12. Opisthobranchs of the Gulf of Eilat and the Red Sea: an account of similarities and differences

(color plates pp. 189-196)

Yonow, N.

Opisthobranchs are fascinating molluscs: their evolution has led them to lose their shell completely in the more evolved genera, but each step in this evolutionary loss has been successful and examples survive today (Fig. 1. p 189). The lack of a protective shell has led to active methods of defence, an amazing array of body forms, and to flamboyant colors and patterns. Opisthobranchs form part of the class Gastropoda. They are highly evolved, and almost all species are marine. They range in size from 2 to 500 millimetres, and are characterized by a soft and flexible body with a dorsal fold of skin known as the mantle. The sub-class Opisthobranchia includes more than 6000 species (Behrens and Hermosillo, 2005). The prosobranchs (sea snails and shells) are certainly more numerous, but the opisthobranchs surpass them with the diversity of flamboyancy, behavior, dietary requirements, and by their extraordinary methods of defence.

Introduction to Opisthobranchs

Opisthobranchs are predators and grazers, occurring in all marine habitats, and their diet can be extremely specialized. They may display seasonal population explosions, dramatically affecting the reef ecosystems. In the Gulf of Eilat, for example, increased numbers are frequently due to eutrophication of the sea around industrial regions. These high incidences have facilitated the study of swimming and feeding behaviors of *Notarchus indicus* Schweigger and *Melibe rangi* Bergh by Schuhmacher (1973), for example. Studies on the diet and acid secretion of *Berthellina citrina* (Rüppell and Leuckart) have also been conducted in the Gulf of Eilat (Marbach and Tsurnamal, 1973). And, it has been shown that *Aplysia oculifera* Adams and Reeve hatches, metamorphoses, grows, reproduces and dies in less than one year in this location (Plaut *et al.*, 1995, 1998).

Opisthobranchs keep their predators at bay with remarkable powers of attack and defence. Some species (aeolids) incorporate the stinging cells – nematocysts – they recover from the tentacles of their cnidarian (e.g., jelly fish, sea anemone, coral and hydroid) prey, to use them for their own defence. Other species (pleurobranchs) secrete acids in a mono-layer of epidermal cells, situated in the mantle, foot and rhinophores as a defence. Finally, there are all those species which are capable of

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concentrating toxic substances extracted from the sponges upon which they feed, which render them inedible and poisonous. In these cases, the sea slugs advertise their colors boldly, acting as warning signs to would-be predators (chromodorids and phyllidiids; aposematic coloration). Indeed, they often have extra appendages where they concentrate these toxins (*Ceratosoma*). Opisthobranchs which feed on sponges can also be cryptic, adopting the same colors as their sponge food or the surrounding substrate. They are often very flattened and usually rough to touch, because they can take the spicules from the sponge and incorporate them into their own skin, giving them a second line of defence if their camouflage fails. Some even have pits which imitate the sponge's osculae (*Aldisa* sp., *Sclerodoris tuberculata* Eliot) and make them virtually invisible to predators. Other particularly vigorous species will not hesitate to attack marine molluses, including other sea slugs.

All opisthobranchs are hermaphrodite, the reproductive organs of both sexes being present in the adults, on the right side of the body. Copulation is usually reciprocal, but aplysiids, for instance, will mate in long chains, injecting the one in front with sperm. Eggs are usually laid in a characteristic spiral or string, often brightly colored: they contain the toxins acquired by the adult and do not need camouflage (Pawlik et al. 1988). Larval development type is related to egg size: numerous small eggs hatch into veligers with a longer planktonic phase, and fewer larger eggs either hatch into a short-lived veliger or a benthic larva with no planktonic stage. Most species of opisthobranchs have a planktonic stage which can survive in the open ocean for many months if the trigger for metamorphosis (usually the adult food) is unavailable. The lifespan of opisthobranchs is generally less than one year but in more primitive species, larger species, species which live in temperate and cold water, and species living at depths may live longer, but probably not more than a few years. The longest-lived tropical sea slug is Dolabella auricularia (Lightfoot), living for five years in an aquarium in Hawaii (Hadfield and Switzer-Dunlap, 1984), a similar lifespan to the Antarctic species Philine gibba Strebel (Seager, 1982).

Opisthobranchs are distributed among ten orders; only five are considered here as the others are specialized and have their own literature and two are unknown in the Red Sea. Of the Cephalaspidea, the more primitive bubble shells, for example *Bulla* and *Hydatina*, rummage in the mud whereas the colorful chelidonurids (*Chelidonura* and *Philinopsis*) have only a small internal cap-like shell and glide along on the sea bottom and among corals in search of prey. Some are herbivorous, but the majority are active predators: *Philinopsis cyanea* (Martens) was found to feed on its cousins *Bulla ampulla* (Linnaeus), *Ringicula acuta* Philippi and *Chelidonura sandrana* Rudman in the Red Sea and the Maldives (Yonow, 1992). The order Anaspidea, otherwise known as sea hares, have the shell reduced to a small internal plate. They are herbivorous, feeding on a variety of algae, often changing species composition as they grow. Many can produce a purple ink when disturbed, and some species swim gracefully, using their parapodia as undulating wings. The Pleurobranchida may also retain a cap-like shell internally, and have a large gill present on the right side. They are a relatively sparse order and all species are carnivorous, feeding on a variety of sessile invertebrates. Most species can secrete acid as a defence mechanism. The Sacoglossa are almost exclusively herbivorous, and have developed a uniseriate radula for piercing algal cells. Species have a variety of shells, bulloid as in *Oxynoe viridis* (Pease), bivalved (*Julia exquisite* Gould), cap-like, or it may be lacking altogether (*Elysia, Thuridilla, Plakobranchus*). Body form covers the whole range, from simple and leaf-like to covered in cerata as in *Cyerce elegans* (Bergh) and *Polybranchia orientalis* (Kelaart) or with parapodia like the aplysiids (*Elysia trilobata* Heller and Thompson).

The order Nudibranchia has the greatest number of species which are the most specialized (see Fig. 1, colour plate p. 189). They owe their evolutionary success to the total loss of a shell and the resulting development of other mechanisms of defence. The majority of species hatch from an egg with a larval shell, which is retained in the more primitive Opisthobranchia but lost during stages of metamorphosis in the Nudibranchia. There are four sub-orders, each with a range of species which exploit every niche and many prey species. The Doridina are the most numerous and probably the most varied in body form, followed by the Dendronotina, with tree-like gills present on the dorsum for respiration. Many species of Aeolidina store nematocysts for their own defence, and are species-specific when it comes to diet. The Arminina are the least well known, being the least frequently encountered, and have traditionally included species which did not fit in any other grouping.

Variety of body form in nudibranchs is influenced directly by diet organism and mode of life of the species. Nudibranchs are carnivorous without exception, and feed on a wide range of marine invertebrates, from sessile epifauna such as barnacles and ascidians to highly mobile polychaetes, gastropods and crustaceans. Some have also specialized to feed on egg masses, such as Favorinus japonicus Baba, now found in the Red Sea (Yonow, 2008). In numerous sub-orders and families, the mantle bears digitiform appendages. As well as being used for respiration, they may also be used to store sponge toxins in the Chromodorididae or unexploded nematocysts from cnidarian prey for the sea slug's own defence in the aeolids. In fact, two species Pteraeolidia ianthina (Angas) and Glaucilla marginata Bergh can give a nasty sting if roughly handled. Most dorids feed on sponges, but others feed on other opisthobranchs, soft corals, or ascidians (see review, Yonow, 1996). They, and indeed all other opisthobranchs, except the Dendrodorididae and Phyllidiidae, have a rasping "tongue," or radula covered with small hook-shaped teeth, as varied in number and shape as the prey items they feed on. All have numerous rows of teeth on the radula, and the number in the rows can vary from five to several hundred. The phyllidiids feed on sponges, like most dorids, but by everting their mouthparts; they have no teeth but an eversible pharynx with glands attached (internally) which dissolve the sponge in situ, and they then suck up the nutritious juices. The chemical substances and terpenoids produced by the sponge to protect it are contained in this juice (Gunthorpe and Cameron, 1987).

History of Opisthobranch Research in Red Sea

The Red Sea is an important marine biogeographical region, but to date the opisthobranch fauna has only been described sporadically. Despite the ancient connection between the Red Sea and the Mediterranean, the fauna and flora of the Red Sea are derived from the Indian Ocean. The Red Sea is considered part of the Western Indian Ocean Province, of the Indo-West Pacific zoogeographical region. Scientific expeditions to these regions date back nearly 250 years. The first European scientific expedition specifically to the Red Sea was the ill-fated Danish "Arabia Felix" expedition from 1761 - 1767, which spent October 1762 to August 1763 in the Red Sea, final destination Yemen. Peter Forsskål, a Swedish botanist and student of Linnaeus, was on the trip, and one of the six of seven scientists who died. The extensive mollusc collection of Forsskål with detailed field notes was published posthumously by the leader and sole survivor of the expedition, Carsten Niebuhr, in 1775 (Forsskål 1775). Niebuhr was in fact the first geographer to accurately measure and map the Red Sea, and his chart was used by the British Royal Navy 100 years later. His measurements and calculations of the Gulf of Suez were so precise that they were used in the building of the Suez Canal (Maempel 1992). The history of the collection and the taxonomic status of the species described by Forsskål, including three opisthobranchs, have recently been reviewed by Yaron et al. (1986).

The second important expedition was Napoleon I's great "Expedition d'Egypte" during 1798 - 1801, with the natural historian J.C. de Savigny, among others. Savigny's beautiful engraved plates of marine organisms, including 15 opisthobranchs, were published in 1817, but he became blind before he was able to write the accompanying descriptions. Audouin's interpretation of the molluscs (1826) was completed by Issel (1869) and Pallary (1926). Bouchet and Danrigal (1982) published copies of the plates, with a history of both the plates and the collections.

Ehrenberg and Hemprich made extensive invertebrate collections between 1820 and 1826, which were published by Ehrenberg (Hemprich died in Ethiopia) as 10 colored plates in 1828 followed by the accompanying text in 1831. Eduard Rüppell also travelled in the Red Sea during this period and published beautiful coloured plates and monographs, the opisthobranchs in 1828 and 1830 with Leuckart, including 16 new species. Reproductions of these beautiful plates are presented for the first time in a monograph of benthic opisthobranchs of the Red Sea (Yonow, 2008).

An important review of all records of molluscs published since the time of Linnaeus was published in a book of Red Sea molluscs by Issel (1869) in Italian. Expeditions to the Red Sea continued, and the Austrian research vessel "Pola" cruised the northern Red Sea in 1895 - 1896, and the southern half in 1897 - 1898. Sturany published the gastropod material, including a few opisthobranchs, in 1903. In 1904 Cyril Crossland, a British zoologist with tropical experience, was sent to the Sudanese Red Sea, and his collections of opisthobranchs were described in two publications by Eliot (1908, 1911). Albert Vayssière, who was the director of the marine laboratory in Marseille, published descriptions and plates of opisthobranchs, approximately 45

species, collected by Jousseaume during five voyages from 1887 - 1894 to Aden, Djibouti, and throughout the Red Sea and by Gravier in an expedition in 1904 to Djibouti (1906, 1911a, 1911b, 1912).

The next major work on the Red Sea opisthobranchs resulted from the Cambridge Expedition to the Suez Canal in 1924, its main aim to assess the effects of migration since the opening of the Suez Canal. O'Donoghue's large work describes four cephalaspideans and 15 nudibranchs, together with a list of all species recorded from the Red Sea (then a total of 82 species) and a list of all species recorded in the Mediterranean for comparison (O'Donoghue, 1929). The French "Mission Robert Ph. Dollfus en Egypte" (December 1927 - March 1929, s/s "Al Sayad") collected opisthobranchs in the northern Red Sea and in Lake Timsah in the Suez Canal. Alice Pruvot-Fol (Museum National d'Histoire Naturelle, Paris) published the findings on their collection of opisthobranchs in 1933. Many of her species have not been found subsequently, and many are too poorly described to be recognized. One species, *Hypselodoris dollfusi* Pruvot-Fol, was long considered *incertae sedis*, but it has been rediscovered recently in the Gulf of Oman (Gosliner and Behrens, 2000) and the Gulfs of Eilat and Suez (Yonow, 2008).

The John Murray Expedition (1933 - 1934) was the third expedition in the first half of that century to collect a significant number of opisthobranchs (23 species) in the Red Sea and Gulf Aden (and the Kuria Muria Islands, off the coast of Oman). These were published by Nellie Eales (British Museum [Natural History]) in 1938. Heller and Thompson (1983) published some of the results of the British Suakin expedition, describing eight new species and re-describing ten species. Barash and Danin worked from the Gulf of Eilat, and documented the fauna, especially in comparison with migrations to and from the Red Sea (1972, 1977, 1980). Gohar and his colleagues, based at the Marine Station of Al Ghardaqa (= Hurghada) on the Egyptian coast, detailed the biology, spawning and development of numerous species (1948-1967).

During the summer of 1983, the m/v "Ibn Batuta" circumnavigated the Red Sea, with the express purpose of collecting bivalves, prosobranchs and opisthobranchs. This voyage, supplemented by further collections, culminated in two books (Sharabati, 1984; Yonow, 2008) and a series of taxonomic papers (Yonow, 2000 *passim*). Dekker and Orlin (2000) published a valuable assessment and list of all mollusc species recorded from the Red Sea. Since the "Ibn Batuta" trip, I have been collecting and compiling a record of benthic opisthobranchs; published records from the Red Sea now exceed 250 (compared with 82 in 1929) excluding the Thecosomata, Gymnosomata, and Acochlida. Additional specimen and photographic records have been provided from further collections by the author and by few reliable and trusted sources, bringing the total number of species recorded to 300: these are listed in the appendix (including species without names) with an assessment for presence or absence in the Gulf of Eilat and geographical distribution. Descriptions and illustrations of the majority of these species can be found in Yonow (2008).

Zoogeography and Endemism

The real extent of endemism in the Red Sea, and indeed anywhere, is difficult to estimate due to the extensive collections and comparisons of faunas necessary. However, some taxa have been well studied and in fish, an endemism level of 17% in the Red Sea appears to be accurate (Ormond and Edwards, 1987). However, this is higher for butterfly fish, 41% (Righton *et al.*, 1996), cardinal fish, 22% (Randall, 2003) and gobies, 27% (Goren, pers. comm.). Levels of 8% are reported for scleractinian corals (De Ventier *et al.*, 2000), but 56% for Xeniidae (Reinicke, 1997); 14% for echinoderms and 9% for algae have also been estimated (Head 1987, Campbell 1987). Polychaetes are estimated to have a 7.5% endemic population (calculated from Wehe and Fiege, 2002). Cowries show higher endemism levels of 35% (calculated from Foin and Ruebush, 1969) as do echinoids (Nebelsick, 1996). Endemism of the molluscan fauna is estimated at 16% (Dekker and Orlin, 2000). Of the 300 species of sea slugs recorded from the Red Sea, 80 are found only here, representing 27% endemism.

The distribution of molluscs is not homogeneous, with more species being present in the southern region of the Red Sea. This distribution pattern also occurs with echinoids, 25 species recorded from the northern part of the Red Sea and 43 recorded from the southern region (Nebelsick, 1996). Nineteen species of cowries are present in the northern Red Sea, 24 in the central Red Sea, and 37 in the south (Foin and Ruebush, 1969). A study of the Polyplacophora reveals 23 species for the Red Sea with only two endemic species (9%), five of which are limited to the southern Red Sea (Strack, 1993). Pteropods are also more speciose in the southern Red Sea (Almogi-Labin, pers. comm.). While molluscs in general show a trend of higher diversity in the southern half of the Red Sea, opisthobranchs follow an opposite trend with 51 species found only in the Gulf of Eilat but not in the southern Red Sea. It remains to be seen whether these observations are real (possibly due to differences in habitat, shipping or other causes) or artificial (and may decrease with increased taxonomic effort). This "anomalous" distribution of opisthobranchs will certainly change as numerous species are not vet formally described and others recently described may yet be found in the Indian Ocean or further afield. However, others have been shown to be different from their Indian Ocean counterparts, demonstrating a real difference with the Indian Ocean.

No recent surveys have been carried out in the Gulf of Suez following the Cambridge and Dollfus expeditions in the 1920s. A study based on collections by the Hebrew University and Smithsonian Institute (1967 - 1969) provided only six opisthobranch species: of these, four species were previously recorded from the Gulf of Suez and two species were Indo-Pacific species already known from the Red Sea. However, *Diniatys dentifer* (A. Adams in Sowerby) is a west Pacific species and was only recorded from the Gulf of Suez but just recently it has been recorded from the Gulf of Eilat (Yonow, 2008). Opisthobranchs also differ from the general molluscan rule of greatest affinity to the Gulfs of Aden and Arabia and the western Indian Ocean: a few opisthobranch species which were thought to be endemic have recently been found in the Gulf of Oman and the Gulf of Aden. There are only 8.5% in common with the western Indian Ocean compared to 31% affinity with the broader Indo-West Pacific region, in contrast to other groups for which information is available. For example, cluster analysis of coral species show greatest affinities with the Gulf of Aden and the Persian Gulf species, and this cluster shows affinities with the western Indian Ocean fauna (Shepard and Sheppard, 1991). Polychaetes show a very high affinity with the nearby regions of the Gulf of Aden, Arabian Sea, and Arabian Gulf, and Wehe

Region	Opisthobran	ch numbers	Opisthobrand	ch percentages
Region	Red Sea	Gulf of Eilat	Red Sea	Gulf of Eilat
Endemic	80	21	26.67%	41.18%
Western Indian Ocean	25	3	8.33%	5.88%
Indo-West Pacific	5	1	1.67%	1.96%
West Pacific-Pacific	93	11	31.00%	21.57%
Circumtropical	44	10	14.67%	19.61%
	12	0	4.00%	0.00%
Mediterranean	4	1	1.33%	1.96%
Not assessed-NW Atlantic	37	4	12.33%	7.84%
Total	300	51	100.00%	100.00%

Table 1. Zoogeographical Affinities of Red Sea and Gulf of Eilat Opisthobranchs

and Fiege (2002) calculate 20% endemism for the "Arabian region" alone. Chitons also show similar affinities with all the non-endemic species also recorded from the western Indian Ocean (Strack, 1993).

Of the 300 species recorded in total in the Red Sea (see list on pp. 192 - 196) 51 species are found only in the Gulf of Eilat while 249 species are common to both areas. Of the 51 species recorded only in the Gulf of Eilat, ten are western Pacific species but not found in the Red Sea or Indian Ocean, and only three are western Indian Ocean species (and one Indian Ocean species). None of the 21 species recorded only from the Gulf of Eilat are named. As many as 15 are possibly new species; the remainder belong to groups in need of taxonomic revision or whose food items may be more widely distributed. The Indian Ocean and Indo-West Pacific species are not problematic, in view of marine species assemblages. The ten western Pacific species not recorded in the Red Sea or Indian Ocean pose more of a problem. One probable suggestion is that the larvae travelled in the ballast tanks of large ships. This appears to have been the distribution of many invasive species all over the world. Another possibility is transport via the hulls of these ships: a whole invertebrate community can exist on these hulls, and it is feasible that opisthobranch larvae, most likely nudibranchs, could settle and metamorphose on the invertebrate hull community as the ship travels, growing and eventually laying before dying. The egg mass would develop and the larvae hatch weeks later in a different sea. Enquiries were made to the Eilat Port Authority to obtain an idea of the number of ships entering the port (there was no information available for the port of Aqaba). In 2004, 2.3 million tons of cargo arrived on ships compared to 2.8 million tons for 1995. Although most ships come to unload cargo, a few come in ballast, and these are the ones of interest for larval transfer. Israel has already placed safety measures on ballast water (IMO 1999) but if none of the other countries bordering the Red Sea adheres to this, alien species will certainly continue to invade the Red Sea. Colour photographs of some sea slugs from the Gulf are presented on two colour plates, pp 190-191.

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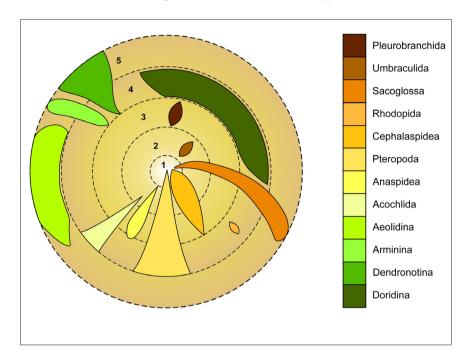
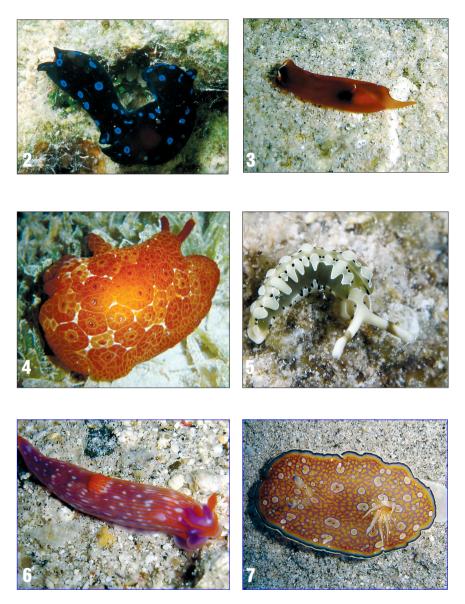
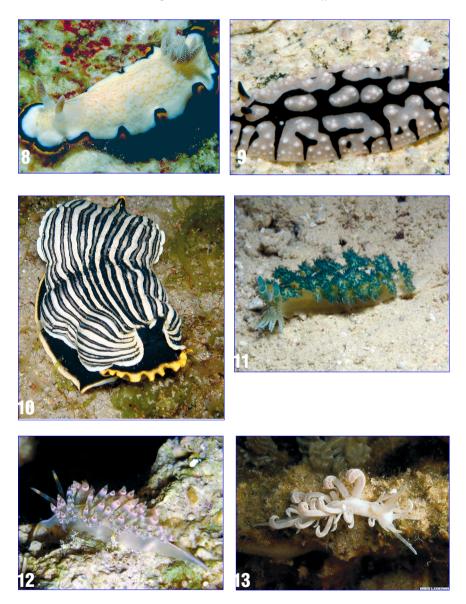


Figure 1. The levels or stages in the evolution of opisthobranchs from the heavily shelled species to the naked ones. The area of each group approximates the number of species it contains . 1 animals retain a spiral shell and operculum. 2 reduced shell, displays enlargement of body. Slug-like body, internal shell, single gill remaining. 4 bilaterally symmetrical bodies, no shell, secondary gills developing. 5 digestive gland extending into cerata (adapted from Willan & Morton 1984, drawn by Anna Ratcliffe).



- Figure 2. Chelidonura livida Yonow, 1994 50 mm (photo J. Kuchinke)
 Figure 3. Chelidonura sp. 10 mm (photo B. & S. Koretz)
 Figure 4. Pleurobranchus forskalii Rüppell & Leuckart, 1828 100 mm (photo B. & S. Koretz)
 Figure 5. Thuridilla decorata (Heller & Thompson, 1983) 20 mm (photo O. Lederman)
 Figure 6. Analogium sp. 35 mm (photo B. & S. Koretz)
 Figure 7. Chromodoris charlottae (Schrödl, 1999) 40 mm (photo B. & S. Koretz)



- Figure 8. Chromodoris obsoleta Rüppell & Leuckart, 1828 50 mm (photo G. Smith)
 Figure 9. Phyllidiopsis sinaiensis (Yonow, 1988) 85 mm (photo N. Yonow)
 Figure 10. Marioniopsis cyanobranchiata (Rüppell & Leuckart, 1828) 45 mm (photo J. Kuchinke)
 Figure 11. Armina sp. 100 mm (B. & S. Koretz)
 Figure 12. "Facelina" sp. 25 mm (photo B. & S. Koretz)
 Figure 13. Phyllodesmium colemani 20 mm (photo O. Lederman)

LIST OF SPECIES RECORDED FROM THE RED SEA

The version published here is modified from a similar list including all published records of every species recorded in the Red Sea (Yonow 2008). This list has been coded to show endemic species, geographical distribution of species, and their presence (or absence) in the Gulf of Eilat. Some of the unnamed species in the list are misidentifications from the older literature, a dozen are present as specimens in collections (N. Yonow), but most are photographic records only, of which a number, especially most of those recorded from the Gulf of Eilat, can be found at http://www.koretz.net/Eilat/Invertebrates/ Molluscs/Opistobranchs.htm. A plethora of websites now exist illustrating diving holiday snaps and have not been included here due to the unreliability of locality data, but imply that there may be still more species recorded from the Red Sea.

Key: Endemic Species. Western Indian Ocean.	Indian Ocean. Indo-West Pacific.	West pacific. Circumtropical.
Mediterranean. ???unkown.		

Mediterranean. ???unkown.	
Also recorded in Eilat.	Only recorded in Eilat.
Order Cephalaspidea	
i	00000
Acteon sp.	<u>??????</u> ??????
Pupa affinis (A. Adams, 1855)	277777 777777
Pupa tessellata (Reeve, 1842)	
Pupa cf. solidula (Linnaeus, 1758)	<u>999999</u>
Pupa sp.	22222
Hydatina physis (Linnaeus, 1758)	Circumtropical
Hydatina zonata (Lightfoot, 1786)	Indo-West Pacific
Ringicula acuta Philippi, 1849	??????
Colpodaspis thompsoni Brown, 1979	Indian Ocean
Acteocina fusiformis (A. Adams, 1850)	<u>??????</u>
Acteocina inconspicua (H. Adams, 1852)	222222
Acteocina involuta (Nevill, 1871)	??????
Acteocina simplex (A. Adams, 1850)	222222
Utriculastra mucronata (Philippi, 1849)	Lessepsian migrant
Cylichna mongii (Audouin 1826)	
Cylichna villersii Audouin, 1826	
Roxania lithiensis (Sturany, 1903)	
Retusa desgenettii (Audouin, 1826)	
<i>Pyrunculus fourierrii</i> (Audouin, 1826)	Indo-West Pacific (Lessepsian migrant)
Retusa tarutana Smythe, 1979	Western Indian Ocean
Ventomnestia girardi (Audouin, 1826)	West Pacific (Lessepsian migrant)
Philine vaillanti Issel, 1869	
Chelidonura flavolobata Heller & Thompson, 1983	
Chelidonura fulvipunctata Baba, 1938	Indo-West Pacific (Lessepsian migrant)
Chelidonura livida Yonow, 1994	Western Indian Ocean
Chelidonura punctata Eliot, 1903	Western Indian Ocean
Chelidonura sandrana Rudman, 1973	Western Indian Ocean
Philinopsis cyanea (Martens, 1879)	Indo-West Pacific
Philinopsis esticulata (Eliot, 1904)	Western Indian Ocean
Odontoglaja guamensis Rudman, 1978	West Pacific
Chelidonura sp.1	west I achie
Chelidonura sp.2	
Chelidonura sp.2 Chelidonura sp.3	22222
	<u>???????</u>
Chelidonura sp.4	??????
Philinopsis sp.1	277777
Philinopsis sp.2	
Sagaminopteron cf. ornatum Tokiota & Baba, 1964	West Pacific 4 Western Indian Ocean
Sagaminopteron cf. psychedelicum Carlson & Hoff, 197-	4 western mulan Ocean
Siphopteron sp.	
Atys cylindricus (Helbling, 1779)	Indo-West Pacific
Atys ehrenbergi Issel, 1869	
Atys naucum (Linnaeus, 1758)	Indo-West Pacific
Atys semistriatus Pease, 1860	West Pacific
Diniatys dentifer (A. Adams, 1850)	West Pacific
Diniyatys dubius (Schepman, 1913)	West Pacific
Haminoea cyanocaudata Heller & Thompson, 1983	
Haminoea cyanomarginata Heller & Thompson, 1983	
Haminoea pemphis (Philippi, 1847)	???????
Liloa curta (A. Adams, 1850)	West Pacific
Phanerophthalmus albocollaris Heller & Thompson, 1	1983
Phanerophthalmus olivaceus (Ehrenberg, 1828)	Indo-West Pacific (+ P. smaragdina)
Smaragdinella calyculata (Broderip & Sowerby, 1829)	Indo-West Pacific
Smaragdinella sieboldi A. Adams, 1864	West Pacific

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Bulla ampulla Linnaeus, 1758	Indo-West Pacific (Lessepsian migrant)
Order Anaspidea	
Akera soluta (Gmelin, 1791)	Indo-West Pacific
Aplysia cornigera (Sowerby, 1869)	Indo-West Pacific
Aplysia dactylomela (Rang, 1828)	Circumtropical
Aplysia fasciata (Poiret, 1789)	Mediterranean
Aplysia oculifera (Adams & Reeves, 1850)	Indo-West Pacific
Aplysia parvula (Guilding in Morch, 1863)	Circumtropical
Aplysia sp.	Western Indian Ocean
Dolabella auricularia (Lightfoot, 1786)	Indo-West Pacific
Dolabrifera dolabrifera (Curvier, 1817)	Circumtropical
<i>Phyllaplysia</i> sp.	??????
Petalifera petalifera Rang, 1828	Mediterranean
Petalifera ramosa Baba, 1959	West Pacific
Syphonota geographica (Adams & Reeve, 1850)	Circumtropical
Bursatella leachii savignana (Audouin, 1826)	Circumtropical
Notarchus indicus (Schweigger, 1820)	Circumtropical
Stylocheilus longicauda (Quoy & Gaimard, 1824)	Circumtropical
Stylocheilus striatus (Quoy & Gaimard, 1832)	Circumtropical
Orden Dieuwehmen ehide	
Order Pleurobranchida	Malianna (L. 1997)
Berthella stellata (Risso, 1826)	Mediterranean (+ circumtropical)
Berthella citrina (Ruppell & Leuckart, 1828) Berthella saidensis (O'Donoghue, 1929)	Indo-West Pacific
	Indo-West Pacific
Pleurobranchus albiguttatus (Bergh, 1905) Pleurobranchus forskalii (Ruppell & Leuckart, 1828)	Indo-West Pacific (Lessepsian migrant)
Pleurobranchus grandis Pease, 1868	West Pacific
Pleurobranchus cf. grandis (Pease, 1868)	west I achie
Euselenops luniceps (Cuvier, 1817)	Indo-West Pacific
Euserenops (eurier, 1017)	
Order Sacoglossa	
Oxynoe viridis (Pease, 1861)	Indo-West Pacific
Julia exquisita (Gould, 1862)	Indo-West Pacific
Elysia tomentosa Jensen, 1997	Indo-West Pacific
Elysia trilobata Heller & Thompson, 1983	
Elysia sp. l	
Elysia sp.2	
Elysia sp.3	Inda West Destes
Elysiella pusilla Bergh, 1872	Indo-West Pacific 222222
Elysiella sp. Thuridilla decorata (Heller& Thompson, 1983)	
<i>Thuridilla</i> ? indopacifica Gosliner, 1995	Indian Ocean
Thuridilla sp.	Indian Occan
Plakobranchus ocellatus van Hasselt, 1824	Indo-West Pacific
Cyerce elegans Bergh, 1870	West Pacific
Polybranchia orientalis (Kelaart, 1858)	Indo-West Pacific
Stiliger ornatus Ehrenberg, 1828	Indo-West Pacific
Order Nudibranchia Doridina - Phanerobranchs	
Goniodoridella savignyi Pruvot-Fol, 1933	Indo-West Pacific
Goniodoris joubini (Risbec, 1928)	West Pacific
Goniodoris castanea (Alder & Hancock, 1845)	north-western Europe
Trapania toddi Rudman, 1987	West Pacific
Goniodorid	??????
Analogium striatum (Eliot, 1908) Analogium sp.	Indo-West Pacific
Gymnodoris alba (Bergh, 1877)	West Pacific
Gymnodoris alba (Beigh, 1877) Gymnodoris ceylonica (Kelaart, 1858)	The state of the s
Gymnodoris citrina (Bergh, 1875)	Indo-West Pacific
<i>Gymnodoris impudica</i> (Ruppell & Leuckart, 1828)	Indo-West Pacific (= G. rubropapulosa)
<i>Gymnodoris inornata</i> (Bergh, 1880)	Indo-West Pacific
<i>Gymnodoris cf. nigricolor</i> (Baba, 1960)	West Pacific
Gymnodoris sp.1	
<i>Gymnodoris</i> sp.2	Indo-West Pacific
Nembrotha megalocera Yonow, 1990	

$T = 1$: $C = (T_{1}) + 1004$	Lada Wast Daste
Tambja affinis (Eliot, 1904)	Indo-West Pacific Indo-West Pacific
Tambja limaciformis (Eliot, 1908)	Indo-West Facilic Indo-West Pacific
Thecacera pacifica (Bergh, 1883) Kaloplocamus sp.	222222
Plocamopherus indicus (Bergh, 1890)	Indo-West Pacific
Plocamopherus ocellatus (Ruppell & Leuckart, 1828)	Lessepsian migrant
Aegires citrinus Pruvot-Fol, 1930	Indo-West Pacific
Aegires villosus Farran, 1905	Indo-West Pacific
Hexabranchus sanguineus (Ruppell & Leuckart, 1830)	indo west i denie
Hexabranchus sp.	West Pacific
1	
Doridina - Cryptobranchs	
Actinocyclus sordida (Ruppell & Leuckart, 1830)	
Actinocyclus verrucosus Ehrenberg, 1831	Indo-West Pacific
Hallaxa indecora (Bergh, 1905)	Indo-West Pacific
Hallaxa sp.	
Discodoris fragilis (Alder & Hancock, 1864)	Indo-West Pacific (Lessepsian migrant)
Discodoris granulata (Ehrenberg, 1831)	W (D)
Discodoris schmeltziana (Bergh, 1880)	West Pacific
Discodoris sp.1	?????? ??????
Discodoris sp.2	Indo-West Pacific
Paradoris erythraeensis (Vayssiere, 1912) Sebadoris nubilosa (Pease, 1871)	Western Indian Ocean
Aldisa sp.1	Western Indian Ocean
Aldisa sp.2	Western Indian Ocean
Archidoris vayssierea O'Donoghue, 1929	
Archidoris sp.	??????
Atagema ornata (Ehrenberg, 1831)	Indo-West Pacific (= Trippa intecta)
Atagema spongiosa (Kelaart, 1858)	Indo-West Pacific (= Trippa areolata)
Atagema sp.1	
Atagema sp.2	??????
Doriopsis granulosa Pease, 1860	Indo-West Pacific
Doris sp.1	222222
Doris sp.2	22222 22222
Doris sp.3 Doris sp.4	<u>???????</u>
Hoplodoris grandiflora (Pease, 1860)	Indo-West Pacific
Hoplodoris pustulata Abraham, 1877	Indo-West Pacific (+ <i>H. bifurcata</i>)
Hoplodoris sp.	??????
Hoplodoris cf. armata (Baba, 1993)	???????
Sclerodoris apiculata (Alder & Hancock, 1864)	Indo-West Pacific
Sclerodoris tuberculata Eliot, 1904	Indo-West Pacific
Sclerodoris cf. rugosa (Vayssiere, 1912)	??????
Spongiodoris sp.	
Artachaea clavata Eliot, 1908	
Artachaea verrucosa Eliot, 1908	Indo West Decife
Asteronotus cespitosus (van Hasselt, 1824)	Indo-West Pacific
Asteronotus denticulata (Eliot, 1908) Halgerda willeyi Eliot, 1904	Indo-West Pacific
Halgerda sp.	indo-west i acine
Jorunna funebris (Kelaart, 1858)	Indo-West Pacific
Jorunna pantherina Angas, 1864	West Pacific
Jorunna sp.1	Western Indian Ocean
Jorunna sp.2	??????
Platydoris scabra (Cuvier, 1804)	Indo-West Pacific
Platydoris striata (Kelaart, 1858)	Western Indian Ocean
Platydoris sp.	??????
Ardeadoris egretta Rudman, 1984	West Pacific
Cadlinella ornatissima (Risbec, 1928)	Indo-West Pacific
Cadlinella sp.	
Chromodoris africana Eliot, 1904	Western Indian Ocean
Chromodoris annulata Eliot, 1904	Western Indian Ocean
Chromodoris aspersa (Gould, 1852) Chromodoris charlottae (Schrodl, 1999)	Indo-West Pacific
Chromodoris charlottae (Schrodi, 1999) Chromodoris fidelis (Kelaart, 1858)	Indo-West Pacific
Chromodoris giaetis (Keldali, 1858) Chromodoris geminus Rudman, 1987	Indian Ocean
Chromodoris geminus Rudinan, 1987 Chromodoris obsoleta (Ruppell & Leuckart, 1830)	
Chromouoris obsoletii (Ruppell & Leuckart, 1050)	

Chromodoris auadricolor (Ruppell & Leuckart, 1830) Chromodoris strigata Rudman, 1980 Chromodoris tennentana (Kelaart, 1859) Chromodoris tinctoria (Ruppell & Leuckart, 1830) Chromodoris verrieri (Crosse, 1875) Chromodoris sp. Durvilledoris lemniscata (Quoy & Gaimard, 1832) Durvilledoris pusilla (Bergh, 1874) Glossodoris atromarginata (Cuvier, 1804) Glossodoris cincta (Bergh, 1888) Glossodoris electra Rudman, 1990 West Pacific Glossodoris hikuerensis (Pruvot-Fol, 1954) Glossodoris pallida (Ruppell & Leuckart, 1830) Glossodoris plumbea (Pagenstecher, 1877) Glossodoris symmetricus Rudman, 1990 ?????? Glossodoris sp. Hypselodoris dollfusi (Pruvot-Fol, 1833) Hypselodoris infucata (Ruppell & Leuckart, 1830) Hypselodoris maculosa (Pease, 1871) Hypselodoris maridadilus Rudman, 1977 Hypselodoris nigrostriata (Eliot, 1904) Mexichromis katalexis Yonow, 2001 Noumea alboannulata Rudman, 1986 West Pacific Noumea flava (Eliot, 1904) Noumea norba Marcus & Marcus, 1907 Noumea simplex (Pease, 1871) West Pacific Noumea sudanica Rudman, 1985 Noumea sp. Risbecia ghardaqana (Gohar & Aboul-Ela, 1957) Risbecia pulchella (Ruppell & Leuckart, 1830) Thorunna africana Rudman, 1984 Ceratosoma tenue Abraham, 1876 Ceratosoma trilobatum (Gray, 1827) Miamira magnifica Eliot, 1910 Orodoris miamirana Bergh, 1875 Doridina - Porostomes Dendrodoris coronata Kay & Young, 1969 Dendrodoris cuprea (Ehrenberg, 1831) West Pacific Dendrodoris elongata Baba, 1936 Dendrodoris fumata (Ruppell & Leuckart, 1830) Dendrodoris jumata (Rappen & Ledekar, 10 Dendrodoris immaculata (Audouin, 1826) Dendrodoris jousseaumei (Vayssiere, 1912) Dendrodoris leptopus Ehrenberg, 1831 Dendrodoris lugubris Ehrenberg, 1831 Dendrodoris nigra (Stimpson, 1955) Dendrodoris nigropunctata (Vayssiere, 1912) Dendrodoris tigrina (Audouin, 1826) Dendrodoris tuberculosa (Quoy & Gaimard, 1832) Dendrodoris sp. ?????? Doriopsilla sp. Fryeria rueppelii (Bergh, 1869) Phyllidia multifaria Yonow, 1986 Phyllidia schupporum Fahrner & Schrodl, 2000 Phyllidia undula Yonow, 1986 Phyllidia varicosa Lamarck, 1801 Phyllidia sp. Phyllidiella pustulosa (Cuvier, 1804) Phyllidiopsis cardinalis Bergh, 1873 Phyllidiopsis dautzenbergi (Vayssiere, 1912) Phyllidiopsis monacha (Yonow, 1988) Phyllidiopsis sinaiensis (Yonow, 1988)

Western Indian Ocean Indo-West Pacific Western Indian Ocean Arabian Gulf (assessed as WIO) Indo-West Pacific Western Indian Ocean **Indo-West Pacific** Arabian Gulf (assessed as WIO) Indo-West Pacific (Lessepsian migrant) Indo-West Pacific Western Indian Ocean Western Indian Ocean Indo-West Pacific Indo-West Pacific Indo-West Pacific Indo-West Pacific Western Indian Ocean Western Indian Ocean Indo-West Pacific Indo-West Pacific Western Indian Ocean Indo-West Pacific Indo-West Pacific Indo-West Pacific Indo-West Pacific Indo-West Pacific Western Indian Ocean Western Indian Ocean Indo-West Pacific

Indo-West Pacific

Indo-West Pacific

Dendronotina

Marioniopsis glama (Ruppell & Leuckart, 1828) Marioniopsis cyanobranchiata (Ruppell & Leuckart, 1828)

Marioniopsis levis (Eliot, 1904)	Western Indian Ocean
Marioniopsis rubra (Ruppell & Leuckart, 1828)	
Marioniopsis viridescens (Eliot, 1904)	Western Indian Ocean
Tritoniopsis elegans (Audouin, 1826)	Indo-West Pacific
tritonid sp.1	??????
tritonid sp.2	???????
	Indo-West Pacific
Bornella stellifer (Adams & Reeve in Adams, 1848)	muo-west racinc
Bornellopsis kabretiana O'Donoghue, 1929	
Doto orcha Yonow, 2000	
Lomanotus vermiformis Eliot, 1908	West Pacific
Scyllaea pelagica Linnaeus, 1758	Circumtropical
Crosslandia viridis Eliot, 1902	Indo-West Pacific
Melibe bucephala Bergh, 1902	Indo-West Pacific
Melibe rangii Bergh, 1875	West Pacific (= M. engeli)
Arminina	
Armina semperi (Bergh, 1860)	West Pacific
Armina sp.1 cf. major Baba, 1955	
Armina sp.1 cj. major Duota, 1955	
Dermatobranchus albus (Eliot, 1904)	Western Indian Ocean
	western mulan Ocean
Dermatobranchus glabrus (Eliot, 1908)	W/ (D 10
Dermatobranchus gonatophorus van Hasselt, 1824	West Pacific
Dermatobranchus ornatus (Bergh, 1874)	West Pacific
Dermatobranchus striatus van Hasselt, 1824	West Pacific
Dermatobranchus sp.1	
Dermatobranchus sp.2	
	Ind. West Dester
Madrella ferruginosa Alder & Hancock, 1864	Indo-West Pacific
Janolus sp.	
Aeolidina	
Flabellina bicolor (Kelaart, 1858)	Indo-West Pacific
<i>"Flabellina"</i> sp.	
Flabellina bilas Gosliner & Willan, 1991	West Pacific
	Indo-West Pacific (Lessespian migrant)
Flabellina rubrolineata (O'Donoghue, 1929)	
Flabellina rhodopos Yonow, 2000	Western Indian Ocean
Flabellina rhodopos Yonow, 2000 "Facelina" sp.	
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