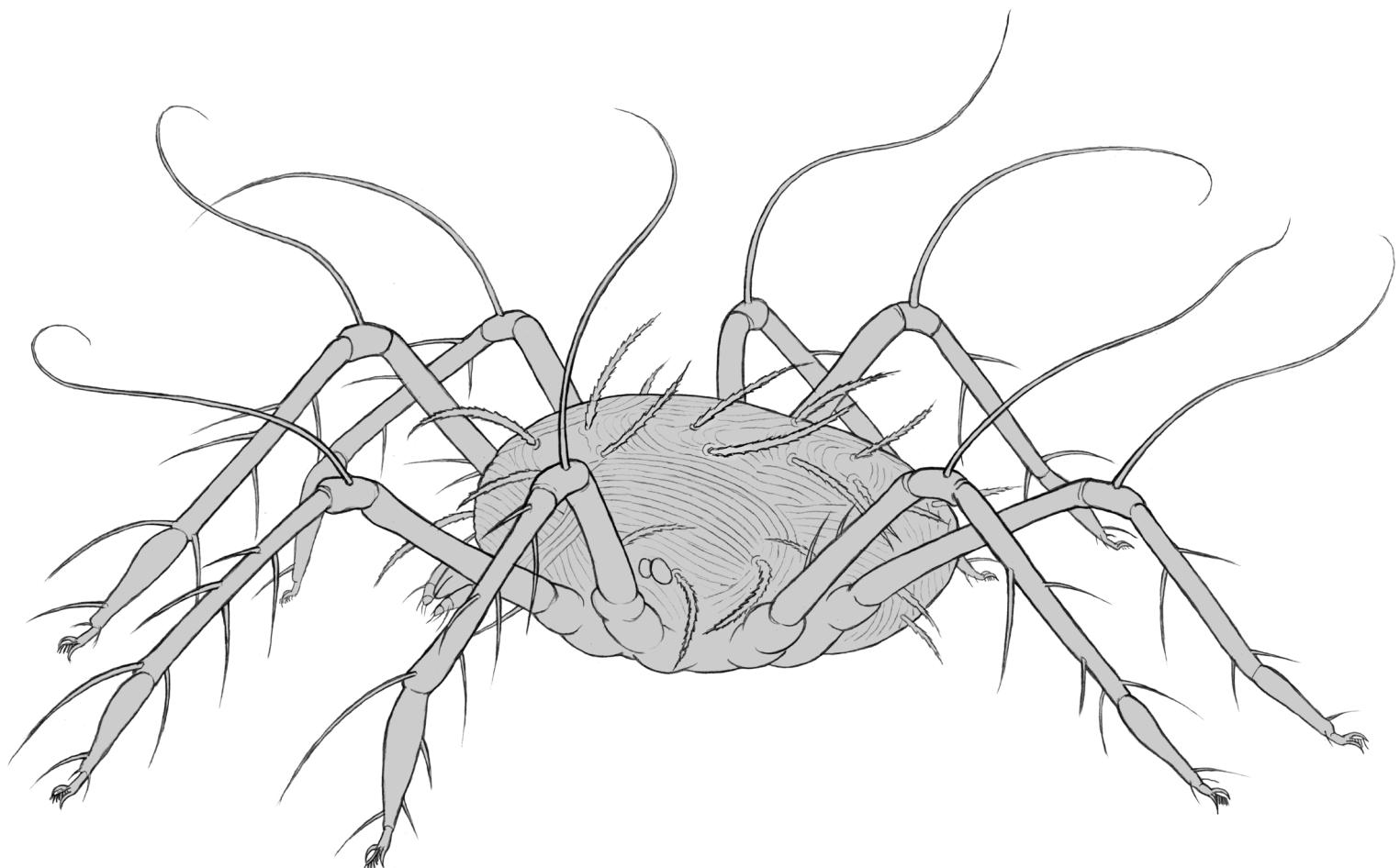


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ACARI – Bibliographia Acarologica – now in its 18th year – aims to advance and help propagate acarological knowledge. To this end, each year we compile all internationally available papers published on Acari, as far as they become known to us. As always, two major taxon groups are excluded from this bibliography on the paraphyletic Actinedida – the Eriophyidae and the paraphyletic “Hydracarina” since literature databanks of these groups are available elsewhere.

With more than 300 papers recorded this year, the present bibliography is somewhat less than the last few years, but on average for actinedid research in the last decade. Research on 38 families is reported in this issue, of the more than 160 families known for Actinedida. Almost half of the papers once again deal with the economically important Tetranychidae, with much work also on other health and food-safety relevant taxa such as Tenuipalpidae, Tarsonemidae and Trombiculidae. Strongly represented this year are as usual Heterostigmata (ca. 17%) and Parasitengona (14% of all papers, the majority dealing with chiggers and Erythraeidae). Stigmaeidae are represented by 10 the publications (about 4%). Endeostigmata are again not reported at all this year. It is interesting to observe how the taxonomic distribution of actinedid research remains very similar from year to year, representing not only economic interests, but probably mostly the taxonomic expertise of current researchers. As the number of taxonomists declines worldwide, the diversity of taxonomic research and thus our knowledge on biodiversity continues to become increasingly depauperate.

Economically important research dominates actinedid research; this year general plant (crop) protection topics – i.e., acarine-pest biology, biological mite control (including predator-prey relationships) and the ecology/biology of plant pests – again account for the majority (over 40%) of all papers. Systematics and taxonomy remain the second-most important topic (39% of all papers), with over 100 descriptions of new species and 13 new genera in about 80 papers. Although the number of morphological papers has increased somewhat, the number of species descriptions has steadily declined in the last five years, again indicating how basic taxonomic research of this mite group is steadily deteriorating. On the other hand, molecular biological research on Actinedida continues to increase (20 papers this year), almost all related to the economically important Tetranychidae.

Taxonomic revisions and determination keys still remain sorely needed for most soil-living families and genera, their availability will help promote ecological field research on Actinedida. The present bibliography does include a number of keys, i.e. for both Serbian and Saudi Arabian Tetranychidae or the subfamily Pteridopodinae (Chyzeriidae; including a revision). Most keys are for species of specific genera, such as *Neobonzia* (Cunaxidae), Larval *Balaustium* (Erythraeidae), *Eryngiopus* and *Agistemus* (Stigmaeidae) as well as *Obuloides* and *Terminalichus* (Tenuipalpidae). A new Brazilian database on Tetranychidae is reported by Flechtmann et al. 2018.

We once again point out the lack of general ecological research, considering that Actinedida represent one of the most abundant soil-microarthropod groups. Research relating to general biology of these mites continues to decline from year to year. Only three papers in the present volume deal explicitly with Actinedida in the soil fauna. Nonetheless, a number of checklists are reported in this volume, which can be valuable for general biodiversity studies, including checklists for marine littoral mangroves, Israeli Tenuipalpidae, Hungarian Tetranychidae and Tenuipalpidae, but also for African and Brazilian chiggers.

Due to the new European data-protection legislation, we no longer collect the addresses and e-mails of the first authors. This information can normally be found in the publications themselves. We therefore can no longer report

on the geographic distribution of Actinedida research around the globe. However, it remains notable how much research originates from Middle-Eastern and Asian countries, increasing the global taxonomic knowledge of actinedid mites as these areas continue to be a centre of recent basic acarological taxonomic research.

The acarological literature collection and databank in Görlitz is one of the largest in the world. The databank of Actinedid literature cited in ACARI has now accumulated 8,699 papers on 3,944 species of actinedid mites. The databank as well as previous issues of ACARI can be accessed via <http://www.senckenberg.de/Acari>.

Reprints of the majority of cited papers are present in the Chelicerata Department of the Senckenberg Museum of Natural History in Görlitz. The registration of all recent publications on actinedid mites is a daunting and time-consuming task, which cannot be undertaken without the aid of all acarologists worldwide. We expressly thank all authors who have assisted this goal and sent reprints of their papers. We nonetheless ask for your continued help by sending reprints or copies of all your papers on actinedid mites. As with any journal, mistakes and omissions are unavoidable. Critique and suggestions are welcome and explicitly called for. Please inform us if we have failed to list any of your publications in the Bibliographia and we will include them in later volumes..

Acarological literature

Literature citations printed in bold type contain descriptions of new species. Titles marked with “*” were only found as a citation or abstract.

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Nomina nova

The names of new taxa are listed here as far as we have received the papers. Their validity was not examined here. The authors of new combinations and new synonyms are written in [brackets].

Type-material information as follows:

Zetziella kamali Akyol & Güл, 2018 (Page: 464¹) –
TYPES: HT² + PT² - CBZM³

1 – first page of the description

2 – holotype (HT), paratypes (PT) or allotypes (AT)

3 – abbreviations of the places of storage of new types, as far as they were cited in the publications

Abbreviations of the places of storage of new types

ABUH - Al-Baath University, Department of Plant Protection, Faculty of Agriculture, Homs, Syria

ACASI - Acarological Collection, Acarological Society of Iran, University of Tehran, Karaj, Iran

AETMU - Acarological Collection, Department of Entomology, Faculty of Agriculture, Tarbiat Modares University, Tehran, Iran

ALUG - Acarology Laboratory, Department of Plant Protection, University of Guilan, Guilan, Iran

ALUM - Acarology Laboratory, Department of Plant Protection, University of Maragheh, Maragheh, Iran

AMU - Adam Mickiewicz University, Natural History Collection, Poznań, Poland

ANIC - Australian National Insect Collection, CSIRO Division of Entomology, Canberra, Australia

BASU - Bu-Ali Sina University, Acarology Laboratory, Hamedan, Iran

BZOL - BiologieZentrum des Oberösterreichischen Landesmuseums, Linz, Austria

CALBS - Collection of the Acarology Laboratory, Uni-

versity of Bu-Ali Sina, Hamadan, Iran

CBD - Collection of Burkhard Dietrich, Sarstedt, Germany

CBZM - Celal Bayar University, Zoological Museum, Manisa, Turkey

CECOUAL - Centro de Investigación de Colecciones Científicas de la Universidad de Almería, Almeria, Spain

CHG - Collection of Heinrich Grabenhorst, Wienhausen, Germany

CNC - Canadian National Collection of Insects, Arachnids and Nematodes, Ottawa, Canada

DATE - Department of Animal Taxonomy and Ecology, Adam Mickiewicz University, Poznań, Poland

DZSJRP - Departamento de Zoologia, Campus de S.J. do Rio Preto, Universidade Estadual Paulista, Sao Paulo, Brazil

DZUB - Department of Zoology, University of Brasilia, Brasilia, Brazil

ESALQ/USP - Escola Superior de Agricultura “Luiz de Queiroz”, Universidade de São Paulo, Departamento de Entomologia e Acarologia, Piracicaba, Brazil

FAFU - Fujian Agricultural and Forestry University, Department of Plant Protection, Fuzhou, China

FMNH - Field Museum of Natural History, Chicago, USA

FUM - Department of Plant Protection, Ferdowsi University of Mashhad, Mashhad, Iran

GPIH - Geologisch-Paläontologisches Institut der Universität Hamburg, Hamburg, Germany

GUGC - Guizhou University, Institute of Entomology, Guiyang, Guizhou, China

GXU - GuangXi University, Department of Plant Protection, Nanning, China

HUM - Hokkaido University Museum, Sapporo, Japan

IPCAS - Institute of Parasitology, Biology Centre Academy of Sciences of the Czech Republic, České Budějovice, Czech Republic

IRSNB - L'Institut Royal des Sciences Naturelles, Bruxelles, Belgium	Beltsville, USA
JAZM - Jalal Afshar Zoological Museum, Acarological Collection, University of Tehran, Karaj, Iran	NMNST - National Museum of Nature and Science, Tsukuba, Japan
KFUG - Karl-Franzens-University, Institute of Zoology, Graz, Austria	NSMT - National Museum of Nature and Science, formerly National Science Museum, Tokyo, Japan
KSMA - King Saud University Museum of Arthropods, Riyadh, Saudi Arabia	NWUP - North-West University, Potchefstroom, South Africa
MAI - Museum of Amber Inclusions, Department of Invertebrate Zoology and Parasitology, University of Gdańsk, Gdańsk, Poland	NZAC - New Zealand Arthropod Collection, Landcare Research, Auckland, New Zealand
MCN - Museu de Ciencias Naturais da Univates Centro Universitário, Lajeado, Brazil	NZMC - National Zoological Museum of China, Institute of Zoology, Chinese Academy of Sciences, Beijing, China
MIZ - Museum and Institute of Zoology, Polish Academy of Sciences, Warszawa, Poland	ONUDZ - I.I. Mechnikov Odessa National University, Department of Zoology, Odessa, Ukraine
MNCN - Museo Nacional de Ciencias Naturales, Madrid, Spain	OSAL - Ohio State University, Museum of Biological Diversity, Acarology Laboratory, Columbus, Ohio, USA
MNHNP - Museum of Natural History, Podgorica, Montenegro	QM - Queensland Museum, South Brisbane, Queensland, Australia
MNHWU - Museum of Natural History, Wrocław University of Environmental and Life Sciences, Wrocław, Poland	SBUK - Collection of the Acarology Laboratory, Shahid Bahonar University of Kerman, Kerman, Iran
MPMT - Meguro Parasitological Museum, Tokyo, Japan	SMNG - Senckenberg Museum für Naturkunde Görlitz, Görlitz, Germany
MZLQ - Museu de Zoologia da Escola Superior de Agricultura "Luiz de Queiroz", Piracicaba, São Paulo, Brazil	TSUMZ - Tyumen State University Museum of Zoology, Tyumen, Russia
NCA-PPRI - South Africa National Collection of Arachnida (Acar), Plant Protection Research Institute, Pretoria, South Africa	TUAC - Tabriz University, Department of Plant Protection, Acarological Collection, Tabriz, Iran
NHME - Natural History Museum of Erfurt, Erfurt, Germany	UESC - Universidade Estadual de Santa Cruz, Laboratório de Entomologia, Ilhéus, Bahia, Brazil
NJAU - Nanjing Agricultural University, Department of Entomology, Jiangsu Province, Nanjing, China	UFLA - Universidade Federal de Lavras, Lavras, Minas Gerais, Brazil
NMB - National Museum Bloemfontein, Bloemfontein, South Africa	UFMG - Universidade Federal de Minas Gerais, Departamento de Zoologia, Colecao de Acarologia, Belo Horizonte, Brazil
NMK - National Museums of Kenya, Nairobi, Kenya	UGDIZP - University of Gdańsk, Department of Invertebrate Zoology and Parasitology, Gdańsk, Poland
NMNH - National Insect and Mite Collection, National Museum of Natural History, Smithsonian Institution,	

UMMZ - University of Michigan, Museum of Zoology, Ann Arbor, USA	<i>Cheiracus costus</i> Tan, Yang, Lai & Wang, 2015 (Page: 778) – TYPES: HT + PT - GXU
UNESP - Universidade Estadual Paulista, Campus de Sao José do Rio Preto, Sao Paulo, Brazil	<i>Chenaphila marmoronetta</i> Zmudzinski & Unsoeld, 2017 (Page: 480) – TYPES: HT + PT - AMU, PT - ZSM
UPLB - University of Philippines Los Banos, Museum of Natural History, Laguna, Philippinen	<i>Cheyletus rashtiensis</i> Salarzehi, Hajizadeh & Ueckermann, 2018 (Page: 641) – TYPES: HT + PT - ALUG, PT - NCA-PPRI
USNM - United States National Museum of Natural History, Washington, USA	<i>Cicaditrombium lorestanensis</i> Noei, 2017 (Page: 174) – TYPES: HT + PT - JAZM, PT - ACASI
ZISP - Zoological Institute, Russian Academy of Sciences, St. Petersburg, Russia	<i>Collemboerythraeus izadpanahi</i> Noei, Sabbori, Sedghi & Hakimitabar, 2017 (Page: 2) – TYPES: HT + PT - JAZM, PT - ACASI
ZSM - Zoologische Staatssammlungen, München, Germany	<i>Collemboerythraeus vosoughae</i> Noei, Saboori & Hakimitabar, 2017 (Page: 1259) – TYPES: HT + PT - JAZM, PT - ACASI

New species

<i>Abrolophus Marianopolicus</i> Haitlinger & Šundić, 2018 (Page: 354) – TYPES: HT + PT - MNHWU, PT - BZOL, MNHP	<i>Cretessenia roccamenaica</i> Haitlinger & Šundić, 2018 (Page: 421) – TYPES: HT - MNHWU
<i>Acarhis kleinhoviae</i> Tan, Yang, Lai & Wang, 2015 (Page: 770) – TYPES: HT + PT - GXU	<i>Cunaxa iranica</i> Paktinat-Saeij, Bagheri & Castro, 2017 (Page: 1278) – TYPES: HT + PT - ALUM, PT - JAZM, ACASI
<i>Agistemus jawadi</i> Rehman, Kamran & Alatawi, 2018 (Page: 1053) – TYPES: HT + PT - KSMA	<i>Cyta kreiteri</i> Barbar & Ueckermann, 2017 (Page: 1090) – TYPES: HT + PT - ABUH
<i>Armascirus raulzito</i> Rocha & Argolo, 2017 (Page: 114) – TYPES: HT - ESALQ/USP, PT - UESC	<i>Demodex bandicotae</i> Izdebska, Rolbiecki, Morand & Ribas, 2017 (Page: 1911) – TYPES: HT - UGDIZP, PT - MIZ
<i>Athyreacarus primitivus</i> Khaustov & Frolov, 2017 (Page: 502) – TYPES: HT + PT - UNESP, PT - TSUMZ, ZISP	<i>Demodex fusiformis</i> Izdebska & Rolbiecki, 2015 (Page: 608) – TYPES: HT - UGDIZP, PT - MIZ
<i>Bakerdania minuta</i> Khaustov & Minor, 2018 (Page: 290) – TYPES: HT + PT - NZAC, PT - TSUMZ	<i>Demodex marculus</i> Izdebska & Rolbiecki, 2015 (Page: 607) – TYPES: HT - UGDIZP, PT - MIZ
<i>Balaustium akramii</i> Noei, 2017 (Page: 2219) – TYPES: HT + PT - JAZM, PT - ACASI	<i>Diptacus genusetosus</i> Gol, Namaghi & Lillo, 2018 (Page: 1044) – TYPES: HT + PT - FUM
<i>Caesarodispus delabiei</i> Silva, Khaustov & Oliveira, 2018 (Page: 1258) – TYPES: HT - UESC, PT - ESALQ/USP	<i>Diptacus longiscatuber</i> Gol, Namaghi & Lillo, 2018 (Page: 1046) – TYPES: HT + PT - FUM
<i>Callidosoma cassiculophylla</i> Oliveira Bernardi, Wohltmann, Lornezon & Ferreira, 2017 (Page: 461) – TYPES: HT + PT - UFLA, PT - MZLQ, SMNG, UFMG	<i>Dividilobus parvifolius</i> Xu & Xue, 2017 (Page: 1401) – TYPES: HT + PT - NJAU, PT - NZMC
<i>Castosyringophilus meropis</i> Skoracki, Hromada & Sikora, 2017 (Page: 2) – TYPES: HT + PT - AMU, PT - IPCAS, ZSM, NMK	<i>Dolichocybe sibiricensis</i> Khaustov, 2017 (Page: 1679) – TYPES: HT - TSUMZ

- Eotetranychus spinophilus* Zhang, Beard & Seeman, 2017 (Page: 497) – TYPES: HT + PT - QM, PT - ANIC, USNM
- Excelsotarsonemus cabrucae* Sousa, Lofego & Ochoa, 2018 (Page: 71) – TYPES: HT + PT - DZSJRP, PT - NMNH, UESC, ESALQ/USP
- Fessonnia grabenhorsti* Bartel, Konkiewicz, Makol, Wohltmann & Dunlop, 2015 (Page: 653) – TYPES: HT - CHG, PT - CBD
- Fessonnia groehni* Bartel, Konkiewicz, Makol, Wohltmann & Dunlop, 2015 (Page: 655) – TYPES: HT - GPIH
- Fessonnia wunderlichi* Bartel, Konkiewicz, Makol, Wohltmann & Dunlop, 2015 (Page: 649) – TYPES: HT - MAI, PT - GPIH
- Formicomotes brasiliensis* Khaustov & Frolov, 2018 (Page: 394) – TYPES: HT + PT - UNESP, PT - TSUMZ, ZISP
- Herpetacarus junkeri* Stekolnikov, 2018 (Page: 271) – TYPES: HT + PT - IRSNB
- Hirstiosoma amfilohijei* Haitlinger & Šundić, 2017 (Page: 940) – TYPES: HT + PT - MNHWU
- Hoplocheylus evansi* Arjomandi & Hajiqanbar, 2018 (Page: 21) – TYPES: HT + PT - AETMU
- Iberochyzeria fornileesi* Mayoral, Welbourn & Barranco, 2018 (Page: 1130) – TYPES: HT + PT - MNCN, PT - CECOUAL, USNM
- Larvacarus iranicus* Mahdavi, Latifi, Asadi & Seeman, 2018 (Page: 217) – TYPES: HT + PT - SBUK, PT - QM
- Ledermuelleriopsis aminiae* Nazari & Khanjani, 2017 (Page: 194) – TYPES: HT + PT - BASU
- Leptus candangus* Šundić, Haitlinger & Pompermaier, 2017 (Page: 90) – TYPES: HT - DZUB, PT - MNHP, MNHWU
- Leptus darvishi* Saboori, Hakimitabar & Khademi, 2018 (Page: 90) – TYPES: HT + PT - JAZM, PT - NHME
- Lobotarsonemus betulae* Khaustov & Abramov, 2017 (Page: 1081) – TYPES: HT + PT - TSUMZ, PT - ZISP
- Makwacarus petrodromi* Stekolnikov, 2018 (Page: 275) – TYPES: HT + PT - IRSNB
- Microdispodides moseri* Khaustov, 2018 (Page: 442) – TYPES: HT + PT - TSUMZ
- Microtrombicula livingstonei* Stekolnikov, 2018 (Page: 281) – TYPES: HT + PT - IRSNB
- Nasutidania orientalis* Khaustov & Frolov, 2018 (Page: 285) – TYPES: HT + PT - ZISP, PT - TSUMZ
- Neobakerdania pilosa* Khaustov & Minor, 2018 (Page: 283) – TYPES: HT + PT - NZAC, PT - TSUMZ
- Neobonzia ermilovi* Corpuz-Raros, Naredo & Garcia, 2017 (Page: 916) – TYPES: HT + PT - UPLB, PT - OSAL
- Neobonzia neomalookensis* Chen, Guo, Yi & Jin, 2018 (Page: 105) – TYPES: HT + PT - GUGC
- Neophyllobius abiegnus* Khaustov & Abramov, 2017 (Page: 1386) – TYPES: HT + PT - TSUMZ
- Nothrypnacus buettnerus* Tan, Yang, Lai & Wang, 2015 (Page: 773) – TYPES: HT + PT - GXU
- Neoscirula klompeni* Corpuz-Raros, Naredo & Garcia, 2017 (Page: 923) – TYPES: HT + PT - UPLB, PT - OSAL
- Neoscirula lagunaensis* Corpuz-Raros, Naredo & Garcia, 2017 (Page: 929) – TYPES: HT + PT - UPLB, PT - OSAL
- Neoscirula lambatina* Corpuz-Raros, Naredo & Garcia, 2017 (Page: 933) – TYPES: HT + PT - UPLB
- Nothrotrombidium birjandensis* Noei, 2017 (Page: 162) – TYPES: HT + PT - JAZM
- Obuloides crinitus* Maake & Ueckermann, 2018 (Page: 996) – TYPES: HT + PT - NCA-PPRI
- Odontoscirus tixieri* Barbar & Ueckermann, 2017 (Page: 1095) – TYPES: HT + PT - ABUH
- Oligonychus hamedaniensis* Khanjani, Khanjani & Seeman, 2018 (Page: 235) – TYPES: HT + PT - CALBS, PT - QM
- Oligonychus shojaeii* Khanjani, Khanjani & Seeman, 2018 (Page: 225) – TYPES: HT + PT - CALBS, PT - QM
- Paracarophenax triplaxophilus* Khaustov & Abramov, 2018 (Page: 333) – TYPES: HT + PT - TSUMZ, PT - ZISP
- Pavania africana* Khaustov & Frolov, 2018 (Page: 134)

- TYPES: HT + PT - ZISP
- Pavania neotropica* Khaustov & Frolov, 2017 (Page: 508)
– TYPES: HT + PT - UNESP, PT - TSUMZ, ZISP
- Petalomium braziliensis* Silva, Khaustov & Oliveira, 2018
(Page: 1255) – TYPES: HT + PT - UESC, PT - DZSJRP,
ESALQ/USP
- Petalomium megasolenidiatum* Silva, Khaustov & Oliveira,
2018 (Page: 297) – TYPES: HT + PT - UESC, PT -
ESALQ/USP, UNESP, TSUMZ
- Petalomium verenae* Silva, Khaustov & Oliveira, 2017
(Page: 1804) – TYPES: HT + PT - UESC, PT - TSUMZ,
ESALQ/USP
- Podotarsonemus australiensis* Seeman, Lindquist & Husband, 2018 (Page: 10) – TYPES: HT + PT - QM, PT -
CNC, UMMZ
- Podotarsonemus boliviensis* Seeman, Lindquist & Husband, 2018 (Page: 18) – TYPES: HT + PT - UMMZ,
PT - CNC, QM
- Podotarsonemus ecuadorensis* Seeman, Lindquist & Husband, 2018 (Page: 26) – TYPES: HT + PT - UMMZ,
PT - CNC, QM
- Podotarsonemus indicus* Seeman, Lindquist & Husband,
2018 (Page: 30) – TYPES: HT + PT - UMMZ, PT -
CNC, QM
- Podotarsonemus nipponicus* Seeman, Lindquist & Husband, 2018 (Page: 35) – TYPES: HT + PT - NMNS, PT -
CNC, QM, UMMZ
- Podotarsonemus queenslandensis* Seeman, Lindquist & Husband, 2018 (Page: 40) – TYPES: HT + PT - QM,
PT - CNC, UMMZ
- Podotarsonemus zuluensis* Seeman, Lindquist & Husband,
2018 (Page: 42) – TYPES: HT + PT - UMMZ, PT -
CNC, QM
- Protobakerdania diseta* Khaustov & Minor, 2018 (Page:
278) – TYPES: HT + PT - NZAC, PT - TSUMZ
- Protohylomysobia erinaceophilus* Sidorchuk & Bochkov,
2018 (Page: 5) – TYPES: HT + PT - GPIH
- Punicodoxa termitophila* Azhari & Hajiqanbar, 2018 (Page:
469) – TYPES: HT + PT - AETMU
- Riccardoella (Proriccardoella) tokyoensis* Waki &
Shimano, 2018 (Page: 165) – TYPES: HT + PT - MPMT,
PT - NSMT
- Scolotydaeus uralensis* Khaustov, 2017 (Page: 168) –
TYPES: HT + PT - TSUMZ, PT - ZISP
- Scutacarus pseudoplurisetus* Khaustov, 2017 (Page: 594)
– TYPES: HT + PT - TSUMZ
- Scutacarus tarifae* Baumann, 2018 (Page: 146) – TYPES:
HT - MNCN, PT - KFUG
- Scutacarus termitophilus* Khaustov, Hugo-Coetze &
Ermilov, 2018 (Page: 59) – TYPES: HT + PT - TSUMZ,
PT - ZISP, NMB
- Scutopalus acaraje* Rocha & Argolo, 2017 (Page: 110) –
TYPES: HT - ESALQ/USP, PT - UESC
- Sonotetranychus madinahensis* Alatawi & Kamran, 2018
(Page: 433) - TYPES: HT + PT - KSMA
- Spatulaphorus altaicus* Khaustov & Trach, 2018 (Page:
124) – TYPES: HT + PT - TSUMZ, PT - ZISP, ONUDZ
- Spatulaphorus brasiliensis* Khaustov & Frolov, 2017 (Page:
513) – TYPES: HT + PT - UNESP, PT - TSUMZ, ZISP
- Spatulaphorus enoplotruporum* Khaustov & Frolov, 2018
(Page: 278) – TYPES: HT + PT - ZISP, PT - TSUMZ
- Sphaerotarsus quercus* Yazdanpanah, Saboori & Hakimtabar,
2016 (Page: 869) – TYPES: HT + PT - JAZM,
PT - ACASI
- Stigmaeopsis continentalis* Saito & Lin, 2018 (Page: 420)
– TYPES: HT + PT - NZMC
- Stigmaeopsis sabelisi* Saito & Sato, 2018 (Page: 416) –
TYPES: HT + PT - HUM
- Stigmaeus grandis* Khaustov, 2017 (Page: 1414) – TYPES:
HT + PT - NWUP, PT - TSUMZ
- Stigmaeus pampaensis* Da-Costa & Johann, 2018 (Page:
716) – TYPES: HT - ESALQ/USP, PT - MCN
- Tanytydaeus cubanus* Khaustov, 2017 (Page: 195) –
TYPES: HT + PT - TSUMZ, PT - ZISP
- Tanytydaeus kethleyi* Khaustov, 2017 (Page: 202) – TYPES:
HT + PT - FMNH

- Tarsenomus lenticulatus* Gheblealivand, Haddad & Mągowski, 2018 (Page: 15) – TYPES: HT + PT - TUAC, PT - DATE
- Tenuipalpoides iraniensis* Mahdavi & Asadi, 2017 (Page: 151) – TYPES: HT + PT - SBUK, PT - ACASI
- Tenuipalpus gneti* Xu, Fan, Huang & Zhang, 2018 (Page: 540) – TYPES: HT + PT - NZMC, PT - FAFU
- Tenuipalpus iranicus* Khadem, Asadi & Seeman, 2018 (Page: 512) – TYPES: HT + PT - SBUK, PT - ACASI, QM
- Tenuipalpus kermanicus* Khadem, Asadi & Seeman, 2018 (Page: 519) – TYPES: HT + PT - SBUK, PT - ACASI
- Tenuipalpus maai* Xu, Fan, Huang & Zhang, 2018 (Page: 561) – TYPES: HT + PT - NZMC, PT - FAFU
- Terminalichus simplex* Castro, Ochoa & Feres, 2017 (Page: 1432) – TYPES: HT + PT - USNM, PT - DZSJRP
- Tetranychus salicornicus* Alatawi & Kamran, 2018 (Page: 433) - TYPES: HT + PT – KSMA
- Thaumatopelvis lorasiacus* Navabi & Hajiqanbar, 2018 (Page: 361) – TYPES: HT + PT - AETMU
- Troxodania minuta* Khaustov & Minor, 2018 (Page: 286) – TYPES: HT + PT - NZAC, PT - TSUMZ
- Tuckerella parsi* Khadem & Asadi, 2018 (Page: 16) – TYPES: HT + PT - SBUK
- Tuckerella weiterschani* Sidorchuk & Khaustov, 2018 (Page: 101) – TYPES: HT - GPIH
- Vimola samarangensis* Xu & Xue, 2017 (Page: 1405) – TYPES: HT + PT - NJAU, PT - NZMC
- Vimola scutellata* Tan, Yang, Lai & Wang, 2015 (Page: 775) – TYPES: HT + PT - GXU
- Xenotarsonemus quiriri* Lofego, Cavalcante & Demite, 2018 (Page: 271) – TYPES: HT + PT - ESALQ/USP
- Xenotarsonemus scorpius* Lofego, Cavalcante & Demite, 2018 (Page: 274) – TYPES: HT + PT - ESALQ/USP
- Zetziella kamili* Akyol & Güll, 2018 (Page: 464) – TYPES: HT + PT - CBZM

New genera

- Collemboerythraeus* Noei, Saboori & Hakimitabar, 2017 (Page: 1258) – Typ. sp.: *Collemboerythraeus vosoughae* Noei, Saboori & Hakimitabar, 2017
- Dividilobus* Xu & Xue, 2017 (Page: 1400) – Typ. sp.: *Dividilobus parvifolius* Xu & Xue, 2017
- Iberochyzeria* Mayoral, Welbourn & Barranco, 2018 (Page: 1130) – Typ. sp.: *Iberochyzeria forniesi* Mayoral, Welbourn & Barranco, 2018
- Lobotarsonemus* Khaustov & Abramov, 2017 (Page: 1080) – Typ. sp.: *Lobotarsonemus betulae* Khaustov & Abramov, 2017
- Makwacarus* Stekolnikov, 2018 (Page: 274) – Typ. sp.: *Makwacarus petrodromi* Stekolnikov, 2018
- Nasutidania* Khaustov & Frolov, 2018 (Page: 283) – Typ. sp.: *Nasutidania orientalis* Khaustov & Frolov, 2018
- Neobakerdania* Khaustov & Minor, 2018 (Page: 282) – Typ. sp.: *Neobakerdania pilosa* Khaustov & Minor, 2018
- Podotarsonemus* Seeman, Lindquist & Husband, 2018 (Page: 7) – Typ. sp.: *Podotarsonemus boliviensisminis* Seeman, Lindquist & Husband, 2018
- Promicrodispus* Khaustov, 2017 (Page: 487) – Typ. sp.: *Brennandania pumilis* Sevastianov, 1975
- Protobakerdania* Khaustov & Minor, 2018 (Page: 277) – Typ. sp.: *Pygmephorus togatus* Willmann, 1942
- Protohylomysobia* Sidorchuk & Bochkov, 2018 (Page: 4) – Typ. sp.: *Protohylomysobia erinaceophilus* Sidorchuk & Bochkov, 2018
- Pseudomicrodispus* Khaustov, 2017 (Page: 481) – Typ. sp.: *Diversipes (Microdispus) setosus* Evans, 1952
- Rhombomicrodispus* Khaustov, 2017 (Page: 486) – Typ. sp.: *Microdispus equisetosus* Mahunka, 1970

New combinations

- Austracarus wittebolsi* (Vercammen-Grandjean, 1959) – [Stekolnikov, 2018: 43]

- Blanciella deschiensi* (Vercammen-Grandjean, 1956) – [Stekolnikov, 2018: 125] *Hypotrombidium felinum* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 139]
- Coleopygmephorus bogenschutzi* (Mahunka & Moser, 1982) – [Khaustov, 2017: 598] *Hypotrombidium geli* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 139]
- Endotrombicula vanmoli* (Vercammen-Grandjean & Benoit, 1971) – [Stekolnikov, 2018: 56] *Hypotrombidium legaci* (André, 1950) – [Stekolnikov, 2018: 139]
- Ericotrombidium accraense* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 130] *Hypotrombidium meleagridae* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 140]
- Ericotrombidium chabaudi* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 131] *Hypotrombidium mourrae* (Taufflieb & Mouchet, 1962) – [Stekolnikov, 2018: 141]
- Ericotrombidium gerardi* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 132] *Hypotrombidium psammodromi* (Taufflieb, 1959) – [Stekolnikov, 2018: 141]
- Ericotrombidium gilliardi* (Vercammen-Grandjean & Langston, 1959) – [Stekolnikov, 2018: 131] *Hypotrombidium ruziziense* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 141]
- Ericotrombidium marcandrei* (Taufflieb, 1960) – [Stekolnikov, 2018: 132] *Hypotrombidium subquadratum* (Lawrence, 1951) – [Stekolnikov, 2018: 142]
- Ericotrombidium oguni* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 132] *Hyracarus claviglis* (Vercammen-Grandjean, 1955) – [Stekolnikov, 2018: 44]
- Ericotrombidium rheinwaldi* (Kolebinova, 1979) – [Stekolnikov, 2018: 133] *Hyracarus lawrencei* (Radford, 1948) – [Stekolnikov, 2018: 44]
- Ericotrombidium rodhaini* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 133] *Hyracarus lemniscomyia* (Vercammen-Grandjean, 1957) – [Stekolnikov, 2018: 45]
- Ericotrombidium scotophilum* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 133] *Marcandreae boaedonia* (Jadin & Vercammen-Grandjean, 1952) – [Stekolnikov, 2018: 144]
- Ericotrombidium spatzi* (Kolebinova, 1980) – [Stekolnikov, 2018: 134] *Marcandreae fromonti* (Jadin & Vercammen-Grandjean, 1960) – [Stekolnikov, 2018: 145]
- Ericotrombidium tarentolae* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 134] *Microtrombicula abyssinica* (Radford, 1947) – [Stekolnikov, 2018: 145]
- Ericotrombidium turdi* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 134] *Microtrombicula agamae* (Lawrence, 1949) – [Stekolnikov, 2018: 146]
- Ericotrombidium ugandaense* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 135] *Microtrombicula bruynoghei* (Jadin & Vercammen-Grandjean, 1952) – [Stekolnikov, 2018: 147]
- Hypotrombidium buttneri* (Vercammen-Grandjean & Taufflieb, 1959) – [Stekolnikov, 2018: 138] *Microtrombicula celiae* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 148]
- Hypotrombidium clamatori* (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 139] *Microtrombicula centropi* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 148]

- Microtrombicula draconensis* (Lawrence, 1949) – [Stekolnikov, 2018: 149] *Microtrombicula rhodesiensis* (Lawrence, 1949) – [Stekolnikov, 2018: 167]
- Microtrombicula dschangi* (Taufflieb & Mouchet, 1959) – [Stekolnikov, 2018: 150] *Microtrombicula rosamonda* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 169]
- Microtrombicula evilla* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 151] *Microtrombicula sicei* (André, 1951) – [Stekolnikov, 2018: 170]
- Microtrombicula felis* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 151] *Microtrombicula smithi* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 171]
- Microtrombicula gerrhosauri* (Lawrence, 1949) – [Stekolnikov, 2018: 151] *Microtrombicula sporopipia* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 171]
- Microtrombicula hyracis* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 153] *Microtrombicula streptopelia* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 172]
- Microtrombicula lumsdeni* (Radford, 1953) – [Stekolnikov, 2018: 156] *Microtrombicula tardaea* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 173]
- Microtrombicula mastomyia* (Radford, 1942) – [Stekolnikov, 2018: 158] *Microtrombicula tragardhi* (Oudemans, 1910) – [Stekolnikov, 2018: 173]
- Microtrombicula microps* (Lawrence, 1951) – [Stekolnikov, 2018: 159] *Microtrombicula tropidosauri* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 174]
- Microtrombicula mini* (Vercammen-Grandjean & Brennan, 1957) – [Stekolnikov, 2018: 159] *Microtrombicula ugandae* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 174]
- Microtrombicula montensis* (Lawrence, 1949) – [Stekolnikov, 2018: 161] *Microtrombicula yangi* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 176]
- Microtrombicula myonacis* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 161] *Neotrombiculoides abonnenci* (Taufflieb, 1960) – [Stekolnikov, 2018: 183]
- Microtrombicula nivaria* (Lawrence, 1949) – [Stekolnikov, 2018: 162] *Neotrombiculoides claviglicola* (Lawrence, 1949) – [Stekolnikov, 2018: 183]
- Microtrombicula pachydactyli* (Lawrence, 1949) – [Stekolnikov, 2018: 163] *Neotrombiculoides elegantissima* (Kolebinova, 1981) – [Stekolnikov, 2018: 184]
- Microtrombicula pembaensis* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 165] *Oudemansidium howelli* (Goff, 1983) – [Stekolnikov, 2018: 184]
- Microtrombicula polymorpha* (Vercammen-Grandjean, 1965) – [Stekolnikov, 2018: 166] *Parawenhoekia aginapaica* (Haitlinger, 1999) – [Mayoral, Welbourn & Barranco, 2018: 1129]
- Microtrombicula quasigiroudi* (Jadin & Vercammen-Grandjean, 1954) – [Stekolnikov, 2018: 166] *Pentidionis maura* (Taufflieb, 1960) – [Stekolnikov, 2018: 185]
- Microtrombicula quasiiscei* (Taufflieb, 1958) – [Stekolnikov, 2018: 167] *Pentidionis meridialis* (Taufflieb, 1960) – [Stekolnikov, 2018: 185]

- Promicrodispus fageus* (Rack, 1965) – [Khaustov, 2017: 489]
- Promicrodispus pumilis* (Sebastianov, 1975) – [Khaustov, 2017: 490]
- Protobakerdania aperta* (Rack & Kaliszewski, 1985) – [Khaustov & Minor, 2018: 278]
- Protobakerdania arvorum* (Jacot, 1936) – [Khaustov & Minor, 2018: 278]
- Protobakerdania arvorum nodulosa* (Mahunka, 1969) – [Khaustov & Minor, 2018: 278]
- Protobakerdania baloghi* (Mahunka, 1969) – [Khaustov & Minor, 2018: 278]
- Protobakerdania crenata* (Mahunka, 1969) – [Khaustov & Minor, 2018: 278]
- Protobakerdania pristinus* (Mahunka, 1968) – [Khaustov & Minor, 2018: 278]
- Protobakerdania randae* (Sebastianov & Zahida Al Douri, 1989) – [Khaustov & Minor, 2018: 278]
- Protobakerdania togatus* (Willmann, 1942) – [Khaustov & Minor, 2018: 278]
- Pseudomicrodispus hastatus* (Mahunka, 1981) – [Khaustov, 2017: 483]
- Pseudomicrodispus montanus* (Khaustov & Minor, 2016) – [Khaustov, 2017: 483]
- Pseudomicrodispus setosus* (Evans, 1952) – [Khaustov, 2017: 483]
- Rhombomicrodispus equisetosus* (Mahunka, 1970) – [Khaustov, 2017: 487]
- Schoengastiella adami* (Taufflieb, 1964) – [Stekolnikov, 2018: 27]
- Schoengastiella berriti* (Taufflieb, 1964) – [Stekolnikov, 2018: 27]
- Schoengastiella chippauxi* (Taufflieb, 1964) – [Stekolnikov, 2018: 28]
- Schoengastiella combesi* (Taufflieb, 1964) – [Stekolnikov, 2018: 29]
- Schoengastiella meyai* (Taufflieb, 1964) – [Stekolnikov, 2018: 30]
- Schoengastiella pauliani* (Taufflieb, 1964) – [Stekolnikov, 2018: 31]
- Schoengastiella petteri* (Taufflieb, 1964) – [Stekolnikov, 2018: 32]
- Schoengastiella rickenbachi* (Taufflieb, 1964) – [Stekolnikov, 2018: 32]
- Schoengastiella vattierae* (Taufflieb, 1964) – [Stekolnikov, 2018: 33]
- Scolotydaeus alexanderi* (Baker, 1949) – [Khaustov, 2017: 154]
- Scolotydaeus lootsi* (Theron, Meyer & Ryke, 1969) – [Khaustov, 2017: 163]
- Scolotydaeus tauricus* (Kuznetsov, 1973) – [Khaustov, 2017: 157]
- Straellesia monosetosa* (Brown, 2006) – [Stekolnikov, 2018: 19]
- Susa hexasternalaea* (Vercammen-Grandjean, 1960) – [Stekolnikov, 2018: 115]
- Tanytydaeus simplex* (Delfinado & Baker, 1974) – [Khaustov, 2017: 186]
- Tauffliebiella mailloti* (Taufflieb & Abonnenc, 1957) – [Stekolnikov, 2018: 115]
- Trombicula crassipalpis* (André, 1958) – [Stekolnikov, 2018: 192]
- Walchia acutalis* (Jadin, Vercammen-Grandjean, Herman, Thienpoint & Fain, 1954) – [Stekolnikov, 2018: 34]
- Walchia felis* (Vercammen-Grandjean & Fain, 1957) – [Stekolnikov, 2018: 34]
- Walchia manis* (Vercammen-Grandjean & Fain, 1957) – [Stekolnikov, 2018: 35]
- Whartonacarus sulae* (Oudemans, 1910) – [Stekolnikov, 2018: 188]
- Willmannium natalense* (Lawrence, 1949) – [Stekolnikov, 2018: 189]

Willmannium suswaensis (Vercammen-Grandjean & Langston, 1976) – [Stekolnikov, 2018: 189]

Paratydeus Baker, 1949 – [Khaustov, 2017: 154]
= *Scolotydaeus* Berlese, 1910

New synonyms

Agistemus layyahensis Khan, Afzal, Bashir, Farooq & Ghaffar, 2016 – [Rehman, Kamran & Alatawi, 2018: 1066]
= *Agistemus garrulus* Chaudhri, Akbar & Rasool, 1974

Agistemus rafaqi Khan, Afzal & Kamran, 2005 – [Rehman, Kamran & Alatawi, 2018: 1066]
= *Agistemus garrulus* Chaudhri, Akbar & Rasool, 1974

Agistemus rawalpindiensis Khan, Bashir, Farooq & Khan, 2010 – [Rehman, Kamran & Alatawi, 2018: 1066]
= *Agistemus garrulus* Chaudhri, Akbar & Rasool, 1974

Anasuscuta (Brown, 2009) – [Stekolnikov, 2018: 18]
= *Straelenia* Vercammen-Grandjean & Kolebinova, 1968

Gahrliepia (Giroudia) traubi Audy, Lawrence & Vercammen-Grandjean, 1961 – [Stekolnikov, 2018: 21]
= *Gahrliepia brennani* (Jadin & Vercammen-Grandjean, 1952)

Herpetacarus (Lukoschuskaia) Kolebinova & Vercammen-Grandjean, 1980 – [Stekolnikov, 2018: 65]
= *Herpetacarus (Herpetacarus) Vercammen-Grandjean, 1960*

Napassenia Haitlinger, 1999 – [Mayoral, Welbourn & Baranco, 2018: 1129]
= *Parawenhoekia* Paoli, 1937

Sacotydeus Baker, 1949 – [Khaustov, 2017: 154]
= *Scolotydaeus* Berlese, 1910

Schoutedenichia tanzaniaensis Goff, 1983 – [Stekolnikov, 2018: 278]
= *Schoutedenichia musaranei* Taufflieb, 1966

Walytydeus Kuznetsov, 1973 – [Khaustov, 2017: 154]
= *Scolotydaeus* Berlese, 1910

New names

Microtrombicula squirrelis Stekolnikov, 2018 pro *Eltonella (Eltonella) myonacis heliosciuri* Vercammen-Grandjean, 1965 – [Stekolnikov, 2018: 171]

New tribus

Podotarsonemini Seeman, Lindquist & Husband, 2018
(Page: 6) – Typ. gen.: *Podotarsonemus* Seeman, Lindquist & Husband, 2018

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18 (3) · 2018

Russell, D. & K. Franke

Actinedida No. 17	1–28
Acarological literature 2	
Publications 2018	2
Publications 2017	9
Publications, additions 2016	17
Publications, additions 2015	18
Publications, additions 2014	18
Publications, additions 2013	18
Nomina nova 19	
New species	21
New genera	24
New combinations	24
New synonyms	27
New names	28
New tribus	28