

# OUTLINE OF FUNGI

*Outlineoffungi.org is a website dedicated to the taxonomy and classification of the Fungi.*

## Outlineoffungi.org – Note 8 (cut-off date 31.12.2020)

**Makbule Erdoğdu**, Department of Landscape Architects, Faculty of Agriculture, Kırşehir Ahi Evran University, Kırşehir, Turkey.

### **Shiva Prakash Nedle**

ICAR-Central Plantation Crops Research Institute (ICAR-CPCRI), Regional Station, Vittal - 574243, Karnataka, India.

### **Josiane S. Monteiro**

Coordenação de Botânica, Museu Paraense Emílio Goeldi, Avenida Perimetral, 1901, Terra Firme, 66077-530, Belém, Pará, Brazil.

### **Andrei Tsurykau**

F. Skorina Gomel State University, Department of Biology, Sovetskaja Str. 104, Gomel 246019, Belarus. Samara National Research University, Institute of Natural Sciences, Department of Ecology, Botany and Nature Protection, Moskovskoye shosse 34, Samara 443086, Russia.

Edited by **Ramesh K. Saxena, Kevin D. Hyde, Andre Aptroot, and Irina S. Druzhinina**

### **Abbreviations:**

*benA* – nuclear gene encoding  $\beta$ -tubulin protein

*cal* – nuclear gene encoding calmodulin protein

*cytB* – mitochondrial gene encoding cytochrome B

IGR – nuclear intergenic region of the rRNA gene cluster

ITS – nuclear internal transcribed spacers 1 and 2 of the rRNA gene cluster, includes 5.8S rRNA gene

LSU – nuclear large subunit, a gene encoding 28S rRNA

mtSSU – mitochondrial small subunit of the rRNA gene cluster

*rpb1* – nuclear gene encoding RNA polymerase subunit B I of RPB1 protein

*rpb2* – nuclear gene encoding RNA polymerase subunit B II of RPB2 protein

rRNA – ribosomal RNA

SSU – nuclear small subunit, a gene encoding 18S rRNA

*tub* – nuclear gene encoding  $\beta$ -tubulin protein

	<b>Statistics</b>
<b>Orders</b>	<b>11</b>
<b>Families</b>	<b>13</b>
<b>Genera</b>	<b>91</b>

## Table of Contents

<b>Orders.....</b>	<b>4</b>
<i>Aulographales</i> Crous, Spatafora, Haridas & I.V. Grig.....	4
<i>Aureoconidiellales</i> Hern.-Restr. & Crous.....	4
<i>Catabotryales</i> K.D. Hyde & Senan.....	4
<i>Cladosporiales</i> Abdollahz. & Crous.....	4
<i>Comminutisporales</i> Abdollahz. & Crous.....	4
<i>Coniosporiales</i> Crous, Spatafora, Haridas & I.V. Grig.....	4
<i>Heitmaniales</i> Q.M. Wang & F.Y. Bai.....	4
<i>Lineolatales</i> Crous, Spatafora, Haridas & I.V. Grig.....	4
<i>Neophaeothecales</i> Abdollahz. & Crous.....	5
<i>Racodiales</i> Abdollahz. & Crous.....	5
<i>Rosetozymales</i> Q.M. Wang & F.Y. Bai.....	5
<b>Families.....</b>	<b>6</b>
<i>Aureoconidiellaceae</i> Hern.-Restr. & Crous.....	6
<i>Coniosporiaceae</i> Crous, Spatafora, Haridas & I.V. Grig.....	6
<i>Cylindrosympodiaceae</i> Crous, M. Shen & Y. Zhang ter.....	6
<i>Foliocryphiaceae</i> C.M. Tian, N. Jiang & Crous.....	6
<i>Mastigosporiaceae</i> C.M. Tian, N. Jiang &.....	6
<i>Neocryphonectria</i> C.M. Tian, N. Jiang & Crous.....	6
<i>Heitmaniaceae</i> Q.M. Wang & F.Y. Bai.....	7
<i>Jianyuniaceae</i> Q.M. Wang & F.Y. Bai.....	7
<i>Lineolataceae</i> Crous, Spatafora, Haridas & I.V. Grig.....	7
<i>Neoantennariellaceae</i> Abdollahz. & Crous.....	7
<i>Neophaeothecaceae</i> Abdollahz. & Crous.....	7
<i>Readeriellipsoidaceae</i> Abdollahz. & Crous.....	7
<i>Rhizodiscinaceae</i> Crous, Spatafora, Haridas & I.V. Grig.....	7
<i>Rosetozymaceae</i> Q.M. Wang & F.Y. Bai.....	7
<i>Rubikioideae</i> Lücking, M. Cáceres & Aptroot.....	8
<i>Tricladiaceae</i> P.R. Johnst. & Baschien.....	8
<b>Genera.....</b>	<b>9</b>
<i>Acidotalaromyces</i> Houbraken, Frisvad & Samson.....	9
<i>Aphanodesmium</i> Rėblová & Hern.-Restr.....	9
<i>Arboricolonus</i> S. Bien & Damm.....	9
<i>Ascospirella</i> Houbraken, Frisvad & Samson.....	9
<i>Atrophysma</i> T. Sprib.....	9
<i>Aureoconidiella</i> Hern.-Restr. & Crous.....	10
<i>Baidera</i> Ertz & Diederich.....	10
<i>Batnamyces</i> Noumeur.....	10
<i>Begerowomyces</i> Q.M. Wang & F.Y. Bai.....	10
<i>Bellamyces</i> Crous, Coppins & U. Braun.....	10
<i>Bergerella</i> Diederich & Lawrey.....	10
<i>Boekhoutia</i> Q.M. Wang & F.Y. Bai.....	11
<i>Bolbea</i> Buaya & Thines.....	11
<i>Bryoclavula</i> H. Masumoto & Y. Degawa.....	11
<i>Burrowsia</i> Fryday & I. Medeiros.....	11
<i>Carneothele</i> Fryday, T. Sprib. & M. Svenss.....	11
<i>Cladocillium</i> Chun-Hao Chen & R. Kirschner.....	11
<i>Commelinaceomyces</i> E. Tanaka.....	12
<i>Corylicola</i> Wijesinghe, Camporesi, Yong Wang bis & K.D. Hyde.....	12
<i>Cryphognomonina</i> C.M. Tian & N. Jiang.....	12
<i>Diabolocovidia</i> Crous.....	12
<i>Emmanuelia</i> Ant. Simon, Lücking & Goffinet.....	12
<i>Erichansenia</i> S.Y. Kondr., Kärnefelt & A. Thell.....	12
<i>Evansstolkia</i> Houbraken, Frisvad & Samson.....	13
<i>Fagicola</i> Crous, M. Shen & Y. Zhang ter.....	13
<i>Fraxinicola</i> Crous, M. Shen & Y. Zhang ter.....	13
<i>Fulgogasparrea</i> S.Y. Kondr., N.-H. Jeong, Kärnefelt, Elix, A. Thell & Hur.....	13
<i>Fuscohilum</i> Crous, M. Shen & Y. Zhang ter.....	13
<i>Gamsomyces</i> Hern.-Restr. & Rėblová.....	13
<i>Halocryptosphaeria</i> Dayarathne, Devadatha, V.V. Sarma & K.D. Hyde.....	14
<i>Halotestudina</i> Dayarathne & K.D. Hyde.....	14
<i>Italiofungus</i> Crous.....	14
<i>Jennwenomyces</i> Goh & C.H. Kuo.....	14
<i>Jocatoa</i> R. Miranda.....	15
<i>Juncomyces</i> Crous.....	15

<i>Kaseifertia</i> Réblová & Hern.-Restr. & J. Fourn.....	15
<i>Keithomyces</i> Samson, Luangsa-ard & Houbraken.....	15
<i>Kosmimatamyces</i> Bianchin., Reinoso-Fuentealba, Rodr.-Andr., Cano & Stchigel.....	15
<i>Kukwaea</i> Suija, Motiej. & Zhurb.....	15
<i>Lendemeriella</i> S.Y. Kondr.....	16
<i>Linoporopsis</i> Voglmayr & Beenken.....	16
<i>Longiseptatispora</i> L.W. Hou & Crous.....	16
<i>Marquandomyces</i> Samson, Houbraken & Luangsa-ard.....	16
<i>Meniscomyces</i> Q.M. Wang & F.Y. Bai.....	16
<i>Muriphila</i> Jurjevic, Cmoková & Hubka.....	17
<i>Neoantennariella</i> Abdollahz. & Crous.....	17
<i>Neoasbolisia</i> Abdollahz. & Crous.....	17
<i>Neofusicladium</i> Crous, M. Shen & Y. Zhang ter.....	17
<i>Neolamproconium</i> Crous & Akulov.....	17
<i>Neophaeothea</i> Abdollahz. & Crous.....	17
<i>Neoshiraia</i> H.A. Ariyaw.....	17
<i>Neosorocybe</i> Crous & Akulov.....	18
<i>Neotorrubiella</i> Tasan., Thanakitp. & Luangsa-ard.....	18
<i>Nothoseiridium</i> Crous.....	18
<i>Nothoseptoria</i> Crous & Bulgakov.....	18
<i>Ochraceocephala</i> Voglmayr & Aiello.....	18
<i>Papiliomyces</i> Luangsa-ard, Samson & Thanakitp.....	18
<i>Parafusicladium</i> Crous, M. Shen & Y. Zhang ter.....	19
<i>Parathyridariella</i> Prigione, A. Poli, E. Bovio & Varese.....	19
<i>Parvomorbus</i> Wen Wang & S.F. Chen.....	19
<i>Patellariopsisaceae</i> Karun., Camporesi & K.D. Hyde.....	19
<i>Penicillaginaceae</i> Houbraken, Frisvad & Samson.....	19
<i>Petchia</i> Thanakitp., Mongkols. & Luangsa-ard.....	19
<i>Pinaceicola</i> Crous, M. Shen & Y. Zhang ter.....	20
<i>Pisutiella</i> S.Y. Kondr., Lokös & Farkas.....	20
<i>Protographum</i> Le Renard, Upchurch, Stockey & Berbee.....	20
<i>Pruniphilomyces</i> Crous & Bulgakov.....	20
<i>Pseudobactrodesmium</i> H. Zhang, W. Dong & K.D. Hyde.....	20
<i>Pseudohamigera</i> Houbraken, Frisvad & Samson.....	20
<i>Pseudosterigmatospora</i> Q.M. Wang & F.Y. Bai.....	21
<i>Purpureomyces</i> Luangsa-ard, Samson & Thanakitp.....	21
<i>Rhagadodidymellosis</i> Fern.-Brime, Gaya, Llimona & Nav.-Ros.....	21
<i>Robertozyma</i> Q.M. Wang & F.Y. Bai.....	21
<i>Rosettozyma</i> Q.M. Wang & F.Y. Bai.....	21
<i>Sajamaea</i> Flakus, Piatek & Rodr. Flakus.....	21
<i>Serusiaxia</i> Ertz & Diederich.....	22
<i>Serusiaxiella</i> S.H. Jiang, Lücking & J.C. Wei.....	22
<i>Siphulopsis</i> Kantvilas & A.R. Nilsen.....	22
<i>Sterigmatospora</i> Q.M. Wang & F.Y. Bai.....	22
<i>Sterila</i> Crous, M. Shen & Y. Zhang ter.....	22
<i>Sungia</i> Luangsa-ard, Samson & Thanakitp.....	22
<i>Synnematotriadelphia</i> Chuasechar., Somrith., Nuankaew & Boonyuen.....	23
<i>Teunia</i> Q.M. Wang & F.Y. Bai.....	23
<i>Trichophoma</i> Magaña-Dueñas, Cano & Stchigel.....	23
<i>Triseptata</i> Boonmee & Phookamsak.....	23
<i>Tyloclitostomum</i> van den Boom & Magain.....	23
<i>Vandijkomycella</i> Hern.-Restr., L.W. Hou, L. Cai & Crous.....	24
<i>Vesiculozygosporium</i> Crous.....	24
<i>Wetmoreana</i> Arup, Søchting & Frödén.....	24
<i>Yosiokobayasia</i> Samson, Luangsa-ard & Thanakitp.....	24
<b>References.....</b>	<b>25</b>

## Orders

### ***Aulographales* Crous, Spatafora, Haridas & I.V. Grig.**

Haridas et al. (2020) introduced this order to accommodate the families *Aulographaceae* and *Rhizodiscinaceae* based on phylogenetic analysis.

### ***Aureoconidiellales* Hern.-Restr. & Crous**

Hernández-Restrepo et al. (2020) introduced this order to accommodate the family *Aureoconidiellaceae* based on phylogenetic analysis.

### ***Catabotryales* K.D. Hyde & Senan.**

The analysis of LSU, SSU, *tef1*, and *rpb2* gene data by Hyde et al. (2017) has led to the description of seven subclasses including the order *Catabotryales*. On the basis of evolutionary data, Hyde et al. (2017) has proposed this order resulting in 15 sub-orders and 65 families in the subclass *Diaporthomycetidae*, having its stem age estimated 172 MYA. *Catabotryales* comprises saprobic species on dead leaves and stems of tropical monocotyledons. This order is distinct from its sister orders in having astromatic ascomata, broad cylindrical asci and ellipsoidal to cylindrical ascospores without a mucilaginous sheath (Hyde et al. 2020).

### ***Cladosporiales* Abdollahz. & Crous**

Based on the phylogenetic results, combined with morphology and ecology, Abdollahzadeh et al. (2020) introduced this new order to accommodate the family *Cladosporiaceae*, which was previously placed in *Capnodiales*.

### ***Comminutisporales* Abdollahz. & Crous**

Based on the phylogenetic results, combined with morphology and ecology, Abdollahzadeh et al. (2020) introduced this new order to accommodate the family *Comminutisporaceae*.

### ***Coniosporiales* Crous, Spatafora, Haridas & I.V. Grig.**

Haridas et al. (2020) introduced this order to accommodate the family *Coniosporiaceae* based on phylogenetic analysis.

### ***Heitmaniales* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tef1*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Lineolatales* Crous, Spatafora, Haridas & I.V. Grig.**

Haridas et al. (2020) introduced this order to accommodate the family *Lineolataceae* based on phylogenetic analysis.

### ***Neophaeothecales* Abdollahz. & Crous**

Abdollahzadeh et al. (2020) introduced this order to accommodate the family *Neophaeothecaceae* based on the phylogenetic results, combined with morphology and ecology.

### ***Racodiales* Abdollahz. & Crous**

The typification of the genus *Racodium* Fr. was discussed earlier by Hawksworth et al. (2011). Based on the results of Bayesian analysis of combined LSU, *tef1* and *rpb2* sequences of 193 taxa representing orders *Capnodiales* sens. lat., *Myrangiiales* and *Dothideales*, the new order *Racodiales* was introduced by Abdollahzadeh et al. (2020) to include family *Racodiaceae* (based on genus *Racodium*).

### ***Rosettozyma* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tef1*, and *cytB*), and physiological comparisons (Li et al. 2020).

## Families

### ***Aureoconidiellaceae* Hern.-Restr. & Crous**

Hernández-Restrepo et al. (2020) introduced this new family to accommodate the type genus *Aureoconidiella* Hern.-Restr. & Crous based on phylogenetic analysis.

### ***Coniosporiaceae* Crous, Spatafora, Haridas & I.V. Grig.**

Haridas et al. (2020) introduced this new family to accommodate the type genus *Coniosporium* Link based on phylogenetic analysis.

### ***Cylindrosympodiaceae* Crous, M. Shen & Y. Zhang ter**

Shen et al. (2020) introduced *Cylindrosympodiaceae* to accommodate the genera *Cylindrosympodium* W.B. Kendr. & R.F. Castañeda, *Pseudoamungitea* Crous, *Sympodiella* W.B. Kendr. and *Tothia* Bat. within the *Venturiales*.

### ***Foliocryphiaceae* C.M. Tian, N. Jiang & Crous**

Revaluation of families in the pathogenic fungal order *Diaporthales* and the family *Cryphonectriaceae* has resulted in the separation of the latter into two subclades, 21 genera and 55 species, based on the phylogenetic studies on combined sequence data of ITS, LSU, *tef1*, and *rpb2* genes. The family *Foliocryphiaceae* is distinguished by its phylogeny and dimorphic conidia while *Mastigosporellaceae* can be differentiated by owning apical conidial appendages. A new genus, *Neocryphonectria* has also been described in the family *Foliocryphiaceae* which is characterized by fusoid, aseptate macroconidia. In total, 31 families have been validated under the order *Diaporthales* (Jiang et al. 2020).

### ***Mastigosporellaceae* C.M. Tian, N. Jiang & Crous**

Revaluation of families in the pathogenic fungal order *Diaporthales* and the family *Cryphonectriaceae* has resulted in the separation of the latter into two subclades, 21 genera and 55 species, based on the phylogenetic studies on combined sequence data of ITS, LSU, *tef1*, and *rpb2* genes. The family *Foliocryphiaceae* is distinguished by its phylogeny and dimorphic conidia while *Mastigosporellaceae* can be differentiated by owning apical conidial appendages. A new genus, *Neocryphonectria* has also been described in the family *Foliocryphiaceae* which is characterized by fusoid, aseptate macroconidia. In total, 31 families have been validated under the order *Diaporthales* (Jiang et al. 2020).

### ***Neocryphonectria* C.M. Tian, N. Jiang & Crous**

Revaluation of families in the pathogenic fungal order *Diaporthales* and the family *Cryphonectriaceae* has resulted in the separation of the latter into two subclades, 21 genera and 55 species, based on the phylogenetic studies on combined sequence data of ITS, LSU, *tef1*, and *rpb2* genes. The family *Foliocryphiaceae* is distinguished by its phylogeny and dimorphic conidia while *Mastigosporellaceae* can be differentiated by owning apical conidial appendages. A new genus, *Neocryphonectria* has also been described in the family *Foliocryphiaceae* which is characterized by fusoid, aseptate macroconidia. In total, 31 families have been validated under the order *Diaporthales* (Jiang et al. 2020).

### ***Heitmaniaceae* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including

newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Jianyuniaceae* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Lineolataceae* Crous, Spatafora, Haridas & I.V. Grig.**

Haridas et al. (2020) introduced this new family to accommodate the type genus *Lineolata* Kohlm. & Volkm.-Kohlm. based on phylogenetic analysis.

### ***Neoantennariellaceae* Abdollahz. & Crous**

Based on the phylogenetic results, combined with morphology and ecology, Abdollahzadeh et al. (2020) introduced *Neoantennariellaceae* to accommodate the genera *Fumiglobus* D.R. Reynolds & G.S. Gilbert, *Neoantennariella* Abdollahz. & Crous and *Neoasbolisia* Abdollahz. & Crous within the *Capnodiales*.

### ***Neophaeothecaceae* Abdollahz. & Crous**

Abdollahzadeh et al. (2020) introduced this new family to accommodate the type genus *Neophaeotheca* Abdollahz. & Crous.

### ***Readerielliopsisaceae* Abdollahz. & Crous**

Abdollahzadeh et al. (2020) introduced *Readerielliopsisaceae* to accommodate the coelomycetous genera *Phaeoxyphiella* Bat. & Cif., *Readerielliopsis* Crous & Decock, *Scolecoxyphium* Cif. & Bat. and *Scorias* Fr. within the *Capnodiales*.

### ***Rhizodiscinaceae* Crous, Spatafora, Haridas & I.V. Grig.**

Based on phylogenetic analysis, Haridas et al. (2020) introduced this new family to accommodate the type genus *Rhizodiscina* Hafellner.

### ***Rosetozymaceae* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Rubikioideae* Lücking, M. Cáceres & Aptroot**

Cáceres et al. (2020) reanalyzed the phylogenetic placement of two non-lichenized fungi, namely *Furcaspora eucalypti* Crous & Verkley and *Rubikia evansii* Crous (*Graphidaceae*). Maximum likelihood tree of *Graphidaceae* based on the mtSSU, LSU, and *rpb2* concatenated markers showed the positions of *Furcaspora* Bonar and *Rubikia* H.C. Evans & Minter as separate, basally diverging clade. The new subfamily, *Rubikioideae*, was described by the authors.

### ***Tricladiaceae* P.R. Johnst. & Baschien**

While circumscribing the family *Solenopezaceae*, a novel family has been established which include the genera that have aquatic hyphomycete-like asexual morphs and sexual morphs comprising of shiny apothecia; such as *Cudoniella* Sacc., *Geniculospora* Sv., *Graddonia* Dennis, *Halenospora* E.B.G. Jones, *Mycofalcella* Marvanova, Om-Kalth. & J. Webster, *Spirosphaera* Beverw., and *Tricladium* Ingold. However, the family *Solenopezaceae* comprised of *Lasiobelonium* Ellis & Everh., *Solenopezia* Sacc., *Trichopeziza* Fuckel, and *Trichopezizella* Dennis ex Raitv. all showing sexual morphs with smooth-walled hairs. The current proposed novel family has been evidenced by the phylogenetic analysis revealing the new family has monophyletic clade having relationships with other families *Pleuroascaceae* and *Helotiaceae* (Johnston & Baschien 2020).

## Genera

### ***Acidotalaromyces* Houbraken, Frisvad & Samson**

An updated review of the generic, subgeneric, sectional classification of the order *Eurotiales* have resulted in the description of new family *Penicillaginaceae* and few genera, mainly based upon the sequence-based studies using GenBank accession numbers of ITS, *benA*, *calI* and *rpb2* gene sequences. In total, the order *Eurotiales* under *Ascomycetes* comprise of five accepted families, 28 genera and 1187 species (Houbraken et al. 2020).

### ***Aphanodesmium* Réblová & Hern.-Restr.**

With evidence from the phylogenetic studies, Réblová et al. (2020) placed the four species of *Bactrodesmium* in three unrelated novel genera, viz., *Gamsomyces* (*Sclerococcales*), *Kaseifertia* (*Pleosporales*) and *Aphanodesmium* (*Helotiales*) the last of which had been originally described in Koukol & Kolářová (2010) and made several novel combinations viz., *A. gabretae* (Koukol & Kolářová) Réblová & Hern.-Restr., *G. longisporus* (M.B. Ellis) Hern.-Restr. & Réblová, *G. stilboideus* (R.F. Castaneda & G.R.W. Arnold) Hern.-Restr. & Réblová, *Helicoascotaiwania farinosa* (Linder) Réblová, Hern.-Restr. & J. Fourn., *K. cubense* (R.F. Castaneda & G.R.W. Arnold) Réblová, Hern.-Restr. & J. Fourn. The new sequence data for *B. longisporum* M.B. Ellis and *B. stilboideum* R.F. Castañeda & G.R.W. Arnold placed them in *Sclerococcales* (*Eurotiomycetes*), *B. cubense* (R.F. Castañeda & G.R.W. Arnold) Zucconi & Lunghini in *Dothideomycetes*, and *B. gabretae* Koukol & Kolářová in *Leotiomycetes* (Réblová et al. 2020).

### ***Arboricolonus* S. Bien & Damm**

Multi-locus phylogenetic studies (LSU, ITS, *tub*, *tef1*) of fungi associated with wood necroses of *Prunus* L. in Germany, has led to the description of novel species, in a new genus *Arboricolonus* with the type *A. simplex* S. Bien & Damm, belonging in *Helotiales* (*Leotiomycetes*) with a collophorina-like asexual morph along with seven new species of *Cadophora* Lagerb. & Melin. The authors found the novel genus has similarities with *Glutinomyces* Nor Nakam., *Chalara* (Corda) Rabenh., *Hyalodendriella* Crous and *Polyphilus* D.G. Knapp, Ashrafi, W. Maier & Kovacs with ITS (90% for LC218288 with the strains of *Glutinomyces vulgaris*) and LSU (97% for EU040232 with the ex-type strain of *Hyalodendriella betulae*) similarity (Bein & Damm 2020) and has distinguishable asexual morphs.

### ***Ascospirella* Houbraken, Frisvad & Samson**

An updated review of the generic, subgeneric, sectional classification of the order *Eurotiales* have resulted in the description of new family *Penicillaginaceae* and few genera, mainly based upon the sequence-based studies using GenBank accession numbers of ITS, *benA*, *calI* and *rpb2* gene sequences. In total, the order *Eurotiales* under *Ascomycetes* comprise of five accepted families, 28 genera and 1187 species (Houbraken et al. 2020).

### ***Atrophysma* T. Sprib.**

The genus was introduced by Spribille et al. (2020) to accommodate a single species of a cyanolichen which differs from *Placynthium* by simple ascospores, and from *Leciophysma* by producing dark blue-black pigments in the apothecia. Multi-locus DNA sequencing placed the new genus in *Pannariaceae*. So far, no genus with black pigments was known within *Pannariaceae*.

### ***Aureoconidiella* Hern.-Restr. & Crous**

The monotypic genus *Aureoconidiella* was introduced by Hernández-Restrepo et al. (2020) with *A. foliicola* Hern.-Restr. & Crous as the type species. This genus is characterized by unbranched conidiophores, cicatrized and sympodial conidiogenous cells with thickened scars, producing sub-globose, verruculose, and golden-brown conidia.

### ***Baidera* Ertz & Diederich**

*Baidera* containing a single species, *B. mauritiana*, was described from Mauritius by Diederich & Ertz (2020). A maximum-likelihood analysis based on the two *rpb2* sequences (MN989868 for Ertz 21443-hymenium, MN989869 for Ertz 21443-thallus) was used to place *Baidera* in a phylogeny of *Arthoniales*. The new genus forms a distinct lineage in *Roccellaceae*, being somewhat related to *Gyrographa* Ertz & Tehler and *Sigridea* Tehler. *Baidera* differs from all genera of *Roccellaceae* by a thick crustose, not byssoid thallus, lirelliform ascomata with a carbonized excipulum not covered by a thalline layer, a carbonized hypothecium extending down to the substrate, a pruinose and atomentose hymenial disc, ascospores without a distinct gelatinous sheath, and a chemistry with psoromic acid as a major substance.

### ***Batnamyces* Noumeur**

A new endophytic genus *Batnamyces* belonging to *Chaetomiaceae* was described based on the morphological and molecular studies. It is phylogenetically related to *Stolonocarpus* X. Wei Wang & Houbraken and *Madurella* Brumpt. Besides the phylogenetic studies, biochemical analyses were also undertaken using NMR spectroscopy, HRESIMS and ECD techniques resulting in eight diketopiperazines. The strain has been isolated from the medicinal plant *Globularia alypum* L. collected in Algeria (Noumeur et al. 2020).

### ***Begerowomyces* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Bellamyces* Crous, Coppins & U. Braun**

*Bellamyces*, with *B. quercus* Crous, Coppins & U. Braun as the type species, was proposed by Shen et al. (2020) based on the multigene phylogenetic analysis, morphological and ecological characteristics. The genus is characterized by having solitary, and transversely multiseptate, rarely oblique conidia.

### ***Bergerella* Diederich & Lawrey**

A new genus and species discovered from Austria, *Bergerella* with the species *B. atrofusca* Diederich & Lawrey belonging to the family *Hydnaceae* in the order *Cantharellales*. The phylogenetic studies by analyzing ITS and nuLSU rRNA revealed that novel species has similarities with the genus *Minimedusa* Weresub & P.M. LeClair of the same order *Cantharellales*. However, it was quite unique in neither having lichen association nor bulbilliferous species of the order (Lawrey et al. 2020).

### ***Boekhoutia* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Bolbea* Buaya & Thines**

A novel holocarpic parasitoid oomycetes genus readily cultivable on malt-extract agar medium, belonging to the early *Saprolegniomycetes* in the order *Leptomitales*, has been reported from the Rebstock lake, Frankfurt, Germany. The absence of typical sacculate hyphal swellings and the “elephant hyphae” and the weak clustering with *Atkinsielladubia* (D. Atkins) Vishniac has resulted in describing this taxon as a novel genus. Phylogenetic analyses based on partial 18S rRNA gene (SSU) sequences were not conclusive in deriving contrasting topologies in minimum evolution (ME) and maximum likelihood (ML) analyses (Buaya & Thines 2020).

### ***Bryoclavula* H. Masumoto & Y. Degawa**

The genus was introduced by Masumoto & Degawa (2020) for a species, *Bryoclavula phycophila* H. Masumoto & Y. Degawa, described from Japan. Maximum-likelihood analysis of LSU (D1/D2) sequences placed the new genus in a new lineage within *Cantharellales*. Morphologically the genus is similar to lichenized genus *Multiclavula* R.H. Petersen but differs by lacking a globular or bulbil-like thallus.

### ***Burrowsia* Fryday & I. Medeiros**

The monotypic genus *Burrowsia* is proposed by Fryday et al. (2020) to accommodate the South African species *B. cataractae* Fryday & I. Medeiros. Maximum likelihood analysis of concatenated ITS, LSU, *rpb1* and *rpb2* sequences places the new species in the *Caliciaceae* outside related buellioid genera. Morphologically, *Burrowsia* is characterized by its pigmented, submuriform ascospores and asci with an apical tube-like structure.

### ***Carneothele* Fryday, T. Sprib. & M. Svenss.**

Spribile et al. (2020) introduced this genus based on morphological traits. The single successful DNA extraction yielded ITS and 28S rRNA sequences which affinities to *Dothideomycetes* and *Sarea* Fr., respectively. However, these sequences could derive from another fungal material. Morphologically, the genus is close to *Thelocarpon* Nyl. by its minute ascomata with the occasional presence of a yellow pruina, and multi-spored asci that gradually taper to a narrow apex but differs by having red-brown ascomata with the wall pigment forming magenta crystals in 10% KOH.

### ***Cladocillium* Chun-Hao Chen & R. Kirschner**

Based on evidence from DNA sequence data and morphology, Chen et al. (2020) introduced *Cladocillium* as a monotypic genus in *Mycosphaerellaceae*. The genus is typified by *C. musae* Chun-Hao Chen & R. Kirschner collected from leaves of *Musa itinerans* Cheesman in Taiwan. The genus is characterized by having macronematous, single, erect, unbranched or rarely branched, but with conidial chains arising in verticils along the conidiophore stipe, pigmented conidiophores, intercalary and terminal conidiogenous cells, in branched acropetal chains, pigmented, rough conidia (Chen et al. 2020).

### ***Commelinaceomyces* E. Tanaka**

A novel false-smut fungal genus has been described from Japan that infects the flowers of *Murdannia keisak* (Hasskarl) Handel-Mazzetti (*Commelinaceae*) belonging to the family *Clavicipitaceae* of the order *Hypocreales* (*Ascomycota*) based on the multi-locus phylogenetic analysis of DNA sequences (18S, 28S, *tefl*, *rpb1*, and *rpb2*). The novel taxa varied from its close relative *Ustilago* (Pers.) Roussel in having conidia development at the apex of conidiogenous cells (Eiji et al. 2020).

### ***Corylicola* Wijesinghe, Camporesi, Yong Wang bis & K.D. Hyde**

Wijesinghe et al. (2020) introduced the genus *Corylicola* in the family *Bambusicolaceae* (*Pleosporales*), to accommodate *C. italica* Wijesinghe, Camporesi, Yong Wang bis & K.D. Hyde isolated from *Corylus avellana* L. in Italy. This genus is characterized by scattered, globose to subglobose and ostiolate ascomata; anastomosing and branching pseudoparaphyses; cylindrical asci with a well-developed ocular chamber and short furcate pedicel; and single-septate ascospores. The coelomycetous asexual morph of *Corylicola* has holoblastic, phialidic conidiogenous cells and light brown conidia (Wijesinghe et al. 2020).

### ***Cryphognomonia* C.M. Tian & N. Jiang**

A polyphasic approach including morphological, cultural data, and phylogenetic analyses of partial ITS, LSU, *tefl*, *rpb2*, and *tub2* gene sequences has resulted in description of novel species of *Cryphognomonia*, *C. pini* C.M. Tian & N. Jiang belonging to *Gnomoniaceae* from China. The new species is characterized by developed pseudostromata and ascospores with obvious hyaline sheath (Yang et al. 2020).

### ***Diabolocovidia* Crous**

*Diabolocovidia* is the genus described during the lockdown period due to the COVID-19 pandemic from Gainesville, Florida, USA on the leaves of the palm *Serenoa repens* (W. Bartram) Small based on the collection of M J Wingfield. It is allied to *Vamsapriya* Gawas & Bhat in having brown, synnematus conidiophores, mono- to polytretic conidiogenous cells, and dark brown, septate conidia arranged in acropetal chains (Crous et al. 2020a).

### ***Emmanuelia* Ant. Simon, Lücking & Goffinet**

The genus *Emmanuelia* was proposed to comprise twelve species previously recognized within polyphyletic subfamily *Lobarioideae* (*Peltigeraceae*) and restricted to the New World. Molecular phylogenetic analyses using ITS and mtSSU sequences show that *Emmanuelia* forms a well-supported monophyletic group within the *Lobaria sensu lato* clade. Morphologically the genus is similar to *Ricasolia* D. Not., but differs by its apothecia, rimmed by overarching and often crenulate to lobulate margins, with the parathecium and the amphithecium apically separated and of a different structure (Simon et al. 2020).

### ***Erichansenia* S.Y. Kondr., Kärnefelt & A. Thell,**

Results of the three gene phylogeny of the *Teloschistaceae* based on ITS, LSU and mtSSU sequences revealed the genus *Shackletonia* Söchting, Frödén & Arup to be polyphyletic and the new genus *Erichansenia* was proposed by Kondratyuk et al. (2020) to accommodate three species, namely *Caloplaca epithallina* Lynge, *S. cryodesertorum* Garrido-Ben., Söchting & Pérez-Ort. and *S. sauronii* (Söchting & Øvstedal) Söchting, Frödén & Arup. Morphologically, the new genus is similar to *Shackletonia*, but differs in having better developed thallus, black apothecia with a dark greenish blue outer exciple, *Lecidea*-green pigment in the cortex of thallus and exciple, as well as in geographic distribution.

### ***Evansstolkia* Houbraken, Frisvad & Samson**

An updated review of the generic, subgeneric, sectional classification of the order *Eurotiales* have resulted in the description of new family *Penicillaginaceae* and few genera, mainly based upon the sequence-based studies using GenBank accession numbers of ITS, *benA*, *calI* and *rpb2* gene sequences. In total, the order *Eurotiales* under *Ascomycetes* comprise of five accepted families, 28 genera and 1187 species (Houbraken et al. 2020).

### ***Fagicola* Crous, M. Shen & Y. Zhang ter**

*Fagicola* was proposed by Shen et al. (2020) based on *F. fagi* (Crous & de Hoog) Crous, M. Shen & Y. Zhang ter (as *Fusicladium fagi* Crous & de Hoog), which was saprotrophic on leaves of *Fagus sylvatica* L. collected in the Netherlands.

### ***Fraxinicola* Crous, M. Shen & Y. Zhang ter**

Shen et al. (2020) introduced this genus to accommodate two new species and two new combinations based on the multigene phylogenetic analysis, morphological and ecological characteristics. Currently, the genus comprises four species viz. *F. europaea* Crous, M. Shen & Y. Zhang ter, *F. fraxini* (Aderh.) Crous, M. Shen & Y. Zhang ter (the type species), *F. italica* Crous, M. Shen & Y. Zhang ter and *F. orni* (M. Ibrahim, M. Schlegel & T.N. Sieber) Crous, M. Shen & Y. Zhang ter (Shen et al. 2020).

### ***Fulgogasparrea* S.Y. Kondr., N.-H. Jeong, Kärnefelt, Elix, A. Thell & Hur**

The genera *Fulgogasparrea* S.Y. Kondr., N.-H. Jeong, Kärnefelt, Elix, A. Thell & Hur and *Wetmoreana* Arup, Søchting & Frödén were introduced in 2013 by Kondratyuk et al. (2013) and Arup et al. (2013), respectively, for the *Caloplaca decipioides* group. The latter genus was accepted by Wijayawardene et al. (2020) while the former was considered to be a synonym. However, Mishra et al. (2020) accepted both *Fulgogasparrea* and *Wetmoreana* revealing genetical distance based on ITS sequences as well as morphological differences.

### ***Fuscohilum* Crous, M. Shen & Y. Zhang ter**

Shen et al. (2020) introduced this genus within *Symptoventuriaceae* to accommodate *Fusicladium rhodense* Crous & M.J. Wingf. and *F. sicilianum* Koukol. Since *F. rhodense* and *F. sicilianum* formed a separate generic clade within *Symptoventuriaceae*, these two species were phylogenetically assigned to a new genus, *Fuscohilum*.

### ***Gamsomyces* Hern.-Restr. & Réblová**

With evidence from the phylogenetic studies, Réblová et al. (2020) placed the four species of *Bactrodesmium* in three unrelated novel genera, viz., *Gamsomyces* (*Sclerococcales*, *Kaseifertia* (*Pleosporales*) and *Aphanodesmium* (*Helotiales*), the last of which had been originally described in Koukol & Kolářová (2010) and made several novel combinations such as *A. gabretae* (Koukol & Kolářová) Réblová & Hern.-Restr., *G. longisporus* (M.B. Ellis) Hern.-Restr. & Réblová, *G. stilboideus* (R.F. Castaneda & G.R.W. Arnold) Hern.-Restr. & Réblová, *Helicoascotaiwania farinosa* (Linder) Réblová, Hern.-Restr. & J. Fourn., *K. cubense* (R.F. Castaneda & G.R.W. Arnold) Réblová, Hern.-Restr. & J. Fourn. The new sequence data for *B. longisporum* M.B. Ellis and *B. stilboideum* R.F. Castañeda & G.R.W. Arnold placed them in *Sclerococcales* (*Eurotiomycetes*), *B. cubense* (R.F. Castañeda & G.R.W. Arnold) Zucconi & Lunghini in *Dothideomycetes*, and *B. gabretae* Koukol & Kolářová in *Leotiomycetes* (Réblová et al. 2020).

### ***Halocryptosphaeria* Dayarathne, Devadatha, V.V. Sarma & K.D. Hyde**

Two new genera, viz., *Halocryptosphaeria*; *Halotestudina* belonging the families *Diatrypaceae* (*Xylariales*) and *Testudinaceae* (*Pleosporales*), respectively have been described from mangrove forests of India and Thailand. The genus *Halocryptosphaeria* shows similarities with *Cryptosphaeria* Ces. & De Not. however, differs morphologically from *Cryptosphaeria* in having light-brown ascospores containing oil droplets versus pale yellow ascospores; and dwelling in marine environment (Rappaz, 1987). The other genus *Halotestudina* differs from all other genera of the family in having brown muriform ascospores that are constricted at each septum. These genera have been described mainly on the morphological aids supported by phylogenetic evidences (Dayarathne et al. 2020).

### ***Halotestudina* Dayarathne & K.D. Hyde**

Two new genera, viz., *Halocryptosphaeria*; *Halotestudina* belonging the families *Diatrypaceae* (*Xylariales*) and *Testudinaceae* (*Pleosporales*), respectively have been described from mangrove forests of India and Thailand. The genus *Halocryptosphaeria* shows similarities with *Cryptosphaeria* Ces. & De Not. however, differs morphologically from *Cryptosphaeria* in having light-brown ascospores containing oil droplets versus pale yellow ascospores; and dwelling in marine environment (Rappaz, 1987). The other genus *Halotestudina* differs from all other genera of the family in having brown muriform ascospores that are constricted at each septum. These genera have been described mainly on the morphological aids supported by phylogenetic evidences (Dayarathne et al. 2020).

### ***Italiofungus* Crous**

A total of seven new genera, 26 new species, 10 new combinations have been described from different parts of the world by Crous et al. (2020b) providing genera and species descriptions. All of these genera have been recorded on *Phillyrea latifolia* L., *Lonicera tatarica* L., *Tilia* sp., *Pinus sylvestris* L., *Caragana arborescens* Lam., *Prunus cerasus* L., *Muntingia calabura* L. respectively. The morphological studies were supported by molecular phylogeny analysis using the LSU and ITS (Crous et al. 2020b).

### ***Jennwenomyces* Goh & C.H. Kuo**

A newly segregated hyphomycete genus *Jennwenomyces* has been described based on the morphological and molecular data from the freshwater streams of Taiwan. The novel genus is characterized by the presence of versicolored, straight, navicular to cylindrical euseptate phragmospores borne on multiple percurrently extending, annellate conidiophores. The phylogenetic analysis showed similarities with the species of *Dictyospora* Abdel-Aziz, *Junewangia* W.A. Baker & Morgan-Jones, and *Sporidesmiella* P.M. Kirk in the family *Junewangiaceae* clustered together under *Sordariomycetes* (Goh & Kuo 2020).

### ***Jocatoa* R. Miranda**

The new genus *Jocatoa* was introduced by Miranda-González et al. (2020) to accommodate the single species *J. agminalis* (Nyl.) Lücking, Herrera-Camp. & R. Miranda previously included in the polyphyletic genus *Medusulina* Müll. Arg. which is no longer valid. The new monospecific genus strongly resembles species of *Diorygma* Eschw. but maximum likelihood analysis of SSU, LSU, and *rpb2* sequences put the genus *Jocatoa* outside the *Diorygma* clade. From *Diorygma* the new genus differs by having simple paraphyses tips that do not form an apothecium.

## ***Juncomyces* Crous**

Crous et al. (2020a) introduced *Juncomyces* to accommodate *J. californiensis* Crous (the type species) collected from leaves of *Juncus effuses* L. This genus is closely related to *Graminopassalora* U. Braun, C. Nakash., Videira & Crous but differs in having solitary conidiophores (rarely fascicles of 2–3), and multiseptate, obclavate conidia.

## ***Kaseifertia* Réblová & Hern.-Restr. & J. Fourn.**

With evidence from the phylogenetic studies, Réblová et al. (2020) placed the four species of *Bactrodesmium* in three unrelated novel genera, viz., *Gamsomyces* (*Sclerococcales*), *Kaseifertia* (*Pleosporales*) and *Aphanodesmium* (*Helotiales*) the last of which had been originally described in Koukol & Kolářová (2010) and made several novel combinations viz., *A. gabretae* (Koukol & Kolářová) Réblová & Hern.-Restr., *G. longisporus* (M.B. Ellis) Hern.-Restr. & Réblová, *G. stilboideus* (R.F. Castaneda & G.R.W. Arnold) Hern.-Restr. & Réblová, *Helicoascotaiwania farinosa* (Linder) Réblová, Hern.-Restr. & J. Fourn., *K. cubense* (R.F. Castaneda & G.R.W. Arnold) Réblová, Hern.-Restr. & J. Fourn. The new sequence data for *B. longisporum* M.B. Ellis and *B. stilboideum* R.F. Castañeda & G.R.W. Arnold placed them in *Sclerococcales* (*Eurotiomycetes*), *B. cubense* (R.F. Castañeda & G.R.W. Arnold) Zucconi & Lunghini in *Dothideomycetes*, and *B. gabretae* Koukol & Kolářová in *Leotiomycetes* (Réblová et al. 2020).

## ***Keithomyces* Samson, Luangsa-ard & Houbraken**

A phylogenetic framework is reconstructed for the *Clavicipitaceae* focusing on *Metarhizium* Sorokin through increased taxon-sampling using five genomic loci (SSU, LSU, *tefl*, *rpb1*, *rpb2*) and the barcode marker ITS rRNA resulted in describing six new genera of monophyletic clades subtending the core *Metarhizium* clade, 19 new species and several novel combinations (Mongkolsamrit et al. 2020).

## ***Kosmimatamyces* Bianchin., Reinoso-Fuentealba, Rodr.-Andr., Cano & Stchigel**

Crous et al. (2020a) introduced *Kosmimatamyces* as a monotypic genus in *Capnodiaceae*. The genus is typified by *K. alatophylus* Bianchin., Reinoso-Fuentealba, Rodr.-Andr., Cano & Stchigel collected from soil of the salt marsh in Argentina.

## ***Kukwaea* Suija, Motiej. & Zhurb.**

On the basis of morphological and phylogenetic studies, a novel genus, *Kukwaea*, found associated with *Cetraria islandica* (L.) Ach. has been described from Russia. The newly described fungus, characterized by its cupulate, brown ascomata with grey to blackish disc surrounded by brownish grey pubescence, exciple of textura angularis type, with crystals in the lower part, with granulose excipular hairs obtuse at the tips, simple to forked paraphyses, *Calycina*-type asci, and hyaline, aseptate ascospores. The genetic analysis proved that it belongs to the order *Helotiales* (Suija et al. 2020).

### ***Lendemeriella* S.Y. Kondr.**

The genus *Lendemeriella* was proposed by Kondratyuk et al. (2020) for seven species of the *Caloplaca reptans* Lendemer & B.P. Hodk. group on the basis of the analysis of ITS, LSU and mtSSU sequences. *Lendemeriella* forms the separate robust branch in sister position to the genera *Olegblumia* S.Y. Kondr., Lökös & Hur, *Rufoplaca* Arup, Søchting & Frödén and *Usnochroma* Søchting, Arup & Frödén. Authors mentioned that some contradictive data exist, and additional molecular data are needed to clarify position of these and other possible *Lendemeriella* species.

### ***Linosporopsis* Voglmayr & Beenken**

A new genus *Linosporopsis* belonging to the order *Xylariales* has been described based on the basis of molecular phylogenetic and morphological studies from the leaves of *Fagus sylvatica* L. The multi-locus matrix of partial ITS-LSU rRNA, *rpb2* and *tub2* gene sequences as well as morphological investigations revealed that both species, *Linospora ischnotheca* (Desm.) Sacc. and *L. ochracea* Sacc. are unrelated to the diaphothalean genus *Linospora* Fuckel, but belong to *Xylariaceae* sens. str. Thus, along with *Linospora carpini* J. Schrot. and *Linospora magnagutiana* Speg. these four species have been accommodated in the new genus (Voglmayr & Beenken 2020).

### ***Longiseptatispora* L.W. Hou & Crous**

Crous et al. (2020b) introduced this genus to accommodate two new species based on the DNA phylogeny and morphological characters. The genus is typified by *L. curvata* Crous & Bulgakov collected from leaves of *Lonicera tatarica* L. in Russia.

### ***Marquandomyces* Samson, Houbraken & Luangsa-ard**

A phylogenetic framework is reconstructed for the *Clavicipitaceae* focusing on *Metarhizium* Sorokin through increased taxon-sampling using five genomic loci (SSU, LSU, *tef1*, *rpb1*, *rpb2*) and the barcode marker ITS rRNA resulted in describing six new genera of monophyletic clades subtending the core *Metarhizium* clade, 19 new species and several novel combinations (Mongkolsamrit et al. 2020).

### ***Meniscomyces* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tef1*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Muriphila* Jurjevic, Cmoková & Hubka**

Crous et al. (2020a) introduced *Muriphila* as a monotypic genus in *Teratosphaeriaceae*. This genus is characterized by velvety, dark olivaceous to black, convex, radially wrinkled to crateriform colonies, septate, with smooth to verrucous walls hyphae, rectangular to asymmetrical, and disintegrates into fragments at maturity hyphal cells (Crous et al. 2020a).

### ***Neoantennariella* Abdollahz. & Crous**

The monotypic genus *Neoantennariella* was erected by Abdollahzadeh et al. (2020) with *N. phyllicae* Abdollahz. & Crous as the type species. Morphologically similar to *Antennariella* Bat. & Cif., but different in conidiomatal and conidium morphology (Abdollahzadeh et al. 2020).

### ***Neoasbolisia* Abdollahz. & Crous**

Based on the phylogenetic results, combined with morphology and ecology, the monotypic genus *Neoasbolisia* was erected by Abdollahzadeh et al. (2020) with *N. phyllicae* Abdollahz. & Crous as the type species.

### ***Neofusicladium* Crous, M. Shen & Y. Zhang ter**

Shen et al. (2020) introduced this genus within *Sympoventuriaceae* to accommodate three new combinations. The genus is typified by *N. eucalypti* (Crous & R.G. Shivas) Crous, M. Shen & Y. Zhang ter collected from leaves of *Eucalyptus* sp.

### ***Neolamproconium* Crous & Akulov**

A total of seven new genera, 26 new species, 10 new combinations have been described from different parts of the world by Crous et al. (2020b) providing genera and species descriptions. All of these genera have been recorded on *Phillyrea latifolia* L., *Lonicera tatarica* L., *Tilia* sp., *Pinus sylvestris* L., *Caragana arborescens* Lam., *Prunus cerasus* L., *Muntingia calabura* L. respectively. The morphological studies were supported by molecular phylogeny analysis using the LSU and ITS (Crous et al. 2020b).

### ***Neophaeothea* Abdollahz. & Crous**

The monotypic genus *Neophaeothea* was erected by Abdollahzadeh et al. (2020) with *N. salicorniae* (Crous & Roets) Abdollahz. & Crous as the type species based on the phylogenetic results, combined with morphology and ecology.

### ***Neoshiraia* H.A. Ariyaw.**

Ariyawansa et al. (2020) introduced this new genus in *Shiraiaceae* to accommodate two coelomycetous species isolated from *Camellia sinensis* L. based on multi-gene phylogenetic inference and morphological interpretations. *Shiraia* Henn. can be distinguished from *Neoshiraia* in having fusiform, muriform, asymmetrical, hyaline to light brown conidia whereas *Neoshiraia* possesses hyaline, aseptate, obovoid to ellipsoidal conidia (Ariyawansa et al. 2020).

### ***Neosorocybe* Crous & Akulov**

A total of seven new genera, 26 new species, 10 new combinations have been described from different parts of the world by Crous et al. (2020b) providing genera and species descriptions. All of these genera have been recorded on *Phillyrea latifolia* L., *Lonicera tatarica* L., *Tilia* sp., *Pinus sylvestris* L., *Caragana arborescens* Lam., *Prunus cerasus* L., *Muntingia calabura* L. respectively. The morphological studies were supported by molecular phylogeny analysis using the LSU and ITS (Crous et al. 2020b).

### ***Neotorrubiella* Tasan., Thanakitp. & Luangsa-ard**

Two new fungal genera, *Neotorrubiella* and *Pechia*, on insects has been described in the *Hypocreales* from Thailand. A multi-locus molecular phylogenetic analysis involved SSU, LSU, *tef1*, *rpb1*, and *rpb2* together with the nuclear intergenic region (IGR) for 67 sequences.

*Neotorrubiella* resembles *Torrubiella* Boud. in the absence of a stipe and can be distinguished by the production of whole ascospores, which are not commonly found in *Torrubiella* (Thanakitpipattana et al. 2020).

### ***Nothoseiridium* Crous**

A novel pathogenic fungus infecting *Podocarpus* L'Heritier ex Persoon has been discovered and described from Cape Town, South Africa. This novel genus produces black-colored, round, flattened acervular conidiomata with many brownish wall layers. The conidiophores, reduced to conidiogenous cells are colorless, smooth, subcylindrical to ampuliform, annellidic and are arising from the base of the stroma. The conidia are fusoid, slightly curved, smooth-walled, guttulate, pale brown, unequally 4-euseptate; basal cell obconic with truncate hilum, hyaline; median cells pale brown; apical cell obtuse, hyaline; apical and basal appendage filiform, flexuous, unbranched, excentric. Type species: *N. podocarpi* Crous (Crous et al. 2020a).

### ***Nothoseptoria* Crous & Bulgakov**

Crous et al. (2020b) introduced this new genus in the family *Mycosphaerellaceae* (*Mycosphaerellales*), to accommodate *N. caraganae* (Henn.) Crous & Bulgakov collected from living leaves of *Caragana arborescens* Lam. in Russia.

### ***Ochraceocephala* Voglmayr & Aiello**

Aiello et al. (2020) described the monotypic genus *Ochraceocephala* to accommodate *O. foeniculi* Voglmayr & Aiello within *Leptosphaeriaceae*. It is phylogenetically closely related to *Plenodomus* Preuss, from which it deviates substantially in morphology. *Plenodomus* species are characterized by pycnidial phoma-like asexual morphs, and while in two *Plenodomus* species [*P. chrysanthemi* (Zachos, Constantinou & Panag.) Gruyter, Aveskamp & Verkley, *P. tracheiphilus* (Petri) Gruyter, Aveskamp & Verkley] simple hyphomycetous, phialophora-like synanamorphs have been recorded (Boerema et al. 1994), these are very different from the complex conidiophores of the present fennel pathogen (Aiello et al. 2020).

### ***Papiliomyces* Luangsa-ard, Samson & Thanakitp.**

A phylogenetic framework is reconstructed for the *Clavicipitaceae* focusing on *Metarhizium* Sorokin through increased taxon-sampling using five genomic loci (SSU, LSU, *tefl*, *rpb1*, *rpb2*) and the barcode marker ITS rRNA resulted in describing six new genera of monophyletic clades subtending the core *Metarhizium* clade, 19 new species and several novel combinations (Mongkolsamrit et al. 2020).

### ***Parafusicladium* Crous, M. Shen & Y. Zhang ter**

Based on a multigene phylogenetic analysis, Shen et al. (2020) introduced the genus *Parafusicladium* with three new combinations in *Sympoventuriaceae*. Currently, the genus comprises three species viz. *P. amoenum* (R.F. Castañeda & Dugan) Crous, M. Shen & Y. Zhang ter (the type species), *P. intermedium* (Crous & W.B. Kendr.) Crous, M. Shen & Y. Zhang ter and *P. paraamoenum* (Crous, Jacq. Edwards & P.W.J. Taylor) Crous, M. Shen & Y. Zhang ter (Shen et al. 2020).

### ***Parathyridariella* Prigione, A. Poli, E. Bovio & Varese**

Poli et al. (2020) introduced the genus *Parathyridariella*, typified by the new species *P. dematiacea* Prigione, A. Poli, E. Bovio & Varese within *Thyridariaceae*. This genus is characterized by septate, hyaline to lightly pigmented hyphae; numerous, mostly in chain,

intercalary or solitary, globose to subglobose, from brownish to dark brown chlamydospores (Poli et al. 2020).

### ***Parvomorbus* Wen Wang & S.F. Chen**

Studies on the pathogenic fungal family *Cryphonectriaceae* in the southern China on different members of higher plants of the order *Myrtales* has resulted in the discovery of eight species of four genera and a novel genus *Parvomorbus* with two novel species. The novelty has been supported by DNA sequences of the partial LSU, ITS, two regions of the  $\beta$ -tubulin (*tub2/tub1*) gene, and *tefl* gene region (Wang et al. 2020).

### ***Patellariopsidaceae* Karun., Camporesi & K.D. Hyde**

A new family under the polyphyletic *Ascomycetes* order *Helotiales* has been described from Italy on the basis of morphology and molecular phylogeny. The novel family is represented with an asexual morph of *Patellariopsis atrovinosa* (A. Bloxam ex Curr.) Dennis while a new record of *Cheirospora botryospora* (Mont.) Berk. & Broome (*Vibrisseaceae*) on *Fagus sylvatica* L. (*Fagaceae*) has also been reported. Phylogenetic analyses based on a combined sequence dataset of LSU and ITS were used to infer the phylogenetic relationships within the *Helotiales* and thus confirmed its identity (Karunarathna et al. 2020)

### ***Penicillaginaceae* Houbraken, Frisvad & Samson**

An updated review of the generic, subgeneric, sectional classification of the order *Eurotiales* have resulted in the description of new family *Penicillaginaceae* and few genera, mainly based upon the sequence-based studies using GenBank accession numbers of ITS, *benA*, *calI* and *rpb2* gene sequences. In total, the order *Eurotiales* under *Ascomycetes* comprise of five accepted families, 28 genera and 1187 species (Houbraken et al. 2020).

### ***Petchia* Thanakitp., Mongkols. & Luangsa-ard**

Two new fungal genera, *Neotorrubiella* and *Petchia*, on insects has been described in the *Hypocreales* from Thailand. A multi-locus molecular phylogenetic analysis involved SSU, LSU, *tefl*, *rpb1*, and *rpb2* together with the nuclear intergenic region (IGR) for 67 sequences. *Neotorrubiella* resembles *Torrubiella* Boud. in the absence of a stipe and can be distinguished by the production of whole ascospores, which are not commonly found in *Torrubiella* (Thanakitpipattana et al. 2020).

### ***Pinaceicola* Crous, M. Shen & Y. Zhang ter**

Based on a multigene phylogenetic analysis, Shen et al. (2020) proposed the genus *Pinaceicola* with two new combinations in *Sympoventuriaceae*. Currently, the genus comprises two species viz. *P. pini* (Crous & de Hoog) Crous, M. Shen & Y. Zhang ter (the type species) and *P. cordae* (Koukol) Crous, M. Shen & Y. Zhang ter (Shen et al. 2020).

### ***Pisutiella* S.Y. Kondr., Lokös & Farkas**

The phylogenetic analysis of the *Teloschistaceae* based on ITS, LSU and mtSSU sequences supported recognition new genus, *Pisutiella*, to accommodate six species of the *Caloplaca conversa* (Kremp.) Jatta group (Kondratyuk et al. 2020). The genus *Pisutiella* is positioned as a strong branch between *Rufoplaca* Arup, Søchting & Frödén and *Usnochroma* Søchting, Arup & Frödén.

### ***Protographum* Le Renard, Upchurch, Stockey & Berbee**

The fossil fungus *Protographum* was described by Le Renard et al. (2020) from cuticle of unidentified conifer. *P. luttrellii* Le Renard, Upchurch, Stockey & Berbee is distinguished from other fossil taxa by pseudomonopodial dichotomous branching in a scutellum initiated by multiple generator hyphae (Le Renard et al. 2020).

### ***Pruniphilomyces* Crous & Bulgakov**

Crous et al. (2020b) introduced this monotypic genus *Pruniphilomyces* to accommodate the plant pathogenic fungus *P. circumscissus* (Sacc.) Crous & Bulgakov collected from living leaves of *Prunus cerasus* L. in Russia.

### ***Pseudobactrodesmium* H. Zhang, W. Dong & K.D. Hyde**

A couple of new species resembling *Bactrodesmium longisporum* M.B. Ellis were collected from the Greater Mekong Subregion and is described under new genus *Pseudobactrodesmium*. They have fasciculate conidiophores, enteroblastic conidiogenous cells and subulate to fusiform, phragmoseptate conidia with a tapering apical cell and sheath. On the basis of analyses of LSU, SSU, ITS and *rpb2* sequence data, the novel genus is placed under the family *Dactylosporaceae* (*Eurotiomycetes*) (Dong et al. 2020).

### ***Pseudohamigera* Houbraken, Frisvad & Samson**

An updated review of the generic, subgeneric, sectional classification of the order *Eurotiales* have resulted in the description of new family *Penicillaginaceae* and few genera, mainly based upon the sequence-based studies using GenBank accession numbers of ITS, *benA*, *calI* and *rpb2* gene sequences. In total, the order *Eurotiales* under *Ascomycetes* comprise of five accepted families, 28 genera and 1187 species (Houbraken et al. 2020).

### ***Pseudosterigmatospora* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tef1*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Purpleomyces* Luangsa-ard, Samson & Thanakitp.**

A phylogenetic framework is reconstructed for the *Clavicipitaceae* focusing on *Metarhizium* Sorokin through increased taxon-sampling using five genomic loci (SSU, LSU, *tef1*, *rpb1*, *rpb2*) and the barcode marker ITS rRNA resulted in describing six new genera of monophyletic clades subtending the core *Metarhizium* clade, 19 new species and several novel combinations (Mongkolsamrit et al. 2020).

### ***Rhagadodidymellopsis* Fern.-Brime, Gaya, Llimona & Nav.-Ros.**

The lichenicolous fungus *Rhagadodidymellopsis* growing on the thallus of the terricolous lichen *Endocarpon pusillum* Hedw. was introduced by Fernández-Brime et al. (2020) with *R. endocarpi* Fern.-Brime, Gaya, Llimona & Nav.-Ros. as the type species. *R. endocarpi* is characterized by its almost completely superficial stromatic ascomata with a coarse and irregular surface, and an ascomatal wall of very irregular thickness, and ascospores smaller than those of *Didymellopsis perigena* (Nyl.) Grube & Hafellner (Fernández-Brime et al. 2020).

### ***Robertozyma* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Rosettozyma* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Sajamaea* Flakus, Piatek & Rodr. Flakus**

The mycoparasitic fungus *Sajamaea* was proposed by Piatek et al. (2020) with *S. mycophila* Flakus, Piatek & Rodr. Flakus as the type species within *Dictyosporiaceae* based on molecular and morphological characters.

### ***Serusiauxia* Ertz & Diederich**

The genus accommodates a single species, *S. inexpectata* Ertz & Diederich, found on Mauritius. Maximum likelihood and Bayesian inference analyses based on the four mtSSU sequences put the fungus in a clade including *Lithothelium septemseptatum* (R.C. Harris) Aptroot, the genus *Anthracotheceum* Hampe ex A. Massal. and several species of *Pyrenula* Ach. (including type species, *P. nitida* Weigel) Ach. Based on the unique morphological and chemical characters, namely a soreciate thallus and producing the gyrophoric acid, the genus *Serusiauxia* was proposed by Diederich & Ertz (2020).

### ***Serusiauxiella* S.H. Jiang, Lücking & J.C. Wei**

The genus *Serusiauxiella* was introduced by Jiang et al. (2020) to accommodate three species, all new to science and known from (sub-)tropical Asia. Maximum likelihood analysis based on four markers (SSU, LSU, *tefl*, and *rpb2*) revealed a well-supported novel lineage within *Strigula sens. lat.* for which the name *Serusiauxiella* was proposed. From *Strigula sens. str.* the genus differs in the *Trentepohlia* Mart. photobiont and the extremely long macroconidial appendages.

### ***Siphulopsis* Kantvilas & A.R. Nilsen**

Maximum likelihood analyses based on combined ITS and LSU sequences indicated that *Icmadophila* is not monophyletic. The new genus *Siphulopsis* was introduced to comprise a single rare Australian species, *S. queenslandica* (Kantvilas) Kantvilas & A.R. Nilsen, which had previously been placed in *Knightiella* Müll. Arg. Morphologically it resembles a species of *Siphula* Fr. but differs by lacking the basal rhizines characteristic of the genus (Ludwig et al. 2020).

### ***Sterigmatospora* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in

the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Sterila* Crous, M. Shen & Y. Zhang ter**

Shen et al. (2020) introduced this new genus in the family of *Sympoventuriaceae* (*Venturiales*), to accommodate *S. eucalypti* Crous, M. Shen & Y. Zhang ter collected from leaves of a *Eucalyptus* sp. based on the multigene phylogenetic analysis, morphological and ecological characteristics.

### ***Sungia* Luangsa-ard, Samson & Thanakitp.**

A phylogenetic framework is reconstructed for the *Clavicipitaceae* focusing on *Metarhizium* Sorokin through increased taxon-sampling using five genomic loci (SSU, LSU, *tefl*, *rpb1*, *rpb2*) and the barcode marker ITS rRNA resulted in describing six new genera of monophyletic clades subtending the core *Metarhizium* clade, 19 new species and several novel combinations (Mongkolsamrit et al. 2020).

### ***Synnematotriadelphia* Chuaseehar., Somrith., Nuankaew & Boonyuen**

Based on the molecular phylogenetic analyses of nuclear ribosomal DNA and partial sequence of *rpb2*, the species of *Triadelphia* Shearer & J.L. Crane bifurcated into two monophyletic clades having the type species along with different others, viz., *T. heterospora* Shearer & J.L. Crane, *T. diversa* Tzean & J.L. Chen, *T. loudetiae* Maggi, Bartoli & Rambelli, *T. hexaformispora* Chuaseehar., Somrith. & Boonyuen, and *T. romanica* Constant. & Samson while the other clade comprised of *T. moubasheri* Abdel-Sater & Soliman, *T. pulvinata* Maggi, Bartoli & Rambelli, and *T. disseminata* Madrid & Edathodu leaving the other two species, *Triadelphia stilboidea* Mercado & R.F. Castañeda and *T. synnematofera* Matsush. in a separate clade for which the authors assigned them under the proposed novel genus (Chuaseeharonnachai et al. 2020).

### ***Teunia* Q.M. Wang & F.Y. Bai**

On the basis of multi-locus phylogenetic analyses combined with the clustering optimisation analysis revealed that there has been tremendous diversity of the Basidiomycetous yeasts in the Chinese soils. An exhaustive list of 107 new species have also been described including newly proposed two orders, three families and eight new genera and three combinations. The phylogenetic studies involved analyses of D1/D2 domain of LSU, SSU, ITS, and protein-coding genes (*rpb1*, *rpb2*, *tefl*, and *cytB*), and physiological comparisons (Li et al. 2020).

### ***Trichophoma* Magaña-Dueñas, Cano & Stchigel**

*Trichophoma* was proposed by Crous et al. (2020a) with *T. cylindrospora* Magaña-Dueñas, Cano & Stchigel as the type species. Based on the phylogenetic analysis of the ITS, LSU and *tefl* combined dataset, the closest relative of *T. cylindrospora* is *Forliomyces uniseptata* Phukhams., Camporesi & K.D. Hyde. *F. uniseptata* differs from *T. cylindrospora* in producing shorter and broader conidia (10–15 × 5–8 µm vs 18–20 × 2–3 µm), which are brown-coloured when mature (hyaline in *T. cylindrospora*) (Phukhamsakda et al. 2016).

### ***Triseptata* Boonmee & Phookamsak**

Boonmee et al. (2020) introduced this new genus in the family *Latoruaceae* based upon the morphological and phylogenetic evidences. This new genus is characterized by immersed, uni- to multi-loculate ascomata, filamentous pseudoparaphyses, bitunicate, fissitunicate,

cylindrical-clavate asci and 3-septate, light brown spores, with asymmetrical ends, in its sexual form. The LSU, ITS and SSU showed that *Triseptata* has distinct lineage than other genera of the family.

### ***Tylocliostomum* van den Boom & Magain**

The Bayesian analysis of ITS, mtSSU and *rpb1* sequences confirmed the genus *Bacidina* Vězda to be paraphyletic (see Kistenich et al. 2018) and the new genus *Tylocliostomum* was introduced by van den Boom & Magain (2020) to accommodate a single species, *T. viridifarinosum* van den Boom & Magain. Within the *Ramalinaceae*, *Tylocliostomum* forms a well-supported group with *Tylothallia biformigera* (Leight.) P. James & H. Kiliyas and two accessions of *Cliostomum haematommatis* (Keissl.) D. Hawksw., Earl.-Benn. & Coppins, which most probably do not represent *Cliostomum* sens. str. as it was assumed by authors.

### ***Vandijkomycella* Hern.-Restr., L.W. Hou, L. Cai & Crous**

Hou et al. (2020) introduced the genus *Vandijkomycella* to accommodate two new species isolated from soil samples within *Didymellaceae* based on morphological and molecular data. *V. joseae* Hern.-Restr., L.W. Hou, L. Cai & Crous is characterized by producing pycnidia with longer whitish hyphal outgrowths, and with elongated necks.

### ***Vesiculozygosporium* Crous**

A total of seven new genera, 26 new species, 10 new combinations have been described from different parts of the world by Crous et al. (2020b) providing genera and species descriptions. All of these genera have been recorded on *Phillyrea latifolia* L., *Lonicera tatarica* L., *Tilia* sp., *Pinus sylvestris* L., *Caragana arborescens* Lam., *Prunus cerasus* L., *Muntingia calabura* L. respectively. The morphological studies were supported by molecular phylogeny analysis using the LSU and ITS (Crous et al. 2020b).

### ***Wetmoreana* Arup, Søchting & Frödén**

The genera *Fulgogasparrea* S.Y. Kondr., N.-H. Jeong, Kärnefelt, Elix, A. Thell & Hur and *Wetmoreana* Arup, Søchting & Frödén were introduced in 2013 by Kondratyuk et al. (2013) and Arup et al. (2013), respectively, for the *Caloplaca decipioides* group. The latter genus was accepted by Wijayawardene et al. (2020) while the former was considered to be a synonym. However, Mishra et al. (2020) accepted both *Fulgogasparrea* and *Wetmoreana* revealing genetical distance based on ITS sequences as well as morphological differences.

### ***Yosiokobayasia* Samson, Luangsa-ard & Thanakitp.**

A phylogenetic framework is reconstructed for the *Clavicipitaceae* focusing on *Metarhizium* Sorokin through increased taxon-sampling using five genomic loci (SSU, LSU, *tef1*, *rpb1*, *rpb2*) and the barcode marker ITS rRNA resulted in describing six new genera of monophyletic clades subtending the core *Metarhizium* clade, 19 new species and several novel combinations (Mongkolsamrit et al. 2020).

## References

- Abdollahzadeh, J; Groenewald, JZ; Coetzee, MPA; Wingfield, MJ; Crous, PW. 2020. Evolution of lifestyles in *Capnodiales*. *Studies in Mycology* 95: 381–414.
- Aiello, D; Vitale, A; Polizzi, G; Voglmayr, H. 2020. *Ochraceocephala foeniculi* gen. et sp. nov., a new pathogen causing crown rot of fennel in Italy. *Mycology* 66: 1–22.
- Ariyawansa, HA; Tsai, I; Thambugala, KM; Chuang, WY; Lin, SR; Hozzein, WN; Cheewangkoon, R. 2020. Species diversity of pleosporalean taxa associated with *Camellia sinensis* (L.) Kuntze in Taiwan. *Scientific Reports* 10(12762): 1–20.
- Arup, U; Søchting U; Frödén P. 2013. A new taxonomy of the family *Teloschistaceae*. *Nordic Journal of Botany* 31: 16–83.
- Bien, S; Damm, U. (2020) *Arboricolonus simplex* gen. et sp. nov. and novelties in *Cadophora*, *Minutiella* and *Proliferodiscus* from *Prunus* wood in Germany. *Mycology* 63: 119–161. <https://doi.org/10.3897/mycokeys.63.46836>
- Boerema, GH; de Gruyter, J; van Kesteren, HA (1994). Contributions towards a monograph of *Phoma* (*Coelomycetes*) – III. 1. Section *Plenodomus*: Taxa often with a *Leptosphaeria* teleomorph. *Persoonia* 15: 431–487.
- Boonmee, S; Calabon, MS; Phookamsak, R; Elgorban, AM; Hyde, KD. (2020). *Triseptata sexualis* gen. et sp. nov. in *Latoruaceae* (*Pleosporales*). *Phytotaxa* 447(4): 252–264.
- Buaya, AT; Thines, M. (2020). *Bolbea parasitica* gen. et sp. nov., a cultivable holocarpic parasitoid of the early-diverging *Saprolegniomycetes*. *Fungal Evolution and Systematics* 6: 129–137.
- Cáceres, MES; Lücking, R; Schumm, F; Aptroot, A. 2020. A lichenized family yields another renegade lineage: *Papilionovela albohallina* is the first non-lichenized, saprobic member of *Graphidaceae* subfam. *Graphidoideae*. *The Bryologist* 123(2): 144–154.
- Chen, C; Hsieh, S; Yeh, Y. et al. 2020. *Cladocilliummusae*, a new genus and species of cercosporoid fungi (*Mycosphaerellaceae*) on wild banana in Taiwan. *Mycological Progress* 19: 837–843.
- Chuaseeharonnachai, C; Suetrong, S; Nuankaew, S; Somrithipol, S; Hongsanan, S; Srikitikulchai, P; Jones, EBG; Boonyuen, N. 2020. *Synnematotriadelphia* gen. nov. (*S. stilboidea* comb. nov. and *S. synnematofera* comb. nov.) and *Triadelphia hexaformispora* sp. nov. in the family *Triadelpiaceae*. *Mycological Progress* 19(2): 127–137.
- Crous, PW; Wingfield, MJ; Chooi, YH; Gilchrist, CLM; Lacey, E; Pitt, JI; Roets, F; Swart, WJ; Cano-Lira, JF; Valenzuela-Lopez, N; Hubka, V; Shivas, RG; Stchigel, AM; Holdom, DG; Jurjević, Ž; Kachalkin, AV; Lebel, T; Lock, C; Martín, MP; Tan, YP; Tomashevskaya, MA; Vitelli, JS; Baseia, IG; Bhatt, VK; Brandrud, TE; De Souza, JT; Dima, B; Lacey, HJ; Lombard, L; Johnston, PR; Morte, A; Papp, V; Rodríguez, A; Rodríguez-Anrade, E; Semwal, KC; Tegart, L; Abad, ZG; Akulov, A; Alvarado, P; Alves, A; Andrade, JP; Arenas, F; Asenjo, C; Ballarà, J; Barrett, MD; Berná, LM; Berraf-Tebbal, A; Bianchinotti, MV; Bransgrove, K; Burgess, TI; Carmo, FS; Chávez, R; Čmoková, A; Dearnaley, JDW; de A. Santiago, ALCM; Freitas-Neto, JF; Denman, S; Douglas, B; Dovana, F; Eichmeier, A; Esteve-Raventós, F; Farid, A; Fedosova, AG; Ferisin, G; Ferreira, RJ; Ferrer, A; Figueiredo, CN; Figueiredo, YF; Reinoso-Fuentealba, CG; Garrido-Benavent, I; Cañete-Gibas, CF. 2020a. Fungal Planet description sheets: 1042–1111. *Persoonia* 44: 301–459.
- Crous, PW; Wingfield, MJ; Schumacher, RK; Akulov, A; Bulgakov, TS; Carnegie, AJ; Jurjević, Ž; Decock, C; Denman, S; Lombard, L; Lawrence, DP; Stack, AJ; Gordon, TR; Bostock, RM; Burgess, T; Summerell, BA; Taylor, PWJ; Edwards, J; Hou, LW; Cai, L; Rossman, AY; Wöhner, T; Allen, WC; Castlebury, LA; Visagie, CM;

- Groenewald, JZ. 2020b. New and Interesting Fungi. 3. Fungal Systematics and Evolution 6: 157–231.
- Dayarathne, MC; Jones, EBG; Maharachchikumbura, SSN; Devadatha, B; Sarma, VV; Khongphinitbunjong, K; Chomnunti, P; Hyde, KD (2020). Morpho-molecular characterization of microfungi associated with marine based habitats. *Mycosphere* 11(1): 1–188, Doi 10.5943/mycosphere/11/1/1.
- Diederich, P; Ertz, D. 2020. First checklist of lichens and lichenicolous fungi from Mauritius, with phylogenetic analyses and the description of new taxa. *Plant and Fungal Systematics* 65(1): 13–75.
- Dong, W; Hyde, KD; Doilom, M; Yu, XD; Bhat, DJ; Jeewon, R; Boonmee, S; Wang, GN; Nalumpang, S; Zhang, H. 2020. *Pseudobactrodesmium* (*Dactylosporaceae*, *Eurotiomycetes*, *Fungi*) a Novel Lignicolous Genus. *Frontiers in Microbiology* 11 (no. 465): 1–13.
- Tanaka, E; Shrestha, B; Shivas, RG. (2020). *Commelinaceomyces*, gen. nov., for four clavicipitaceous species misplaced in *Ustilago* that infect *Commelinaceae*, *Mycologia*, 112: 3, 649–660, doi: <https://doi.org/10.1080/00275514.2020.1745524>.
- Fernández-Brime, S; Gaya, E; Llimona, X; Wedin, M; Navarro-Rosinés, P. 2020. *Rhagadodidymellopsis endocarpi* gen. et sp. nov. and *Arthopyrenia symbiotica* (*Dothideomyceta*), two lichenicolous fungi growing on *Endocarpon* species. *Plant and Fungal Systematics* 65(1): 176–184.
- Fryday, AM; Medeiros, ID; Siebert, SJ; Pope, N; Rajakaruna, N. 2020. *Burrowsia*, a new genus of lichenized fungi (*Caliciaceae*), plus the new species *B. cataractae* and *Scoliciosporum fabisporum*, from Mpumalanga, South Africa. *South African Journal of Botany* 132: 471–481.
- Goh, T; Kuo, C. 2020. *Jennwenomyces*, a new hyphomycete genus segregated from *Belemnospora*, producing versicolored phragmospores from percurrently extending conidiophores. *Mycol. Progress* 19: 869–883 <https://doi.org/10.1007/s11557-020-01602-7>
- Haridas, S; Albert, R; Binder, M; Bloem, J; LaButti, K; Salamov, A; Andreopoulos, B; Baker, SE; Barry, K; Bills, G; Bluhm, BH; Cannon, C; Castanera, R; Culley, DE; Daum, C; Ezra, D; González, JB; Henrissat, B; Kuo, A; Liang, C; Lipzen, A; Lutzoni, F; Magnuson, J; Mondo, SJ; Nolan, M; Ohm, RA; Pangilinan, J; Park, HJ; Ramírez, L; Alfaro, M; Sun, H; Tritt, A; Yoshinaga, Y; Zwiers, LH; Turgeon, BG; Goodwin, SB; Spatafora, JW; Crous, PW; Grigoriev. 2020. 101 *Dothideomycetes* genomes: A test case for predicting lifestyles and emergence of pathogens. 2020. *Studies in Mycology*. 96: 141–153.
- Hawksworth, DL; Santesson, R; Tibell, L. 2011. *Racoleus*, a new genus of sterile filamentous lichen-forming fungi from the tropics, with observations on the nomenclature and typification of *Cystocoleus* and *Racodium*. *IMA Fungus* 2: 71–79.
- Hernández-Restrepo, M; Giraldo, A; van Doorn, R; Wingfield, MJ; Groenewald, JZ; Barreto, RW; Colmán, AA; Mansur, PSC; Crous, PW. 2020. The Genera of Fungi – G6: *Arthrographis*, *Kramasamuha*, *Melnikomyces*, *Thysanorea*, and *Verruconis*. *Fungal Systematics and Evolution* 6: 1–24.
- Hou, LW; Hernández-Restrepo, M; Groenewald, JZ; Cai, L; Crous, PW. 2020. Citizen science project reveals high diversity in *Didymellaceae* (*Pleosporales*, *Dothideomycetes*). *MycKeys* 65: 49–99.
- Houbraken, J; Kocsubé, S; Visagie, CM; Yilmaz, N; Wang, XC, Meijer, M; Kraak, B; Hubka, V; Bensch, K; Samson, RA; Frisvad, JC. 2020. Classification of *Aspergillus*, *Penicillium*, *Talaromyces* and related genera (*Eurotiales*): An overview of families, genera, subgenera, sections, series and species. *Studies in Mycology* 95: 5–169.

- Hyde, KD; Maharachchikumbura, SSN; Hongsanan, S; Samarakoon, MC et al. (2017). The ranking of fungi: a tribute to David L. Hawksworth on his 70th birthday. *Fungal Diversity* 84: 1–23.
- Hyde, KD; Norphanphoun, C; Maharachchikumbura, SSN; Bhat DJ et al. (2020). Refined families of *Sordariomycetes*. *Mycosphere* 11(1): 305–1059, <https://doi.org/10.5943/mycosphere/11/1/7>.
- Jiang, N; Fan, XL; Tian, CM; Crous, PW. 2020. Reevaluating *Cryphonectriaceae* and allied families in *Diaporthales*. *Mycologia* 112 (2): 267–292.
- Jiang, SH; Lücking, R; Xavier-Leite, AB; Cáceres, MES; Aptroot, A; Portilla, CV; Wei, JC. (2020). Reallocation of foliicolous species of the genus *Strigula* into six genera (lichenized *Ascomycota*, *Dothideomycetes*, *Strigulaceae*). *Fungal Diversity* 102: 257–291.
- Johnston, PR; Baschien, C. (2020). *Tricladiaceae* fam. nov. (*Helotiales*, *Leotiomycetes*). *Fungal Systematics and Evolution* 6: 233–242.
- Karunarathna, A; Peršoh, D; Ekanayaka, AH; Jayawardena, RS; Chethana, KWT; Goonasekara, ID; Cheewangkoon, R; Camporesi, E; Hyde, KD; Lumyong, S; Karunarathna, SC. (2020). *Patellariopsidaceae* Fam. Nov. with Sexual-Asexual Connection and a New Host Record for *Cheirospora botryospora* (*Vibrissaceae*, *Ascomycota*). *Frontiers in Microbiology* 11 (no. 906): 1–13.
- Kistenich, S; Timdal E; Bendiksby M; Ekman S. (2018). Molecular systematics and character evolution in the lichen family *Ramalinaceae* (*Ascomycota*: *Lecanorales*). *Taxon* 67: 871–904.
- Kondratyuk, SY; Lokös, L; Farkas, E; Kärnefelt, I; Thell, A; Yamamoto, Y; Hur, JS. 2020. Three new genera of the *Teloschistaceae* proved by three gene phylogeny. *Acta Botanica Hungarica* 62(1-2): 109–136.
- Kondratyuk, SY; Jeong, M-H; Yu, N-H; Kärnefelt, I; Thell, A; Elix, JA; Kim, J; Kondratyuk, AS; Hur, J-S. 2013. Four new genera of teloschistoid lichens (*Teloschistaceae*, *Ascomycota*) based on molecular phylogeny. *Acta Botanica Hungarica* 55(3-4): 251–274.
- Koukol O; Kolářová, Z. 2010. *Bactrodesmium gabretae* (anamorphic *Helotiales*), a new sporodochial species described from spruce needles. *Nova Hedwigia* 91: 243–248.
- Lawrey, JD; Sikaroodi, M; Gillevet, PM; Diederich, P. 2020. A new species of bulbil-forming lichenicolous fungi represents an isolated clade in the *Cantharellales*. *The Bryologist* 123(2): 155–162. <https://doi.org/10.1639/0007-2745-123.2.155>.
- Le Renard, L; Stockey, RA; Upchurch, G; Berbee, M. 2020. A new epiphyllous fly-speck fungus from the Early Cretaceous Potomac Group of Virginia (125–112 Ma): *Protographum luttrellii*, gen. et sp. nov. *Mycologia* 112(3): 504–518.
- Li, AH; Yuan, FX; Groenewald, M; Bensch, K; Yurkov, AM; Li, K; Han, PJ; Guo, LD; Aime, MC; Sampaio, JP; Jindamorakot, S; Turchetti, B; Inacio, J; Fungsin, B; Wang, QM; Bai, FY. 2020. Diversity and phylogeny of basidiomycetous yeasts from plant leaves and soil: Proposal of two new orders, three new families, eight new genera and one hundred and seven new species. *Studies in Mycology* 96: 17–140.
- Ludwig, LR; Kantvilas, G; Nilsen, AR; Orlovich, DA; Ohmura, Y; Summerfield, TC; Wilk, K; Lord, JM. 2020. A molecular-genetic reassessment of the circumscription of the lichen genus *Icmadophila*. *The Lichenologist* 52(3): 213–220.
- Masumoto, H; Degawa, Y. 2020. *Bryoclavula phycophila* gen. et sp. nov. belonging to a novel lichenized lineage in *Cantharellales* (*Basidiomycota*). *Mycological Progress* 19(7): 705–714.
- Miranda-González, R; Lücking, R; Barcenás-Peña, A; Herrera-Campos, MA. 2020. The new genus *Jocatoa* (*Lecanoromycetes*: *Graphidaceae*) and new insights into subfamily *Redonographoideae*. *The Bryologist* 123(2): 127–143.

- Mishra, GK; Upreti, DK; Nayaka, S; Thell, A; Kärnefelt; I; Lökös, L; Hur, J-S; Sinha, GP; Kondratyuk SY. 2020. Current taxonomy of the lichen family Teloschistaceae from India with descriptions of new species. *Acta Botanica Hungarica* 62(3–4): 309–391.
- Mongkolsamrit, S; Khonsanit, A; Thanakitpipattana, D; Tسانathai, K; Noisriboom, W; Lamlertthon, S; Himaman, W; Houbraken, J; Samson, RA; Luangsa-ard, J. (2020). Revisiting *Metarhizium* and the description of new species from Thailand. *Studies in Mycology* 95: 171–251.
- Noumeur, SR, Teponno, RB, Helaly, SE et al. (2020). Diketopiperazines from *Batnamyces globulariicola*, gen. & sp. nov. (*Chaetomiaceae*), a fungus associated with roots of the medicinal plant *Globularia alypum* in Algeria. *Mycological Progress* 19, 589–603. <https://doi.org/10.1007/s11557-020-01581-9>
- Phukhamsakda, C; Ariyawansa, HA; Phillips, AJL et al. 2016. Additions to *Sporormiaceae*: Introducing two novel genera, *Sparticola* and *Forliomyces*, from *Spartium*. *Cryptogamie, Mycologie* 37: 75–97.
- Piatek, M; Rodriguez-Flakus, P; Domic, A; Palabral-Aguilera, AN; Gómez, MI; Flakus, A. 2020. Phylogenetic placement of *Leptosphaeria polylepidis*, a pathogen of Andean endemic *Polylepis tarapacana*, and its newly discovered mycoparasite *Sajamaea mycophila* gen. et sp. nov. *Mycological Progress* 19(1): 1–14.
- Poli, A; Bovio, E; Ranieri, L; Varese, GC; Prigione V. 2020. News from the Sea: A New Genus and Seven New Species in the Pleosporalean families *Roussoellaceae* and *Thyridariaceae*. *Diversity* 12(4, no. 144): 1–18.
- Rappaz, F. (1987) Taxonomie et nomenclature des Diatrypacées à asquesoctosporées. *Mycologia Helvetica* 2, 285–648.
- Réblová, M; Hernández-Restrepo, M; Fournier, J; Nekvindová, J. 2020. New insights into the systematics of *Bactrodesmium* and its allies and introducing new genera, species and morphological patterns in the *Pleurotheciales* and *Savoryellales* (*Sordariomycetes*). *Studies in Mycology* 95: 415–466. <https://doi.org/10.1016/j.simyco.2020.02.002>.
- Shen, M; Zhang, JQ; Zhao, LL; Groenewald, JZ; Crous, PW; Zhang, Y. 2020. *Venturiales*. *Studies in Mycology* 96: 185–308.
- Simon, A; Lücking, R; Moncada, B; Mercado-Díaz, JA; Bungartz, F; Cáceres, MES; Gumboski, EL; Martins, SMA; Spielmann, AA; Parker, D; Goffinet, B. 2020. *Emmanuelia*, a new genus of lobaroid lichen-forming fungi (*Ascomycota: Peltigerales*): phylogeny and synopsis of accepted species. *Plant and Fungal Systematics* 65(1): 76–94.
- Spribille, T; Fryday, AM; Pérez-Ortega, S; Svensson, M; Tønsberg, T; Ekman, S; Holien, H; Resl, P; Schneider, K; Stabenheiner, E; Thüs, H; Vondrák, J; Sharman, L. 2020. Lichens and associated fungi from Glacier Bay National Park, Alaska. *The Lichenologist* 52(2): 61–181.
- Suija, A; Zhurbenko, MP; Stepanchikova, ISS; Himelbrant, DE; Kuznetsova, ES; Motiejunaite, J. (2020). *Kukwaea pubescens* gen. et sp. nova (*Helotiales*, incertae sedis), a new lichenicolous fungus on *Cetraria islandica*, and a key to the lichenicolous fungi occurring on *Cetraria* s. str. *Phytotaxa* 459 (1): 39–50.
- Thanakitpipattana, D; Tسانathai, K; Mongkolsamrit, S; Khonsanit, A; Lamlertthon, S; Luangsa-ard, JJ. 2020. Fungal pathogens occurring on *Orthopterida* in Thailand. *Persoonia* 44: 140–160.
- van den Boom, PPG; Magain, N. 2020. Three new lichen species from Macaronesia belonging in *Ramalinaceae*, with the description of a new genus. *Plant and Fungal Systematics* 65(1): 167–175.
- Voglmayr, H and Beenken, L. 2020. *Linosporopsis*, a new leaf-inhabiting scolecosporeous genus in *Xylariaceae*. *Mycological Progress* 19(3): 205–222.

- Wang, W; Li, GQ; Liu, QL; Chen, SF. 2020. *Cryphonectriaceae* on *Myrtales* in China: phylogeny, host range, and pathogenicity. *Persoonia* 45: 101–131.
- Wijesinghe, SN; Wang, Y; Camporesi, E; Wanasinghe, DN; Boonmee, S; Hyde, KD. 2020. A new genus of *Bambusicolaceae* (*Pleosporales*) on *Corylus avellana* (*Fagales*) from Italy. *Biodiversity Data Journal* 8: e55957, 1–19.
- Yang, Q; Jiang, N; Tian, CM. (2020). Tree inhabiting gnomoniaceous species from China, with *Cryphogonomonia* gen. nov. proposed. *MycoKeys* 69: 71–89.