Open Access

Population Genetic Structures and Demographic Expansion of the Exotic Jellyfish *Carybdea brevipedalia* in Korean Coasts Inferred from Mitochondrial *COI* Analysis

Buhari Lawan Muhammad¹, Yoseph Seo¹, Jinho Chae², and Jang-Seu Ki^{1,*}

¹Department of Biotechnology, Sangmyung University, Seoul 03016, South Korea.

*Correspondence: E-mail: kijs@smu.ac.kr (Ki). Phone: +82-2-2287-5449, Fax: +82-2-2287-0070. E-mail: buharilawan20@gmail.com (Muhammad); akdldytpq12@gmail.com (Seo) ²Marine Environmental Research and Information Laboratory, Gunpo 15850, South Korea. E-mail: jinhochae@gmail.com (Chae)

Received 28 January 2022 / Accepted 30 May 2022 / Published 11 October 2022 Communicated by James D. Reimer

Carybdea brevipedalia Kishinouye, 1891 is a poisonous jellyfish that usually occurs only in Japanese coastal regions. However, it was recently found on the Korean coast, thus expanding its known geographical range. In this study, we analyzed the population genetics and demographic histories of 113 *C. brevipedalia* specimens from the southern and eastern coastal regions of Korea by sequencing mitochondrial DNA cytochrome *c* oxidase subunit I (*COI*). We identified 42 *C. brevipedalia COI* haplotypes with high genetic diversity and a significant genetic structure. Populations were highly differentiated based on geographic location and distinctly divided into A and B clades. The results of Mantel tests indicated that geographic distance influenced the genetic distance between the two clades. Moreover, demographic analyses (neutrality tests) and the star-like profile of the Templeton, Crandall, and Sing (TCS) haplotype network indicated that *C. brevipedalia* had recently expanded into the southern and eastern coastal regions of Korea. These findings suggest that *C. brevipedalia* populations along the Korean coast have significant genetic differentiation that could be influenced by geographic isolation and subsequent adaptation to regional ecological conditions.

Key words: *Carybdea brevipedalia*, Cytochrome *c* oxidase subunit I (*COI*), Exotic species, Genetic differentiation, Population expansion.

BACKGROUND

The class Cubozoa, commonly called box jellyfish, is one of the smallest groups in the phylum Cnidaria; it comprises the monophyletic orders Carybdeida and Chirodropida and contains at least 49 known species (Collins and Jarms 2021). It has a metagenetic life cycle in which a benthic polyp can reproduce asexually by budding into multiple polyps, each of which metamorphoses into a single medusa (Werner et al. 1971; Toshino et al. 2018). The scientific community has recently become more interested in Cubozoans since their discovery in 1810 due to their unique characteristics such as complex eyes (Coates 2003; Nilsson et al. 2005), exceptional courtship and reproductive behavior (Lewis and Long 2005), superior swimming abilities (Gordon and Seymour 2008), and highly toxic venom (Chung et al. 2001; Kintner et al. 2005) that pose a serious threat to public health, particularly among fishermen and swimmers (Fenner et al. 1996). Various venomous species of box jellyfish such as *Chironex fleckeri* and *Chironex yamaguchii* are

Citation: Muhammad BL, Seo Y, Chae J, Ki JS. 2022. Population genetic structure and demographic expansion of the exotic jellyfish *Carybdea brevipedalia* (Cubozoa, Cnidaria) into the Korean coast. Zool Stud **61:**48. doi:10.6620/ZS.2022.61-48.

largely restricted to the tropical Indo-Pacific region (Fenner and Hadok 2002; Lewis and Bentlage 2009). Nonetheless, various species of box jellyfish are widely distributed in tropical and subtropical oceans (Gershwin and Gibbons 2009; Straehler-Pohl et al. 2017).

Péron and Lesueur (1810) originally identified Carybdea as the first cubozoan, and numerous species of box jellyfishes were assigned to this genus after recent taxonomic revisions (Bentlage and Lewis 2012; Acevedo et al. 2019). Ten species in this genus (Collins and Jarms 2021) are characterized by a heart-shaped rhopaliar niche ostium (Bentlage et al. 2010; Acevedo et al. 2019). Most Carybdea species inhabit warm waters worldwide (Straehler-Pohl et al. 2017). Some species are globally distributed, whereas others are endemic to specific geographical regions (Acevedo et al. 2019). For example, Carybdea rastoni commonly inhabits the warm waters around Hawaii, Australia, Japan, and the Philippines (Scripps Institution of Oceanography 2021). In addition, C. sivickisi inhabits the western Pacific Ocean from Japan to New Zealand (Hoverd 1985). However, some Carybdea species are restricted to specific regions, such as C. branchi in the oceanic areas of Namibia and South Africa (Gershwin and Gibbons 2009; Branch et al. 2010), C. brevipedalia in Japan, and C. marsupialis in the Mediterranean Sea (Acevedo et al. 2019; Rodríguez-García et al. 2021).

Carybdea brevipedalia Kishinouye, 1891 was originally identified in the western coastal waters of Japan (Kishinouye 1891; Uchida 1929 1970) and was misclassified as C. rastoni because of similar morphology (Kramp 1961; Ueno 2003; Nagai 2003). However, recent taxonomic and molecular findings of Cubozoa (Straehler-Pohl et al. 2017; Acevedo et al. 2019) have revealed that the C. rastoni in Japanese coastal waters should be regarded as C. brevipedalia. This species is considered one of the most problematic species among all coastal regions of Japan, especially for fishermen and swimmers (Uchida 1970; Ueno 2003). Their stings can cause severe pain that might be associated with a longer discharge of nematocyst tubules (Kitatani et al. 2015). The poison of this jellyfish has the high hemolytic activity of purified protein toxins (Nagai et al. 2000). Carybdea brevipedalia has historically been distributed only in the coastal waters of Japan (Ueno 2003; Bentlage and Lewis 2012; Acevedo et al. 2019). Despite being undetectable elsewhere, it was recently discovered in Korean coastal waters (Chae et al. 2017) when hundreds of people were stung on the southern coastline and Jeju Island (W. D. Yoon; personal communication). Since then, many such incidents have occurred annually in the southern coastal region (SCR). Although the Korean government recognized the need for studies on countermeasures (Chae et al.

2017), blooms appeared for the first time in the eastern coastal region (ECR) during 2020. This suggested that the geographical range of *C. brevipedalia* is expanding in Korea and probably elsewhere. In the face of increasing socio-economic damage, the *C. brevipedalia* population that has invaded the Korean coast should be investigated to reveal the genetic structure and determine connectivity between recently emerged populations in the Korean SCR and ECR. In addition, understanding the genetic structure and phylogeography of this jellyfish species is imperative to predict its origin and geographical expansion patterns for effective bloom management.

Mitochondrial genes such as cytochrome coxidase subunit I (COI), cytochrome B (cvtB), and 16S rDNA (Glynn et al. 2016; Seo et al. 2021a) have been investigated to determine the population genetics and phylogeography of jellyfish. In particular, COI is a powerful molecular marker of the genetic variation and structure of populations in various organisms (Palraju et al. 2018; Choi et al. 2020). This is because it has the advantages of rapid evolution, elevated polymorphism, and is easily amplified and sequenced (Hu et al. 2008; Xu et al. 2011; Palraju et al. 2018). To date, 89 DNA sequences identified from Carybdea nuclear rDNA and mitochondrial COI listed in the National Center for Biotechnology Information (NCBI) database have mostly been used for evolutionary studies of box jellyfish (Collins 2002; Bentlage et al. 2010). However, the population genetics of box jellyfish have not been investigated using these sequences.

Therefore, we analyzed the mitochondrial *COI* gene to reveal the population genetic structure and phylogeographic profiles of *C. brevipedalia* populations sampled from the SCR and ECR of Korea. We also assessed genetic relationships between native and recently emerged populations in the SCR and ECR. We resolved the expansion profiles and genetic differentiation among *C. brevipedalia* populations along the Korean coast.

MATERIALS AND METHODS

Sampling and morphological identification

Carybdea brevipedalia specimens were collected in August 2020 from four different coastal regions located in the southern coastal region (SCR; St. 1 and 2) and eastern coastal region (ECR; St. 3 and 4) of Korea (Fig. 1A). A total of 113 specimens (41 at St. 1, 12 at St. 2, 40 at St. 3, and 20 at St. 4) were collected using a hand-net, immediately fixed with 100% ethanol and transported to the laboratory where they were stored at 4°C for further analysis.

Morphological observations (Fig. 1B) were carried out following Chae et al. (2017). In particular, we observed the morphology using a high-resolution camera (D810; Nikon, Tokyo, Japan) with close-up lenses (Micro-Nikkor 60 mm f2.8; Nikon and Makroplanar 100 mm f2.0; Carl Zeiss, Oberkochen, Germany) and a close-up tube (PK-13, Nikon, Tokyo, Japan).

DNA extraction, amplification and sequencing

Preserved specimens were washed individually with distilled water to remove all ethanol, and then genomic DNA (gDNA) was extracted using the modified cetyltrimethylammonium bromide (CTAB) DNA extraction protocol (Richards et al. 1994).

Polymerase chain reaction (PCR) amplifications of *COI* fragments were done using a newly designed primer pair (Cb-F1 5'-GTTCTACAAACCAC AAAGATATAGG-3', and Cb-R1 5'-TATGGCTAACA TAGCATAAACCAT-3'). In brief, the reaction solution (total volume: 20 μ L) was prepared for the PCR amplification. It consisted of 2 μ L template DNA, 2 μ L Ex Taq buffer, 2 μ L dNTP mix, 1 μ L forward primer, 1 μ L reverse primer, 0.2 μ L Ex Taq Polymerase (TaKaRa Bio Inc, Shiga, Japan), and 11.8 μ L of distilled water. PCR reaction conditions were as follows: 94°C for 3 min, followed by 40 cycles at 94°C for 30 s, 55°C for 30 s, and 72°C for 60 s, and finally 72°C for 5 min. All PCR amplified samples were confirmed in a 1% agarose gel using MIDORI green dye (Nippon Genetics Europe GmbH, Düren, Germany) as a fluorescent source. Confirmed PCR products were purified using the PCR Cleanup S & V Kit (Bionics, Seoul, Korea) according to the manufacturer's instructions.

Purified PCR amplicons were sequenced by Bionics Inc. (Seoul, Korea). DNA sequencing was performed using the BigDyeTM Terminator v3.1 Cycle Sequencing Kit (Thermo Fisher Scientific), and the synthesized sequences were analyzed using the

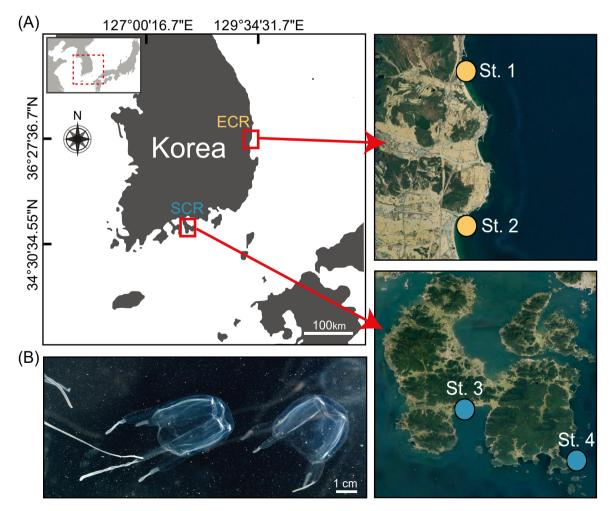


Fig. 1. A map showing four sampling locations (St. 1, St. 2, St. 3, and St. 4; (A) of *C. brevipedalia* specimens (B) collected from the southern coastal regions (SCR) and eastern coastal regions (ECR) of Korea.

Applied Biosystems[™] 3730xl DNA Analyzer (Applied Biosystems, Foster City, CA). Editing and contig assembly of the *COI* sequence fragments were carried out using Sequencher v5.1 (Gene Codes, Ann Arbor, MI). All the sequences determined in this study were deposited into the GenBank database with accession numbers (OM108321–OM108433).

Phylogenetic analysis

To determine the taxonomic relationship between C. brevipedalia and other jellyfishes, we constructed maximum likelihood (ML) and Bayesian inference (BI) analyses based on the 113 COI sequences determined in this study and 27 COI sequences of other medusozoans (including cubozoans, scyphozoans, and hydrozoans) retrieved from the GenBank database. The ML analysis was inferred using MEGA X (Kumar et al. 2018) with the General Time-Reversible model (GTR + I + G) and 1,000 bootstrap values. BI analyses were built with MrBayes v3.2.6 (Huelsenbeck and Ronquist 2001) using the GTR model with a gamma distribution for the remaining sites. One million generations were run until the standard deviation of the split frequencies was < 0.01. Trees were sampled every 1,000 generations, with a burn-in of 250 trees. The output file containing trees with posterior probabilities (PP) was shown in FigTree v1.4.4 (http://tree.bio.ed.ac.uk/). The tree resulting from the ML analysis with bootstrap values (BS) was compared with the BI tree with posterior probability, and the 50% majority-rule consensus tree was summarized with PP and BS as nodal support. The tree was edited using Adobe Illustrator CS6 (Adobe Systems, San Jose, CA, USA). A list of all the sequences used for the phylogenic analysis is provided in supplementary material tables S1 and S3.

Estimates of genetic diversity and differentiation

The assembled 113 *COI* sequences (689 bp) of *C. brevipedalia* were used to calculate the standard genetic diversity indices, including the number of haplotypes (Nh), the number of polymorphic sites (Ps), haplotype diversity (*h*), and nucleotide diversity (π) using Arlequin v.3.5.2.2 (Excoffier and Lischer 2010). In addition, neutrality, Tajima's *D* (Tajima 1989), and Fu's *F*_s tests (Fu 1997) were performed to reveal molecular evidence for past demographic changes using DnaSP v.6 (Rozas et al. 2017). To determine genetic variation among different clusters, the 113 *COI* sequences were imported into Arlequin v.3.5.2.2 where Analysis of Molecular Variance (AMOVA) was assessed. In addition, a pairwise *F*_{ST} statistical analysis (with 10,000 permutations; Wright 1969) was used to test for genetic differentiation among populations across two different regions.

TCS haplotype network and phylogenetic analyses

To explore the phylogeographic patterns of the C. brevipedalia haplotypes in the Korean coasts, we constructed a Templeton, Crandall, and Sing (TCS) haplotype network (Clement et al. 2002) and a Maximum-Likelihood (ML) phylogenetic tree for the 113 COI sequences. The haplotype network was constructed using PopART v.1.7 (Leigh and Bryant 2015) with default settings, while the maximum likelihood (ML) and Bayesian inference (BI) phylogenetic trees were constructed in MEGA X and MrBayes v3.2.6, respectively, with a General Time-Reversible model (GTR + I + G) and 1000 bootstrap values. The tree was visualized with Mega X Tree Explorer and was edited in Adobe Illustrator CS6 (Adobe Systems, San Jose, CA). The haplotype frequency of the 42 unique haplotypes is presented in supplementary table S2.

Furthermore, we used the non-parametric Mantel test to investigate the relationship between genetic distance and geographic distance (Peakall and Smouse 2012). It was evaluated using two distance measures between all 42 distinct haplotype pairs. In the Mantel test, the pairwise genetic distance between specimens was calculated as the number of nucleotide differences between the *COI* sequences. In addition, the pairwise geographic distance was calculated as a direct physical distance based on their longitudinal and latitudinal coordinates that were transformed logarithmically. The Mantel test was conducted using GenAlEx v.6.5 with statistical significance derived based on 1,000 permutations (Peakall and Smouse 2012).

RESULTS

Identification and phylogenetic relationships of *C. brevipedalia*

We confirmed the identities of 113 box jellyfish (*C. brevipedalia*) collected from four sites in the SCR and ECR of Korea according to their evident morphology (Fig. 1B). A sharp cylindrical bell with an almost flat apex was slightly narrower and rounder towards the end. The box-shaped part of the bell in adult specimens averaged 3.3 (2.6–4.2) cm in height and four tentacles were attached to each corner.

The molecular identity of *COI* fragments was impossible to determine because *C. brevipedalia COI*

sequences were not available in any public databases. Therefore, we sequenced 113 COI fragments of 689-bp from C. brevipedalia for the first time and inferred phylogenetic relationships among cubozoans, with an emphasis on Carybdea species (Fig. 2). Our phylogenetic tree showed that the cubozoans were separated from Scyphozoans and Hydrozoans and that they formed a monophyletic clade with 87% bootstrap support, but not by posterior probability (PP) < 50%. The order Carybdeida in the Cubozoan clade formed polyphyletic clades with the order Chirodropida nested between the Carybdeida clades. The Alatina, Carybdea, Morbakka, and Tamoya genera differed from one another. The C. brevipedalia sequences determined herein clustered together and were clearly separated from those of the Carybdea species, C. arborifera, and C. xavmacana, with moderate support (PP 69% and BS 55%).

Genetic diversity and neutrality test

The genetic diversity assessed in 113 *COI* sequences identified 53 polymorphic sites (Ps) and 42 unique haplotypes across all populations (Table 1). The estimated haplotype (*h*) and nucleotide (π) diversities were 0.8428 ± 0.0248 and 0.005139 ± 0.002915, respectively. The genetic diversity (*h* and π) was higher for SCR than ECR (0.6944 and 0.004132 vs. 0.6691 and 0.001573, respectively).

The neutrality of the *COI* haplotypes of all populations was tested. The Tajima *D* (-2.03, p = 0.003) and Fu F_s (-26.092, p = 0.000) were negative and statistically significant for the overall population. The Tajima *D* values and Fu F_s scores for the ECR and SCR

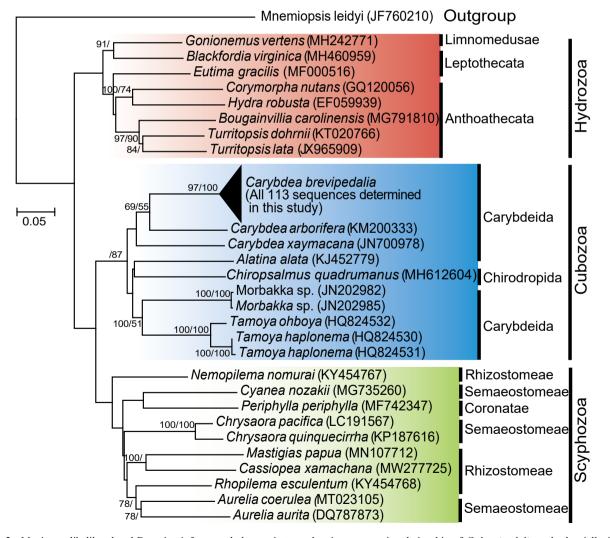


Fig. 2. Maximum likelihood and Bayesian inference phylogenetic tree, showing taxonomic relationship of *C. brevipedalia* and other jelly fish species. Numbers separated by a slash above each branch are posterior probabilities (on the left side) followed by bootstrap values (on the right side). Nodal support of less than 50% is not shown.

were significantly negative (Table 1).

Genetic differentiation

The population differentiation was determined using analyses of molecular variance (AMOVA). Most genetic variation (59.95%, p < 0.001) occurred between the regions, whereas 40.05% occurred within the entire population (Table 2). The fixation index ($F_{\rm ST}$), calculated to assess population differentiation caused by genetic structure, was 0.599 (p = 0).

Phylogeographic structures of C. brevipedalia

The phylogenetic profiles of C. brevipedalia COI haplotypes on the Korean coast were determined using the TCS haplotype network and phylogenetic analysis (Fig. 3). The results showed that the 42 unique haplotypes detected herein were differentiated into supported clades A and B based on their geographical locations. The TCS network revealed that clade A comprised 18 haplotypes unique to the SCR, with C01 being the most frequent in 33 individuals (Fig. 3A). Clade B contained 19 haplotypes, of which 17 and 2 (C23 and C24) were unique to the ECR and SCR, respectively. Haplotype C25 in clade B served as the intermediate haplotype connecting the two clades and was shared between the SCR and ECR. Four haplotypes (C19-22) unique to SCR were separated from both clades by 4-8 mutational steps and did not belong to either. Clades A and B were shaped like stars in the TCS network, in which most haplotypes were linked to those that were the most frequent (C01 in SCR and C30 in ECR) with one or two mutational steps, indicating recent population expansion. The unrooted phylogenetic tree similarly revealed two distinct clades with four intermediate haplotypes (CB19–22) that were clearly distinguishable (Fig. 3B). Table S2 shows the frequency and distribution of the 42 unique haplotypes.

Mantel correlation analyses of genetic and geographic distances uncovered a significant positive correlation ($R^2 = 0.128$, p = 0.001; Fig. 4).

DISCUSSION

Genetic structures of C. brevipedalia

Several outbreaks of jellyfish and the organisms that caused them to spread globally due to ocean currents and anthropogenic activities have recently been reported (Condon et al. 2012). For example, C. brevipedalia is a toxic jellyfish species native to Japanese coastal waters that has recently been identified in Korean coastal waters where it has caused socioeconomic damage (Chae et al. 2017; Acevedo et al. 2019). Population genetic analyses can help to determine the invasion and transmission routes of harmful marine organisms such as jellyfish Aurelia and ascidians (Dawson 2005; Zhan et al. 2010 2015). This is fundamental to understanding the population dynamics of certain species (Kingsford et al. 2021). We found high genetic diversity with significant differentiation in the overall C. brevipedalia population along the

 Table 1. Genetic diversity indices and neutrality test for mitochondrial COI of 113 specimens of Carybdea brevipedalia populations

| Location | N | Nh | Ps | Tajima's D | Fu's $F_{\rm s}$ | h | π |
|---------------------------------------------------------------|----------|----------|----------|----------------------|-------------------------|-----------------------------------------------------------------------|-------------------------------------------------------------------------------|
| Eastern coastal region (ECR) Southern coastal region (SCR) | 53 60 | 18 25 | 20 39 | -2.38*** -2.182** | -18.56*** -16.763*** | $\begin{array}{c} 0.6691 \pm 0.0709 \\ 0.6944 \pm 0.0682 \end{array}$ | $\begin{array}{c} 0.001573 \pm 0.001166 \\ 0.004132 \pm 0.002446 \end{array}$ |
| Total | 113 | 42 | 53 | -2.03** | -26.092*** | 0.8428 ± 0.0248 | 0.005139 ± 0.002915 |

Abbreviations: N, Number of sample; Nh, Number of haplotype; Ps, Number of polymorphic site; D, Tajima's D; Fs, Fu's F_s ; h, Haplotype diversity; π , Nucleotide diversity; **, p < 0.05; ***; p < 0.001.

 Table 2. Analysis of molecular variance (AMOVA) for populations of Carybdea brevipedalia in Korean coasts based on mitochondrial COI

| Source | d.f. | Sum of squares | Variance components | Percentage of variation | Fixation indices |
|-------------------|------|----------------|---------------------|-------------------------|-----------------------------------|
| Among Populations | 1 | 86.507 | 1.51897 | 59.95% | $F_{\rm ST} = 0.59947$ (p = 0) |
| Whole Populations | 111 | 112.652 | 1.01488 | 40.05% | (p - 0) |

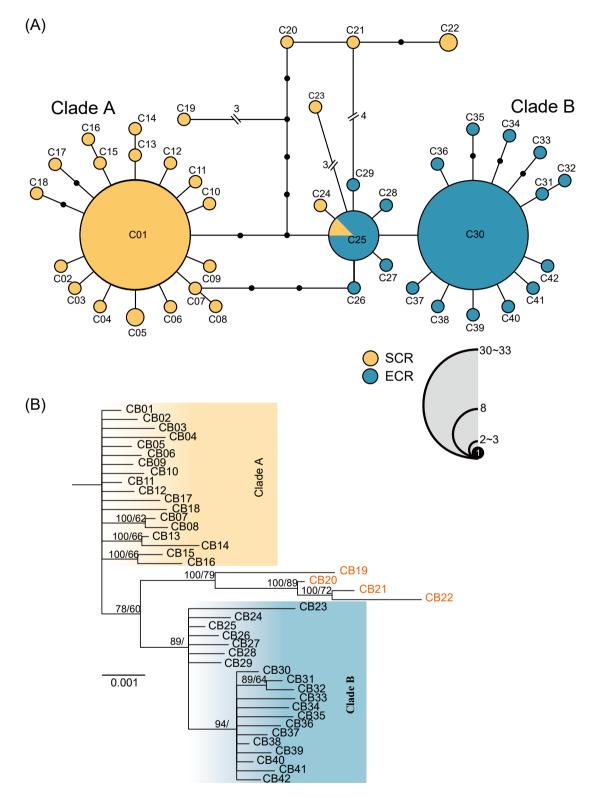


Fig. 3. The haplotype TCS network (A) and Bayesian inference and maximum likelihood phylogenetic tree (B) show phylogeography and relationships among the 42 *COI* haplotypes of *C. brevipedalia* collected from the southern (SCR; orange colors) and eastern coastal regions (ECR; blue colors) of Korea. Each circle on the haplotype network represents individual haplotypes, and the size of the cycle is the proportion of haplotype frequency. The colors indicate the geographical origin of the haplotypes. Each line between haplotypes represents a 1-nucleotide mutational change. Small black dots indicate unsampled haplotypes. On the phylogenetic tree, numbers on branches are Bayesian posterior probabilities/bootstrap support values.

Korean coast. This is consistent with previous findings, which showed that genetic differentiation is commonly intraspecific in marine jellyfish (*e.g.*, Holland et al. 2004; Dawson 2005; Ramšak et al. 2012; Lee et al. 2013). Similar *COI* genetic variation and significant genetic structure were also found in the jellyfish *Aurelia coerulea* (Seo et al. 2021a b) and the sea squirt *Ciona savignyi* (Yi and Kim 2020). Our results showed that the Korean *C. brevipedalia* population also consists of two genetically distinct populations.

Genetic differentiation among relatively distant populations is primarily caused by geographical isolation and adaptation to the ecological conditions of specific regions (Coyne and Orr 2004; Zhan et al. 2009; Liu et al. 2019). The Mantel test results of the COI sequences (Fig. 4) showed that genetic differentiation and gene flow between the populations might be greatly influenced by the geographical distance between the Korean SCR and ECR. This agrees with the findings of other Korean marine organisms, such as the Asian shore crab (Hong et al. 2012), ascidians (Kim et al. 2012; Yi and Kim 2020), barnacles (Yoon et al. 2013), and disk abalone (Nam et al. 2021). The Asian shore crab (Hemigrapsus sanguineus), for example, lacks geographically-associated haplotypes and a genetic structure within and among populations in the Korean coasts. However some degree of genetic differentiation occurred between populations of the ECR and other coastal regions (Hong et al. 2012). A population of disk abalone (Haliotis discus) sampled from the ECR was also genetically separated from those in the SCR and in western coastal regions (WCR) (Nam et al. 2021). These findings suggested strong genetic structures among marine creatures, including C. brevipedalia along the Korean coast (Hong et al. 2012; Kim et al. 2012; Yoon

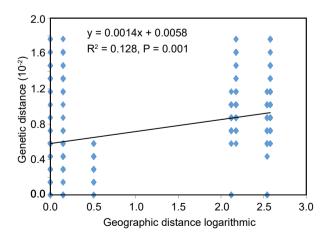


Fig. 4. Mantel test showing relationships between genetic distance and the logarithm of geographic distance between all pairs of 42 *COI* haplotypes of *C. brevipedalia* populations from two coastal regions of Korea.

et al. 2013; Yi and Kim 2020, and the present study). In addition, genetic structures might have been influenced by geographical distance and other environmental factors, such as ocean current, temperature, and salinity (Hong and Cho 1983; Rebstock and Kang 2003).

Introduction and demographic expansion of *C. brevipedalia* on Korean coasts

The first blooms of C. brevipedalia in Korea were recently identified in the SCR (Chae et al. 2017). However, whether the species was recently introduced or native to the Korean coast has remained unknown. In general, a significant geographic structure is considered a major distinguishing factor between native and recently introduced populations (Hellberg et al. 2002; Geller et al. 2010; Zhan et al. 2010). Geographically differentiated populations can reflect species persistence in a region, which would allow for sufficient gene flow isolation and the accumulation of distinct genetic variability (Avise et al. 1987; Yi and Kim 2020), whereas the introduced population is usually genetically homogeneous (Zhan et al. 2010). However, diverse sources might result in genetic variations within introduced populations (Pineda et al. 2016). Here, we applied the Tajima D and Fu F_s neutrality tests to test the hypothesis that C. brevipedalia has recently emerged and expanded along the Korean coast. These tests are generally based on the distribution of pairwise differences between sequences within populations and have been implemented to detect population growth (Ramos-Onsins and Rozas 2002; De Jong et al. 2011). Positive and negative values indicate bias towards intermediate-frequency and rare alleles, respectively, and the negative value is a sign of recent population expansion (Tajima 1989; Fu 1997). The Tajima D and Fu $F_{\rm s}$ values determined herein were negative in the overall population and significantly so in the SCR and ECRs. These findings implied recent population expansion in the overall population, as well as in both SCR and ECR.

The recent demographic expansion closely corresponded with the *COI* haplotype network. The profiles of the SCR and ECR haplotypes (Fig. 3A) were both star-shaped and their compositions were similar. The star-like patterns reflect a recent appearance and rapid population growth (Slatkin and Hudson 1991; Rogers and Harpending 1992; Avise 2000; Froufe et al. 2016). The phylogenetic tree (Fig. 3B) was also consistent with the haplotype network that showed the connectivity in the population and that the ancestral clade was A. In addition, the high haplotype (0.6691) and lower nucleotide diversity (0.001573) in the ECR indicated that the populations might have rapidly emerged from a small effective ancestral population (Grant and Bowen 1998; Avise 2000; De Jong et al. 2011). The profiles of population expansion were similar in the Asian shore crab *Hemigrapsus sanguineus* along the Korean coast (Hong et al. 2012). Considering these findings, our results suggested that *C. brevipedalia* populations recently appeared and thrived in Korean coastal regions, thus increasing population in these regions (Avise 2000; Froufe et al. 2016).

Inflow pathway of *C. brevipedalia* into Korean coasts

The genetic relationships between the Korean and Japanese *C. brevipedalia* populations remain unknown because *COI* sequences for the latter are not available. However, considering the present results, oceanographic characteristics, and earlier findings (Hong et al. 2012; Kim et al. 2012; Yoon et al. 2013; Yi and Kim 2020; Nam et al. 2021), the emergence and expansion of *C. brevipedalia* in the Korean coasts could be explained as follows (Fig. 5). The Tsushima Warm Current branches off the Kuroshio Current (Fig. 5A) and is a factor for the emergence and/or expansion of marine species. It is characterized by high water temperatures and high densities (Rebstock and Kang 2003). Some parts of the

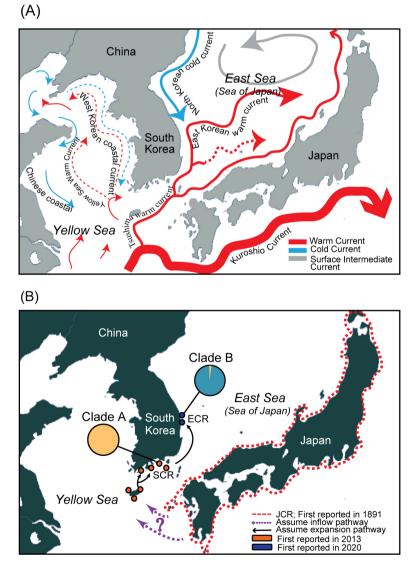


Fig. 5. A major ocean current flow pattern in Korean waters (A) and a possible scenario for the introduction and expansion of *C. brevipedalia* in the Korean coasts (B). The big cycles represent the haplotype compositions sampled in the present study, while the small cycles indicate the current distribution of *C. brevipedalia* in Korean waters. The big orange cycle represents clade A haplotypes while the blue cycle represents clade B haplotypes. Moreover, the orange color in clade B represents the shared haplotypes between southern coastal regions (SCR) and eastern coastal regions (ECR). JCR; Japanese coastal regions.

current run into the Yellow Sea, and the main part flows into the East Sea along the Korean Peninsula (Hong and Cho 1983; Rebstock and Kang 2003). The Tsushima Warm Current might have transported *C. brevipedalia* from its native habitat in Japan to Jeju Island and to the Korean SCR (Fig. 5B). However, we inferred from the present findings that the dispersal and gene flow of *C. brevipedalia* from the Korean SCR to the ECR are highly restricted. Nevertheless, due to the genetic drift (founder effect) caused by water currents and/or humanrelated vectors, new populations were established in the ECR, most likely by haplotype C25, which was shared between the SCR and ECR and intermediate between the two populations. As a result, haplotype C30 adapted to the ECR then rapidly disseminated.

The ECR population was located at the sub-polar front, where the North Korean cold (low temperature with relatively low salinity) and East Korean warm (high temperature, high salinity) currents converge (Rebstock and Kang 2003). Temperature and salinity influence the metamorphosis of polyps into medusae in *Carybdea* spp. (Canepa et al. 2014; Toshino et al. 2018). Thus, the balanced effects of cold and warm water currents in the Eastern Sea might explain the genetic differentiation of *C. brevipedalia* between the ECR and SCR populations. Such hydrographic differentiation profiles can change the dispersal and population structures of *C. brevipedalia* and other marine species on the Korean coast (Hong et al. 2012; Kim et al. 2012; Yoon et al. 2013; Nam et al. 2021).

Our demographic history results support the recent appearance and rapid expansion of *C. brevipedalia* populations in Korean coastal waters. We presented a possible scenario for the introduction and profile of the gene flow of this species in these environments. However, this perspective remains ambiguous, and genetic parameters could not be directly compared because molecular data for *C. brevipedalia* from its native region are not available. Further sampling from Japan is necessary to clarify the historical influences and to determine the population connectivity and inflow pathways of this species in Korean coastal waters. Understanding population connectivity is critical for risk assessment and to design appropriate management strategies for invasive species (Hampton et al. 2004).

CONCLUSIONS

We analyzed the population genetic structure and demographic history of *Carybdea brevipedalia* populations using specimens that have massively bloomed in Korean coastal waters. Our results revealed high genetic diversity and strong genetic differentiation between geographically distant populations (SCR and ECR), most likely caused by geographical isolation and other ecological and hydrographic factors. The demographic history indicated that the C. brevipedalia populations in Korea underwent a recent population expansion. Our findings of the first genetic analysis of C. brevipedalia populations could serve as baseline data for future phylogeographic and demographic studies, especially in the native and other recently emergent regional populations. Furthermore, our findings provide a more in-depth understanding of the population structure of this species in Korea, thus aiding in the management of these potentially lethal organisms. Future studies should broaden the sampling to include native populations to compare genetic characteristics, which could provide useful information about how the species was introduced and its general gene flow profiles.

Acknowledgments: We thank Dr. S. Abassi for the critical comments and English editing on the early version of the manuscript. This research was supported by a part of the project titled "Improvement of management strategies on marine disturbing and harmful organisms" and a part of the project titled "Development of hull adherent organism management technology (20210651)" funded by the Ministry of Oceans and Fisheries, Korea.

Authors' contributions: Muhammad BL: analyses, writing original draft, writing-review & editing. Seo Y: conceptualization, investigation, analyses. Chae J: Investigation. Ki JS: Conceptualization, Writing review & editing.

Competing interests: The authors declare no conflict of interest.

Availability of data and materials: All the data presented in this study are available in the supporting information (Supplementary materials). All sequences determined were deposited into the GenBank database under the accession numbers OM108321–OM108433.

Consent for publication: Not applicable.

Ethics approval consent to participate: Not applicable.

REFERENCES

Acevedo MJ, Straehler-Pohl I, Morandini AC, Stampar SN, Bentlage B, Matsumoto GI, Yanagihara A, Toshino S, Bordehore C, Fuentes VL. 2019. Revision of the genus *Carybdea* (Cnidaria: Cubozoa: Carybdeidae): clarifying the identity of its type species *Carybdea marsupialis*. Zootaxa **4543:**515–548. doi:10.11646/ zootaxa.4543.4.3.

- Avise JC. 2000. Phylogeography: the history and formation of species. Harvard University Press, Cambridge, UK.
- Avise JC, Arnold J, Ball RM, Bermingham E, Lamb T, Neigel JE, Reeb CA, Saunders NC. 1987. Intraspecific phylogeography: the mitochondrial DNA bridge between population genetics and systematics. Annu Rev Ecol Evol Syst 18:489–522. doi:10.1146/ annurev.es.18.110187.002421.
- Bentlage B, Cartwright P, Yanagihara AA, Lewis C, Richards GS, Collins AG. 2010. Evolution of box jellyfish (Cnidaria: Cubozoa), a group of highly toxic invertebrates. Proc Royal Soc B 277:493–501. doi:10.1098/rspb.2009.1707.
- Bentlage B, Lewis C. 2012. An illustrated key and synopsis of the carybdeid box jellyfishes (Cnidaria: Cubozoa: Carybdeida), with emphasis on the "Irukandji family" (Carukiidae). J Nat Hist 46:2595–2620. doi:10.1080/00222933.2012.717645.
- Branch GM, Branch ML, Griffiths CL, Beckley LE. 2010. Two Oceans: a guide to the marine life of southern Africa. Random House Struik; (New edn). ISBN 978-1-77007-772-0.
- Canepa A, Purcell JE, Belmar MB, Acevedo M, Gentile M, Fuentes V. 2014. Salinity effects on asexual reproduction of *Carybdea* sp. (Cnidaria: Cubozoa). J Plankton Res 36:585–590. doi:10.1093/ plankt/fbt124.
- Chae J, Yoon WD, Kim B, Ki JS. 2017. First record of box jellyfish, *Carybdea brevipedalia* (Cnidaria: Cubozoa: Carybdeidae) from Korean coastal waters: morphology and molecular descriptions. Anim Syst Evol Divers **33:**8–16. doi:10.5635/ASED.2017.33.1. 059.
- Choi EH, Kim G, Cha SH, Lee JS, Ryu SH, Suk HY, Lee YS, Baek SY, Hwang UW. 2020. Molecular phylogenetic, population genetic and demographic studies of *Nodularia douglasiae* and *Nodularia breviconcha* based on *CO1* and *16S rRNA*. Sci Rep 10:1–14. doi:10.1038/s41598-020-72015-5.
- Chung JL, Ratnapala L, Cooke IM, Yanagihara AA. 2001. Partial purification and characterization of a hemolysin (CAH1) from Hawaiian box jellyfish (*Carybdea alata*) venom. Toxicon **39:**981–990. doi:10.1016/s0041-0101(00)00237-3.
- Clement M, Snell Q, Walke P, Posada D, Crandall K. 2002. TCS: estimating gene genealogies. *In*: Proc. 16th Int Symp Parallel Distrib Comput **2**:184.
- Coates MM. 2003. Visual ecology and functional morphology of Cubozoa (Cnidaria). Integr Comp Biol 43:542–548. doi:10.1093/ icb/43.4.542.
- Collins AG. 2002. Phylogeny of Medusozoa and the evolution of cnidarian life cycles. J Evol Biol 15:418–432. doi:10.1046/ j.1420-9101.2002.00403.x.
- Collins AG, Jarms G. 2021. World List of Cubozoa. Cubozoa. Accessed through: World Register of Marine Species at: https:// www.marinespecies.org/aphia.php?p=taxdetails&id=135219. Accessed on 20 Aug. 2021.
- Condon RH, Graham WM, Duarte CM, Pitt KA, Lucas CH, Haddock SH, Sutherland KR, Robinson KL, Dawson MN, Decker MB, Purcell JE, Malej A, Mianzan H, Uye S, Gelcich S, Madin PL, Mills CE. 2012. Questioning the rise of gelatinous zooplankton in the world's oceans. BioScience 62:160–169. doi:10.1525/ bio.2012.62.2.9.
- Coyne JA, Orr HA. 2004. Speciation. Sinauer Associates, Sunderland, pp. 1–545.
- Dawson MN. 2005. Incipient speciation of *Catostylus mosaicus* (Scyphozoa, Rhizostomeae, Catostylidae), comparative phylogeography and biogeography in south-east Australia. J Biogeogr **32:**515–533. doi:10.1111/j.1365-2699.2004.01193.x.

- De Jong MA, Wahlberg N, Van Eijk M, Brakefield PM, Zwaan BJ. 2011. Mitochondrial DNA signature for range-wide populations of *Bicyclus anynana* suggests a rapid expansion from recent refugia. PLoS ONE 6:e21385. doi:10.1371/journal.pone.0021385.
- Excoffier L, Lischer HE. 2010. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. Mol Ecol Resour 10:564–567. doi:10.1111/j.1755-0998.2010.02847.x.
- Fenner PJ, Hadok JC. 2002. Fatal envenomation by jellyfish causing Irukandji syndrome. Med J Aust **177:3**62–363. doi:10.5694/ j.1326-5377.2003.tb05109.x.
- Fenner PJ, Williamson JA, Burnett JW. 1996. Clinical aspects of envenomation by marine animals (Abstract). Toxicon **34:**145.
- Froufe E, Prié V, Faria J, Ghamizi M, Gonçalves DV, Gürlek ME, Sousa R. 2016. Phylogeny, phylogeography, and evolution in the Mediterranean region: news from a freshwater mussel (*Potomida*, Unionida). Mol Phylogenet Evol **100:**322–332. doi:10.1016/ j.ympev.2016.04.030.
- Fu YX. 1997. Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. Genetics 147:915–925. doi:10.1093/genetics/147.2.915.
- Geller JB, Darling JA, Carlton JT. 2010. Genetic perspectives on marine biological invasions. Annu Rev Mar Sci 2:367–393. doi:10.1146/annurev.marine.010908.163745.
- Gershwin LA, Gibbons MJ. 2009. Carybdea branchi, sp. nov., a new box jellyfish (Cnidaria: Cubozoa) from South Africa. doi:10.11646/zootaxa.2088.1.5.
- Glynn F, Houghton JD, Bastian T, Doyle TK, Fuentes V, Lilley MK, Provan J. 2016. High-resolution genetic analysis reveals extensive gene flow within the jellyfish *Pelagia noctiluca* (Scyphozoa) in the North Atlantic and Mediterranean Sea. Biol J Linn Soc **117**:252–263. doi:10.1111/bij.12654.
- Gordon MR, Seymour JE. 2008. Quantifying movement of the tropical Australian cubozoan *Chironex fleckeri* using acoustic telemetry. Hydrobiologia 616:87–97. doi:10.1007/s10750-008-9594-7.
- Grant WAS, Bowen BW. 1998. Shallow population histories in deep evolutionary lineages of marine fishes: insights from sardines and anchovies and lessons for conservation. J Hered 89:415– 426. doi:10.1093/jhered/89.5.415.
- Hampton JO, Spencer PBS, Alpers DL, Twigg LE, Woolnough AP, Doust J. 2004. Molecular techniques, wildlife management and the important of genetic population structure and dispersal: a case study with feral pigs. J Appl Ecol 41:735–743. doi:10.1111/ j.0021-8901.2004.00936.x.
- Hellberg ME, Burton RS, Neigel JE, Palumbi SR. 2002. Genetic assessment of connectivity among marine populations. B Mar Sci 70:273290.
- Holland BS, Dawson MN, Crow GL, Hofmann DK. 2004. Global phylogeography of *Cassiopea* (Scyphozoa: Rhizostomeae): molecular evidence for cryptic species and multiple invasions of the Hawaiian Islands. Mar Biol 145:1119–1128. doi:10.1007/ s00227-004-1409-4.
- Hong CH, Cho K. 1983. The northern boundary of the Tsushima Current and its fluctuations. J Oceanol Soc Korea 18:1–9.
- Hong SE, Kim JK, Yu JN, Kim KY, Lee CI, Hong KE, Park KY, Yoon MG. 2012. Genetic variation in the Asian shore crab *Hemigrapsus sanguineus* in Korean coastal waters as inferred from mitochondrial DNA sequences. Fish Aquat Sci 15:49–56. doi:10.5657/FAS.2012.0049.
- Hoverd WA. 1985. Occurrence of the order Cubomedusae (Cnidaria: Scyphozoa) in New Zealand: collection and laboratory observations of *Carybdea sivickisi*. New Zeal J Zool 12:107– 110. doi:10.1080/03014223.1985.10428267.
- Hu J, Zhang JL, Nardi F, Zhang RJ. 2008. Population genetic structure

of the melon fly, *Bactrocera cucurbitae* (Diptera: Tephritidae), from China and Southeast Asia. Genetica **134:**319. doi:10.1007/s10709-007-9239-1.

- Huelsenbeck JP, Ronquist FR. 2001. MRBAYES: Bayesian inference of phylogenetic trees. Bioinformatics 17:754–755. doi:10.1093/ bioinformatics/17.8.754.
- Kim WJ, Lee CI, Kim HS, Han HS, Jee YJ, Kong HJ, Nam BH, Kim YO, Kim KK, Kim BS, Lee SJ, Hong KE, Yu JN, Yoon MG. 2012. Population genetic structure and phylogeography of the ascidian, *Halocynthia roretzi*, along the coasts of Korea and Japan, inferred from mitochondrial DNA sequence analysis. Biochem Syst Ecol 44:128–135. doi:10.1016/j.bse.2012.04.020.
- Kintner AH, Seymour JE, Edwards SL. 2005. Variation in lethality and effects of two Australian chirodropid jellyfish venoms in fish. Toxicon 46:699–708. doi:10.1016/j.toxicon.2005.07.015.
- Kingsford MJ, Schlaefer JA, Morrissey SJ. 2021. Population Structures and Levels of Connectivity for Scyphozoan and Cubozoan Jellyfish. Diversity 13:174. doi:10.3390/d13040174.
- Kishinouye K. 1891. Zwei neue Medusen von Charybdea (Ch. brevipedalia n. sp., Ch. latigenitalia n. sp.). Dobutsugaku Zasshi 3:437–440. (in Japanese with German diagnoses)
- Kitatani R, Yamada M, Kamio M, Nagai H. 2015. Length is associated with pain: jellyfish with painful sting have longer nematocyst tubules than harmless jellyfish. PLoS ONE 10:e0135015. doi:10.1371/journal.pone.0135015.
- Kramp PL. 1961. Synopsis of the medusae of the world. Mar Biol Ass UK 40:1–469. doi:10.1017/S0025315400007347.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. Mol Biol Evol 35:1547–1549. doi:10.1093/molbev/ msy096.
- Lee PL, Dawson MN, Neill SP, Robins PE, Houghton JD, Doyle TK, Hays GC. 2013. Identification of genetically and oceanographically distinct blooms of jellyfish. J R Soc Interface 10:20120920. doi:10.1098/rsif.2012.0920.
- Leigh JW, Bryant D. 2015. POPART: full-feature software for haplotype network construction. Methods Ecol Evol 6:1110– 1116. doi:10.1111/2041-210X.12410.
- Lewis C, Bentlage B. 2009. Clarifying the identity of the Japanese Habu-kurage, *Chironex yamaguchii*, sp. nov. (Cnidaria: Cubozoa: Chirodropidae). Zootaxa **2030**:59–65. doi:10.11646/zootaxa. 2030.1.5.
- Lewis C, Long TAF. 2005. Courtship and reproduction in *Carybdea sivickisi* (Cnidaria: Cubozoa). Mar Biol **147**:477–483. doi:10.1007/s00227-005-1602-0.
- Liu Y, Dietrich CH, Wei C. 2019. Genetic divergence, population differentiation and phylogeography of the cicada *Subpsaltria yangi* based on molecular and acoustic data: an example of the early stage of speciation? BMC Evol Biol **19:**1–17. doi:10.1186/ s12862-018-1317-8.
- Nagai H. 2003. Recent progress in jellyfish toxin study. J Health Sci 49:337–340. doi:10.1248/jhs.49.337.
- Nagai H, Takuwa K, Nakao M, Ito E, Miyake M, Noda M, Nakajima T. 2000. Novel proteinaceous toxins from the box jellyfish (sea wasp) *Carybdea rastoni*. Biochem Biophys Res Commun 275:582–588. doi:10.1006/bbrc.2000.3353.
- Nam BH, Kim H, Seol D, Kim H, Noh ES, Kim EM, Kwak W. 2021. Genotyping-by-Sequencing of the regional Pacific abalone (*Haliotis discus*) genomes reveals population structures and patterns of gene flow. PLoS ONE 16:e0247815. doi:10.1371/ journal.pone.0247815.
- Nilsson D, Gislen L, Coates MM, Skogh C, Garm A. 2005. Advanced optics in a jellyfish eye. Nature 435:201–205. doi:10.1038/ nature03484.

Palraju M, Paulchamy R, Sundaram J. 2018. Population genetic

- Peakall R, Smouse PE. 2012. GenAlEx 6.5: Genetic analysis in Excel. Population genetic software for teaching and research— An update. Bioinformatics 28:2537–2539. doi:10.1093/ bioinformatics/bts460.
- Péron F, Lesueur C. 1810. Tableau des caractères génériques et spécifiques de toutes les espèces de méduses connues jusqu'à ce jour. Ann Mus Hist Nat Paris **14:**325–366.
- Pineda MC, Lorente B, Lopez-Legentil S, Palacín C, Turon X. 2016. Stochasticity in space, persistence in time: Genetic heterogeneity in harbour populations of the introduced ascidian *Styela plicata*. PeerJ **4**:e2158. doi:10.7717/peerj.2158.
- Ramos-Onsins SE, Rozas J. 2002. Statistical properties of new neutrality tests against population growth. Mol Biol Evol 19:2092–2100. doi:10.1093/oxfordjournals.molbev.a004034.
- Ramšak A, Stopar K, Malej A. 2012. Comparative phylogeography of meroplanktonic species, *Aurelia* spp. and *Rhizostoma pulmo* (Cnidaria: Scyphozoa) in European Seas. *In*: Pitt KA, Lucas CH (ed) Jellyfish blooms. Springer, Dordrecht, pp. 69–80. doi:10.1007/s10750-012-1053-9.
- Rebstock GA, Kang YS. 2003. A comparison of three marine ecosystems surrounding the Korean peninsula: Responses to climate change. Prog Oceanogr 59:357–379. doi:10.1016/ j.pocean.2003.10.002.
- Richards E, Reichardt M, Rogers S. 1994. Preparation of genomic DNA from plant tissue. Curr Protoc Mol Biol **27:**2–3. doi:10.1002/0471142727.mb0203s27.
- Rodríguez-García C, Sanz-Fernández V, Muñoz-Lechuga R, Gutiérrez-Martínez M, Cabrera-Castro R. 2021. First records and presence over time of the cubozoan *Carybdea marsupialis* (Linnaeus, 1758) on the southwestern Spanish coast (Northeast Atlantic). Reg Stud Mar Sci 44:101712. doi:10.1016/j.rsma.2021.101712.
- Rogers AR, Harpending H. 1992. Population growth makes waves in the distribution of pairwise genetic differences. Mol Biol Evol 9:552–569. doi:10.1093/oxfordjournals.molbev.a040727.
- Rozas J, Ferrer-Mata A, Sánchez-DelBarrio JC, Guirao-Rico S, Librado P, Ramos Onsins SE, Sánchez-Gracia A. 2017. DnaSP 6: DNA sequence polymorphism analysis of large data sets. Mol Biol Evol 34:3299–3302. doi:10.1093/molbev/msx248.
- Scripps Institution of Oceanography. 2021. Carybdea rastoni Zooplankton Guide. Available at: http://sio-legacy.ucsd.edu/ zooplanktonguide/species/carybdea-rastoni. Accessed on 5 Nov. 2021.
- Seo Y, Muhammad BL, Chae J, Ki JS. 2021a. Population genetic structures of the jellyfish *Aurelia coerulea* polyps on Korean coasts and implications as revealed by Mitochondrial *COI*. Zool Stud **60:**63. doi:10.6620/zs.2021.60-63.
- Seo Y, Muhammad BL, Chae J, Ki JS. 2021b. Genetic Structure and Diversity of the Moon Jellyfish *Aurelia coerulea* Polyp Population in Jaran Bay, Korea, Revealed by Mitochondrial *COI* and 16S rRNA Genes. Ocean Sci J 56:99–105. doi:10.1007/ s12601-021-00008-0.
- Slatkin M, Hudson RR. 1991. Pairwise comparisons of mitochondrial DNA sequences in stable and exponentially growing populations. Genetics 129:555–562. doi:10.1093/genetics/129.2.555.
- Straehler-Pohl I, Matsumoto GI, Acevedo MJ. 2017. Recognition of the Californian cubozoan population as a new species— *Carybdea confusa* n. sp. (Cnidaria, Cubozoa, Carybdeida). Plankton Benthos Res 12:129–138. doi:10.3800/pbr.12.129.
- Tajima F. 1989. Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. Genetics **123:**585–595. doi:10.1093/genetics/123.3.585.
- Toshino S, Miyake H, Shibata H. 2018. Development of Carybdea

brevipedalia Kishinouye, 1891 (Cnidaria: Cubozoa: Carybdeida: Carybdeidae) collected from northern Japan. Plankton Benthos Res **13**:116–128. doi:10.3800/pbr.13.116.

- Uchida T. 1929. Studies on the Stauromedusae and Cubomedusae, with special reference to their metamorphosis. Jpn J Zool 2:103–193.
- Uchida T. 1970. Revision of Japanese Cubomedusae. Publ Seto Mar Biol Lab 17:289–297. doi:10.5134/175610.
- Ueno S. 2003. *Carybdea rastoni* Haacke. *In*: Tohru, Y. (ed.) Sea UFO jellyfish Occurrence, Ecology and Countermeasures. Kouseishakouseikaku, Tokyo, Japan, pp. 206. (in Japanese)
- Werner B, Cutress CE, Studebaker JP. 1971. Life cycle of *Tripedalia cystophora* Conant (Cubomedusae). Nature 232:582–583. doi:10.1038/232582a0.
- Wright S. 1969. Evolution and the genetics of populations, Volume 2: The theory of gene frequencies. University of Chicago Press, Chicago, pp. 520.
- Xu ZH, Chen JL, Cheng DF, Liu Y, Frederic F. 2011. Genetic variation among the geographic population of the grain aphid, *Sitobiona venae* (Hemiptera: Aphididae) in China inferred from mitochondrial *COI* gene sequence. Agric Sci China 10:1041– 1048. doi:10.13057/biodiv/d200933.
- Yi CH, Kim W. 2020. Assessing cryptic invasion state: fine-scale genetic analysis of *Ciona savignyi* population in putative native habitat of the Korean Coast. Ocean Sci J 55:99–113. doi:10.1007/s12601-019-0041-7.
- Yoon M, Jung JY, Kim DS. 2013. Genetic diversity and gene flow patterns in *Pollicipes mitella* in Korea inferred from mitochondrial DNA sequence analysis. Fish Aquat Sci 16:243– 251. doi:10.5657/FAS.2013.0243.
- Zhan A, Briski E, Bock DG, Ghabooli S, MacIsaac HJ. 2015. Ascidians as models for studying invasion success. Mar Biol 162:2249–2470. doi:10.1007/s00227-015-2734-5.
- Zhan A, Hu J, Hu X, Zhou Z, Hui M, Wang S, Bao Z. 2009. Fine-scale population genetic structure of Zhikong scallop (*Chlamys farreri*): do local marine currents drive geographical differentiation? Mar Biotechnol 11:223–235. doi:10.1007/ s10126-008-9138-1.
- Zhan A, Macisaac HJ, Cristescu ME. 2010. Invasion genetics of the *Ciona intestinalis* species complex: from regional endemism to global homogeneity. Mol Ecol **19:**4678–4694. doi:10.1111/ j.1365-294X.2010.04837.x.

Supplementary materials

Table S1. List of all the *Carybdea brevipedalia COI*

 sequences determined in this study. (download)

Table S2. Frequency and distribution of the 42 unique haplotypes detected in the Eastern coastal region (ECR) and Southern coastal region (SCR) of South Korea. (download)

Table S3. List of other jellyfish species accessions retrieved from the GenBank database and used in the phylogenetic analysis. (download)