

DETECTION OF MYCOPLASMA BOVIGENITALIUM ANTIBODIES BY INDIRECT-ELISA IN COWS WITH VARIOUS REPRODUCTIVE DISORDERS

P. CHANDER, Y. SINGH¹ and D. N. GARG

Department of Veterinary Public Health, College of Veterinary Sciences
CCS Haryana Agricultural University, Hisar-125 004

ABSTRACT

During present investigation, a total of 135 cows comprising 85 diseased (15 abortion/still-birth, 17 metritis, 6 cervicitis, 25 repeat breed, 22 anoestrus) and 50 apparently healthy, were tested for *Mycoplasma bovis* infection by indirect-ELISA. *M. bovis* specific ELISA-titre ($\geq 1:160$) was detected in 30 out of 85 (35.3%) diseased and 11 out of 50 (22%) apparently healthy cows. Higher percentage of cows having anoestrus (45.55%) and abortion/still-birth (80%) proved positive for *M. bovis* ELISA-Ab as compared to repeat breed, cervicitis and metritis cases (16 to 17.64%).

Key words: *Mycoplasma bovis*, cows, indirect-ELISA, reproductive disorders

Mycoplasma bovis, a potential genital pathogen of bovines, is known to cause a variety of reproductive disorders viz. granular vulvovaginitis, abortion/still-birth, seminal vesiculitis and sperm abnormalities. This organism was first isolated from genital tract of infertile cows having bursitis and salpingitis (Edward *et al.*, 1947). In India, it was first reported from the genital tract of cow having the history of repeat breeding (Jayaraman, 1961) and has been reported continuously from various bovine reproductive disorders (endometritis, cervico-vaginitis, cervicitis, repeat breed, abortion, low fertile bull-semen) as well as from apparently healthy bovines (Volintir *et al.*, 1970, Misra *et al.*, 1975, Ball *et al.*, 1978, Pal *et al.*, 1982, 1984, Garg *et al.*, 1988, Sharma *et al.*, 1997). In the present study, cows having reproductive disorders as well as healthy cows were tested for *M. bovis* infection by indirect-ELISA.

MATERIALS AND METHODS

Specimen: Serum samples from 135 cows including 85 diseased (16 abortion/still birth, 17 metritis, 6 cervicitis, 25 repeat breed, 22 anoestrus) and 50 apparently healthy cows were

collected. In case of abortion/still birth cases, paired serum samples were collected from dam at an interval of three weeks.

Production of hyperimmune serum: A polyclonal hyperimmune serum against *M. bovis* (PG-11) was raised in rabbits (New Zealand white) and bovine calves (4-6 months age), respectively (Chander, 1999).

Indirect ELISA: To perform this test, the procedure described earlier by Garg *et al.* (1997) was followed. The sonicated whole cell *M. bovis* (PG-11) antigen diluted (1:25) in carbonate-bicarbonate buffer (pH 9.2) was centrifuged at 10,000 g for 30 minutes and protein of supernatant was adjusted so that each well receives 4.8 μ g protein. After adding 100 μ l of *M. bovis* antigen (1:100) in each well, the microtitre plates were incubated at 37°C for 2 h followed by overnight incubation at 4°C. After three washings with PBS-tween-20, diluted test serum (100 μ l) was added to each well and incubated at 37°C for 1 h. The phosphate buffer saline (pH 7.6) containing 0.05% Tween-20 was used as serum and conjugate diluent. The horse-radish-peroxidase (type VI) labelled rabbit anti-cow IgG (M/s Dakopatts, Denmark) was used as enzyme-antiglobulin conjugate. Optimum dilution of coating conjugate was 1:2000. The substrate used was orthophenylene diamine-

¹ Corresponding author

Table 1
Seroprevalence of *M. bovis genitalium* ELISA- Ab in cows with various reproductive disorders and apparently healthy

ELISA titre	Number of animals							
	Apparently Healthy	Diseased						Total
		Anoestrus	Repeat breeders	Cervicitis	Metritis	Abortion/Still-birth		
					1-3D	21D		
0 to 1:40	29	10	14	3	9	6	2	38 (49.35)*
1:80	10	2	7	2	5	2	1	17 (20.00)
1:160	5	6	0	0	1	3	2	09 (10.58)
1:320	6	2	4	0	2	4	3	11 (12.94)
1:640	0	1	0	1	0	0	5	07 (8.23)
1:1280	0	1	0	0	0	0	2	03 (3.52)
ELISA+ve	11 (22)	10 (45.55)	4 (16)	1 (16.66)	3 (17.64)	7 (46.66)	12 (80)	30 (35.30)

* Figure in parenthesis shows percentage, D = day

dihydrochloride (OPD, Sigma, USA), which was prepared fresh in 30 % hydrogen peroxide solution with 0.05M citrate buffer. Absorbance values were measured at 492 nm in a Organon Teknika Reader 530. With each test plate, known positive and known negative sera as controls were included. The maximum absorbance of reference negative serum was considered as cut-off value for calculating ELISA end titre. Eliminating a cross-reacting titres upto 1:80, the serum samples showing $\geq 1:160$ ELISA titre were considered positive.

RESULTS AND DISCUSSION

The results of seroprevalence of *M. bovis genitalium* antibodies in cows with various reproductive disorders and normal health are depicted in the Table 1. *M. bovis genitalium* specific ELISA-titre ($\geq 1:160$) was detected in 30 out of 85 diseased (35.30%) and 11 out of 50 (22%) apparently healthy cows. A higher rate of prevalence of *M. bovis genitalium* ELISA-Ab was recorded in genitally diseased cows in comparison to that in apparently healthy cows. Data regarding seroprevalence against *M. bovis genitalium* using ELISA in cows with various reproductive disorders is scanty. Higher seroprevalence of specific *M. bovis genitalium* ELISA antibodies in genitally diseased cows (19.1%) and mastitic cows (9.4%) than that in healthy cows (8.2%) has been recorded earlier (Kumar and Garg, 1996, Garg *et al.*, 1999). Disease-wise specific seroprevalence recorded

against *M. bovis genitalium* in diseased cows ranged from 16 to 80% where the highest incidence was seen in cows showing anoestrus (45.45%) or abortion/still birth (80%) conditions as compared to repeat breed, cervicitis and metritis cases (16-17%). These findings are in agreement with Garg *et al.* (1999) who also reported higher incidence of *M. bovis genitalium* antibodies using ELISA in aborted cows (30.5%) as compared to repeat breed condition (11%). The results of present study indicate that amongst various reproductive disorders in cows, seroprevalence of *M. bovis genitalium* infection was more in cases of anoestrus and abortion.

Acknowledgements

The authors thankfully acknowledge the financial support of the Indian Council of Agricultural Research, New Delhi under a National Fellow Project awarded to Dr. D. N. Garg.

REFERENCES

- Ball, H.J., Neill, S.D., Ellis, W.A., O'Brien, J.J., Ferguson, H.W. (1978). The isolation of Mycoplasma from bovine fetuses and their dams. *Br. Vet. J.* **134**: 584-589.
- Chander, P. (1999). Studies on genital mycoplasmosis in cows with special emphasis on *M. bovis genitalium*. M.V.Sc thesis. CCS HAU, Hisar.
- Edward, D.G.F., Hancock, J.L., Hignett, S.L. (1947). Isolation of pleuropneumonia like organisms from the bovine genital tract. *Vet. Rec.* **59**: 329-330.
- Garg, D.N., Singh, Y. and Kapoor, P.K. (1988). Mollicutes organisms in the genital tract of male bovines. *Indian*

- J. Anim. Sci.* **58**: 1407.
- Garg, D.N., Singh, Y. and Kapoor, P.K. (1997). In: Laboratory Manual of Enzyme-immuno assays in Veterinary Microbiology' Eds. S.C. Tewari and G. Prasad, pp 29-35. ICAR-CAS, Dept. Vet. Microbiol., CCS Haryana Agri. University, Hisar.
- Garg, D.N., Kapoor, P.K. and Singh, Y. (1999). Detection of mycoplasmal antibodies in bovines with reproductive disorders. *Indian J. Comp. Microbiol. Immunol. Infect. Dis.* **20** : 32-35
- Jayaraman, M.S. (1961). Isolation of *Mycoplasma bovis genitalium* from the genital tract of a cow. *Indian Vet. J.* **38**: 17.
- Kumar, A. and Garg, D.N. (1996). Detection of serum antibodies to *M. bovis* and *M. bovis genitalium* in mastitic cows and buffaloes by ELISA, IHA and AGIPT. *Indian Vet. J.* **73**: 603-606.
- Mishra, P.K., Panda, S.N., Kar, B.C., Misra, B. and Nayak, B.C. (1975). Isolation and characterization of *Mycoplasma bovis genitalium* from bovine vaginal tract. *Indian J. Anim. Hlth.* **14**: 103-107.
- Pal, B.C., Singh, P.P. and Pathak, R.C. (1982). Involvement of *Mycoplasma bovis genitalium* in the genital tract of female buffaloes. *Indian J. Vet. Med.* **2**: 37-40.
- Pal, B.C., Singh, P.P. and Pathak, R.C. (1984). Mycoplasmas from genital tract of aborting buffaloes. *Indian Vet. J.* **61**: 1-3.
- Sharma, V., Dhanesar, N.S. and Mehra, K.N. (1997). *Indian J. Comp. Microbiol. Immunol. Infect. Dis.* **18**:157.
- Volintir, V., Popescu, I., Moga-Minzat, R., Constantinescu, S. and Purcel-Vlah, M. (1970). Study of *Mycoplasma* infection of uterus in cows. *Lucrari-Stiintifice-Institutul-Agronomic-Timisoara, Seria. Medicina Veterinara* **13**: 189-197 (Abst. Vet. Bull. **43**: 3308).

ATTENTION

Membership of the Haryana Veterinarian is open on a nominal subscription to all professionals actively engaged in the veterinary practice or associated activities such as research, extension and teaching in veterinary profession. The subscription of Rs 800/- for membership for 10 years and @ 100/- per annum may be sent to Dean, College of Vety. Sciences, Hisar (Haryana) through Demand Draft/Money order with intimation to the Editor.

Editors

With Best Compliments from:

CHHABRA MEDICAL AGENCIES

*A Leading House of Veterinary Health
Products Poultry/Canine Vaccines etc.*

DISTRIBUTOR FOR:

- ⇒ Sarabhai Chemicals (Vety. Division)

- ⇒ Glaxo India Ltd. (Vety. Division)

- ⇒ Ranbaxy FineChemical Ltd. (Vety. Division)

- ⇒ Merind Ltd. (Vety. Division)

- ⇒ Intervet India Ltd. (Vety. Division)

- ⇒ Pfizer Ltd. (Vety. Division)

- ⇒ Alembic Chemicals (Vety. Division)

- ⇒ Novartis India Ltd. (Vety. Division)

**Near Haryana Agricultural University,
Gate No. 1, Prem Nagar, HISAR (Haryana)
Ph.: 237481, 238844**