

A New Species of Genus *Plakortis* Schulze 1880 (Porifera: Homoscleromorpha) from Badabalu, Andaman and Nicobar Islands, India

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Vibha V. Ubare and PM. Mohan (2016) Recently revealed Class: Homoscleromorpha play a significant role in sponge systematics. The smaller size, cryptic habits and unresolved species complex are greater obstacles for the taxonomic identification of this class. Out of < 100 species, only 6 species have been described from the Indian region. Till date, only one species of the genus *Plakortis*, that is, *P. simplex* has been described from this region. The present study provides a detailed description of new species of the genus *Plakortis*, Schulze 1880. *P. badabaluensis* sp. nov. is characterized by the two size classes of diods (one is thick and the other is thin), triods and smooth microrhabds; with well differentiated ectosome, presence of subectosomal lacunae and confused choanosome. The identification of this new species has increased the number of plakinid species from one to two in the Andaman and Nicobar Islands, India. Further detailed studies are required to explore this type of cryptic species in this region.

Key words: Homoscleromorpha, Plakortis, Taxonomy, Badabalu, Pongibalu, Andaman Islands.

BACKGROUND

Marine sponges are very important and dominant in the benthic communities of oceans. They are one of the most primitive multicellular animals grouped under the phylum Porifera. Sponges are the most ancient multicellular living animal on earth today (Hooper and Van Soest 2002). Most zoologists are of the opinion that sponges diverge earlier than the main line of evolution and gave rise to other group of animals (Jacob 1973; Knoll 2011; Brain et al. 2012; Riesgo et al. 2014).

Homoscleromorpha Bergquist, 1978 is a small group of Porifera with unique traits such as: flagellated pinacocytes, basement membrane linings such as choanoderm and pinacoderm, oval to spherical choanocyte chambers with large choanocyte, and a viviparous cinctoblastula

larva (Van Soest et al. 2012, 2014). The homoscleromorpha is the only sponge lineage in which cell layers are very similar to those of eumetazoans, both in larvae and adults, and can be considered as a true epithelium (Ereskovsky et al. 2009). Further, recent molecular analyses also clearly separated the Homoscleromorpha from the Demospongiae clade (Gazave et al. 2012). Previously, homoscleromorpha was under a family or a suborder of the subclass Tetractinellida, within the class Demospongiae. At present, it is the fourth major lineage composed of two families, Oscarllidae Lendenfeld, 1887 and Plakinidae Schulze, 1880 (Gazave et al. 2010). It contains < 100 described species. Based on morphological traits, they have been traditionally subdivided into 8 genera Corticium, Oscarella, Placinolopha, Plakina, Plakinastrella, Plakortis, Pseudocorticium and Tetralophophora. All are exclusively marine

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and mainly located in shallow waters ranging from 8 to 60 m, but few are also recorded from abyssal depths, that is, up to 2460 m (Murciy and Diaz 2002; Gazave et al. 2010). The genera are distinguished by four morphological characters: 1) the presence of a siliceous skeleton; 2) presence of a cortex associated with leuconoid aquiferous system and well developed mesohyl or a sylleibid aguiferous system with poorly developed mesohyl and ectosome; 3) number of spicule size classes; and 4) the presence of different types of ramification in the actines of calthrops (tetractinal spicules) with three distinct general morphology have been recognized (Murciy and Diaz 2002; Gazave et al. 2010, 2012; Ereskovsky et al. 2013). Some of the morphological features are shared with the eumetazoans, which are absent in other groups of sponges. It is a taxonomically poorly studied group among the sponge clade as a result of insufficient sampling due to small size, rarity and cryptic habits of many species, as well as the presence of unresolved species complexes.

The Indian coastline runs over a distance of 7500 km, even with the vast coastal region, the only area that has been studied in detail by various spongologist is the Gulf of Mannar (Burton 1930). In addition, few occasional reports are available in certain parts of India. Most of the studies on marine sponges of India are from Southern India (Ali 1956; Thomas 1970a, b and c; 1973a, 1976a and b, 1984), Lakshadweep and Minicoy Islands (Thomas 1973b, 1979, 1980a and b, 1989), North-west India from the Gulf of Kutch and Cambay (Gulf of Khambat) (Dendy 1916; Thomas 1976b; Thomas et al. 1996). The fauna of Andaman and Nicobar Islands is very distinct and diverse (Ramakrishna et al. 2010). The taxonomic information of the sponges of Andaman and Nicobar Islands is far from complete and prompted the necessity of this work (Pattnayak 2006). Few researchers like Burton (1928), Burton and Rao (1932), Thomas (1977) and Pattnayak (2006) are the foremost contributors in the history of sponges from the Andaman and Nicobar Islands. Also, the recent reports of Immanuel and Raghunathan (2011), Krishnan et al. (2012) and Vinod et al. (2012) are acknowledged.

Till date, from the Class Homoscleromorpha, the species *Plakina monolopha* Schulze, 1880, *Plakina trilopha* Schulze, 1880, *Placinolopha acantholopha* (Thomas, 1970), *Corticium acanthastrum* Thomas, 1968, and *Corticium candelabrum* Schmidt, 1862 have been identified and described in India (Thomas 1968 and 1970b).

Only one species: *Plakortis simplex* Schulze, 1880 was check listed by Krishnan et al. (2012) in the Andaman and Nicobar Islands. This genus is well known world-wide for its several interesting natural products and bioactive compounds. This chemical diversity makes the genus *Plakortis* an interesting target group for pharmacological studies (Muricy 2011).

The aforementioned studies on sponge in the Andaman Islands have paved the way for further exploration of this subject. Therefore, an attempt has been made to describe more sponges from this region. During the process of identification of sponges in this region, the present new species evolved and has been described as a new report.

MATERIALS AND METHODS

The specimens were collected by Skin Diving from both regions: Badabalu (Lat. 11°30'37.04"N, Long. 092°41'09.28"E) and Pongibalu (Lat. 11°30'51.88"N, Long. 092°39'22.44"E) (Fig. 1). All underwater photographs were taken using underwater Sony Cyber Shot 13.6 Mega Pixel Camera. The collected specimens were fixed in 70% ethanol and deposited in the Zoological Survey of India (ZSI), Regional center of Andaman and Nicobar Islands and Department of Ocean Studies and Marine Biology (DOSMB), Pondicherry University, Port Blair. Spicule slides were prepared by dissociating small fragment of the sponge in boiling Nitric acid. Thick hand cut sections were made using surgical blade for skeleton and were observed under light stereomicroscope Leica M205C with inbuilt camera. Five to 20 Spicules of each kind were measured per individual through light microscope Carl Zeiss AXIO Vert.A1 with inbuilt camera. Spicule measurements were expressed as range length/ range width in μm (number of measurements). The description has been compared with the existing literature (Table 1).

RESULTS

Systematics

Class - Homoscleromorpha Bergquist 1978

Order - Homosclerophorida Dendy 1905

Family - Plakinidae Schulze 1880

Genus - Plakortis Schulze 1880

Definition: The skeleton formed by diods and triods is either in variable abundance or may be absent. Different types of microscleres such as microrhabds, quasiamphiasters, spined diods and spheres were also present in some species. Calthrops are absent in this genus (Muricy 2011).

Plakortis badabaluensis sp. nov.(LSID urn:lsid:zoobank.org:act:D746456C-2AEB-43B3-

926C-706EC11EF12C) Specimen examined (4). Holotype: ZSI/ ANRC 12102, Badabalu, depth 4.5 m, coll. M. Muruganantham, 15 February 2014. Paratypes: ZSI/ ANRC 12103, Pongibalu, depth 4.0 m, coll. PM. Mohan, 01 March 2014; DOSMB-00107: Badabalu, depth 3.5 m, coll. PM. Mohan; 15 February 201; DOSMB-00108, depth 5.5 m, coll. M. Muruganantham; 15 February 2015.

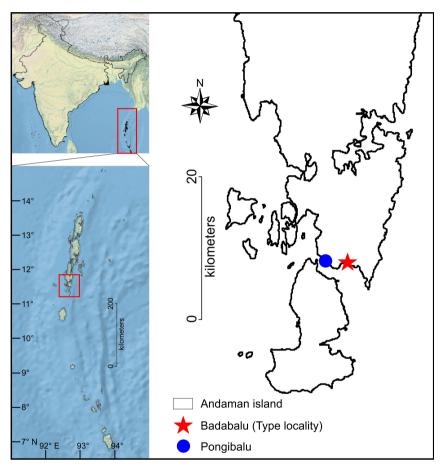


Fig. 1. Study Area.

Table 1. Comparison of the morphological and anatomical features of *Plakortis badabaluensis* sp. nov. with other species of Plakortis genus

	Form	Colour	Oscule Shape	Surface	Consistency	Diods
P. simplex	Incrusting	Bright rosy lavender	Not visible	Smooth/ roughend	Soft	(Centrotylote) 60- 150/3-6
P. nigra	Thickly encrusting	Black	Few	Smooth/ porous	Firm	Thin, 20-90
P. copiosa	Cushion shaped	Brown outside, cream inside	-	_	Fragile	55-100
P. erythraena	-	Brown/tan	-	-	-	10-90/1-22
P. galapagensis	-	Beige (alcohol)	-	Smooth	Soft	126-165/4-8 and 27 92/1.5-4

Table 1. (continued)

	Form	Colour	Oscule Shape	Surface	Consistency	Diods
P. kenyensis	Massive	Buff-cream	-	-	Tough	80-260/2-7
P. zyggompha	Lamellate	Blue(alcohol)	-	Smooth	Cheesy	-
P. halicondriodes	Cake-shaped	Chocolate-brown/ lighter	Small excentrically	Even, pilose	Dense and firm	160/4-5
P. angulospiculatus	Thickly encrusting	Dark brown, grey to yellow or greenish	Flush with surface with rim	Smooth with papillae	Compressible	25-205/1-7
P. japonica	Thin encrusting	Pinkish-white (dry)	Not visible	Smooth	-	40-200/2-8
P. albicans	Thinly to massively	White to ivory,	Circular shaped,	Smooth and	Compressible	12-137.5/1.3-7.5
	encrusting	small purple pathches	slightly elevated	sculptured	but firm	
P. insularis	Thickly encrusting to massive	Chocolate-brown/ light brown	Flush with surface	Smooth	Soft	25-125/1-4
P. microrhabdifera	Massively encrusting	Light brown with dark brown patches	Conspicuous, Circular	Smooth	Compressible	50-138/2-8
P. lita	Thickly encrusting	Black, dark brown, grey brown/brown or grey-brown	Terminal oscules	Smooth, slimy	Soft	23-145/0.5-8.0
P. quasiamphiaster	Thickly encrusting to massive	Reddish-brown, paler	Rounded	Smooth	Compressible	28.7-179.6/0.8-7
P. communis	Thickly encrusting to cushion-shaped	Dark or light brown, khaki- brown, greyish- brown	Flush or elevated	Smooth	Firm to soft	27-143/1-6
P. bergquistae	Thickly encrusting	Light brown to orange(alcohol)	Contracted	Smooth	Firm, cartilaginous	2 types: 91-163/2-6; 202-356/5-11
P. fromontae	Thickly encrusting to cushion-shaped	Black/cream	Contracted	Smooth	Firm, cartilaginous	25-223/2-10
P. hooperi	Thinly encrusting to cushion shaped	Brown-beige	Large, contracted in alcohol	Smooth	Slightly compressible	79-148/2.5
P. myrae	Thinly encrusting to cushion shaped	Light brown	Elevated with rim	Smooth	Soft, compressible	66.6-119.0/2-4
P. edwardsi	Thickly encrusting to massive	Light to dark brown with different patches	Flush with surface	Smooth	Soft, compressible	2 types: 110-128/2.6- 3.0; 22.4-31.1/0.56- 1.09
P. dariae	Thinly encrusting to cushion shaped	Light green with thin brownish patches	Flush with surface	Smooth	Soft, compressible	2 types: 67.3-112.2/1.6-2.8; 30-59.5/0.8-2.1
P. petrupaulensis	Thinly encrusting	Light brown with dark patches/ yellowish cream	Circular or elliptical with rim	Smooth	Firm, cartilaginous, compressible	28-76/1-3
P. spinalis	Thinly encrusting	Grey	Contracted	Irregular, rugose, coriaceous	Cartilaginous	29-103/1-4
P. potiguarensis	Thickly encrusting	Brown with beige patches	Circular, contracted with light rim		Cartilaginous	32-89/1-3
P. clarionensis	Encrusting to cushion shaped	Dark and light brown on the surface and choanosome pale yellow (alcohol)	Circular shaped	Uneven, rugose with rounded bumps	Cartilaginous but internally fleshy	17.5-77.5/1.25-3.0
Plakortis badabaluensis sp. nov.	Thickly encrusting to cushion-shaped	Dark chocolate brown/light brown	Circular with thin rim	Smooth	Soft, fragile	2 types: 64.1-133.1/3.4 6.3; 77.9-116.6/1.3-5.0

Diagnosis: Plakortis was dark chocolate brown in colour externally and light brown internally. The choanosome formed elliptically shaped meshes of approximately 103 × 63 - 325 \times 200 μm in diameter. Ectosome differentiated

with subectosomal lacuna. The ectosome and choanosome were both pigmented. Choanosome confused and vaguely reticulated. Microstrongyloid microrhabd was found to be present.

Description: Sponge is thickly encrusting,

Table 1. (continued)

	Triods	Other spicules	Ectosome	Choanosome	Reference
P. simplex	25-50/3-6	-	Thin fleshy dermis	-	Schulze 1880
P. nigra	absent	-	-	-	Lévi 1953
P. copiosa	18-37/4-4.5	-	-	-	Pulitzer-Finali 1993
P. erythraena	20-25/1-2	-	-	-	Lévi 1958
P. galapagensis	17-36/1.5-4	-	-	-	Desqueyroux-
					Faúndez and Van Soest 1997
P. kenyensis	Rare 30-60	-	-	-	Pulitzer-Finali 1993
P. zyggompha	absent	-	Dense	-	de Laubenfels 1934
P. halicondriodes	absent	-	-	-	Wilson 1902
P. angulospiculatus	Rare,15-80/1-3	-	Distinguished, multi spicular tract	Dense, confused	Carter 1879; Moraes and Muricy 2003
P. japonica	30 × 4	-	Not distinguished	_	Hoshino 1977
P. albicans	Rare-6.3-47.5	-	Alveolar arrangement	Dense, confused	Cruz-Barraza and Carballo 2005
P. insularis	20-50/1-3	-	Loose, confused	Loose, confused	Moraes and Muricy 2003
P. microrhabdifera	absent	Microrhabds, 3-12/1-2	Distinct, 50-250 thick	Confused, dense	Moraes and Muricy 2003
P. lita	1-66/0.5-2.5	Microrhabds, 1-25/0.5- 2.0	Dense, irregular reticulation	Confused	de Laubenfels 1954; Muricy 2011
P. quasiamphiaster	10.5-78.4/1-5	Quasiamphiaster, spheres	Distinct, 100-1012	Dense, confused	Díaz and Van Soest 1994; Muricy 2011
P. communis	30-60/1-5	Spheres	Tangential reticution, well differentiated	Confused or vaguely reticulate	
P. bergquistae	30-122/5-20	-	Reticulate, multispicular tracts	Poorly differentiated, denser	Muricy 2011
P. fromontae	Rare-6-44/0.5-5.0	Spheres	Double reticulation	Confused	Muricy 2011
P. hooperi	17-61/2-3	Microrhabds, 2-8/0.5- 1.0, Spheres	Confused,undifferntiated	Confused	Muricy 2011
P. myrae	17.8-53.5/1.8-3.3	•	Distinct	Dense, confused	Ereskovsky et al. 2013
P. edwardsi	28.1-59.4/2-2.6	Absent	Loose, confused	Dense, confused	Ereskovsky et al. 2013
P. dariae	Rare-20-43.5/2.2- 2.7	Absent	Distinctly reticulate	Confused, reticulate	Ereskovsky et al. 2013
P. petrupaulensis	absent	Microrhabds, 1-18/0.5-3.0, spheres	Disorganized, vaguly reticulate	Disorgnaized	Domingos et al. 2013
P. spinalis	24.7-49.4/1-5		Differentiated, reticulate	Dense, disorgnized	Domingos et al. 2013
P. potiguarensis	22-32/1-3	Microrhabds, 4-25/1-2	Disorganized, single mesh reticulation	Scares, disorgnized	Domingos et al. 2013
P. clarionensis	10-30	Absent	Nondetachable, pigmentated, differentiated	-	Cruz-Barraza et al. 2014
Plakortis badabaluensis sp. nov.	16.1-61.4/1.9-5.2	Microrhabds, 13.3- 43.8/3.8-6.1	Differentiated, dense	Confused with round to elliptical mesh	Present study

cushion shaped, with size up to 10-15 cm wide and 1-2 cm thick. Colour *in vivo* is dark chocolate brown externally and light brown internally (Fig. 2A); when preserved in ethanol the specimen was black in colour externally and light brown internally. The surface is smooth, oscules are circular in shape with thin rim after fixation contracted, and 1-4 mm in diameter. Consistency is soft, fragile, crumbly and easy to tear. Neither exudates nor smell were observed.

Spicules: Diods abundant (Figs. 2B1, B2), 2 size classes- thick and thin. Thick diods are

irregular, smooth, slightly curved, the central region is thick, sinuous, s-bent, and sometimes show a well-developed protuberance; while the endings are acerate 64.1-106.8-133.1/3.4-6.3 μ m (N = 20). Thin diods with significant quantity are almost straight, some are slightly curved in the central region; the endings are pointed 77.9-96.2-116.6/1.3-5.0 μ m (N = 20). Styloid modification is also present in significant number (Fig. 2B3).

However, Triods are rare, have irregular Y-shaped base with sharp to rounded ends. Actines $16.1-37.0-61.4/1.9-5.2 \mu m$ (N = 10; Fig. 2C1).

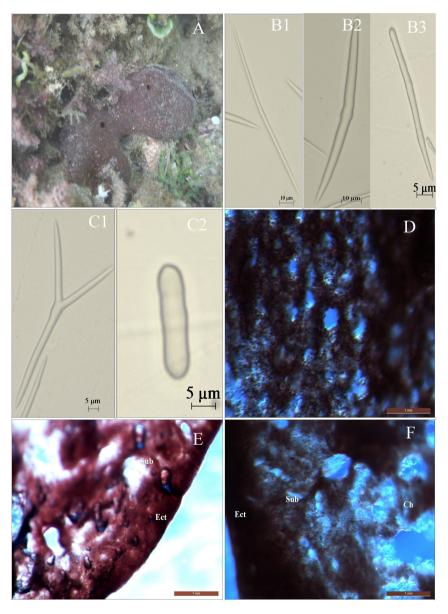


Fig. 2. (A) Specimen *in situ*. (B1) Thin Diods. (B2) Thick Diods. (B3) Styloid shaped Diods. (C1) Y-shaped Triods. (C2) Microrhabds. (D) T.S Showing Ectosomal Meshes. (E) L.S. Showing Ectosome (Ect) and Subectosomal Lacune (Sub). (F) L.S. Showing Ectosome (Ect), Subectosomal Lacune (Sub) and Choanosome (Ch).

Microrhabds are significant in number, smooth, microstrongyloid 13.3-24.3-43.8/3.8-6.1 μ m (N =7; Fig. 2C2).

Skeleton: Ectosomal skeleton: Ectosome is well differentiated, dense and darker than choanosome. In longitudinal sections, the ectosome was observed to be 0.2-0.3 mm thick with dark pink and black colour pigments. In transverse sections, the skeleton is dense with round to elliptical meshes (Figs. 2D, E). Subectosomal lacunae are present and it is higher than wide in shape, 41-208/38-147 μ m (Fig. 2E).

Choanosomal skeleton: It is confused, with round to elliptical meshes and measures 103-325/63-200 μm in diameter. It also shows dark pink and black colour pigmentation (Fig. 2F).

Ecology: This species was found in a bay of the Andaman Sea, which extended to mix with the Bay of Bengal. The whole area has a rocky bottom with patches of sand seen everywhere. The depth of this species occurrence was noticed from is about 3 - 5 m and attached to the rocky outcrop. This sponge was surrounded by corallimorphs, algae, ascidians and corals (Fig. 2A).

Distribution: Badabalu and Pongibalu are situated around the Port Blair town, of the Andaman and Nicobar Islands, India.

Etymology: The name badabaluensis refers to the type locality of the species in Badabalu, Andaman and Nicobar Islands, India.

DISCUSSION

The morphological and anatomical characters of P. badabaluensis sp. nov were compared with all the twenty six species available worldwide under this genera (Table 1). Plakortis badabaluensis sp. nov. is grouped under *Plakortis* with microrhabds, which include 7 species, that is, Plakortis lita de Laubenfels, 1954; Plakortis microrhabdifera Moraes and Muricy, 2003; Plakortis hooperi Muricy, 2011; Plakortis myrae Ereskovsky Lavrov and Willenz, 2014; Plakortis petrupaulensis Domingos, Moraes and Muricy 2013; Plakortis potiguarensis Domingos, Moraes and Muricy 2013; and *Plakortis* spinalis Domingos, Moraes and Muricy 2013. The new species is mostly similar to P. lita in the dark brown colour, thickly encrusting, and with smooth surface. Subectosomal lacunae are present, with choanosomal skeleton forming reticulation with elliptical meshes. Moreover, Diods are abundant, but triods are rare. They differ by the slimy surface and sometimes show large tubes with small

terminal oscules and smaller size of diods (23-145 μm/0.5-0.8 μm), styloid modification is rare and irregular, strongyloid microrhabds is smaller in size (1-25 µm/ 0.5-2.0 µm), while the choanosomal meshes are large (200-600 μm in diameter) (Muricy 2011). P. microrhabdifera is distinguished by light brown colour with exudate, microrhabds are irregularly twisted, triods are absent, but with well-developed tangential alveolar arrangement of spicule tracts in the ectosome (Moraes and Muricy 2003). P. hooperi is thinly encrusting, brownbeige in colour with irregular surface, releases abundant mucous and has acetone like smell, deformed microstrongyles, and its ectosome is undifferentiated (Muricy 2011). P. myrae differed by light brown colour, abundant triods, the microrhabds are irregularly twisted, the presence of choanosome with alveolar arrangement of diods (60-80 μm in diameter) (Ereskovsky et al. 2013). P. petrupaulensis is distinguished by thinly encrusting, light brown in colour, the diods are small (28-76 µm /1-3 µm) and triods are absent, microrhabds are tuberculate, absence of subectosomal lacunae, and the ectosome and choanosome are undifferentiated. P. potiguarensis differed by small diods (37-89 μ m/1-3 μ m), the microrhabds are irregular, and shows absence of subectosomal lacunae. Similarly, P. spinalis is thinly encrusting, gray in colour, the surface is irregular, it is rugose, and coriaceous. The diods are small and with spines (29-103 μ m/ 1-2 μ m), while the microrhabds are highly tuberculate (Domingos et al. 2013).

CONCLUSION

The Andaman and Nicobar group of Islands are highly diverse and are present in the Bay of Bengal, but the phylum Porifera is inadequately revised. Not only poriferans but also other marine organisms are yet to be taxonomically classified.

The genus *Plakortis* is considered cosmopolitan and has very simple identification characters, but its uniformity in external characters made this genus taxonomically very difficult to resolve the species complex. The present study has increased the number of species reported from the Andaman and Nicobar Islands.

List of abbreviations

ZSI/ ANRC- Zoological Survey of India/ Andaman and Nicobar Regional Center.

DOSMB- Department of Ocean Studies and

Marine Biology, Pondicherry University, Port Blair, Andaman and Nicobar

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