The early clinical development of a multicomponent vaccine against meningococcal serogroup B

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The development of meningococcal serogroup B vaccines has been a worldwide public health priority based on continuing disease burden, combined with the scientific challenges associated with antigen identification. A new multicomponent vaccine, 4CMenB, is currently being evaluated for global licensure. The multicomponent strategy accounts for multiple surface antigens and, therefore, provides varied opportunities to induce bactericidal antibodies. 4CMenB contains four components: outer membrane vesicles from the New Zealand MenB outbreak strain, fHbp, NadA and NHBA. In early clinical studies protective antibody levels with acceptable tolerability outcomes were observed in persons who received 4CMenB starting at as young as 2 months of age. A meningococcal antigen-typing system has been developed to bridge clinical trial data with circulating strains. This article describes the early clinical development program and the rationale for Phase III study design and effectiveness evaluations.

Keywords: fHbp • MenB vaccine • meningococcal disease • meningococcal serogroup B • NadA • NHBA • PorA

The development of vaccines against Neisseria meningitidis serogroup B (MenB) has long been viewed by experts as a global public health priority, based on continuing disease burden combined with the scientific challenges associated with antigen identification [1-3]. Invasive MenB disproportionately affects infants in the developed world and has been associated with extended epidemics and outbreaks. MenB is the leading cause of meningococcal meningitis and sepsis in European infants, and attack rates in the UK have long been comparable to those that led to universal vaccination against meningococcal serogroup C (Figure 1) [4-12]. With the recent introduction of a low-cost serogroup A conjugate vaccine (MenAfriVac^{*}) for use in Africa [13,14], experts note that MenB remains the last major hurdle to controlling invasive meningococcal disease (IMD) globally [2,3]. A multicomponent MenB vaccine, 4CMenB, has been submitted for consideration by licensing bodies [1,15-17]. We consider a recent body of literature on the topic of MenB vaccines, including recent review work, to provide a broader context and discussion about the early development program and the design of the late-stage clinical trials of 4CMenB, including plans for bridging clinical trial data against current regional epidemiology [1,15-21].

Rationale for vaccination against MenB

Several factors inform the rationale for establishing immunization programs against MenB disease, including symptoms, epidemiology, persistent case fatality rates and other undesirable outcomes [1,18,22,23]. IMD generally presents as rapidly progressive meningitis and/or septicemia, although small numbers of meningo-coccal pneumonia cases and other rare syndromes are reported in the literature

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Figure 1. Regions and countries with reports of serogroup B meningococcal disease.

[7,22,24]. Initial symptoms are indistinguishable from mild respiratory illnesses and progression from nonspecific signs to permanent disability or death is generally rapid and difficult to control once sepsis or shock develop. The advent of antibiotic therapies reduced the case fatality rate from IMD to approximately 10% overall, although the rates in septicemia patients (which can exceed 30%), or in the developing world or during epidemics, are often higher. The case fatality rate has remained relatively stable despite advances in therapeutic options [6,25] and the risks of death and disability in the developing world are higher relative to those in the developed world [26]. Delays in differential diagnosis can contribute to negative outcomes in some patients, particularly if culture confirmation is required to refine treatment choices [23,27]. Most people who develop IMD are otherwise healthy and have no commonly recognized risk factors, which contributes to public anxiety and bolsters epidemiologic and clinical rationales for preventive measures, such as vaccination [27].

While chemoprophylaxis of close contacts of IMD cases can be effective in limiting outbreaks [27,28,201],

vaccination is the primary prevention strategy [7,23,29]. Conjugate vaccines against serogroups A, C, W-135 and Y have robust safety and immunogenicity profiles in all age groups and have been included in routine immunization programs in many countries for infants, children, adolescents and those at risk of developing meningococcal disease [30-33], In North America, adolescents, a primary reservoir for asymptomatic meningococcal carriage, commonly receive quadrivalent conjugate vaccine, which is also increasingly recommended for Hajj and Umrah [9,18,34]. Investigations into universal or broad-coverage MenB vaccines have been hampered by the poor immunogenicity of the capsular polysaccharide [2,7]. Three licensed wild-type (wt) outer membrane vesicle (OMV) vaccines were tailor-made to address specific pathogenic strains, and have successfully limited clonal outbreaks and epidemics [1,3,35-38]. Cuba has the only routine recommendation for serogroup B IMD immunization [7,18,38], using a tailor-made wtOMV vaccine that protects against strains bearing a specific PorA serosubtype [7,36,37]. However, the epidemiology and incidence of sporadic disease and the

possibility of new epidemics spurred the development of vaccines to address MenB more comprehensively [1,15,16,35].

MenB vaccines

The poor immunogenicity of the MenB capsular polysaccharide results because it is identical to the polysialic acid in the fetal neural cell-adhesion molecule $_{[19,35-37]}$. Although this has raised concerns about the possibility that autoimmune responses could be induced by MenB conjugate vaccines, no immunogenic formulation has been developed to confirm or disprove this hypothesis $_{[1,2,7,19,39]}$. All ongoing clinical vaccine development includes subcapsular antigens that exhibit considerable antigenic diversity and, therefore, pose significant challenges for vaccine development $_{[1-3,7,39-41]}$, including concerns about poor efficacy and the development of escape variants or mutants $_{[41]}$

The three licensed wtOMV vaccines (VA-MENGOC -BC^{*}, MenBVac^{*} and MeNZB^{*}) employ detergent-extracted OMV or culture-released blebs [1,36]. In contrast, recombinant OMVs are produced through genetic modification. To maintain the surface expression of proteins in a stable membrane, lipopolysaccharide (LPS), a known reactogen, is necessary to ensure the integrity of the OMV phospholipid bilayer. Various factors, such as detergent or aluminum adjuvant use, are known to limit LPS reactogenicity in vaccines. Furthermore, membrane-bound LPS is much less reactogenic than the free form [36,42].

All OMVs contain various proteins that can theoretically contribute to immune responses. Such proteins include PorA, PorB, RmpM and OpcA invasin (Figure 2), each of which has been suggested as a possible vaccine antigen candidate [1,15,16,35,43]. Iron-regulated proteins may also be evident on wtOMVs. Based on the results of clinical studies, PorA is considered to be the immunodominant constituent of OMV in infants, who are the target population for MenB disease [36,37,44–46]. Nevertheless, it has been suggested that additional proteins, such as PorB may also elicit antibody formation [46].

Licensed wtOMV vaccines

The wtOMV vaccines were developed to limit longterm clonal MenB epidemics in Cuba, Norway and New Zealand (Table 1). MeNZB, which was implemented in New Zealand, was associated with marked reductions in IMD incidence during a serosubtype B:4:P1.7b,4 ST-41/44 (strain NZ 98/254) outbreak that reached an incidence of 17.4/100,000 of the population overall and more than 200/100,000 among some indigenous populations [47]. Initial vaccine effectiveness was 70–80% [48] and declined over time [49,50]. VA-MENGOC-BC and MenBVac have been used to prevent or control epidemic MenB IMD in Brazil, Chile and France caused by epidemic strains carrying PorA serosubtypes matched to the vaccines [36,50–53]. In infants, clinical data show that wtOMV vaccines do not provide cross protection against strains expressing heterologous PorA [7,36,37]. Furthermore, while these wtOMV vaccines may interrupt the acquisition of new carriage, only limited effects have been observed on existing carriage, suggesting that wtOMV vaccines will not provide similar herd-protection effects compared with conjugate vaccines, even for the target strains [4,23,36].

Millions of doses of wtOMV vaccine have been administered, yielding extensive safety outcomes data. Self-limited reactogenicity has been observed with wtOMV vaccines. Adults tend to report headache, injection site pain and swelling and generalized arm pain, while transient fever lasting 48 h or less and irritability has been noted in infants and young children. Results of several placebo-controlled studies suggest that aluminum-containing adjuvants may contribute to these reactogenicity outcomes [36,54-57].

The wtOMV vaccines have also been investigated in combination with additional components. For example, MeNZB has been studied in combination with a serogroup C conjugate vaccine and with MenBVac [51]. Recombinant OMVs designed to overexpress relevant surface proteins or purified proteins derived from OMVs have also been developed as a means to help overcome some limitations of wt vaccines (Table 2) [1,15,35-37,44,46,58]. However, PorA proteins, an important feature of the recombinant OMV vaccines, exhibit great variability, which can limit their potential usefulness as vaccine antigens [40]. In the USA, 20 different PorA variants would be needed to cover 80% of circulating MenB strains. In addition, preliminary clinical trials of hexavalent and nonavalent PorA vaccines have shown stronger immunogenicity for some PorA variants than others [59-61]. The addition of different antigens is a means of broadening protection and a vaccine with six PorA and five FetA variants has been suggested to address hypervirulent lineages [40,59].

Investigational vaccines against MenB

Most experimental MenB vaccines against serogroup B continue to use one or more OMV component, in part because the wtOMV vaccines are the only products to have demonstrated effectiveness. In addition, OMV may provide immunomodulatory or adjuvant effects beyond the primary immunogenicity contributed by PorA, as observed with a combination



Figure 2. Identification of various proteins within a typical wild-type outer membrane vesicle used in vaccine development. (A) Shows a representative Coomassie gel and a representative lane electropherogram, shown with the corresponding gel lane for a single protein. **(B)** Protein pattern composition, relative ratio of outer membrane proteins and total purity are monitored by SDS-PAGE. A typical SDS-PAGE separation for outer membrane vesicle is presented in **(A)**. Evaluation of the presence and relative quantities of eight proteins was performed by densitometric evaluation of gel lane electropherograms.

Table 1. Outer membrane vesicle vaccines licensed for clinical use.				
OMV type	Trade name	Countries employing	Number of doses administered	
Finlay 4:P1.19, 15	VA-MENGOC-BC®	Cuba, Chile and Brazil	2	
NIPH 15:P1.7, 16	MenBVac [®]	Norway and France	3	
NIPH/Novartis 4:P1.7–2,4	MeNZB®	New Zealand	3	
OMV: Outer membrane vesicle.				

Table 2. Examples of subcapsular proteins under investigation for use in meningococcal serogroup B vaccines.				
Protein(s)	Type of vaccine	Latest research stage initiated		
Single PorA	wtOMV	Licensed for use against specific clonal groups		
Multiple PorA	Recombinant OMV	Early clinical trials		
ОрсА	OMV	Early clinical trials		
Bivalent PorA with fHbp and NadA	Multicomponent	Early clinical trials		
Bivalent fHbp	Multicomponent	Advanced clinical trials		
fHbp with NadA and NHBA (rMenB)	Multicomponent	Early clinical trials		
rMenB with Norway strain OMV	Multicomponent	Early clinical trials		
4CMenB (rMenB with New Zealand strain OMV)	Multicomponent	Application for licensure		
MenB: Neisseria meningitidis serogroup B; OMV: Outer membrane vesicle; wtOMV: Wild-type OMV.				

OMV and outer membrane protein vaccine, which provides an additional rationale for inclusion in novel formulations [62].

Bivalent, hexavalent and nonavalent vaccines based on recombinant OMVs that express multiple different PorA serosubtypes have entered trials in adults [63,64,202] and are described based on the number of PorA variants, not necessarily the number of OMV components [60,61,65]. The hexavalent and nonavalent vaccines contain two or three OMVs, respectively, each expressing three different PorA variants. The nonavalent vaccine has also been tested in combination with a pneumococcal conjugate vaccine [61]. Additional OMV vaccines, including formulations containing OpcA with or without overexpressed fHbp, have also been studied [66,67]. Vaccines based on OMV from genetically detoxified or mutant lipooligosaccharides that eliminate the detergent extraction step that can affect protein conformation, have also entered clinical trials [15,16,36]. It was hoped that an OMV vaccine derived from Neisseria lactamica, which has surface proteins similar to those on the meningococcus, would be able to induce crossprotective antibodies for both species, but this has

not progressed to late-stage clinical development [1,68]. Additional vaccines have induced bactericidal activity against OpcA and fHbp with good tolerability in adults [2,35,66,67].

Vaccine formulations containing fHbp, either alone, in combination with additional recombinant antigens, or in multicomponent formulations containing OMV, have been shown to generate immune responses [15,16,35,66,67,69]. As mentioned above, one multicomponent vaccine, 4CMenB, has completed late-stage clinical trials and is under consideration for licensure. It contains four components: OMV from the New Zealand outbreak strain, fHbp, NadA and NHBA.

Reverse vaccinology: antigen identification for novel MenB vaccines

The fHbp, NadA and NHBA included in 4CMenB were identified via reverse vaccinology. Whole genome sequences represent a list of virtually all the proteins that a pathogen can express at any time and reverse vaccinology uses bioinformatic algorithms to 'mine' these sequences for potential vaccine antigens. The first group of targets was identified for MenB strain

Table 3. Major antigens included in 4CMenB.					
Antigen	Full name	Number of variants	Biological function (example)	Presented as	
fHbp	Factor H binding protein	Three major non- crossreactive variants⁺	Recruits host molecules to the bacterial surface, facilitating survival in host tissues	Fusion protein with GNA 2091	
NadA	Neisserial [‡] adhesin A	Five major variants; variants 1–3 are crossreactive	Mediates adhesion and invasion of host cells	Self (nonfused)	
NHBA	Neisserial [‡] heparin binding antigen	Over 24 crossreactive peptides	Recruits host molecules to the bacterial surface, aiding survival	Fusion protein with GNA 1030	
PorA	Porin A	Multiple non-crossreactive serosubtypes	Participates in transport into and out of the cell membrane	OMV	
[†] An alternate nomenclature groups variants 2 and 3 into subfamily A and terms variant 1 subfamily B, based on genetic information. [†] Alternative spellings have appeared in the literature					

OMV: Outer membrane vesicle.

MC58 [70] and yielded 28 proteins that could be expressed as recombinant proteins in Escherichia coli, were surface-expressed and also induced bactericidal antibody responses in animals. The most immunogenic of these fHbp, NadA and NHBA were selected for inclusion in a multicomponent vaccine (Table 3) to provide broad strain coverage and minimize the potential for immune evasion and the development of escape mutants. To enhance immunogenicity and facilitate large-scale manufacturing, NHBA was fused to a meningococcal accessory protein (GNA1030) and fHbp was fused to GNA2091. These fusion proteins, formulated with aluminum hydroxide, induced more potent bactericidal antibodies than those induced by the individual antigens [71]. NadA was included as a single protein because it did not perform well in fusion, which may have altered its native trimeric organization [72]. These antigens were adsorbed to aluminum hydroxide [71] for use in clinical trials [15,16,70,73].

fHbp

Meningococcal fHbp is a 27 kDa membrane-bound lipoprotein that binds human factor H, a downregulator of the alternative complement pathway. Thus, fHbp aids bacterial survival in human blood by permitting complement evasion [20,74,75]. Surface expression of fHbp can be high, intermediate or low [76,77]; only a few strains (1% in the USA) lack fHbp expression altogether [78]. Such strains may employ alternative systems for recruiting human factor H, such as NspA [79]. Three fHbp sequence variants,



Figure 3. Simplified dendrograms showing the major variants and subvariants of fHbp, NadA and NHBA. (A) Three variants exist for fHbp; fHbp-induced antibodies are crossprotective against strains carrying homologous, but not distantly related alleles. (B) Five variants have been identified for NadA. The three main variants (NadA-1, NadA-2 and NadA-3) are crossprotective regardless of sequence variations. (C) NHBA is structured into a substantial number of peptides. Antibodies induced by NHBA peptide 2 are crossprotective against most strains tested thus far. Courtesy of S Bambini (Novartis Vaccines, Siena, Italy). with intravariant conservation of 91.6–100% and between-variant conservation as low as 62.8%, have been identified. An alternate classification system divides fHbp into subfamilies A (variants 2 and 3) and B (variant 1), based on genetic relationships between the variants [80]. A bivalent fHbp vaccine is being developed based on the hypothesis that cross-reactivity from two subvariants corresponding to the subfamily classification, would be sufficiently cross-reactive to provide protection against a majority of pathogenic strains; limited clinical data have been published describing the effects of this vaccine [81].

The three fHbp variants (termed 1, 2 and 3) induce bactericidal antibodies that exhibit evidence of intraand inter-variant cross reactivity that appears to vary by age (Figure 3). Within all variants, crossprotection by anti-fHbp antibodies is strongest for strains carrying homologous alleles and weaker against isolates harboring distantly related variants or subvariants [76]. The fHbp subvariant 1.1 fusion protein included in 4CMenB has been shown to induce bactericidal antibodies in 90-100% of infant, adolescent and adult vaccinees [82-86]. When tested against a panel of isogenic serogroup B strains engineered to express ten different fHbp variant 1 subvariants [87], sera from adults who received two to four doses of 4CMenB and 13-monthold toddlers vaccinated at 2, 4, 6 and 12 months of age were crossreactive against all ten subvariants. However, sera from 7-month-old infants who received three doses of 4CMenB were cross protective for closely related subvariants only [87].

NadA

NadA is an 'Oca' (oligomeric coiled-coil adhesin) bacterial trimeric autotransporter adhesin [88]. These proteins characteristically form trimers on the bacterial surface and mediate meningococcal adhesion to, and entry into, human epithelial cells [72]. NadA is commonly considered a meningococcal protein. No known strains of *Neisseria gonorrhea* or the commensal species *N. lactamica* and *Neisseria cinerea* harbor the *nadA* gene [88]. Recent analysis using a noninvasive *Yersinia enterocolitica* mutant engineered to express NadA revealed that β -1 integrin is a likely NadA receptor [89].

Unlike fHbp and NHBA, which occur on nearly all meningococcal strains, NadA is found primarily in a substantial subset of pathogenic strains and is associated with three of the four known hypervirulent lineages of MenB and MenC. In the USA, the *nadA* gene was identified in 39% of strains in a panel of 650 MenB isolates collected between 2000 and 2008 [90]. In this panel, NadA was more common in pathogenic strains

than in strains associated primarily with isolates from asymptomatic carriers or 'carriage strains.'

In clones where it is present, NadA is well conserved. Five variant alleles have been identified [88,91,92] and the three most common variants (NadA-1, NadA-2 and NadA-3) induce crossreactive antibody activity in animals and humans, regardless of sequence variations (Figure 3). NadA-4 occurs more rarely and is strongly associated with 'carriage strains' [77,91], while NadA-5 is generally associated with strains from a single clonal complex [92].

The *nadA* gene is both subject to complex regulatory controls and highly dependent on environmental signals, leading to highly variable expression in different conditions [88,93,94]. During bacterial growth, NadA is expressed maximally in stationary phase [88]. Phase variation occurs because of a tetranucleotide tract (TAAA) that is located upstream of the nadA promoter [93] and controlled by transcriptional regulator, NadR, which represses surface exposure in vitro [94]. 4-hydroxyphenyl acetate (4-HPA), aromatic amino acid catabolites secreted in human saliva, can de-repress NadA transcription in vitro [95], promoting surface expression at high levels and rendering strains highly susceptible to killing in a serum bactericidal assay [96]. In clinical studies, NadA formulated with fHbp and NHBA or in 4CMenB has been shown to induce high levels of bactericidal antibodies in infants, toddlers, adolescents and adults [82-86].

NHBA

NHBA is a *Neisseria*-specific surface-exposed lipoprotein with a predicted molecular weight of 50.5 kDa that binds heparin *in vitro*. This property correlates with increased survival of the unencapsulated bacterium in human serum [97]. Serum antibodies from mice immunized with recombinant NHBA elicited complement-mediated bactericidal activity against diverse MenB strains [70,71]. Anti-NHBA antibody also elicited deposition of human C3b on the bacterial surface and passively protected infant rats against bacteremia after challenge [98].

NHBA is present in all meningococcal strains tested [21,77,92,99] and is structured into a substantial number of peptides that have some association with clonal complexes and sequence types. While gene-sequence analysis of genetically diverse MenB strains reveals variable segments of NHBA, its amino- and carboxyl-terminal regions are highly conserved. 4CMenB contains NHBA peptide 2, which was shown to be the most common in a recent molecular epidemiology study (Figure 3) [95]. As with the other antigens included in 4CMenB, NHBA formulated with fHbp and NadA or in 4CMenB induced bactericidal antibodies in all age groups studied [82-86].

4CMenB: a multicomponent vaccine against meningococcal disease

To date, 4CMenB is the only broadly protective serogroup B vaccine that has completed Phase III trials and is under review by several regulatory authorities. The multicomponent approach might create the possibility of antigen cooperativity or synergy that could augment protection provided on the individual contributions of each vaccine component. Cooperative serum bactericidal activity between human antibodies raised against fHbp and NHBA has been detected [100]. Functional data on these antigens suggest that antibodies induced by 4CMenB could act in two ways: directly, by activating classical complement pathway, or indirectly, by interfering with adhesion and colonization and/or preventing fH binding on the bacterial surface, increasing susceptibility to killing by the alternative pathway.

Early clinical development

Evaluating novel vaccines requires extensive planning efforts, which can include the development or adaptation of end point measures for use in clinical trials. In disease states that occur at a relatively low incidence, such as meningococcal disease, clinical development programs must employ serological measures that correlate to clinical protection. For meningococcal disease, the original accepted correlate of protection, a titer ≥ 4 in the serum bactericidal assay using human complement (hSBA) was established by Goldschneider et al. and has subsequently been used widely in clinical trials, including studies of wtOMV vaccines [101,102]. Studies of 4CMenB have employed hSBA titers ≥ 4 , ≥ 5 (which ensures that the lower bound of a 95% CI is \geq 4), \geq 8 (a more conservative measure) and fourfold rises in hSBA titer above pre-existing protective levels of antibody as measures of immunogenicity. ELISAs for antigenspecific antibody have also been obtained in some studies [82-86]. Although such measures would be considered sufficient for polysaccharide vaccines, conjugate vaccines and wtOMV vaccines, assessments of broad strain coverage by 4CMenB required additional measures to bridge clinical trial data to circulating strain epidemiology, as discussed below [58].

Early clinical studies of vaccine formulations containing fHbp, NadA and NHBA evaluated immunogenicity against strain panels that were selected in part to assess and identify possible reference strains for use in Phase III studies to evaluate the individual contribution of different vaccine antigens (Tables 4 &

Table 4. Early clinical results with various formulations containing fHbp, NadA and NHBA.					
			Percentage of vaccinees with hSBA titers ≥4		
Strain	Phenotype	Sequence type	4CMenB	rMenB with Norway strain OMV	rMenB alone
5/99	B:2b:P1.5,2	1349	100	100	100
2996	B:2b:P1.5,2	540	>80+	>70	>50
M6190	B:2b:P1.5,2	1988	-	_	-
M01240013	B:2b:P1.5,2	11	>70	>60	>70
95N477	B:2b:P1.2	475	-	_	-
44/76	B:15:P1.7,16	32	>90	100	100
MC58	B:15:P1.17,16b	74	100	100	100
CU385	B:4:P1.15	33	100	100	100
M4105	B:4,7:P1.7,4	154	100+	>50	-
NZ98/254	B:4:P1.7-2,4	154	>90+	>70	>50
M1390	B:15:P1.7,4	41	>90	>90	>90
1000	B:NT:P1.5	20	>80	>70	>50
M4458	B:NT:P1.3	6161	70	>70	>75
M01240364	B:NT:P1.22,9	275	100	>90	>90
M3812	B:NT:P1.5	60	>70	70	>80
1					

'Strains for which a possible immunogenic advantage was identified for 4CMenB relative to rMenB or rMenB with a different OMV component. Of note, both formulations with an OMV showed similar effects against the Norway strain 44/77.

Dashes indicate that fewer than half of participants generated protective hSBA titers

OMV: Outer membrane vesicle

5) [73,84-86].

In a Phase I study in 70 healthy adults, Toneatto et al. observed that 4CMenB provided good evidence of immunogenicity against a panel of 15 genetically heterologous MenB strains, including three of the four strains later chosen for evaluation in Phase III studies (Table 5) [84]. Compared with rMenB alone or formulated with OMV from the Norwegian outbreak strain, 4CMenB induced protective hSBA titers against more strains, and also provided crossprotection in exceess of that expected based on PorA serosubtypes. For example, both vaccines seemed to induce similarly protective effects against strain 44/76, the source for the Norwegian OMV component. This finding may indicate the activity of another protein in the New Zealand strain OMV, synergystic effects among vaccine antigens, or crossprotection afforded by PorA 1.4 [84]. In the case of strain 44/76, the primary 4CMenB

immunogen contributing to killing in the hSBA was later found to be fHbp [73].

Two Phase II studies of 4CMenB were conducted in healthy infants, whose sera were tested in the hSBA against a six-strain panel, which included three of the four strains later chosen for evaluation in Phase III studies [15,16,82,83]. In these studies, infants received 4CMenB or rMenB at either 2, 4 and 6 months of age or at 6-8 months of age and 2 months later. All infants received a booster dose at 12 months of age. Infants enrolled in these studies were found to mount robust immune responses to genetically heterologous MenB strains. However, these antibody responses tracked closely to strains whose surface antigens were closely related to the vaccine variants. In other words, responses to genetically heterologous strains appeared to have resulted from the activity of multiple components as opposed to cross-protection afforded by individual

Table 5. Meningococcal serogroup B strains used for Phase III studies of 4CMenB.					
Strain	Phenotype	Sequence type	PorA type	hSBA killing by antibodies against	
NZ98/254	B:4:P1.7-2,4	42	P1.4	PorA	
44/76-SL	B:15:P1.7,16	32	P1.16	fHbp	
5/99	B:2b:P1.5,2	8	P1.2	NadA	
M10713	B:NT:P1.17,16-3	136	P1.16-3	NHBA	

components. Thus, the induction of protective antibodies was not evident against strains that expressed only antigen variants that were distantly related to the vaccine components. These findings were further supported by *in vitro* assessment of a genetically engineered meningococcal strain that expressed multiple fHbp variant 1 subvariants. In this study, post-vaccination sera from 7-month-old infants covered only closely related fHbp variants on the recombinant strain. However, the same sera were capable of killing wt strains harboring distantly related fHbp variant 1 subvariants, which was considered likely due to the contribution of the other components in the vaccine [87].

The Phase II studies in infants also include an evaluation of safety and tolerability parameters. The infants aged 2, 4 and 6 months of age at study immunization also received Pediacel[®] (diphtheria, tetanus, acellular pertussis, inactivated poliovirus, Haemophilus influenzae type B conjugate vaccine; Sanofi Pasteur) and Prevnar (7-valent pneumococcal conjugate vaccine; Pfizer) [82]. In the study in older infants, Menitorix[®] (H. influenzae type B, N. meningitidis group C polysaccharide conjugate vaccine; GlaxoSmithKline) was concomitantly administered at the 12-month booster vaccination with 4CMenB [83]. The concomitant use of 4CMenB with these routinely used vaccines did not markedly affect immunogenicity or tolerability outcomes for any of the vaccines. In both studies, researchers observed that all vaccine regimens were generally well tolerated by the enrolled infants and that no unexpected side effects occurred [82,83].

Selection of reference strains for Phase IIb & III studies

The late-stage clinical development program for 4CMenB was designed to provide a means of evaluating the individual immunogenic contributions of each major vaccine component and then to bridge these findings against strain epidemiology to predict the likelihood that 4CMenB would provide adequate strain coverage to limit endemic MenB disease in a given area or population. Reference strains were chosen to evaluate the individual ability of the major antigenic protein in each 4CMenB component to induce bactericidal antibodies. Therefore, reference strains were selected because they strongly express only one 4CMenB antigen and either lack the gene, have very low surface expression of, or express a mismatched variant of the other antigens (Table 5). Researchers genetically characterized a large panel of MenB strains and then identified strains that are killed by antibodies against a single 4CMenB antigen using a competitive inhibition serum bactericidal assay [73]. The reference strains for fHbp, NadA and PorA P1.4 (the immunodominant protein in the

OMV) have been described [73]. Of note, although an initial NHBA strain was proposed, logistical difficulties prevented its use in the hSBA and strain M10713 was later selected as the NHBA reference strain for pediatric studies [86].

Late-stage trials

Data from a large-scale study in 1885 healthy infants indicated a promising immunogenicity and safety profile for 4CMenB when administered at 2, 3 and 4, or 2, 4 and 6 months of age [85]. Prevnar[®] (7-valent pneumococcal conjugate vaccine) and Infanrix[®] hexa (diphtheria, tetanus, acellular pertussis, inactivated poliovirus, hepatitis B and H. influenzae type B combination vaccine) were administered to infants either with 4CMenB or alone. One schedule alternated routine vaccines with 4CMenB at separate study visits. Based on hSBA titers ≥5 against a panel of MenB reference strains, 4CMenB was immunogenic in all dosing schedules. No evidence of clinical interference was observed for the comparator vaccines with concomitant administration of 4CMenB [85]. Results of the booster dose in this study, as well as the results of a large-scale safety trial [103-105] and a toddler 'catch-up' schedule, are pending publication. Specific outcomes such as fever, which are commonly associated with wtOMV vaccines, warrant further consideration across the large dataset collected in these trials.

In adolescents, 4CMenB induced robust protective-antibody responses in the vast majority of vaccines, with acceptable safety, using various dosing schedules. Overall, 92–97% of subjects had hSBA titres ≥4 against a panel of three MenB reference strains after one dose of 4CMenB, as did 99–100% of those who received two or three doses. Evidence of waning of antibodies in persons without pre-existing titers against MenB strains was observed after a single dose, but not after two doses of 4CMenB [86].

In order to assess immunogenicity in adults (aged 18-50 years) who have increased exposure to N. meningitidis relative to the general population, a trial in laboratory workers was conducted at one center in Italy and one center in Germany (n = 54), 4CMenB was administered at 0, 2 and 6 months. Although baseline titres were high, as might be expected in such a population, fourfold rises in hSBA titres against a panel of MenB reference strains were observed in 64-88% of subjects after one dose, and 69-100% after three doses. A follow-up vaccination with conjugate quadrivalent meningococcal vaccine was also immunogenic. Pain was reported following 4CMenB administration by every participant, consistent with previous reports of pain following OMV vaccines in this age group. Three participants experienced transient fever [106]. Additional clinical studies of 4CMenB are ongoing.

Strain coverage

Numerous genetically diverse MenB strains cause IMD each year; therefore, assessments consider protective antibodies for all circulating strains. However, performing hSBA against all circulating strains poses logistical and ethical hurdles, particularly in infants, as large volumes of sera would be required. Strain coverage could be predicted by examining antigens on circulating strains. Since established typing recommendations, such as multi-locus sequence typing, do not account for fHbp, NadA and NHBA, a new system was developed [107].

The meningococcal antigen typing system (MATS) was developed to assess the expression, degree of crossreactivity and antigenicity of fHbp, NadA, and NHBA in meningococcal strains and to estimate strain coverage based on those findings in conjunction with the PorA serosubtype of the strains [58]. The use of the MATS method to predict MenB global and national strain coverage by 4CMenB is underway.

In MATS, the genotype of the variable region 2 of PorA is determined using conventional PCR. For fHbp, NadA and NHBA, a sandwich ELISA is used to test expression and antigenicity. Strains are considered covered by 4CMenB if the relative potency of fHbp, NadA or NHBA in the MATS ELISA is above a value that predicts killing in the hSBA. This value, the positive bactericidal threshold, conservatively estimates the minimum level of expression and antigenicity for fHbp, NadA or NHBA above which at least 80% of strains will be killed in the hSBA. Any strain with a relative





potency above the positive bactericidal threshold for fHbp, NadA or NHBA or a strain expressing PorA P1.4 is considered covered by 4CMenB.

The initial MATS predictions were confirmed by hSBA testing against large strain panels designed to over-represent strains that did not strongly express the vaccine antigens. Confirmatory hSBA testing showed that MATS predictions were conservative for toddlers, adolescents and adults because a substantial proportion of strains predicted not to be covered were in fact killed according to the hSBA. Very few strains predicted to be killed by MATS survived confirmatory hSBA testing, showing that MATS had a high positive predictive value for strain killing and a moderate negative predictive value for strains that would not be killed in the hSBA (Figure 4) [58]. Additional work to confirm whether MATS results are conservative against epidemiologically representative strain panels is pending publication.

The MATS ELISA has been transferred to national reference laboratories in Europe, North America and Australia and may be transferred to other laboratories as well. MATS testing of MenB strain panels collected during the last full defined epidemiological year(s) for which strain collections were available have been under-taken. Based on presentations at scientific meetings, the overall conservatively predicted coverage of 4CMenB is between 70 and 90% for individual countries.

The antigens in 4CMenB can occur in all serogroups [69,81,90]. Most circulating serogroup C, Y and W-135 strains express or have the genes coding for fHbp (99%) and NHBA (>90%), and a majority of serogroup C strains also express NadA [81,90]. In the USA, approximately half of strains with fHbp express variant 1, and most of these harbor subvariant 1.1, which is included in 4CMenB. Many different subvariants of NadA, NHBA and fHbp were detected in circulating strains [90]. Assessment of 4CMenB coverage in additional serogroups is also of interest.

Expert commentary

Conjugate vaccines against meningococcal serogroups A, C, W-135 and Y are commonly used and becoming more widely available. One rate-limited factor in the implementation of these vaccines in the developing world was expense, thus, with the advent of the new low-cost vaccine MenAfriVac^{*} (Meningitis Vaccine Project), to prevent disease caused by serogroup A in the African 'meningitis belt', vaccination against MenB has become the most important public health need for IMD. Since MenB disproportionately affects infants in the developed world, new vaccines should be designed to be included in existing routine immunization programs. Various investigational vaccines have been

proposed and one, 4CMenB, has completed Phase III trials and is being considered for approval in Europe and elsewhere.

The genetic diversity and mutability of *N. meningitidis* relies on various mechanisms, including horizontal gene transfer, for all meningococcal serogroups. The dynamic epidemiology of this organism poses special challenges for MenB, which lacks a universal antigen. The necessity for using subcapsular proteins is therefore well established. The multicomponent strategy is promising because it can address genetic diversity and the potential for future mutations. 4CMenB was developed to provide protection against circulating strains over time by employing surface antigens that are immunogenic and conserved across pathogenic and carriage isolates. Additional characterization of MenB strains over time and across geographic regions is ongoing.

Clinical studies of 4CMenB demonstrated protective antibody levels with acceptable tolerability outcomes in persons as young as 2 months of age. In adults, two- and three-dose schedules generally provided similar outcomes. Data are needed to assess persistence over time and the publication of the major datasets for Phase III is pending. Published safety findings indicate that 4CMenB outcomes were generally similar to those in studies of the OMV component alone. Studies in infants also indicate robust immune effects with a primary series and a booster dose, and tolerability that was comparable to published data describing the OMV vaccine in that age group. The most common solicited reactions to 4CMenB in adolescents and adults were injection site pain and swelling, malaise and headache while infants and toddlers experienced tenderness and erythema at the injection site, fever and irritability. These reactions were generally transient and self-limiting, most commonly resolving within 48 h of onset (Table 6).

The assessment of strain coverage is of special interest for MenB. The MATS methodology, which accounts for genetic expression and antigenicity, may prove to be a valuable tool for assessing strain protection and surveillance efforts. Where it has been implemented, MenAfriVac is having a dramatic impact on serogroup A IMD and is revealing the impact of serogroup W-135 and X disease. Although these serogroups cause only a fraction of MenA reports before MenAfriVac was introduced, disease incidence warrants additional efforts to develop protective vaccines. Perhaps with the existing complement of quadrivalent conjugate vaccines, new lowcost vaccines against serogroups A, X and W-135 may become available. With rises in serogroup Y disease reported in some regions that currently recommend only serogroup C vaccines, quadrivalent vaccines may become more widely used in developed nations as well.

The substantial body of available data describing the effects of 4CMenB is currently being evaluated by the EMA and other regulatory agencies. If introduced into national immunization programs, 4CMenB could markedly reduce MenB-related morbidity and mortality. Post-licensure studies will be crucial to help define 4CMenB safety and effectiveness, the extent of herd protection, persistence of antibodies, need for a booster, degree of strain coverage and potential coverage of nonserogroup B meningococcal strains.

It is likely that, in addition to the usual immunogenicity and safety studies required to assess recently licensed products, ongoing epidemiologic surveillance of circulating strains using MATS will be required to monitor MenB. Regional epidemiology and surveillance will be important to address the possibility for strain replacement or serogroup replacement, which has been reported with pneumococcal vaccines. Possible influence on commensal species, such as N. lactamica, should also be considered. Effective databases that can track molecular epidemiology of IMD and nasopharyngeal flora worldwide are needed. Of particular interest is new information about the surface features of pathogenic and carriage meningococcal isolates, which could prove important in future public health decision making. Development of additional combination and multicomponent vaccines could draw on more comprehensive information on the characterization of pathogenic meningococcal strains, regardless of capsular serogroup.

5-year view

Table 6. Solicited reactogenicity outcomes.						
Study	n	Age	Local reactogenicity rate (mainly erythema and pain) (%)	Systemic reactogenicity rate (%)	Ref.	
Snape <i>et al</i> . (2010)	60	Infants	10–26	1–18	[83]	
Findlow et al. (2010)	147	Infants	20–100	12	[82]	
Kimura <i>et al</i> . (2011)	54	Adults	40–100	10-50	[106]	
Santolaya <i>et al</i> . (2012)	1631	Adolescents	40-85	5–50	[86]	
Gossger <i>et al</i> . (2012)	1885	Infants	10–69	1–79	[85]	

Future perspective

We anticipate that much of the work initiated to develop 4CMenB – the first broad coverage MenB vaccine to complete sufficient Phase III clinical trials – will likely contribute to the reduction of IMD worldwide. The multicomponent strategy, which accounts for multiple surface antigens and therefore provides varied opportunities to induce bactericidal antibodies, could prove a vital addition to the public-health armamentarium. In addition, the MATS method could provide a means

Executive summary

Background

- The development of vaccines against *Neisseria meningitidis* serogroup B (MenB) has been a global public-health priority based on continuing disease burden combined with the scientific challenges associated with antigen identification.
- Several factors inform the rationale for establishing immunization programs against MenB, including disease characteristics, epidemiology and persistent case fatality rates.
- The poor immunogenicity of the MenB capsular polysaccharide arises from its structural similarity to the polysialic acid in the fetal neural cell-adhesion molecule.

MenB vaccines

- Licensed wild-type outer membrane vesicle (OMV) vaccines (VA-MENGOC-BC[®], MenBVac[®] and MeNZB[®]) are protective against strains bearing the same PorA serosubtype.
- Further vaccine development includes the wild-type OMV vaccine combinations, recombinant OMVs containing multiple surface proteins and purified proteins.
- One multicomponent vaccine 4CMenB has completed late-stage clinical trials and is under consideration for licensure.
- 4CMenB contains four components: OMV from the New Zealand outbreak strain, fHbp, NadA and NHBA.

Reverse vaccinology: antigen identification for novel MenB vaccines

- Reverse vaccinology uses bioinformatic algorithms to 'mine' the genomic sequences for potential vaccine antigens.
- The first group of such targets was identified for MenB strain MC58 and led to the selection of purified proteins included in 4CMenB.

4CMenB: a multicomponent vaccine against meningococcal disease

- Clinical studies employed accepted correlates of protection and safety outcomes.
- Clinical data in infants, adolescents and adults support the immunogenicity of 4CMenB.
- Safety and tolerability findings for 4CMenB were promising in all studies and age groups.
- Strain coverage assessments will be based on a meningococcal antigen-typing system method that accounts for genotypic expression and antigenicity.

of future MenB surveillance and could be adapted for use with additional pathogens. Should 4CMenB be licensed, it will likely result in a reduction of disease burden where implemented, particularly in areas with a high or relatively high incidence of MenB disease. Further investigation of the possible effects of this vaccine against other meningococcal serogroups could yield data to support additional applications for this vaccine. Overall, we hope that in the next 5 years, the development of 4CMenB will have provided valuable information for the community working to eliminate meningococcal disease.

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References

 Sadarangani M, Pollard AJ. Serogroup B meningococcal vaccines – an unfinished story. *Lancet Infect. Dis.* 10(2), 112–124

(2010).

3

- Zollinger WD, Poolman JT, Maiden MC.
 Meningococcal serogroup B vaccines: will they live up to expectations? *Expert Rev. Vaccines* 10(5), 559–561 (2011).
- Holst J. Strategies for development of universal vaccines against meningococcal serogroup B disease: the most promising options and the challenges evaluating them. *Human Vacc.* 3(6), 290–294 (2007).
- 4 Ramsay ME, Andrews NJ, Trotter CL *et al.* Herd immunity from meningococcal serogroup C conjugate vaccination in England: database analysis. *BMJ* 326(7385), 365–366 (2003).
- 5 Halperin SA, Bettinger JA, Greenwood B et al. The changing and dynamic epidemiology of meningococcal disease. Vaccine doi:10.1016/j.vaccine.2011.12.032 (2011) (Epub ahead of print).
- 6 Harrison LH, Trotter CL, Ramsay ME. Global epidemiology of meningococcal disease. *Vaccine* 275, B51–B63 (2009).

- 7 Tan LK, Carlone GM, Borrow R. Advances in the development of vaccines against Neisseria meningitidis. N. Engl. J. Med. 362(16), 1511–1520 (2010).
- 8 Koroleva IS, Demina AA. The serosubtyping of serogroup B meningococci. Zh. Mikrobiol. Epidemiol. Immunobiol. 5, 41–44 (1995).
- 9 Khalil MK, Borrow R. Serogroup B meningococcal disease during Hajj: preparing for the worst scenario. *Travel. Med. Infect. Dis.* 7, 231–234 (2009).
- 10 Ceyhan M, Yildrim I, Balmer P et al. A prospective study of etiology of childhood acute bacterial meingitis, Turkey. Emerg. Infect. Dis. 14(7), 1089–1096 (2008).
- 11 Gil-Prieto R, García-García L, Alvaro-Meca A, González-Escalada A, Viguera Ester P, Gil De Miguel A. The burden of hospitalizations for meningococcal infection in Spain (1997–2008). Vaccine 29(34), 5765–5770 (2011).
- 12 Safadi MAP and Cintra OAL. Epidemiology of meningococcal disease in Latin America: current situation and opportunities for prevention. *Neurol. Res.* 32(3), 263–271 (2010).
- 13 Sow SO, Okoko BJ, Diallo A et al. Immunogenicity and safety of a meningococcal A conjugate vaccine in Africans. N. Engl. J. Med. 364(24), 2293–2304 (2011).
- 14 LaForce FM, Konde K, Viviani S, Préziosi MP. The Meningitis Vaccine Project. *Vaccine* 3(25 Suppl. 1), A97–A100 (2007).
- 15 Bai X, Findlow J, Borrow R. Recombinant protein meningococcal serogroup B vaccine combined with outer membrane vesicles. *Expert Opin. Biol. Ther.* 11(7), 969–985 (2011).
- 16 Su EL, Snape MD. A combination recombinant protein and outer membrane vesicle vaccine against serogroup B meningococcal disease. *Expert Rev. Vaccines* 10(5), 575–588 (2011).
- Panatto D, Amicizia D, Lai PL, Gasparini R. Neisseria meningitidis B vaccines. *Expert Rev. Vaccines* 10(9), 1337–1351 (2011).
- 18 Safadi MAP, McIntosh EDG. Epidemiology and prevention of meningococcal disease: a critical appraisal of vaccine policies. *Expert Rev. Vaccines* 10(12), 1717–1730 (2011).
- 19 Bruge J, Bouvert-Le Cam N, Danve B et al. Clinical evaluation of a group B meningococcal N-propionylated polysaccharide conjugate vaccine in adult, male volunteers. Vaccine 22(9–10), 1087–1096 (2004).

- 20 Seib KL, Serruto D, Oriente F *et al.* Factor H-binding protein is important for meningococcal survival in human whole blood and serum and in the presence of the antimicrobial peptide LL-37. *Infect. Immun.* 77(1), 292–299 (2009).
- 21 Jacobsson S, Thulin S, Mölling P *et al.* Sequence constancies and variations in genes encoding three new meningococcal vaccine candidate antigens. *Vaccine* 24(12), 2161–2168 (2006).
- 22 Stephens DS, Greenwood B, Brandtzaeg P. Epidemic meningitis, meningococcaemia, and *Neisseria meningitidis*. *Lancet* 269(9580), 2196–2210 (2007).
- 23 Gardner P. Clinical practice. Prevention of meningococcal disease. N. Engl. J. Med. 355(14), 1466–1473 (2006).
- Rosenstein NE, Perkins BA, Stephens DS et al. Meningococcal disease. N. Engl. J. Med. 344(18), 1378–1388 (2001).
- 25 Cohn AC, MacNeil JR, Harrison LH et al. Changes in Neisseria meningitidis disease epidemiology in the United States, 1998–2007: implications for prevention of meningococcal disease. Clin. Infect. Dis. 50, 184–191 (2010).
- 26 Pelkonen T, Roine I, Monteiro L et al. Acute childhood bacterial meningitis in Luanda, Angola. Scand. J. Infect. Dis. 40(11-12), 859-866 (2008).
- 27 De Gaudio

M, Chiappini E, Galli L M, De Martino M. Therapeutic management of bacterial meningitis in children: a systematic review and comparison of published guidelines from a European perspective. J. Chemother. 22(4), 226–237 (2010).

- 28 Bröker M, Cooper B, DeTora LM, Stoddard JJ. Critical appraisal of a quadrivalent CRM197 conjugate vaccine against meningococcal serogroups A, C, W-135, and Y (Menveo') in the context of treatment and prevention of invasive disease. *Infect. Drug Resist.* 4, 1–11 (2011).
- 29 Harrison LH, Pelton SI, Wilder-Smith A et al. The Global Meningococcal Initiative: recommendations for reducing the global burden of meningococcal disease. Vaccine 29, 3363–3371 (2011).
- 30 Gray SJ, Trotter CL, Ramway ME et al. Epidemiology of meningococcal disease in England and Wales 1993/94 to 2003/04: contribution and experiences of the Meningococcal Reference Unit. J. Med. Microbiol. 55(7), 887–896 (2006).
- 31 de Voer RM, Mollema L, Schepp RM et al. Immunity against Neisseria meningiditis serogroup C in the Dutch population before and after introduction of the meningococcal

C conjugate vaccine. *PLoS One* 5, e12144 (2010).

- 32 de Greeff SC, Spanjaard L, Dankert J et al. Underreporting of meningococcal disease incidence in The Netherlands: results from a capture-recapture analysis based on three registration sources with correction for false positive diagnoses. E. J. Epidemiol. 21, 315–321 (2006).
- 33 Bettinger JA, Scheifele DW, Le Saux N, Halperin SA, Vaudry W, Tsang R, and the members of the Canadian Immunization Monitoring Program, Active (IMPACT). The impact of childhood meningococcal serogroup C conjugate vaccine programs in Canada. *Pediatr. Infect. Dis. J.* 28, 220–224 (2009).
- 34 Memish ZA, Shibl AM. Consensus building and recommendations based on the available epidemiology of meningococcal disease in Gulf Cooperation Council States. *Travel Med. Infect. Dis.* 9(2), 60–66 (2011).
- Granoff DM. Review of meningococcal group B vaccines. *Clin. Infect. Dis.* 50(Suppl. 2), S54–S65 (2010).
- 36 Holst J, Martin D, Arnold R *et al.* Properties and clinical performance of vaccines containing outer membrane vesicles from *Neisseria meningitidis. Vaccine* 27S, B3–B12 (2009).
- 37 Tappero JW, Lagos R, Ballesteros AM et al. Immunogenicity of 2 serogroup B outermembrane protein meningococcal vaccines. A randomized controlled trial in Chile. JAMA 281(16), 1520–1527 (1999).
- 38 Sotolongo Padrón F, Campa H, Casanueva Gil V *et al.* Cuban meningococcal BC vaccine: experiences and contributions from 20 years of application. *MEDICC Rev.* 9, 16–22 (2007).
- 39 Granoff DM, Bartoloni A, Ricci S *et al.* Bactericidal monoclonal antibodies that define unique meningococcal B polysaccharide epitopes that do not cross-react with human polysialic acid. *J. Immunol.* 160(10), 5028–5036 (1998).
- 40 Rinaudo CD, Telford JL, Rappuoli R, Seib KL. Vaccinology in the genome era. J. Clin. Investig. 119(9), 2515–2525 (2009).
- 41 Urwin R *et al.* Distribution of surface protein variants among hyperinvasive meningococci: implications for vaccine design. *Infect. Immun.* 72(10), 5955–5962 (2004).
- 42 Frasch CE, van Alphen L, Holst J et al. Outer membrane protein vesicle vaccines for meningococcal disease. *Methods Mol. Med.* 66, 81–107 (2001).
- 43 Black S, Pizza M, Nissum M, Rappuoli R. Toward a meningitis-free world. *Sci. Transl. Med.* 4(123), 123ps5 (2012).

- Lewis S, Sadarangani M, Hoe JC, Pollard AJ. Challenges and progress in the development of a serogroup B meningococcal vaccine. *Expert Rev. Vaccines* 8(6), 729–745 (2009).
- 45 Holst J, Feiring B, Naess LM *et al.* The concept of 'tailor-made', protein-based, outer membrane vesicle vaccines against meningococcal disease. *Vaccine* 23, 2202–2205 (2005).
- 46 Martin DR, Ruijne N, McCallum L *et al.* The VR2 epitope on the PorA P1.7–2,4 protein is the major target for the immune response elicited by the strain-specific group B meningococcal vaccine MeNZB. *Clin. Vaccine Immunol.* 13(4), 486–491 (2006).
- 47 Thornton V, Lennon D, Rasanathan K et al. Safety and immunogenicity of New Zealand strain meningococcal serogroup B OMV vaccine in healthy adults: beginning of epidemic control. *Vaccine* 24(9), 1395–1400 (2006).
- 48 Galloway Y, Stehr-Green P, McNicholas A, O'Hallahan J. Use of an observational cohort study to estimate the effectiveness of the New Zealand group B meningococcal vaccine in children aged under 5 years. *Int. J. Epidemiol.* 38, 413–418 (2009).
- 49 Lennon D, Jackson C, Wong S et al. Fast tracking the vaccine licensure process to control an epidemic of serogroup B meningococcal disease in New Zealand. Clin. Infect. Dis. 49, 597–605 (2009).
- 50 Caron F, Du Châtelet IP, Leroy JP et al. From tailor-made to ready-to-wear meningococcal B vaccines: longitudinal study of a clonal meningococcal B outbreak. Lancet Infect. Dis. 11(6), 455–463 (2011).
- 51 Sandbu S, Feiring B, Oster P *et al.* Immunogenicity and safety of a combination of two serogroup B meningococcal outer membrane vesicle vaccines. *Clin. Vaccine Immunol.* 14(9), 1062–1069 (2007).
- 52 Wedege E, Bolstad K, Aase A *et al*. Functional and specific antibody responses in adult volunteers in new zealand who were given one of two different meningococcal serogroup B outer membrane vesicle vaccines. *Clin. Vaccine Immunol.* 14(7), 830–838 (2007).
- 53 Wong S, Lennon D, Jackson C *et al*. New Zealand epidemic strain meningococcal B outer membrane vesicle vaccine in children aged 16–24 months. *Pediatr. Infect. Dis. J.* 26(4), 345–350 (2007).
- 54 Oster P, O'Hallahan J, Aaberge I *et al.* Immunogenicity and safety of a strainspecific MenB OMV vaccine delivered to under 5-year olds in New Zealand. *Vaccine* 25(16), 3075–3079 (2007).
- 55 Nøkleby H, Aavitsland P, O'Hallahan J et al. Safety review: two outer membrane vesicle (OMV) vaccines against systemic Neisseria

meningitidis serogroup B disease. *Vaccine* 25(16), 3080–3084 (2007).

- 56 Jackson C, Lennon DR, Sotutu VT et al. Phase II meningococcal B vesicle vaccine trial in New Zealand infants. Arch. Dis. Child. 94, 745–751 (2009).
- 57 Thornton V, Lennon D, Rasanathan K et al. Safety and immunogenicity of New Zealand strain meningococcal serogroup B OMV vaccine in healthy adults: beginning of epidemic control. *Vaccine* 24(9), 1395–1400 (2006).
- 58 Donnelly J, Medini D, Boccadifuoco G et al. Qualitative and quantitative assessment of meningococcal antigens to evaluate the potential strain coverage of protein-based vaccines. Proc. Natl Acad. Sci. USA 107(45), 19490 –19495 (2010).
- 59 Tondella MLC, Popovic T, Rosenstein NE et al. Distribution of Neisseria meningitidis serogroup B serosubtypes and serotypes circulating in the United States. J. Clin. Microbiol. 38(9), 3323–3328 (2000).
- 60 van der Voort ER, van Dijken H, Kuipers B et al. Human B- and T-cell responses after immunization with a hexavalent PorA meningococcal outer membrane vesicle vaccine. *Infect. Immun.* 65, 5184–5190 (1997).
- 61 van den Dobbelsteen GP, van Dijken HH, Pillai S, van Alphen L. Immunogenicity of a combination vaccine containing pneumococcal conjugates and meningococcal PorA OMVs. *Vaccine* 25(13), 2491–2496 (2007).
- 62 Sanders H, Feavers IM. Adjuvant properties of meningococcal outer membrane vesicles and the use of adjuvants in *Neisseria meningitidis* protein vaccines. *Expert Rev. Vaccines* 10(3), 323–334 (2011).
- 63 Borrow R, Balmer P. The future of meningococcal vaccines. *Pediatric Health* 1(1), 51–61 (2007)
- 64 van de Waterbeemd B, Streefland M, van der Ley P *et al.* Improved OMV vaccine against Neisseria meningitidis using genetically engineered strains and a detergent-free purification process. *Vaccine* 28(30), 4810–4816 (2010).
- 65 Boutriau D, Poolman J, Borrow R et al. Immunogenicity and safety of three doses of a bivalent (B:4:p1.19,15 and B:4:p1.7–2,4) meningococcal outer membrane vesicle vaccine in healthy adolescents. Clin. Vaccine Immunol. 14(1), 65–73 (2007).
- 66 Keiser PB, Gibbs BT, Coster TS *et al.* A Phase 1 study of a group B meningococcal native outer membrane vesicle vaccine made from a strain with deleted *lpxL2* and *synX* and stable expression of *opcA. Vaccine* 228, 6970–6976 (2010).
- 67 Keiser PB, Biggs-Cicatelli S, Moran EE *et al.* A phase 1 study of a meningococcal native

outer membrane vesicle vaccine made from a group B strain with deleted *lpxL1* and *synX*, over-expressed factor H binding protein, two PorAs and stabilized OpcA expression. *Vaccine* 29(7), 1413–1420 (2011).

- 68 Gorringe AR, Taylor S, Brookes C et al. Phase I safety and immunogenicity study of a candidate meningococcal disease vaccine based on *Neisseria lactamica* outer membrane vesicles. *Clin. Vaccine Immunol.* 16(8), 1113–1120 (2009).
- 69 Beernink PT, Shaughnessy J, Ram S, Granoff DM. Impaired immunogenicity of a meningococcal factor H-binding protein vaccine engineered to eliminate factor H binding. *Clin. Vaccine Immunol.* 17(7), 1074–1078 (2010).
- 70 Pizza M, Scarlato V, Masignani V et al. Identification of vaccine candidates against serogroup B meningococcus by wholegenome sequencing. *Science* 287(5459), 1816–1820 (2000).
- 71 Giuliani MM, Adu-Bobie J, Commanducci M et al. A universal vaccine for serogroup B meningococcus. Proc. Natl Acad. Sci. USA 103(29), 10834–10839 (2006).
- 72 Capecchi B, Adu-Bobie J, Di Marcello F et al. Neisseria meningitidis NadA is a new invasion which promotes bacterial adhesion to and penetration into human epithelial cells. Mol. Micro. 55(3), 687–698 (2005).
- 73 Giuliani MM, Biolchi A, Serruto D et al. Measuring antigen-specific bactericidal responses to a multicomponent vaccine against serogroup B meningococcus. Vaccine 28, 5023–5030 (2010).
- 74 Madico G, Welsch J, Lewis LA *et al*. The meningococcal vaccine candidate GNA1870 binds the complement regulatory protein factor H and enhances serum resistance. *J. Immunol.* 177(1), 501–510 (2006).
- 75 Schneider MC, Exley RM, Chan H et al. Functional significance of factor H binding to Neisseria meningitidis. J. Immunol. 176(12), 7566–7575 (2006).
- 76 Masignani V, Comanducci M, Giuliani MM et al. Vaccination against Neisseria meningitidis using three variants of the lipoprotein GNA1870. J. Exp. Med. 197(6), 789–799 (2003).
- Bambini S, Muzzi A, Olcen P *et al.* Distribution and genetic variability of three vaccine components in a panel of strains representative of the diversity of serogroup B meningococcus. *Vaccine* 11(21), 2794–2803 (2009).
- 78 Lucidarme J, Tan L, Exley RM *et al.* Characterization of *Neisseria meningitidis* isolates that do not express the virulence factor and vaccine antigen factor H binding protein. *Clin. Vaccine Immunol.* 18(6), 1002–1014 (2011).
- 79 Lewis LA, Ngampasutadol J, Wallace R et al.

The meningococcal vaccine candidate neisserial surface protein A (NspA) binds to factor H and enhances meningococcal resistance to complement. *PLoS Pathog.* 6(7), e1001027 (2010).

- 80 Fletcher LD, Bernfield L, Barniak V et al. Vaccine potential of the Neisseria meningitidis 2086 lipoprotein. Infect. Immun. 72(4), 2088–2100 (2004).
- 81 Harris SL, Zhu D, Murphy E et al. Preclinical evidence for the potential of a bivalent fHbp vaccine to prevent Neisseria meningitidis serogroup C disease. Hum. Vaccin. 7, 68 –74 (2011).
- 82 Findlow J, Borrow R, Snape MD et al. Multicenter, open-label, randomized Phase II controlled trial of an investigational recombinant Meningococcal serogroup B vaccine with and without outer membrane vesicles, administered in infancy. Clin. Infect. Dis. 51(10), 1127–1137 (2010).
- 83 Snape MD, Dawson T, Oster P *et al.* Immunogenicity of two investigational serogroup B meningococcal vaccines in the first year of life. *Pediatr. Infect. Dis. J.* 29, e71–e79 (2010).
- 84 Toneatto D, Ismaili S, Ypma E *et al.* The first use of an investigational multicomponent meningococcal serogroup B vaccine (4CMenB) in humans. *Human Vacc.* 7(6), 646–653 (2011).
- 85 Gossger N, Snape MD, Yu LM *et al.* Immunogenicity and tolerability of recombinant serogroup B meningococcal vaccine administered with or without routine infant vaccinations according to different immunization schedules: a randomized controlled trial. *JAMA* 307(6), 573–582 (2012).
- 86 Santolaya ME, O'Ryan ML, Valenzuela MT et al. Immunogenicity and tolerability of a multicomponent meningococcal serogroup B (4CMenB) vaccine in healthy adolescents in Chile: a Phase 2b/3 randomised, observer-blind, placebo-controlled study. Lancet 379(9816), 617–624. (2012).
- Brunelli B, Del Tordello E, Palumbo E et al. Influence of sequence variability on bactericidal activity sera induced by Factor H binding protein variant 1.1. Vaccine 29, 1072–1081 (2011).
- 88 Comanducci M, Bambini S, Brunelli B et al. NadA, a novel vaccine candidate of Neisseria meningitidis. J. Exp. Med. 195(11), 1445–1454 (2002).
- 89 Nägele V, Heesemann J, Schielke S *et al.* Neisseria meningitidis adhesin NadA targets β 1 integrins: functional similarity to Yersinia invasin. J. Biol. Chem. 286(23), 20536–20546 (2011).
- 90 Wang X, Cohn A, Comanducci M et al. Prevalence and genetic diversity of candidate vaccine antigens among invasive Neisseria meningitidis isolates in the

United States. *Vaccine* 29(29–30), 4739–4744 (2011).

- 91 Comanducci M, Bambini S, Caugant DA et al. NadA diversity and carriage in Neisseria meningitidis. Infect. Immun. 72(7), 4217–4223 (2004).
- 92 Lucidarme J, Comanducci M, Findlow J et al. Characterization of fHbp, nhba (gna2132), nadA, porA, and sequence type in group B meningococcal case isolates collected in England and Wales during January 2008 and potential coverage of an investigational group B meningococcal vaccine. Clin. Vaccine Immunol. 17(6), 919–929 (2010).
- 93 Martin P, Sun L, Hood HW, Moxon ER. Involvement of genes of genome maintenance in the regulation phase variation frequencies in *Neisseria meningitidis*. *Microbiology* 150(Pt 9), 3001–3012 (2004).
- 94 Metruccio MM, Pigozzi E, Roncarati D et al. A novel phase variation mechanism in the meningococcus driven by a ligandresponsive repressor and differential spacing of distal promoter elements. PLoS Pathog. 5(12), e1000710 (2009).
- 95 Bambini S, Muzzi A, Comandi S et al. Variation of the Neisseria heparin binding antigen of meningococcus. Presented at: Annual Meeting of the European Monitorian Group for Meningococci (EMGM). Ljubljana, Slovenia, 18–20 May 2011.
- 96 Biolchi A, Fagnocchi L, Pigozzi E et al. In vitro levels of NadA expression may underestimate the potential effectiveness of immune responses against nadA in vivo. Presented at: Annual Meeting of the European Monitorian Group for Meningococci (EMGM). Ljubljana, Slovenia, 18–20 May 2011.
- 97 Serruto D, Spadafina T, Ciucchi L *et al. Neisseria meningitidis* GNA2132, a heparin-binding protein that induces protective immunity in humans. *Proc. Natl Acad. Sci. USA* 107, 3880–3775 (2010).
- 98 Welsch JA, Moe GR, Rossi R et al. Antibody to genome-derived Neisserial antigen 2132, a *Neisseria meningitidis* candidate vaccine, confers protection against bacteremia in the absence of complement-mediated bactericidal activity. J. Infect. Dis. 188(11), 1730–1740 (2003).
- 99 Lucidarme J, Comanducci M, Findlow J et al. Characterization of fHbp, nhba (gna2132), nadA, porA, sequence type (ST), and genomic presence of IS1301 in group B meningococcal ST269 clonal complex isolates from England and Wales. J. Clin. Microbiol. 47(11), 3577–3585 (2009).
- 100 Vu DM, Wong TT, Granoff DM. Cooperative serum bactericidal activity between human antibodies to meningococcal factor H

binding protein and Neisserial heparin binding antigen. *Vaccine* 29(10), 1968–1973 (2011).

- 101 Frasch CE, Borrow R, Donnelly J. Bactericidal antibody is the immunologic surrogate of protection against meningococcal disease. *Vaccine* 27(Suppl. 2), B112–B116 (2009).
- 102 Borrow R, Carlone GM, Rosenstein N et al. Neisseria meningitidis group B correlates of protection and assay standardization: international meeting report Emory University, Atlanta, Georgia, United States, 16–17 March 2005. Vaccine 24(24), 5093–5107 (2006).
- Esposito S, Vesikari T, Kimura A et al.
 Tolerability of a three-dose schedule of an investigational, multicomponent, meningococcal serogroup B vaccine and routine infant vaccines in a lot consistency trial. Presented at: 17th International Pathogenic Neisseria Conference. Banff, Alberta, Canada, 11–16 September 2010.
- 104 Vesikari T, Esposito S, Kimura A et al. Immunogenicity of an investigational, multicomponent, meningococcal serogroup B vaccine in healthy infants at 2, 4, and 6 months of age. Presented at: 17th International Pathogenic Neisseria Conference. Banff, Alberta, Canada, 11–16 September 2010.
- 105 Vesikari T, Esposito S, Prymula R *et al.* Use of an investigational multicomponent meningococcal serogroup B vaccine (4CMenB) in a clinical trial in 3630 infants. *Arch. Dis. Child.* 96(3), 11–16 (2011).
- 106 Kimura A, Toneatto D, Kleinschmidt A et al. Immunogenicity and safety of a multicomponent meningococcal serogroup B vaccine and a quadrivalent meningococcal CRM₁₉₇ conjugate vaccine against serogroups A, C, W-135, and Y in adults who are at increased risk for occupational exposure to meningococcal isolates. *Clin. Vaccine Immunol.* 18(3), 483–486 (2011).
- 107 Maiden MCJ, Bygraves JA, Feil E et al. Multilocus sequence typing: a portable approach to the identification of clones within populations of pathogenic microorganisms. Proc. Natl Acad. Sci. USA 95, 3140–3145 (1998).
- Websites
- 201 NICE. Bacterial meningitis and meningococcal septicemia: management of bacterial meningitis and meningococcal septicaemia in children and young people younger than 16 years in primary and secondary care.
 www.nice.org.uk/nicemedia/ live/13027/49339/49339.pdf (Accessed 6 June 2011)
- 202 Netherlands Vaccine Institute. Scientific report, 2006. www.nvi-vaccin.com/Publications (Accessed 10 June 2011)