

THE RESPONSES OF TURBIDITY, CDOM, BENTHIC MICROALGAE,
PHYTOPLANKTON AND ZOOPLANKTON TO VARIATION IN SEASONAL
FRESHWATER INFLOW TO THE CALOOSAHATCHEE ESTUARY

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1.0 INTRODUCTION

Increasing human population and concomitant changes in land and water use alter the timing and amount of freshwater delivered to estuaries. Though coastal counties represent 17% of the total land area of the U.S. (excluding Alaska), they account for 53% of the total population (Crossett et al. 2004). Regionally, Florida led the nation in coastal population growth (75%), normalized to percent change, from 1980 to 2003 (Crossett et al. 2004). This concentration of population along the coastline results in an intensification of environmental problems in coastal watersheds. Urbanization and development of coastal watersheds increases the area of impervious surfaces. As a result, runoff leaves the landscape more quickly and enters estuaries downstream in a pulse: peak flows become higher than under predevelopment conditions and the lag between rainfall and runoff is greatly reduced (Hopkinson & Vallino 1995), rendering tidal tributaries “flashy.” In estuaries where seasonal rainfall often determines the degree of freshwater inflow (i.e., relatively low groundwater contributions), estuarine tributaries become more flashy with increasing watershed development.

Increases in the maximum freshwater inflow rate influence estuarine ecosystems by increasing nutrient availability, decreasing benthic primary production as a result of decreased light penetration (i.e., turbidity, chlorophyll, colored dissolved organic matter), relocating the chlorophyll maximum downstream, and reducing residence time of planktonic organisms through advective processes. Freshwater inflow has been identified as a significant landscape process shaping community structure within estuaries (Mannino & Montagna 1997, Sklar & Browder 1998, Palmer et al. 2002). High levels of freshwater inflow can decrease the abundance of both estuarine residents and marine species that utilize estuaries as nurseries (Garcia et al. 2004). Extreme fluctuations in salinity often elicit stress responses or emigration, resulting in a reduction of biodiversity (Sklar & Browder 1998). Altered freshwater delivery can greatly influence salinity distribution within an estuary both spatially, through the establishment of salinity gradients, and seasonally, through the relocation of isohalines. Biotic responses to these changes can restructure communities associated with affected habitats such as seagrasses (e.g., Matheson et al. 1999, Biber & Irlandi 2006) and oyster reefs (e.g., Maurer & Watling 1973, Tolley et al. 2005, 2006). In addition, suspended sediment and chlorophyll *a* concentration, along with their associated light attenuation, are important factors affecting the health of a variety of

organisms, including oysters, clams and seagrasses (e.g., Lawson et al. 2007, Wilber & Clarke 2001).

Survival of fish stocks depends on the survival of juveniles within nursery habitats. On Florida's west coast, young estuarine-dependent fishes (e.g., bay anchovy, yellowfin menhaden, red drum, sand seatrout) congregate at specific locations along the estuarine gradient. These congregations typically occur within the interiors of riverine estuaries, where 90% of individuals can occur within a few km of river reach (Peebles et al. 2007). Benthic or hyperbenthic invertebrates, some of which are prey for these fishes, also co-occur here. These areas have mud or sand bottoms and are generally devoid of seagrasses and other macrophytes. Predators (fishes) and potential prey (invertebrates) move upstream and downstream with changing freshwater inflows (Flannery et al. 2002, Greenwood et al. 2007). However, salinity and inflow are not proximal habitat determinants for these animals, as the animals congregate at significantly different salinities in different estuaries (Peebles et al. 2007), and, over time, individual species' locations are sometimes better correlated with each other than they are with either freshwater inflow or salinity.

Longitudinal in-situ fluorometry transects, synchronized with standardized fish and invertebrate surveys, have identified associations between fish and chlorophyll *a* peaks within the Alafia River, Florida, provided that the peaks were located within depositional, normoxic habitats (English et al. 2007). Time-averaged fish distributions correlate with sediments that have small grain size and high organic content.

In summary, analyses of diet, prey distribution, salinity fidelity, in-situ fluorometry, and sediment characteristics indicate that fish are congregating in and deriving biomass from benthic habitats where microalgae are either produced (benthic microalgae) or deposited (sedimented phytoplankton). The dynamics of microalgal production and deposition in these areas are poorly understood, particularly as they relate to freshwater inflow.

This project represents a multidisciplinary approach to understanding the influence of freshwater inflow on estuarine production and ecosystem condition and examines both phytoplankton and benthic microalgal biomass. In addition, the project examines an important estuarine function that often goes overlooked when assessing ecosystem condition—the production of prey organisms (i.e., zooplankton and hyperbenthos) for fishes that use estuaries as nursery habitat.

1.1 Objectives

The overall goal of this project is to establish linkages between variability in freshwater inflow and ecosystem condition by characterizing and quantifying the responses of estuarine phytoplankton, zooplankton, and benthic microalgae to variation in freshwater inflow and by identifying linkages among these responses. The resulting relationships can be used to assess ecosystem change, to predict future environmental impact based on projected inflow alterations and to develop targets for ecosystem restoration or enhancement.

Specifically, this project addresses the following objectives:

1. Characterize seasonal (wet/dry) differences in phytoplankton and fish-prey distribution, abundance, and community structure; benthic microalgal biomass and condition; CDOM and the ETM.
2. Investigate how freshwater inflow influences abundance, distribution, and community structure of phytoplankton and zooplankton, benthic microalgal biomass and condition, CDOM and the ETM along the estuarine salinity gradient.
3. Investigate freshwater inflow influences on linkages among CDOM, ETM, benthic microalgae, phytoplankton and zooplankton.

2.0 METHODS

2.1 Study Area

The study area consists of the tidal Caloosahatchee River and Estuary (Fig. 1). The Caloosahatchee watershed is highly altered and highly managed. The historic Caloosahatchee watershed was augmented in the 19th century through a man-made direct connection with Lake Okeechobee and thereby an indirect connection to the Kissimmee River watershed (Antonini et al. 2002). As a result, the Caloosahatchee derives its freshwater both from its own watershed and from the Lake (Doering & Chamberlain 1999). As part of the Okeechobee Waterway that traverses the state, the upper Caloosahatchee has been converted from a meandering river to a canal over much of its length, and water in the river is impounded behind a series of control structures. The W.P. Franklin Lock and Dam is the water control structure located farthest downstream on the Caloosahatchee and represents the upper boundary of the study area. Significant freshwater releases into the river from Lake Okeechobee occur as a means of flood prevention during the rainy season and as a result of periodic drawdowns in the Lake to manage aquatic vegetation and habitat.

2.2 Survey Design

A total of seven zones were sampled from Point Ybel, Sanibel in San Carlos Bay to just below the Franklin Lock and Dam, with two stations (downstream and upstream) sampled within each zone for a total of 14 stations per sampling transect (Table 2.1.1). The use of zones was not based on the identification of strata along the estuarine gradient but simply facilitated station location and sampling along the approximately 47-km transect. Zooplankton and phytoplankton sampling sites were fixed for all collections, with water quality stations being located at approximately the mid-point of each plankton-net tow. The position of the collection vessel was recorded at the beginning of each zooplankton tow using a GPS chartplotter. Mean distance between adjacent sampling sites was 3.26 (\pm 2.01) km. This systemic sampling approach

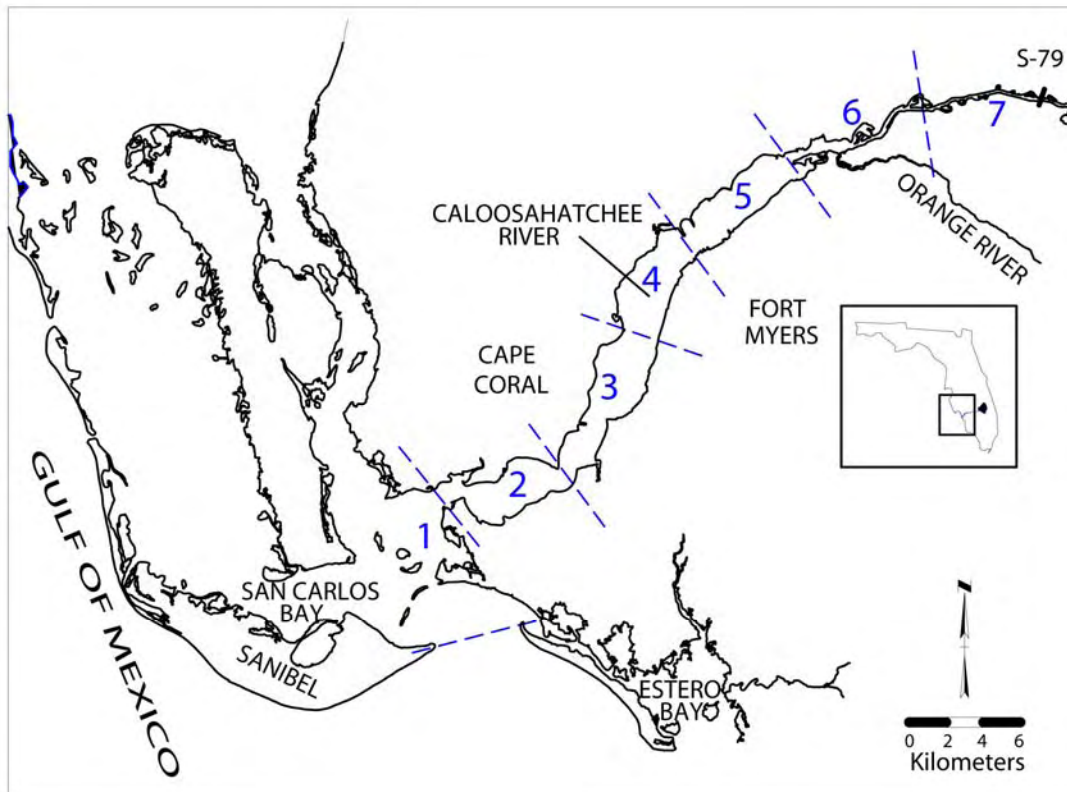


Figure 2.1.1. Map of the tidal Caloosahatchee River and Estuary indicating sampling zones. Two stations (downstream and upstream) were sampled within each zone for a total of 14 sites. S-79 represents the W.P. Franklin Lock and Dam, the upstream extent of the Caloosahatchee Estuary.

(equidistant sampling locations) is preferred over stratified-random sampling for surveying estuarine water quality (Jassby et al. 1997). The following landmarks were used to demarcate zones:

- Zone 1: Inside Point Ybel, Sanibel Island to upstream end of Big Shell Island
- Zone 2: Upstream end of Big Shell Island to Redfish Point
- Zone 3: Redfish Point to Fourmile Point (Midpoint Bridge)
- Zone 4: Midpoint Bridge to Caloosahatchee Bridge
- Zone 5: Caloosahatchee Bridge to Beautiful Island
- Zone 6: Beautiful Island to Trout Creek (just upstream of S.R. 31 Bridge)
- Zone 7: Trout Creek to W.P. Franklin Lock & Dam

Sampling and data collection were conducted at night during a flood tide when larval fishes and invertebrates are generally more abundant in the water column (Wilkins and Lewis 1971, King 1971, Olmi 1994, Morgan 1995a, 1995b). Organisms that selectively occupy the water column during flood tides tend to move upstream, and organisms that occupy the water column during all tidal stages tend to have little net horizontal movement other than that caused by net estuarine outflow (Cronin 1982, McCleave and Kleckner 1982, Olmi 1994). The zooplankton catch was therefore likely biased toward organisms that were either invading the Caloosahatchee River and Estuary or were attempting to maintain position within the system. Furthermore, transects were typically conducted on two consecutive nights in order to ensure that sampling and data collection fell on a similar tide (Table 2.1.2).

Table 2.1.1. Sampling sites for biological and water quality data. Sites were sampled May 2008 through April 2010. Depth represents the mean maximum water depth recorded at each site during biological sampling. D = downstream and U = upstream stations within each zone, with zones as described above.

| Zone | Station | Location (rkm) | Latitude (N) | Longitude (W) | Mean depth (m) |
|------|---------|-------------------|-----------------|------------------|-------------------|
| 1 | D | -5.9 | 26.47760 | 82.01157 | 2.92 |
| | U | -3.6 | 26.49721 | 82.01514 | 3.09 |
| 2 | D | 2.5 | 26.53089 | 81.98688 | 3.96 |
| | U | 5.2 | 26.52616 | 81.96375 | 2.91 |
| 3 | D | 7.6 | 26.54528 | 81.94169 | 4.06 |
| | U | 10.6 | 26.56413 | 81.92283 | 3.88 |
| 4 | D | 16.2 | 26.60805 | 81.90220 | 3.73 |
| | U | 20.0 | 26.64585 | 81.88743 | 2.53 |
| 5 | D | 24.2 | 26.66452 | 81.85461 | 1.97 |
| | U | 26.9 | 26.68076 | 81.83474 | 2.03 |
| 6 | D | 30.2 | 26.69704 | 81.80800 | 2.97 |
| | U | 34.4 | 26.70864 | 81.77011 | 4.38 |
| 7 | D | 37.1 | 26.71587 | 81.75259 | 3.83 |
| | U | 41.0 | 26.72397 | 81.71516 | 1.64 |

Table 2.1.2. Sampling dates during the study period. Sampling efforts typically consisted of two back-to-back sampling nights to ensure that samples were collected on an incoming tide.

| Month | Day | | |
|-----------|-------|-------|-------|
| | 2008 | 2009 | 2010 |
| January | | 21–22 | 26-27 |
| February | | 20–21 | 25 |
| March | | 20–21 | 25-26 |
| April | | 20-21 | 22-23 |
| May | 28–29 | 19–20 | |
| June | 26–27 | 17–19 | |
| July | 27–28 | 16–17 | |
| August | 24–25 | 13–14 | |
| September | 24–25 | 13–14 | |
| October | 25–26 | 10–11 | |
| November | 19–20 | 13 | |
| December | 16–18 | 11-12 | |

2.3 Water Quality Determination

A water quality profile was taken from a second vessel at each site along the sampling transect in conjunction with each biological sampling effort (Fig. 2.3.1). Water quality data acquired are identified in Table 2.3.1. The Seabird 19+ CTD measured conductivity, temperature, and depth at a frequency of 4 Hz and allowed for an examination of water-column structure at high vertical resolution. Attached to the CTD was a Downing Optical Backscatter (OBS) that measured turbidity at high vertical resolution. Bulk water samples were collected using a Geosub purge pump affixed to the CTD instrument at the same level as the OBS. One liter was collected for total suspended sediment concentration (TSS) and percent organic matter analyses at near bottom (1 meter above) and near surface (1 meter below) depths for each cast. Bulk water samples were also used to determine chlorophyll *a* concentration in order to calibrate the fluorometry data.

Table 2.3.1. Details of water quality determination. Unless otherwise specified, parameters were measured throughout the water column at each sampling site. Depth resolution was estimated using mean sampling frequency for each instrument.

| Instrument | Parameters | Depth resolution | No. Sites | Sampling frequency |
|-----------------------------|---|---------------------------|------------------|---------------------------|
| Sea Bird CTD | Conductivity, temperature, depth | ~ 8 mm | 14 | Monthly |
| Downing Optical Backscatter | Turbidity | ~ 8 mm | 14 | Monthly |
| YSI multiparameter sonde | Salinity, temperature, DO, pH, chlorophyll <i>a</i> | ~ 0.30 m | 14 | Monthly |
| WET Labs ECO Triplet | CDOM, turbidity (red backscatter) | ~ 0.15 m | 14 | Monthly |
| Diving-PAM Fluorometer | Photosynthetic efficiency (effective quantum yield) | Bottom | 14 | Monthly |
| Bulk water samples (pumped) | Chlorophyll <i>a</i> concentration | mid-depth, 1 m off bottom | 14 | Monthly |

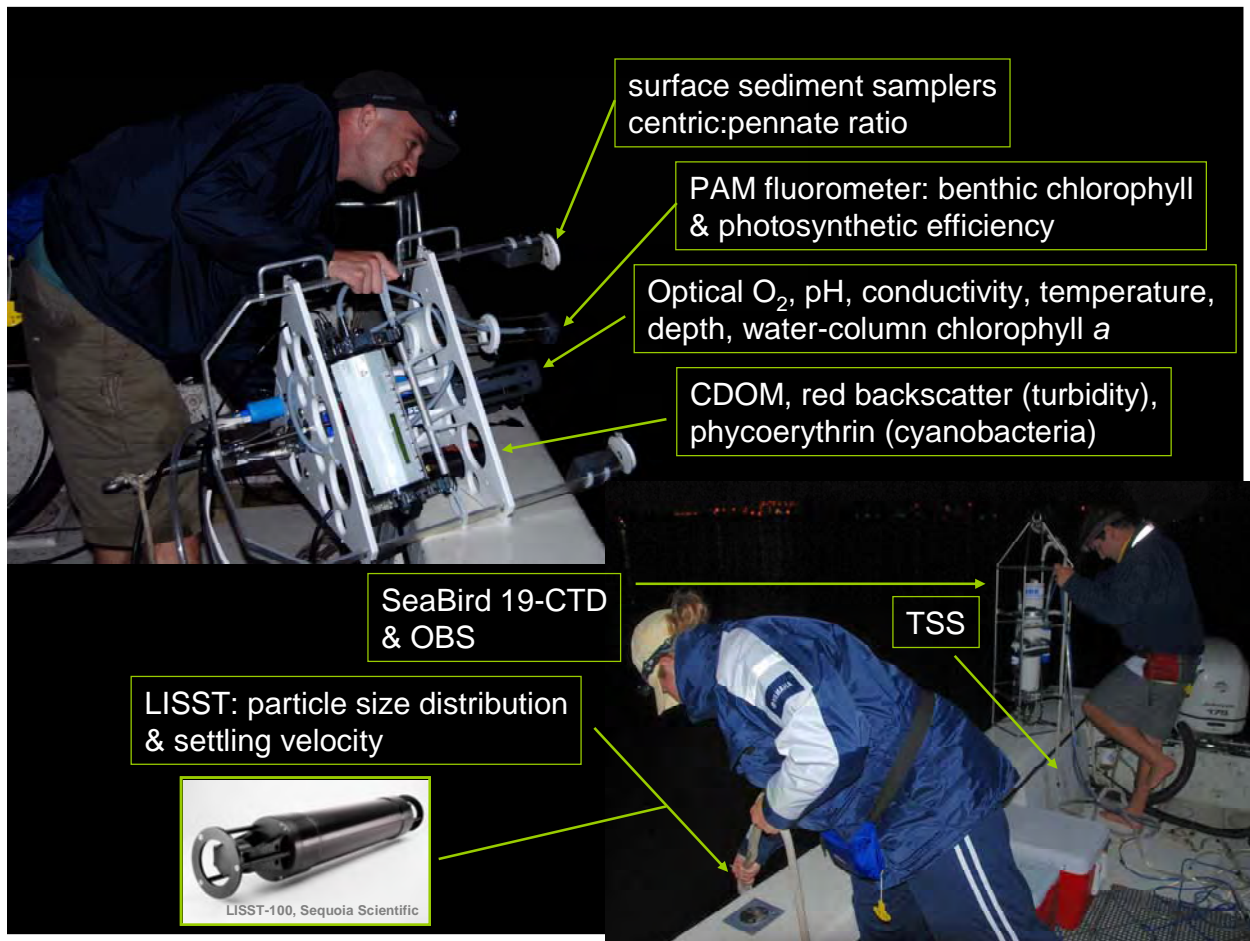


Figure 2.3.1. Water quality instrumentation and associated sampling devices employed in conjunction with biological sampling.

2.4 Biological Sample Collection

In addition to deployment of the above instrumentation at each sampling site, zooplankton (500- μm mesh), phytoplankton (10- μm mesh), and nanoplankton (bulk water) samples were collected (Fig. 2.4.1). Benthic microalgae were collected using glass microscope slides that were smeared with silicone sealant and pressed against surface sediments within 3 min of sealant application (i.e., prior to sealant skinning). The microscope slides were subsequently examined microscopically to identify the relative contributions of planktonic (primarily centric diatoms) and benthic (primarily pennate diatoms) microalgae to the surface sediment.

2.4.1 Phytoplankton Sample Collection

Vertical net-tow samples were collected monthly at each of the sampling sites. A 10- μm mesh, 30-cm diameter, conical (5:1 aspect ratio) plankton net, similar to that described below for zooplankton sampling, was lowered to the near bottom and then retrieved upward through the water column to collect phytoplankton. The plankton-net sample was then transferred to a 125-ml bottle. Each sample was immediately preserved with 0.5% glutaraldehyde to prevent cell lysis and deterioration that could otherwise hinder identification and measurement. Samples were stored on ice for transport back to the laboratory for processing.

2.4.2 Phytoplankton Sample Analysis

Selected phytoplankton vertical net tow samples were examined using an Olympus BX51 microscope under transmitted light and differential interference contrast optics when necessary. Samples were selected from the stations exhibiting the highest chlorophyll concentrations according to integrated YSI chlorophyll profiles. Small aliquots (0.06 ml) of each sample were transferred to a calibrated Phycotech[®] Nannochamber counting slide.

Phytoplankton specimens were identified to the lowest taxonomic level practical according to Tomas (1997), Horner (2002), and Wehr and Sheath (2003). Other publications were consulted as necessary. Phytoplankton taxa were also grouped into their respective phytoplankton classes to provide a more robust measure of phytoplankton composition. All phytoplankton cells $>10\ \mu\text{m}$ were enumerated and measured. Filamentous and other colonial forms comprised of smaller cells were also counted if the colony was $>10\ \mu\text{m}$. Dimensions of all autotrophs and heterotrophs encountered were measured with a calibrated eyepiece micrometer on the microscope. These measurements were used to calculate biovolume (μm^3) based on appropriate geometric formulas (Hillebrand et al. 1999).

2.4.3 Phytoplankton Data Analysis

Phytoplankton abundance was calculated as cells L^{-1} and cell volume L^{-1} (μm^3) for each class and taxon of phytoplankton encountered. Average phytoplankton size was calculated by

dividing the total volume of phytoplankton by the total number of phytoplankton cells. Multi-dimensional scaling (MDS) analysis was used to determine how phytoplankton assemblages changed over time and as water flow and water chemistry changed. Phytoplankton classes and average cell size were also compared to cumulative river flow for the 30 days prior to sampling, as past research has demonstrated that phytoplankton (measured as chlorophyll) are best correlated with a 30-day average flow (Doering et al. 2006).

2.4.4 Chlorophyll data analysis

The ascending profiles from the YSI Sonde casts for each station from each sampling trip were averaged to obtain vertically-integrated values for water chemistry parameters including temperature, salinity, dissolved oxygen (% saturation and concentration), CDOM, pH, and chlorophyll *a*. All parameters except chlorophyll *a* were used in the MDS analysis above. The salinity and chlorophyll *a* data were analyzed against averaged and lagged river flow with additional analyses examining relationships between chlorophyll *a* changes at each station versus river flow (30-d lag) and sampling date. For the flow analyses, the upstream and downstream chlorophyll values were averaged to provide a zone average to provide clearer interpretation of changes observed.

2.4.5 Zooplankton Sampling

Zooplankton samples were also collected monthly at each sampling site. Standard zooplankton collection gear consisted of a 500- μm Nitex mesh, 0.5-m mouth diameter, conical (3:1 aspect ratio) plankton net, equipped with a three-point bridle, 1-liter cod-end jar, 20 kg of weight suspended from the mouth ring, and a General Oceanics model 2030R flowmeter suspended at the center of the net's mouth. Deployment at each site consisted of a three-step oblique tow that divided fishing time equally between bottom, mid-depth, and surface waters. Tow duration was 5 min with tow speed estimated at 1.0-1.5 m/s. Net position in the water column was regulated using a gunwale-mounted winch with metered tow line. Flowmeter readings were recorded before and after deployment so that catch could be normalized to the

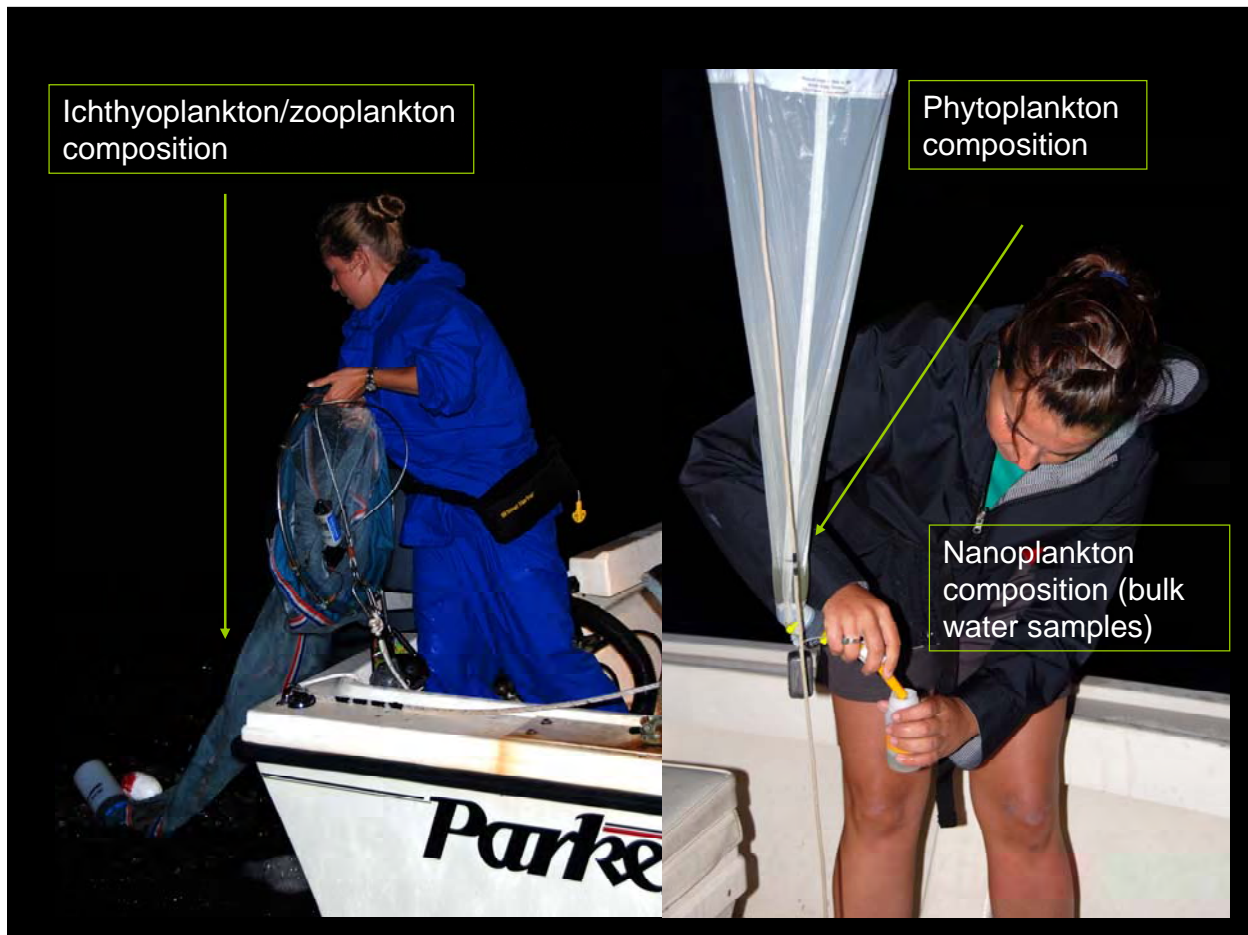


Figure 2.4.1. Biological sampling conducted at 14 stations along the Caloosahatchee River and Estuary.

volume of water filtered during each tow. Upon retrieval, the net was suspended vertically and rinsed from the outside using ambient water delivered by a high pressure, on-demand washdown pump. The final volume within the cod-end jar was reduced by draining excess water through a small area of mesh in the throat of the net, followed by a second rinse of the throat area using approximately 500-800 ml of water. The sample was then preserved by adding 50 ml of formalin (37% formaldehyde) to the 500-800 ml sample volume. One zooplankton net tow was made at each site from May 2008 through April 2010 (14 sites x 24 months = 336 samples).

To reduce both processing time in the field and the potential for contamination, samples were preserved in the plastic cod-end jars in which they were collected. Each cod-end jar was redundantly marked with a unique identifier to facilitate sample handling and track sample custody: an adhesive label placed on the cod-end jar lid and the same unique identifier copied onto the side of the jar using permanent ink. All jars were pre-labeled in the laboratory.

Fixed zooplankton samples were stored in ice chests as collected and were transported back to the laboratory at USF, where they were stored in a fume hood. After a minimum of three days in formalin, plankton-net samples were transferred to a 250- μm mesh sieve (one half the mesh size opening of the collection gear), rinsed with tap water, and then preserved in their original cod-end jars using a rinse of 50% isopropanol in deionized water.

2.4.6 Zooplankton Sample Processing

Plankton-net samples were divided into two size fractions using two stacked sieves of 4-mm and 250- μm mesh openings. This process separated larger items to prevent them from obscuring smaller items during analysis. Sample analysis consisted of determining taxonomic composition, expressed as the number of each taxon per unit volume of water filtered. The >4-mm fraction could often be analyzed without the aid of a microscope. After sieving, the contents of the 4-mm sieve were returned to the original cod-end jar and the contents of the 250- μm sieve were transferred to a numbered beaker. The unique sample identifier (the sample number), the unique number of the beaker containing the 250 μm –4 mm fraction, and the taxonomic composition of the >4-mm fraction were recorded.

A zoom stereomicroscope was used by the Primary Taxonomist to analyze the 250 μm –4 mm fraction of the sample. Successive aliquots were poured from the numbered beaker into a 90-mm square Petri dish delineated into 6 x 6, 13.5 mm square grids. Each aliquot was methodically examined by the Primary Taxonomist, who used the grid delineations to define the processing path. After each aliquot had been processed, the contents of the dish were rinsed into a 1000-ml graduated cylinder using 50% isopropanol in deionized water. Taxonomic quality control was achieved through maintenance of a photographic reference collection. Ichthyoplankton and other zooplankton were identified to the lowest practical taxonomic level.

Taxonomic data for each sample were entered into a specially constructed spreadsheet directly from the microscope workstation. As each sample was processed, a list of encountered taxa was constructed by the Primary Taxonomist within a specialized template area of the spreadsheet. The list was compiled as a subset of a master lookup table for taxonomic descriptions and associated numeric codes. Spreadsheet macros allowed the Primary Taxonomist to select individual taxonomic descriptions/codes from the lookup table for copying to the

template area. The template area was divided into two work areas: a direct enumeration work area and a split-sample work area. The Primary Taxonomist determined which taxa were abundant enough in a given sample to warrant enumeration after splitting and used a macro to construct a list of these taxa within the split-sample work area. All other taxa in the sample were added to a list in the direct enumeration work area as they were encountered. The Primary Taxonomist counted these non-abundant and difficult-to-identify taxa but was responsible for making a list of all taxa in the sample. With few exceptions, the Primary Taxonomist was able to process the entire sample by methodically examining successive aliquots. Each aliquot was rinsed into a 1000-ml graduated cylinder after examination and enumeration within the aliquot were complete. After all aliquots had been processed, the graduated cylinder was covered and inverted multiple times, and a 30-50 ml aliquot was immediately decanted. Beginning and ending volumes within the cylinder were entered into specific cells within the split-sample work area, and a multiplier value was automatically calculated. The split-sample work area of the template was printed, complete with the multiplier value.

The printout of the split-sample work area was then transferred to the Secondary Taxonomist along with the measured aliquot. The Secondary Taxonomist then enumerated the listed taxa in the measured aliquot using a second zoom stereomicroscope. At the computer, the Primary Taxonomist examined the two lists for errors and then executed a macro that transferred the lists to the database region of the spreadsheet. Counts from the direct enumeration of the large and small fractions were included in this data transfer. After the Secondary Taxonomist had processed the measured aliquot, it was recombined with the remainder of the sample in the original cod-end jar and the numbers estimated for the split sample taxa were entered into the database. All template areas were automatically cleared of data after transfer.

All fishes were classified according to developmental stage as follows:

preflexion larva = period between hatching and notochord flexion; The tip of the straight notochord is the most distal osteological feature.

flexion larva = period during notochord flexion; The upturned notochord or urostyle is the most distal osteological feature.

postflexion larva = period between completion of flexion and the juvenile stage; The hypural bones are the most distal osteological feature.

juvenile = period beginning with attainment of meristic characters and body shape comparable to adult fish and ending with sexual maturity

adult = period beginning with sexual maturity

In many fish species, the juvenile stage was difficult to distinguish. At its lower limit, the juvenile stage often lacked a clear developmental juncture that distinguished it from the postflexion stage. Likewise, more than one length at maturity has often been reported for a single species, or the reported length at maturity may differ between males and females. To avoid inconsistency in the staging process, length-based staging conventions were applied to the more common taxa. These staging conventions agree with stage designations used by the U.S. Fish and Wildlife Service (e.g., Jones et al. 1978).

Decapod larvae were classified as zoea, megalopa, or mysis stage. These are terms of convenience and should not be interpreted as technical definitions. Planktonic larvae belonging to Anomura and Brachyura (crabs) were called zoea and individuals from these groups displaying planktonic to benthic transitional morphologies were classified as megalopae. All other decapod larvae (shrimps) were classified as mysis stage until the uropods differentiated into exopods and endopods (5 total elements in the telsonic fan), after which they were classified as postlarvae until they reached the juvenile stage. The juvenile stage was characterized by resemblance to small (immature) adults. Under this system, the juvenile shrimp stage (e.g., for *Palaemonetes* spp.) is equivalent to the postlarval designation used by some authors.

2.5 Water Quality Data Processing

For each water-quality transect, salinity and temperature were used to estimate the vertical density structure at each profile. Bulk water samples were filtered and weighed to determine TSS concentrations. Bulk water samples were also filtered and run through an Aminco-Bowman fluorometer using standard DEP procedures to determine chlorophyll *a* concentrations.

The profiles from each transect were combined in Matlab® to produce a 2D vertical and longitudinal high resolution representation of the salinity and density structure of the

Caloosahatchee River and Estuary. Mass concentration of TSS was estimated by calibrating the OBS results to the bulk water sample results. The calibrated OBS results were then used to produce a 2D vertical and longitudinal high resolution representation of the suspended sediment distribution

2.5.1 Total Suspended Sediment

Total suspended sediment concentrations were determined gravimetrically from the bulk water samples. First, 47-mm glass fiber filters were baked at 525 °C then pre-weighed. Approximately 1 L of the bulk water sample was vacuum filtered, dried at 80 °C for 24 h, weighed, then dried another 24 h and weighed again. Once the weight stabilized, these values were used to derive the TSS concentrations. In order to estimate the organic content of the suspended sediment, the filters were then baked in a muffle furnace at 525 °C for 4 h. Subsequently, the filters were weighed to determine the loss on ignition from volatile carbon.

2.6 Data Analysis

2.6.1 Freshwater Inflow

Data for freshwater inflow from May 2008 through April 2010 were obtained from the U.S. Army Corps of Engineers through the DBHYDRO database maintained by the South Florida Water Management District (SFWMD 2009). Data represent inflow measured at the W.P. Franklin Lock and Dam (S-79) and are reported as cubic feet per second (cfs). Missing values were estimated using polynomial regressions. Inflow data are presented as mean \pm standard deviation unless otherwise specified. Rainfall data were also obtained for S-79 using DBHYDRO.

2.6.2 Organism Center of Abundance

Organism center of abundance was calculated as the weighted mean

$$rkm_U = \frac{\sum (km \cdot U)}{\sum U}$$

where U is organism density (no. m^{-3}) and rkm is distance in river kilometers from the river mouth extending along a conveyance channel or likely path of flow. Positive values represent distances associated with stations located upstream within the Caloosahatchee River, and negative values represent distances associated with stations located downstream of the mouth of the Caloosahatchee River.

2.6.3 Organism Number

Using plankton-net data, the total number of organisms collected (N) in the survey area during each sampling effort (month) was estimated by summing the products of mean organism density (U , as no. m^{-3}) and tide-corrected water volume (V) for each sampling zone of the Caloosahatchee River and Estuary. The study area was first divided into segments representative of each sampling zone (Fig. 2.6.3.1) and the area of each segment was determined using GIS. Bathymetry data taken from digital NOAA charts (zones 1D-6U or derived from local knowledge (zones 6U-7U) were averaged, and the product of area and mean depth for each zone were summed as an estimate of total volume at Mean Lower Low Water. This volume was then adjusted to sampling data using water stage data (NGVD 29) recorded at Shell Point and I-75 stations along the river and provided by SFWMD.

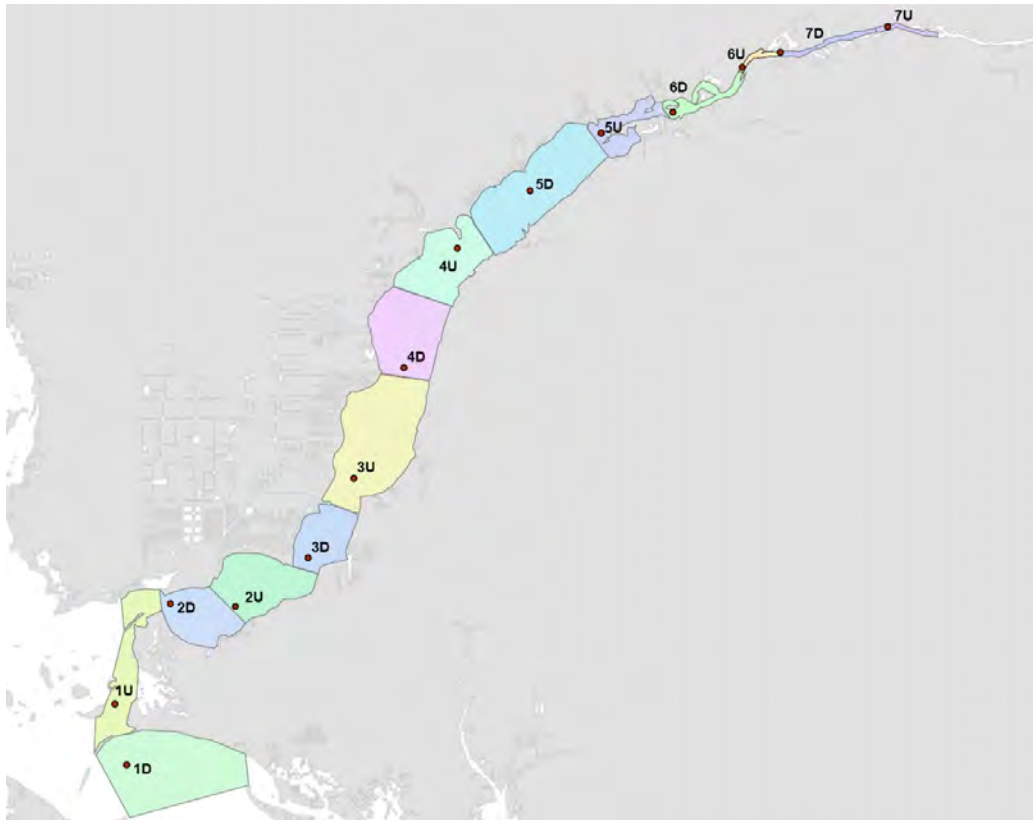


Figure 2.6.3.1. Sampling zones constructed to calculate total volume of the study area. Each color shaded area represents a different sampling zone (1D-7U) and each red circle represents the downstream starting point of each zooplankton net tow.

2.6.4 Identification of Truncated Zooplankton Distributions

When organisms shift their distributions upstream in response to decreased freshwater inflows and an upper abundance percentile (e.g., the 90th percentile) is calculated to be near a fixed barrier to upstream movement (e.g., S-79), then habitat compression due to barrier impingement is likely (Peebles and Greenwood 2009). Abundance, represented as organism density (no. m⁻³), was summed for each monthly survey to produce a total monthly value. Monthly density at individual stations was then summed sequentially in the upstream direction, and the resulting sums were expressed as a percentage of total monthly density. This process is analogous to creating a cumulative distribution curve or function, except that it sums sequential density values from successive stations along a transect instead of summing data-class

frequencies. The location (rkm) of the 90th percentile (the upper decile) of total monthly density was interpolated linearly. These linear interpolations were always made between the station with the highest percentile <90 and the next station upstream. Monthly surveys were excluded from this analysis if >10% of the catch was encountered at the downstream-most station, or if there were fewer than three stations with non-zero densities.

2.6.5 Biodiversity

Biodiversity was estimated using taxa richness. Taxa richness is the number of taxa present in a sample, where an individual taxon reflects both an organism's taxonomic designation (to the lowest level practical) and its developmental stage. The minimum species present in each sample was identified by pooling organisms of a single taxonomic designation (e.g., species) representing different developmental stages and by pooling organisms of different taxonomic levels (e.g., species, genus, family, etc.) regardless of developmental stage.

Center of diversity was calculated as the weighted mean

$$rkm_r = \frac{\sum (km \cdot T)}{\sum T}$$

where T is taxa richness (no. taxa) and rkm is distance in river kilometers from the river mouth extending along a conveyance channel or likely path of flow. Positive rkm values represent distances associated with stations located upstream within the Caloosahatchee River, and negative values represent distances associated with stations located downstream of the mouth of the Caloosahatchee River.

3.0 RESULTS AND DISCUSSION

3.1 Inflow Status

During the period of May 1 through April 30, 2010, freshwater inflow to the Caloosahatchee, as recorded at the W.P. Franklin Lock and Dam (S-79), averaged 1,451 ($\pm 2,159$) cfs with a median inflow of 704 cfs. Inflow varied considerably between wet and dry seasons: mean inflow for May 2008 was 0 (± 0) cfs; however, mean inflow during August 2008 average 5,855 ($\pm 5,997$) cfs (Fig. 3.1.1). In 2008, wet season rains arrived in the study area in early June, with increased inflow present by mid-June. In mid-August, seasonal precipitation was augmented by basin runoff caused by Tropical Storm Fay, resulting in a peak inflow of 18,139 cfs at S-79 on August 20th, in the midst of a 10-d sustained inflow of $>10,000$ cfs (Figs. 3.1.1-3.1.2). In 2009, wet season rains arrived early, beginning in May; however, daily rainfall events >1 in. did not occur until June (Fig. 3.1.1-3.1.2). Freshwater inflow in 2009 peaked at 10,616 cfs on July 3. Although seasonal rains diminished by the end of September of each year, the El Niño event of 2009-2010 resulted in a wetter than normal dry season, triggering releases in Spring 2010 by the U.S. Army Corps of Engineers to reduce water levels in Lake Okeechobee.

In addition to these general patterns of rainfall and inflow recorded in the study area, a series of pulsed releases from S-79 was made in early 2009 to increase freshwater inflow to the estuarine portion of the system and to reduce the need for larger regulatory releases later in the year (Fig. 3.1.1).

3.2 Water Quality

Summary statistics for water quality data collected are presented in Table 3.2.1. Dissolved oxygen at depth ranged from 1.81-11.38 mg L⁻¹ (25.7-118.9 % air saturation) and was generally ≥ 6.0 mg L⁻¹, with 75% of the values ≥ 6.75 mg L⁻¹. However, during the wet season, minimum DO values were consistently low upstream (Fig. 3.2.1) in bottom waters near rkm 34.4. Although water quality measurements were not made in the main channel at deeper depths, hypoxia (<2.0 mg L⁻¹) was encountered once at this location during May 2009.

3.2.1 Density Structure

Variations in vertical and longitudinal salinity structure were strongly related to wet and dry season conditions (Fig. 3.2.1.1; Appendix A). The highest salinity near the mouth (38 psu) was recorded in May 2008. At this time the mouth was well mixed and there was some slight stratification at the head of tidal intrusion where salinity was ~20 psu (Fig. 3.2.1.1). With the onset of higher freshwater discharge in June 2008 salinity was still around 36 psu at the mouth, but stronger stratification was set up at the Franklin Lock and Dam, where salinity ranged from 18 psu near bottom to 9 psu at the surface. By August 2009, as freshwater discharge continued, a highly stratified salt wedge type structure developed near the mouth with maximum salinities around 18 psu (Fig. 3.2.1.1). Thereafter, throughout the decreasing freshwater discharge

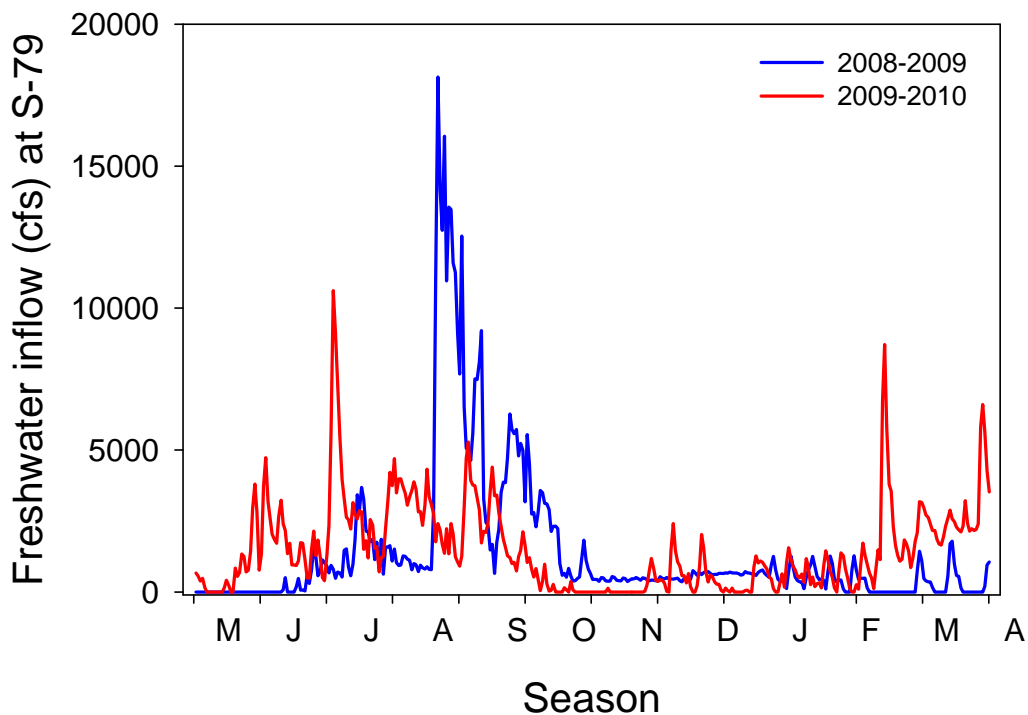


Figure 3.1.1. Freshwater inflow to the Caloosahatchee River and Estuary as recorded at the W.P. Franklin Lock and Dam, May 1, 2008 – April 30, 2010. Note pulsed releases from S-79 in the winter and spring of 2009.

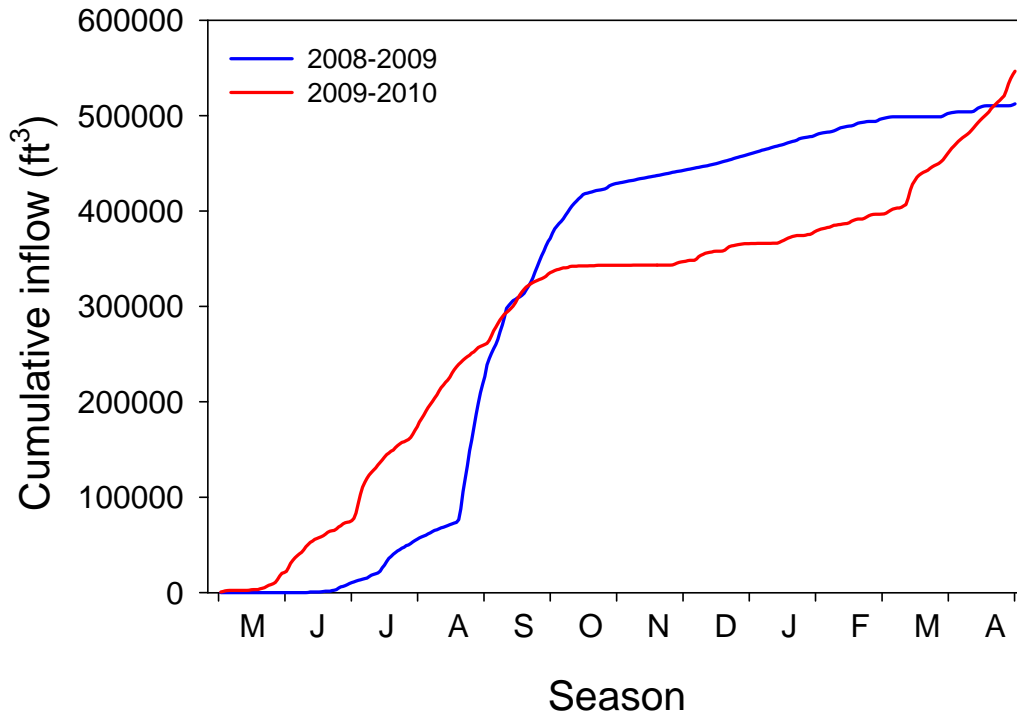


Figure 3.1.2. Cumulative freshwater inflow delivered to the Caloosahatchee River and Estuary, May 1, 2008 – April 30, 2010.

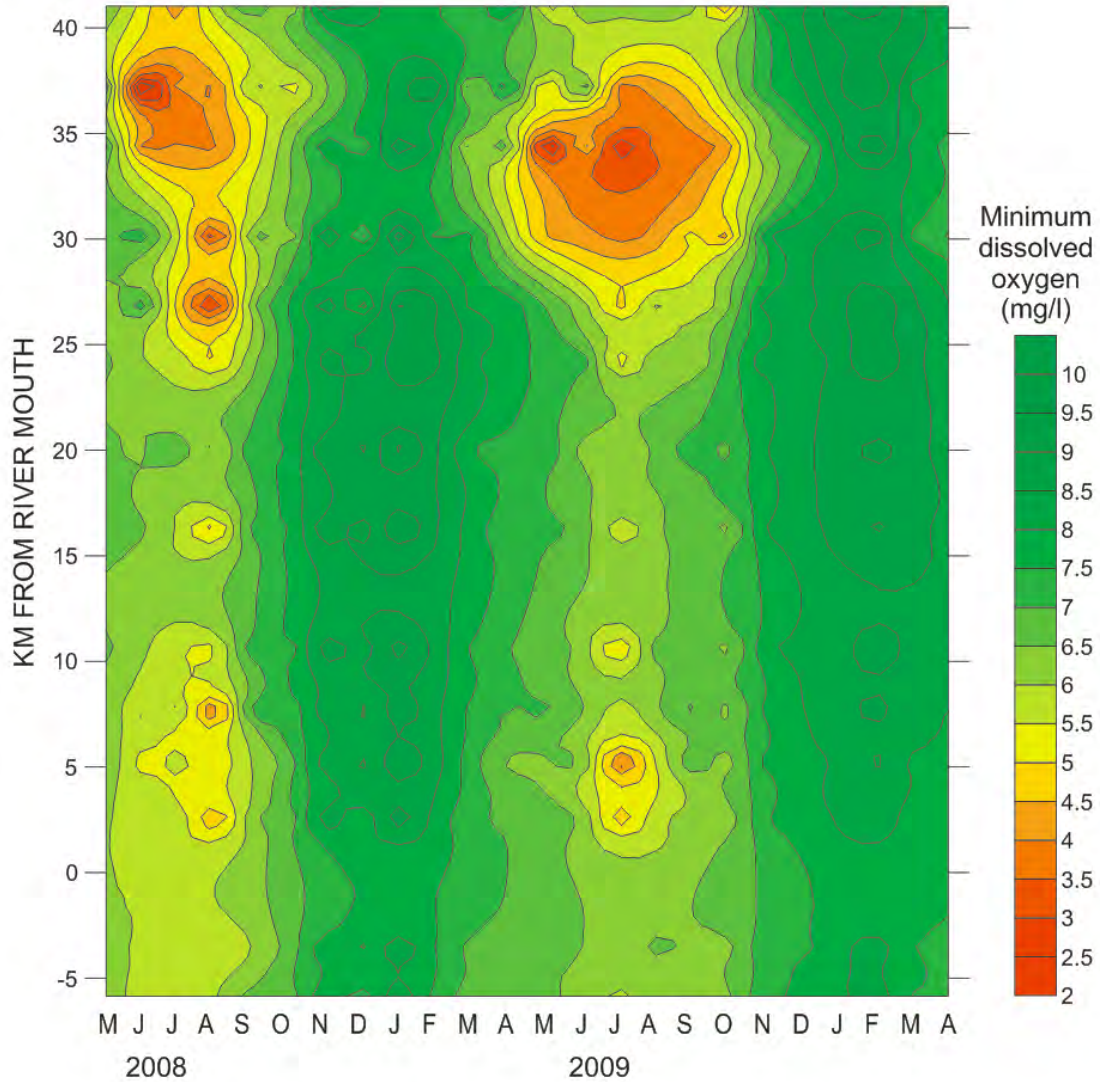


Figure 3.2.1. Time series of minimum dissolved oxygen concentration (mg L^{-1}) sampled along the length of the study area, May 2008–April 2010.

of fall and dry season of 2009 salt water crept back up the estuary and remained generally well mixed during the surveys, which were always conducted during flood tides.

With the onset of the wet season in 2009 stratification increased at the mouth again and salinity dropped. As in the previous year, by October salinity began to intrude back into the estuary, and the water column remained generally well mixed throughout the dry season of 2010.

The general seasonal pattern is of a rapid transition of well mixed salty water during dry seasons to wet season conditions. Freshwater moves rapidly down the estuary at the onset of seasonal rains, as can be seen by noting the rapid movement down the estuary of the location of the 2 psu isohaline (Fig. 3.2.1.2). The freshwater region is well mixed, but there is usually a region of strong stratification at the extent of the salinity intrusion. As freshwater discharge decreases during fall, the 2 psu isohaline rapidly moves back up the estuary, accompanied by a region of high stratification, with well mixed waters both up and downstream of the salinity intrusion (Figs. 3.2.1.3). An exception to this trend occurs around August of 2009 when there was a moderate degree of density stratification upstream of the 2 psu isohaline. This stratification was due to vertical temperature gradients rather than vertical salinity gradients, shown in Fig. 3.2.1.4, demonstrating that the highest salinity gradients occurred just downstream of the 2 psu isohaline. Comparisons of absolute values of surface and bottom salinities for all stations and months show the generally well mixed nature of the Caloosahatchee, with the exceptions predominantly occurring during the wet season (Fig. 3.2.1.5). There are also interesting annual trends in salinity. As noted earlier, the maximum salinity occurs at the end of the 2008 dry season. By following the 20 psu contour, one can observe that mean salinities decreased during each consecutive dry season. Conversely, by following the 2 psu isohaline, one can observe that the extent of freshwater also decreased from the 2008 wet season to the 2009 wet season. The pattern results in a decreasing annual range of salinity at any given spot along the river and may have an effect on the ecology of the river.

3.2.2 Turbidity and Suspended Sediment Characteristics

General Characteristics

Suspended sediment concentrations in the Caloosahatchee are very low compared to most temperate estuaries, rarely rising above 0.025 g l^{-1} . Figures 3.2.2.1–3.2.2.4 show median concentrations and percent organic content of water samples collected from all stations along the transect for each deployment month and depth (i.e., near surface and near bottom). Median concentrations are very low, ranging from $0.003\text{-}0.016 \text{ g l}^{-1}$ at the bottom and $2\text{-}10 \text{ mg l}^{-1}$ at the surface (excluding an anomolous value at 15 mg l^{-1}). In 2008 there was a trend from higher

Table 3.2.1. Summary statistics for water quality data collected May 2008–April 2010. Data represent means (\pm SD) calculated from vertically averaged data from each sampling effort.

| Location (rkm) | Temperature (°C) | Salinity (‰) | pH | DO % | DO mg L ⁻¹ | chl <i>a</i> * (µg L ⁻¹) | Turbidity (OBS volts) | CDOM (absorption ₄₄₀ [m ⁻¹]) |
|-------------------|---------------------|-----------------|----------------|----------------|--------------------------|---|--------------------------|--|
| -5.9 | 24.76 (5.38) | 31.44 (5.36) | 7.77 (0.41) | 101.7 (3.8) | 7.11 (0.72) | 3.9 (1.6) | 2.8 (2.0) | 1.50 (2.72) |
| -3.6 | 24.77 (5.27) | 29.52 (6.77) | 8.03 (0.14) | 101.0 (4.2) | 7.13 (0.70) | 4.7 (2.4) | 2.7 (2.0) | 2.14 (3.30) |
| 2.5 | 24.86 (5.01) | 21.79 (9.32) | 8.08 (0.12) | 99.9 (4.5) | 7.37 (0.82) | 6.4 (3.9) | 1.9 (1.8) | 4.12 (4.24) |
| 5.2 | 24.73 (5.10) | 20.03 (8.41) | 8.07 (0.09) | 98.6 (6.7) | 7.36 (0.84) | 8.8 (5.5) | 2.8 (2.1) | 4.31 (3.66) |
| 7.7 | 24.89 (4.87) | 17.06 (9.45) | 8.11 (0.16) | 99.9 (6.0) | 7.56 (0.84) | 9.8 (6.0) | 2.3 (1.6) | 5.08 (4.41) |
| 10.6 | 24.73 (4.95) | 14.71 (9.19) | 8.13 (0.12) | 100.2 (5.4) | 7.71 (0.89) | 10.8 (6.1) | 2.1 (1.5) | 5.49 (4.36) |
| 16.2 | 24.64 (5.06) | 11.68 (8.68) | 8.16 (0.23) | 101.1 (6.7) | 7.93 (0.99) | 13.3 (6.4) | 2.7 (1.4) | 5.82 (3.99) |
| 20.0 | 24.59 (5.01) | 10.06 (8.44) | 8.14 (0.19) | 101.2 (4.4) | 8.02 (0.89) | 12.2 (2.8) | 2.6 (1.6) | 6.14 (4.07) |
| 24.2 | 25.14 (5.00) | 7.39 (7.05) | 8.20 (0.25) | 100.0 (7.2) | 7.97 (1.04) | 12.1 (2.8) | 3.6 (3.2) | 6.28 (3.69) |

| | | | | | | | | |
|------|-----------------|----------------|----------------|-----------------|----------------|---------------|--------------|----------------|
| 26.9 | 25.13 (5.01) | 6.55 (6.99) | 8.06 (0.22) | 98.7 (6.4) | 7.92 (1.02) | 12.4 (2.7) | 5.2 (5.7) | 6.60 (3.79) |
| 30.2 | 26.61 (4.31) | 4.53 (5.32) | 8.00 (0.21) | 95.40 (9.80) | 7.53 (1.15) | 12.6 (3.7) | 2.9 (2.2) | 7.10 (3.83) |
| 34.4 | 25.40 (4.33) | 4.39 (5.44) | 7.91 (0.27) | 90.8 (12.8) | 7.36 (1.46) | 13.7 (3.9) | 2.3 (2.3) | 7.36 (4.00) |
| 37.1 | 25.00 (4.59) | 3.92 (5.02) | 7.92 (0.20) | 93.8 (10.9) | 7.65 (1.26) | 12.6 (3.2) | 1.9 (0.9) | 7.58 (4.13) |
| 41.0 | 24.41 (4.54) | 3.63 (4.87) | 7.94 (0.16) | 95.6 (7.4) | 7.91 (1.13) | 11.9 (3.5) | 2.4 (1.7) | 7.59 (4.08) |

* Chlorophyll *a* data from May 2009-April 2010 only.

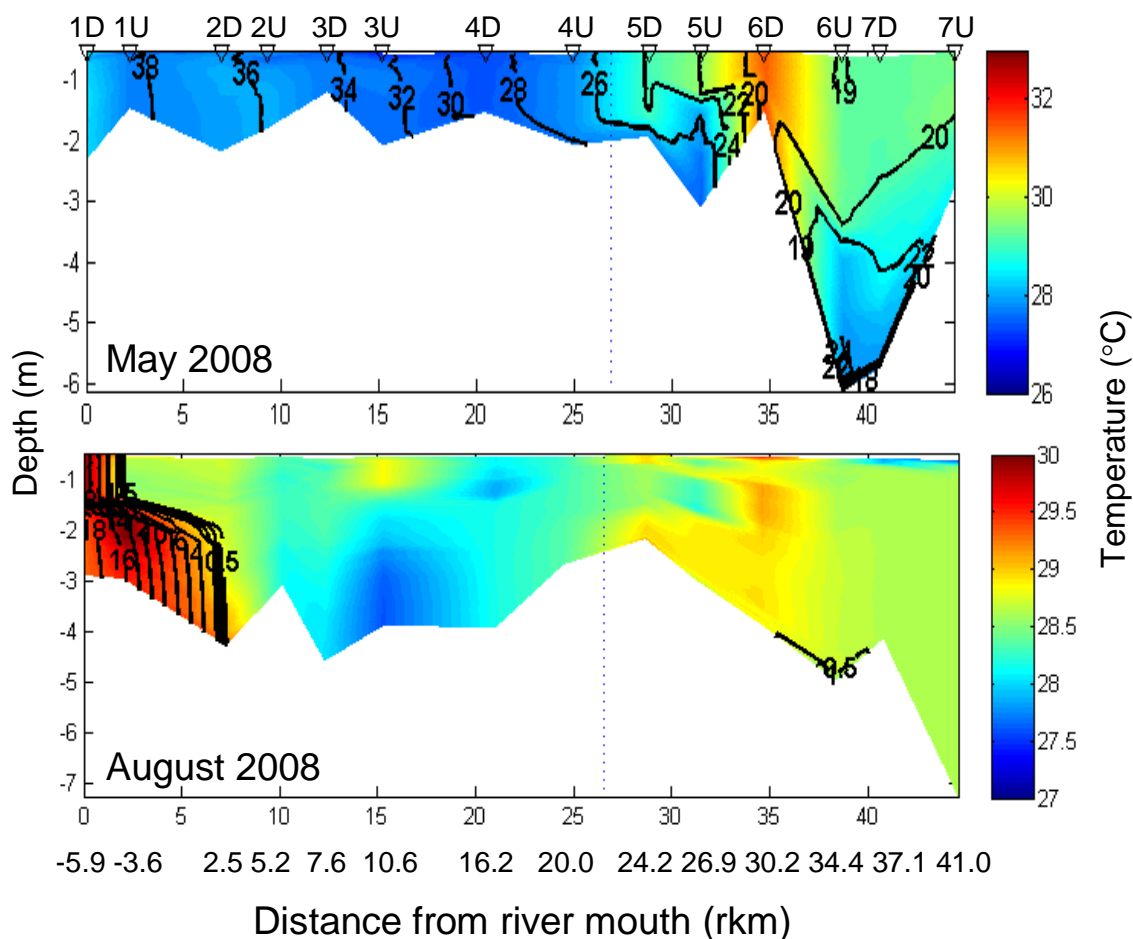


Figure 3.2.1.1. Longitudinal temperature distribution with isohalines illustrating seasonal extremes in salinity structure (May: dry season; August: wet season). Triangles denote locations of profiles along the Caloosahatchee River and Estuary and are labeled according to site.

values during the wet season, decreasing as the wet season waned, at both surface and bottom water samples. In contrast, there was little seasonal trend in 2009, with both surface and bottom concentrations remaining relatively constant. The median percentage of organic material trended from low values (~25–40 %) during the early dry season (February–April), then increasing throughout the rest of the year (February and March 2010 samples were incinerated due to a malfunctioning muffle furnace). The maximum transect median percentage of organics was around 82% at the surface and 57% at the bottom. Surface suspended material was generally higher in organic material than that at the bottom.

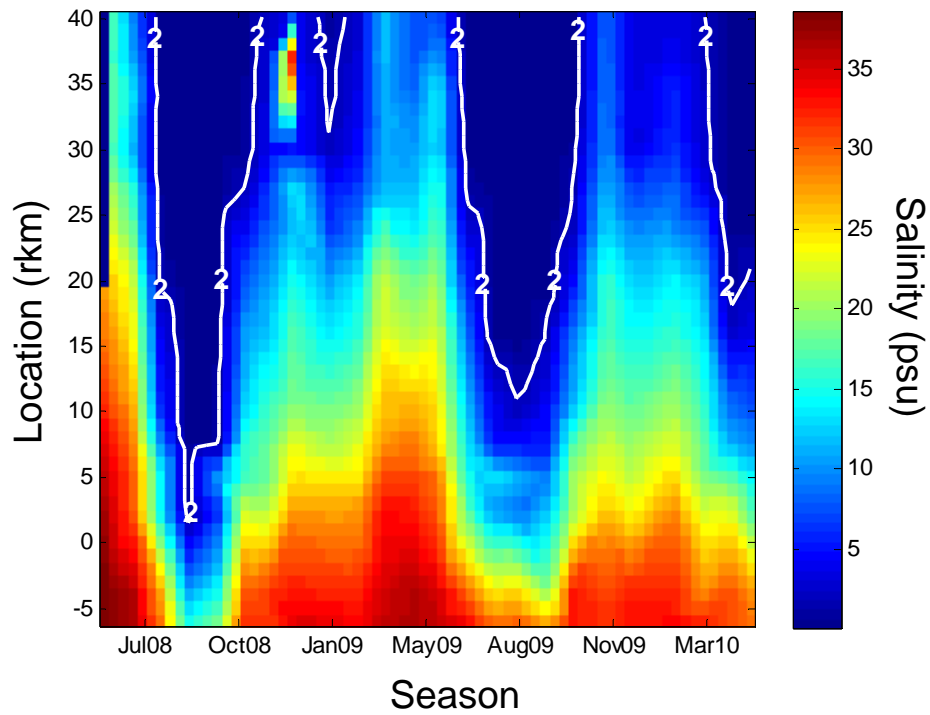


Figure 3.2.1.2. Time series of vertically averaged salinity. White line represents the 2 psu isohaline.

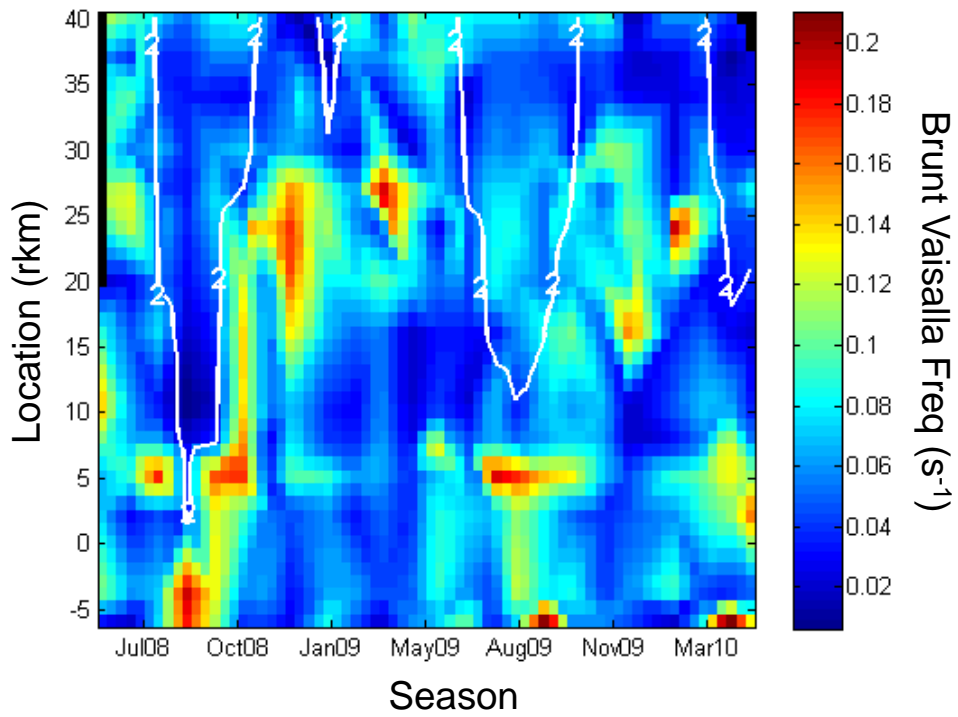


Figure 3.2.1.3. Time series of density stratification. More stratified regions are in warm colors, well mixed regions in cool colors.

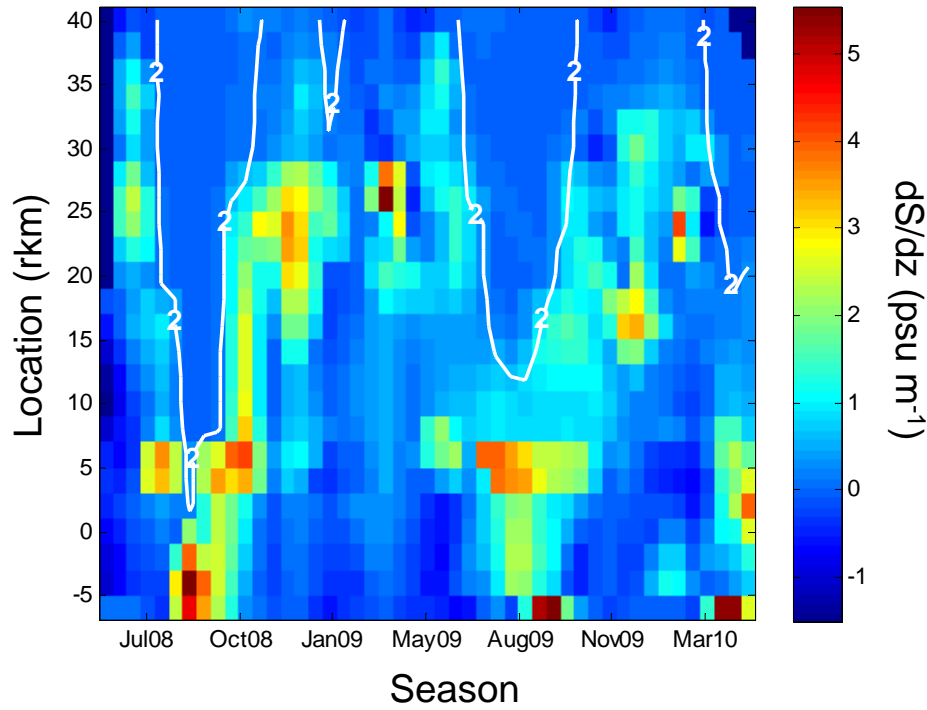


Figure 3.2.1.4. Time series of vertical salinity gradient (difference between surface and bottom salinities, dS, and depth, dz). Stratified regions are in warm colors, well mixed regions in cool colors.

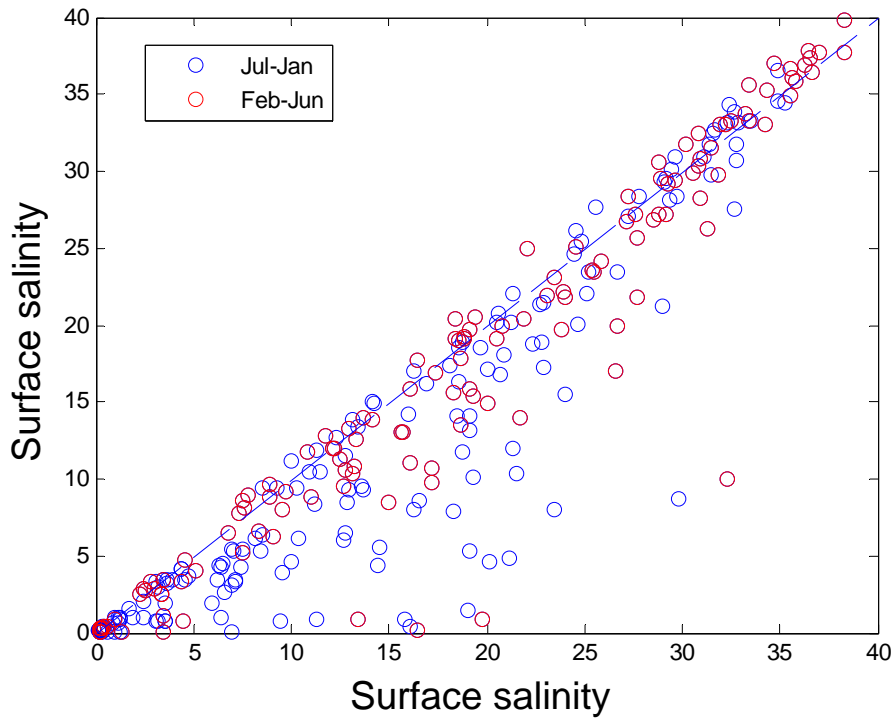


Figure 3.2.1.5. Surface and bottom salinities for all months and stations. Dashed line represents a slope of one.

Temporal and Longitudinal Distributions

Turbidity levels peak regularly near the 2 psu isohaline, as might be expected from classically formed estuarine turbidity maxima (ETMs) (Fig. 3.2.2.5). During the dry seasons, peak turbidity levels develop around rkm 27, a region where the Caloosahatchee becomes restricted, and resuspension was probably due to the increasing currents in the shallow narrow channel. Additionally, turbidity peaks also formed just outside of the mouth (-5 to 0 rkm) and appeared to be dispersed into the mouth by tidal oscillations. Two large peaks in these locations occurred during the wet season of 2008 and the end of the dry season in 2010. Total mass concentrations track the turbidity peaks well (Figs. 3.2.2.6-3.2.2.7). The majority of the peaks in turbidity were due to increases in inorganic mass concentrations of suspended sediments. Near bottom total organic concentrations peaked during three distinct periods around river km 27, during the wet season of 2008, the dry season and beginning of wet season 2009, and the dry season of 2010 (Figs. 3.2.2.8-3.2.2.9). High organic content was responsible for the upriver turbidity peak during the wet season of 2008, which was accompanied by a classical ETM downstream of the 2 psu isohaline. Time series of the percentage of organic material in the water column reveals inter- and intra-annual patterns in the nature of the suspended sediment (Figs. 3.2.2.10-3.2.2.11). Surface material tends to have a higher percentage of organic material, as expected. There was a tendency for higher organic content upriver of the 2 psu halocline during the rainy seasons. In both 2008 and 2009, high organic content material reached all the way down river in the early wet season. As each season continued, the percentage of organic material decreased first at the downstream extent and then decreased going upriver. Although some of the data from late 2009 and 2010 is missing, there appears to be an intra-annual decreasing trend in percentage of organic material. This decrease may be associated with the extent of freshwater discharge during the rainy seasons, which also is greatest in 2008 and decreases each year thereafter.

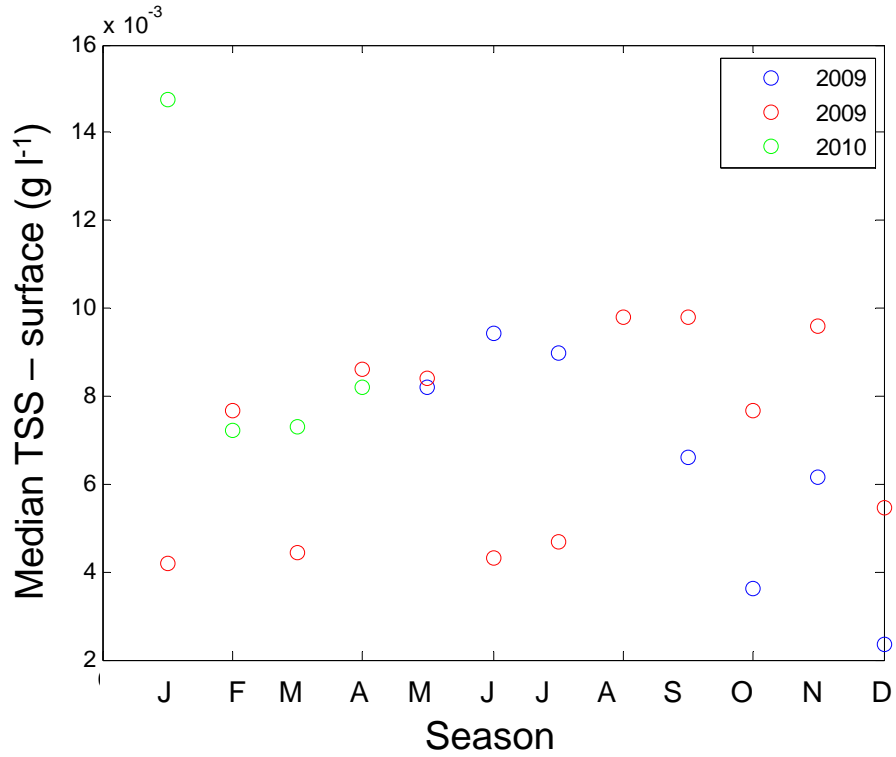


Figure 3.2.2.1. Seasonal distribution of total suspended solids in water samples collected near the surface.

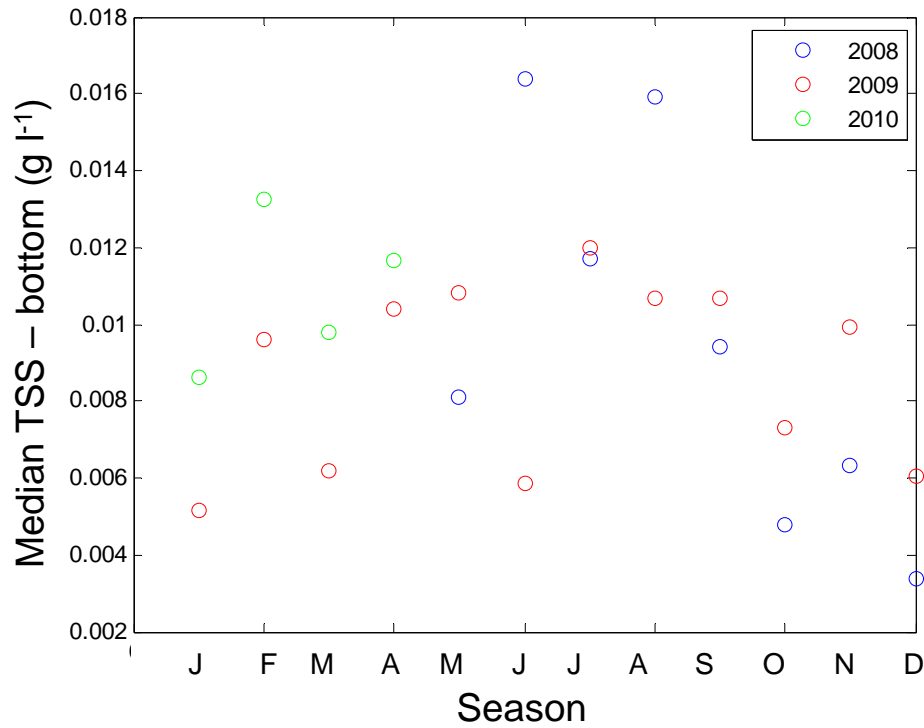


Figure 3.2.2.2. Seasonal distribution of total suspended solids in water samples collected near the bottom.

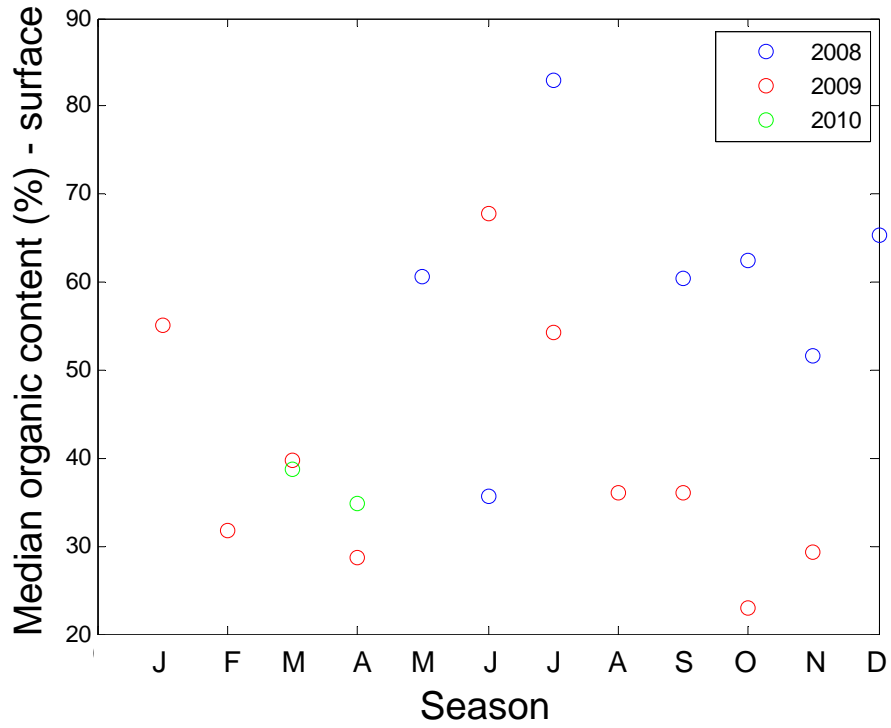


Figure 3.2.2.3. Seasonal distribution of organic content from water samples collected near the surface.

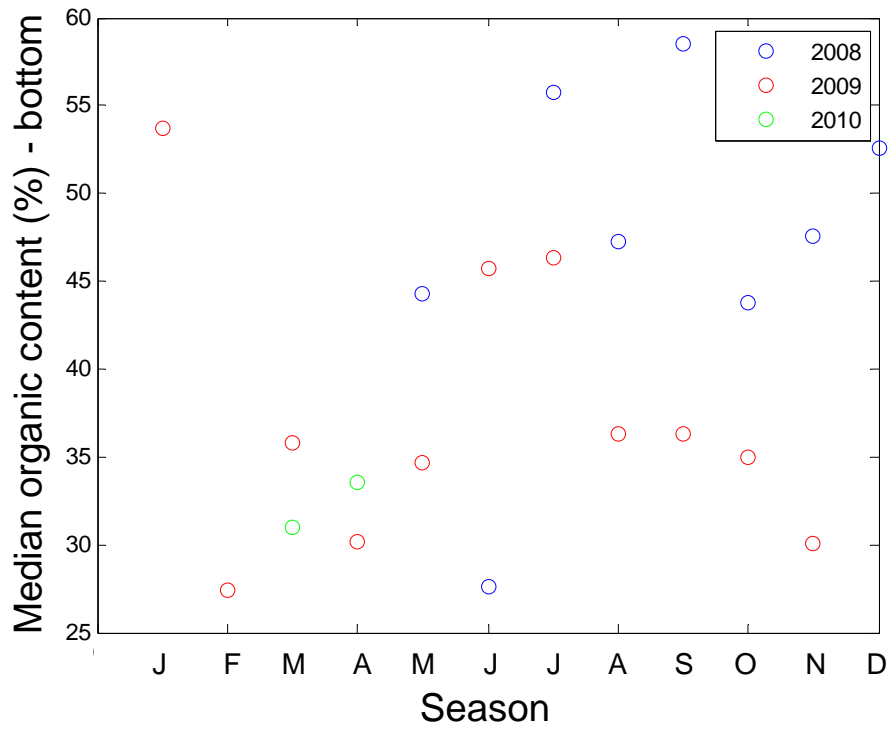


Figure 3.2.2.4. Seasonal distribution of organic content from water sampled collected near the bottom.

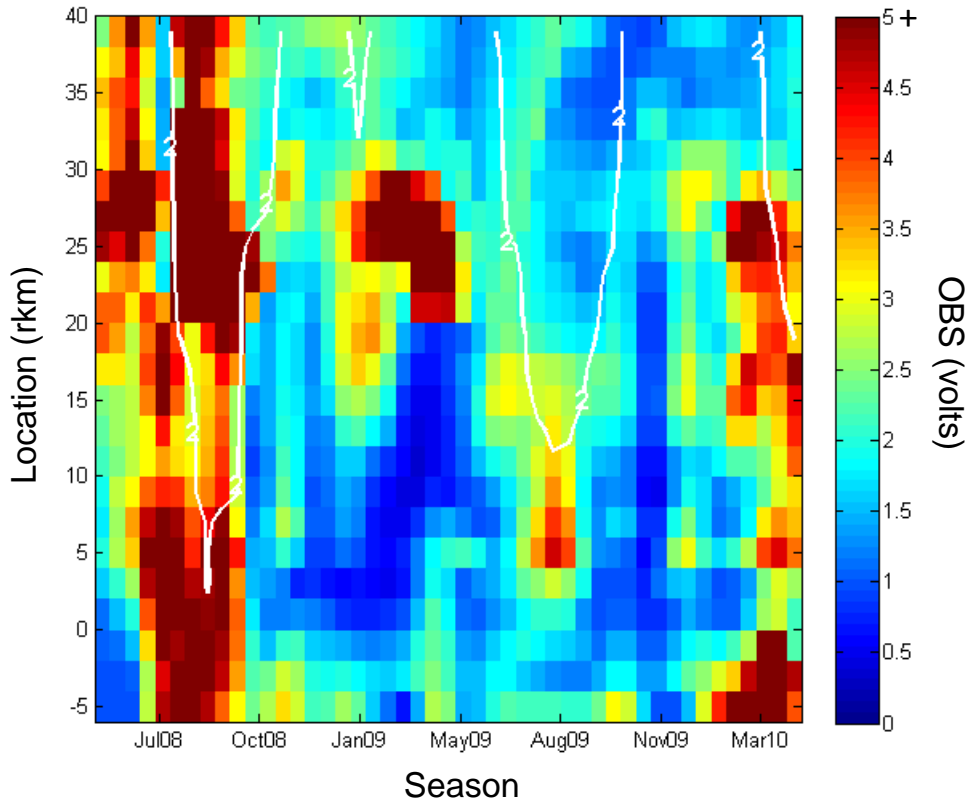


Figure 3.2.2.5. Time series of turbidity distribution as measured using optical back scatter.

3.2.3 Colored Dissolved Organic Matter

Colored dissolved organic matter (CDOM) was determined fluorometrically using a WET Labs ECO Triplet instrument and was converted to absorption coefficient via calibration of bulk water samples (collected at each site for each sampling period) for which absorption at 440 nm was measured using a spectrometer. The distribution of CDOM throughout the study period suggested an upstream source that was diluted by mixing downstream (Figs. 3.2.3.1-3.2.3.4). CDOM values were relatively low during dry months and increased substantially with the onset of the wet season. During periods of high freshwater inflow (e.g., August 2008, Fig. 3.2.3.1), high levels of CDOM were present throughout the tidal portion of the river.

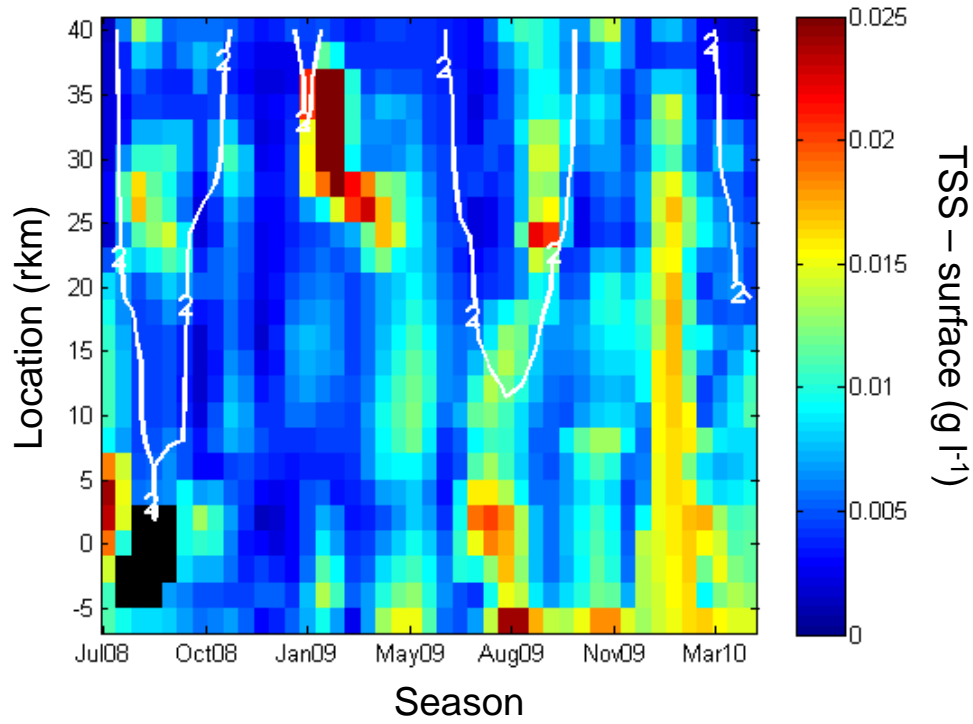


Figure 3.2.2.6. Time series of total suspended solids estimated from water samples taken near the surface.

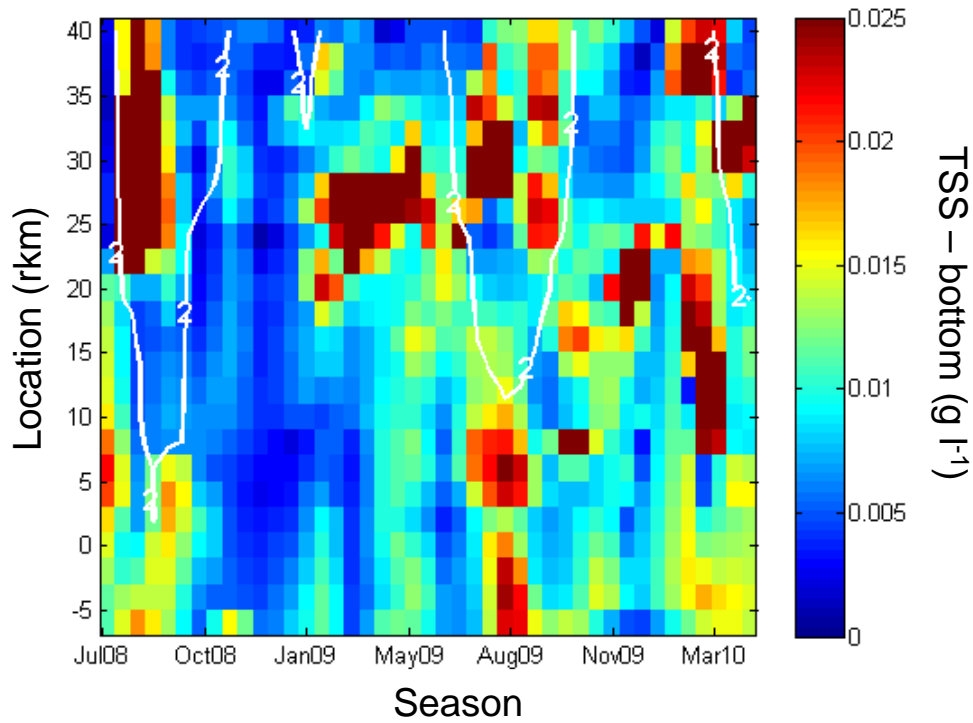


Figure 3.2.2.7. Time series of total suspended solids estimated from water samples taken near the bottom.

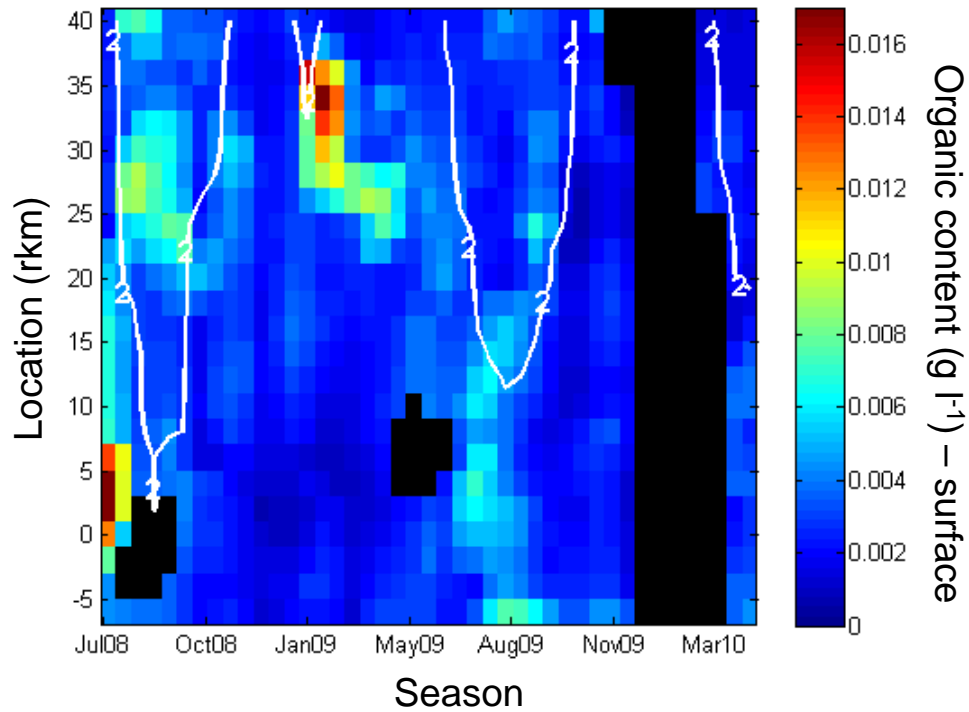


Figure 3.2.2.8. Time series of organic matter estimated from water samples collected near the surface.

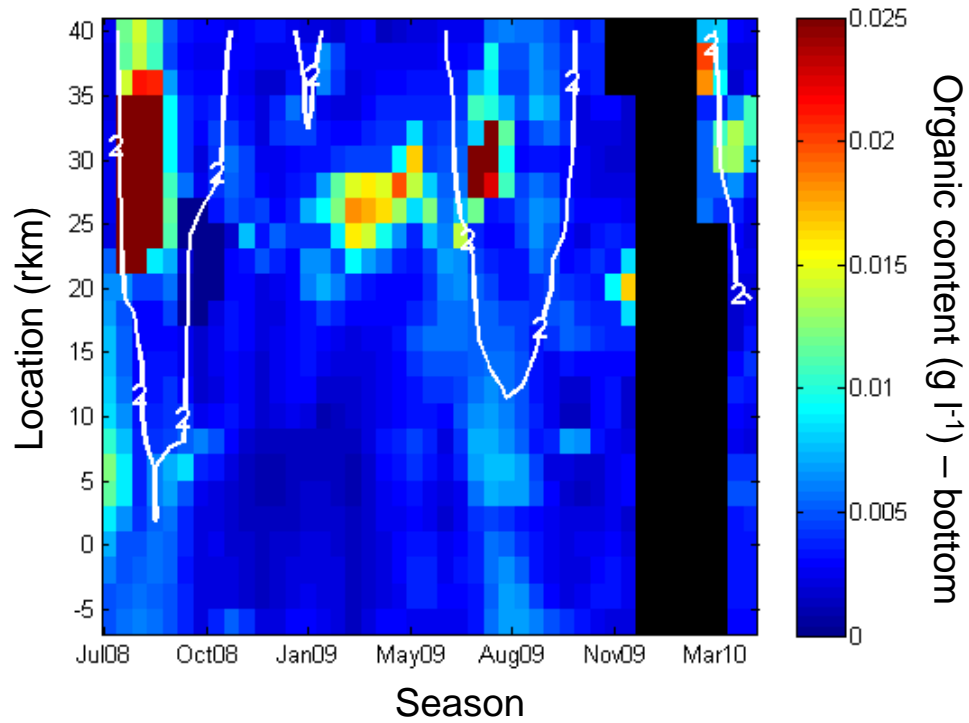


Figure 3.2.2.9. Time series of organic matter estimated from water samples collected near the bottom.

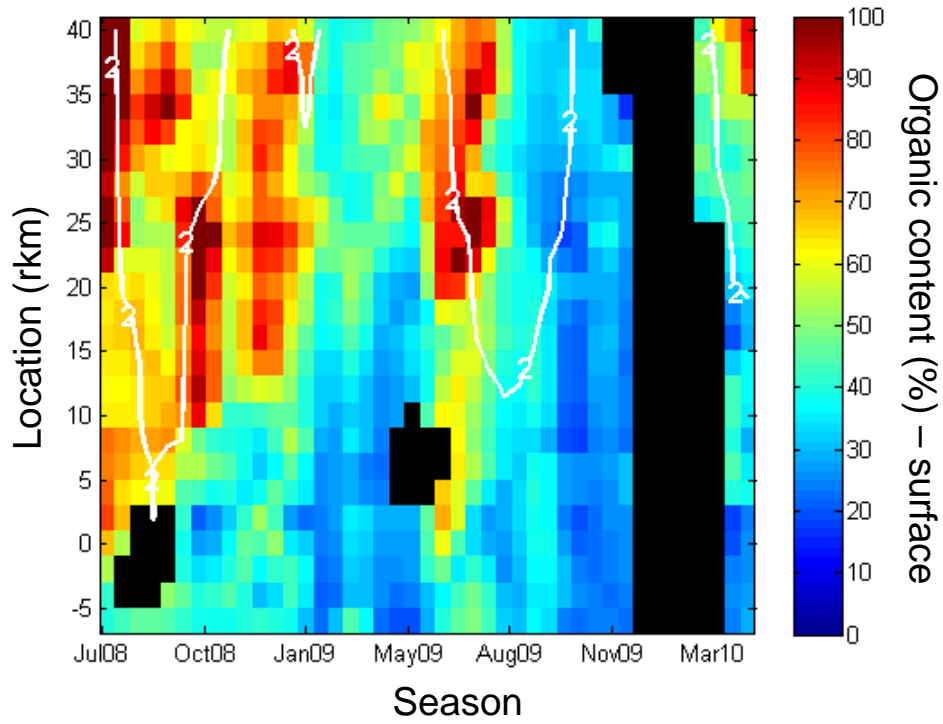


Figure 3.2.2.10. Time series of relative organic content determined from water samples collected near the surface.

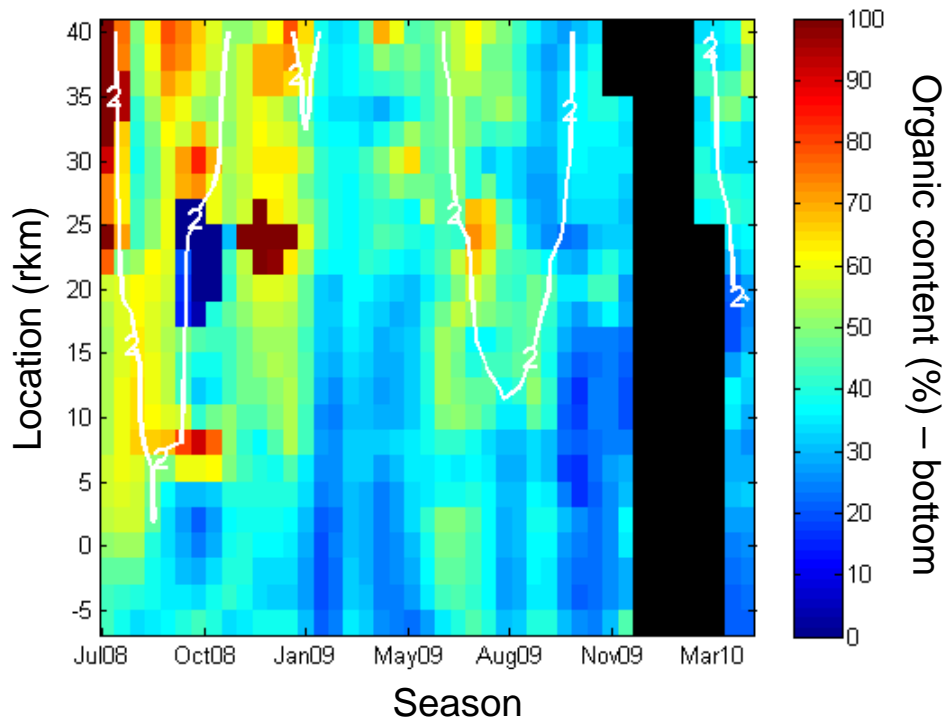


Figure 3.2.2.11. Time series of relative organic content determined from water samples collected near the bottom.

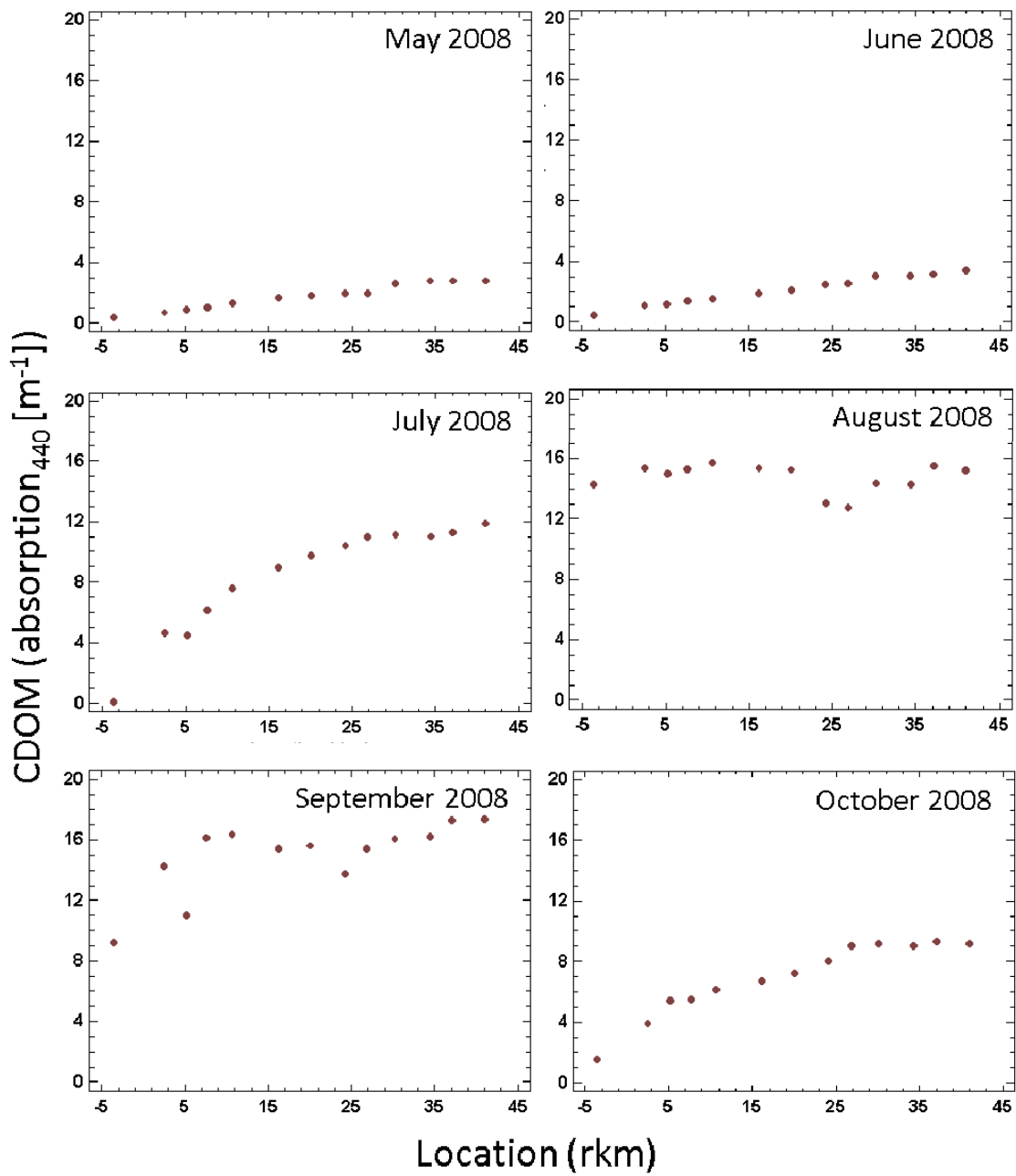


Figure 3.2.3.1. Vertically averaged colored dissolved organic matter (CDOM) calculated along the length of the study area May–October 2008.

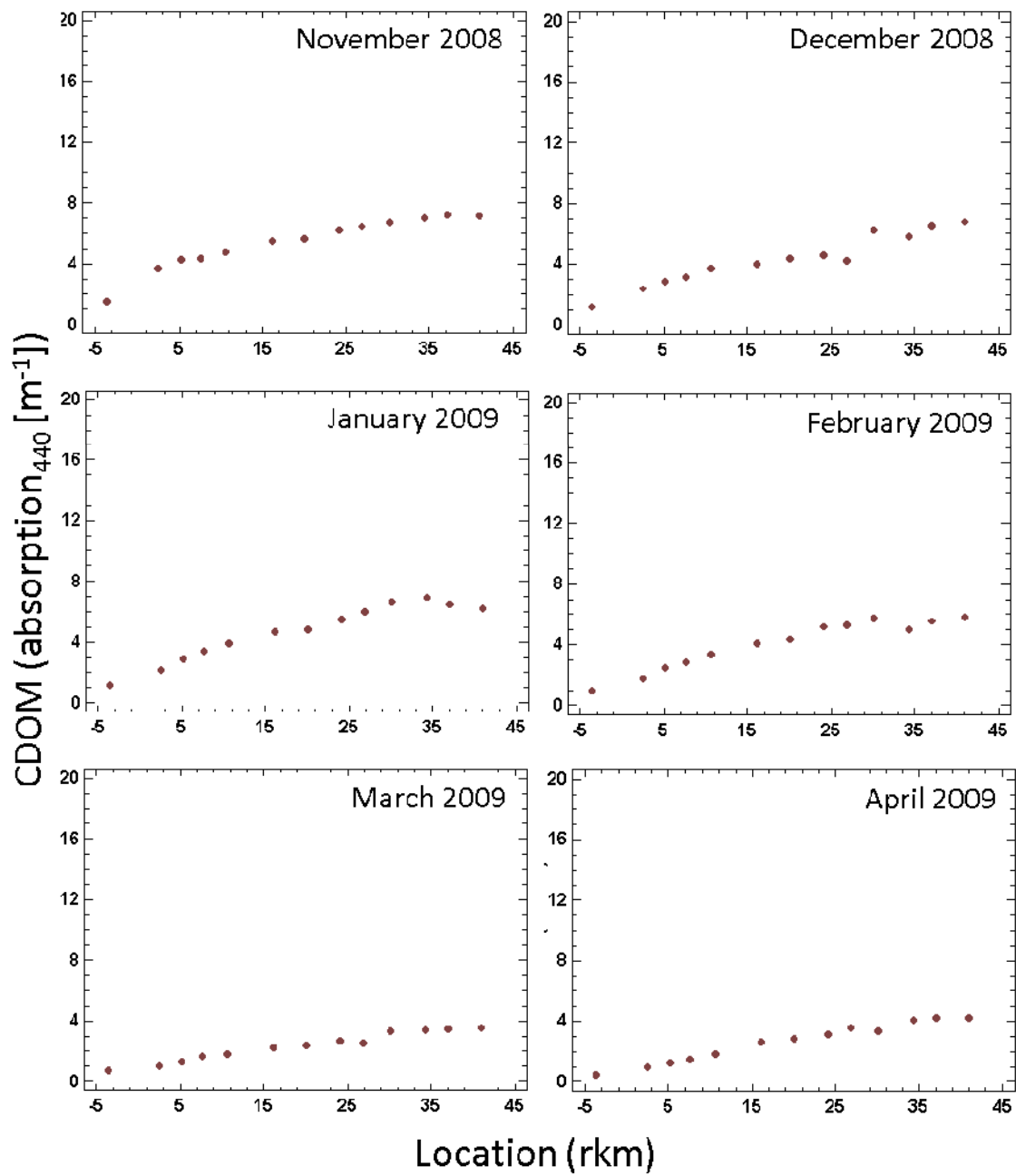


Figure 3.2.3.2. Vertically averaged colored dissolved organic matter (CDOM) calculated along the length of the study area November 2008–April 2009.

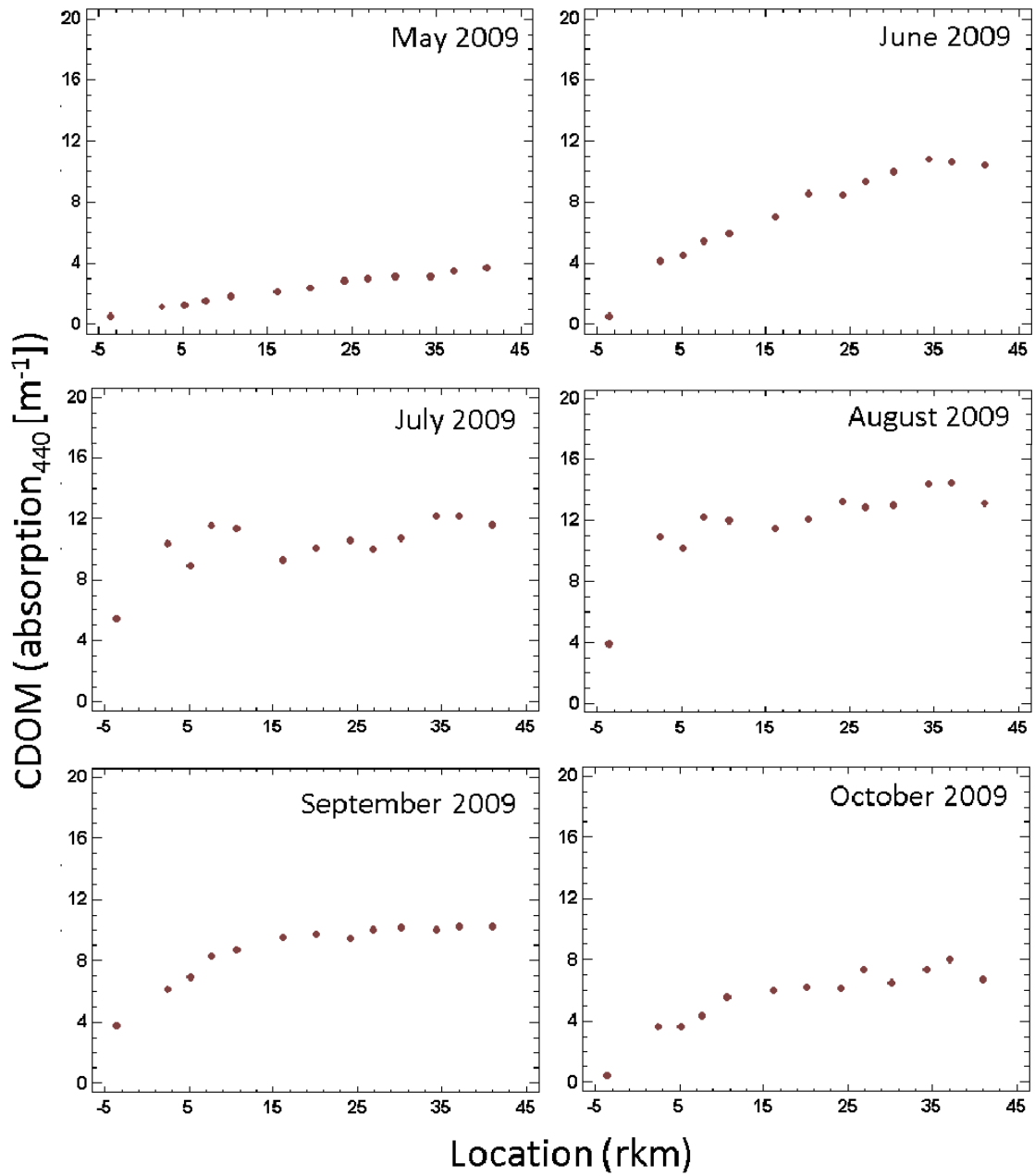


Figure 3.2.3.3. Vertically averaged colored dissolved organic matter (CDOM) calculated along the length of the study area May–October 2009.

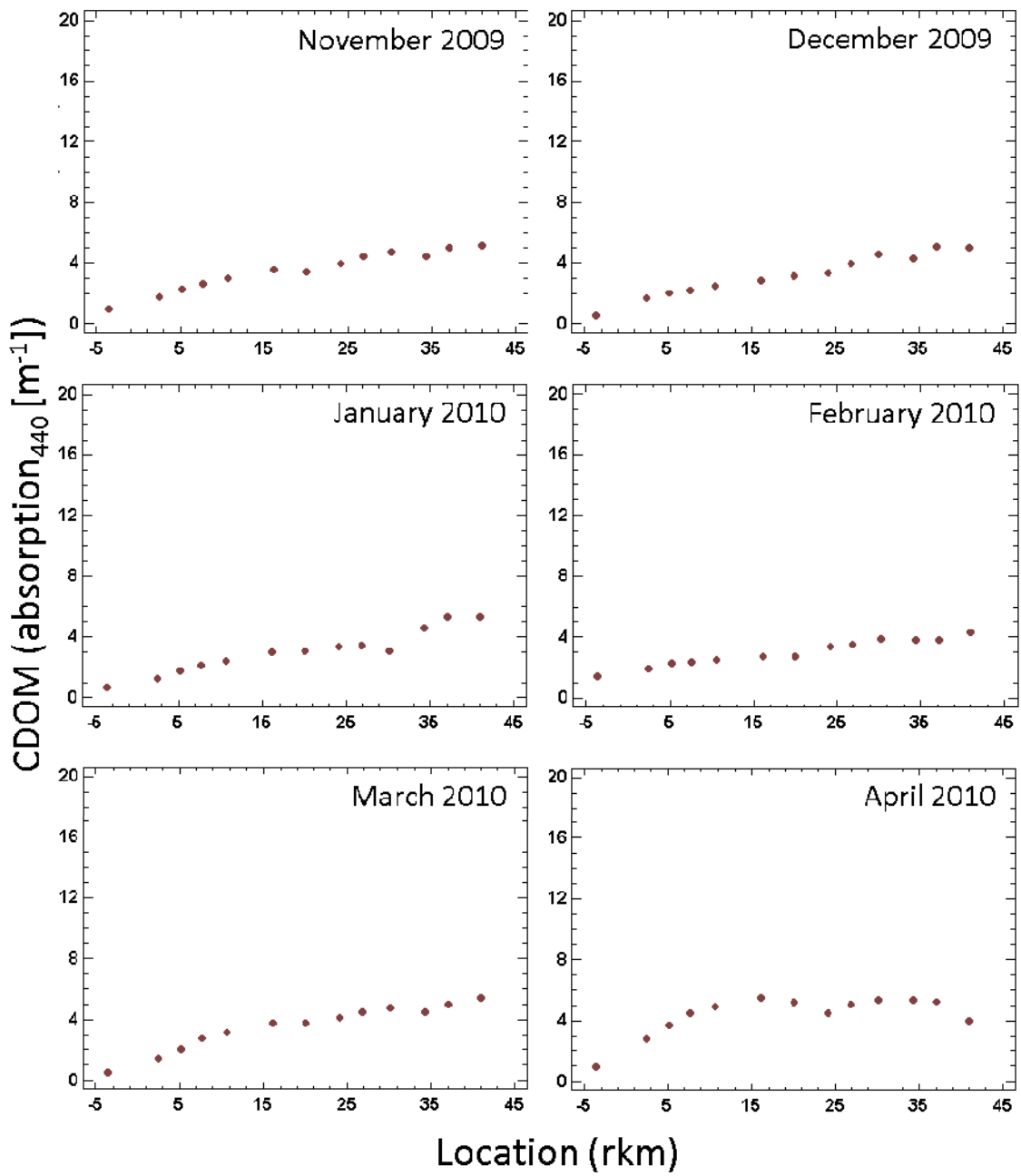


Figure 3.2.3.4. Vertically averaged colored dissolved organic matter (CDOM) calculated along the length of the study area November 2009–April 2010.

3.3 Water Column and Benthic Primary Production

Mean chlorophyll *a* concentration for each sampling station is presented in Table 3.2.1. In general, chlorophyll *a* concentrations were higher upstream, especially above 10.6 rkm, where mean concentrations were typically $>10 \mu\text{g L}^{-1}$.

A comparison of water column and benthic primary production along the length of the study area was made by examining monthly mean chlorophyll *a* concentration and maximum fluorescence (F_{max}) measured using the diving PAM fluorometer (Figs. 3.3.1–3.3.6) for the period May 2009–April 2010. These figures also illustrate that chlorophyll *a* concentrations were typically higher upstream and tended to decrease downstream, the exceptions occurring in July 2009 when concentrations were highest just inside the mouth of the river. In contrast, F_{max} values tended to be highest downstream, typically below the river mouth in San Carlos Bay. These downstream peaks in F_{max} were associated almost exclusively with pennate diatoms; when F_{max} peaks occurred upstream, they were more likely to be associated with large percentages of centric diatoms. These results suggest that production downstream was truly benthic in nature, with pennate diatoms dominating in the clear waters below the river mouth. Benthic production upstream was likely the result of phytoplankton sedimentation.

Comparison of water column chlorophyll *a* concentration (estimated using maximum fluorescence, relative fluorescence units) and effective quantum yield suggests that even though phytoplankton biomass was typically greater upstream, phytoplankton were often healthier downstream (Figs. 3.3.7–3.3.18). Although increased values of quantum yield often accompanied the chlorophyll maximum, especially during dry months (Fig. 3.3.13, 3.3.15–3.3.16), higher values of quantum yield, indication healthy phytoplankton populations, were often present downstream relative to the chlorophyll maximum during wet months (Figs. 3.3.10–3.3.12). These results suggest that phytoplankton occurring upstream during the wet season may have experienced salinity stress, with associated quantum yield values falling below 0.3.

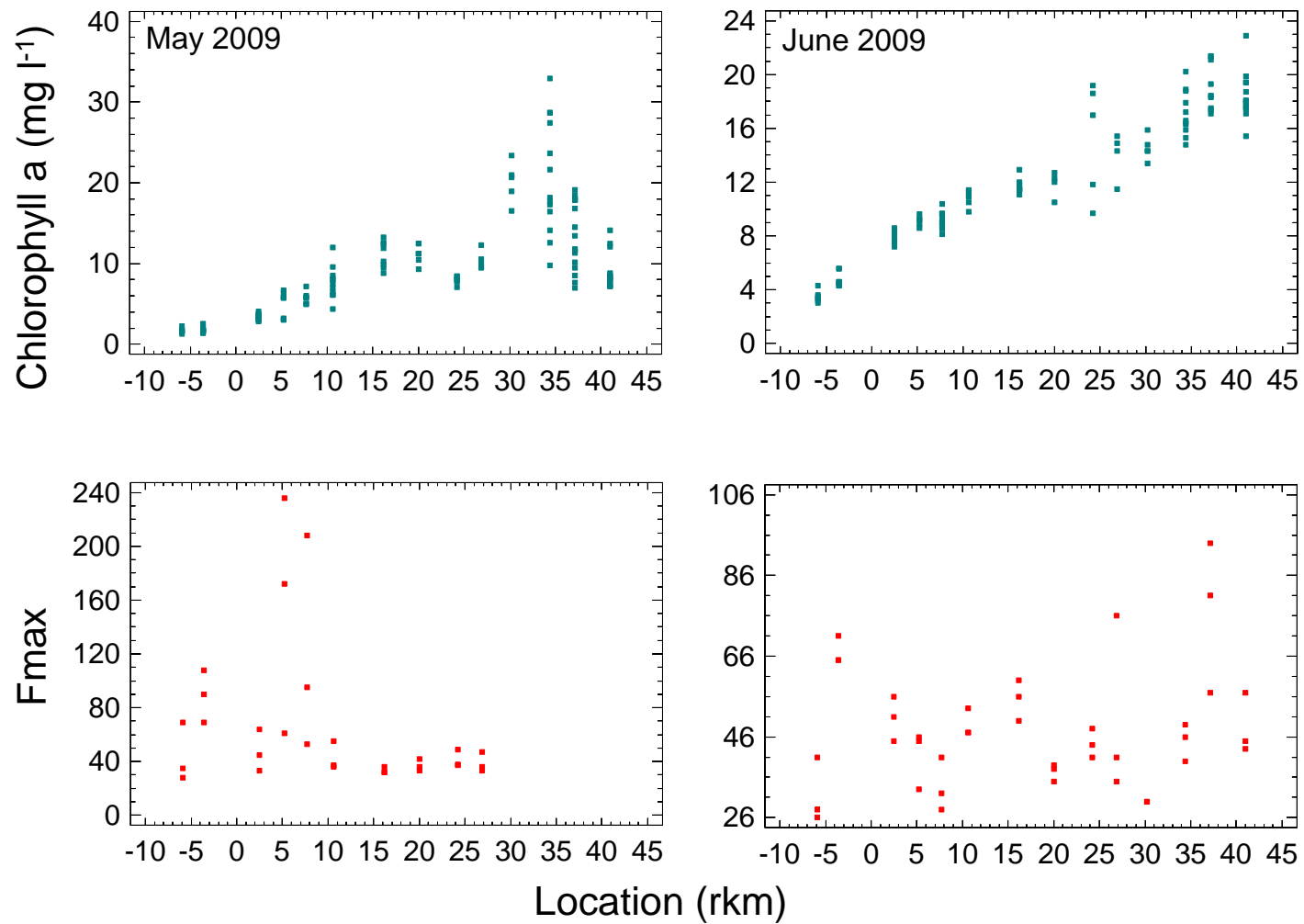


Figure 3.3.1. Comparison of water column (chlorophyll *a* concentration) and benthic primary production (F_{\max}) along the study area, May–June 2009.

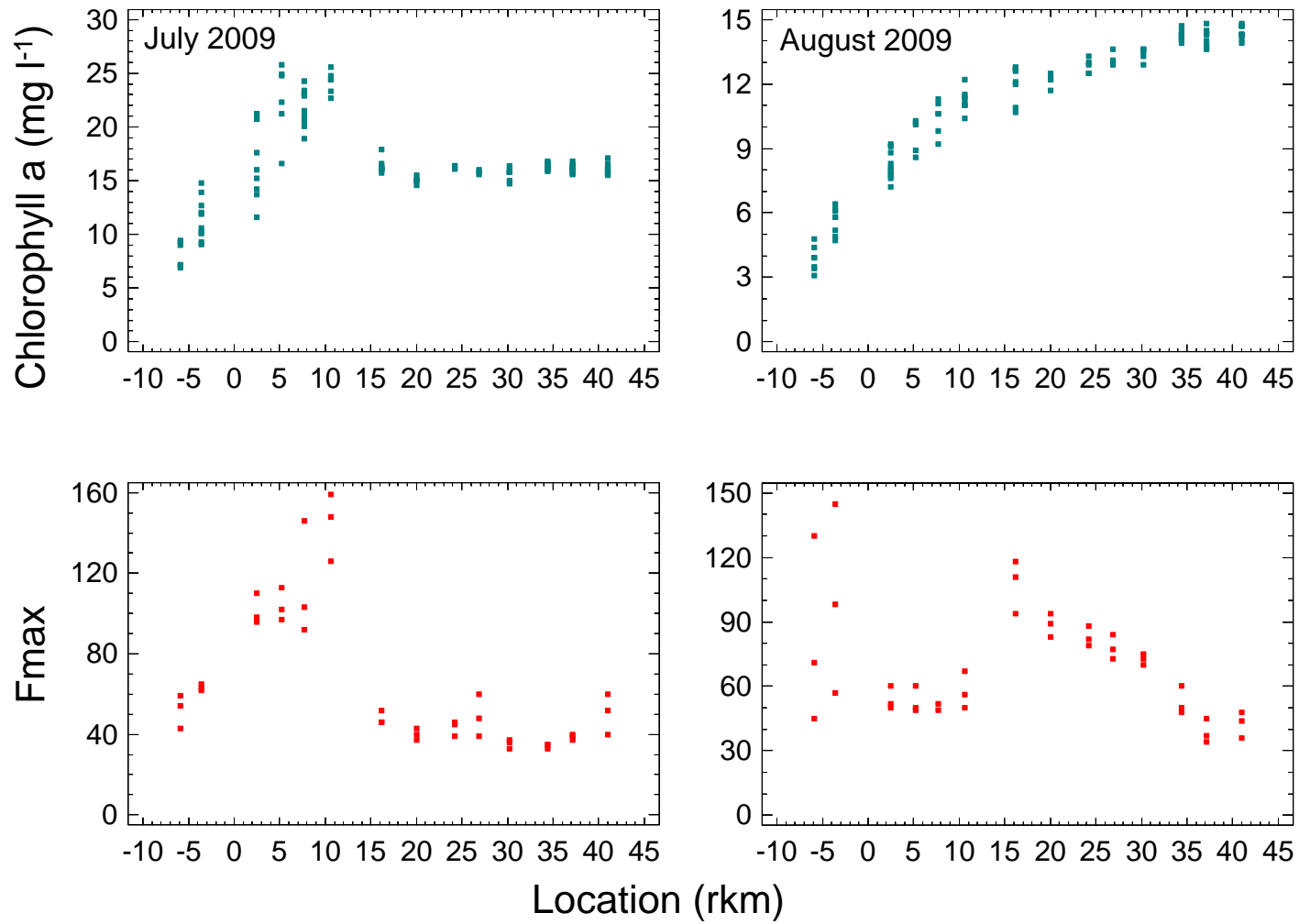


Figure 3.3.2. Comparison of water column (chlorophyll *a* concentration) and benthic primary production (F_{\max}) along the study area, July–August 2009.

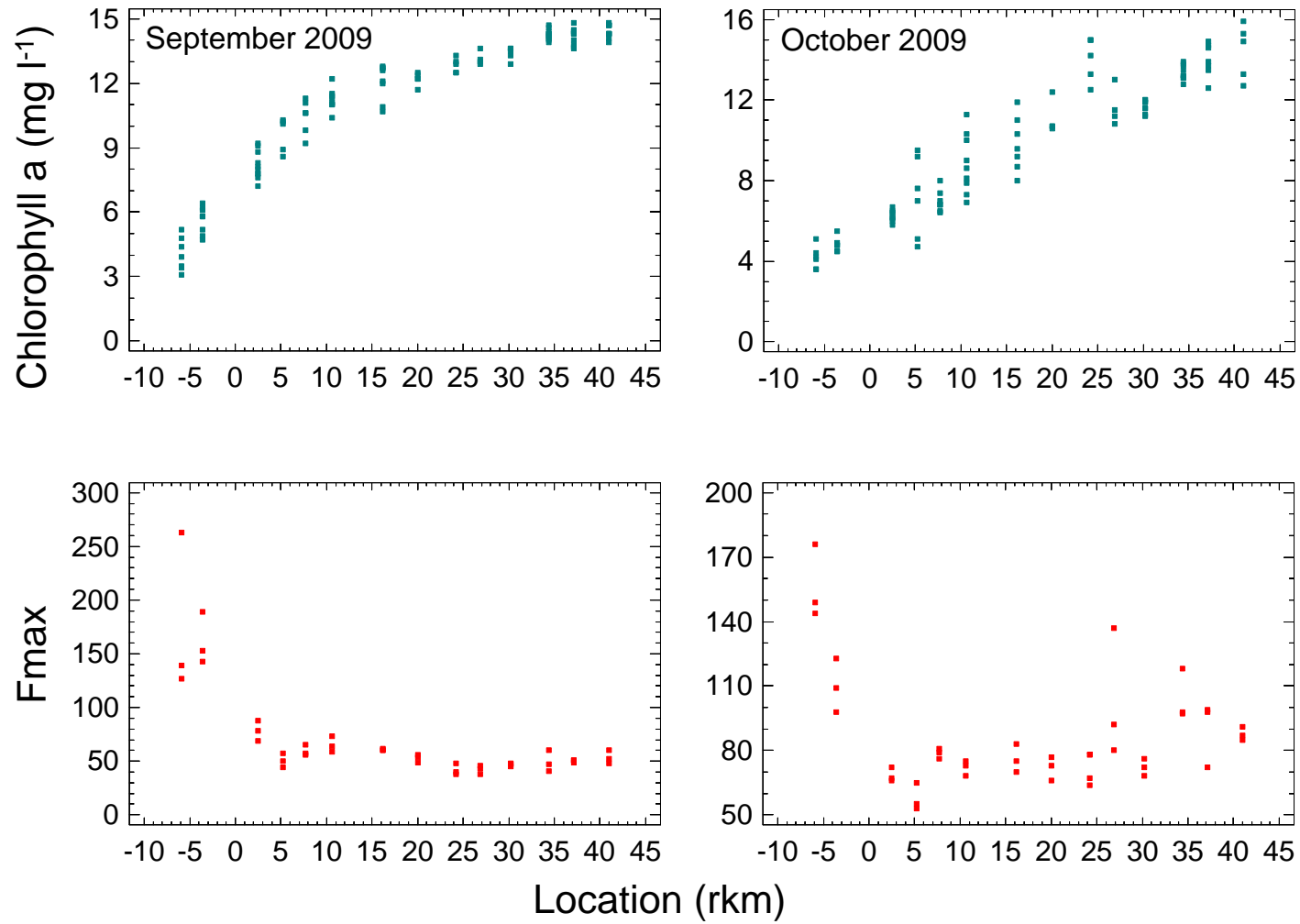


Figure 3.3.3. Comparison of water column (chlorophyll *a* concentration) and benthic primary production (F_{\max}) along the study area, September–October 2009.

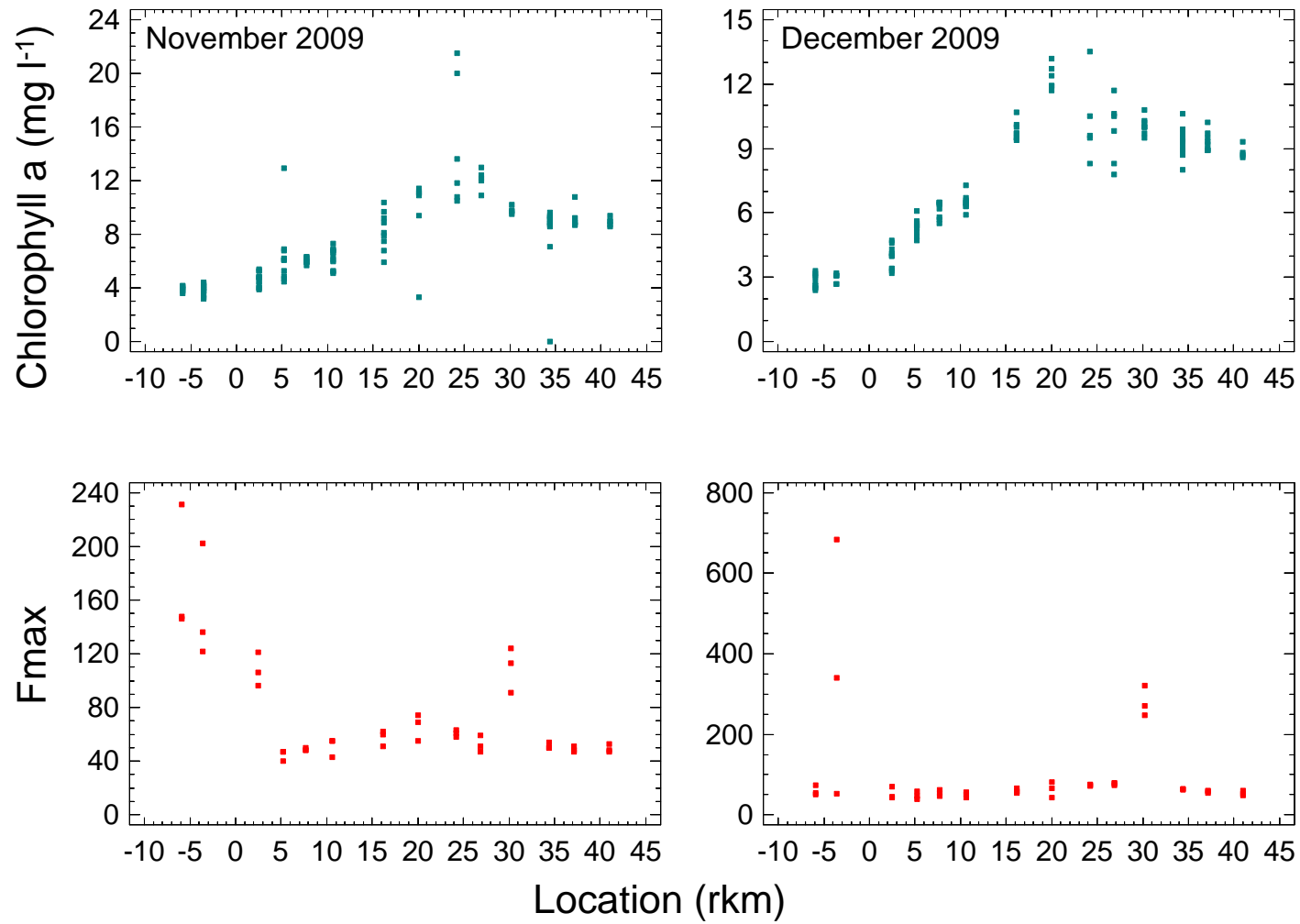


Figure 3.3.4. Comparison of water column (chlorophyll *a* concentration) and benthic primary production (F_{\max}) along the study area, November–December 2009.

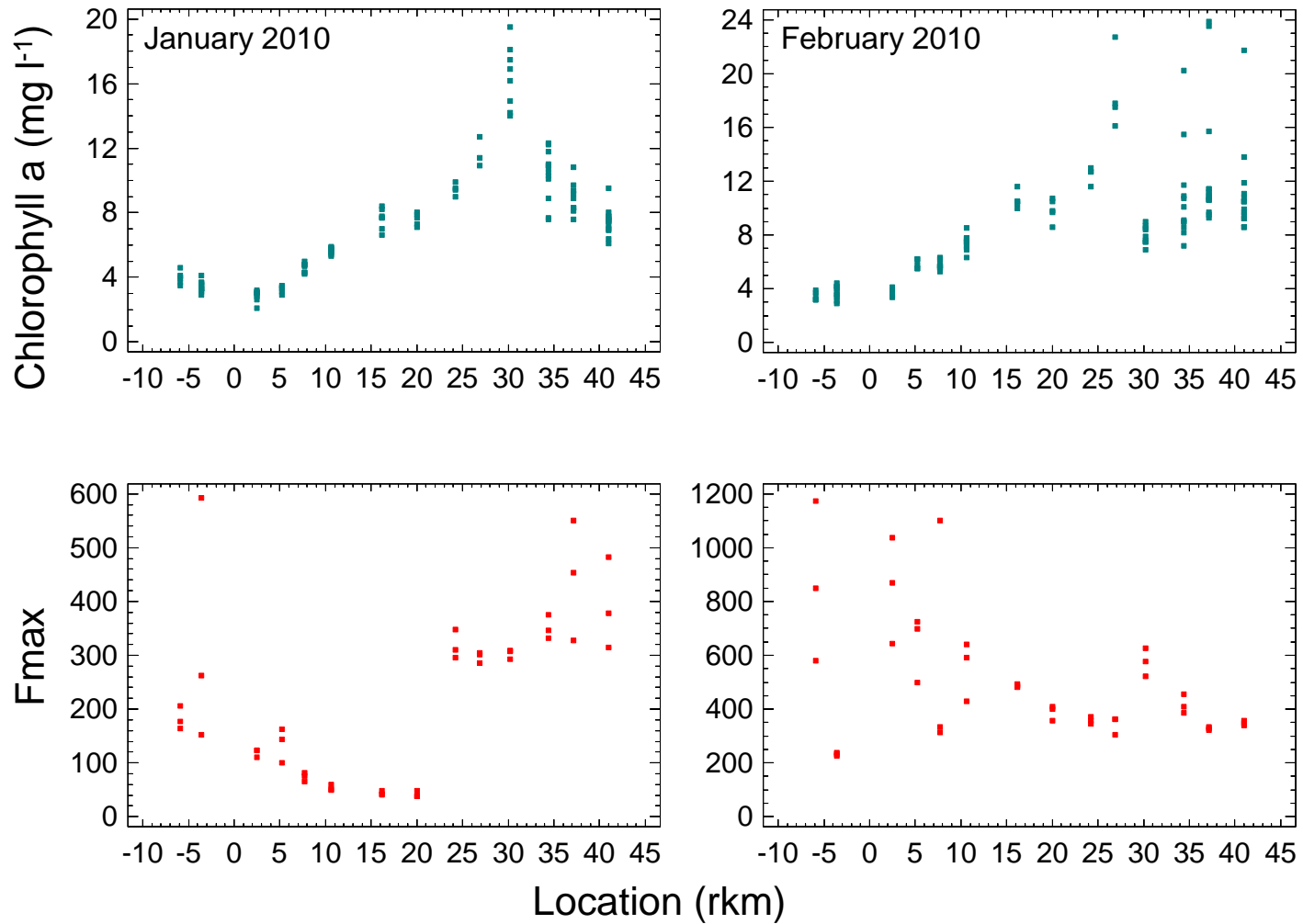


Figure 3.3.5. Comparison of water column (chlorophyll *a* concentration) and benthic primary production (F_{\max}) along the study area, January–February 2010.

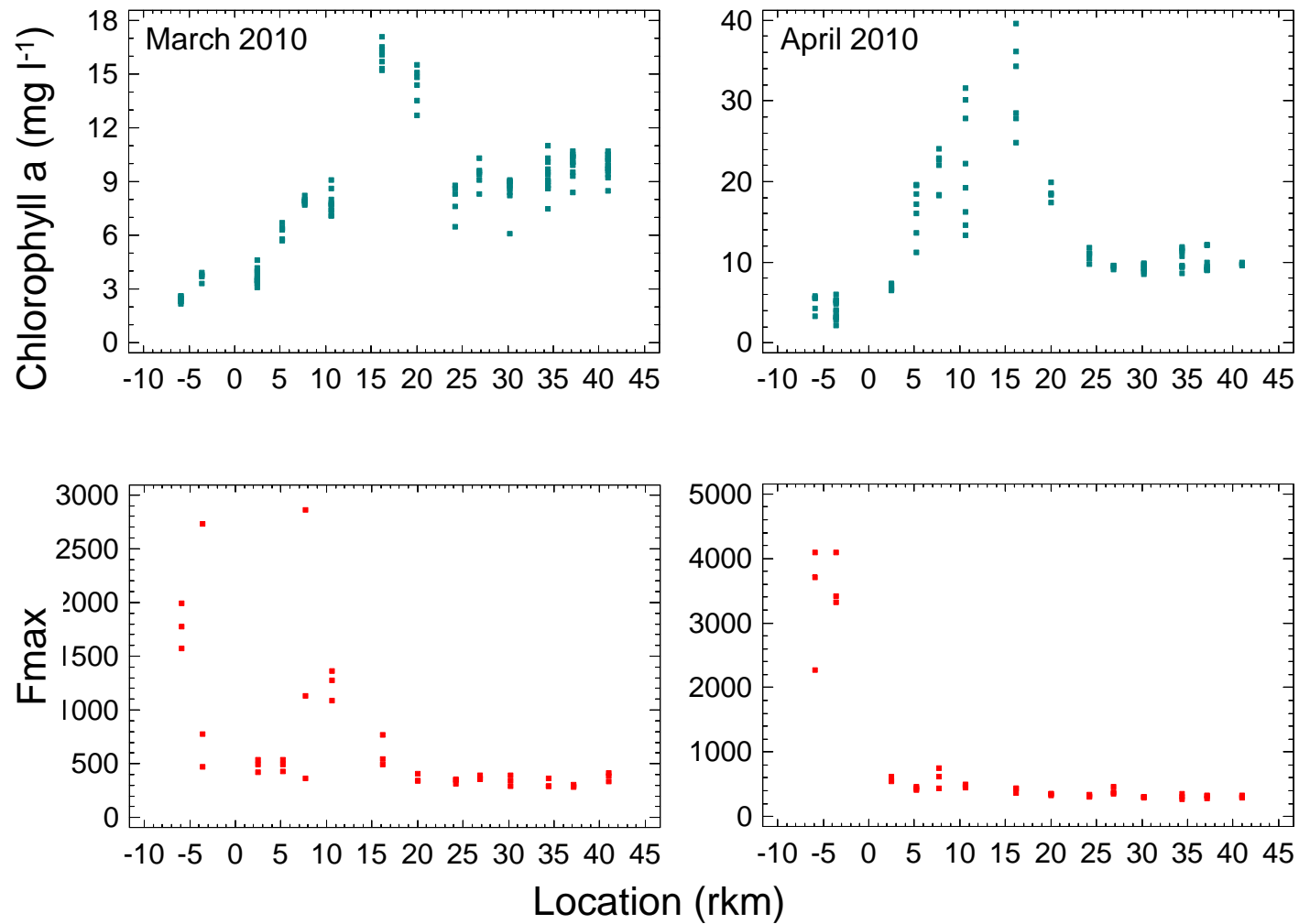


Figure 3.3.6. Comparison of water column (chlorophyll *a* concentration) and benthic primary production (F_{\max}) along the study area, March–April 2010.

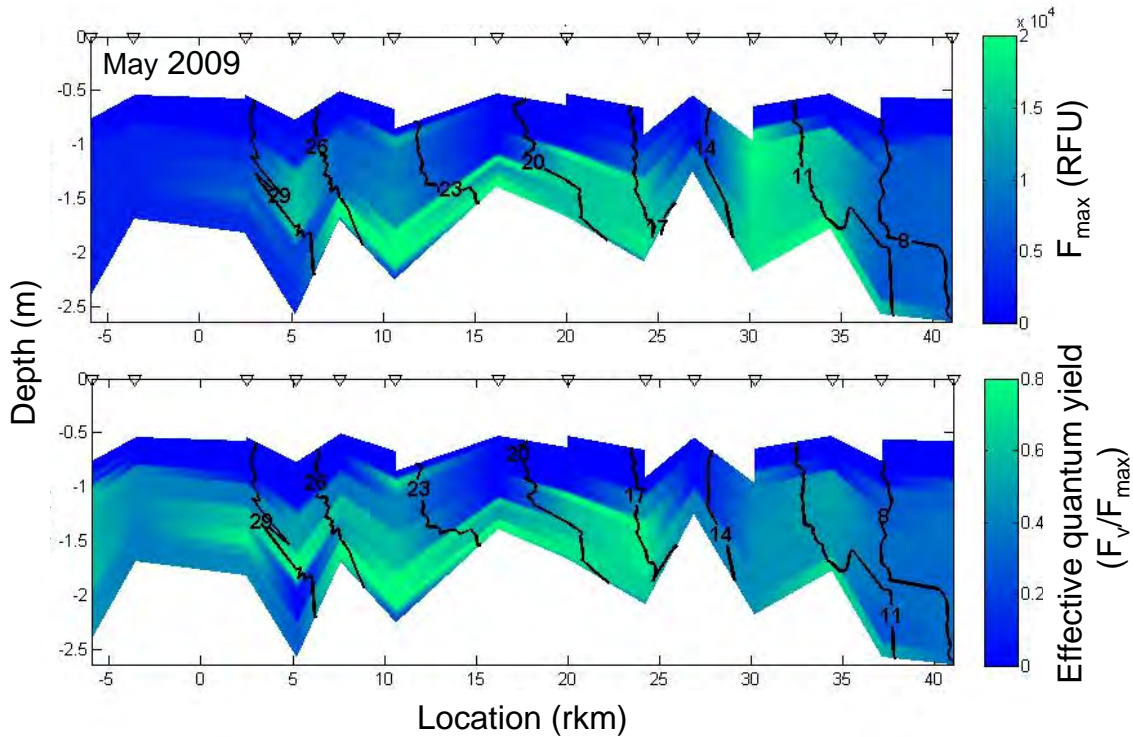


Figure 3.3.7. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for May 2009. Triangles denote profile locations and lines represent isohalines.

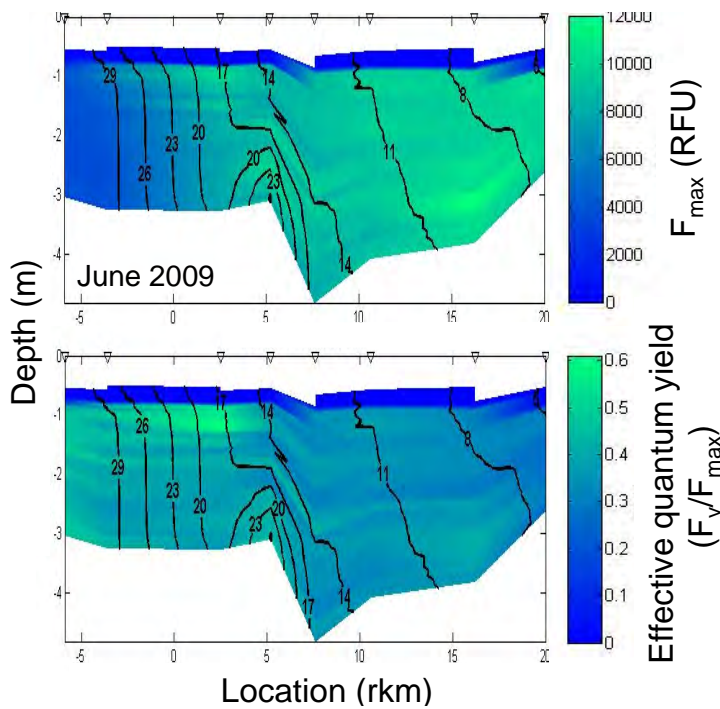


Figure 3.3.8. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for June 2009. Triangles denote profile locations and lines represent isohalines. Data are lacking in the upper portion of Caloosahatchee due to instrument failure.

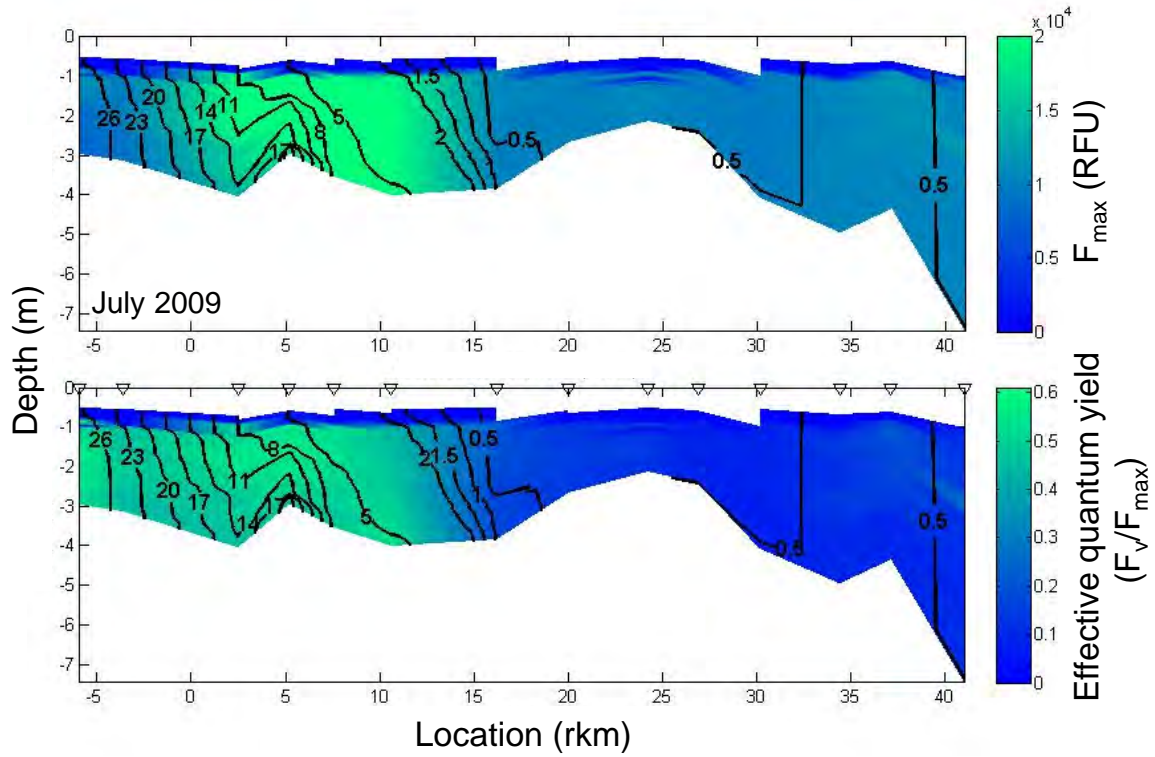


Figure 3.3.9. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for July 2009. Triangles denote profile locations and lines represent isohalines.

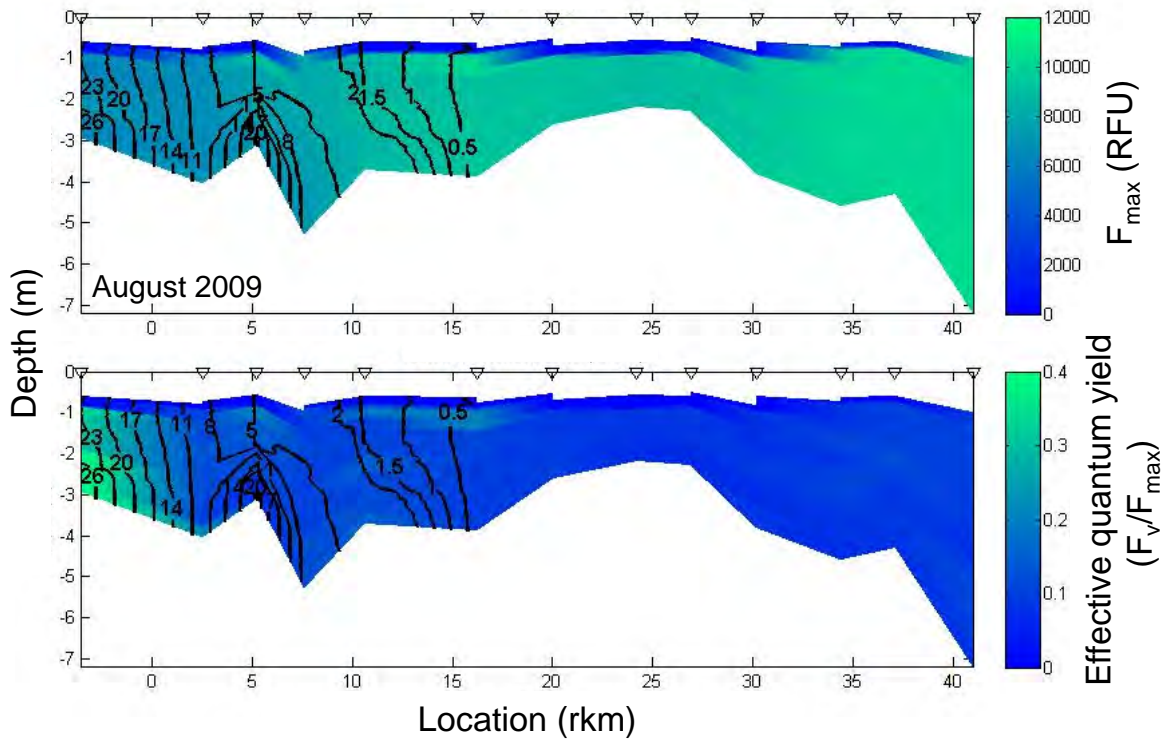


Figure 3.3.10. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for August 2009. Triangles denote profile locations and lines represent isohalines.

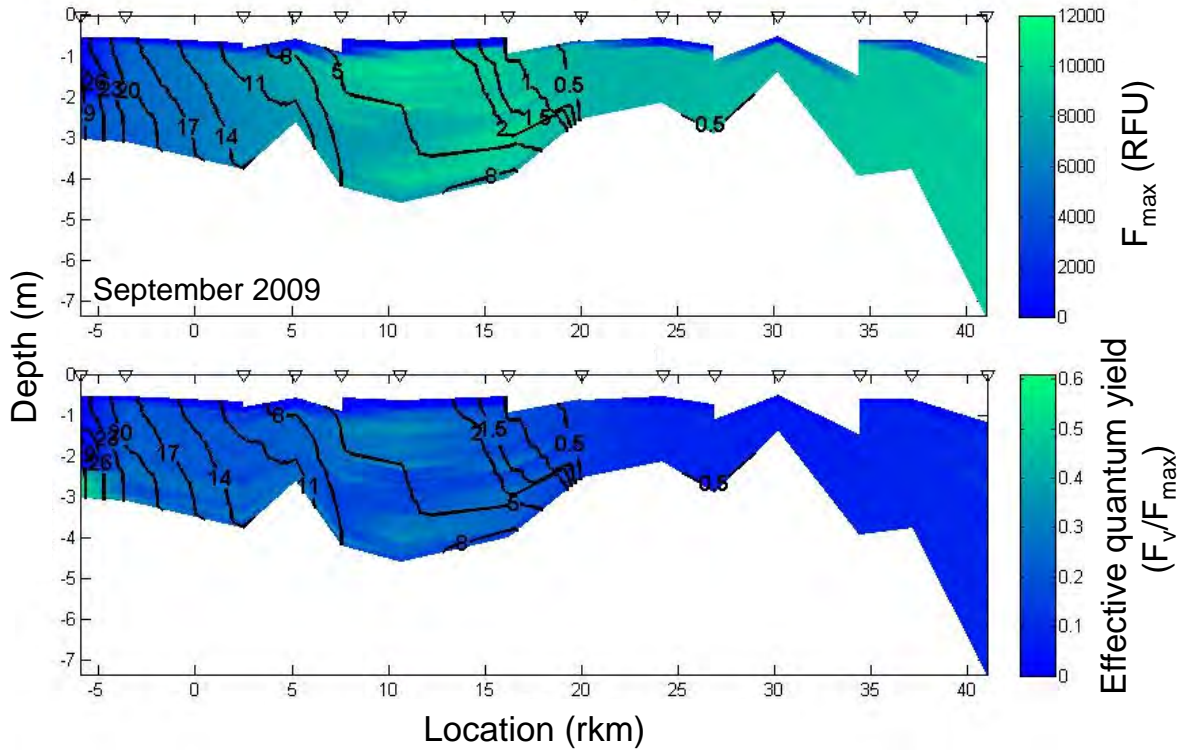


Figure 3.3.11. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for September 2009. Triangles denote profile locations and lines represent isohalines.

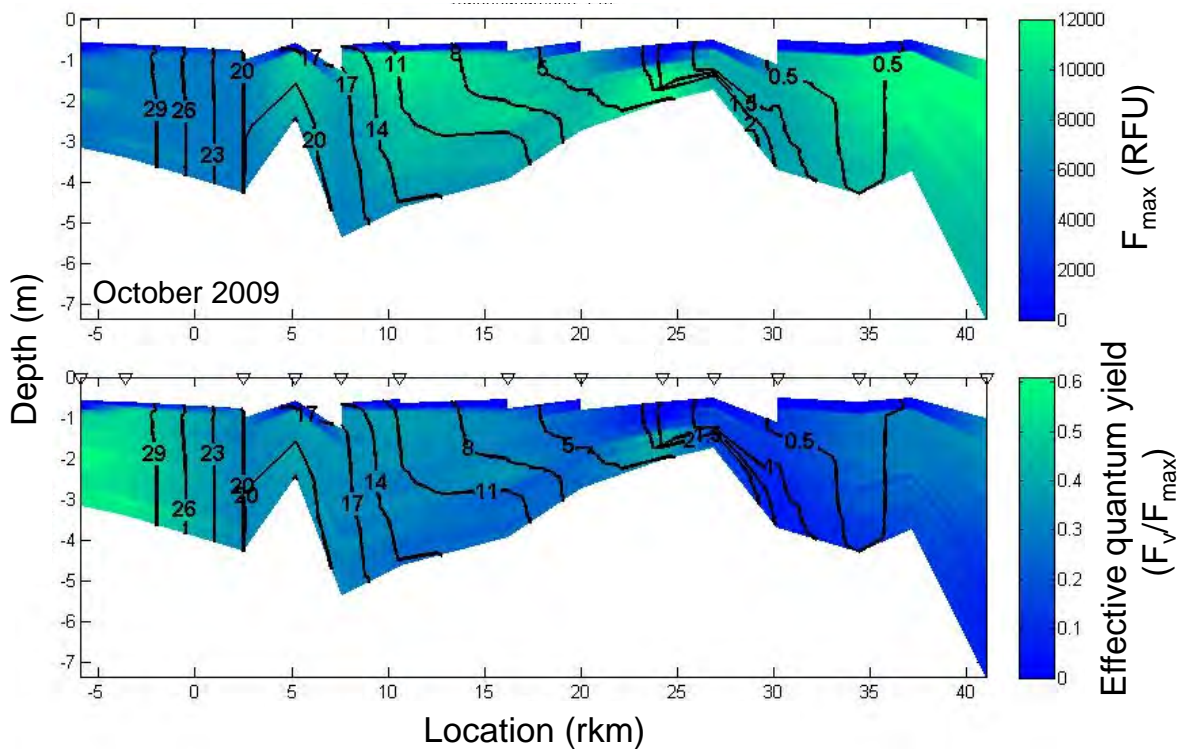


Figure 3.3.12. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for October 2009. Triangles denote profile locations and lines represent isohalines.

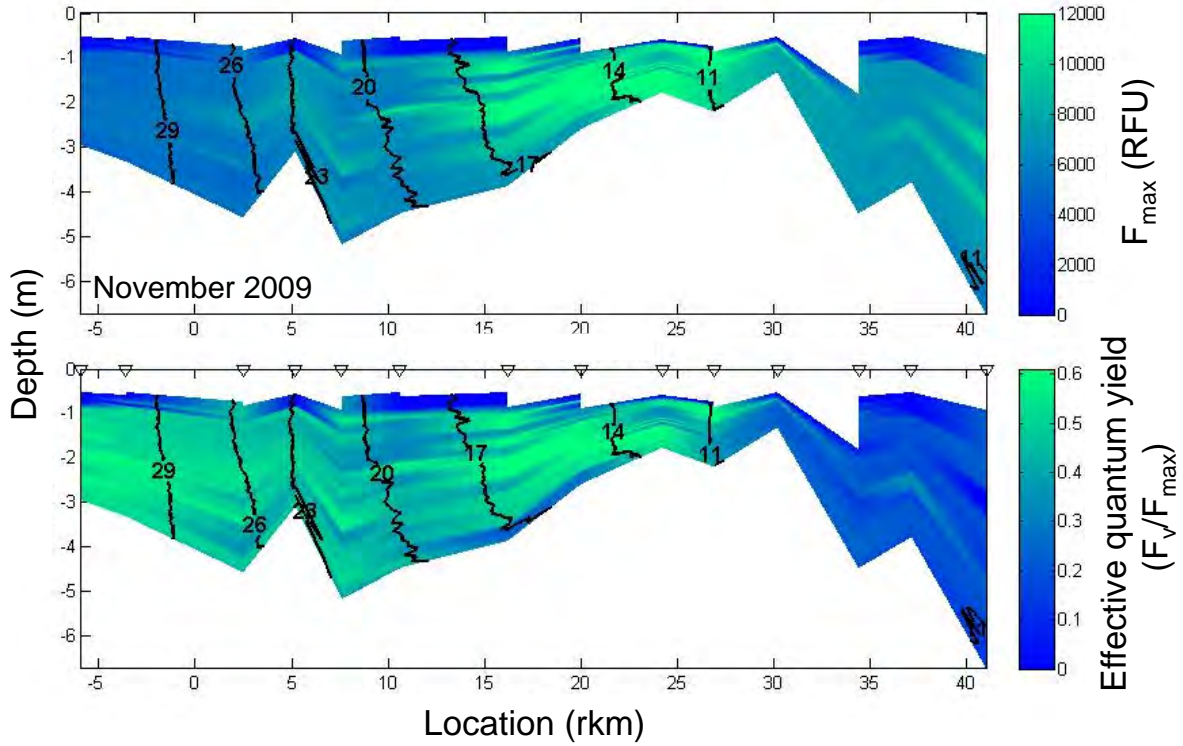


Figure 3.3.13. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for November 2009. Triangles denote profile locations and lines represent isohalines.

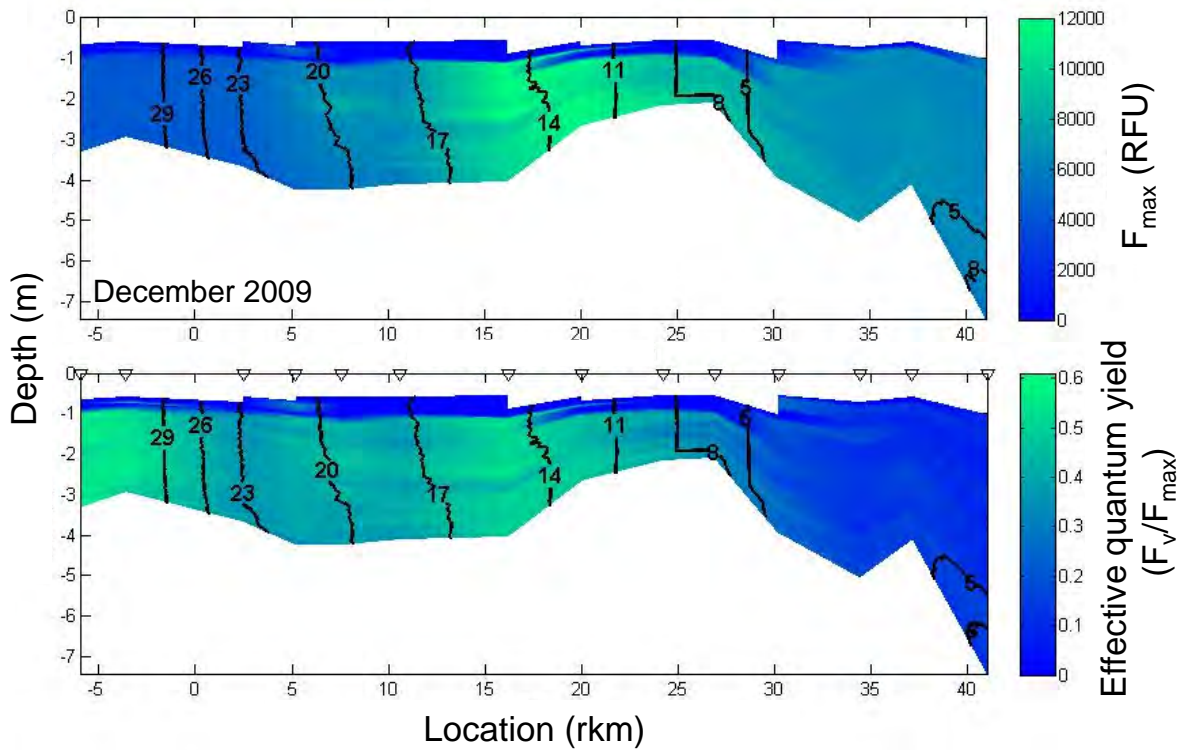


Figure 3.3.14. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for December 2009. Triangles denote profile locations and lines represent isohalines.

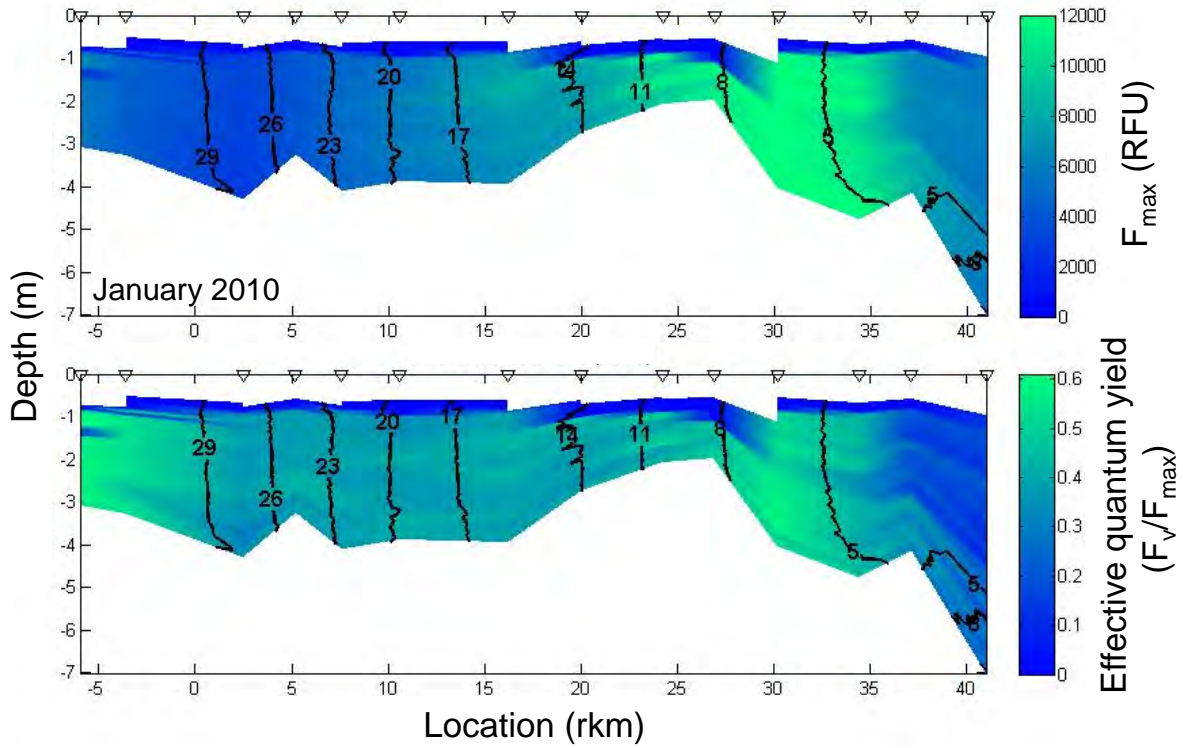


Figure 3.3.15. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for January 2010. Triangles denote profile locations and lines represent isohalines.

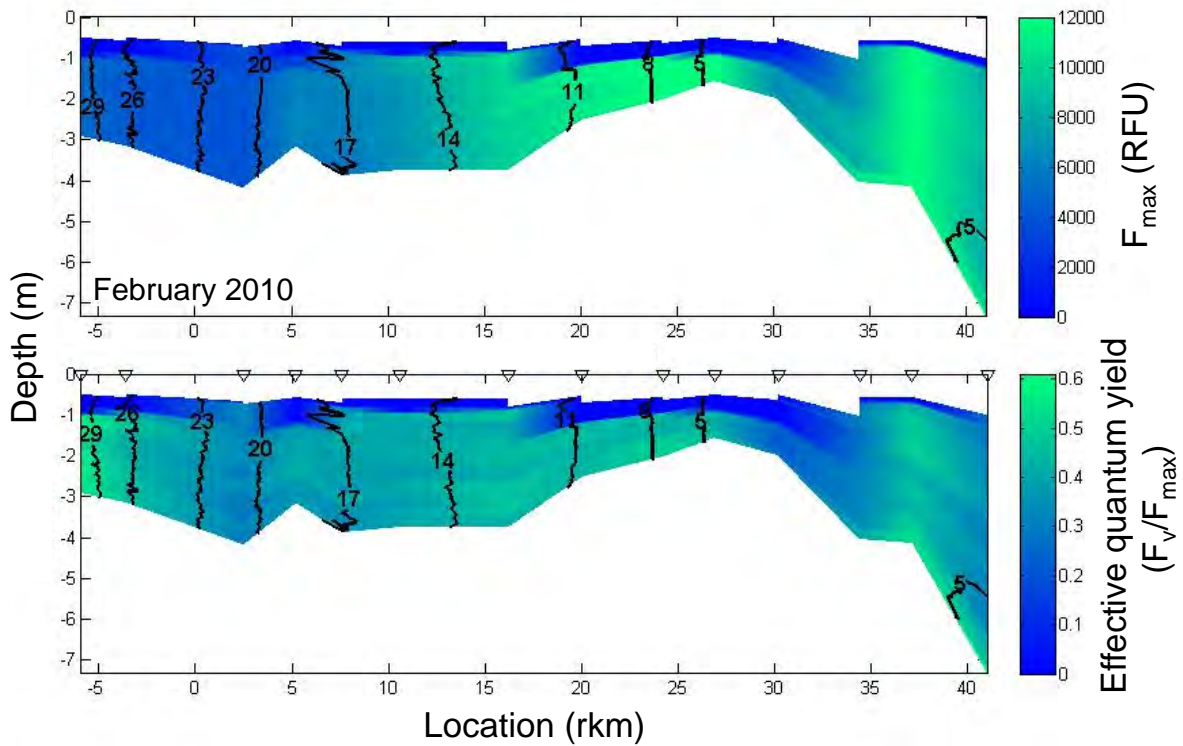


Figure 3.3.16. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for February 2010. Triangles denote profile locations and lines represent isohalines.

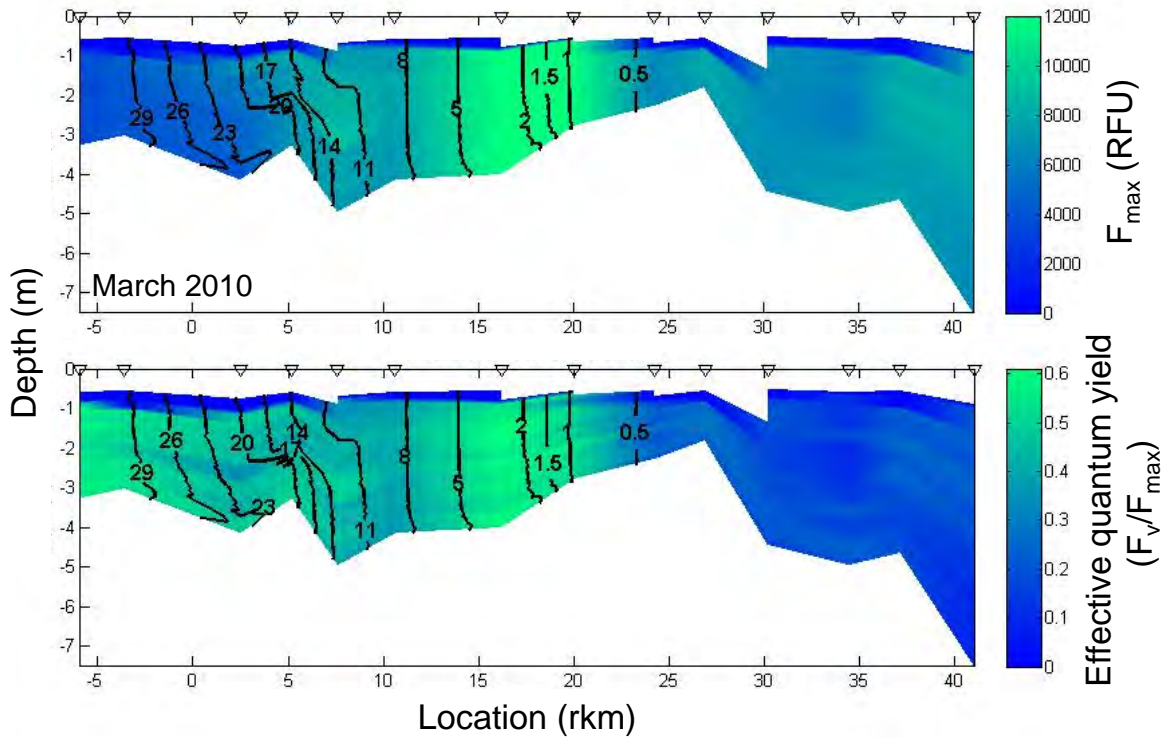


Figure 3.3.17. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for March 2010. Triangles denote profile locations and lines represent isohalines.

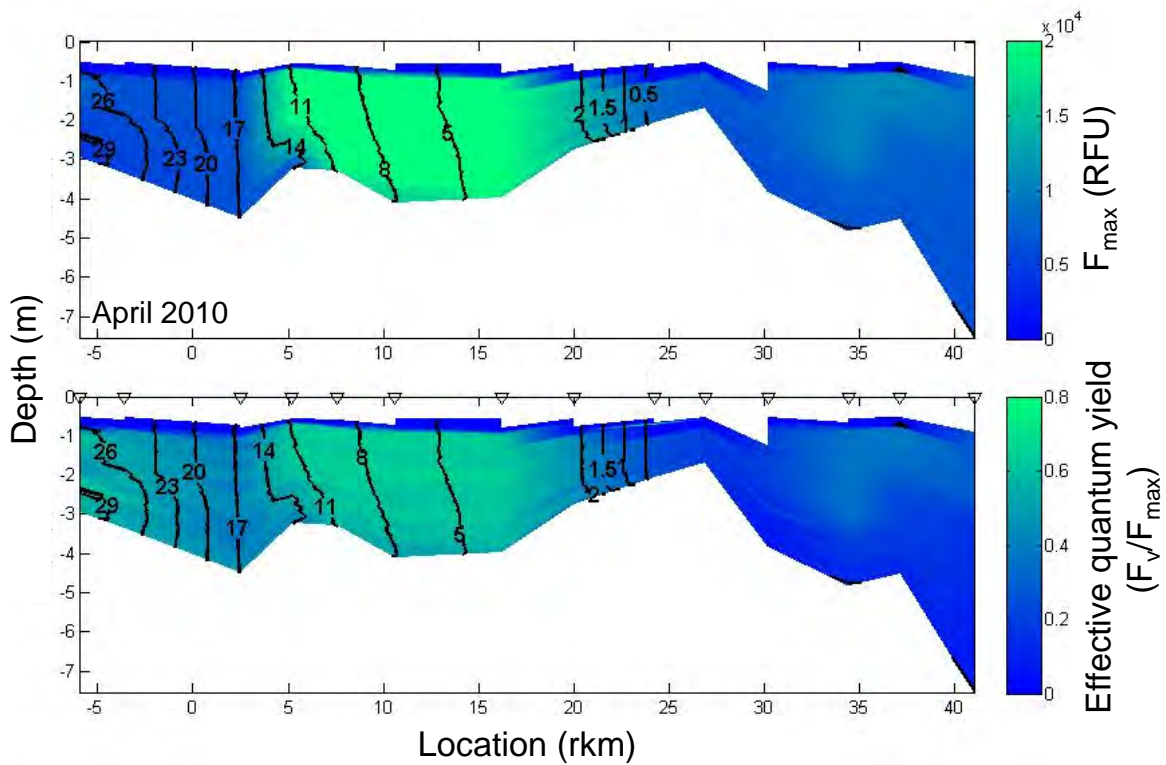


Figure 3.3.18. Longitudinal distribution of maximum fluorescence (top) and quantum yield (bottom) for April 2010. Triangles denote profile locations and lines represent isohalines.

3.4 Phytoplankton Composition and Changes in Community Structure

A total of 93 net tow samples were analyzed for this project. Data analysis focused on stations with the highest vertically-averaged chlorophyll concentrations from each sampling trip ($n = 24$) based on descending profiles. An unforeseen lag in the data collection/processing of the YSI (12-second integration) resulted in incomplete data collection during the descent of the YSI, resulting in erroneous chlorophyll estimates. Consequently, many of the 24 samples selected for further analysis did not represent chlorophyll peaks from each sampling trip. Comparisons with chlorophyll data from water grab samples collected and analyzed from the stations, however, indicates that 29% of the samples were confirmed from the chlorophyll peaks, whereas another 33% of samples were from a station adjacent to the location of maximum chlorophyll concentration. Therefore, 62% of samples were at least proximal to the chlorophyll peak and the associated maximal phytoplankton biomass. Subsequent chlorophyll data analyses (see below) used ascending profiles of the YSI, which alleviated some of the concerns with the lagged collection of chlorophyll data.

There were 181 different phytoplankton taxa encountered in this study, 86 (47%) of which were diatoms (Table 3.4.1; Appendices B-C). Cyanobacteria and diatoms were most abundant in terms of cells per liter of water (Table 3.4.2), although the dinoflagellate *Akashiwo sanguineum* was the most abundant phytoplankter based on total cell volume (i.e., there were fewer cells, but these cells were large). The composite group, cyanobacteria filament, is likely composed of multiple, non-descript species similar in appearance to *Jaaginema*. This group will require further taxonomic (and molecular) work to identify the taxa present. *Skeletonema subsalsum* was the most commonly occurring phytoplankter, being present in 83% of the samples (Table 3.4.3). Generally speaking, diatoms and cyanobacteria were the most abundant groups of phytoplankton.

Table 3.4.1. Classes or phyla of phytoplankton encountered and the number of species within each group. The abbreviation for each group is given in parenthesis.

| Class/Phylum | # of species |
|-----------------------|--------------|
| Bacillariophyceae (D) | 86 |
| Chlorophyta (CH) | 38 |
| Dinophyta (I) | 16 |
| Cyanophyta (CY) | 16 |
| Prymnesiophyceae (F) | 8 |
| Euglenophyta (E) | 7 |
| Ciliophora (CI) | 4 |
| Cryptophyta (CX) | 3 |
| Haptophyta (CR) | 2 |
| Phaeophyta (BR) | 1 |
| TOTAL | 181 |

Table 3.4.2. Top ten phytoplankton taxa encountered in terms of cells L⁻¹ abundance. Sampling event, date, and station correspond to occurrence of maximal abundance for each taxon. Class (or phylum) is also noted for each taxon (see Table 1).

| Sampling Event | Sampling Date | Station | species | cells/L | class |
|----------------|---------------|---------|---------------------------------------|----------|-------|
| 17 | Sep 2009 | 6U | cyanobacteria filament | 2262433 | CY |
| 24 | Apr 2010 | 4D | <i>Skeletonema subsalsum</i> | 1690646 | D |
| 17 | Sep 2009 | 6U | <i>Planktolyngbya</i> sp. | 269229.5 | CY |
| 18 | Oct 2009 | 5U | <i>Aulacoseira</i> sp. | 178149.6 | D |
| 15 | Jul 2009 | 3U | <i>Melosira varians</i> | 149457.2 | D |
| 1 | May 2008 | 7D | <i>Pseudo-nitzschia pungens</i> group | 128800 | D |
| 2 | Jun 2008 | 6D | <i>Planktothrix</i> sp. | 118657.2 | CY |
| 16 | Aug 2009 | 7U | <i>Skeletonema tropicum</i> | 101650.2 | D |
| 16 | Aug 2009 | 7U | <i>Fragilaria</i> 5-10 | 72607.26 | D |
| 14 | Jun 2009 | 6U | <i>Microcystis</i> sp. | 48920.36 | CY |

Table 3.4.3. Top ten phytoplankton taxa encountered in terms of cell volume (μm^3). Sampling event, date, and station are noted for when the maximal abundance was recorded. Class (or phylum) is also noted for each taxon (see Table 1).

| Sampling Event | Sampling Date | Station | Species | Total volume (μm^3) | Class |
|----------------|---------------|---------|---------------------------------------|----------------------------------|-------|
| 9 | Jan 2009 | 5U | <i>Gymnodinium sanguineum</i> | 2.22E+08 | I |
| 7 | Nov 2008 | 5U | <i>Skeletonema subsalsum</i> | 1.53E+08 | D |
| 23 | Mar 2010 | 4D | <i>Surirella</i> sp. | 1.41E+08 | D |
| 19 | Nov 2009 | 5U | <i>Cerataulina bicornis</i> | 1.1E+08 | D |
| 1 | May 2008 | 7D | <i>Pseudo-nitzschia pungens</i> group | 94836753 | D |
| 16 | Aug 2009 | 7U | <i>Skeletonema tropicum</i> | 29938420 | D |
| 7 | Nov 2008 | 5U | armored auto dino >30 | 27525874 | I |
| 15 | Jul 2009 | 3U | <i>Melosira varians</i> | 18341137 | D |
| 18 | Oct 2009 | 5U | <i>Aulacoseira</i> sp. | 17489783 | D |
| 2 | Jun 2008 | 6D | <i>Planktothrix</i> sp. | 13105275 | CY |

Results of multidimensional scaling (Fig. 3.4.1) indicate that variations in the phytoplankton assemblage were significantly related ($r = 0.56$; $p < 0.01$) to salinity and river flow (averaged over previous 30 d). Specifically, diatom abundance decreased and cyanobacteria abundance increased as river flow increased (Fig. 3.4.2). Accompanying this shift in phytoplankton, cell sizes also decreased with increasing flow (Fig. 3.4.3), likely reflecting this shift from diatoms to cyanobacteria.

Accompanying this shift towards smaller, cyanobacteria cells were changes in the location of the chlorophyll maximum within the river and estuary. In May 2008, the chlorophyll maximum was located in zone 7 (near S-79), likely reflecting the lack of flow through the structure and the accumulation of biomass in the upper reaches of the estuary due to tidal forcing. As flows began to increase through June and July 2008, the chlorophyll maximum moved downstream and appears to have been flushed out by August (Fig. 3.4.4).

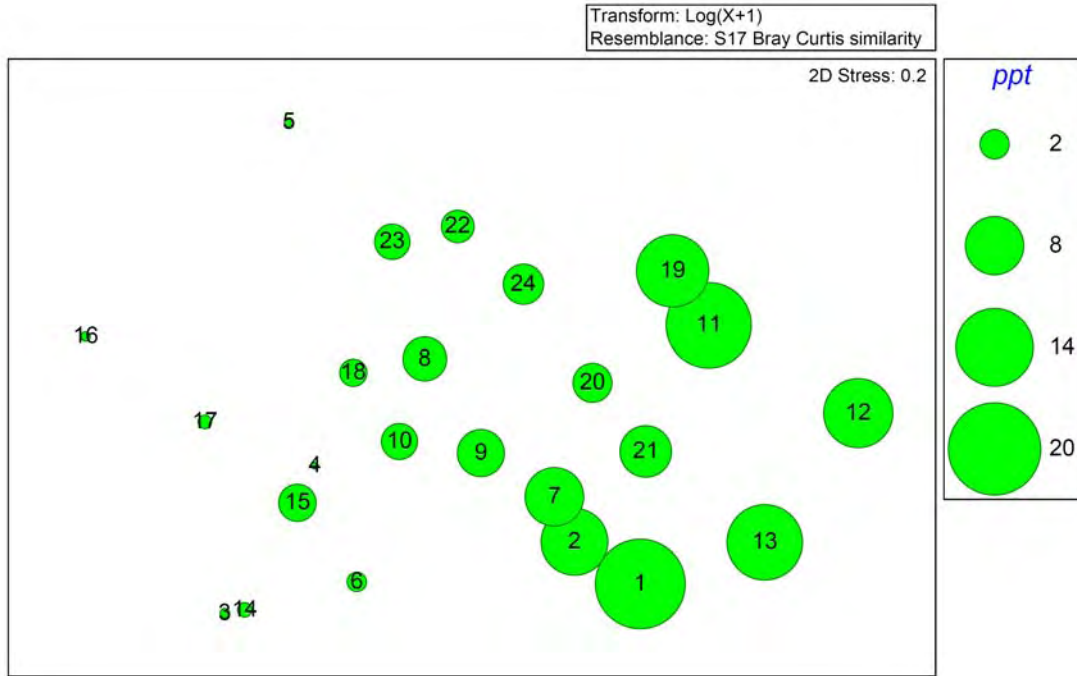


Figure 3.4.1. MDS plot of phytoplankton species >10% occurrence. Sampling month (1-24) is depicted and vertically-averaged salinity is noted via the bubble plot.

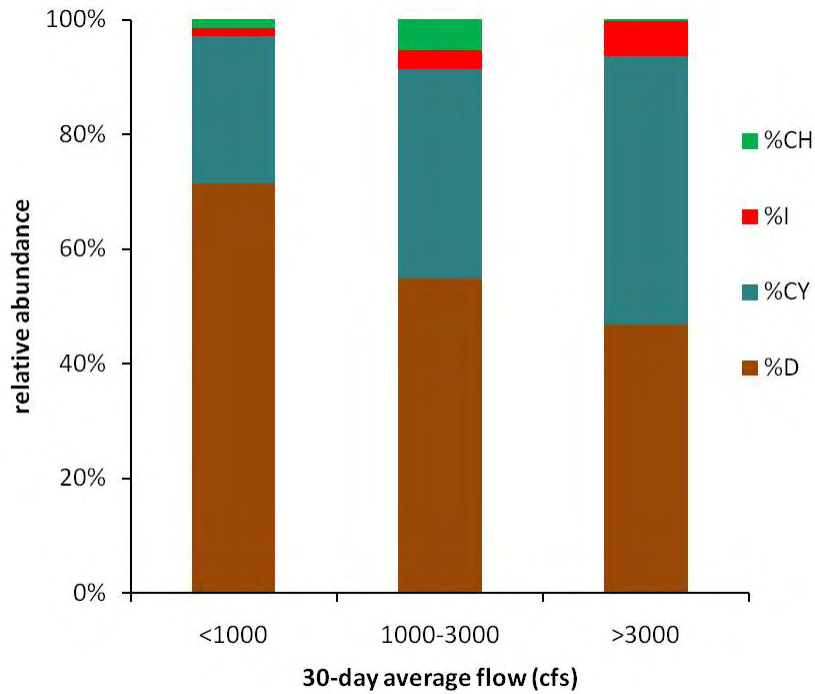


Figure 3.4.2. Abundance of cyanobacteria (CY), diatoms (D), dinoflagellates (I), and chlorophytes (CH) relative to mean freshwater inflow through S-79 for previous 30 days.

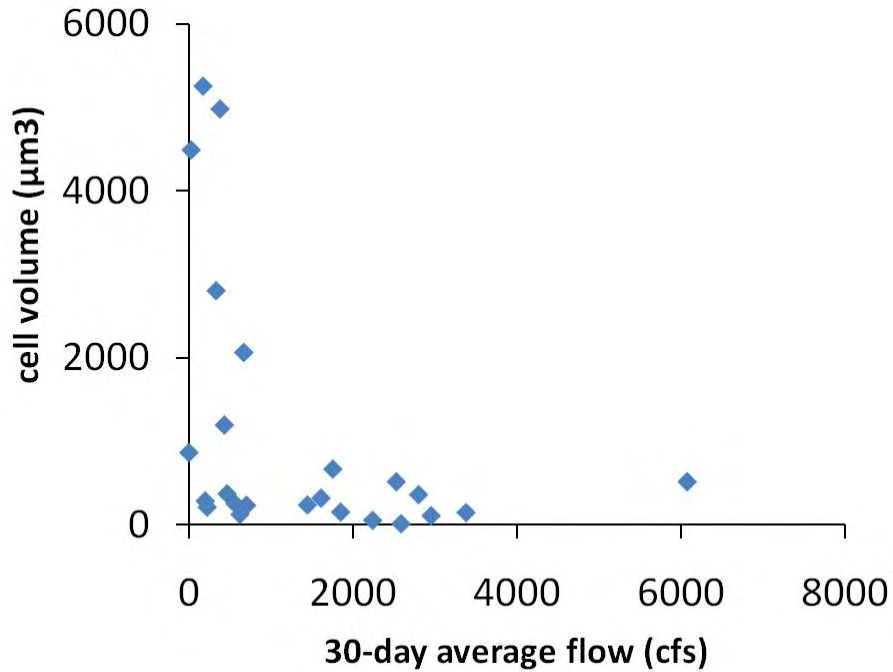


Figure 3.4.3. Mean phytoplankton cell volume (μm^3) versus mean freshwater inflow through S-79 for previous 30 days.

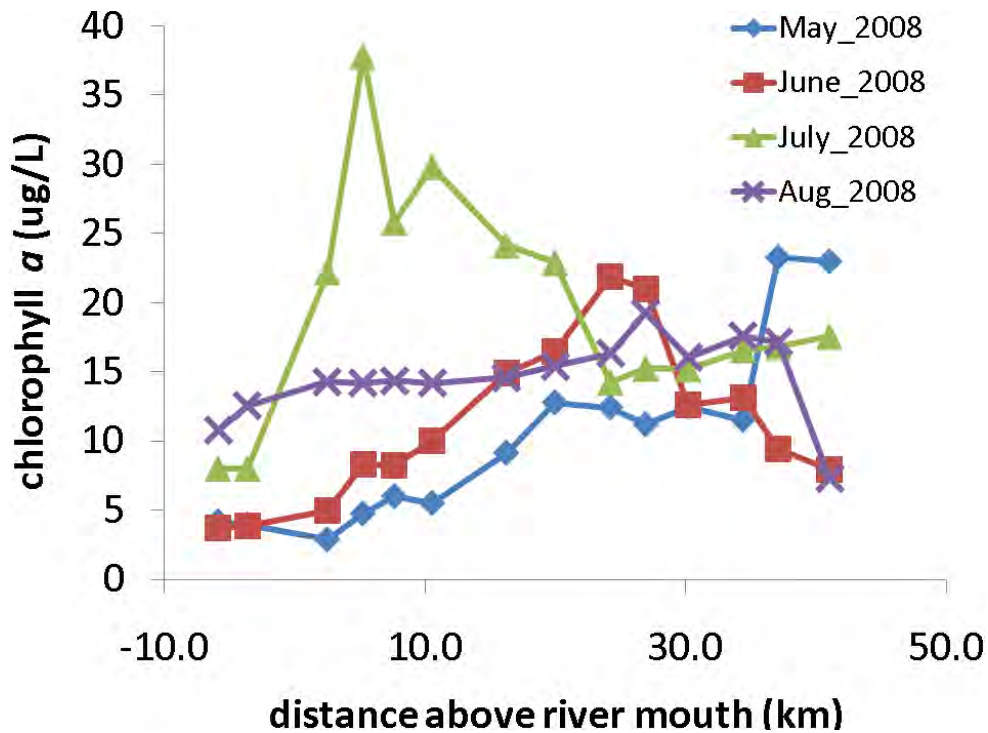


Figure 3.4.4. Chlorophyll *a* concentration versus distance upstream from the river mouth, May through August 2008.

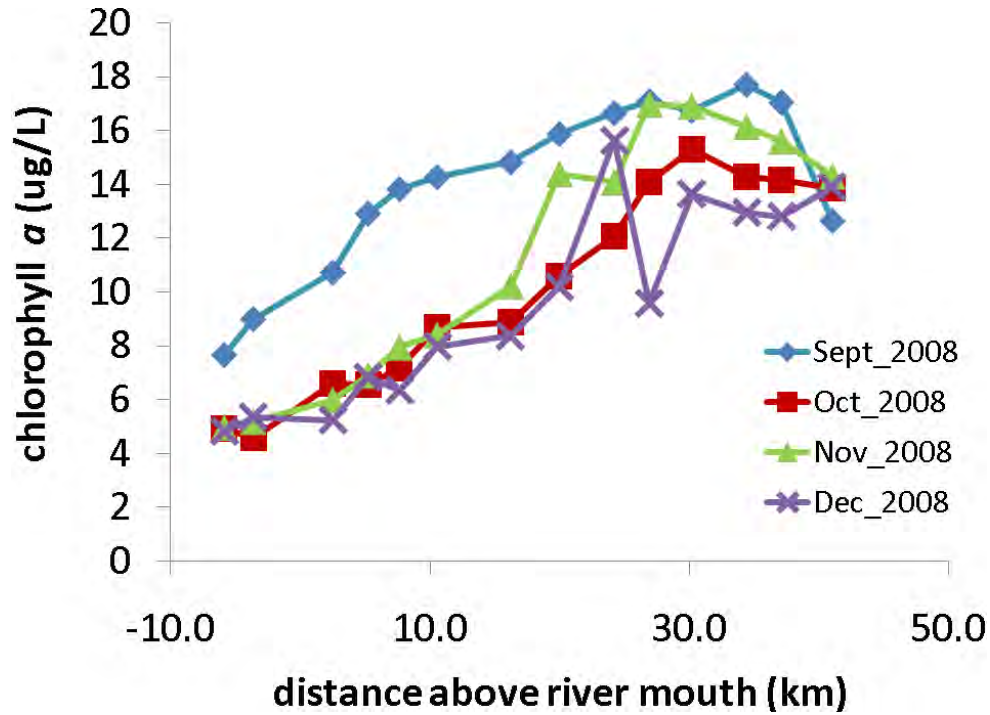


Figure 3.4.5. Chlorophyll *a* concentration versus distance upstream from the river mouth, September through December 2008.

Phytoplankton did not build up biomass over the course of the wet season (Fig. 3.4.5) and decreased downstream from September through December 2008. Between January and May 2009, chlorophyll concentrations increased in the upper zones (Figs. 3.4.6-3.4.7: stations 5U and 6D), but chlorophyll again decreased with the onset of the wet season in June, being reduced downstream through August 2009 (Fig. 3.4.7). This trend continued into September 2009 (Fig. 3.4.8), followed by the observed chlorophyll maximum moving up the estuary at the end of 2009 (Fig. 3.4.8) and back down the estuary in the first four months of 2010 (Fig. 3.4.9).

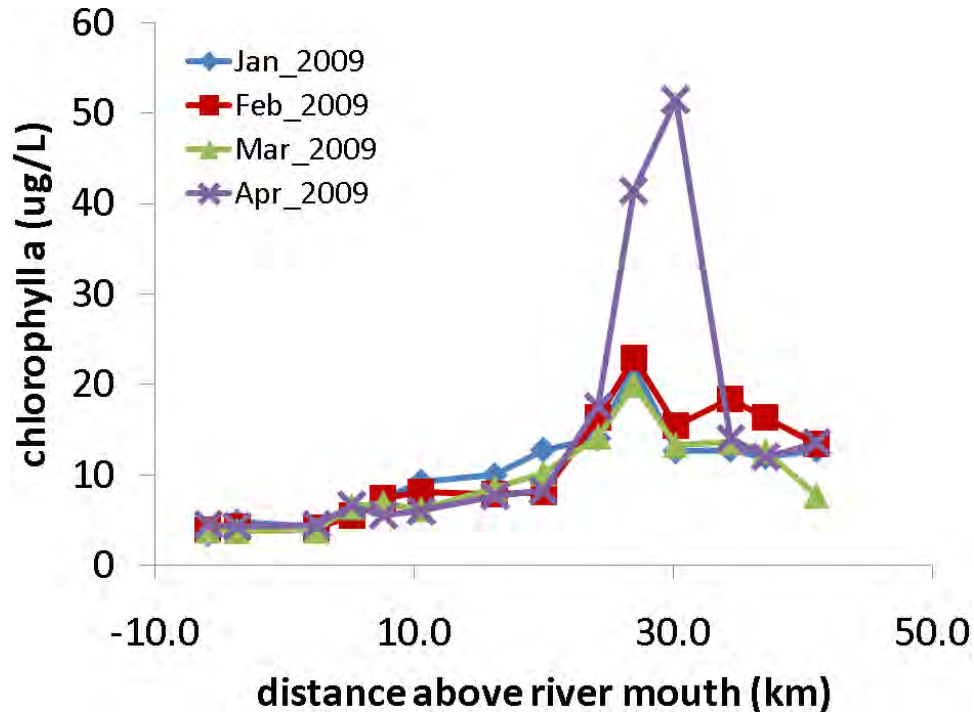


Figure 3.4.6. Chlorophyll *a* concentration versus distance upstream from river mouth, January through April 2009.

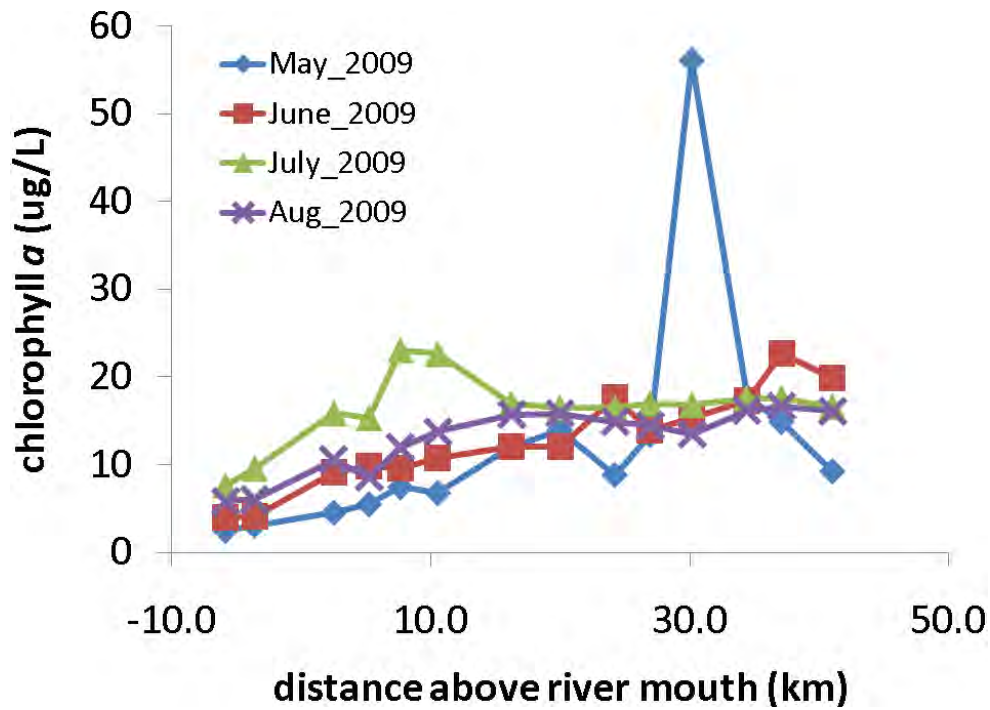


Figure 3.4.7. Chlorophyll *a* concentration versus distance upstream from river mouth, May through August 2009.

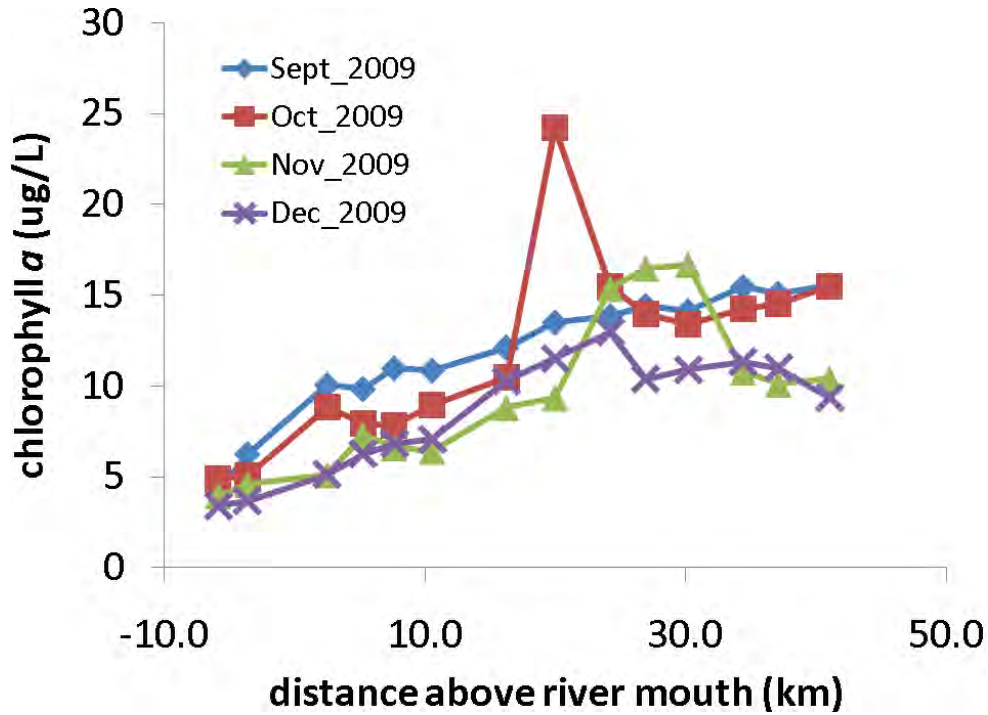


Figure 3.4.8. Chlorophyll *a* concentration versus distance upstream from river mouth, September through December 2009.

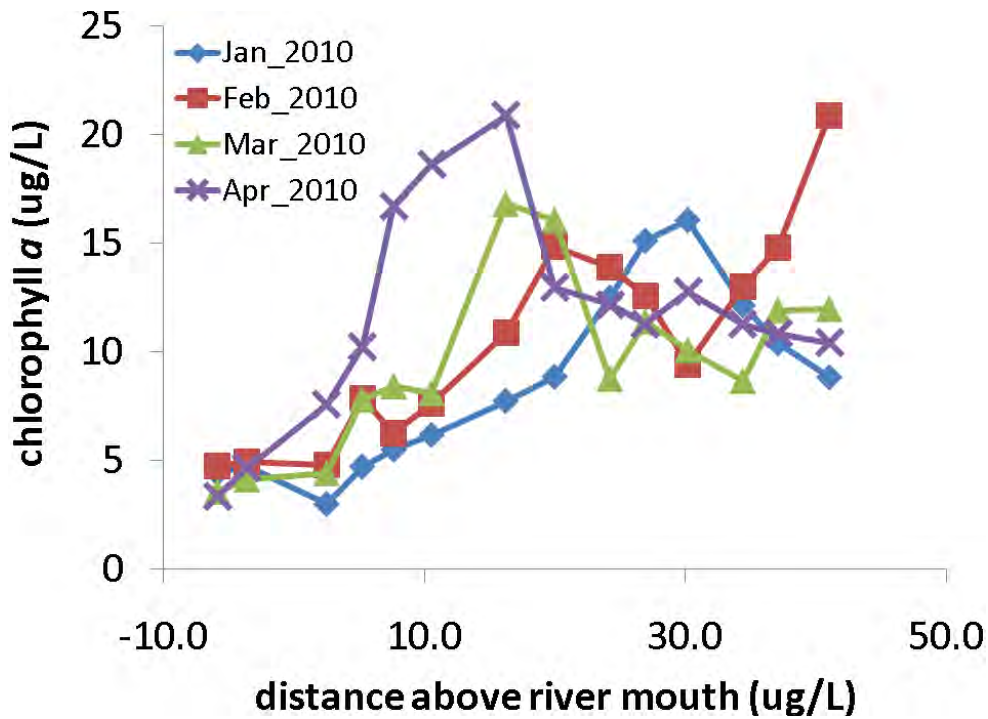


Figure 3.4.9. Chlorophyll *a* concentration versus distance upstream from river mouth, January through April 2010.

The above dynamics likely result from changing discharges through S-79. Chlorophyll *a* concentrations tended to increase as flow increased in zones 1-3, with peaks seen at flows of approximately 1,500 cfs in zones 2 and 3, but fell off above 3,500 cfs, likely indicating flushing (Fig. 3.4.10). Chlorophyll *a* responses were much different in the upper zones (4-7), where the highest concentrations were seen at low flows (<1,000 cfs) in zones 5 and 6, with no apparent response to increasing flow otherwise (indicating flushing; Fig. 3.4.11).

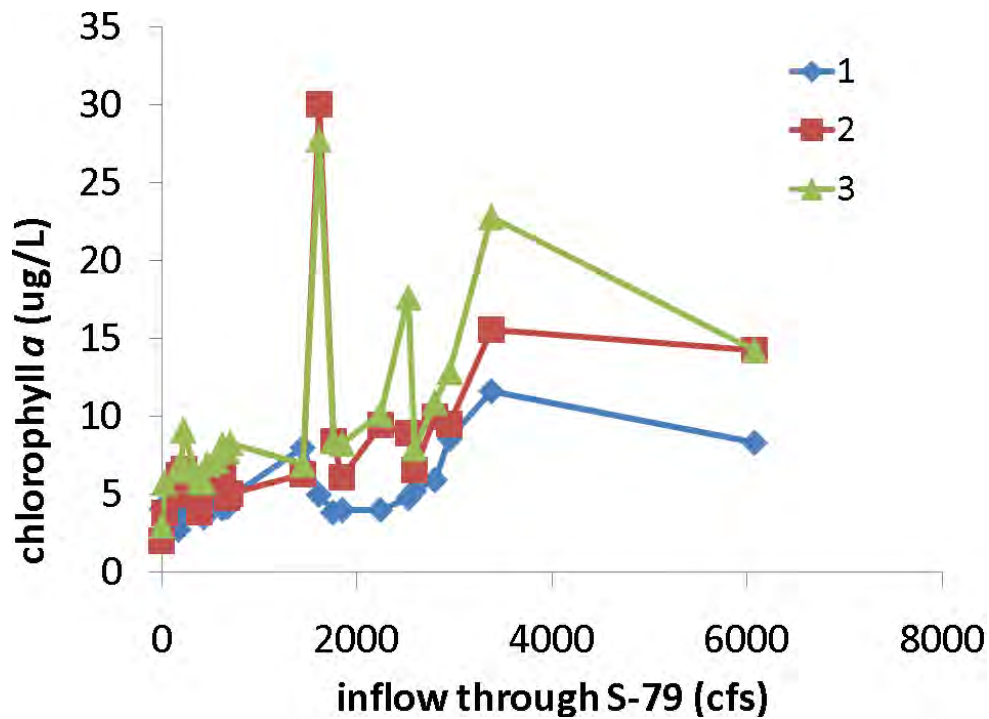


Figure 3.4.10. Chlorophyll *a* concentration versus freshwater inflow through S-79, downstream stations 1-3.

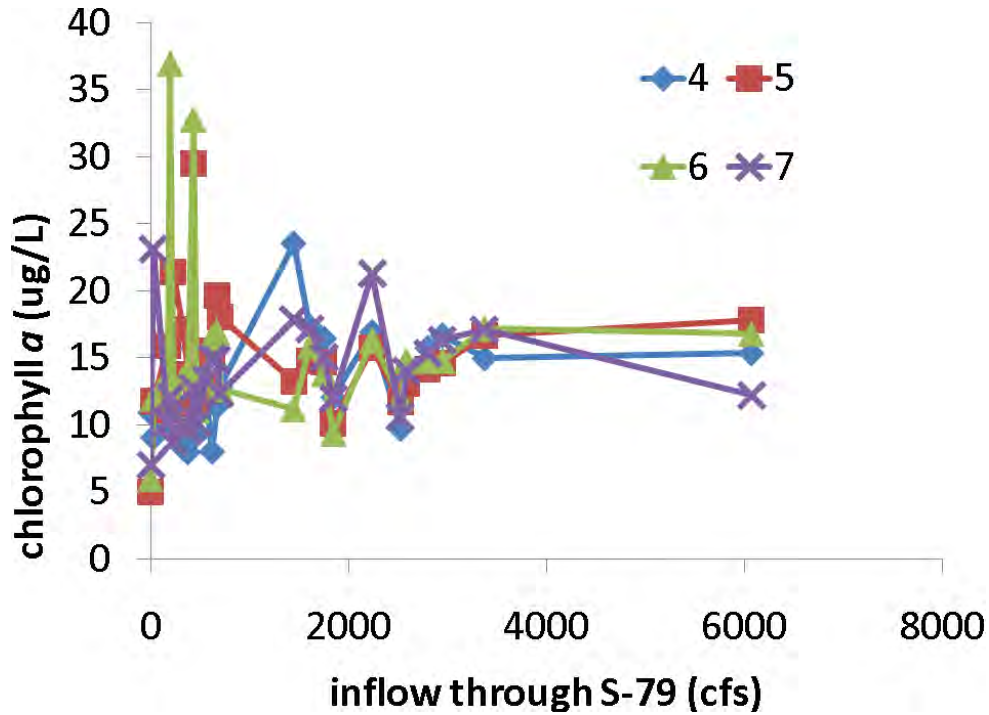


Figure 3.4.11. Chlorophyll *a* concentration versus freshwater inflow through S-79, upstream stations 4-7.

All zones (except zone 1) exhibited similar correlations between observed salinity and river flow (Fig. 3.4.12), with the highest correlations between salinity and river flows averaged over the previous 10–25 d. In terms of chlorophyll, however, zones 2–4 had similar correlations with river flow averaged over the previous 15–25 days, whereas the upper zones (5–7) exhibited weak correlations, and zone 1 has the best correlation with flows <10 days (Fig. 3.4.13).

Phytoplankton clearly responded to changing inflow: low flow conditions (<1000 cfs) generally corresponded to larger cells, predominately diatoms, and as flows increased the assemblage shifted towards smaller cells, predominately cyanobacteria. The shift is likely in response to reduced salinities and lower residence times associated with higher inflows, favoring faster growing forms such as cyanobacteria. Both of these scenarios present benefits and concerns. Diatoms, more dominant in low flow conditions, are considered beneficial nourishment for grazers (Officer and Ryther 1980): cell sizes tend to be larger when diatoms dominate, leading to shorter food chains and greater energy transfer to fishes (Lalli and Parsons 1997). However, many diatoms (including *Skeletonema* sp.) are “sinkers,” settling on the bottom and resulting in low dissolved oxygen levels as bacteria breakdown the diatom biomass (Dortch

et al. 1992). Doering et al. (2006) reported a negative relationship between water column chlorophyll *a* concentration and bottom dissolved oxygen concentration, possibly demonstrating this sinking phenomenon. Conversely, cyanobacteria are smaller and many contain gas vacuoles that inhibit sinking. Though cyanobacteria may not cause low dissolved oxygen levels in bottom waters, they are not considered to be a high quality food source for grazers (Martin-Creuzburg and Von Elert 2009). Therefore, a shift to cyanobacteria may reduce the quality of the (planktonic) food source for grazers, impacting higher trophic levels.

Although phytoplankton are responding to changing flow, the upper zones (e.g., 5 and 6) are more susceptible to phytoplankton biomass build-up when flows are low (<1,000 cfs), probably due to increased residence time that allows for longer incubations. Lower zones (1-3) are more susceptible to higher chlorophyll concentrations as flow increases, but only to a point. Flows >3,500 cfs result in phytoplankton being flushed out (probably more quickly than their generation times of <1–4 days), whereas flows of ~1,500 cfs may cause chlorophyll maxima in zones 2 and 3. Doering et al. (2006) noted similar inflection points in the relationship between chlorophyll and flow.

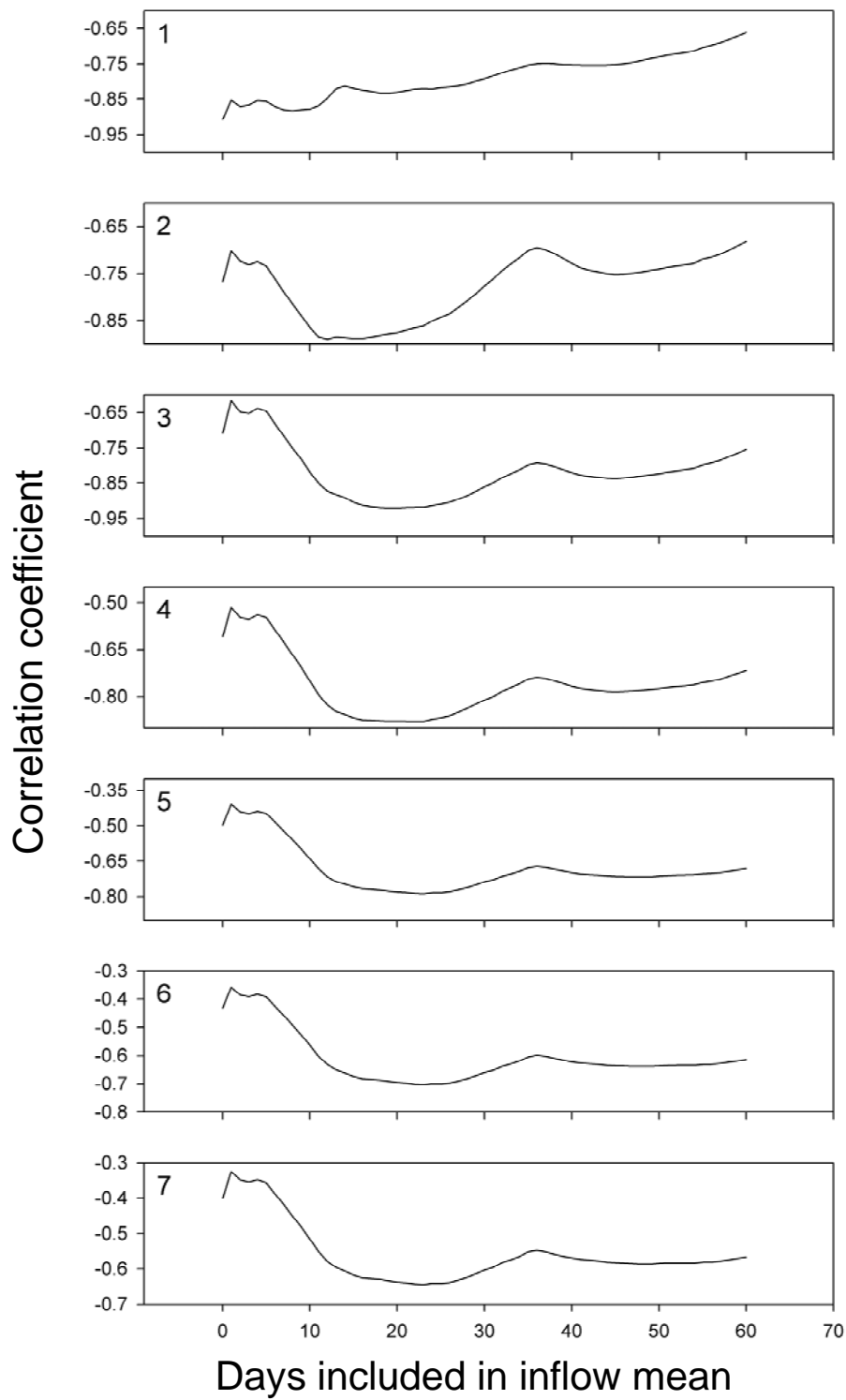


Figure 3.4.12. Correlation coefficients for relationships between salinity and lagged freshwater inflow for each sampling zone. Zones are identified in the upper left corner of each panel.

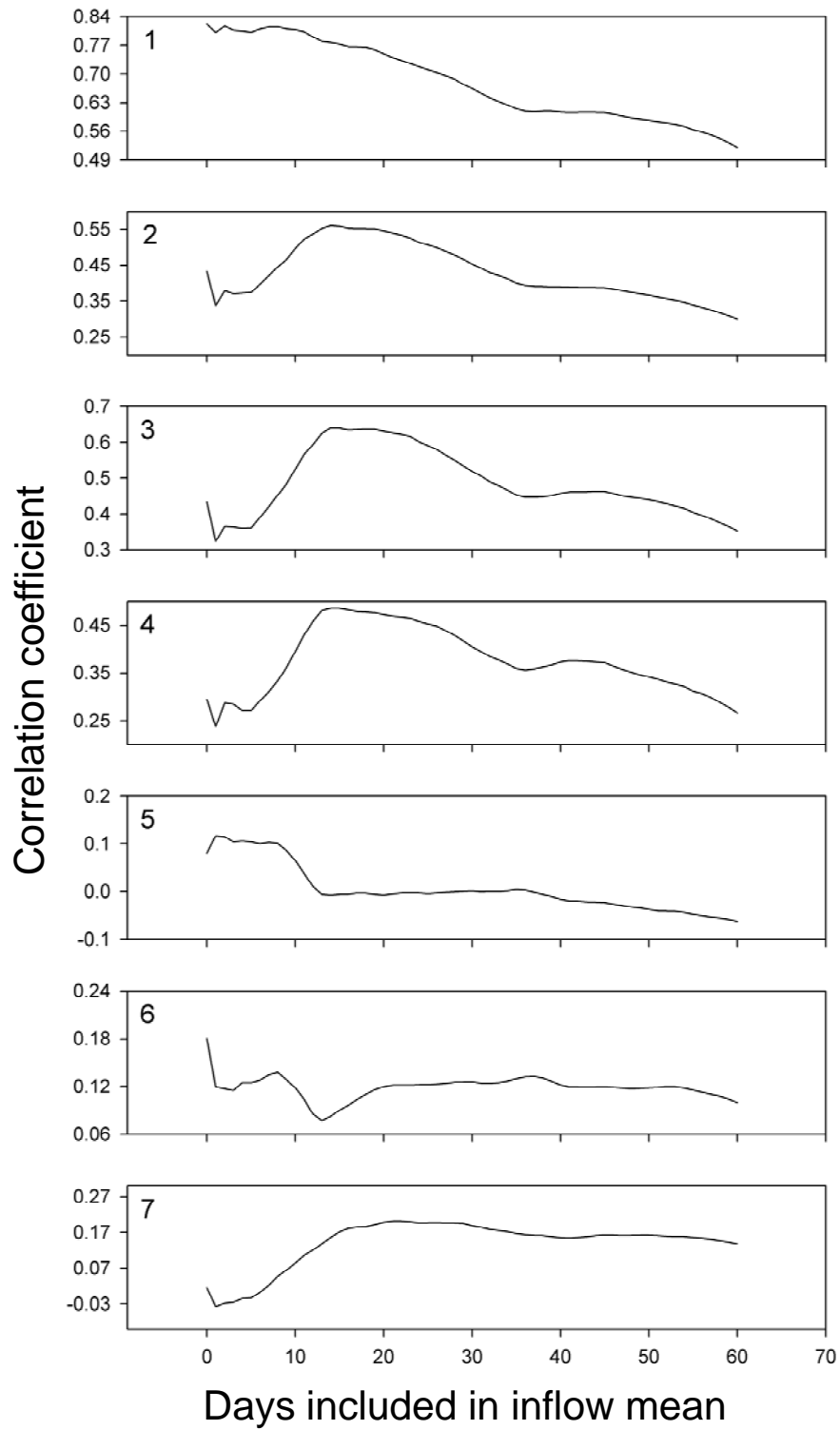


Figure 3.4.13. Correlation coefficients for relationships between chlorophyll *a* and lagged freshwater inflow for each sampling zone. Zones are identified in the upper left corner of each panel.

3.5 Zooplankton Catch Composition

A total of 3,278,566 individuals representing 321 zooplankton taxa and an estimated minimum of 208 species were identified from samples collected May 2008–April 2010. Zooplankton samples were dominated by decapod crustaceans, representing 51.6% of the total catch (Appendix D). Other major groups in order of abundance were amphipods (13.6%: primarily gammarideans), copepods (8.1%), fishes (6.4%), cumaceans (6.2%), and cladocerans (4.9%). Less abundant groups of interest represented a minor component of the total catch and included gelatinous predators (3.2%), mysids (2.4%), and the larvacean *Oikopleura dioica* (1.7%). Together all these groups comprised 98.2% of the total zooplankton catch.

Among the decapods, unidentified zoeae, mysis stage larvae, and megalopae comprised 93.6% of the total. Important decapods identified that were collected in numbers greater than 10,000 included the mud shrimp *Upogebia* spp., the sergestid *Lucifer faxoni*, snapping shrimp of the family Alpheidae, zostera shrimp *Hippolyte zostera*, and hermit crabs. A total of 49 decapod taxa were identified from the samples, representing a minimum of 25 species.

Ichthyoplankton were dominated by the eggs of both unidentified percomorph fishes (47.4%) and bay anchovies *Anchoa mitchilli* (33.4%), and by larvae of anchovies (11.0%) and gobies (3.2%) (Appendix D). Anchovies were dominated by the bay anchovy *Anchoa mitchilli*, gobies of the genus *Gobiosoma* were found in greater numbers than those of the genus *Microgobius*, and the Florida blenny *Chasmodes saburrae* was the most abundant identifiable blenny. Fishes were relatively diverse, with a total of 137 fish taxa identified from the samples, representing a minimum of 57 species.

Among other taxonomic groups of interest, the copepods were represented by 26 species, with *Acartia tonsa*, *Labidocera aestiva*, and *Mesocyclops edax* dominating in order of abundance; isopods were represented by 15 species, though only *Edotia triloba* could be considered abundant; and mysids were represented by 11 species and were dominated numerically by juvenile *Americamysis* spp. and by adult *Americamysis almyra* and *Bowmaniella brasiliensis*; and (Appendix D).

3.6 Zooplankton Seasonal and Spatial Trends

In general, total zooplankton densities peaked spring/early summer in both sampling years, with a secondary peak in October 2009 (Fig. 3.6.1; Appendix E). Throughout the sampling periods, decapod zoeae and mysis stage shrimps were the dominant species associated with many of the peaks in zooplankton abundance; however, there were some notable exceptions (Figs. 3.6.2-3.6.5). Gammaridean amphipods were the dominant taxa associated with zooplankton peaks in August 2008 and in October and December 2009; cumaceans were associated with a peak downstream (rkm 16.2) in March 2009; *Anchoa mitchilli* eggs were associated with one of many peaks present in April 2009; the larvacean *Oikopleura dioica* was associated with a peak in abundance at rkm 20.0 in May 2009; and *Daphnia* spp. comprised the dominant taxon present at peaks upstream in July 2009 (Appendix F).

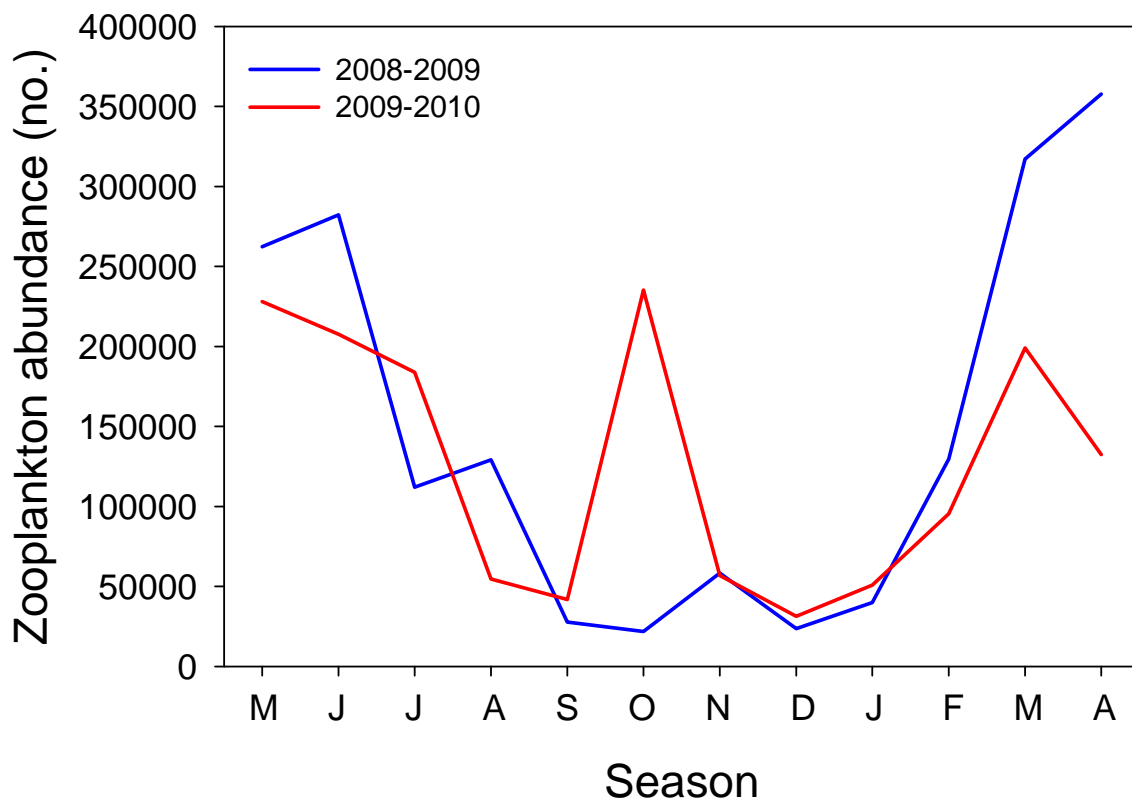


Figure 3.6.1. Seasonal variation in zooplankton abundance collected at all stations.

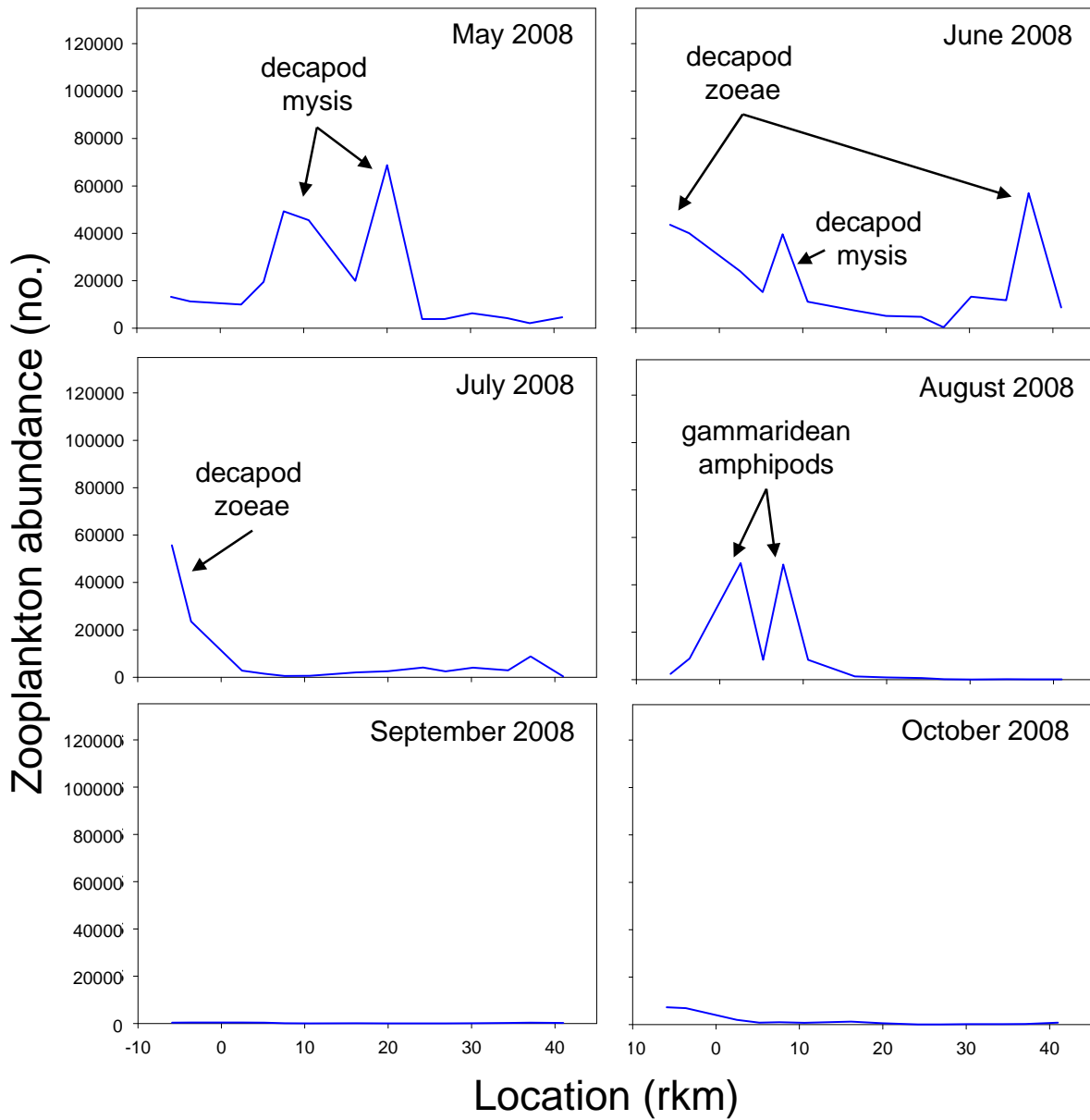


Figure 3.6.2. Monthly variation in zooplankton abundance along the study area, May–October 2008. Arrows identify dominant species associated with peaks in zooplankton abundance.

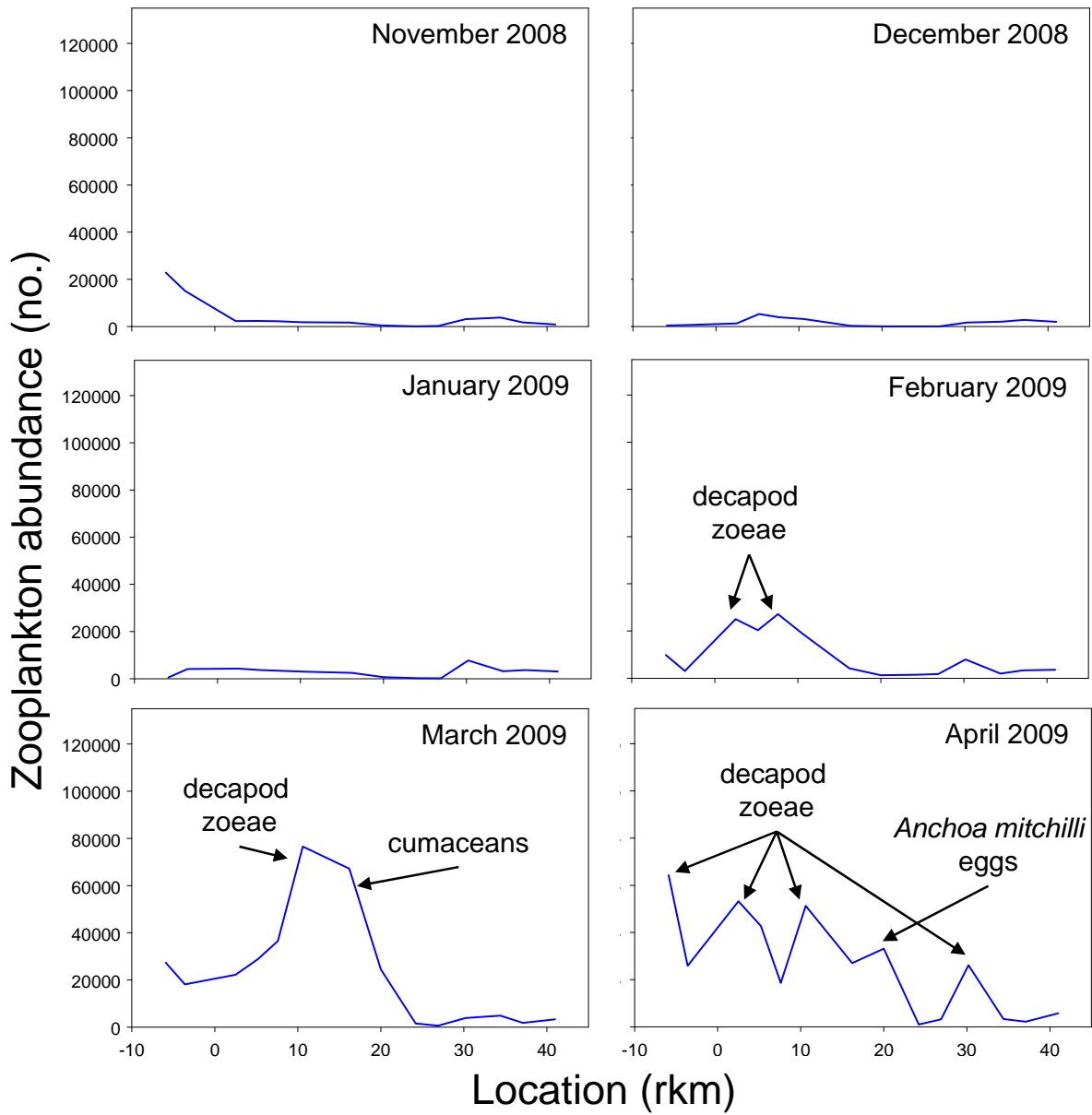


Figure 3.6.3. Monthly variation in zooplankton abundance along the study area, November 2008–April 2009. Arrows identify dominant species associated with peaks in zooplankton abundance.

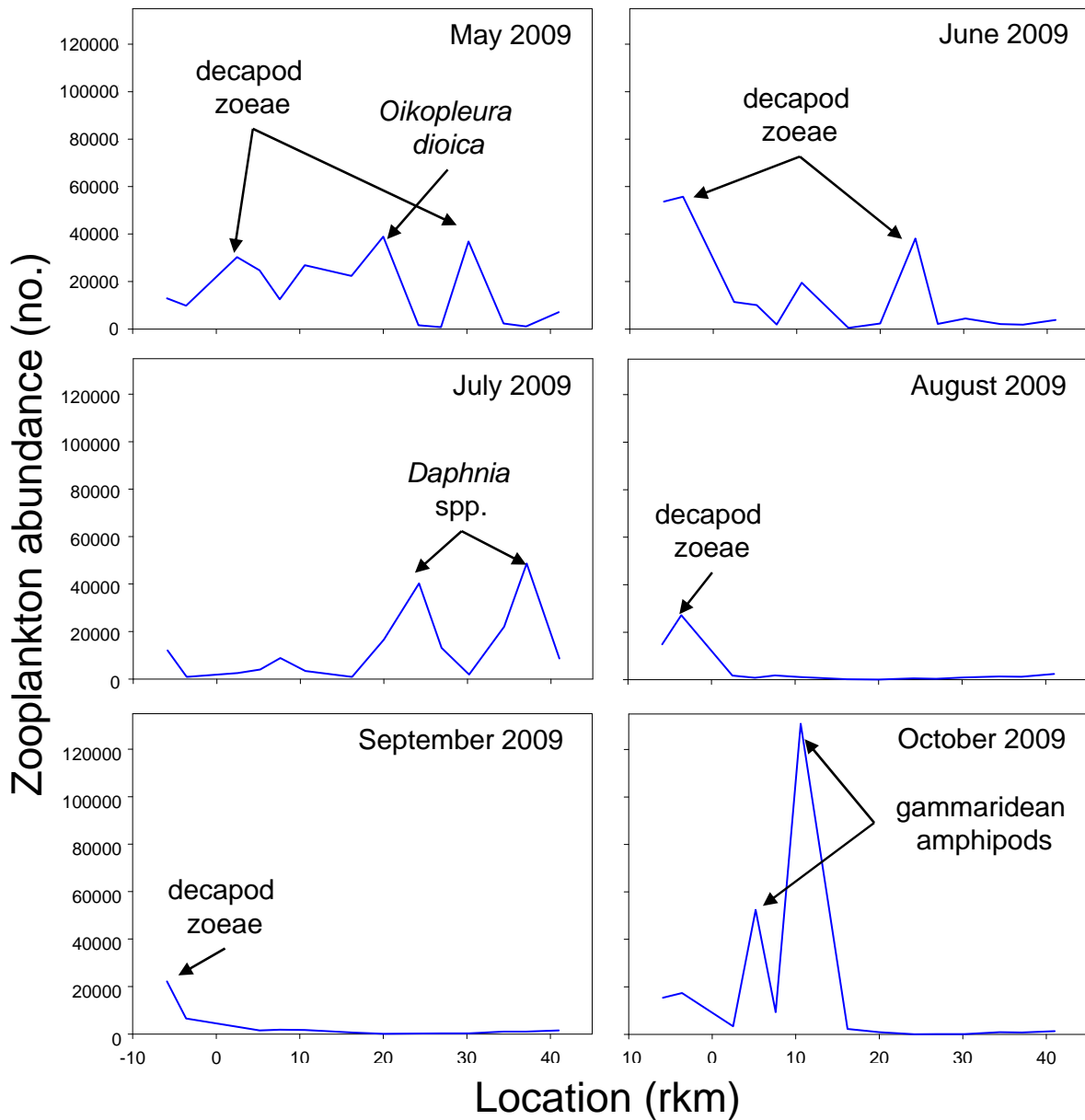


Figure 3.6.4. Monthly variation in zooplankton abundance along the study area, May–October 2009. Arrows identify dominant species associated with peaks in zooplankton abundance.

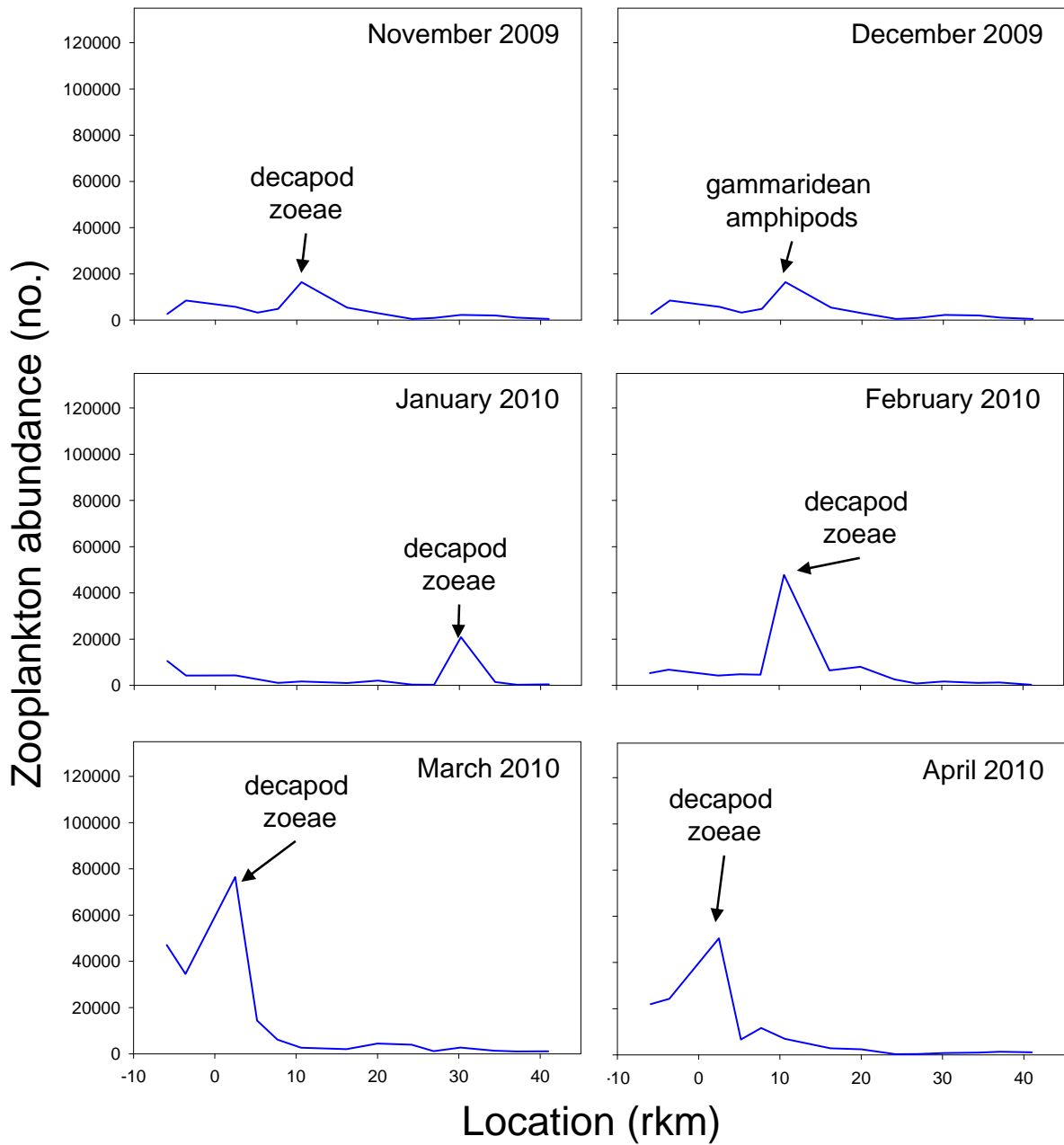


Figure 3.6.5. Monthly variation in zooplankton abundance along the study area, November 2009–April 2010. Arrows identify dominant species associated with peaks in zooplankton abundance.

A number of taxa exhibited marked seasonal changes in their centers of abundance, responding to reduced flows by moving upstream and relocating downstream during periods of higher inflow (Figs. 3.6.6–3.6.7). For example, in May 2008 (dry) the copepod *Acartia tonsa*, the chaetognaths *Sagitta* spp., and larval gobies were located well inside the river, but during September 2008 (wet) were likely most abundant downstream of the study area. During the dry season, the hyperbenthic animals *Edotia triloba* and *Americamysis almyra*, along with juvenile bay anchovies, were located well upstream above Beautiful Island, where the river narrows considerably; during the wet season, the former species were located well downstream but still inside the river proper and the latter was likely most abundant downstream of the study area.

Taxa richness, an estimate of biodiversity, and the estimated minimum number of species present also suggested that the system was more diverse in spring/early summer, with peak taxa richness occurring in May–July 2008 and March–June 2009 (Figs. 3.6.8–3.6.9). A second peak in taxa richness was also present in November 2008. In general, biodiversity was greater downstream, with a broad region of increased taxa richness (>40 taxa) present below rkm 20.0 in May 2008 and below 16.2 in April 2009 (Figs. 3.6.10–3.6.11). With the onset of the wet season, peak taxa richness was relocated downstream toward the mouth of the river or beyond (Figs. 3.6.10–3.6.13). It is important not to oversell the relative importance of these trends in biodiversity, as the largest portion of the zooplankton catch (47.7%) was identified only as larval decapods, and it is not known how much biodiversity this taxon actually comprised.

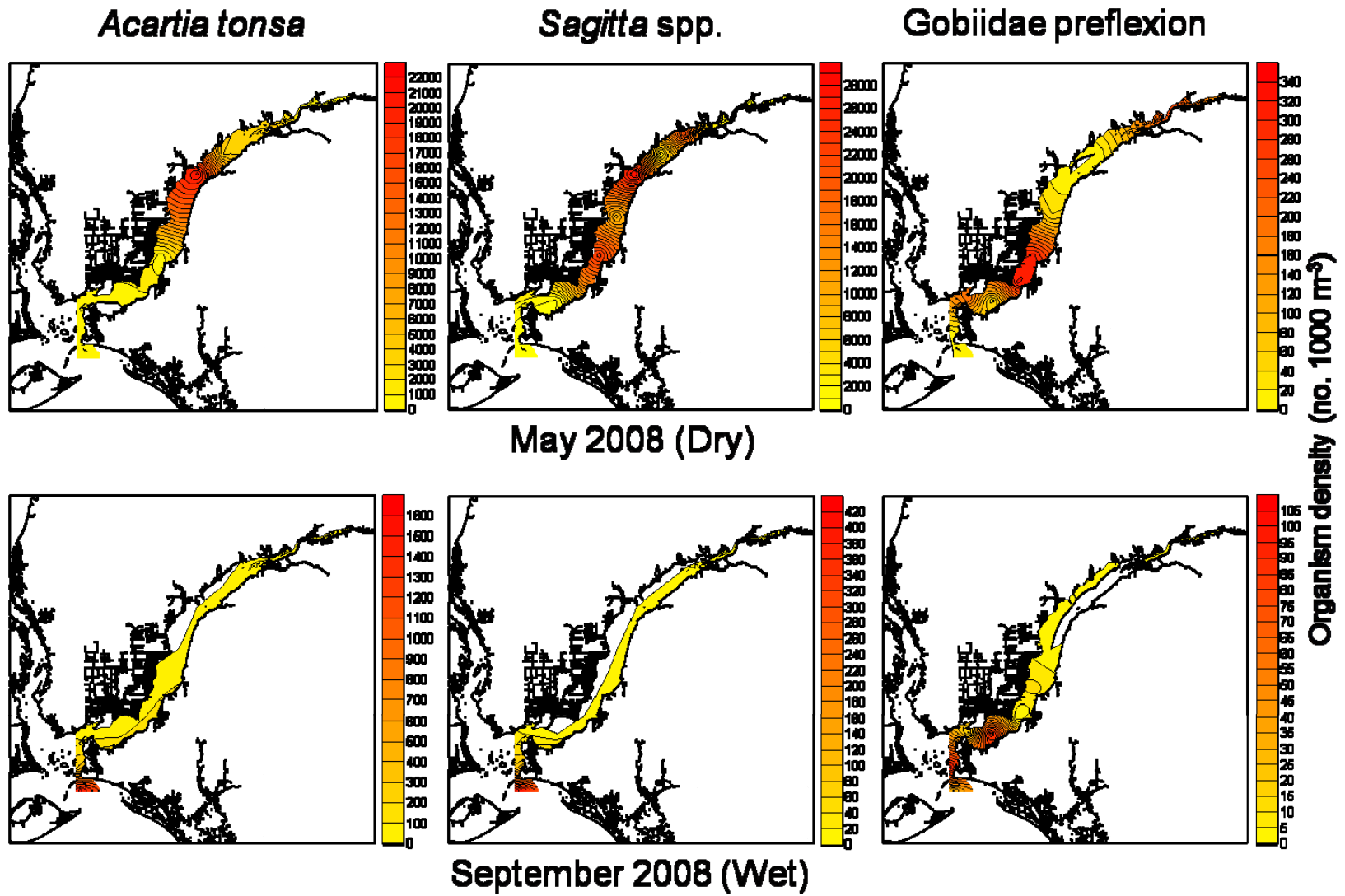


Figure 3.6.6. Seasonal changes (dry vs. wet) in the distribution of select zooplankton species.

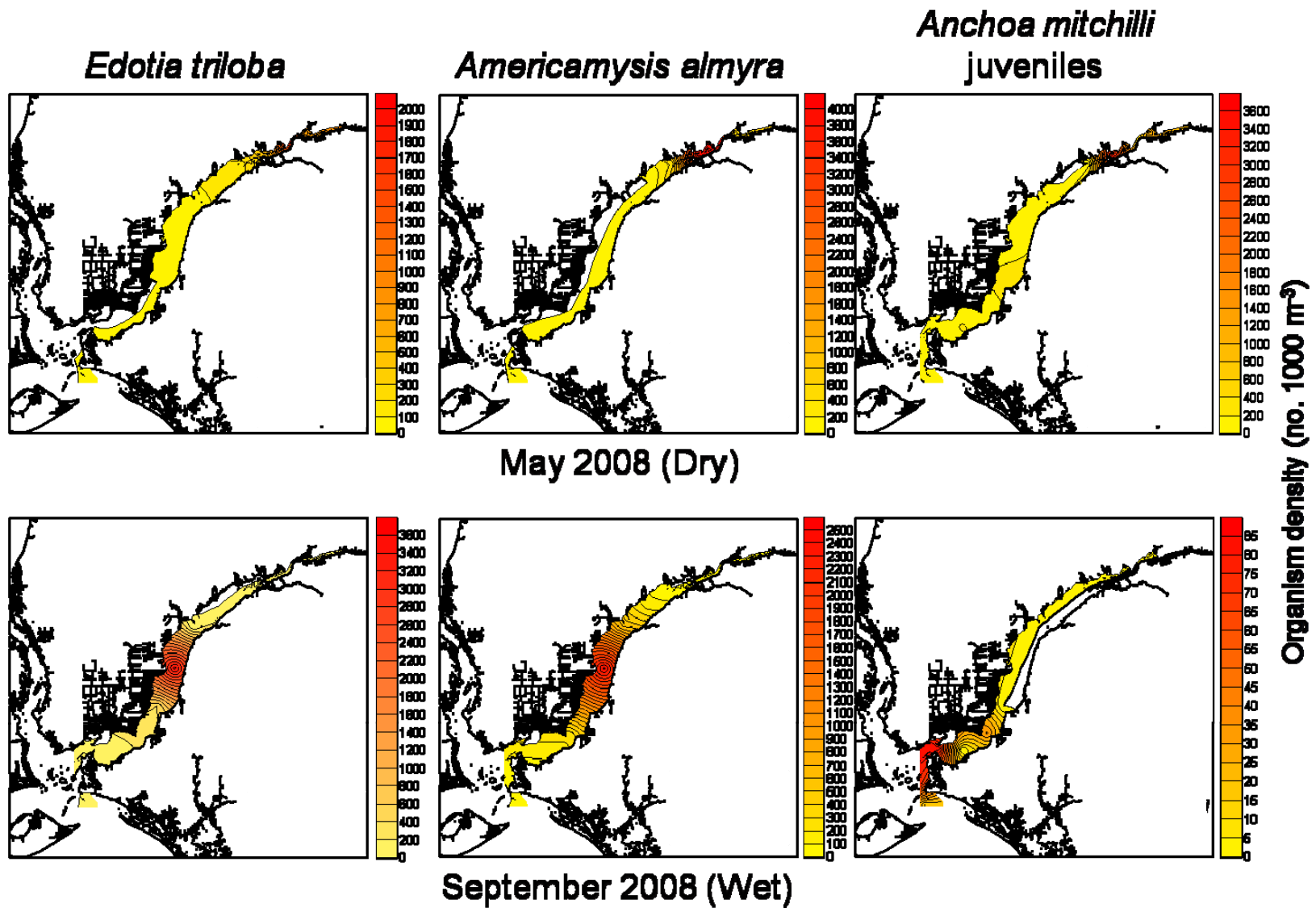


Figure 3.6.7. Seasonal changes (dry vs. wet) in the distribution of select hyperbenthos and juvenile bay anchovy.

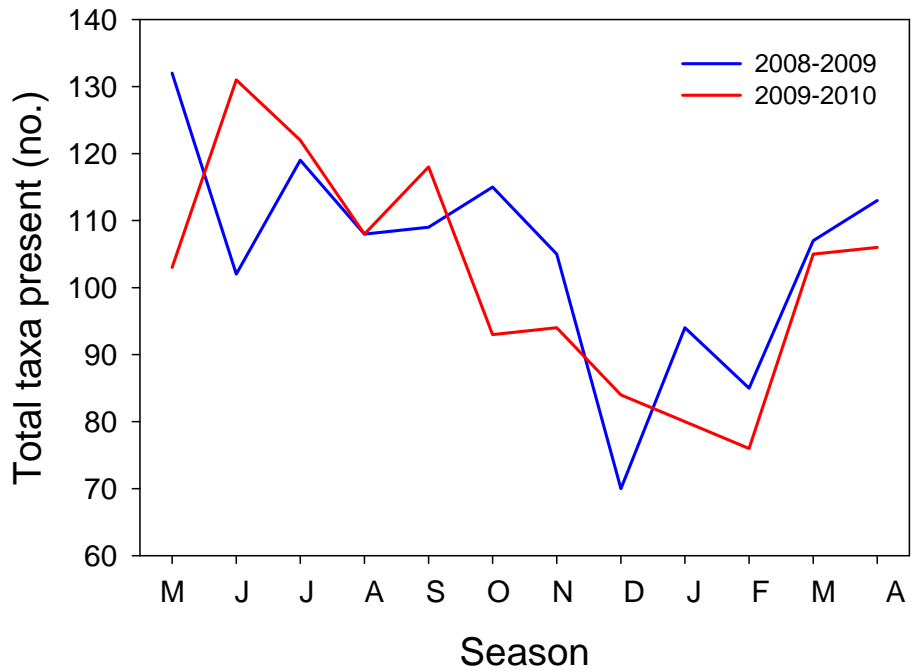


Figure 3.6.8. Seasonal variation in taxa richness for the Caloosahatchee River and Estuary (all stations combined).

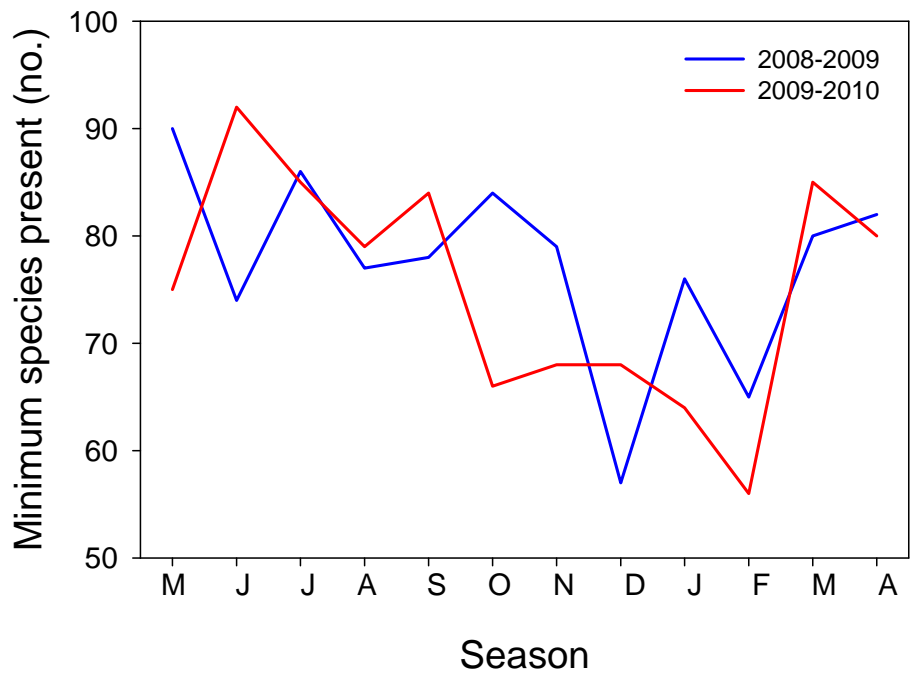


Figure 3.6.9. Seasonal variation in the minimum number of species present for the Caloosahatchee River and Estuary (all stations combined).

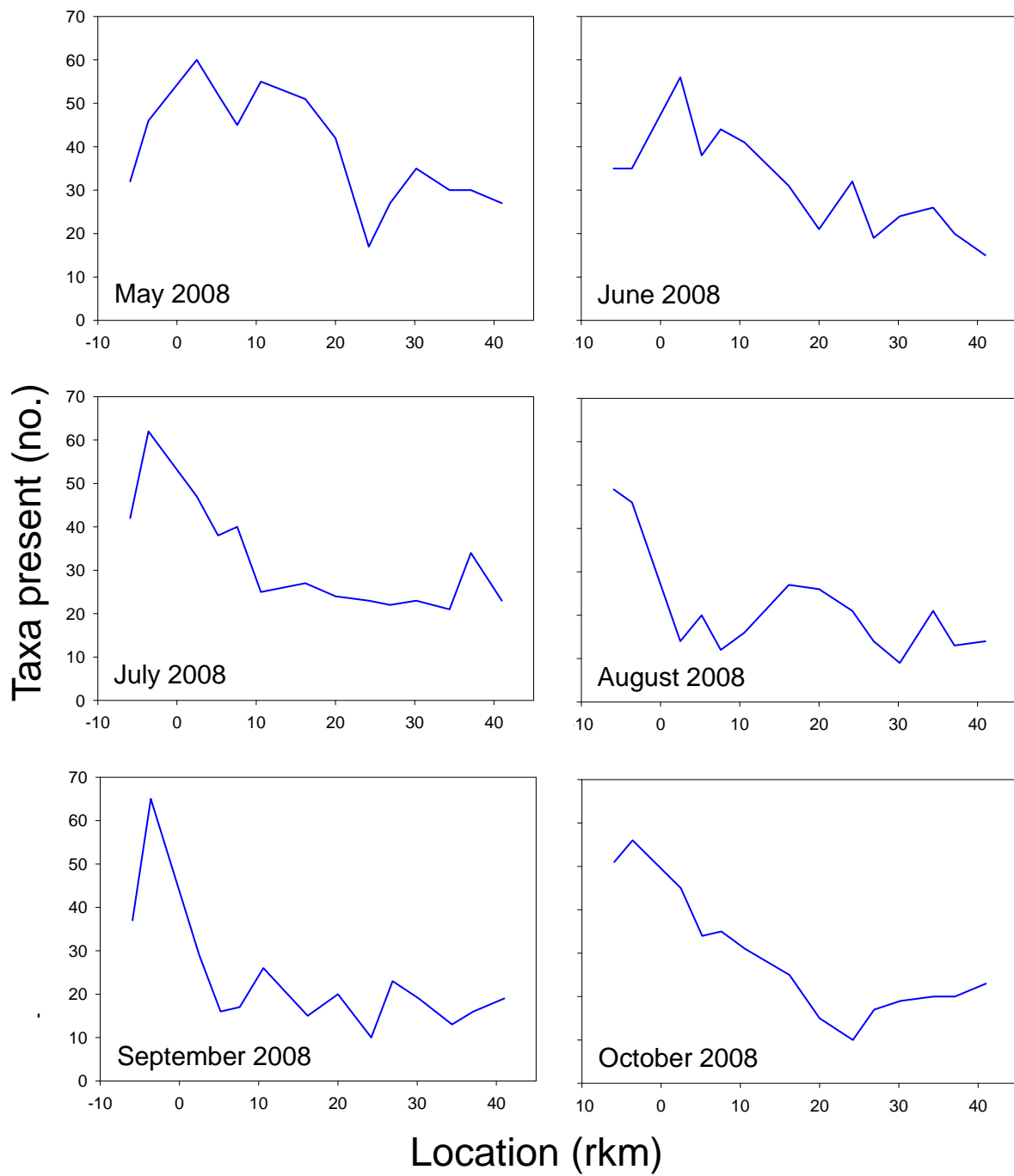


Figure 3.6.10. Monthly variation in taxa richness along the study area, May–October 2008.

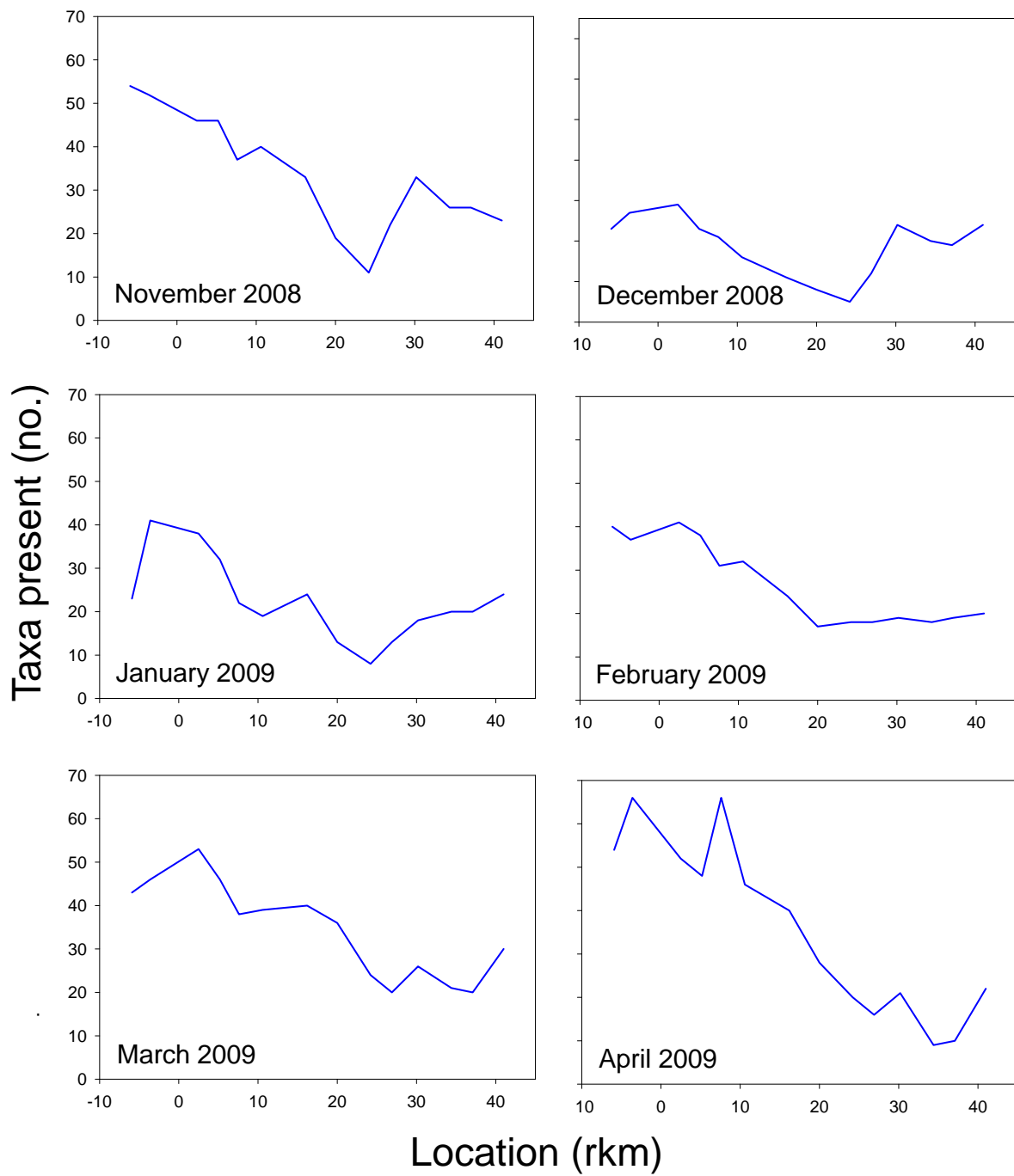


Figure 3.6.11. Monthly variation in taxa richness along the study area, November 2008–April 2009.

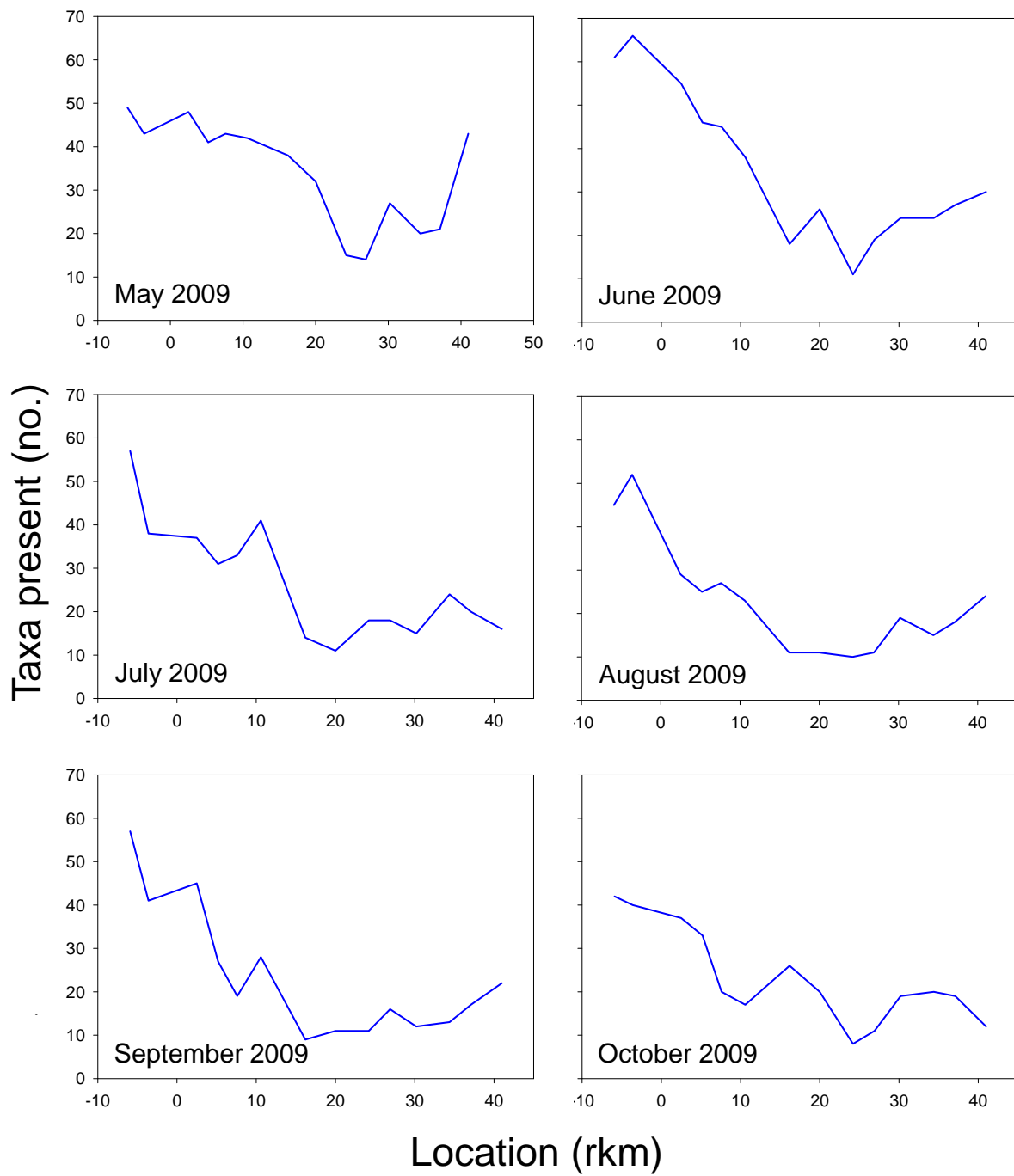


Figure 3.6.12. Monthly variation in taxa richness along the study area, May–October 2009.

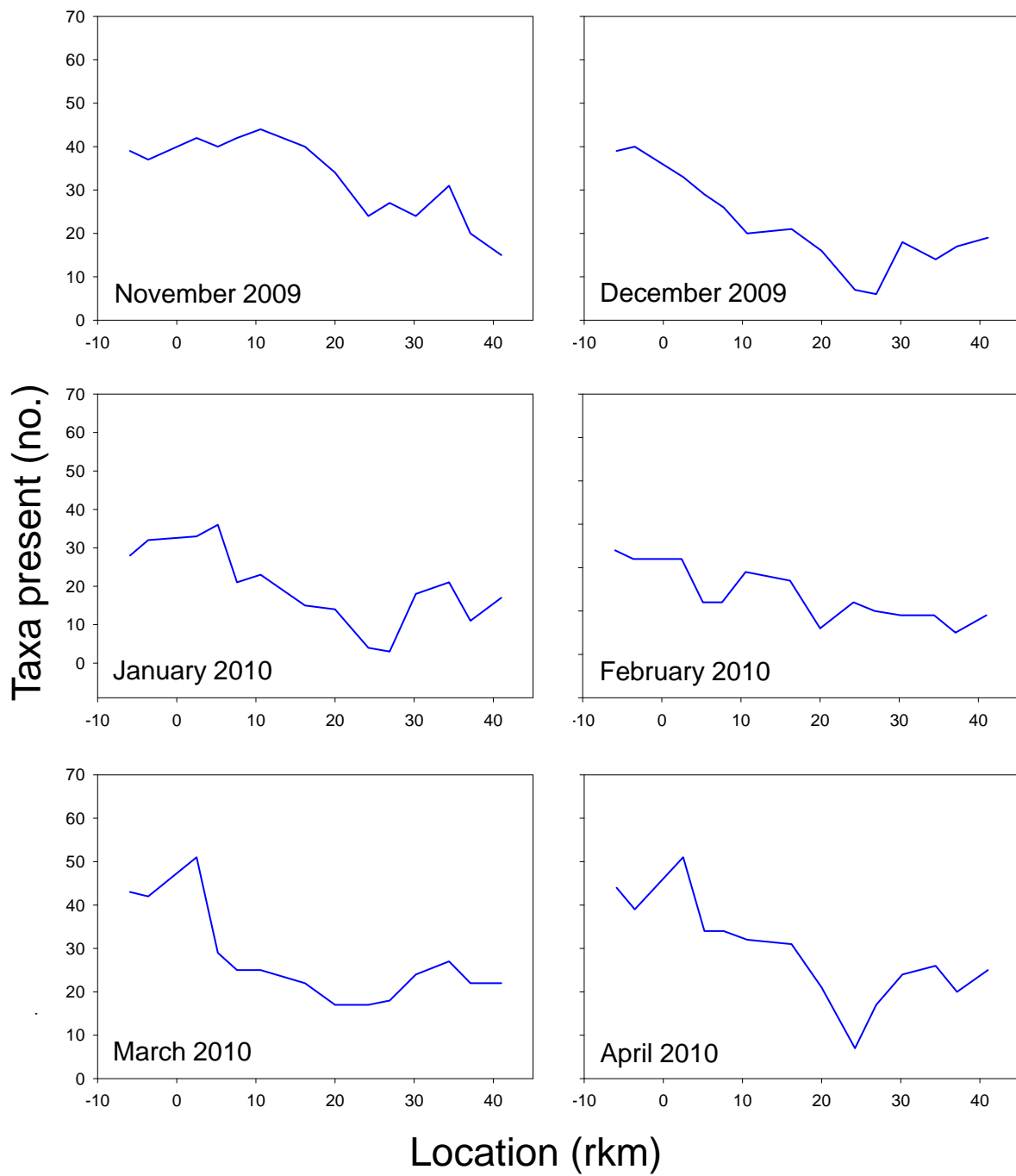


Figure 3.6.13. Monthly variation in taxa richness along the study area, November 2009–April 2010.

3.7 Freshwater Inflow Relationships

3.7.1 Centers of Abundance

A number of taxa demonstrated significant relationships between center of abundance and freshwater inflow (natural log transformed) calculated at time lags (i.e., number of consecutive days prior to sampling used to calculate mean inflow) of 1-120 days (Table 3.7.1.1). All, save one, were negative responses, with animals moving upstream as inflows decreased. Upstream movement is the typical response to reduced inflows seen in riverine estuaries on Florida's west coast (Peebles 2002, MacDonald et al. 2005, Greenwood et al. 2006). Time lags associated with the responses were variable, but most taxa (92%) responded to inflow on a scale of <2 mo, and 32% of the responses corresponded to lags of 6-8 wk. In contrast, the tanaidacean *Hargeria rapax* exhibited a position relationship between center of abundance and freshwater inflow.

An examination of plots comparing organism center of abundance and non-transformed freshwater inflow (Appendix G) indicates that the centers of distribution for a number of estuarine taxa were located immediately downstream of S-79. These taxa all had centers of distribution at >35 rkm following periods of low inflow, typically at the end of the dry season: preflexion larval silversides *Menidia* spp.; postlarval gobies *Microgobius* spp. and hogchokers *Trinectes maculatus*; juveniles pipefish *Syngnathus louisianae*; juvenile *Americamysis* spp. and adult mysids *Americamysis almyra*; and isopods *Edotia triloba* (see Fig. 3.7.1.1 for example).

Of particular interest are the inflows required to remove predatory hydromedusae (*Clytia* sp.) and ctenophores (*Mnemiopsis leidyi*) from this region of the Caloosahatchee. Such inflows appear to be near the low end of the observed inflow range (Fig. 3.7.1.2, Appendix H). These gelatinous zooplankton are known to be voracious predators/competitors of larval fishes.

In addition to these taxa-specific responses to freshwater inflow, overall diversity, estimated using taxa richness, was negatively related to freshwater inflow at a lag of 24 d, though the relationship was not strong (Fig. 3.7.1.3).

Table 3.7.1.1. Response of center of abundance (km_U) to freshwater inflow using natural-log transformed inflow values for inflow data recorded at S-79. Regression statistics are sample size (n), intercept (a), slope (b), significance (p), and coefficient of determination r^2 as %. The Durbin-Watson statistic (DW) was used to identify possible serial correlation (x indicates $p < 0.05$). D is the number of consecutive daily inflow values used to calculate mean inflow. For taxa exhibiting relationships within inflow at multiple values of D , data are presented for the regression with the lowest p value.

| Taxon | n | a | b | p | r^2 | DW | D |
|---|-----|-------|--------|---------|-------|------|-----|
| <i>Clytia</i> sp. | 8 | 96.03 | -12.78 | 0.0002 | 79 | | 50 |
| <i>Microgobius</i> spp. postflexion larvae | 17 | 79.30 | -8.81 | 0.0033 | 63 | | 55 |
| <i>Lironeca</i> sp. (isopod) | 24 | 69.99 | -7.84 | <0.0001 | 56 | | 51 |
| <i>Gobiosoma</i> spp. postflexion larvae | 20 | 62.12 | -7.22 | 0.0022 | 41 | | 57 |
| <i>Bowmaniella brasiliensis</i> | 24 | 65.56 | -7.20 | <0.0001 | 64 | x | 50 |
| <i>Pseudodiaptomus pelagicus</i> | 22 | 55.30 | -6.90 | <0.0001 | 69 | | 57 |
| <i>Menidia</i> spp. preflexion larvae | 17 | 75.09 | -6.81 | 0.0005 | 57 | | 67 |
| <i>Edotia triloba</i> (isopod) | 24 | 65.25 | -6.33 | <0.0001 | 65 | x | 51 |
| Gobiidae preflexion larvae | 24 | 51.73 | -5.94 | 0.0004 | 44 | x | 59 |
| <i>Americamysis almyra</i> | 24 | 63.51 | -5.35 | 0.0001 | 49 | x | 51 |
| <i>Americamysis</i> spp. juveniles | 24 | 59.76 | -5.22 | 0.0002 | 48 | x | 50 |
| <i>Menticirrhus</i> spp. preflexion larvae | 15 | 34.70 | -4.85 | 0.0003 | 64 | | 58 |
| <i>Oithona</i> spp. | 20 | 29.97 | -4.49 | <0.0001 | 62 | | 46 |
| <i>Acartia tonsa</i> | 24 | 30.65 | -4.42 | <0.0001 | 72 | | 45 |
| Gobiidae flexion larvae | 23 | 39.45 | -4.31 | 0.0309 | 20 | x | 66 |
| Caligoid copepods | 24 | 29.69 | -4.20 | <0.0001 | 64 | | 52 |
| <i>Oikopleura dioica</i> | 23 | 25.66 | -4.06 | <0.0001 | 63 | x | 44 |
| Cumaceans | 24 | 37.13 | -4.04 | <0.0001 | 66 | | 48 |
| <i>Syngnathus louisianae</i> juveniles | 12 | 37.21 | -3.96 | 0.0075 | 53 | | 1 |
| <i>Mesocyclops edax</i> | 19 | 64.23 | -3.88 | 0.0001 | 63 | | 4 |
| <i>Sagitta</i> spp. | 24 | 25.41 | -3.80 | <0.0001 | 70 | | 45 |
| <i>Anchoa</i> spp. preflexion larvae | 22 | 27.15 | -3.62 | <0.0001 | 70 | | 16 |
| Gerreidae preflexion larvae | 13 | 24.66 | -3.60 | <0.0001 | 86 | | 49 |
| <i>Microgobius</i> spp. flexion larvae | 18 | 43.00 | -3.59 | 0.0314 | 26 | | 80 |
| <i>Anchoa</i> spp. flexion larvae | 16 | 30.84 | -3.33 | <0.0001 | 71 | | 21 |
| Caprellid amphipods | 18 | 23.19 | -3.33 | 0.0010 | 50 | | 74 |
| <i>Diaptomus</i> spp. | 14 | 58.74 | -3.28 | 0.0062 | 48 | | 4 |
| Alpheidae postlarvae | 21 | 19.64 | -2.97 | <0.0001 | 66 | | 46 |
| <i>Trinectes maculatus</i> postflexion larvae | 12 | 33.48 | -2.87 | 0.0118 | 49 | | 2 |
| Paguridae juveniles | 15 | 14.74 | -2.55 | 0.0006 | 61 | | 26 |
| <i>Hippolyte zostericola</i> juveniles | 18 | 15.90 | -2.53 | <0.0001 | 67 | | 43 |
| <i>Mnemiopsis leidy</i> | 8 | 35.53 | -2.53 | 0.0006 | 88 | | 1 |
| Paracalanid copepods | 15 | 15.90 | -2.50 | 0.0033 | 50 | | 21 |
| <i>Argulus</i> sp. (branchiuran) | 16 | 42.19 | -2.39 | 0.0197 | 33 | | 7 |
| Gammaridean amphipods | 24 | 28.76 | -2.37 | 0.0393 | 18 | x | 92 |
| <i>Labidocera aestiva</i> | 24 | 13.18 | -2.31 | <0.0001 | 74 | x | 43 |

Table 3.7.1.1. Continued

| Taxon | <i>n</i> | <i>a</i> | <i>b</i> | <i>p</i> | <i>r</i> ² | DW | <i>D</i> |
|--|----------|----------|----------|----------|-----------------------|----|----------|
| <i>Temora turbinata</i> | 15 | 16.57 | -2.29 | 0.0140 | 38 | | 22 |
| <i>Palaemonetes</i> spp. postlarvae | 24 | 20.51 | -2.29 | 0.0015 | 37 | | 3 |
| <i>Bathygobius soporator</i> preflexion | 14 | 18.26 | -2.26 | 0.0038 | 52 | | 14 |
| <i>Chasmodes saburrae</i> flexion larvae | 13 | 18.88 | -2.13 | 0.0063 | 51 | | 24 |
| <i>Anchoa mitchilli</i> postlarvae | 17 | 20.86 | -2.09 | 0.0001 | 67 | | 2 |
| <i>Monstrilla</i> sp. | 18 | 14.02 | -2.09 | 0.0023 | 45 | | 51 |
| <i>Eutima</i> sp. | 17 | 10.28 | -1.97 | <0.0001 | 79 | | 39 |
| <i>Americamysis bahia</i> | 20 | 17.36 | -1.92 | <0.0001 | 65 | | 7 |
| <i>Upogebia</i> spp. juvenile | 10 | 13.59 | -1.87 | 0.0007 | 78 | | 8 |
| <i>Anchoa mitchilli</i> adult | 24 | 40.01 | -1.82 | 0.0496 | 16 | | 17 |
| <i>Upogebia</i> spp. postlarvae | 20 | 9.86 | -1.73 | <0.0001 | 71 | x | 12 |
| Medusa sp. E | 12 | 8.44 | -1.72 | 0.0062 | 54 | | 41 |
| <i>Chaoborus punctipennis</i> | 22 | 47.76 | -1.56 | 0.0153 | 26 | | 7 |
| Paguridae postlarvae | 20 | 7.61 | -1.40 | 0.0003 | 53 | | 14 |
| <i>Parasterope pollex</i> | 20 | 10.98 | -1.38 | 0.0491 | 20 | | 50 |
| Penaeidae metamorphs | 14 | 11.70 | -1.38 | 0.0483 | 29 | x | 8 |
| <i>Tozeuma carolinense</i> postlarvae | 16 | 5.90 | -1.16 | 0.0023 | 50 | | 5 |
| Processidae postlarvae | 22 | 4.48 | -1.11 | <0.0001 | 58 | | 2 |
| <i>Hippolyte zostericola</i> postlarvae | 24 | 4.97 | -1.03 | 0.0001 | 50 | x | 2 |
| <i>Erichsonella attenuata</i> | 21 | 12.45 | -0.96 | 0.0262 | 23 | | 37 |
| <i>Cynoscion nebulosus</i> preflexion larvae | 11 | 3.03 | -0.95 | 0.0004 | 77 | | 2 |
| <i>Lucifer faxoni</i> adults | 23 | 2.40 | -0.84 | 0.0001 | 51 | x | 14 |
| <i>Obelia</i> spp. | 12 | 0.82 | -0.82 | 0.0072 | 53 | | 1 |
| <i>Centropages velificatus</i> | 12 | -2.92 | -0.26 | 0.0132 | 30 | | 3 |
| <i>Hargeria rapax</i> | 21 | -2.58 | 1.24 | 0.0280 | 23 | | 52 |

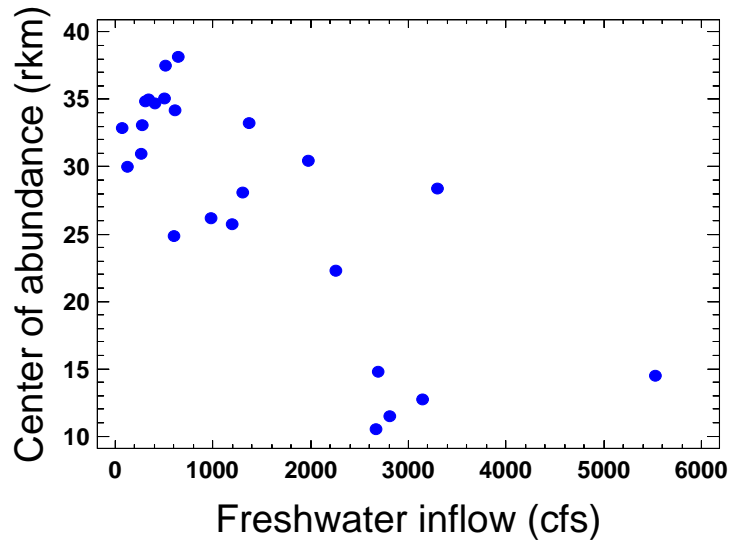


Figure 3.7.1.1. Center of abundance for the mysid *Americamysis almyra* in relation to freshwater inflow.

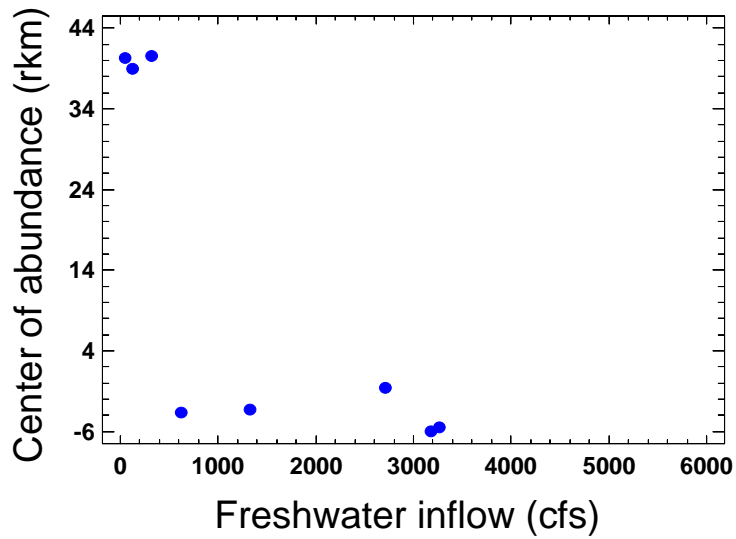


Figure 3.7.1.2. Center of abundance for the gelatinous predator *Clytia* sp. in relation to freshwater inflow.

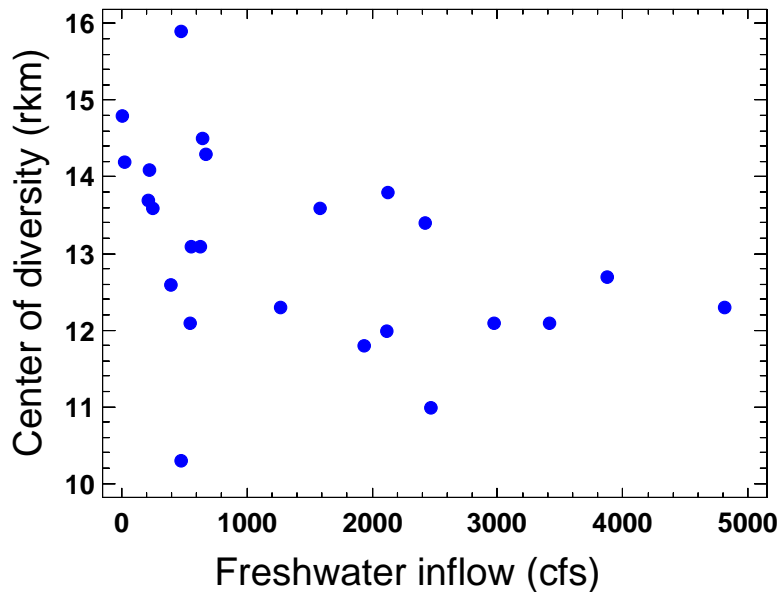


Figure 3.7.1.3. Response of density-weighted center of diversity, calculated using taxon richness, to freshwater inflow ($r = -0.49$, $p = 0.0158$, $n = 24$: natural log transformed inflow data).

3.7.2 Total Organism Abundance and Inflow

A number of taxa also demonstrated significant relationships between total organism abundance (natural log transformed) estimated for the entire study area and freshwater inflow (natural log transformed) calculated at time lags (i.e., number of consecutive days prior to sampling used to calculate mean inflow) of 1-120 days (Table 3.7.2.1; Appendix). Of these, the majority (72%) were negative responses, with fewer numbers of animals associated with greater levels of freshwater inflow. These results suggest that species associated with these taxa either experienced reduced spawning or downstream movement through the study area and out into the Gulf of Mexico in response to high levels of inflow. Plots of non-transformed abundance-inflow data exposed potential break points in this relationship that were much less apparent in plots that used natural log transformed data (Figs. 3.7.2.1-3.7.2.2; Appendix I). An examination of these break points in the data suggests that reduced abundances were often associated with inflows greater than 1,000 cfs (Appendix E).

Table 3.7.2.1. Response of total organism abundance to freshwater inflow using natural-log transformed abundance and inflow values (recorded at S-79). Regression statistics are sample size (n), intercept (a), slope (b), significance (p), and coefficient of determination r^2 as %. The Durbin-Watson statistic (DW) was used to identify possible serial correlation (x indicates $p < 0.05$). D is the number of consecutive daily inflow values used to calculate mean inflow. For taxa exhibiting relationships within inflow at multiple values of D , data are presented for the regression with the lowest p value. Asterisk denotes those cases where a lognormal model applied to non-transformed data provided a better fit.

| Taxon | n | a | b | p | r^2 | DW | D |
|--|-----|-------|-------|---------|-------|------|-----|
| <i>Palaemon floridanus</i> | 10 | 30.81 | -2.05 | 0.0028 | 69 | | 93 |
| <i>Upogebia</i> sp. postlarvae* | 20 | 27.41 | -1.46 | 0.0111 | 31 | | 120 |
| <i>Anchoa mitchilli</i> postflexion larvae | 17 | 23.60 | -1.16 | 0.0010 | 52 | | 120 |
| <i>Clytia</i> sp.* | 8 | 23.48 | -1.15 | 0.0191 | 63 | | 120 |
| Alpheidae postlarvae | 21 | 24.86 | -1.14 | 0.0082 | 31 | | 120 |
| Gerreidae preflexion larvae | 13 | 22.80 | -1.10 | 0.0160 | 42 | | 119 |
| Cumaceans* | 24 | 26.91 | -1.05 | 0.0033 | 33 | x | 80 |
| <i>Palaemonetes</i> spp. postlarvae | 24 | 22.72 | -1.05 | 0.0042 | 32 | x | 120 |
| <i>Parasterope pollex</i> * | 20 | 22.67 | -1.05 | 0.0001 | 59 | | 67 |
| <i>Anchoa</i> spp. flexion larvae* | 16 | 21.08 | -0.94 | 0.0167 | 34 | x | 120 |
| Caprellid amphipods | 18 | 20.95 | -0.91 | 0.0023 | 45 | | 120 |
| <i>Gobiosoma</i> spp. postflexion larvae* | 20 | 20.81 | -0.86 | 0.0377 | 22 | | 120 |
| <i>Eusarsiella zostericola</i> | 20 | 19.70 | -0.81 | 0.0013 | 44 | | 81 |
| Paracalanid copepods* | 15 | 19.72 | -0.76 | 0.0286 | 32 | x | 104 |
| <i>Harrieta faxoni</i> | 13 | 18.33 | -0.74 | 0.0183 | 41 | | 120 |
| <i>Chasmodes saburrae</i> postflexion larvae | 12 | 19.00 | -0.72 | 0.0196 | 44 | | 57 |
| <i>Erichsoniella attenuata</i> | 21 | 17.30 | -0.55 | 0.0120 | 29 | x | 120 |
| <i>Upogebia</i> sp. juveniles | 10 | 17.29 | -0.47 | 0.0205 | 51 | | 43 |
| <i>Syngnathus scovelli</i> juveniles | 19 | 16.67 | -0.43 | 0.0380 | 23 | | 99 |
| <i>Temora turbinata</i> | 15 | 16.62 | -0.37 | 0.0132 | 39 | | 1 |
| <i>Centropages velificatus</i> | 20 | 17.61 | -0.35 | 0.0042 | 37 | | 1 |
| <i>Sagitta</i> spp. | 24 | 21.58 | -0.34 | 0.0015 | 38 | | 1 |
| <i>Anchoa</i> spp. preflexion larvae | 22 | 19.06 | -0.32 | 0.0448 | 19 | x | 1 |
| Harpacticoid copepods | 17 | 15.65 | -0.30 | 0.0308 | 27 | x | 1 |
| <i>Acartia tonsa</i> | 24 | 21.89 | -0.26 | 0.0204 | 22 | | 1 |
| <i>Pseudodiaptomus coronatus</i> | 22 | 17.09 | -0.24 | 0.0334 | 21 | | 2 |
| Blenniidae preflexion larvae | 24 | 16.79 | -0.19 | 0.0497 | 16 | x | 1 |
| Caligoid copepods | 24 | 14.78 | -0.18 | 0.0123 | 25 | | 1 |
| <i>Anchoa mitchilli</i> adults | 24 | 12.43 | 0.16 | 0.0375 | 18 | | 2 |
| <i>Cynoscion nebulosus</i> preflexion larvae | 12 | 13.18 | 0.21 | 0.0452 | 34 | x | 1 |
| <i>Americamysis bahia</i> * | 20 | 12.99 | 0.28 | 0.0382 | 22 | | 4 |
| Penaeidae metamorphs | 14 | 13.48 | 0.35 | <0.0001 | 77 | | 2 |
| <i>Argulus</i> sp. (branchiuran) | 16 | 8.72 | 0.44 | 0.0211 | 32 | | 51 |

| | | | | | | | |
|--|----|-------|------|--------|----|---|----|
| <i>Penilia avirostris</i> * | 17 | 14.32 | 0.46 | 0.0258 | 29 | x | 5 |
| <i>Cyathura polita</i> | 21 | 8.22 | 0.70 | 0.0001 | 57 | x | 15 |
| <i>Menidia</i> spp. preflexion larvae* | 17 | 3.43 | 1.21 | 0.0047 | 42 | x | 41 |
| <i>Chaoborus punctipennis</i> | 22 | 2.66 | 1.83 | 0.0001 | 54 | x | 27 |
| <i>Diaptomus</i> spp.* | 14 | 0.63 | 1.89 | 0.0053 | 49 | | 20 |
| <i>Mesocyclops edax</i> | 19 | -0.46 | 2.34 | 0.0001 | 62 | | 22 |

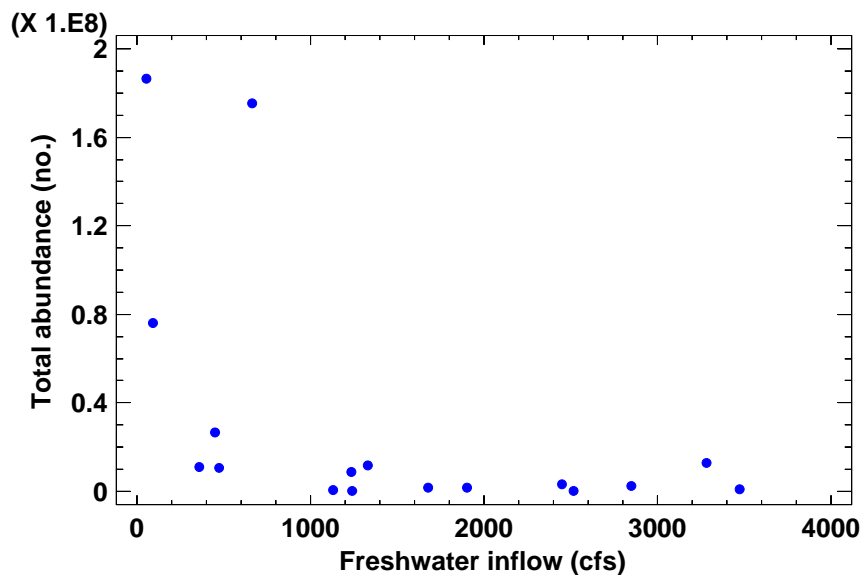


Figure 3.7.2.1. Negative relationship between total abundance of *Anchoa mitchilli* postflexion larvae and freshwater inflow (120-d lag).

Time lags associated with the responses were variable and in most cases significant correlations existing for multiple lags (Appendix J), but most taxa responded to inflow on either end of the lag spectrum, at scales of days or scales of several months. In some cases, a lognormal model applied to non-transformed data provided a better fit (based on adjusted R^2) than the linear model applied to transformed data (Fig. 3.7.2.3; Appendix I).

In contrast, eleven taxa, including preflexion larvae of the spotted seatrout *Cynoscion nebulosus* exhibited increased abundance in response to freshwater inflow. Again, associated lags were highly variable, but correlation coefficients with lowest associated p values were all at time scales of <51 days (Table 3.7.2.1). Based on the criteria of identifying non-freshwater

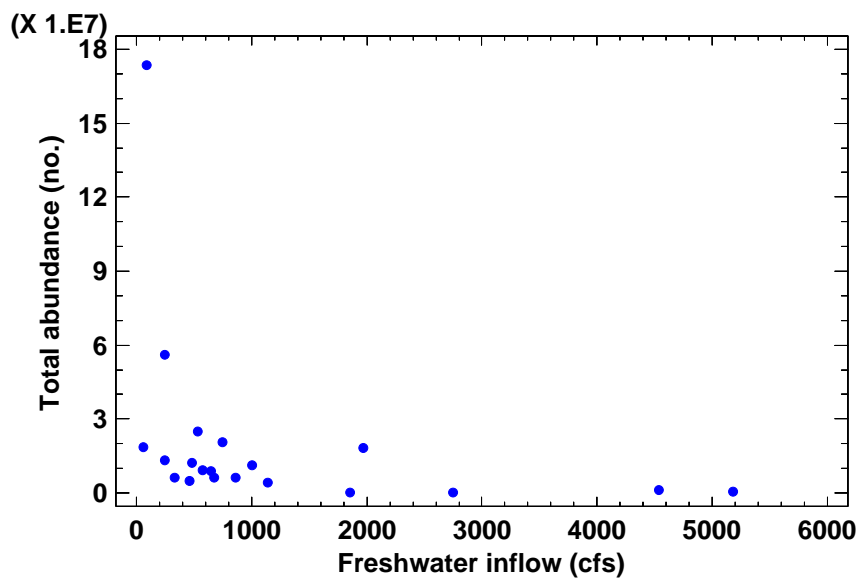


Figure 3.7.2.2. Negative relationship between total abundance of the ostracod *Parasterope pollex* and freshwater inflow (67-d lag).

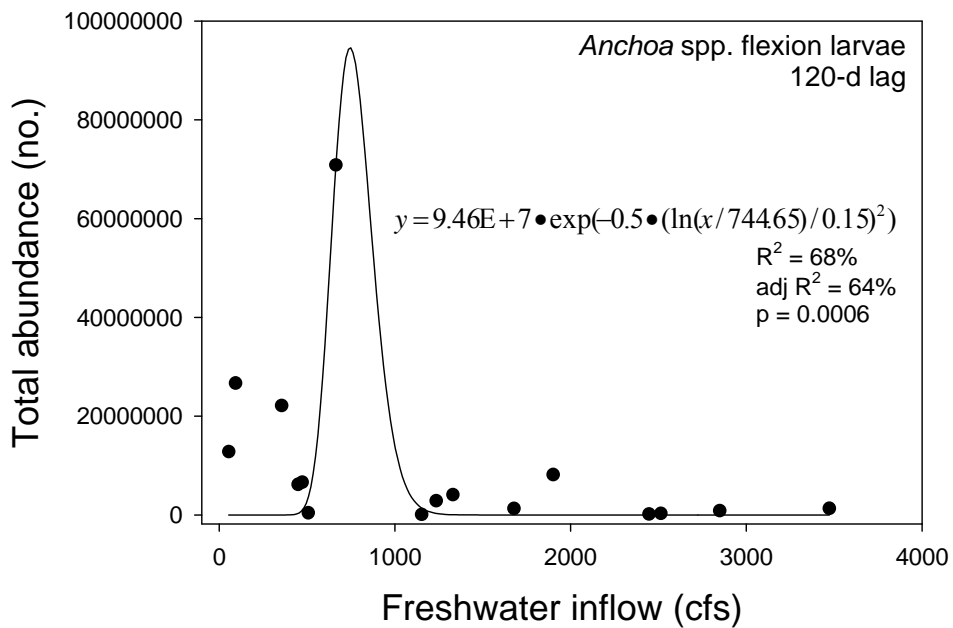


Figure 3.7.2.3. Lognormal relationship between total abundance of *Anchoa* spp. flexion larvae and freshwater inflow (120-d lag).

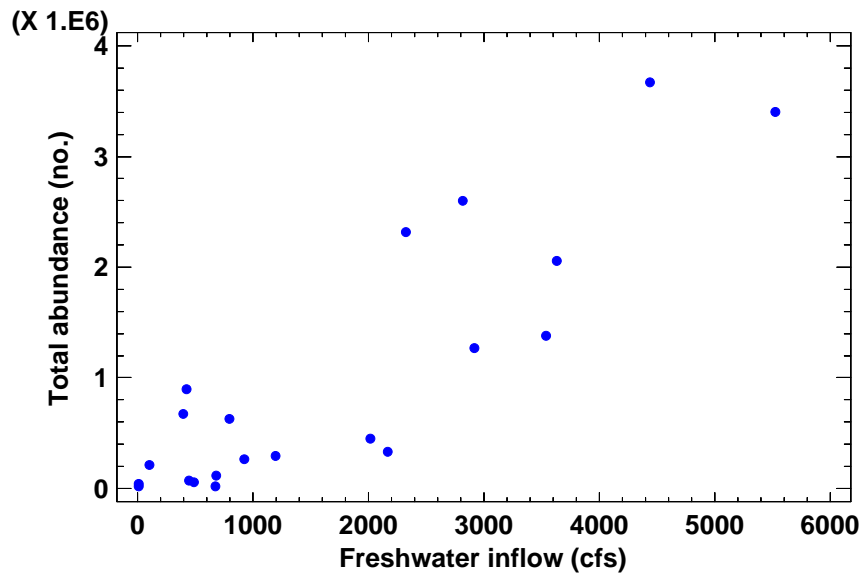


Figure 3.7.2.4. Positive relationship between total abundance of the isopod *Cyathura polita* and freshwater inflow (15-d lag).

species with a positive abundance relationship to inflow, seven taxa were identified as potential indicators of freshwater inflow: *Anchoa mitchilli* adults (bay anchovy), *Cynoscion nebulosus*, *Americamysis bahia* (mysid), Penaeidae metamorphs (commercial shrimp), *Penilia avirostris* (cladoceran), *Cyathura polita* (isopod), and *Menidia* spp. preflexion larvae (silversides).

However, only the relationship for *C. polita* exhibited a coefficient of determination of >50% (Fig. 3.7.2.4).

3.8 Identifying Truncated Distributions

A plot of the 90th percentiles of cumulative organism density for each sampling survey (month) suggests that distributions of juvenile bay anchovies and adult *Americamysis almyra* may be truncated due to the presence of the W.P. Franklin Lock and Dam (Fig. 3.7.1). Such blockage of further upstream migration is inferred when data converge and become unresponsive to flow (upper left data points of Fig. 3.8.1). Bin Din and Gunter (1986) reported that *Americamysis* (= *Mysidopsis*) *almyra* was a major component in the diet of bay anchovy sampled from Biloxi Bay, Mississippi, and Peebles et al. (2007) noted that the distributions of bay anchovy and *A. almyra* tracked one another closely in the Alafia River estuary, even though

4.0 CONCLUSIONS

Synthesis of the preceding information suggests that there are six principal project findings that are relevant to the establishment of water reservations for the Caloosahatchee River and Estuary.

1. It appears that phytoplankton biomass can accumulate and/or proliferate under various inflow scenarios, with inflow playing a role in determining both phytoplankton location and composition. Lower flows can cause larger phytoplankton (diatoms) to grow in the upper zones (5 and 6); although this may benefit higher trophic levels (shorter food chains, higher nutritional value) it may also lead to reduced dissolved oxygen levels in bottom waters as the diatom biomass sinks and decomposes. Higher flows support growth farther down the estuary (zones 2 and 3) and are accompanied by an increase in cyanobacteria. This scenario may have negative food web implications (longer food chains, lower nutritional value) and possible harmful algal bloom consequences (many cyanobacteria produce phycotoxins, e.g., *Microcystis*). Very high inflows (>3,500 cfs) can reduce phytoplankton biomass within the system, causing problems for those species requiring some level of salinity and flushing nutrients out into the Gulf where algal blooms may occur in coastal waters. Therefore, it appears that moderate flows (1,000 to 2,000 cfs) may be best, although potential blooms in zones 2 and 3 should be monitored in case the apparent stimulation at 1,500 cfs is a consistent and predictable response.

2. Water-column chlorophyll *a* concentrations were typically greater upstream, especially during the dry season, with peak values generally located in the region of 30-37 rkm. During the wet season, freshwater copepods *Mesocyclops edax* and *Diaptomus spp.*, cladocerans *Daphnia spp.*, and larvae of the phantom midge *Chaoborus punctipennis* all exhibited centers of abundance in this region. These species are known to feed on phytoplankton or on phytoplankton grazers (Chimney et al. 1981, Williamsons and Magnien 1982).

During the dry season, however, a number of estuarine species moved into this region from centers of abundance farther downstream, including estuarine-dependent fishes (e.g., *Trinectes maculatus* postlarvae, and *Anchoa mitchilli* juveniles) and important prey of fishes including the hyperbenthic *Americamysis spp.* juveniles, *A. almyra* adults, and *Edotia triloba*.

These species were likely associated with high phytoplankton abundances, whether suspended in the water column (high chlorophyll *a* concentrations) or sedimented on the bottom (high F_{\max} values associated with centric diatoms). This is not to say that these potential prey organisms were grazing on phytoplankton directly: for example, hyperbenthic species often feed on plant detritus or benthic microalgae during the day, only to migrate vertically into the water column at need to feed on zooplankton (e.g., Kouassi et al. 2006).

The combination of high primary production and the concentration of trophic-dependent species in this portion of the river is of concern, for this region is also prone to low dissolved oxygen concentrations (2-4 mg L⁻¹) in its bottom waters, especially during wet months. The potential positioning of a hyperbenthic prey base in regions of low DO could result in a substantial loss of important food resources for fishes and invertebrates that feed on these species. In addition, hypoxia has been identified as a source of habitat compression (Eby and Crowder 2002, Prince and Goodyear, Bograd et al. 2008, Diaz and Rosenberg 2008), a phenomenon that can increase competitive interactions among species and that can potentially result in dietary shifts and reduced growth rates (Eby and Crowder 2002). Because all of the taxa listed above demonstrate significant inflow responses, the release of freshwater at S-79 during dry months (when these taxa are most likely to concentrate in the upper tidal river) would result in the relocation of these taxa, downstream and away from this region prone to the development of low DO. Based on inflow relationships of hyperbenthic prey, it appears that releases of 1,000–1,200 cfs would achieve this result.

3. Based on the results of the spatial abundance quartile analysis, it appears that juvenile bay anchovy and their prey *Americamysis almyra* are blocked from moving farther upstream during the dry season by the presence of the W.P. Franklin Lock and Dam. This pattern indicates habitat compression for these organisms, as they are not only concentrated in a relatively narrow portion of the river but also are truncated in their upstream distribution. By obstructing the movement of estuarine organisms, the Lock and Dam causes habitat impingement for some species, preventing them from expanding their range upstream. Based on the 90th percentile abundance plot, it appears that releases of freshwater on the order of 1,000 cfs at S-79 would be sufficient to release these organisms from the impingement caused by the Lock and Dam structure itself.

4. As mentioned previously, a number of estuarine species collected during this study (including fishes, mysids, commercial shrimp, and isopods) responded to reduced freshwater inflow by moving upstream into the narrow portion of the tidal Caloosahatchee above Beautiful Island during the dry season. Although these organisms may experience greater overall dispersion as a result of this movement, Peebles and Greenwood (2009) cautioned that such gains in habitat extent may be offset by losses in habitat volume due to the inherent geomorphology (i.e., funnel shape) of tidal rivers. This upstream movement in response to reduced inflow into the more restricted region of the tidal Caloosahatchee resulted in a potential second source of habitat compression. By concentrating aquatic organisms in a reduced volume of habitat, habitat compression increases competitive interactions among and within species and enhances predator-prey encounters in the same habitat (Prince and Goodyear 2006). This can result in habitat overlap, higher predation rates, decreased prey availability, and reduced growth rates (Crowder 1986, Eby and Crowder 2002). Based on the compiled regressions of organism center of abundance vs. freshwater inflow, it appears that a number of these species would be relocated downstream of this restricted portion of the tidal river at inflows of 800–1,000 cfs.

5. In addition to changes in center of distribution in response to variable inflow, some taxa exhibited significant changes in total abundance in response to lagged inflow. The majority of these responses were negative, indicating reduced system-wide abundances at higher levels of inflow. Based on the examination of non-transformed data, it appears that, for many of these taxa, greatest abundances would be maintained at inflows $\leq 1,000$ cfs.

6. Gelatinous predators can negatively impact fish populations by feeding directly on developing eggs and larvae or by competing with larvae for zooplankton prey (Purcell and Arai 2001). Centers of abundance calculated for most of the gelatinous predators identified from Caloosahatchee zooplankton samples were located well downstream, either in the lower tidal river or downstream of the river mouth in San Carlos Bay; however, two species were notably present in the upper river during periods of reduced freshwater inflow. Populations of the hydromedusa *Clytia* sp. and the ctenophore *Mnemiopsis leidyi* were centered at ~40 rkm and 32-39 rkm, respectively, during the dry season when abundances were high. As reported previously, a number of estuarine zooplankton also moved into the upper tidal river during the dry season.

Of these two gelatinous predators, *Clytia* sp. exhibited a very sensitive response to freshwater inflow (i.e., slope of the regression). Considering both species, freshwater releases of 1,000 cfs or less should result in substantial movement downstream, relocating these gelatinous predators away from areas in the upper tidal river where potential zooplankton prey (including larval fishes) are concentrated, thus reducing predation and competition pressure.

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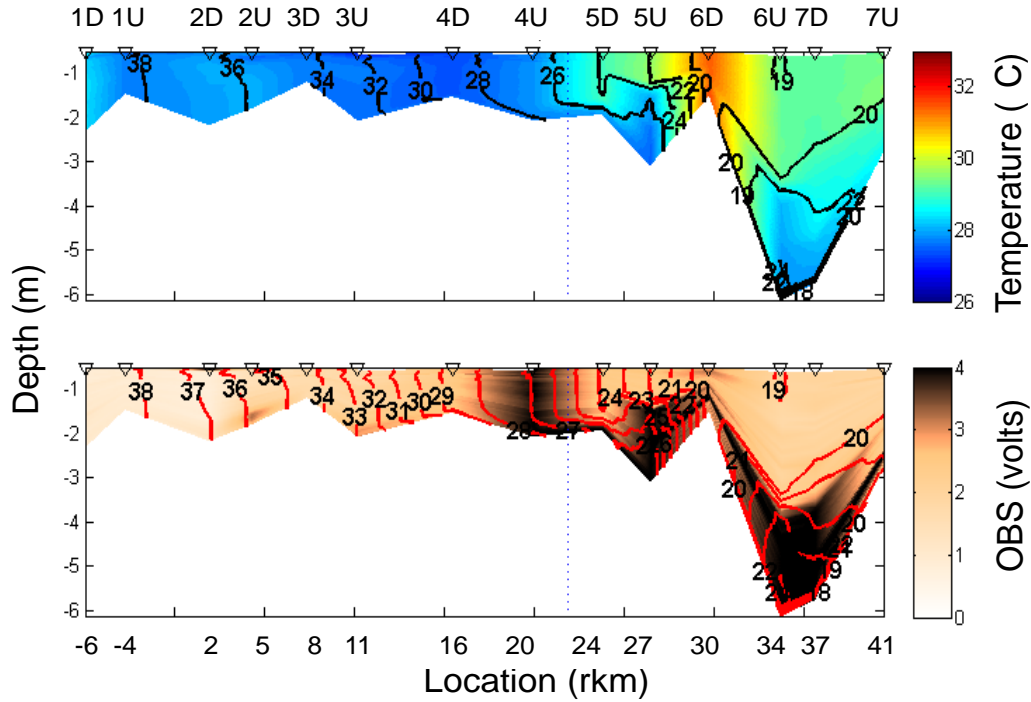
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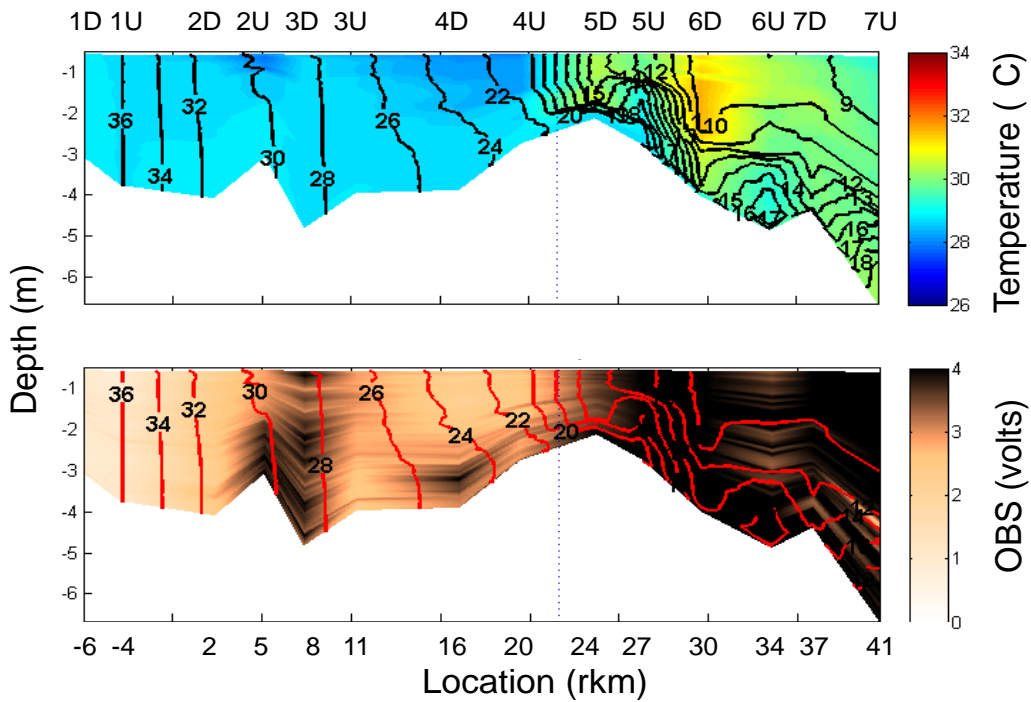
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Appendix A

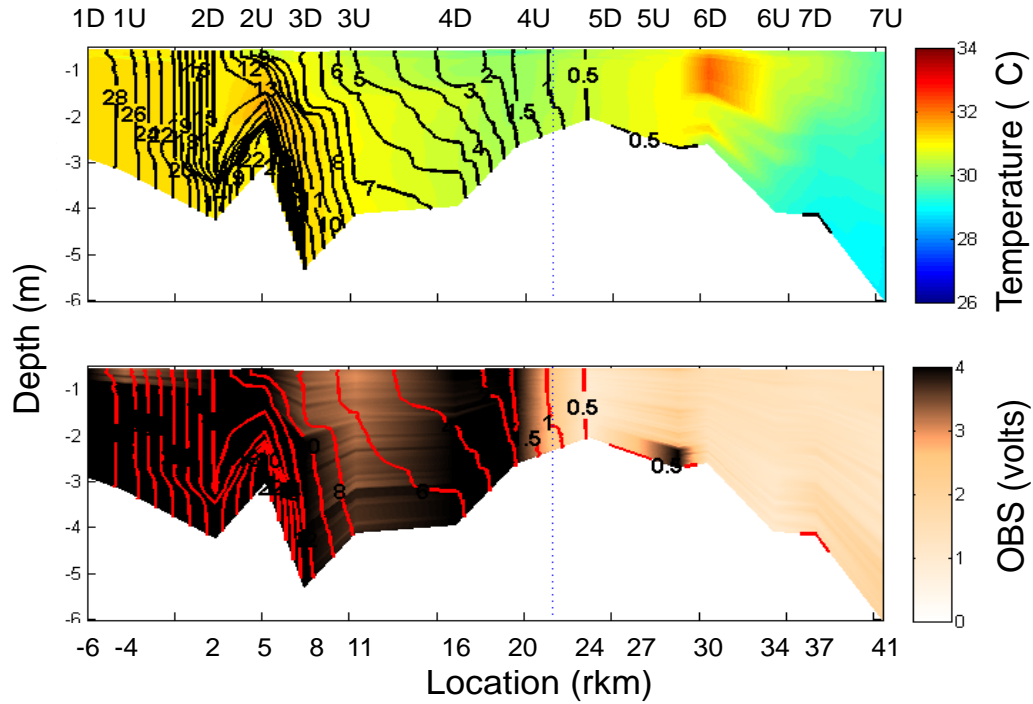
Monthly longitudinal temperature distribution with isohalines



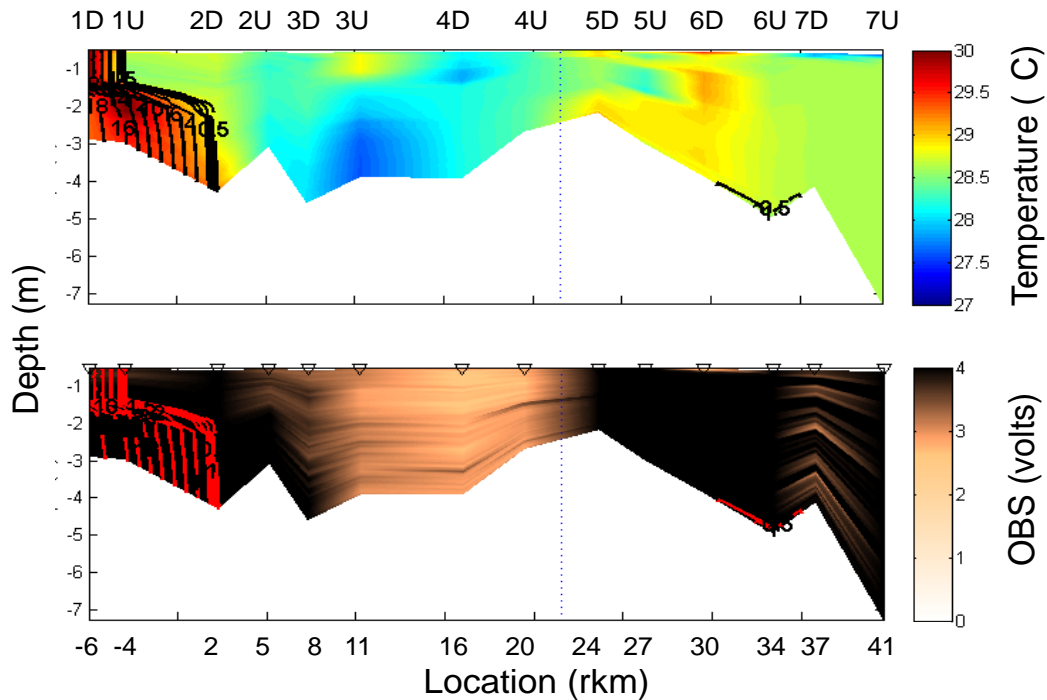
Top: Longitudinal temperature distribution with isohalines for May 2008. Bottom: Turbidity distribution (optical back scatter) with isohalines for May 2008. Stations are labeled according to site.



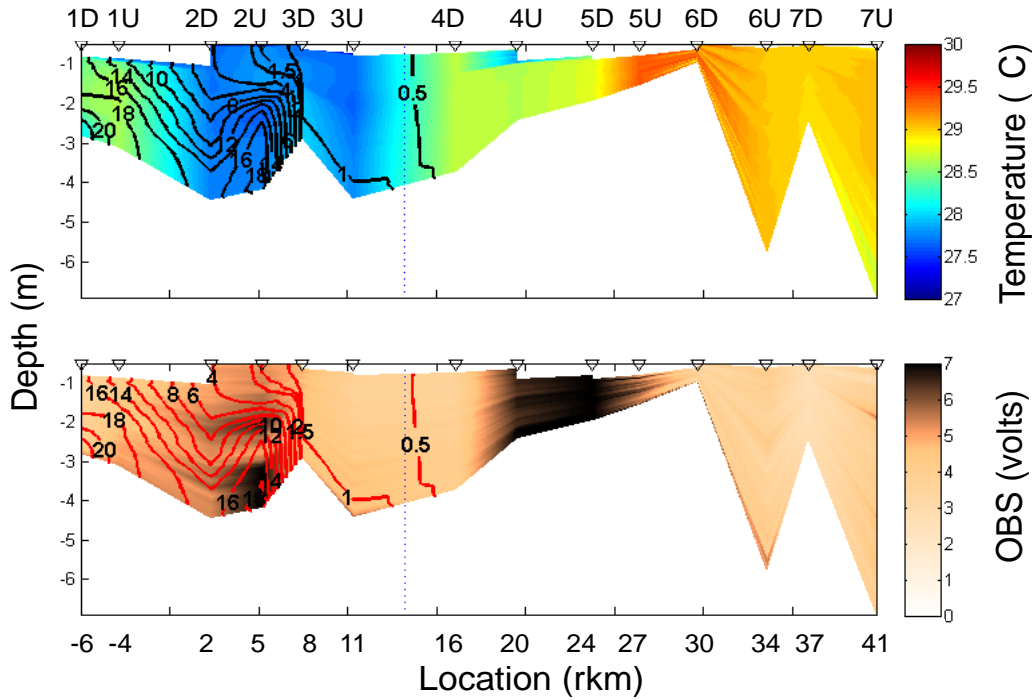
Top: Longitudinal temperature distribution with isohalines for June 2008. Bottom: Turbidity distribution (optical back scatter) with isohalines for June 2008. Stations are labeled according to site.



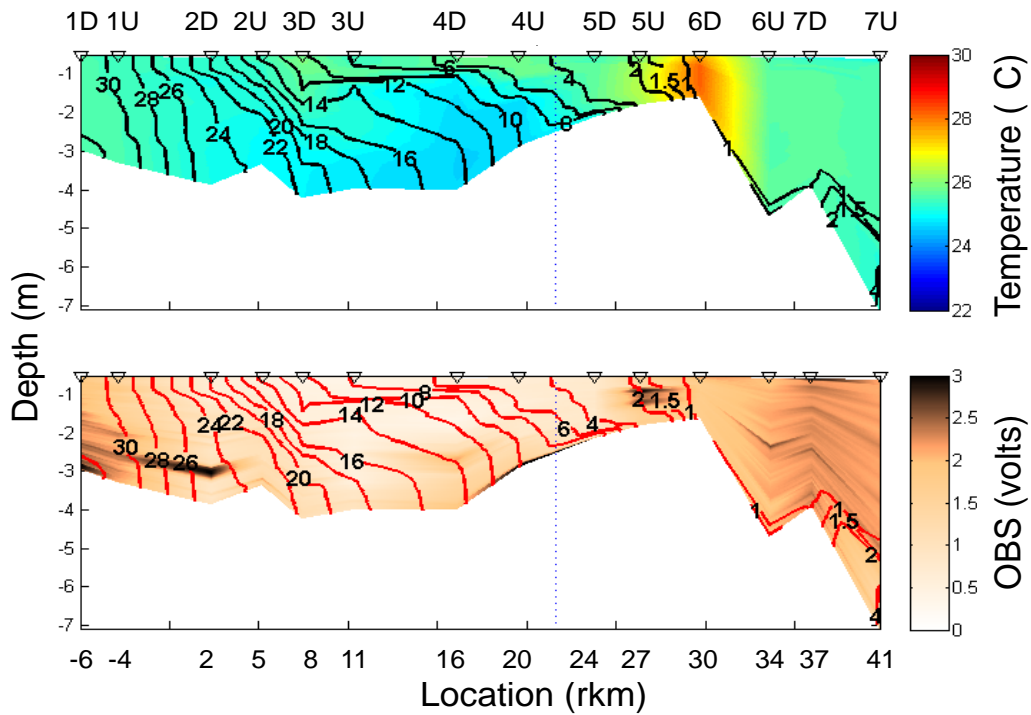
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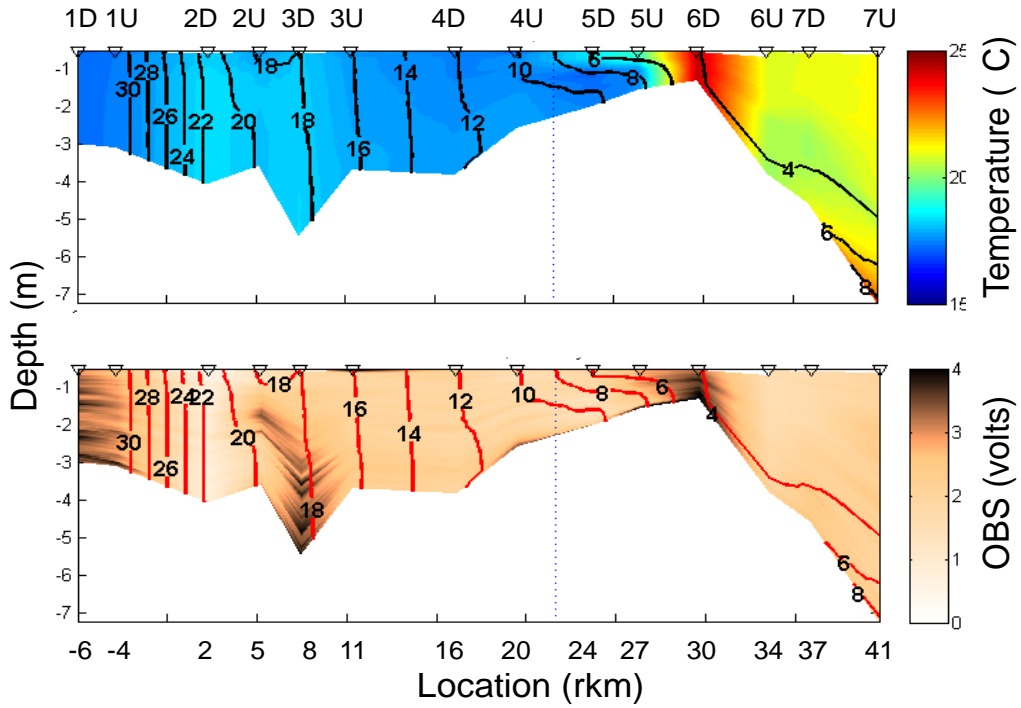
Top: Longitudinal temperature distribution with isohalines for August 2008. Bottom: Turbidity distribution (optical back scatter) with isohalines for August 2008. Stations are labeled according to site.



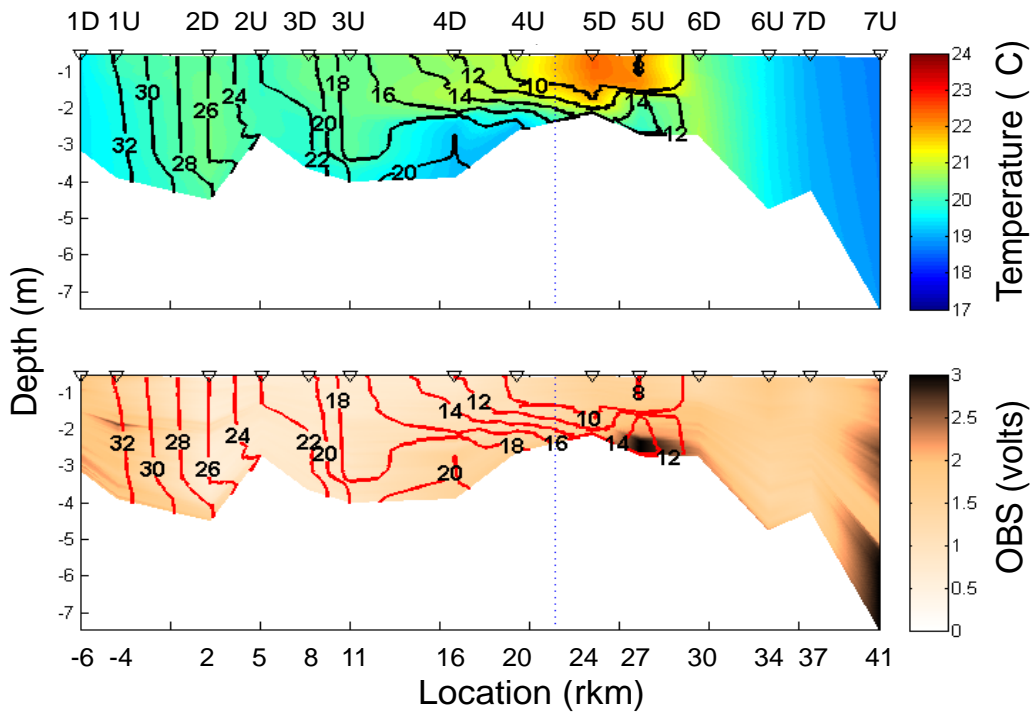
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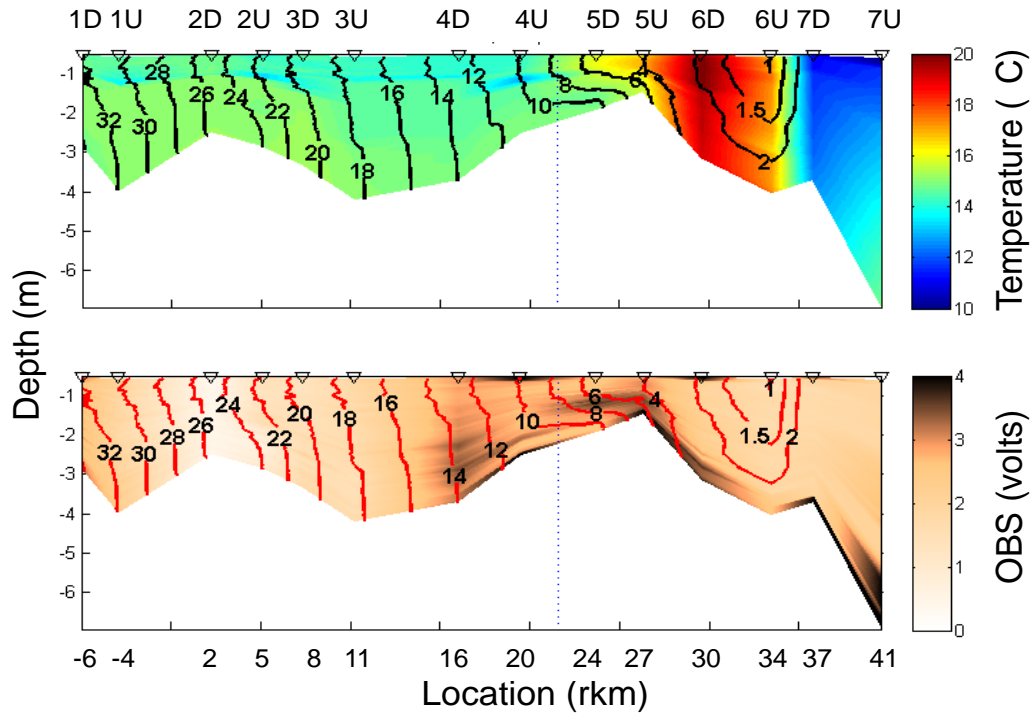
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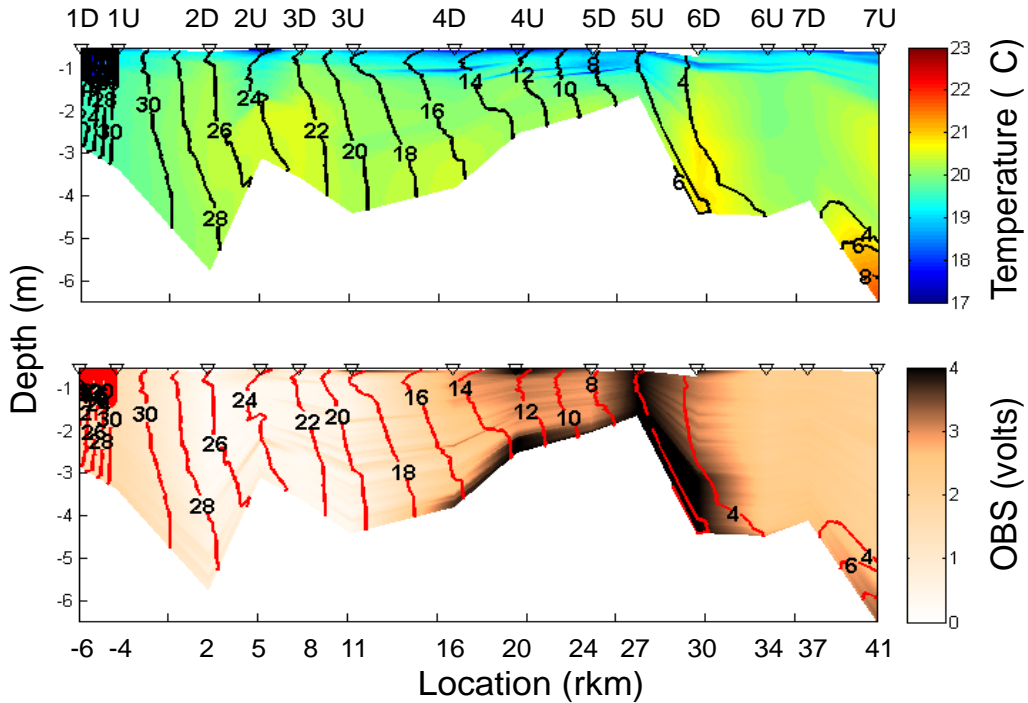
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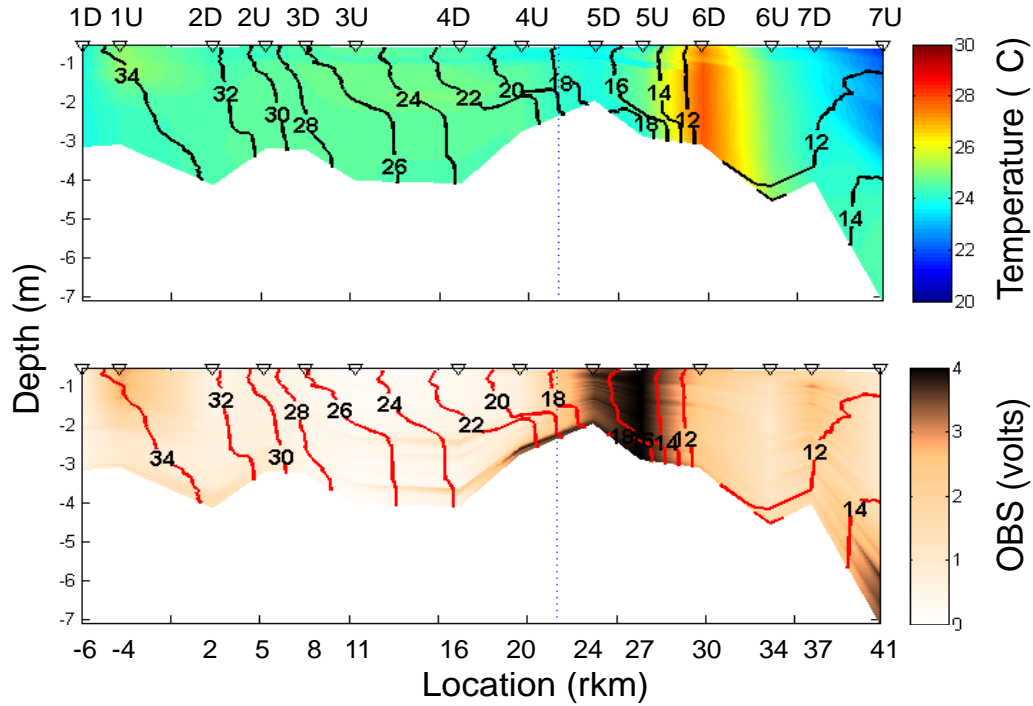
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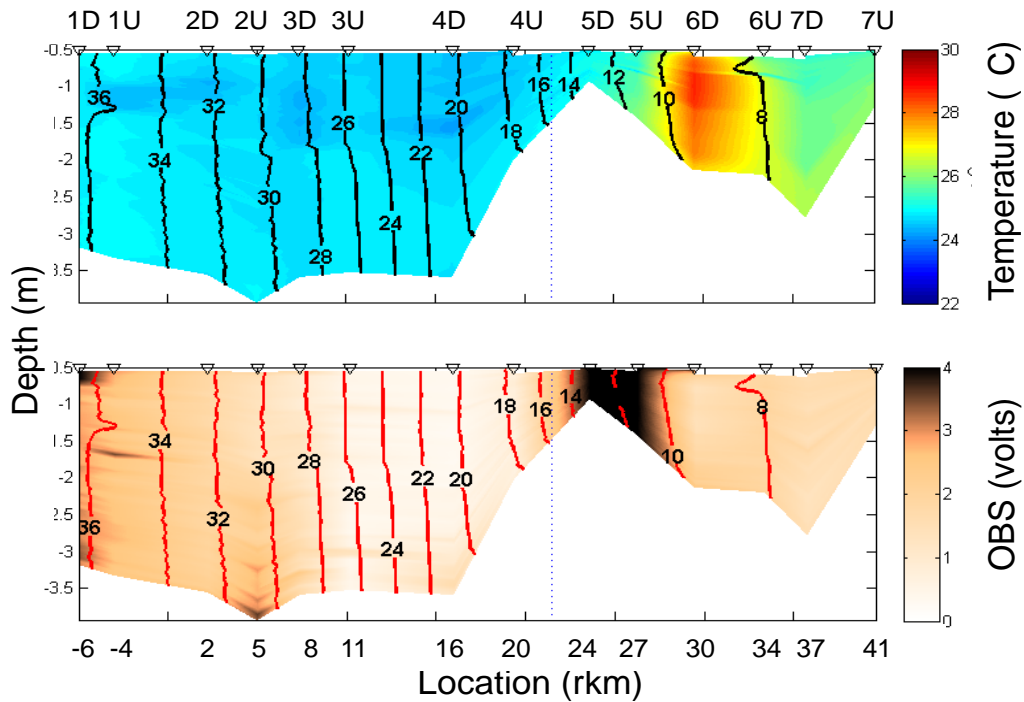
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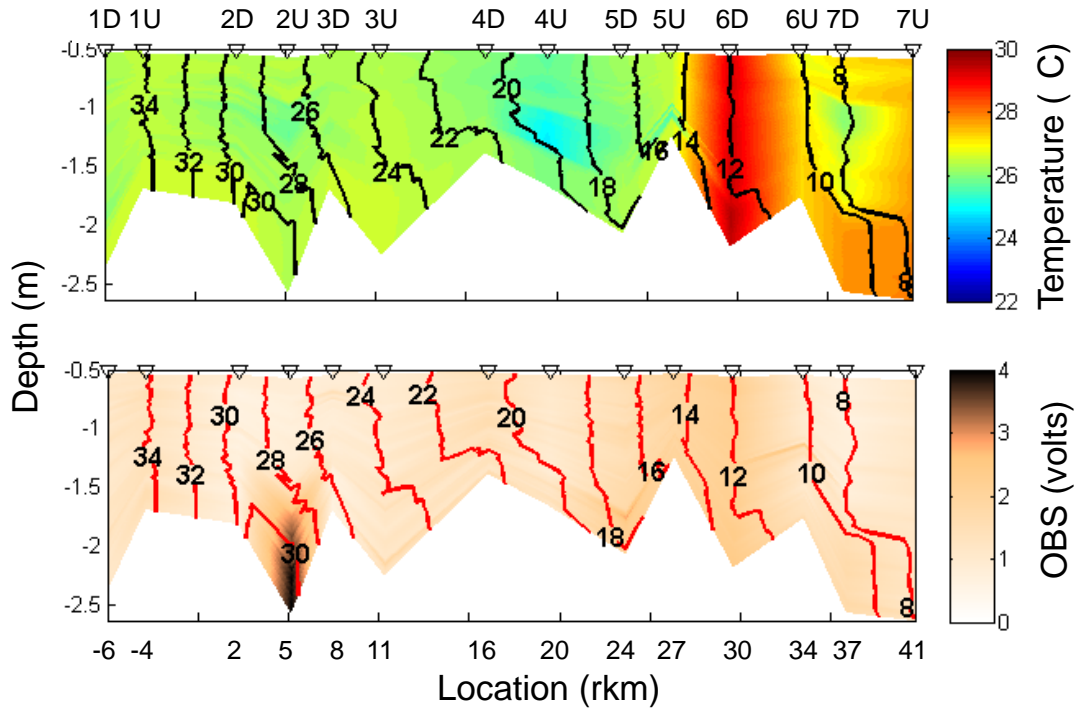
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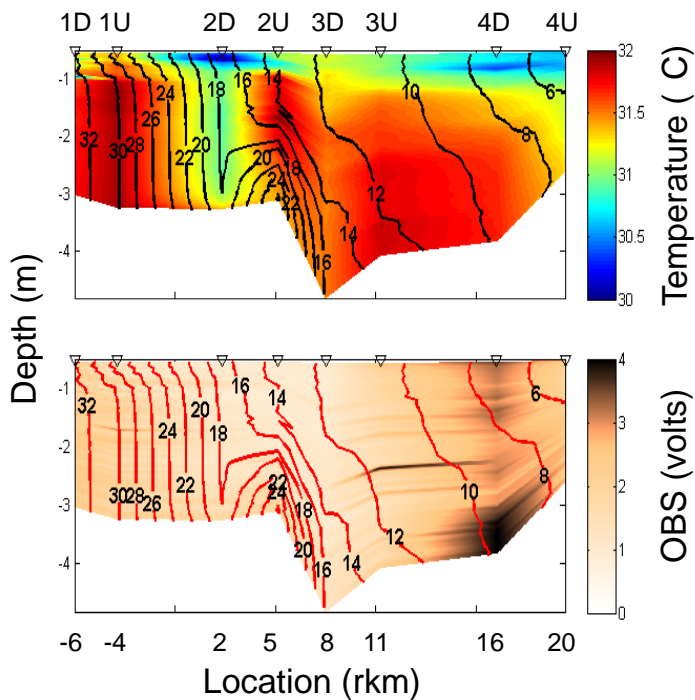
Top: Longitudinal temperature distribution with isohalines for March 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for March 2009. Stations are labeled according to site.



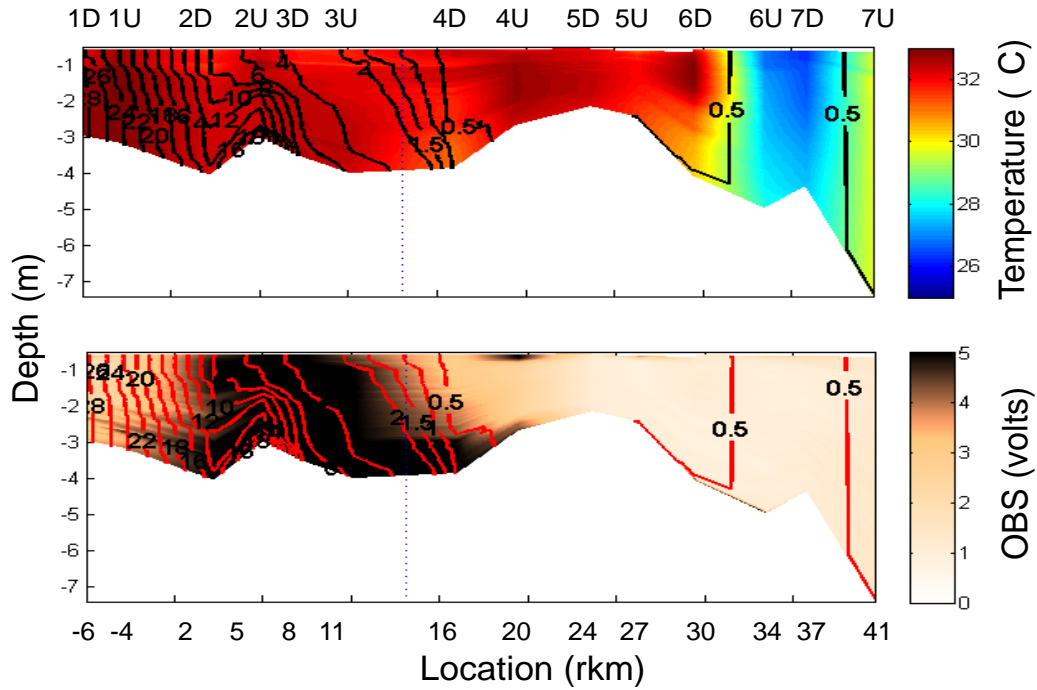
Top: Longitudinal temperature distribution with isohalines for April 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for April 2009. Stations are labeled according to site



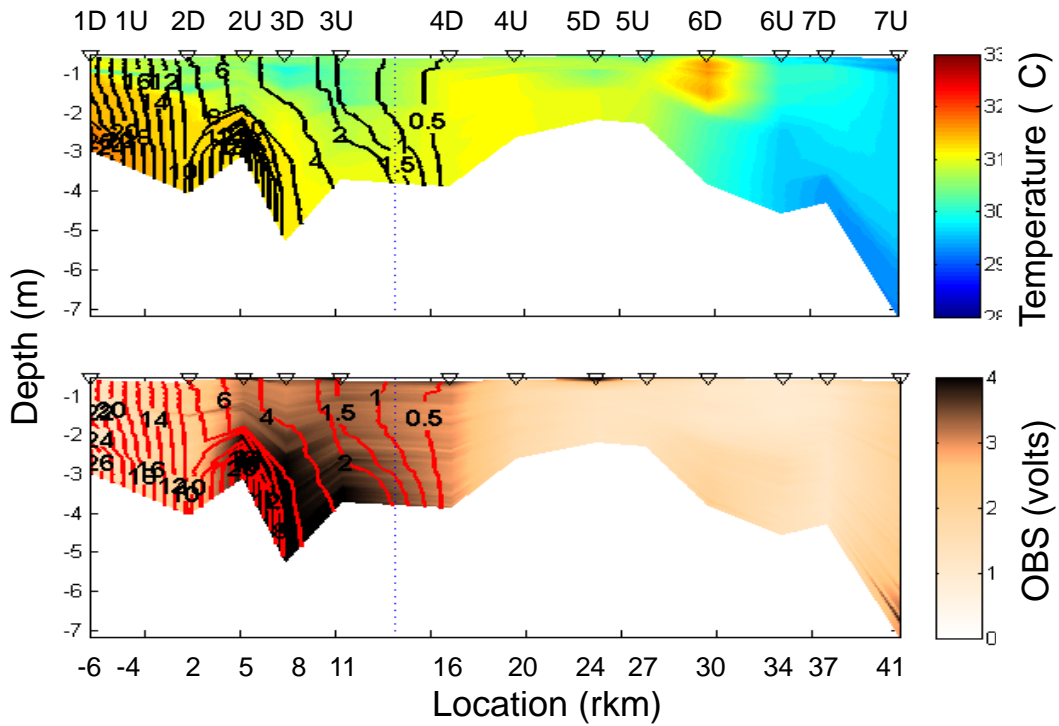
Top: Longitudinal temperature distribution with isohalines for May 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for May 2009. Stations are labeled according to site.



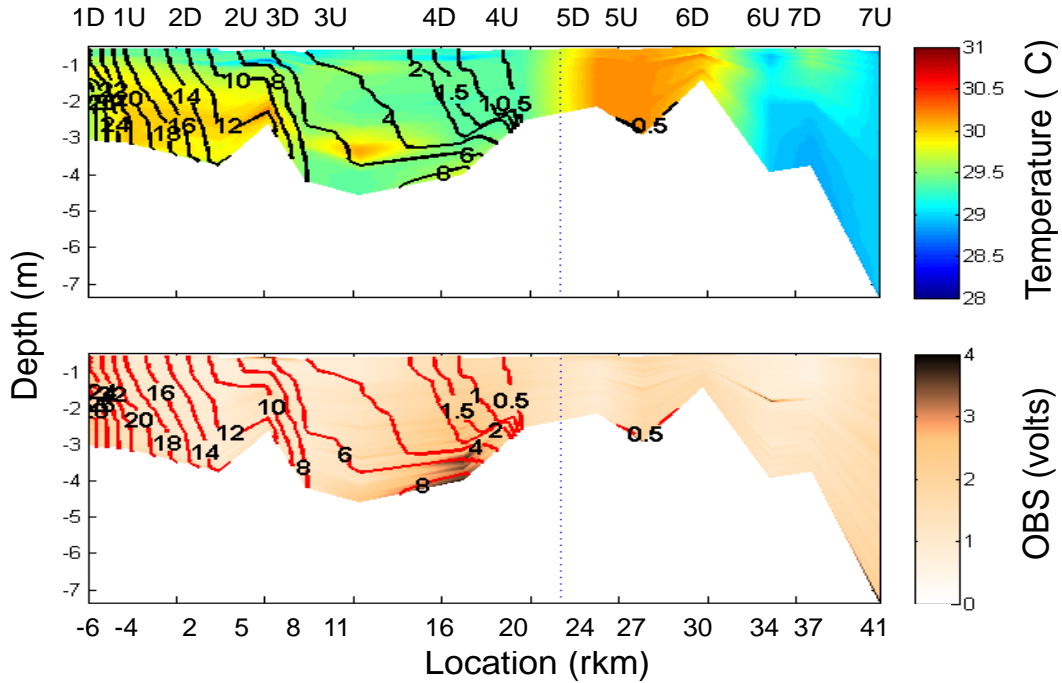
Top: Longitudinal temperature distribution with isohalines for June 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for June 2009. Stations are labeled according to site. Upper stations were not sampled due to instrument failure.



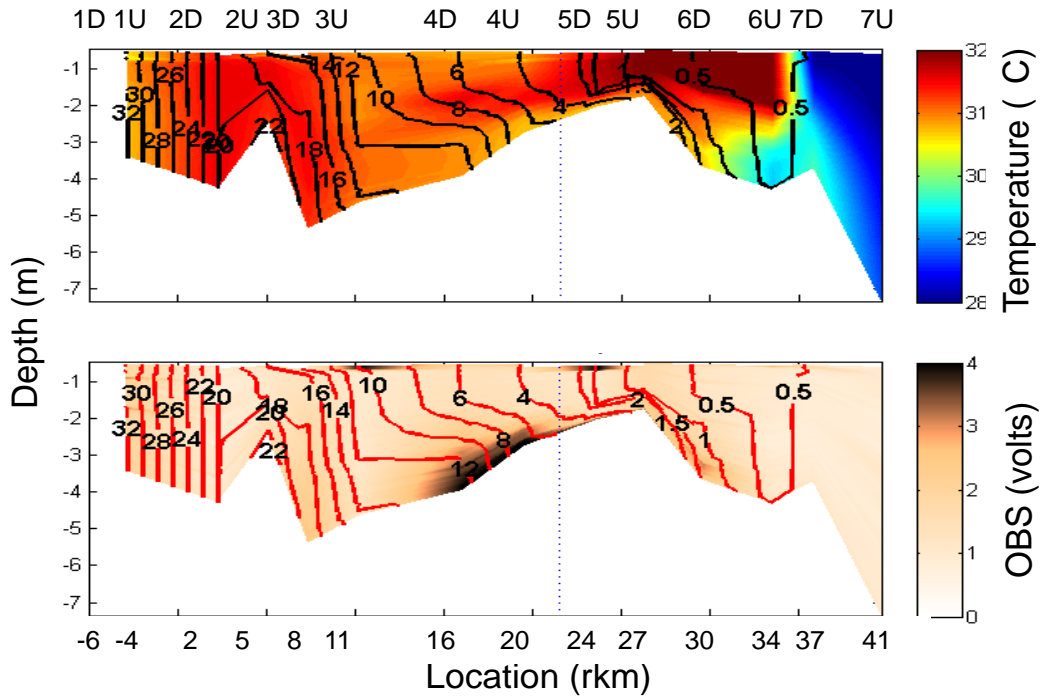
Top: Longitudinal temperature distribution with isohalines for July 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for July 2009. Stations are labeled according to site.



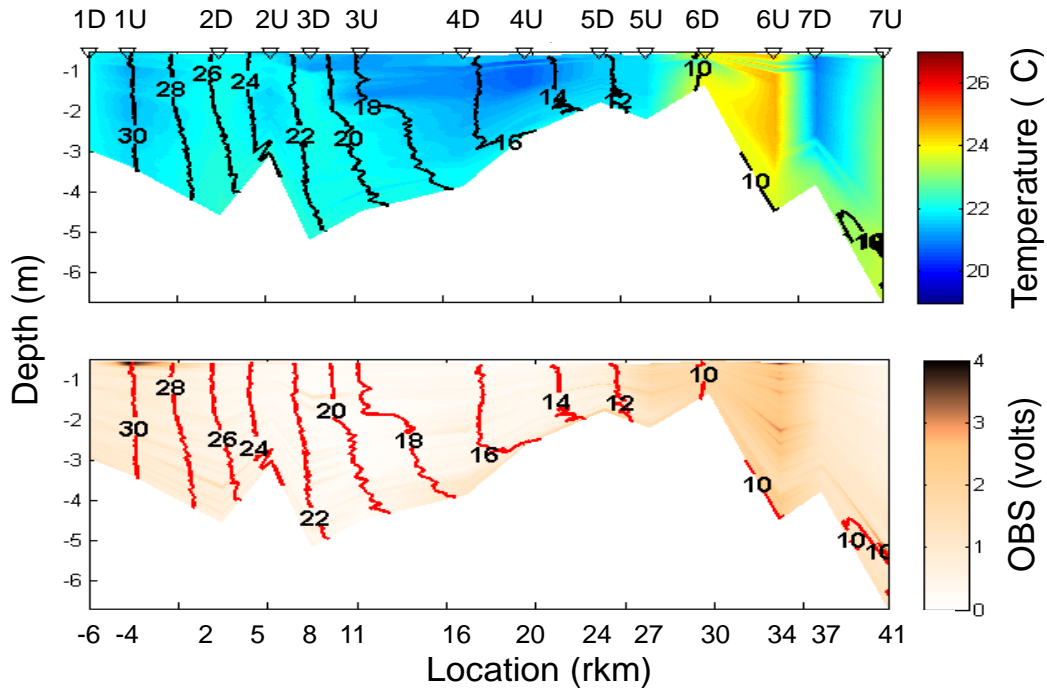
Top: Longitudinal temperature distribution with isohalines for August 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for August 2009. Stations are labeled according to site.



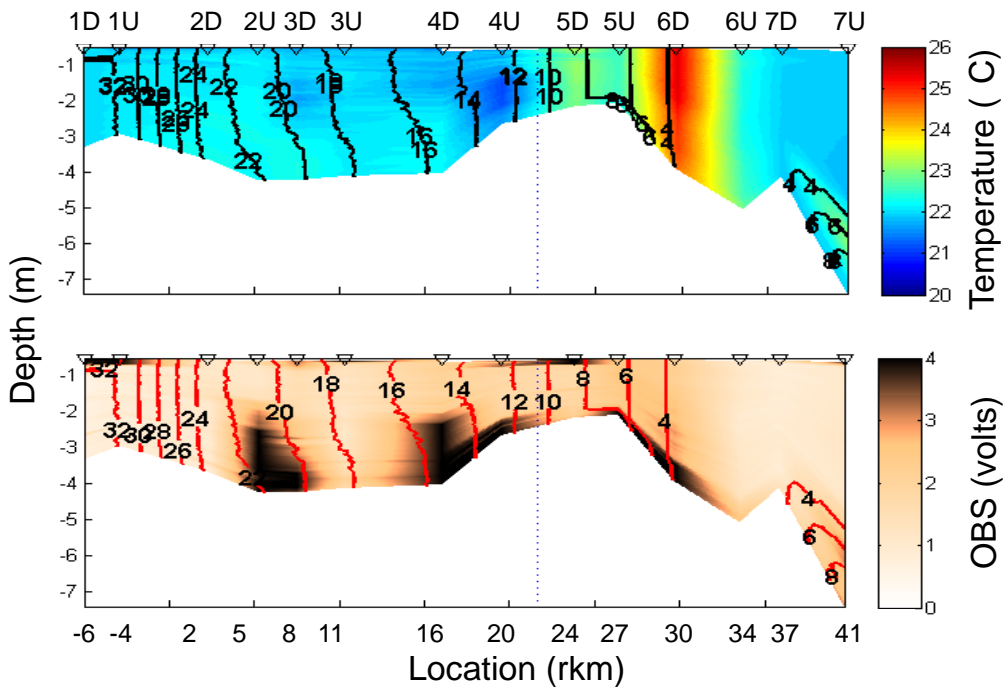
Top: Longitudinal temperature distribution with isohalines for September 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for September 2009. Stations are labeled according to site.



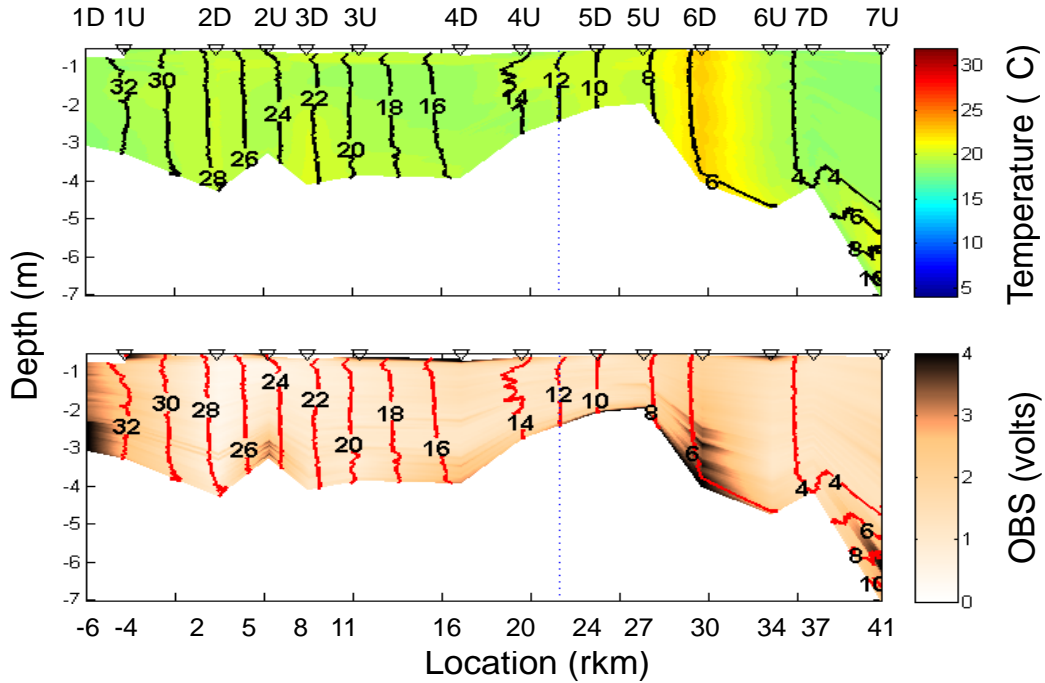
Top: Longitudinal temperature distribution with isohalines for October 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for October 2009. Stations are labeled according to site.



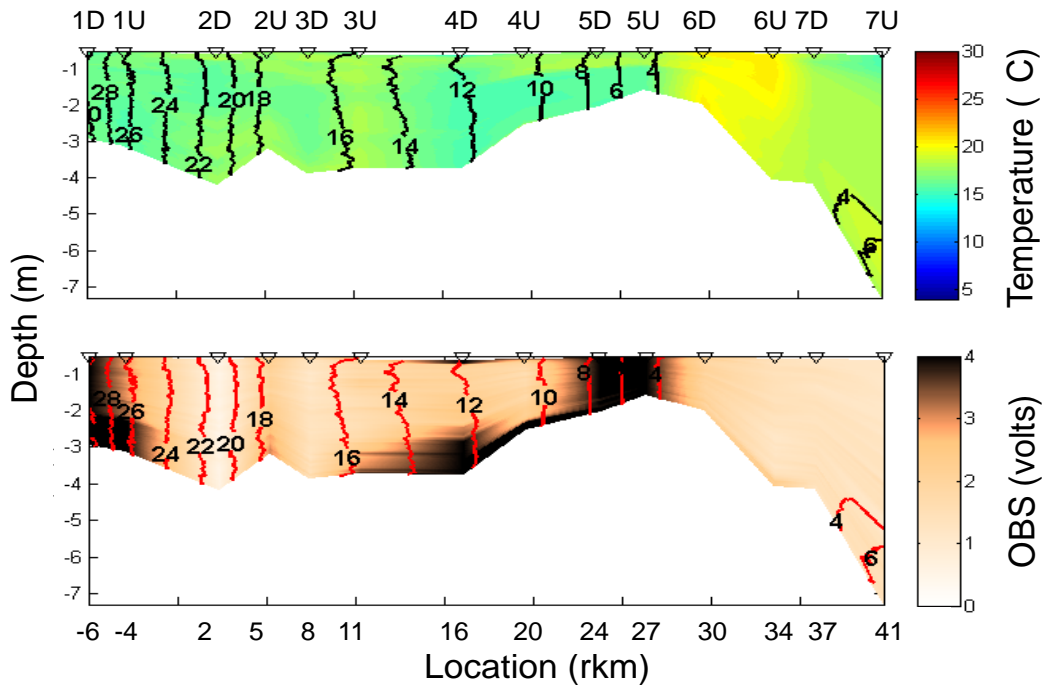
Top: Longitudinal temperature distribution with isohalines for November 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for November 2009. Stations are labeled according to site.



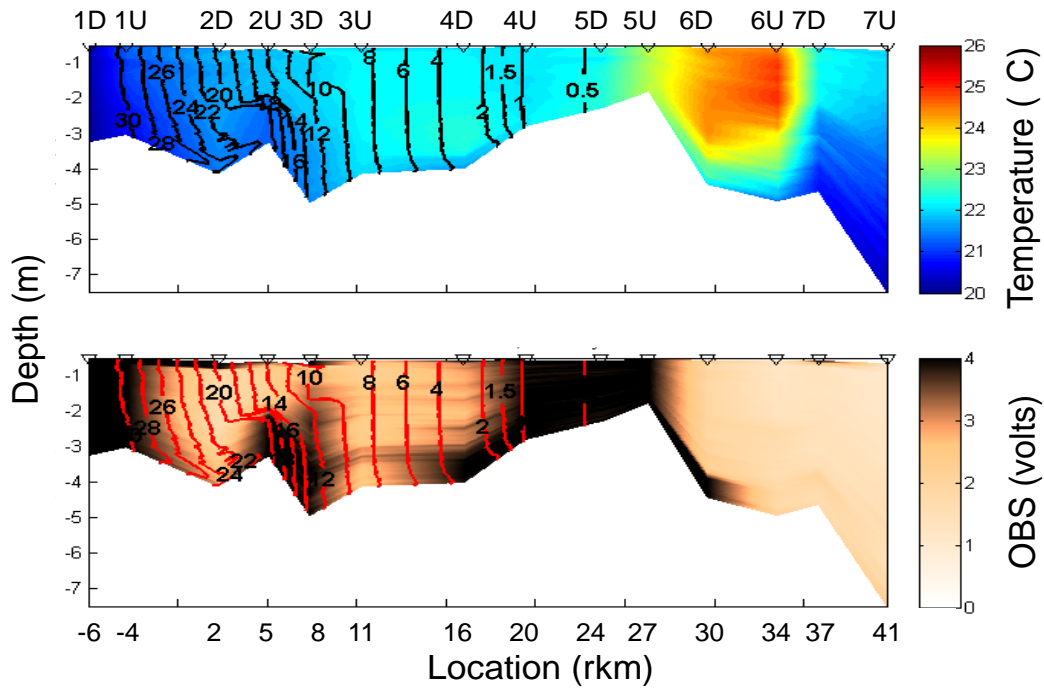
Top: Longitudinal temperature distribution with isohalines for December 2009. Bottom: Turbidity distribution (optical back scatter) with isohalines for December 2009. Stations are labeled according to site.



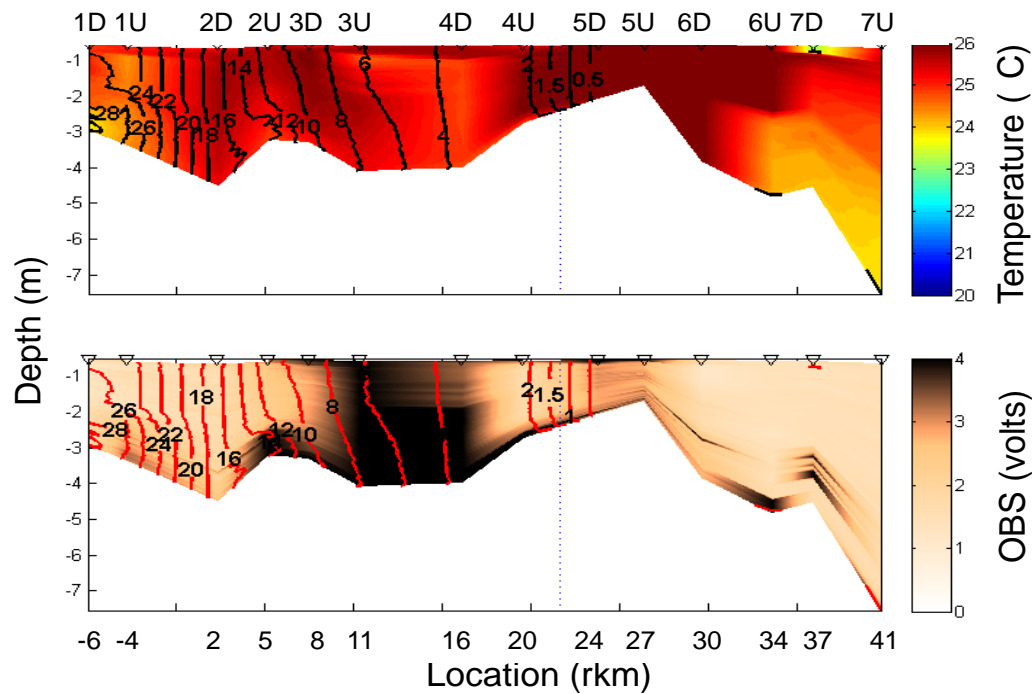
Top: Longitudinal temperature distribution with isohalines for January 2010. Bottom: Turbidity distribution (optical back scatter) with isohalines for January 2010. Stations are labeled according to site.



Top: Longitudinal temperature distribution with isohalines for February 2010. Bottom: Turbidity distribution (optical back scatter) with isohalines for February 2010. Stations are labeled according to site.



Top: Longitudinal temperature distribution with isohalines for March 2010. Bottom: Turbidity distribution (optical back scatter) with isohalines for March 2010. Stations are labeled according to site.



Top: Longitudinal temperature distribution with isohalines for March 2010. Bottom: Turbidity distribution (optical back scatter) with isohalines for March 2010. Stations are labeled according to site.

Appendix B

Phytoplankton abundance (cells L⁻¹) for selected samples

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 7D | 6D | 7U | 5U | 5D | 6D |
|---|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2008 | June_2008 | July_2008 | Aug_2008 | Sept_2008 | Oct_2008 |
| <i>Actinastrum</i> | 0 | 0 | 7727 | 0 | 0 | 101 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 68 | 43 | 0 |
| <i>Amphora</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Anabaena</i> | 0 | 1416 | 9421 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 24 | 141 | 0 | 0 | 110 |
| <i>Apedinella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 0 | 795 | 3105 | 0 | 0 |
| armored auto dino >30 | 196 | 94 | 0 | 0 | 0 | 7 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 304 | 0 |
| armored auto dino 20-30 | 0 | 376 | 212 | 0 | 0 | 22 |
| <i>Asterocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | 0 | 0 | 0 | 177 | 196 | 0 |
| <i>Aulacoseira</i> fine | 0 | 0 | 106 | 177 | 500 | 55 |
| autotrophic ciliate 10-15 | 0 | 0 | 7 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 196 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 196 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| centric >30 | 785 | 0 | 0 | 89 | 87 | 0 |
| centric 10-15 | 0 | 94 | 0 | 0 | 413 | 0 |
| centric 15-20 | 1767 | 94 | 0 | 89 | 22 | 0 |
| centric 20-30 | 785 | 6 | 0 | 0 | 0 | 14 |
| centric 5-10 | | 0 | 0 | 0 | 87 | 0 |
| <i>Centritractus</i> | 0 | 0 | 0 | 0 | 0 | 55 |
| <i>Cerataulina bicornis</i> | 5498 | 94 | 0 | 0 | 0 | 0 |
| <i>Cerataulina</i> resting spore | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 0 | 94 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 41 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 61 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 393 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 230 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 196 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 0 | 12 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 27 | 565 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 0 | 12 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 0 | 565 | 0 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 1178 | 753 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 94 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 12 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 94 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 15-20 | 0 | 94 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 7 |
| chlorophyte coccoid 5-10 | 0 | 84 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 7D | 6D | 7U | 5U | 5D | 6D |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2008 | June_2008 | July_2008 | Aug_2008 | Sept_2008 | Oct_2008 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 319 | 177 | 0 | 144 |
| chlorophyte coccoid colonial 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte filamentous | 0 | 0 | 318 | 0 | 0 | 57 |
| chlorophyte flagellated colonial | 0 | 0 | 82 | 181 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 22 | 0 |
| <i>Chroococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 22 | 0 |
| <i>Closterium</i> | 0 | 0 | 106 | 23 | 0 | 55 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> >80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 94 | 0 | 0 | 0 | 7 |
| <i>Coscinodiscus granii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cosmarium</i> | 0 | 0 | 0 | 1380 | 0 | 0 |
| cryptomonad auto 10-15 | | 6 | 0 | 0 | | 0 |
| cryptomonad auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 565 | 1905 | 0 | 0 | 1822 |
| cyanobacteria filament | 0 | 3293 | 3070 | 44356 | | 2111 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 724 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 22 |
| <i>Cyclotella</i> 10-15 | 0 | 282 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 15-20 | 0 | 0 | 0 | 89 | 0 | 110 |
| <i>Cyclotella</i> 20-30 | 196 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 177 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 0 | 0 | 0 | 7 |
| <i>Cylindrotheca</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 20-30 | 20 | 6 | 0 | 0 | 0 | 0 |
| <i>Desmodesmus</i> | 0 | 0 | 318 | 1419 | 87 | 65 |
| <i>Diacanthos belenophorus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 7 |
| <i>Dinophysis ovum</i> | 0 | 36 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Entomoneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 0 | 1694 | 0 | 0 | 0 |
| <i>Euglena acus</i> | 0 | 12 | 0 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto >30 | 7 | 0 | 0 | 45 | 0 | 55 |
| euglenoid auto 10-15 | 0 | 94 | 0 | 0 | 0 | 0 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 0 | 0 | 621 | 174 | 50 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 7D | 6D | 7U | 5U | 5D | 6D |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2008 | June_2008 | July_2008 | Aug_2008 | Sept_2008 | Oct_2008 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Golenkinia radiata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 0 | 94 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax spinifera</i> | 0 | 188 | 0 | 0 | 0 | 0 |
| <i>Guinardia flaccida</i> | 636 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 0 | 188 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Haptophyte | 0 | 0 | 7 | 23 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 7657 | 54 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Mallomonas</i> | 0 | 0 | 106 | 23 | 0 | 497 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira varians</i> | 0 | 24 | 423 | 2218 | 0 | 2539 |
| <i>Merismopedia</i> | 0 | 360 | 0 | 0 | 783 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Microcystis</i> | 0 | 0 | 2532 | 0 | 0 | 0 |
| <i>Microspora</i> | 0 | 0 | 0 | 0 | 0 | 55 |
| naked auto dino >30 | 0 | 188 | 106 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 0 | 106 | 0 | 0 | 0 |
| naked auto dino 15-20 | 0 | 94 | 1799 | 0 | 22 | 0 |
| naked auto dino 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 5-10 | 196 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula lanceolate</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia lanceolate</i> >30 | 0 | 12 | 0 | 89 | 22 | 110 |
| <i>Nitzschia</i> ovoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 45 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 0 | 0 | 115 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pediastrum</i> | 0 | 0 | 200 | 1863 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | 22 | 0 |
| pennate 10-20 | 0 | 188 | 0 | 0 | 0 | 0 |
| pennate 20-40 | 196 | 6 | 0 | 23 | 22 | 221 |
| pennate 40-80 | 0 | 0 | 0 | 89 | 0 | 7 |
| <i>Phacus</i> | 0 | 0 | 0 | 45 | 22 | 0 |
| Phaeophyte filamentous | 0 | 0 | 56101 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 7D | 6D | 7U | 5U | 5D | 6D |
|---------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2008 | June_2008 | July_2008 | Aug_2008 | Sept_2008 | Oct_2008 |
| phytoflagellate 10-15 | 0 | 0 | 106 | 266 | 0 | 276 |
| phytoflagellate 15-20 | 0 | 0 | 635 | 0 | 0 | 276 |
| phytoflagellate 20-30 | 0 | 6 | 847 | 89 | 0 | 0 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktolyngbya</i> | 0 | 0 | 15772 | 14105 | 0 | 129 |
| <i>Planktothrix</i> | 0 | 118657 | 19265 | 1471 | 0 | 373 |
| <i>Platydorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 0 | 0 | 89 | 0 | 0 |
| <i>Prorocentrum micans</i> | 785 | 1694 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 659 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia brasiliiana</i> | 108 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 128800 | 0 | 0 | 89 | 0 | 0 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 7 |
| <i>Radiococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 0 | 710 | 0 | 136 |
| <i>Scrippsiella</i> | 0 | 18 | 0 | 0 | 0 | 0 |
| silicoflagellate | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 8246 | 2352 | 0 | 2307 | 0 | 386 |
| <i>Skeletonema tropicum</i> | 7068 | 0 | 0 | 45 | 0 | 1325 |
| <i>Snowella</i> | 0 | 565 | 0 | 0 | 0 | 0 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 65 | 0 |
| <i>Spirulina</i> | 0 | 0 | 661 | 0 | 0 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 1109 | 57 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 15 | 45 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 7 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 23 | 0 | 0 |
| <i>Strombidium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Surirella</i> | 0 | 0 | 0 | 0 | 9 | 0 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 89 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 2749 | 12 | 0 | 0 | 0 | 55 |
| <i>Thalassiosira</i> 20-40 | 0 | 12 | 318 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|---|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| <i>Actinastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 0 | 17 | 0 |
| <i>Amphora</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Anabaena</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apedinella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 0 | 0 | 316 | 0 | 0 |
| armored auto dino >30 | 1095 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino 20-30 | 0 | 5 | 0 | 8 | 492 | 0 |
| <i>Asterocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> fine | 0 | 73 | 41 | 173 | 0 | 2 |
| autotrophic ciliate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 6024 | 21 | 0 | 0 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| centric >30 | 0 | 0 | 0 | 16 | 123 | 0 |
| centric 10-15 | 548 | 0 | 1029 | 553 | 0 | 0 |
| centric 15-20 | 17524 | 355 | 882 | 79 | 246 | 0 |
| centric 20-30 | 3833 | 39 | 0 | 0 | 123 | 18 |
| centric 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Centritractus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cerataulina bicornis</i> | 4381 | 0 | 0 | 0 | 0 | 5 |
| <i>Cerataulina</i> resting spore | 38 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 869 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 1643 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 2738 | 0 | 0 | 316 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 7667 | 0 | 0 | 632 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 2190 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 1643 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 226 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 5 | 0 | 79 | 0 | 0 |
| chlorophyte coccoid 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 237 | 0 | 0 |
| chlorophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte flagellated colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chroococcus</i> | 0 | 21 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Closterium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> >80 | 0 | 0 | 0 | 0 | 68 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 17 | 0 |
| <i>Coscinodiscus</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus granii</i> | 0 | 0 | 0 | 0 | 861 | 0 |
| <i>Cosmarium</i> | 0 | 39 | 0 | 0 | 0 | 0 |
| cryptomonad auto 10-15 | 0 | 0 | 0 | 0 | 369 | 804 |
| cryptomonad auto 15-20 | 0 | 0 | 147 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 8 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 0 | 516 | 0 | 0 | 0 |
| cyanobacteria filament | 155521 | 6632 | 7498 | 1264 | 0 | 0 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 10-15 | 1095 | 0 | 147 | 158 | 0 | 0 |
| <i>Cyclotella</i> 15-20 | 1643 | 0 | 1029 | 0 | 17 | 0 |
| <i>Cyclotella</i> 20-30 | 3833 | 0 | 147 | 0 | 0 | 124 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 79 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 20-30 | 0 | 0 | 0 | 8 | 0 | 0 |
| <i>Desmodesmus</i> | 0 | 39 | 83 | 316 | 0 | 0 |
| <i>Diacanthos belenophorus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dinophysis ovum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 0 | 5 | 0 | 0 | 0 | 0 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 17 | 0 |
| <i>Entomoneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena acus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto >30 | 0 | 0 | 0 | 0 | 0 | 124 |
| euglenoid auto 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 20-30 | 38 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 79 | 0 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 1733 |
| <i>Golenkinia radiata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 75 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 124 |
| <i>Gonyaulax spinifera</i> | 38 | 0 | 0 | 0 | 0 | 18 |
| <i>Guinardia flaccida</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 0 | 0 | 4116 | 0 | 34 | 1609 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Haptophyte | 1095 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 0 | 0 | 0 | 948 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 0 | 395 | 984 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 492 | 0 |
| <i>Mallomonas</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira varians</i> | 0 | 0 | 0 | 711 | 0 | 0 |
| <i>Merismopedia</i> | 0 | 0 | 0 | 754 | 0 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 0 | 147 | 0 | 123 | 62 |
| <i>Microcystis</i> | 0 | 0 | 0 | 4330 | 0 | 0 |
| <i>Microspora</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 0 | 0 | 0 | 0 | 124 |
| naked auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 20-30 | 0 | 16 | 0 | 0 | 0 | 0 |
| naked auto dino 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 548 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> 20-30 | 548 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula lanceolate</i> >30 | 38 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia lanceolate</i> >30 | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Nitzschia</i> ovoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 17 | 0 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 79 | 0 | 0 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pediastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | 1 | 0 |
| pennate 10-20 | 548 | 39 | 147 | 711 | 0 | 0 |
| pennate 20-40 | 38 | 16 | 0 | 31 | 0 | 0 |
| pennate 40-80 | 548 | 0 | 0 | 8 | 0 | 0 |
| <i>Phacus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Phaeophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|---------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| phytoflagellate 10-15 | 548 | 10 | 294 | 79 | 1476 | 0 |
| phytoflagellate 15-20 | 0 | 21 | 0 | 79 | 123 | 0 |
| phytoflagellate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 62 |
| <i>Planktolyngbya</i> | 4928 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktothrix</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Platydorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum micans</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 0 | 0 | 0 | 1107 | 124 |
| <i>Pseudo-nitzschia brasiliiana</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 0 | 0 | 0 | 0 | 0 | 18 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Radiococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 75 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scrippsiella</i> | 0 | 0 | 0 | 0 | 123 | 248 |
| silicoflagellate | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 922614 | 29876 | 101746 | 164283 | 186275 | 6188 |
| <i>Skeletonema tropicum</i> | 0 | 10262 | 0 | 0 | 0 | 0 |
| <i>Snowella</i> | 0 | 0 | 0 | 1580 | 0 | 0 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Spirulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Strombidium</i> | 548 | 0 | 0 | 79 | 0 | 0 |
| <i>Surirella</i> | 38 | 0 | 0 | 0 | 0 | 0 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | 1 | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 0 | 5 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 62 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|---|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| <i>Actinastrum</i> | 0 | 2592 | 462 | 0 | 0 | 1987 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Amphora</i> 20-30 | 0 | 0 | 0 | 0 | 6787 | 0 |
| <i>Anabaena</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apedinella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 407 | 0 | 0 | 0 | 0 |
| armored auto dino >30 | 0 | 0 | 0 | 1926 | 0 | 0 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino 20-30 | 0 | 324 | 0 | 0 | 0 | 248 |
| <i>Asterocapsa</i> | 0 | 0 | 1365 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> fine | 0 | 379 | 2622 | 3853 | 0 | 178150 |
| autotrophic ciliate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 249 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| centric >30 | 0 | 0 | 807 | 0 | 866 | 0 |
| centric 10-15 | 0 | 972 | 2622 | 963 | 0 | 0 |
| centric 15-20 | 124 | 0 | 1412 | 0 | 289 | 41 |
| centric 20-30 | 0 | 0 | 403 | 0 | 0 | 81 |
| centric 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Centritractus</i> | 0 | 324 | 0 | 0 | 0 | 20 |
| <i>Cerataulina bicornis</i> | 166 | 0 | 0 | 0 | 0 | 0 |
| <i>Cerataulina</i> resting spore | 0 | 0 | 202 | 0 | 0 | 0 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 41 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 55 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 0 | 0 | 40 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 663 | 0 | 0 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 41 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 972 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 15-20 | 0 | 0 | 40 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 5-10 | 0 | 0 | 3227 | 21782 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 3240 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 0 | 0 | 162 |
| chlorophyte filamentous | 0 | 0 | 0 | 963 | 0 | 0 |
| chlorophyte flagellated colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chroococcus</i> | 0 | 1296 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Closterium</i> | 0 | 70 | 0 | 963 | 0 | 0 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> >80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 40-60 | 41 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus granii</i> | 41 | 0 | 0 | 0 | 0 | 0 |
| <i>Cosmarium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 26242 | 11900 | 0 | 18099 | 0 |
| cyanobacteria filament | 0 | 0 | 8107 | 50825 | 2262433 | 10431 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 10-15 | 0 | 1620 | 202 | 963 | 0 | 0 |
| <i>Cyclotella</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 20-30 | 124 | 324 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 20 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 10-15 | 41 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 20-30 | 41 | 56 | 202 | 0 | 0 | 0 |
| <i>Desmodesmus</i> | 0 | 1296 | 807 | 7705 | 13575 | 41 |
| <i>Diacanthos belenophorus</i> | 0 | 14 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dinophysis ovum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 41 | 0 | 0 | 963 | 0 | 20 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Entomoneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 6480 | 0 | 0 | 0 | 487 |
| <i>Euglena acus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto >30 | 5 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 20-30 | 5 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 0 | 0 | 0 | 289 | 0 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 58086 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 7261 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 72607 | 11312 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 161 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Golenkinia radiata</i> | 0 | 324 | 20 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax spinifera</i> | 290 | 0 | 0 | 0 | 0 | 0 |
| <i>Guinardia flaccida</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 1408 | 14 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 289 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Haptophyte | 0 | 70 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 0 | 1242 |
| <i>Mallomonas</i> | 0 | 1620 | 202 | 0 | 2262 | 745 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 0 | 0 | 20 |
| <i>Melosira varians</i> | 0 | 9395 | 149457 | 0 | 15837 | 0 |
| <i>Merismopedia</i> | 0 | 0 | 12707 | 0 | 0 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Microcystis</i> | 0 | 48920 | 10488 | 0 | 0 | 0 |
| <i>Microspora</i> | 0 | 28 | 0 | 0 | 0 | 0 |
| naked auto dino >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 648 | 0 | 0 | 289 | 0 |
| naked auto dino 15-20 | 0 | 324 | 0 | 0 | 0 | 0 |
| naked auto dino 20-30 | 0 | 0 | 20 | 0 | 0 | 0 |
| naked auto dino 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula lanceolate</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> >30 | 5 | 0 | 20 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 169 | 40 | 0 | 0 | 0 |
| <i>Nitzschia lanceolate</i> >30 | 0 | 0 | 202 | 1926 | 0 | 20 |
| <i>Nitzschia</i> ovoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 223 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 0 | 20 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 0 | 4617 | 689 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 20 |
| <i>Pediastrum</i> | 0 | 0 | 1614 | 2889 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate 10-20 | 0 | 0 | 0 | 36304 | 577 | 0 |
| pennate 20-40 | 0 | 0 | 0 | 0 | 0 | 101 |
| pennate 40-80 | 0 | 0 | 40 | 0 | 0 | 0 |
| <i>Phacus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Phaeophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|---------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| phytoflagellate 10-15 | 0 | 324 | 0 | 0 | 0 | 20 |
| phytoflagellate 15-20 | 0 | 324 | 0 | 0 | 0 | 0 |
| phytoflagellate 20-30 | 0 | 2268 | 20 | 0 | 0 | 20 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktolyngbya</i> | 0 | 17495 | 27632 | 0 | 269230 | 446 |
| <i>Planktothrix</i> | 0 | 0 | 0 | 0 | 0 | 1744 |
| <i>Platydorina</i> | 0 | 13207 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 28 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum micans</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia brasiliiana</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 5508 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Radiococcus</i> | 0 | 913 | 60 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 462 | 0 | 0 | 0 |
| <i>Scrippsiella</i> | 0 | 0 | 0 | 963 | 0 | 0 |
| silicoflagellate | 0 | 28 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 41 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 14333 | 0 | 6656 | 0 | 22624 | 36259 |
| <i>Skeletonema tropicum</i> | 0 | 0 | 0 | 101650 | 0 | 0 |
| <i>Snowella</i> | 0 | 20410 | 5244 | 0 | 0 | 466 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 0 | 5215 |
| <i>Spirulina</i> | 0 | 0 | 0 | 0 | 10100 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Strombidium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Surirella</i> | 0 | 0 | 0 | 0 | 0 | 248 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 28 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 1296 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 0 | 1296 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 20-40 | 124 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|---|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| <i>Actinastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 128 | 0 | 0 |
| <i>Amphora</i> 20-30 | 0 | 41 | 0 | 0 | 0 | 0 |
| <i>Anabaena</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apedinella</i> | 0 | 0 | 165 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino >30 | 0 | 0 | 30 | 64 | 0 | 0 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 1017 |
| armored auto dino 20-30 | 0 | 12 | 330 | 128 | 638 | 1017 |
| <i>Asterocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> fine | 0 | 0 | 0 | 19954 | 8300 | 0 |
| autotrophic ciliate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 0 | 0 | 119 | 64 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | 0 | 678 |
| centric >30 | 19 | 0 | 0 | 443 | 30646 | 678 |
| centric 10-15 | 137 | 0 | 661 | 1774 | 9577 | 0 |
| centric 15-20 | 0 | 290 | 1156 | 0 | 7023 | 0 |
| centric 20-30 | 38 | 0 | 330 | 192 | 3192 | 1017 |
| centric 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Centritractus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cerataulina bicornis</i> | 22726 | 83 | 69 | 0 | 0 | 0 |
| <i>Cerataulina</i> resting spore | 137 | 207 | 0 | 443 | 0 | 339 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 0 | 17 | 991 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 0 | 0 | 2973 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 0 | 0 | 1156 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 0 | 0 | 330 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 0 | 23 | 30 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 0 | 0 | 0 | 638 | 0 |
| chlorophyte coccoid 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 0 | 0 | 5321 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 15-20 | 0 | 17 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte flagellated colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chroococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Closterium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 5083 |
| <i>Coscinodiscus</i> >80 | 958 | 0 | 50 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 40-60 | 0 | 0 | 59 | 887 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus granii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cosmarium</i> | 0 | 0 | 0 | 0 | 0 | 678 |
| cryptomonad auto 10-15 | 411 | 41 | 330 | 0 | 0 | 0 |
| cryptomonad auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria filament | 4655 | 983 | 3303 | 183132 | 47246 | 128757 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 10-15 | 0 | 0 | 0 | 0 | 2554 | 187 |
| <i>Cyclotella</i> 15-20 | 0 | 0 | 826 | 0 | 0 | 0 |
| <i>Cyclotella</i> 20-30 | 0 | 41 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 3104 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 0 | 0 | 0 | 140 |
| <i>Cylindrotheca</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 339 |
| <i>Cylindrotheca</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 339 |
| <i>Desmodesmus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diacanthos belenophorus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dinophysis ovum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Entomoneis</i> | 0 | 47 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena acus</i> | 0 | 12 | 165 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 90 | 0 |
| euglenoid auto >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 10-15 | 0 | 0 | 0 | 0 | 0 | 47 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 1774 | 0 | 0 |
| euglenoid auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 12 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 447 | 0 | 0 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|---------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| phytoflagellate 10-15 | 0 | 41 | 1486 | 0 | 0 | 339 |
| phytoflagellate 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| phytoflagellate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktolyngbya</i> | 0 | 157 | 0 | 4878 | 0 | 0 |
| <i>Planktothrix</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Platydorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum micans</i> | 0 | 6 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 0 | 1486 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia brasiliiana</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Radiococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 137 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scrippsiella</i> | 0 | 0 | 109 | 0 | 0 | 0 |
| silicoflagellate | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 373 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 2601 | 2072 | 14203 | 32813 | 144931 | 1690646 |
| <i>Skeletonema tropicum</i> | 0 | 0 | 4459 | 0 | 0 | 0 |
| <i>Snowella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Spirulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Strombidium</i> | 137 | 12 | 20 | 0 | 0 | 93 |
| <i>Surirella</i> | 0 | 0 | 0 | 64 | 638 | 0 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 373 | 0 | 0 | 0 | 0 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 67767 |

Phytoplankton Appendix 1. Number of cells per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Golenkinia radiata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax spinifera</i> | 0 | 0 | 165 | 0 | 0 | 0 |
| <i>Guinardia flaccida</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 137 | 41 | 1156 | 443 | 0 | 0 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 6 | 0 | 0 | 0 | 0 |
| Haptophyte | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 59 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 0 | 280 |
| <i>Mallomonas</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 128 | 179 | 0 |
| <i>Melosira varians</i> | 0 | 0 | 0 | 0 | 2598 | 0 |
| <i>Merismopedia</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 17 | 0 | 64 | 0 | 0 |
| <i>Microcystis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Microspora</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 0 | 0 | 443 | 0 | 0 |
| naked auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 0 | 0 | 0 | 0 | 0 | 339 |
| <i>Navicula</i> 20-30 | 0 | 0 | 0 | 0 | 638 | 0 |
| <i>Navicula lanceolate</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> >30 | 0 | 0 | 10 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> 20-30 | 0 | 41 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia lanceolate</i> >30 | 0 | 0 | 0 | 887 | 0 | 0 |
| <i>Nitzschia</i> ovoid >30 | 0 | 6 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 90 | 47 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pediastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate 10-20 | 274 | 83 | 0 | 1330 | 0 | 0 |
| pennate 20-40 | 137 | 83 | 165 | 887 | 638 | 0 |
| pennate 40-80 | 0 | 0 | 10 | 383 | 0 | 0 |
| <i>Phacus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Phaeophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |

Appendix C

Phytoplankton abundance (cell volume: μm^3) for selected samples

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station species\month | 7D May_2008 | 6D June_2008 | 7U July_2008 | 5U Aug_2008 | 5D Sept_2008 | 6D Oct_2008 |
|---|----------------|-----------------|-----------------|----------------|-----------------|----------------|
| <i>Actinastrum</i> | 0 | 0 | 1777980 | 0 | 0 | 19742 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 285776 | 124898 | 0 |
| <i>Amphora</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Anabaena</i> | 0 | 69515 | 462437 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 9426 | 13851 | 0 | 0 | 5419 |
| <i>Apedinella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 0 | 39003 | 19052 | 0 | 0 |
| armored auto dino >30 | 4065981 | 1047845 | 0 | 0 | 0 | 88839 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 732309 | 0 |
| armored auto dino 20-30 | 0 | 1870533 | 509201 | 0 | 0 | 132200 |
| <i>Asterocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | 0 | 0 | 0 | 97980 | 115291 | 0 |
| <i>Aulacoseira</i> fine | 0 | 0 | 31176 | 34837 | 85934 | 17613 |
| autotrophic ciliate 10-15 | 0 | 0 | 2916 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 54213 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 19276 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| centric >30 | 72284 | 0 | 0 | 11975 | 8006 | 0 |
| centric 10-15 | 0 | 2887 | 0 | 0 | 60848 | 0 |
| centric 15-20 | 683085 | 13856 | 0 | 29394 | 1068 | 0 |
| centric 20-30 | 2081782 | 1473 | 0 | 0 | 0 | 4230 |
| centric 5-10 | | 0 | 0 | 0 | 1601 | 0 |
| <i>Centritractus</i> | 0 | 0 | 0 | 0 | 0 | 4065 |
| <i>Cerataulina bicornis</i> | 25299439 | 810564 | 0 | 0 | 0 | 0 |
| <i>Cerataulina</i> resting spore | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 0 | 73898 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 24894 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 23898 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 231309 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 304700 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 6024 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 0 | 18851 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 17924 | 346395 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 0 | 18851 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 0 | 124702 | 0 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 578273 | 13856 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 36949 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 7069 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 2887 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 15-20 | 0 | 64660 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 44067 |
| chlorophyte coccoid 5-10 | 0 | 4124 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 7D | 6D | 7U | 5U | 5D | 6D |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2008 | June_2008 | July_2008 | Aug_2008 | Sept_2008 | Oct_2008 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 125391 | 26128 | 0 | 14101 |
| chlorophyte coccoid colonial 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte filamentous | 0 | 0 | 187053 | 0 | 0 | 2820 |
| chlorophyte flagellated colonial | 0 | 0 | 4010 | 8887 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 8540 | 0 |
| <i>Chroococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 6005 | 0 |
| <i>Closterium</i> | 0 | 0 | 811863 | 319936 | 0 | 601548 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> >80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 15588 | 0 | 0 | 0 | 5288 |
| <i>Coscinodiscus granii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cosmarium</i> | 0 | 0 | 0 | 1058815 | 0 | 0 |
| cryptomonad auto 10-15 | | 589 | 0 | 0 | | 0 |
| cryptomonad auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 5196 | 1461 | 0 | 0 | 11177 |
| cyanobacteria filament | 0 | 2526 | 14126 | 34021 | | 1619 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 35548 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 135373 |
| <i>Cyclotella</i> 10-15 | 0 | 138558 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 15-20 | 0 | 0 | 0 | 52256 | 0 | 146323 |
| <i>Cyclotella</i> 20-30 | 12047 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 29394 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 0 | 0 | 0 | 2820 |
| <i>Cylindrotheca</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 20-30 | 13443 | 1473 | 0 | 0 | 0 | 0 |
| <i>Desmodesmus</i> | 0 | 0 | 5845 | 209025 | 1601 | 6346 |
| <i>Diacanthos belenophorus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 9518 |
| <i>Dinophysis ovum</i> | 0 | 1298981 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Entomoneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 0 | 10392 | 0 | 0 | 0 |
| <i>Euglena acus</i> | 0 | 18851 | 0 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto >30 | 53107 | 0 | 0 | 391032 | 0 | 3294968 |
| euglenoid auto 10-15 | 0 | 25980 | 0 | 0 | 0 | 0 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 0 | 0 | 381034 | 12009 | 44419 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 7D | 6D | 7U | 5U | 5D | 6D |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2008 | June_2008 | July_2008 | Aug_2008 | Sept_2008 | Oct_2008 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Golenkinia radiata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 0 | 3670632 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax spinifera</i> | 0 | 1154650 | 0 | 0 | 0 | 0 |
| <i>Guinardia flaccida</i> | 5990435 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 0 | 4752539 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Haptophyte | 0 | 0 | 45564 | 568774 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 8457241 | 42416 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Mallomonas</i> | 0 | 0 | 113661 | 39992 | 0 | 164613 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira varians</i> | 0 | 7953 | 41567 | 255157 | 0 | 249290 |
| <i>Merismopedia</i> | 0 | 2209 | 0 | 0 | 38430 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Microcystis</i> | 0 | 0 | 15537 | 0 | 0 | 0 |
| <i>Microspora</i> | 0 | 0 | 0 | 0 | 0 | 86710 |
| naked auto dino >30 | 0 | 1501045 | 1646458 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 0 | 51959 | 0 | 0 | 0 |
| naked auto dino 15-20 | 0 | 101032 | 2583673 | 0 | 38430 | 0 |
| naked auto dino 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 5-10 | 19276 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> lanceolate >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> capitata >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> capitata 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> lanceolate >30 | 0 | 74227 | 0 | 161667 | 21350 | 25742 |
| <i>Nitzschia</i> ovoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 59988 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 0 | 0 | 5641 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pediastrum</i> | 0 | 0 | 118101 | 5555482 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | 144113 | 0 |
| pennate 10-20 | 0 | 209569 | 0 | 0 | 0 | 0 |
| pennate 20-40 | 119269 | 15906 | 0 | 6665 | 12009 | 75871 |
| pennate 40-80 | 0 | 0 | 0 | 533448 | 0 | 11281 |
| <i>Phacus</i> | 0 | 0 | 0 | 870936 | 1067507 | 0 |
| Phaeophyte filamentous | 0 | 0 | 172115 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 7D | 6D | 7U | 5U | 5D | 6D |
|---------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2008 | June_2008 | July_2008 | Aug_2008 | Sept_2008 | Oct_2008 |
| phytoflagellate 10-15 | 0 | 0 | 2598 | 58788 | 0 | 108387 |
| phytoflagellate 15-20 | 0 | 0 | 707295 | 0 | 0 | 365806 |
| phytoflagellate 20-30 | 0 | 1178 | 3787829 | 108867 | 0 | 0 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktolyngbya</i> | 0 | 0 | 145161 | 86549 | 0 | 6346 |
| <i>Planktothrix</i> | 0 | 13105275 | 2127731 | 27078 | 0 | 9166 |
| <i>Platydorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 0 | 0 | 81650 | 0 | 0 |
| <i>Prorocentrum micans</i> | 4626183 | 11971409 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 1163887 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia brasiliiana</i> | 26553 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 94836753 | 0 | 0 | 60966 | 0 | 0 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 16922 |
| <i>Radiococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 0 | 30483 | 0 | 5024 |
| <i>Scrippsiella</i> | 0 | 222682 | 0 | 0 | 0 | 0 |
| silicoflagellate | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 404791 | 64949 | 0 | 382123 | 0 | 113806 |
| <i>Skeletonema tropicum</i> | 1561337 | 0 | 0 | 71097 | 0 | 219484 |
| <i>Snowella</i> | 0 | 10392 | 0 | 0 | 0 | 0 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 4611628 | 0 |
| <i>Spirulina</i> | 0 | 0 | 4055 | 0 | 0 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 435543 | 5641 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 72902 | 53323 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 2820 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 138861 | 0 | 0 |
| <i>Strombidium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Surirella</i> | 0 | 0 | 0 | 0 | 716303 | 0 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 714167 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 337326 | 3314 | 0 | 0 | 0 | 32516 |
| <i>Thalassiosira</i> 20-40 | 0 | 5302 | 535830 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|---|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| <i>Actinastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 0 | 5824 | 0 |
| <i>Amphora</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Anabaena</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apedinella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 0 | 0 | 1939 | 0 | 0 |
| armored auto dino >30 | 27525874 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino 20-30 | 0 | 31822 | 0 | 24686 | 869684 | 0 |
| <i>Asterocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> fine | 0 | 14256 | 22276 | 50916 | 0 | |
| autotrophic ciliate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 6061606 | 85538 | 0 | 0 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | | 0 |
| centric >30 | 0 | 0 | 0 | 27772 | 15099 | 0 |
| centric 10-15 | 15120 | 0 | 25258 | 20354 | 0 | 0 |
| centric 15-20 | 645138 | 549355 | 32475 | 3877 | 10569 | 0 |
| centric 20-30 | 188165 | 2422 | 0 | 0 | 6794 | 4339 |
| centric 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Centritractus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cerataulina bicornis</i> | 48734774 | 0 | 0 | 0 | 0 | |
| <i>Cerataulina</i> resting spore | 266613 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 63971 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 645138 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 3150086 | 0 | 0 | 3877 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 4704129 | 0 | 0 | 46524 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 5927202 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 1008028 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 599879 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 2037 | 0 | 60579 | 0 | 0 |
| chlorophyte coccoid 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 11631 | 0 | 0 |
| chlorophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte flagellated colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chroococcus</i> | 0 | 3437 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Closterium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> >80 | 0 | 0 | 0 | 0 | 63230 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 3328 | 0 |
| <i>Coscinodiscus</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus granii</i> | 0 | 0 | 0 | 0 | 137398 | 0 |
| <i>Cosmarium</i> | 0 | 8720 | 0 | 0 | 0 | 0 |
| cryptomonad auto 10-15 | 0 | 0 | 0 | 0 | 36237 | 98721 |
| cryptomonad auto 15-20 | 0 | 0 | 66302 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 6172 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 0 | 3164 | 0 | 0 | 0 |
| cyanobacteria filament | 119283 | 20346 | 5751 | 15508 | 0 | 0 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 10-15 | 302408 | 0 | 4510 | 19385 | 0 | 0 |
| <i>Cyclotella</i> 15-20 | 70562 | 0 | 606191 | 0 | 2496 | 0 |
| <i>Cyclotella</i> 20-30 | 3010642 | 0 | 259796 | 0 | 0 | 6835 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 36347 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 20-30 | 0 | 0 | 0 | 1929 | 0 | 0 |
| <i>Desmodesmus</i> | 0 | 969 | 12150 | 38770 | 0 | 0 |
| <i>Diacanthos belenophorus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dinophysis ovum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 0 | 24440 | 0 | 0 | 0 | 0 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 159738 | 0 |
| <i>Entomoneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena acus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto >30 | 0 | 0 | 0 | 0 | 0 | 546765 |
| euglenoid auto 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 20-30 | 29624 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|------------------------------------|----------|----------|-----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 8723 | 0 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 85052 |
| <i>Golenkinia radiata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 3299336 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 334894 |
| <i>Gonyaulax spinifera</i> | 947957 | 0 | 0 | 0 | 0 | 218706 |
| <i>Guinardia flaccida</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 0 | 0 | 222270085 | 0 | 1830331 | 44424654 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Haptophyte | 8944565 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 0 | 0 | 0 | 20354 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 0 | 26655 | 42276 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 6039 | 0 |
| <i>Mallomonas</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira varians</i> | 0 | 0 | 0 | 157020 | 0 | 0 |
| <i>Merismopedia</i> | 0 | 0 | 0 | 4629 | 0 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 0 | 194847 | 0 | 905921 | 17086 |
| <i>Microcystis</i> | 0 | 0 | 0 | 26567 | 0 | 0 |
| <i>Microspora</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 0 | 0 | 0 | 0 | 27338 |
| naked auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 20-30 | 0 | 12220 | 0 | 0 | 0 | 0 |
| naked auto dino 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 913945 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> 20-30 | 37801 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula lanceolate</i> >30 | 51841 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia lanceolate</i> >30 | 0 | 0 | 0 | 0 | | 0 |
| <i>Nitzschia</i> ovoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 1060760 | 0 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 30858 | 0 | 0 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pediastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | | 0 |
| pennate 10-20 | 20161 | 15260 | 5412 | 26170 | 0 | 0 |
| pennate 20-40 | 16663 | 18330 | 0 | 37030 | 0 | 0 |
| pennate 40-80 | 210006 | 0 | 0 | 27772 | 0 | 0 |
| <i>Phacus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Phaeophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 7D | 5U | 6U | 5D | 6D |
|---------------------------------------|-----------|----------|----------|----------|----------|----------|
| species\month | Nov_2008 | Dec_2008 | Jan_2009 | Feb_2009 | Mar_2009 | Apr_2009 |
| phytoflagellate 10-15 | 20161 | 4073 | 64949 | 7754 | 36237 | 0 |
| phytoflagellate 15-20 | 0 | 16293 | 0 | 2908 | 33972 | 0 |
| phytoflagellate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 3038 |
| <i>Planktolyngbya</i> | 30241 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktothrix</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Platydorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum micans</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 0 | 0 | 0 | 3478738 | 191368 |
| <i>Pseudo-nitzschia brasiliiana</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 0 | 0 | 0 | 0 | 0 | 52073 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Radiococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 214772 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scrippsiella</i> | 0 | 0 | 0 | 0 | 672647 | 972027 |
| silicoflagellate | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 152849240 | 8799292 | 19665693 | 18144497 | 30860211 | 593278 |
| <i>Skeletonema tropicum</i> | 0 | 7870715 | 0 | 0 | 0 | 0 |
| <i>Snowella</i> | 0 | 0 | 0 | 77541 | 0 | 0 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Spirulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Strombidium</i> | 11357111 | 0 | 0 | 2564655 | 0 | 0 |
| <i>Surirella</i> | 11464360 | 0 | 0 | 0 | 0 | 0 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 0 | 891 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 7594 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|---|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| <i>Actinastrum</i> | 0 | 111322 | 90622 | 0 | 0 | 60954 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Amphora</i> 20-30 | 0 | 0 | 0 | 0 | 2998534 | 0 |
| <i>Anabaena</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apedinella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 20001 | 0 | 0 | 0 | 0 |
| armored auto dino >30 | 0 | 0 | 0 | 23828538 | 0 | 0 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino 20-30 | 0 | 779253 | 0 | 0 | 0 | 672020 |
| <i>Asterocapsa</i> | 0 | 0 | 66982 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> fine | 0 | 93106 | 450484 | 945577 | 0 | 17489783 |
| autotrophic ciliate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 1464073 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| centric >30 | 0 | 0 | 84156 | 0 | 254977 | 0 |
| centric 10-15 | 0 | 214692 | 1287097 | 189115 | 0 | 0 |
| centric 15-20 | 5338 | 0 | 60642 | 0 | 42496 | 5971 |
| centric 20-30 | 0 | 0 | 27227 | 0 | 0 | 19903 |
| centric 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Centritractus</i> | 0 | 214692 | 0 | 0 | 0 | 31845 |
| <i>Cerataulina bicornis</i> | 3458873 | 0 | 0 | 0 | 0 | 0 |
| <i>Cerataulina</i> resting spore | 0 | 0 | 1504914 | 0 | 0 | 0 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 32535 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 48740 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 0 | 0 | 3940 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 249096 | 0 | 0 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 763 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 23855 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 15-20 | 0 | 0 | 23641 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 5-10 | 0 | 0 | 59404 | 1069229 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 9939 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 0 | 0 | 7961 |
| chlorophyte filamentous | 0 | 0 | 0 | 585076 | 0 | 0 |
| chlorophyte flagellated colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chroococcus</i> | 0 | 15903 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Closterium</i> | 0 | 341390 | 0 | 661904 | 0 | 0 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> >80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 40-60 | 5592 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus granii</i> | 7880 | 0 | 0 | 0 | 0 | 0 |
| <i>Cosmarium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 80510 | 73018 | 0 | 55528 | 0 |
| cyanobacteria filament | 0 | 0 | 198975 | 1871151 | 6941052 | 32001 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 10-15 | 0 | 636125 | 61261 | 472788 | 0 | 0 |
| <i>Cyclotella</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 20-30 | 219611 | 397578 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 6895 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 10-15 | 1271 | 0 | 0 | 0 | 0 | 0 |
| <i>Cylindrotheca</i> 20-30 | 2288 | 16552 | 11138 | 0 | 0 | 0 |
| <i>Desmodesmus</i> | 0 | 397578 | 66830 | 756462 | 166585 | 3981 |
| <i>Diacanthos belenophorus</i> | 0 | 5517 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dinophysis ovum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 48802 | 0 | 0 | 756462 | 0 | 44783 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Entomoneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 159031 | 0 | 0 | 0 | 71652 |
| <i>Euglena acus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto >30 | 202182 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 20-30 | 18052 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 0 | 0 | 0 | 254977 | 0 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 2138459 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 1960254 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 12028829 | 3470526 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 7880 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Golenkinia radiata</i> | 0 | 5964 | 7880 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax spinifera</i> | 1729436 | 0 | 0 | 0 | 0 | 0 |
| <i>Guinardia flaccida</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 83655515 | 297940 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 3187218 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Haptophyte | 0 | 1765571 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 0 | 15239 |
| <i>Mallomonas</i> | 0 | 1749343 | 138610 | 0 | 1776909 | 585160 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 0 | 0 | 95536 |
| <i>Melosira varians</i> | 0 | 1152976 | 18341137 | 0 | 2623718 | 0 |
| <i>Merismopedia</i> | 0 | 0 | 77968 | 0 | 0 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Microcystis</i> | 0 | 150086 | 8044 | 0 | 0 | 0 |
| <i>Microspora</i> | 0 | 60691 | 0 | 0 | 0 | 0 |
| naked auto dino >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 286256 | 0 | 0 | 1770677 | 0 |
| naked auto dino 15-20 | 0 | 11927 | 0 | 0 | 0 | 0 |
| naked auto dino 20-30 | 0 | 0 | 123128 | 0 | 0 | 0 |
| naked auto dino 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> lanceolate >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> capitata >30 | 2482 | 0 | 10835 | 0 | 0 | 0 |
| <i>Nitzschia</i> capitata 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 66209 | 15760 | 0 | 0 | 0 |
| <i>Nitzschia</i> lanceolate >30 | 0 | 0 | 412119 | 8321077 | 0 | 111459 |
| <i>Nitzschia</i> ovoid >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 591131 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 0 | 75633 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 0 | 226647 | 33836 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 3981 |
| <i>Pediastrum</i> | 0 | 0 | 79206 | 10637740 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate 10-20 | 0 | 0 | 0 | 12028829 | 382466 | 0 |
| pennate 20-40 | 0 | 0 | 0 | 0 | 0 | 24879 |
| pennate 40-80 | 0 | 0 | 252166 | 0 | 0 | 0 |
| <i>Phacus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Phaeophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 6U | 6U | 3U | 7U | 6U | 5U |
|---------------------------------------|----------|-----------|-----------|----------|-----------|----------|
| species\month | May_2009 | June_2009 | July_2009 | Aug_2009 | Sept_2009 | Oct_2009 |
| phytoflagellate 10-15 | 0 | 11927 | 0 | 0 | 0 | 15923 |
| phytoflagellate 15-20 | 0 | 13915 | 0 | 0 | 0 | 0 |
| phytoflagellate 20-30 | 0 | 4007586 | 15760 | 0 | 0 | 3981 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktolyngbya</i> | 0 | 161019 | 339101 | 0 | 1651970 | 21894 |
| <i>Planktothrix</i> | 0 | 0 | 0 | 0 | 0 | 342339 |
| <i>Platydorina</i> | 0 | 777954553 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 11035 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum micans</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia brasiliiana</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 439324 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Radiococcus</i> | 0 | 44829 | 79787 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 203900 | 0 | 0 | 0 |
| <i>Scrippsiella</i> | 0 | 0 | 0 | 5909856 | 0 | 0 |
| silicoflagellate | 0 | 62071 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 31772 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 2374544 | 0 | 326725 | 0 | 3748168 | 2669793 |
| <i>Skeletonema tropicum</i> | 0 | 0 | 0 | 29938420 | 0 | 0 |
| <i>Snowella</i> | 0 | 8015171 | 2059356 | 0 | 0 | 34333 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 0 | 16000 |
| <i>Spirulina</i> | 0 | 0 | 0 | 0 | 247895 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Strombidium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Surirella</i> | 0 | 0 | 0 | 0 | 0 | 51372173 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 75864 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 214692 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 0 | 572512 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 20-40 | 9150 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|---|-----------|----------|----------|----------|----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| <i>Actinastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Amphora</i> >30 | 0 | 0 | 0 | 1143520 | 0 | 0 |
| <i>Amphora</i> 20-30 | 0 | 69932 | 0 | 0 | 0 | 0 |
| <i>Anabaena</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apatococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Apedinella</i> | 0 | 0 | 3547 | 0 | 0 | 0 |
| <i>Aphanocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| armored auto dino >30 | 0 | 0 | 419915 | 677641 | 0 | 0 |
| armored auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 1684036 |
| armored auto dino 20-30 | 0 | 36536 | 1037711 | 784307 | 3917562 | 2444971 |
| <i>Asterocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> coarse | | 0 | 0 | 0 | 0 | 0 |
| <i>Aulacoseira</i> fine | 0 | 0 | 0 | 4407668 | 560211 | 0 |
| autotrophic ciliate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic ciliate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| autotrophic flagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Bacillaria paxillifer</i> | 0 | 0 | 134140 | 65882 | 0 | 0 |
| <i>Campylodiscus clypeus</i> | 0 | 0 | 0 | 0 | 0 | 77964634 |
| centric >30 | 9257 | 0 | 0 | 43533 | 2632602 | 58214 |
| centric 10-15 | 4200 | 0 | 20268 | 54416 | 352581 | 0 |
| centric 15-20 | 0 | 12461 | 49656 | 0 | 301652 | 0 |
| centric 20-30 | 7406 | 0 | 22295 | 37647 | 176290 | 68609 |
| centric 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Centritractus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cerataulina bicornis</i> | 110439503 | 897163 | 489901 | 0 | 0 | 0 |
| <i>Cerataulina</i> resting spore | 1088670 | 1557574 | 0 | 3308471 | 0 | 3368072 |
| <i>Chaetoceros</i> c.f. <i>subtilis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros compressus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros constrictus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros curvisetus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros karianus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lacinosus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros lorenzianus</i> | 0 | 41103 | 6128979 | 0 | 0 | 0 |
| <i>Chaetoceros minimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros seiracanthus</i> | 0 | 0 | 2052114 | 0 | 0 | 0 |
| <i>Chaetoceros simplex</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros</i> sp. | 0 | 0 | 1596088 | 0 | 0 | 0 |
| <i>Chaetoceros subtilis</i> | 0 | 0 | 16214 | 0 | 0 | 0 |
| <i>C. subtilis</i> var. <i>abnormis</i> f. <i>simplex</i> | 0 | 2854 | 4374 | 0 | 0 | 0 |
| <i>Chaetoceros tenuissimus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chaetoceros wighamii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 10-15 | 0 | 0 | 0 | 0 | 250724 | 0 |
| chlorophyte coccoid 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial <5 | 0 | 0 | 0 | 110192 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| chlorophyte coccoid colonial 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 15-20 | 0 | 1284 | 0 | 0 | 0 | 0 |
| chlorophyte coccoid colonial 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte flagellated colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| chlorophyte triangular | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chroococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Chrysochromulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Closterium</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coelastrum microporum</i> | 0 | 0 | 0 | 0 | 0 | 623717 |
| <i>Coscinodiscus</i> >80 | 235206 | 0 | 4704609 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus</i> 40-60 | 0 | 0 | 32077 | 5920423 | 0 | 0 |
| <i>Coscinodiscus</i> 60-80 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Coscinodiscus granii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cosmarium</i> | 0 | 0 | 0 | 0 | 0 | 83162 |
| cryptomonad auto 10-15 | 90722 | 5086 | 57003 | 0 | 0 | 0 |
| cryptomonad auto 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| cryptomonad auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria coccoid colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| cyanobacteria filament | 14280 | 3015 | 22801 | 561842 | 36237 | 98755 |
| <i>Cyanotetras</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 10-15 | 0 | 0 | 0 | 0 | 376086 | 164967 |
| <i>Cyclotella</i> 15-20 | 0 | 0 | 403456 | 0 | 0 | 0 |
| <i>Cyclotella</i> 20-30 | 0 | 32550 | 0 | 0 | 0 | 0 |
| <i>Cyclotella</i> 5-10 | 0 | 0 | 0 | 514228 | 0 | 0 |
| <i>Cylindrotheca</i> >30 | 0 | 0 | 0 | 0 | 0 | 54989 |
| <i>Cylindrotheca</i> 10-15 | 0 | 0 | 0 | 0 | 0 | 10395 |
| <i>Cylindrotheca</i> 20-30 | 0 | 0 | 0 | 0 | 0 | 66530 |
| <i>Desmodesmus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diacanthos belenophorus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dicellula</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Dinophysis ovum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Diploneis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Ditylum brightwellii</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Entomoneis</i> | 0 | 164411 | 0 | 0 | 0 | 0 |
| <i>Eudorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Euglena acus</i> | 0 | 20551 | 456025 | 0 | 0 | 0 |
| <i>Euglena oblonga</i> | 0 | 0 | 0 | 0 | 1266412 | 0 |
| euglenoid auto >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| euglenoid auto 10-15 | 0 | 0 | 0 | 0 | 0 | 36659 |
| euglenoid auto 15-20 | 0 | 0 | 0 | 685637 | 0 | 0 |
| euglenoid auto 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 10-15 | 0 | 10276 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Fragilaria</i> 20-30 | 0 | 0 | 0 | 247057 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|------------------------------------|----------|----------|----------|----------|----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| <i>Fragilaria</i> 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gloeocapsopsis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Golenkinia radiata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax polygramma</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax</i> sp. | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gonyaulax spinifera</i> | 0 | 0 | 2979365 | 0 | 0 | 0 |
| <i>Guinardia flaccida</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gymnodinium sanguineum</i> | 32508890 | 1822807 | 71951664 | 22590657 | 0 | 0 |
| <i>Gyrosigma fasciolata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Gyrosigma/Pleurosigma</i> | 0 | 114175 | 0 | 0 | 0 | 0 |
| Haptophyte | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus danicus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> | 0 | 0 | 20413 | 0 | 0 | 0 |
| <i>Leptocylindrus minimus</i> thin | 0 | 0 | 0 | 0 | 0 | 5155 |
| <i>Mallomonas</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Melosira nummuloides</i> | 0 | 0 | 0 | 941169 | 4447832 | 0 |
| <i>Melosira varians</i> | 0 | 0 | 0 | 0 | 255041 | 0 |
| <i>Merismopedia</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Mesodinium rubrum</i> | 0 | 38534 | 0 | 112940 | 0 | 0 |
| <i>Microcystis</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Microspora</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 10-15 | 0 | 0 | 0 | 110192 | 0 | 0 |
| naked auto dino 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| naked auto dino 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Navicula</i> >30 | 0 | 0 | 0 | 0 | 0 | 280673 |
| <i>Navicula</i> 20-30 | 0 | 0 | 0 | 0 | 141032 | 0 |
| <i>Navicula lanceolate</i> >30 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> >30 | 0 | 0 | 75818 | 0 | 0 | 0 |
| <i>Nitzschia capitata</i> 20-30 | 0 | 76289 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> colonial | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia lanceolate</i> >30 | 0 | 0 | 0 | 8184114 | 0 | 0 |
| <i>Nitzschia</i> ovoid >30 | 0 | 34966 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> ovoid 10-15 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Nitzschia</i> sigmoid >30 | 0 | 0 | 0 | 0 | 131918 | 109978 |
| <i>Onychonema filiforme</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pandorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Parapedinella reticulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pediastrum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate >100 | 0 | 0 | 0 | 0 | 0 | 0 |
| pennate 10-20 | 6720 | 10172 | 0 | 40812 | 0 | 0 |
| pennate 20-40 | 83162 | 36555 | 136808 | 2133093 | 1692387 | 0 |
| pennate 40-80 | 0 | 0 | 3888 | 2032924 | 0 | 0 |
| <i>Phacus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| Phaeophyte filamentous | 0 | 0 | 0 | 0 | 0 | 0 |

Phytoplankton Appendix 2. Volume of cells (μm^3) per liter encountered in the 24 samples examined. The numbers following the species refer to cell sizes in micrometers.

| station | 5U | 6U | 6D | 7D | 4D | 4D |
|---------------------------------------|----------|----------|----------|----------|-----------|----------|
| species\month | Nov_2009 | Dec_2009 | Jan_2010 | Feb_2010 | Mar_2010 | Apr_2010 |
| phytoflagellate 10-15 | 0 | 16275 | 583712 | 0 | 0 | 259882 |
| phytoflagellate 15-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| phytoflagellate 20-30 | 0 | 0 | 0 | 0 | 0 | 0 |
| phytoflagellate 5-10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Planktolyngbya</i> | 0 | 5780 | 0 | 44893 | 0 | 0 |
| <i>Planktothrix</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Platydorina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Polyedriopsis spinulosa</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum micans</i> | 0 | 92481 | 0 | 0 | 0 | 0 |
| <i>Prorocentrum minimum</i> | 0 | 0 | 4669699 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia brasiliiana</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudo-nitzschia pungens</i> group | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Pseudostaurastrum lobulatum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Radiococcus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Rhizosolenia setigera</i> | 403211 | 0 | 0 | 0 | 0 | 0 |
| <i>Scenedesmus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Scrippsiella</i> | 0 | 0 | 668268 | 0 | 0 | 0 |
| silicoflagellate | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema auxospore</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Skeletonema grethae</i> | 0 | 36619 | 0 | 0 | 0 | 0 |
| <i>Skeletonema subsalsum</i> | 430932 | 343302 | 871515 | 2416055 | 16007157 | 82989299 |
| <i>Skeletonema tropicum</i> | 0 | 0 | 1751137 | 0 | 0 | 0 |
| <i>Snowella</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Spirogyra</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Spirulina</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Stanieria</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum anatinum</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurastrum</i> other | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Staurodesmus cuspidatus</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Strombidium</i> | 493934 | 71359 | 279944 | 0 | 0 | 2469920 |
| <i>Surirella</i> | 0 | 0 | 0 | 2117629 | 140922533 | 0 |
| <i>Tabularia tabulata</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Terpsinoe</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassionema nitzschioides</i> | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> <10 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 10-20 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 20-40 | 0 | 0 | 0 | 0 | 0 | 0 |
| <i>Thalassiosira</i> 40-60 | 0 | 43485 | 0 | 0 | 0 | 0 |
| <i>Trichodesmium</i> | 0 | 0 | 0 | 0 | 0 | 831623 |

Appendix D

Zooplankton Summary Catch Statistics

Table A1, page 1 of 5.

Plankton-net catch statistics (May 2008 through October 2009, n = 252 samples)

Organisms are listed in phylogenetic order.

| Taxon | Common Name | Number Collected | Collection Frequency | Kmu (km) | Su (psu) | Mean CPUE (No./10 ³ m ³) | Max CPUE (No./10 ³ m ³) |
|------------------------------------|-------------------------|------------------|----------------------|----------|----------|---|--|
| foraminiferans | foraminiferans | 4 | 1 | -3.6 | 36.3 | 0.17 | 43.13 |
| medusa, Bougainvillia sp. | hydromedusa | 626 | 22 | -5.4 | 35.9 | 31.52 | 2913.71 |
| Clytia sp. | hydromedusa | 14,653 | 22 | 39.1 | 10.9 | 925.40 | 109555.38 |
| medusa, Obelia sp. | hydromedusa | 548 | 22 | -3.3 | 30.9 | 27.65 | 2008.26 |
| medusa, Eutima sp. | hydromedusa | 1,110 | 37 | -3.8 | 29.6 | 53.27 | 3444.43 |
| Liriope tetraphylla | hydromedusa | 81 | 9 | 12.4 | 31.3 | 4.83 | 275.78 |
| medusa sp. a | hydromedusa | 16 | 2 | -3.9 | 34.1 | 0.84 | 200.08 |
| medusa sp. d | hydromedusa | 73 | 13 | 21.6 | 19.7 | 3.84 | 421.67 |
| medusa sp. e | hydromedusa | 248 | 26 | -2.4 | 31.4 | 12.37 | 464.26 |
| Craspedacusta sowerbii | hydromedusa | 52 | 3 | 32.7 | 10.0 | 2.72 | 496.25 |
| siphonophores | siphonophores | 33 | 4 | -5.5 | 35.9 | 1.69 | 306.79 |
| Aurelia aurita | moon jellyfish | 2 | 2 | 9.0 | 28.1 | 0.12 | 15.67 |
| Beroe ovata | sea walnut, ctenophore | 2 | 1 | 34.4 | 14.1 | 0.21 | 52.92 |
| Mnemiopsis mccradyi | comb jelly, ctenophore | 7,125 | 30 | 32.7 | 10.4 | 429.83 | 21091.19 |
| turbellarians | flatworms | 5 | 2 | 13.8 | 21.2 | 0.28 | 58.93 |
| nematodes | roundworms, threadworms | 72 | 15 | -2.4 | 27.0 | 3.87 | 307.43 |
| polychaetes | sand worms, tube worms | 1,935 | 127 | 6.6 | 26.6 | 114.57 | 4314.68 |
| oligochaetes | freshwater worms | 383 | 13 | 35.2 | 0.2 | 19.00 | 2941.47 |
| hirudinoideans | leeches | 7 | 6 | 31.6 | 2.7 | 0.31 | 23.82 |
| gastropods, prosobranch | snails | 947 | 69 | 20.4 | 13.3 | 47.50 | 2489.77 |
| gastropods, opisthobranch | sea slugs | 34 | 11 | 3.8 | 24.1 | 1.78 | 56.93 |
| Lolliguncula brevis juveniles | bay squid | 31 | 7 | 6.4 | 29.9 | 1.80 | 202.45 |
| pelecypods | clams, mussels, oysters | 119 | 27 | 21.9 | 13.8 | 7.83 | 559.92 |
| Limulus polyphemus larvae | horseshoe crab | 1 | 1 | 5.2 | 18.4 | 0.06 | 13.89 |
| acari | water mites | 29 | 15 | 35.8 | 1.0 | 1.39 | 65.03 |
| pycnogonids | sea spiders | 123 | 17 | 4.6 | 31.9 | 6.42 | 343.49 |
| cladocerans, unidentified | water fleas | 2 | 1 | 20.0 | 0.1 | 0.12 | 29.67 |
| Latona setifera | water flea | 43 | 11 | 36.5 | 0.4 | 1.93 | 156.34 |
| Latonopsis fasciculata | water flea | 2,745 | 10 | 40.6 | 7.2 | 135.48 | 33127.39 |
| Penilia avirostris | water flea | 15,270 | 43 | -4.3 | 31.8 | 799.11 | 74039.22 |
| Sida crystallina | water flea | 17 | 7 | 24.9 | 8.8 | 0.83 | 62.19 |
| Diaphanosoma brachyurum | water flea | 136 | 5 | 37.0 | 0.3 | 7.07 | 1312.85 |
| Bosmina sp. | water flea | 39 | 4 | 37.9 | 0.3 | 2.01 | 301.96 |
| Kurzia longirostris | water flea | 1 | 1 | 26.9 | 0.3 | 0.05 | 12.89 |
| Leydigia sp. | water flea | 22 | 9 | 29.3 | 0.2 | 1.09 | 68.15 |
| Ceriodaphnia sp. | water flea | 607 | 16 | 36.1 | 0.3 | 30.94 | 3859.79 |
| cladocerans, Daphnia spp. | water fleas | 139,852 | 45 | 31.0 | 0.2 | 6113.73 | 516915.93 |
| Simocephalus vetulus | water flea | 233 | 28 | 32.3 | 0.5 | 12.50 | 474.71 |
| Ilyocryptus sp. | water flea | 377 | 14 | 32.6 | 0.2 | 19.97 | 1729.43 |
| Psuedevadne tergestina | water flea | 898 | 13 | -5.3 | 28.8 | 42.44 | 7044.69 |
| Pleopsis polyphemoides | water flea | 2 | 1 | -3.6 | 32.1 | 0.10 | 26.00 |
| cirriped nauplius stage | barnacles | 1,721 | 46 | -3.6 | 31.2 | 83.59 | 4734.81 |
| cirriped cypris stage | barnacles | 128 | 13 | 19.9 | 12.3 | 6.55 | 1063.28 |
| branchiurans, Argulus spp. | fish lice | 25 | 23 | 24.4 | 5.8 | 1.22 | 29.73 |
| Acartia tonsa | copepod | 120,665 | 190 | 6.4 | 27.1 | 6500.92 | 143723.97 |
| Centropages velificatus | copepod | 1,470 | 42 | -4.0 | 34.2 | 73.46 | 3921.81 |
| Osphranticum labronectum | copepod | 3 | 1 | 26.9 | 0.1 | 0.18 | 45.41 |
| Diaptomus spp. | copepods | 4,465 | 53 | 33.6 | 0.3 | 216.58 | 7061.86 |
| Pseudodiaptomus coronatus | copepod | 3,098 | 87 | 14.7 | 21.6 | 173.35 | 22612.47 |
| Eucalanus sp. | copepod | 12 | 5 | -4.4 | 29.5 | 0.61 | 59.71 |
| paracalanids | copepods | 217 | 25 | -1.9 | 31.1 | 11.17 | 940.07 |
| Calanopia americana | copepod | 857 | 24 | 0.1 | 21.3 | 45.49 | 3118.93 |
| Labidocera aestiva | copepod | 43,595 | 91 | 1.4 | 31.5 | 2254.50 | 140044.39 |
| Labidocera acutifrons | copepod | 2 | 1 | -5.9 | 32.6 | 0.09 | 22.96 |
| Eurytemora affinis | copepod | 2 | 1 | 41.0 | 1.0 | 0.09 | 22.74 |
| Temora turbinata | copepod | 309 | 38 | -0.1 | 26.6 | 16.47 | 680.08 |
| Temora stylifera | copepod | 1 | 1 | -3.6 | 24.5 | 0.05 | 12.06 |
| Temora longicornis | copepod | 13 | 4 | -4.1 | 24.6 | 0.61 | 59.71 |
| unidentified freshwater cyclopoids | copepods | 67 | 22 | 35.0 | 1.0 | 3.46 | 113.34 |
| Cyclops spp. | copepods | 20 | 3 | 37.9 | 0.4 | 0.95 | 124.38 |
| Eucyclops speratus | copepod | 135 | 19 | -3.7 | 33.8 | 7.09 | 561.43 |
| Macrocyclus albidus | copepods | 18 | 6 | 13.6 | 8.0 | 0.98 | 75.68 |
| Mesocyclops edax | copepod | 21,278 | 79 | 35.6 | 0.5 | 987.02 | 24733.74 |

Table A1, page 2 of 5.

Plankton-net catch statistics (May 2008 through October 2009, n = 252 samples)

Organisms are listed in phylogenetic order.

| Taxon | Common Name | Number Collected | Collection Frequency | Kmu (km) | Su (psu) | Mean CPUE (No./10 ³ m ³) | Max CPUE (No./10 ³ m ³) |
|---|-------------------------|------------------|----------------------|----------|----------|---|--|
| Mesocyclops leuckarti | copepod | 8 | 1 | 41.0 | 0.1 | 0.69 | 173.50 |
| Oithona spp. | copepods | 544 | 32 | 3.9 | 28.5 | 30.06 | 1586.01 |
| Alteutha sp. | copepod | 67 | 10 | 13.9 | 24.2 | 3.50 | 256.48 |
| unidentified harpacticoids | copepods | 216 | 34 | -0.4 | 29.4 | 11.20 | 467.29 |
| Saphirella spp. | copepods | 6 | 5 | -4.0 | 22.7 | 0.35 | 34.06 |
| siphonostomatids | parasitic copepods | 164 | 48 | 3.7 | 27.8 | 8.17 | 204.87 |
| Monstrilla sp. | copepod | 252 | 35 | 0.0 | 32.1 | 13.56 | 591.35 |
| myodocopod sp. a | ostracod, seed shrimp | 5 | 4 | 20.2 | 11.6 | 0.23 | 24.30 |
| Parasterope pollex | ostracod, seed shrimp | 1,893 | 65 | 5.6 | 28.5 | 108.58 | 13615.23 |
| Sarsiella zostericola | ostracod, seed shrimp | 274 | 32 | 4.8 | 20.8 | 14.74 | 1951.48 |
| ostracods, podocopid | ostracods, seed shrimps | 29 | 12 | 34.0 | 3.7 | 1.63 | 86.75 |
| Squilla empusa larvae | mantis shrimp | 155 | 18 | 5.1 | 32.6 | 8.71 | 435.76 |
| mysid sp. A | opossum shrimps, mysids | 1 | 1 | 2.5 | 22.4 | 0.05 | 12.94 |
| (Heteromysis sp.) | opossum shrimp, mysid | 99 | 7 | 5.1 | 31.2 | 5.69 | 575.13 |
| Americamysis almyra | opossum shrimp, mysid | 28,362 | 138 | 29.7 | 5.4 | 1390.08 | 27446.24 |
| Americamysis bahia | opossum shrimp, mysid | 511 | 42 | 2.9 | 19.9 | 29.21 | 1737.06 |
| unidentified Americamysis juveniles | opossum shrimps, mysids | 36,014 | 180 | 29.2 | 6.5 | 1809.45 | 47257.92 |
| Americamysis stucki | opossum shrimp, mysid | 90 | 12 | -0.9 | 27.3 | 4.99 | 421.94 |
| Bowmaniella dissimilis | opossum shrimp, mysid | 5,208 | 132 | 17.8 | 11.3 | 256.70 | 4368.78 |
| Brasilomysis castroi | opossum shrimp, mysid | 1 | 1 | 10.6 | 16.3 | 0.06 | 14.33 |
| Mysidopsis furca | opossum shrimp, mysid | 21 | 5 | 3.8 | 26.9 | 1.00 | 88.47 |
| Mysidopsis mortenseni | opossum shrimp, mysid | 1 | 1 | -3.6 | 32.3 | 0.06 | 14.50 |
| Spelaeomysis sp. | opossum shrimp, mysid | 3 | 2 | 7.7 | 8.6 | 0.14 | 24.51 |
| Taphromysis bowmani | opossum shrimp, mysid | 23 | 13 | 31.5 | 4.5 | 1.12 | 65.03 |
| amphipods, gammaridean | amphipods | 412,464 | 235 | 8.6 | 11.1 | 20275.76 | 1431192.05 |
| amphipods, caprellid | skeleton shrimps | 416 | 51 | 5.2 | 28.7 | 23.82 | 825.35 |
| Lestrigonus bengalensis | hyperiid amphipod | 2 | 1 | -3.6 | 30.1 | 0.10 | 25.46 |
| isopod sp. a | isopod | 2 | 1 | -3.6 | 38.7 | 0.11 | 27.96 |
| Cyathura polita | isopod | 201 | 48 | 24.4 | 4.3 | 9.63 | 351.12 |
| Xenanthura brevitelson | isopod | 7 | 4 | 8.6 | 16.5 | 0.31 | 34.66 |
| Anopsilana jonesi | isopod | 58 | 14 | 32.7 | 13.6 | 3.65 | 312.21 |
| cymothoid sp. a (Lironeca) juveniles | isopod | 290 | 77 | 21.7 | 12.8 | 15.09 | 496.25 |
| Cassidinidea ovalis | isopod | 18 | 11 | 27.7 | 6.8 | 0.84 | 52.62 |
| Sphaeromatinae | isopods | 46 | 7 | 2.8 | 29.2 | 2.69 | 436.05 |
| Harrieta faxoni | isopod | 79 | 13 | 13.3 | 22.5 | 4.38 | 320.37 |
| Sphaeroma quadridentata | isopod | 3 | 2 | 9.8 | 27.3 | 0.16 | 30.33 |
| Sphaeroma terebrans | isopod | 2 | 1 | 30.2 | 5.1 | 0.10 | 24.74 |
| Gnathia sp. larvae | isopod | 1 | 1 | 7.7 | 23.4 | 0.05 | 12.21 |
| Munna reynoldsi | isopod | 79 | 21 | -0.1 | 31.8 | 3.88 | 204.87 |
| Edotia triloba | isopod | 8,889 | 130 | 24.5 | 10.1 | 449.71 | 19548.87 |
| Erichsonella attenuata | isopod | 100 | 38 | 6.7 | 22.3 | 5.08 | 188.01 |
| Hoplomachus propinquus | tanaid | 1 | 1 | 2.5 | 8.8 | 0.05 | 11.52 |
| Hargeria rapax | tanaid | 2,868 | 63 | 5.4 | 22.9 | 137.58 | 4875.23 |
| Sinelobus stanfordi | tanaid | 7 | 6 | 21.0 | 9.8 | 0.34 | 26.34 |
| Apseudes sp. | tanaid | 289 | 17 | 6.2 | 25.8 | 16.63 | 2099.47 |
| cumaceans | cumaceans | 167,177 | 178 | 13.1 | 22.4 | 9369.28 | 452677.51 |
| penaeid mysis larvae | penaeid shrimps | 7 | 2 | -4.3 | 11.6 | 0.44 | 75.78 |
| penaeid postlarvae | penaeid shrimps | 86 | 11 | -5.0 | 27.0 | 4.06 | 479.32 |
| penaeid metamorphs | penaeid shrimps | 649 | 60 | 2.2 | 17.7 | 32.83 | 971.68 |
| Farfantepenaeus duorarum juveniles | pink shrimp | 38 | 25 | 12.4 | 9.4 | 1.90 | 65.72 |
| Trachypenaeopsis mobilispinus juveniles | shrimps | 5 | 1 | -5.9 | 33.4 | 0.26 | 65.77 |
| Lucifer faxoni juveniles and adults | shrimp | 26,765 | 92 | -3.3 | 32.5 | 1361.73 | 32530.03 |
| Lucifer faxoni mysis | shrimp | 41 | 12 | -4.3 | 34.5 | 2.10 | 91.84 |
| shrimps, unidentified postlarvae | shrimps | 24 | 6 | -2.3 | 29.6 | 1.12 | 97.74 |
| decapod zoeae | crab larvae | 943,417 | 221 | 8.2 | 25.2 | 49480.46 | 818828.25 |
| decapod megalopae | post-zoea crab larvae | 10,308 | 142 | 4.8 | 21.0 | 510.37 | 15664.45 |
| decapod mysis | shrimp larvae | 286,668 | 182 | 7.0 | 30.4 | 16085.32 | 644584.99 |
| Leptocheila serratorbita postlarvae | comblaw shrimp | 4 | 1 | -5.9 | 36.5 | 0.19 | 48.87 |
| Palaemon floridanus postlarvae | Florida grass shrimp | 1,613 | 31 | 2.1 | 28.9 | 79.30 | 4465.37 |
| Periclimenes longicaudatus juveniles | longtail grass shrimp | 6 | 2 | 3.5 | 20.1 | 0.26 | 42.45 |
| Periclimenes spp. postlarvae | shrimps | 6 | 4 | -0.5 | 22.6 | 0.35 | 43.41 |
| Periclimenes spp. juveniles | shrimps | 46 | 12 | 0.6 | 20.8 | 2.42 | 189.45 |
| Palaemonetes paludosus juveniles | grass shrimp | 94 | 4 | 2.5 | 19.9 | 3.98 | 923.27 |

Table A1, page 3 of 5.

Plankton-net catch statistics (May 2008 through October 2009, n = 252 samples)

Organisms are listed in phylogenetic order.

| Taxon | Common Name | Number Collected | Collection Frequency | Kmu (km) | Su (psu) | Mean CPUE (No./10 ³ m ³) | Max CPUE (No./10 ³ m ³) |
|---|---------------------------|------------------|----------------------|----------|----------|---|--|
| Palaemonetes pugio juveniles | daggerblade grass shrimp | 33 | 14 | 6.7 | 15.4 | 1.64 | 95.73 |
| Palaemonetes pugio adults | daggerblade grass shrimp | 2 | 2 | 16.5 | 4.5 | 0.09 | 11.72 |
| Palaemonetes spp. postlarvae | grass shrimps | 6,204 | 108 | 11.0 | 29.4 | 375.65 | 64714.38 |
| Palaemonetes spp. juveniles | grass shrimps | 3 | 1 | 34.4 | 0.1 | 0.20 | 49.66 |
| Palaemonetes vulgaris juveniles | grass shrimp | 1 | 1 | 2.5 | 28.0 | 0.05 | 11.45 |
| alphaeid postlarvae | snapping shrimps | 12,097 | 86 | 6.5 | 30.2 | 680.57 | 23929.75 |
| alphaeid juveniles | snapping shrimps | 37 | 7 | -1.5 | 23.2 | 1.89 | 279.50 |
| Alpheus estuariensis juveniles | snapping shrimp | 4 | 3 | 1.9 | 18.4 | 0.19 | 22.88 |
| Hippolyte zostericola postlarvae | zostera shrimp | 4,650 | 88 | -0.7 | 28.2 | 233.86 | 11108.56 |
| Hippolyte zostericola juveniles | zostera shrimp | 99 | 23 | -2.6 | 29.4 | 5.07 | 152.67 |
| Hippolyte zostericola adults | zostera shrimp | 15 | 4 | -4.4 | 24.1 | 0.83 | 108.89 |
| Latreutes parvulus postlarvae | sargassum shrimp | 136 | 2 | 8.7 | 33.7 | 7.93 | 1293.23 |
| Tozeuma carolinense postlarvae | arrow shrimp | 952 | 35 | -1.2 | 33.4 | 47.73 | 2227.15 |
| Tozeuma carolinense juveniles | arrow shrimp | 82 | 12 | -4.7 | 25.4 | 4.26 | 352.41 |
| Ogyrides alphaerostriis postlarvae | estuarine longeye shrimp | 4 | 2 | 5.8 | 30.6 | 0.21 | 40.06 |
| processid postlarvae | night shrimps | 3,321 | 65 | -1.4 | 32.2 | 167.56 | 14100.31 |
| processid juveniles | night shrimps | 1 | 1 | 2.5 | 22.4 | 0.05 | 12.94 |
| Ambidexter symmetricus juveniles | shrimp | 16 | 9 | -0.3 | 25.4 | 0.80 | 45.90 |
| Upogebia spp. postlarvae | mud shrimps | 29,179 | 75 | 4.1 | 30.8 | 1514.83 | 40144.04 |
| Upogebia spp. juveniles | mud shrimps | 212 | 21 | 5.7 | 27.9 | 11.46 | 1104.45 |
| Euceramus praelongus juveniles | olivepit porcelain crab | 18 | 3 | -2.5 | 33.1 | 0.88 | 131.54 |
| Petrolisthes armatus juveniles | porcelain crab | 174 | 13 | -1.6 | 32.8 | 7.98 | 679.32 |
| Crab, Emerita sp. juveniles | mole crab | 65 | 3 | 7.6 | 34.5 | 3.66 | 801.24 |
| paguroid megalops larvae | hermit crabs | 7,514 | 47 | -2.8 | 31.4 | 394.06 | 50837.08 |
| paguroid juveniles | hermit crabs | 188 | 24 | -3.0 | 30.5 | 9.81 | 626.11 |
| Portunus sp. juveniles | swimming crab | 9 | 2 | -5.1 | 31.5 | 0.49 | 81.67 |
| xanthid juveniles | mud crabs | 4 | 2 | 2.4 | 10.9 | 0.23 | 43.72 |
| Pinnixa sayana juveniles | pea crab | 1 | 1 | -3.6 | 14.6 | 0.06 | 14.57 |
| Uca spp. juveniles | fiddler crabs | 1 | 1 | 16.2 | 28.9 | 0.06 | 14.03 |
| ephemeropteran larvae | mayflies | 256 | 34 | 25.0 | 0.6 | 13.89 | 1498.29 |
| odonates, anisopteran larvae | dragonflies | 6 | 6 | 30.5 | 0.2 | 0.30 | 14.89 |
| odonates, zygopteran larvae | damsel flies | 27 | 9 | 23.6 | 0.1 | 1.46 | 133.02 |
| hemipterans, corixid adults | water boatmen | 19 | 8 | 23.6 | 0.8 | 1.08 | 94.38 |
| hemipterans, corixid juveniles | water boatmen | 2 | 2 | 28.5 | 0.2 | 0.09 | 11.55 |
| coleopterans, dytiscid adults | predaceous diving beetles | 1 | 1 | 34.4 | 5.7 | 0.05 | 11.50 |
| coleopterans, curculionid adults | beetles | 1 | 1 | 26.9 | 14.1 | 0.05 | 11.90 |
| coleopterans, elmid larvae | riffle beetles | 1 | 1 | 41.0 | 0.5 | 0.05 | 11.83 |
| coleopterans, elmid adults | riffle beetles | 4 | 4 | 31.3 | 4.2 | 0.18 | 11.95 |
| dipterans, pupae | flies, mosquitoes | 3,898 | 89 | 36.3 | 0.7 | 185.60 | 4883.81 |
| dipterans, tabanid larvae | deer flies | 2 | 1 | 20.0 | 5.9 | 0.09 | 23.82 |
| dipterans, ceratopogonid larvae | biting midges | 7 | 5 | 31.3 | 2.0 | 0.36 | 29.77 |
| dipteran, Chaoborus punctipennis larvae | phantom midge | 25,046 | 94 | 36.0 | 0.4 | 1220.03 | 57030.33 |
| dipterans, chironomid larvae | midges | 72 | 32 | 32.0 | 2.1 | 3.90 | 198.64 |
| trichopteran larvae | caddisflies | 11 | 6 | 30.5 | 0.3 | 0.70 | 82.54 |
| brachiopod, Glottidia pyramidata larvae | lamp shell | 7 | 2 | -5.9 | 33.2 | 0.35 | 48.87 |
| chaetognaths, sagittid | arrow worms | 55,660 | 128 | 6.0 | 27.8 | 3136.78 | 78078.57 |
| ascidiacean larvae | tunicate larvae | 24 | 2 | 2.5 | 31.2 | 1.48 | 342.61 |
| appendicularian, Oikopleura dioica | larvacean | 54,511 | 69 | 11.3 | 24.8 | 3468.74 | 259900.09 |
| Elops saurus postflexion larvae | ladyfish | 26 | 10 | 23.6 | 10.5 | 1.22 | 126.03 |
| Myrophis punctatus juveniles | speckled worm eel | 1 | 1 | 5.2 | 24.2 | 0.04 | 11.18 |
| clupeid eggs | herrings | 1,308 | 31 | 3.9 | 30.9 | 71.00 | 6317.69 |
| clupeid preflexion larvae | herrings | 615 | 25 | 3.6 | 29.7 | 34.88 | 1694.17 |
| Brevoortia spp. postflexion larvae | menhaden | 21 | 11 | 21.9 | 11.4 | 0.98 | 79.37 |
| Brevoortia spp. metamorphs | menhaden | 2 | 2 | -4.8 | 31.7 | 0.10 | 13.15 |
| Harengula jaguana eggs | scaled sardine | 1 | 1 | 2.5 | 36.9 | 0.06 | 15.16 |
| Harengula jaguana flexion larvae | scaled sardine | 1 | 1 | 2.5 | 36.9 | 0.06 | 15.16 |
| Harengula jaguana postflexion larvae | scaled sardine | 4 | 1 | -3.6 | 38.7 | 0.22 | 55.92 |
| Harengula jaguana metamorphs | scaled sardine | 1 | 1 | 5.2 | 35.6 | 0.06 | 14.53 |
| Harengula jaguana juveniles | scaled sardine | 3 | 3 | 6.7 | 17.6 | 0.16 | 14.61 |
| Opisthonema oglinum flexion larvae | Atlantic thread herring | 23 | 7 | 8.1 | 29.4 | 1.19 | 97.74 |
| Opisthonema oglinum postflexion larvae | Atlantic thread herring | 22 | 5 | 9.1 | 28.0 | 1.09 | 88.64 |
| Opisthonema oglinum juveniles | Atlantic thread herring | 2 | 1 | 10.6 | 3.9 | 0.09 | 22.56 |
| Sardinella aurita flexion larvae | Spanish sardine | 8 | 3 | -4.4 | 33.6 | 0.43 | 54.44 |

Table A1, page 4 of 5.

Plankton-net catch statistics (May 2008 through October 2009, n = 252 samples)

Organisms are listed in phylogenetic order.

| Taxon | Common Name | Number Collected | Collection Frequency | Kmu (km) | Su (psu) | Mean CPUE (No./10 ³ m ³) | Max CPUE (No./10 ³ m ³) |
|--|---------------------------|------------------|----------------------|----------|----------|---|--|
| Sardinella aurita postflexion larvae | Spanish sardine | 9 | 4 | 8.5 | 30.6 | 0.52 | 80.03 |
| Anchoa spp. preflexion larvae | anchovies | 18,940 | 127 | 11.6 | 23.9 | 1246.57 | 81346.98 |
| Anchoa spp. flexion larvae | anchovies | 858 | 70 | 13.2 | 19.0 | 45.94 | 1239.34 |
| Anchoa hepsetus eggs | striped anchovy | 114 | 11 | 12.4 | 24.9 | 6.17 | 946.75 |
| Anchoa hepsetus postflexion larvae | striped anchovy | 2 | 1 | 5.2 | 14.5 | 0.11 | 27.10 |
| Anchoa hepsetus adults | striped anchovy | 1 | 1 | 24.2 | 13.0 | 0.05 | 12.46 |
| Anchoa mitchilli eggs | bay anchovy | 59,180 | 46 | 16.2 | 19.5 | 3024.18 | 274794.27 |
| Anchoa mitchilli postflexion larvae | bay anchovy | 2,570 | 94 | 14.6 | 22.2 | 143.08 | 9668.61 |
| Anchoa mitchilli juveniles | bay anchovy | 1,374 | 115 | 29.8 | 11.1 | 73.51 | 3430.90 |
| Anchoa mitchilli adults | bay anchovy | 387 | 75 | 30.3 | 4.6 | 19.33 | 607.28 |
| Notemigonus crysoleucas preflexion larvae | golden shiner | 3 | 1 | 41.0 | 0.2 | 0.13 | 32.22 |
| Hoplosternum littorale juveniles | brown hoplo catfish | 2 | 2 | 38.1 | 0.1 | 0.15 | 21.69 |
| Liposarcus spp. juveniles | suckermouth catfish | 6 | 5 | 36.4 | 0.2 | 0.32 | 26.26 |
| Opsanus beta juveniles | gulf toadfish | 7 | 4 | 5.5 | 0.2 | 0.39 | 40.96 |
| Labidesthes sicculus preflexion larvae | brook silverside | 1 | 1 | 41.0 | 7.4 | 0.05 | 12.36 |
| Labidesthes sicculus flexion larvae | brook silverside | 1 | 1 | 41.0 | 7.4 | 0.05 | 12.36 |
| Membras martinica preflexion larvae | rough silverside | 9 | 5 | 14.9 | 16.2 | 0.49 | 59.17 |
| Membras martinica flexion larvae | rough silverside | 3 | 1 | 2.5 | 30.3 | 0.17 | 42.55 |
| Membras martinica juveniles | rough silverside | 6 | 5 | 27.2 | 5.8 | 0.30 | 24.88 |
| Membras martinica adults | rough silverside | 4 | 4 | 13.5 | 9.5 | 0.19 | 12.21 |
| Menidia spp. preflexion larvae | silversides | 26 | 19 | 21.9 | 3.9 | 1.21 | 63.50 |
| Menidia spp. flexion larvae | silversides | 1 | 1 | 41.0 | 0.3 | 0.05 | 12.59 |
| Menidia spp. juveniles | silversides | 15 | 13 | 31.4 | 3.4 | 0.75 | 26.66 |
| Gambusia holbrooki juveniles | eastern mosquitofish | 6 | 6 | 37.8 | 0.2 | 0.34 | 21.69 |
| Synodus foetens postflexion larvae | inshore lizardfish | 2 | 2 | -0.4 | 28.6 | 0.10 | 12.48 |
| Hippocampus erectus juveniles | lined seahorse | 11 | 6 | 1.5 | 35.1 | 0.62 | 45.49 |
| Hippocampus zosterae juveniles | dwarf seahorse | 1 | 1 | -3.6 | 27.3 | 0.05 | 12.15 |
| Cosmocampus hildebrandi juveniles | dwarf pipefish | 28 | 16 | 29.9 | 5.2 | 1.31 | 54.98 |
| Syngnathus floridae juveniles | dusky pipefish | 1 | 1 | 26.9 | 6.5 | 0.05 | 12.92 |
| Syngnathus louisianae juveniles | chain pipefish | 39 | 19 | 20.4 | 17.0 | 2.10 | 61.78 |
| Syngnathus scovelli juveniles | gulf pipefish | 104 | 41 | 9.2 | 21.6 | 5.46 | 97.74 |
| Syngnathus scovelli adults | gulf pipefish | 1 | 1 | 20.0 | 1.6 | 0.05 | 12.95 |
| Prionotus spp. postflexion larvae | searobins | 1 | 1 | -3.6 | 14.6 | 0.06 | 14.57 |
| Mugil cephalus juveniles | striped mullet | 4 | 4 | 23.2 | 9.9 | 0.19 | 12.48 |
| fish eggs, percomorph | sciaenid eggs (primarily) | 88,022 | 98 | 15.2 | 24.9 | 5319.30 | 214589.22 |
| Lepomis macrochirus juveniles | bluegill | 6 | 3 | 35.6 | 0.1 | 0.43 | 49.66 |
| Lepomis spp. juveniles | sunfishes | 2 | 2 | 35.7 | 0.3 | 0.09 | 12.45 |
| Chloroscombrus chrysurus preflexion larvae | Atlantic bumper | 33 | 5 | -5.5 | 30.8 | 1.60 | 191.07 |
| Chloroscombrus chrysurus flexion larvae | Atlantic bumper | 12 | 2 | -5.9 | 27.2 | 0.54 | 100.32 |
| Chloroscombrus chrysurus postflexion larvae | Atlantic bumper | 4 | 2 | 16.9 | 24.5 | 0.23 | 48.67 |
| Chloroscombrus chrysurus juveniles | Atlantic bumper | 1 | 1 | 10.6 | 0.9 | 0.05 | 12.40 |
| Oligoplites saurus preflexion larvae | leatherjack | 15 | 4 | 3.8 | 22.6 | 0.73 | 79.59 |
| Oligoplites saurus flexion larvae | leatherjack | 7 | 2 | 0.2 | 23.4 | 0.32 | 45.90 |
| Oligoplites saurus postflexion larvae | leatherjack | 1 | 1 | -3.6 | 19.9 | 0.06 | 16.02 |
| Oligoplites saurus juveniles | leatherjack | 3 | 3 | 4.7 | 16.6 | 0.16 | 13.98 |
| Uraspis secunda juveniles | cottonmouth jack | 1 | 1 | 10.6 | 27.1 | 0.05 | 13.65 |
| Lutjanus griseus juveniles | gray snapper | 1 | 1 | -5.9 | 29.2 | 0.05 | 13.36 |
| Eucinostomus spp. postflexion larvae | mojarras | 14 | 6 | -0.2 | 20.9 | 0.66 | 72.91 |
| Eucinostomus spp. juveniles | mojarras | 6 | 2 | -4.3 | 15.1 | 0.39 | 85.15 |
| Eugerres plumieri postflexion larvae | striped mojarra | 11 | 3 | 14.4 | 29.6 | 0.69 | 97.34 |
| gerreid preflexion larvae | mojarras | 689 | 35 | 5.5 | 30.1 | 36.77 | 1096.00 |
| gerreid flexion larvae | mojarras | 9 | 3 | 13.7 | 27.6 | 0.56 | 97.34 |
| Archosargus probatocephalus postflexion larvae | sheepshead | 1 | 1 | -3.6 | 32.1 | 0.05 | 13.00 |
| Archosargus probatocephalus juveniles | sheepshead | 39 | 7 | -1.5 | 30.7 | 2.09 | 263.85 |
| Bairdiella chrysoura preflexion larvae | silver perch | 91 | 12 | -1.2 | 28.5 | 4.43 | 588.98 |
| Bairdiella chrysoura flexion larvae | silver perch | 7 | 3 | 3.5 | 22.6 | 0.37 | 46.72 |
| Bairdiella chrysoura postflexion larvae | silver perch | 12 | 5 | 10.8 | 17.7 | 0.62 | 52.62 |
| Bairdiella chrysoura juveniles | silver perch | 2 | 1 | 30.2 | 19.3 | 0.15 | 36.69 |
| Cynoscion arenarius preflexion larvae | sand seatrout | 124 | 15 | 7.8 | 25.5 | 8.68 | 957.41 |
| Cynoscion arenarius flexion larvae | sand seatrout | 10 | 4 | -1.8 | 17.6 | 0.50 | 55.74 |
| Cynoscion arenarius postflexion larvae | sand seatrout | 41 | 10 | 7.5 | 11.8 | 2.27 | 197.03 |
| Cynoscion arenarius juveniles | sand seatrout | 7 | 6 | 31.5 | 7.9 | 0.37 | 25.54 |
| Cynoscion nebulosus preflexion larvae | spotted seatrout | 142 | 18 | -3.3 | 28.1 | 6.75 | 378.70 |

Table A1, page 5 of 5.

Plankton-net catch statistics (May 2008 through October 2009, n = 252 samples)

Organisms are listed in phylogenetic order.

| Taxon | Common Name | Number Collected | Collection Frequency | K _{mu} (km) | S _u (psu) | Mean CPUE (No./10 ³ m ³) | Max CPUE (No./10 ³ m ³) |
|--|-----------------------|------------------|----------------------|----------------------|----------------------|---|--|
| Cynoscion nebulosus flexion larvae | spotted seatrout | 6 | 2 | -0.2 | 20.7 | 0.27 | 35.12 |
| Cynoscion nebulosus postflexion larvae | spotted seatrout | 6 | 3 | 0.6 | 19.0 | 0.29 | 35.12 |
| Leiostomus xanthurus postflexion larvae | spot | 4 | 1 | -3.6 | 36.3 | 0.17 | 43.13 |
| Menticirrhus spp. preflexion larvae | kingfishes | 389 | 31 | 4.1 | 28.1 | 20.90 | 620.16 |
| Menticirrhus spp. postflexion larvae | kingfishes | 9 | 4 | 2.8 | 21.4 | 0.46 | 46.72 |
| Pogonias cromis postflexion larvae | black drum | 6 | 3 | -3.1 | 10.3 | 0.37 | 45.47 |
| Pogonias cromis juveniles | black drum | 2 | 1 | -5.9 | 14.6 | 0.14 | 34.06 |
| Chaetodipterus faber preflexion larvae | Atlantic spadefish | 25 | 9 | -1.4 | 31.4 | 1.40 | 93.44 |
| Chasmodes saburrae flexion larvae | Florida blenny | 215 | 28 | 8.2 | 27.1 | 11.94 | 607.42 |
| Chasmodes saburrae postflexion larvae | Florida blenny | 178 | 25 | 7.9 | 27.8 | 9.62 | 310.65 |
| Chasmodes saburrae juveniles | Florida blenny | 2 | 1 | -3.6 | 27.3 | 0.10 | 24.30 |
| Lupinoblennius nicholsi flexion larvae | highfin blenny | 1 | 1 | -3.6 | 32.3 | 0.06 | 14.50 |
| Lupinoblennius nicholsi postflexion larvae | highfin blenny | 6 | 2 | 13.3 | 24.0 | 0.34 | 47.00 |
| blenniid preflexion larvae | blennies | 1,653 | 106 | 6.3 | 26.9 | 84.74 | 2522.12 |
| Bathygobius soporator preflexion larvae | frillfin goby | 377 | 39 | 11.0 | 23.7 | 20.68 | 784.02 |
| Bathygobius soporator flexion larvae | frillfin goby | 64 | 15 | 7.2 | 24.0 | 3.16 | 93.67 |
| Bathygobius soporator postflexion larvae | frillfin goby | 26 | 8 | 3.0 | 25.2 | 1.28 | 58.93 |
| Gobionellus spp. flexion larvae | gobies | 1 | 1 | 2.5 | 14.3 | 0.05 | 12.56 |
| Gobionellus spp. postflexion larvae | gobies | 2 | 1 | -3.6 | 27.3 | 0.10 | 24.30 |
| Gobiosoma bosc juveniles | naked goby | 19 | 9 | 35.3 | 0.8 | 0.95 | 54.48 |
| Gobiosoma robustum juveniles | code goby | 1 | 1 | 10.6 | 3.9 | 0.04 | 11.28 |
| Gobiosoma spp. postflexion larvae | gobies | 2,008 | 97 | 16.8 | 19.4 | 109.53 | 3846.58 |
| Microgobius gulosus juveniles | clown goby | 92 | 17 | 38.5 | 1.4 | 5.01 | 642.24 |
| Microgobius gulosus adults | clown goby | 1 | 1 | 34.4 | 0.2 | 0.05 | 13.20 |
| Microgobius spp. flexion larvae | gobies | 105 | 36 | 17.9 | 15.7 | 5.57 | 131.75 |
| Microgobius spp. postflexion larvae | gobies | 285 | 43 | 22.9 | 11.5 | 14.57 | 456.57 |
| Microgobius thalassinus juveniles | green goby | 2 | 2 | 12.0 | 0.1 | 0.12 | 14.78 |
| gobiid preflexion larvae | gobies | 2,685 | 158 | 9.9 | 23.6 | 144.21 | 2079.93 |
| gobiid flexion larvae | gobies | 851 | 82 | 8.1 | 24.2 | 44.02 | 1190.38 |
| gobiid postflexion larvae | gobies | 2 | 1 | -5.9 | 14.6 | 0.14 | 34.06 |
| Gobiesox strumosus preflexion larvae | skilletfish | 130 | 35 | 2.1 | 28.7 | 6.45 | 194.06 |
| Gobiesox strumosus flexion larvae | skilletfish | 11 | 8 | 18.5 | 16.2 | 0.60 | 61.48 |
| Gobiesox strumosus postflexion larvae | skilletfish | 8 | 2 | 28.4 | 11.5 | 0.44 | 61.48 |
| Achirus lineatus preflexion larvae | lined sole | 96 | 16 | -4.3 | 29.6 | 4.68 | 222.94 |
| Achirus lineatus flexion larvae | lined sole | 25 | 8 | -3.2 | 28.1 | 1.21 | 89.18 |
| Achirus lineatus postflexion larvae | lined sole | 6 | 2 | -4.6 | 32.4 | 0.32 | 46.60 |
| Trinectes maculatus preflexion larvae | hogchoker | 148 | 33 | -0.3 | 26.3 | 7.46 | 286.60 |
| Trinectes maculatus flexion larvae | hogchoker | 20 | 9 | 24.2 | 14.7 | 1.05 | 86.96 |
| Trinectes maculatus postflexion larvae | hogchoker | 43 | 22 | 16.9 | 14.9 | 2.14 | 101.79 |
| Trinectes maculatus juveniles | hogchoker | 98 | 32 | 33.7 | 3.7 | 4.92 | 167.67 |
| Trinectes maculatus adults | hogchoker | 1 | 1 | 24.2 | 0.2 | 0.06 | 15.06 |
| Symphurus plagiosa postflexion larvae | blackcheek tonguefish | 5 | 1 | -5.9 | 33.4 | 0.26 | 65.77 |
| Symphurus plagiosa juveniles | blackcheek tonguefish | 5 | 3 | -2.9 | 16.2 | 0.28 | 43.72 |
| Chilomycterus schoepfi preflexion larvae | striped burrfish | 6 | 1 | -5.9 | 36.6 | 0.29 | 73.25 |
| Chilomycterus schoepfi juveniles | striped burrfish | 2 | 2 | -3.6 | 17.0 | 0.11 | 15.16 |
| Sphoeroides spp. preflexion larvae | puffers | 2 | 1 | -3.6 | 38.7 | 0.11 | 27.96 |
| tetraodontid preflexion larvae | puffers | 92 | 10 | 0.4 | 32.9 | 4.89 | 466.85 |
| tetraodontid postflexion larvae | puffers | 1 | 1 | 16.2 | 28.9 | 0.06 | 14.03 |
| unidentified preflexion larvae | fish | 11 | 4 | -5.0 | 32.7 | 0.55 | 59.71 |

Appendix E

Month-specific Zooplankton Catch

Table A2. Page 1 of 7.

Plankton net catch by month (May 2008 to October 2009). Number of monthly samples is indicated in parentheses.

| Taxon | Common Name | Jan (14) | Feb (14) | Mar (14) | Apr (14) | May (28) | Jun (28) | Jul (28) | Aug (28) | Sep (28) | Oct (28) | Nov (14) | Dec (14) |
|-------------------------------|-------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| foraminiferans | foraminiferans | | | | 4 | | | | | | | | |
| medusa, Bougainvillia sp. | hydromedusa | | | 44 | 235 | 218 | 47 | 29 | 34 | 10 | 2 | 7 | |
| Clytia sp. | hydromedusa | 3 | | | | 1821 | 12795 | 11 | 5 | | 5 | 13 | |
| medusa, Obelia sp. | hydromedusa | | | 19 | 31 | 39 | 311 | 13 | 44 | 4 | 6 | 81 | |
| medusa, Eutima sp. | hydromedusa | | | 11 | 120 | 26 | 351 | 111 | 29 | 325 | 107 | 30 | |
| Liriope tetraphylla | hydromedusa | | | | | 81 | | | | | | | |
| medusa sp. a | hydromedusa | | | 15 | | | | | | | | 1 | |
| medusa sp. d | hydromedusa | 1 | | 36 | | 11 | 3 | 6 | | | | 16 | |
| medusa sp. e | hydromedusa | | | 16 | 57 | 22 | 38 | 13 | 8 | 14 | 65 | 15 | |
| Craspedacusta sowerbii | hydromedusa | | | | 39 | 13 | | | | | | | |
| siphonophores | siphonophores | | | 25 | 8 | | | | | | | | |
| Aurelia aurita | moon jellyfish | | | | | | 2 | | | | | | |
| Beroe ovata | sea walnut, ctenophore | | | | | 2 | | | | | | | |
| Mnemiopsis mccradyi | comb jelly, ctenophore | 3 | 3 | 282 | 5259 | 1228 | 350 | | | | | | |
| turbellarians | flatworms | | | | | 4 | | | | | | 1 | |
| nematodes | roundworms, threadworms | | 2 | 2 | | 33 | 12 | 5 | 13 | | | 4 | 1 |
| polychaetes | sand worms, tube worms | 85 | 91 | 85 | 47 | 436 | 606 | 126 | 166 | 59 | 81 | 93 | 60 |
| oligochaetes | freshwater worms | | | | | | | | 54 | 329 | | | |
| hirudinoideans | leeches | 1 | 1 | | | | 4 | | 1 | | | | |
| gastropods, prosobranch | snails | 220 | 8 | 5 | 100 | 140 | 26 | 67 | 98 | 128 | 5 | 106 | 44 |
| gastropods, opisthobranch | sea slugs | 2 | 9 | | 1 | 3 | 4 | | | | | 15 | |
| Lolliguncula brevis juveniles | bay squid | | 2 | | | 6 | 23 | | | | | | |
| pelecypods | clams, mussels, oysters | 10 | | | | 24 | 5 | | | | | 28 | 8 |
| Limulus polyphemus larvae | horseshoe crab | | | | | | | 8 | 19 | 3 | 12 | 1 | |
| acar | water mites | | | | | 1 | 6 | 3 | 6 | 11 | 2 | | |
| pycnogonids | sea spiders | | | 5 | 79 | 28 | 9 | | 2 | | | | |
| cladocerans, unidentified | water fleas | | | | | | | | 2 | | | | |
| Latona setifera | water flea | 2 | | | | | 5 | 2 | | 8 | 26 | | |
| Latonopsis fasciculata | water flea | | | | | 2681 | | 2 | 61 | | 1 | | |
| Penilia avirostris | water flea | 51 | | 2 | 151 | 211 | 6818 | 813 | 6687 | 427 | 77 | 33 | |
| Sida crystallina | water flea | | | | | | 3 | 2 | | 10 | 2 | | |
| Diaphanosoma brachyurum | water flea | | | | | | | 134 | 2 | | | | |
| Bosmina sp. | water flea | | | | | | | 39 | | | | | |
| Kurzia longirostris | water flea | | | | | | | 1 | | | | | |
| Leydigia sp. | water flea | | | | | | 1 | | 6 | 15 | | | |
| Ceriodaphnia sp. | water flea | | | | | | 34 | 458 | 8 | 107 | | | |
| cladocerans, Daphnia spp. | water fleas | | | | | 713 | 323 | 138423 | 93 | 125 | 175 | | |
| Simocephalus vetulus | water flea | | | | | 1 | 30 | 29 | 109 | 63 | 1 | | |
| llyocryptus sp. | water flea | | | | | | 13 | 18 | 92 | 254 | | | |
| Pseudevadne tergestina | water flea | 15 | | | | | 22 | 29 | 820 | 10 | 2 | | |
| Pleopsis polyphemoides | water flea | | 2 | | | | | | | | | | |
| cirriped nauplius stage | barnacles | 2 | 586 | 4 | 4 | 18 | 1 | 35 | 65 | 11 | 787 | 206 | 2 |
| cirriped cypris stage | barnacles | | 1 | | | 5 | 3 | 7 | 7 | 5 | 106 | 1 | |
| branchiurans, Argulus spp. | fish lice | | | | | 2 | 2 | 5 | 4 | 3 | 6 | 2 | 1 |
| Acartia tonsa | copepod | 1721 | 8727 | 27977 | 30183 | 13936 | 15806 | 4412 | 1868 | 208 | 2556 | 13097 | 174 |
| Centropages velificatus | copepod | 6 | | 175 | 554 | 383 | 20 | 3 | 2 | 73 | 220 | 34 | |
| Osphranticum labronectum | copepod | | | | | | | | 3 | | | | |
| Diaptomus spp. | copepods | 10 | | | | 29 | 305 | 2385 | 75 | 1632 | 27 | | 2 |

Table A2. Page 2 of 7.

Plankton net catch by month (May 2008 to October 2009). Number of monthly samples is indicated in parentheses.

| Taxon | Common Name | Jan (14) | Feb (14) | Mar (14) | Apr (14) | May (28) | Jun (28) | Jul (28) | Aug (28) | Sep (28) | Oct (28) | Nov (14) | Dec (14) |
|--------------------------------------|-------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Pseudodiaptomus coronatus | copepod | 15 | 110 | 102 | 2289 | 246 | 66 | 23 | 48 | 11 | 39 | 113 | 36 |
| Eucalanus sp. | copepod | 2 | | | | | | 5 | | | 5 | | |
| paracalanids | copepods | | 24 | 78 | 17 | 7 | 50 | 21 | 9 | 1 | 3 | 6 | 1 |
| Calanopia americana | copepod | | | | 4 | 3 | 51 | 5 | 5 | 211 | 516 | 62 | |
| Labidocera aestiva | copepod | 12 | 118 | 14831 | 7956 | 10259 | 3869 | 247 | 3121 | 1522 | 1538 | 101 | 21 |
| Labidocera acutifrons | copepod | | | | | | | | | | 2 | | |
| Eurytemora affinis | copepod | 2 | | | | | | | | | | | |
| Temora turbinata | copepod | 5 | 2 | 60 | 37 | | 8 | 8 | 7 | 30 | 24 | 127 | 1 |
| Temora stylifera | copepod | | | | | | | 1 | | | | | |
| Temora longicornis | copepod | | | | | | | 5 | 5 | 3 | | | |
| unidentified freshwater cyclopoids | copepods | 1 | | | | 7 | 9 | 32 | 14 | | 4 | | |
| Cyclops spp. | copepods | | | | | | 2 | | | 18 | | | |
| Eucyclops speratus | copepod | 6 | 4 | 76 | 9 | 11 | | 4 | | | 2 | 21 | 2 |
| Macrocyclops albidus | copepods | | 1 | 3 | | 1 | | | 8 | 5 | | | |
| Mesocyclops edax | copepod | 2521 | 80 | | | 302 | 2937 | 5405 | 3255 | 5493 | 1272 | 5 | 8 |
| Mesocyclops leuckarti | copepod | | | | | | | | 8 | | | | |
| Oithona spp. | copepods | 1 | 123 | 113 | 158 | 80 | | 15 | 34 | 6 | 7 | 7 | |
| Alteutha sp. | copepod | | 17 | 4 | | | 46 | | | | | | |
| unidentified harpacticoids | copepods | | 13 | 49 | 40 | 57 | | 16 | 22 | 1 | 6 | 2 | 10 |
| Saphirella spp. | copepods | 1 | | | | | | | 2 | 1 | | | 2 |
| siphonostomatids | parasitic copepods | 2 | 8 | 5 | 50 | 31 | 16 | 10 | 5 | 3 | 13 | 20 | 1 |
| Monstrilla sp. | copepod | | 31 | 37 | 1 | 62 | 42 | 17 | 3 | 6 | 50 | | 3 |
| myodocopod sp. a | ostracod, seed shrimp | | | | | | | 2 | 3 | | | | |
| Parasterope pollex | ostracod, seed shrimp | 71 | 73 | 31 | 42 | 160 | 1221 | 145 | 3 | 6 | 3 | 84 | 54 |
| Sarsiella zostericola | ostracod, seed shrimp | | 5 | 6 | 8 | 36 | 42 | 157 | 5 | 1 | 4 | 3 | 7 |
| ostracods, podocopid | ostracods, seed shrimps | | | 5 | | | | 3 | 9 | | 7 | 5 | |
| Squilla empusa larvae | mantis shrimp | | | 47 | 26 | 64 | 18 | | | | | | |
| mysid sp. A | opossum shrimps, mysids | | | | | | | | | | 1 | | |
| (Heteromysis sp.) | opossum shrimp, mysid | 1 | | | | 18 | 80 | | | | | | |
| Americamysis almyra | opossum shrimp, mysid | 4578 | 1956 | 2322 | 4097 | 808 | 2269 | 6372 | 559 | 506 | 334 | 1717 | 2844 |
| Americamysis bahia | opossum shrimp, mysid | 155 | 10 | 1 | | 28 | 54 | 48 | 112 | 6 | 34 | 63 | |
| unidentified Americamysis juveniles | opossum shrimps, mysids | 3475 | 3584 | 3140 | 5124 | 1558 | 1543 | 10353 | 488 | 383 | 379 | 3276 | 2711 |
| Americamysis stucki | opossum shrimp, mysid | 2 | | | | 9 | | | | 6 | 1 | 72 | |
| Bowmaniella dissimilis | opossum shrimp, mysid | 540 | 174 | 223 | 315 | 366 | 483 | 557 | 110 | 1166 | 409 | 466 | 399 |
| Brasiliomysis castroi | opossum shrimp, mysid | | | | | | | | | | | 1 | |
| Mysidopsis furca | opossum shrimp, mysid | 10 | 5 | 2 | 4 | | | | | | | | |
| Mysidopsis mortenseni | opossum shrimp, mysid | 1 | | | | | | | | | | | |
| Spelaeomysis sp. | opossum shrimp, mysid | | | | | | | | 2 | | 1 | | |
| Taphromysis bowmani | opossum shrimp, mysid | 1 | | | | 2 | 8 | | 3 | 2 | 5 | 2 | |
| amphipods, gammaridean | amphipods | 2554 | 6488 | 3701 | 22771 | 18387 | 14219 | 10364 | 115691 | 12766 | 193584 | 1117 | 10822 |
| amphipods, caprellid | skeleton shrimps | 1 | 44 | 17 | 49 | 144 | 114 | 17 | 10 | 10 | 8 | | 2 |
| Lestrigonus bengalensis | hyperiid amphipod | | | | | | | | | | 2 | | |
| isopod sp. a | isopod | | | | | 2 | | | | | | | |
| Cyathura polita | isopod | 1 | 4 | 1 | 19 | 8 | 16 | 58 | 27 | 53 | 9 | 3 | 2 |
| Xenanthura brevitelson | isopod | | 2 | | | | 3 | | | 2 | | | |
| Anopsilana jonesi | isopod | | | | 1 | 35 | 9 | 7 | 3 | | | 3 | |
| cymothoid sp. a (Lironeca) juveniles | isopod | 12 | 23 | 10 | 61 | 61 | 18 | 60 | 15 | 5 | 4 | 13 | 8 |
| Cassinidea ovalis | isopod | 1 | | | 4 | | 1 | 4 | 2 | 1 | 4 | | 1 |

Table A2. Page 3 of 7.

Plankton net catch by month (May 2008 to October 2009). Number of monthly samples is indicated in parentheses.

| Taxon | Common Name | Jan (14) | Feb (14) | Mar (14) | Apr (14) | May (28) | Jun (28) | Jul (28) | Aug (28) | Sep (28) | Oct (28) | Nov (14) | Dec (14) |
|---|--------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Sphaeromatinae | isopods | | | | | | 38 | 8 | | | | | |
| Harrieta faxoni | isopod | 2 | | | 21 | 28 | 5 | 19 | | 4 | | | |
| Sphaeroma quadridentata | isopod | | | | | 2 | | 1 | | | | | |
| Sphaeroma terebrans | isopod | | | | | | | | | | | | 2 |
| Gnathia sp. larvae | isopod | | | | | | | | | | | | 1 |
| Munna reynoldsi | isopod | | 8 | 9 | 19 | 10 | 7 | | | 1 | 2 | 4 | 19 |
| Edotia triloba | isopod | 124 | 270 | 367 | 2779 | 2030 | 495 | 987 | 350 | 810 | 454 | 137 | 86 |
| Erichsonella attenuata | isopod | 1 | 7 | 7 | 11 | 13 | 24 | 25 | 1 | 3 | 5 | 2 | 1 |
| Hoplomachus propinquus | tanaid | | | | | | | | 1 | | | | |
| Hargeria rapax | tanaid | 25 | 594 | 293 | 331 | 331 | 697 | 327 | | 9 | 12 | 95 | 154 |
| Sinelobus stanfordi | tanaid | | | 2 | | | | | | 2 | 1 | | 2 |
| Apseudes sp. | tanaid | | | | | 33 | 197 | 54 | 5 | | | | |
| cumaceans | cumaceans | 10766 | 33461 | 75597 | 19568 | 11156 | 10282 | 1206 | 1940 | 162 | 522 | 1629 | 888 |
| penaeid mysis larvae | penaeid shrimps | | | | | | | | 7 | | | | |
| penaeid postlarvae | penaeid shrimps | | | | 4 | 3 | 8 | 2 | 2 | 50 | 17 | | |
| penaeid metamorphs | penaeid shrimps | | | | 4 | 9 | 236 | 99 | 147 | 101 | 28 | 25 | |
| Farfantepenaeus duorarum juveniles | pink shrimp | 1 | | | | 2 | 2 | 13 | 3 | 10 | 1 | 5 | 1 |
| Trachypenaeopsis mobilispinus juveniles | shrimps | | | | | | 5 | | | | | | |
| Lucifer faxoni juveniles and adults | shrimp | 37 | 259 | 36 | 3855 | 3392 | 5930 | 2064 | 182 | 345 | 7465 | 2939 | 261 |
| Lucifer faxoni mysis | shrimp | 1 | | 4 | 8 | 7 | 3 | | | | 12 | 4 | 2 |
| shrimps, unidentified postlarvae | shrimps | | | | 12 | | 10 | | | | 2 | | |
| decapod zoeae | crab larvae | 11312 | 55985 | 111677 | 150715 | 151848 | 268812 | 81869 | 30444 | 30158 | 33670 | 14034 | 2893 |
| decapod megalopae | post-zoea crab larvae | 1 | | 405 | 622 | 647 | 2932 | 2342 | 994 | 1936 | 213 | 210 | 6 |
| decapod mysis | shrimp larvae | 176 | 5911 | 25106 | 27568 | 128860 | 84892 | 4428 | 5329 | 440 | 3272 | 536 | 150 |
| Leptochela serratorbita postlarvae | combclaw shrimp | | | | 4 | | | | | | | | |
| Palaemon floridanus postlarvae | Florida grass shrimp | | | 128 | 671 | 211 | 534 | 5 | 25 | | 38 | 1 | |
| Periclimenes longicaudatus juveniles | longtail grass shrimp | | | | | | | | | | 6 | | |
| Periclimenes spp. postlarvae | shrimps | | | | | 2 | | | | 4 | | | |
| Periclimenes spp. juveniles | shrimps | 3 | | | | | | | | 13 | 12 | 15 | |
| Palaemonetes paludosus juveniles | grass shrimp | | | 1 | | | | | | 4 | 87 | 2 | |
| Palaemonetes pugio juveniles | daggerblade grass shrimp | 1 | 1 | | | | | 5 | | 9 | 1 | 15 | 1 |
| Palaemonetes pugio adults | daggerblade grass shrimp | | | | | | | | 1 | 1 | | | |
| Palaemonetes spp. postlarvae | grass shrimps | 2 | 13 | 226 | 125 | 4544 | 883 | 52 | 227 | 29 | 56 | 42 | 5 |
| Palaemonetes spp. juveniles | grass shrimps | | | | | | | | 3 | | | | |
| Palaemonetes vulgaris juveniles | grass shrimp | | | | | | | | | | | | 1 |
| alphaeid postlarvae | snapping shrimps | 3 | 61 | 2866 | 1373 | 3791 | 2781 | 172 | 489 | 402 | 135 | 24 | |
| alphaeid juveniles | snapping shrimps | | | | | 3 | | 25 | 1 | 8 | | | |
| Alpheus estuariensis juveniles | snapping shrimp | | | | | | | | | 1 | 3 | | |
| Hippolyte zostericola postlarvae | zostera shrimp | 31 | 252 | 698 | 157 | 194 | 1624 | 246 | 544 | 241 | 121 | 483 | 59 |
| Hippolyte zostericola juveniles | zostera shrimp | 7 | 7 | 18 | 2 | 1 | 7 | 5 | 10 | 1 | 8 | 20 | 13 |
| Hippolyte zostericola adults | zostera shrimp | | | | | | | | 3 | | | 12 | |
| Latreutes parvulus postlarvae | sargassum shrimp | | | | | 136 | | | | | | | |
| Tozeuma carolinense postlarvae | arrow shrimp | 2 | | 9 | 375 | 121 | 396 | 26 | 15 | | | 8 | |
| Tozeuma carolinense juveniles | arrow shrimp | | | | | | | 39 | 10 | 5 | 13 | 15 | |
| Ogyrides alphaerostris postlarvae | estuarine longeye shrimp | | | 3 | 1 | | | | | | | | |
| processid postlarvae | night shrimps | 3 | 4 | 277 | 397 | 275 | 1671 | 42 | 115 | 89 | 224 | 220 | 4 |
| processid juveniles | night shrimps | | | | | | | | | | 1 | | |
| Ambidexter symmetricus juveniles | shrimp | | | | 1 | 1 | 4 | 4 | | 2 | | 4 | |

Table A2. Page 4 of 7.

Plankton net catch by month (May 2008 to October 2009). Number of monthly samples is indicated in parentheses.

| Taxon | Common Name | Jan (14) | Feb (14) | Mar (14) | Apr (14) | May (28) | Jun (28) | Jul (28) | Aug (28) | Sep (28) | Oct (28) | Nov (14) | Dec (14) |
|---|---------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Upogebia spp. postlarvae | mud shrimps | 1 | 49 | 4777 | 6902 | 7680 | 6697 | 2005 | 483 | 370 | 191 | 24 | |
| Upogebia spp. juveniles | mud shrimps | | | 4 | 15 | 90 | 38 | 29 | 5 | 31 | | | |
| Euceramus praelongus juveniles | olivepit porcelain crab | | | | 8 | | 10 | | | | | | |
| Petrolisthes armatus juveniles | porcelain crab | | | | 112 | | 47 | 3 | | 11 | 1 | | |
| Crab, Emerita sp. juveniles | mole crab | | | | | 65 | | | | | | | |
| paguroid megalops larvae | hermit crabs | 12 | 69 | 770 | 1374 | 17 | 310 | 66 | 26 | 49 | 25 | 4796 | |
| paguroid juveniles | hermit crabs | | | 20 | 9 | | 30 | 34 | | 5 | 16 | 74 | |
| Portunus sp. juveniles | swimming crab | | | | | | | | | | | 9 | |
| xanthid juveniles | mud crabs | | | | | | | | | 4 | | | |
| Pinnixa sayana juveniles | pea crab | | | | | | | | | 1 | | | |
| Uca spp. juveniles | fiddler crabs | | | | | 1 | | | | | | | |
| ephemeropteran larvae | mayflies | | | | | | 3 | 17 | 179 | 23 | 33 | 1 | |
| odonates, anisopteran larvae | dragonflies | | | | | | | | 2 | 4 | | | |
| odonates, zygopteran larvae | damsel flies | | | | | | | 4 | 19 | 4 | | | |
| hemipterans, corixid adults | water boatmen | | | | | | | | 13 | 1 | 5 | | |
| hemipterans, corixid juveniles | water boatmen | | | | | | | 1 | 1 | | | | |
| coleopterans, dytiscid adults | predaceous diving beetles | | | | | | | | | | | | 1 |
| coleopterans, curculionid adults | beetles | | | | | | | | | | | | 1 |
| coleopterans, elmid larvae | riffle beetles | | | | | | | | | | 1 | | |
| coleopterans, elmid adults | riffle beetles | | | | | | | 1 | 1 | | 1 | | 1 |
| dipterans, pupae | flies, mosquitoes | 43 | 26 | 2 | | 127 | 143 | 963 | 890 | 1163 | 484 | 33 | 24 |
| dipterans, tabanid larvae | deer flies | | | | | | 2 | | | | | | |
| dipterans, ceratopogonid larvae | biting midges | | | 1 | | | | | | 3 | 3 | | |
| dipteran, Chaoborus punctipennis larvae | phantom midge | 53 | 7 | | 242 | 26 | 1848 | 12060 | 4090 | 4923 | 1769 | 13 | 15 |
| dipterans, chironomid larvae | midges | 2 | 1 | 1 | | | 10 | 12 | 22 | 18 | 5 | | 1 |
| trichopteran larvae | caddisflies | | 1 | | | | 1 | | 8 | 1 | | | |
| brachiopod, Glottidia pyramidata larvae | lamp shell | | | | 4 | | | 3 | | | | | |
| chaetognaths, sagittid | arrow worms | 987 | 2738 | 4049 | 9562 | 16241 | 5129 | 2417 | 206 | 563 | 3690 | 8472 | 1606 |
| ascidiacean larvae | tunicate larvae | | | | | 2 | 22 | | | | | | |
| appendicularian, Oikopleura dioica | larvacean | 15 | 640 | 266 | 7786 | 39593 | 24 | 255 | 1243 | 621 | 1271 | 2791 | 6 |
| Elops saurus postflexion larvae | ladyfish | 18 | 4 | 4 | | | | | | | | | |
| Myrophis punctatus juveniles | speckled worm eel | | 1 | | | | | | | | | | |
| clupeid eggs | herrings | | | 70 | 528 | 401 | 220 | 19 | 26 | 8 | 36 | | |
| clupeid preflexion larvae | herrings | | | 82 | 63 | 419 | 39 | 8 | | | | 4 | |
| Brevoortia spp. postflexion larvae | menhaden | 7 | 13 | 1 | | | | | | | | | |
| Brevoortia spp. metamorphs | menhaden | | | | | | 2 | | | | | | |
| Harengula jaguana eggs | scaled sardine | | | | | 1 | | | | | | | |
| Harengula jaguana flexion larvae | scaled sardine | | | | | 1 | | | | | | | |
| Harengula jaguana postflexion larvae | scaled sardine | | | | | 4 | | | | | | | |
| Harengula jaguana metamorphs | scaled sardine | | | | | 1 | | | | | | | |
| Harengula jaguana juveniles | scaled sardine | 2 | | | | | | | 1 | | | | |
| Opisthonema oglinum flexion larvae | Atlantic thread herring | | | | 12 | 8 | 3 | | | | | | |
| Opisthonema oglinum postflexion larvae | Atlantic thread herring | | | | 12 | 7 | 3 | | | | | | |
| Opisthonema oglinum juveniles | Atlantic thread herring | | | | | | | 2 | | | | | |
| Sardinella aurita flexion larvae | Spanish sardine | | | 3 | | 1 | | | | | | 4 | |
| Sardinella aurita postflexion larvae | Spanish sardine | 1 | | | | 7 | | | | 1 | | | |
| Anchoa spp. preflexion larvae | anchovies | 5 | 295 | 1927 | 996 | 11507 | 2268 | 862 | 788 | 69 | 174 | 49 | |
| Anchoa spp. flexion larvae | anchovies | 1 | | 3 | 53 | 206 | 479 | 69 | 32 | 7 | 8 | | |

Table A2. Page 5 of 7.

Plankton net catch by month (May 2008 to October 2009). Number of monthly samples is indicated in parentheses.

| Taxon | Common Name | Jan (14) | Feb (14) | Mar (14) | Apr (14) | May (28) | Jun (28) | Jul (28) | Aug (28) | Sep (28) | Oct (28) | Nov (14) | Dec (14) |
|---|---------------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Anchoa hepsetus eggs | striped anchovy | | 19 | 87 | 6 | 2 | | | | | | | |
| Anchoa hepsetus postflexion larvae | striped anchovy | | | | | | | 2 | | | | | |
| Anchoa hepsetus adults | striped anchovy | | | | 1 | | | | | | | | |
| Anchoa mitchilli eggs | bay anchovy | | 3137 | 19495 | 23138 | 1498 | 11906 | | | | 6 | | |
| Anchoa mitchilli postflexion larvae | bay anchovy | | | | 84 | 1024 | 1139 | 208 | 18 | 19 | 24 | 54 | |
| Anchoa mitchilli juveniles | bay anchovy | 13 | 2 | | 39 | 497 | 193 | 191 | 92 | 126 | 84 | 122 | 15 |
| Anchoa mitchilli adults | bay anchovy | 42 | 36 | 9 | 8 | 5 | 16 | 53 | 20 | 75 | 22 | 92 | 9 |
| Notemigonus crysoleucas preflexion larvae | golden shiner | | | | | | | 3 | | | | | |
| Hoplosternum littorale juveniles | brown hoplo catfish | | | | | | | | 2 | | | | |
| Liposarcus spp. juveniles | suckermouth catfish | | | | | | | 5 | 1 | | | | |
| Opsanus beta juveniles | gulf toadfish | | | | | | | | 7 | | | | |
| Labidesthes sicculus preflexion larvae | brook silverside | | | | | 1 | | | | | | | |
| Labidesthes sicculus flexion larvae | brook silverside | | | | | 1 | | | | | | | |
| Membras martinica preflexion larvae | rough silverside | | | 5 | | | | | | 3 | 1 | | |
| Membras martinica flexion larvae | rough silverside | | | | | 3 | | | | | | | |
| Membras martinica juveniles | rough silverside | 1 | | | | 1 | | | | 3 | | | |
| Membras martinica adults | rough silverside | | 2 | | | | | 1 | | 1 | | | |
| Menidia spp. preflexion larvae | silversides | 1 | | 1 | | | 1 | 1 | 1 | 16 | 3 | 1 | 1 |
| Menidia spp. flexion larvae | silversides | | | | | | | 1 | | | | | |
| Menidia spp. juveniles | silversides | | | | 2 | | 2 | 2 | 3 | | 3 | 2 | 1 |
| Gambusia holbrooki juveniles | eastern mosquitofish | | | | | | | 2 | 3 | | 1 | | |
| Synodus foetens postflexion larvae | inshore lizardfish | 1 | | | | | 1 | | | | | | |
| Hippocampus erectus juveniles | lined seahorse | 1 | | | 1 | 8 | | 1 | | | | | |
| Hippocampus zosterae juveniles | dwarf seahorse | | | | | | | 1 | | | | | |
| Cosmocampus hildebrandi juveniles | dwarf pipefish | 1 | | 1 | | 2 | | 2 | 6 | 10 | 3 | | 3 |
| Syngnathus floridae juveniles | dusky pipefish | | 1 | | | | | | | | | | |
| Syngnathus louisianae juveniles | chain pipefish | | 1 | 8 | | 10 | 8 | 6 | 2 | 3 | | 1 | |
| Syngnathus scovelli juveniles | gulf pipefish | 7 | 1 | | 21 | 18 | 12 | 19 | 2 | 20 | | 2 | 2 |
| Syngnathus scovelli adults | gulf pipefish | | | | | | | 1 | | | | | |
| Prionotus spp. postflexion larvae | searobins | | | | | | | | | 1 | | | |
| Mugil cephalus juveniles | striped mullet | 3 | 1 | | | | | | | | | | |
| fish eggs, percomorph | sciaenid eggs (primarily) | 5 | 2906 | 12315 | 16449 | 46765 | 8686 | 309 | 302 | 112 | 154 | 1 | 18 |
| Lepomis macrochirus juveniles | bluegill | | | | | | | | 6 | | | | |
| Lepomis spp. juveniles | sunfishes | | | | | | | 1 | | 1 | | | |
| Chloroscombrus chrysurus preflexion larvae | Atlantic bumper | | | | | 3 | 6 | 16 | 6 | | 2 | | |
| Chloroscombrus chrysurus flexion larvae | Atlantic bumper | | | | | | | 3 | | 9 | | | |
| Chloroscombrus chrysurus postflexion larvae | Atlantic bumper | | | | | 3 | | | | 1 | | | |
| Chloroscombrus chrysurus juveniles | Atlantic bumper | | | | | | | | | 1 | | | |
| Oligoplites saurus preflexion larvae | leatherjack | | | | | | 14 | 1 | | | | | |
| Oligoplites saurus flexion larvae | leatherjack | | | | | | 7 | | | | | | |
| Oligoplites saurus postflexion larvae | leatherjack | | | | | | | | | 1 | | | |
| Oligoplites saurus juveniles | leatherjack | | | | | 1 | | 1 | 1 | | | | |
| Uraspis secunda juveniles | cottonmouth jack | | | | | | 1 | | | | | | |
| Lutjanus griseus juveniles | gray snapper | | | | | | | 1 | | | | | |
| Eucinostomus spp. postflexion larvae | mojarra | | | | | | | 12 | 1 | 1 | | | |
| Eucinostomus spp. juveniles | mojarra | | | | | | | | 5 | | | 1 | |
| Eugerres plumieri postflexion larvae | striped mojarra | | | 2 | | 9 | | | | | | | |
| gerreid preflexion larvae | mojarra | | 25 | 182 | 111 | 251 | 65 | 5 | 31 | 18 | 1 | | |

Table A2. Page 6 of 7.

Plankton net catch by month (May 2008 to October 2009). Number of monthly samples is indicated in parentheses.

| Taxon | Common Name | Jan (14) | Feb (14) | Mar (14) | Apr (14) | May (28) | Jun (28) | Jul (28) | Aug (28) | Sep (28) | Oct (28) | Nov (14) | Dec (14) |
|--|--------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| gerreid flexion larvae | mojaras | | | 2 | | 6 | | | | 1 | | | |
| Archosargus probatocephalus postflexion larvae | sheepshead | | 1 | | | | | | | | | | |
| Archosargus probatocephalus juveniles | sheepshead | 33 | | | | | | | | | | | 6 |
| Bairdiella chrysoura preflexion larvae | silver perch | | | 4 | 5 | 9 | 19 | 3 | 51 | | | | |
| Bairdiella chrysoura flexion larvae | silver perch | | | | | | 6 | 1 | | | | | |
| Bairdiella chrysoura postflexion larvae | silver perch | | | | 4 | | 3 | 5 | | | | | |
| Bairdiella chrysoura juveniles | silver perch | | | | | 2 | | | | | | | |
| Cynoscion arenarius preflexion larvae | sand seatrout | | | | | 69 | 5 | 8 | | 41 | 1 | | |
| Cynoscion arenarius flexion larvae | sand seatrout | | | | | | | | 3 | 6 | 1 | | |
| Cynoscion arenarius postflexion larvae | sand seatrout | 1 | | | 4 | | | 1 | 13 | 6 | 16 | | |
| Cynoscion arenarius juveniles | sand seatrout | | | | 1 | 1 | 1 | | 1 | | | 3 | |
| Cynoscion nebulosus preflexion larvae | spotted seatrout | | | 2 | 1 | 11 | 55 | 26 | 32 | 15 | | | |
| Cynoscion nebulosus flexion larvae | spotted seatrout | | | | | | 3 | | | 3 | | | |
| Cynoscion nebulosus postflexion larvae | spotted seatrout | | | | | | 3 | 2 | | 1 | | | |
| Leiostomus xanthurus postflexion larvae | spot | | | | 4 | | | | | | | | |
| Menticirrhus spp. preflexion larvae | kingfishes | | 1 | 38 | 58 | 167 | 26 | 42 | 18 | 34 | 1 | 4 | |
| Menticirrhus spp. postflexion larvae | kingfishes | | | | | | 3 | 2 | 3 | 1 | | | |
| Pogonias cromis postflexion larvae | black drum | | | | | | | | 6 | | | | |
| Pogonias cromis juveniles | black drum | | | | | | | | 2 | | | | |
| Chaetodipterus faber preflexion larvae | Atlantic spadefish | | 2 | 2 | | 2 | 14 | | | 5 | | | |
| Chasmodes saburrae flexion larvae | Florida blenny | | 9 | 108 | 33 | 24 | 38 | | | 1 | 1 | 1 | |
| Chasmodes saburrae postflexion larvae | Florida blenny | | 6 | 58 | 47 | 27 | 35 | 1 | 3 | 1 | | | |
| Chasmodes saburrae juveniles | Florida blenny | | | | | | | 2 | | | | | |
| Lupinoblennius nicholsi flexion larvae | highfin blenny | 1 | | | | | | | | | | | |
| Lupinoblennius nicholsi postflexion larvae | highfin blenny | | | | 3 | | 3 | | | | | | |
| blenniid preflexion larvae | blennies | 44 | 342 | 446 | 360 | 161 | 130 | 31 | 20 | 53 | 36 | 19 | 11 |
| Bathygobius soporator preflexion larvae | frillfin goby | | | 130 | 91 | 43 | 65 | 8 | 29 | 8 | 1 | 2 | |
| Bathygobius soporator flexion larvae | frillfin goby | | | 10 | 15 | 11 | 19 | 3 | 3 | 3 | | | |
| Bathygobius soporator postflexion larvae | frillfin goby | | | | | 11 | 8 | 1 | 3 | 3 | | | |
| Gobionellus spp. flexion larvae | gobies | | | | | | | 1 | | | | | |
| Gobionellus spp. postflexion larvae | gobies | | | | | | | 2 | | | | | |
| Gobiosoma bosc juveniles | naked goby | | | | | | 3 | 4 | 12 | | | | |
| Gobiosoma robustum juveniles | code goby | | | | | | | 1 | | | | | |
| Gobiosoma spp. postflexion larvae | gobies | 5 | | 57 | 361 | 228 | 1027 | 242 | 33 | 35 | 9 | 11 | |
| Microgobius gulosus juveniles | clown goby | | | | | 8 | 5 | 61 | 17 | | | 1 | |
| Microgobius gulosus adults | clown goby | | | | | | | | | 1 | | | |
| Microgobius spp. flexion larvae | gobies | 1 | 1 | 6 | 4 | 26 | 45 | 3 | | 13 | 6 | | |
| Microgobius spp. postflexion larvae | gobies | | | 3 | 24 | 24 | 157 | 59 | 4 | 9 | 3 | 2 | |
| Microgobius thalassinus juveniles | green goby | | | | | | | | 2 | | | | |
| gobiid preflexion larvae | gobies | 35 | 86 | 641 | 336 | 407 | 634 | 257 | 77 | 75 | 72 | 30 | 35 |
| gobiid flexion larvae | gobies | 9 | 7 | 60 | 119 | 102 | 393 | 86 | 13 | 30 | 25 | 6 | 1 |
| gobiid postflexion larvae | gobies | | | | | | | | 2 | | | | |
| Gobiesox strumosus preflexion larvae | skilletfish | 8 | 53 | 27 | 14 | 7 | | | | 1 | 8 | 5 | 7 |
| Gobiesox strumosus flexion larvae | skilletfish | 1 | 1 | 5 | 1 | 1 | | | | | | 2 | |
| Gobiesox strumosus postflexion larvae | skilletfish | | | 8 | | | | | | | | | |
| Achirus lineatus preflexion larvae | lined sole | | | | 8 | 4 | 18 | 20 | 24 | 20 | 2 | | |
| Achirus lineatus flexion larvae | lined sole | | | | | 1 | 7 | 2 | 5 | 9 | 1 | | |
| Achirus lineatus postflexion larvae | lined sole | | | | | | 3 | | | 3 | | | |

Table A2. Page 7 of 7.

Plankton net catch by month (May 2008 to October 2009). Number of monthly samples is indicated in parentheses.

| Taxon | Common Name | Jan (14) | Feb (14) | Mar (14) | Apr (14) | May (28) | Jun (28) | Jul (28) | Aug (28) | Sep (28) | Oct (28) | Nov (14) | Dec (14) |
|--|-----------------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|
| Trinectes maculatus preflexion larvae | hogchoker | | | | | 20 | 30 | 47 | 3 | 27 | 21 | | |
| Trinectes maculatus flexion larvae | hogchoker | | | | | 9 | 2 | 3 | | 1 | 5 | | |
| Trinectes maculatus postflexion larvae | hogchoker | | | | 9 | 2 | 13 | 3 | 1 | 5 | 9 | 1 | |
| Trinectes maculatus juveniles | hogchoker | | | | 1 | 7 | 13 | 13 | 23 | 3 | 22 | 13 | 3 |
| Trinectes maculatus adults | hogchoker | | | | | | | | | 1 | | | |
| Symphurus plagiusa postflexion larvae | blackcheek tonguefish | | | | | | 5 | | | | | | |
| Symphurus plagiusa juveniles | blackcheek tonguefish | | | | | | | | | 4 | | 1 | |
| Chilomycterus schoepfi preflexion larvae | striped burrfish | | | | | | 6 | | | | | | |
| Chilomycterus schoepfi juveniles | striped burrfish | | | | | | | | 2 | | | | |
| Sphoeroides spp. preflexion larvae | puffers | | | | | 2 | | | | | | | |
| tetraodontid preflexion larvae | puffers | | | 78 | 7 | 7 | | | | | | | |
| tetraodontid postflexion larvae | puffers | | | | | 1 | | | | | | | |
| unidentified preflexion larvae | fish | | 2 | | | 1 | 3 | 5 | | | | | |

Appendix F

Location-specific Zooplankton Catch

Table A3, page 1 of 6. Location specific plankton-net catch .

Data are presented as mean number per 1,000 cubic meters.

Organisms are listed in phylogenetic order.

| Description | Common Name | Location (km from mouth) | | | | | | | | | | | | | |
|-------------------------------|-------------------------|--------------------------|----------|---------|---------|---------|----------|----------|---------|----------|---------|--------|----------|----------|---------|
| | | -5.9 | -3.6 | 2.5 | 5.2 | 7.7 | 10.6 | 16.2 | 20.0 | 24.2 | 26.9 | 30.2 | 34.4 | 37.1 | 41.0 |
| foraminiferans | foraminiferans | 0.00 | 2.40 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| medusa, Bougainvillia sp. | hydromedusa | 391.16 | 38.03 | 10.56 | 0.81 | 0.66 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Clytia sp. | hydromedusa | 18.47 | 3.12 | 3.80 | 4.75 | 0.00 | 2.67 | 5.46 | 9.91 | 0.00 | 0.00 | 21.62 | 499.99 | 4892.76 | 7492.99 |
| medusa, Obelia sp. | hydromedusa | 153.43 | 184.46 | 27.29 | 10.02 | 4.62 | 0.00 | 7.34 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| medusa, Eutima sp. | hydromedusa | 408.78 | 251.29 | 48.07 | 11.65 | 3.34 | 17.52 | 4.38 | 0.00 | 0.74 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Liriope tetraphylla | hydromedusa | 4.52 | 9.32 | 7.58 | 6.46 | 0.00 | 0.00 | 10.13 | 15.32 | 4.21 | 9.03 | 1.02 | 0.00 | 0.00 | 0.00 |
| medusa sp. a | hydromedusa | 11.12 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.71 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| medusa sp. d | hydromedusa | 2.22 | 8.52 | 11.99 | 0.00 | 0.73 | 1.25 | 0.79 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 3.55 | 24.73 |
| medusa sp. e | hydromedusa | 82.00 | 58.87 | 4.77 | 10.82 | 8.59 | 2.67 | 0.00 | 5.41 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Craspedacusta sowerbii | hydromedusa | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 27.57 | 2.94 | 0.00 | 7.55 |
| siphonophores | siphonophores | 19.76 | 3.85 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Aurelia aurita | moon jellyfish | 0.00 | 0.00 | 0.00 | 0.00 | 0.87 | 0.76 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Beroe ovata | sea walnut, ctenophore | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.94 | 0.00 | 0.00 |
| Mnemiopsis mccradyi | comb jelly, ctenophore | 0.00 | 0.00 | 1.93 | 30.62 | 0.00 | 47.38 | 254.52 | 69.98 | 226.72 | 874.07 | 562.95 | 1766.68 | 1505.89 | 676.95 |
| turbellarians | flatworms | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 3.27 | 0.00 | 0.00 | 0.00 | 0.00 | 0.65 | 0.00 | 0.00 | 0.00 |
| nematodes | roundworms, threadworms | 22.58 | 28.30 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.84 | 0.00 | 0.00 | 0.00 | 1.72 | 0.00 | 0.69 |
| polychaetes | sand worms, tube worms | 394.92 | 368.58 | 107.16 | 84.89 | 155.81 | 143.52 | 21.55 | 47.61 | 7.29 | 10.60 | 4.06 | 242.22 | 4.03 | 11.81 |
| oligochaetes | freshwater worms | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.46 | 4.95 | 17.48 | 22.64 | 5.30 | 8.07 | 168.00 | 37.11 |
| hirudinoideans | leeches | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.32 | 0.00 | 0.00 | 1.24 | 0.00 | 0.00 | 1.84 |
| gastropods, prosobranch | snails | 114.55 | 44.78 | 48.11 | 7.29 | 3.66 | 4.63 | 54.12 | 36.23 | 65.85 | 4.79 | 43.85 | 24.01 | 13.35 | 199.85 |
| gastropods, opisthobranch | sea slugs | 6.04 | 2.55 | 4.36 | 3.11 | 2.44 | 1.59 | 4.79 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Lolliguncula brevis juveniles | bay squid | 0.00 | 0.00 | 12.33 | 2.23 | 2.61 | 5.70 | 2.34 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| pelecypods | clams, mussels, oysters | 2.23 | 1.45 | 0.00 | 0.00 | 28.81 | 13.14 | 0.79 | 0.00 | 6.59 | 0.00 | 18.15 | 35.84 | 0.65 | 1.92 |
| Limulus polyphemus larvae | horseshoe crab | 0.00 | 0.00 | 0.00 | 0.77 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| acari | water mites | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.82 | 2.47 | 0.00 | 0.68 | 0.74 | 0.00 | 3.26 | 11.51 |
| pycnogonids | sea spiders | 14.08 | 10.54 | 29.33 | 11.97 | 14.26 | 0.00 | 2.34 | 0.00 | 0.00 | 0.00 | 7.32 | 0.00 | 0.00 | 0.00 |
| cladocerans, unidentified | water fleas | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.65 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Latona setifera | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.47 | 11.92 | 7.01 | 6.67 |
| Latonopsis fasciculata | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.82 | 18.13 | 10.49 | 9.05 | 0.00 | 5.52 | 12.25 | 1840.41 |
| Penilia avirostris | water flea | 3753.31 | 7339.76 | 35.61 | 37.08 | 12.27 | 9.44 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Sida crystallina | water flea | 2.03 | 0.00 | 0.00 | 1.27 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.18 | 0.66 | 0.00 | 1.95 | 3.45 |
| Diaphanosoma brachyurum | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.50 | 0.00 | 0.00 | 0.00 | 0.00 | 3.62 | 4.15 | 72.94 | 16.78 |
| Bosmina sp. | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.87 | 0.00 | 4.84 | 3.65 | 16.78 |
| Kurzia longirostris | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.72 | 0.00 | 0.00 | 0.00 | 0.00 |
| Leydigia sp. | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 6.41 | 3.75 | 1.28 | 0.00 | 1.20 | 2.68 |
| Ceriodaphnia sp. | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.64 | 2.47 | 0.00 | 43.94 | 14.71 | 36.09 | 255.89 | 78.35 |
| cladocerans, Daphnia spp. | water fleas | 0.00 | 0.00 | 0.00 | 0.00 | 4.93 | 26.57 | 38.75 | 8194.81 | 22353.78 | 7559.79 | 828.09 | 12337.70 | 28956.69 | 5291.06 |
| Simoccephalus vetulus | water flea | 0.00 | 0.00 | 0.00 | 0.73 | 0.00 | 3.24 | 26.37 | 17.48 | 15.06 | 9.23 | 20.27 | 30.18 | 52.50 | 0.00 |
| Ilyocryptus sp. | water flea | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 3.28 | 17.31 | 38.29 | 17.72 | 12.83 | 101.60 | 53.82 | 34.74 | 0.00 |
| Pseudevadne tergestina | water flea | 437.08 | 155.92 | 0.00 | 1.20 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Pleopsis polyphemoides | water flea | 0.00 | 1.44 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| cirriped nauplius stage | barnacles | 515.24 | 586.49 | 5.69 | 3.88 | 13.84 | 23.90 | 1.58 | 9.06 | 1.47 | 7.13 | 0.00 | 0.00 | 0.58 | 1.42 |
| cirriped cypris stage | barnacles | 17.76 | 3.42 | 0.00 | 0.00 | 0.00 | 0.00 | 3.49 | 0.00 | 0.00 | 59.07 | 0.00 | 4.58 | 0.71 | 2.61 |
| branchiurans, Argulus spp. | fish lice | 0.00 | 0.67 | 0.60 | 0.66 | 1.32 | 0.69 | 1.27 | 2.17 | 0.00 | 0.78 | 2.61 | 1.26 | 4.33 | 0.66 |
| Acartia tonsa | copepod | 18586.50 | 11202.27 | 7164.24 | 5632.92 | 7360.71 | 15619.07 | 15957.53 | 7656.74 | 480.15 | 416.10 | 413.22 | 291.17 | 152.74 | 79.53 |
| Centropages velificatus | copepod | 631.84 | 272.79 | 45.02 | 53.67 | 16.28 | 8.84 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Osphranticum labronectum | copepod | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.52 | 0.00 | 0.00 | 0.00 | 0.00 |
| Diaptomus spp. | copepods | 0.00 | 0.00 | 0.72 | 0.00 | 0.00 | 2.19 | 117.81 | 56.95 | 145.86 | 401.69 | 267.16 | 570.58 | 893.83 | 575.28 |
| Pseudodiaptomus coronatus | copepod | 39.03 | 66.73 | 20.88 | 40.29 | 83.32 | 434.75 | 1522.52 | 51.30 | 46.00 | 23.87 | 53.83 | 36.69 | 3.71 | 3.95 |
| Eucalanus sp. | copepod | 5.66 | 2.12 | 0.00 | 0.70 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| paracalanids | copepods | 60.38 | 65.53 | 7.87 | 2.55 | 1.55 | 9.26 | 7.59 | 1.68 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Calanopia americana | copepod | 79.85 | 248.15 | 175.93 | 61.39 | 51.76 | 11.87 | 7.98 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Labidocera aestiva | copepod | 5467.25 | 12734.90 | 4394.56 | 2090.22 | 731.88 | 2679.70 | 1038.73 | 2418.86 | 0.00 | 3.42 | 0.00 | 0.00 | 2.07 | 1.42 |

Table A3, page 2 of 6. Location specific plankton-net catch .

Data are presented as mean number per 1,000 cubic meters.

Organisms are listed in phylogenetic order.

| Description | Common Name | Location (km from mouth) | | | | | | | | | | | | | |
|--------------------------------------|-------------------------|--------------------------|---------|----------|----------|----------|----------|---------|---------|---------|---------|---------|---------|---------|---------|
| | | -5.9 | -3.6 | 2.5 | 5.2 | 7.7 | 10.6 | 16.2 | 20.0 | 24.2 | 26.9 | 30.2 | 34.4 | 37.1 | 41.0 |
| Labidocera acutifrons | copepod | 1.28 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Eurytemora affinis | copepod | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.26 |
| Temora turbinata | copepod | 77.08 | 75.66 | 16.90 | 20.60 | 8.59 | 16.27 | 0.79 | 11.78 | 1.47 | 1.42 | 0.00 | 0.00 | 0.00 | 0.00 |
| Temora stylifera | copepod | 0.00 | 0.67 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Temora longicornis | copepod | 5.17 | 2.06 | 1.28 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| unidentified freshwater cyclopoids | copepods | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 3.21 | 4.26 | 6.78 | 7.76 | 13.54 | 12.90 |
| Cyclops spp. | copepods | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.26 | 0.00 | 0.00 | 0.00 | 5.19 | 6.91 |
| Eucyclops speratus | copepod | 50.89 | 39.64 | 1.99 | 0.00 | 0.66 | 6.02 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Macrocyclops albidus | copepods | 0.00 | 0.00 | 3.12 | 2.23 | 0.00 | 3.44 | 0.00 | 0.00 | 0.68 | 4.20 | 0.00 | 0.00 | 0.00 | 0.00 |
| Mesocyclops edax | copepod | 0.00 | 0.00 | 36.41 | 3.98 | 41.11 | 8.19 | 125.80 | 258.37 | 304.69 | 661.91 | 1273.97 | 2744.50 | 4146.08 | 4213.23 |
| Mesocyclops leuckarti | copepod | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 9.64 |
| Oithona spp. | copepods | 140.31 | 88.19 | 3.90 | 7.36 | 0.66 | 73.28 | 52.33 | 52.79 | 0.00 | 0.00 | 2.04 | 0.00 | 0.00 | 0.00 |
| Alteutha sp. | copepod | 0.00 | 2.89 | 0.00 | 4.35 | 5.29 | 4.33 | 17.14 | 14.25 | 0.74 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| unidentified harpacticoids | copepods | 66.82 | 50.92 | 4.75 | 16.34 | 2.63 | 0.00 | 3.64 | 0.00 | 3.60 | 2.80 | 4.07 | 0.00 | 0.58 | 0.60 |
| Saphirella spp. | copepods | 4.26 | 0.00 | 0.00 | 0.00 | 0.68 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| siphonostomatids | parasitic copepods | 9.98 | 41.00 | 10.99 | 17.10 | 12.00 | 7.69 | 3.03 | 8.28 | 0.74 | 0.90 | 2.64 | 0.00 | 0.00 | 0.00 |
| Monstrilla sp. | copepod | 35.89 | 44.81 | 92.73 | 8.75 | 2.40 | 2.60 | 0.00 | 2.70 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| myodocopod sp. a | ostracod, seed shrimp | 0.00 | 1.35 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.68 | 0.58 | 0.61 |
| Parasterope pollex | ostracod, seed shrimp | 29.16 | 187.60 | 200.81 | 181.21 | 799.80 | 54.86 | 60.64 | 0.75 | 2.98 | 0.00 | 0.00 | 0.00 | 1.65 | 0.66 |
| Sarsiella zostericola | ostracod, seed shrimp | 4.30 | 17.62 | 13.72 | 140.08 | 10.84 | 16.16 | 2.15 | 0.75 | 0.74 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| ostracods, podocopid | ostracods, seed shrimps | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.70 | 0.00 | 0.82 | 3.69 | 0.00 | 3.30 | 1.46 | 3.44 | 9.42 |
| Squilla empusa larvae | mantis shrimp | 31.30 | 4.04 | 12.52 | 6.13 | 37.05 | 14.76 | 0.78 | 15.32 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| mysid sp. A | opossum shrimps, mysids | 0.00 | 0.00 | 0.72 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| (Heteromysis sp.) | opossum shrimp, mysid | 0.00 | 0.81 | 22.20 | 35.99 | 19.15 | 1.52 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Americamysis almyra | opossum shrimp, mysid | 2.71 | 15.16 | 298.67 | 342.84 | 523.53 | 767.63 | 550.35 | 1151.33 | 1115.95 | 637.73 | 5151.32 | 3267.56 | 2765.28 | 2871.09 |
| Americamysis bahia | opossum shrimp, mysid | 99.53 | 47.90 | 80.52 | 46.57 | 74.57 | 11.13 | 45.95 | 2.08 | 0.71 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| unidentified Americamysis juveniles | opossum shrimps, mysids | 38.66 | 102.63 | 149.71 | 553.95 | 722.52 | 1117.41 | 642.72 | 659.04 | 3093.45 | 1422.07 | 6066.30 | 4257.78 | 3308.92 | 3197.09 |
| Americamysis stucki | opossum shrimp, mysid | 23.44 | 24.07 | 1.99 | 5.33 | 9.97 | 5.06 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Bowmaniella dissimilis | opossum shrimp, mysid | 24.50 | 25.99 | 400.11 | 276.45 | 384.47 | 709.70 | 370.65 | 56.47 | 57.01 | 62.28 | 588.11 | 265.94 | 246.83 | 125.30 |
| Brasilomysis castroi | opossum shrimp, mysid | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.80 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Mysidopsis furca | opossum shrimp, mysid | 0.00 | 2.40 | 4.35 | 4.92 | 0.00 | 2.41 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Mysidopsis mortenseni | opossum shrimp, mysid | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Spelaeomysis sp. | opossum shrimp, mysid | 0.00 | 0.00 | 0.00 | 0.00 | 1.94 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Taphromysis bowmani | opossum shrimp, mysid | 0.00 | 0.81 | 0.00 | 0.77 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.52 | 2.41 | 2.47 | 1.82 | 4.90 |
| amphipods, gammaridean | amphipods | 820.19 | 2718.66 | 47107.32 | 58113.62 | 57022.28 | 94438.14 | 6060.19 | 5180.03 | 1739.67 | 718.78 | 5280.80 | 2057.11 | 1025.58 | 1578.27 |
| amphipods, caprellid | skeleton shrimps | 42.00 | 55.33 | 44.76 | 57.41 | 31.69 | 35.73 | 56.66 | 6.07 | 2.23 | 0.90 | 0.00 | 0.62 | 0.00 | 0.00 |
| Lestrigonus bengalensis | hyperiid amphipod | 0.00 | 1.41 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| isopod sp. a | isopod | 0.00 | 1.55 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Cyathura polita | isopod | 0.00 | 0.00 | 17.78 | 2.86 | 0.58 | 0.00 | 0.74 | 2.43 | 25.08 | 33.83 | 36.84 | 9.02 | 2.55 | 3.07 |
| Xenanthura brevitelson | isopod | 0.00 | 0.00 | 2.51 | 1.24 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.60 |
| Anopsilana jonesi | isopod | 1.89 | 1.35 | 0.00 | 0.00 | 0.66 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 4.22 | 19.27 | 19.99 | 3.67 |
| cymothoid sp. a (Lironeca) juveniles | isopod | 6.74 | 15.20 | 17.73 | 7.42 | 6.98 | 13.39 | 7.51 | 5.81 | 8.42 | 12.96 | 60.28 | 22.88 | 15.04 | 10.89 |
| Cassidinidea ovalis | isopod | 0.00 | 0.00 | 0.00 | 0.00 | 0.68 | 0.00 | 2.92 | 0.00 | 3.24 | 0.61 | 0.00 | 2.46 | 1.92 | 0.00 |
| Sphaeromatinae | isopods | 0.00 | 3.38 | 24.92 | 5.95 | 3.34 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Harrieta faxoni | isopod | 1.86 | 11.07 | 17.95 | 3.23 | 3.02 | 2.67 | 0.00 | 0.00 | 0.71 | 0.00 | 0.00 | 17.80 | 0.00 | 3.05 |
| Sphaeroma quadridentata | isopod | 0.00 | 0.00 | 1.68 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.60 | 0.00 | 0.00 | 0.00 |
| Sphaeroma terebrans | isopod | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.37 | 0.00 | 0.00 | 0.00 |
| Gnathia sp. larvae | isopod | 0.00 | 0.00 | 0.00 | 0.00 | 0.68 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Munna reynoldsi | isopod | 5.20 | 33.32 | 1.94 | 7.91 | 3.80 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.69 | 1.46 | 0.00 | 0.00 |
| Edotia triloba | isopod | 1.86 | 2.72 | 6.59 | 37.44 | 556.60 | 478.21 | 654.03 | 811.47 | 166.89 | 576.86 | 1993.28 | 382.13 | 186.45 | 441.42 |
| Erichsonella attenuata | isopod | 0.00 | 4.64 | 10.18 | 22.88 | 19.85 | 8.79 | 1.56 | 0.65 | 1.68 | 0.90 | 0.00 | 0.00 | 0.00 | 0.00 |
| Hoplomachus propinquus | tanaid | 0.00 | 0.00 | 0.64 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Hargeria rapax | tanaid | 2.03 | 21.17 | 762.92 | 528.79 | 299.78 | 289.06 | 18.36 | 0.00 | 3.35 | 0.00 | 0.00 | 0.62 | 0.00 | 0.00 |

Table A3, page 3 of 6. Location specific plankton-net catch .

Data are presented as mean number per 1,000 cubic meters.

Organisms are listed in phylogenetic order.

| Description | Common Name | Location (km from mouth) | | | | | | | | | | | | | |
|---|--------------------------|--------------------------|-----------|----------|----------|----------|----------|----------|----------|----------|---------|----------|----------|----------|---------|
| | | -5.9 | -3.6 | 2.5 | 5.2 | 7.7 | 10.6 | 16.2 | 20.0 | 24.2 | 26.9 | 30.2 | 34.4 | 37.1 | 41.0 |
| Sinelobus stanfordi | tanaid | 0.00 | 0.00 | 0.58 | 0.00 | 0.00 | 1.38 | 0.00 | 0.66 | 0.00 | 0.00 | 0.69 | 1.46 | 0.00 | 0.00 |
| Apseudes sp. | tanaid | 4.73 | 6.02 | 22.78 | 49.56 | 135.37 | 14.33 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| cumaceans | cumaceans | 593.18 | 1680.59 | 5475.80 | 7768.22 | 25417.10 | 32022.94 | 41196.21 | 2808.03 | 1906.59 | 1483.19 | 9777.29 | 718.50 | 255.93 | 66.39 |
| penaeid mysis larvae | penaeid shrimps | 1.89 | 4.21 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| penaeid postlarvae | penaeid shrimps | 43.53 | 10.00 | 3.36 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| penaeid metamorphs | penaeid shrimps | 77.77 | 73.99 | 102.14 | 127.05 | 36.96 | 30.39 | 7.22 | 0.72 | 0.00 | 0.68 | 2.69 | 0.00 | 0.00 | 0.00 |
| Farfantepenaeus duorarum juveniles | pink shrimp | 0.74 | 0.81 | 2.39 | 3.48 | 7.74 | 3.39 | 1.65 | 1.44 | 0.71 | 0.00 | 1.34 | 1.31 | 1.65 | 0.00 |
| Trachypenaeopsis mobilispinus juveniles | shrimps | 3.65 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Lucifer faxoni juveniles and adults | shrimp | 8223.14 | 8032.81 | 1359.70 | 806.69 | 362.04 | 210.06 | 13.99 | 50.41 | 0.00 | 3.42 | 0.00 | 0.00 | 0.67 | 1.32 |
| Lucifer faxoni mysis | shrimp | 20.53 | 6.44 | 1.65 | 0.00 | 0.00 | 0.00 | 0.78 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| shrimps, unidentified postlarvae | shrimps | 6.15 | 5.65 | 0.00 | 3.90 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| decapod zoeae | crab larvae | 166114.20 | 117315.14 | 69928.18 | 42275.55 | 44806.02 | 57840.04 | 22048.52 | 16770.87 | 27047.40 | 3888.72 | 47716.05 | 20350.19 | 48520.82 | 8104.80 |
| decapod megalopae | post-zoea crab larvae | 1308.93 | 2008.15 | 771.46 | 973.56 | 352.83 | 495.42 | 209.94 | 271.10 | 28.55 | 16.64 | 202.69 | 217.11 | 186.47 | 102.36 |
| decapod mysis | shrimp larvae | 32964.71 | 11790.86 | 24760.00 | 25591.61 | 48076.92 | 40749.22 | 3065.38 | 37535.83 | 223.59 | 63.12 | 134.95 | 124.58 | 59.08 | 54.60 |
| Leptochela serratorbita postlarvae | combcraw shrimp | 2.71 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Palaemon floridanus postlarvae | Florida grass shrimp | 206.00 | 168.32 | 308.28 | 180.74 | 106.90 | 122.99 | 16.92 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Periclimenes longicaudatus juveniles | longtail grass shrimp | 0.00 | 0.00 | 2.36 | 1.27 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Periclimenes spp. postlarvae | shrimps | 2.41 | 0.81 | 0.84 | 0.00 | 0.00 | 0.00 | 0.78 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Periclimenes spp. juveniles | shrimps | 0.00 | 13.12 | 16.84 | 1.47 | 2.46 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Palaemonetes paludosus juveniles | grass shrimp | 0.00 | 0.00 | 55.78 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Palaemonetes pugio juveniles | daggerblade grass shrimp | 0.00 | 4.86 | 9.73 | 4.56 | 0.64 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.24 | 0.63 | 1.30 |
| Palaemonetes pugio adults | daggerblade grass shrimp | 0.00 | 0.00 | 0.64 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.65 | 0.00 | 0.00 | 0.00 |
| Palaemonetes spp. postlarvae | grass shrimps | 71.88 | 206.75 | 120.65 | 204.22 | 253.73 | 3695.39 | 75.34 | 434.18 | 20.38 | 14.04 | 25.59 | 29.23 | 40.53 | 67.16 |
| Palaemonetes spp. juveniles | grass shrimps | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.76 | 0.00 | 0.00 |
| Palaemonetes vulgaris juveniles | grass shrimp | 0.00 | 0.00 | 0.64 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| alphaeid postlarvae | snapping shrimps | 1161.17 | 722.30 | 1406.33 | 1593.67 | 1066.74 | 1987.21 | 421.56 | 1156.76 | 1.68 | 2.42 | 3.06 | 0.00 | 3.72 | 1.31 |
| alphaeid juveniles | snapping shrimps | 0.00 | 21.23 | 1.98 | 0.60 | 0.00 | 2.67 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Alpheus estuariensis juveniles | snapping shrimp | 0.00 | 0.81 | 0.59 | 1.27 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Hippolyte zostericola postlarvae | zostera shrimp | 1009.29 | 1259.07 | 301.28 | 173.75 | 151.90 | 251.58 | 67.42 | 38.08 | 3.28 | 1.81 | 0.61 | 0.00 | 0.67 | 15.35 |
| Hippolyte zostericola juveniles | zostera shrimp | 18.12 | 40.18 | 6.26 | 3.00 | 1.32 | 1.30 | 0.78 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Hippolyte zostericola adults | zostera shrimp | 6.05 | 4.84 | 0.66 | 0.00 | 0.00 | 0.00 | 0.00 | 4.80 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Latreutes parvulus postlarvae | sargassum shrimp | 0.00 | 0.00 | 0.00 | 0.00 | 71.85 | 39.13 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Tozeuma carolinense postlarvae | arrow shrimp | 211.64 | 166.95 | 230.57 | 24.40 | 6.09 | 25.65 | 2.92 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Tozeuma carolinense juveniles | arrow shrimp | 31.06 | 27.80 | 0.72 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Ogyrides alphaerostris postlarvae | estuarine longeye shrimp | 0.00 | 0.00 | 0.00 | 2.23 | 0.66 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| processid postlarvae | night shrimps | 1149.87 | 475.20 | 295.77 | 125.51 | 68.70 | 111.53 | 85.25 | 31.36 | 0.82 | 1.81 | 0.00 | 0.00 | 0.00 | 0.00 |
| processid juveniles | night shrimps | 0.00 | 0.00 | 0.72 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Ambidexter symmetricus juveniles | shrimp | 0.00 | 7.02 | 1.98 | 0.81 | 0.66 | 0.80 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Upogebia spp. postlarvae | mud shrimps | 3860.16 | 3414.50 | 3502.49 | 2288.76 | 3385.61 | 2047.97 | 331.44 | 2374.58 | 0.74 | 0.00 | 0.00 | 0.00 | 0.00 | 1.40 |
| Upogebia spp. juveniles | mud shrimps | 6.86 | 26.41 | 35.24 | 3.23 | 18.18 | 61.36 | 6.40 | 2.70 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Eucерamus praelongus juveniles | olivepit porcelain crab | 7.31 | 2.40 | 0.00 | 0.00 | 2.64 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Petrolisthes armatus juveniles | porcelain crab | 26.14 | 53.96 | 14.16 | 11.49 | 0.00 | 3.09 | 2.92 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Crab, Emerita sp. juveniles | mole crab | 0.00 | 0.00 | 0.00 | 4.04 | 44.51 | 2.67 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| paguroid megalops larvae | hermit crabs | 3126.23 | 1033.72 | 659.94 | 246.50 | 403.98 | 42.44 | 3.29 | 0.71 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| paguroid juveniles | hermit crabs | 59.68 | 52.67 | 11.70 | 6.54 | 0.66 | 6.07 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Portunus sp. juveniles | swimming crab | 4.54 | 2.31 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| xanthid juveniles | mud crabs | 0.00 | 2.43 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.83 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Pinnixa sayana juveniles | pea crab | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Uca spp. juveniles | fiddler crabs | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.78 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| ephemeropteran larvae | mayflies | 0.00 | 0.00 | 0.00 | 0.00 | 0.73 | 2.20 | 36.13 | 84.07 | 0.00 | 10.52 | 10.84 | 5.36 | 13.09 | 31.52 |
| odonates, anisopteran larvae | dragonflies | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.65 | 0.00 | 0.00 | 0.65 | 0.00 | 0.63 | 1.30 |
| odonates, zygopteran larvae | damselflies | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 7.39 | 6.59 | 0.00 | 1.50 | 0.00 | 0.62 | 1.89 | 2.41 |
| hemipterans, corixid adults | water boatmen | 0.00 | 0.00 | 0.00 | 0.60 | 0.00 | 0.00 | 1.64 | 4.12 | 6.03 | 0.00 | 0.74 | 0.67 | 0.00 | 1.31 |

Table A3, page 4 of 6. Location specific plankton-net catch .

Data are presented as mean number per 1,000 cubic meters.

Organisms are listed in phylogenetic order.

| Description | Common Name | Location (km from mouth) | | | | | | | | | | | | | |
|---|---------------------------|--------------------------|---------|---------|----------|---------|----------|---------|----------|--------|---------|---------|---------|---------|---------|
| | | -5.9 | -3.6 | 2.5 | 5.2 | 7.7 | 10.6 | 16.2 | 20.0 | 24.2 | 26.9 | 30.2 | 34.4 | 37.1 | 41.0 |
| hemipterans, corixid juveniles | water boatmen | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.64 | 0.60 | 0.00 | 0.00 | 0.00 |
| coleopterans, dytiscid adults | predaceous diving beetles | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.64 | 0.00 | 0.00 |
| coleopterans, curculionid adults | beetles | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.66 | 0.00 | 0.00 | 0.00 | 0.00 |
| coleopterans, elmid larvae | rifle beetles | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.66 |
| coleopterans, elmid adults | rifle beetles | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.66 | 0.00 | 0.00 | 0.62 | 0.62 | 0.00 | 0.66 |
| dipterans, pupae | flies, mosquitoes | 0.80 | 0.71 | 3.79 | 4.58 | 15.09 | 10.73 | 27.89 | 16.85 | 29.71 | 60.33 | 195.22 | 681.71 | 528.07 | 1022.89 |
| dipterans, tabanid larvae | deer flies | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.32 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| dipterans, ceratopogonid larvae | biting midges | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.65 | 0.00 | 0.75 | 0.00 | 0.67 | 0.00 | 2.02 |
| dipteran, Chaoborus punctipennis larvae | phantom midge | 8.48 | 5.54 | 35.50 | 39.54 | 56.24 | 16.75 | 67.50 | 261.89 | 551.21 | 425.28 | 1853.67 | 3682.38 | 3488.37 | 6588.09 |
| dipterans, chironomid larvae | midges | 0.00 | 0.00 | 2.33 | 0.00 | 0.73 | 0.59 | 0.00 | 2.98 | 2.61 | 3.39 | 5.80 | 13.53 | 14.26 | 8.40 |
| trichopteran larvae | caddisflies | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.64 | 1.65 | 0.00 | 0.00 | 0.65 | 0.00 | 5.20 | 0.67 |
| brachiopod, Glottidia pyramidata larvae | lamp shell | 4.94 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| chaetognaths, sagittid | arrow worms | 9376.98 | 5413.52 | 3070.62 | 4891.25 | 4180.66 | 6280.99 | 5501.39 | 3578.18 | 136.26 | 1242.94 | 173.74 | 41.99 | 6.12 | 20.32 |
| ascidiacean larvae | tunicate larvae | 0.00 | 0.00 | 20.72 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| appendicularian, Okolpeura dioica | larvacean | 3016.81 | 2241.09 | 1425.75 | 10782.96 | 1332.65 | 5821.61 | 9309.40 | 14449.46 | 156.01 | 21.19 | 2.51 | 2.94 | 0.00 | 0.00 |
| Elops saurus postflexion larvae | ladyfish | 0.00 | 0.00 | 0.54 | 1.32 | 0.00 | 2.93 | 0.00 | 0.65 | 0.00 | 7.00 | 1.38 | 1.99 | 0.00 | 1.26 |
| Myrophis punctatus juveniles | speckled worm eel | 0.00 | 0.00 | 0.00 | 0.62 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| clupeid eggs | herrings | 113.38 | 204.29 | 139.52 | 111.53 | 59.23 | 366.07 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| clupeid preflexion larvae | herrings | 111.94 | 29.12 | 89.31 | 54.74 | 78.27 | 104.74 | 20.18 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Brevoortia spp. postflexion larvae | menhaden | 1.57 | 0.00 | 1.08 | 0.62 | 0.00 | 0.00 | 0.81 | 0.00 | 0.66 | 2.55 | 5.10 | 0.00 | 0.69 | 0.67 |
| Brevoortia spp. metamorphs | menhaden | 0.73 | 0.64 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Harengula jaguana eggs | scaled sardine | 0.00 | 0.00 | 0.84 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Harengula jaguana flexion larvae | scaled sardine | 0.00 | 0.00 | 0.84 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Harengula jaguana postflexion larvae | scaled sardine | 0.00 | 3.11 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Harengula jaguana metamorphs | scaled sardine | 0.00 | 0.00 | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Harengula jaguana juveniles | scaled sardine | 0.00 | 0.81 | 0.00 | 0.00 | 0.68 | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Opisthonema oglinum flexion larvae | Atlantic thread herring | 5.43 | 2.40 | 2.60 | 0.00 | 0.78 | 0.00 | 0.00 | 2.70 | 0.00 | 0.00 | 0.00 | 0.00 | 1.42 | 1.37 |
| Opisthonema oglinum postflexion larvae | Atlantic thread herring | 2.71 | 1.55 | 7.52 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 3.43 |
| Opisthonema oglinum juveniles | Atlantic thread herring | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.25 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Sardinella aurita flexion larvae | Spanish sardine | 5.25 | 0.00 | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Sardinella aurita postflexion larvae | Spanish sardine | 0.00 | 0.00 | 1.28 | 0.00 | 1.56 | 4.45 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Anchoa spp. preflexion larvae | anchovies | 950.53 | 554.97 | 934.10 | 1790.09 | 1407.80 | 3497.84 | 6076.67 | 1328.32 | 167.11 | 291.24 | 336.58 | 90.34 | 22.86 | 3.53 |
| Anchoa spp. flexion larvae | anchovies | 13.28 | 44.30 | 81.93 | 62.36 | 65.34 | 113.25 | 70.03 | 32.28 | 43.97 | 32.62 | 48.33 | 23.79 | 1.65 | 10.09 |
| Anchoa hepsetus eggs | striped anchovy | 3.66 | 2.17 | 7.31 | 0.00 | 5.29 | 12.47 | 52.60 | 2.85 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Anchoa hepsetus postflexion larvae | striped anchovy | 0.00 | 0.00 | 0.00 | 1.51 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Anchoa hepsetus adults | striped anchovy | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.69 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Anchoa mitchilli eggs | bay anchovy | 9.52 | 23.20 | 135.87 | 2148.51 | 1668.73 | 10108.77 | 3614.06 | 24302.72 | 131.84 | 3.70 | 3.46 | 109.74 | 55.14 | 23.31 |
| Anchoa mitchilli postflexion larvae | bay anchovy | 58.07 | 94.48 | 143.75 | 129.49 | 176.04 | 356.38 | 130.01 | 592.03 | 113.81 | 20.62 | 50.59 | 60.13 | 43.49 | 34.24 |
| Anchoa mitchilli juveniles | bay anchovy | 3.02 | 9.83 | 21.51 | 40.56 | 35.29 | 30.21 | 20.37 | 24.38 | 33.77 | 14.50 | 298.10 | 144.56 | 159.73 | 193.29 |
| Anchoa mitchilli adults | bay anchovy | 0.00 | 5.62 | 7.71 | 3.02 | 1.28 | 0.00 | 2.35 | 5.67 | 9.88 | 79.65 | 14.71 | 64.69 | 39.55 | 36.46 |
| Notemigonus crysoleucas preflexion larvae | golden shiner | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.79 |
| Hoplosternum littorale juveniles | brown hoplo catfish | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.92 | 0.00 | 1.20 |
| Liposarcus spp. juveniles | suckermouth catfish | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.60 | 0.62 | 2.61 | 0.70 | 0.00 |
| Opsanus beta juveniles | gulf toadfish | 0.00 | 0.00 | 2.28 | 0.80 | 1.62 | 0.75 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Labidesthes sicculus preflexion larvae | brook silverside | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.69 |
| Labidesthes sicculus flexion larvae | brook silverside | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.69 |
| Membras martinica preflexion larvae | rough silverside | 0.00 | 0.81 | 0.00 | 0.00 | 1.21 | 0.00 | 4.10 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.71 |
| Membras martinica flexion larvae | rough silverside | 0.00 | 0.00 | 2.36 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Membras martinica juveniles | rough silverside | 0.00 | 0.00 | 0.00 | 0.00 | 0.64 | 0.00 | 0.00 | 0.84 | 0.00 | 1.39 | 0.00 | 0.00 | 0.00 | 1.38 |
| Membras martinica adults | rough silverside | 0.00 | 0.00 | 0.00 | 0.63 | 0.67 | 0.00 | 0.67 | 0.00 | 0.68 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Menidia spp. preflexion larvae | silversides | 0.00 | 0.00 | 1.30 | 0.00 | 0.61 | 4.22 | 1.27 | 1.45 | 1.28 | 1.40 | 2.03 | 0.00 | 0.00 | 3.40 |
| Menidia spp. flexion larvae | silversides | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.70 |
| Menidia spp. juveniles | silversides | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.82 | 0.64 | 0.00 | 3.49 | 0.65 | 1.24 | 0.61 | 3.04 |

Table A3, page 5 of 6. Location specific plankton-net catch .

Data are presented as mean number per 1,000 cubic meters.

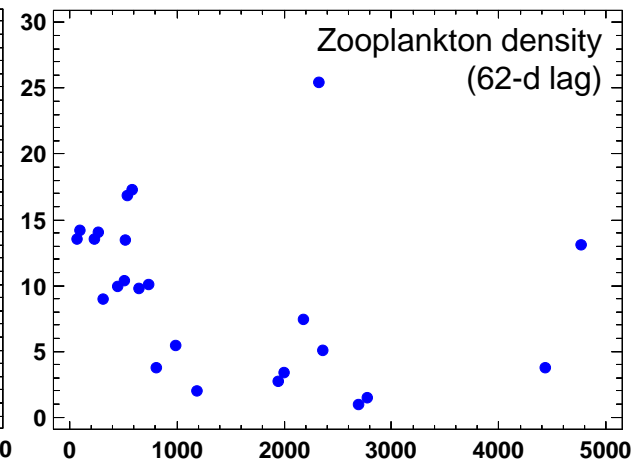
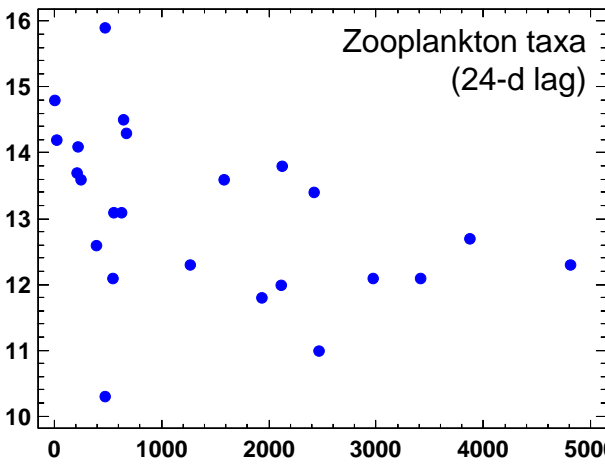
Organisms are listed in phylogenetic order.

| Description | Common Name | Location (km from mouth) | | | | | | | | | | | | | |
|--|---------------------------|--------------------------|---------|---------|---------|---------|---------|----------|----------|---------|--------|-------|--------|--------|-------|
| | | -5.9 | -3.6 | 2.5 | 5.2 | 7.7 | 10.6 | 16.2 | 20.0 | 24.2 | 26.9 | 30.2 | 34.4 | 37.1 | 41.0 |
| Gambusia holbrooki juveniles | eastern mosquitofish | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.54 | 1.26 | 1.90 |
| Synodus foetens postflexion larvae | inshore lizardfish | 0.00 | 0.64 | 0.69 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Hippocampus erectus juveniles | lined seahorse | 0.00 | 3.03 | 2.53 | 2.42 | 0.66 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Hippocampus zosterae juveniles | dwarf seahorse | 0.00 | 0.68 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Cosmocampus hildebrandi juveniles | dwarf pipefish | 0.00 | 1.37 | 0.00 | 0.69 | 0.00 | 0.00 | 0.00 | 0.00 | 0.82 | 3.34 | 0.69 | 6.34 | 1.89 | 3.18 |
| Syngnathus floridae juveniles | dusky pipefish | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.72 | 0.00 | 0.00 | 0.00 | 0.00 |
| Syngnathus louisianae juveniles | chain pipefish | 3.85 | 3.26 | 2.57 | 0.00 | 0.68 | 0.73 | 3.29 | 0.00 | 0.00 | 0.00 | 0.00 | 5.94 | 4.18 | 4.95 |
| Syngnathus scovelli juveniles | gulf pipefish | 18.14 | 7.91 | 10.18 | 7.64 | 4.62 | 5.72 | 4.04 | 0.72 | 0.71 | 3.00 | 1.39 | 4.97 | 4.62 | 2.80 |
| Syngnathus scovelli adults | gulf pipefish | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.72 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Prionotus spp. postflexion larvae | searobins | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Mugil cephalus juveniles | striped mullet | 0.00 | 0.00 | 0.69 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.64 | 0.63 | 0.68 | 0.00 | 0.00 |
| fish eggs, percomorph | sciaenid eggs (primarily) | 1240.97 | 3377.07 | 1768.32 | 2630.03 | 4083.64 | 4079.15 | 30022.39 | 23677.13 | 2604.91 | 130.36 | 13.20 | 353.15 | 422.27 | 67.60 |
| Lepomis macrochirus juveniles | bluegill | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.87 | 0.00 | 0.00 | 2.76 | 0.00 | 2.41 |
| Lepomis spp. juveniles | sunfishes | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.69 | 0.63 | 0.00 |
| Chloroscombrus chrysurus preflexion larvae | Atlantic bumper | 18.22 | 4.13 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Chloroscombrus chrysurus flexion larvae | Atlantic bumper | 7.56 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Chloroscombrus chrysurus postflexion larvae | Atlantic bumper | 0.00 | 0.00 | 0.58 | 0.00 | 0.00 | 0.00 | 0.00 | 2.70 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Chloroscombrus chrysurus juveniles | Atlantic bumper | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.69 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Oligoplites saurus preflexion larvae | leatherjack | 3.65 | 0.67 | 0.00 | 0.00 | 0.00 | 5.94 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Oligoplites saurus flexion larvae | leatherjack | 0.00 | 2.55 | 0.00 | 1.95 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Oligoplites saurus postflexion larvae | leatherjack | 0.00 | 0.89 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Oligoplites saurus juveniles | leatherjack | 0.00 | 0.78 | 0.00 | 0.00 | 0.73 | 0.75 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Uraspis secunda juveniles | cottonmouth jack | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.76 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Lutjanus griseus juveniles | gray snapper | 0.74 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Eucinostomus spp. postflexion larvae | mojarra | 1.99 | 4.05 | 0.70 | 0.66 | 0.00 | 1.84 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Eucinostomus spp. juveniles | mojarra | 4.73 | 0.00 | 0.00 | 0.77 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Eugerres plumieri postflexion larvae | striped mojarra | 0.00 | 0.00 | 1.65 | 0.00 | 0.00 | 2.67 | 0.00 | 5.41 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| gerreid preflexion larvae | mojarra | 73.80 | 61.67 | 104.90 | 8.92 | 65.07 | 128.14 | 26.76 | 45.59 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| gerreid flexion larvae | mojarra | 0.80 | 0.00 | 1.65 | 0.00 | 0.00 | 0.00 | 0.00 | 5.41 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Archosargus probatocephalus postflexion larvae | sheepshead | 0.00 | 0.72 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Archosargus probatocephalus juveniles | sheepshead | 14.66 | 1.61 | 10.81 | 0.70 | 0.68 | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Bairdiella chrysoura preflexion larvae | silver perch | 38.78 | 4.46 | 2.77 | 3.83 | 0.66 | 8.25 | 3.29 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Bairdiella chrysoura flexion larvae | silver perch | 0.00 | 0.00 | 4.52 | 0.00 | 0.00 | 0.63 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Bairdiella chrysoura postflexion larvae | silver perch | 0.00 | 0.00 | 0.60 | 0.00 | 3.25 | 1.88 | 2.92 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Bairdiella chrysoura juveniles | silver perch | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 2.04 | 0.00 | 0.00 | 0.00 |
| Cynoscion arenarius preflexion larvae | sand seatrout | 33.53 | 0.81 | 5.47 | 3.83 | 14.02 | 6.55 | 54.00 | 3.36 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Cynoscion arenarius flexion larvae | sand seatrout | 3.10 | 2.53 | 0.58 | 0.00 | 0.00 | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Cynoscion arenarius postflexion larvae | sand seatrout | 1.86 | 12.57 | 0.69 | 1.27 | 0.67 | 0.63 | 10.52 | 0.00 | 0.68 | 2.83 | 0.00 | 0.00 | 0.00 | 0.00 |
| Cynoscion arenarius juveniles | sand seatrout | 0.00 | 0.00 | 0.00 | 0.66 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 1.02 | 1.26 | 1.42 | 0.77 | 0.00 |
| Cynoscion nebulosus preflexion larvae | spotted seatrout | 43.37 | 34.48 | 12.28 | 0.63 | 3.78 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Cynoscion nebulosus flexion larvae | spotted seatrout | 1.86 | 0.00 | 0.00 | 1.95 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Cynoscion nebulosus postflexion larvae | spotted seatrout | 0.00 | 2.16 | 0.00 | 1.95 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Leiostomus xanthurus postflexion larvae | spot | 0.00 | 2.40 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Menticirrhus spp. preflexion larvae | kingfishes | 76.49 | 14.68 | 52.62 | 43.39 | 26.25 | 37.75 | 24.00 | 17.40 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Menticirrhus spp. postflexion larvae | kingfishes | 0.00 | 2.06 | 2.60 | 0.00 | 0.00 | 1.84 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Pogonias cromis postflexion larvae | black drum | 1.89 | 2.53 | 0.00 | 0.80 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Pogonias cromis juveniles | black drum | 1.89 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Chaetodipterus faber preflexion larvae | Atlantic spadefish | 7.02 | 3.48 | 7.42 | 1.61 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Chasmodes saburrae flexion larvae | Florida blenny | 13.06 | 10.87 | 35.35 | 4.95 | 3.50 | 56.22 | 33.55 | 8.97 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.71 |
| Chasmodes saburrae postflexion larvae | Florida blenny | 14.57 | 9.39 | 11.11 | 2.23 | 29.66 | 36.79 | 24.61 | 6.27 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Chasmodes saburrae juveniles | Florida blenny | 0.00 | 1.35 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Lupinoblennius nicholsi flexion larvae | highfin blenny | 0.00 | 0.81 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |
| Lupinoblennius nicholsi postflexion larvae | highfin blenny | 0.00 | 0.00 | 0.00 | 0.00 | 2.61 | 0.00 | 0.00 | 2.21 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 | 0.00 |

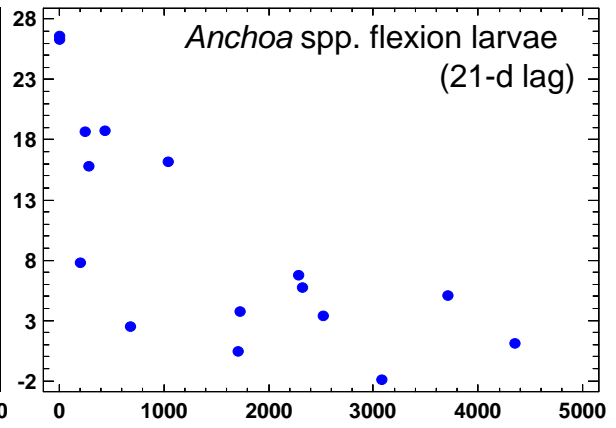
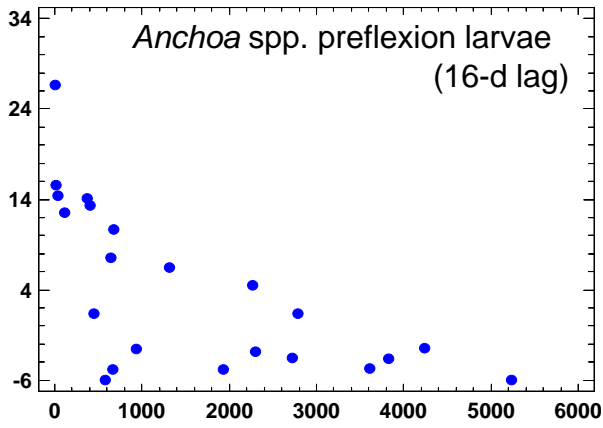
Appendix G

Location (center of abundance) responses to inflow

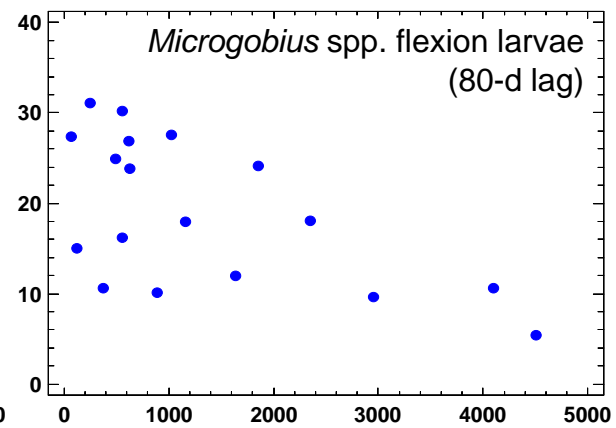
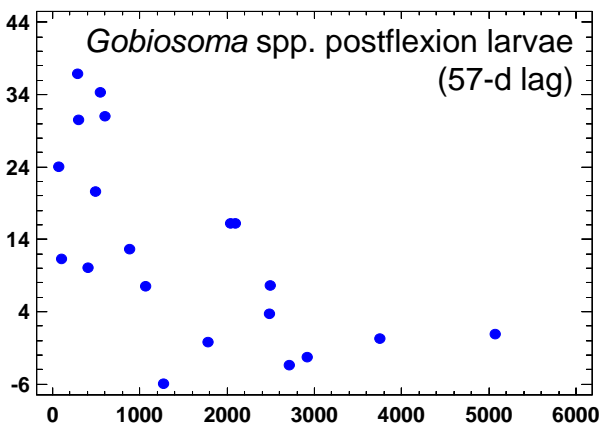
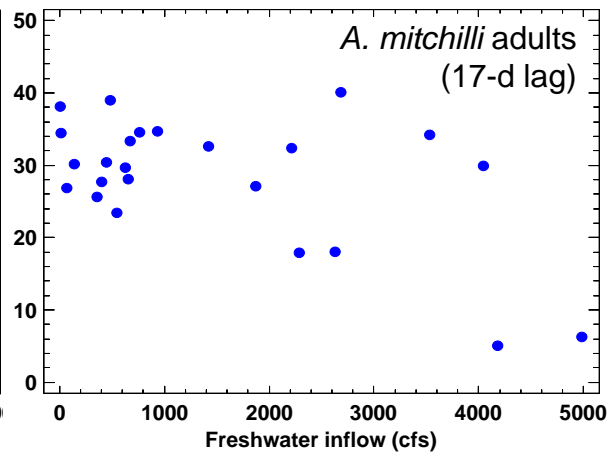
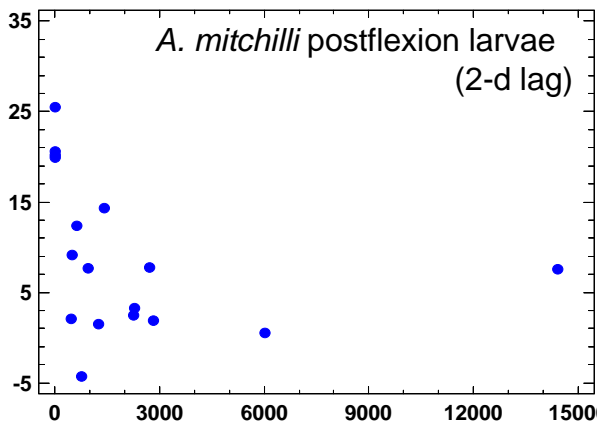
Center of abundance (rkm)



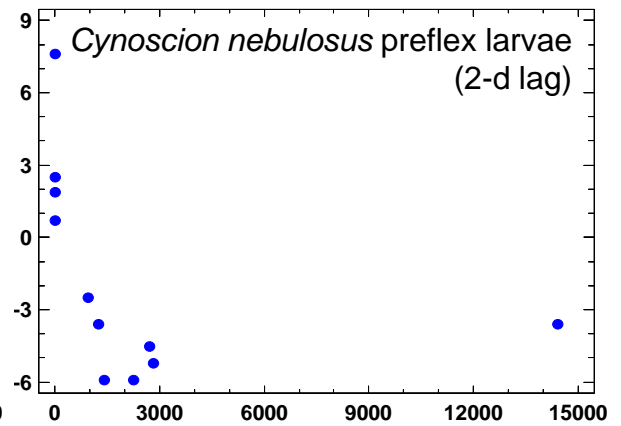
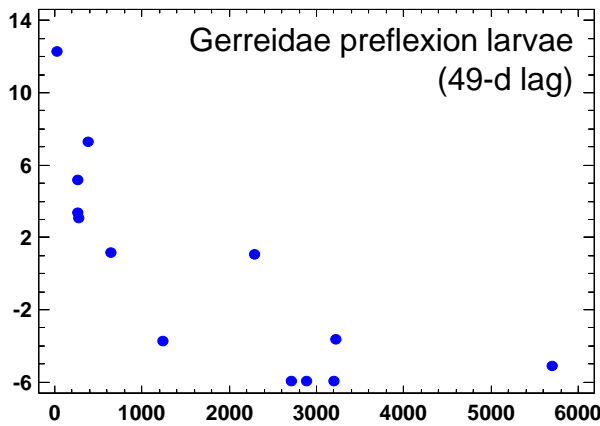
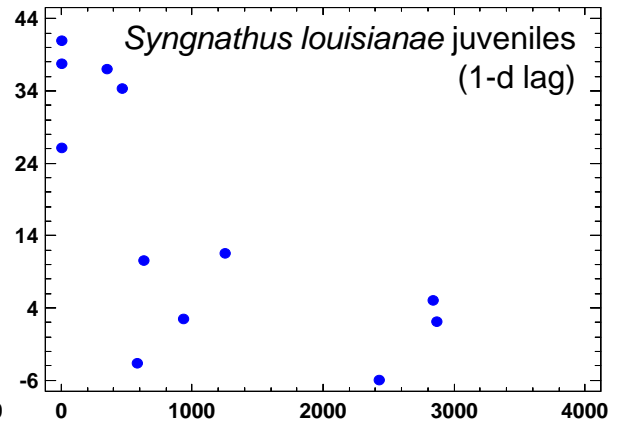
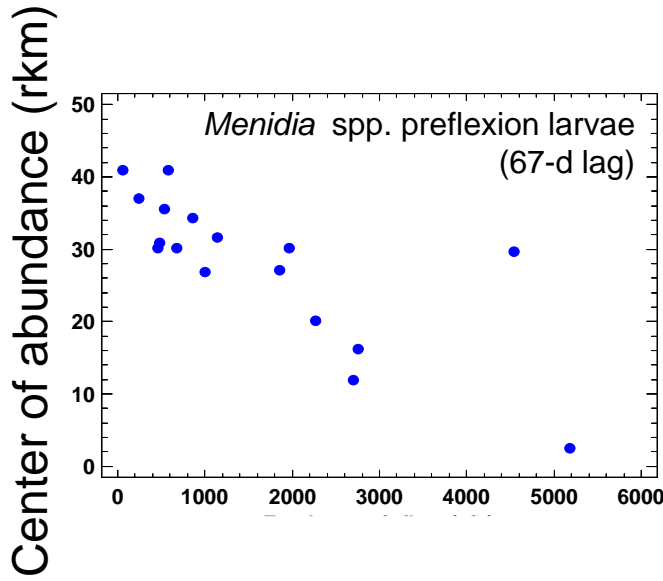
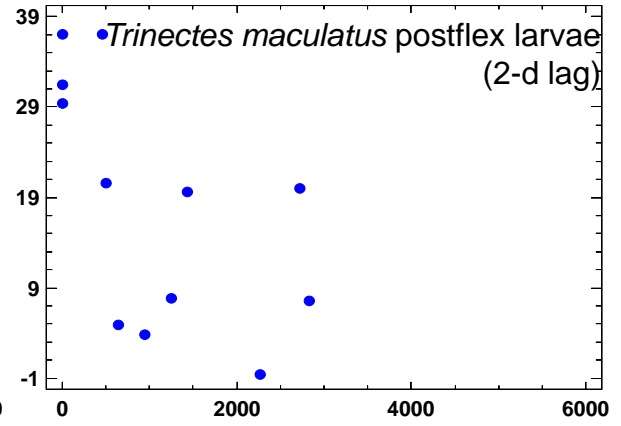
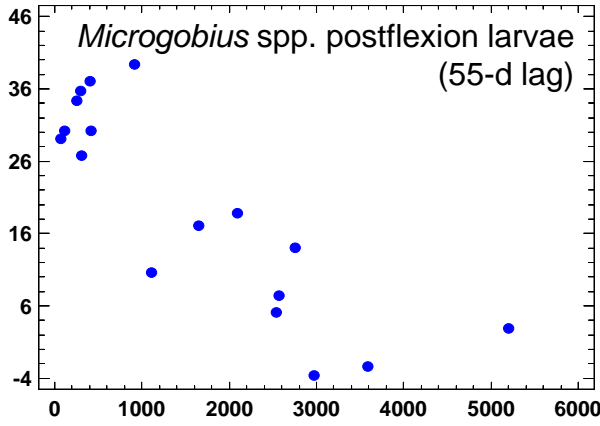
Freshwater inflow (cfs)



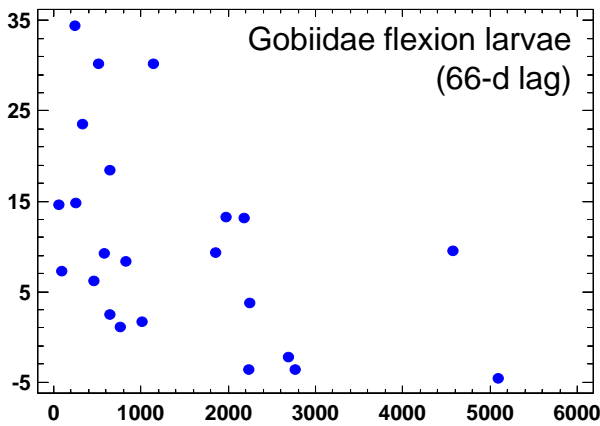
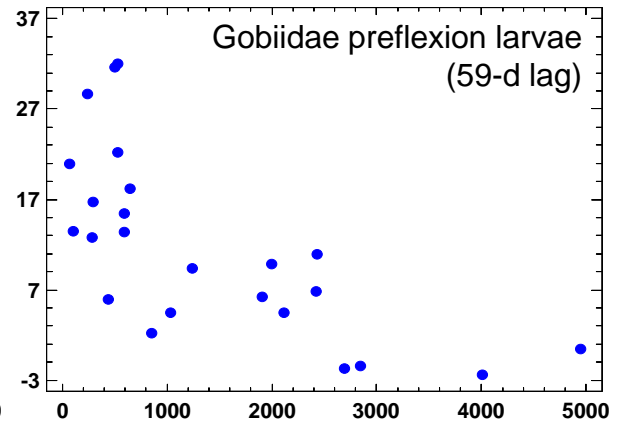
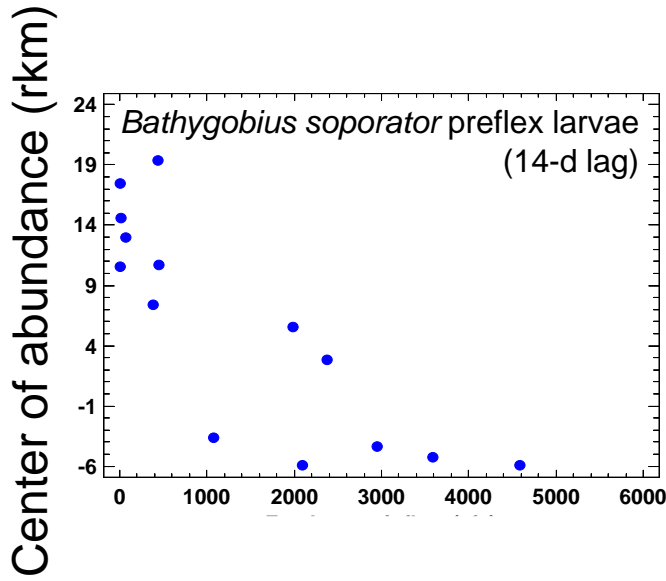
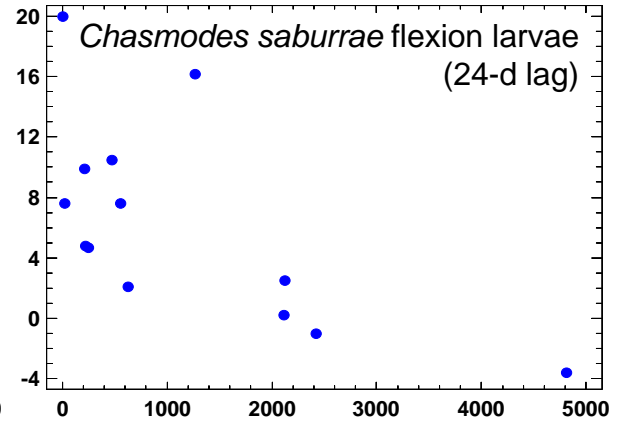
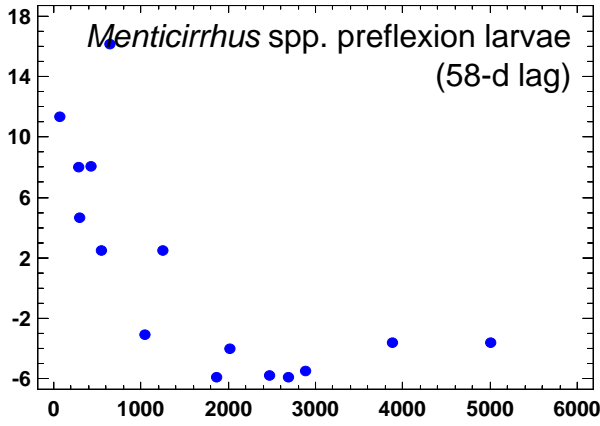
Center of abundance (rkm)



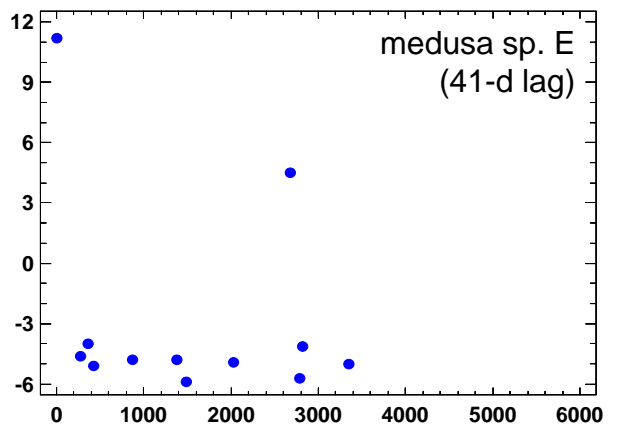
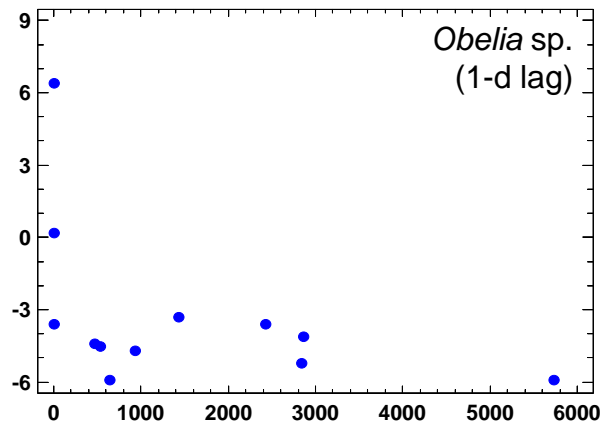
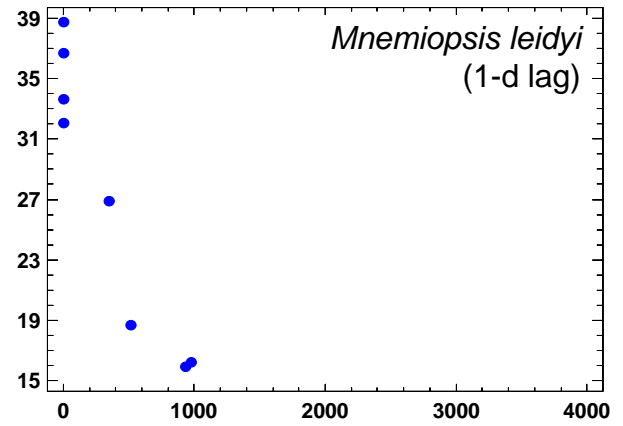
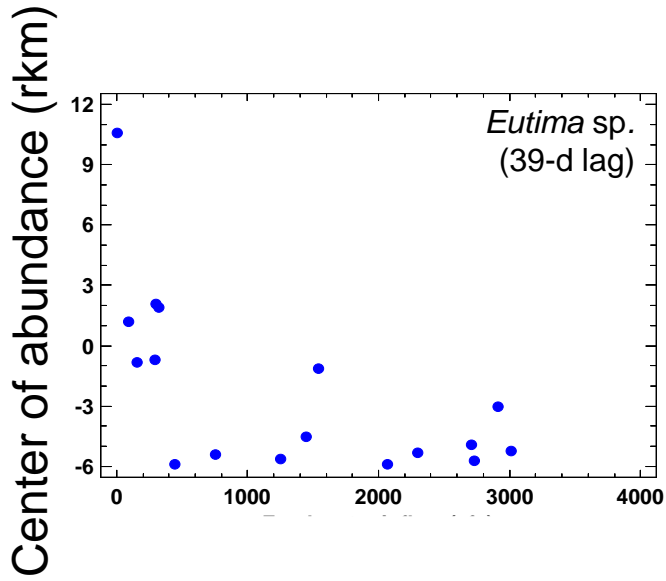
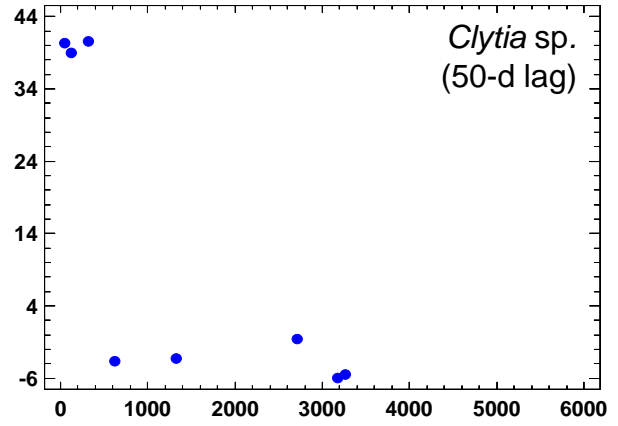
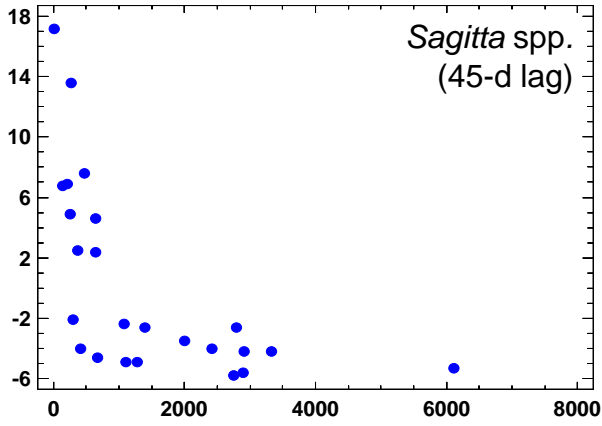
Freshwater inflow (cfs)



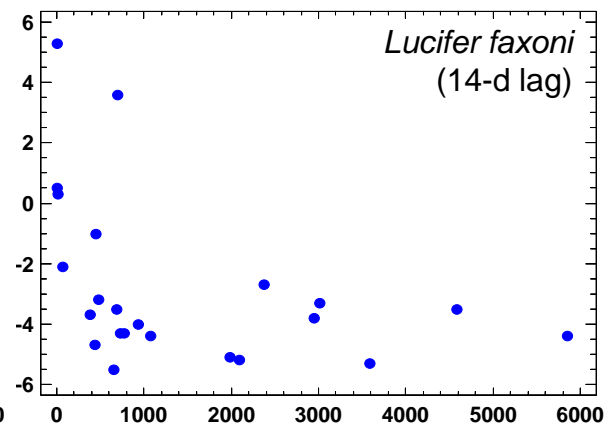
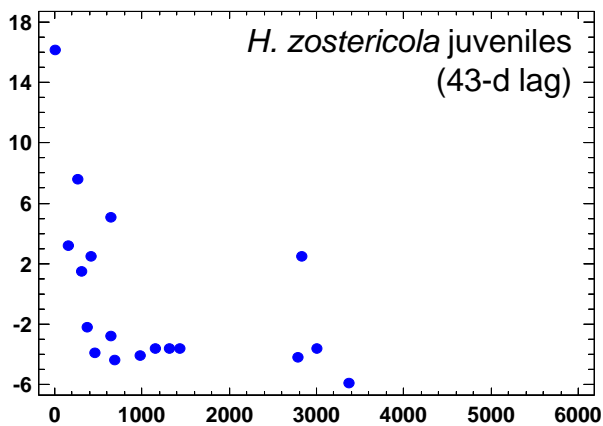
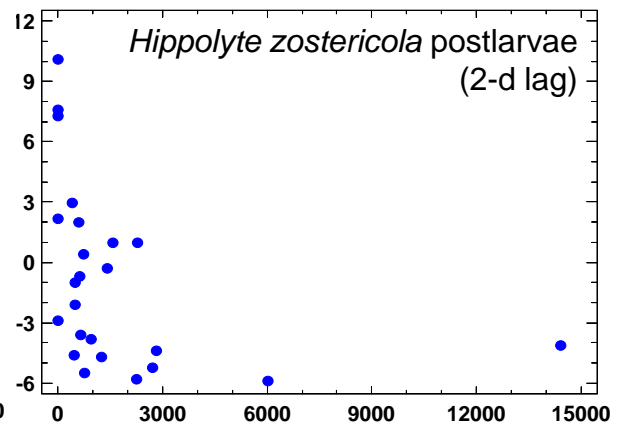
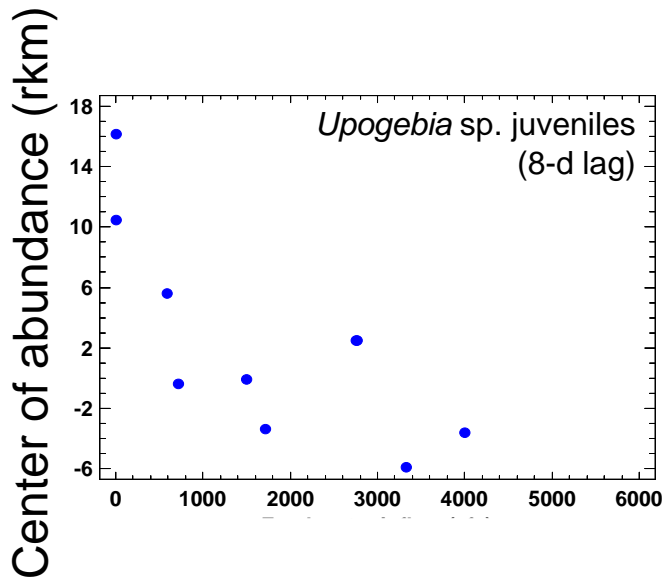
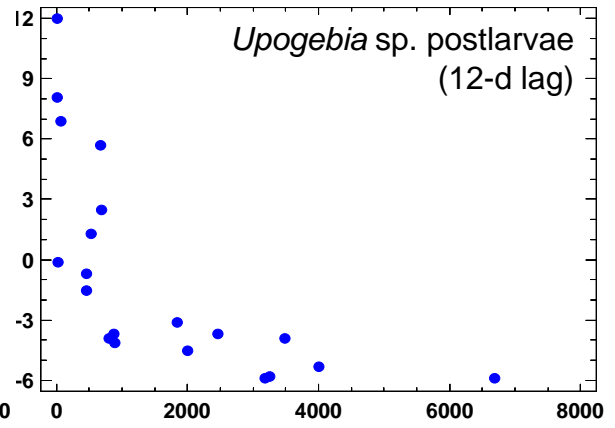
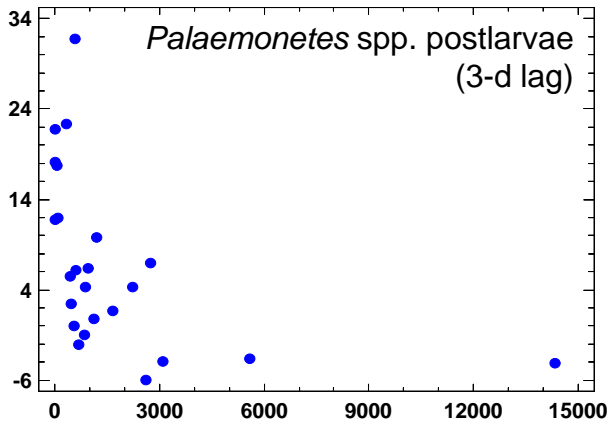
Freshwater inflow (cfs)



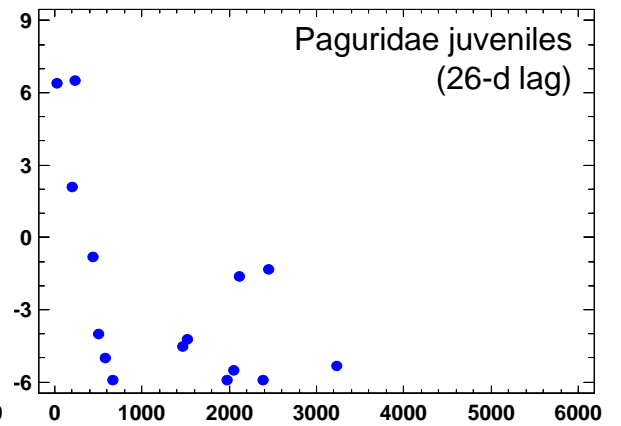
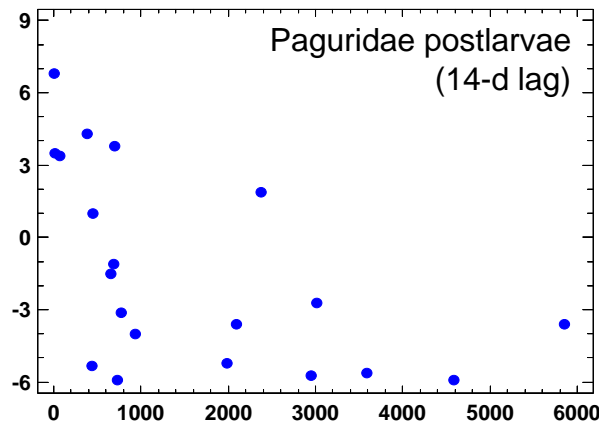
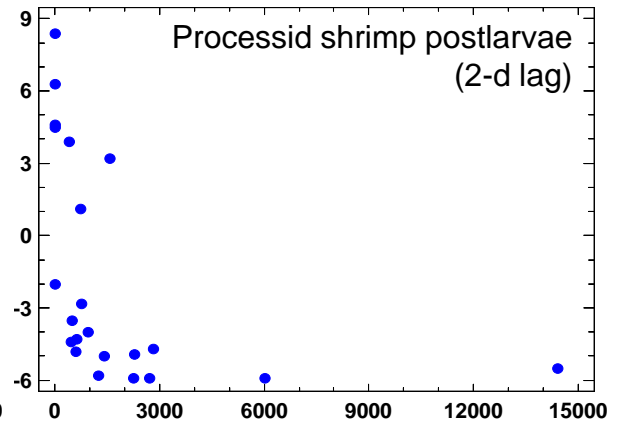
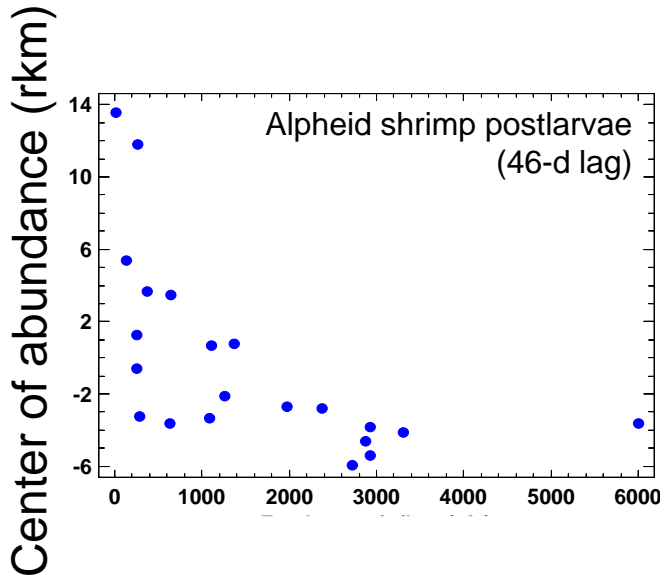
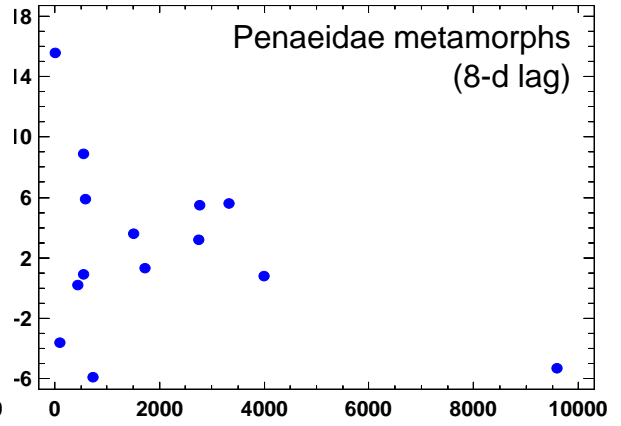
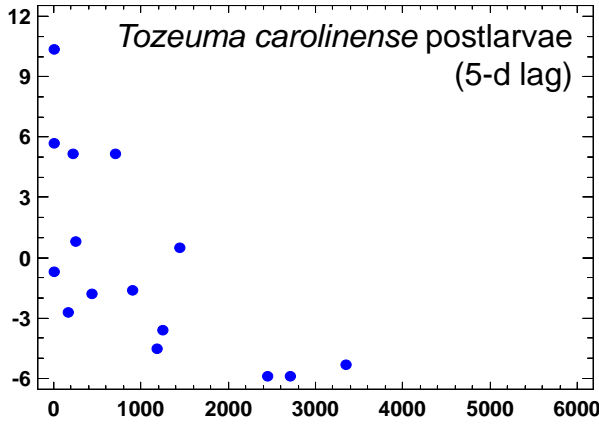
Freshwater inflow (cfs)



Freshwater inflow (cfs)

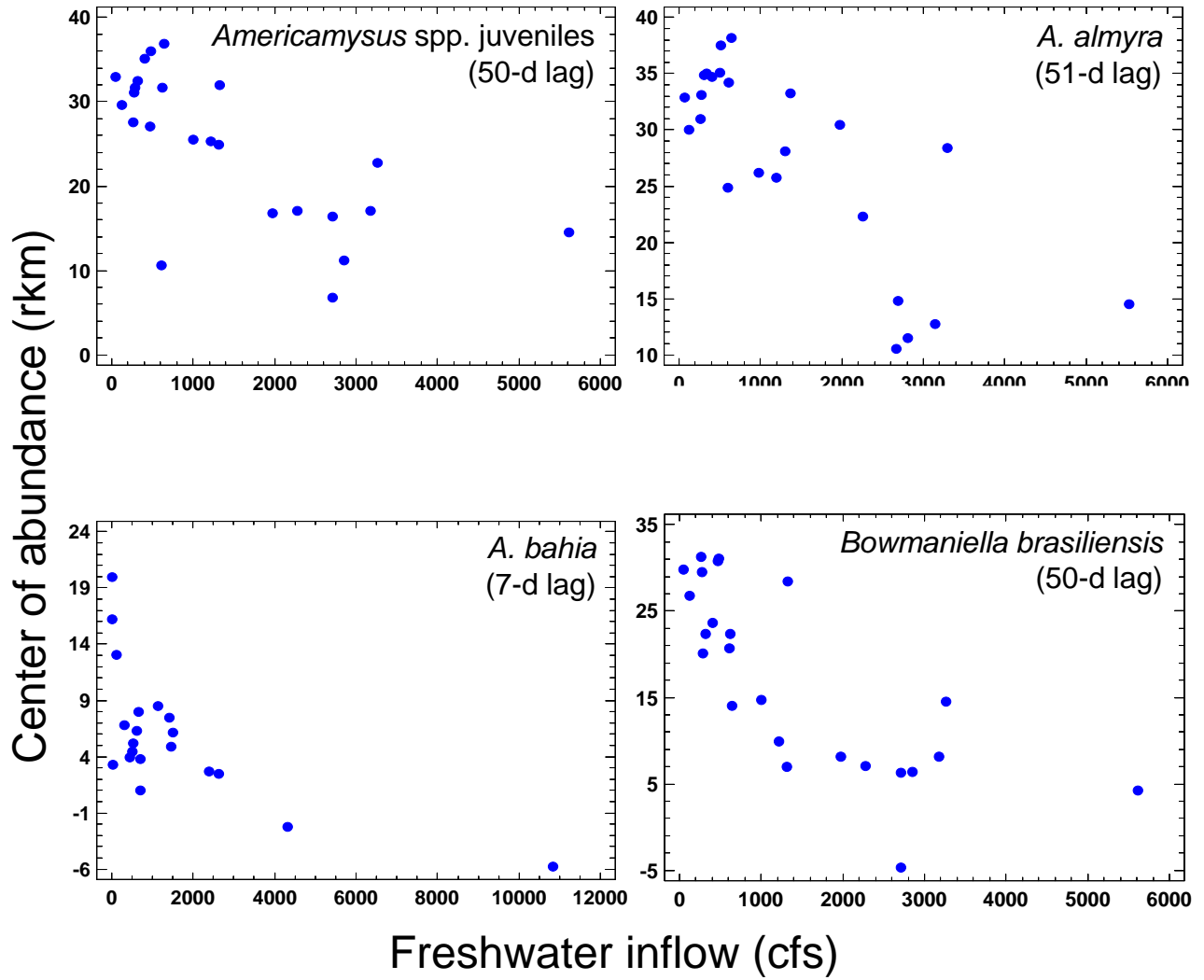


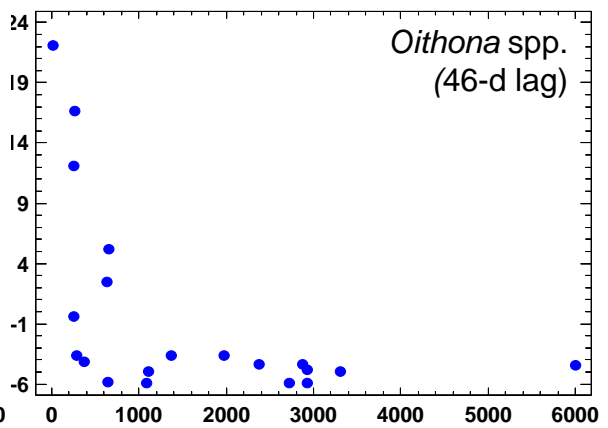
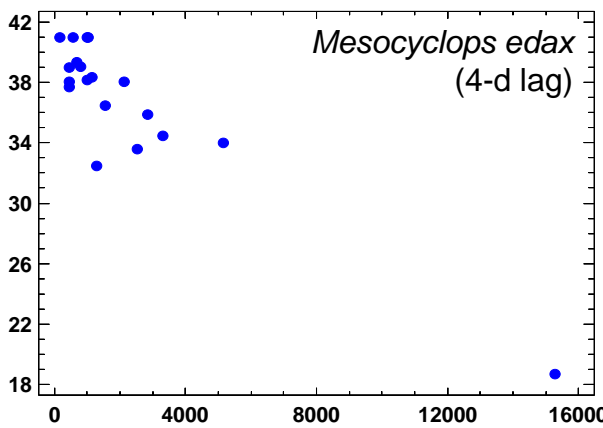
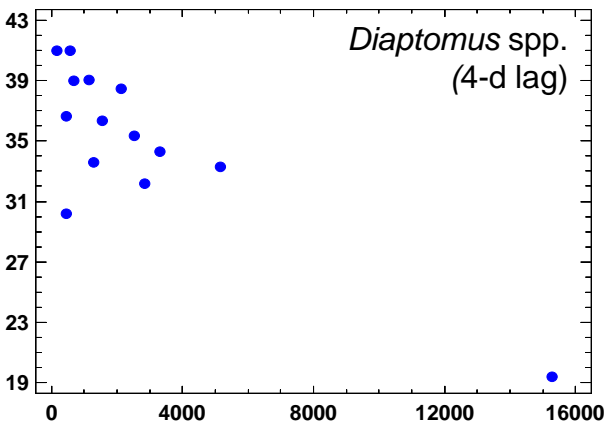
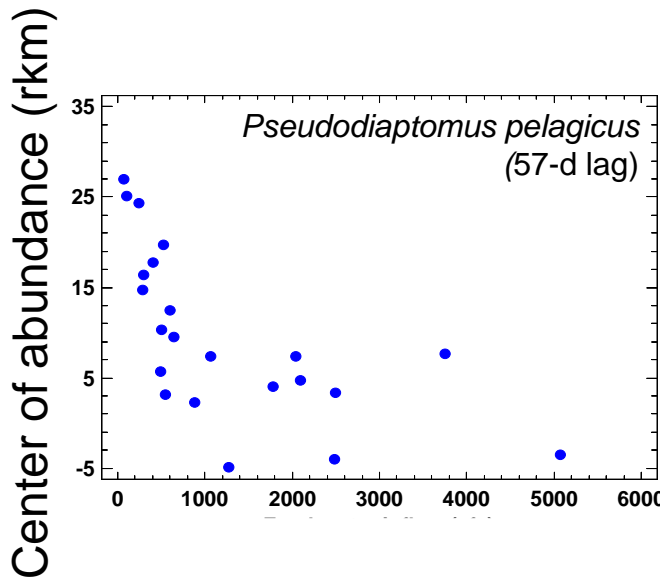
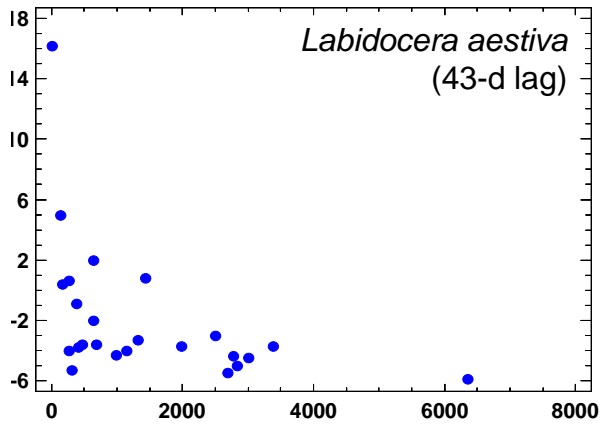
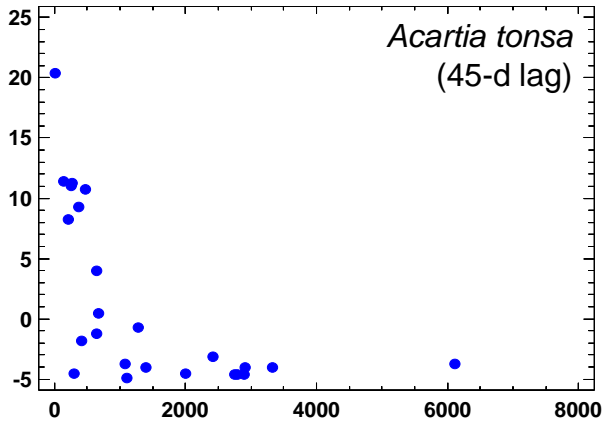
Freshwater inflow (cfs)



Freshwater inflow (cfs)

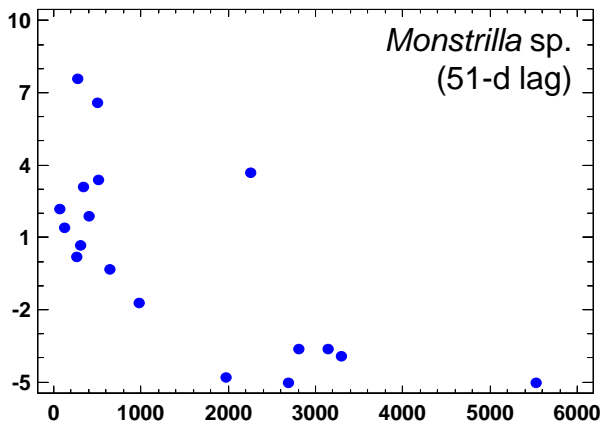
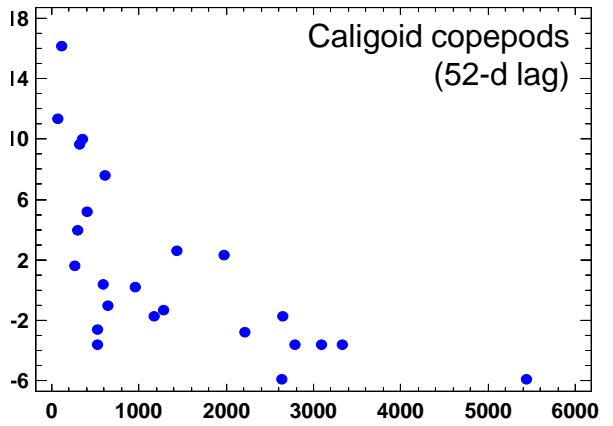
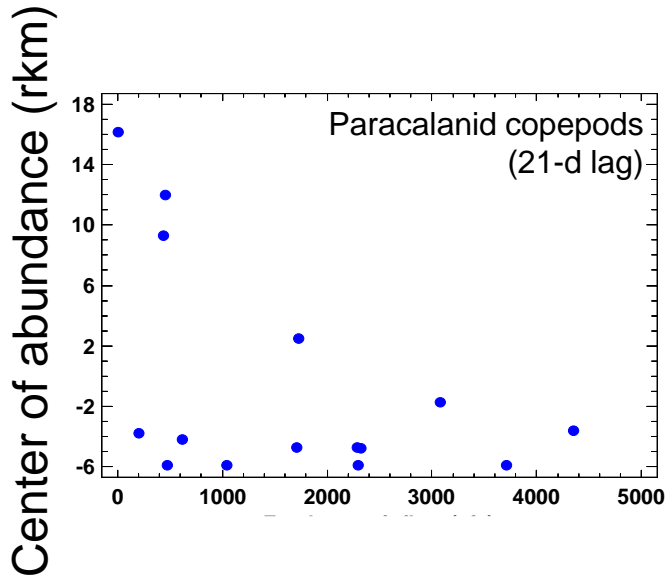
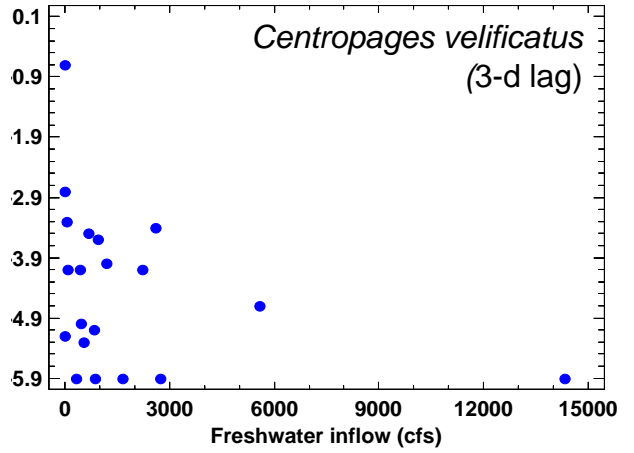
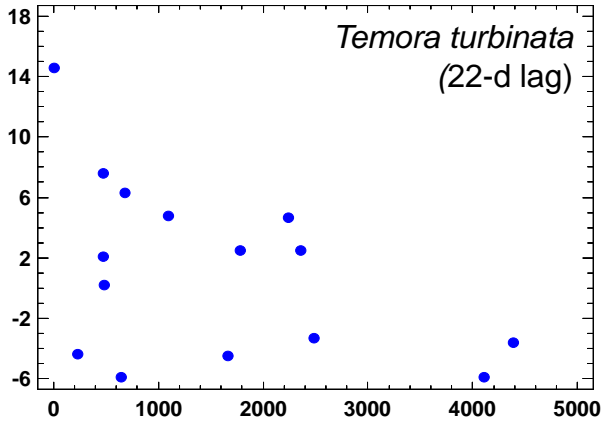
Center of abundance (rkm)



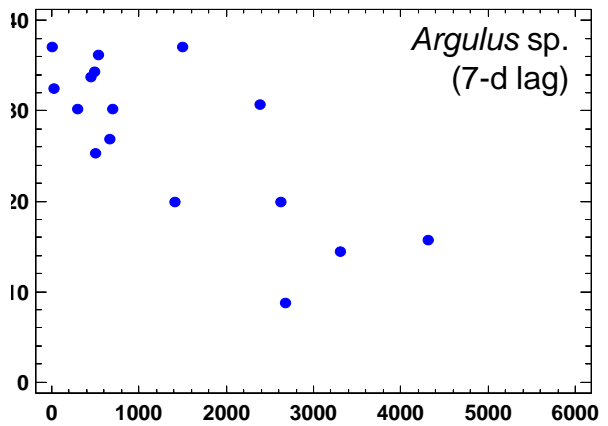
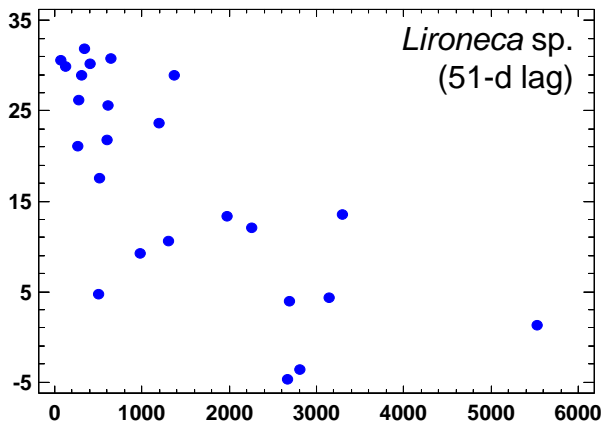
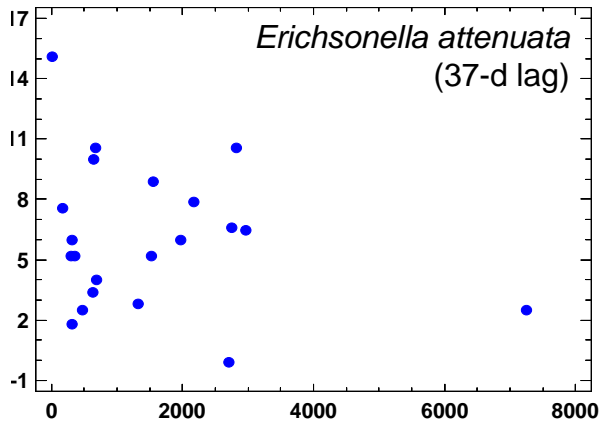
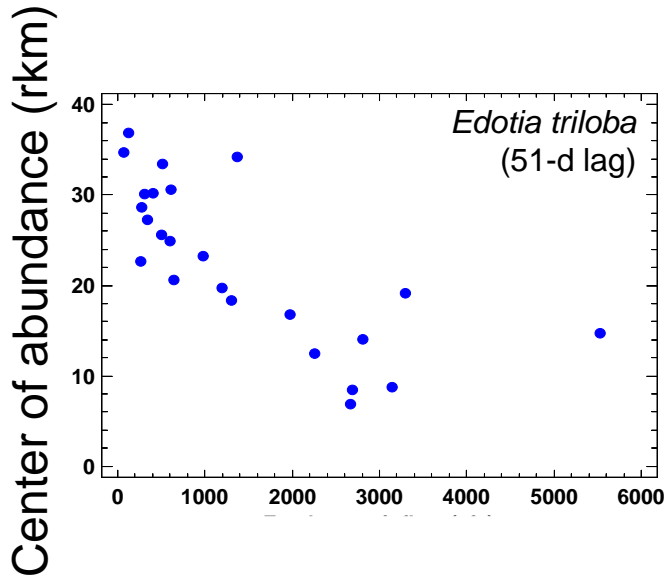
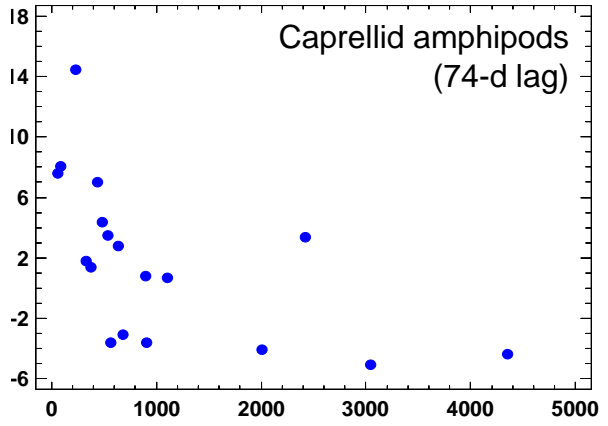
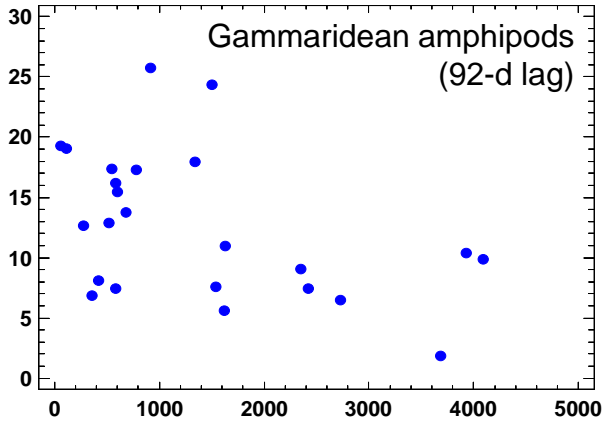


Center of abundance (rkm)

Freshwater inflow (cfs)

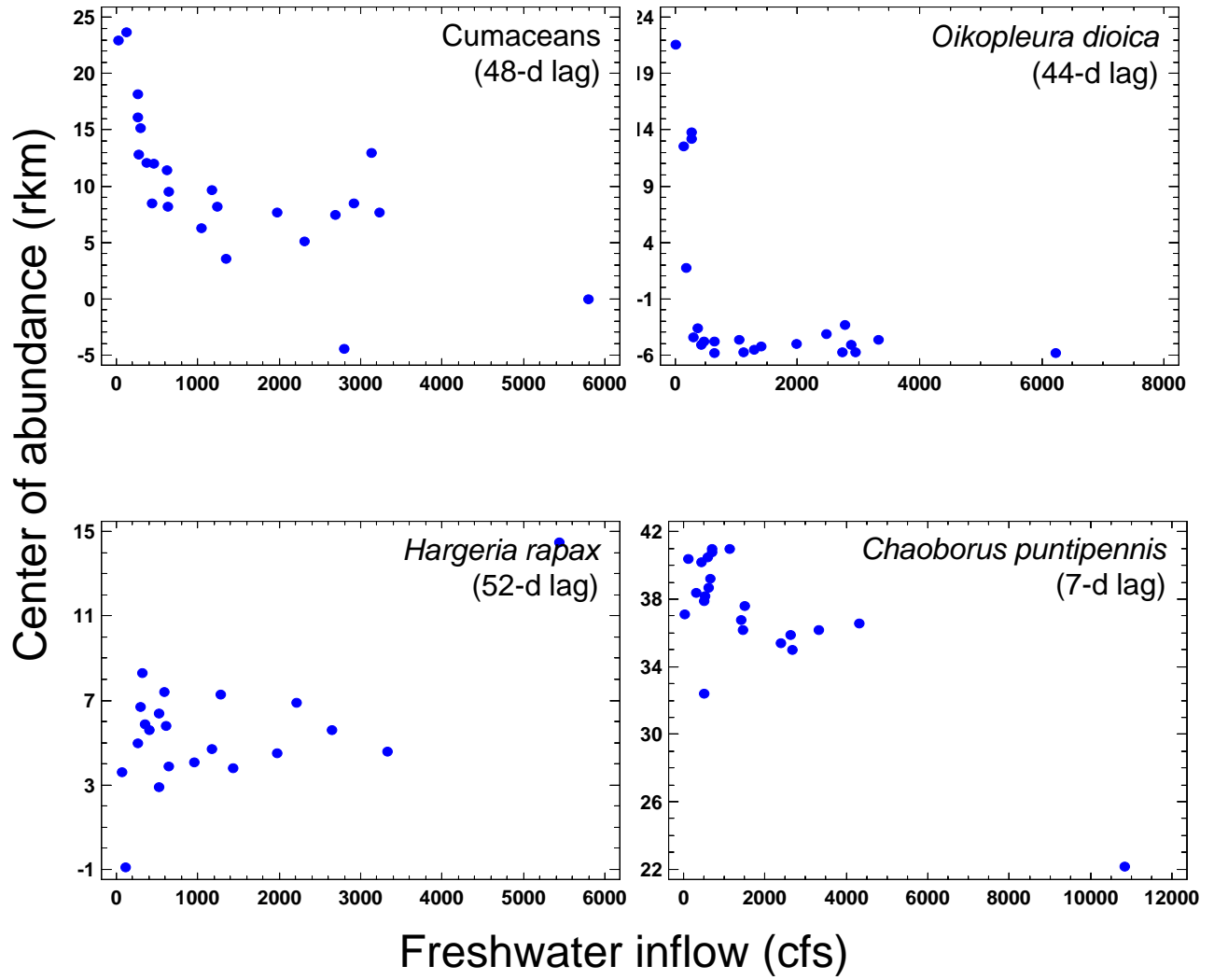


Freshwater inflow (cfs)



Freshwater inflow (cfs)

Center of abundance (rkm)

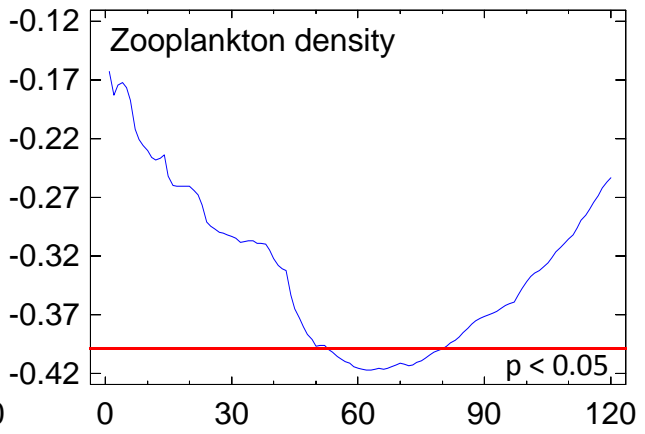
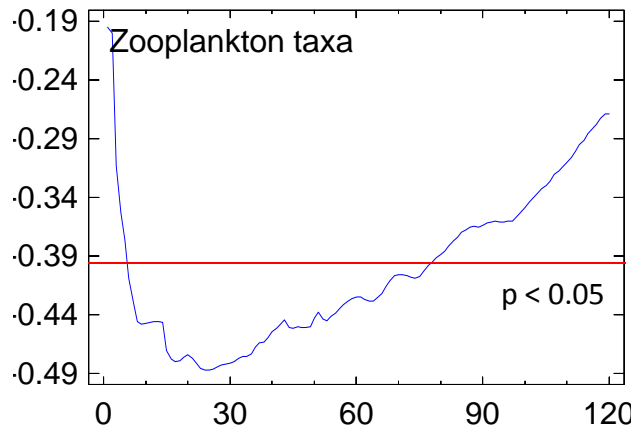


Appendix H

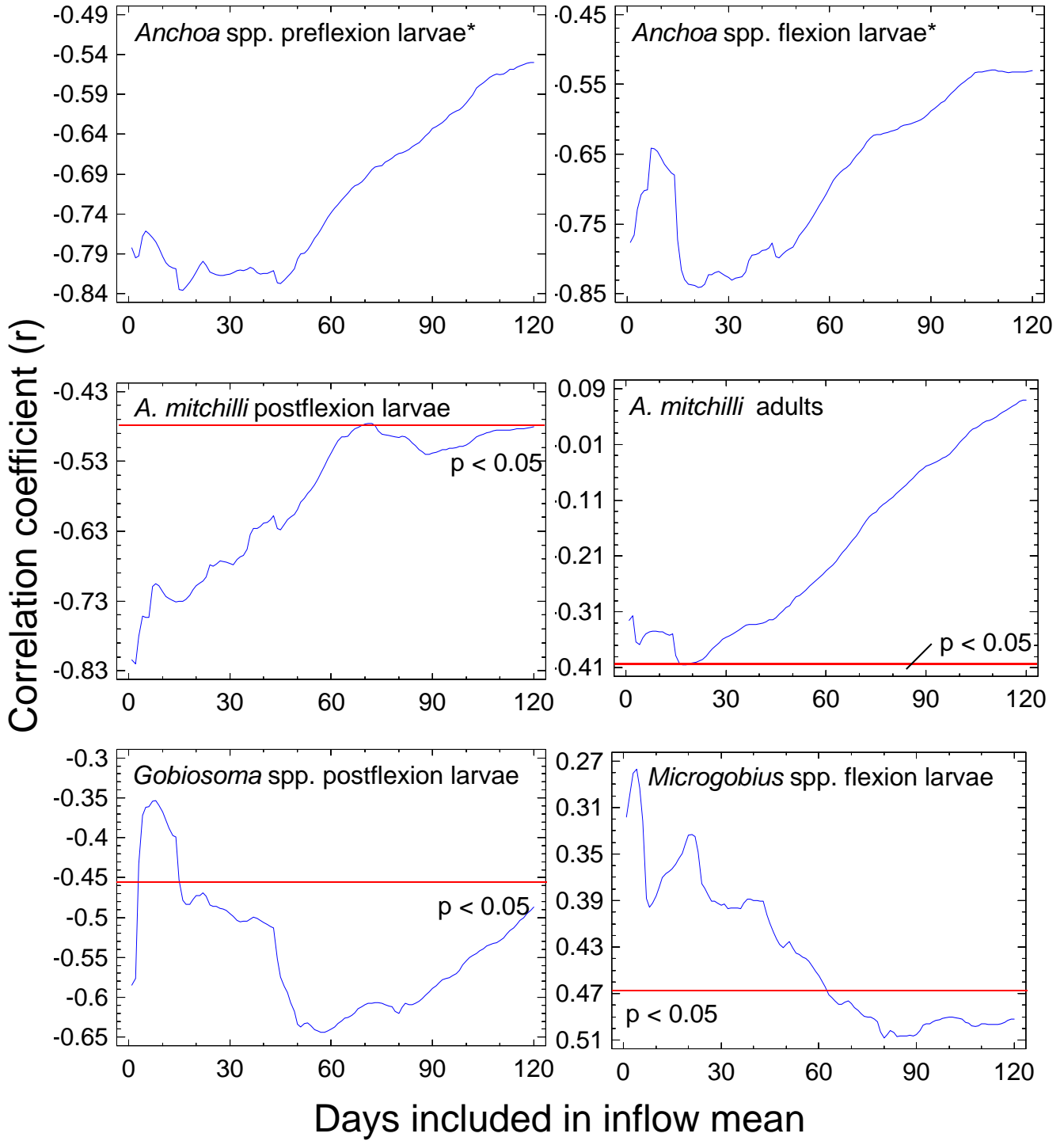
Correlation coefficients vs. lag time for relationships

between center of abundance and inflow

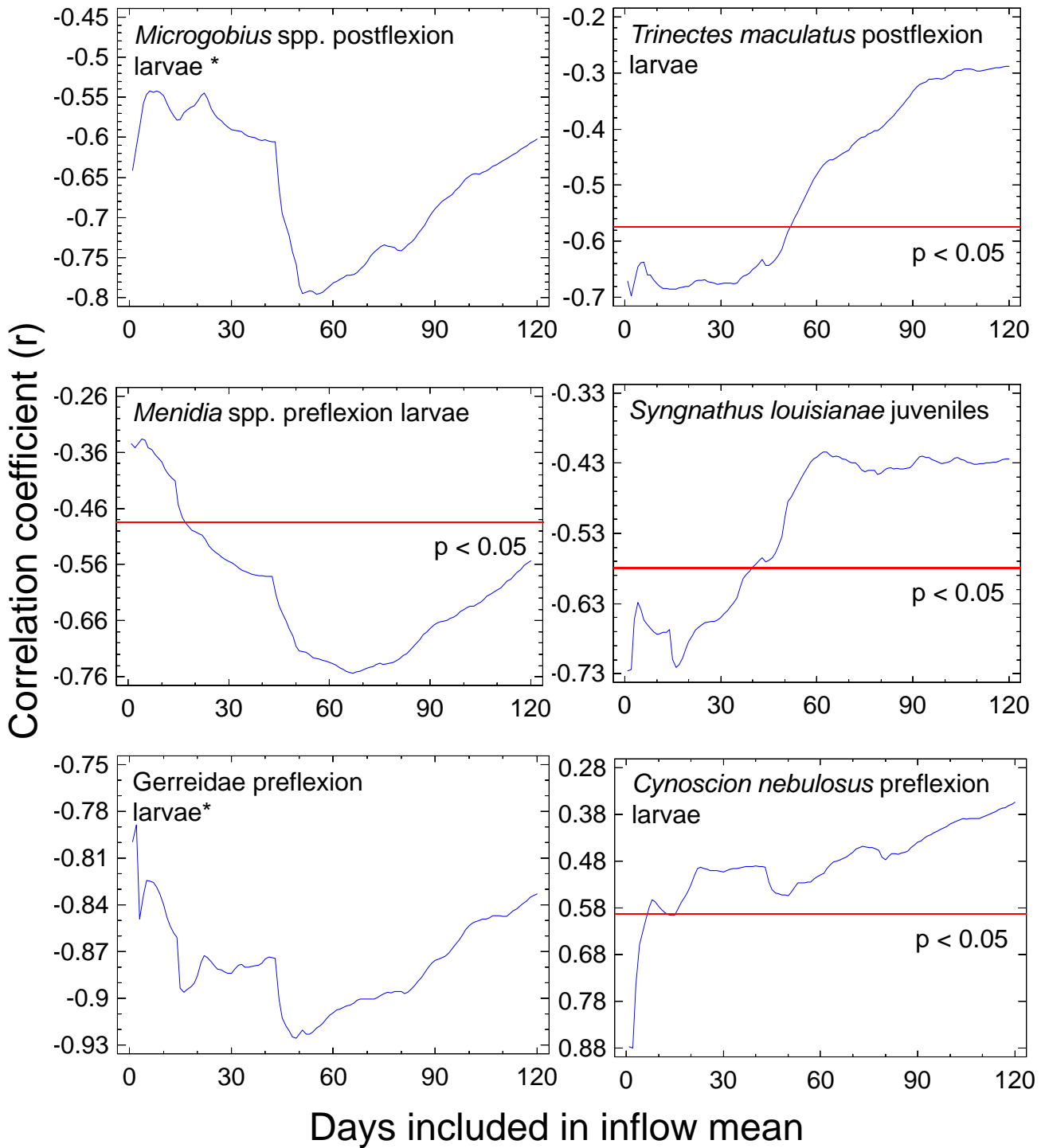
Correlation coefficient (r)



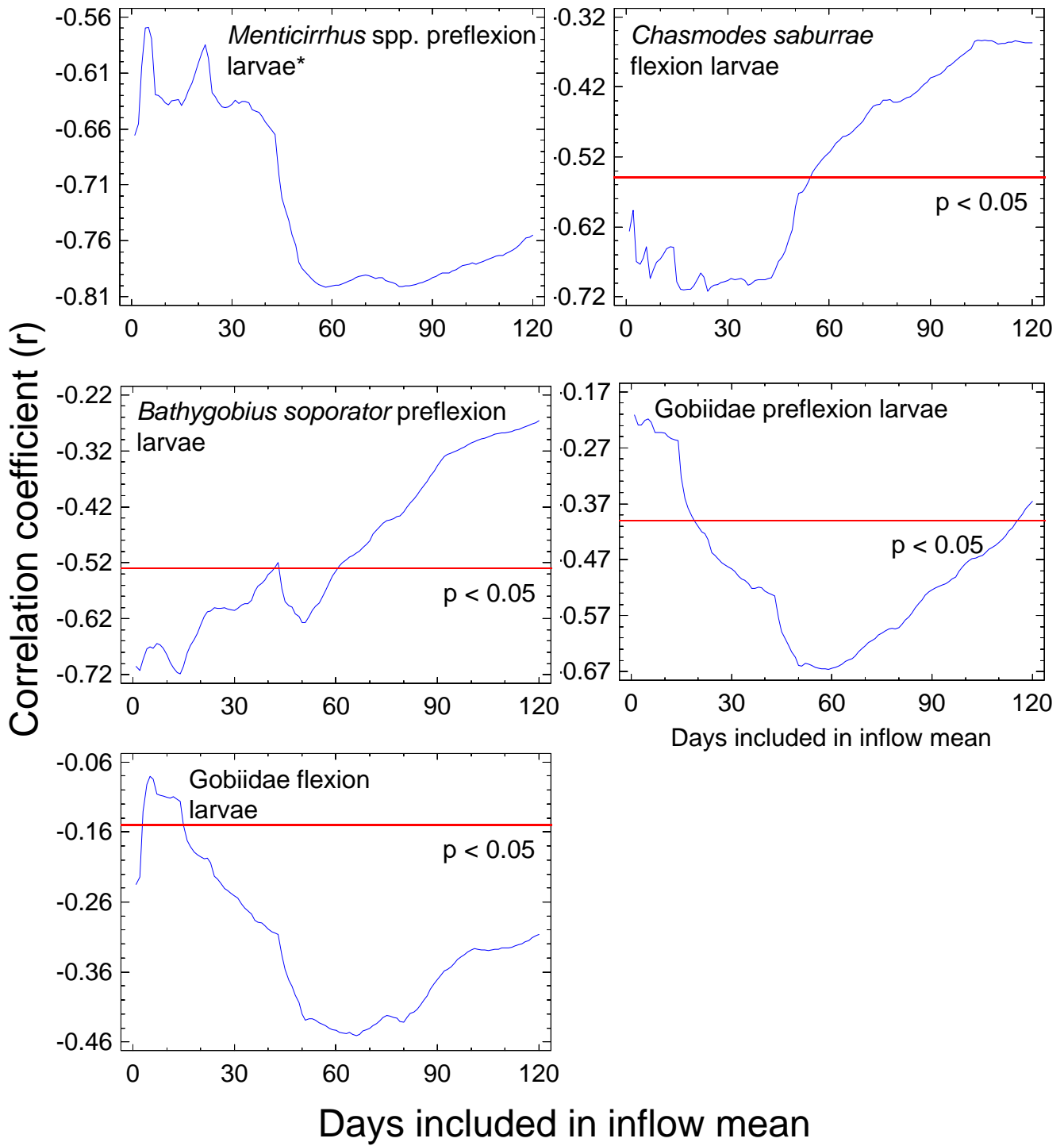
Days included in inflow mean



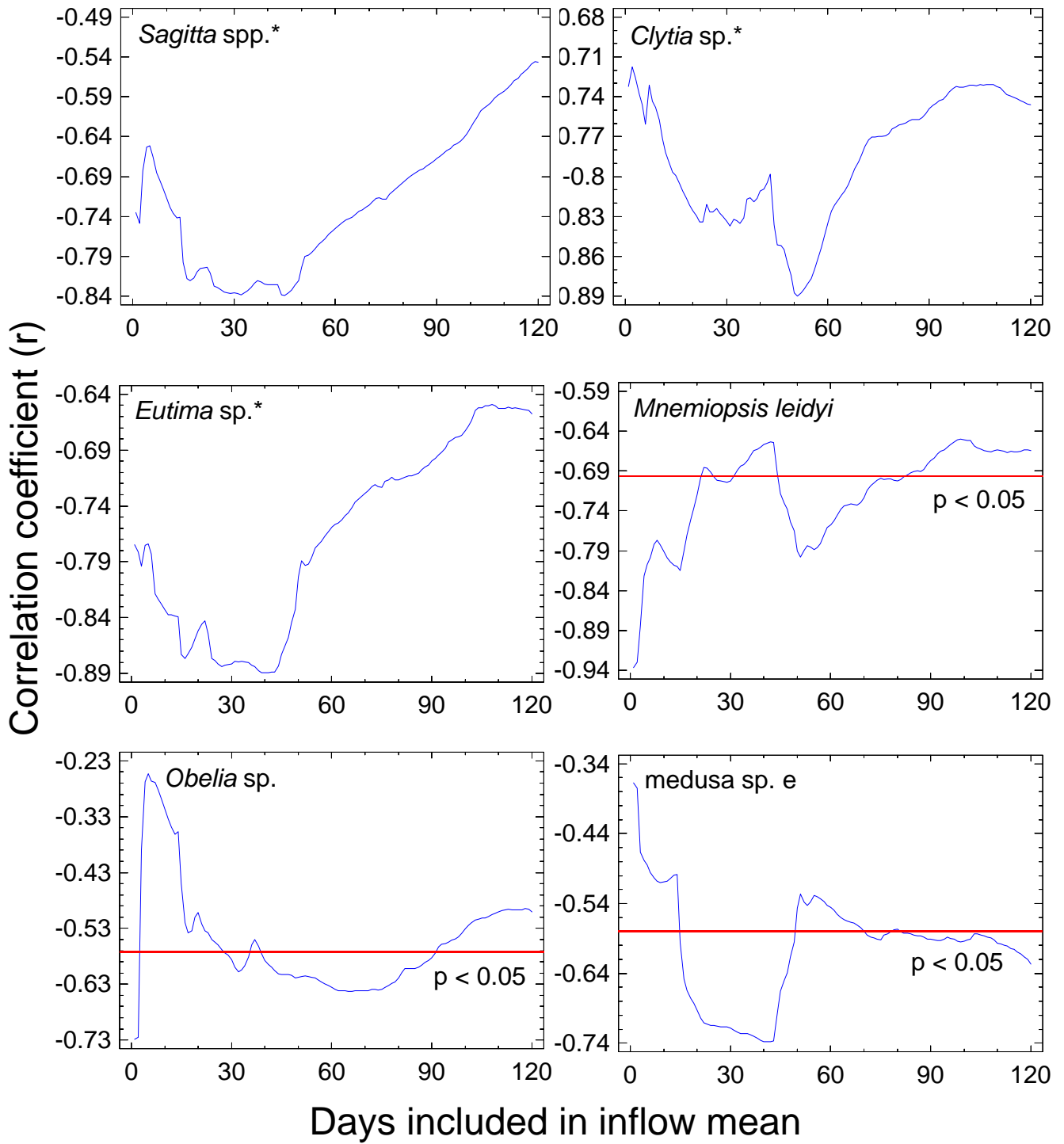
* All r values significant at $p < 0.05$



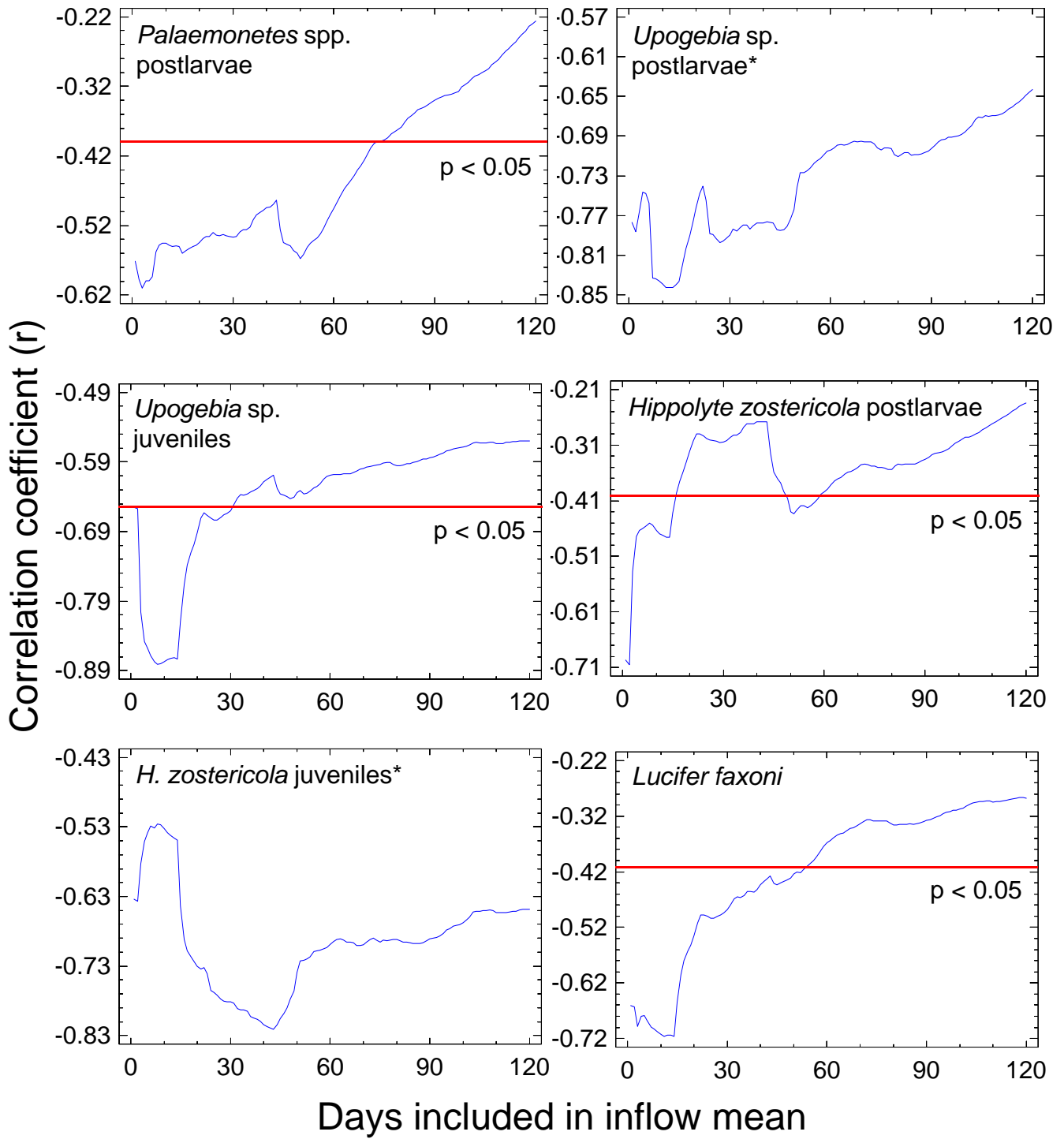
* All r values significant at $p < 0.05$



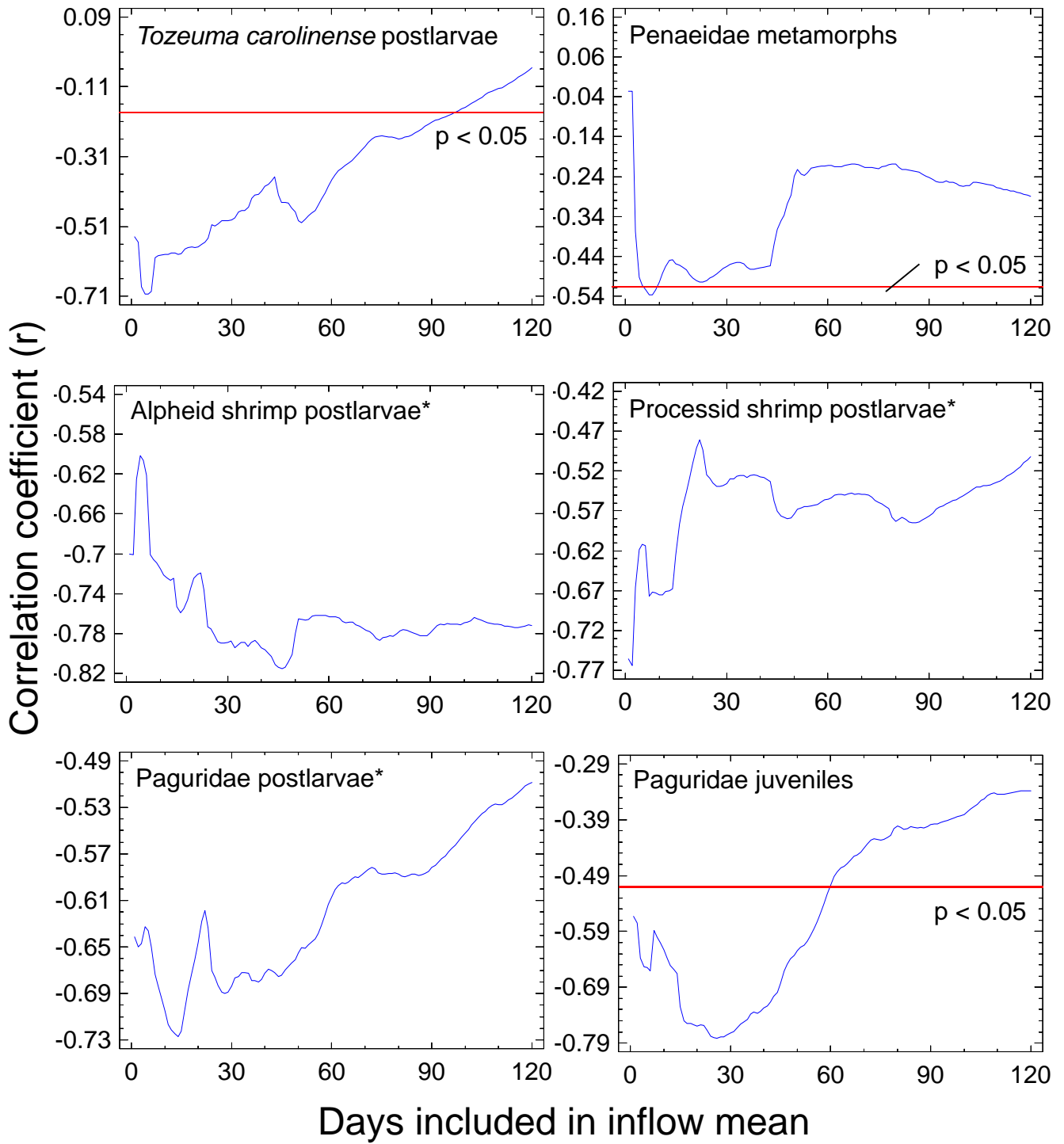
* All r values significant at $p < 0.05$



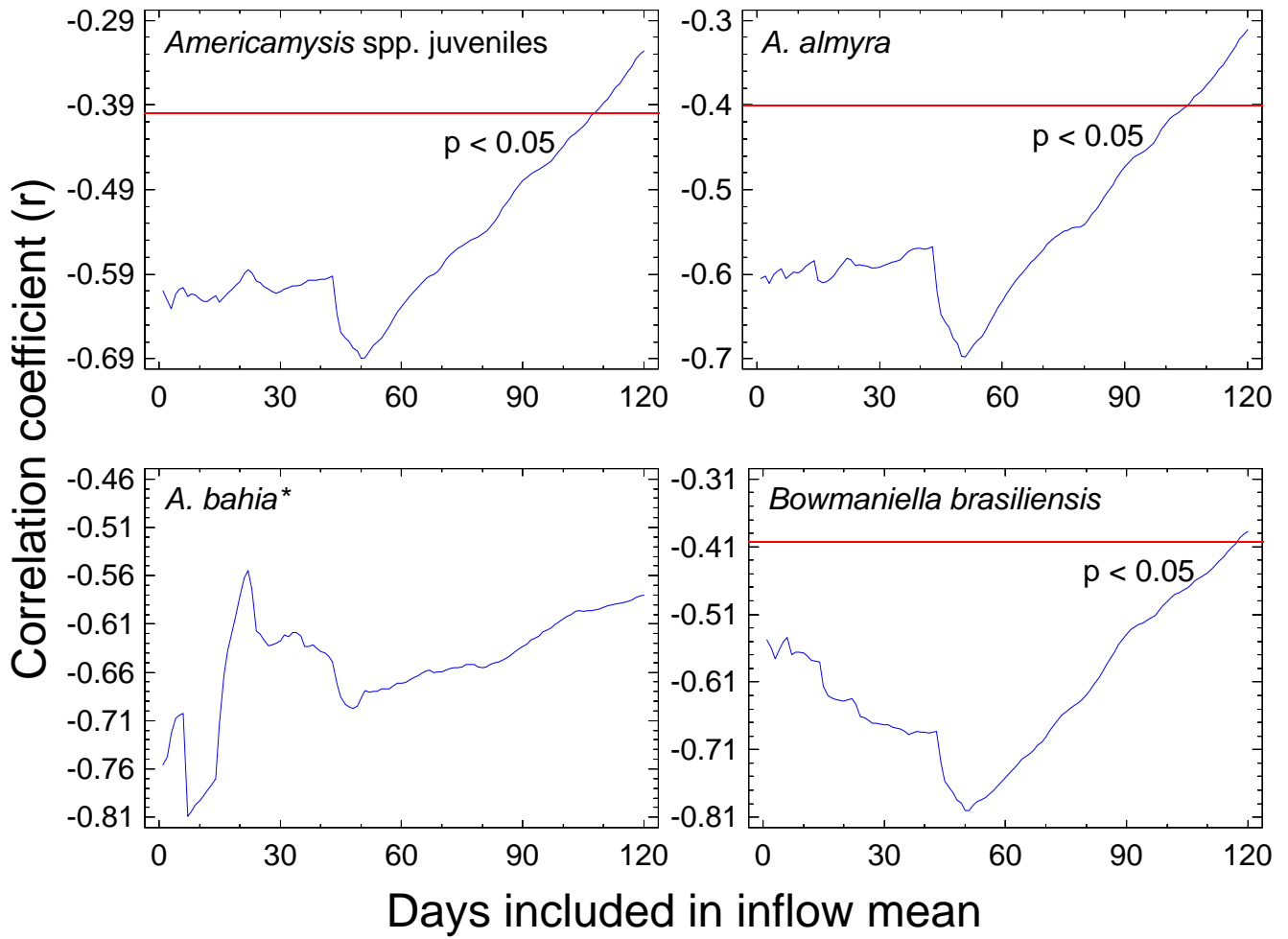
* All r values significant at $p < 0.05$



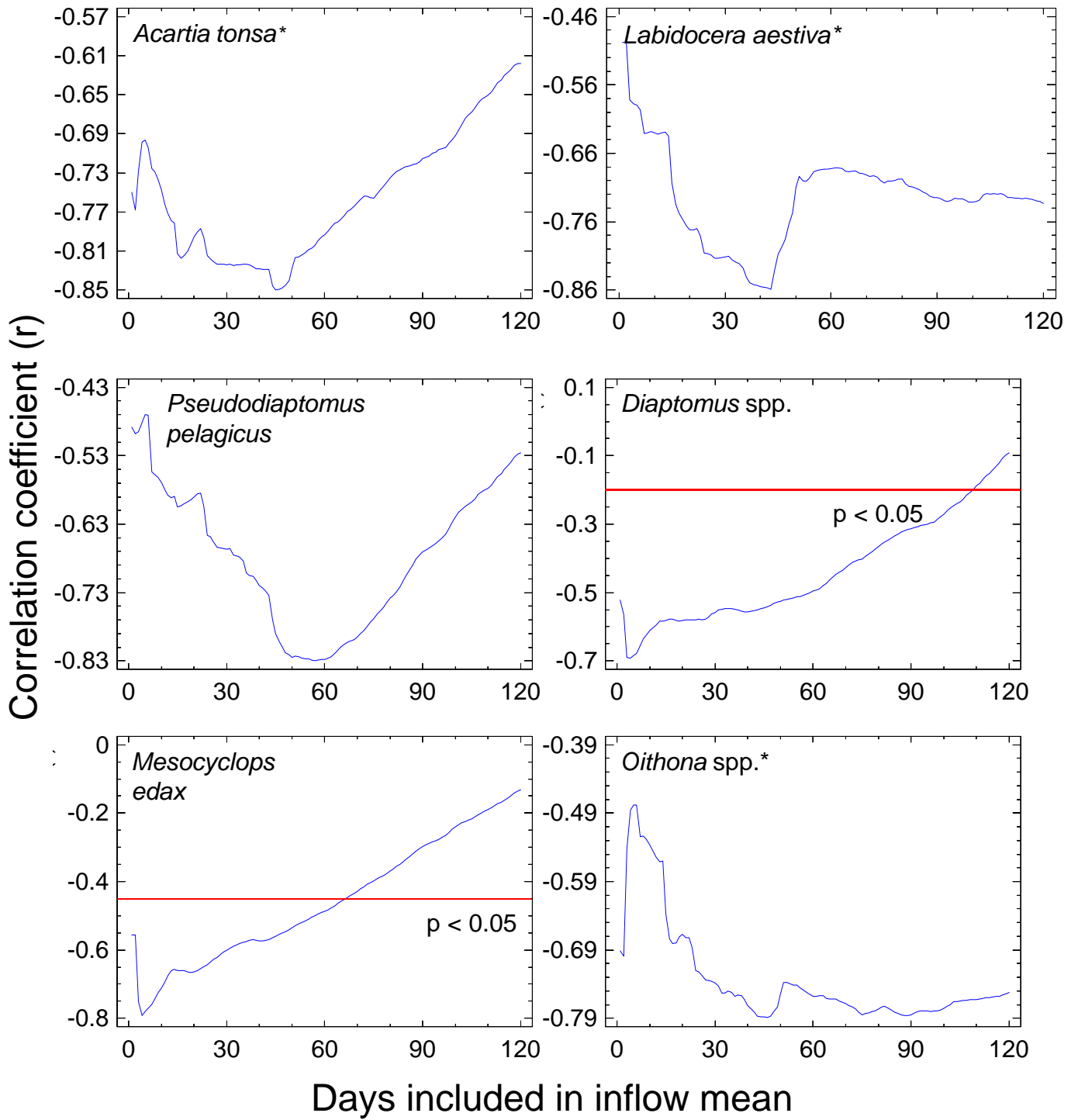
* All r values significant at $p < 0.05$



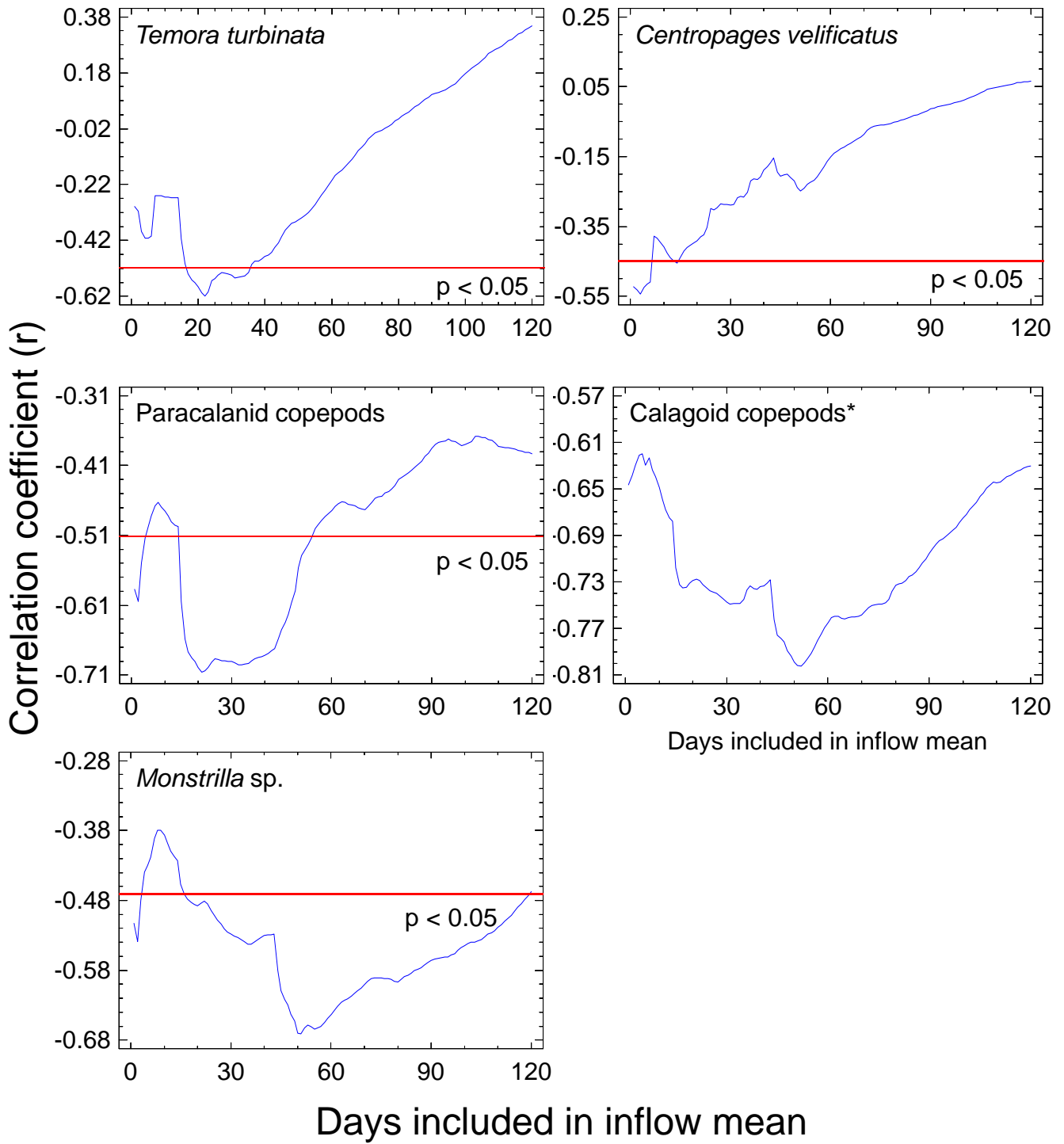
* All r values significant at $p < 0.05$



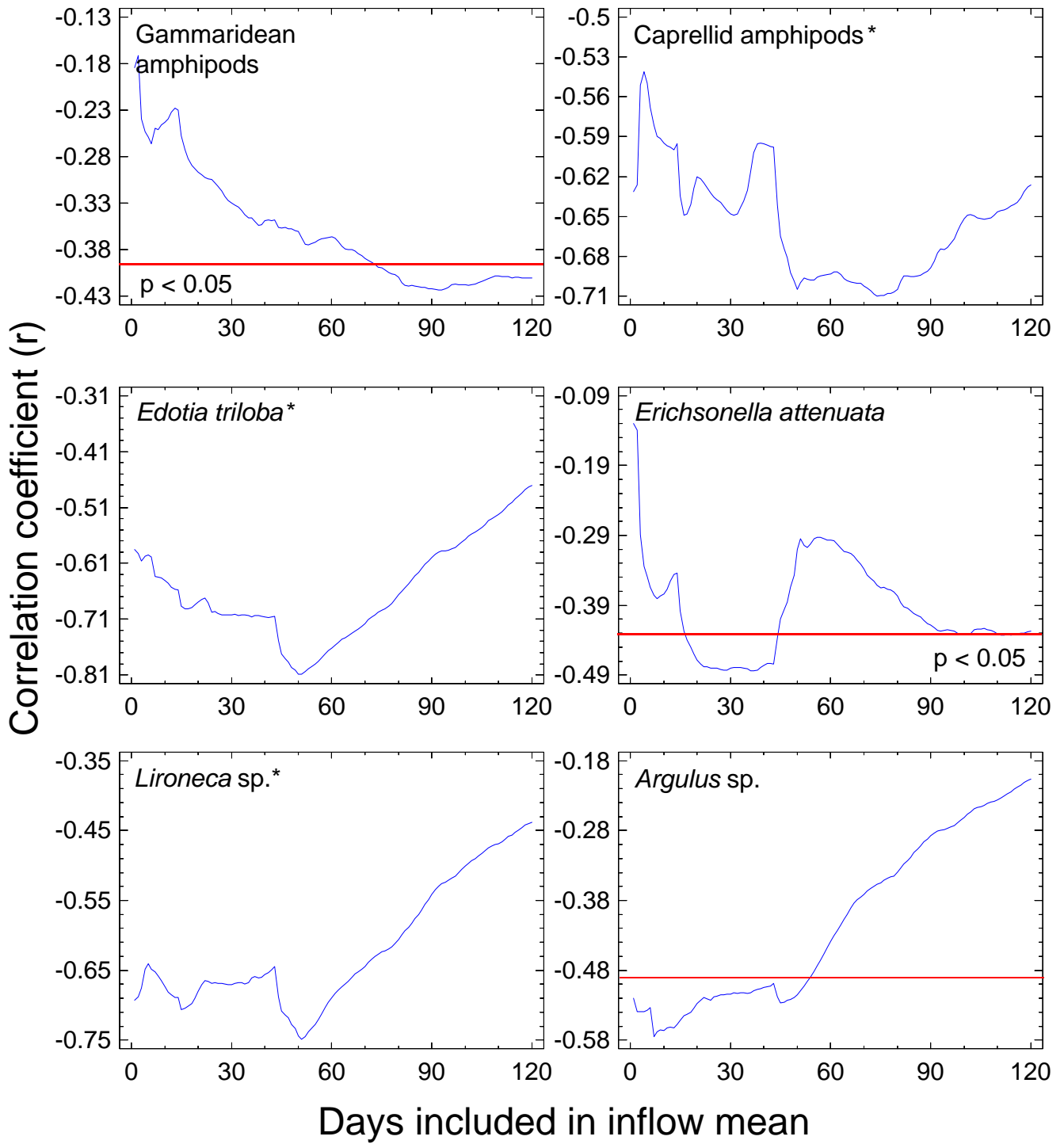
* All r values significant at $p < 0.05$



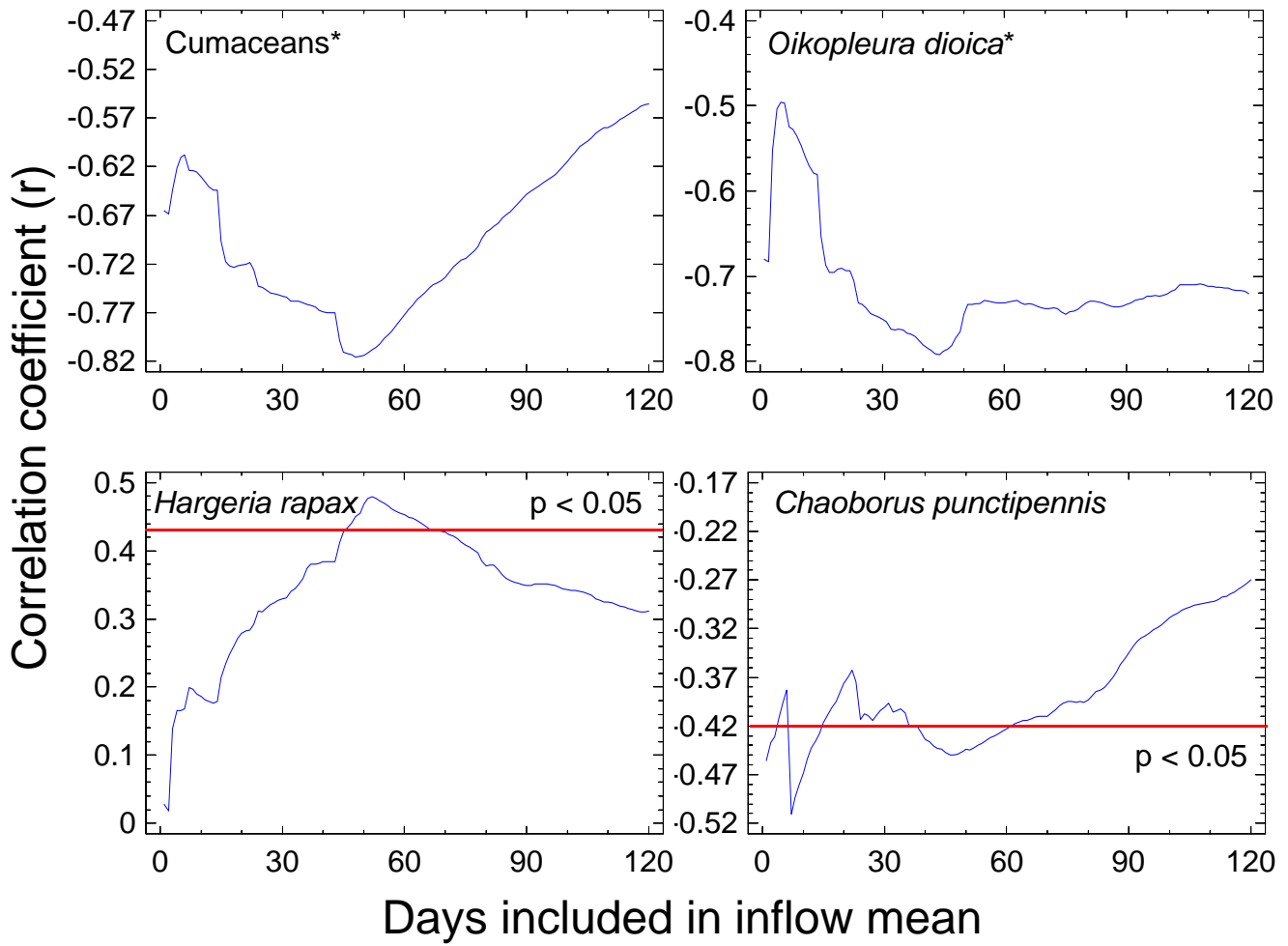
* All r values significant at $p < 0.05$



* All r values significant at $p < 0.05$



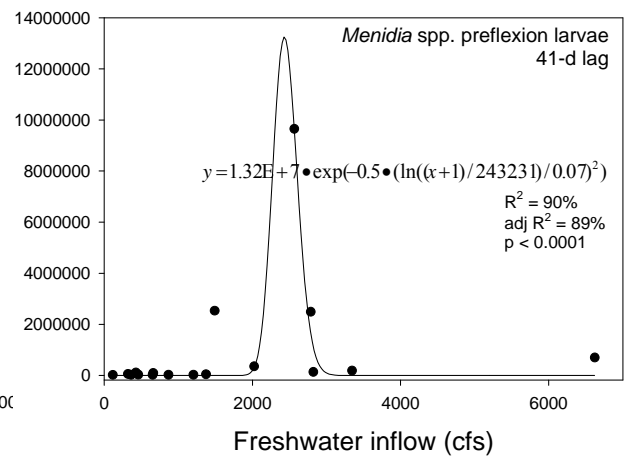
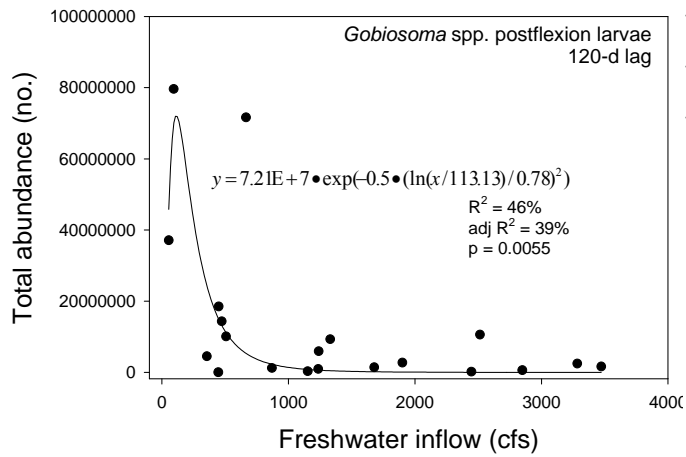
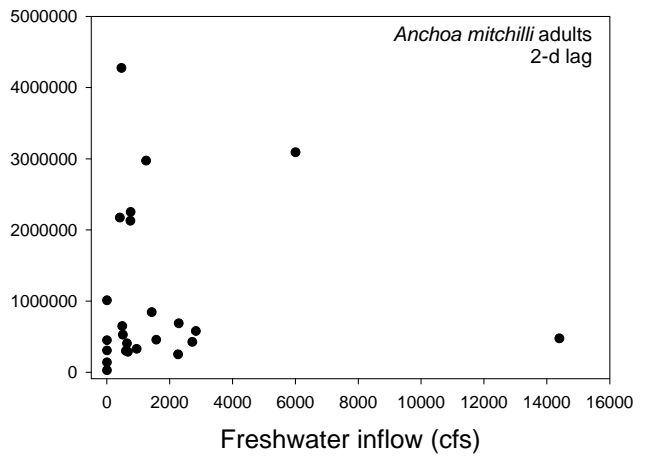
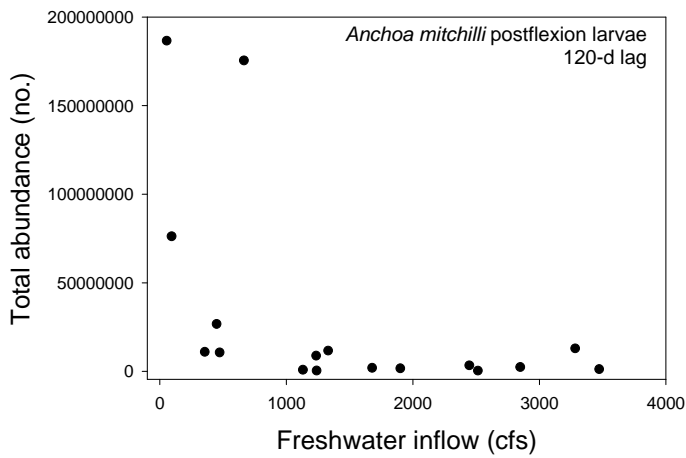
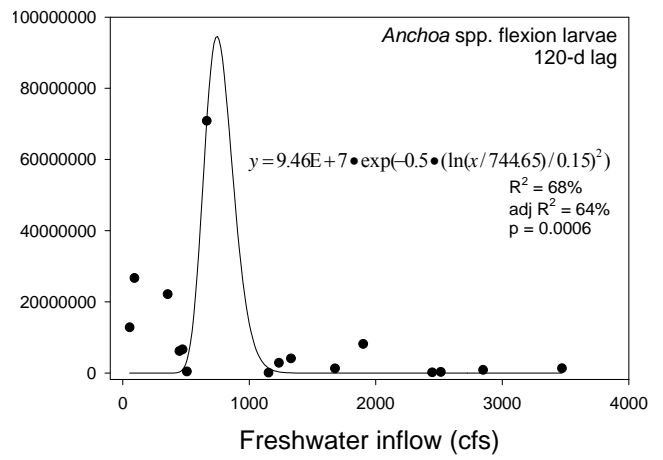
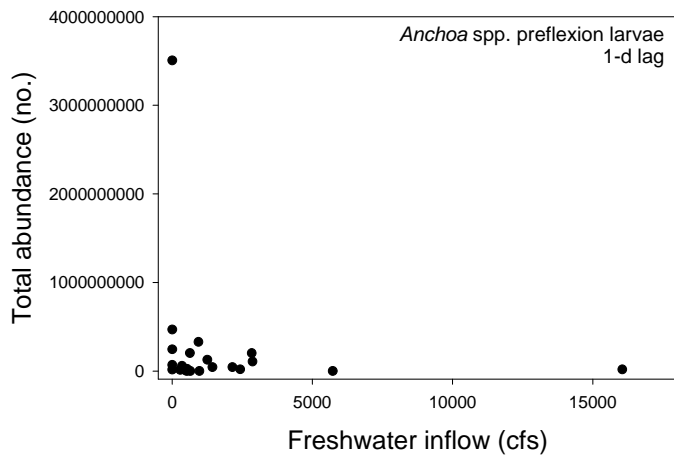
* All r values significant at $p < 0.05$

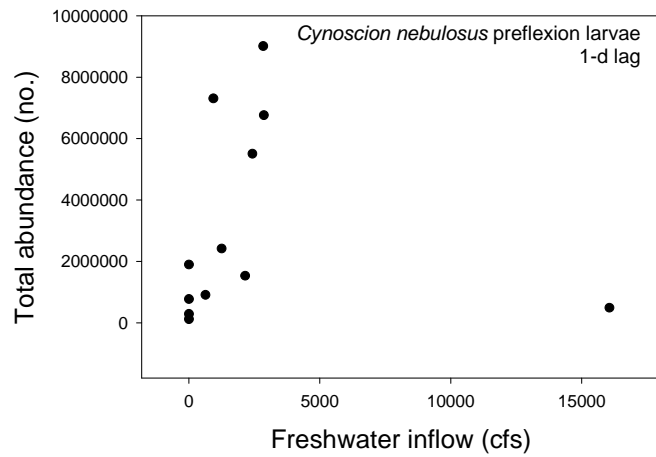
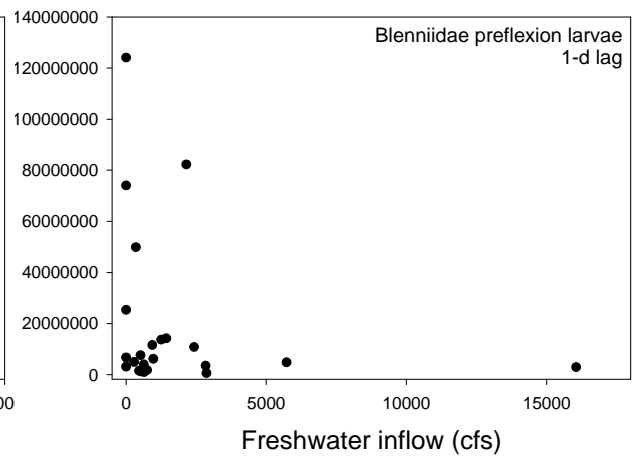
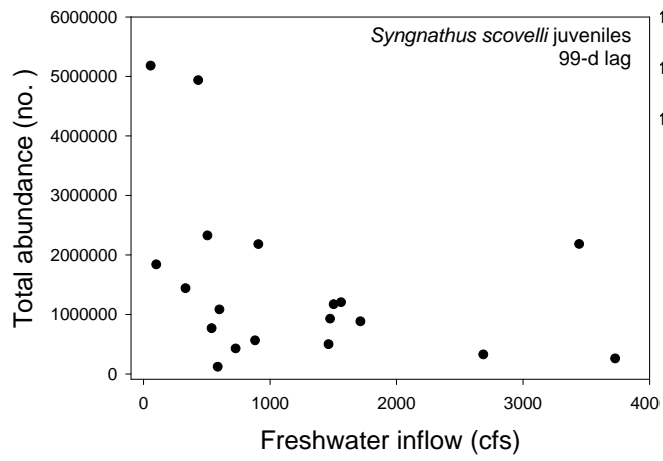
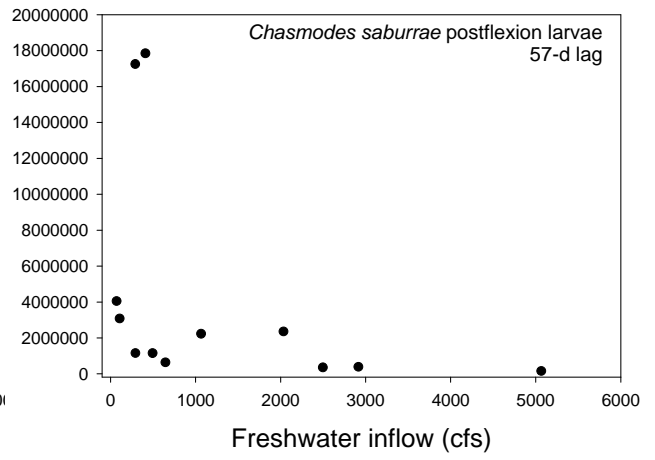
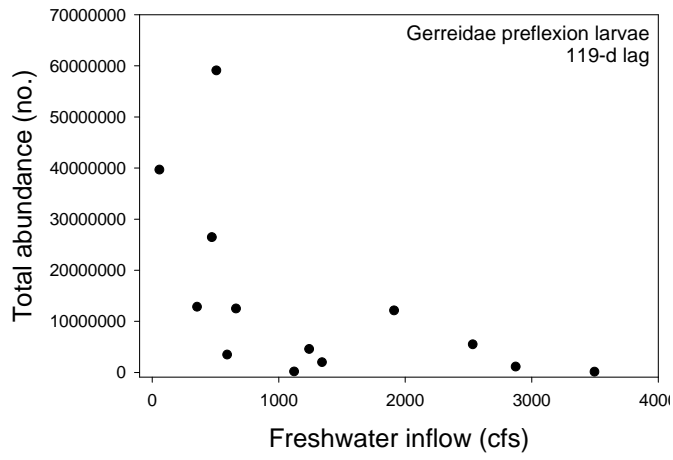


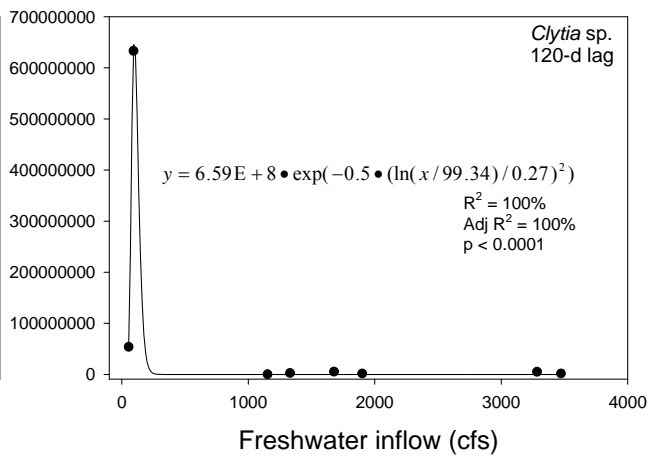
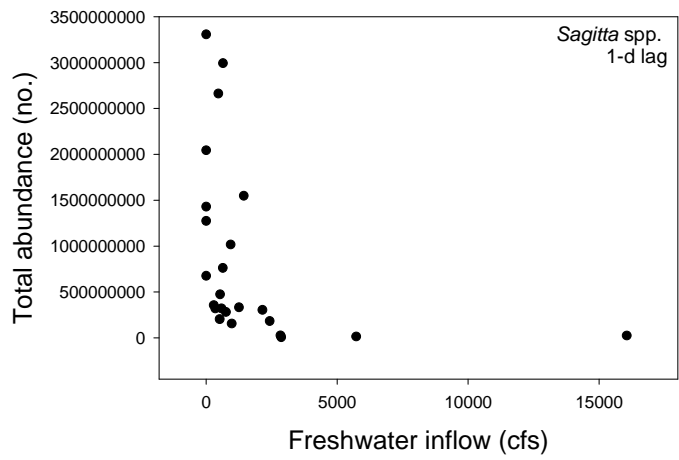
* All r values significant at $p < 0.05$

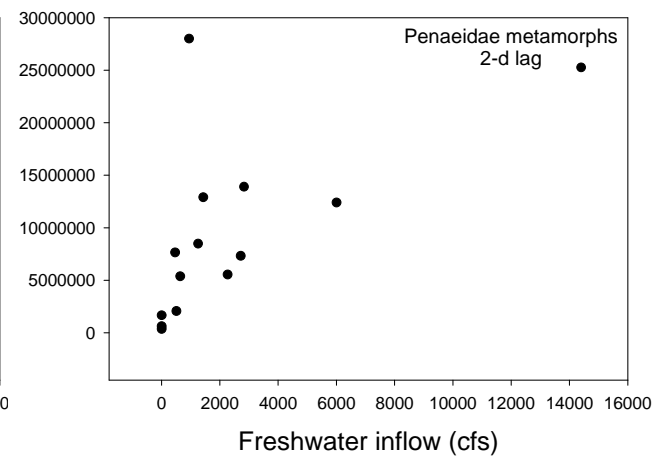
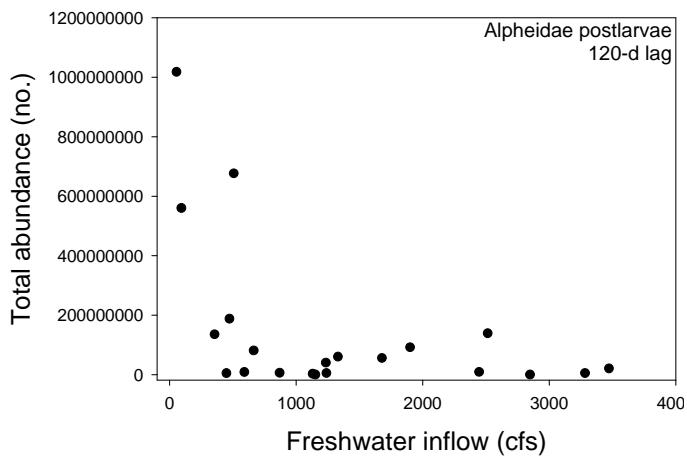
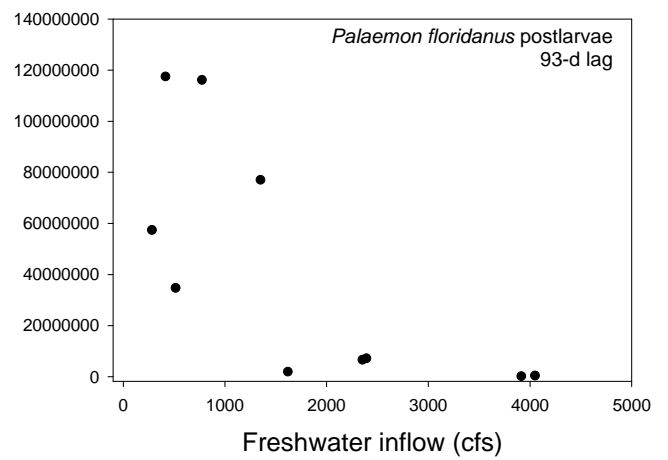
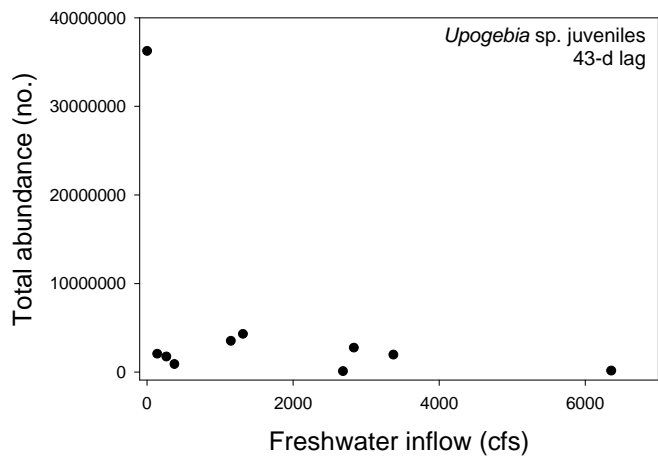
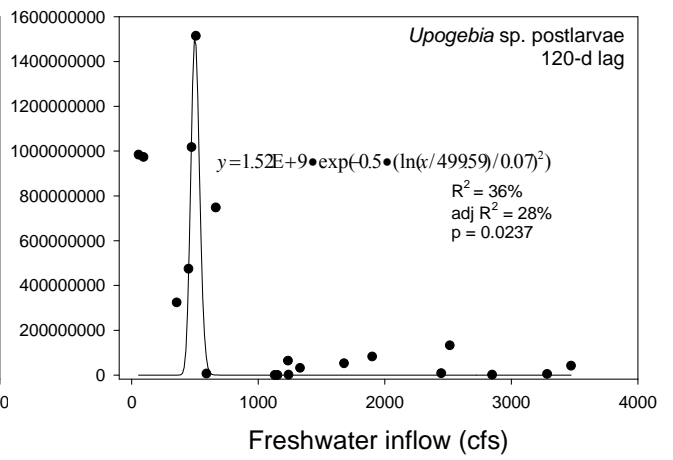
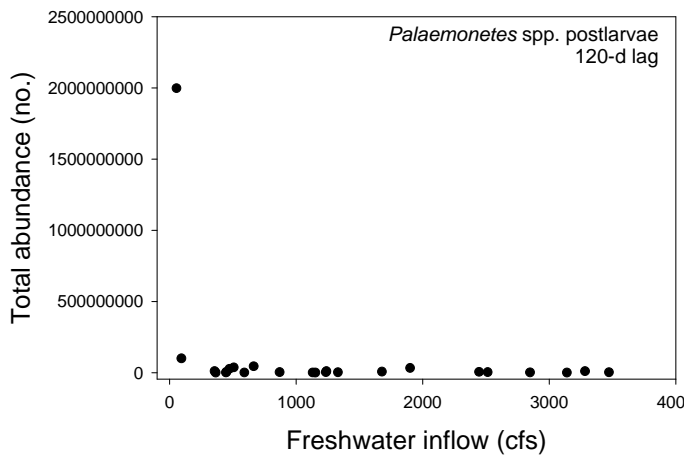
Appendix I

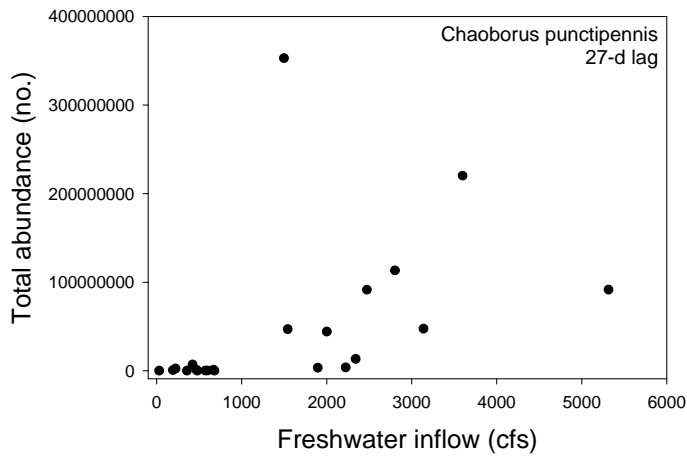
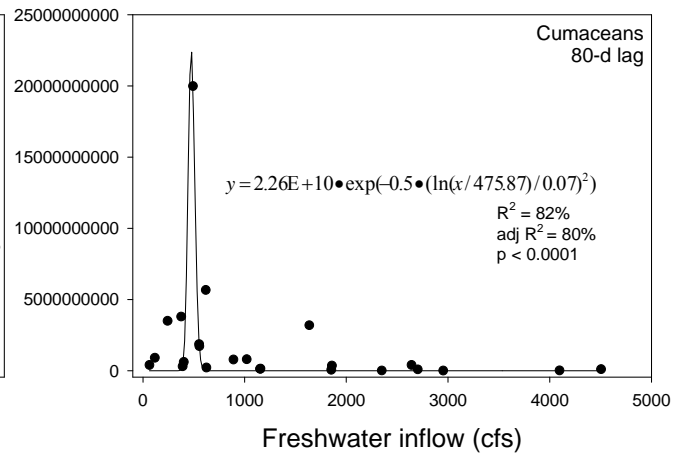
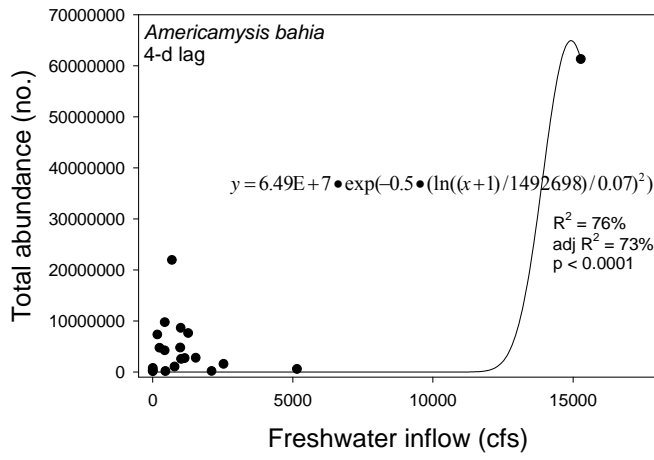
Total organism abundance responses to inflow

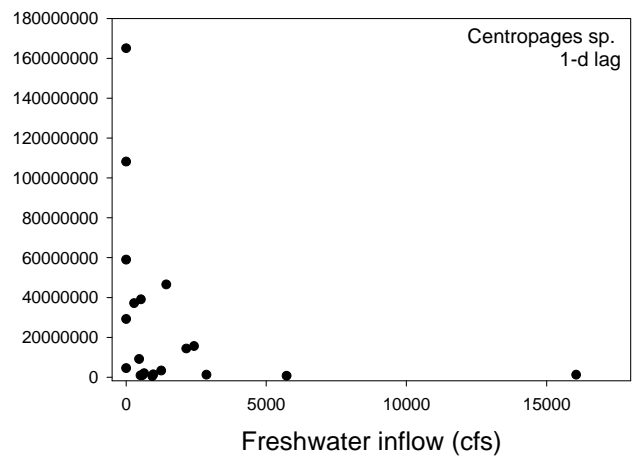
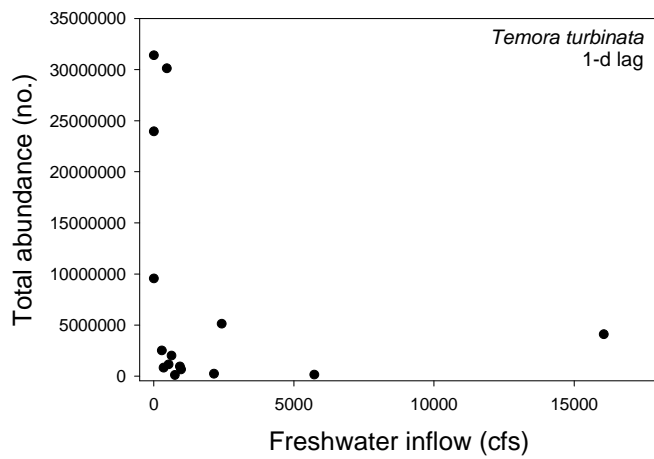
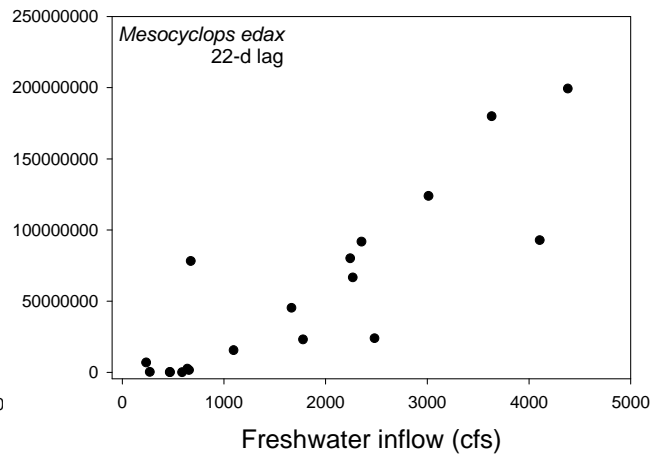
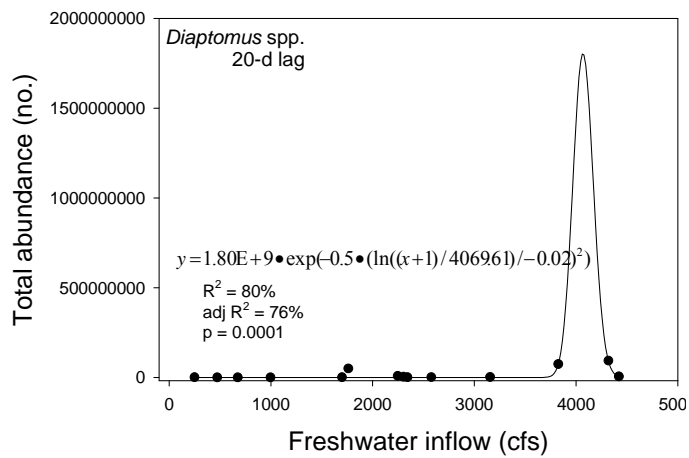
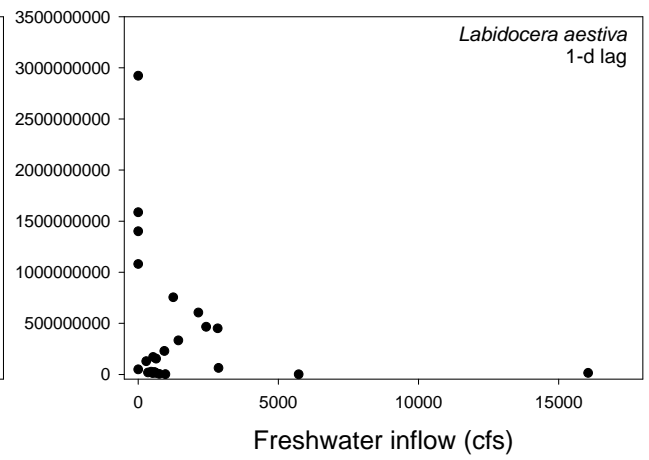
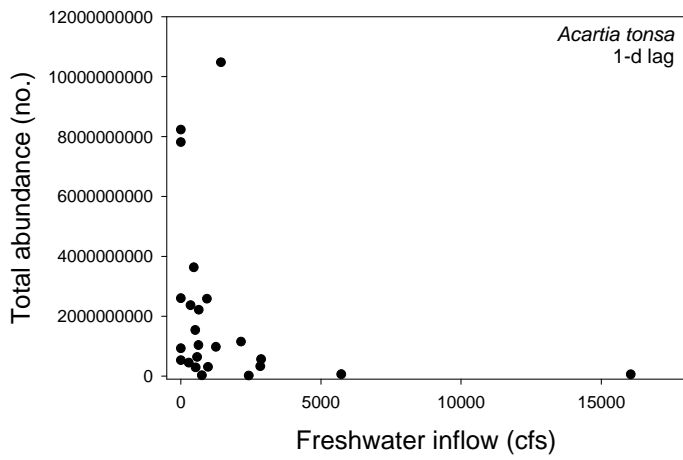


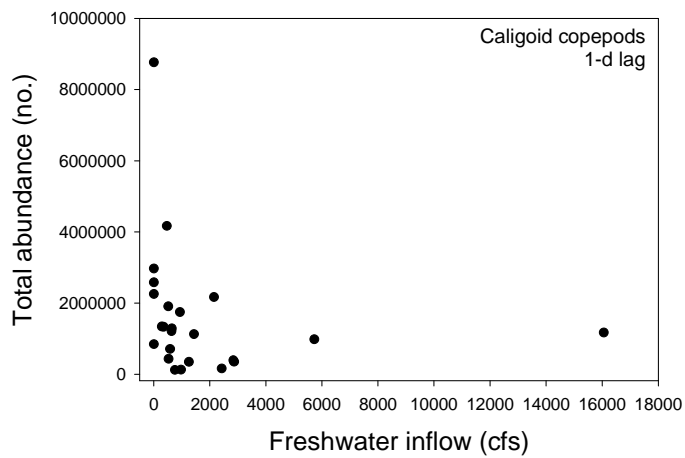
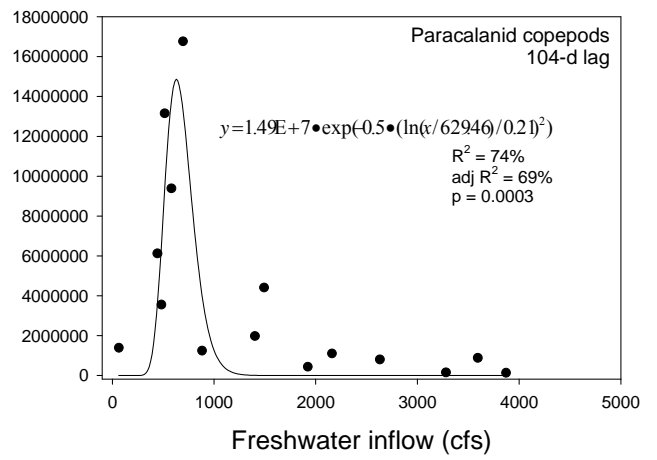
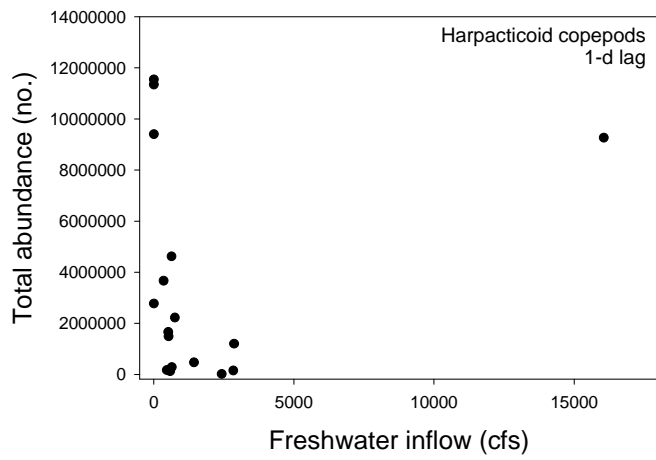


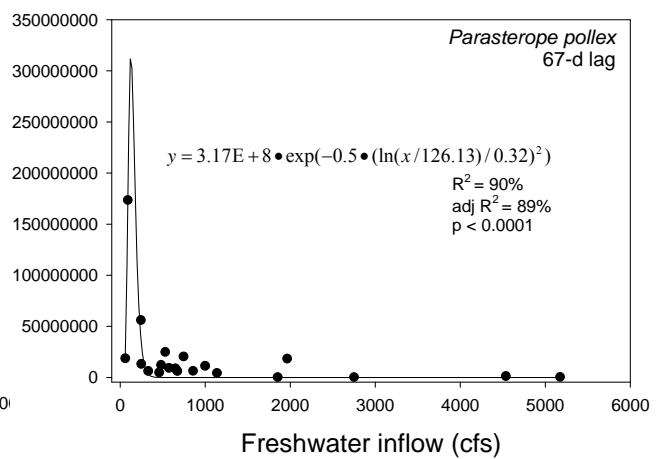
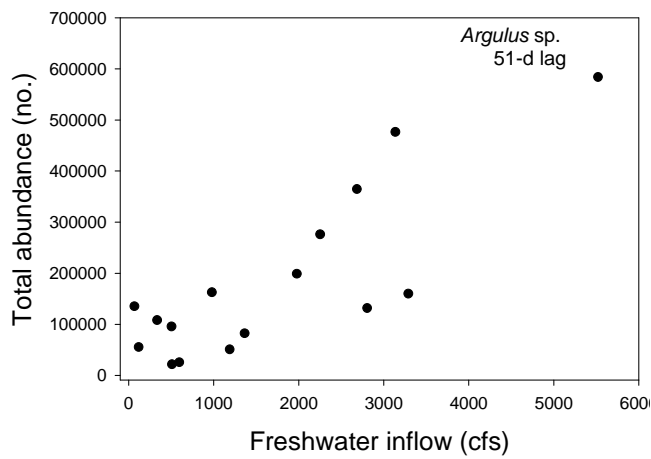
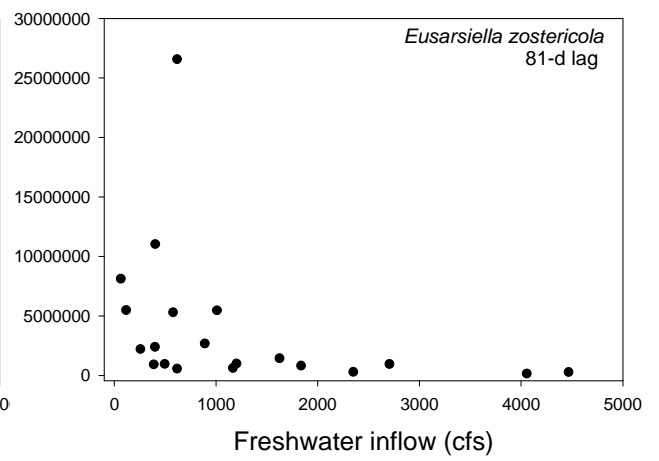
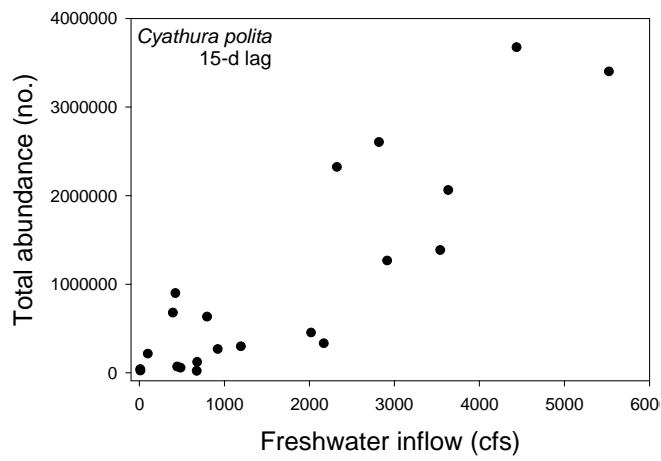
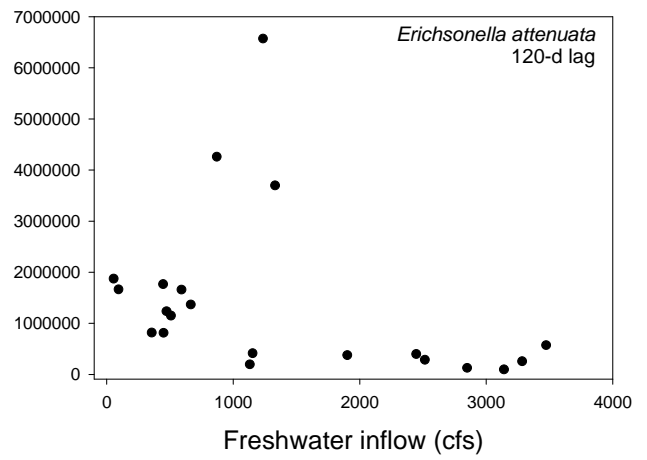
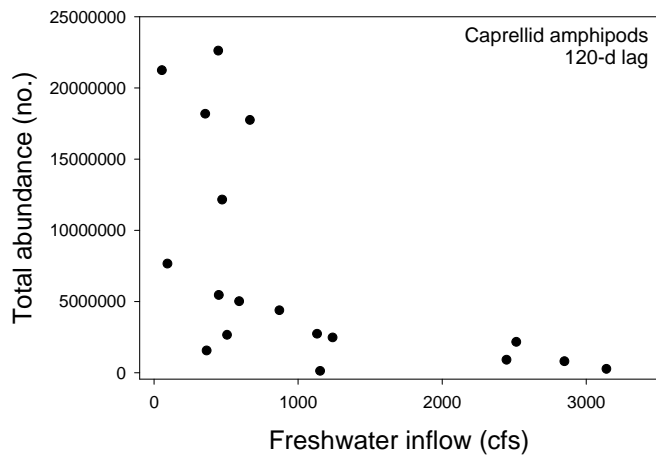


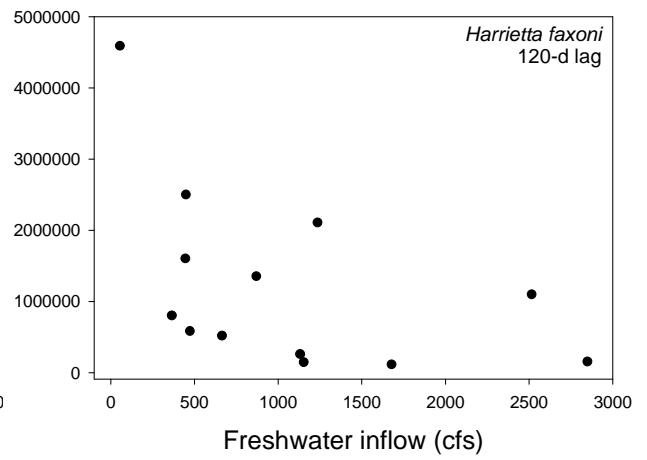
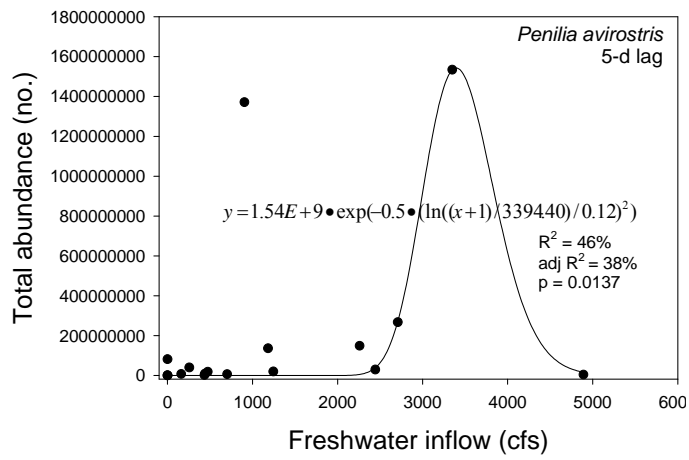












Appendix J

Correlation coefficients vs. lag time for relationships
between total abundance and inflow

