

Using multi-locus sequence data for addressing species boundaries in commonly accepted lichen-forming fungal species

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Abstract Accurate species delimitations are of great importance for effectively characterizing biological diversity. Our criteria for delimiting species have changed dramatically over the last decades with the increasing availability of molecular data and improvement of analytical methods to evaluate these data. Whereas reciprocal monophyly is often seen as an indicator for the presence of distinct lineages, recently diverged species often fail to form monophyletic groups. At the same time, cryptic species have repeatedly been detected in numerous organismal groups. In this study, we addressed the species delimitation in the crustose lichen-forming fungal genus *Diploschistes* using multilocus sequence data from specimens representing 16 currently accepted species. Our results indicate the presence of previously undetected, cryptic species-level lineages in the subgenus *Limborina*. In the subgenus *Limborina*, samples from different continents currently classified under the same species were shown to be only distantly related. At the same time, in parts of subgen. *Diploschistes* characterized by short branches, none of the currently accepted species formed monophyletic groups. In spite of the lack of

monophyly in phylogenetic reconstructions, a multispecies coalescent method provided support for eight of the nine accepted species in subgen. *Diploschistes* as distinct lineages. We propose to reduce *D. neutrophilus* to synonymy with *D. diacapsis* and point out that additional sampling will be necessary before accepting additional species in subgen. *Limborina*.

Keywords BPP · *Diploschistes* · Graphidaceae · Molecular phylogeny · Species delimitation

Introduction

In a broad sense, species delimitation is the process of identifying how individuals and populations fit into natural, species-level clusters, which are not simply constructs of classification (Carstens et al. 2013). Species are generally considered to represent a fundamental unit in biology and provide valuable context for organizing, evaluating, and communicating important biological concepts and principles (Coyne and Orr 2004; Mayr 1963). Therefore, accurate species circumscriptions are integral to interpreting biological patterns and processes across a wide range of sub-disciplines in biology.

Over the last decade, multi-locus sequence data have increasingly been used for assessing species (Camargo and Sites 2013), and the ongoing development of empirical approaches facilitates more objective species delimitation using molecular data (Leavitt et al. 2015b, 2016). DNA sequence data have revealed previously unrecognized species-level lineages hidden within nominal taxa in all organismal groups studied to date. Furthermore, many traditional taxonomic concepts conflict to various degrees with species-level lineages circumscribed using molecular sequence data. The bulk of species delimitation research highlights the fact that finding

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and applying the appropriate character sets and analytical tools remains one of the greatest challenges with empirical species delimitation (Lumbsch and Leavitt 2011). The phylogenetic species criterion is widely used for species delimitation research using molecular sequence data (Nixon and Wheeler 1990; Sites and Marshall 2004; Fujisawa and Barraclough 2013). Although reciprocal monophyly is intuitively appealing, largely because it provides a utilitarian approach with broad applicability, it may fail to accurately delimit species boundaries, particularly for species that recently diverged (Knowles and Carstens 2007). Hence, we argue that species delimitation using DNA sequence data is best viewed as a statistical inference problem given the stochastic nature of the coalescent and process of sequence evolution (Rannala 2015). Here, we use statistical inference under the multi-species coalescent (MSC) model (Kingman 1982; Degnan and Rosenberg 2009; Yang and Rannala 2010a) to test species boundaries.

Similar to other biological groups, molecular sequence data have been central to improving hypotheses of species boundaries in lichen-forming fungi (Crespo and Lumbsch 2010). Traditionally, differences in morphological, chemical, and ecological features have been the predominant source of diagnostic taxonomic characters for circumscribing lichen-forming fungal species (Printzen 2010). However, lichenized fungi generally display few taxonomically useful characters, and varying levels of intraspecific variation among different species groups may confound accurate taxonomic circumscriptions. Therefore, molecular genetic data play an increasingly prominent role in delimiting fungal species and understanding evolutionary relationships in lichenized fungi.

The lichen-forming genus *Diploschistes* Norman (Lecanoromycetes: Ostropales: Graphidaceae) (Lumbsch and Huhndorf 2010) currently includes about 30 crustose species (Jaklitsch et al. 2016), which grow on rocks, soil, or over mosses and other lichens (Fig. 1). Traditionally, *Diploschistes* has been characterized by having a carbonized proper excipulum with lateral paraphyses, *Trebouxia* Puymary as its photobiont, and by the absence of a columella (Lumbsch 1989). Morphologically, *Diploschistes* was regarded as consisting of three main groups: the *D. actinostomus* group with perithecioid ascomata, the *D. scrupopus* group with urceolate ascomata, and the *D. ocellatus* group with lecanoroid ascomata. These informal groups were supported as monophyletic clades in phylogenetic studies using phenotypical (Lumbsch and Tehler 1998) or molecular data (Fernández-Brime et al. 2013; Martín et al. 2003).

The monophyly of the genus *Diploschistes* has been discussed in the literature. Morphologically, two species were unique in the genus: *D. bisporus* (Bagl.) J. Steiner and *D. ocellatus* (Vill.) Norman, which both lack lateral paraphyses (Lumbsch 1989). Morphological studies already revealed that the former species is distinct in numerous characters and

consequently it was separated at the generic level as *Ingvariella bispora* (Bagl.) Guderley & Lumbsch (Guderley et al. 1997). Recent studies showed that this genus is not only unrelated to other *Diploschistes* but belongs to a different family in Ostropales (Fernandez-Brime et al. 2011). *Diploschistes ocellatus* was also found to be distinct from other *Diploschistes* species with some studies showing the genus being non-monophyletic (Parnmen et al. 2013; Rivas Plata et al. 2013; Kraichak et al. 2014). Consequently, *D. ocellatus* has been segregated as *Xalocoa ocellata* (Kraichak et al. 2014). Currently, *Diploschistes* includes species placed in the *actinostomus* group, which is now recognized as subgenus *Limborina* Fdez.-Brime, Gaya & Llimona, and the *scrupopus* group or subgenus *Diploschistes* (Fernández-Brime et al. 2013).

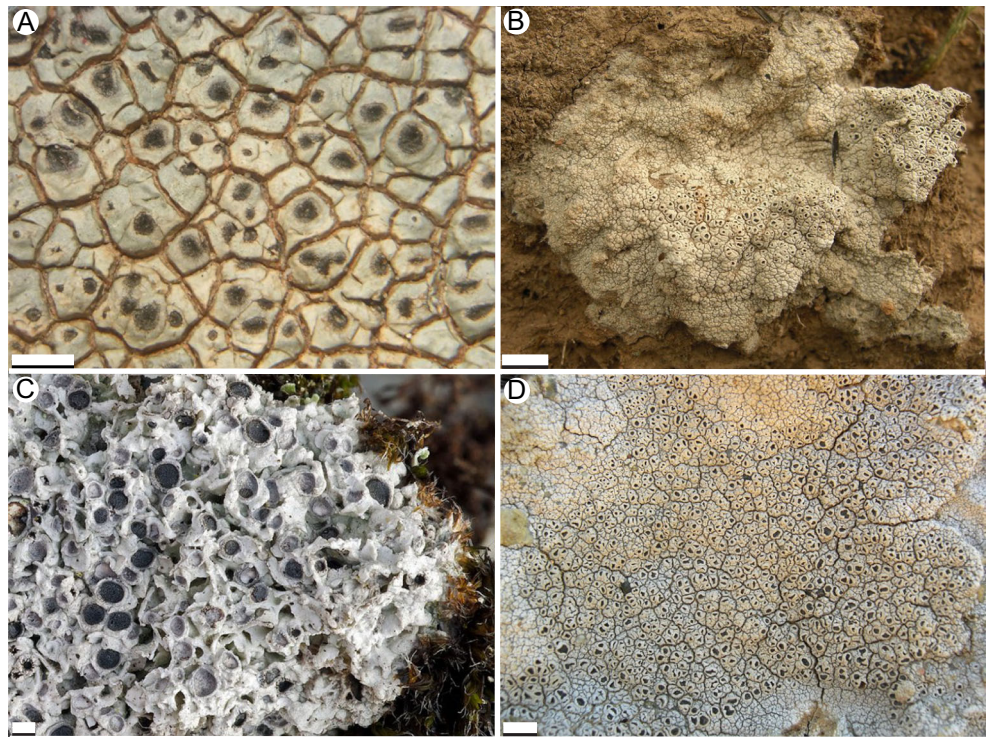
While recent molecular studies yielded in a better understanding of the circumscription and phylogenetic placement of *Diploschistes* and the phylogeny of major clades within the genus, the delimitation of species is still unresolved. Previous studies (Fernández-Brime et al. 2013; Martín et al. 2003) indicated that the circumscription of some species needs re-examination and the species delimitation based on phenotypic characters differs between authors (Clauzade and Roux 1985; Llimona Pages 1974; Lumbsch 1989, 1988; Pant and Upreti 1993). Hence, we assembled a dataset consisting of six loci (two nuclear ribosomal, one mitochondrial ribosomal, and three nuclear protein-coding genes) to address the species delimitations within *Diploschistes* and test morphology-based hypotheses.

Methods and materials

Taxon sampling

Our sampling of *Diploschistes* species included a total of 93 specimens representing 16 currently recognized species (Table 1). Species of the subgenus *Diploschistes* included the following: *D. cinereocaesius*, *D. diacapsis*, *D. gypsaceus*, *D. interpediens*, *D. muscorum*, *D. neutrophilus*, *D. rampoddensis*, *D. scrupopus*, and *D. thunbergianus*. Taxa included in this study that belong to subgenus *Limborina* included the following: *D. actinostomus*, *D. caesioplumbeus*, *D. candidissimus*, *D. diploschistoides*, *D. elixii*, *D. euganeus*, and *D. sticticus*. Based on previous studies, the tree was rooted with subgenus *Limborina* (Martín et al. 2003; Fernández-Brime et al. 2013). We attempted to sample specimens across the range of each species' distribution, and overall specimens from Africa, Australia, Central America, Europe, North America, and South America were selected. Additional specimens were selected from GenBank to improve our taxonomic sampling.

Fig. 1 Habit photographs of *Diploschistes* species. **a** *D. actinostomus*. **b** *D. diacapsis*. **c** *D. muscorum*. **d** *D. scruposus*. Photographs kindly provided by Jason Hollinger and Richard Doker, and the “Ways of enrichment” (“www.waysofenrichment.net/#”)



Molecular data

Sample preparation, DNA isolation, PCR, and direct sequencing were performed as described previously (Fernández-Brime et al. 2013; Leavitt et al. 2012). Molecular data were generated for six loci: the internal transcribed spacer (ITS), nuclear large subunit (nuLSU), mitochondrial small subunit (mtSSU), minichromosome maintenance complex component 7 (MCM7), the largest subunit of the RNA polymerase II gene (RPB1), and the second largest subunit of RNA polymerase II gene (RPB2). Primers and PCR cycling parameters used for amplifying the six loci are listed in Table 2.

Sequence alignments

New sequences were assembled and edited using the program Sequencher v4.10 (Gene Code Corporation, Ann Arbor, MI) and were subjected to BLAST searches for a first verification of their identities. Sequences of each locus were aligned using the program MAFFT v7 (Katoh et al. 2009) with settings appropriate for the variability of each locus. For ITS sequences, we used the L-ING-i alignment algorithm with the remaining parameters set to default values. For nuLSU, G-ING-i algorithm and “leave gappy regions” were selected. Then, we used E-ING-i algorithm for mtSSU and RPB1, and G-ING-i algorithm for MCM7 and RPB2, with the remaining parameters set to default values. The alignments were adjusted manually to exclude missing data and concatenated.

Ambiguous positions of the ITS and mtSSU alignments were removed using the Gblocks web server (Castresana 2000), implementing all the options for a less stringent selection.

Phylogenetic analysis

Exploratory phylogenetic analyses of individual loci revealed a general pattern of poorly resolved topologies. Therefore, the six single-locus alignments were concatenated in Geneious v6.1.2 (Biomatters Ltd., Auckland, NZ) for subsequent phylogenetic analyses. Only specimens that were represented by at least two of six targeted loci were included in the concatenated data matrix (Table 1). A maximum likelihood (ML) analysis was carried out on the multilocus matrix using the locus-specific model partitions (ITS, nuLSU, mtSSU, MCM7, RPB1, and RPB2) in RAxML v8.1.24 (Stamatakis 2006). A search combining 200 separate ML searches was conducted, implementing a GTRGAMMA model, and 1000 pseudoreplicates to evaluate bootstrap support for each node. In addition to the ML analysis, a Bayesian analysis with MrBayes v3.2.3 (Ronquist et al. 2012) was also used for phylogenetic inference from our multilocus dataset. The most appropriate nucleotide substitution model for each of the six loci was selected using the Akaike information criterion in jModelTest v2.1.7 (Darriba et al. 2012). The Bayesian analysis was run for 10,000,000 generations with four independent chains and sampling every 1000th tree. All model parameters were unlinked. Two independent Bayesian runs were conducted to ensure that stationarity was reached and the runs converged at the

Table 1 Information of the species and sequences used in this paper

Species	Isolation; herbarium number	Locality	ITS	nuLSU	mtSSU	RPB1	RPB2	MCM7
<i>Diploschistes actinostomus</i>	Lumbsch3089 (F)	Kenya	KF688486	KF688496	KF688510	KF688530		
<i>Diploschistes actinostomus</i>	SFB004; 0016461 (DUKE)	USA, North Carolina	KC166972		KC167025			
<i>Diploschistes caesioplumbus</i>	SFB028; 19325 (BCN-LICH)	Spain, Girona	KC166973		KC167026		KX545647	
<i>Diploschistes caesioplumbus</i>	SFB030; 19323 (BCN-LICH)	Spain, Girona	KC166974		KC167027		KX545634	
<i>Diploschistes candidissimus</i>	SFB008; 0144447 (DUKE)	USA, Delaware	KC166976		KC167028		KX545633	KX545555
<i>Diploschistes candidissimus</i>	SFB086; 19340 (BCN-LICH)	Spain, Lleida	KC166977		KC167029	KX545630		
<i>Diploschistes cinereocaesius</i>	AFTOL-ID290	Ecuador	KJ542542	KX545505	KX545587	KJ766850		
<i>Diploschistes cinereocaesius</i>	Luecking 15540 (F)	Costa Rica	KF688481	KF688491	JX421026	KF688518		
<i>Diploschistes cinereocaesius</i>	Lumbsch 19303 (F)	Peru	KF688479	KF688490	KF688502	KF688516		
<i>Diploschistes cinereocaesius</i>	AFTOL-ID328	Costa Rica, San Jose	HQ650715	DQ883799	DQ912306	DQ883742	DQ883755	
<i>Diploschistes diacapsis</i>	AFTOL-ID4860			KJ766550	KJ766384			
<i>Diploschistes diacapsis</i>	SFB001; 0030912 (DUKE)	Spain, Zaragoza	KC166978		KC167030			
<i>Diploschistes diacapsis</i>	SFB052 (BCN)	Spain, Zaragoza	KX545483				KX545643	KX545538
<i>Diploschistes diacapsis</i>	SFB077(BCN)	Spain, La Rioja	KX545484		KX545585	KX545612	KX545675	KX545516
<i>Diploschistes diacapsis</i>	SFB002; 0130126 (DUKE)	USA, Arizona	KC166979		KC167031			
<i>Diploschistes diacapsis</i>	SFB133; 19347 (BCN-LICH)	Spain, Lleida	KC166980		KC167032			
<i>Diploschistes diacapsis</i>	ALM58	Spain, Almeria	KX545469	KX545512	KX545559	KX545614	KX545654	KX545549
<i>Diploschistes diacapsis</i>	ALM59	Spain, Almeria	KX545503	KX545511	KX545561	KX545625	KX545639	KX545552
<i>Diploschistes diploschistoides</i>	Lumbsch 19073b (F)	Australia, Queensland		AY605076	KF688500	KF688513		
<i>Diploschistes diploschistoides</i>	SFB006; 0144445 (DUKE)	Australia, Western Australia	KC166984		KC167036		KX545636	
<i>Diploschistes diploschistoides</i>	SFB010	Australia, Queensland	KC166985		KC167037		KX545635	
<i>Diploschistes elixii</i>	Elix 32450 (F)	Australia, Western Australia	KF688482	EU126644	KF688504	KF688520		
<i>Diploschistes euganeus</i>	Lumbsch 20603 (F)	Switzerland, Ticino			KF688507	KF688527		
<i>Diploschistes euganeus</i>	Lumbsch 20605g (F)	Switzerland, Ticino	KF688485	KF688494	KF688508	KF688528		
<i>Diploschistes euganeus</i>	SFB029; 0144451 (DUKE)	Australia, Western Australia	KC166986		KC167038		KX545632	
<i>Diploschistes gypsaceus</i>	SFB088; 19324 (BCN-Lich)	Spain, Lleida	KC166988		KC167039			
<i>Diploschistes gypsaceus</i>	SFB020; 17180 (BCN-Lich)	Spain, Lleida	KC166987		KX545557	KX545596	KX545644	KX545532
<i>Diploschistes gypsaceus</i>	SFB139; 19340 (BCN-Lich)	Spain, Lleida	KC166991		KC167042			
<i>Diploschistes gypsaceus</i>	SFB127; 19345 (BCN-Lich)	Spain, Lleida	KC166989		KC167040			
<i>Diploschistes gypsaceus</i>	SFB131; 19346 (BCN-Lich)	Spain, Lleida	KC166990		KC167041			
<i>Diploschistes interpediens</i>	Lumbsch 20605d (F)	Switzerland, Ticino		KC167075	KF688509	KF688529		
<i>Diploschistes interpediens</i>	SFB057 (BCN)	Spain, La Rioja		KF688495	KX545563	KX545592	KX545650	
<i>Diploschistes interpediens</i>	SFB064 (BCN)	Spain, La Rioja	KX545488				KX545679	
<i>Diploschistes interpediens</i>	SFB065; 14751 (BCN-Lich)	Spain, La Rioja	KX545475			KX545593	KX545659	KX545523
<i>Diploschistes interpediens</i>	SFB021; 19317 (BCN-Lich)	Portugal, Braganca	KX545473			KX545591	KX545651	KX545522

Table 1 (continued)

Species	Isolation; herbarium number	Locality	ITS	nuLSU	mtSSU	RPB1	RPB2	MCM7
<i>Diploschistes interpediens</i>	SFB026; 18007 (BCN-Lich)	Spain, Santiago de Compostela	KX545485		KC167047	KX545619	KX545678	KX545553
<i>Diploschistes interpediens</i>	SFB056 (BCN)	Portugal, Braganca	KX545487		KX545580	KX545620	KX545676	KX545548
<i>Diploschistes interpediens</i>	SFB025; 19319 (BCN-Lich)	Spain, Girona	KX545480	KC167076	KC167046	KX545621	KX545660	KX545546
<i>Diploschistes interpediens</i>	SFB035; 19350 (BCN-Lich)	Spain, Barcelona, Dosrius	KX545479		KC167048	KX545607	KX545662	KX545547
<i>Diploschistes interpediens</i>	SFB036 (BCN)	Spain, Barcelona	KX545477		KX545588	KX545609	KX545661	KX545547
<i>Diploschistes interpediens</i>	SFB082(BCN)	Spain, Cordoba	KX545478		KX545565	KX545608		KX545525
<i>Diploschistes interpediens</i>	SFB078 (BCN)	Spain, La Rioja	KX545476		KX545582	KX545590	KX545658	KX545520
<i>Diploschistes muscorum</i>	9038 (BCN)	Spain			KX545584	KX545631	KX545666	KX545529
<i>Diploschistes muscorum</i>	DMUS140 (BCN)	Spain	KX545481		KX545570	KX545617	KX545657	
<i>Diploschistes muscorum</i>	DMUS4500B (BCN)	Spain	KX545482		KX545570			
<i>Diploschistes muscorum</i>	SFB125; 19344 (BCN-Lich)	Spain, Lleida	KC167007		KC167058			
<i>Diploschistes muscorum</i>	SFB140; 19334 (BCN-Lich)	Spain, Barcelona	KC167008		KC167059			
<i>Diploschistes muscorum</i>	SFB045; 19333 (BCN-Lich)	Italy, Sardinia	KC167005		KC167056			
<i>Diploschistes muscorum</i>	SFB079; 14435 (BCN-Lich)	Spain, La Rioja	KC167006		KC167057	KX545624	KX545652	KX545528
<i>Diploschistes muscorum</i>	SFB013 (BCN)	USA, Arizona	KX545472		KX545578	KX545618	KX545656	KX545530
<i>Diploschistes muscorum</i>	SFB003; 0016462 (DUKE)	USA, Connecticut	KC167004	KC167077	KC167055			
<i>Diploschistes neutrophilus</i>	ALM55 (BCN)	Spain, Almeria	KX545471		KX545579	KX545615	KX545665	KX545550
<i>Diploschistes neutrophilus</i>	ALM56 (BCN)	Spain, Almeria	KX545504		KX545560	KX545628	KX545642	KX545539
<i>Diploschistes neutrophilus</i>	ALM57 (BCN)	Spain, Almeria	KX545470	KX545506		KX545616	KX545653	KX545551
<i>Diploschistes neutrophilus</i>	SFB118; 19329 (BCN-Lich)	Spain, Girona	KC166983		KC167035			
<i>Diploschistes neutrophilus</i>	SFB063; 19357 (BCN-Lich)	Spain, Girona	KC166982		KC167034	KX545627	KX545641	KX545541
<i>Diploschistes neutrophilus</i>	SFB044; 19338 (BCN-Lich)	Spain, Illes Balears	KC166981		KC167033	KX545626	KX545638	KX545540
<i>Diploschistes neutrophilus</i>	SFB146 (BCN)	Spain, Illes Balears			KX545566	KX545629	KX545640	KX545514
<i>Diploschistes rampoddensis</i>	SFB101; 18011 (BCN-Lich)	Spain, Girona	KC166993		KC167044			
<i>Diploschistes rampoddensis</i>	SFB068; 18009 (BCN-Lich)	Spain, Girona	KC166992		KC167043	KX545622	KX545637	KX545521
<i>Diploschistes rampoddensis</i>	SFB073; 18008 (BCN-Lich)	Spain, Girona	KJ542543			KX545623		KX545556
<i>Diploschistes scruposus</i>	9042 (BCN)	Spain					KX545670	KX545519
<i>Diploschistes scruposus</i>	9044 (BCN)	Spain	KX545499		KX545569		KX545673	
<i>Diploschistes scruposus</i>	9045 (BCN)	Spain			KX545572		KX545685	
<i>Diploschistes scruposus</i>	Schmitt1376 (F)	Germany, Hesse	KF688478	KF688489	KF688501	KF688515		
<i>Diploschistes scruposus</i>	Schmitt 1375 (F)	Germany, Hesse	KF688477	KF688488	KF688501	KF688514		
<i>Diploschistes scruposus</i>	9040 (BCN)	Spain	KX545491		KX545571		KX545672	KX545518
<i>Diploschistes scruposus</i>	AFTOL-ID4813	USA		KJ766552	KJ766385			
<i>Diploschistes scruposus</i>	SFB014 (BCN)	Canada, Ontario	KX545492				KX545674	KX545537
<i>Diploschistes scruposus</i>	SFB058; 14227 (BCN-Lich)	Spain, La Rioja	KC167016		KC167066	KX545611	KX545684	

Table 1 (continued)

Species	Isolation; herbarium number	Locality	ITS	nuLSU	mtSSU	RPB1	RPB2	MCM7
<i>Diploschistes seruposus</i>	SFB066; 14398 (BCN-Lich)	Spain, La Rioja	KC167020		KC167070	KX545594	KX545664	KX545527
<i>Diploschistes seruposus</i>	SFB076 (BCN)	Spain, La Rioja	KX545474		KX545562	KX545589	KX545649	KX545524
<i>Diploschistes seruposus</i>	SFB080 (BCN)	Spain, La Rioja	KX545486				KX545677	
<i>Diploschistes seruposus</i>	SFB059 (BCN)	Spain, Girona	KJ542545		KX545586	KX545597	KX545648	KX545533
<i>Diploschistes seruposus</i>	SFB060; 19326 (BCN-Lich)	Spain, Girona	KC167017		KC167067	KX545606	KX545646	KX545535
<i>Diploschistes seruposus</i>	SFB024; 19328 (BCN-Lich)	Spain, Lleida	KC167014	KC167078	KC167064	KX545610	KX545680	KX545536
<i>Diploschistes seruposus</i>	SFB034 (BCN)	Spain, Barcelona	KX545501		KX545567	KX545602	KX545668	KX545543
<i>Diploschistes seruposus</i>	SFB037; 19351 (BCN-Lich)	Spain, Barcelona	KC167015		KC167065	KX545604	KX545669	KX545544
<i>Diploschistes seruposus</i>	SFB039 (BCN)	Spain, Barcelona	KX545502		KX545564	KX545603	KX545667	KX545545
<i>Diploschistes seruposus</i>	SFB069; 19354 (BCN-Lich)	Spain, Barcelona	KC167021		KC167071	KX545595	KX545663	KX545526
<i>Diploschistes seruposus</i>	SFB147 (BCN)	Sweden, Norrbotten	KX545493	KX545508	KX545573			
<i>Diploschistes seruposus</i>	SFB148 (BCN)	Sweden, Norrbotten	KX545494		KX545583			
<i>Diploschistes seruposus</i>	SFB149 (BCN)	Sweden, Norrbotten	KX545495		KX545574			
<i>Diploschistes seruposus</i>	SFB150 (BCN)	Sweden, Norrbotten	KX545500	KX545510	KX545575	KX545598	KX545671	KX545517
<i>Diploschistes seruposus</i>	SFB151 (BCN)	Sweden, Norrbotten	KX545496	KX545509	KX545576	KX545599	KX545483	KX545554
<i>Diploschistes seruposus</i>	SFB155 (BCN)	Norway, Finnmark	KX545498	KX545513	KX545577	KX545600	KX545682	KX545542
<i>Diploschistes seruposus</i>	SFB157 (BCN)	Norway, Finnmark	KX545497	KX545507	KX545581	KX545601	KX545681	KX545515
<i>Diploschistes sticticus</i>	Lumbsch 19109b (F)	Australia, Queensland		JX421482	KF688499	KF688512		
<i>Diploschistes sticticus</i>	Lumbsch 3056 (F)	Kenya			KF688511	KF688531		
<i>Diploschistes thunbergianus</i>	Eldridge 3800 (F)	Australia, New South Wales	AI458289	AF274095	AF431955			
<i>Diploschistes thunbergianus</i>	Lumbsch 10728d (F)	Australia, South Australia	AI458290					
<i>Diploschistes</i> sp.	DMUS159 (BCN)	Spain	KX545490		KX545558	KX545613	KX545645	KX545534
<i>Diploschistes</i> sp.	DSPY138 (BCN)	Spain	KX545489		KX545568	KX545605	KX545655	KX545531

Newly generated sequences are in bold

same log-likelihood level (Nylander et al. 2008). After discarding the burn-in, the remaining 7500 trees of each run were pooled to calculate a 50% majority rule consensus tree. Clades that received bootstrap support $\geq 70\%$ under ML and posterior probabilities ≥ 0.95 were considered significant. Phylogenetic trees were visualized using FigTree v1.4.2 (Rambaut 2009).

Species delimitation analysis

For a subgroup of species in the *D. scruposus* group, the multispecies coalescent model implemented in the program BPP v3.2 (Yang and Rannala 2010b, 2014; Rannala and Yang 2013) was used to infer support for the separation of the sampled *Diploschistes* species. The *D. scruposus* group has recently been shown to have a recent diversification history (Rivas Plata 2011; Kraichak et al. 2015), and recently diverged species may not be recovered as monophyletic due to incomplete lineage sorting (Knowles and Carstens 2007). Given the recent diversification history for the *D. scruposus* group, lack of resolution and short branches in phylogenetic

reconstructions for this group (see Results), and support from phenotypic and ecological evidence (Lumbsch and Tehler 1998), it may be reasonable to assume that traditionally circumscribed species in the *D. scruposus* group represent distinct evolutionary lineages, in spite of their lack of monophyly in phylogenetic reconstructions. Therefore, for the BPP analyses, which accounts for incomplete lineage sorting within a multispecies coalescent framework, specimens within the *D. scruposus* group were assigned to candidate species based on phenotype-based identifications.

BPP incorporates coalescent theory and phylogenetic uncertainty into parameter estimation, and the posterior distribution for species delimitation models is sampled using a reversible-jump Markov Chain Monte Carlo (rjMCMC) chain.

We used the unguided species delimitation analysis “A11” (Yang 2015), which explores different species delimitation models and different species phylogenies, with fixed specimen assignments to populations. Specimens were assigned to nine currently accepted species: *D. cinereocaesius*, *D. diacapsis*, *D. gypsaceus*, *D. interpediens*, *D. muscorum*,

Table 2 Primer information and PCR settings used for this paper

Loci/PCR info	ITS	nucLSU	mtSSU	MCM7	RPB1	RPB2
PCR primers	ITS1 ^a	AL1R ^c	mrSSU1 ^f	LecMCM7 ^h	gRPB1a ^j	RPB2-6 ^d
	ITS4a ^b	AL2R ^d	mrSSU2 ^r	LecMCM7 ^r	fRPB1c ^k	RPB2-7 ^{cr}
		LR5 ^e	mrSSU3 ^r	MCM7-709 ^f		
		LR6 ^e	MSU1 ^g	MCM7-1348 ^r		
			MSU7 ^g			
Initial denaturation	95 °C 5 min	95 °C 5 min	94 °C 10 min	94 °C 10 min	94 °C 10 min	94 °C 10 min
Phase 1	10 cycles	10 cycles	34 cycles	34 cycles	34 cycles	34 cycles
	95 °C 30s	95 °C 30s	95 °C 45 s	94 °C 45 s	94 °C 45 s	94 °C 45 s
	66 °C 30s	66 °C 30s	50 °C 45 s	50 °C 50s	50 °C 50s	50 °C 50s
	72 °C 1 min 30s	72 °C 1 min 30s	72 °C 1 min 30s	72 °C 1 min	72 °C 1 min	72 °C 1 min
Phase 2	34 cycles	34 cycles	None	None	None	None
	95 °C 30s	95 °C 30 sec				
	56 °C 30 sec	56 °C 30 sec				
	72 °C 1 min 30 sec	72 °C 1 min 30 sec				
Final extension	72 °C 10 min	72 °C 10 min	72 °C 10 min	72 °C 5 min	72 °C 5 min	72 °C 5 min

^a Gardes and Bruns (1993)

^b Larena et al. (1999)

^c Döring et al. (2000)

^d Mangold et al. (2008)

^e Vilgalys and Hester (1990)

^f Zoller et al. (1999)

^g Zhou and Stanosz (2001)

^h Leavitt et al. (2011)

ⁱ Schmitt et al. (2009)

^j Stiller and Hall (1997)

^k Matheny et al. (2002)

^l Liu et al. (1999)

D. neutrophilus, *D. rampoddensis*, *D. scruposus*, and *D. thunbergianus*. Using analysis A11, the algorithm attempts to merge populations into one species and uses the nearest neighbor interchange (NNI) or subtree pruning and regrafting (SPR) algorithms to change the species tree topology (Yang and Rannala 2014). Analysis “A00” (Yang 2015), a within-model inference, was used to generate the posterior distribution of the parameters theta (θ s) and tau (τ s) under the multi-species coalescent (MSC) model to infer a reasonable combination of priors given the data (Rannala 2015). Based on the results from the A00 analyses, the gamma prior G for θ was set to $\sim G(1, 85)$, and the gamma prior G for tau (τ) was set to $\sim G(1, 200)$. Under the unguided species delimitation model, A11, we used two different search algorithms (algorithm 0 or 1), equal probabilities for the labeled histories, to assign probabilities to the models, rates were allowed to vary among loci (locus rate = 1), and the analyses were set for automatic fine-tune adjustments. The rjMCMC analysis was run for 100,000 generations, sampling every 2 generations discarding the first 10% as burn-in. The analysis was run twice to confirm consistency between runs.

Results

Molecular data

For this study, 217 new sequences were generated (Table 1). The multilocus matrix we used in this study was deposited in TreeBase (ID# pending). The concatenated, six-locus matrix consisted of 93 individuals and 5074 aligned nucleotide position characters (Table 3). A summary of alignment information for the multilocus dataset was also provided in Table 3.

Phylogenetic analysis

Phylogenies derived from the ML and B/MCMC analyses were generally concordant. Minor differences in the arrangement of some terminals occurred, but relationships at deeper nodes and in well-supported clades were identical. We chose to present the ML topology, with nodal support values from

both ML bootstrap analysis and posterior probabilities from the Bayesian inference (Fig. 2).

The two subgenera of *Diploschistes* were recovered in our phylogenetic trees with strong support (both are BS = 98, PP = 1.0; Fig. 2). In our ML tree, subgen. *Diploschistes* was shown to include several clades. *Diploschistes cinereocaesius* (four specimens) was strongly supported as a monophyletic group forming a strongly supported sister-group (BS = 100, PP = 1.0) with the monophyletic *D. rampoddensis* (three specimens). These two species formed an unsupported sister-group to the monophyletic *D. thunbergianus* (two specimens). Seven specimens of *D. muscorum* formed a strongly supported (BS = 91, PP = 1.0) group, which formed an unsupported relationship with an unsupported clade including five specimens of *D. diacapsis* and seven specimens of *D. neutrophilus*. The bulk of specimens of subgen. *Diploschistes* clustered in an unsupported clade containing five species—*D. diacapsis* (three specimens), *D. gypsaceus* (five specimens), *D. interpediens* (12 specimens), *D. muscorum* (two specimens), and *D. scruposus* (26 specimens), and two samples which could not be identified with certainty, with seven supported internodes. Within subgen. *Limborina*, most relationships were also unresolved, and only two internodes were supported. In this clade, only *D. diploschistoides* (BS = 75, PP = 1.0) was recovered as monophyletic with strong support, whereas all other species were either not monophyletic or their monophyly was not strongly supported. This includes *D. actinostomus* with the samples from North America and Africa being separated, *D. euganeus* with samples from Australia and Europe not clustering together, and *D. sticticus* with samples from Australia and Africa not forming a monophyletic group and *D. caesioplumbeus* forming an unsupported clade.

Branch lengths between clades in subgen. *Limborina* and *D. cinereocaesius*, *D. rampoddensis*, and *D. thunbergianus* differed considerably from those in the other parts of subgen. *Diploschistes*. Branch lengths were generally short, and support for clades was overall low in the latter. Further, species in this part of the phylogenetic tree did not form monophyletic groups. However, their monophyly could not be rejected using alternative topology tests (data not shown). Hence, we employed multispecies coalescent species delimitation using

Table 3 The alignment information for the multilocus dataset

Alignments	ITS	nucLSU	mtSSU	RPB1	RPB2	MCM7	Total
Number of sequences	82	27	84	57	56	43	349
Newly generated sequences	36	9	33	43	54	43	218
Number of sites (including gaps)	629	1404	820	837	768	616	5074
Missing sequences/ the percentages	11/12%	66/71%	9/10%	36/39%	37/40%	50/54%	209/37%
Nucleotide substitution models	GTR + I + G	HKY + G	K80 + G	HKY + I + G	SYM + G	K80 + G	

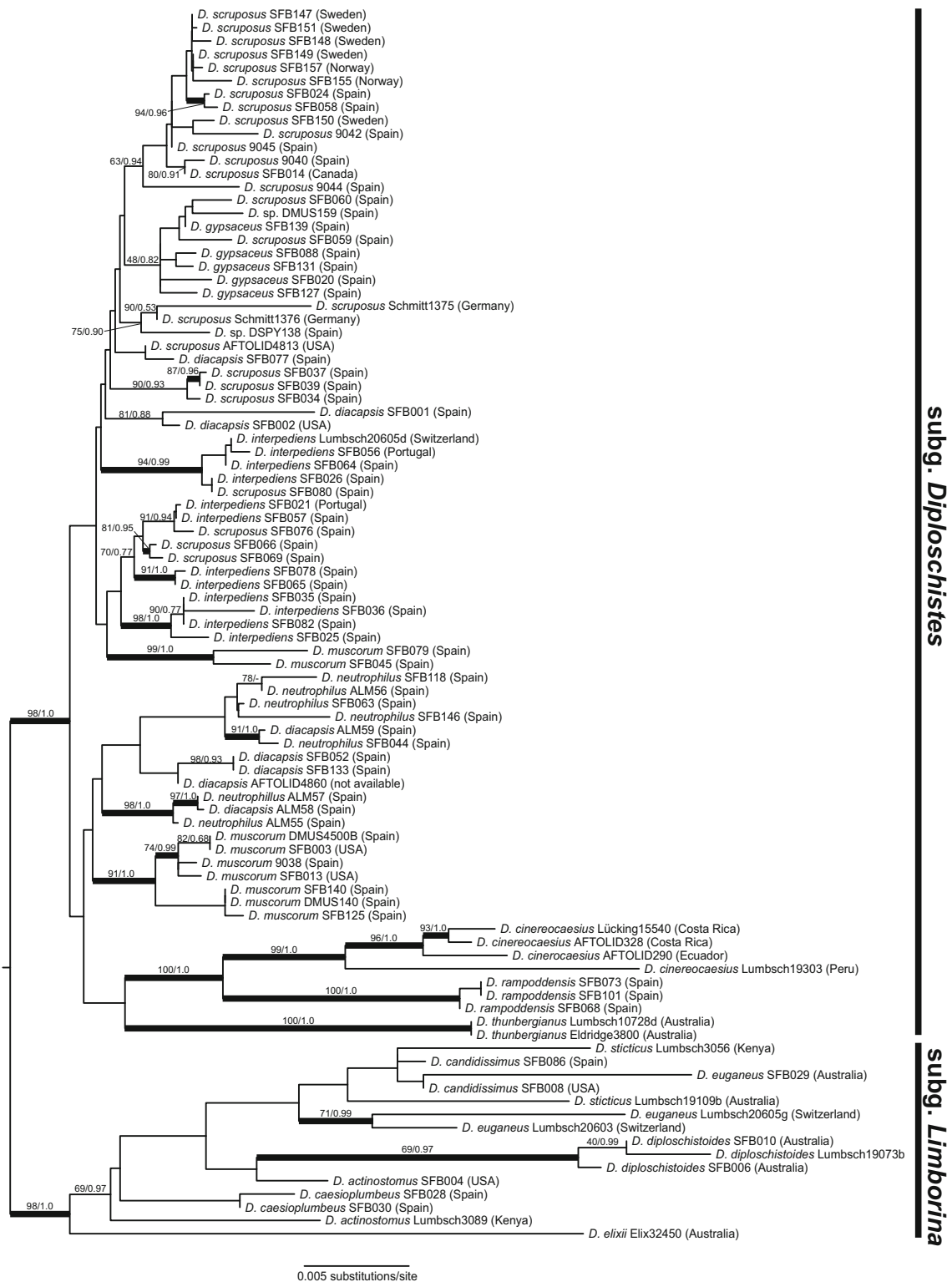


Fig. 2 Maximum likelihood (ML) phylogenetic relationships of *Diploschistes* taxa inferred from a combined 6-locus analysis. Values at each node indicate nonparametric bootstrap support (BS)/posterior

probability (PP), branches in bold received maximum likelihood bootstrap support values equal or above 70 and posterior probabilities equal or above 0.95

BPP to evaluate separation of currently accepted species in subgenus *Diploschistes*. An 8-species delimitation scenario had the highest probability, followed by a 7-species scenario

(Table 4). All other species delimitation models had probabilities <0.05. Currently accepted species in subgenus *Diploschistes* received the highest supported, with the

Table 4 Species delimitation scenarios and their posterior probabilities inferred under the multispecies coalescent model using the program BPP

Posterior probability	# of inferred species	Taxa supported in each scenario.
0.570	8	(<i>D. cinereoaeisus</i> ; <u><i>D. diacapsis</i></u> = <u><i>D. neutrophilus</i></u> ; <i>D. gypsaceus</i> ; <i>D. interpediens</i> ; <i>D. muscorum</i> ; <i>D. rampoddensis</i> ; <i>D. scruposus</i> ; <i>D. thunbergianus</i>)
0.367	7	(<i>D. cinereoaeisus</i> ; <u><i>D. diacapsis</i></u> = <u><i>D. neutrophilus</i></u> ; <u><i>D. gypsaceus</i></u> = <u><i>D. scruposus</i></u> ; <i>D. interpediens</i> ; <i>D. muscorum</i> ; <i>D. rampoddensis</i> ; <i>D. thunbergianus</i>)
0.041	7	(<i>D. cinereoaeisus</i> ; <u><i>D. diacapsis</i></u> = <u><i>D. neutrophilus</i></u> ; <i>D. gypsaceus</i> = <i>D. interpediens</i> ; <i>D. muscorum</i> ; <i>D. rampoddensis</i> ; <i>D. scruposus</i> ; <i>D. thunbergianus</i>)
0.011	9	(<i>D. cinereoaeisus</i> ; <i>D. diacapsis</i> ; <i>D. gypsaceus</i> ; <i>D. interpediens</i> ; <i>D. muscorum</i> ; <i>D. neutrophilus</i> ; <i>D. rampoddensis</i> ; <i>D. scruposus</i> ; <i>D. thunbergianus</i>)

Cases where multiple taxa were collapsed into a single species are underlined. Only species delimitation models with posterior probabilities >0.01 are reported

exception of *D. diacapsis* and *D. neutrophilus*, which were collapsed into a single species with high probability (Table 5).

Discussion

We used a six-locus dataset including three ribosomal (ITS, nuLSU, mtSSU) and three protein-coding markers (MCM7, RPB1, RPB2) of 93 specimens representing 16 currently accepted species to test the species delimitation in the genus *Diploschistes*. Our results indicate both the presence of previously undetected, cryptic species, and difficulties in separating species using molecular markers. Species in this genus have largely been separated based on ascomatal characters, such as

apothecial morphology, exciple thickness, number of ascospores per ascus, ascospore-size, -form and -amyloidity, secondary metabolites, and thallus morphology (Lumbsch 1989; Lumbsch and Elix 1989; Rivas Plata et al. 2010). Whereas species delimitation based on these phenotypical characters have largely been in agreement among authors, variability of number of ascospores, thallus morphology, and ecology of species of the subgenus *Diploschistes* have differed somewhat among authors (Clauzade and Roux 1985; Llimona Pages 1974; Lumbsch 1988, 1989; Pant and Upreti 1993).

Among species of the subgenus *Limborina*, samples from different continents currently classified under the same species were often only distantly related, including *D. actinostomus*, *D. euganeus*, and *D. sticticus*. These results suggest that phenotypically similar specimens occurring on different continents in fact represent distinct lineages. Our taxon sampling is insufficient to address species delimitation in subgen. *Limborina*, but these results demonstrate that additional studies are necessary to better understand species delimitation in this subgenus. However, the presence of distinct lineages within nominal species on different continents has repeatedly been shown in other groups of lichen-forming fungi (Amo de Paz et al. 2012; Arguello et al. 2007; Divakar et al. 2010; Hodkinson and Lendemer 2011; Thell et al. 2009; Otálora et al. 2010; Parmen et al. 2012; Alors et al. 2016; Leavitt et al. 2015a; Zhao et al. 2015; Cornejo and Scheidegger 2015) and further alternative topology tests rejected monophyly of those species significantly (data not shown). A study addressing the species delimitation in this subgenus will require additional sampling.

In subgen. *Diploschistes*, only three species were monophyletic and strongly supported. In contrast, *D. diacapsis*, *D. interpediens*, *D. muscorum*, *D. neutrophilus*, and *D. scruposus* were not monophyletic, and the relationships among samples of those species were inferred with short branches and mostly poorly supported. Monophyly is not a prerequisite of a species, and thus, a lack thereof is not necessarily evidence that these lineages are conspecific (Leavitt et al. 2016). For example, in North American species of the genus

Table 5 Delimited species and their posterior probabilities

Probability	Delimited species
1.000	<i>D. cinereoaeisus</i>
1.000	<i>D. muscorum</i>
1.000	<i>D. rampoddensis</i>
1.000	<i>D. thunbergianus</i>
0.979	<i>D. diacapsis</i> = <i>D. neutrophilus</i>
0.958	<i>D. interpediens</i>
0.624	<i>D. scruposus</i>
0.582	<i>D. gypsaceus</i>
0.376	<i>D. gypsaceus</i> = <i>D. scruposus</i>
0.042	<i>D. gypsaceus</i> = <i>D. interpediens</i>
0.021	<i>D. diacapsis</i>
0.021	<i>D. neutrophilus</i>
<0.001	<i>D. diacapsis</i> = <i>D. gypsaceus</i> = <i>D. neutrophilus</i>
<0.001	<i>D. rampoddensis</i> = <i>D. thunbergianus</i>
<0.001	<u><i>D. cinereoaeisus</i></u> = <u><i>D. thunbergianus</i></u>

Cases where multiple taxa were collapsed into a single species are underlined

Xanthoparmelia, recent diversification during Pliocene and Pleistocene was estimated, and independent species-level lineages were not supported by concordant evolutionary histories across multiple, independent loci (Leavitt et al. 2011a, b, 2013).

In spite of our attempt to reconstruct phylogenetic relationships for the *D. scruposus* group using multilocus sequence data, species boundaries and relationships within this group remained unresolved. Therefore, we used the program BPP to delimit species boundaries in this group within a statistical framework modeled under the multispecies coalescent. This analysis strongly supported eight of the nine accepted species in subgen. *Diploschistes* as separate species. Previously, it has been shown that the genus *Diploschistes*, and especially subgen. *Diploschistes*, has diversified recently (Rivas Plata 2011; Kraichak et al. 2015). We hypothesize that the difficulties in separating species in some species of subgen. *Diploschistes* are due to recent diversification. The multispecies coalescent analysis, which accounts for incomplete lineage shorting, supports that most of them are in fact distinct species. The only exception is *D. neutrophilus*, which was supported to belong to *D. diacapsis*. Consequently, we propose to reduce *D. neutrophilus* to synonymy with *D. diacapsis* below.

The coalescent-based BPP program accounts for ancestral polymorphisms and incomplete lineage sorting. However, also factors, such as occasional gene flow, hybridization, and recombination, are other evolutionary factors potentially influencing species delimitation inferences. While BPP performs quite robustly under a range of scenarios, speciation probabilities decrease with increasing levels of gene flow (Camargo et al. 2012; Zhang et al. 2011).

BPP also requires a priori assignment of individuals to candidate species, and the impact of incorrectly assigned specimens remains unclear. While traditionally accepted nominal species have been well-studied morphologically (Lumbsch 1989; Lumbsch and Elix 1989; Rivas Plata et al. 2010), our multilocus phylogenetic reconstructions failed to provide strong support either for or against the traditionally, phenotype-based species in the *D. scruposus* group. Therefore, we based our specimen assignments to candidate species for the BPP analysis on phenotype-based identifications. Arguably, as the use of genome-wide molecular data becomes more commonplace in lichen research, species boundaries and evolutionary relationships in lineages with recent diversification histories, including the *D. scruposus* group, will better understood. In the meantime, we propose that the 8-species model inferred for the *D. scruposus* group (Table 4) represents a useful working hypothesis of species boundaries for this group.

Taxonomic conclusions

Diploschistes diacapsis (Ach.) Lumbsch

Lichenologist 20: 20 (1988).—*Urceolaria diacapsis* Ach., *Lich. Univ.*: 339 (1810); lectotype (selected by Lumbsch, *Lichenologist* 20: 20, 1988); Spain, Lagasca (S!).

= *Diploschistes neutrophilus* (Clauzade & Cl. Roux) Fdez.-Brime & Llimona, *Taxon* 62: 275 (2013); holotype: France, Provence, Bouches-du-Rhône Crau, 10 km from Fos-sur-Mer, Clor de Tenque, on neutral clayey sandy soil, 25 Apr 1980, C. Roux (Hb. Claude Roux no. 99!).

In a recent revision of *Diploschistes* (Fernández-Brime et al. 2013), the phylogeny supported two separate clades corresponding to the morphological concepts of *D. diacapsis* and *D. diacapsis* subsp. *neutrophilus*. The authors also noticed morphological and ecological differences: *D. diacapsis* has thicker and convex thalli (up to 2 mm), which were detached from the substrate and is growing on gypsiferous or highly calcareous soils in continental areas, whereas *D. diacapsis* subsp. *neutrophilus* has thin and flat thalli completely attached to the substratum and grows on decarbonized soils in coastal areas. Based on these results, Fernández-Brime et al. (2013) raised *D. diacapsis* ssp., *neutrophilus* to species level. Our present study, however, includes a larger number of samples and loci, and our phylogeny (Fig. 2) shows that specimens with *D. diacapsis* and *D. neutrophilus* morphologies do not form distinct clades. Furthermore, several samples identified as *D. neutrophilus* were collected in the Tabernas desert (Table 1), a typical inland semi-arid *D. diacapsis* locality. In the light of these results based on more data, and finding the thallus thickness and shape a much more inconsistent character than previously believed, we formally synonymize *D. neutrophilus* with *D. diacapsis*.

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References

- Alors, D., Lumbsch, H. T., Divakar, P. K., Leavitt, S. D., & Crespo, A. (2016). An integrative approach for understanding diversity in the *Punctelia rudenta* species complex (Parmeliaceae, Ascomycota). *PloS One*, 11(2). doi:10.1371/journal.pone.0146537.
- Amo de Paz, G., Crespo, A., Cubas, P., Elix, J. A., & Lumbsch, H. T. (2012). Transoceanic dispersal and subsequent diversification on separate continents shaped diversity of the *Xanthoparmelia pulla* group (Ascomycota). *PloS One*, 7(6), e39683.
- Arguello, A., Del Prado, R., Cubas, P., & Crespo, A. (2007). *Parmelia quercina* (Parmeliaceae, Lecanorales) includes four phylogenetically supported morphospecies. *Biological Journal of the Linnean Society*, 91(3), 455–467.
- Camargo, A., Morando, M., Avila, L. J., & Sites, J. W., Jr. (2012). Species delimitation with ABC and other coalescent-based methods: a test of accuracy with simulations and an empirical example with lizards of the *Liolaemus darwini* complex (Squamata: Liolaemidae). *Evolution*, 66, 2834–2849, doi:10.1111/j.1558-5646.2012.01640.x.

- Camargo, A., & Sites, J. (2013). Species delimitation: a decade after the renaissance, the species problem - ongoing issues. In I. Pavlinov (Ed.), *InTech*, ISBN: 978-953-51-0957-0. doi:10.5772/52664. Available from: <http://www.intechopen.com/books/the-species-problem-ongoing-issues/species-delimitation-a-decade-after-the-renaissance>.
- Carstens, B. C., Pelletier, T. A., Reid, N. M., & Satler, J. D. (2013). How to fail at species delimitation. *Molecular Ecology*, 22(17), 4369–4383. doi:10.1111/mec.12413.
- Castresana, J. (2000). Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution*, 17(4), 540–552.
- Clauzade, G., & Roux, C. (1985). Likenoj de Okcidenta Euro. Ilustrita Determinlibro: Bulletin de la Societe Botanique du Centre-Ouest, Nouvelle Serie, Numero Special 7. Royan, France.
- Cornejo, C., & Scheidegger, C. (2015). Multi-gene phylogeny of the genus *Lobaria*: evidence of species-pair and allopatric speciation in East Asia. *American Journal of Botany*, 102(12), 2058–2073.
- Coyne, J. A., & Orr, H. A. (2004). *Speciation*. Sunderland: Sinauer Associates.
- Crespo, A., & Lumbsch, H. T. (2010). Cryptic species in lichen-forming fungi. *IMA Fungus*, 1, 167–170.
- Darriba, D., Taboada, G. L., Doallo, R., & Posada, D. (2012). jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods*, 9(8), 772.
- Degnan, J. H., & Rosenberg, N. A. (2009). Gene tree discordance, phylogenetic inference and the multispecies coalescent. *Trends in Ecology & Evolution*, 24(6), 332–340. doi:10.1016/j.tree.2009.01.009.
- Divakar, P. K., Figueras, G., Hladun, N. L., & Crespo, A. (2010). Molecular phylogenetic studies reveal an undescribed species within the north American concept of *Melanelixia glabra* (Parmeliaceae). *Fungal Diversity*, 42, 47–55.
- Döring, H., Clerc, P., Grube, M., & Wedin, M. (2000). Mycobiont-specific PCR primers for the amplification of nuclear ITS and LSU rDNA from lichenized ascomycetes. *Lichenologist*, 32, 200–204.
- Fernandez-Brime, S., Llimona, X., Molnar, K., Stenroos, S., Hognabba, F., Bjoerk, C., et al. (2011). Expansion of the Stictidaceae by the addition of the saxicolous lichen-forming genus *Ingvariella*. *Mycologia*, 103(4), 755–763. doi:10.3852/10-287.
- Fernández-Brime, S., Llimona, X., Lutzoni, F., & Gaya, E. (2013). Phylogenetic study of Diploschistes (lichen-forming Ascomycota: Ostropales: Graphidaceae), based on morphological, chemical, and molecular data. *Taxon*, 62(2), 267–280.
- Fujisawa, T., & Barraclough, T. G. (2013). Delimiting species using single-locus data and the generalized mixed yule coalescent (GMYC) approach: a revised method and evaluation on simulated datasets. *Systematic Biology*, 62(5), 707–724. doi:10.1093/sysbio/syt033.
- Gardes, M., & Bruns, T. D. (1993). ITS primers with enhanced specificity for basidiomycetes - Application to the identification of mycorrhizae and rusts. *Molecular Ecology*, 2, 113–118.
- Guderley, R., Lumbsch, H. T., & Feige, G. B. (1997). *Ingvariella*, a new genus in the Thelotremales (lichenized Ascomycotina). *Nova Hedwigia*, 64(1–2), 147–154.
- Hodkinson, B. P., & Lendemer, J. C. (2011). Molecular analyses reveal semi-cryptic species in *Xanthoparmelia tasmanica*. *Bibliotheca Lichenologica*, 106, 108–119.
- Jaklitsch, W. M., Baral, H. O., Lücking, R., & Lumbsch, H. T. (2016). Ascomycota. In W. Frey (Ed.), *Syllabus of plant families - Adolf Engler's syllabus der Pflanzenfamilien* (Vol. 1/2, 13th ed., p. 150). Stuttgart: Gebr. Borntraeger Verlagsbuchhandlung.
- Katoh, K., Asimenos, G., & Toh, H. (2009). Multiple alignment of DNA sequences with MAFFT. *Methods in Molecular Biology*, 537, 39–64.
- Kingman, J. (1982). The coalescent. *Stochastic Processes and their Applications*, 13, 235–248.
- Knowles, L. L., & Carstens, B. C. (2007). Delimiting species without monophyletic Gene trees. [article]. *Systematic Biology*, 56(6), 887–895. doi:10.1080/10635150701701091.
- Kraichak, E., Parmmen, S., Lücking, R., & Lumbsch, H. T. (2014). *Gintarasia* and *Xalocoa*, two new genera to accommodate temperate to subtropical species in the predominantly tropical Graphidaceae (Ostropales, Ascomycota). *Australian Systematic Botany*, 26, 466–474.
- Kraichak, E., Divakar, P. K., Crespo, A., Leavitt, S. D., Nelsen, M. P., Lücking, R., et al. (2015). A tale of two hyper-diversities: diversification dynamics of the two largest families of lichenized fungi. *Scientific Reports*, 5, e10028.
- Larena, I., Salazar, O., González, V., Julián, M. C., & Rubio, V. (1999). Design of a primer for ribosomal DNA internal transcribed spacer with enhanced specificity for ascomycetes. *Journal of Biotechnology*, 75, 187–194.
- Leavitt, S. D., Fankhauser, J. D., Leavitt, D. H., Porter, L. D., Johnson, L. A., & St Clair, L. L. (2011). Complex patterns of speciation in cosmopolitan "rock posy" lichens - Discovering and delimiting cryptic fungal species in the lichen-forming Rhizoplaca melanophthalma species-complex (Lecanoraceae, Ascomycota). *Molecular Phylogenetics and Evolution*, 59, 587–602. doi:10.1016/j.ympev.2011.03.020.
- Leavitt, S. D., Johnson, L., & St Clair, L. L. (2011a). Species delimitation and evolution in morphologically and chemically diverse communities of the lichen-forming genus *Xanthoparmelia* (Parmeliaceae, Ascomycota) in western North America. *American Journal of Botany*, 98(2), 175–188. doi:10.3732/ajb.1000230.
- Leavitt, S. D., Johnson, L. A., Goward, T., & St. Clair, L. L. (2011b). Species delimitation in taxonomically difficult lichen-forming fungi: an example from morphologically and chemically diverse *Xanthoparmelia* (Parmeliaceae) in North America. *Molecular Phylogenetics and Evolution*, 60, 317–332.
- Leavitt, S. D., Esslinger, T. L., Divakar, P. K., & Lumbsch, H. T. (2012). Miocene and Pliocene dominated diversification of the lichen-forming fungal genus *Melanohalea* (Parmeliaceae, Ascomycota) and Pleistocene population expansions. *BMC Evolutionary Biology*, 12, 176.
- Leavitt, S. D., Lumbsch, H. T., Stenroos, S., & St Clair, L. L. (2013). Pleistocene speciation in north American lichenized fungi and the impact of alternative species circumscriptions and rates of molecular evolution on divergence estimates. *PLoS One*, 8(12). doi:10.1371/journal.pone.0085240.
- Leavitt, S. D., Divakar, P. K., Ohmura, Y., Wang, L. S., Esslinger, T. L., & Lumbsch, H. T. (2015a). Who's getting around? Assessing species diversity and phylogeography in the widely distributed lichen-forming fungal genus *Montanelia* (Parmeliaceae, Ascomycota). *Molecular Phylogenetics and Evolution*, 90, 85–96. doi:10.1016/j.ympev.2015.04.029.
- Leavitt, S. D., Moreau, C. S., & Lumbsch, H. T. (2015b). The dynamic discipline of species delimitation: progress toward effectively recognizing species boundaries in natural populations. In D. K. Upreti, P. K. Divakar, V. Shukla, & R. Bajpai (Eds.), *Recent advances in lichenology* (pp. 11–44). India: Springer.
- Leavitt, S. D., Divakar, P. K., Crespo, A., & Lumbsch, H. T. (2016). A matter of time - understanding the limits of the power of molecular data for delimiting species boundaries. *Herzogia*, submitted. *Herzogia*, 29, submitted.
- Liu, Y. J., Whelen, S., & Hall, B. D. (1999). Phylogenetic relationships among ascomycetes: evidence from an RNA polymerase II subunit. *Molecular Biology and Evolution*, 16, 1799–1808.
- Llimona Pages, X. (1974). *Las Comunidades de Liqueenes de los Yesos de Espana: Universidad de Barcelona*. Barcelona: Secretariado de Publicaciones.

- Lumbsch, H. T. (1988). The identity of *Diploschistes gyphaceus*. *The Lichenologist*, 20(1), 19–24.
- Lumbsch, H. T. (1989). Die holarktischen Vertreter der Flechtengattung *Diploschistes* (Thelotremales). *Journal of the Hattori Botanical Laboratory*, 66, 133–196.
- Lumbsch, H. T., & Elix, J. A. (1989). Taxonomy of some *Diploschistes* spp (lichenized ascomycetes, Thelotremales) containing gyrophoric acid. *Plant Systematics and Evolution*, 167(3–4), 195–199.
- Lumbsch, H. T., & Huhndorf, S. M. (2010). Myconet volume 14. Part one. Outline of Ascomycota - 2009. *Fieldiana (Life and Earth Sciences)*, 1, 1–42.
- Lumbsch, H. T., & Leavitt, S. D. (2011). Goodbye morphology? A paradigm shift in the delimitation of species in lichenized fungi. *Fungal Diversity*, 50, 59–72.
- Lumbsch, H. T., & Tehler, A. (1998). A cladistic analysis of the genus *Diploschistes* (Ascomycotina, Thelotremales). *Bryologist*, 101(3), 398–403.
- Mangold, A., Martin, M. P., Lücking, R., & Lumbsch, H. T. (2008). Molecular phylogeny suggests synonymy of Thelotremales within Graphidaceae (Ascomycota : Ostropales). *Taxon*, 57, 476–486.
- Martín, M. P., LaGreca, S., & Lumbsch, H. T. (2003). Molecular phylogeny of *Diploschistes* inferred from ITS sequence data. *The Lichenologist*, 35(1), 27–32.
- Matheny, P. B., Liu, Y. J., Ammirati, J. F., & Hall, B. D. (2002). Using RPB1 and RPB2 nucleotide sequences (Inocybe; Agaricales). *American Journal of Botany*, 89, 688–698.
- Mayr, E. (1963). *Animal species and evolution*. Cambridge: Harvard University Press.
- Nixon, K., & Wheeler, Q. (1990). An amplification of the phylogenetic species concept. *Cladistics*, 6, 211–223.
- Nylander, J. A. A., Wilgenbusch, J. C., Warren, D. L., & Swofford, D. L. (2008). AWTY (are We there yet?): a system for graphical exploration of MCMC convergence in Bayesian phylogenetics. *Bioinformatics*, 24, 581–583.
- Otálora, M. A. G., Martínez, I., Aragón, G., & Molina, M. C. (2010). Phylogeography and divergence date estimates of a lichen species complex with a disjunct distribution pattern. *American Journal of Botany*, 97, 216–223.
- Pant, G., & Upreti, D. K. (1993). The lichen genus *Diploschistes* in India and Nepal. *The Lichenologist*, 25(1), 33–50.
- Parnmen, S., Rangsiruji, A., Mongkolsuk, P., Boonpragob, K., Nutakki, A., & Lumbsch, H. T. (2012). Using phylogenetic and coalescent methods to understand the species diversity in the *Cladia aggregata* Complex (Ascomycota, Lecanorales). *PloS One*, 7(12), e52245. doi:10.1371/journal.pone.0052245.
- Parnmen, S., Cáceres, M. E. S., Lücking, R., & Lumbsch, H. T. (2013). *Myriochapsa* and *Nitidochapsa*, two new genera in Graphidaceae (Ascomycota: Ostropales) for chroodiscoid species in the *Ocellularia* clade. *Bryologist*, 116, 127–133.
- Printzen, C. (2010). Progress in botany. In U. Lüttge, W. Beyschlag, B. Büdel, & D. Francis (Eds.), *Lichen systematics: the role of morphological and molecular data to reconstruct phylogenetic relationships* (Vol. 71, pp. 233–275). Botany: Progress in Botany.
- Rambaut, A. (2009). FigTree 1.2.2. <http://tree.bio.ed.ac.uk/software/figtree/>.
- Rannala, B. (2015). The art and science of species delimitation. *Current Zoology*, 61(5), 846–853.
- Rannala, B., & Yang, Z. (2013). Improved reversible jump algorithms for Bayesian species delimitation. *Genetics*, 194, 245–253.
- Rivas Plata, E. (2011). *Historical biogeography, ecology and systematics of the family Graphidaceae (lichenized Ascomycota: Ostropales)*. Chicago: University of Illinois at Chicago.
- Rivas Plata, E., Lücking, R., Sipman, H. J. M., Mangold, A., & Lumbsch, H. T. (2010). A world-wide key to the thelotremoid Graphidaceae, excluding the *Ocellularia-Myriotrema-Stegobolus* clade. *The Lichenologist*, 42, 139–185.
- Rivas Plata, E., Parnmen, S., Staiger, B., Mangold, A., Frisch, A., Weerakoon, G., et al. (2013). A molecular phylogeny of Graphidaceae (Ascomycota: Lecanoromycetes: Ostropales) including 437 species. *MycKeys*, 6, 55–94.
- Ronquist, F., Teslenko, M., van der Mark, P., Ayres, D. L., Darling, A., Höhna, S., et al. (2012). MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology*, 61, 539–542.
- Schmitt, I., Crespo, A., Divakar, P. K., Fankhauser, J., Herman-Sackett, E., Nelsen, M. P., et al. (2009). New primers for single-copy protein-coding genes for fungal systematics. *Persoonia - Molecular Phylogeny and Evolution of Fungi*, 23, 35–40.
- Sites, J. W., & Marshall, J. C. (2004). Operational criteria for delimiting species. *Annual Review of Ecology, Evolution, and Systematics*, 35(1), 199–227. doi:10.1146/annurev.ecolsys.35.112202.130128.
- Stamatakis, A. (2006). RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics*, 22, 2688–2690.
- Stiller, J. W., & Hall, B. D. (1997). The origin of red algae: implications for plastid evolution. *Proceedings of the National Academy of Sciences of the United States of America*, 94, 4520–4525.
- Thell, A., Elix, J. A., & Söchting, U. (2009). *Xanthoparmelia lineola* s. l. in Australia and North America. *Bibliotheca Lichenologica*, 99, 393–404.
- Vilgalys, R., & Hester, M. (1990). Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology*, 172, 4238–4246.
- Yang, Z. (2015). The BPP program for species tree estimation and species delimitation. *Current Zoology*, 61(5), 854–865.
- Yang, Z., & Rannala, B. (2010a). Bayesian species delimitation using multilocus sequence data. *Proceedings of the National Academy of Sciences*, 107(20), 9264–9269. doi:10.1073/pnas.0913022107.
- Yang, Z., & Rannala, B. (2010b). Bayesian species delimitation using multilocus sequence data. *Proceedings of the National Academy of Sciences of the United States of America*, 107, 9264–9269.
- Yang, Z., & Rannala, B. (2014). Unguided species delimitation using DNA sequence data from multiple loci. *Molecular Biology and Evolution*, 31, 3125–3135.
- Zhang, C., Zhang, D. X., Zhu, T., & Yang, Z. (2011). Evaluation of a Bayesian coalescent method of species delimitation. *Systematic Biology*, 60, 747–761.
- Zhao, X., Zhang, L. L., Zhao, Z. T., Wang, W. C., Leavitt, S. D., & Lumbsch, H. T. (2015). A molecular phylogeny of the lichen genus *Lecidella* focusing on species from mainland China. *PloS One*, 10(9). doi:10.1371/journal.pone.0139405.
- Zhou, S., & Stanosz, G. R. (2001). Primers for amplification of mt SSU rDNA, and a phylogenetic study of *Botryosphaeria* and associated anamorphic fungi. *Mycological Research*, 105, 1033–1044.
- Zoller, S., Scheidegger, C., & Sperisen, C. (1999). PCR primers for the amplification of mitochondrial small subunit ribosomal DNA of lichen-forming ascomycetes. *Lichenologist*, 31, 511–516.