Nereididae) is a complex of species, J Mar Biol Ass U.K. 2000; 80:735-736.
- 22. Claparède RE. Les annélides chétopodes du Golfe de Naples. Supplément, Société de physique et d'histoire naturelle de Genève, Genève et Bale, 1870, 236.
- 23. Fauvel P. Quatrième note préliminaire sur les Polychètes provenant des compagnes de l'Hirondelle et de la Princesse-Alice ou déposées dans le musée océanographique de Monaco. Bulletin océanographique de Monaco 1913; 253-273:1-80.
- 24. Fauvel P. Polychètes errantes. Faune de France 5, Paul Le Chevaliers, Paris 1923, 488.
- 25. Bakken T, Glasby CJ, Wilson RS. A review of paragnath morphology in Nereididae (Polychaeta), Zoosymposia 2009; 2:305-316.
- Imajima M, Gamô S. Polychaetous annelids from the intrtidal zone of manazourou, Kanagawa Prafecture. Science Reports of the Yokohama National University 1970; 16:1-18.
- 27. Glasby CJ, Wei NWV, Gibb KS. Cryptic species of

- Nereididae (Annelida: Polychaeta) on Australian coral reefs. Invertebrate Systematics 2013; 27:245-264.
- 28. Anderson DT, Kerkut GA. Embryology and phylogeny in annelids and arthropods. International series of monographs in pure and applied biology, Pargamon Press, 50 New York, 1973, 495.
- 29. Rouabah A, Scaps P. Life Cycle and Population dynamics of the polychaete Perinereis cultrifera from the Algerian Mediterranean Coast Marine Ecology 2003; 24(2):85-99.
- 30. Eckelbarger KJ. Oogenesis and oocytes Hydrobiologia 2005; 535(536):179-198.
- 31. Zribi S, Zghal F, Tekaya S. Ovogenèse de Perinereis macropus Claparède 1870 (Annélide, Polychète) dans le golfe de Gabès (Tunisie), Comptes Rendus Biologies 2007; 330(3):199-204.
- 32. Snani M, Meghlaoui Z, Maamcha O, Daas T, Scaps P. Laying period and biomarkers of the polychaete Perinereis cultrifera from the eastern coast of Algeria subjected to marine pollution, Journal of Entomology and Zoology Studies. 2015; 3(3):249-254.
- 33. Fischer A, Dorresteijn A. The polychaete Platynereis dumerilii (Annelida): a laboratory animal with spiralian cleavage, lifelong segment proliferation and a mixed benthic/pelagic life cycle BioEssays 2004; 26:314-325.
- 34. Daas T, Younsi M, Daas-Maamcha O, Gillet P, Scaps P. Reproduction, population dynamics and production of Nereis falsa (Nereididae: Polychaeta) on the rocky coast of El Kala National Park, Algeria Helgoland Marine Research 2010; 65(2):165-173.
- 35. Dhainaut A. Contribution à l'étude de la gamétogénèse des Néreidiens dans les conditions naturelles et en l'absence d'hormone cérébrale, Doctoral thesis, University of Lille, Lille, France, 1970.
- 36. Fidalgoe Costa P. The oogenic cycle of *Nereis diversicolor* (O. F. Müller, 1776) (Annelida: Polychaeta) in shallow water environments in southwestern Portugal. Boletín del Instituto Español de Oceanografía 2003; 19:17-29.
- 37. Fischer A. Reproductive and developmental phenomena in annelids: a source of exemplary research problems Hydrobiologia 1999; 402:1-20.
- 38. Hébert Chatelain E, Breton S, Lemieux H, Blier PU. Epitoky in Nereis (Neanthes) virens (Polychaeta: Nereididae): A story about sex and death. Comparative Biochemistry and Physiology 2008; 149(Part B):202-208.
- 39. Rouabah L, Rouabah A, Ferroudj S, Scaps P. Comparison of the life cycles of two populations of the polychaete Perinereis cf. cultrifera from the Bay of Algiers (Mediterranean Sea) Scientia Marina 2008; 72(4):769-778.