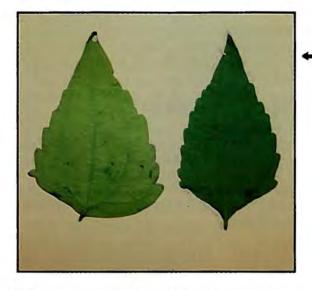
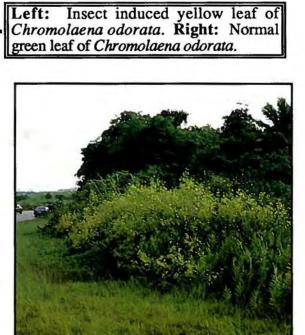
CHROMOLAENA ODORATA Newsletter

Number 2

October 1988



Insect induced yellowing in the bushes of Chromolaena odorata.



Chromolaena odorata defoliated by Pareuchaetes pseudoinsulata on Rota (the same area as in the bottom figure).



Chromolaena odorata infested pasture area in Rota.



A NOTE FROM THE EDITOR; R. Muniappan, College of Agriculture and Life Sciences, University of Guam, Mangilao, Guam 96923 U.S.A.

PROCEEDINGS IN DEMAND!

Proceedings of the First International Workshop on Biological Control of Chromolaena odorata was prepared and mailed in July-August, 1988 to scientists involved or indicated interest in this subject area. Few copies are still available and will be distributed upon request.to R. Muniappan, College of Agriculture and Life Sciences, University of Guam, Mangilao, Guam 96923, U.S.A.

SHIPMENTS OF PAREUCHAETES PSEUDOINSULATA

P. pseudoinsulata has been shipped to Yap and Ponape in the Caroline Islands, Thailand and South Africa from Guam since the last newsletter.

ACKNOWLEDGEMENTS

The secretariat wishes to recognize Mr. Patrick E.Q. Perez for typesetting and formatting and thank the Cooperative Extension Service, College of Agriculture and Life Sciences, University of Guam for funding the publication of this newsletter.

C. ODORATA IN HAINAN, CHINA; F.D. Bennett, Department of Entomology and Nematology, University of Florida, Gainesville, Florida 32611-0143 U.S.A.

While on Hainan Island, People Republic of China, 1-3 June 1988 to survey for natural enemies of certain citrus pests in the company of Dr. Ren Hui, Guangdong Entomological Institute, Guangzhou, I frequently observed solid stands of the following three Neotropical weeds: Chromolaena odorata, Parthenium hysterophorus and Lantana camara. Light damage to C. odorata by chrysomalid adult

feeding, larvae of a polyphagous Lepidoptera and a green aphid was noted, but these appeared to have little impact on plant growth. Although a few plants have sustained heavy attack of aphids which caused leaf curling and distortion of the terminal growth, aphids were attacked by coccinellids, (2 spp.), syrphids, chamaemyiids (Leucopis sp.), chrysopids and hemerobiids.

SOME ADDITIONAL REFERENCES ON C. ODORATA

- Dove, M.R. 1986. The practical reason of weeds in Indonesia: Peasant vs. State views of *Imperata* and *Chromolaena*. Human Ecology. 14(2): 163-190.
- Ooi, P.A.C., Sim, C.H. and Tay, E.B. 1988. Status of the arctiid moth introduced to control Siam weed in Sabah, Malaysia. Planter, Kuala Lumpur. 64: 298-304.
- Seibert, T.F. 1988. Biological control of the weed, *Chromolaena odorata* (Asteraceae), by *Pareuchaetes pseudoinsulata* (Lepidoptera: Arctiidae) on Guam and the Northern Mariana Islands. Entomophaga (in press).

APPLICATION OF SOILS INFORMATION IN THE STUDY OF THE DISTRIBUTION OF CHROMOLAENA ODORATA

Hari Eswaran, SMSS, USDA, P.O. Box 2890 Washington, D.C. 20013 U.S.A.

Siam weed (Chromolaena odorata) is wide spread in Southeast Asia and spreading to other parts of the tropics. Its resilience is so great that it has emerged as one of the most obnoxious weeds of the tropics. It has the capability of even smothering out alang alang (Imperata cylinderica). The recent workshop on 'Biological Control of Chromolaena odorata', has addressed the issue of its spread, not only historical but also the potential.

This note is based on observations of the author and is not substantiated by any kind of study. Neither is the author an expert on this weed. The purpose of the note is to stimulate soil scientists to be involved in this potential problem of global significance as the distribution of the weed is controlled by soil and climate.

Preliminary observations suggest that the weed prefers acid soils, though it has been reported on base rich soils. It proliferates in areas with a mean annual soil temperature of more than 22°C and it does not seem to tolerate prolonged mositure stress. Even if these environmental conditions prevail, it appears to be concentrated in open areas and not under shade. In rubber and oil-palm plantations in Malaysia, if the canopy is thick, there is practically no weeds; but when light penetrates the canopy or at the edges of the fields, the weed is rampent.

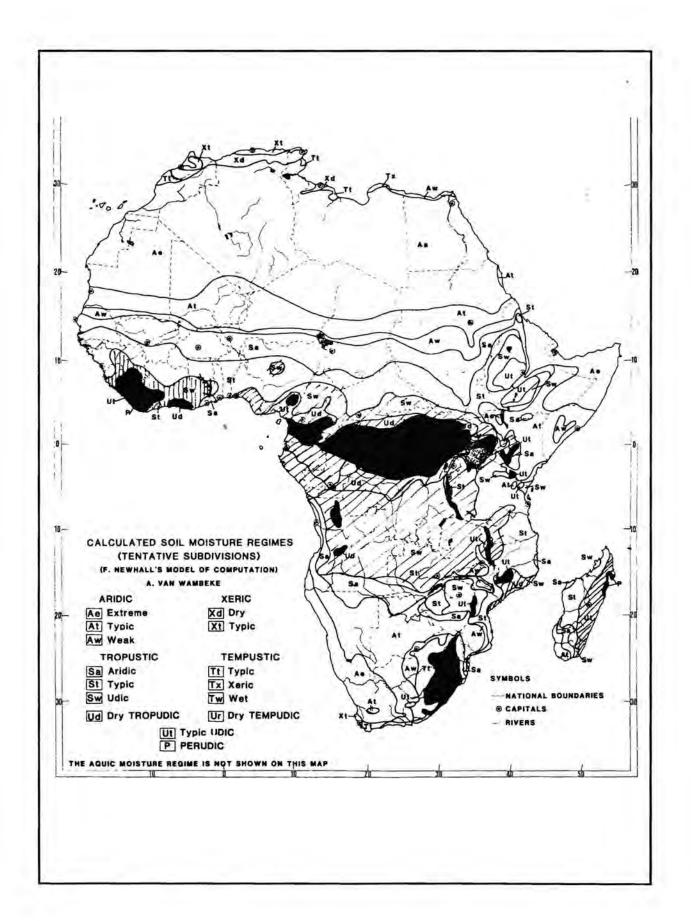
Soil scientists consider soil moisture and soil temperature regimes as soil properties. Both the Soil Moisture Regimes (SMR) and Soil Temperature Regimes (STR) may be computed from atmospheric data and the storage capacity of the soil and the regimes can be plotted on a map, as shown in the map of Africa.

The units, 'Ut and P' represent moist and humid areas of the warm tropics and these are also the areas where C. odorata is prevalent in Africa. The units 'Tw and Sw' and more specifically the latter, are the subunits of the regions with a dry season. These subunits, however, have a much longer moist season than the typical (Sa and St). The units 'Sw and Ud' represent areas which will be threatened in the near future with respect to spread of the weed. Although they are not ideal for the crop, the weed can survive under these conditions.

The map thus shows current and potential areas of spread of *C. odorata*. The map can be further refined if other soil properties are taken into consideration. The present scale of the map will not permit it but this can be done for any given country. Similar maps could be drawn for other parts of the world and this would present a more comprehensive picture of the situation. The agronomic requirements of the weed needs more attention and would contribute to its control.

Occurence of this weed is one of the first indicators of soil degradation. Colonization by C. odorata takes place upon deforestation or on abandoned land. It can be prevented or retarded by establishing a cover crops such as Pueraria sp. or Centrosema sp. Colonization also takes place on land which has been severely eroded and the acid subsoil is exposed to the surface. As it is tolerant to high soil acidity and aluminum saturation, it establishes before any other plant can establish. Once it is established, it has the advantage of reducing soil loss through erosion; this is a beneficial role. However, permanent eradiction is a problem, particularly in low-input agriculture and this is the major source of concern.

¹February 29 through March 4, 1988, Bangkok, Thailand. Proc. Publ. Agric. Exp. Station, Guam U.S.A.



PHYTOPHAGOUS INSECTS RECORDED FROM C. ODORATA*

R.E. Cruttwell McFadyen, Queensland Department of Lands Alan Fletcher Research Station, P.O. Box 36 Sherwood, 4075, Queensland, Australia

Thysanoptera

Elaphothrips sp. nr. angusticips (Crawf.), Yucatan, Mexico.

Haplothrips gowdeyi Franklin, India (Anon., 1983-1984).

Leptothrips sp., Yucatan, Mexico.

Orthoptera

Acrididae:

Abracis obliqua (Thun.), Yucatan, Mexico.

Chromacris miles (Drury), Venezuela (Guagliumi, 1966). Osmilia flavolineata (deG), Yucatan and Veracruz, Mexico.

Patanga succincta R., Thailand (Pholboon, 1965).

Sitalces trinitatis Bruner, feeding on many other plants,

Trinidad.

Zonocerus variegatus in Nigeria (Iheagwan 1983, Chapman

et al., 1986).

Pyrgomorphidae:

? Calamacris sp., Veracruz, Mexico.

Neorthacris acuticeps (Bolivar), India (Muniappan and

Viraktamath, 1986)

Tettigoniidae:

Conocephalus sp. ? cinereus Thun., Turrialba, Costa Rica.

Conocephalus ictus (Scudd), Veracruz, Mexico.

Gryllidae:

Nisitra vittata Scop., Sumatra (Naezer & Meer Mohr, 1953).

Hemiptera

Membracidae:

Acanophora concolor (Walk.), Trinidad.

Acutalis fusconervosa (Fairm.), Costa Rica.

Amastris sp., Costa Rica.

Bolbonota inaequalis (Fabr.) Costa Rica.

Bolbonota pictipennis Fairm., also on cacao and other

plants, Trinidad, Mexico and Yucatan.

Campylenchia hastata Fabr., also on pigeon pea, Trinidad. Ceresa vitulis (Fabr.), also on sugarcane, Trinidad and

Costa Rica.

Cocostrephus sp., India (Anon., 1983).

C. minutus (Fabricius), India (Muniappan and Viraktamath,

1986).

Cyphonia clavata F., also on pigeon pea and other plants,

Trinidad.

Cyphonia flavovittata Stal., Trinidad.

^{*}All records from Cruttwell, 1974 except where another reference is given.

Enchophyllum dubium Fowler, Yucatan and Costa Rica.
Entylia sp., also on guava, Trinidad.
Entylia carinata (Forster), Costa Rica.
Entylia gemmata (Germ), Venezuela (Guagliumi, 1966).
Hypsoprora coronata (Fabr.), Costa Rica.
Leptocentrus sp., India (Anon., 1983).
Membracis humilis Fowl., Trinidad.
Membracis tectigera (Stoll), Trinidad.
Micrutalis calva (Say), Trinidad, also on many plants in the U.S.A. (Beirne, 1959).
Micrutalis epihippium (Burmeister), Costa Rica.
Poppea capricornis Fowl., Trinidad and Costa Rica.
Sphongonophorus sp., Yucatan.
Sphongonophorus guerini Fairm., Trinidad.
Stegaspis viridis Funkh., also on other plants, Trinidad.

Plataspidae:

Coptosoma sp., India (Anon., 1983-1984).
Coptosoma siamicum Walk., Sumatra (Naezer & Meer

Mohr, 1953), also on crops in Ceylon (Hutson, 1930). Sepontia nigrofusca Dist., India (Anon., 1983-1984).

Pentatomidae:

Acrosternum marginatum (Beauvois), Mexico. Antestia anchora Thunb., Sumatra (Naezer & Meer Mohr,

1953), also on coffee in Asia.

Antiteuchus t. tripterus Fabr., also on Thunbergia, Trinidad. Edessa meditabunda F., also on other plants, Trinidad.

Edessa rufomarginata (DeGeer), Costa Rica. Euschistus obscurus (Beauvois), Mexico.

Podisus sagitta (Fabr.), Mexico.

Proxys punctulatus (Beauvois), Mexico.

Coreidae:

Anoplocnemis curvipes in Nigeria (Iheagwan 1983).

Anoplocnemis phasianus F., Sumatra (Naezer & Meer Mohr, 1953), also on numerous crop plants in Asia (Maheswariah & Puttarudriah, 1956).

Archimerus sp., Costa Rica.

Hypselonotus atratus Dist., Costa Rica.

Leptocorisa acuta (Thunberg), India (Anon., 1983-1984).

Mictis longicornis Westw., Sumatra (Naezer & Meer Mohr, 1953).

Riptortus pedestris (Fabricius), India (Anon., 1983-1984). Serinetha abdominalis Fabr., Thailand (Pholboon, 1965). Zicca taenida (Dallas), Costa Rica.

Lygaeidae:

Ligyrocoris abdominalis (Guerin), Yucatan, Mexico.
Ligyrocoris litigiosus (Sta.), Yucatan, Mexico.
Ochrimnus mimulus (Stal), Costa Rica.
Ochrostomus poeyi (Guerin), Mexico.
Ochrostomus verecundus (Distant), Mexico.
Ortholomus scolopax (Say), Costa Rica.
Pachybrachius bilobata (Say), Costa Rica.
Paromius longulus (Dallas), Yucatan, Mexico.

Pyrrhocoridae:

Dysdercus sp., Yucatan, Mexico.

Dysdercus cingulatus F., Sumatra (Naezer & Meer Mohr,

1953), also on many Malvaceae in India.

Dysdercus delauneyi Lett., St. Vincent, W.I., also on many

plants including cotton (Sands, 1917).

Dysdercus koenegii Fabricius, India (Anon., 1983-1984).

Dysdercus mimus (Say), Mexico and Costa Rica. Dysdercus obscuratus garskei Schmidt, Costa Rica. Dysdercus obscuratus obscuratus Distant, Mexico.

Jadera sanguinolenta Fabr., Trinidad.

Largus sp., Trinidad.

Macrocerca grandis Gray, Trinidad.

Tingidae:

Phymata simulans Stal. ssp. recifensis Ker., on flowers,

Trinidad.

Miridae:

Collaria oleosa (Dist.), Venezuela (Guagliumi, 1966).

Helopeltis? theivora Wat., Sumatra (Naezer & Meer Mohr,

1953), also on tea in India. Lopidea sp., Costa Rica.

Cercopidae:

Sphenorhina (Tomaspis) rubra (L.), also on Eupatorium sp.,

Demerara (Urich, 1914).

Cicadellidae:

Agallia sp., Mexico.

Agallia sp., Venezuela (Guagliumi, 1966). Agrosoma placetis Medler, Costa Rica.

Agrosoma sp., Costa Rica.

Catagonalia marginella Fabr., Yucatan, Mexico. Diedrocephala variegata (Fab.), Costa Rica.

Graphocephala sp., Costa Rica.

Gypona sp., Costa Rica.

Metascarta coeruleovittata (Sign), Venezuela (Guagliumi,

1966).

Omcometopia clarior (Walker), Yucatan, Mexico.

Omcometopia sp., Costa Rica. Parathona cayennensis G., Trinidad.

Poeciloscarta sp., Costa Rica.

Tettigella ceylonica Melich, India (Anon., 1983-1984).

Jassidae:

Sibovia occatoria Say, Trinidad, also on many other plants

in Mexico and Costa Rica.

Cixiidae:

Bothriocera sp., Yucatan, Mexico and Costa Rica.

Delphacidae:

Peregrinus maidis (Ashmead), Yucatan, Mexico.

Flatidae:

Gen. near *Docerus* sp. indet., Veracruz, Mexico. Euhyloptera corticalis Fenn., Trinidad.

Lawana conspersa Walk., Sumatra (Naezer & Meer Mohr,

1953), also on tea in Malaya (Corbett, 1935). Monoflata (sensu latu) sp., Veracruz, Mexico.

Poeciloptera phalaenoides L., Trinidad.

Acanaloniidae:

Acanalonia sp., Trinidad.

Acanalonia sp., Veracruz, Mexico.

Aleurodidae:

Aleurodicus trinidadensis Q & B., also on coconut,

Trinidad.

Bemisia tabaci Genn., Sumatra (Naezer & Meer Mohr,

1953), also on cotton and other crops (Laan, 1940).

Aphididae:

Aphis gossypii Glov., Trinidad, Nigeria (Iheagwam 1983), also on many plants in Thailand (Patch, 1938; Pholboon,

1965).

Aphis spiraecola Patch., Trinidad, also on many plants in

India (Bennett & Rao, 1968; Patch, 1938).

Brachycaudatus helichrysi (Kaltenbach), India (Joy et. al.,

1979).

Dactynotus ambrosiae (Thos.), Costa Rica, also on many

other plants (Patch, 1938).

Rhopalosiphum maidis Fitch, India (Ganguli and

Raychaudhuri, 1980).

Toxoptera odinae (v.d. Goot), India (Yadav et. al., 1981).

Orthezidae:

Orthezia insignis Browne, also on many crops, Trinidad.

Orthezia pseudinsignis Morrison, Mexico.

Coccidae:

Ceroplastes sp., Trinidad.

Saissetia sp., India (Muniappan and Viraktamath, 1986). Saissetia oleae Bern., also on many crops, Trinidad.

Pseudococcidae:

Dysmicoccus sp., Mexico.

Phenacoccus gossypii Townsend & Cockerell, Mexico, also

on many plants (McKenzie, 1967).

Pseudococcus sp., Sumatra (Naezer & Meer Mohr, 1953).

Lepidoptera

Tineidae:

Recurvaria sp. Adults reared in Trinidad from larvae feeding in flowers of C. odorata and Condylidium iresinoides (H.B.K.) K & R, which are only present December_May; other plants probably attacked during remaining months. Eggs laid singly in flower-heads, larvae feed in developing seeds. Mature, larvae pupate in flower-head without a cocoon; adult emerges in one to two weeks. Each larva destroys the seeds in one flower-head.

Larvae parasitised by a Eulophid, Euderus sp.

Lyonetidae:

Bucculatrix sp. Larvae collected mining leaves of C. odorata in Mexico and of E. hookerianum in Tucuman, Argentina. A similar species occurs in Trinidad attacking C. iresinoides but not C. odorata. Larvae are solitary and pupate in the mines.

Stenomidae:

Antaeotricha sp., from pupa on leaf, Costa Rica.

Gelechiidae

Dichomeris (Trichotaphe) sp. nr. eupatoriella Cham., leafroller, also on C. ivaefolia, Trinidad. Adults reared from leaf-rolling larvae on C. odorata were identified as a new species of Dichomeris in the delotella sub-group. For biology and host range see Cruttwell, 1973b.

Larvae parasitised by the solitary endoparasitic Braconids Xanthomicrogaster seres Nixon and Apanteles sp., and pupae by a third solitary endoparasitic Braconid as yet unidentified.

Dichomeris sp. nov. 2. Adults reared from leaf-rolling larvae on C. odorata in Belem, Brazil were identified as a second new species of Dichomeris, Life-history similar to Dichomeris sp. 1.

Tortricidae:

Amorbia catenana Wals., leaf-roller, Trinidad, also on banana in Brazil, Antilles and Central America (Da Costa Lima, 1951).

Amorbia emigratella Busck., leaf-roller, Veracruz, Mexico. Archips micaceanus (Walker), India (Muniappan and Viraktamath, 1986).

Platynota sp., leaf-roller, Yucatan.

Platynota rostrana (Wlk.) leaf-roller, Mexico.

Sparganothis restitutana Wkr. Reared from larvae feeding on the Composites Wedelia trilobata (L.), W. caracasana DC., Wulffia baccata (L.), Synedrella nodiflora (L.) Gaertn., C. odorata, C. ivaefolia and C. iresinoides.

Eggs laid in clusters on leaves, newly hatched larvae disperse and feed initially on leaf buds. Larger larvae feed on leaves or flowers, living and pupating in a loose leafroll or between leaves or flowers fastened together with silk.

Larvae parasitised by a Braconid and an Ichneumonid as yet unidentified, and pupae by another Ichneumonid and by Spilochalcis sp.

Cochlidae:

Phalonidia multistrigata Wals., feeding in flowers, Trinidad.

Pterophoridae:

Adaina bipunctata Moeschl. Trinidad, reared from flowers of C. odorata and C. iresinoides. Recorded from U.S.A. (McDunnough, 1939) and Puerto Rico, where "larvae were intercepted on Pluchea purpurascus" (Wolcott, 1948). Flowers of other Composites probably attacked when C. odorata not in flower. Larvae stout and cream-colored, feeding in flower-heads, entering new ones by chewing through involucral bracts. Mature larva pupates inside flower-head without a cocoon, adult emerges in a few days.

Larvae attacked by a solitary endoparasite, *Bracon* sp. nr. vulgaris Ashm.

Adaina sp., larvae in hollow stems, Veracruz, Mexico.

Thyrididae:

Dysodia oculataria Clem., leaf-roller, Yucatan, Mexico.

Palthis sp. nr. agroteralis (Guen.), leaf-roller, Veracruz,
Mexico.

Pyralidae:

Mescinia parvula (Zeller). Larvae shoot-borers attacking C. odorata and C. ivaefolia in Trinidad; similar larvae collected on C. odorata in Veracruz, Mexico and Belem, Brazil, and on E. hookerianum in Tucuman, Argentina. The biology and host-range of the species have already been described (Cruttwell, 1977a), under the name Mescinia sp. nr. parvula.

Larvae in Trinidad attacked by eight hymenopterous and one Tachinid parasite. One species, a gregarious ectoparasite, *Hormius* sp. nov., also attacks small larvae of *Hypsipyla grandella* (Zell.) in mahogany and cedar in Trinidad. There are four other ectoparasites, *Ipobracon* sp., *Hormius* sp., *Euderus* sp., *Horismenus* sp., and two solitary egg-larval endoparasites emerging from the prepupal larvae, *Phanerotoma* sp., and *Microchelonus* sp. *Parasierola* sp. and the Tachinid, as yet unidentified, are both solitary endoparasites of the larvae.

Herpetogramma sp. ? bipunctalis (Fabr.), pupa on leaf, Trinidad.

? Hyalospila sp., larvae feeding in open gall in stem tips, in Costa Rica, and Veracruz, Mexico.

Laetilia portoricensis (Dyer), from withered stem, Puerto Rico (Wolcott, 1948).

Loxostege new sp., leaf-roller, Veracruz, Mexico and Costa Rica.

Pionea (Hapalia) upalusalis Wkr. Reared from larvae feeding on C. odorata, C. ivaefolia, Fleischmannia microstemon, Austroeupatorium inulaefolium and Ageratum conyzoides in Trinidad. Also recorded from Puerto Rico and West Indies generally and from Venezuela (Wolcott, 1948).

Green spherical eggs laid in groups of one to three on the underside of the leaves, newly hatched larvae live in a silken tube on the underside and older larvae in a leaf-roll, usually in the leaf centre. Larvae pale green in colour until mature, when they become pink with white lines and pupate in the tube. No parasites known.

Psara ambitalis Reb., defoliator, Sumatra (Naezer & Meer Mohr, 1953), also on tomatoes and tobacco in Sumatra (Laan, 1940).

Geometridae:

Apicia asterica Druce, defoliator, Morelos, Mexico.

Chloropteryx languescens Warr., feeding on flowers,

Trinidad.

Eupithecia sp., reared from gall in stem tip, Trinidad.

Eupithecia sp. nr. maleformata Warr., feeding on flowers, Trinidad.

Hyposidra talaca (Walker), India (Muniappan and Viraktamath, 1986).

Racheospila rufilineata Warr., feeding on flowers, Trinidad.

Synchlora sp. ? frondaria Gn., defoliator, Trinidad.

Sphingidae:

Pholus labruscae Moss., defoliator, Trinidad (Moss, 1912), also on Vitis, Cissus and Ampelopsis in Guadeloupe

(D'Aguilar, 1966).

Noctuidae:

Chrysodeixis chalcites (Esper), India (Muniappan and

Viraktamath, 1986).

Perigea albigera Guen., defoliator, also on F. microstemon, Trinidad, Costa Rica and Yucatan, and on

Chrysanthemum, Barbados (Bourne, 1921).

Spodoptera (Prodenia) latifascia Wkr., defoliator, Trinidad, also on tomatoes and other crops in Puerto Rico (Wolcott,

1948).

Arctiidae:

Diacrisia (Spilosoma) alcumena Berg., feeding on C. odorata, Bolivia, also on mango, Venezuela (Guagliumi, 1966).

Diacrisia obliqua Walker, India (Anon., 1983-1984).

Paraeuchetes pseudoinsulata Rego Barros (Incorrectly recorded as Ammalo insulata Walker in Bennett and Cruttwell 1973 and Cruttwell 1974), defoliator on C. odorata and C. ivaefolia in Trinidad.

Pareuchetes insulata (Walker) defoliator on C. odorata and Ageratum in Florida, Central America and Venezuela

(Cock and Holloway 1982).

Pericallia ricini (Fabricius), India (Anon., 1983-1984).

Riodinidae:

Calephelis laverna G. & S. This species occurs in Brazil, Venezuela and Central America, as well as in Trinidad (Barcant, 1970) where larvae have been collected feeding on C. odorata, C. ivaefolia, F. microstemon and Hebeclinium macrophyllum (L.) DC.

Larvae solitary and sluggish, pale green, covered with long silky green hairs. They feed on the leaves then pupate inside a silk cocoon attached to leaves or stem.

Larvae parasitised by a gregarious Apanteles sp.

Lycaenidae:

Thecla palegon Cr., feeding on flowers, also on C.

iresinoides, Trinidad.

Acraeidae:

Actinote anteas Doubleday, defoliator, Costa Rica. Recorded from Trinidad (Barcant, 1970).

Danaidae:

Pteronymia lincera H-S., Venezuela (Guagliumi, 1966).

Diptera

Ceratopogonidae:

Forcipomyia sp., reared from nail-gall on leaf, Turrialba, Costa Rica.

Cecidomyidae: (see Gagne, 1977)

Asphondylia corbulae Mohn, (see Gagne 1977) reared from flowers of Eupatorium sp. in El Salvador (Mohn, 1960) and of C. odorata and F. microstemon in Trinidad. Larvae feed singly inside developing achenes which swell to a gall two to three mm in diameter. Two or three galls form per flower-head and few seeds develop. Parasitised by Tenuipetiolus sp. (Eurytomid), Horismenus sp., Galeopsmyia sp. 3, Tetrastichus dimachus Walk. and Leptacis sp. 2 (Eulophidae).

Contarinia sp. nr. perfoliata Felt, reared from flowers of C. odorata, C. ivaefolia and F. microstemon in Trinidad. Larvae live and feed in corollas or between achenes, causing little apparent damage. Pupation occurs in the flowers and the adults emerge in a few days.

Dasyneura corollae Gagne, larvae singly within petal-tubes

of young flowers, Trinidad and Bolivia.

Clinodiplosis (Hyperdiplosis) eupatorii (Felt), reared from conical nail-galls on upper surface of leaves of Eupatorium sp. in St. Vincent, W.I. (Felt, 1911) and of C. odorata and C. ivaefolia in Trinidad, Costa Rica and Belem, Brazil.

Larvae in Trinidad parasitised by Horismenus sp., Galeopsymia sp. 1, Achrysocharis new sp. and Leptacis sp. 1, the last two also occurring in Belem, Brazil.

Clinodiplosis sp. Adults reared from bud-galls in C. odorata in Costa Rica. Larvae occur singly in hollow pear-shaped galls, three to five mm long, in stem tips or axillary buds, with several small leaves developed without internodes beneath the gall, giving a 'rosette' appearance.

No parasites are known.

Clinodiplosis sp., from bud-galls in C. odorata and C. ivaefolia in Trinidad. one to three larvae live between the bud leaves of stem tips or axillary buds, destroying the tissue and preventing further growth. The bud leaves swell slightly and become red and densely covered with hairs. Mature larvae leave the gall and pupate just below the soil surface; adults emerge in 11 to 18 days. The species is widespread and abundant in Trinidad, breeding throughout the year.

Larvae are attacked by the predatory cecidomyid Lestodiplosis callipus Gagne and by the parasites Tetrastichus sp. and Patasson sp.

Neolasioptera cruttwellae Gagne. Adults reared from stem galls in C. odorata and C. ivaefolia in Trinidad and Bolivia. Galls develop in young shoots and when mature reach about one cm diameter. one to three larvae feed and pupate in tunnels in each gall.

Larvae in Trinidad parasitised by a solitary parasite Metanopedias brunneipes (Ashm.) and by a gregarious ectoparasitic Ceraphronid as yet unidentified, and in Bolivia by four species of Hymenoptera, Leptacis sp. (Eulophid), Aphariogmus sp. (Ceraphronid), Rhoptrocentrus sp. (Braconid) and Eupelmus sp. (Eupelmid).

Neolasioptera frugivora Gagne, adults reared from flowers of C. odorata, and adults probably of this species from F. microstemon, both in Trinidad. Larvae singly inside the achenes, each consuming one achene and pupating inside. No external damage caused. No parasites known.

Perasphondylia reticulata Mohn. Adults reared from budgalls on C. odorata and Eupatorium sp. in El Salvador (Mohn, 1960) and from C. odorata and C. ivaefolia in Trinidad, in Belem, Brazil, and in Bolivia. Larvae occur singly in a hollow pear-shaped gall, seven to nine mm long and five to six mm wide, in stem tips and axillary buds. Species scarce and confined to the cooler valleys in Trinidad but in Brazil and Bolivia galls were more common.

The following Eulophids were reared from galls in Trinidad: Tetrastichus valerus Walker, Paragaleopsymia sp., Galeopsymia sp. 1 and Eurytoma sp. from pupae. In Bolivia, Galeopsymia sp. 2 and Rileya sp. were reared from galls.

Procecidochares new sp., reared from stem-galls in C. odorata in Veracruz, Mexico, in Belem, Brazil, and in Bolivia. Similar adults from stem-galls in C. laevigata in Bolivia did not attack C. odorata. Eggs inserted into the stem tip by the female; abnormal growth of the stem starts before the eggs hatch.

Larvae feed in curved tunnels in the gall tissue, one to seven larvae in separate tunnels in the gall. Mature larvae pupate in the tunnel below an epidermal 'window' through which the adult emerges. Galls slow and distort but do not arrest further growth of the stem.

Trypetidae:

Larvae in Brazil parasitised by a gregarious Braconid Heterospilus pallidipes Ashm. and by Heterospilus sp. nr. humeralis Ashm.; in Bolivia by Heterospilus sp. 1 and Syntomosphyrum sp.; in Mexico by Torymus umbilicatus (Gahan), Eupelmus sp., Neocatolaccus sp. and an indet. Pteromalid. Larvae and pupae from C. laevigata in Bolivia were parasitised by Heterospilus sp. 1, Dimeromicrus cecidomyidae Ashm. and Eupelmus sp.

Cecidochares fluminensis (Lima). Larvae of this species, previously recorded from S.E. Brazil (Aczel, 1953), feed in the flowers of C. odorata and C. ivaefolia in Trinidad. In December and January eggs are inserted singly into the flower buds, and the fat, white pilose larvae feed on the developing achenes in one flowerhead, pupating in the cavity formed. Puparia oval, black and pilose; adults emerge in 10-14 days, probably remain in sexual diapause for nine to ten months until C. odorata flowers again.

A Pteromalid, *Pseudocatolaccus* sp., has been reared from the pupae.

Euaresta? bellula Snow., from flowers, Trinidad.

Polymorphomyia basilica Snow, galls stems of C. odorata in Puerto Rico (Wolcott, 1948).

Trupanea sp. from galls, Venezuela (Guagliumi, 1966).

Xanthaciura insecta (Loew.), larvae in flowers, also of F. microstemon, Ageratum conyzoides, Wedelia caracasana, Trinidad, from C. odorata, Bolivia, from flowers of Bidens pilosa, Florida (Needham, 1946).

Caliope sp., from flowers, also of C. ivaefolia, Trinidad. Sapromyza sp., from pupae on leaves, Trinidad.

Olcella pleuralis Becker, from flowers, also of C. ivaefolia and C. iresinoides, F. microstemon, Ageratum conyzoides, Aspilia verbesinoides, Wedelia caracasana and Wulffia baccata, Trinidad.

Agromyza eupatoriae Mall., mines leaves, U.S.A. (Frost, 1924).

Calycomyza flavinotum Frick., mines leaves, Trinidad, also on E. purpureum, Viburnum, Arctium and Alomia, U.S.A. (Frick, 1959).

Calycomyza jucunda (Wulp.), mines leaves, also of other

plants, Puerto Rico (Wolcott, 1948).

Melanagromyza eupatoriella Spencer, from stem tips of C. odorata and C. ivaefolia in Trinidad, in Belem, Brazil and in Bolivia. Larvae occur singly in young shoots, tunnelling spirally down the stem, destroying the conducting tissue and killing the shoot for about 6 cm. Mature larvae pupate in the hollow stem after cutting an epidermal window through which the adults emerge.

Lauxanidae:

Oscellinae:

Agromyzidae:

Breeding is continuous throughout the year; the species is abundant generally.

In Trinidad, larvae attacked by Euderus spp., and an indet. Pteromalid, and by two larval-pupal parasites, Eurytoma walshii How. and Tropideucoila rufipes Ashm. In Bolivia larvae attacked by Tropideucoila sp., and in Brazil by Euderus sp., Eurytoma sp. and Opius sp.

Melanagromyza longicaudalis Mall., on flowers, Bolivia and Jamaica (Spencer, 1963).

Melanagromyza mallochi (Kendel), bores in stem, Puerto

Rico (Spencer, 1963).

Melanagromyza minima Mall., reared from flowers of C. odorata, C. ivaefolia, C. iresinoides and Wedelia trilobata in Trinidad. one to three larvae in a flower-head, young larvae feeding within single achenes and older larvae between achenes. Each larva destroys 20 to 30 achenes and pupates in the cavity left.

Two parasites, *Tetrastichus* sp. and an unidentified Chalcid, reared from pupae.

Coleoptera

Lampyridae: Cantharidae: ? Psilocladus sp., Morelos, Mexico. Belotus sp., Morelos, Mexico. Canthari sp., Turrialba, Costa Rica. Silis sp., Veracruz, Mexico.

Elateridae:

Aelolus sp. nr. facetus Candeze, Veracruz, Mexico.

Glyphonyx sp., Veracruz, Mexico.

Helodidae:

Cychon sp., Turrialba, Costa Rica.

Languridae:

Langurites lineata (Cast), Veracruz, Mexico.

Anobiidae:

Cryptorama sp., Veracruz, Mexico.

Lamiidae:

Aerenica hirticornis Klug., recorded from Argentina and Central and Southern Brazil (Guerin, 1953); collected from C. odorata in Trinidad and in Santa Cruz, Bolivia. No other host known.

Adults 10-15 mm long, pale buff colour with darker brown markings; when resting in typical position with the head on the plant stem and the body projecting up at an angle, closely resemble the dried flower-heads. Adults present in Trinidad from June -August, feed by scraping tissue from stem tips, killing these. Eggs laid singly near stem tips, larvae feed in the pith. Larvae full grown and have hollowed one to two metres of stem by October or November. As only the pith destroyed, stem growth not

affected. Frass ejected through holes along the stem. Larvae remain in stem until May when they pupate near the stem base. Adults emerge seven to ten days later, remain quiescent for a further two to three weeks; adult activity may be initiated by the rains which commence at this season.

In Bolivia, south of the Equator, half and full_grown larvae were present in April and May. Life-cycle synchronized with the host, with active stages present in the season of maximum plant growth.

In Trinidad, young larvae are attacked by a solitary endoparasitic Eulophid.

Cerambycidae:

Ataxia sp., larvae boring in stem, Yucatan, Mexico.
Lophalia sp. nr. cyanicollis (Dupont), adults feeding inside stem base, Yucatan, Mexico.

Chlamisidae:

Aulocochlamys sp. Adults, black, 1.8-2.5 mm long, and larvae feed by scraping stems and leaf petioles of C. odorata and C. ivaefolia. Eggs laid singly in cylindrical ribbed cases of faecal matter. These form the apex of the larvae case, being gradually enlarged into a conical case 3.5-3.7 mm long and 1.6 mm maximum diameter. Mature larvae attach the case to the stem, pupate inside, and adults emerge in one to two weeks. Widespread in Trinidad, occasionally abundant in the valleys of the Northern Range.

No parasites known.

Chlamisus insularis Jac. Recorded from Mexico and Panama (Blackwelder, 1957); adults black with golden markings, 3.3-4.3 mm long and 2.5-3.0 mm maximum width. Widespread in Trinidad throughout the year but not abundant.

Life-history similar to Aulocochlamys sp.; egg case 1.4 mm long and 1.0 mm diameter, mature larval case conical with a rough surface, six to seven mm long. Feeding adults collected on C. odorata, C. ivaefolia and Bidens pilosa.

A black Eulophid reared as a solitary egg parasite.

Hispidae:

Pentispa explanata Chap. Recorded from Mexico to Colombia (Blackwelder, op. cit.), and Venezuela where it is recorded on Pithecoctenium sp. (Bignoniaceae) (Maulik, 1937). In Trinidad adults collected on C. odorata and C. ivaefolia would not feed on Pithecoctenium echinatum Jacq. when tested.

Adults, present throughout the year, feed by scraping away the tissues of the leaf from below, leaving characteristic scars. March-April, congregate and mate in groups of six to 20. Eggs laid April-July, inserted singly under the leaf epidermis, covered with a faecal plug. Larvae hatch in 12 days and mine the leaves, forming irregular blotch mines two to three cm in diameter when full size, 20-25 days later. Mature larvae pupate in the mine; adults emerge in five to eight days. Newly-emerged adults disperse and feed on the leaves but do not breed until the next year.

P. explanata occurs on C. odorata throughout Trinidad but is rare except in the northern valleys. Adults avoid open sunlight; in the laboratory, adults survive but do not breed in cages exposed to the sun.

Larvae parasitised by a solitary ectoparasitic Elasmid, Austelasmus sp., and are taken by predatory wasps especially Polistes and Polybia species.

Chrysomelidae:

Antipus? mutabilis Lac., Morelos, Mexico. Cephaloleia? limonensis Uhman, Turrialba, Costa Rica. Chelymorpha sp., Morelos, Mexico. Colaspis sp., Morelos, Mexico. Colaspoides batesi Jac., Turrialba, Costa Rica. Corynodes sp., India (Anon., 1983-1984). Cryptocephalus? trizonatus Suffr., Veracruz, Mexico. Cryptocephalus 18-punctatus Suffr., Veracruz, Mexico. Ctenochira cumulata (Bog), adults defoliators, also on Citrus, Coffea, and other plants, Costa Rica. Ctenochira? ferranti Spaeth, Veracruz, Mexico. Diabrotica sp., Turrialba, Costa Rica. Disonycha sp., Veracruz, Mexico. Disonycha sp. nr. glabrata Fab., Turrialba, Costa Rica. Disonycha sp. nr. politula Horn., Turrialba, Costa Rica. Exema sp., Veracruz, Mexico. Glyptoscelis sp., adults defoliators, also on other plants, Trinidad.

? Hecataeus sp., Yucatan, Mexico.
? Maecolaspis spp., Veracruz, Mexico.
? Malacosoma sp., Yucatan, Mexico.
Mesomphalia sp., Veracruz, Mexico.
Metacycla marginata Chap., Yucatan, Mexico.
Metriona sp. nr. tuberculata F., Veracruz, Mexico.
? Monolepta sp., Veracruz, Mexico.
Nodonota sp., Veracruz, Mexico and Turrialba, Costa Rica.
Omophoita sp., Veracruz, Mexico.
Pachybrachys sp., Morelos, Mexico.
Physonota sp. nr. alutacea Boh., Veracruz, Mexico.
Plectrotreta? clarkei Jac., Guanacaste, Costa Rica.
Plectrotreta? dogrni Jac., Veracruz, Mexico.
? Rhabdopterus sp., Veracruz, Mexico.
Saxinis sp., Veracruz, Mexico.

Zygogramma sp., Veracruz, Mexico.

Bruchidae:

Acanthoscelides oblongoguttatus (Fahreus), also larvae in seeds of Acacia spp., Mexico.

Caryedon sp., India (Anon., 1983-1984).

Curculionidae:

Amblyrrhinus sp., India (Anon., 1983-1984). Apion sp., India (Anon., 1983-1984).

Apion brunneonigrum B.B. Recorded from Venezuela and Argentina (Blackwelder, 1957) as well as Trinidad; the only hosts known are C. odorata and C. ivaefolia.

Biology of this weevil has already been described (Cruttwell, 1973a). In Trinidad, C. odorata flowers in late December; weevils oviposit in flower-buds December-January, larvae feed and pupate in the flowers, adults emerge February-March. Newly-emerged adults initially remain on the flower-heads, then disperse to plants in shaded areas where they feed on buds and young leaves. Adults remain sexually immature until October, when the reproductive system begins to develop. In November when flower-buds are forming on the host plant, the weevils congregate on suitable plants, and mating followed by oviposition begins.

Astycus aurovittatus Heller, India (Anon., 1983-1984).

Rhodobaenus sp. nr. cariniventris Champ. Adults feed on stems and leaf petioles of B. pilosa, C. odorata, C. ivaefolia and A. inulaefolium, and larvae in the stems of the three latter species. Adults often found together with Rhodobaenus 13-punctatus Ill. whose larvae do not feed in these 3 species, but have been collected from stems of B. pilosa in Trinidad, and in the U.S.A. are recorded from several Composites including Eupatoriadelphus purpureus (Satterthwaite, 1948).

Life-history of R. sp. nr. cariniventris described by Bennett (1955). Eggs are laid in stem tips and larvae feed in stems. When mature, larva cuts off the tip of the hollow stem; the piece about 2 cm in length containing the larva falls to the ground. The open ends are plugged with frass and the larvae pupates within.

Rhodobaenus ypsilon Cheor., reared from larvae collected in stems of C. odorata in Vera Cruz, Mexico. Adults feed on stems and foliage.

Adults of the following Curculionids feed on the flowers, buds and leaves of *C. odorata*:

Antenistes attennuatus (Fabr.), Trinidad. Anthomus sp., on flowers, Trinidad.

Apion sp., Morelos, Mexico.

Baris sp., common on flowers and leaf-buds, also attacks

other Composites, Trinidad.

Brachyomus octotuberculatus (F.), on leaves, also attacks crop and garden plants, Trinidad and Venezuela (Guagliumi, 1966).

Centrinaspis spp., on flower and leaf-buds, Trinidad and

Costa Rica.

Coleocerus ? setosus Boh., Morelos, Mexico.

Compsus simoni Faust., on leaves, also attacks other plants, Trinidad and Venezuela (Guagliumi, op.cit.).

Derosomus sp., Yucatan, Mexico.

Eustylus puber Oliv., on leaves, also attacks crop plants, Trinidad.

Exopthalmus jekelianus White, Turrialba, Costa Rica.

Glyptobaris? viduata (F.), Trinidad.

Hoplopactus sp., adults defoliators, Trinidad.

Hypomeces squamosus F., Sumatra (Naezer & Meer Mohr, 1953), also attacks crop plants (Hung, 1966).

Lixus sp., adults defoliators, also attacks other Composites, Trinidad.

Lixus sp. ? impressicollis Boh., Porto Alegre, Brazil. Lixus sp. nr. nigrinus Champ., Yucatan, Mexico.

Myrmex sp., Yucatan, Mexico.

Myrmex sp. nr mexicanus (Chevr.), Veracruz, Mexico.

Pantomorus spp., Veracruz, Mexico.

Promecops sp., on buds and leaves, Trinidad.

Sibinia sp., Trinidad.

Meloidae:

Mylabris sp., India (Anon., 1983-1984).

Acarina

Eriophyidae:

Acalitus adoratus Keifer, causing erineum growth on leaves

and stems, Trinidad, Brazil and Bolivia.

Calacarus sp., India (Muniappan and Viraktamath, 1986).

Phyllocoptes cruttwellae Keifer, on leaves, Trinidad.

Biology of both mites described in Cruttwell 1977b.

Oribatidae:

Eremulus flagellifer Berlese, India (Ramani and Haq, 1983).

Galumna sp., India (Ramani and Haq, 1983).

Lamellobatus palustris Hammer, India (Ramani and Haq, 1983).

Parolamellobates bengalensis Bhaduri and Raychaudhuri, India (Ramani and Haq, 1983).

Pelokylla malabarica Clement and Haq, India (Ramani and Haq, 1983).

Scheloribates sp., India (Ramani and Haq, 1983).

Tarsonemidae:

Polyphagotarsonemus latus Banks, India (Muniappan and

Viraktamath, 1986).

Tetranychidae:

Tetranychus sp., India (Muniappan and Viraktamath, 1986).

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