

E-ISSN: 2347-5129 P-ISSN: 2394-0506 (ICV-Poland) Impact Value: 5.62 (GIF) Impact Factor: 0.549 IJFAS 2017; 5(3): 297-305 © 2017 IJFAS www.fisheriesjournal.com Received: 10-03-2017 Accepted: 11-04-2017

El-Damhougy Faculty of Science, Al-Azhar University, Cairo.

El-Sayed Faculty of Science, Al-Azhar University, Cairo.

Aboul Ezz National Institute of Oceanography and Fisheries, Alexandria.

Abu Husein MS Faculty of Science, Al-Azhar University, Cairo.

Tintinnida (Ciliophora) as bio-indicator for certain pollutants at al-max area, Alexandria, Egypt

El-Damhougy KA, El-Sayed AA, Aboul Ezz SM and Abu Husein MS

Abstract

Tintinnids community was studied seasonally at Al-Max area during the period from spring 2014 to winter 2015. This study was aimed to throw light on the occurrence of tintinnid species that considered as bio-indicators for certain environmental factors. Fifty three species of tintinnids belong to 21 genera were recorded. From them eight species viz. *Eutintinnus pingius, Proplectella angistior, Favella ehrenbergii, Ascampbelliiella sp, Helicostomella subulata, Tintinnopsis nudicauda, Favella panamensis* and *Tintinnopsis campanula* are considered as bio-indicators for dissolved oxygen, salinity and pH. While *Favella panamensis, Favella ehrenbergii* and *Tintinnopsis campanula* showed that for nitrites and *Favella panamensis, Favella ehrenbergii* and *Tintinnopsis campanula* for nitrates. However, *Tintinnopsis nudicauda* is positively correlated for temperature and the other seven species are negatively correlated with temperature. Furthermore, *Tintinnopsis radix, Tintinnopsis campanula, Tintinnopsis nudicauda* and *Favella adriatica* are considered as euryhaline species.

Keywords: Tintinnids, bio-indicator, Al-Max Bay, Pollution & Environmental conditions

1. Introduction

Tintinnids are unicellular organisms. They form a group of loricate ciliates (Montagnes, 2013)^[25]. They inhabit marine and freshwaters (McManus and Santoferrara, 2013)^[24] and play a vital role in aquatic food chains which feed on phytoplankton and bacteria, in turn they act as food for larger organisms such as copepods and fish larvae (Stoecker, 2013)^[35].

Tintinnids are part of the microzooplankton that have many advantages as a favorable bioindicator to evaluate environmental stress and anthropogenic impacts in aquatic ecosystems (Jiang *et al.*, 2011, 2013a, 2013b, Xu *et al.*, 2011a, b, c, 2014) ^[19, 18, 20, 39, 40, 41, 42] and to monitor sea water quality (Wu *et al.* 2016) ^[38]. Owing to their short life cycle and delicate pellicles, they may respond more quickly to environmental changes than any other metazoans (Coppellotti and Matarazzo, 2000, Ismael and Dorgham, 2003) ^[8, 17]. Many ciliated microbiota can tolerate extreme of environmental hazards than macrofauna (Xu *et al.* 2011a, b) ^[39, 40].

Tintinnids are important components of the aquatic ecosystem and play a crucial role in transferring elements and energy from low trophic levels (pico- and nano-phytoplankton) to high one such as copepods (Crawford *et al.* 1997 and Corliss, 2002)^[10, 9]. This study aims to:

- Investigate distribution and abundance of tintinnids at Al-Max area.
- Determine the correlation coefficient between tintinnid density and certain environmental parameters.
- Clear the differentiation between different sites using diversity indices and similarity.
- Show the tintinnid species that considered as bio-indicators.

Materials and Methods

Study area: This study was carried out on tintinnid assemblages in Al-Max area during the period from spring 2014 to winter 2015. Four sites were selected at the study area to represent different habitats. The coordinates and local names are shown in Table (1) and Figure (1).

Correspondence El-Damhougy Faculty of Science, Al-Azhar University, Cairo.

Table (1): Show position of the selected sites at Al-Max area during the present study.

	9	North	East		
Ι	Al - Umoum Drain	31° 8' 56"	29° 50' 42"		
II	Al - Umoum Drain outlet	31° 9' 7.18"	29° 50' 27.04"		
III	Al-Max Bay (~ 1.2 Km far of the outlet)	31° 9' 22.56"	29° 49' 55.00"		
IV	Al-Max Bay (~ 4 Km far of the outlet)	31°10' 39.34"	29° 48' 52.71"		



Fig1: Map shows sites of collection at Al-Max area during the study period.

Values of both water temperature and dissolved oxygen were measured using TRANS digital dissolved oxygen meter model HD3030. Seawater salinity was determined by using ADWA digital Salinometer model AD 410, while pH values were measured using ADWA pH digital meter model AD 11. Dissolved nitrites, phosphates and COD were measured according to APHA (1995)^[7]. Dissolved nitrates were measured according to Mullin and Riley (1956)^[26].

Escherichia coli samples were collected by filling 10 ml sterilized glass bottles directly from the water followed by rapidly and tightly closing. They were kept in an ice box, and then estimated (as colonies per ml) through 48 h. in the center laboratory of drinking water Company at Abu Hommos, El-Behaira Governorate, Egypt.

Sample collection and treatment

Plankton samples were collected by filtering known volumes of water, using 55µm-mesh plankton¹net. These samples were preserved in 5% buffer formalin solution. In the laboratory, the sample volume has adjusted to 100 ml. Sub-samples of 0.5–1 ml were transferred to a counting chamber and then examined under a compound binocular and trinocular microscopes. For identification, many taxonomic guides have been used such as Tregouboff and Rose (1957) ^[36], Marchall (1969) ^[23], Paulier (1997) ^[28] and WORMS database ^[37].

The tintinnids standing crop was calculated as individuals per cubic meter according to Santhanam and Srinivasan (1994) ^[32] formula as following:

N = n (v/V) * 1000

Where, N= Total number of individuals per cubic meter; n= Average number of individuals in one ml of the sample; v= Volume of zooplankton concentrate (ml) and V= Volume of total water filtered (L).

Statistical analyses

Correlation analyses were carried out by computer Excel program and MINITAB 14 statistical program, while diversity indices similarity were done by PRIMER 5 statistical program.

Results and Discussion 1. Environmental conditions

Figure (2) exhibits that all variables showed distinct spatial differences except temperature. These results clarified that, water temperature ranged between 17.58 ± 0.08 °C at site IV during winter and 29.80 ± 0.70 °C at site I during summer. This pattern agrees with that noticed by Aboul Ezz *et al.* (2014) ^[5] and Shreadah *et al.* (2014) ^[34] at Al-Max Bay.

Salinity increases gradually from site I to site IV. It ranged between $2.47\pm0.09\%$ at site I in spring and $27.65\pm0.78\%$ at site IV in winter (Figure 2). This remarkable wide range in salinity may be due to the effects of huge volume of the discharged fresh water from Al-Umoum Drain into the bay and the evaporation by high temperature. These findings coincide with that showed by Nessim *et al.* (2010) ^[27], Aboul Ezz *et al.* (2014) ^[5], Abou Zaid *et al.* (2014) ^[3], and Shreadah *et al.* (2014) ^[34].

Values of pH ranged between 7.50 \pm 0.0 at site I during summer and 9.30 \pm 0.0 at site III during spring (Figure 2). Then pH occurred on alkaline side, this is related to high photosynthetic activity (Hammer, 1971 and Hegab, 2015) ^[14, 15].

Dissolved oxygen levels increased remarkably from 1.26 ± 0.16 mg/l at site II to 9.9 ± 0.17 mg/l at sites III and IV in spring (Figure 2). This is related to the effect of direct draining especially sewage at the former sites, and is in agreement with Heneash *et al.* (2015) ^[16].

The lowest value of COD was 5.47 ± 5.10 mg/l at site IV in winter, increased sharply into the highest average of 22.93 ± 5.10 mg/l at the same site in spring and 22.67 ± 0.92 mg/l at site II in winter (Figure 2). These values were greatly varied from site to another but their annual averages exhibited the maxima at sites II and I. The present results indicated that, the high level of COD at sites I and II is attributed to the high content of organic matters and wastes discharged from the surrounding factories within these two sites. The present findings are similar to that noticed by Aboul-Ezz *et al* (2014) ^[5] at Al-Max Bay.

The minimum concentrations of nitrite were 15.27 \pm 2.93 and 15.48 \pm 1.87 µg/l, at site II in spring and site I in summer,

respectively, increased to the maximum levels being 411.61±58.77µg/l at site I in autumn. The highest annual average was 124.79±95.89 µg/l at site I while the lowest one was 66.19±22.78 µg/l at site IV (Figure 2). These results are similar to that recorded by Shreadah et al. (2014) [34]. They reported that, nitrites ranged from 107.52 µg/l at Al-Umoum Drain outlet to 35 μ g/l at the highest offshore stations. Also, Nessim et al. (2010)^[27] recorded nitrites from 25.2 to 158.2 µg/l at Al-Max Bay. These changes are due to the increasing of agricultural wastes and fertilizers in the drain and high rate

of nitrate reduction and ammonia oxidation forming nitrite as intermediate state by the action of denitrifying bacteria. Nitrate values ranged between 0 at both sites III and IV in spring and 480.09±45.51µg/l at site II in autumn (Figure 2). This pattern of results coincides with Shreadah et al. (2014) ^[34] at Al-Max Bay. Furthermore, the biological consumption, particularly the phytoplankton uptake could be low leading to increase nitrate content in the medium (Satpathy et al. 2010a) [33]



Fig 2: Spatial and temporal variations in the environmental factors at Al-Max area selected sites from spring 2014 to winter 2015. ~ 299 ~

Phosphate concentrations were measured during summer, autumn and winter only. The average values of phosphates were highly fluctuated between 38.33 ± 00 and $564.99\pm00 \ \mu g/l$ at sites IV and I, respectively during winter. Usually, seawater serves as the main source of phosphates in estuarine and coastal waters except those receive fresh water loaded with domestic wastes contaminated by detergent runoff from fields rich with phosphate-phosphorous fertilizer (Satpathy *et al.* 2010a) ^[33].

In general, both sites I and II exhibited higher nutrient content, compared with the other sites. The enrichment is due to agricultural runoff, sewage effluents and untreated drainage.

Escherichia coli bacteria were counted during summer, autumn and winter only, their numbers at sites I and II were higher than that at sites III and IV. The values of this bacteria fluctuated sharply between 160 colonies/ml at site IV and 250000 colonies/ml at site I in summer (Figure 2). It is worth to mention that, *Escherichia coli* are found in the lowest intestine of warm-blooded organisms, so they were chosen to indicate the sewage pollution. The highest increase at Al-Umoum Drain especially in summer mostly was attributed to direct discharge from surrounding houses in addition to daily discharging semi and untreated sewage effluents.

2. Tintinnid faunal composition

During this work, fifty three (53) species in addition to cysts of tintinnids were recorded belonging to 11 families and 21 genera. Tintinnid species were divided into agglutinated and non-agglutinated (hyaline) forms on the basis of morphological characteristics. The agglutinated genus Tintinnopsis was the dominant one with a maximum 11 species forming 20.8% of total tintinnid species number and 67.66% (5300 ind./m³) of total tintinnid densities. These results are similar to that postulated for genus Tintinnopsis by previous authors as Abdel-Aziz (2004) [1] who listed 10 species, Dorgham et al. (2009) [11] who collected 14 species, Abou Zaid and Hellal (2012)^[4] who recorded 12 species, Abo-Taleb et al. (2016)^[2] who listed 17 species and Rakshit et al. (2017)^[29] who collected 13 species. But these results are higher than that reported by Aboul Ezz et al. (1990)^[6] and Dorgham et al. (2013) ^[12] recorded 3 species and 4 species, respectively.

Tintinnopsis followed by genera coxliella with 5 species, Codonellopsis, Eutintinnus, Favella (4 species for each),

Helicostomella and *Undella* (3 species for each), *Epiplocylis*, *Liprotintinnus*, *Metacylis*, *Parafavella* and *Proplectella* (2 species for each). The remaining nine genera were represented by a single species for each one.

The dominance of *Tintinnopsis* in estuarine and coastal waters during the present work agrees with that reported by Feng *et al.* (2015) ^[13] and Jiang *et al.* (2011) ^[19]. This may be due to their unique flexible adaptive strategies by which they reached their maximum abundance; or to the eurythermal and euryhaline nature of *Tintinnopsis* which can sustain in different aquatic environments. These data coincide with that observed by Krinsic (1987) ^[22] and Reynolds (1997) ^[30]. In contrast, non-agglutinated genera, such as *Favella, Metacylis, Eutintinnus, Amphorellopsis* and *Helicostomella* were recorded in low density (about 32% of the total tintinnid community).

3. Spatial distribution and abundance

Results in Table (2) show that, the tintinnid communities in the studied sites were varied greatly between sites. Sites IV and III recorded the highest species number (42 and 39 species, respectively), with high density forming 15175 and 11327 inds./m³ respectively. On the other hand, the lowest number of species was recorded at sites I and II, listed 1 and 5 species, respectively, forming 139 and 4695 inds./m³ respectively. The tintinnid biodiversity in this region has been largely affected by anthropogenic activities as well as natural catastrophic events. It is assumed that the prevalent site-specific variations in biodiversity might be related to the environmental conditions (Ismael and Dorgham, 2003) ^[17] and in other sites as mentioned by Coppellotti and Matarazzo (2000) ^[8].

Tintinnopsis nudicauda was the most dominant species at sites II, III & IV, represented 76.92, 51.98 and 50.18 %, respectively. While *Favella panamensis* dominated at sites III and IV amounted 15.61 and 13.63%, respectively. At the same time, *Favella ehrenbergii* dominated at site IV only, and had a low density attained 12.96%. However, only one species, *Tintinnopsis radix* was reported at all sites (Table 2). The other collected species considered rare. This refers to their high tolerance to different environmental parameters and in congruence with that noticed by Abo-Taleb *et al.* (2016) ^[2] who listed the last three species (*Favella panamensis, Favella ehrenbergii* and *Tintinnopsis radix*) within the most dominant species.

 Table 2: Spatial distribution, average density (ind./ m³) and relative abundance (RA) of the recorded species at Al-Max area during the study period from spring 2014 to winter 2015.

	Sites and density	I		п		III		IV	
	Species	D±SE	RA	D±SE	RA	D±SE	RA	D±SE	RA
	Family: Codonellidae (Kent, 1882)								
1	Tintinnopsis beroidea Stein, 1867	0	0.00	0	0.00	186±139	1.64	204±139	1.34
2	Tintinnopsis campanula (Ehrenberg, 1840)	0	0.00	139±139	2.96	1011±684	8.92	534±311	3.52
3	Tintinnopsis cylindrica Daday,1887	0	0.00	0	0.00	0	0.00	18±18	0.12
4	Tintinnopsis gracilis Kofoid & Campbell, 1929	0	0.00	0	0.00	26±21	0.23	11±11	0.07
5	Tintinnopsis karajacensis Brandt, 1896	0	0.00	0	0.00	211±211	1.86	179±179	1.18
6	Tintinnopsis levigataKofoid& Campbell, 1929	0	0.00	0	0.00	0	0.00	11±11	0.07
7	Tintinnopsis mortenseni Schmidt, 1901	0	0.00	0	0.00	44±44	0.39	28±28	0.19
8	Tintinnopsis nana Lohmann, 1908	0	0.00	0	0.00	13±13	0.11	46±46	0.30
9	Tintinnopsis nudicauda Paulmier, 1997	0	0.00	3611±3611	76.92	5888 ± 5831	51.98	7615±7391	50.18
10	Tintinnopsis radix Brandt, 1907	139±13 9	100.00	278±278	5.92	652±325	5.76	281±129	1.85
11	Tintinnopsis sp	0	0.00	0	0.00	14±14	0.12	62±25	0.41
12	Codonella sp	0	0.00	0	0.00	4±4	0.04	0	0.00
	Codonellopsis indica Kofoid & Campbell, 1929								

13		0	0.00	0	0.00	146+146	1 29	47+47	0.31
15	Codonellonsis americana Kofoid&Campbell	0	0.00	0	0.00	140±140	1.27	4/_4/	0.51
14		0	0.00	0	0.00	0	0.00	298±298	1.97
15	Codonellonsis sn	0	0.00	0	0.00	0	0.00	37+37	0.25
16	Codonellopsis sp	0	0.00	0	0.00	61+52	0.54	141+141	0.23
10	Eamily: Eninlocylididae	0	0.00	0	0.00	01±52	0.54	141-141	0.75
17	Fninlocylis atlantica Kofoid & Campbell 1929	0	0.00	0	0.00	18+13	0.16	8+8	0.05
18	Epipiocytis anamica Koloid & Campbell, 1929	0	0.00	0	0.00	0	0.10	11+11	0.05
10	Epipiocyiis sp Family: Metacylididae	0	0.00	0	0.00	0	0.00	11-11	0.07
19	Climacocylis scalaria (Brandt 1906)	0	0.00	0	0.00	4+4	0.04	0	0.00
20	Metacylis lucasensis	0	0.00	0	0.00	0	0.04	8+8	0.00
20	Metacylis annulifera (Ostenfeld&Schmidt 1901)	0	0.00	0	0.00	28+28	0.00	94+56	0.62
21	Coxliella annulata (Daday, 1886)	0	0.00	0	0.00	0	0.25	11+11	0.02
22	Coxligila holivari Osorio Tafall 19/1	0	0.00	0	0.00	15+15	0.00	0	0.07
23	Coxliella fasciata (Kofoid 1905)	0	0.00	0	0.00	15±15	0.13	0+0	0.00
24	Coxtiella Jasciala (Rotold, 1905)	0	0.00	0	0.00	4±4	1.00	9±9	0.00
23	Coxtietta tacintosa (Brandt, 1900)	0	0.00	0	0.00	125±105	1.09	10+10	0.00
20	Coxitetta sp Helioostomolla edentate (Equité Erémiet, 1008)	0	0.00	0	0.00	4	0.05	19±19	0.12
27	Helicostometia edentate (Faule-Fielmet, 1908)	0	0.00	0	0.00	4±4	0.04	25 + 15	0.00
28	Helicostomella kilensis (Laackmann, 1906)	0	0.00	0	0.00	4 ± 4	0.03	25±15	0.10
29	Helicostomella subulata (Enrenberg, 1833)	0	0.00	0	0.00	68±45	0.60	276±172	1.82
30	Family: Ascampbelliellidae	0	0.00	279,279	5.02	0	0.00	0	0.00
	Ascampbelliella sp	0	0.00	278±278	5.92	0	0.00	0	0.00
21	Family: Ptychocyhdidae	0	0.00	200 - 200	8 20	25 1 4	0.22	42 - 42	0.28
22	<i>Faveila aarianca</i> Jorgensen, 1924	0	0.00	<u>389±389</u>	8.29	25 ± 14	0.22	42±42	0.28
32	Favella companula (Schmidt, 1901)	0	0.00	0	0.00	35±29	0.30	1/9±1/9	1.18
33	Favella enrenbergii (Claparede&Lachmann,	0	0.00	0	0.00	631±316	5.57	1967±1222	12.96
24	1636)	0	0.00	0	0.00	17(9+1170	15 (1	20(0) 1524	12 (2
54	Favena panamensis Kololux Callipbell, 1929	0	0.00	0	0.00	1/08±11/0	13.01	2009±1324	15.05
	Phahdonallongis longiagulis Vofoid & Comphall								
35		0	0.00	0	0.00	0	0.00	11±11	0.07
	Eamily: Tintinnidae								
36	Amphoralopsis tatragona (Jörgensen, 1924)	0	0.00	0	0.00	15+15	0.13	60+50	0.30
	Eutentinnus tereascans (Kofoid & Campbell	0	0.00	0	0.00	15±15	0.15	00±30	0.57
37	1929)	0	0.00	0	0.00	0	0.00	8 ± 8	0.05
38	Eutintinnus alagans Kofoid & Campbell 1929	0	0.00	0	0.00	11+11	0.10	22+22	0.15
30	Eutintinnus lusus undas (Entr. 1885)	0	0.00	0	0.00	11 ± 11 20+20	0.10	25+15	0.15
40	Eutintinnus ningius (Kofoid & Campbell 1020)	0	0.00	0	0.00	20 <u>+</u> 20 83+83	0.17	307+261	2.61
40	Liprotintinnus bottnicus (nordavist 1800)	0	0.00	0	0.00	36+22	0.73	0	0.00
41	Liprotintinuus vordavisti (Brandt 1906)	0	0.00	0	0.00	<u> </u>	0.32	40+27	0.00
42	Exproventing the second	0	0.00	0	0.00	44_44	0.39	40±27	0.20
43	Undella clevi Jörgensen 1924	0	0.00	0	0.00	0	0.00	8+8	0.05
44	Undella homisphaorica Lapokmann, 1000	0	0.00	0	0.00	4+4	0.00	0	0.05
44	Undella ostanfaldi Kofoid&Campbell 1020	0	0.00	0	0.00	4 <u>+</u> 4	0.03	0	0.00
45	Undellopsis sp	0	0.00	0	0.00	4 <u>+</u> 4	0.04	0	0.00
40	Proplectalla angistion (Jörgenson, 1024)	0	0.00	0	0.00	10 ± 13 70 ± 70	0.10	234+234	1.54
47	Duorlootalla an	0	0.00	0	0.00	19±19	0.70	10+10	0.12
40	Eamily: Xystonellidae	0	0.00	0	0.00	0	0.00	19±19	0.12
49	Parafavalla evindriag (Jörgensen, 1800)	0	0.00	0	0.00	0	0.00	11+11	0.07
50	Parafavella en	0	0.00	0	0.00	4+4	0.00	22+22	0.07
51	Parundalla massinensis (Drondt 1006)	0	0.00	0	0.00	4 <u>±</u> 4 12,⊥12	0.04	23±23	0.13
51	Family Detalatriabidas	0	0.00	U	0.00	15±15	0.11	U	0.00
50	Patalotricha ampulla (Ecl. 1991)	0	0.00	0	0.00	0	0.00	72-72	0.47
52	<i>Cymatocylis calyciformis</i> (Lasekmann, 1000)	0	0.00	0	0.00	0	0.00	1/ <u></u> 1/ <u></u> 1/	0.47
53	Cymulocyus curycijornus (Laackinanii, 1909)	0	0.00	0	0.00	U 12-12	0.00	14-14	0.09
54	Cysis of unumuas	0	0.00	U	0.00	12112	0.11	0	0.00
	Total No. of individuals	139	100.00	4695	100.00	11327	100.00	15174	100.00

4. Temporal Distribution and Abundance

The highest number of species (32 species) was recorded in summer, from them five species dominated the others viz, *Eutintinnus pingius, Proplectella angistior, Favella ehrenbergii, Ascampbelliiella sp* and *Helicostomella subulata.* These species were represented by 17.38, 15.16, 14.04, 13.49 and 10.94%, respectively. In autumn, *Tintinnopsis nudicauda was the only dominant species (88.21%).* While in winter *Favella panamensis, Favella ehrenbergii* and *Tintinnopsis campanula* were dominated by 36.73, 22.21 and 14.88% of

total individuals, respectively. During spring, the same three species (*F. panamensis*, *F. ehrenbergii* and *T. campanula*) mentioned above during winter were dominated by 41.76, 23.15 and 22.31%, respectively. At the same time, *Favella ehrenbergii* and *Tintinnopsis radix* appeared all the year round. However, *Favella panamensis*, *Tintinnopsis beroidea* and *Tintinnopsis campanula* appeared at all seasons except autumn, while *Tintinnopsis gracilis* and *Tintinnopsis nudicauda* occurred all the year round except winter. While *Amphorelopsis tetragona* was recorded during all seasons

except spring. In addition *Tintinnopsis sp.* disappeared only in summer. These results showed that, 16 species occurred only in two seasons, eight of them were listed in summer and autumn. Regarding the rest 29 species, all of them were noticed only during one season (Table 3).

Favella panamensis and *Favella ehrenbergii* were abundant especially during winter, forming 58.94% of the total individuals during this study. In addition to their frequency during spring, summer and winter for the former and all the

year round for the later. These species were noticed only at offshore sites (III & IV). This refers to their less tolerance to fresh water and other pollutants, and coincides with that reported by Dorgham *et al.* (2013) ^[12] where they recorded *Favella ehrenbergii* at less stressed area.

Table (3): Temporal average density (inds./ m^3) and relative abundance (RA) of the recorded species at Al-Max area during the study period from spring 2014 to winter 2015.

Seasons		Spring		Summer		Autumn		Winter		An AV	An DA
	Species	D±SE	RA	D±SE	RA	D±SE	RA	D±SE	RA	All. Av	AII, KA
1	Tintinnopsis beroidea	79±46	3.35	12±7	0.56	0	0.00	300±173	3.86	97	1.24
2	Tintinnopsis campanula	525 ± 304	22.31	4±4	0.19	0	0.00	1155±630	14.88	421	5.37
3	Tintinnopsis cylindrica	0	0.00	0	0.00	0	0.00	18 ± 18	0.24	5	0.06
4	Tintinnopsis gracilis	11±11	0.48	4±4	0.19	22±22	0.11	0	0.00	9	0.11
5	Tintinnopsis karajacensis	0	0.00	0	0.00	0	0.00	390±226	5.02	97	1.24
6	Tintinnopsis levigata	11±11	0.48	0	0.00	0	0.00	0	0.00	3	0.04
7	Tintinnopsi smortenseni	0	0.00	0	0.00	72±43	0.38	0	0.00	18	0.23
8	Tintinnopsis nana	0	0.00	0	0.00	0	0.00	59±44	0.75	15	0.19
9	Tintinnopsis nudicauda	42±42	1.79	170±120	8.26	16902±6453	88.21	0±0	0.00	4279	54.62
10	Tintinnopsis radix	34±34	1.43	71±57	3.44	787±227	4.10	458±290	5.90	337	4.30
11	Tintinnopsis sp	11±11	0.48	0	0.00	24±24	0.12	41±26	0.53	19	0.24
12	Codonella sp	0	0.00	4±4	0.19	0	0.00	0	0	1	0.01
13	Codonellopsis indica	0	0.00	0	0.00	194±138	1.01	0	0	48	0.61
14	Codonellopsis americana	0	0.00	0	0.00	298±298	1.56	0	0	75	0.96
15	Codonellopsis sp	0	0.00	0	0.00	37±37	0.20	0	0	9	0.11
16	Codonellopsis sp	0	0.00	8±8	0.38	194±133	1.01	0	0	51	0.65
17	Epiplocylis atlantica	0	0.00	12±7	0.57	0	0.00	14±14	0.18	6	0.08
18	Epiplocylis sp	11±11	0.48	0	0.00	0	0.00	0	0.00	3	0.04
19	Clymacocylis scalaria	0	0.00	4 <u>+</u> 4	0.19	0	0.00	0	0	1	0.01
20	Metacylis lucasensis	0	0.00	<u>8±8</u>	0.38	0	0.00	0	0.00	2	0.03
21	Metacylis annulifera	0	0.00	66±39	3.23	56±56	0.29	0	0.00	31	0.40
22	Coxliella annulata	11±11	0.48	0	0.00	0	0.00	0	0	3	0.04
23	Coxliella bolivari	0	0.00	0	0.00	0	0.00	15±15	0.2	4	0.05
24	Coxliella fasciata	0	0.00	4±4	0.19	9±9	0.05	0	0.00	3	0.04
25	Coxliella laciniosa	0	0.00	109±109	5.29	0	0.00	14±14	0.18	31	0.40
26	Coxliella sp	0	0.00	4±4	0.19	19±19	0.10	0±0	0.00	6	0.08
27	Helicostomella edentate	0	0.00	4±4	0.19	0	0.00	0	0.00	1	0.01
28	Helicostomella kilensis	0	0.00	19±15	0.94	9±9	0.05	0	0.00	/	0.09
29	Helicostomella subulata	0	0.00	225±169	10.94	119±93	0.62	0	0.00	86	1.10
30	Ascampbelliella sp	0	0.00	2/8±2/8	13.49	0	0.00	0	5 72	09 114	0.88
31	Favella darianca	0	0.00	0	0.00	11±11	0.06	445 ± 372	5.75	52	1.40
32	Favella companula	0	0.00	4±4	0.19	0	0.00	210±171	2.7	55	0.08
24		344±317	23.13	289±255	0.10	39±23	0.21	$1/24\pm1310$	22.21	049	0.20
25	Phate day all an air law air and	982±370	41.70	4±4	0.19	0	0.00	2831±1073	30.73	939	12.24
33	Rhabaonellopsis longicaulis	11±11	0.48	0	0.00	52,52	0.00	0	0.00	3	0.04
36	Amphorelopsis tetragona	0	0.00	8±8	0.38	52±52	0.27	15±15	0.2	19	0.24
37	Eutentinnus tergescens	0	0.00	8±8	0.38	0	0.00	0	0.00	2	0.03
38	Eutintinnus elegens	34±21	1.43	0	0.00	0	0.00	0	0.00	8	0.10
39	Eutintinnus lusus-undae	0	0.00	35±21	1.71	9±9	0.05	0	0.00	11	0.14
40	Eutintinnus pingius	0	0.00	358±259	17.38	122±122	0.64	0	0.00	120	1.53
41	Liprotintinnus bottnicus	0	0.00	0	0.00	22±22	0.12	14 ± 14	0.18	9	0.11
42	Liprotintinnus nordqvisti	11±11	0.48	0	0.00	73±44	0.38	0	0.00	21	0.27
43	Undella clevi	0	0.00	8 ± 8	0.38	0	0.00	0	0.00	2	0.03
44	Undella hemisphaerica	0	0.00	4±4	0.19	0	0.00	0	0.00	1	0.01
45	Undella ostenfeldi	0	0.00	4+4	0.19	0	0.00	0	0.00	1	0.01
46	Undellonsis sn	0	0.00	1=1	0.19	0	0.00	1/1+1/	0.18	5	0.06
47	Proplectella angistion	0	0.00	312+220	15.16	0	0.00	0	0.00	78	1.00
19	Proplectella sp	0	0.00	0 0	0.00	10+10	0.00	0	0.00	5	0.06
40	Parafavella cylindrica	11+11	0.00	0	0.00	0	0.10	0	0.00	3	0.00
	Parafavella sp	23+23	0.40	<u></u>	0.00	0	0.00	0	0.00	7	0.04
51	Parundella messinensis	0	0.90	0	0.19	0	0.00	13+13	0.00	3	0.09
52	Petalotricha ampulla	0	0.00	0	0.00	72+72	0.38	0	0.00	18	0.23
53	Cymatocylis calyciformis	0	0.00	0	0.00	0	0.00	14+14	0.18	3	0.04
54	Cysts of tintinnids	0	0.00	12+12	0.58	0	0.00	0+0	0.00	3	0.04
~ 1	Total individuals	2352	100.00	2060	100.00	19161	100.00	7763	100.00	8414	100.00
	No. of species	16	100.00	32	100.00	23	100.00	19	100.00	5.11	54
	r seres	10		52		25		./			

5. Correlation Coefficients

Table (4) exhibits tintinnid densities which have significant positive correlation coefficients all the year round with salinity (r = 0.88, 0.68, 0.78 and 0.84), pH (r = 0.88, 0.83, 0.78 and 0.84) and dissolved oxygen (r = 0.90, 0.78, 0.65 and 0.76) in spring, summer, autumn and winter respectively. This means that the abundant tintinnid species during these seasons viz, *Eutintinnus pingius*, *Proplectella angistior*, *Favella ehrenbergii*, *Ascampbelliiella sp Helicostomella subulata*, *Tintinnopsis nudicauda*, *Favella panamensis* and *Tintinnopsis campanula* can tolerate high values of salinity, pH and DO, and then they prefer alkaline and oxygenated water. This agrees with that mentioned by Heneash *et al.* (2015) ^[16].

At the same time, it significantly correlated positively with nitrites in spring (r = 0.59) which shows that the abundant tintinnid species in this season viz, *Favella panamensis*, *Favella ehrenbergii* and *Tintinnopsis campanula* can tolerate

high nitrite concentration. Also tintinnid species were correlated significantly in positive pattern with nitrates in winter (r = 0.78), which indicates that the abundant tintinnid species in winter viz, *Favella panamensis*, *Favella ehrenbergii* and *Tintinnopsis campanula* can tolerate high nitrate concentration. This is related to the feeding habits of tintinnids which consume algae that need nutrients through their growing (Kamiyama and Arima, 2001, Rosetta and McManus, 2003 and Montagnes, 2013) ^[21, 31, 25].

At the same time, it is revealed that, tintinnid species had significantly positive correlation with temperature in autumn (r = 0.59). On the other hand, negative correlation was signified with phosphates during all the three studied seasons (r = -0.68, -0.79 and -0.84) in summer, autumn and winter, respectively. This means that more concentrations of phosphates affect negatively tintinnid densities.

 Table 4: Multiple correlation coefficient values (r) between tintinnid densities and physic-chemical parameters at Al-Max area during the study period from spring 2014 to winter 2015.

Seasons	Parameters	r	p-Value	Seasons	Parameters	r	p-Value
	Tem.	-0.73	0.007		Tem.	0.59	0.04
	Salinity	0.88	0.0001		Salinity	0.78	0.003
	pН	0.87	0.0002		pН	0.78	0.003
	DO	0.90	0.00006		DO	0.65	0.02
Spring	COD	0.32	0.31	Autumn	COD	-0.34	0.28
	Nitrites	0.59	0.045		Nitrites	-0.79	0.002
	Nitrates	-0.33	0.29		Nitrates	0.09	0.79
	Phosphates				Phosphates	-0.79	0.002
	E. coli				E. coli	-0.24	0.756
	Tem.	-0.72	0.009		Tem.	-0.85	0.0007
	Salinity	0.68	0.01		Salinity	0.84	0.001
	pН	0.83	0.0009		pН	0.84	0.001
	DO	0.78	0.003		DO	0.76	0.004
Summer	COD	-0.10	0.75	Winter	COD	-0.65	0.02
	Nitrites	0.04	0.90		Nitrites	0.2	0.52
	Nitrates	0.01	0.98		Nitrates	0.78	0.003
	Phosphates	-0.68	0.02		Phosphates	-0.84	0.001
	E. coli	-0.65	0.354		E. coli	-0.62	0.379

6. Similarity index



Fig 3: Cluster diagram showing similarity between different sites at Al-Max area during the study period from spring 2014 to winter 2015.

7. Biodiversity measures

Figure (4) shows that species richness has the lower values 0 and 0.47 at site I and II, respectively. This reflects the high pollution at those sites. On contrast, the highest richness values were 4.3 and 4.1, recorded at sites IV and III. These results mean that sites III and IV may be subjected to moderate pollution. However, Shannon diversity exhibited the lowest values 1.16×10^{-15} and 0.8 at sites I and II, respectively. This shows that these sites are polluted while the highest values were 1.9 and 1.8 at sites IV and III respectively, which mean moderate pollution. Furthermore, evenness index revealed that there is no evenness value at site I due to presence of *Tintinnopsis radix* only, while amounted 0.5 at the other three sites, which means equitability at those sites.



Fig 4: Spatial variations in species richness, evenness and Shannon diversity of tintinnids at Al-Max area sites during the study period from spring 2014 to winter 2015.

Conclusion

- 1. -Fifty three species of tintinnids were recorded during this work; they were dominated by genus *Tintinnopsis*, from them some species are considered as bio-indicator for some physico-chemical parameters as the following:
- a. Eutintinnus pingius, Proplectella angistior, Favella ehrenbergii, Ascampbelliiella sp, Helicostomella subulata, Tintinnopsis nudicauda, Favella panamensis and Tintinnopsis campanula for dissolved oxygen, salinity and pH.
- b. *Favella panamensis*, *Favella ehrenbergii* and *Tintinnopsis campanula* for nitrite.
- c. Favella panamensis, Favella ehrenbergii and Tintinnopsis campanula for nitrate.
- d. *Tintinnopsis nudicauda* for high temperature while all above species except *Tintinnopsis nudicauda* are considered as bio-indicator for low temperature.
- 2. *Tintinnopsis radix, Tintinnopsis campanula, Tintinnopsis nudicauda* and *Favella adriatica* considered as euryhaline species.

References

- Abdel-Aziz NE. Changes of zooplankton community in a chronic eutrophic Bay on Alexandria coast, Egypt. Bulletin of Faculty. Of Science, Alexandria University. 2004; 43(1&2):203-220.
- 2. Abo-Taleb HA, Abdel Aziz NE, AboulEzz SM, El Raey M, Abou Zaid MM. Study of Chromista and Protozoa in a Hotspot area at the Mediterranean Coast with Special Reference to the Potentiality to Use It as Bio-indicators. International Journal of Marine Science, 2016; 6(53):1-17.
- 3. Abou Zaid MM, El Raey M, AboulEzz SM, Abdel Aziz NE, Abo-Taleb HA. Diversity of Copepoda in a Stressed Eutrophic Bay (El-Mex Bay), Alexandria, Egypt. Egyptian Journal of Aquatic Research. 2014; 40: 143-162.
- Abou Zaid MM, Hellal AM. Tintinnids (Protozoa: Ciliata) from the coastof Hurghada Red Sea, Egypt. Egyptian Journal of Aquatic Research. 2012; 38: 249– 268.
- 5. Aboul Ezz SM, Abdel Aziz NE, Abou Zaid MM, El Raey M, Abo-Taleb HA. Environmental assessment of El-Mex Bay, Southeastern Mediterranean by using Rotifera as a plankton bio-indicator. Egyptian Journal of Aquatic Research. 2014; 40: 43-57.
- 6. Aboul Ezz SM, Hussein MM, Sallam NA. Effect of domestic sewage discharge on the distribution of zooplankton organisms in the Eastern harbor of Alexandria (Egypt). The bulletin of the high institute of public health. 1990; XX (4): 861-874.

- 7. APHA (American Public Health Association). Standard methods for the examination of water and wastewater. New York. 1995, 1193.
- 8. Coppellotti O, Matarazzo P. Ciliate colonization of artificial substrates in the lagoon of Venice. Journal of Marine Biology Association, UK. 2000; 80:419–427.
- 9. Corliss JO. Biodiversity and biocomplexity of the protists and an overview of their significant roles in maintenance of our biosphere. Acta Protozoologica. 2002; 41:199– 220.
- 10. Crawford DW, Purdie DA, Lockwood APM, Weissman P. Recurrent red-tides in the Southampton water estuary caused by the phototrophic ciliate mesodiniumrubrum. Estuarine, Coastal and Shelf Science. 1997; 45: 799-812.
- 11. Dorgham MM, Abdel-Aziz NE, El-Ghobashy A, El-Tohamy WS. Preliminary study on protozoan community in Damietta Harbor, Egypt. Global Veterinaria, 2009; 3(6):495-502.
- 12. Dorgham MM, El-Tohamy WS, Abdel-Aziz NE, El-Ghobashy A, Qin JG. Protozoa in a stressed area of the Egyptian Mediterranean coast of Damietta, Egypt. Oceanologia, 2013; 55(3):733-750.
- Feng M, Zhang W, Wang W, Zhang G, Xiao T, Xu H. Can tintinnids be used for discriminating water quality status inmarine ecosystems? Marine Pollution Bulletin. 2015; 101:549-555.
- Hammer UI. Limnological studies of the lakes and streams of the Upper Qu' Appelle River System, Saskatchewan, Canada. Hydrobiologia. 1971; 37:437-504.
- 15. Hegab MH. Effect of some environmental parameters on distribution of zooplankton in Lake Nasser. Ph.D Thesis, Department of zoology, Faculty of Science, Al-Azhar University, Cairo, 2015, 263.
- 16. Heneash AMM, Tadrose HRZ, Hussein MMA, Hamdona SK, Abdel-Aziz N, Gharib SM. Potential effects of abiotic factors on the abundance and distribution of the plankton in the Western Harbour, south-eastern Mediterranean Sea, Egypt. Oceanologia, 2015; 57(1):61-70.
- 17. Ismael AA, Dorgham MM. Ecological indices as a tool for assessing pollution in El-Dekhaila Harbour (Alexandria, Egypt). Oceanologia. 2003; 45:121-31.
- 18. Jiang Y, Xu H, Hu X, Warren A, Song W. Functional groups of marine ciliated protozoa and their relationships to water quality. Environmental Science and Pollution Research. 2013; 20:5272-5280.
- 19. Jiang Y, Xu H, Hu X, Zhu M, Al-Rasheid KA, Warren A. An approach to analyzing spatial patterns of planktonic ciliate communities for monitoring water quality in Jiaozhou Bay, northern China. Marine Pollution Bulletin.

2011; 62:227-235.

- 20. Jiang Y, Yang EJ, Min JO, Kang SH, Lee SH. Using pelagic ciliated microzooplankton communities as an indicator for monitoring environmental condition under impact of summer sea-ice reduction in western Arctic Ocean. Ecological Indicators, 2013; 34:380-390.
- 21. Kamiyama T, Arima S. Feeding characteristics of two tintinnid ciliate species on phytoplankton including harmful species: effects of prey size on ingestion rates and selectivity. Journal of Experimental Marine Biology and Ecology, 2001; 257(2):281-296.
- 22. Krinsic F. On the ecology of tintinnines (Ciliata– Oligotrichida, Tintinnina) in the open waters of the south Adriatic. Marine Biology. 1987; 68:83-90.
- 23. Marshall SM. Protozoa, order Tintinnia. Fiches d'indentification de Zooplancton.Conseil International pour l'Exploration de la Mer, Copenhagen. 1969; 117-127.
- McManus GB, Santoferrara LF. Tintinnids in microzooplankton communities. In: Dolan JR, Montagnes DJS, Agatha S, Coats WD, Stoecker DK, editors. The biology and ecology of tintinnid ciliates: models for marine plankton. West Sussex, Wiley-Blackwell, 2013; 198-213.
- 25. Montagnes DJS. Ecophysiology and behavior of tintinnids. In: Dolan JR, Montagnes DJS, Agatha S, Coats WD, Stoecker DK, editors. The biology and ecology of tintinnid ciliates: models for marine plankton. Wiley-Blackwell, West Sussex, 2013; 85-121.
- 26. Mullin JB, Riely JP. The spectrophotometric determination of nitrate in natural waters, with particular references to see water. Analytica Chimica Acta, 1956; 12:479-480.
- 27. Nessim RB, Bassiouny AR, Zaki HR, Moawad MN, Kandeel KM. Environmental studies at El-Mex Region (Alexandria, Egypt) during 2007-2008. World Applied Science Journal. 2010; 9(7):779-787.
- 28. Paulmier G. Tintinnides (Ciliophora, Oligotrichida, Tintinna) DE L'Atlantique Boréal, De L'océan Indien Et De Quelques Mers Adjacentes: Méditerrannée, Mer Caraïbe, Mer Rouge. Inventaire Et Distribution. Observations Basées Sur Les Loricas. Station IFREMER Place du Séminaire, France, 1997, 135.
- 29. Rakshit D, Sahu G, Mohanty AK, Satpathy KK, Jontathan MP, Murugan K Sarkar SK. Bioindicator role of tintinnid (Protozoa: Ciliophora) for water quality monitoring in Kalpakkam, Tamil Nadu, south east coast of India. Marine Pollution Bulletin. 2017; 114:134-143.
- Reynolds CS. Vegetation Processes in the Pelagic: a model for ecosystem theory. Ecology Institute, Oldendorf/Luhe, Germany, 1997; 77:371.
- 31. Rosetta CH, McManus GB. Feeding by ciliates on two harmful algal bloom species, *Prymnesium parvum* and *Prorocentrum minimum*. Harmful Algae. 2003; 2(2): 109-126.
- 32. Santhanam R, Srinivasan A. A Manual of Marine Zooplankton. Oxford & IBH Publishing Co. New Delhi, 1994, 160.
- 33. Satpathy KK, Mohanty AK, Natesan U, Prasad MVR, Sarkar SK. Seasonal variations in physicochemical properties of coastal waters of Kalpakkam, east coast of India with special emphasis on nutrients. Environmental Monitoring and Assessment, 2010; 164:153-171.
- 34. Shreadah MA, Masoud MS, Khattab AM, El Zokm GM.

Impacts of different drains on the seawater quality of El-Mex Bay (Alexandria, Egypt). Journal of Ecology and the Natural Environment. 2014; 8(8):287-303.

- 35. Stoecker DK. Predators of tintinnids. In: Dolan JR, Montagnes DJS, Agatha S, Coats WD, Stoecker DK, editors. The biology and ecology of tintinnid ciliates: models for marine plankton. Wiley-Blackwell, West Sussex, 2013; 122-144.
- 36. Tregouboff G, Rose M. Manuel De Planctonologie Mediterraneenne. Centre National De La Recherche Scientifique, Paris, 1957, 587.
- 37. WORMS, http://www.marinespecies.org/
- Wu F, Huang J, Dai M, Liu H, Huang H. Using ciliates to monitor different aquatic environments in Daya Bay, South China Sea. Canadian Journal of Zoology, 2016; 94(4):265-273.
- 39. Xu H, Jiang Y, Al-Rasheid KA, Al-Farraj SA, Song W. Application of an indicator based on taxonomic relatedness of ciliated protozoan assemblages for marine environmental assessment. Environmental Science and Pollution Research, 2011; 18:1213-1221.
- 40. Xu H, Zhang W, Jiang Y, Min GS, Choi JK. An approach to identifying potential surrogates of periphytic ciliate communities for monitoring water quality of coastal waters. Ecological Indicators. 2011b; 11:1228-1234.
- 41. Xu H, Zhang W, Jiang Y, Zhu M, Al-Rasheid KA, Warren A, *et al.* An approach to determining the sampling effort for analyzing biofilm-dwelling ciliate colonization using an artificial substratum in coastal waters. Biofouling. 2011c; 27:357-366.
- 42. Xu H, Zhang W, Jiang Y, Yang EJ. Use of biofilmdwelling ciliate communities to determine environmental quality status of coastal waters. Science of the Total Environment. 2014; 470:511-518.